

# Reconsidering the species problem in downy mildews – where are we now?

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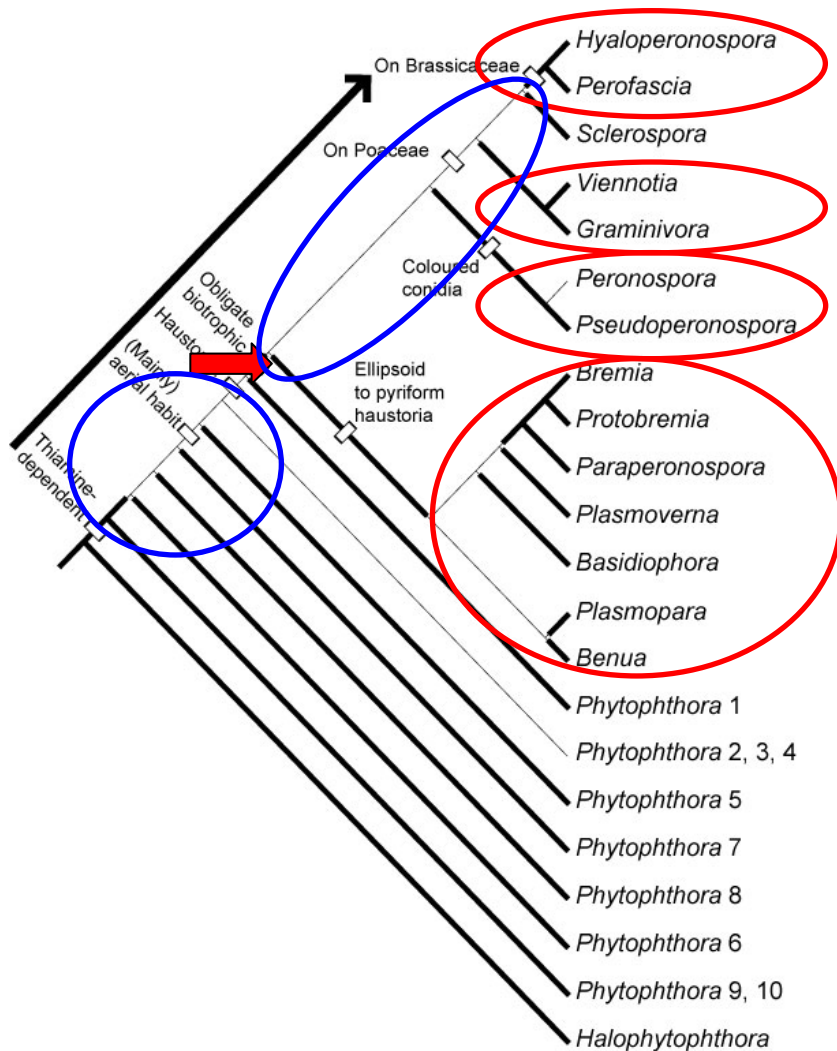
# The current talk will focus on...

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- historical competing species concepts of downy mildews, their implications and influence
- influence of recent (mainly molecular) evidence for the debate on a revised species concept in downy mildews
- the current status and shortcomings of knowledge on biodiversity of downy mildews
- the status of a molecular barcoding system

# Multigene analysis of downy mildews

LSU (D1-D3, D7-D8), *cox2*,  $\beta$ -tubulin, NADH (3921 bp)



Peronosporaceae

Phytophthora

- downy mildews (Peronosporaceae) are likely to be monophyletic
- downy mildews are rooted within a paraphyletic *Phytophthora*
- circumscription of important genera mostly resolved
- various subgroups of downy mildews highly supported
- relationships between these groups remains mostly unresolved

# What are important species problems in downy mildews?

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- except for economically important species, most described species are little known and investigated
- lack of sound contemporary investigations on biodiversity
- lack of a sound reference for species identification
- uncritical use of species names
- species identification solely based on host association
- uncertainties about the host ranges of species
- how to delimit and define the species – is a narrow or wide species concept more appropriate (splitting vs. lumping)?
- is the popular and commonly applied “one host family – one parasite” concept appropriate?

# What are the main reasons for the species problem?

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- comparatively few morphological features available for species delimitation
- few morphological features are commonly variable and overlapping
- cryptic speciation appears to be common (genetically distinct entities lack morphological distinction) – shall they be formally classified?
- obligate parasites – cannot be cultured and investigated on artificial media
- experimentally difficult- many biological experiments which can be carried out in *Phytophthora* cannot practically be applied to downy mildews (crossing experiments, recognition reactions, nutrition requirements,...)
- host range can only be examined by time-consuming inoculation experiments

# How many species do we have in downy mildews?

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- species circumscription was in the past rather based on personal opinion than on facts
- highly deviating species estimates on downy mildews, depending on the species concept (narrow versus wide):
  - *Peronospora*: from c. 60 to more than 350
  - *Plasmopara*: from c. 80 to 120
  - *Bremia*: from 1 to c. 15

# Brief history of species concepts in downy mildews

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- **morphological (morphometric) species concept:**
- species delimitation based on morphological features/differences
- problem: few morphological features available; often no clear-cut morphological differences, but a morphological continuum/overlap
- morphological features often influenced by environment
- only few „species“ morphologically distinguishable
- each of these morphological „species“ would have a wide host range
- but: experimental data indicated narrow host range!
- due to these problems, practically, a purely morphological species concept was never applied in downy mildews

# Brief history of species concepts in downy mildews

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- **Gäumann's (1918, 1923) „biological“ species concept:**
- high host specialisation is considered the most important biological feature of species
- species delimitation primarily based on host species/genus, with a combination of morphological features/differences
- result: narrow “one host – one species“ concept - leads to a high number of accepted species (splitting approach)
- problems:
  - host specificity often not experimentally proven
  - morphometric differences between species given by Gäumann often very small, based on few (often single) specimens
  - species cannot be identified if host is unknown, new or unidentified
  - misleads to species identification only by host species
- not widely accepted by plant pathologists who preferred a wide species concept
- more widely applied by investigators of biodiversity



# Brief history of species concepts in downy mildews

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- **Yerkes & Shaw's (1959) “one host family-one species“ concept:**
- accessions from the same host family are classified within a single species if not morphologically clearly distinct
- accessions from different host families are classified as distinct species, even if morphologically not clearly distinct
- result: wide “one host family – one species” concept – leads to few accepted species (lumping approach)
- convenient approach and therefore popular and widely accepted amongst plant pathologists and still commonly used
- problems:
  - misleads to species identification only by host family
  - untested assumption that downy mildews from the same host family are closely related – if not, non-related entities are classified under a single species
  - confusion about host ranges, inoculation sources, incomparable experiments etc.

# Modern species concepts

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- **Biological species concept (Ernst Mayr, 1982)**
- species are considered/defined as reproductive communities and separated by reproductive isolation
- practically not applicable in obligate parasitic downy mildews due to methodological difficulties (not culturable!)
  
- **Phylogenetic species concept**
- nowadays the dominant concept due to rapid progress in DNA sequencing techniques
- phylogenies (trees) are used for defining species
- species are defined as distinct, monophyletic entities
- reproductive isolation is mirrored by genetic distance

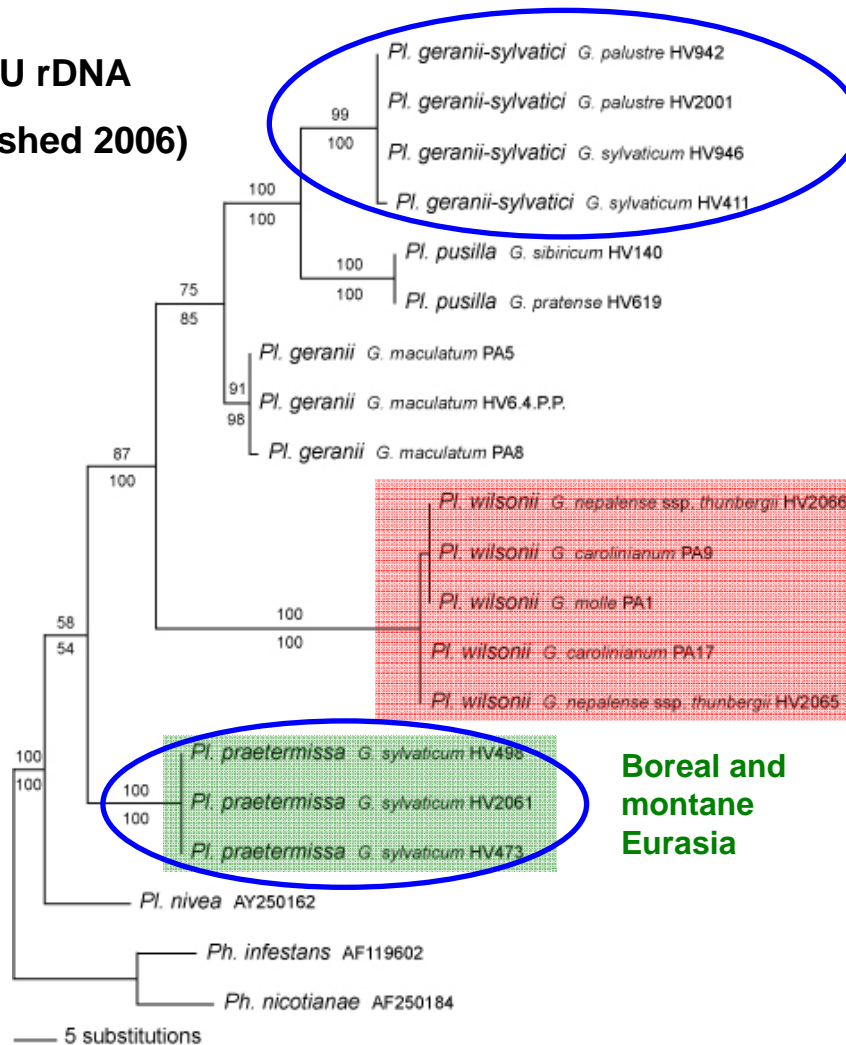
# Evidence from recent investigations

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- (1) detection of new, morphologically clearly distinct species by thorough re-investigations

# Plasmopara on Geranium

nu LSU rDNA  
(published 2006)



North  
America &  
Eastern  
Asia

Boreal and  
montane  
Eurasia

- 2 new species were revealed, which were clearly distinct
- new species are quite common, widespread and sympatric with already described species
- different species can infect the same host species even on the same host individual!
- remained undetected despite clear morphological differences - due to uncritical species determination based solely on host association!

# Evidence from recent investigations

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- (1) detection of new, clearly distinct species by thorough re-investigations
- (2) inappropriate species classification by uncritical adherence to the “one host family – one species” concept

# The identity of the downy mildew of sweet basil (*Ocimum* spp.)

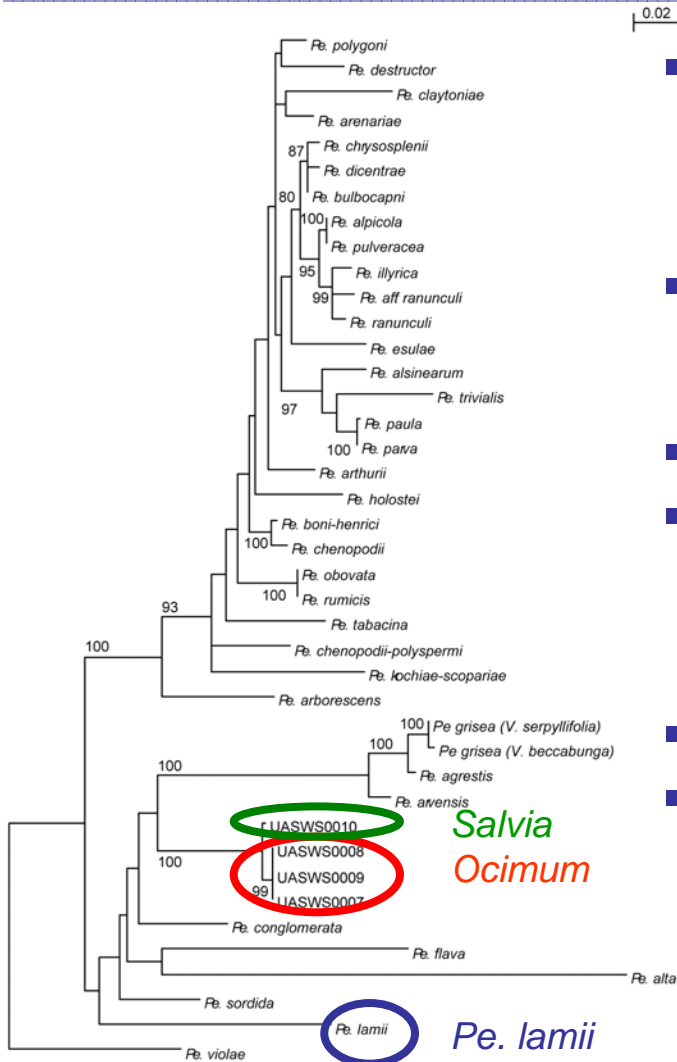


from Heller & Baroffio, [http://www.db-acw.admin.ch/pubs/wa\\_cma\\_03\\_pub\\_492\\_d.pdf](http://www.db-acw.admin.ch/pubs/wa_cma_03_pub_492_d.pdf)



- severe outbreaks of downy mildew of basil world-wide from about 2000 onwards.
- Identified as *Peronospora lamii* primarily on host family (Lamiaceae)
- *Peronospora lamii* supposed to be the sole species on Lamiaceae; type host: *Lamium*
- based on distribution records of *Peronospora lamii* on the various hosts, the pathogen was considered to be indigenous in most European, Asian and North American countries
- therefore, the sweet basil pathogen was not included in quarantine lists, promoting rapid spread via infected seed lots

# The identity of the downy mildew of sweet basil (*Ocimum* spp.)



- molecular phylogenetic analyses (ITS rDNA) showed the *Ocimum-Peronospora* to be markedly distinct from *Peronospora lamii*! (Belbahri et al., 2005)
- close but probably not conspecific with the *Peronospora* from *Salvia* - should represent a distinct species
- the species could not be given a name
- problem: altogether, more than 30 *Peronospora* species were described from 23 genera of Lamiaceae, for which no molecular data are available!
- pathogen origin unclear (?Africa)
- recent outbreak on Painted Nettle (*Solenostemon scuttelarioides*), followed by rapid spread

tree from Belbahri et al. (2005), Mycological Research 109: 841-848.

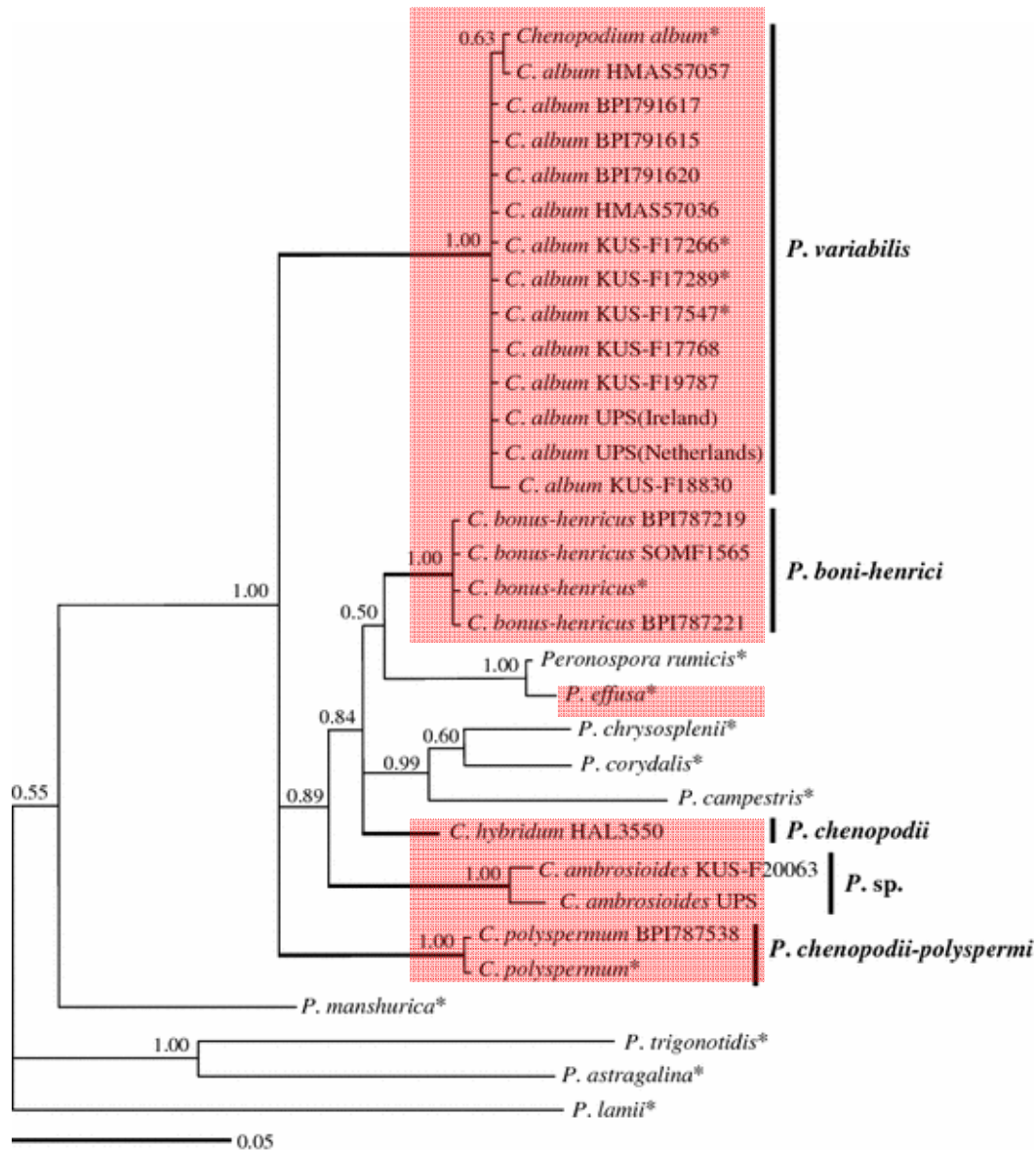
# Evidence from recent investigations

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- (1) detection of new, clearly distinct species by thorough re-investigations
- (2) inappropriate species classification by uncritical adherence to the “one host family – one species” concept
- (3) molecular evidence for a narrow species concept and the re-establishment of previously lumped species



# Peronospora on Chenopodiaceae

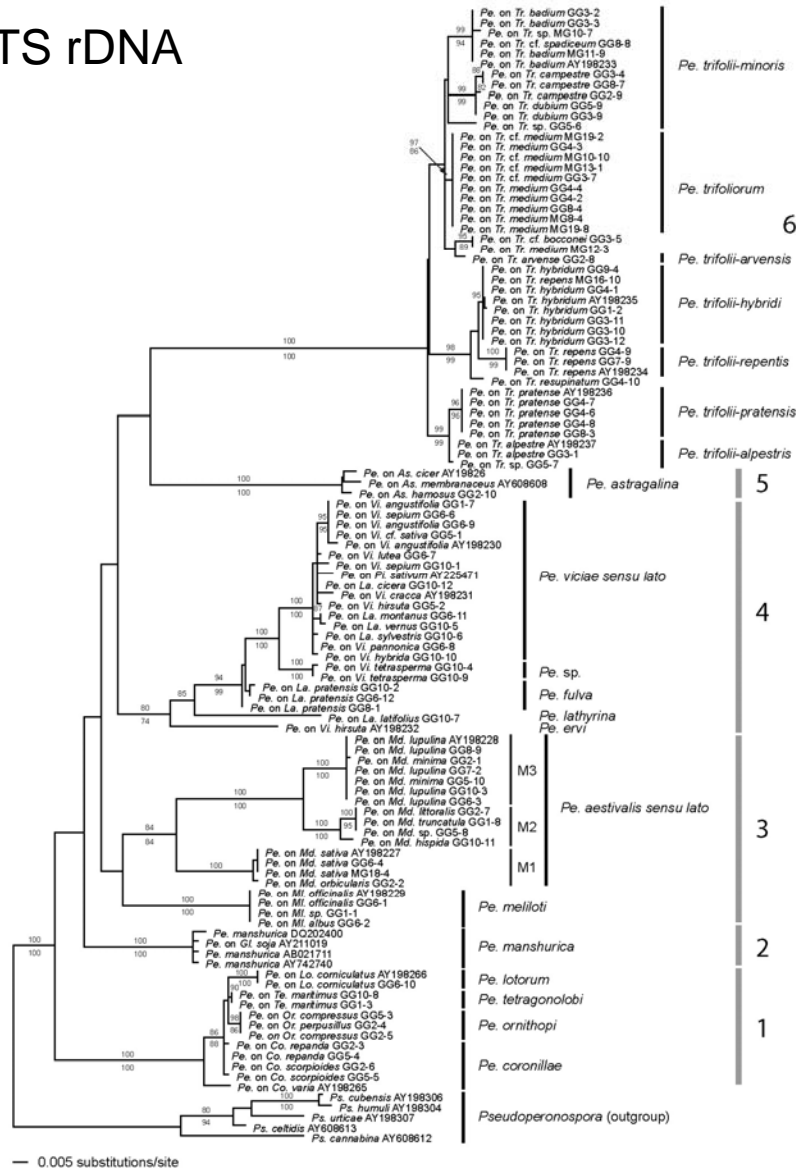


- *Peronospora* on Chenopodiaceae commonly treated as a single species (*Pe. farinosa*), following the concept of Yerkes & Shaw (1959)
- in phylogenetic analyses of DNA data, accessions from Chenopodiaceae are polyphyletic and not closely related
- high genetic distances between accessions from different hosts – evidence for high host specificity
- some subtle morphological differences present
- classification as a single species (*Pe. farinosa*) not tenable

tree from Choi et al. (2008), Mycopathologia 165: 155-164.

# Peronospora on Fabaceae

## ITS rDNA



- commonly two species accepted (*Pe. trifoliorum*, *Pe. viciae*) (de Bary 1864)
- accessions from different host genera/species are genetically distinct
- accessions from the same host are genetically homogeneous
- species on Fabaceae are not monophyletic
- high host specificity corroborated
- narrow species concept corroborated
- some nomenclatural problems require additional investigations

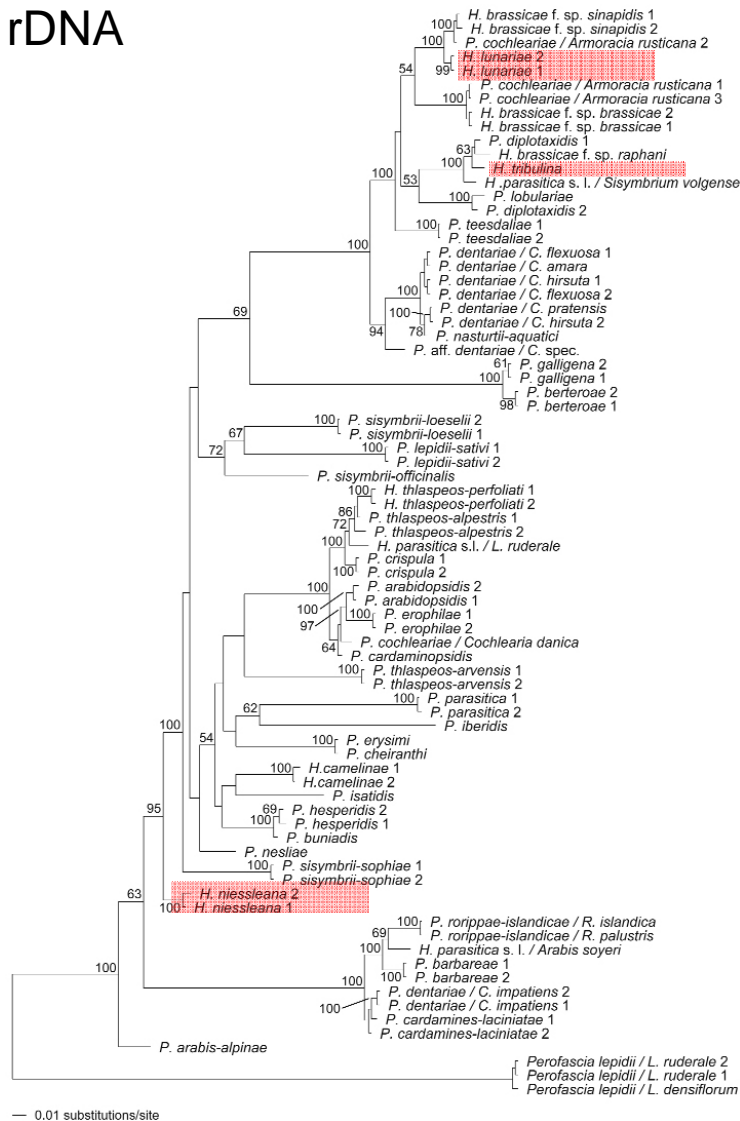
# *Hyaloperonospora* – a case study for downy mildew speciation

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- recently split from the genus *Peronospora* (Constantinescu & Fatehi 2002), recognising 6 morphologically distinct species
- numerous host species affected, mainly from *Brassicaceae*
- disagreement about the number of species (from 1 to more than 100!). Gäumann (1918, 1923) applied excessive splitting, whereas Yerkes & Shaw (1959) accepted only one species
- morphological delimitation often impossible
- often lumped into a single species (*H. parasitica*)
- species boundaries and host specificity often unclear
- ideal model group for investigating host-parasite cospeciation

# Hyaloperonospora

ITS rDNA

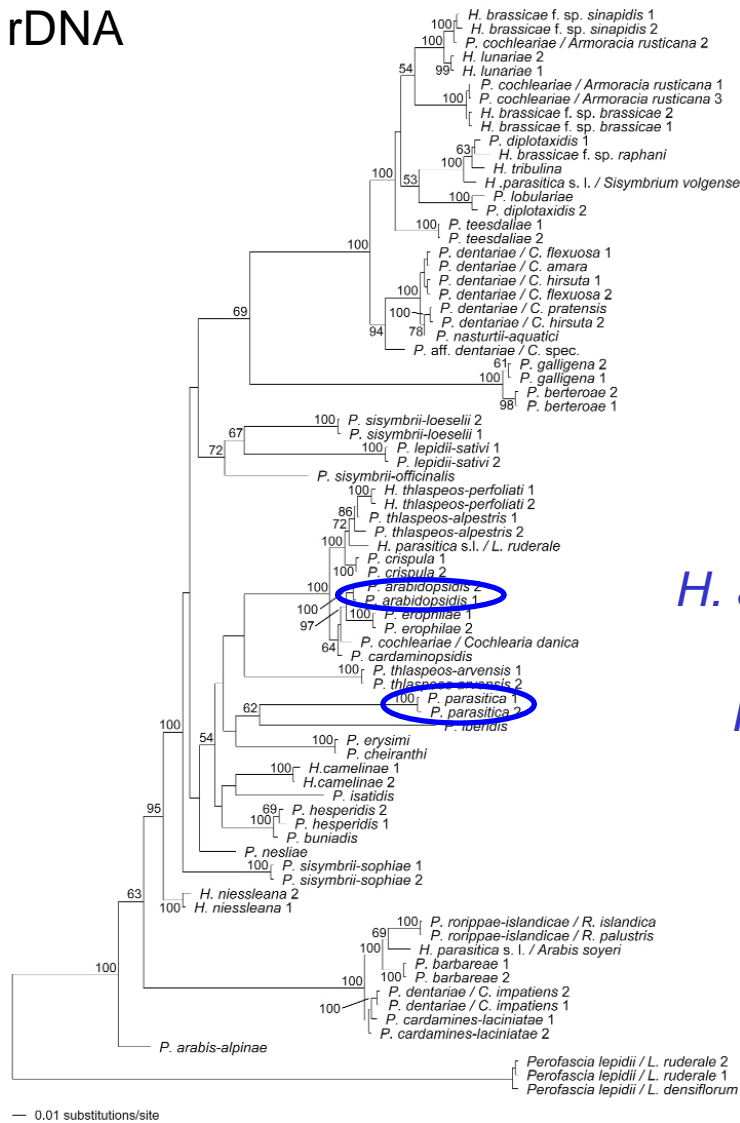


- Göker et al. (2004): morphologically clearly distinct taxa sensu Constantinescu & Fatehi (2002) are embedded within a paraphyletic “*H. parasitica*”
- accessions within a host species/genus genetically uniform
- genetic distances between host specific groups high and consistent
- evidence supports narrow species concept of Gäumann, but investigation included comparatively few accessions

from Göker & al. (2004), Phylogeny of *Hyaloperonospora* based on nuclear ribosomal internal transcribed spacer sequences

# Hyaloperonospora

ITS rDNA



Example for genetically distinct entities:

*H. parasitica*: on *Capsella bursa-pastoris* (type host)

*H. arabidopsidis*: on *Arabidopsis thaliana* (important species on a genetic model plant)

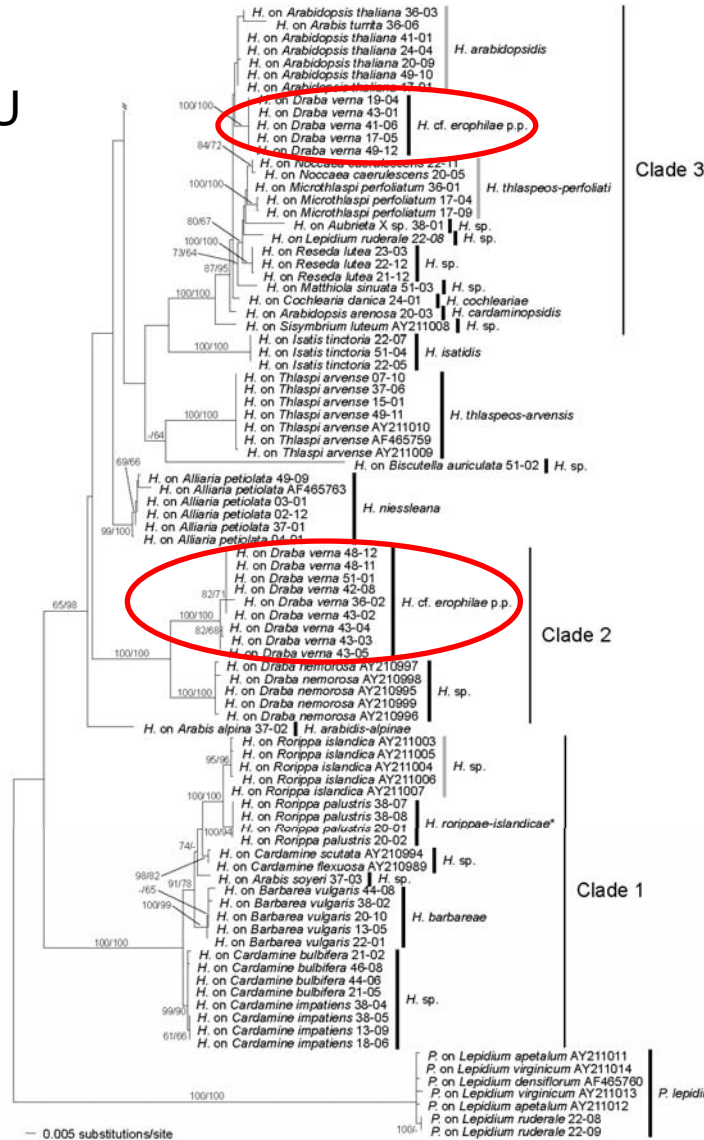
*H. arabidopsidis*

*H. parasitica sensu stricto*

from Göker & al. (2004), *Mycological Progress* 3: 83-94.

# Hyaloperonospora

ITS +  
nuLSU  
rDNA



- extensive investigation using more accessions and sequence data (Göker et al., submitted) support previous results
- narrow species delimitation corroborated – high internal support
- the same host can be parasitised by more than one species (e.g. *Draba verna*)
- no evidence for hybridisation
- evidence for several undescribed species

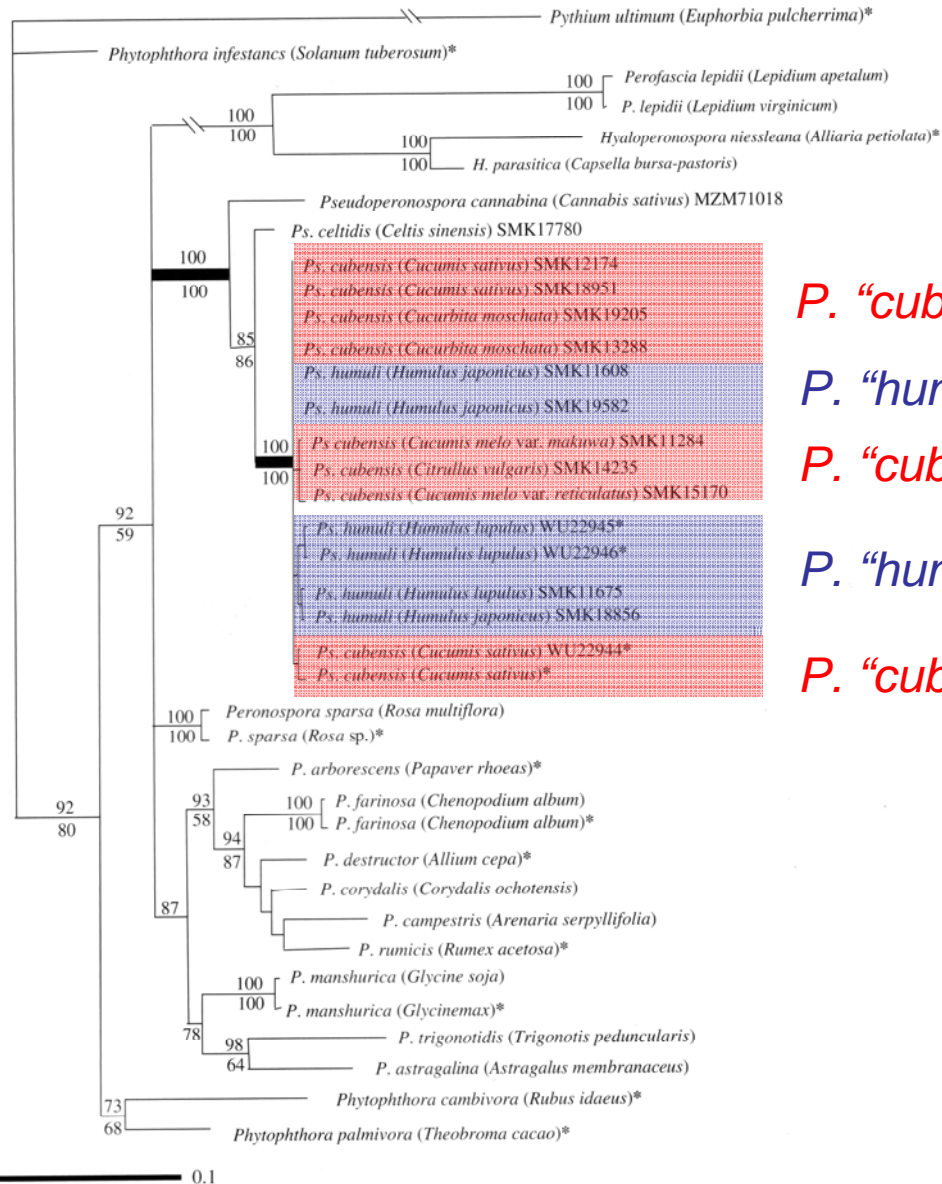
from Göker & al. (submitted), Species delimitation in downy mildews: the case of *Hyaloperonospora* in the light of nuclear ribosomal internal transcribed spacer and large subunit sequences

# Evidence from recent investigations

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- (1) detection of new, clearly distinct species by thorough re-investigations
- (2) inappropriate species classification by uncritical adherence to the “one host family – one species” concept
- (3) molecular evidence for a narrow species concept and the re-establishment of previously lumped species
- (4) molecular evidence for a wide species concept and the lumping of species from different host families

# Reevaluation of species: *Pseudoperonospora*



- in *Pseudoperonospora* species were delimited based on host families
- *Pseudoperonospora humuli* on hop (Cannabaceae)
- *Pseudoperonospora cubensis* on cucumber/melon/pumpkin (Cucurbitaceae)
- little genetic and morphological differences between accessions from these 2 non-related families
- molecular evidence for conspecificity

ITS tree from Choi et al. (2005), *Mycological Research* 109: 841-848.



# Species identification and molecular barcoding

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- identification by molecular tools (sequences) highly reliable
- problem: there is still no consensus about the sequence region of choice
- the ITS rDNA region, a commonly used barcoding region for fungi and also *Phytophthora*, works well in *Peronospora* and *Hyaloperonospora* (especially ITS2)
- however, ITS cannot be used universally for downy mildews due to length polymorphism and presence of numerous repeats in some lineages, in combination with amplification and sequencing problems (e.g. in *Plasmopara*, *Bremia*).
- mitochondrial DNA has better candidates (e.g. *cox*): high resolution, high number of copies – can be amplified even in historic collections
- however, sequence data on mitochondrial DNA still highly fragmentary and not yet optimised. For downy mildews, specific well-working primers need to be developed for routine use
- species boundaries need to be clarified before a barcoding system can be implemented to avoid taxonomic confusion, which necessitates thorough taxonomic revisions

# Conclusions

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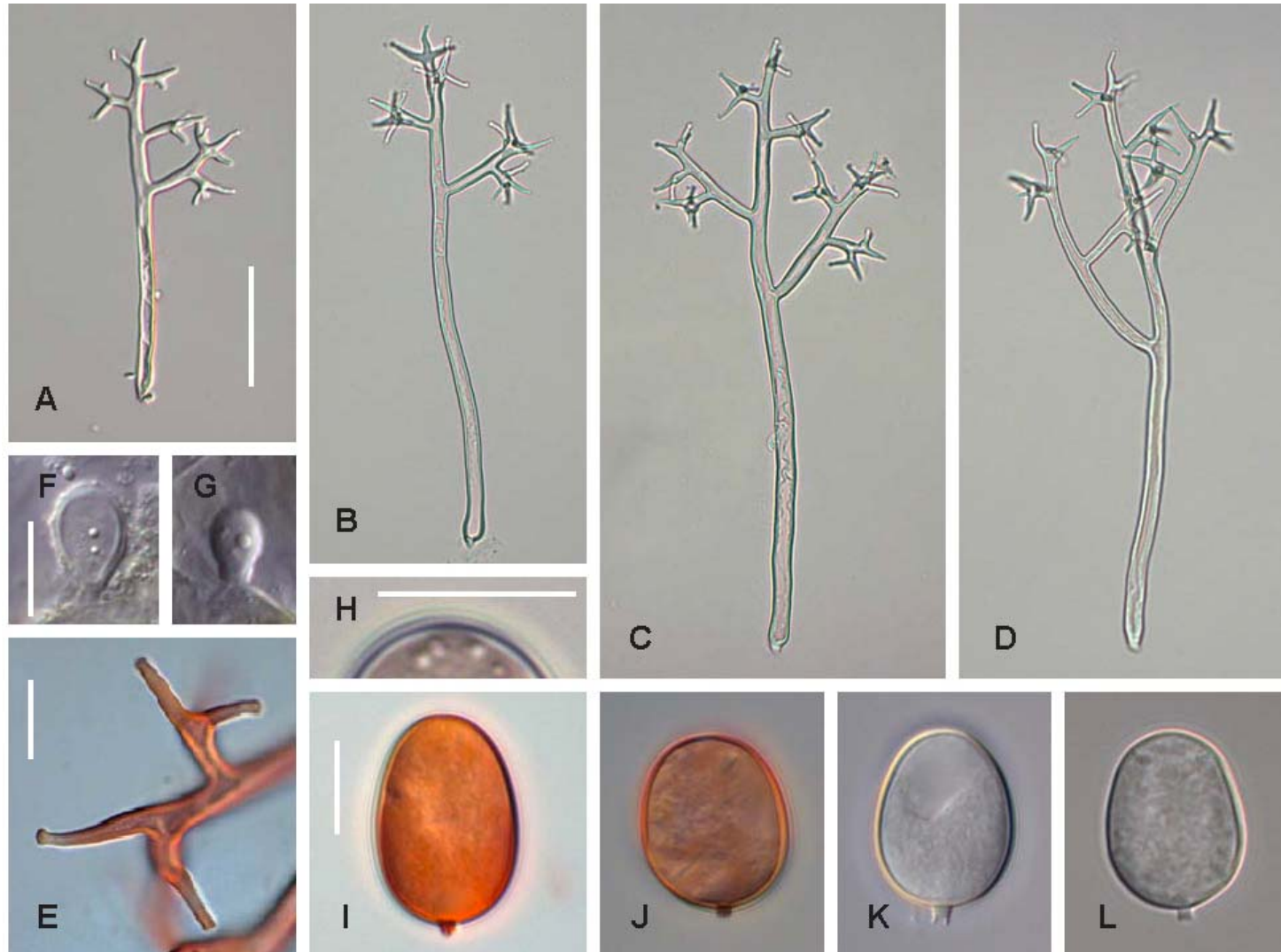
- applying a phylogenetic species concept, a narrow species circumscription seems to be more appropriate in most cases
- narrow host range should be a central factor for genetic isolation and speciation in downy mildews – strong genetic isolation barriers due to host specificity (no evidence for hybridisation, high genetic change)
- host jumps to unrelated hosts occurred frequently, followed by rapid genetic change
- the popular “one host family-one species” concept does not conform with a modern phylogenetic species concept. In addition, uncritical adherence to it can have severe practical consequences and problems (e.g. *Peronospora* on sweet basil)
- more appropriate to formally classify cryptic species
- methodologically, we currently rely on indirect evidence for genetic isolation by molecular data (mainly sequences). Most investigations are based on a single or few sequence regions

# Conclusions

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- for identification, molecular tools are most reliable and indispensable for downy mildews. For development of a reliable identification system (“barcoding”), additional investigations are needed
- for barcoding, the most important step is the choice of the region to be primarily used. As ITS is inappropriate for some important groups, mitochondrial DNA (cox?) may be a good candidate, which needs additional investigations
- a barcoding approach must be accompanied by thorough taxonomic revisions in order to clarify and stabilise species nomenclature
- additional investigations are also needed to appropriately document the biodiversity of downy mildews. Most detailed biodiversity investigations are more than 50 years old. Numerous distinct species still await description.

# Thank you for your attention!



*Plasmopara euphrasiae* Voglmayr & Constantinescu (2008)

# Literature

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- Belbahri, L., Calmin, G., Pawlowski, J. & Lefort F. (2005). Phylogenetic analysis and Real Time PCR detection of a presumably undescribed *Peronospora* species on sweet basil and sage. *Mycological Research*, 109, 1276-1287.
- Choi, Y.-J., Hong, S.-B. & Shin, H.-D. (2005). A reconsideration of *Pseudoperonospora cubensis* and *P. humuli* based on molecular and morphological data. *Mycological Research*, 109, 841-848.
- Choi, Y.-J., Denchev, C. M. & Shin, H.-D. (2008). Morphological and molecular analyses support the existence of host-specific *Peronospora* species infecting *Chenopodium*. *Mycopathologia*, 168, 155-164.
- Choi, Y.-J., Hong, S.-B. & Shin, H.-D. (2007c). Re-consideration of *Peronospora farinosa* infecting *Spinacia oleracea* as distinct species, *Peronospora effusa*. *Mycological Research*, 110, 381-391.
- Gäumann, E. (1918). Über die Formen der *Peronospora parasitica* (Pers.) Fries. *Beihefte zum Botanischen Centralblatt*, 35, 395-533.
- Gäumann, E. (1923). Beiträge zu einer Monographie der Gattung *Peronospora* Corda. *Beiträge zur Kryptogamenflora der Schweiz*, 5, 1-360.
- García-Blázquez, G., Göker, M., Voglmayr, H., Martín, M. P., Tellería, M. T. & Oberwinkler F. (2008). Phylogeny of *Peronospora*, parasitic on Fabaceae, based on ITS sequences. *Mycological Research*, 112, 502-512.
- Göker, M., Riethmüller, A., Voglmayr, H., Weiß, M. & Oberwinkler, F. (2004). Phylogeny of *Hyaloperonospora* based on nuclear ribosomal internal transcribed spacer sequences. *Mycological Progress* 3, 83-94.
- Göker, M., Voglmayr, H., Riethmüller, A. & Oberwinkler, F. (2007). How do obligate parasites evolve? A multi-gene phylogenetic analysis of downy mildews. *Fungal Genetics and Biology*, 44, 105-122.
- Voglmayr, H., Fatehi, J. & Constantinescu, O. (2006). Revision of *Plasmopara* (Chromista, Peronosporales) parasitic on Geraniaceae. *Mycological Research*, 110, 633-645.
- Voglmayr, H., Riethmüller, A., Göker, M., Weiß, M. & Oberwinkler, F. (2004). Phylogenetic relationships of *Plasmopara*, *Bremia* and other genera of downy mildews with pyriform haustoria based on Bayesian analysis of partial LSU rDNA sequence data. *Mycological Research*, 108, 1011-1024.
- Yerkes, W. D. & Shaw, C. G. (1959). Taxonomy of *Peronospora* species on Cruciferae and Chenopodiaceae. *Phytopathology*, 49, 499-507.