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# CHEMISTRY AND SOME BIOLOGICAL ACTIVITIES OF BENZOQUINONES OF EMBELIA SCHIMPERI.

ΒY

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A thesis submitted in partial fulfilment for the Degree of Master of Science (Chemistry) of the University of Nairobi. UNIVERSITY OF NAIROBI LIBRARY II P. O. Box 30197 NAIROBI

This thesis is my original work and has not been presented for a degree in any University.

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# OUR BELOVED DAUGHTERS, CHEBET AND CHEPNG'ETICH.

## THIS THESIS IS DEDICATED TO MY BELOVED WIFE, LINNER AND

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#### ABSTRACT.

Embelia schimperi and Embelia keniensis are among the five medicinal Myrsinaceae plants found in Kenya which find a wide range of application in ethnopharmacology as antimicrobials and anthelmintics. The berries and root bark of <u>E</u>. schimperi were extracted with cold ethyl acetate and found to contain mostly benzoquinones. The level of benzoquinones was found to be higher in the berries relative to root bark. Chromatographic separation of the bioassay selected fractions on oxalic acid impregnated silica gel (soaking in 3% oxalic acid in methanol) using various solvent systems for <u>E</u>. schimperi led to the isolation and identification of embelin (10), methylvilangin (21), myrsinaquinone (56), decylvilangin (76) and decylanhydrovilangin (77), all of which are benzoquinones. <u>Embelia keniensis</u> was only screened (with aid of TLC analysis) for the presence of the benzoquinone pigments and found to contain trace amounts of embelin (10). All these structures were established using physical and spectroscopic (UV, MS, <sup>1</sup>H, <sup>13</sup>C and 2-D NMR) data. Two compounds were not characterized.

Biological activity tests such as insect anti-feedant, brine shrimp lethality, larvicidal and anti-microbial were carried out with the pure compounds.

Insect anti-feedant tests were determined with <u>Locusta migratoria</u> using a concentration of 100  $\mu$ g/ml of each sample which was applied on sucrose treated Whatman No.1 filter paper. In the control experiments, the filter papers had sucrose only. Embelin (10) and 21 were found to have relative anti-feedant percentage (RAP) of 96% and 33% respectively.

The larvicidal tests were done on mosquito larvae, <u>Aedes aegvpti</u>. Compound 21 was the only one tested for this activity and was found to be a growth retardant.

Brine shrimp lethality test was done for 21, 76, KCP-02 and KCP-06. The  $LC_{50}$  values were found to be 120, 51, 54 and 0  $\mu$ g/ml respectively.

Stored products pests test was performed using <u>Sitophilus zeamais</u> (maize weevil) and <u>Anthoscelides obtectus</u> (bean weevil) for embelin (10) and it was found that the total number of emerged progeny and reproductibility of the two insects were reduced significantly at all concentrations.

Anti-microbial activity test was performed using <u>Candida albicans</u>. <u>Trichophyton</u> <u>metagrophyte</u>, <u>Microsporium gypsum</u> and <u>Escherichia coli</u> for 10, 21, 56, 76, 77, KCP-02 and KCP-06 and it was found that all the compounds had less than significant activity against the micro-organisms.

#### CHAPTER ONE.

#### 1.0.0. INTRODUCTION.

In the tropical third world, crop losses either in the field or in storage are tremendous and measures must be taken to effectively protect field and harvested crop. In Kenya it is estimated that 30 % of the crops are lost due to the pest attack (KARI, 1991). Such pests include insects, fungi, bacteria. nematodes and grazing higher animals. For this reason, integrated pest management is desirable. Among the methods used include introduction of natural predators (biological control), pesticides use, crop rotation and introduction of pest resistant varieties.

A survey conducted in 1989 indicated that the usage of pesticides based on their cash value is concentrated in the developed world with US accounting for 24%, Western Europe 24%, Far east 9%, Eastern Europe and the then USSR 8% while the whole of Africa and the rest of the third world was only 7% (Cremlyn, 1991). This is an unfortunate situation since these latter countries contain 49% of the world's population and therefore need the greatest protection of crops. On a world scale, pests destroy about half the annual crop but in Africa and most other parts of the third world these losses may be up to 70% of everything produced; despite the need to accelerate food production to cater for the ever increasing population in these countries (Chander and Ahmed.1983).

In Kenya, pesticide cost in 1989 was approximately 42 million Kenya pounds (KARI, 1989). This represented quite a big foreign exchange allocation yet overall this was still not adequate. Nearly all the conventional pesticides for the control of field crops and their respective stored products are manufactured in the developed world. These pesticides are therefore expensive. For this reason the use of pesticides in Kenya and other third world countries is still low. In view of this, self-reliance in agro-chemical development and production is the only way out for Kenya and other third world countries.

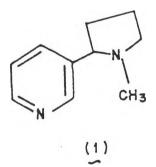
Furthermore, petrochemical based pesticides are potential health hazards due to toxic residues. The main petrochemical based pesticides include organochlorides, organoarsenicals, dinitrophenols, organic thiocyanates.

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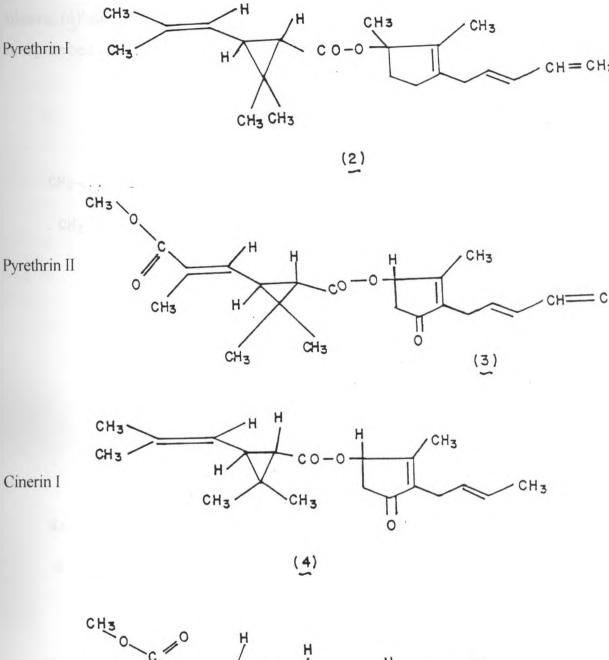
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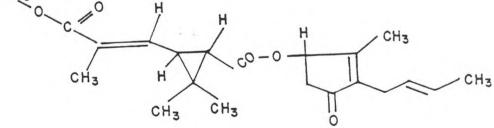
organophosphorus and carbamate compounds. The organo-phosphates and carbamate compounds, even though devoid of persistence, are highly toxic and need to be applied under strict supervision by trained manpower, an aspect which is lacking in the third world.

Over the years man has used a large number of plants as insecticides. These plants contain natural deterrent principles which have and can still be utilized to develop environmentally friendly pesticides. Among the botanical insecticides still in use today include pyrethrum, avermeetins, derris (rotenone) and nicotine. Nicotine (1), an active principle in tobacco, is an alkaloid which exists naturally as a salt of citric and malic acids (Fuchs and Shroeder, 1983). It is only the levorotatory natural nicotine that is an insecticidal principle. Although this insecticide which is used against aphids, capsids, leaf minor, codling moth and thrips is non persistent contact insecticide, it has high mammalian toxicity and therefore its use is declining.



Pyrethrin is a contact insecticide obtained from the flower heads of <u>Chrvsanthemum cinerariafolium</u>. Pyrethrum is safe for household use because it has low mammalian toxicity and has no persistence in addition to the tremendous knockdown characteristics on flying insects (Woods, 1974). Pyrethrins are the active principles in pyrethrum and are four in number (2-5).

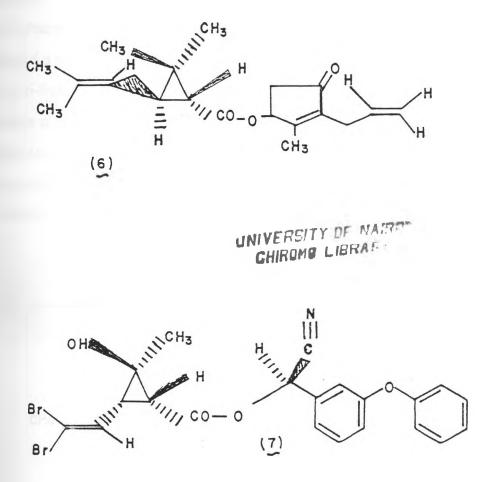




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Cinerin II

Natural models of pyrethroids have been used to develop synthetic ones. Allethrin ( $\underline{6}$ ) and decamethrin ( $\underline{7}$ ) which have strong insecticidal properties are among the best examples (Martin and Woodcock, 1983).

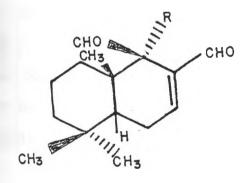


More plants with efficacious properties are being searched and identified. These properties are anti-feedant or feeding deterrents, repellents and pesticidal effects. Anti-feedants are ecologically appropriate since they only cut down on rather than eliminate insect population and therefore leaving useful non pest insects alone.

The Indian Neem tree. <u>Azadirachta indica</u> which grows in Kenya is a higher plant which is known to be resistant to many varieties of insects. Complex tetranor triterpenoids have been isolated from it (Ley, 1985). These include the

most active anti-feedant against lepidopterous species known, azadirachtin. Azadirachtin is a highly complex molecule and cannot economically be produced by synthetic process. <u>Azadirachta indica</u> produces enough quantities of this compound and its analogues and there are programs to propagate it in several parts of the world.

The Meliaceae triterpenoids are apparently intrinsically anti-feedant. <u>Melia</u> <u>volkensii</u> ( a species indigenous to Kenya) and <u>Melia azedarach</u> have also proved to be anti-feedant towards <u>Schistocerca gregaria</u> (Mwangi, 1985). Less complex chemicals also show anti-feedant activity. (-) - Polygodial (§) from <u>Polygonum</u> <u>hvdropiper</u> and warbuganal (9) from <u>Warbugia ugandensis</u> are typical simple azadirachtin mimicking substances; they are effective as anti-aphids and anti-African army worm.

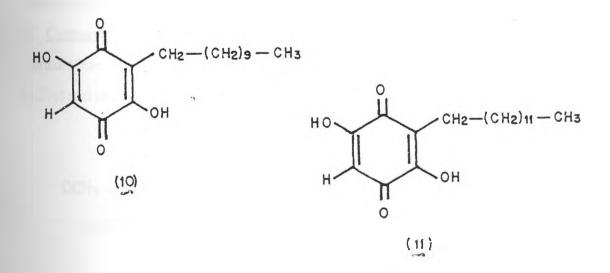


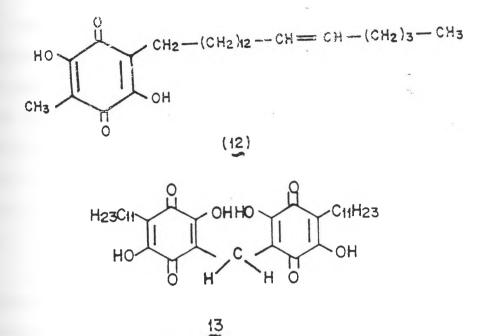
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(9) R = OH

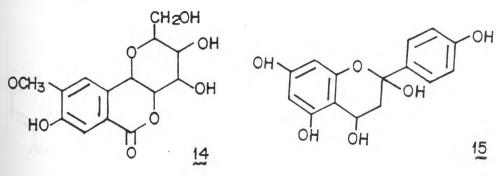
#### 1.0.1. LITERATURE REVIEW

The Myrsinaceae plants are distributed all over the world. The family has received extensive chemical investigations since the beginning of the century. It has been pointed out that the presence of alkylated hydroxybenzoquinone derivatives and a number of triterpenoid based on oleanane/ursen skeleton are a chemotaxonomical characteristics of this family (Ogawa and Natori 1968). Embelin (10), rapanone (11), maesaquinone (12) and bisbenzoquinone, vilangin (13) are among the long chain alkyl-1,4-benzoquinones isolated from some Myrsinaceae species occurring in Japan (Ogawa and Natori, 1968). A chemical study of Embelia ribes was done by Paranipe and Gokhale (1932). They isolated embelin (10) as the major constituent. Four years later, Krishna and Varma (1936) reported the isolation of the same compound, embelin, from petroleum ether and chloroform extracts of Myrsine africana berries. In a separate study, Kawamura and Hokoku (1937) isolated an orange crystalline substance from the bark and woody portions of Rapanea maximowiczii (Koidz) and named it rapanone (11). Earlier, Stather (1931) screened Myrsinaceae plants of Rapanea and Myrsine genera for their tannin compositions. It was discovered that in the stem bark of Rapanea and /or Myrsine, the percentage of tannin was 8-9%.



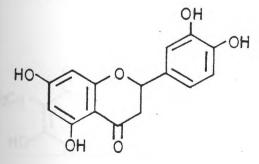


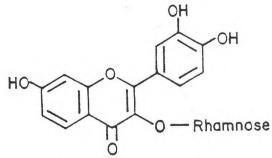
A chemical investigation of <u>Maesa japonica</u> (Moritzii), a plant found in Japan revealed the presence of maesaquinone (12) up to 1.5 % yield (Harimoto, 1939). Similarly from <u>Myrsine africana</u> seeds thought to be originating from British Somaliland (present day Djibouti) but probably originating from Abyssinia (present day Ethiopia) was found to contain 4.8% of embelin. Merian and Schlittler (1948) also reported the isolation of rapanone from <u>Rapanea neurophylla</u>. <u>E. kilimandscharica</u> and <u>M. africana</u>. In a separate report rapanone(11) together with bergenin (14) and leucopelargonidin (15) have been isolated from the roots of <u>Connarus monocarpus</u>, an ever-green shrub belonging to the family Connaraceae. The roots were found to constitute a very good source of rapanone (1.2%) and of bergenin (1.5%).

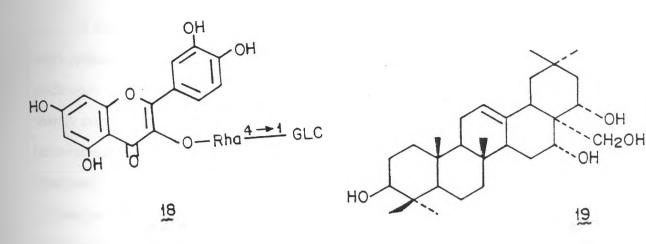


Paris and Rabenoro (1950) carried out a phytochemical evaluation of two Madagascar Myrsinaceae plants namely; <u>E</u>. <u>berbevana</u> and <u>Maesa emirnensis</u>. Their study revealed the presence of sterols, traces of essences, a bitter principle, tannins, saponins and quinones. The fruits were found to contain up to 2% (w/w) embelin. A similar study performed using various parts of <u>R</u>. <u>pulchra</u> showed the existence of embelin up to 0.39% (w/w) in the berries, rapanone 2.8% (w/w) in the root and bark while the stem bark was composed of 1.2% embelin (Wilkinson, 1961).

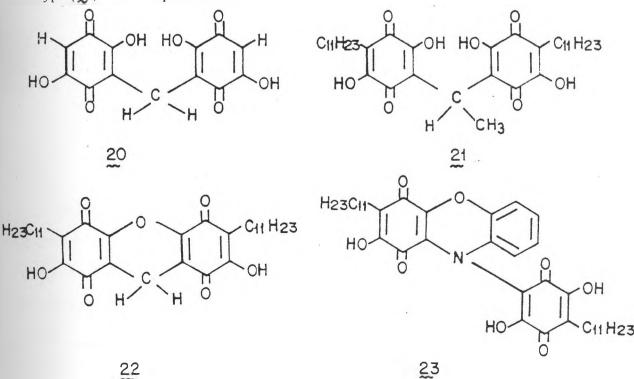
In India, <u>Embelia ribes</u> has been regarded as one of the most important crude drugs of indigenous medicine since ancient times. The berries of this plant known as "Vidanga" are recommended for relieving headache, rhinitis, hemicrania, epilepsy and insomnia. As part of the study on the chemistry of this plant. a new benzoquinone pigment vilangin (13) (a name taken from the vernacular Telugu name "Viyuvilanga") together with embelin have been isolated from the plant. The new compound constitution was confirmed by synthesis and structural modifications (Rao and Venkateswarlu, 1961). In another study, Cambie and Couch (1967) reported the isolation of vilangin and (+)-quercitol (§) from the flowers of <u>Myrsine australis</u>, a Myrsinaceae plant common to both northern and southern parts of New Zealand. Further chemical investigation of the same plant showed that embelin (10) and two leucoanthocyanins were present in the fruits while from the leaves quercetin (16), (17), rutin(18) and triterpenoid (19) were isolated.



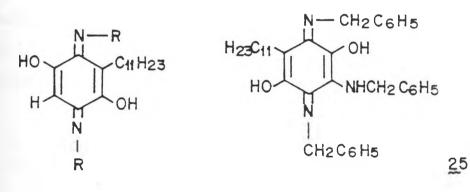


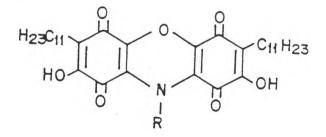


Rao and Venkateswarlu (1962) in a separate publication communicated the synthesis of vilangin by condensing 2,5-dihydroxy-1,4-benzoquinone with formaldehyde to give methylene-bis (2,5-dihydroxy-3,6- benzoquinone) (20) which was subsequently alkylated using dilauroyl peroxide in dioxane solution to give the vilangin. In addition to this, they also revealed that embelin undergoes condensation reactions with various aldehydes to give analogues of vilangin (21) and anhydrovilangin (22). They further noted that with acetic, propionic and benzaldehydes both products are obtained and only the anhydro (22) was achieved with the case of other aromatic aldehydes. In the case of salicylaldehyde, a product of type (23) was also produced.

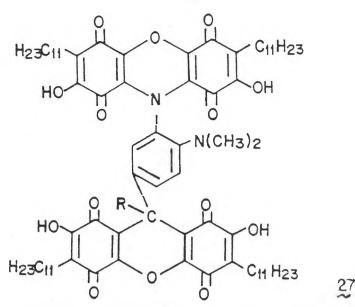


It has also been reported that embelin undergoes condensation reactions with primary amines forming the corresponding quinone di-imine (24) which undergoes decomposition in boiling water or with concentrated HCl, forming mainly polymeric products and affording only traces of embelin. In the case of benzylamine two products are obtained; the normal quinone di-imine (24) and 3-benzylamino (bis desoxy-bis(benzylimino)-embelin (25). Further condensation reactions of embelin (2 moles) with various nitroso compounds (one mole) to give corresponding N-bis (anhydrobenzoquinone) (26) was also communicated (Rao and Venkateswarlu, 1964). In the condensation with p-dimethylamino-m-nitrosobenzaldehyde, a more complicated reaction takes place with the formation of (27) (Jeney and Sohnai, 1955).

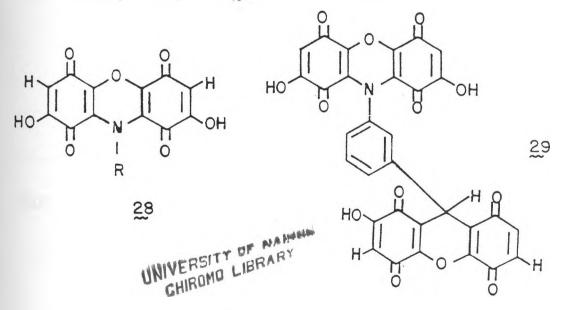




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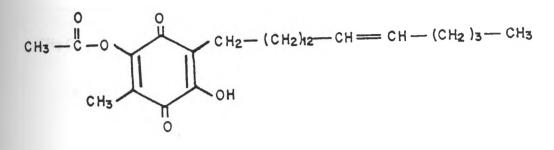


While continuing with their study, Rao and Venkateswarlu (1964) extended their work to 2,5-dihydroxy-1,4-benzoquinones which readily underwent condensation with nitroso compounds producing the corresponding N-bis (anhydro-2,5-dihydroxy-3,6-benzoquinone) (28). Similarly with p-dimethyl-m-nitrosobenzaldehyde compound (29) was readily formed.

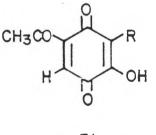


In Japan, the eleven species of Myrsinaceae plants were grouped into three genera namely, <u>Ardisia</u>, <u>Myrsine</u> and <u>Maesa</u>. They grow chiefly in Southern Japan (Ohwi, 1964). A discovery of the biological importance of benzoquinone derivatives , mostly associated with the family, (Morton, 1965) led Ozawa<u>et al</u> (1964) and Ogawa and Natori (1968) to work on the biochemical examinations of hydroxybenzoquinone derivatives and the screening of the distribution of these

secondary metabolites among the Japanese Myrsinaceae plants. As a result embelin (10), rapanone (11) maesaquinone (12) acetylmaesaquinone (30), 2-hydroxy-5methoxy-3-pentadecenyl (tridecenyl, tridecyl)-benzoquinone (31) and bis(benzoquinonyl)-olefines (ardisaquinones) (32), (33) and (34) were isolated. The other methoxy derivatives 35,36,37 and the pigments 32,33, and 34 were found to be restricted to <u>Ardisia</u> species (Ogawa <u>et al.</u> 1968 a,b).

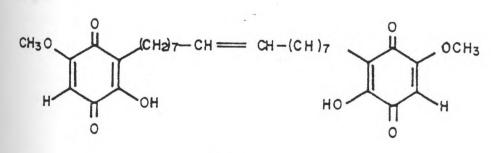


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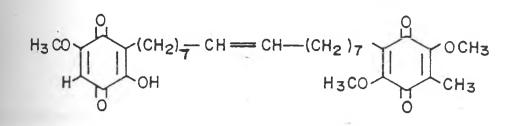


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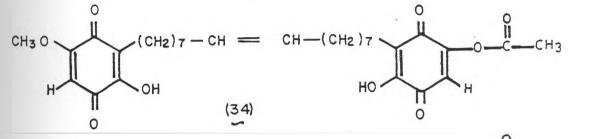
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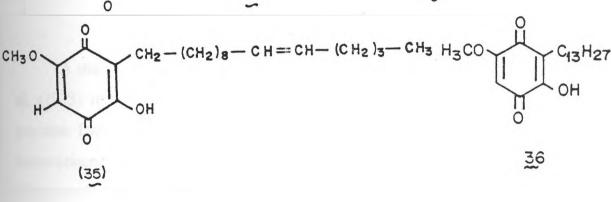


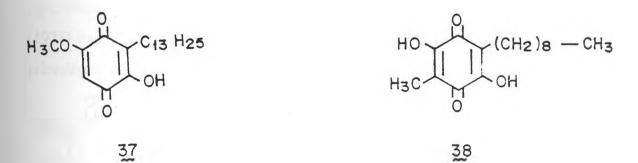
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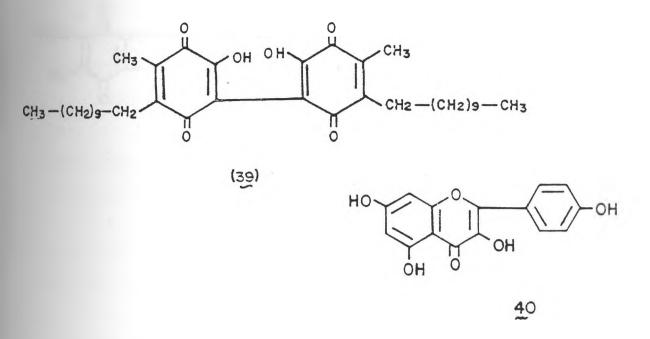




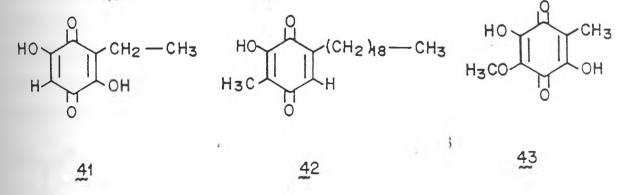


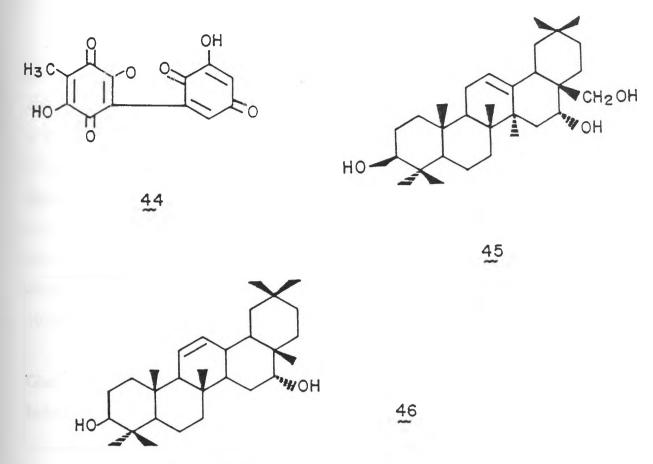


A Nepalese <u>Maesa macrophylla</u>. Wall, a plant commonly used in the remedy of fever, cough and ulcers, was found to contain a new benzoquinone pigment, 2,5-dihydroxy-3-methyl-6-nonyl-1.4-benzoquinone (38) up to 2.5 % (w/w) in the leaves (Chandrasekhar <u>et al.</u> 1970). Further chemical analysis by Prabu <u>et al.</u> (1971) showed the presence of another novel benzoquinone, macrophyllin (39) along with (16) and kaempferol (40) in the leaves.



A study on <u>Rapanea umbellata</u>, a Brazilian Myrsinaceae plant, by Bauer <u>et</u> <u>al</u>, (1973) revealed the presence of embelin (10) as the chief benzoquinone pigment. Dallacker and Lohnert (1972) reported the synthesis of a number of benzoquinone pigments and confirmed their structures which included 3,6dihydroxy-2-ethyl-1,4-benzoquinone (41), dihydromaesaquinone (42), spinulosin (43), oesporein (44), embelin (10), rapanone (11) and vilangin (13). Heltz <u>et al</u>, (1973) found the shoots and leaves of <u>Embelia concinna</u> contain the triterpene primulagenin (45) and embeliagenin (46).

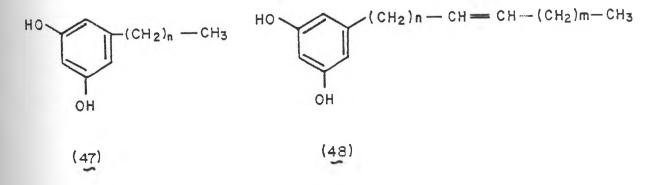




In India, Desai <u>et</u>. <u>al</u> (1975) in the course of their chemical investigations of many families of the Indian flora for their biological activities. isolated maesaquinone (12) from Maesa indica. and embelin (10) from Connarus ritchiei.

Another study of a Taiwanese plant, <u>Maesa formosana</u> by Russel <u>et al</u>, (1976) revealed (12) and acetylmaesaquinone (30) as constituents.

Phenolic compounds have also been isolated from fruits of the Myrsinaceae plants. These compounds especially those with side chains are biologically active. Madrigal <u>et al.</u> (1977) reported the isolation and characterization of a series of such phenolic compounds from <u>Rapanea</u> <u>laetevirens</u>. Examples of such compounds are 5-n-alkylresorcinols (47) and 5-n-alkenylresorcinols (48).



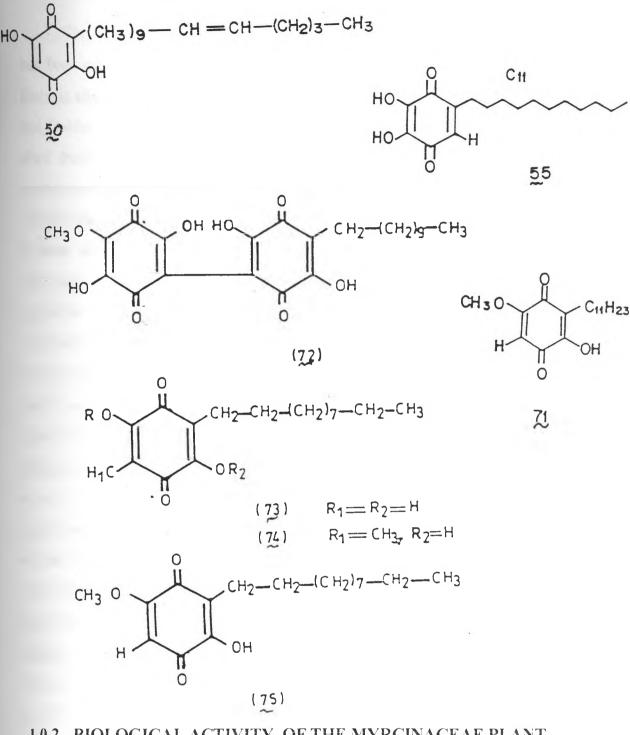
Midiwo <u>et al</u> (1988) reported the Kenyan Myrsinaceae to show a marked chemical relationship within the group which correlates with their morphological classification. His paper also indicated the existence of a remarkable discontinuity in the distribution of the benzoquinone pigments among the four species found in Kenya namely; <u>Embelia schimperi</u>. <u>Maesa lanceolata</u>. <u>Mvrsine africana</u> and <u>Rapanea melanphloes</u>. During the separation of extract from <u>Maesa lanceolata</u> to retrieve compounds for anti-fertility test. Midiwo <u>et al</u> (1990) also isolated two new dihydroxylated benzoquinones which are related to maesanin (35): 2,5-dihydroxy-3-pentadecyl-1,4-benzoquinone(49) and 2,5-dihydroxy-3-(pentadec-10'z-enyl)-1,4-benzoquinone (50).

In their continued analysis of the Kenyan Myrsinaceae, Midiwo and Ghebremeskel (1993) examined <u>Rapanea</u> <u>melanphloes</u> and isolated a bisbenzoquinone- 6,6'-biembelin (72).

While continuing investigation on the Kenyan Myrsinaceae, Midiwo and Arot (1990) reported two new natural benzoquinones, 2,5-dihydroxy-3- methyl-6undecyl-1,4-benzoquinone (73, muketanin ) and 2-O-methyl-muketanin (74) from fruits of <u>Myrsine africana</u> and <u>Maesa lanceolata</u> respectively.

In the process of further careful analysis of <u>M</u>. <u>africana</u>. Midiwo <u>et al</u> (1992) isolated a new dihyroxylated benzoquinone, myrsinone (55) which differs from embelin in the manner of hydroxy group arrangement around the benzoquinone moiety (in that they are 2,3-placed) and 5-O-methyl embelin (75) which was previously reported in Aegiceras corniculatum (Gomez <u>et al</u> 1989)

(CH2)14 --- CH3



# 1.0.2. <u>BIOLOGICAL ACTIVITY OF THE MYRCINACEAE PLANT</u> <u>EXTRACTS - REVIEW.</u>

A review of Myrsinaceae plants indicates that quite a number of them and their derived compounds find a wide range of biological uses. These include; anthelmintic, anti-malarial, anti-bacterial, analgesic, anti-allergic, larvicidal in grain storage, anti-cancer and fertility regulation.

The ethno-medical implication of plants from this family as anthelmintic has been extensively researched and documented. A pharmacological study of Embelia ribes (Burm) and Embelia robusta (Roxb) was carried out by Paranjpe and Gokhale (1932). They observed that the gelanic preparation of the plants' dried fruits were effective against tapeworm. Further research evaluated the constituent embelin (10) as the active anthelmintic principle. Doses in the range of 660-800 mg were observed to have no toxicity in man or dogs. In a similar respect, Krishna and Varma (1936) communicated that both petroleum ether and chloroform extracts of Myrsine africana berries exhibited activity against ringworm and other skin related diseases. The efficacy of embelin isolated from the dried berries of E.robusta (Roxb), M semiserrata (wall) and M. capitellata (wall) in this respect was also expressed (Krishna and Varma, 1937). In the same year Kawamura and Hokoku (1937) evaluated rapanone (11) and found it to be a potent anthelmintic principle. Two Myrsinaceae plants from Madagascar, Maesa emirnensis and Embelia berbevana have been used locally as anthelmintic and the active principle was embelin (10) (Paris and Rabenoro, 1950). The berries of E. ribes known in as "Vidanga" in India are formulated into a paste and applied on the skin as a treatment for skin infections. The powder from berries when taken with milk followed by a purgative has been one of the ancient remedies to get rid of tapeworm (Haris , 1987). E. ribes, a Myrsinaceae plant native to India and Hagenia abyssinica (Rosaceae) are commonly used in Eastern Africa where in Ethiopia they are known locally as "Enkoko" and "Kosso" respectively (Gupta et al. 1977; Pankhurst, 1972). They are used as arbortifacients in traditional medicine (Casy, 1960; Farnsworth et al. 1975; Seshadri et al, 1978).

A review of past work on Myrsinaceae plants presents information that they have been widely used as anti-bacterial (Duclox and Awschalom, 1926). Harimoto (1939) tested extracts from the fruits of <u>Maesa japonica</u> (Moritzii) against pathogenic Gram-positive and Gram-negative bacteria and found it to be active against both. A pharmacological evaluation of two Madagascan Myrsinaceae plants namely <u>M. emirnensis</u> and <u>E. berbevana</u> revealed that the infusion of

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M.emirnensis fruits killed paramecia in dilution of 1:24,000 in one hour while daphnia was killed by 5% infusion in 45 minutes. A sub-cutaneous injection of powdered leaf suspension at a dose of 10 g per Kg killed 100% of mice. The deaths as a result of the injection of stem, root and fruit suspensions were 40%, 70% and 50% respectively. A hemolytic effect of the plant was due to saponins present. It was further reported that the preparations shortened the blood coagulation time and also caused contraction of the isolated guinea pig gut. On the other hand extracts from the roots of E. berbevana killed paramecia at 1:4000 dilution in 5 minutes. The decoction was found to be toxic to epinochus (Gasterosteus aculeatus) but had very little effect on daphnia. The LD<sub>50</sub> in Mice was 1.g per Kg (Paris and Rabenoro, 1950). Joshi and Magar (1952) reported that decoction of dried fruits of E. ribes is used for treating fever and diseases that cause pain in the chest. They further noted that the drug exhibits significant antibacterial activity. Earlier, Merian and Schlittler (1948) had reported that an infusion of the decoction was used in the treatment of cough and diarrhoea. The roots of Ardisia solanaceae have been used in folk medicine as febrifuge, antidiarrhoeic and anti-rheumatic (Chopra et al. 1956). Huang.et. al (1980), reported the inhibition of growth of Mycobacterium tuberculosis in vitro by E. ribes seeds extract. They observed that the active principle was embelin (10). Other active compounds which were noted along with embelin for this activity were barganin and quercetin. Okazaki and Ishikawa (1976) studied the antibacterial action of auraptene and rapanone against Gram-positive and Gramnegative bacteria, pathogenic fungi and acid fast bacillus. It was observed that auraptene was fairly active against these micro-organisms while rapanone was found to be extremely antagonistic to the sulphur compounds.

Structural modification of natural embelin was attempted using halogenation and esterification to get different pharmacological profiles (Tukannen et al, 1983). These were found to be active against bacteria and fungi up to 10  $\mu$ g/ml. It was found that 6-iodoembelin was more effective as an anti-bacterial than embelin while diacetate of haloembelin showed high efficacy against fungi.

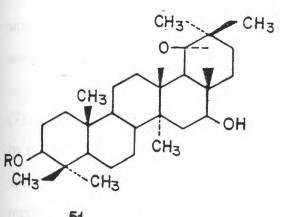
Embelin compounds substituted with aromatic amines were also reported to show a good anti-fungal activity (Rao <u>et al</u>, 1984).

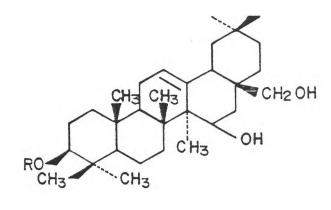
M.lanceolata is a Myrsinaceae plant native to Eastern Africa and hot water decoction of its fruits is drunk to prevent cholera infection (Kubo, 1982). Previous chemotherapeutic evaluation of the plant by Taniguchi. et al (1978) implicated methanol extract of the fresh fruits as anti-bacterial agents. Kubo, et al (1983), further carried out a more detailed bioassay using both crude extract and pure isolates and showed that maesanin (35) is the active principle which involved a non-specific host defence reaction in that mice treated with a single dose 5mg/Kg were significantly protected from normal lethal Escherichia coli infection. Similarly anti-amoebic activity of benzoquinone especially with Entamoeba histolytica showed that rapanone (11) is the active principle in this respect (Shan, 1984). Shan also confirmed the efficacy of rapanone against Trichomonas vaginalis. A screening program in search of biologically active compounds from natural sources for novel anti-parasitic agents such as anti- amoebic and antitrichomonas was carried out by De Souza (1986). In this case plant selection criteria was based on traditional medicine and ethnotherapeutic reports. The CH<sub>2</sub>Cl<sub>2</sub>, MeOH and aqueous extracts of the selected plants were screened for their activity against Entamoeba histolytica and Trichomonas vaginalis in vitro and in vivo models (Chatterjee et al 1984; et al 1985). Plant extracts that displayed activity were those of Chonimorpha fragans (moon). Apocynaceae and Ardisia oxyphlla (Burm), and Myrsinaceae. Chonemorphine was described as the antiamoebic principle of C. fragans and rapanone containing a small amount of embelin and its homologue  $C_{21}H_{34}O_4$  as the anti-trichomonad principle of A. oxyphlla. The compound rapanone(11) also displayed in vitro activity against  $\underline{E}$ . histolytica at 50 µg/ml. Chonemorphine dihydrochloride displayed in vitro activity against <u>T</u>. vaginalis (200  $\mu$ g/ml) and <u>E.histolytica</u> (25 mg/ml) and *in vivo* activity against Hepatic amoebiasis in golden hamster (cured/treated=14/14, 100mg/Kg X 4 doses) and intestinal amoebiasis in weaning instar rats (200mg/Kg X 4 doses).

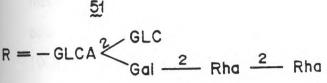
A phytochemical analysis of Kenyan Myrsinaceae plants by Midiwo and Manguro (1988) afforded benzoquinones, embelin (10), rapanone (11) maesaquinone (12), acetylmaesaquinone (30) and maesanin (35) as the isolates. An anti-bacterial study using the compounds against Gram-positive bacteria Bacillus subtilis, Staphyllococus aureus, Streptococcus mutans and Gram-negative Escheria coli implicated embelin (10) and rapanone (11) as the only active metabolites. They were active against Streptococci (Manguro, 1988). In a separate study Mavi, et al (1993) investigated into the molluscicidal and antifungal properties of R.melanphloes (Mez) and found that methanol extract of the plant leaves showed activity in both assays. The extract exhibited a molluscicidal activity at 50 ppm against the schistosomiasis transmitting snail Biomphalaria glabrata and also displayed anti-fungal activity against the plant pathogenic fungus Cladosporium cucumerinum in a TLC assay (Hostettmann, 1980). Activity guided fractionation of the extract using different chromatographic techniques afforded compounds 51, 52, 53, and 54 in yields of 0.25, 0.25, 0.015 and 0.025% respectively.

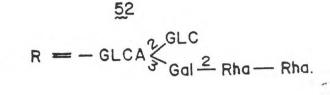
Midiwo <u>et al</u> (1996) conducted mosquito larvicidal tests on embelin (10), myrsinone (55) and myrsinaquinone (56) from <u>Rapanea melanphloes</u> using  $2^{nd}$ instar <u>Aedes aegypti</u> larvae. In this test which was done according to methods of Zebitz (1986) and Mwangi and Rembold (1986). ten larvae were introduced and immediately treated with potassium salts of either (10), (55) or (56). The salts were made by adding an equivalent amount of the compounds to potassium hydroxide solution and then transferred into the jars to achieve the required concentrations of the salts. Control jars were not treated with the salt solutions. Larval growth was inhibited and the survivors took longer time to mature than in the controls. The LD<sub>50</sub> in 48 hours worked out from plots of log of concentration against probits of frequency of embelin, myrsinone and myrsinaquinone were 2.40, 2.54 and 2.69 µg/ml respectively.

\*

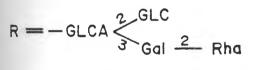


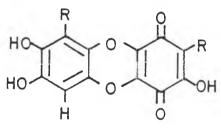






53





54

 $R = - GLCA \stackrel{2}{<} GLC$ 

 $R \equiv C_{11} H_{23}$  $R \equiv C_{13} H_{27}$ 

56

While continuing with their work on insect bioactive compounds from <u>Rapanea melanphloes</u>. Midiwo <u>et al</u> (1996) carried out insect antifeedant test using <u>Schistocerca gregaria</u> females done according to Butterworth and Morgan (1971). Each experimental group of animals consisted of eight mid-5<sup>th</sup> instar females which were starved for 24 hours before the experiment. Whatman # 1 filter papers (2.75x2.75 cm) were then dipped in 100  $\mu$ g/ml solution of the test compounds while the control papers were not treated further. The filter papers were presented to the test animals after drying and left in the cages for 8 hours in a no-choice

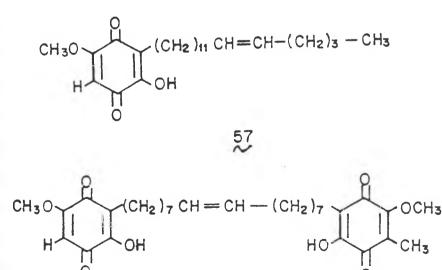
situation. The relative antifeedant percentages (RAP) values were 88.4%, 90.2% and 44.5% for the three compounds, respectively, when tested against S. gregaria.

The effect of natural embelin in prevention of plaque has been discussed (Tsuneo, et al. 1985). In the course of basic studies on the dental carries prevention by traditional medicine, various crude drugs used in Sri-Lanka were screened for anti- plaque action against <u>Streptococcus mutans</u>. It was observed that the methanol extract of the fruits of <u>E</u>. ribes potentially inhibited adherence of viable cells of <u>S</u>. mutans to smooth surfaces with a 50% inhibitor concentration (LC<sub>50</sub>) of 10-30 µg/ml. They further noted that the extract exhibited anti-bacterial activity against <u>S</u>. mutans and anti-enzymic action against glucosyltranferase. The active principle in this case was embelin, which inhibited the bacterial growth with a minimum inhibitory concentration of 62.5 µg/ml and the glucan synthesis with an LC<sub>50</sub> of 125 µg/ml. A year later Tsuneo <u>et al</u> (1986) reported that embelin incorporated in tooth paste prevented formation of dental plaque and acid on teeth.

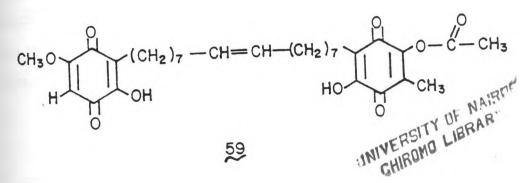
Profile chemical characteristics of benzoquinones in relation to identification of anti-malarial compounds has been extensively studied (Sastry et al 1986). Complexion of anti-malarial drugs such as amodiaquine, chloroquine, primaquine, trimethoprim and pyiremthamine with benzoquinones resulted in formation of colored complex compounds; thus benzoquinones could be used in the detection of anti-malarials. Larvicidal tests using mosquito larvae <u>Aedes aevpti</u> were carried out by Ghebremeskel (1991) and it was observed that embelin (10) and myrsinone (55) had LD<sub>50</sub> of 2.4  $\mu$ g/ml and 2.54  $\mu$ g/ml respectively. Myrsinaquinone (56) on the other hand had LD<sub>50</sub> of 2.69  $\mu$ g/ml

Embelin has been studied for analgesic activity on rats and mice . Alat. (1977) in his study on the analgesic property of embelin with rats and mice reported the affective oral administration of the drug and the results obtained were comparable to those of morphine but he pointed out that the action of the drug was different from that of opiates. Gupta, et al (1977) also reported on ten of embelin disalts which showed a remarkable analgesic activities in rats when administered through intraperitoneal injection. A further research by Neth (1977) showed amine

salts of embelin to have similar effects. Dhar (1980) tested potassium salt of embelin on rats for its analgesic activity and found that the drug did not have any effect on serotonin or histamine levels but levels of adrenaline in cerebro-spinal fluid increased while the activity of acetylcholinesterase decreased. In a separate study, embelin was found to be analgesic by oral administration and central routes and the results compared with morphine. Although the drug acts centrally to produce analgesia, its effect is not antagonized by naloxone indicating that a central site of action is different from opiates. Also no precipitation of abstrience syndrome was observed as with morphine. A peripheral site of action of the drug was ruled out as it lacked any demonstrable anti-inflammatory action. However, the high oral efficacy and non-narcotic properties makes embelin more accepted than morphine (Atal, et al 1984). Similarly, Kokai (1985) in his pharmacological studies of benzoquinones, evaluated methanol extracts of Ardisia japonica as arachidonate-5-lipoxy-genase inhibitor. The extract was fractionated by silica gel column chromatography to give the benzoquinones (57) and (58) respectively as the arachidonate-5-lipoxygenase inhibitors and are also effective in treatment of asthma and inflammation.

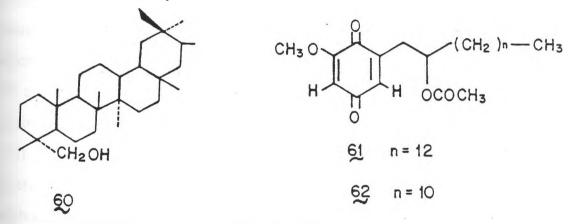


There are benzoquinone derivatives with wide range of applications as antiallergic compounds that have also been reported in the literature. Hideyuki (1984) studied the anti-allergic activities of the derivatives of ardsiaquinones (32), (58) and (59) and found them to be effective and could therefore be used as substitutes for anti-allergy compounds. In the following year, a study on the synthetic benzoquinone derivatives by Iwaki (1985) indicated that the compounds have antiasthmatic and anti-histamine activities.



A comparative study of fungicidal quinones and natural embelin from <u>E</u>. ribes as protectants of wheat seeds against the stored product insect pests; <u>Sitophilus orvzae</u> (L), <u>Rhvzopertha dominica</u> (F) and <u>Ephestia contella</u> (Walker) was done (Chander and Ahmed, 1989). Embelin and chloranil at concentrations of 0.0125 and 0.025% (w/w) respectively in wheat produced mortalities of 77.8 % and 68.7 % in adult <u>S</u>. <u>orvzae</u> after 14 days of exposure, while dichlone produced 82.8% mortality at 0.01% concentration level. When applied against <u>R</u>. <u>dominica</u>, dichlone even at 0.0125 % concentration level gave 78 % adult mortality after 14 days, whereas the same mortality was observed with embelin and chloranil at 0.025 % and 0.05 % concentration levels respectively. Dichlone did not exhibit any chemosterilant action on the adults of <u>S</u>. <u>orvzae</u> or <u>R</u>. <u>dominica</u> or any inhibitory effect on oviposition as with the controls.

Some Myrsinaceae plants have been screened for anti-cancer activity. Kupchan <u>et al.</u> (1969) tested <u>Myrsine africana</u> (L) leaves for anti-tumor activity against "Walker" intramuscular carcinosarcoma-256 in rats. The chief component of the leaf hydrolysate was lucynoside (60). Information on certain methylated benzoquinone derivatives as having sensitizing activity has been reported (Shultz et al, 1979). In a similar respect <u>Ardisia cornudentata</u> (Mez), a species used in folk medicine in the south eastern part of China as anti-inflammatory/analgesic medication to improve general blood circulation and also as antidote for snake and insect bites, was subjected to phytochemical investigation by Chang et al. (1987) to identify the active components. In this study, root extracts of the plant were evaluated in several <u>in vitro</u> receptor and enzymatic assays. The methylene chloride extract was found to inhibit the binding of leukotriene,  $D_4$  (LTD<sub>4</sub>),to a receptor preparation from a guinea pig lung tissue. Using the LTD<sub>4</sub> binding assay as a guide, two benzoquinones, ardisianone (61) and cornudentanone (62) were isolated as the active components.



In recent years, researchers have been interested in the development of a safe and reversible effective anti-fertility agent of plant origin. Das <u>et al</u> (1976) developed an oral contraceptive by mixing crushed <u>Embelia ribes</u> fruits, <u>Piper</u> longum, asafoetida and purified borax in the ratio 1:1; 1:2 respectively. The drug was administered orally to female rats while not abstaining the test animals from sexual intercourse. This drug was active in preventing implantation of the fertilized ovum. There was no side effect associated with the drug. In continuing with research for anti-fertility agent from plants, embelin isolated from the berries of <u>E</u>. ribes (Burm) has been reported to induce functional sterility in mice, rats and dogs (Munishi and Rao, 1972; Munishi. <u>et al</u>, 1972; Prakash and Mathur 1979; Dixit and Bhargava, 1983). "Garbhanivarana aushadam", an ayurvedic anti-fertility drug agent in India composed of <u>Piper longum</u>, <u>E</u>. ribes and borax, when administered to female rats at a dose of 10% of the food in their diet for 16 days or fed (0.5g per

day for 16 days) to guinea pigs, showed a slight prolongation of the oestrus cycle in both species. Histochemical studies revealed that alkaline phosphatase activity in the uteri of treated animals was enhanced when compared to the control (Geeta <u>et</u> al, 1976).Greep (1976), in his comparative analysis of studies in male and female reproductive physiology noted that the understanding of males' reproductive physiology lags behind that of the females.

While continuing with research on anti-fertility, Kholkute (1978) studied the activity of E. ribes berries and its extract on female rats. The drug was incorporated into their diet and administered at various dose levels. It was found to prolong the diestrus phase of the oestrus cycle and inhibit fertility in 62% of the animals. Serially hot extracted petroleum ether and methanol extracts affected cyclicity and prevented pregnancy in 75% of the rats. Benzene extract showed 51% anti-fertility whereas that of chloroform gave 37% anti-fertility. However, embelin in this case failed to reveal any anti-fertility activity (Kholkute et al, 1978). In male rats, daily administration of 75-300mg/Kg of embelin showed a reduction of testis and prostrate gland weights and indicated an impairment in the metabolic functions of these organs. Acid and alkaline phosphate activities were elevated (Chanhan et al, 1979). Semen analysis and hormonal levels in bonnet monkeys treated with E. ribes berries's extract was studied by Purandare et al, (1979). When the extract was administered orally for 3 months at a dose of 100 mg per day to male monkeys, the quality and quantity of semen was adversely affected. Circulating testosterone levels were also reduced but the luteinizing hormone (LH) levels was however not affected . Testicular biopsy revealed normal spermatogenesis. The reduction in the testosterone level in the circulation may be responsible for the reduced secretory activity of the accessory glands which in turn resulted in a decrease in the volume of semen. Prakash, (1980) studied the effect of embelin on Corpora lutea of cycling guinea pigs and showed that increase in ovarian weight depended on the dosage given to the animals. He also observed that embelin was neither antizygotic nor blastotoxic despite its anti-implantation and abortifacient activities. However, the drug was observed to show an antioestrogenic activity but not anti-progesterone activity (Prakash, 1980). In a previous report Dixit <u>et al.</u>(1978) showed that embelin administered at 80mg/Kg for 100 days to male dogs caused azoospermia and the animals recovered 250 days later. Similarly embelin from the seeds of <u>E</u>. <u>ribes</u> administered at 100 mg/kg and 50mg/Kg to female rats showed 57.9% and 55.5% anti-fertility respectively (Mohana <u>et al.</u> 1981).

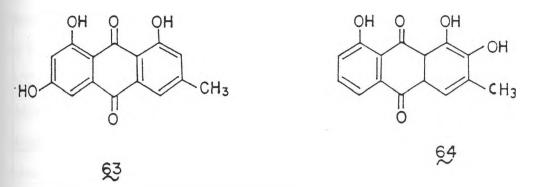
Prakash, (1981) in his continuation with the study of embelin on fertility regulation administered the drug orally to ovariectomized rats and found that the drug altered the activity of various enzymes in the uterine tissue. However, when the drug was given to oestradiol primed rats, a decline of uterine weight was noted. The drug did not bring any reasonable change in the uterine biochemistry of progesterone primed rats. Antispermatogenic effect studies of embelin by Seth, (1983) showed that embelin significantly reduced the sperm count, and the weight of the experimental albino rats. Embelin fed at 50-200mg/Kg body weight for 15 days reduced the weight of the testis significantly and also effectively reduced the sperm count significantly. Studies by Bharghara et al, (1984 and 1985) showed that embelin when given to rats at one to five days of pregnancy at 20-50mg/Kg inhibited implantation of the blastocyst. Agarwal et al. (1986) reported that embelin altered the rat testis histology, glycogen levels, sperm count and accessory sex glands. Effects of embelin on the activity of uterine ß-gluconidase in ovariectomized rats (Prakash and Sukhla, 1987) showed a marginal increase in enzyme activity. In another development, subcutaneous exposure of male albino rats to embelin (20mg/Kg body weight) for 15 to 30 days revealed significant impairment in carbohydrate metabolism in the primary and secondary reproductive tissues. Alterations of the enzyme activities of glycolysis. Krebs's cycle, NAD and NADP dependent enzymes, transaminases and lipogenesis, phosphatases are noted on embelin treatment in testis, epididymis, seminal vesicles and ventral prostate as well as in spermatozoal suspension. Reduction in fertility parameters such as pregnancy attainment and uteri sizes obtained was also noted on embelin treatment. All these changes in embelin treated animals are restored

back to the normal biochemical make up of the control animals, once the drug is withdrawn and the animals are allowed to recover for another 15 to 30 days (Gupta et al. 1989). In a similar study Makawiti and Midiwo (1990) in their preliminary studies have indicated that in white New Zealand male rabbits, embelin dose of 40-80mg/Kg body weight/day for 15 days led to the decrease in testosterone levels. They observed a concomitant correlation in the rise of progesterone and a fall in testosterone level. Luteinizing hormone (LH) showed a 26 % rise in level apparently responding to decreasing testosterone levels. Health parameters such as packed cell volume and weight were monitored and were found to be normal except in animals that were under maesaquinone (56) treatment. It is apparent that the progesteronal activity exhibited by embelin was probably due to its ability to raise the concentrations of progesterone. Further research studies suggested that embelin causes the elevation of progesterone levels by inhibiting the17hydroxylase enzyme in the testosterone biosynthetic pathway. In their continuation with the study they observed that histological studies showed a major difference between the embelin treated and control rabbits. In the case of treated animals, the epididymis was devoid of spermatozoa and the seminiferous tubules were atrophied. The germ cells in the basal lining appeared normal but the adluminal cells were disorganized, extensively vacuolated and appeared to be sloughing off. It was also observed that some of the meiotic cells and spermatids fell off to the lumen of the seminiferous tubule. The sertoli cells were also found to be highly vacuolated and hyalinised.

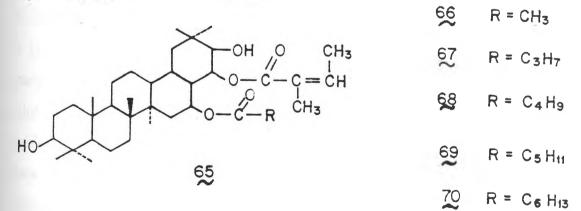
The effect of embelin from <u>E</u>. <u>ribes</u> and oleanolic acid from <u>Parthenium</u> <u>hvsterophorus</u> on maize and cow pea germination has been investigated (Sinha <u>et</u> <u>al</u> 1981) at 50-520ppm at 22°C. Oleanolic acid markedly reduced the germination percentage 48 hrs after seed treatment. Embelin reduced germination at 100 ppm and 250ppm but stimulated germination at 50 ppm. Data on respiratory rates and catalase activity indicated that both compounds inhibited respiration.

Bioactivity of ethanol extract of the roots of <u>M</u>. africana using brine shrimp lethality test was studied by Mclaughlin and Xia-Hua (1989). From the study,

emodin (63) and 2-hydroxychrysophanol (64) were implicated as the cytotoxic components.



A number of medicinal uses have been reported for <u>M</u>. <u>lanceolata</u>, a tree growing in many African countries. In Kenya, the plant is used against cholera (Kubo <u>et al</u>, 1983). Anti-bacterial and anti-viral activity tests using methanol extract of the plant showed that the extract only exhibited a virucidal affect against herpes simplex type 1, vesicular stomatitis and semliki forest A<sub>7</sub> viruses. A bioassay guided fractionation of the extract implicated the compound (65)(16-0propionyl-22-0-angeloyl-3,21-dihdroxyl-12-oleanen-28-al) as the active principle. Other compounds reported to have been isolated in minor quantities along with (65) are (66-70) (Sindanambiwe <u>et al</u>, 1993).



Compounds and their derivatives from Myrsinaceae plants have shown only a few adverse effects. However, the areas sighted have not been fully investigated. <u>E. ribes</u> and <u>Hagenia abyssinica</u> have been reported to cause optic atrophy among Ethiopian population (Low <u>et al.</u> 1985). In their study they investigated the possible toxic effects of ingested <u>E. ribes. H. abyssinica</u> and embelin on visually guided behavior in chicks. followed by histological

examinations of their retina. A high dose of E. ribes was administered for 1 to 5 days while that for H. abyssinica was administered for 1 to 9 days. For the low dose of both E. ribes and H. abyssinica, the dosing regime was for a period of 1, 4 or 9 days. Embelin, the active principle of E, ribes was administered at a dose of 0.02g/Kg per day for 9 days. Control chicks were fed on an equivalent amount of chick feed. From the experiment, they observed that treatment with E. ribes or H. abyssinica significantly reduced the ability of the chicks to detect a moving bead introduced into the peripheral field of vision. They further noted that the degree of constriction of the visual field for detection was dependent upon the total amount of drug administered. Performance on a visual discrimination task which required discrimination of feed grains from pebbles was also impaired in chicks with a total dose of 0.20g and 0.25g of E. ribes or H. abvssinica. Thus, the extent of deficit in visually guided task was found to be dose-dependent. However, the visual deficits observed in E. ribes treated chicks were mimicked by embelin suggesting that embelin may be responsible for the visual defects. Anatomical evidence of degeneration of ganglion cells was found in retinae exposed to high doses of  $\underline{E}$ . ribes (1.25g) and H. abyssinica (2.25g). However, no retinal lesions were detected in chicks following treatment with cumulative doses of less than 0.25g of E. ribes or H. abyssinica. In a separate study the crude extract of twigs and stem of the mangrove plant. Aegiceras corniculatum exhibited toxicity to juvenile fish, Tilapia nilotica at a concentration of 1ppm within a period of 75 minutes. A fractionated guided assay implicated a derivative of embelin, 5-0-methylembelin (71) as the toxic substance in this respect. At the same time, the growth fungus, Pythium ultimum was inhibited by the same compound with an inhibition zone of 8mm diameter with a solution of 1mg of the compound in chloroform (Gomez et al. 1989).

The efficacy of natural embelin (10) against the red flour beetle <u>Tribolium</u> casterenum (Herbst) was studied by Chander and Ahmed (1985). They observed that when the drug was mixed with wheat samples at 0.19% concentration, a high efficacy of action was reported which brings about adult mortality even after 8

months of storage. Reproductivity (progeny/adult day) of the insect was also reduced significantly in the treated samples as compared to the controls at different interval of storage. After 8 months of storage, significant reduction in progeny at the lowest concentration was mainly due to larvicidal action of embelin. Embelin, however, did not inhibit any chemosterilant action or contact toxicity to the adult beetle. A further research on the drug by Chander and Ahmed (1987) revealed that the drug is mutagenic to this strain of insect only when one or two hydroxyl groups on it are methyl substituted. Two years later the proven efficacy of embelin was evaluated as a protectant against a variety of insect pests of wheat storage. The drug showed 50% mortality on both <u>Sitophilus orvzae</u> and <u>Rhysopertha dominica</u> adults at 0.025% concentration (Chander and Ahmed, 1987). Previous studies by Tukannen, (1983) in this respect showed that natural naphthoquinones and benzoquinones were mutagenic to <u>S</u>. typhimunum strain TA 2637 with metabolic activation. Ardisiaquinones (32), (58) & 59 were implicated for this activity.

#### **BROAD OBJECTIVE.**

The broad objective of this study was to extensively examine the chemical composition of extracts from berries and root bark of *Embelia schimperi* and *Embelia keniensis*.

#### Specific Objectives.

- a) To isolate bioactive compounds from Embelia schimperi and Embelia keniensis.
- b) Perform toxicity assays of the isolated compounds using brine shrimp.
- c) Perform anti-feedant and grain protectant tests on the compounds found to be positive on the brine shrimp assay.

#### **CHAPTER TWO**

#### 2.0.0. RESULTS AND DISCUSSION.

Dry ground root bark and fruit of <u>Embelia schimperi</u> and <u>Embelia keniensis</u> were extracted with cold ethyl acetate and the solvent removed <u>in vacuo</u> leaving a dark gummy solid. A preliminary spot-test survey was carried out to determine the quinonoid pigments in two plants using thin layer chromatography (t.1.c). A solvent system consisting of n-hexane, ethyl acetate, acetic acid (80:15:5) revealed that the root bark of <u>Embelia schimperi</u> had components with Rf 0.9, 0.55, 0.53 and 0.47 while the fruit had components with Rf 0.9, 0.64, 0.62, 0.55 and 0.49. All the spots turned permanently pink on exposure to conc. ammonia vapor except the spot with Rf value of 0.47 which absorbed UV radiation. <u>Embelia keniensis</u> had components occurring in very trace amounts and thus was not followed.

The extracts from each of the two parts of Embelia schimperi were preabsorbed on dry de-activated silica gel and subjected to column chromatography in a column packed under n-hexane with the de-activated silica gel. The column was exhaustively eluted with n-hexane, dichloromethane and ethyl acetate respectively and a fraction of each solvent was collected separately. Each eluate was tested against Artemia salina (brine shrimp) nauplii (larvae). The dichloromethane and ethyl acetate eluate were both found to be active against the organism. However, from t.l.c. analysis, the ethyl acetate eluate was found to contain mainly the more polar components occuring in trace amounts and for this reason, it was not followed. The dichloromethane eluent of the root bark was found to contain the embelin (10), Rf 0.55 (solvent system: n-hexane, ethyl acetate and acetic acid-80:15:5) in larger amounts than others and therefore masked the minor ones on tlc plate. This prompted repeated fractional cyrstallization of 10 until very little remained in the mother liquor. The solvent was removed from the mother liquor leaving behind a residue which was pre-adsorbed on de-activated silica gel and subjected to column chromatography using de-activated silica gel. The polarity of the system was increased with increased addition of dichloromethane upto 100%.

The column was finally washed with 5% methanol in dichloromethane. Further purification of the compounds was done using preparative t.l.c, gravity column chromatography and fractional crystallization. The compounds obtained from the root bark were Rf 0.55, embelin (10), Rf 0.9, decylanhydrovilangin (77), Rf 0.53, myrsinaquinone (56) and KCP-02. The compounds were characterized using their melting points and spectroscopic data. Isolation of components in the dichloromethane eluate of E.schimperi fruit was also done using column chromatography. The eluting solvents were n-hexane, dichloromethane, ethyl acetate or their combinations. Purification of the compounds was done using preparative t.l.c. gravity chromatography and fractional crystallization. The compounds obtained from this part of the plant included Rf 0.55, embelin (10), Rf 0.9, decylanhydrovilangin(77), Rf 0.53, myrsinaquinone (56), Rf 0.62, methylvilangin (21), Rf 0.49, KCP-06 and Rf 0.64, decvlvilangin (76). The compounds were characterized using their melting points and spectroscopic data. Each of the pure compounds was tested for the biological activities shown in section 2.0.1

#### 2.0.1. BIOASSAYS.

#### 2.0.2. Brine shrimp lethality test.

This method utilizes brine shrimp (<u>Artemia salina LEACH</u>) and is proposed as a simple bioassay for possible bioactive compounds. The procedure determines  $LC_{50}$  values in µg/ml of active compounds and the extracts in the brine medium. Activities of a broad range of known active compounds are manifested as toxicity to the brine shrimp nauplii (Mclauglin <u>et al</u> 1982). The method is rapid, reliable, inexpensive and convenient as an-in-house general bioassay tool.

Methylvilangin (21), decylvilangin (76), KCP-02 and KCP-06 were subjected to this test and their  $LC_{50}$  (summarized in Table 1) values are 120, 51, 54 and 0 respectively. These results indicate that 76 and KCP-02 have a moderate activity. Embelin (10) and myrsinaquinone (56) were not subjected to this test 35

because they had been tested ealier (Ghebremeskel, 1991) while  $\frac{77}{2}$  was not tested due to lack sample.

#### Table 1

Effects of various benzoquinones on Artemia salina (shrimp)

Compound	(	% Deaths at 24 hrs				LC <sub>50</sub>		
Conc.(µg/ml)	5	10	15	20	25	50	100	
КСР-02	10	17	20	27	31	33	62	54
21	10	10	20	23	26	30	36	120
КСР-06	0	0	0	0	0	0	0	0
76	7	13	16	25	32	50	55	51

#### 2.0.3. Anti-feedant test

In the recent years research scientists have focused attention on feeding deterrents for control of insect pests. In 1983, Kraus <u>et al</u> for instance conducted research on anti-feedant effects of Meliaceae extracts. In view of the continued interest focused on feeding deterrents, embelin (10) and methylvilangin (21) were screened for anti- feedant activities using <u>Locusta migratoria migratorioides</u> using "choice" test in which whatman No. 1 filter papers previously soaked in 0.25 M sucrose solution were sprayed with 100  $\mu$ g/ml of each sample. The relative antifeedant percentage for each compound was calculated using the formula given below and were found to be 96 and 33 for 10 and 21 respectively. The results (summarized in Table 2) indicate that that embelin (10) offers a significant level of protection to field crops against destruction by locusts. Compounds 56, 76, 77, KCP-02 and KCP-06 were not tested because of lack of samples.

#### Table 2

Relative Anti-feedant Percentages (RAP) for embelin (10) and methylvilangin (21).

Compound	Average eaten area	Average uneaten	RAP(%)	
1.0	0.08	6.25	96	
21	2.38	3.88	33	

	Average eaten areas of		Average eaten areas of	
	control filter paper	-	sample treated filter paper	
RAP =				X 100
	Average eaten areas of	+	Average eaten areas of	
	control filter paper		sample treated filter paper	

#### 2.0.4. Anti-Microbial tests.

An attempt was made to determine anti-microbial activities of 77, 56, 21. 10 and KCP-06. Compound KCP-02 was not available for this test. Concentrations of 25, 50 and 100  $\mu$ g/ml of each compound were prepared and tested against <u>Candida albicans. Trichophyton metagrophyte. Microsporium gypsum</u> and <u>Escherichia coli</u> and all were found to be inactive against the micro-organisms.

#### 2.0.5. Larvicidal test.

In spite of many years of research into malarial drugs, malaria still remains the leading killer disease mainly in Africa. Plasmodium (malaria causing parasite) has been found to be resistant to most drugs in the market today. In view of this worrying trend, an attempt was made to investigate 21 for larvicidal activity using concentrations ranging from 25  $\mu$ g/ml to 100  $\mu$ g/ml in a pond containing twenty (20) 2<sup>nd</sup> instar larvae of *Aedes aeypti*. Compounds 10 and 56 had been subjected to this test earlier (Ghebremeskel, 1991) while KCP-02, KCP-06, 76 and 77 were not available for testing. Compound (21) was found to disrupt metamorphosis without causing any mortality. In fact, when 100  $\mu$ g/ml of 21 was applied into a pond containing the 2<sup>nd</sup> instar mosquito larvae, only a few underwent metamorphosis to the 3<sup>rd</sup> instar larvae and none attained the adult stage. This biological activity can be used to reduce population of mosquitoes. The results are summarized in Table 3.

#### Table 3

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Concentration	Population				
	2 <sup>nd</sup> instar	3 <sup>rd</sup> instar	4 <sup>th</sup> instar	Pupae	Adults
100 μg/ml	14	5	1	0	-
75 μg/ml	10	6	3	1	-
50 μg/ml	5	7	6	-	2
25 µg/ml	3	3	2	8	4
10 μg/ml	-	2	3	2	13

Larvicidal effect of methylvilangin (21)

#### 2.0.6. Stored Products Pests Test.

In view of tremendous losses due to pest attack in Kenya (*vide supra*), an attempt was made to investigate the level of protection that 10 can offer to stored beans and maize grains. Concentrations of embelin ranging from 0.04% to 0.2% were introduced to jars containing twenty (20) mixed adults of *Anthoscelides* 

obtectus and Sitophilus zeamais. Each experiment was, performed in three replicates.

Results on the effectiveness of embelin against <u>S</u>. <u>zeamais</u> (Figure 1) indicated that even after 5 months of storage, the number of emerged adult progeny and reproductivity reduced significantly at all the concentrations in comparison with the control. After this period of storage, only the highest concentration of 0.1% gave the highest mortality.

Data on the effectiveness of embelin against <u>A</u>. <u>obtectus</u> (figure 2) similarly revealed that the total number of emerged progeny and reproducibility were reduced significantly at all concentrations as compared with controls. The significant reduction in the progeny of <u>S</u>.zeamais and <u>A</u>. <u>obtectus</u> at all concentrations of embelin could possibly be due to the effect of embelin as a larvicide.

It can be concluded from the foregoing discussion that the effectiveness of embelin as a larvicide is retained even after 5 months of storage, indicating stability and slow rate of degradation under the experimental conditions. In general, the effectiveness of embelin was greater against <u>A. obtectus</u> compared to <u>S. zeamais.</u>

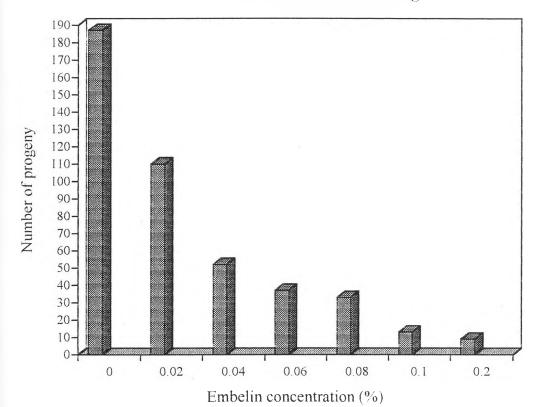


Figure 1: Variation of *Sitophilus zeamais* progeny with concentration of embelin after 5 months storage.

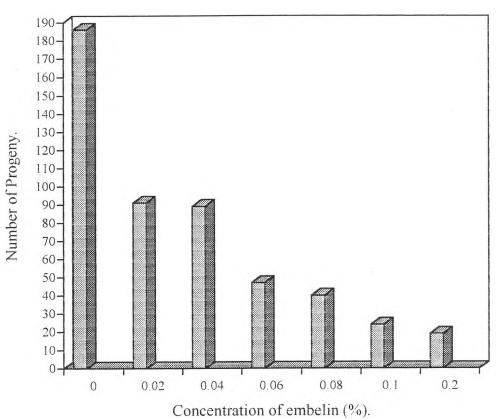


Figure 2: Variation of *Anthoscelides obtectus* progeny with concentration of embelin.

#### 2.1.0. Structural determination of dihydroxyalkyl-1,4-benzoquinones.

During the course of the investigation on the secondary metabolites, embelin (10), which had been characterized from the same plant before, was isolated in high yield from the fruit and root bark relative to other components. It is a golden-yellow compound with Rf 0.55 (eluent: n-hexane - ethyl acetate - acetic acid - 80:15:5) and m.p 141-142°C (same as that reported in the literature by Ogawa and Natori, 1968). The highest concentration of the compound was realized in the fruits which co**ntai**ned 10.2% w/w while the root bark contained 8.1% w/w. Its characteristics matched those of the same compound isolated from the same plant previously in our laboratory.

Chromatographic separation of the dichloromethane eluent of E. schimperi gave an orange compound (21) (solvent system: same as that used for 10) with m.p. 129-131°C. This compound turned pink on exposure to conc. ammonia vapor. Its UV-Vis spectrum displayed a peak at 430 nm and 290 nm thus indicating the chromophore of the compound as a 2,5-dihydroxy-1,4-benzoquinone. The compound showed six signals in the <sup>1</sup>H NMR spectrum. There was no signal corresponding to the quinonoid proton which usually appears at 6.50-5.0 ppm. This suggests that the quinonoid ring is fully substituted. A singlet peak due to phenolic hydroxyl hydrogens bonded to carbonyl group appeared at 7.82 ppm. The spectrum also contained peaks at 4.39 ppm (H. q, J=7.50 Hz) and 1.59 ppm(3 H, d, J=7.50Hz) which are unusual for the usual alkyl substituted 1,4-benzoquinones from Myrsinaceae. The peak at 4.39 ppm suggests the presence of a proton sitting on the same carbon atom with a methyl group whereas the doublet peak at 1.59 ppm could be due to a methyl group next to a proton. Infact, a methyl group substituting the ring displays a sharp singlet at approximately 1.90 ppm whereas the quinonoid proton appears either as a singlet or a multiplet at 6.5-5.0 ppm (Ogawa and Natori, 1968; Chandrasekhar, 1970; Thomson, 1971). Irradiation of the peak at 4.39 ppm collapsed the doublet at 1.59 ppm into a singlet suggesting

that the proton and the methyl group are adjacent to each other and possibly sitting on the same carbon atom. The triplet at 2.40 (J=7.1 Hz) ppm was assigned to the benzylic methylene protons. Integration of this peak relative to the set at 4.39 ppm correspond to four protons calculated for two methylenes. This therefore suggests that the compound under consideration probably consists of two dihydroxyalkyl-1.4-benzoquinones bridged by a carbon holding a proton and a methyl group. A multiplet at 1.43-1.25 ppm on integration relative to the peak at 4.39 ppm corresponded to 36 protons calculated for 18 methylenes whereas the terminal methyl protons which appeared at 0.9 ppm similarly on integration relative to the peak at 4.39 ppm revealed the existence of six protons thus accounting for two methyl groups. The <sup>13</sup>C NMR showed a C-6 peak at 115.86 ppm which corresponds to a quartenary carbon bearing the bridging carbon holding a methyl and a proton. This seems to confirm the presence of two alkyldihydroxy-1,4benzoquinones in (21). The mass specrum showed significant fragments at m/e 320, 294, 180, 154, 139 and 125. The molecular ion did not show. However, the most striking feature of the mass spectrum of this compound was the peak at m/e 294 nm which evidenced the existence of embelin (10) and/or myrsinone (55). The latter was ruled out as additional peaks due to long range coupling between the alkyl side chain and the quinonoid proton were absent in the <sup>1</sup>H NMR spectrum. Also evident in the mass spectrum was a base peak at m/e 154 nm and a low abundance peak at m/e 153 nm. Such fragmentation patterns have been observed in embelin (10) and myrsinone (55) both of which have a quinonoid proton in the ring. Having ruled out 55 with the help of <sup>1</sup>H NMR data, mass spectral data stongly favoured the existence of embelin moiety as part of the molecule. The spectroscopic data confirm 21 as methylvilangin which according to literature has been synthesized from embelin (10) and acetaldehyde by Rao and Venkateswarlu (1964)

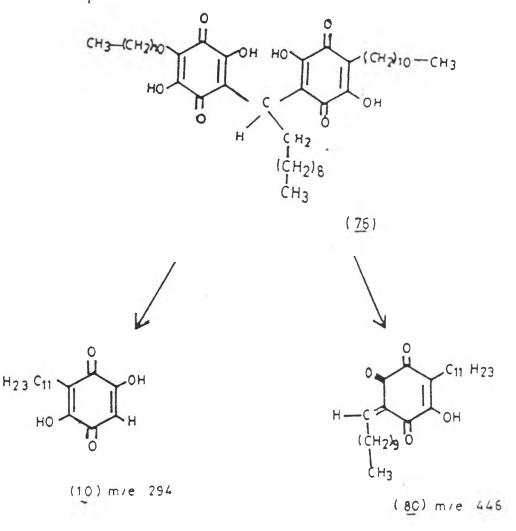
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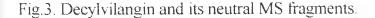
Atom	<sup>1</sup> H (ppm)	<sup>13</sup> C (ppm)
1,1'	-	-
2,2'	-	-
3,3'	-	116.35
4,4'	-	-
5,5'	-	-
6,6'	-	116.09
H- <u>C</u> -CH <sub>3</sub>	4,39 (q. J=7.50 Hz)	28.20
H-C- <u>C</u> H <sub>3</sub>	1.59 (d, J=7.50 Hz)	16.75
7,7'	2.40 (t J=7.1Hz)	29.60
8,8',9,9',10,10',		32.13,29.92,29.87,
11,11',12,12'13,13'14,14',1		29.84,29.81,29.77,
5,15',	1.43 -1.25 (m)	29.55,28.18,26.87,
16,16'		26.87,22.90,22.76
17,17'	0.9 (t, J=13.2 .6.3 Hz)	
		14.32
2,5, 2'5'- OH	7.82 (s. D <sub>2</sub> O exchangeable)	-

 Table 4: <sup>1</sup>H and <sup>13</sup>C NMR (CDCl<sub>3</sub>) for 21

Compound (76) Rf 0.62 (eluent: n-hexane, ethyl acetate and acetic acid-85:10:5) m.p 121-123°C was also isolated from the fruit. It turned permanently pink on exposure to concentrated ammonia vapor on t.l.c. Its UV-Vis spectrum displayed two peaks at 430 nm (2.6) and 290 nm (4.5) thus indicating the chromophore of the compound as a dihydroxy-1,4-benzoquinone. The <sup>1</sup>H NMR of the compound exhibited peaks at 7.80 ppm which was assigned to phenolic hydroxyl hydrogen bonded to a carbonyl group. Peaks were also displayed at 4.27 ppm (H, t, J=7.92 Hz), 2.05 ppm (m) and 1.29 to 1.45 ppm (m) which are unusual for dihydroxy-1,4-benzoquinone. Integration of the peak at 7.80 ppm with respect to that at 4.27 ppm corresponded to four hydroxyl groups. The proton-proton correlation (homonuclear COSY) experiments showed coupling between proton absorbing at 4.27 ppm and the multiplet at 2.05 ppm which in turn coupled with the multiplet at 1.29 to 1.45 ppm. Integration of the peak at 2.05 ppm with respect to that at 4.27 ppm corresponded to two protons. The peak at 4.27 ppm suggested the presence of a proton sitting on the same carbon with a methylene group while the multiplet at 2.05 ppm could possibly be assigned to a methylene group sitting on the same carbon atom with a proton. The multiplet at 1.29 to 1.45 ppm could be due to the remaining methylene groups in the decyl chain. As expected for alkylated dihyroxy-1.4-benzoquinones, there was a triplet at 2.40 ppm (J=7.44 Hz) which was assigned to the benzylic methylene protons. Integration of this peak relative to the set at 4.27 ppm corresponded to four protons calculated for two methylenes. This suggested that the compound probably consisted of two alkyl substituted dihydroxy-1,4-benzoquinones bridged by a carbon holding a decyl group and a proton. Furthermore, irradiating the benzylic peak at 2.40 ppm did not affect the triplet at 4.27 ppm and the multiplet at 2.05 ppm, proving that neither the proton nor the decyl group and the alkyl side chain are vicinal Integration of the multiplet at 1.29 to 1.45 ppm relative to the peak at 4.27 ppm corresponded to 52 protons calculated for 26 methylenes. The terminal methyl groups appeared at 0.88 ppm. Integration of this peak relative to that at 4.27 ppm revealed the existence of nine protons and thus accounting for three methyl groups. On comparing the <sup>13</sup>C NMR spectral data of (76) with that of embelin (10), there was only one significant difference for the signal due to C-6. The C-6 peak, which in (10) is centred at 103.80 ppm appeared at 115.1 ppm, corresponding to a quartenary carbon bearing the bridging carbon holding the decyl group and the proton. This seems to confirm the presence of two alkyl dihydroxy-1,4-benzoquinones in (76). The mass spectrum showed significant fragments ions at 320, 294, 180, 139 and 125. The mass spectrum did not show molecular ion peak. However, it showed a base peak at m/e 154 indicating the presence of alkyldihydroxy-1,4-benzoquinone in the

molecule. The peak at m/e 294 evidenced the existence of either embelin (10) or myrsinone (55) with the latter being ruled out as additional peaks due to long range coupling between the alkyl side chain and the quinonoid proton were absent in the <sup>1</sup>H NMR spectrum. Having ruled out (55) with the help of <sup>1</sup>H NMR data, mass spectral data strongly favoured the existence of embelin unit as part of the molecule. In fact the fragment ion peak at m/e 446 seems to be due to the thermally propagated reaction in the ionization chamber in the manner reported by Rao and Venkateswarlu (1961) which fragments the compound leading to the formation of (10), m/e 294 and (80), m/e 446 as shown in figure 3. The spectral data strongly support 76 as decylvilangin - a compound that is being encountered for the first time as a natural product.





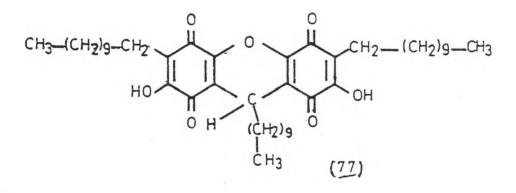
Atom	<sup>1</sup> H (ppm)	<sup>13</sup> C (ppm)
1,1'	-	-
2,2'	-	-
3,3'	-	116.33
4.4'	-	-
5,5'	-	-
6,6'	-	115.10
H- <u>C</u> -CH <sub>2</sub> -(CH <sub>2</sub> ) <sub>8</sub> -CH <sub>3</sub>	4.27 (t,J=7.92 Hz)	28.63
H-C- <u>C</u> H <sub>2</sub> -(CH <sub>2</sub> ) <sub>8</sub> -CH <sub>3</sub>	2.05 (m)	28.18
H-C-CH <sub>2</sub> -(CH <sub>2</sub> ) <sub>8</sub> - <u>C</u> H <sub>3</sub>	0.9 (t J=6.3 H)	14.33
7,7'	2.40 (t,J=7.44 Hz)	29.60
H-C-CH <sub>2</sub> ( <u>C</u> H <sub>2</sub> ) <sub>8</sub> -CH <sub>3</sub> ,		32.33,32.13,30.43,
8,8',9,9'.10,10'.		29.84,29.77,29.74,
11,11',12,12',13,13',	1.45 - 1.25 (m)	29.55,22.90,22.79
14,14',15,15',16,16'		
17,17'	0.9 (t J=6.3 Hz)	14.33
2.5, 2',5'- OH	7.84 (s, D <sub>2</sub> O exchangeable)	-

Table 5: <sup>1</sup>H and <sup>13</sup>C NMR (CDCl<sub>3</sub>) for 76

The other compound was designated as decylanhydrovilangin, (77) Rf 0.9 (eluent: n-hexane, ethyl acetate and acetic acid-85:10:5) was isolated from the two parts of the plant. It is orange in color with m.p 119-121°C. Like (10), it was realized in highest concentration in fruits 1.9% w/w and a lower concentration in the root bark 0.5% w/w. The compound turned permanently pink on exposure to conc. ammonia vapor which is a positive test for dihydroxy-1,4-benzoquinones. The UV-Vis spectrum displayed absorption bands at 292 nm and 450 nm which is a characteristic feature of a dihydroxy-1,4-benzoquinone moiety (Nakata et al.

Ogawa and Natori, 1968). The <sup>1</sup>H NMR of the compound exhibited six signals. There was no signal corresponding to the quinonoid proton which usually appears at 6.50 - 5.0 ppm. This suggests that the quinonoid ring is fully substituted. A singlet peak due to phenolic hydroxyl protons bonded to carbonyl appeared at 7.84 ppm. Peaks were also displayed at 4.27 ppm (H, t, J=7.92 Hz), 2.06 ppm (m) and 1.26 to 1.46 ppm (m) which are unusual for dihydroxy-1,4-benzoquinones. Integration of the peak at 7.84 ppm with respect to that at 4.27 ppm corresponded to two hydroxyl groups. The proton-proton correlation (homonuclear COSY) experiments showed coupling between proton absorbing at 4.27 ppm and the multiplet at 2.06 ppm which in turn coupled with the multiplet at 1.26 to 1.46 ppm. Integration of the peak 2.05 ppm with respect to that at 4.27 ppm corresponded to two protons. The peak at 4.27 ppm suggested the presence of a proton sitting on the same carbon with a methylene group while the multiplet at 2.06 ppm could possibly be assigned to a methylene group sitting on the same carbon atom with a proton. The multiplet at 1.26 to 1.46 ppm could be due to the remaining methylene groups in the decyl chain. Furthermore, irradiating the peak due to benzylic methylene protons at 2.41 ppm did not affect the triplet at 4.27 ppm and the multiplet at 2.06 ppm, proving that neither the proton nor the decyl group and the alkyl side chain are vicinal. As expected for alkylated dihyroxy-1,4-benzoquinones, there was a triplet at 2.41 ppm (J=7.44 Hz) which was assigned to the benzylic methylene protons. Integration of this peak relative to the set at 4.27 ppm corresponded to four protons calculated for two methylenes. This suggested that the compound probably consists of two alkyl substituted dihydroxy-1,4benzequinones bridged by a carbon holding a decyl group and a proton. Integration of the multiplet at 1.26 to 1.46 ppm relative to the peak at 4.27 ppm corresponded to 52 protons calculated for 26 methylenes. The terminal methyl protons appeared at 0.89 ppm. Integration of this peak relative to that at 4.27 ppm revealed the existence of nine protons and thus accounting for three methyl groups. On comparing the  ${}^{13}$ C NMR spectral data of (77) with that of embelin (10), there was only one significant difference for the signal due to C-6. The C-6 peak, which in

(10) is centred at 103.80 ppm appeared at 115.1 ppm, corresponding to a quartenary carbon bearing the bridging carbon holding the decyl group and the proton. This seems to confirm the presence of two alkyl dihydroxy-1,4benzoquinones in (77). Compound 77 differs from 76 in that it has got two hydroxyl groups less than the latter. It is reasonable to assume that 77 is formed from 76 due to loss of a molecule of water. The mass spectrum showed significant fragments ions at 320, 294, 180, 139 and 125. The peak at m/e 294 evidenced the existence of either embelin (10) or myrsinone (55) at part of the molecule with the latter being ruled out as additional peaks due long range coupling between the alkyl side chain and the quinonoid proton were absent in the <sup>1</sup>H NMR spectrum. Also evident in the mass spectrum was the strong fragment peak at m/e 154 and low abundance peak at m/e 153. Such fragmentation patterns are charateristics of embelin (10) and myrsinone (55). Having ruled out (55) with the help of  ${}^{1}H$  NMR data, mass spectral data strongly favoured the existence of two embelin moeities joined together by a carbon bearing a proton and a decyl group. The spectral data strongly support the compound as decylanydrovilangin (77)



Atom	<sup>1</sup> H (ppm)	<sup>13</sup> C (ppm)
1,1'	-	-
2,2'	-	-
3,3'	-	116.32
4,4'	-	-
5,5'	-	-
6,6'	-	115.10
H- <u>C</u> -CH <sub>2</sub> -(CH <sub>2</sub> ) <sub>8</sub> -CH <sub>3</sub>	4.27 (t,J=7.92 Hz)	28.63
H-C- <u>C</u> H <sub>2</sub> -(CH <sub>2</sub> ) <sub>8</sub> -CH <sub>3</sub>	2.05 (m)	28.18
H-C-CH <sub>2</sub> -(CH <sub>2</sub> ) <sub>8</sub> - <u>C</u> H <sub>3</sub>	0.9 (t J=6.3 H)	14.33
7,7'	2.40 (t,J=7.44 Hz)	29.55
H-C-CH <sub>2</sub> ( <u>C</u> H <sub>2</sub> ) <sub>8</sub> -CH <sub>3</sub> ,		32.33,32.15.30.43,
8,8',9,9',10,10',		29.84,29.77,29.74,
11,11',12,12',13,13',	1.45 - 1.25 (m)	29.55,22.90,22.79
14,14',15,15',16,16'		
17,17'	0.9 (t J=6.3 Hz)	14.33
2, 2' - OH	7.84 (s, D <sub>2</sub> O exchangeable)	_

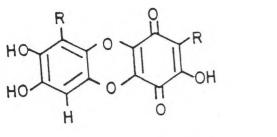
Table 6:  ${}^{1}$ H and  ${}^{13}$ C NMR (CDCl<sub>3</sub>) for 77

The next compound was a red substance (56) Rf 0.53 (solvent system: same as that for 10), m.p 166- 167°C was isolated from the  $CH_2Cl_2$  eluent. It turned pink on exposure to conc. ammonia vapor and therefore could be a dihydroxy-1,4-benzoquinone. Its UV-Vis spectrum displayed a peak at 273.5 nm, 375 nm and 460 nm. The <sup>1</sup>H NMR exhibited a triplet for the alkane terminal methyl protons at 0.90 ppm which upon integration corresponded to six protons thus suggesting the presence of two methyl groups. The signal at 1750-1.20 ppm was assigned to the

chain methylene protons and it corresponded to thirty six protons. There was a triplet at 2.40 (J=7.2 Hz) ppm which could be attributed to the benzylic methylene protons on the quinonoid ring and another triplet at 2.74 (J=7.2) ppm which probably originated from the methylene attached to the aromatic benzenoid system. The compound also exhibited a singlet peak at 7.26 ppm which upon integration corresponded to one proton. The mass spectrum showed the parent molecular ion peak at m/e 570 nm accounting for  $C_{34}H_{50}O_7$ . The presence of three hydroxyl groups in the molecule was suggested by the characteristic <sup>1</sup>H NMR signals at 7.05, 6.02, 5.84 ppm which were D<sub>2</sub>O exchangeable. All these spectroscopic data suggest that (56) is a fused bisalkylbenzoquinone system in which one of the quinonoid rings has been aromatised.

Since the chemical shift of the side chain alkyl group on the quinonoid ring of (56) is quite similar to that of (10), it can be assumed that it has adjacent hydroxyl groups. Considering that there is a single aromatic proton in the <sup>1</sup>H NMR of (56), it can comfortably be concluded that it is not vicinal to the alkyl side chain because such an arrangement leads to long range coupling as in the <sup>1</sup>H NMR spectrum of myrsinone (55). The <sup>13</sup>C NMR showed peaks at 146.90 (C-5), 149.30 (C-6), 119.0 (C-11) and 108.0 ppm suggesting a dioxane type of fusion between the benzenoid and quinonoid rings. This was further supported by calculating the carbon chemical shifts around the aromatic ring basing on the fact that the benzenoid and the quinonoid rings are joined to two oxygen atoms as in structure (56) which gave the following results: C1-118.30, C2-146.40, C3-139.10, C4-106.50, C5-142.30 and C6-143.10 ppm. These results could be matched with peaks in the spectrum at 114.32, 149.40, 141.0, 108.34, 146.90 and 114.30 ppm respectively. A heteronuclear COSY experiment showed a three-bond interaction between the ring proton and carbon atoms absorbing at 149.40 and 149.30 ppm the two carbons C-2 and C-6 which can be interchanged. Complete assignment for all the carbon atoms were made as C7-169.40, C8-148.10, C9-107.90, C10173.30, C11-119.00 and C12-108.00 ppm. The spectroscopic data seem to support 56 as myrsinaquinone.

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 $R \equiv C_{11} H_{23}$  $R \equiv C_{13} H_{27}$ 

Atom	<sup>1</sup> H (ppm)	<sup>13</sup> C (ppm)
1	-	114.33
2	-	149.40
3	-	141.00
4	7.26 (s)	108.34
5	-	146.90
6	-	149.30
7	-	169.40
8	-	148.10
9	-	107.90
10	-	173.30
11	-	111.90
12	-	108.00
13	2.74 (t, J=15.0 Hz)	29.60
13'	2.40 (t,J=7.2 Hz)	29.40
14,14',15,15'.16,16' 17,17',18,18',19,19' 20,20',21,21'	1.49 - 1.25 (m)	31.80, 29.30,29.10, 23.80, 22.60
22, 22'	0.90 (t,J=6.3 Hz)	14.00
2 - OH	6.02 (s, D <sub>2</sub> O exchangeable)	1020
3 - OH	5.54 (s, $D_2O$ exchangeable)	-
8 - OH	7.05 (s, $D_2O$ exchangeable)	-

**Table 7:** <sup>1</sup>H and <sup>13</sup>C NMR data (CDCl<sub>3</sub>) of 56.

#### CHAPTER THREE.

3.0.0. EXPERIMENTAL.

#### 3.1.0. GENERAL.

#### 3.1.1. Instruments and spectra.

Plant materials were ground into powder using a Willey mill. Gallenkhamp melting point apparatus was used for melting point determination.

The Ultra Violet/Visible spectra were obtained using Beckman DU-50 spectrophotometer. High resolution <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were obtained from University of Botswana. Mass spectra were obtained from International Center of Insect Physiology and Ecology (ICIPE).

#### 3.1.2. Plant materials.

Parts of <u>Embelia schimperi</u> were collected from Kericho which is 2400m above mean sea level and 260 Km west of Nairobi while parts of <u>Embelia keniensis</u> were collected from Limuru which is 35 Km west of Nairobi. The plant species were identified at the herbarium in the department of Botany, University of Nairobi. The fruits and roots' bark brought to the laboratory were cut-up dried and ground into powder before solvent extraction.

#### 3.1.3. Qualitative analysis

Thin layer chromatography (TLC) was used analytically to monitor the composition or homogeneity of fractions from columns during column chromatographic separations. The 2,5-dihydroxybenzoquinone compounds are highly unstable on non-deactivated silica gel. For this reason, the commercial analytical TLC plates were de-activated by impregnating with 3% oxalic acid in methanol. This was done by dipping the TLC plates for 2-3 minutes and then drying them for about 5 minutes in an oven at a temperature of 110°C. The plates were developed in one of the following solvent systems; n-hexane, ethyl acetate. acetic acid (85:10:5), n-hexane, ethyl acetate. acetic acid (80:15:5) or n-hexane.

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ethyl acetate, acetic acid (75:20:5). The organic compounds were located on the plates by exposing them to either ammonia vapor, U.V radiation and/or iodine vapor. The Rf value for each compound was then determined and recorded.

#### 3.1.4. Quantitative analysis

Column chromatography was employed for the separation of most of the compounds although in some cases preparative thin layer chromatography had to be used to effect separation of compounds whose Rf values were very close.

The Merck silica gel used was normally de-activated using 3% oxalic acid and gravity eluted using n-hexane, dichloromethane and ethyl acetate (or various combinations of these solvents). Solvents were removed from various fractions in <u>vacuo</u> using rotatory evaporator. In cases of incomplete separation, preparative TLC was employed for further purification. A slurry of silica gel in 3% oxalic acid solution was used to prepare plates of 2 mm thickness on a 20 cm x 20 cm glass slab. These were then air dried overnight and transferred to the oven at a temperature of 110°C for about 30 minutes. De-activation of silica gel for use in column chromatography was achieved by soaking in 3% oxalic acid solution. The solution was decanted as much as possible and dried in an oven at a temperature of 110°C for 30 minutes.

#### 3.2.0. PRELIMINARY WORK

# 3.2.1. Determination of quinonoid pigments in root bark of *Embelia schimperi* and *Embelia keniensis* using Thin Layer Chromatography.

Dry ground root bark of <u>Embelia schimperi</u> and <u>Embelia keniensis</u> were extracted with cold ethyl acetate and the solvent removed in vacuo leaving dark gummy solids. Qualitative TLC analysis of extract using the solvent mixture n-hexane, ethyl acetate, acetic acid (85:10:5) for development revealed the presence of the following components: Rf 0.9, 0.55, 0.53 and 0.47. The first three turned pink when the plate was exposed to ammonia vapor. The fourth compound was not

ammonia sensitive but absorbed UV radiation . Qualitative TLC analysis of the extract from and <u>Embelia keniensis</u> revealed that the components ocurred in very trace amounts and thus was not followed for large scale extraction.

3.2.2. Thin Laver Chromatographic examination of E. schimperi and E. Keniensis fruits for quinonoid pigments.

Extraction of ground fruits of <u>Embelia schimperi</u> and <u>Embelia keniensis</u> were done as in 3.2.1. TLC analysis of the extract from <u>Embelia schimperi</u> showed presence of the following components: Rf 0.9, 0.64, 0.62, 0.55 and 0.49. All the spots turned permanently pink on exposure to ammonia vapor. TLC analysis of the extract from <u>Embelia keniensis</u> showed that components ocurred in very trace amounts and thus was not followed.

### 3.3.0. <u>LARGE SCALE EXTRACTION AND ISOLATION OF</u> COMPOUNDS FROM *EMBELIA SCHIMPERI*.

#### 3.3.1. Extraction of root bark.

A quantity of 2 Kgs of dry ground root bark of <u>E.schimperi</u> was soaked in ethyl acetate (3 liters) in the cold for 48 hrs while mechanically stirring. The root bark residue was further extracted for another 24 hours to ensure that the components were exhausively extracted. The extract was then filtered under suction and concentrated in vacuo using rotary evaporator leaving 97.3 g of a sticky dark paste.

#### 3.3.2. Fractionation of the extract.

A sample of the extract (41 g) was pre-adsorbed on dry de-activated silica gel and subjected to column chromatography in a column packed under n-hexane with 500 g of de-activated silica gel. The column was exhaustively eluted with nhexane, dichloromethane and ethyl acetate respectively and a fraction of each solvent was collected separately. The n-hexane fraction contained mainly fatty acid material; the dichloromethane fraction (with aid of TLC analysis) was found to contain the quinonoid pigments. The ethyl acetate fraction was not followed because the components occured in trace amounts on TLC.

#### 3.3.3. Isolation of compounds from the dichloromethane extract concentrate.

The concentrate (5.93 g) was found to contain a compound, embelin, (10), Rf 0.55 (solvent system: n-hexane, ethyl acetate and acetic acid - 80:15:5) in large quantities than others and therefore masked the minor ones on t.l.c. This prompted repeated fractional crystallization of the extract in dichloromethane until crystals of embelin were no longer in the mother liquor. A quantity of 3.22 g of the concentrate was pre-adsorbed on de-activated silica gel and subjected to a column packed under n-hexane with de-activated silica gel. The polarity of the system was increased with increased addition of dichlororomethane up to 100% dichloromethane. The column was finally washed with 5% methanol in dichloromethane. A total of 32 fractions were collected and the eluate were monitored by t.l.c. Fractions (1-3) from n-hexane gave mainly fatty acids and their derivatives. These were combined, the solvent recovered and the mixture discarded. Eluate (4-11) yielded a yellowish substance which on t.l.c using nhexane, ethylacetate and acetic acid (80:15:5) as a solvent system afforded components with Rf values 0.9, 0.55, and 0.47. The latter component was not ammonia sensitive but absorbed UV radiation. The fractions were combined and subjected to column chromatography. Elution of the column with n-hexane and dichloromethane (70:30) afforded four fractions which when monitored on t.l.c with the above solvent system showed components of Rf values 0.53 and 0.49 both of which occurred in very small amounts. Elution of the column with dichloromethane afforded a component with Rf value 0.53 and traces of two of the earlier components whose Rf values are 0.55 and 0.49 (same solvent system).

#### 3.3.4. Purification of dichloromethane concentrate fractions.

Fractions (4-11) were combined and further subjected to column chromatography on de-activated silica gel with n-hexane - dichloromethane (70:30) as eluent. Fractions of 5 ml each were collected and monitored by t.l.c. The

corresponding fractions were combined, solvent evaporated in vacuo and the respective constituents crystallized from dichloromethane - methanol (9:1) to give components with Rf values 0.9 (77, 16 mg), 0.55 (10, 600 mg) and 0.47 (uncharacterized compound, 120 mg). The first two spots turned permanently pink on exposure to concentrated ammonia vapor while the third was UV sensitive. Fractions eluted with n-hexane-dichloromethane (1:1) were concentrated in vacuo to afford a residue (0.39 g) which was subjected to column chromatography over treated silica gel (20 g). Elution was accomplished with the same solvent system. Components with Rf values of 0.55 (10, 64 mg) and 0.53 (56, 19 mg) were obtained. Both spots changed to pink on exposure to concentrated ammonia vapor. Compound (56) was recrystallized from dichloromethane - methanol (9:1). The eluate obtained from dichloromethane were further resolved by column chromatography over treated silica gel before being subjected to preparative TLC to get rid of traces of more polar compounds from (56). The acid treated preparative t.l.c plates used for separation were developed three times. Additional 10 mg of pure (56) was obtained. Eluate from dichloromethane - methanol (95:5) contained mainly the more polar components occurring in very trace amounts.

Compound (10) - embelin, Rf 0.55 (characterized before from the same plant) was an orange compound with m.p 140-142°C (same as in the literature). The  ${}^{1}$ H ,  ${}^{13}$ C NMR and mass spectra were not obtained.

Compound with Rf 0.9 (77), decylanhydrovilangin is an orange compound with mp 119-121  $^{0}$ C (unreported in the literature). UVA max (MeOH) at 292 nm and 450 nm.  $^{1}$ H and  $^{13}$ C NMR data are displayed in table 6. MS (70 eV): m/e (%) M<sup>-</sup>722, 294(26), 155 (46), 154 (100), 153 (26), 142 (28) 125(11)

Compound (56) - myrsinaquinone is a red compound with Rf 0.53 and mp 166 - 167°C. UVA <sub>max</sub> (MeOH) at 273.5 nm and 430 nm. The 1H and <sup>13</sup>C NMR are in table 7. MS (70 eV): m/e (%) M<sup>+</sup> 570 (100), 598 (3), 552 (1.50), 429 (7), 346 (28), 320 (8), 289 (13), 205 (9). Compound (KCP -02) is a yellowish oil with Rf 0.47 . UVA  $_{max}$  (MeOH) at 275.5 nm and 428nm. This compound was not characterized due to lack of sample. 3.3.5. Extraction of E. schimperi fruit .

A quantity of 800 g of dry ground <u>E</u>. shimperi fruit was soaked in ethyl acetate (2 liters) in the cold for 48 hrs while mechanically stirring. The fruit residue was further extracted for another 24 hrs. The latter extraction was to ensure that all the components were exhaustively extracted. The extract was then filtered under suction and concentrated in vacuo using rotary evaporator leaving 55.78 g of a sticky dark paste.

#### 3.3.6. Fractionation of the extract.

## CHIROMO LIBRAS

A quantity of 55 g of the extract was pre-adsorbed on dry de-activated silica gel and subjected to column chromatography in a column packed under n-hexane with 550 g of de-activated silica gel. The column was exhaustively eluted with hexane, dichloromethane and ethyl acetate respectively and a fraction of each solvent collected separately. The n-hexane fraction contained mainly fatty acid material; the dichloromethane eluent (with aid of t.l.c analysis) was found to contain the quinonoid pigments. The ethyl acetate fraction was not followed because the components occured in very trace amounts.

#### 3.3.7. Isolation of compounds from dichloromethane eluant.

A sample of 30 g was obtained after concentrating the dichloromethane eluant in vacuo. It was found to contain a compound. embelin, (10), Rf 0.55 (solvent system: ethyl acetate , dichloromethane and ethyl acetate - 80:15:5) in larger amounts than others and therefore masked the minor ones on t.l.c. This prompted repeated crystallization of the extract in dichloromethane until crystals of the compound were no longer in the mother liquor. From the residue (i.e after removal of 10), a quantity of 15 g of which was pre-adsorbed on dry de-activated silica gel and subjected to column chromatography in a column packed under nhexane with de-activated silica gel. The polarity of the system was increased with addition of volumes of dichloromethane or ethyl acetate. A total of 29 fractions (200 ml each) were collected and the eluate were monitored with de-activated t.l.c plates.

Fractions 1-5 (eluted with 10% dichloromethane in n-hexane) showed a spot with Rf 0.9 and which turned permanently pink on exposure to conc. ammonia vapor. This was re-purified by crystallizing in dichloromethane methanol (9:1) to give 26 mg of (77). Fractions 6-12 were mainly composed of components with Rf values 0.9, 0.64 and 0.62. These fractions were combined. solvent evaporated and the residue subjected to repeated column chromatography eluting with n-hexane - dichloromethane (30:70) as a mobile phase to give 3 mg more of the component with Rf value of 0.9 (77), 0.64 (76, 5 mg) and 0.62 (21. 110 mg) respectively. Fractions 13-17 afforded a spot which turned permanently pink on exposure to conc. ammonia vapor. Upon crystallization, 750 mg of embelin (10) was obtained. Fractions 18-24 (eluted with 50% dichloromethane in n-hexane) afforded 5 mg of the component with Rf value of 0.62 (21) and a pink compound whose Rf value was 0.49 (KCP-06, 30 mg). Fractions 25-29 (eluted with 100% dichloromethane) were combined and subjected to column chromatography over treated silica gel using n-hexane - ethyl acetate (90:10) followed by the same solvent (80:20). Twelve fractions of 100 ml each were collected and t.l.c analysis revealed a mixture of components. Fractions were combined based on their t.l.c profiles and the solvent evaporated. Repeated preparative t.l.c (on de-activated silica gel) with n-hexane - ethyl acetate (90:10) yielded myrsinaquinone (56, 44 mg). Eluate from n-hexane - ethyl acetate(70:30) contained mainly the more polar components occurring in trace amounts.

Compound (KCP-06) is a pink and amorphous compound with Rf 0.49 (solvent system: n-hexane - ethyl acetate - acetic acid - 80:15:5). UV $A_{max}$  (MeOH) at 280 nm and 425 nm. MS (70 eV): m/e (%) M<sup>+</sup> 530 (47.8), 512 (11.2), 390 (30), 333 (10), 250 (12.4). 249 (13.6). 154 (13.6). 97 (63.6), 83 (64.8), 69 (66), 57 (100), 43 (90), 41 (43). This compound was not characterized because the amount isolated was only enough for biological activity tests.

Compound (76) - decylvilangin, is an orange compound with Rf 0.64 and m.p 121-123°C. Its UVA<sub>max</sub> (MeOH) at 290 nm and 430 nm. The <sup>1</sup>H and <sup>13</sup>C NMR data are displayed in table 5. MS (70 eV): m/e (%) M<sup>+</sup> 740 (19.6), 334 (7.2), 333 (35.6), 294 (43.6), 168 (7.2), 154 (100), 153 (19.6). 142 (20), 125 (7.2).

#### 3.4.0. BIOASSAY.

#### 3.4.1. Brine shrimp lethality test.

These were performed according to McLaughlin <u>et al</u> which briefly was as follows:<u>Artemia salina</u> (Brine shrimp) were hatched into nauplii (larvae) after 48 hours in artificial "sea water" in the dark and allowed to swim to a lighted area through perforations in the hatching vessel. Pure compounds (kcp-02), 21, kcp-06 and 76 were subjected to this test. The test solutions were prepared at concentrations of 5, 10, 15, 20, 25, 50, and 100 µg/ml (1 ml each) in triplicate in 2 ml dram vials in acetone. The acetone was allowed to evaporate and replaced by "sea water" containing nauplii (10 each). The number of nauplii still alive were monitored after 12 and 24 hours. The control experiments were performed in vials with no samples. The data was processed in a simple program on a personal computer to estimate  $LC_{50}$  values with 95 % confidence intervals for statistically significant comparison of potencies. The data collected after 24 hours is summarized in Table 1.

#### 3.4.2. Anti-feedant test.

Anti-feedant activity tests were done on embelin (10), kcp-02 and 21 against Locusta migratoria migratorioides. "Choice" and control experiments were set up with ten mid-5<sup>th</sup> instar nymphs which were starved for 24 hours before feeding them with the test materials (Morgan <u>et al</u>,1968). A 100  $\mu$ g/ml (dissolved in dichloromethane) of each sample was sprayed on a 2.75 cm<sup>2</sup> whatman No.1 filter papers previously soaked in 0.25 M sucrose solution. The control filter papers were only treated with 0.25 M sucrose solution. All the papers were dried in an oven at 40 °C. The data collected after 8 hours is summarized in Table 2, section 2.0.3.

3.4.3. Anti-microbial test.

<u>Candida albicans.</u> <u>Trichophyton metagrophyte Microsporium gypsum.</u> and <u>Escherichia coli</u> was each spread on a petri dish smeared with agar as a growth medium. Concetrations of 50, 100 and 400  $\mu$ g/ml of 77, 56, 21, 10 and KCP-06 were applied into "wells" made in the medium and incubated for a period of between one and three days for any inhibition zones to develop. Results are discussed in section 2.0.4.

#### 3.4.4. Larvicidal Test.

Larvicidal test were done for (21) according to Zebitz (1986) and Mwangi and Rembold (1988). To each jar holding 40 ml of 0.09% sodium chloride, twenty  $2^{nd}$  instar mosquito larvae were introduced and immediately treated with 100, 75, 50, 25 and 10 µg/ml of (21). The results of this test on day 13 are shown on table 3, section 2.0.5.

#### 3.4.5. Stored Products Pests Test.

Natural embelin (10) was tested against <u>Sitophilus zeamais</u> and <u>Anthoscelides</u> obtectus- the two storage pests for maize and beans respectively to evaluate its ability to act as a pesticide. The insects were obtained from standard laboratory cultures maintained in the insectary (Department of zoology). A culture of <u>S.zeamais</u> was maintained on whole maize while <u>A</u> obtectus was maintained on beans. The solid test compound was incorporated into 1 Kg samples of uninfested beans and maize each in 150 x 150 mm jars at rates of 0.02, 0.04, 0.05, 0.08, 0.1, 0.2 and 0.4% (w/w) and twenty (20) mixed adults of each of the two insects were added. The experiments were performed in three replicates. Data (after five months) on adult emergence and productivity (progeny/adult-days) are summarized in fig.1 (in the case of <u>S. zeamais</u>) and fig.2 (in the case of <u>A. obtectus</u>)

#### CHAPTER FOUR.

#### 0.0. COMMENTS AND CONCLUSION.

Embelia schimperi has the ability to accumulate different long alkyl side hain benzoquinone compounds.

A number of biological activity tests were performed using these ompounds and a few cases will be highligted.

Clearly embelin (10) can be developed as a commercial insecticide that can e used to protect stored beans and maize grains from attack by their respective reevils over a long period of time. It can also be applied on field crops to deter ocusts from feeding on them. The anti-feedant activity is demonstrated by the arge relative antifeedant percentage (RAP) value of 10.

Another compound that can be developed for commercial use is nethylvilangin (21). This compound was found to disrupt the process of netamorphosis in the mosquito larvae, *Aedes aegypti*. In fact, when 100  $\mu$ g/ml of 1 was applied into a pond containing 2<sup>nd</sup> instar mosquito larvae, only a few nderwent metamorphosis to the 3<sup>rd</sup> instar larvae and none attained the adult stage. This biological activity can be used to reduce population of mosquitoes.

Since the concentration of the benzoquinone compounds is higher in the erries of  $\underline{E}$ . schimperi it is important to encourage the use of the berries to achieve ptimum results and to avoid damage to the shrub.

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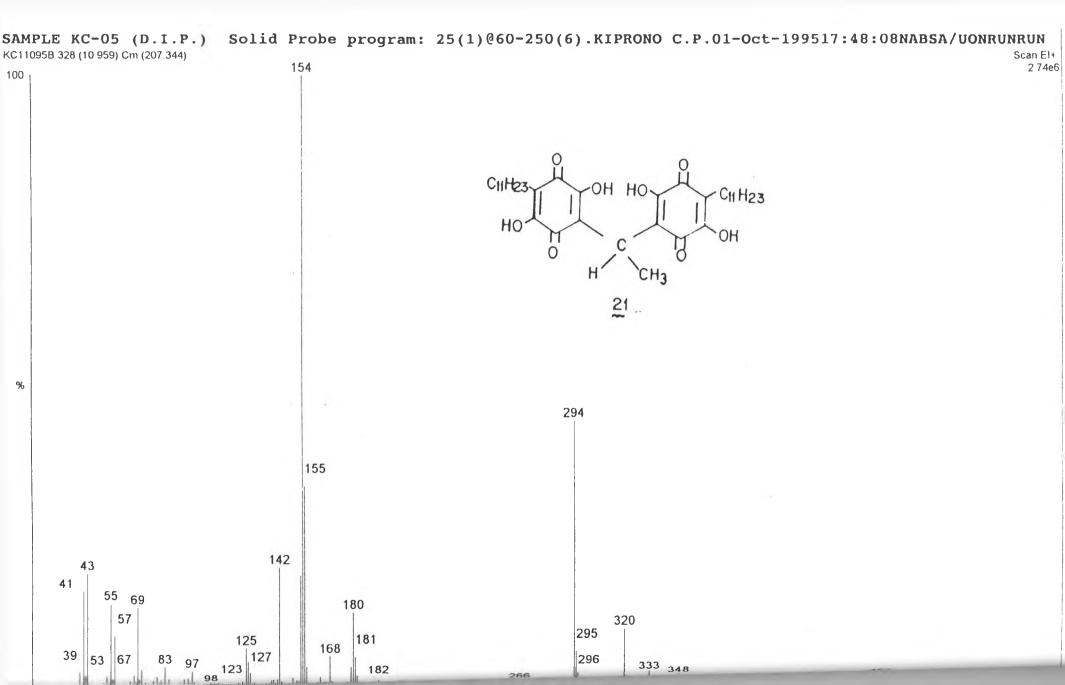
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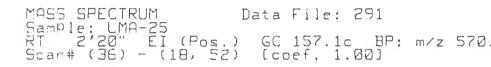
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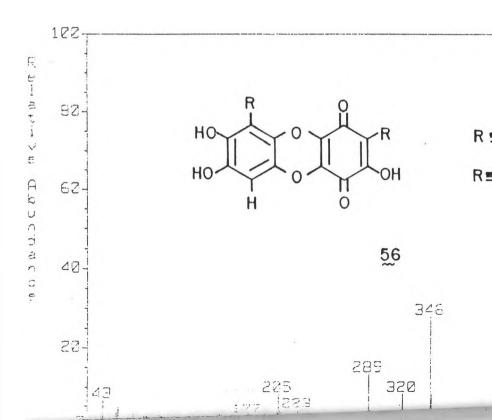
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SPECTRA

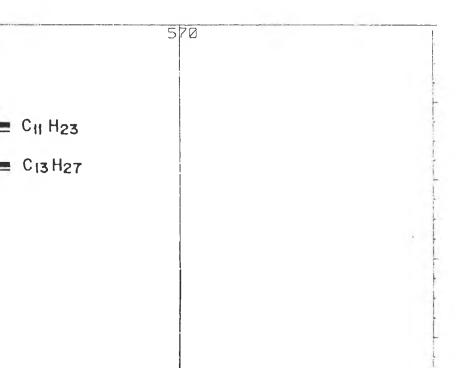
Mass spectra of isolated compounds.

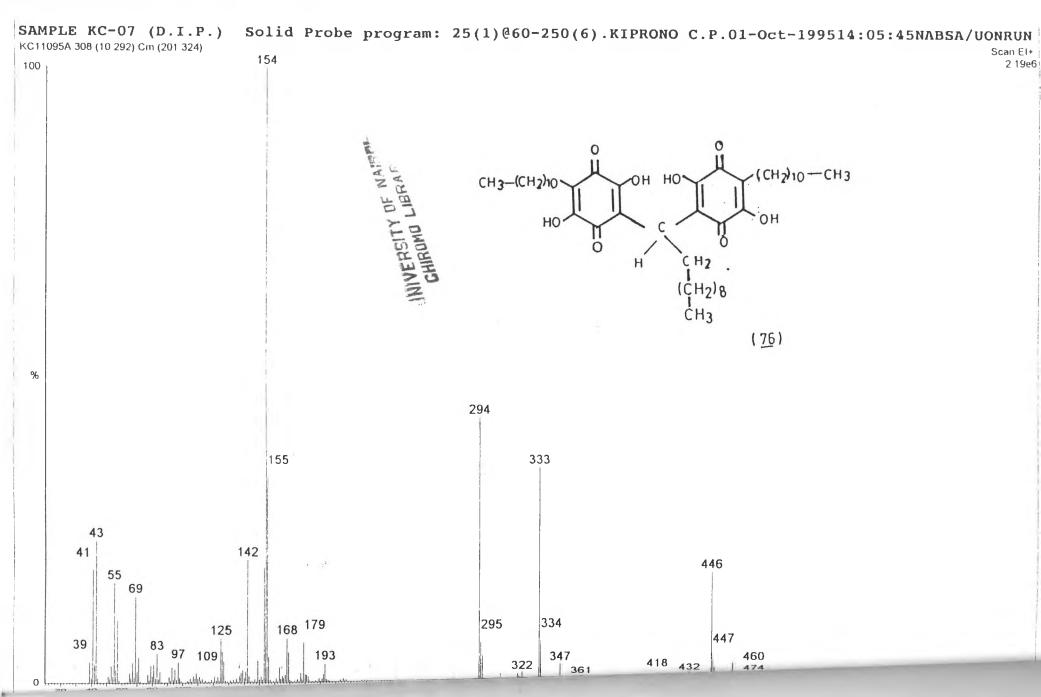


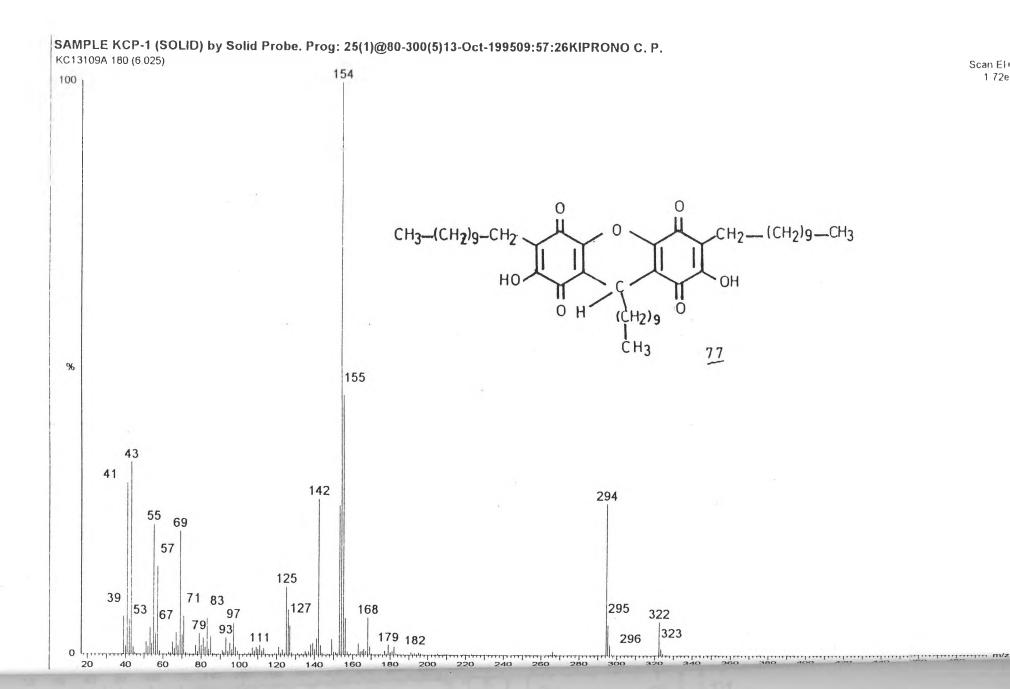




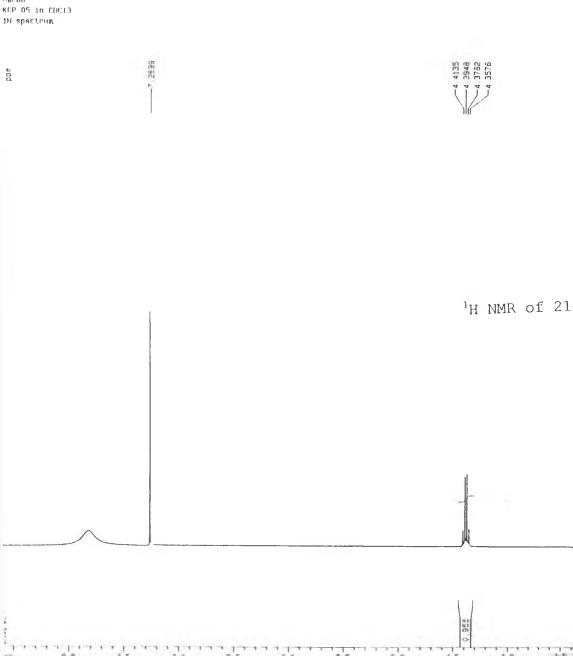
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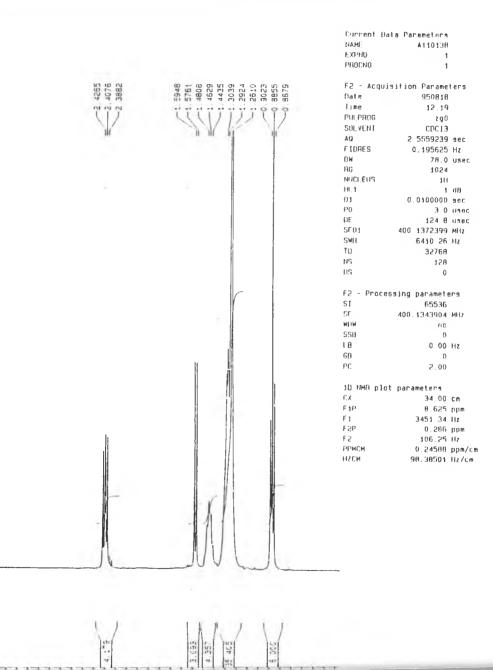


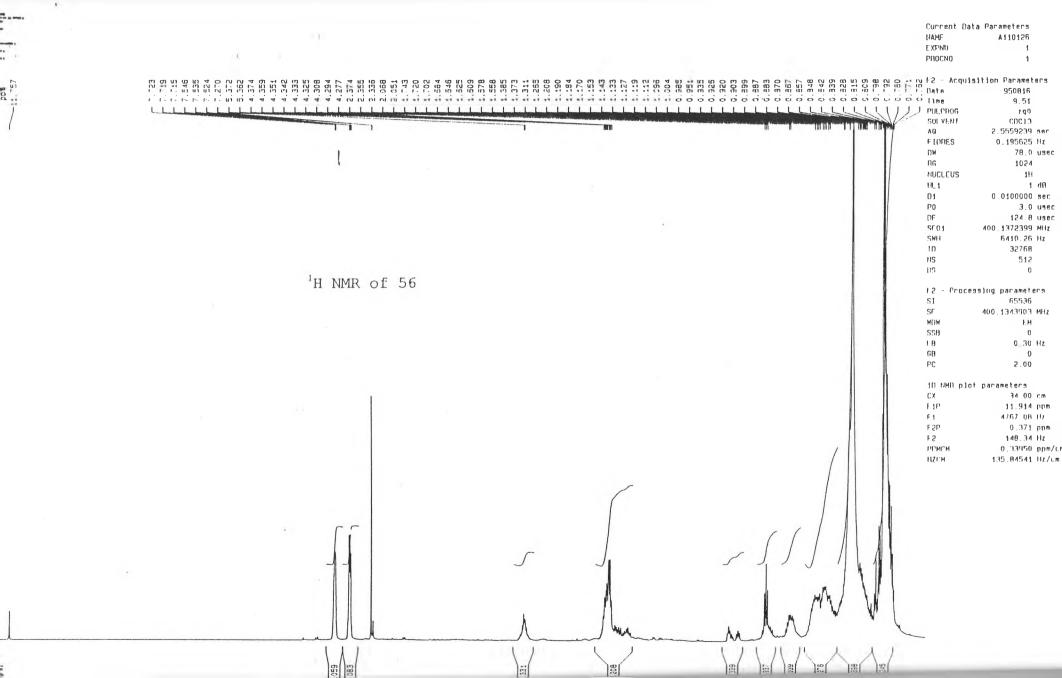


<sup>1</sup>H NMR of isolated compounds.

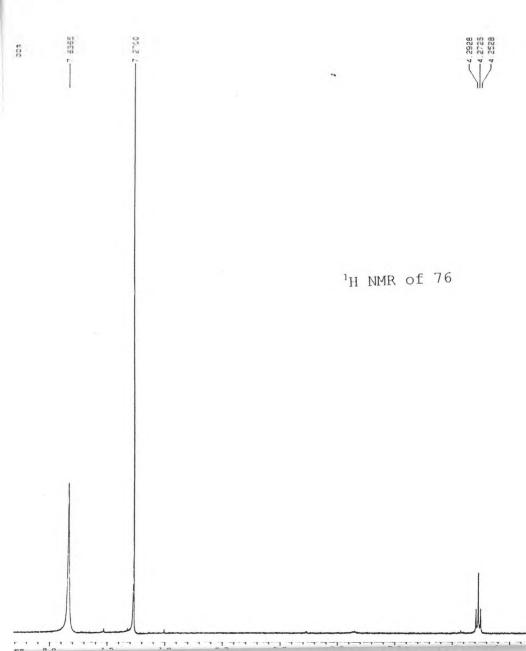


Jacob

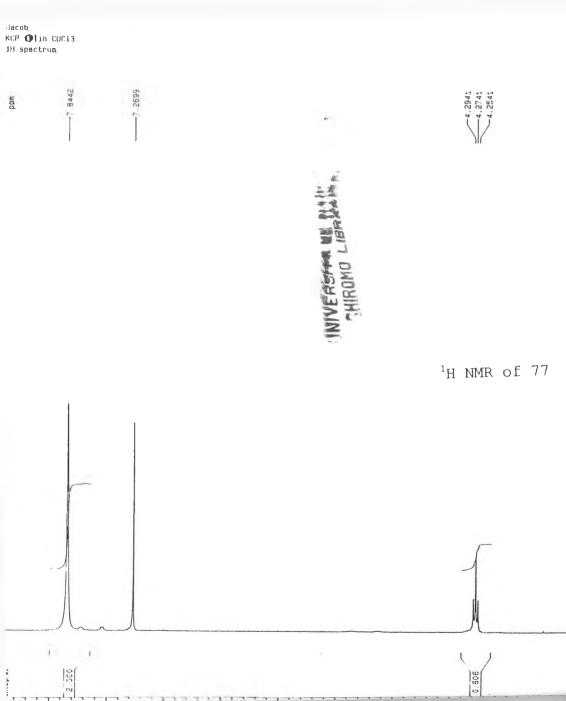


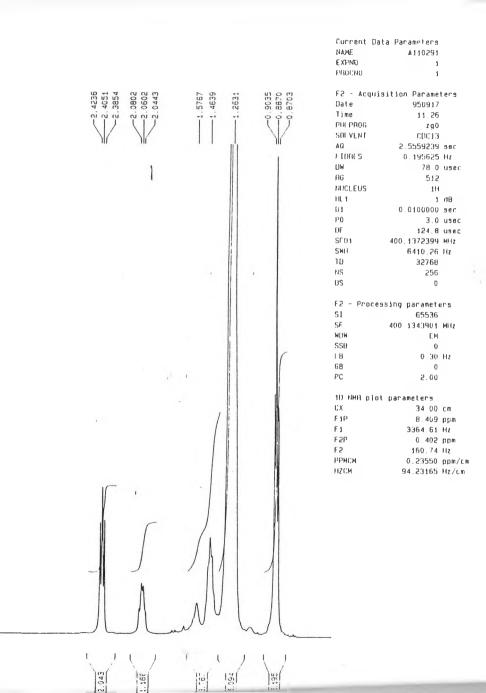


Jacob KCP-07 in CDC13 3H spectrum



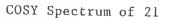
	Current Data Parameters           NAME         A110296           EXPND         1           PROCNO         1           F2 - Acquisition Parameters           Data         950919           limu         8.29           PULPROG         2.00           SOLVENT         CDC13           AQ         2.5559239           PUTS         0.195625           DW         78.0           DW         78.0           PUCLEUS         111           HL1         1           DL1         0.100000           PO         3.0           USC         124.8           USE         SF01           A00         1322399           MUCLEUS         111           HL1         1           DI         0.100000           SF01         400           SF01         400           SC01         20           SWIL         6410.26           IN         3276H           NS         256           IS         0
	F2 - Processing parameters SI 65536 SF 400.1343098 MHz HUW no SSH 0 IB 0.00 Hz GB 0 PC 2.00 ID NMR plot parameters CX 34.00 cm FJP 8.339 ppm F1 3336.63 Hz I 2P 0 367 ppm F2 146.75 Hz PPMCM 0.23447 ppm/cm H2CM 93 82017 Hz/cm

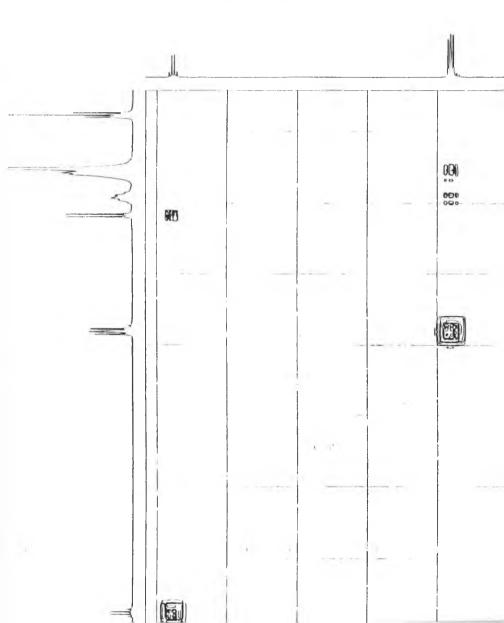


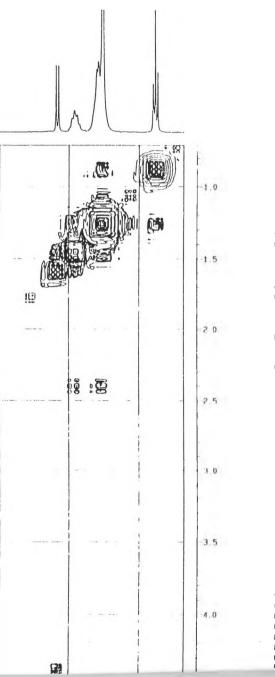


Cosy Spectra of some isolated compounds

Hacob KCP-05 in CUC13 Nocay



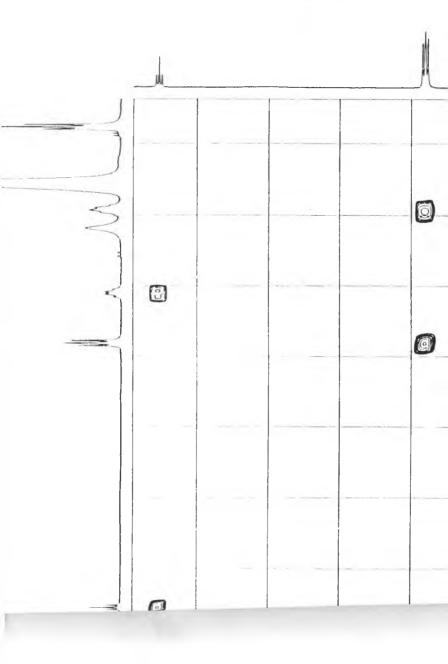


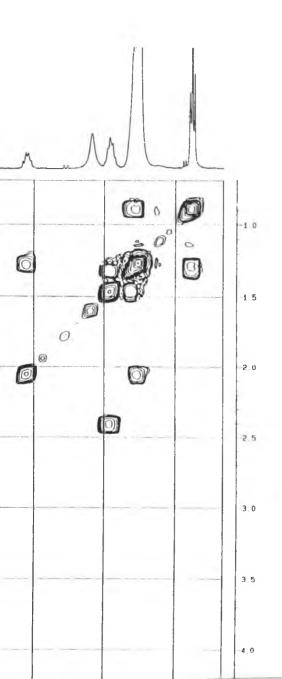


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EXPN0         3           PROCN0         1           F2 - Acquisition Parameters           Date         950818           Time         15.32           PUB PROG         noesytp           Set V.N.         CDC13           AQ         0.1054920 sec           FEDUES         4.740595 H7           DW         103.0 usec           PUB PROG         0.80000 sec           PUG FUS         344           PUG FUS         344           PUG 1         1.08           2.0000000 sec         P1           PUG 0.000000 sec           P1         12.0 usec           D0         0.000000 sec           DF         164.8 usec           SF01         400.1306359 MH2           SMH         4854.37 H2           TU         1024           NG         2           DS         4           NH0         0.000100 mec           F1         Acquisition parameters           SF01         400.1366 MH2           FUBES         18 962279 H2           SM         32.132 ppm           F2 - Processing parameters           SI         1024 </td <td></td> <td></td> <td>ane ter 5</td>			ane ter 5		
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Date         950818           lime         15.32           PUR PROG         noesytp           Set VFN         CDC13           AQ         0.1054920 sec           FIDUES         4.740595 Hz           DW         103.0 usec           HIG FUS         1H           HIT         1 dB           DI         2.000000 sec           PIC FUS         1H           HIT         1 dB           DI         2.000000 sec           PI         12.0 usec           DO         0.000000 sec           DF         164.8 usec           SF01         400.1306359 HHz           SMH         4854.37 Hz           TU         1024           NG         16           US         4           NO         0.000100 nec           F1         Acquisition parameters           NHO         2           DU         255           SF01         400.1366 MHz           FUNES         18 76229 Hz           SN         12.132 opm           F2 - Processing parameters           SI         1024           SN         12.132 opm					
Date         950818           lime         15.32           PUR PROG         noesytp           Set VFN         CDC13           AQ         0.1054920 sec           FIDUES         4.740595 Hz           DW         103.0 usec           HIG FUS         1H           HIT         1 dB           DI         2.000000 sec           PIC FUS         1H           HIT         1 dB           DI         2.000000 sec           PI         12.0 usec           DO         0.000000 sec           DF         164.8 usec           SF01         400.1306359 HHz           SMH         4854.37 Hz           TU         1024           NG         16           US         4           NO         0.000100 nec           F1         Acquisition parameters           NHO         2           DU         255           SF01         400.1366 MHz           FUNES         18 76229 Hz           SN         12.132 opm           F2 - Processing parameters           SI         1024           SN         12.132 opm					
lime       15.32         PUR PROG       noesytp         SURVENT       CUCI3         A0       0.1054920 sec         FIDHES       4.740595 Hz         DM       103.0 unre         HB       256         PUR FUS       1H         HC       1 dB         D1       2.0000000 sec         PIC FUS       1H         HC.1       1 dB         D1       2.0000000 sec         PIC FUS       1H         HC.1       1 dB         D1       2.0000000 sec         D0       0.000000 sec         D0       0.000000 sec         D0       0.000000 sec         D1       102.4 msc         SG1       400.136639 MHz         SWH       484.37 Hz         TU       1024         NO       0.000100 mec         F1       Acquisition parameters         NH0       255         SF01       400.1366 HHz         SNO       2         TD       255         SF01       400.134004 Hz         SNO       12.132 ppm         F2       Proccessing parameters	F.S	Acquisition P	arameters		
PHE PROG         noesytp           SHE VENT         CUCI3           AD         0.1054420 sec           FIDHES         4.740595 Hz           DM         103.0 UNEC           DB         256           PHCFUS         1H           BC         256           PHCFUS         1H           BC         2.000000 sec           PT         12.0 UNEC           DO         0.000000 sec           DB         0.000000 sec           DF         164.8 UNEC           SWH         4854.37 Hz           TU         1024           NO         0.000103 nec           F1         Acquisition parameters           NHO         2           DD         255           SF01         400.134064 HHz           FUDES         18.962379 Hz           SN         12.132 ppm <t< td=""><td></td><td>950818</td><td></td></t<>		950818			
SUM VENT         CDC13           AQ         0.1054920 sec           FIDHES         4.740595 Hz           DW         103.0 UNMC           DB         256           PEGEUS         3H           BULLEUS         3H           BULLEUS         3H           BULL         3 dB           DU         0.000000 sec           P1         12.0 UNDEC           D0         0.000000 sec           D1         2.000000 sec           D0         0.0000000 sec           D1         136539 MHz           SKH         4054.37 Hz           TU         1024           NS         16           DS         4           INO         0.0001010 mm           FLUMES         18 96237 Hz           SM         12.132 ppm           FLUMES         18 96237 Hz           SM         12.132 ppm           F2 - Processing parameters           SI         1024           SN         12.132 ppm           F2 - Processing parameters           SI         1024           SN         12.132 ppm           SGB         0 <tr< td=""><td></td><td></td><td></td></tr<>					
A9     0.1054920 sec       FIDUES     4.740595 H7       DW     103.0 usec       DB     256       PECEFUS     1H       RE     2.000000 sec       P1     1.08       D0     0.000000 sec       DF     164.8 usec       SF01     400.1306359 MHz       SWH     4854.37 Hz       DU     1024       NG     16       US     4       NO     0.000100 sec       DF     164.8 usec       SF01     400.1306359 MHz       SWH     4854.37 Hz       TU     1024       NG     2       DD     256       SF01     400.1366 MHz       FLUES     18 962379 Hz       SW     12.132 ppm       F2 - Processing parameters       SI     1024       SV     12.132 ppm       F2 - Processing parameters       SI     1024       SI     1024       SI     1024       SI     1024       MCM     0SINE       SSB     0       IN     0.00 Hz       GB     0       IN     0.00 Hz       GB     0       CZ     20 00 cm <td></td> <td></td> <td></td>					
FIDHES       4.740595 Hz         DM       103.0 unre         HB       256         HB       256         HB       103.0 unre         HB       256         HB       1.03.0 unre         HB       1.00.00000 sec         P1       12.0 unre         U0       0.000000 sec         U0       0.000000 sec         U0       1.00.00000 sec         U0       0.000000 sec         U0       1.00.00000 sec         U1       1024         SWH       4.051.3659 MHz         SWH       4.01.36639 MHz         SWH       4.00.1366 MHz         V10       255         SF01       400.1366 MHz         FU0       255         SF01       400.1366 MHz         FU0HES       18.962379 Hz         SN       12.132 ppm         F2 - Processing parameters         SI       1024         MUN       0.00 Hz         GD       0         C2					
DW         103.0 usec           DG         256           PPG(FUS)         1H           PL(1)         3 dB           D1         2.000000 sec           P1         12.0 usec           D0         0.000000 sec           P1         12.0 usec           D0         0.000000 sec           DF         164.6 usec           SF01         400.1366359 MHz           SWH         4854.37 Hz           TU         1024           NG         3 16           US         4           TNO         0.0001010 mec           F1         Acquisition parameters           NHO         2           TU         255           F01         400.1364 MHz           NHO         2           TU         275           F01         20.132 Opm           F2         Processing parameters           SI         1024           SS         0           TB         0.00 Hz           GB         0           NOW         0SINE           SSB         0           LB         0.00 Hz           GB					
HG     256       PPRCFUS     3 H       HG     3 dB       HG     1 dB       HG     1 dB       HG     1 2.000000 sec       P1     12.0 usec       D0     0 000030 sec       D8     0.0000000 sec       D8     0.000000 sec       D9     0.000100 sec       SWH     4054.37 Hz       TU     1024       N5     4       D0     0.000100 sec       D5     4       D0     0.000100 sec       SWH     4054.37 Hz       D10     255       SF01     400.1366 MHz       FUBES     18 96237 Hz       SN     12.132 ppm       F2     Processing parameters       SI     1024       SN     12.132 ppm       F2     Processing parameters       SI     1024       SN     12.132 ppm       GB     0       C2     200       F1     Processing parameters       SI     1024       MDN <td< td=""><td></td><td></td><td></td></td<>					
PHOLEUS     1H       HL 1     3 dB       D1     2.000000 sec       P1     12.0 usec       D0     0.000000 sec       D0     0.000000 sec       DF     164.8 usec       SF01     400.136359 HHz       SNH     4854.37 Hz       TU     1024       NG     16       US     4       IN0     0.0001030 sec       SF01     400.1366 HHz       FU     256       SF01     400.1366 HHz       FUBES     18 962379 Hz       SN     12.132 ppm       F2 - Processing parameters       SI     1024       MUM     0.00 Hz       GB     0       C1     Processing parameters       SI     1024       MUM     0.00 Hz       GB     0       C2     1000 Hz       GB     0       C3     0.00 Hz       GB     0       C4     0.00 Hz       GB     0       C4     0.00					
HL1       3 dB         01       2.000000 sec         P1       12.0 usec         00       0.000000 sec         00       0.000000 sec         00       0.000000 sec         0F       164.8 usec         SF01       400.1306359 MHz         SMH       4854.37 Hz         TU       1024         NG       3 dB         US       4         INO       0.000100 sec         US       4         INO       0.000100 sec         SF01       400.1366 MHz         SF01       400.1366 MHz         SF01       400.1366 MHz         SF01       400.1366 MHz         FUHES       18.962379 Hz         SN       12.132 ppm         F2 - Processing parameters         SI       1024         SSB       0         IR       0.000 Hz         GB       0         VDM       QSINE         SSB       0         LR       0.000 Hz         GB       0         QU       NMA plot parameters         SI       1024         MOM       QSINE					
01       2.000000 sec         P1       12.0 usec         00       0.000030 sec         08       0.000000 sec         09       164.8 usec         SF01       400.1366359 HHz         SWH       4054.37 Hz         10       1024         NG       16         US       4         100       0.0001010 mec         F1 = Acquisition parameters         NH0       256         SF01       400.1366 HHz         SF01       40.0.1366 HHz         SF01       40.0.1366 HHz         SF01       40.0.1366 HHz         SF01       40.1364 Hz         SF01       40.1384 NHz         SF01       40.1384 NHz         SF01       40.1384 NHz         SF01       40.1384 NHz         SN       12.132 ppm         F2 - Processing parameters         SI       1024         SSB       0         IB       0.000 Hz         GB       0         NDW       QSINE         SSB       0         LB       0.00 Hz         GB       0         QU       NMB plot par					
P1       12.0 unec         D0       0.000030 sec         D8       0.000000 sec         DF       154.8 unec         SF01       400.1365359 MHz         SWH       4854.37 Hz         TU       1024         NG       16         DS       4         IN0       0.0001010 mm         F1 = Acquisition parameters         NH0       0.0001010 mm         F1 = Acquisition parameters         NH0       255         SF01       400.1366 MHz         FUHES       18.7627 Hz         SN       12.132 ppm         F2 = Processing parameters         SI       1024         SF8       0         IN       0.000 Hz         GB       0         PC       2.00         f1       Processing parameters         SI       1024         SV       2.00         f1       Processing parameters         SI       1024         SV       2.00         f1       Processing parameters         SI       1024         MON       GSINE         SVB       0					
D0         0.000000 sec           DF         164.8 usec           SF01         400.1306359 MHz           SWH         4854.37 Hz           TU         1024           MG         160.0000 sec           US         40.1306359 MHz           SWH         4854.37 Hz           TU         1024           MG         16           US         4           INO         0.000100 nec           F1 = Acquisition parameters           NHO         2           TD         255           SF01         400.1366 MHz           FUBIES         18.962379 Hz           SM         12.132 ppm           F2 = Processing parameters           SI         1024           SM         12.132 ppm           F2 = Processing parameters           SI         1024           SI         400.1343904 MHz           MUM         0SINE           SGB         0           IN         0.00 Hz           GB         0           IN         0.00 Hz           GB         0           SYS         0           LB         0.					
DF         164.8 usec           SF01         400.1366359 MHz           SNH         4094.37 Hz           TU         1024           NG         16           DS         4           TNO         0.0001030 mmc           F1         Acquisition parameters           NHO         2           TD         256           SF01         400.1366 MHz           SF01         400.1366 MHz           SF01         400.1366 MHz           SF01         400.1366 MHz           FTURES         18 962379 Hz           SN         J2.132 ppm           F2         Processing parameters           SI         1024           SF8         0           IB         0.000 Hz           GB         0           VDW         0SINE           SSB         0           IF         J024           MO2         IFPI           SF         400.13A3904 MHz           MDW         QSINE           SSB         0           LB         0.00 Hz           GB         0           QU         NMR plot parameters <t< td=""><td>00</td><td></td><td></td></t<>	00				
SF01     400.1365359 HHz       SH1     4854.37 Hz       TU     1024       NG     16       US     4       IN0     0.0001030 mm       F1 = Acquisition parameters       NH0     2       TD     255       SF01     400.1366 MHz       FUHES     18.46237 Hz       SN     2       TD     255       SF01     400.1366 MHz       FUHES     18.46237 Hz       SN     12.132 dpm       F2 = Processing parameters       SI     1024       SI     0       TI     0.00 Hz       GB     0       PC     2.00       f1     Processing parameters       SI     1024       MUM     0SINE       SSB     0       LR     0.00 Hz       GB     0       CX2     20.00 cm       CX2     20.00 cm       CX2     20.00 cm       CX1     20.00 cm       CX2     20.00 cm       CX1	1)0				
SNH         4894.37 Hz           TU         1024           NG         16           US         4           INO         0.0001030 mmc           F1         Acquisition parameters           NH0         20001030 mmc           F1         Acquisition parameters           NH0         256           SF01         400.1366 HHz           F10         256           SF01         400.1366 HHz           F10         257           SF01         400.1366 HHz           F10         257           SF1         1024           MUM         QSINE           SF2         400.13A004 MHz           MUM         QSINE           SF8         0           C1         0.00 Hz           G0         0           C2         1024           MUM         QSINE           SF4         00.1340904 MHz           MUM         QSINE           SF2         400.1340904 MHz           MUM         QSINE           SF2         000 cm           CX2         20.00 cm           CX2         20.00 cm	DF	164.8	0900		
TU       1024         NG       16         US       4         TN0       0.0001010 mmc         F1 = Acquisition parameters         NH0       2         TD       255         SF01       400.1366 MHz         FUSIES       18.962379 Hz         SM       12.132 ppm         F2 = Processing parameters         SI       1024         SP       400.134094 MHz         MUM       0SINE         SGB       0         LN       0.000 Hz         GB       0         VC2       TPPI         SF8       0         LR       0.000 Hz         GB       0         VDM       QSINE         SF8       0         LR       0.000 Hz         GB       0         VDM       QSINE         SF8       0         LR       0.000 Hz         GB       0         VDM       QCO cm         ZD NMH plot parameters         CX2       20.00 cm         CX2       20.00 cm         CX1       20.00 cm         CX2 <td>SF01</td> <td>400.1366359</td> <td>MHz</td>	SF01	400.1366359	MHz		
N3     16       US     4       UN0     0.0001010 mmc       F1 = Acquisition parameters       NH0     2       TD     255       SF01     400.1366 HHz       FTURES     18.962379 Hz       SN     12.132 ppm       F2 = Processing parameters       S1     1024       S2     400.1343904 HHz       WDW     0SINE       S98     0       IR     0.00 Hz       60     0       PC     2.00       F1     Processing parameters       S1     1024       WDW     0SINE       S98     0       IR     0.00 Hz       60     0       PC     2.00       S1     1024       MDW     QSINE       S58     0       LR     0.00 Hz       60     0       VDN     QSINE       S58     0       LR     0.00 Hz       60     0       20     NMR plot parameters       CX2     20.00 cm       CX2     20.00 cm       CX2     20.00 cm       CX1     20.00 cm       CX2     20.00 cm       CX2 <td>SWH</td> <td>4854.37</td> <td>Hz</td>	SWH	4854.37	Hz		
DS         4           IN0         0.0001030 mmc           F1 = Acquisition parameters           NI0         2           TD         255           SF01         400.1366 MHz           FUBES         18.0223 Hz           SN         12.132 ppm           F2 = Processing parameters           SI         1024           SF         400.1343904 MHz           MOM         0SINE           SGB         0           TA         0.00 Hz           GT         2000           F1         Processing parameters           SSB         0           TA         0.00 Hz           GT         3024           MUN         0SINE           SSB         0           LT         3024           MUN         QSINE           SSB         0           LR         0.00 Hz           GB         0           LR         0.00 Hz           GB         0           CX2         20.00 cm           CX1         20.00 cm           CX2         20.00 cm           CX1         20.00 cm <t< td=""><td>ΤU</td><td>1024</td><td></td></t<>	ΤU	1024			
INO         0.0001030 mmc           F1 = Acquisition parameters           NH0         255           SF01         400.1366 MHz           FLUTES         18.962379 Hz           SN         12.132 ppm           F2 = Processing parameters           SI         1024           SS         400.13404 MHz           MUM         051NE           SGB         0           CD         2.00           F1         Processing parameters           SI         102.4           MUM         051NE           SGB         0           CC         2.00           F1         Processing parameters           SI         1024           MUM         051NE           SGB         0           C2         1004           MUM         051NE           SGB         0           LB         0.00 Hz           GB         0           LB         0.00 Hz           GB         0           LB         0.00 Hz           GB         0           LB         0.00 Cm           CX2         20.00 cm <td></td> <td>16</td> <td></td>		16			
F1 = Acquisition parameters         NH0       2         10       255         SF01       400.1366         FUBES       18 962379         SN       12.132         SN       12.134         SN       0.1343004         SN       0         F1       Processing parameters         SN       00         F2       00         SN       0.00         F3       400.13443004         MDN       QSINE         SSB       0         LB       0.000         SN       0         LB       0.000         LB <td></td> <td></td> <td></td>					
NHO         2           TU         255           SF01         400.1366           SF01         400.1366           TU         235           SF01         400.1366           SN         12.132           SN         12.132           SN         1024           SI         1024           SI         1024           SI         1024           SI         0.01343904           MUN         0STRE           SGB         0           TH         0.00 Hz           GO         0           FC         2.00           f1         Processing parameters           ST         3024           MC2         TPPT           SF         400.1343904           MUN         QSTNE           SVB         0           LR         0.00 Hz           GB         0           CX2         20.00 cm           CX1         20.00 cm           CX2         20.00 cm           CX2         20.00 cm           CX1         20.00 cm           CX2         20.00 cm	1NO	0 0001030	980		
NHO         2           TU         255           SF01         400.1366           SF01         400.1366           TU         235           SF01         400.1366           SN         12.132           SN         12.132           SN         1024           SI         1024           SI         1024           SI         1024           SI         0.01343904           MUN         0STRE           SGB         0           TH         0.00 Hz           GO         0           FC         2.00           f1         Processing parameters           ST         3024           MC2         TPPT           SF         400.1343904           MUN         QSTNE           SVB         0           LR         0.00 Hz           GB         0           CX2         20.00 cm           CX1         20.00 cm           CX2         20.00 cm           CX2         20.00 cm           CX1         20.00 cm           CX2         20.00 cm	£1 -	Acquisition o	arameters		
TD         256           SF01         400.1366         Hi/z           FLUTIES         18.962379         Hz           SN         12.132         ppm           F2 - Processing parameters         SI         1024           SI         1024         SI           SI         400.13A0904         Hiz           MUM         0SINE         SGB           SGB         0         H           GD         0         O           F2         Processing parameters         SI           SGB         0         Hz           GD         0         O           F2         Processing parameters         SI           SGB         0         Hz           GD         0         O           F2         400.1343904         Hiz           MIN         QSINE         SSB           SSB         0         LB           SSB         0         LB           OLD         0.00 Hz           GB         0           CX2         20.00 cm           CX2         20.00 cm           CX1         20.00 cm           CX2         20.0					
FIDHES     18.962379.Hz       SN     J2.132.ppm       F2 - Processing parameters       SI     1024       SF     400.1343904.HHz       WDW     0SINE       SGB     0       IR     0.00 Hz       GB     0       PC     2.00       F1     Processing parameters       SI     J024       MOX     0.00 Hz       GB     0       PC     2.00       F1     Processing parameters       SI     J024       MC2     IPPI       SF     400.1343904.HHz       WDM     QSINE       SSB     0       LR     0.00 Hz       GB     0       CX2     20.00 cm       CX2     20.00 cm       CX1     20.00 cm       CX2     20.00 cm       F2H0     4.567 ppm       F2L0     1033.00 Hz       F2HI     0.664 ppm       F2HI     27.3.42 Hz       F1H2     0.4.557 ppm	ŦD				
SN         12.132 ppm           F2 - Processing parameters           SI         1024           SI         1024           SI         1024           SI         1024           SI         1024           SI         00.1343904 MHz           MUM         0SINE           SGB         0           CD         0           F1         Processing parameters           SI         1024           MC2         IPPI           SF         400.1343904 MHz           MUN         0SINE           SSB         0           LB         0.00 Hz           GB         0           SSB         0           LB         0.00 Hz           GB         0           CX2         20.00 cm           CX1         20.00 cm           CX1         20.00 cm           CX1         20.00 cm           CX2         20.00 cm           CX1         20.00 cm           CX2         20.00 cm           F2HQ         4.563 ppm           F2HQ         664 ppm           F2HQ         4.567 ppm </td <td>SE01</td> <td>400.1366</td> <td>MHZ</td>	SE01	400.1366	MHZ		
F2 - Processing parameters         S1       1024         S4       400.1343004 MHz         MUM       051NE         S38       0         11       0.00 Hz         GB       0         PC       2.00         F1       Processing parameters         S1       1024         MC2       1591         S5       400.1343004 MHz         MC2       1591         S5       400.1343004 MHz         MUM       QSINE         S58       0         LB       0.000 Hz         G8       0         CX2       20.00 cm         CX2       20.00 cm         CX1       20.00 cm         F2PL0       4.581 ppm         F2L0       1833.08 Hz         F2H1       273.42 Hz         F2H1       273.42 Hz	FIDHES	18 962379	Hz		
SI         1024           SI         400.134.004 MHz           WEW         QSINE           SGB         0           LD         0.00 Hz           GD         0           PC         2.00           F1         Processing parameters           SI         1024           MC2         IEPFI           SF         400.134.004 MHz           MDM         QSINE           SF8         0           LB         0.00 Hz           GB         0           2D NMR plot parameters           CX2         20.00 cm           CX1         20.00 cm           F2PI 0         4.564 ppm	SM	15, 135	ព្រុក		
SI         1024           SI         400.134.004 MHz           WEW         QSINE           SGB         0           LD         0.00 Hz           GD         0           PC         2.00           F1         Processing parameters           SI         1024           MC2         IEPFI           SF         400.134.004 MHz           MDM         QSINE           SF8         0           LB         0.00 Hz           GB         0           2D NMR plot parameters           CX2         20.00 cm           CX1         20.00 cm           F2PI 0         4.564 ppm	50	Devenesion			
S/         400.134/3904 MHz           MUM         0SINE           SGB         0           SGB         0           LI         0.00 Hz           GB         0           VC         2.00           F1         Processing parameters           SI         1024           MC2         TFPT           SF         400.134/3904 MHz           MUM         0SINE           SSB         0           LB         0.00 Hz           68         0           CX2         20.00 cm           CX1         20.00 cm           CX2         20.00 cm           CX1         20.00 cm           F2H0         4.567 ppm           F2L0         1633.00 Hz           F2HI         2.73.42 Hz           F2HI         2.73.42 Hz			arameters		
MDM         OSINE           SSB         0           LP         0.00 Hz           GB         0           PC         2.00           F1         Processing parameters           SI         1024           MC2         TEPT           SF         400.1343904 MHz           MUM         QSINE           SSB         0           LB         0.00 Hz           GB         0           CX2         20.00 cm           F2PL0         4.583 ppm           F2U         1633.08 Hz           F2HI         27.3.42 Hz           F2HI         27.3.42 Hz           F1PL0         4.557 ppm			MHZ		
SGB         0           L II         0.00 Hz           GD         0           PC         2.00           f1         Processing parameters           ST         1024           MC2         LPPT           SF         400.1343904           MDM         QSINE           SF8         0           LB         0.00 Hz           GB         0           20 NMR plot parameters           CX2         20.00 cm           CX1         20.00 cm           F2PL0         4.564 ppm           F2L0         1033.00 Hz           F2HT         0.664 ppm           F2HT         27.3.42 Hz           F1H2.0         4.557 ppm					
GB         0           Processing parameters           F1         Processing parameters           S1         3024           MC2         TPP1           SF         400,134304 MHz           MIN         OSINE           S98         0           LB         0.00 Hz           GB         0           CX2         20.00 cm           CX1         20.00 cm           CX1         20.00 cm           F2H0         4.563 ppm           F2L0         1633.00 Hz           F2HII         0.664 ppm           F2HII         27.3.42 Hz           F1HZ0         4.555 ppm					
PC         2:00           E1         Processing parameters           S1         3024           MC2         TPP1           SF         400.1343904 MHz           MUN         QSINE           SSB         0           LB         0.00 Hz           GB         0           20 NMR plot parameters           CX2         20.00 cm           F2PL0         4.583 ppm           F2L0         1833.08 Hz           F2HI         27.3.42 Hz           F2HI         27.3.42 Hz           F1H20         4.557 ppm	LB	0.00	Hiz		
E1         Processing parameters           SI         3024           MC2         EPPT           SF         400.1343904           MUM         QSINE           SF8         0           LB         0.00           B1         0.00           20         NMR plot parameters           CX2         20.00 cm           CX1         20.00 cm           F2PL0         4.561 ppm           F2L0         1033.00 Hz           F2H1         2.7.3.42 Hz           F2H1         2.7.3.42 Hz	60	0			
ST         1024           NC2         TEPT           SF         400.134304 MHz           SF         400.134304 MHz           SFB         0           SFB         0           LB         0.00 Hz           GB         0           CX2         20 00 cm           CX2         20 00 cm           F2H0         4 564 ppm           F2L0         1633.00 Hz           F2HT         0.664 ppm           F2HT         27.3.42 Hz           F2HT         27.4.42 Hz	PC	2.00			
ST         1024           NC2         TEPT           SF         400.134304 MHz           SF         400.134304 MHz           SFB         0           SFB         0           LB         0.00 Hz           GB         0           CX2         20 00 cm           CX2         20 00 cm           F2H0         4 564 ppm           F2L0         1633.00 Hz           F2HT         0.664 ppm           F2HT         27.3.42 Hz           F2HT         27.4.42 Hz					
NC2         IPPI           SF         400.134904         HHz           MUN         QSINE         SSB           SSB         0         LB           GB         0         HZ           GB         0         HZ           CX2         20 00 cm         CX1           CX1         20.00 cm         F2PI 0           F2PI 0         4 SB1 ppm         F2L0           F2HI         0.684 ppm         F2HI           F2HI         27.3.42 Hz         F1HZ           F2HI         27.3.42 Hz         F2HZ			ar anns Cerra		
SF         400.1343904         HHz           MIN         QSINE           SSB         0           LB         0.00 Hz           GB         0           20 NMH plot parameters           CX2         20 00 cm           F2PL0         4 561 ppm           F2L0         1633.00 Hz           F2HI         0.684 ppm           F2HI         27.3.42 Hz           F2HI         27.3.42 Hz					
MUN         QSINE           SSB         0           LR         0.00 Hz           GB         0           2D NMR plot parameters           CX2         20 00 cm           CX1         20.00 cm           F2PL0         4 563 ppm           F2L0         1033.00 Hz           F2H10         0.664 ppm           F2H11         0.664 ppm           F2H11         27.3.42 Hz           F1H20         4.557 ppm			MHz		
SSB         0           LB         0.00 Hz           GB         0           20 NMB plot parameters           CX2         20 00 cm           CX1         20.00 cm           F2PL0         4 563 ppm           F2L0         1633.00 Hz           F2HI         0.663 ppm           F2HI         2/3.42 Hz           F2HI         2/3.42 Hz					
LB 0.00 Hz GB 0 CX2 20 00 cm CX1 20.00 cm F2PL0 4 563 ppm F2L0 5633 pm F2HI 0.663 ppm F2HI 0.663 ppm F2HI 2.13, 42 Hz F2HI 0.4557 ppm					
GB         0           CX2         20 NMR plot parameters           CX2         20 00 cm           CX1         20.00 cm           CX1         20.00 cm           F2PL0         4 587 ppm           F2L0         1633.04 Hz           F2H11         0.663 ppm           F2H11         27.3.42 Hz           F2H11         27.3.42 Hz		0.00	H7		
CX2         20:00 cm           CX1         20.00 cm           F2PL0         4:563 ppm           F2PL0         10:33.08 Hz           F2PHI         0.663 ppm           F2PHI         27.3.42 Hz           F2PHI         27.3.42 Hz					
CX2         20:00 cm           CX1         20.00 cm           F2PL0         4:563 ppm           F2PL0         10:33.08 Hz           F2PHI         0.663 ppm           F2PHI         27.3.42 Hz           F2PHI         27.3.42 Hz					
CX1         20.00 cm           F2PL0         4 563 ppm           F2L0         1633.00 Hz           F2PHI         0.663 ppm           F2HI         27.3.42 Hz           F2HI         24.55 ppm					
F2PL0         4         5BJ         ppm           F2L0         1033.00 Hz         F2HI         0.663.00 Hz           F2HI         0.663.00 Hz         F2HI         F2HI         F2HI           F2HI         2J3.42 Hz         F1HO         F1HO         F157.00 mm					
F2L0 1033.08 Hz F2PHI 0.663 ppm F2HI 273.42 Hz F1PL0 4.557 ppm					
F2141 0.683 ppm F2141 273.42 Hz F119.0 4.557 ppm					
F2H1 273.42 Hz F1P10 4.557 ppm					
f 101.0 4,557 ppm					
C 11,0 1823.60 Hz		c/.1.42 A 557			
		1823.50	112		

Jacob KCP -07 in CDC13 Losy 45

COSY Spectrum of 76

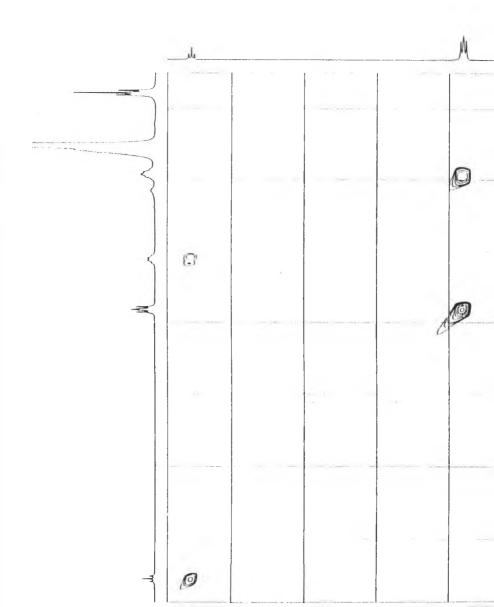


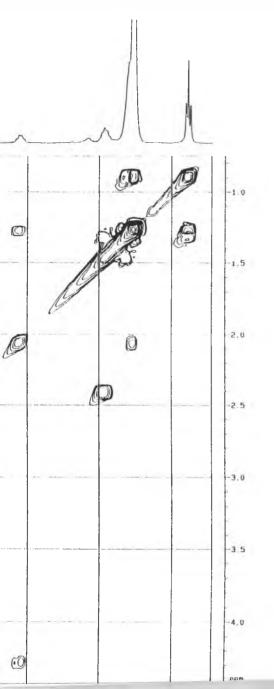


	Current Data Parameters		
NAME	A110296		
EXPNO	2		
PHOCNO	1		
	2 Acquisition Parameters		
Date	950921		
lime PULPROG	10.30		
SOL VEN3			
AQ	0 1085640 sec		
FIDRES	4 606427 Hz		
DW	106.0 usec		
AG	1024		
NUCLEUS	311		
<u>FR 1</u>	1 dB		
01	1 0000000 sec		
P1	12 0 usec		
DO	0.0000 sec		
PO	6.0 usec		
DE	169.6 Usec		
SE 0.1 SWLL	400 1363600 MHz		
10	4716 98 Hz 1024		
NS	1024		
us	4		
INO	0 0002120 sec		
	1 Acquisition parameters		
NDO	1		
10	256		
SF01 FIDHES	400.1354 MHz 18.425707 Hz		
SM	11.788 ppm		
374	11.788 phi		
F	2 Processing parameters		
51	1024		
SF	400.1343898 MHz		
WDW	USINE		
SSB	0		
L B 60	0.00 Hz		
PC	2 00		
1.0	2 00		
F1 - Processing parameters			
SI	1024		
MC2	QF		
SF	400 1343898 MHz		
WDW	QSINE		
SSB	0		
L 8 G 8	0.00 Hz		
60	0		
	2D HMR plot parameters		
CX5	20.00 cm		
CX1	20.00 cm		
F 2PL 0	4.452 ppm		
E SE O	1781.31 Hz		
ESb)(E	0.664 ppm		
E SHI	265.79 Hz		

KCP-10 in CDC13 cosy45

COSY Spectrum of 77





	Current Hata Par	rameters		
NAME	A110291	0		
EXPN0	4			
PHOCNO	1			
	- Acquisition F	parameters		
Date	950920			
Time PUL PRUG	14.42			
	COSY			
SOF VENT	CDE13 0.1167560			
E LORES	4 283169			
DW	4 203109			
BG	114.0			
NUCLEUS	111			
HL1		d9		
01	1.0000000			
P1	12 0			
00	0 0000030			
PO	Б.0	usec		
ĐE	182.4			
SF 0 1	400.1363085	MHz		
SWH	4385.96	Hz		
10	1024			
NS	4			
DS	4			
INO	0.0005590	960		
Fi - Acquisition parameters				
NDO	1			
10	256			
SF01	400.1363			
FIDHES	17.132675			
	10.501	0,000		
	- Processing p			
51	1024			
SE	400.1343901	MHZ		
	GS1NE			
SSB UB	0.00	11-		
68	0.00	112		
PC	2.00			
ru.	2.00			
	- Processing p	arameters		
51	1024			
MC5	0F	1014		
SF MDW	400.1343901 QSINE	mtiz		
MUM SSB	05176			
18	0.00	117		
GB	0.00			
20 NMA plot parameters				
CX5	20 NMH 010T 0ars 20.00			
CX1	20,00			
F2PI.0	4,441			
ESL0	1777 01			
ESBH1	0 727			
F 2H1	290.75			
FIPLO	4 441			
F 11 O	1777.01	Hz		
FIPHI	0.748	() () m		

<sup>13</sup>C NMR of isolated compounds.

