

doi.org/10.3114/fuse.2019.04.05

Setophoma spp. on *Camellia sinensis*

F. Liu¹, J. Wang^{1,2}, H. Li³, W. Wang⁴, L. Cai^{1,2*}

¹State key Laboratory of Mycology, Institute of Microbiology, Chinese Academy of Sciences, Beijing, 100101, China

²College of Life Sciences, University of Chinese Academy of Sciences, Beijing 100049, China

³College of Life Sciences, Hebei University, Baoding, Hebei Province, 071002, China

⁴Shandong Hetian Wang Biological Technology Co., Ltd., WeiFang, 261300, China

*Corresponding author: cail@im.ac.cn

Key words:

five new taxa
fungal pathogen
phylogeny
taxonomy
tea plants

Abstract: During our investigation of *Camellia sinensis* diseases (2013–2018), a new leaf spot disease was found in seven provinces of China (Anhui, Fujian, Guangxi, Guizhou, Jiangxi, Tibet and Yunnan), occurring on both arboreal and terraced tea plants. The leaf spots were round to irregular, brown to dark brown, with grey or tangerine margins. Multi-locus (LSU, ITS, *gapdh*, *tef-1α*, *tub2*) phylogenetic analyses combined with morphological observations revealed four new species belonging to the genus *Setophoma*, i.e. *S. antiqua*, *S. longinqua*, *S. yingyisheniae* and *S. yunnanensis*. Of these four species, *S. yingyisheniae* was found to be present on diseased terraced tea plants in six of the seven sampled provinces (excluding Yunnan). The other three species only occurred on arboreal tea plants in Yunnan Province. In addition to the four species isolated from diseased leaves, *S. endophytica* sp. nov. was isolated from healthy leaves of terraced tea plants.

Effectively published online: 15 May 2019.

INTRODUCTION

During our investigation of diseases of tea plants (*Camellia sinensis*) cultivated in China in 2013, a new leaf spot disease was found to cause severe yield losses in Guangxi Province. The associated leaf spots were circular to irregular, brown to dark brown in colour, with grey or tangerine margins. A subsequent collection effort of similarly affected tea plant leaves has been ongoing in many tea plantations located in six provinces of China, i.e. Anhui, Fujian, Guizhou, Jiangxi, Tibet and Yunnan. Preliminary morphological observations and molecular analyses identified the associated fungi as *Setophoma* spp. To our knowledge, this is the first report of *Setophoma* on tea plants. Considering the potential commercial yield losses in tea plantations and the limited knowledge of this disease, accurate identification of the causal organisms is of great importance.

The genus *Setophoma* (*Phaeosphaeriaceae*) was introduced to accommodate *Phoma terrestris* and *Pyrenochaeta sacchari* (de Gruyter *et al.* 2010). Species of *Setophoma* are characterised as having setose pycnidia, phialidic conidiogenous cells and hyaline, ellipsoidal to subcylindrical, aseptate conidia (de Gruyter *et al.* 2010, Quaedvlieg *et al.* 2013). According to Index Fungorum and MycoBank, four additional *Setophoma* species have been described since the genus was introduced in 2010. The currently recognised species are: *S. chromolaenae*, *S. cyperi*, *S. poaeicola*, *S. sacchari*, *S. terrestris*, and *S. vernoniae*. All except *S. terrestris* are reported to occur on unique host plants (Table 1).

The aim of the present study was to investigate the taxonomic and phylogenetic relationships of *Setophoma* spp. associated with tea plants based on multi-locus phylogenetic analyses,

morphological comparison, host association and geographical distribution.

MATERIALS AND METHODS

Isolates

Isolates were obtained from either diseased or healthy tea plant tissues collected from 18 locations in seven provinces of China, following the single spore isolation and tissue isolation methods described in Liu *et al.* (2015). Type specimens of new species were deposited in the Mycological Herbarium of the Institute of Microbiology, Chinese Academy of Sciences, Beijing, China (HMAS), with the ex-type living cultures being deposited in the China General Microbiological Culture Collection Center (CGMCC).

DNA extraction, PCR amplification and phylogenetic analyses

Total genomic DNA was extracted from fresh mycelia using the CTAB method. Five partial loci including the large subunit of the nrDNA (LSU), the 5.8S nuclear ribosomal RNA gene with the two flanking internally transcribed spacer regions (ITS), partial glyceraldehyde-3-phosphate dehydrogenase (*gapdh*), translation elongation factor 1- α (*tef-1α*) and β -tubulin (*tub2*) were amplified and sequenced using the following primer pairs: ITS1/ITS4 for ITS (White *et al.* 1990), LR0R/LR5 for LSU (Vilgalys & Hester 1990, Rehner & Samuels 1994), *gpd1/gpd2* for *gapdh* (Berbee *et al.* 1999), EF-1/EF-2 for *tef-1α* (O'Donnell *et al.*

Table 1. Host and distribution of *Setophoma* species.

Species	Host	Distribution	References
<i>S. antiqua</i>	<i>Camellia sinensis</i>	China	This study
<i>S. chromolaenae</i>	<i>Chromolaena odorata</i>	Brazil	Quaedvlieg <i>et al.</i> (2013)
<i>S. cyperi</i>	<i>Cyperus sphaerocephalus</i>	South Africa, Eastern Cape	Crous <i>et al.</i> (2016)
<i>S. endophytica</i>	<i>Camellia sinensis</i>	China	This study
<i>S. longinqua</i>	<i>Camellia sinensis</i>	China	This study
<i>S. poaceicola</i>	Grass	Thailand	Thambugala <i>et al.</i> (2017)
<i>S. sacchari</i>	<i>Saccharum officinarum</i>	Brazil	de Gruyter <i>et al.</i> (2010)
<i>S. terrestris</i>	<i>Allium cepa</i>	North America, Senegal	de Gruyter <i>et al.</i> (2010)
	<i>Allium sativum</i>	United States	de Gruyter <i>et al.</i> (2010)
	<i>Brassica</i> sp.	Canada, Alberta	Yang <i>et al.</i> (2017)
	<i>Cucurbita maxima</i>	USA, Oregon	Rivedal <i>et al.</i> (2018)
	<i>Cucurbita moschata</i>	Japan	Ikeda <i>et al.</i> (2012)
<i>S. yunnanensis</i>	<i>Solanum lycopersicum</i>	Canada, Ontario	Johnston-Monje <i>et al.</i> (2017)
	<i>Vernonia polyanthes</i>	Brazil	Crous <i>et al.</i> (2014)
<i>S. yingyisheniae</i>	<i>Camellia sinensis</i>	China	This study
<i>S. yunnanensis</i>	<i>Camellia sinensis</i>	China	This study

Table 2. GenBank accession numbers from NCBI database.

Name	Strain/specimen No. ^a	LSU	ITS
<i>Acericola italica</i>	MFLUCC 13-0609*	MF167429	MF167428
<i>Allophaeosphaeria muriformia</i>	MFLUCC 13-0349*	KP765681	KP765680
<i>Allophaeosphaeria subcylindrospora</i>	MFLUCC 13-0380*	KT314183	KT314184
<i>Amarenomyces ammophilae</i>	CBS 114595	GU301859	KF766146
<i>Ampelomyces quisqualis</i>	CBS 129.79*	EU754128	HQ108038
<i>Bhatiellae rosae</i>	MFLUCC 17-0664*	MG828989	MG828873
<i>Chaetosphaeronema achilleae</i>	MFLUCC 16-0476	KX765266	KX765265
<i>Chaetosphaeronema hispidulum</i>	CBS 216.75	KF251652	KF251148
<i>Coniothyrium carteri</i>	CBS 105.91	KF251712	KF251209
<i>Dactylidina dactylidis</i>	MFLUCC 14-0966*	MG829002	MG828886
<i>Dematiopleospora rosicola</i>	MFLU 16-0232*	MG829006	MG828888
<i>Dematiopleospora salsolae</i>	MFLUCC 17-0828*	MG829007	MG828889
<i>Didymocyrtis consimilis</i>	Gardiennet 12041	KT383796	KT383813
<i>Didymocyrtis ramalinae</i>	Ertz 16399	KT383802	KT383838
<i>Embarria clematidis</i>	MFLUCC 14-0976*	MG828987	MG828871
<i>Galiicola pseudophaeosphaeria</i>	MFLUCC 14-0524*	KT326693	KT326692
<i>Hawksworthiana alliariae</i>	MFLUCC 13-0070	KX494877	KX494876
<i>Hawksworthiana clematidicola</i>	MFLUCC 14-0910*	MG829011	MG828901
<i>Italica achilleae</i>	MFLUCC 14-0959*	MG829013	MG828903
<i>Juncaceicola achilleae</i>	MFLUCC 13-0606*	KX449526	KX449525
<i>Juncaceicola luzulae</i>	MFLUCC 16-0780*	KX449530	KX449529
<i>Juncaceicola typharum</i>	CBS 296.54	KF251695	KF251192
<i>Leptospora galii</i>	KUMCC 15-0521	KX599548	KX599547
<i>Leptospora rubella</i>	CPC 11006	DQ195792	DQ195780
<i>Muriphaeosphaeria galatellae</i>	MFLUCC 14-0614*	KT438329	KT438333
<i>Neosetophoma samarorum</i>	CBS 138.96*	KF251664	KF251160
<i>Neostagonospora caricis</i>	CBS 135092*	KF251667	KF251163
<i>Neostagonospora elegiae</i>	CBS 135101*	KF251668	KF251164

Table 2. (Continued).

Name	Strain/specimen No. ^a	LSU	ITS
<i>Neosulcatispora agaves</i>	CPC 26407*	KT950867	KT950853
<i>Nodosphaeria hirta</i>	MFLUCC 13-0867	KU708845	KU708849
<i>Nodosphaeria scabiosae</i>	MFLUCC 14-1111*	KU708846	KU708850
<i>Ophiobolopsis italica</i>	MFLUCC 17-1791*	MG520959	MG520939
<i>Ophiobolus artemisiae</i>	MFLUCC 14-1156*	KT315509	KT315508
<i>Ophiobolus disseminans</i>	MFLUCC 17-1787*	MG520961	MG520941
<i>Ophiosimulans tanacetii</i>	MFLUCC 14-0525	KU738891	KU738890
<i>Paraophiobolus arundinis</i>	MFLUCC 17-1789*	MG520965	MG520945
<i>Paraophiobolus plantaginis</i>	MFLUCC 17-0245*	KY815010	KY797641
<i>Paraphoma chrysanthemicola</i>	CBS 172.70	KF251669	KF251165
<i>Paraphoma radicina</i>	CBS 102875	KF251677	KF251173
<i>Paraphoma raphiolepidis</i>	CBS 142524*	KY979813	KY979758
<i>Paraphoma vinacea</i>	UMPV001 = BRIP 63684	KU176888	KU176884
<i>Parastagonospora caricis</i>	CBS 135671	KF251680	KF251176
<i>Parastagonospora nodorum</i>	CBS 110109	KF251681	KF251177
<i>Parastagonospora poagena</i>	CBS 136776*	KJ869174	KJ869116
<i>Parastagonospora fallopiae</i>	CBS 135981*	MH460545	MH460543
	CCTU 1151.1	MH460546	MH460544
<i>Phaeosphaeria oryzae</i>	CBS 110110*	KF251689	KF251186
<i>Phaeosphaeria papayae</i>	CBS 135416	KF251690	KF251187
<i>Phaeosphaeriopsis glaucopunctata</i>	CBS 653.86	KF251702	KF251199
<i>Poaceicola arundinis</i>	MFLU 15-0702*	KU058726	KU058716
<i>Poaceicola italica</i>	MFLUCC 13-0267*	KX910094	KX926421
<i>Populocrescentia forlicesenensis</i>	MFLU 15-0651*	KT306952	KT306948
<i>Pseudoophiobolus achilleae</i>	MFLU 17-0925*	MG520966	MG520946
<i>Pseudoophiobolus mathieui</i>	MFLUCC 17-1784	MG520969	MG520949
<i>Pseudophaeosphaeria rubi</i>	MFLUCC 14-0259*	KX765299	KX765298
<i>Sclerostagonospora rosicola</i>	MFLUCC 15-0129*	MG829068	MG828957
<i>Septoriella allojunci</i>	MFLU 15-0701*	KU058728	KU058718
<i>Septoriella phragmitis</i>	CPC 24118 = CBS 140065*	KR873279	KR873251
<i>Setophoma chromolaenae</i>	CBS 135105*	KF251747	KF251244
<i>Setophoma cyperi</i>	CPC 25702 = CBS 141450*	KX228337	KX228286
<i>Setophoma poaeicola</i>	MFLUCC 16-0880*	KY550386	KY568988
<i>Setophoma sacchari</i>	MFLUCC 12-0241	KJ476147	KJ476145
	CBS 333.39*	MH867535	MH856038
<i>Setophoma terrestris</i>	CBS 335.87	KF251750	KF251247
	CBS 377.52	KF251751	KF251248
<i>Setophoma vernoniae</i>	CPC 23123 = CBS 137988*	KJ869198	KJ869141
<i>Sulcispora pleurospora</i>	MFLUCC 14-0995*	KP271444	KP271443
<i>Tintelnotia destructans</i>	CBS 127737*	KY090664	KY090652
<i>Tintelnotia opuntiae</i>	CBS 376.91*	GU238123	KY090651
<i>Vagicola vagans</i>	CBS 604.86	KF251696	KF251193
<i>Wojnowicia lonicerae</i>	MFLUCC 13-0737*	KP684151	KP744471
<i>Wojnowicia rosicola</i>	MFLUCC 15-0128*	MG829091	MG828979
<i>Wojnowiciella eucalypti</i>	CPC 25024*	KR476774	KR476741
<i>Xenoseptoria neosaccardoi</i>	CBS 120.43	KF251783	KF251280
	CBS 128665*	KF251784	KF251281

^aEx-type strains are marked with asterisk *.

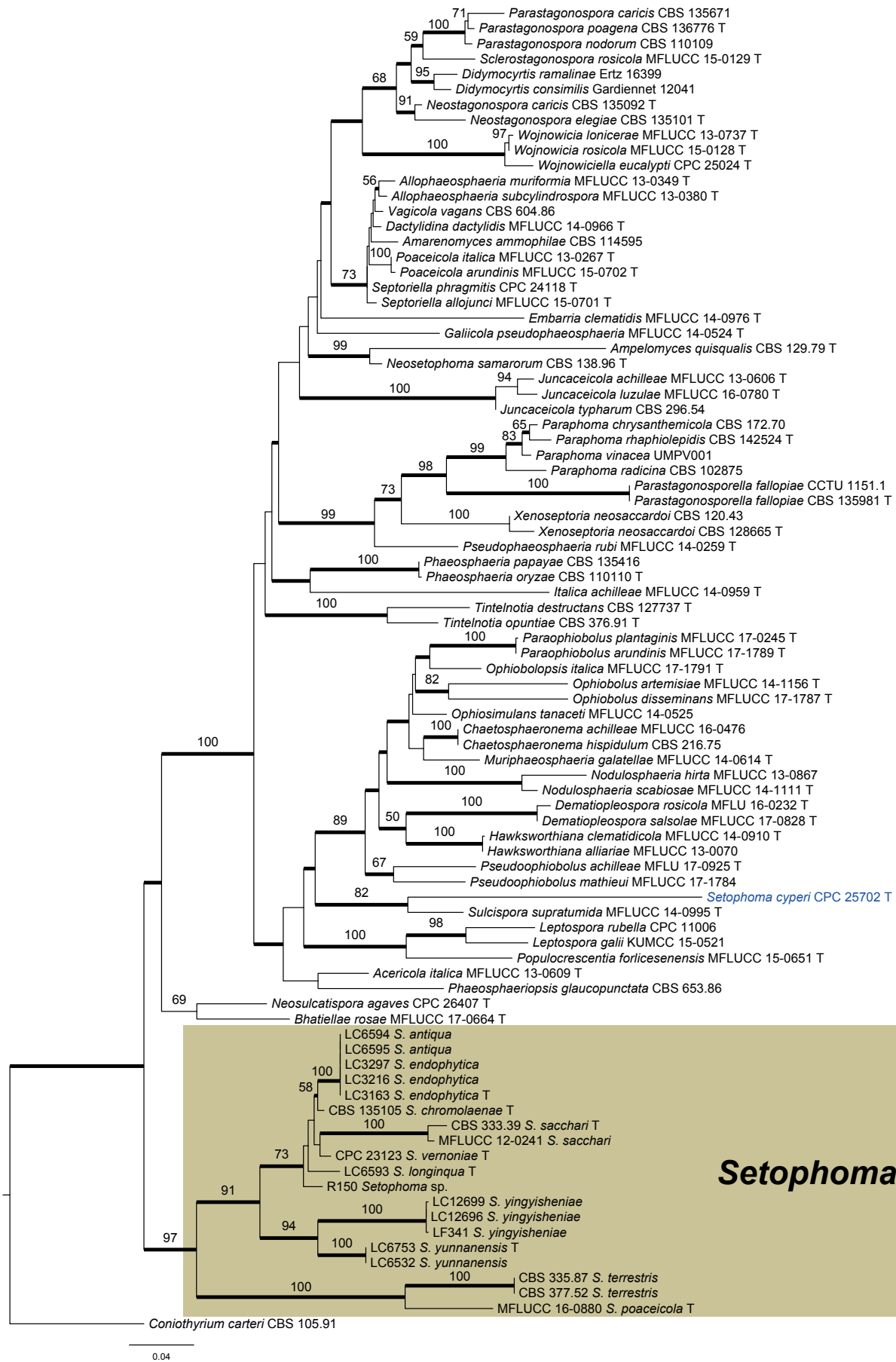


Fig. 1. Overview phylogenetic tree of *Phaeosphaeriaceae* (50 % majority rule consensus) resulting from a Bayesian analysis of the combined LSU and ITS sequence alignment. Bayesian posterior probabilities (PP > 0.95) are emphasized by thickened branches, maximum likelihood bootstrap support values (≥ 50 %) are shown at the nodes. The scale bar represents the expected number of changes per site. Ex-type cultures are indicated with “T” behind the taxa labels. The tree was rooted to *Coniothyrium carteri* (CBS 105.91).

1998) and T1/Bt2b for *tub2* (Glass & Donaldson 1995, O'Donnell & Cigelnik 1997). Amplicons for LSU, ITS, *tef-1α* and *tub2* were generated according to Liu *et al.* (2019) while amplicons for *gapdh* were generated according to Liu *et al.* (2016). Amplicons were sequenced with both forward and reverse primers by the Omegagenetics Company, Beijing, China. MEGA v. 7.0.21 was used to generate consensus sequences.

DNA sequences (ITS and LSU) used to infer generic affiliation within the *Phaeosphaeriaceae* were downloaded from the NCBI database (Table 2). Sequence datasets of ITS, *gapdh*, *tub2* and *tef-1α* (Table 3) were used for species delimitation in *Setophoma*. Sequence alignments were made using MAFFT v. 7 (<http://mafft.cbrc.jp/alignment/server/index.html>) and were then manually edited in MEGA v. 7.0.21. Subsequently, individual alignments of above-mentioned loci were concatenated using Mesquite v. 3.4. The Maximum Likelihood (ML) and Bayesian analysis (BA) methods were used for phylogenetic inferences of both the single gene and concatenated alignments as described in Liu *et al.* (2019). The resulting trees were plotted using FIGTREE v. 1.4.2 (<http://tree.bio.ed.ac.uk/software/figtree>). All the alignments derived from this study were deposited in TreeBASE (<https://treebase.org/>) (S24160).

Morphology

Cultures were cultivated on both 2 % malt extract agar (MEA) and potato dextrose agar (PDA) (Difco™, Becton, Dickinson and Company, Sparks, MD, USA) at 25 °C in a 12 h day/night regime. Growth rates were measured after 10 d in the dark. Colony colours were rated following the colour charts of Rayner (1970). In order to observe wall structures of mature conidiomata, sections were made using a Leica CM1950 freezing microtome. Morphological observations of reproductive structures were made in lactic acid and observed using a Nikon Eclipse 80i microscope using differential interference contrast (DIC) illumination. At least 30 measurements per structure were taken and the mean value, standard deviation and minimum–maximum values are given, with the extreme measurements given in parentheses.

RESULTS

DNA phylogeny

The concatenated alignment of LSU and ITS sequences comprised a total length of 1 397 characters (LSU: 818, ITS: 579) including alignment gaps, 486 of which were unique site patterns. The ML search resolved a best tree with a InL of -14098.620225. Fixed base frequency for LSU and ITS was set in the Bayesian analysis (BA). The BA lasted for 1 305 000 generations and the 50 % consensus tree and posterior probabilities were calculated from 3 908 trees, generated during two runs. The ML tree confirmed the same tree topology as that generated by the Bayesian analysis and it also revealed that strains isolated in this study clustered together with the generic type of *Setophoma* (*S. terrestris*), in the basal clade of Fig. 1. However, the ex-type strain of another known *Setophoma* species, *S. cyperi* (CPC

25702 = CBS 141450) clustered outside *Setophoma s. str.*, and was more closely related to *Sulcispora pleurospora* (MFLUCC 14-0995) (Fig. 1).

To better infer species delimitation in *Setophoma*, sequences of four loci (ITS, *tef-1α*, *gapdh* and *tub2*) were concatenated for further phylogenetic analyses. Strains of *S. sacchari*, *S. vernoniae* and *S. poaceicola* were excluded from this analysis due to their incomplete sequence datasets. The final alignment contained a total of 2 504 characters (ITS: 524, *tef-1α*: 875, *gapdh*: 550, *tub2*: 539) including alignment gaps, of which 872 were unique site patterns. The ML search revealed a best tree with an InL of -10791.633622. Fixed base frequency was set in the Bayesian analysis. The BA lasted for 1 315 000 generations and the 50 % consensus tree and posterior probabilities were calculated from 1 656 trees generated during two runs. The topology of the phylogenetic trees generated from both ML and BA methods were congruent (Fig. 2). Strains associated with tea plants clustered in five different clades.

Taxonomy

A total of 94 isolates from diseased leaves and 17 isolates from healthy leaves were isolated from arboreal and terraced tea plants. Based on ITS sequence similarity and colonial morphology, 35 representative isolates were selected for further microscopic observation and phylogenetic analyses. These 35 strains formed five distinct and novel phylogenetic clades representing five new species (Fig. 2). These are described and illustrated below.

Setophoma Gruyter, Aveskamp & Verkley, *Mycologia* **102**: 1077. 2010. **emend.** F. Liu & L. Cai.

Ascomata immersed or semi-immersed, uniloculate, globose to subglobose, brown to dark brown, solitary or gregarious, centrally ostiolate, papillate, wall of *textura angularis* or *prismatica*. *Pseudoparaphyses* hyaline, frequently septate. *Asci* bitunicate, fissitunicate, cylindrical to cylindrical-clavate, smooth-walled. *Ascospores* fusiform with rounded ends, hyaline, overlapping or irregularly biseriolate, septate, usually widest at the second cell from apex, smooth-walled, guttulate. *Conidiomata* pycnidial, solitary to confluent, on upper surface or submerged in agar, globose to subglobose, setose or not, with papillate ostioles, honey to olivaceous or olivaceous black; *pycnidial wall* of pseudoparenchymatal cells. *Conidiophores* reduced to conidiogenous cells lining inner cavity, or simply branched. *Conidiogenous cells* hyaline, smooth, phialidic, discrete. *Conidia* aseptate, globose, subglobose, ellipsoidal to subcylindrical to subfusoid, guttulate.

Type species: *S. terrestris* (H.N. Hansen) Gruyter *et al.*

Notes: The genus *Setophoma* was introduced to accommodate phoma-like species with setose conidiomata by de Gruyter *et al.* (2010). However, the new species of *S. antiqua*, *S. endophytica* and *S. yunnanensis*, proposed in this study, lack setae. We therefore broadened the generic concept of the genus *Setophoma* to also accommodate species lacking setose conidiomata.

Table 3. Collection details and GenBank accession numbers of isolates included in this study.

Species	Strain No. ^a	Habitat	Location	GenBank accession no. ^b				
				LSU	ITS	tub2	tef-1 α	GAPDH
<i>Didymella pinodella</i>	CBS 531.66	<i>Trifolium pretense</i>	USA	GU238017	FJ427052	FJ427162	MK525067	MK532379
	LC6594	<i>Camellia sinensis</i> , pathogen	Yunnan, China	MK511947	MK511909	MK524999	MK525070	MK525034
<i>Setophoma antiqua</i>	LC6595	<i>Camellia sinensis</i> , pathogen	Yunnan, China	MK511948	MK511910	MK525000	MK525071	MK525035
	CGMCC 3.19525 = LC6596*	<i>Camellia sinensis</i> , pathogen	Yunnan, China	-	MK511911	MK525001	MK525072	MK525036
<i>Setophoma chromolaenae</i>	CBS 135105*	<i>Chromolaena odorata</i>	Brazil	KF251747	KF251244	KF252728	KF253195	-
<i>Setophoma endophytica</i>	LC13538	<i>Camellia sinensis</i> , endophyte	Jiangxi, China	-	MK511923	MK525012	MK525084	MK525047
	LF2067	<i>Camellia sinensis</i> , endophyte	Jiangxi, China	-	MK511924	MK525013	MK525085	MK525048
	CGMCC 3.19528 = LC3163*	<i>Camellia sinensis</i> , endophyte	Jiangxi, China	MK511956	MK511931	MK525020	MK525092	MK525053
	LC3164	<i>Camellia sinensis</i> , endophyte	Jiangxi, China	MK511957	MK511932	MK525021	MK525093	MK525054
	LC3165	<i>Camellia sinensis</i> , endophyte	Jiangxi, China	-	MK511933	MK525022	MK525094	MK525055
	LC3216	<i>Camellia sinensis</i> , endophyte	Jiangxi, China	MK511959	MK511938	MK525026	MK525099	MK525060
<i>Setophoma longinqua</i>	LC3297	<i>Camellia sinensis</i> , endophyte	Jiangxi, China	MK511962	MK511941	MK525029	MK525102	MK525063
	CGMCC 3.19524 = LC6593*	<i>Camellia sinensis</i> , pathogen	Yunnan, China	MK511946	MK511908	MK524998	MK525069	-
	LC13481	<i>Camellia sinensis</i> , pathogen	Yunnan, China	-	MK511925	MK525014	MK525086	-
	LC13482	<i>Camellia sinensis</i> , pathogen	Yunnan, China	-	MK511926	MK525015	MK525087	-
<i>Setophoma</i> sp.	LC12841	Carbonatite in cave	Guizhou, China	-	MK511927	MK525016	MK525088	MK525049
	LC12842	Carbonatite in cave	Guizhou, China	-	MK511928	MK525017	MK525089	MK525050
<i>Setophoma terrestris</i>	CGMCC 3.19526 = LC7511	Carbonatite in cave	Guizhou, China	MK511965	MK511944	MK525032	MK525105	MK525066
	CBS 335.29 = MUCL 9892 = LC6449*	<i>Allium sativum</i>	USA	KF251749	KF251246	KF252729	KF253196	-
	CBS 335.87	<i>Allium cepa</i>	Senegal	KF251750	KF251247	KF252730	KF253197	-
	CBS 377.52	<i>Allium cepa</i>	-	KF251751	KF251248	KF252731	KF253198	-
<i>Setophoma yingyisheniae</i>	LC6739	<i>Camellia sinensis</i> , pathogen	Guizhou, China	-	MK511912	MK525002	MK525073	-
	LC12696	<i>Camellia sinensis</i> , pathogen	Anhui, China	MK511950	MK511914	-	MK525075	MK525038
	LC12699	<i>Camellia sinensis</i> , pathogen	Anhui, China	MK511951	MK511915	MK525004	MK525076	MK525039
	LC13477	<i>Camellia sinensis</i> , pathogen	Fujian, China	MK511952	MK511916	MK525005	MK525077	MK525040
	LC13478	<i>Camellia sinensis</i> , pathogen	Fujian, China	-	MK511917	MK525006	MK525078	MK525041
	CGMCC 3.19527 = LC13479*	<i>Camellia sinensis</i> , pathogen	Guangxi, China	-	MK511918	MK525007	MK525079	MK525042

Table 3. (Continued).

Species	Strain No. ^a	Habitat	Location	GenBank accession no. ^b				
				LSU	ITS	tub2	tef-1 α	GAPDH
	LC13480	<i>Camellia sinensis</i> , pathogen	Guangxi, China	MK511953	MK511919	MK525008	MK525080	MK525043
	LF2026	<i>Camellia sinensis</i> , pathogen	Guangxi, China	-	MK511920	MK525009	MK525081	MK525044
	LF2027	<i>Camellia sinensis</i> , pathogen	Guangxi, China	MK511954	MK511921	MK525010	MK525082	MK525045
	LF2032	<i>Camellia sinensis</i> , pathogen	Guangxi, China	-	MK511922	MK525011	MK525083	MK525046
	LC3133	<i>Camellia sinensis</i> , pathogen	Jiangxi, China	MK511955	MK511929	MK525018	MK525090	MK525051
	LC3137	<i>Camellia sinensis</i> , pathogen	Jiangxi, China	-	MK511930	MK525019	MK525091	MK525052
	LC3176	<i>Camellia sinensis</i> , endophyte	Jiangxi, China	-	MK511934	-	MK525095	MK525056
	LC3181	<i>Camellia sinensis</i> , endophyte	Jiangxi, China	MK511958	MK511935	MK525023	MK525096	MK525057
	LC3185	<i>Camellia sinensis</i> , endophyte	Jiangxi, China	-	MK511936	MK525024	MK525097	MK525058
	LC3197	<i>Camellia sinensis</i> , endophyte	Jiangxi, China	-	MK511937	MK525025	MK525098	MK525059
	LC3236	<i>Camellia sinensis</i> , pathogen	Jiangxi, China	MK511960	MK511939	MK525027	MK525100	MK525061
	LC3276	<i>Camellia sinensis</i> , pathogen	Jiangxi, China	MK511961	MK511940	MK525028	MK525101	MK525062
	LC3334	<i>Camellia sinensis</i> , pathogen	Jiangxi, China	MK511963	MK511942	MK525030	MK525103	MK525064
	LC3499	<i>Camellia sinensis</i> , endophyte	Guangxi, China	MK511964	MK511943	MK525031	MK525104	MK525065
<i>Setophoma yunnanensis</i>	LC6532	<i>Camellia sinensis</i> , pathogen	Yunnan, China	MK511945	MK511907	MK524997	MK525068	MK525033
	CGMCC 3.19529 = LC6753*	<i>Camellia sinensis</i> , pathogen	Yunnan, China	MK511949	MK511913	MK525003	MK525074	MK525037

^aEx-type strains are marked with star *; CBS: Culture collection of the Westerdijk Fungal Biodiversity Institute, Utrecht, The Netherlands; CGMCC: China General Microbiological Culture Collection Center, Institute of Microbiology, Chinese Academy of Sciences, Beijing, China; LC: working collection of Lei Cai, housed at the Institute of Microbiology, Chinese Academy of Sciences, Beijing, China; MUCL: Mycotèque de l'Université catholique de Louvain, Louvain-la-Neuve, Belgium.

^bNewly generated sequences are indicated in bold.

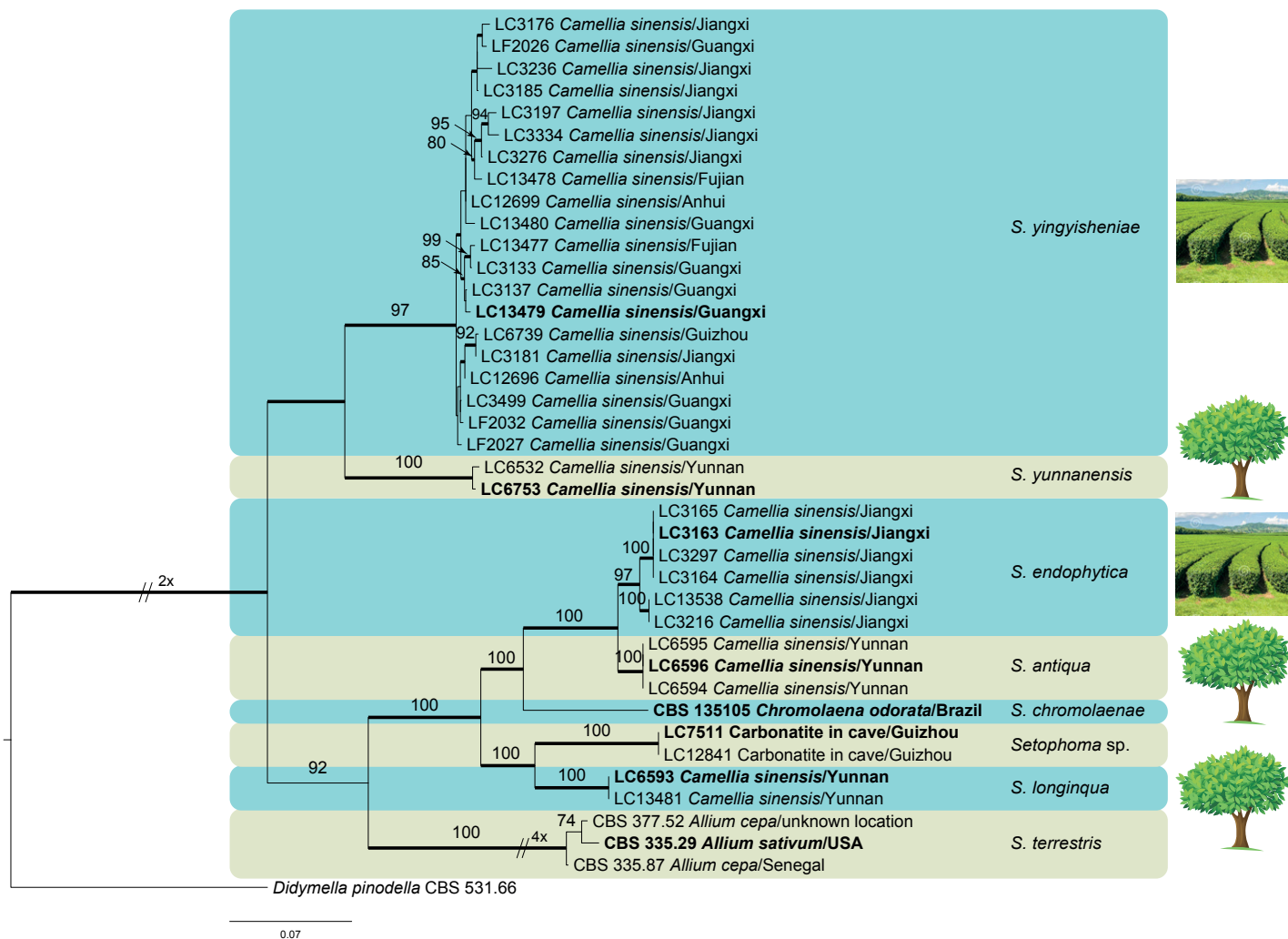


Fig. 2. Phylogenetic tree of *Setophoma* resulting from a Bayesian analysis of the combined ITS, *gapdh*, *tef-1 α* and *tub2* sequence alignment. Bayesian posterior probabilities (PP > 0.95) are emphasized by thickened branches, maximum likelihood bootstrap support values (> 50 %) are shown at the nodes. The scale bar represents the expected number of changes per site. The taxa names consist of strain number, host or substrate, and location. Ex-type strains are represented in bold. *Setophoma yingyisheniae* and *S. endophytica* were isolated from terraced tea plants, *S. yunnanensis*, *S. antiqua* and *S. longinqua* were isolated from arboreal tea plants (illustrated by photos on the right side of the figure). The tree was rooted to *Didymella pinodella* (CBS 531.66).

Setophoma antiqua F. Liu & L. Cai, *sp. nov.* MycoBank MB829900. Fig. 3.

Etymology: From the Latin “*antiquus*” = old, referring to its host, old tea plant.

Sexual morph: Unknown. **Asexual morph:** *Aerial mycelia* hyaline or pale brown, branched, septate. *Conidiomata* pycnidial, scattered or gregarious, immersed or semi-immersed, globose to subglobose, olivaceous, 100–250 μ m diam. *Pycnidial wall* pale to dark brown, 4–15 μ m wide, with 3–4 layers, walls of *textura angularis*. *Conidiophores* reduced to conidiogenous cells lining inner cavity. *Conidiogenous cells* single, phialidic, unbranched, aseptate, hyaline, smooth, ampulliform, 3–5.5 \times 2.5–5 μ m (av. = 3.8 \pm 0.6 \times 3.2 \pm 0.7 μ m). *Conidia* hyaline, aseptate, ellipsoid with rounded ends or allantoid, smooth-walled, 2.5–5 \times 1.5–2.5 μ m (av. = 3.7 \pm 0.5 \times 1.8 \pm 0.2 μ m).

Culture characteristics: On PDA, umbonate with entire edge, luteous in the centre, grey to deep grey at the edge, reverse deep grey, reaching 33–36 mm diam after 10 d at 25 $^{\circ}$ C. On MEA, flat with entire edge, front and reverse pale olivaceous in the

centre and pale grey at the edge, reaching 24–29 mm diam after 10 d at 25 $^{\circ}$ C.

Typus: China, Yunnan Province, Xishuangbanna, Mengla County, on old/arboreal *Camellia sinensis*, 18 Apr. 2015, F. Liu (**holotype** HMAS 248083, culture ex-type CGMCC 3.19525 = LC6596 = LF1239).

Additional materials examined: China, Yunnan Province, Xishuangbanna, Mengla County, on old/arboreal *Camellia sinensis*, 18 Apr. 2015, F. Liu, living cultures LC6594 = LF1237, LC6595 = LF1238.

Notes: Based on the multi-locus phylogeny (Fig. 2), *S. antiqua* is phylogenetically related to *S. endophytica* and has high LSU (100 %), ITS (99 %) and *gapdh* (99 %) sequence similarities with the later. However, both species can be well differentiated from each other based on their *tub2* (502/518, 97 % sequence similarity) and *tef-1 α* (561/613, 92 % sequence similarity) sequences. In addition, the habitat of both species are different. *Setophoma antiqua* was isolated from diseased leaf spots of arboreal tea plant from an unmanaged forest on the mountain in Yunnan Province, while *S. endophytica* was isolated from

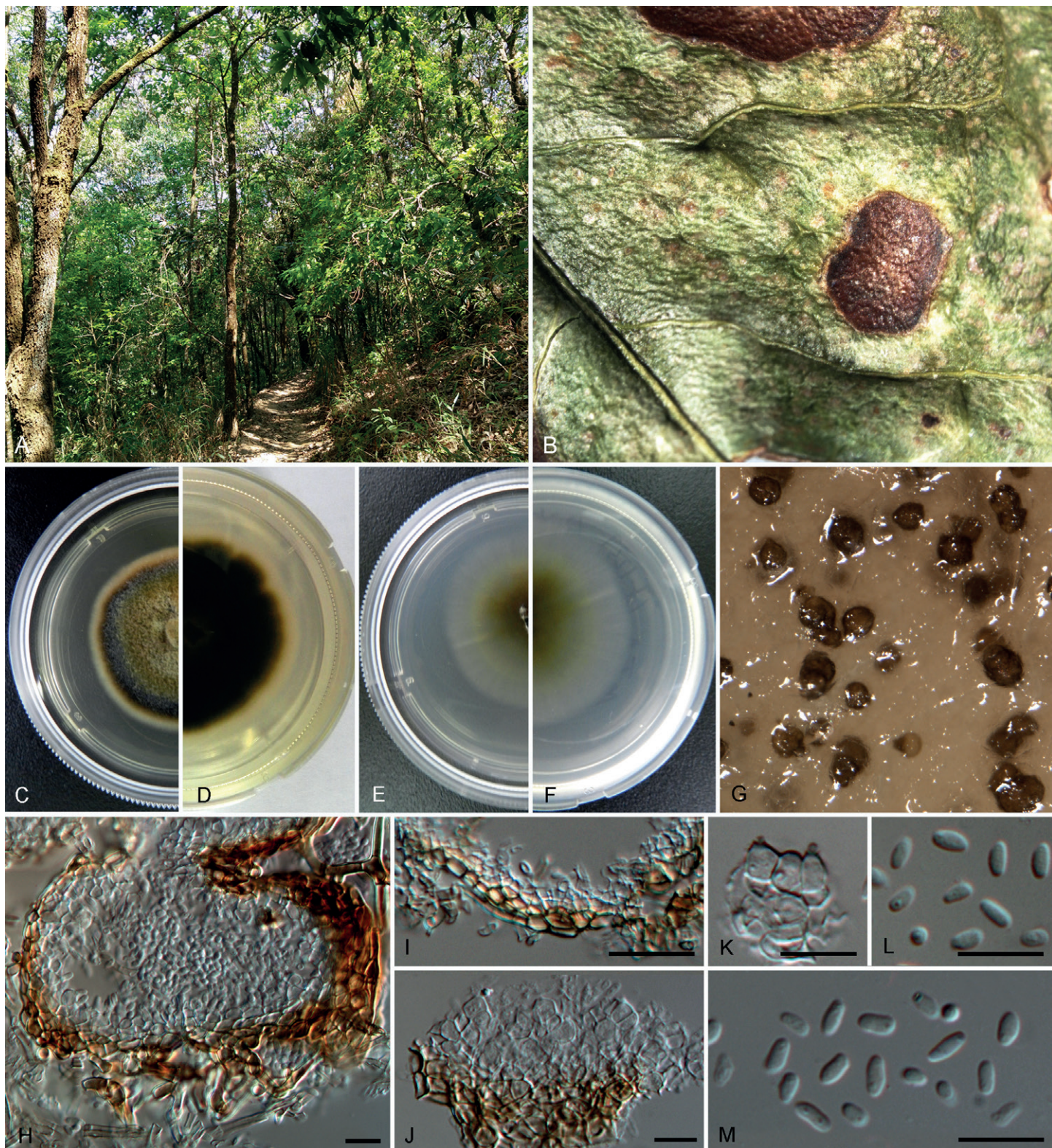


Fig. 3. *Setophoma antiqua* (ex-type CGMCC 3.19525 = LC6596). **A.** Sampling environment. **B.** Symptom of diseased leaf. **C, D.** Front and back of colony on PDA. **E, F.** Front and back of colony on MEA. **G.** Conidiomata. **H–J.** Vertical sections of conidiomata. **K.** Conidiogenous cells. **L, M.** Conidia. Scale bars = 10 μm .

healthy leaves of terraced tea plant from an open national forestry park in Jiangxi Province.

Setophoma endophytica F. Liu & L. Cai, *sp. nov.* MycoBank MB829902. Fig. 4.

Etymology: Named after its original habitat as an endophyte.

Sexual morph: Unknown. *Asexual morph:* Aerial mycelia hyaline, pale brown, smooth, branched, septate. *Conidiomata* immersed, olivaceous, globose to subglobose, scattered, 109–200 μm diam. *Pycnidial wall* pale brown, with 4–5 layers, 15–25 μm wide, walls of *textura angularis*. *Conidiophores* reduced to conidiogenous cells lining inner cavity. *Conidiogenous cells* hyaline, aseptate, smooth, ampulliform, rarely irregular, 3.5–

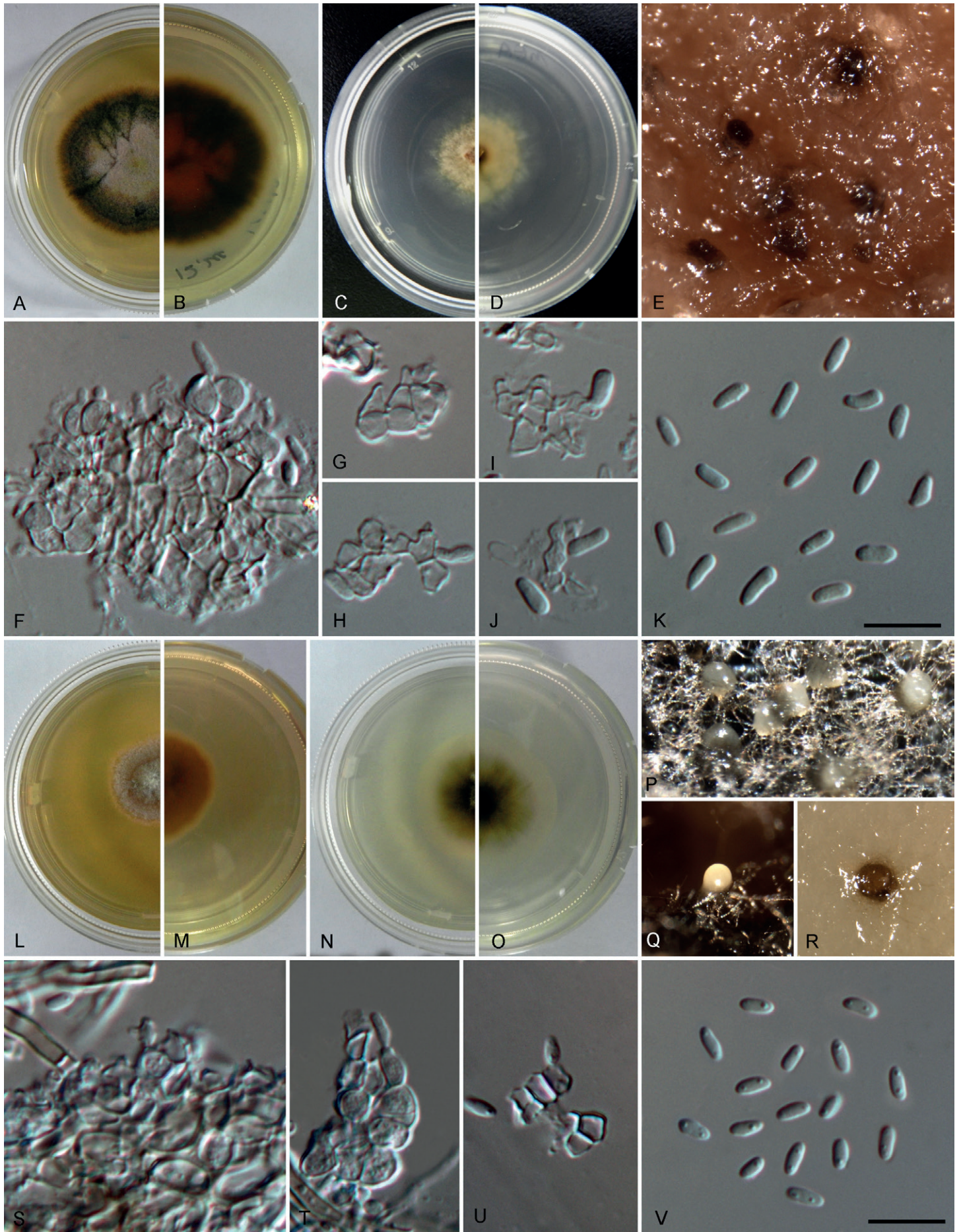


Fig. 4. *Setophoma endophytica* (A–K, ex-type CGMCC 3.19528 = LC3163, L–V, LC3216). A, B, L, M. Front and back of colonies on PDA. C, D, N, O. Front and back of colonies on MEA. E, P–R. Conidiomata. F–J, S–U. Conidiogenous cells and conidia. K, V. Conidia. Scale bars = 10 μm.

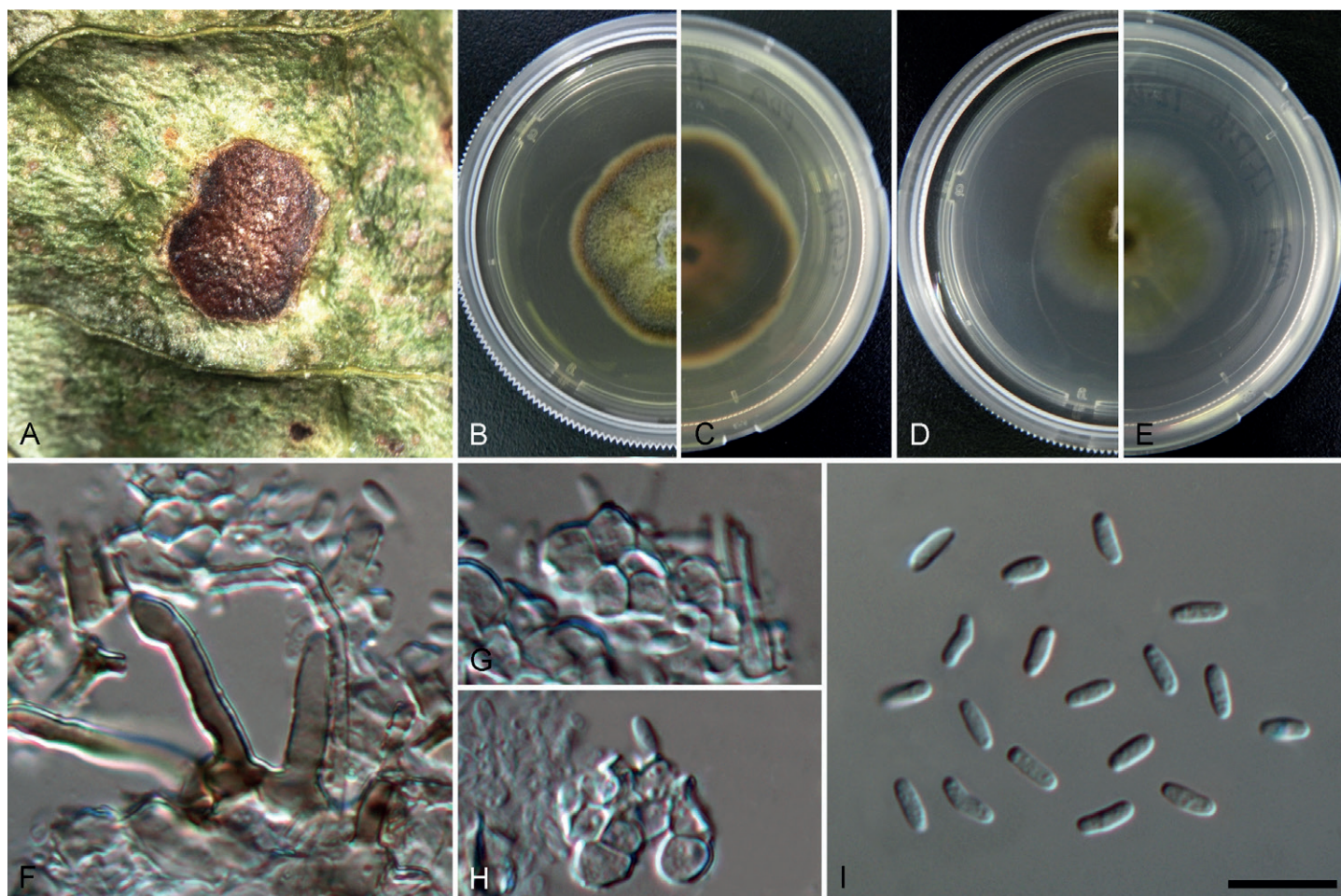


Fig. 5. *Setophoma longinqua* (ex-type CGMCC 3.19524 = LC6593). **A.** Symptom of diseased leaf. **B, C.** Front and back of colony on PDA. **D, E.** Front and back of colony on MEA. **F.** Setae. **G, H.** Conidiogenous cells and conidia. **I.** Conidia. Scale bars = 10 µm.

$6.5 \times 3\text{--}5.5$ µm (av. = $4.9 \pm 0.4 \times 3.6 \pm 0.3$ µm). *Conidia* hyaline, aseptate, smooth, cylindrical to reniform, with rounded ends, straight or slightly curved, $4\text{--}5.5 \times 1.5\text{--}2$ µm (av. = $4.8 \pm 0.2 \times 1.7 \pm 0.1$ µm).

Culture characteristics: On PDA, low convex with entire edge, pale grey in the centre and olivaceous at the edge, reverse dark brown in the centre and black at the edge, reaching 28–30 mm diam after 10 d at 25 °C. On MEA, umbonate with entire edge, front and reverse brown in the centre and buff at the edge, reaching 22–25 mm diam after 10 d at 25 °C.

Typus: **China**, Jiangxi Province, Ganzhou, Yangling National Forest Park, on healthy leaves of *Camellia sinensis*, 24 Apr. 2013, *F. Liu*, YLBE3 (**holotype** HMAS 248081, culture ex-type CGMCC 3.19528 = LC3163 = LF372).

Additional materials examined: **China**, Jiangxi Province, Ganzhou, Yangling National Forest Park, on healthy leaves of *Camellia sinensis*, 24 Apr. 2013, *F. Liu*, YLBE3, living cultures LC3164 = LF373, LC3265 = LF374, LC3297 = LF519; on healthy leaves of *Camellia sinensis*, 24 Apr. 2013, *F. Liu*, YLBE1, living cultures LC3216 = LF428, LC13538 = LF2066.

Notes: *Setophoma endophytica* was isolated from healthy leaves of terraced tea plants from an open national forestry park in Jiangxi Province. Whether it is actually pathogenic to tea plant needs further research. Although strains of *S. endophytica* form

two subclades in the multi-locus phylogeny (Fig. 2), they were recognised as one species because of their same geographical origin, similar morphological characters, and high sequence similarity (100 % in ITS, 99 % in *gapdh*, 98 % in *tef-1α* and *tub2*). For the differences between *S. endophytica* and its closely related species, see the notes under *S. antiqua*.

Setophoma longinqua *F. Liu & L. Cai, sp. nov.* MycoBank MB829904. Fig. 5.

Etymology: From the Latin *longinqua* = remote, refers to the remote mountain where the species was collected.

Sexual morph: Unknown. **Asexual morph:** *Aerial mycelia* hyaline or grey-brown, smooth, branched. *Conidiomata* pycnidial, subglobose to ovoid, with an opening at apex, 100–200 µm diam. *Pycnidial wall* pale brown, with 2–3 layers, walls of *textura angularis*. *Setae* erect, solitary, unbranched, aseptate or septate, smooth, medium-brown to brown, subcylindrical, apex rounded. *Conidiophores* reduced to conidiogenous cells lining the inner cavity. *Conidiogenous cells* phialidic, unbranched, aseptate, hyaline, ampulliform, smooth, $3.5\text{--}4.5 \times 3\text{--}4$ µm (av. = $3.9 \pm 0.2 \times 3.6 \pm 0.2$ µm). *Conidia* aseptate, smooth, hyaline, with small guttules, cylindrical or subcylindrical with round or obtuse ends, sometimes allantoid, $4\text{--}5.5 \times 1.5\text{--}2$ µm (av. = $4.9 \pm 0.3 \times 1.8 \pm 0.2$ µm).

Culture characteristics: On PDA, flat with undulate edge, luteous coloured in the centre and brown at the edge, reverse sepia, reaching 27–28 mm diam after 10 d at 25 °C. On MEA, flat with roughly entire edge, surface and reverse isabelline, reaching 21–22 mm diam after 10 d at 25 °C.

Typus: **China**, Yunnan Province, Xishuangbanna, Mengla County, on *Camellia sinensis*, 18 Apr. 2015, *F. Liu*, GFZCWS001P (**holotype** HMAS 248082, culture ex-type CGMCC 3.19524 = LC6593 = LF1236).

Additional materials examined: **China**, Yunnan Province, Xishuangbanna, Mengla County, on *Camellia sinensis*, 18 Apr. 2015, *F. Liu*, GFZCWS001P, living cultures LC13481 = LF2068, LC13482 = LF2069.

Notes: *Setophoma longinqua* was isolated from the same symptomatic leaf as *S. antiqua*, but they are phylogenetically distinct (Figs 1, 2). It is morphologically distinct from *S. antiqua* by producing seta and longer conidia (av. = $4.9 \times 1.8 \mu\text{m}$ vs. $3.7 \times 1.8 \mu\text{m}$). Based on the multi-locus phylogeny (Fig. 2), the most closely related species to *S. longinqua* is an undescribed carbonatite associated species, but only with low sequence similarities of 95 % on ITS, 92 % on *tef-1 α* , and 90 % on *tub2*.

Setophoma yingyisheniae *F. Liu & L. Cai, sp. nov.* MycoBank MB829903. Fig. 6.

Etymology: Named after the collector, Yingyi Shen, who was also the first to report *Setophoma* leaf spot disease on tea plants.

Sexual morph: Unknown. Sterile on cultural media. Lesions on plant leaves subglobose or irregular, initially off-white to brownish yellow, becoming dark grey in the centre, separated from the healthy tissue by a black margin. *Conidiomata* pycnidial, dark brown or black, globose or subglobose, with an opening at apex, 60–200 μm diam. *Pycnidial wall* brown, walls of *textura angularis*. *Setae* 8.5–10 μm wide, erect, solitary, unbranched, septate, dark brown, apex rounded. *Conidiophores* hyaline, often with two branches. *Conidiogenous cells* hyaline, oblong, $3\text{--}5.5 \times 1.5\text{--}2.5 \mu\text{m}$ (av. = $4.2 \pm 0.6 \times 2.1 \pm 0.2 \mu\text{m}$). *Conidia* hyaline, aseptate, smooth, subglobose, reniform, or cylindrical, $3.5\text{--}6 \times 1.5\text{--}2.5 \mu\text{m}$ (av. = $4.4 \pm 0.5 \times 2.2 \pm 0.2 \mu\text{m}$).

Typus: **China**, Guangxi Province, Guilin, Longsheng County, altitude 1200 m, on *Camellia sinensis*, 21 Sep. 2016, *Y.Y. Shen*, GXGL01a (**holotype** HMAS 248086, culture ex-type CGMCC 3.19527 = LC13479 = LF1986).

Additional materials examined: **China**, Anhui Province, Bengbu County, Jianping Mountain, on *Camellia* sp., 2015, *F. Liu*, AHTEA07, living culture LC12696 = LF1529; on *Camellia* sp., 2015, *F. Liu*, AHTEA08, living culture LC12699 = LF1532; Fujian Province, Wuyishan City, Xingtianpu, 27.31'25"N, 118.1'59"E, 160 m, on *C. sinensis*, 22 Aug. 2016, *F. Liu*, WYSH3, living culture LC13477 = LF1935; on *C. sinensis*, 22 Aug. 2016, *F. Liu*, WYSH4, living culture LC13478 = LF1943; Guangxi Province, Guilin, Longsheng County, altitude 1200 m, on *Camellia sinensis*, 21 Sep. 2016, *Y.Y. Shen*, GXGL02, living cultures LC13480 = LF2017, LF2026; on *C. sinensis*, 21 Sep. 2016, *Y.Y. Shen*, GXGL04, living cultures LF2027, LF2032; Guilin, Tea Science and Research Institute of GuiLin, on healthy twig of *Camellia* sp., Sep. 2013, *T.W. Hou*, living culture LC3499 = LF727; Guizhou Province, on *Camellia* sp., 2015, *F. Liu*, GZSXD001P,

living culture LC6739 = LF1420; Jiangxi Province, Ganzhou City, Yangling National Forest Park, on *C. sinensis*, 24 Apr. 2013, *F. Liu*, YLA2, living culture LC3133 = LF341; on healthy leaves of *C. sinensis*, 24 Apr. 2013, *F. Liu*, YLB5, living culture LC3176 = LF386; Nanchang City, Meiling, on healthy leaves of *C. sinensis*, Apr. 2013, *F. Liu*, MLE002, living culture LC3197 = LF407.

Notes: *Setophoma yingyisheniae* was isolated from both symptomatic and asymptomatic leaves of terraced (shrubby) tea plants from five provinces, *i.e.* Anhui, Fujian, Guangxi, Guizhou and Jiangxi. It is phylogenetically related to *S. yunnanensis* (Fig. 2, 96 % sequence similarity on ITS, 88 % on *gapdh*, 87 % on *tef-1 α* and 93 % on *tub2*), but differs from the later in the host (terraced vs. arboreal tea plant) and habitat (plantation vs. unmanaged forest, Figs 6A, 7B). With respect to the conidiogenesis, the conidiophores of *S. yunnanensis* are simple and often reduced to single and ampulliform or globose conidiogenous cells, while these are branched in *S. yingyisheniae* and produces oblong conidiogenous cells. In addition, *S. yingyisheniae* differs from *S. yunnanensis* by producing seta in culture.

Setophoma yunnanensis *F. Liu & L. Cai, sp. nov.* MycoBank MB829905. Fig. 7.

Etymology: Named after the location where it was collected from, Yunnan Province.

Sexual morph: Unknown. **Asexual morph:** Sterile on cultural media. *Conidiomata* pycnidial, on diseased leaves black, globose or subglobose, with an opening at apex, 100–200 μm diam. *Pycnidial wall* brown, with 3–5 layers, 13–35 μm wide, walls of *textura angularis*. *Conidiophores* reduced to conidiogenous cells lining inner cavity. *Conidiogenous cells* hyaline, smooth, phialidic, ampulliform or globose, aseptate, $2.5\text{--}5 \times 2\text{--}4.5 \mu\text{m}$ (av. = $3.5 \pm 0.5 \times 3.1 \pm 0.8 \mu\text{m}$). *Conidia* hyaline, aseptate, granular to guttulate, ellipsoid or cylindrical, $3.5\text{--}5 \times 2\text{--}3 \mu\text{m}$ (av. = $4.3 \pm 0.4 \times 2.4 \pm 0.2 \mu\text{m}$).

Culture characteristics: On PDA, flat with entire edge, pale grey in the centre, buff at the edge, reverse blackish yellow in the centre and buff at the edge, reaching 31–34 mm diam after 10 d at 25 °C; On MEA, flat with entire edge, front and reverse blackish green, reaching 29 mm diam after 10 d at 25 °C.

Typus: **China**, Yunnan Province, Xishuangbanna, Mengla County, Laomansa, on *Camellia sinensis*, 19 Apr. 2015, *F. Liu*, LMS001P (**holotype** HMAS 248084, culture ex-type CGMCC 3.19529 = LC6753 = LF1434).

Additional material examined: **China**, Yunnan Province, Xishuangbanna, Mengla County, Daqishu, on *Camellia sinensis*, 19 Apr. 2015, *F. Liu*, DQS001P, living culture LC6532 = LF1167.

Notes: The leaf spots of arboreal tea plants where *S. yunnanensis* was isolated from were scattered, grey to brown in colour. *Setophoma yunnanensis* is phylogenetically related to *S. yingyisheniae* (Fig. 2, 96 % sequence similarity on ITS, 88 % on *gapdh*, 87 % on *tef-1 α* and 93 % on *tub2*), but differs both morphologically and geographically from the latter (see notes under *S. yingyisheniae*).

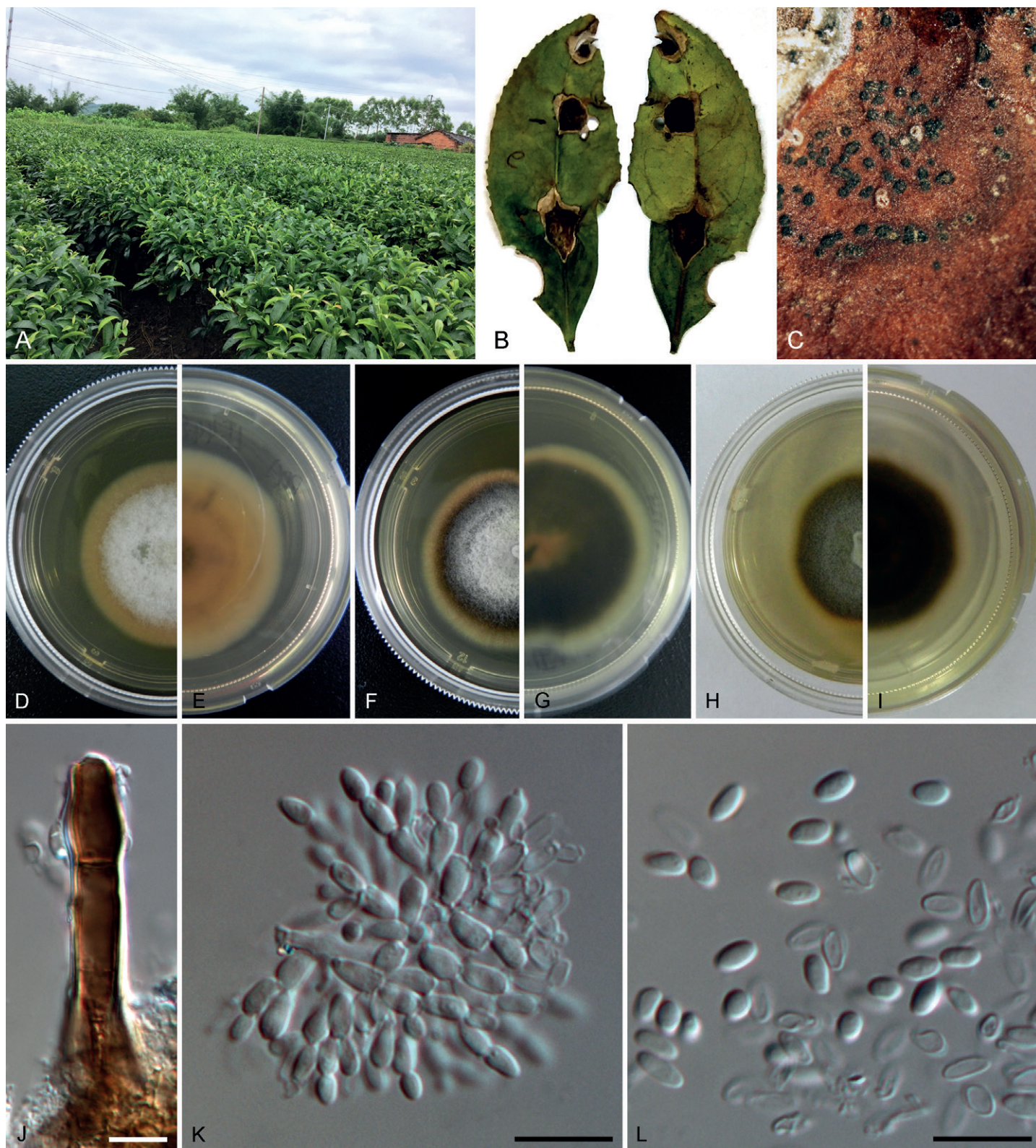


Fig. 6. *Setophoma yingyisheniae* (B, C, J–L, holotype GXGL01a, D, E, LC3236, F, G, LC3334, H, I, LC3133). A. Representative sampling environment—terraced tea plant. B. Front and back of diseased leaves. C. Conidiomata on leaf. D–I. Front and back of colonies on PDA. J. Seta. K. Conidiophores, conidiogenous cells and conidia. L. Conidia. Scale bars = 10 μ m.

DISCUSSION

Leaf spots on tea trees appeared to be caused by several previously undescribed *Setophoma* species. These were found on both old and young tea leaves collected from several Chinese provinces. Phylogenetic analyses provided a clear resolution for these novel *Setophoma* species (Figs 1, 2). Although the genus

Setophoma was only recently proposed and its relatives in *Pleosporales* have already been partly reassessed (e.g. Aveskamp *et al.* 2010, de Gruyter *et al.* 2010, Woudenberg *et al.* 2013), a large number of published phoma-like names still remained untreated. Therefore, in order to avoid proposing new *Setophoma* names for the old published taxa, we compared our species with all known *Pleosporales* species associated with *Camellia*

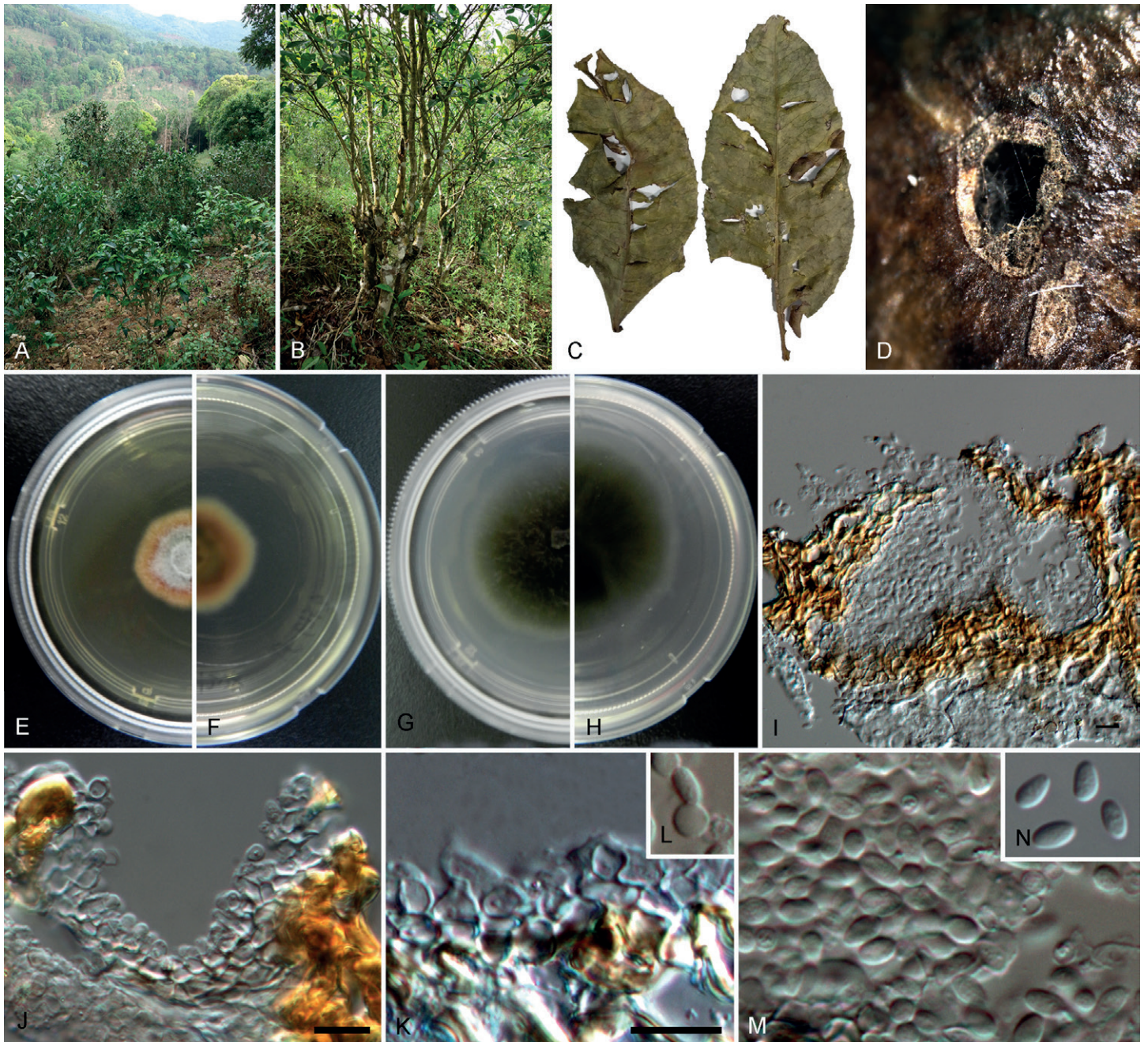


Fig. 7. *Setophoma yunnanensis* (ex-type CGMCC 3.19529 = LC6753). **A, B.** Sampling environment and arboreal tea plants. **C, D.** Symptom of diseased leaves. **E, F.** Front and back of colony on PDA. **G, H.** Front and back of colony on MEA. **I–K.** Vertical section of conidiomata. **L.** Conidiogenous cell and conidium. **M, N.** Conidia. Scale bars = 10 μ m.

from China, i.e. *Alternaria alternata*, *A. longipes*, *A. brassicae*, *Coniothyrium palmarum*, *Deuterophoma* sp., *Fusicladium theae*, *Hendersonia theae*, *Phoma camelliae*, *P. chinensis*, *P. herbarium* var. *herbarum*, *Piggotia* sp., *Remotididymella destructiva* and *Stagonosporopsis cucurbitacearum* (Farr & Rossman 2019). Most of these species are clearly distinctive from *Setophoma* by either morphological or molecular data, but this could unfortunately not be determined for *Phoma camelliae* and *P. chinensis*, as no reliable information regarding type specimen or publication information could be found for these species. These two names were thus ignored in this study and the taxonomic treatments of *P. camelliae* and *P. chinensis* await further study.

Of the four novel species isolated from the tea plant leaf spots, three species (*S. antiqua*, *S. longinqua* and *S. yunnanensis*) were only isolated from old and arboreal tea plants (~100–300-yr-

old, with less disease management and anthropogenic interference, Figs 3A, 7A, B) in Yunnan Province. In contrast, the fourth species, *S. yingyisheniae*, was widely distributed in all investigated provinces except Yunnan, and to our knowledge it only occurs on terraced tea plants (~50-yr-old), with intensive disease management and anthropogenic interference (Fig. 6A). *Setophoma yingyisheniae* has been isolated from both symptomatic and asymptomatic leaves tissues, indicating an alternative lifestyle ranging from endophytic to plant pathogenic. Morphologically, *S. yingyisheniae* differs from other *Setophoma* spp. in producing branched conidiophores, while other species in the genus produce solitary conidiophores. In contrast, *S. endophytica* was only isolated from healthy leaves, and whether it actually causes disease awaits to be determined.

A recently published species, *Setophoma cyperi* (Crous et al.

2016) should be excluded from the genus *Setophoma*, as it does not cluster in *Setophoma s. str.* (Fig. 1). *Setophoma cyperi* appears to be more closely related to *Sulcisporea pleurospora* (Fig. 1). The definitive generic placement of *S. cyperi* should be clarified in future work when a broader sampling of *Phaeosphaeriaceae* species becomes available.

ACKNOWLEDGEMENTS

We thank Yingyi Shen, Zhifeng Zhang, Yongzhao Diao, Peng Zhao, Mengmeng Wang, Dianming Hu, Xiaoming Tan, Qian Chen, Hanxing Zhang, Zuoru He, Haoshan Fu, Xiaoyan Dao and Tianwen Hou for their help in the collection of samples. Dr William Quaedvlieg is thanked for critically revising the English text. This study was financially supported by the Project for Fundamental Research on Science and Technology, Ministry of Science and Technology of China (2014FY120100) and the Frontier Science Research Project of the Chinese Academy of Sciences (QYZDB-SSW-SMC044).

REFERENCES

- Aveskamp MM, De Gruyter J, Woudenberg JHC, *et al.* (2010). Highlights of the *Didymellaceae*: a polyphasic approach to characterise *Phoma* and related pleosporalean genera. *Studies in Mycology* **65**: 1–60.
- Berbee ML, Pirseyedi M, Hubbard S (1999). *Cochliobolus* phylogenetics and the origin of known, highly virulent pathogens, inferred from ITS and glyceraldehyde-3-phosphate dehydrogenase gene sequences. *Mycologia* **91**: 964–977.
- Crous PW, Shivas RG, Quaedvlieg W, *et al.* (2014). Fungal Planet description sheets: 214–280. *Persoonia* **32**: 184–306.
- Crous PW, Wingfield MJ, Richardson DM, *et al.* (2016). Fungal Planet description sheets: 400–468. *Persoonia* **36**: 316–458.
- De Gruyter J, Woudenberg JHC, Aveskamp MM, *et al.* (2010). Systematic reappraisal of species in *Phoma* section *Paraphoma*, *Pyrenochaeta* and *Pleurophoma*. *Mycologia* **102**: 1066–1081.
- Farr DF, Rossman AY (2019). Fungal Databases, U.S. National Fungus Collections, ARS, USDA. <https://nt.ars-grin.gov/fungaldbases/>.
- Glass NL, Donaldson GC (1995). Development of primer sets designed for use with the PCR to amplify conserved genes from filamentous ascomycetes. *Applied and Environmental Microbiology* **61**: 1323–1330.
- Ikeda K, Kuwabara K, Urushibara T, *et al.* (2012). Pink root rot of squash caused by *Setophoma terrestris* in Japan. *Plant Disease* **78**: 372–375.
- Johnston-Monje D, Loewen S, Lazarovits G (2017). Mycobiomes of tomato plants with vine decline. *Canadian Journal of Plant Pathology* **39**: 184–200.
- Liu F, Bonthond G, Groenewald JZ, *et al.* (2019). *Sporocadaceae*, a family of coelomycetous fungi with appendage-bearing conidia. *Studies in Mycology* **92**: 287–415.
- Liu F, Wang M, Damm U, *et al.* (2016). Species boundaries in plant pathogenic fungi: a *Colletotrichum* case study. *British Medical Council Evolutionary Biology* **16**: 81.
- Liu F, Weir BS, Damm U, *et al.* (2015). Unravelling *Colletotrichum* species associated with *Camellia*: employing ApMat and GS loci to resolve species in the *C. gloeosporioides* complex. *Persoonia* **35**: 63–86.
- O'Donnell K, Cigelnik E (1997). Two divergent intragenomic rDNA ITS2 types within a monophyletic lineage of the fungus *Fusarium* are nonorthologous. *Molecular Phylogenetics and Evolution* **7**: 103–116.
- O'Donnell K, Kistler HC, Cigelnik E, *et al.* (1998). Multiple evolutionary origins of the fungus causing Panama disease of banana: concordant evidence from nuclear and mitochondrial gene genealogies. *Proceedings of the National Academy of Sciences of the United States of America* **95**: 2044–2049.
- Phookamsak R, Liu JK, Manamgoda DS, *et al.* (2014). The sexual state of *Setophoma*. *Phytotaxa* **176**: 260–269.
- Quaedvlieg W, Verkley GJM, Shin HD, *et al.* (2013). Sizing up *Septoria*. *Studies in Mycology* **75**: 307–390.
- Rayner RW (1970). *A mycological colour chart*. CMI and British Mycological Society, Kew, Surrey, UK.
- Rehner SA, Samuels GJ (1994). Taxonomy and phylogeny of *Gliocladium* analysed from nuclear large subunit ribosomal DNA sequences. *Mycological Research* **98**: 625–634.
- Rivedal HM, Stone AG, Johnson KB (2018). First report of *Setophoma terrestris* causing pink root rot of winter squash (*Cucurbita maxima*) in Oregon. *Plant Disease* **102**: 2661.
- Thambugala KM, Wanasinghe DN, Phillips AJL, *et al.* (2017). Mycosphere notes 1–50: Grass (*Poaceae*) inhabiting *Dothideomycetes*. *Mycosphere* **8**: 697–796.
- Vilgalys R, Hester M (1990). Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *Journal of Bacteriology* **172**: 4238–4246.
- White TJ, Bruns T, Lee S, *et al.* (1990). Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: *PCR protocols: a guide to methods and applications* (Innes MA, Gelfand DH, Sninsky JJ, *et al.*, eds). Academic Press, USA: 315–322.
- Woudenberg JHC, Groenewald JZ, Binder M, *et al.* (2013). *Alternaria* redefined. *Studies in Mycology* **75**: 171–212.
- Yang Y, Zuzak K, Harding M, *et al.* (2017). First report of pink root rot caused by *Setophoma (Pyrenochaeta) terrestris* on canola. *Canadian Journal of Plant Pathology* **39**: 354–360.