



UNIVERSITI PUTRA MALAYSIA

***GENETIC VARIATION AND CLONAL PROPAGATION OF SUPERIOR
GENOTYPES OF SELECTED *Acacia* SPECIES***

SURES KUMAR MUNIANDI

FH 2016 38



**GENETIC VARIATION AND CLONAL PROPAGATION OF SUPERIOR
GENOTYPES OF SELECTED *Acacia* SPECIES**

By

SURES KUMAR MUNIANDI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

December 2015

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



© COPYRIGHT UPM

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**GENETIC VARIATION AND CLONAL PROPAGATION OF SUPERIOR
GENOTYPES OF SELECTED *Acacia* SPECIES**

By

SURES KUMAR MUNIANDI

December 2015

Chairman : Professor Nor Aini Ab Shukor, PhD
Faculty : Forestry

The *Acacia* species was introduced into Malaysia in the late 1960's for timber production. Plantations of *Acacia* species gain interest as a major timber species in the 1960's especially in Peninsular Malaysia along with *Hevea brasiliensis*. The product in forestry ventures is often sawn timber, which requires trees to be in good conditions such as straight, single-stemmed and defect-free trunks for maximum utilization. Production of such quality wood is another big problem in forestry plantation since trees grown in plantation of some high-value temperate and tropical hardwood species tend to produce low value, short butt logs and bolts due to crooked stems, low fork heights and delayed shedding of lower branches.

In this context, a research study was initiated to select and recommend the best performing tree species or provenances suited for timber production in Malaysia with respect to growth and other characteristics. Species/provenance/progeny test was conducted on four species of *Acacia* namely, *A.mangium*, *A.auriculiformis*, *A.crassicarpa* and *A.aulocarpa* with four provenances for each species from two main regions of Papua New Guinea and Queensland. The growth of the provenances was monitored in terms of some quantitative and qualitative characteristics to evaluate the genetic variation and growth performance of a base breeding population. The study showed that there were significant differences ($p < 0.05$) between species, provenance and progenies for their growth performance. There were also significant differences between provenances within regions and progenies within provenances in all quantitative and qualitative traits tested in this study.

Generally, with regard to the growth, *A.mangium* performs better compared to other *Acacia* species in all of the aspects tested and it was followed by *A.crassicarpa*, *A.aulocarpa* and *A.auriculiformis*. Generally provenances and progenies selected from Papua New Guinea excelled those from Queensland both in quantitative and

qualitative characteristics. Among the top performing progenies of *Acacia* species are CG 1854 of (Bensbach WP) and KN000107 (SW of Boset WP) of *A.mangium*, BVG2609 (Bensbach WP) of *A.crassicarpa*, BVG 00835 (WP Morehead) and MM1016 (Arufi E Morehead WP) of *A.aulococarpa* and JSL363 (Wenlock River) and BVG 2657 (Bansbach) of *A.auriculiformis*.

Three best performing clone (genotypes) were then chosen based on their phenotypic characteristic for clonal propagation of superior tree species through traditional and modern techniques. Vegetative propagation was attempted as initial pretreatment stage of rejuvenation of mature sources through forced flushing, stem cuttings and trunk decapitation. *A. mangium* and *A. auriculiformis* respond well to force flushing by having highest survival percentage (87.7% and 90%, respectively) together with bud breaking and sprout growth. Whereas, *A.aulococarpa* and *A.crassicarpa* only recorded 52.2% and 31.1% of survival percentage. Rooting ability of stem cuttings, feasible and mean root number and root length increase at juvenile stage for all species studied. Rooting ability of mature cuttings decreased and bud breaks occurred only for few days eventually died, and did not respond to the treatments of growth regulators. Rooting ability of young stem cuttings of *A. mangium* (83.3%) and *A.auriculiformis* (76.6%) was better compared to that of *A.crassicarpa* and *A aulococarpa* with only 48% and 68.8%, respectively. Investigation was also done for the use of coppice materials as an alternative source for *in vitro* propagation of mature sources. 12 year-old trees of selected *Acacia* species were felled to the height of 1.0m and 1.5m. Vigorous production of sprout or coppice was noted on the stumps of trees of all species except of *A.crassicarpa*. The greatest coppicing ability in terms of survival rate of stumps was observed on *A.auriculiformis* with 83.8% followed by, *A.mangium*, *A.aulococarpa* and *A.crassicarpa* with 75.0%, 40.0% and 1.67 %, respectively. *A.crassicarpa* produces a very low number of sprouts with mean of 0.03 and mean of 0.09 for sprout length.

Rejuvenated mature explants were further subjected to *in vitro* conditions for mass production of improved materials for establishment of efficient *in vitro* protocol for *Acacia* sp. Decontamination of field collected materials was conducted as an initial stage in shoot initiation stage using some methods optimized in preliminary study. Most effective sterilization in term of average clean culture percentage (>70%) was recorded in 0.1% HgCl₂ for 5 minutes for *A.mangium*, *A.auriculiformis*, *A.mangium* 'Superbulk' and *A. hybrid* and 0.1% HgCl₂ for 10 minutes for both *A.crassicarpa* and *A.aulococarpa*. Incorporation of 0.1g/l of fungicide Benomyl with 50mg/ml of antibiotic streptomycin further enhanced the survival rate and percentage of clean culture up to 80%-100%. Multiple shoot production was obtained from all species of *Acacia* on Murashige and Skoog (MS) medium supplemented with 2.0 mg/l benzyladenine (BA) plus 0.5 mg/l of NAA. It was also noted that greater shoot production occurred with combination of plant growth regulators with additives. The maximum shoot number and shoot length was produced in medium supplemented with 2.0 mg l⁻¹ benzyladenine (BA) + 0.5 mg l⁻¹ of NAA + activated charcoal (0.1% w/v) combined with 100 mg l⁻¹ AdSO₄. It produced maximum number of 9.0 shoots per explant with 3.51 cm in length. Shoots were then elongated and rooted in an optimized condition and further acclimatized to nursery condition.

Another study was initiated to evaluate and identify sequence markers which gave phylogenetic information to be used to infer relationship within *Acacias* at a fine level. Primer designed based on second intron of *LEAFY* gene of *A.mangium* amplified the specific region with single band except for *A. hybrid*. The amplified regions were sequenced to reveal the species relationship within selected *Acacias*. Result revealed that non coding region of the second intron of *LEAFY* gene is more variable and can be used as marker for phylogenetic studies at lower taxonomic levels.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

VARIASI GENETIK DAN PEMBIAKAN KLON GENOTIP TERBAIK SPESIS AKASIA TERPILIH

Oleh

SURES KUMAR MUNIANDI

Disember 2015

Pengerusi : Profesor Nor Aini Ab Shukor, PhD
Fakulti : Perhutanan

Spesies *Akasia* telah diperkenalkan di Malaysia pada lewat tahun 1960-an untuk pengeluaran kayu. *Akasia* dipilih sebagai spesies kayu utama terutama di Semenanjung Malaysia bersama dengan *Hevea brasiliensis*. Produk dalam penghasilan kayu gergaji sering memerlukan pokok dalam keadaan yang baik seperti batang induk yang lurus, serta bebas kecacatan batang untuk penggunaan yang maksima. Pengeluaran kayu berkualiti adalah satu masalah besar di dalam perladangan hutan yang ditanam dengan spesies kayu keras sederhana dan tropika cenderung untuk menghasilkan batang yang bernilai rendah, ruas pendek dan bolt kerana batang yang bengkok, ketinggian cabang rendah dan kelewatan dalam penanggalan cabang rendah.

Kajian ini telah dimulakan untuk memilih dan mencadangkan spesies pokok yang terbaik dengan asal usul yang berkaitan dengan pertumbuhan dan ciri-ciri lain yang sesuai untuk pengeluaran kayu di Malaysia. Ujian spesies / provenan / progeneri telah dijalankan ke atas empat spesies *Akasia* iaitu, *A.mangium*, *A.auriculiformis*, *A.crassicarpa* dan *A. aulococarpa* dengan empat provenan bagi setiap spesies daripada dua kawasan utama Papua New Guinea dan Queensland. Pertumbuhan provenan telah dipantau dari segi beberapa ciri-ciri kuantitatif dan kualitatif untuk menilai perubahan dan pertumbuhan prestasi populasi genetic. Kajian ini menunjukkan bahawa terdapat perbezaan yang ketara ($p < 0.05$) antara spesies, provenan dan progeneri prestasi pertumbuhan mereka. Terdapat juga perbezaan yang ketara antara provenan di dalam kawasan dan progeneri dalam provenan untuk semua sifat-sifat kuantitatif dan kualitatif yang diuji

Secara umumnya, dengan mengambil kira pertumbuhan, *A.mangium* membesar lebih baik berbanding dengan spesies *Akasia* yang lain dalam semua aspek yang diuji dan ia diikuti oleh *A.crassicarpa*, *A.aulococarpa* dan *A.auriculiformis*. Provenan dan progeneri yang dipilih dari kawasan Papua New Guinea adalah lebih baik dari

Queensland dari segi pertumbuhan dalam semua ciri-ciri kuantitatif dan kualitatif. Di antara progeni terbaik spesies Akasia adalah CG 1854 daripada (Bensbach WP) dan KN000107 (SW dari Boset WP) daripada *A.mangium*, BVG2609 (Bensbach WP) daripada *A.crassicarpa*, BVG 00835 (WP Morehead) dan MM1016 (Arufi E Morehead WP) daripada *A.aulococarpa* dan JSL363 (Wenlock River) dan BVG 2657 (Bansbach) daripada *A.auriculiformis*.

Tiga klon berprestasi terbaik (genotip) kemudian dipilih berdasarkan ciri fenotip mereka untuk tujuan pembiakan klon spesies pokok yang unggul melalui teknik pembiakan tradisional dan moden. Pembiakan tampang telah dilakukan sebagai peringkat rawatan awal untuk menjuvinasikan sumber matang melalui 'force flushing', keratan batang dan penebangan batang. *A. mangium* dan *A. auriculiformis* bertindak balas dengan baik untuk 'force flushing' dengan mempunyai peratus kemandirian yang tertinggi (87.7% dan 90%) dengan pertumbuhan putik dan pucuk. Manakala, *A.aulococarpa* dan *A.crassicarpa* hanya merekodkan 52.2% dan 31.1% peratus kemandirian. Keupayaan pengakaran keratan batang boleh dilaksanakan dan purata bilangan akar dan panjang akar dapat ditingkatkan dengan penggunaan sumber yang dijuvenasikan untuk semua spesies Akasia. Keupayaan pengakaran menurun bagi sumber matang dan pertumbuhan putik berlaku hanya untuk beberapa hari dan mati. Ia juga tidak bertindak balas kepada semua rawatan pengalok pertumbuhan. Keupayaan pengakaran keratan batang muda *A. mangium* (83.3%) dan *A.auriculiformis* (76.6%) adalah lebih baik berbanding dengan *A.crassicarpa* dan *A aulococarpa* dengan hanya 48% dan 68.8% masing-masing. Kajian juga telah dijalankan untuk penggunaan sumber dari pucuk muda sebagai sumber alternatif untuk pembiakan *in vitro* sumber matang. Spesies pokok Akasia yang berumur 12 tahun telah dipilih dan ditebang dengan ketinggian 1.0 m dan 1.5 m. Pengeluaran pucuk yang baik diperhatikan pada dari segi kadar kemandirian pada tunggul pokok daripada semua spesies kecuali *A.crassicarpa*. Peratus keupayaan percambahan tunggul dari segi kadar kemandirian yang tinggi diperhatikan pada *A.auriculiformis* dengan 83.8% diikuti oleh, *A.mangium*, *A.aulococarpa* dan *A.crassicarpa* dengan masing-masing mempunyai 75.0%, 40.0% dan 1.67%. *A.crassicarpa* menghasilkan jumlah pucuk yang sangat rendah dengan purata 0.03 dan purata 0.09 untuk kepanjangan pucuk.

Sumber yang dijuvenasikan seterusnya didedahkan kepada keadaan *in vitro* untuk pengeluaran besar-besaran bahan yang terpilih bagi pembentukan protokol *in vitro* yang berkesan untuk spesies Akasia. Pembersihan sumber yang dikumpulkan dari lapangan telah dijalankan sebagai peringkat awal permulaan percambahan pucuk dengan menggunakan beberapa kaedah yang dioptimumkan dalam kajian awal. Pensterilan yang paling berkesan dari segi peratus purata kultur bersih (> 70%) dicatatkan pada 0.1% HgCl₂ selama 5 minit untuk *A.mangium*, *A.auriculiformis*, *A.mangium* 'Superbulk' dan *A. hybrid* dan 0.1% HgCl₂ selama 10 minit untuk *A.crassicarpa* dan *A.aulococarpa*. Penambahan 0.1g / l racun kulat Benomyl dengan 50mg / ml antibiotik streptomycin mempertingkatkan kadar survival dan peratusan purata kultur bersih sehingga 80% -100%. Pengeluaran pucuk yang banyak telah diperolehi daripada semua spesies Akasia pada media Murashige dan Skoog (MS) yang diperkaya dengan 2.0 mg / l benzyladenine (BA) serta 0.5 mg / l. NAA. Didapati pengeluaran pucuk yang terbaik berlaku dengan gabungan pengawal pertumbuhan tumbuhan dengan bahan tambahan. Bilangan pucuk dan panjang pucuk yang

maksimum dihasilkan dalam media ditambah dengan 2.0 mg/l benzyladenine (BA) + 0.5 mg/l-1 NAA + arang aktif (0.1% w / v) digabungkan dengan 100 mg/l AdSO₄. Ia menghasilkan 9.0 bilangan pucuk maksimum per eksplan dengan kepanjangan 3.51 cm. Pucuk telah dipanjangkan dan diakar dalam keadaan optimum dan selanjutnya didedahkan di luar untuk menyesuaikan diri kepada keadaan nurseri.

Satu lagi kajian telah dilakukan untuk menilai dan mengenalpasti penanda urutan maklumat filogenetik yang akan digunakan untuk membuat kesimpulan mengenai hubungan antara Akasia pada tahap yang baik. Primer yang telah direka berdasarkan intron kedua gen *LEAFYA.mangium* telah mengamplifikasi kawasan yang dikehendaki dengan band tunggal kecuali *A. hybrid*. Rantau yang diamplifikasi telah disusun untuk mendedahkan hubungan dalam spesies Akasia yang terpilih. Keputusan ini mendedahkan kawasan bukan pengekod intron kedua gen *LEAFY* ini adalah lebih bervariasi dan boleh digunakan sebagai penanda untuk kajian filogenetik taxanomi pada tahap yang lebih rendah.

ACKNOWLEDGEMENTS

My sincere appreciation and gratitude goes to my supervisor, Prof Dr. Nor Aini Ab Shukor for giving me the opportunity to study under her guidance throughout my PhD programme. My great appreciation also extended to my supervisory committee Prof Dr. Paridah Md Tahir, Assoc. Prof Dr. Parameswari Namasivayam, Assoc. Prof Dr. Chin Chiew Foan and Dr Norwati Muhammad for their guidance, advice, comments and valuable criticism that leads to the completion of this thesis.

I would also like to convey my appreciation to Dr. Sri Rahayu, Prof. Dr Aktar Hussain and Prof Rafiee for their assistance on statistical analysis, all staff's in Faculty of Forestry, Institute of Tropical Forestry and Forest Products and Faculty of Biomolecular Sciences, Mr. Baharum Zainal, Ms Ana Salleza, Ms Kamariah Damanhuri, Ms Nadia Abdullah and Ms Halimah Hussein for their technical support and guidance and all the other members of Universiti Putra Malaysia who have helped me in one way or another. My appreciation also goes to my friends, Dr. Samantha Priyanka, Dr. Yaghoob Tahery, Dr. Zahra Noori, Ms Elnaz Mehdizadeh, Mr Shahidin Ahmad Juffiry, Mr Kazeem Kolapo, Ms Dayana Aisyah, Ms Intan Shafinaz, Ms Azima and Ms Shariffah Sharliza Binti Syed Aualadali for their continuous support and encouragement. My special thanks goes to my dear friend Ms Wafiah Wazer who helped me a lot during field data collection and laboratory works and also encouraged me to complete my studies even though I delt and experienced a lot of difficulties and problems. The completion of this thesis is not possible without her support and guidance.

Lastly, my heartfelt thanks to my beloved family members, my late father Mr. Muniandi a/l Muthaiah, my mother Ms Somawathie d/o John De silva, sister Ms Uma Devi Muniandi and brother Mr. Kumaran Muniandi, for their patience, continuous love, support and their sacrifices in bearing all the shortcomings throughout my study. Encouragement and moral support from my uncle Mr. Ramesh and aunt Ms Uma throughout my study are highly appreciated. Thank you.

I certify that a Thesis Examination Committee has met on 29 December 2015 to conduct the final examination of Sures Kumar on his thesis entitled "Genetic Variation and Clonal Propagation of Superior Genotypes of Selected *Acacia* Species" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Ahmad Ainuddin bin Nuruddin, PhD

Professor
Faculty of Forestry
Universiti Putra Malaysia
(Chairman)

Mohamad Azani bin Alias, PhD

Associate Professor
Faculty of Forestry
Universiti Putra Malaysia
(Internal Examiner)

Mohd Zaki bin Hamzah, PhD

Associate Professor
Faculty of Forestry
Universiti Putra Malaysia
(Internal Examiner)

Jae-Seon Yi, PhD

Professor
Kangwon National University
Republic of Korea
(External Examiner)



ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 24 March 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Nor Aini Ab Shukor, PhD

Professor
Faculty of Forestry
Universiti Putra Malaysia
(Chairman)

Paridah Md Tahir, PhD

Professor
Faculty of Forestry
Universiti Putra Malaysia
(Member)

Parameswari a/p Namasivayam, PhD

Associate Professor
Faculty of Forestry
Universiti Putra Malaysia
(Member)

Chin Chiew Foan, PhD

Associate Professor
Faculty of Science
The University of Nottingham Malaysia Campus
(Member)

Norwati Muhammad, PhD

Forest Research Institute of Malaysia (FRIM)
Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Sures Kumar Muniandi, GS 20626

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of Chairman
of Supervisory
Committee: _____

Signature: _____
Name of Member
of Supervisory
Committee: _____

Signature: _____
Name of Member
of Supervisory
Committee: _____

Signature: _____
Name of Member
of Supervisory
Committee: _____

Signature: _____
Name of Member
of Supervisory
Committee: _____

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iv
ACKNOWLEDGEMENTS	vii
APPROVAL	viii
DECLARATION	x
LIST OF TABLES	xvi
LIST OF FIGURES	xxi
LIST OF ABBREVIATIONS	xxvi
 CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	
2.1 <i>Acacia</i> species	8
2.1.1 <i>Acacia mangium</i> , <i>Acacia auriculiformis</i> , <i>Acacia crassicarpa</i> and <i>Acacia aulococarpa</i>	10
2.1.2 Uses of <i>Acacia</i> species	11
2.1.3 Heart rot disease in <i>A.mangium</i> plantation	12
2.2 Seed sterilization and seedling germination in <i>Acacia</i> species	12
2.3 <i>In vitro</i> propagation in <i>Acacia</i> species	13
2.3.1 Types of explants	19
2.3.2 Types of culture media	20
2.3.3 Growth regulators	20
2.4 Somatic embryogenesis	31
2.4.1 Somatic embryogenesis in <i>Acacia</i> species	32
2.5 Browning of tissue in the culture	32
2.6 Vitrification in tissue culture plantlets	34
3 MATERIALS AND METHODS/METHODOLOGY	
3.1 Description of study scope	38
3.2 Seed germination	38
3.3 Preparation of stock solutions of MS medium	39
3.3.1 Preparation of MS medium (Murashige and Skoog medium, 1962)	39
3.3.2 Preparation of stock solution of growth regulators	40
3.4 Preparation of aseptic conditions	40
3.5 Description of species and explants used in the study	41
3.5.1 Collection techniques of field explants	42
3.6 Standard sterilization protocol	42
3.7 Growth conditions	42
3.8 Observation for evaluation	43

4	PLUS TREE SELECTION OF MATURE EXPLANTS OF SOME SELECTED ACACIA GENOTYPES	
4.1	Introduction	45
4.2	Plus tree selection	45
	4.2.1 Species, provenance and progeny trials	46
	4.2.2 Timber traits for sawn timber production	50
4.3	Materials and methods	55
	4.3.1 Description of study sites and plant materials	55
	4.3.2 Quantitative and qualitative traits	61
4.4	Results and discussion	63
5	REJUVENATION OF SELECTED MATURED ACACIA GENOTYPES FOR MASS PROPAGATION OF ELITE CLONES	
5.1	Introduction	88
5.2	Maturation and rejuvenation in plant species	89
5.3	Rejuvenation techniques in forestry	90
	5.3.1 Shoot sprouts of stumping/epicormic/coppicing	90
	5.3.2 Lateral bud induction and forced flushing from cuttings of mature trees	91
	5.3.3 Chemical treatment of mature branches	92
5.4	Materials and methods	95
	5.4.1 Plant materials	95
	5.4.2 Shoot sprouts of stumping/epicormic/coppicing	95
	5.4.3 Lateral bud induction from cuttings and forced flushing of mature trees	95
	5.4.4 Stem cuttings	96
	5.4.5 Air layering propagation	96
5.5	Results and discussion	97
6	SURFACE STERILIZATION OF MATURE PLANT MATERIALS FROM SELECTED PLUS TREES OF ACACIA SPECIES	
6.1	Introduction	115
6.2	Methods of surface sterilization	117
6.3	Sterilizing agents	117
6.4	Prevention of contamination and pretreatment of the explants	118
6.5	Common sterilization procedures in Acacia species	120
6.6	Fungicide and antibiotic incorporation in basal culture medium	121
	6.6.1 Antibiotics treatments in tissue culture	122
	6.6.2 Fungicide treatments in tissue culture	123
	6.6.3 Combination of fungicide and antibiotic in culture medium	124
6.7	Material and methods	125
	6.7.1 Surface sterilization of explants	125
	6.7.2 Standard sterilization and experimental design	125
	6.7.3 Effects of initial pretreatment on surface sterilization	126

	6.7.4	Effects of antibiotic and fungicide in the medium	127
	6.7.5	Culture condition	128
	6.7.6	Evaluation	128
	6.7.7	Isolation and identification of contaminants in contaminated medium	128
	6.7.7.1	Preparation of PDA for isolation of fungi and bacteria	128
	6.7.7.2	Isolation and identification of fungi	129
	6.7.7.3	Isolation and identification of bacteria	129
	6.7.7.4	Preparation of bacterial slides for identification	130
	6.8	Results and discussion	130
7		OPTIMIZATION OF REGENERATION PROTOCOL FOR SOME SELECTED MATURED ACACIA GENOTYPES	
	7.1	Introduction	157
	7.2	Adenine sulphate and adenine based cytokinin on multiple shoot induction	158
	7.3	Effects of silver nitrate (AgNO ₃) on multiple shoot induction	161
	7.4	Glutamine on multiple shoot induction	163
	7.5	Materials and methods	166
	7.5.1	Surface sterilization of explants	166
	7.5.2	Effects of cytokinin (BAP) and auxin (2,4-D) in shoot induction media (SIM)	166
	7.5.3	Influence of different additives on shoot multiplication (Silver nitrate (AgNO ₃), Adenine sulfate (AdSO ₄) and glutamine)	167
	7.5.4	Effects of the stumped <i>in vitro</i> explants on shooting ability	168
	7.5.5	Elongation, rooting and acclimatization of shoots of <i>in vitro</i> plantlets	168
	7.5.6	Rooting of shoots	169
	7.5.7	Acclimatization	169
	7.5.8	Data analysis	169
	7.6	Results and discussion	169
8		GENETIC VARIATION OF ACACIA SPECIES GENOTYPES REVEALED BY SECOND INTRON OF <i>Acacia mangium</i> LEAFY GENE	
	8.1	Genetic diversity and genetic markers	204
	8.2	Non-coding introns	205
	8.3	LEAFY (FLO/LFY) gene	206
	8.4	Materials and Methods	207
	8.4.1	Plant materials	207
	8.4.2	Sample collection techniques of field explants	207
	8.4.3	Extraction of genomic DNA	210
	8.4.3.1	DNA quantification and analysis of quality of DNA	210
	8.4.4	Design non-coding introns of the oligonucleotide	211

	primers	
	8.4.4.1 Optimization of primer by gradient PCR	211
8.4.5	PCR amplification	211
	8.4.5.1 Gel Electrophoresis	212
	8.4.5.2 Purification of PCR product	212
8.4.6	Sequencing	213
	8.4.6.1 Preparation of competent cells – <i>E. coli</i> DH5 α	213
	8.4.6.2 Ligation of purified PCR product into yT & A cloning vector	213
	8.4.6.3 Transformation of the ligated yT & A vectors into <i>E. coli</i> DH5 α	214
	8.4.6.4 Colony PCR	215
	8.4.6.5 Plasmid DNA extraction	215
	8.4.6.6 Verification by sequencing	215
	8.4.6.7 Phylogenetic analysis	216
8.5	Results and discussion	216
9	SUMMARY, CONCLUSION AND RECOMMENDATIONS	226
	REFERENCES	228
	APPENDICES	279
	BIODATA OF STUDENT	297
	LIST OF PUBLICATIONS	298

LIST OF TABLES

Table		Page
2.1	Number of accepted and described species of <i>Acacia sens.lat.</i> worldwide adapted from Maslin <i>et al.</i> 2003 and Orchard and Maslin, 2003	8
2.2	Summary of micropropagation work on <i>Acacia</i> species in relation to use of different species, explant types, medium and plant growth regulators	15
2.3	Types and age of explants in micropropagation of mature/ elite <i>Acacia</i> species	20
2.4	Summary of information relating previous work on <i>Acacia</i> species on types of PGRs and their concentration for micropropagation, organogenesis and somatic embryogenesis of <i>Acacia</i> species	25
2.5	Examples of somatic embryogenesis production in <i>Acacia</i> species	32
3.1	Tabulation of plant growth regulators, their molecular weight, and their initial solvents	40
4.1	Published provenance-progeny trials for some important quantitative and qualitative growth traits for some economically important tree species around the world	51
4.2	Detail of the 16 provenances of <i>Acacia</i> used in this study	57
4.3	Random distribution of 16 provenances and 80 progenies of four <i>Acacia</i> species in block 1 and 4	59
4.4	Analysis of variance for quantitative growth traits of four 12-year-old <i>Acacia</i> species	64
4.5	Analysis of variance for qualitative growth traits of four 12-year-old <i>Acacia</i> species	65
4.6	Mean values for quantitative growth traits in terms of geographic regions of four 12-year-old <i>Acacia</i> species	66
4.7	Mean values for qualitative traits with regards to geographic regions for four 12-year-old <i>Acacia</i> species	67
4.8	Mean values of quantitative growth traits of four 12-year-old <i>Acacia</i> species	69
4.9	Mean values for qualitative growth traits of four 12-year-old <i>Acacia</i> species	69

4.10	Species ranking based on their quantitative and qualitative growth traits performance	70
4.11	Mean values of quantitative growth traits for provenance within geographic regions of four 12-year-old <i>Acacia</i> species	72
4.12	Mean values of qualitative growth traits for provenance within geographic regions of four 12-year-old <i>Acacia</i> species	73
4.13	Provenance ranking based on their quantitative and qualitative growth traits performance for four 12-years-old <i>Acacia</i> species	74
4.14	Mean values of quantitative growth traits for progenies within provenance and geographic regions of 12-year-old <i>Acacia mangium</i>	75
4.15	Mean values of qualitative growth traits for progenies within provenance and geographic regions of 12-year-old <i>Acacia mangium</i>	76
4.16	Progeny ranking based on their quantitative and qualitative growth traits performance of 12-year-old <i>Acacia mangium</i>	77
4.17	Mean values of quantitative growth traits for progenies within provenance and geographic regions of 12-year-old <i>Acacia auriculiformis</i>	78
4.18	Mean values of qualitative growth traits for progenies within provenance and geographic regions of 12-year-old <i>Acacia auriculiformis</i>	79
4.19	Progeny ranking based on their quantitative and qualitative growth traits performance of 12-year-old <i>Acacia auriculiformis</i>	80
4.20	Mean values of quantitative growth traits for progenies within provenance and geographic regions of four 12-year-old <i>Acacia crassicarpa</i>	81
4.21	Mean values for qualitative growth traits for progenies within provenance and geographic regions of 12-year-old <i>Acacia crassicarpa</i>	82
4.22	Progeny ranking based on their quantitative and qualitative growth traits performance of 12-year-old <i>Acacia crassicarpa</i>	83
4.23	Mean values for quantitative growth traits for progenies within provenance and geographic regions of 12-year-old <i>Acacia aulocarpa</i>	84

4.24	Mean values for qualitative growth traits for progenies within provenance and geographic regions of 12-year-old <i>Acacia aulococarpa</i>	85
4.25	Progeny ranking based on their quantitative and qualitative growth traits performance of 12-year-old <i>Acacia aulococarpa</i>	86
4.26	Total number of candidate trees for <i>Acacia</i> species and selected clone for clonal propagation through macropropagation and micropropagation	87
5.1	Rejuvenation methods in <i>in vitro</i> propagation of mature tree species	93
5.2	Analysis of variance for shoot flushing ability of cuttings of four 12-year-old <i>Acacia</i> species clones	98
5.3	Mean values of flushing ability of cuttings of four 12-year-old <i>Acacia</i> species clones	98
5.4	Mean values for flushing ability of cuttings of four 12-year-old <i>Acacia</i> species clones	99
5.5	Analysis of variance for rooting ability of cuttings of 12-year-old <i>Acacia</i> species	101
5.6	Mean values for rooting ability of cuttings of four 12-year-old <i>Acacia</i> species	102
5.7	Mean values of some rooting ability of cuttings of four 12-year-old <i>Acacia</i> species clones	103
5.8	Mean values of some rooting ability of cutting sources of four 12-year-old <i>Acacia</i> species	104
5.9	Mean values of some rooting ability of treated cutting sources of four 12-year-old <i>Acacia</i> species	106
5.10	Analysis of variance of some coppicing ability of decapitated trunks in four 12-year-old <i>Acacia</i> species	108
5.11	Mean values of some coppicing ability of decapitated trunks in four 12-year-old <i>Acacia</i> species	108
5.12	Mean values of some coppicing ability of decapitated trunks in four 12-year-old <i>Acacia</i> species provenances	110
5.13	Mean values of some coppicing ability of four 12-year-old <i>Acacia</i> species at different height levels	111

6.1	The most common sterilizing agents, their concentration and the explant's exposure time used in micropropagation of woody plant material	118
6.2	Treatments for surface sterilization experiment	126
6.3	Treatments for initial pretreatment prior to surface sterilization experiment	127
6.4	Analysis of variance for surface sterilization of <i>Acacia</i> species explants	132
6.5	Means values of surface sterilization of six selected <i>Acacia</i> species	133
6.6	Analysis of variance for initial pretreatment of selected <i>Acacia</i> species	140
6.7	Mean values for initial pretreatment of six selected <i>Acacia</i> species	141
6.8	Mean values for clean cultures in initial pretreatment of six selected <i>Acacia</i> species clones	142
6.9	Mean values for clean culture in surface sterilization of explant sources of six selected <i>Acacia</i> species	143
6.10	Analysis of variance for incorporation of fungicide and antibiotic in surface sterilization of selected <i>Acacia</i> species	146
6.11	Mean values for incorporation of fungicide and antibiotic in surface sterilization of selected <i>Acacia</i> species	146
7.1	Effects of adenine sulphate (AdSO_4) on morphogenetic response of explants	159
7.2	Effects of silver nitrate (AgNO_3) on morphogenetic response of explants	162
7.3	Effects of glutamine (Gln) on morphogenetic response of explants	164
7.4	Modification and treatment in surface sterilization of young (rejuvenated) and mature field collected explants	166
7.5	Treatment combination in MS medium supplemented with additional 0.1% (w/v) activated charcoal	168
7.6	Analysis of variance on the effects of clone, types of explant, concentration of BA (treatment) and their interaction for shoot induction of six selected <i>Acacia</i> species after 6 weeks of culture incubation	172

7.7	The effects of hormone concentration (BA+ NAA) on mean shoot number and shoot length of six selected <i>Acacia</i> species after 6 weeks of culture incubation	173
7.8	Mean shoot number and shoot length in shoot induction of six selected <i>Acacia</i> species clones after 6 weeks of culture incubation	174
7.9	Mean values for type of explants in shoot induction of six selected <i>Acacia</i> species after 6 weeks of culture incubation	175
7.10	Analysis of variance on the effects of species, clone, concentration of additives (treatment) and their interaction on shoot induction of six selected <i>Acacia</i> species after 6 weeks of culture incubation	181
7.11	The effects of additives on shoot induction of six selected <i>Acacia</i> species after 6 weeks of culture incubation	182
7.12	Analysis of variance on the effects of species, different treatments and their interaction on shoot elongation for six selected <i>Acacia</i> species after 6 weeks of culture incubation	187
7.13	The effects of gibberellic acid concentration on shoot elongation of six selected <i>Acacia</i> species after 6 weeks of culture incubation	187
7.14	Analysis of variance on the effects of species, different treatments and their interaction on root induction for six selected <i>Acacia</i> species after 6 weeks of culture incubation	189
7.15	The effects of auxin concentrations on root induction of six selected <i>Acacia</i> species after 6 weeks of culture incubation	190
7.16	Analysis of variance on the effects of species, clone, different medium and their interaction on acclimatization for six selected <i>Acacia</i> species after 6 weeks of nursery conditions	195
7.17	The effects of acclimatization methods on survival rates of six selected <i>Acacia</i> species after 6 weeks of nursery conditions	195
8.1	Genotypes and their sources used in phylogenetic relationship of <i>Acacia</i> using LEAFY second intron sequences	208
8.2	Primers sequences for amplification of second intron of <i>A. mangium</i> LEAFY gene	211
8.3	PCR conditions used for the amplification of second intron of <i>A. mangium</i> LEAFY gene	212
8.4	Reaction for ligation of PCR product into yT & A cloning vector	214
8.5	The length of second intron of LEAFY gene in <i>Acacia</i> progenies	220

LIST OF FIGURES

Figure		Page
1.1	a) Production of plywood, sawn timber and logs, and b) Production of sawn timber from <i>Hevea</i> logs in Malaysia between 2003-2011	2
1.2	Forest plantation established in Permanent Reserved Forest in Malaysia from 2002-2011	3
1.3	Forest plantation establishment according to species distribution in Malaysia from 2002-2010	5
3.1	A) Types of explants of <i>Acacia</i> sp; and B) Schematic diagram of the plant shoot apex (adapted from Langdale, 1994)	41
3.2	Flow chart of research work	44
4.1	The developmental stage in tree improvement programme	48
4.2	Developmental stages in tree improvement programme for sustainable production of improved materials	49
4.3	Improvement of phenotypic variation among <i>Acacia</i> tree species for production of sawn timber with higher economic value	54
4.4	Main source of variation tested in the trial plot	55
4.5	Experimental design and field layout of <i>Acacia</i> species-provenance-progeny test	56
4.6	<i>Acacia</i> species-provenance-progeny trial location at How Swee Sdn. Bhd. Estate, Kampung Aur Gading, Kuala Lipis, Pahang, Malaysia	58
4.7	Qualitative traits assessment by scoring for shape of stem, shape of bole, branch axis and branching size (ascending assessment)	62
5.1	(A) Coppicing shoots from 12 year-old stump of <i>A.auriculiformis</i> after 1 month ($\leq 2-3$ cm); (B) Coppicing shoots from 12 year-old stump of <i>A.auriculiformis</i> after 3 months; (C) Coppicing shoots from 12 year-old stump of <i>A.mangium</i> after 3 months; and (D) Coppicing shoots from 12 year-old stump of <i>A.aulococarpa</i> after 3 months	113
5.2	(A) Explants for micropropagation from epicormic shoots of <i>A.mangium</i> ; (B) Explants for micropropagation from coppicing materials of <i>A. auriculiformis</i> ; (C) Explants for micropropagation from stem cuttings of <i>A.crassicarpa</i> ; and (D) Explants for micropropagation from forced flushing of <i>A.crassicarpa</i>	114

6.1	Clean cultures percentages in surface sterilization of six selected <i>Acacia</i> species clones	134
6.2	Mean values for clean culture in surface sterilization of explant sources of six selected <i>Acacia</i> species	135
6.3	Mean values for clean cultures in surface sterilization method for six selected <i>Acacia</i> species	137
6.4	Mean values for clean cultures in surface sterilization method within selected <i>Acacia</i> species	144
6.5	Mean values for explant source variation in incorporation of fungicide and antibiotic in surface sterilization of six selected <i>Acacia</i> species	147
6.6	Mean values for explant source variation in incorporation of fungicide and antibiotic in surface sterilization of selected <i>Acacia</i> species	148
6.7	Mean values for treatment variation in incorporation of fungicide and antibiotic in surface sterilization of <i>Acacia mangium</i>	149
6.8	Mean values for treatment variation in incorporation of fungicide and antibiotic in surface sterilization of <i>Acacia auriculiformis</i>	149
6.9	Mean values for treatment variation in incorporation of fungicide and antibiotic in surface sterilization of <i>Acacia crassicarpa</i>	150
6.1	Mean values for treatment variation in incorporation of fungicide and antibiotic in surface sterilization of <i>Acacia aulococarpa</i>	150
6.11	Mean values for treatment variation in incorporation of fungicide and antibiotic in surface sterilization of <i>Acacia mangium</i> 'Superbulk'	151
6.12	Mean values for treatment variation in incorporation of fungicide and antibiotic in surface sterilization of <i>Acacia</i> hybrid	151
6.13	Cumulative mean values for treatment variation in incorporation of fungicide and antibiotic in surface sterilization of <i>Acacia</i> species	152
6.14	Colonies of <i>Mucor</i> (A), <i>Penicillium</i> (B) and <i>Aspergillus</i> (C-F) species incubated for 7 days at 25 °C on PDA + 1% yeast extract	154
6.15	(A and B) <i>Mucor plumbeus</i> ; (C and D) <i>Penicillium chrysogenum</i> ; (E and F) <i>Aspergillus ochraceus</i>	155
6.16	(A and B) <i>Aspergillus niger</i> ; (C and D) <i>Aspergillus flavus</i> ; and (E and F) <i>Aspergillus fumigatus</i>	156

7.1	The effects of hormone concentration (BA+ NAA) on mean shoot number and shoot length of <i>A.mangium</i> after 6 weeks of culture incubation	176
7.2	The effects of hormone concentration (BA+ NAA) on mean shoot number and shoot length of <i>A.auriculiformis</i> after 6 weeks of culture incubation	177
7.3	The effects of hormone concentration (BA+ NAA) on mean shoot number and shoot length of <i>A.crassicarpa</i> after 6 weeks of culture incubation	177
7.4	The effects of hormone concentration (BA+ NAA) on mean shoot number and shoot length of <i>A.aulococarpa</i> after 6 weeks of culture incubation	178
7.5	The effects of hormone concentration (BA+ NAA) on mean shoot number and shoot length of <i>A.mangium</i> 'Superbulk' after 6 weeks of culture incubation	178
7.6	The effects of hormone concentration (BA+ NAA) on mean shoot number and shoot length of <i>A. hybrid</i> after 6 weeks of culture incubation	179
7.7	The effects of hormone concentration (2.0 mg ^l ⁻¹ BA+ 0.1 mg ^l ⁻¹ NAA) plus one of three additives (Silver nitrate: T3-T6, Adenine sulphate: T7-T10 and Glutamine: T11-T14) on mean shoot number and shoot length of <i>A.mangium</i> after 6 weeks of culture incubation	182
7.8	The effects of hormone concentration (3.0 mg ^l ⁻¹ BA+ 0.1 mg ^l ⁻¹ NAA) plus one of three additives (Silver nitrate: T3-T6, Adenine sulphate: T7-T10 and Glutamine: T11-T14) on mean shoot number and shoot length of <i>A.auriculiformis</i> after 6 weeks of culture incubation	183
7.9	The effects of hormone concentration (3.0 mg ^l ⁻¹ BA+ 0.1 mg ^l ⁻¹ NAA) plus one of three additives (Silver nitrate: T3-T6, Adenine sulphate: T7-T10 and Glutamine: T11-T14) on mean shoot number and shoot length of <i>A.crassicarpa</i> after 6 weeks of culture incubation	183
7.10	The effects of hormone concentration (3.0 mg ^l ⁻¹ BA+ 0.1 mg ^l ⁻¹ NAA) plus one of three additives (Silver nitrate: T3-T6, Adenine sulphate: T7-T10 and Glutamine: T11-T14) on mean shoot number and shoot length of <i>A.aulococarpa</i> after 6 weeks of culture incubation	184
7.11	The effects of hormone concentration (2.0 mg ^l ⁻¹ BA+ 0.1 mg ^l ⁻¹ NAA) plus one of three additives (Silver nitrate: T3-T6, Adenine sulphate: T7-T10 and Glutamine: T11-T14) on mean shoot number	184

	and shoot length of <i>A.mangium</i> ‘Superbulk’ after 6 weeks of culture incubation	
7.12	The effects of hormone concentration (2.0 mg ^l ⁻¹ BA+ 0.1 mg ^l ⁻¹ NAA) plus one of three additives (Silver nitrate: T3-T6, Adenine sulphate: T7-T10 and Glutamine: T11-T14) on mean shoot number and shoot length of <i>A. hybrid</i> after 6 weeks of culture incubation	185
7.13	The effects of Gibberellin concentration on shoot elongation of six <i>Acacia</i> species after 6 weeks of culture incubation	188
7.14	The effects of hormone concentration (IBA and IAA) on mean root number and root length of <i>A.mangium</i> after 6 weeks of culture incubation	191
7.15	The effects of hormone concentration (IBA and IAA) on mean root number and root length of <i>A.auriculiformis</i> after 6 weeks of culture incubation	191
7.16	The effects of hormone concentration (IBA and IAA) on mean root number and root length of <i>A.crassicarpa</i> after 6 weeks of culture incubation	192
7.17	The effects of hormone concentration (IBA and IAA) on mean root number and root length of <i>A.aulococarpa</i> after 6 weeks of culture incubation	192
7.18	The effects of hormone concentration (IBA and IAA) on mean root number and root length of <i>A.mangium</i> ‘Superbulk’ after 6 weeks of culture incubation	193
7.19	The effects of hormone concentration (IBA and IAA) on mean root number and root length of <i>A. hybrid</i> after 6 weeks of culture incubation	193
7.20	Mean values for survival rates of seedlings in acclimatization of six selected <i>Acacia</i> species clones	196
7.21	Mean values for survival rates of seedlings in acclimatization methods for six selected <i>Acacia</i> species	197
7.22	Multiple shoot induction of <i>Acacia</i> species in MS medium supplemented with 2.0 mg ^l ⁻¹ + 0.1 mg ^l ⁻¹ NAA + 100.0 mg ^l ⁻¹ AdSO ₄ + 0.1 % activated charcoal after 8 weeks of culture incubation A) <i>A. mangium</i> B) <i>A.mangium</i> ‘Superbulk’ C) <i>A.auriculiformis</i> D) <i>A. hybrid</i> E) <i>A. aulococarpa</i> (third vial from left) and F) <i>A. crassicarpa</i>	198
7.23	Formation of thick, dark green and stunted appearance of leaf disc explant of <i>A.crassicarpa</i> in medium supplemented with 3.0 mg ^l ⁻¹ BA + 0.1 mg ^l ⁻¹ NAA B) Formation of loose, white and friable	199

callus at the cut section of leaf disc of *A. auriculiformis* on BA 3.0 mg^l⁻¹ together with combination of any concentration of NAA. C) Loose patches of undifferentiated friable cell hardened after 8 weeks of culture incubation. D) Formation of adventitious roots directly from leaf segment on medium supplemented with > 3.0 mg^l⁻¹ BA + 0.5 mg^l⁻¹ NAA. E) and F) Formation of shoot directly from leaf segment on medium supplemented with 2.0 mg^l⁻¹ BA + 0.1 mg^l⁻¹ NAA + 100 mg^l⁻¹ AdSO₄

- 7.24 A) Hardened callus of *A. crassicaarpa* produced short and thin roots from base of the callus in medium supplemented with 3.0 mg^l⁻¹ BA + 0.1 mg^l⁻¹ NAA. B) Induction of thin and fibrous root formation on 1.0 mg^l⁻¹ IBA + 0.1 % activated charcoal suitable for acclimatization of *A. hybrid*. (C) Root induction on shoot elongated on 2.0 mg^l⁻¹ GA of *A. aulococarpa* on PGR-free MS medium and on (D) 1.0 mg^l⁻¹ IBA + 0.1 % activated charcoal 200
- 7.25 Root induction of *A. auriculiformis* elongated shoot on (A) 1.0 mg^l⁻¹ IBA + 0.1 % activated charcoal (B) PGR-free MS medium 201
- 7.26 (A) Shoot elongation of *A. aulococarpa* on medium supplemented with GA (0, 0.1, 0.5, 1.0, 2.0 and 3.0 mg^l⁻¹) and (B) root induction of *A. aulococarpa* shoots on medium supplemented with and IBA (0, 0.5, 1.0, 2.0 and 3.0 mg^l⁻¹) 202
- 7.27 Root induction of *A. aulococarpa* shoots on medium supplemented with and IBA (A) 0 mg^l⁻¹ (hormone-free medium) (B) 0.5 mg^l⁻¹ (C) 1.0 mg^l⁻¹ (healthy fibrous roots) and (D) 2.0 mg^l⁻¹ (crowded, thick and short roots which is not suitable for acclimatization) 203
- 8.1 Schematic drawing of the FLO/LFY gene. The minimum and maximum lengths of the exons and introns as found in angiosperms are given in bp. (Grob *et al.* 2004) 206
- 8.2 (A) Gel photograph of optimization for PCR amplification using designed primer of *A. mangium* second intron *LEAFY* for *A. mangium*; and (B) Gel photograph of optimization for PCR amplification using designed primer of *A. mangium* second intron *LEAFY* for *A. crassicaarpa* sample C6 (yielded short and long sequence) 218
- 8.3 (A) Gel photograph of optimization for PCR amplification using designed primer of *A. mangium* second intron *LEAFY* for *A. hybrid* (yielded multiple sequences); and (B) Gel photograph of PCR amplification using designed primer of *A. mangium* second intron *LEAFY* for *A. mangium* (Lane 2-5), *A. aulococarpa* (Lane 6-9), *A. hybrid* (Lane 10 and 11), *A. mangium* 'Superbulk' (Lane 12 and 13) and *A. auriculiformis* (Lane 14-17) 219
- 8.5 Maximum parsimony analysis of taxa 225

LIST OF ABBREVIATIONS

µg/ml	microgram per millilitre
µM	microMolar
°C	degree centigrade
2,4-D	2,4-dichlorophenoxyacetic acid
ABA	abscisic acid
AdS	adenine sulphate (anhydrous)
ANOVA	analysis of variance
BAP	6-benzylamina-purine
cm	centimetre
dbh	diameter at breast height
df	degree of freedom
g	gram
g/l	gram per litre
GA ₃	gibberellic acid
HCl	hydrochloric acid
HgCl ₂	mercuric chloride
IAA	indole-acetic-acid
IBA	indole-butyric-acid
Kin or K	kinetin (6-furfurylaminopurine)
kPa	kilopascal (unit for pressure)
L	litre
LB Agar	Luria Bertani agar
LSD	least significant different
m	metre
M	molar
mg	milligram
mg/l	milligram per litre
min	minute
ml	millilitre
mm	millimetre
Ms	mean of square
MS	Murashige and Skoog medium (1962)

MW	molecular weight
NAA	1-naphthaleneacetic acid
NaOCl	<i>sodium hypochlorite</i>
NaOH	sodium hydroxide
PDA	potato dextrose agar
pH	negative logarithm of the hydrogen concentration
ppm	part per million
SDW	sterile distilled water
UV	ultraviolet
v/v	volume over volume
w/v	weight over volume



CHAPTER 1

GENERAL INTRODUCTION

The tropical rainforest has long been recognized as a repository of genetic resources and is important to world's economy and for preserving the ecosystem. Malaysia, as in any developing countries has converted some of its natural forest areas for agricultural, industrial, recreational and urban development uses, but still retains 44.04 % (5,803,213 ha) of the total land area (13,184,629 ha) under forested land area in Peninsular Malaysia. Out of this, 83.52% of total forested land (4,933,787 hectares) is designated as Permanent Forest Estate (PFE) by legislation which is under sustainable management with 387,828 hectares falls under plantation land area. Approximately 10.84 million hectares of the PFE is production forest with the remaining 3.49 million hectares being protection forest (Forestry Department of Malaysia, 2014).

Due to the rapid shift in the economy of this country and increasing population growth, greater expectation are being placed on the natural forest to function as production area to fulfill the country's economic benefit (generally associated with timber harvesting) and in the meantime as a protection area for conservation of flora and fauna. Concern for the sustainability of natural forest has become a major issue now since Malaysia has traditionally relied on its timber from natural forest. In order to maintain its biodiversity and genetic resources from our natural forest, more timber has to be produced from other resources. Due to its declining trend in the domestic tropical timber production along with conversion of rubber plantation to oil palm, there is a pressing need for Malaysia to ensure wood security for its important and significant wood-based sectors (**Figure 1.1**). Several options such as reliance on the Sustainable Forest Management (SFM) only will not be enough or sufficient to readily provide raw material for drastically developing the industries of the country based on primary and secondary wood. To ensure a long term supply of raw materials for downstream industries, viability of establishing forest plantation is the only practical strategy to overcome this supply and demand paradox.

In addition, the reduction of available harvestable forests from natural distribution of virgin forest due to the above mentioned purposes of land utilization and food security has led to the introduction of several fast-growing tree species for establishment of forest plantation programme. The needs for the establishment of plantations in Malaysia have been emphasised in the National Forestry Policy 1978 (revised 1002) and encouraged the establishment of quality timber forest plantation as a strategy to overcome insufficient raw material for downstream industries.

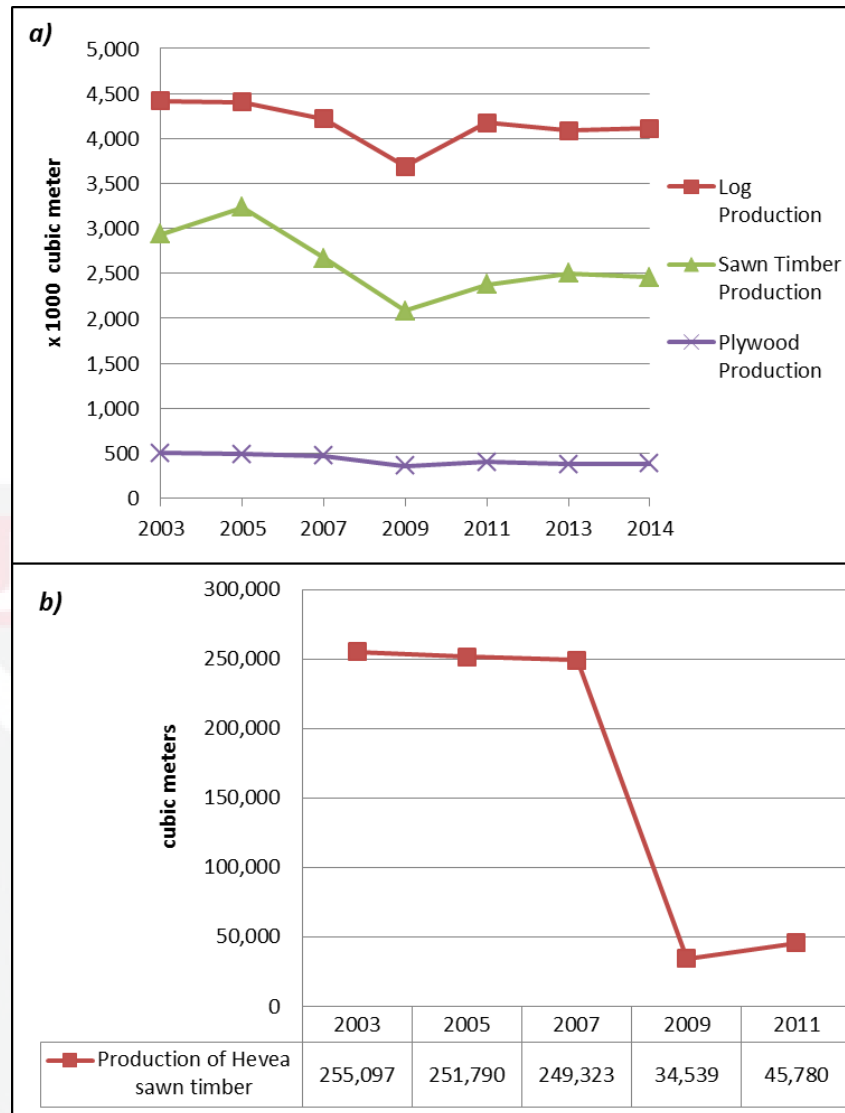


Figure 1.1: a) Production of plywood, sawn timber and logs in Malaysia between 2003-2014; and b) Production of sawn timber from *Hevea* logs in Malaysia between 2003-2011 (Forestry Department of Malaysia, 2014).

The long period of harvestable rotation age hinders private investment for establishing forest plantation. Estimates of present and future merchantable sawntimber volume have been attempted but still the assumption of a particular minimum volume of merchantable sawntimber greatly depends on the supply which genetic and environment interaction and none can predict the phenotype of a particular tree after some prolong period. Given such shortfall, return on the investment will be less than predicted by financial analysis. Furthermore, the genetic diversity of insects herbivory and the low stand density which contribute to the formation of poor quality of harvestable wood need further research to quantify the importance of the interactions between various factors (Scowcroft *et al.* 2010).

Introduction of some species for development of large scale forest plantation area in Malaysia depends totally on the objectives of the plantation and the knowledge as well as the management of the species in forest tree breeding perspective. Forest trees, especially indigenous forest species, pose a long generation period which has been the major obstacle in traditional tree breeding in Malaysia. In addition, the high heterozygosity, especially trees being propagated by seeds, hampers tree improvement through conventional breeding techniques. Thus, mass clonal propagation of superior trees/clones for forest plantation along with accelerated tree improvement programme for sustainable production of wood supplies is necessary for effective reforestation and management of forest resources.

Most of the plantations were established during the first phase of the Compensatory Forest Plantation Programme between 1982 and 1988. Since 1989, annual planting rates have not exceeded 7 000 ha. In order to achieve the target, government provided allocation for the first phase of implementation (2006-2010) and about 40,000 hectares of forest plantation was identified throughout the country in Sabah, Sarawak, Johor and Pahang (Ministry of Palntation Industries and Commodities, 2005). Since then, more forest plantations in Malaysia have been established annually in terms of total area planted with fast growing tree species. A total of 387,828 hectares of forest plantation was successfully established by the year 2014 compared to only 73,957 hectares in 2002 (**Figure 1.2**)

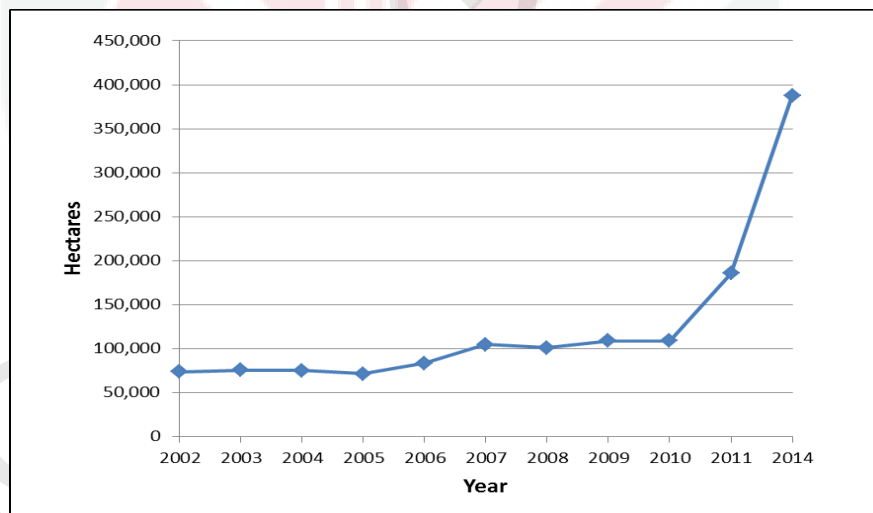


Figure 1.2: Forest plantation established in Permanent Reserved Forest in Malaysia from 2002-2014.

Another important variable for assessing the wood supply and market potential of plantations is species distribution. By thorough perambulation of both public plantation and private field throughout Malaysia, eight plantation tree species have been identified and selected as potential tree species for commercial forest plantation programme by Malaysian Timber Industry Council (MTIB, 2007). One of the main species highlighted in the selection of fast growing hardwood species is *Acacia* sp. (*Acacia* hybrid) among other timber species such as Teak (*Tectona grandis*), Rubber (*Hevea brasiliensis*), Sentang (*Azadiractha excelsa*), African Mahogany (*Khaya*

ivorensis), Kelempayan (*Neolamarckia cadamba*), Batai (*Paraserianthes falcataria*) and Binuang (*Octomeles sumatrana*). For example, of the 6 600 ha established in Johore, 6 470 ha were planted with *Acacia mangium*, whereas another 38 ha, 54 ha and 2 ha were planted with teak (*Tectona grandis*), Sentang (*Azadirachta excelsa*) and rubber respectively.

Malaysian government is striving to establish plantations comprised of fast-growing trees and one of their main goals to achieve this purpose is to establish species, provenance and progeny trial all over the country to choose for appropriate species that fits well with our environmental conditions. There are more than 600 species of tropical timbers in the world where most of them are commercially valuable in the international trade of plywood, roundwood, sawnwood and veneer (Chudnoff 1984; GTWPTN 2011, ITTO 2011; USDA FPL, 2011). Among all the fast growing tropical tree species, *Acacia* species seems to be most viable for plantation forestry and agroforestry needs of tropical Asia and Africa. It has been recommended as one of the most priority species for extensive development in tropical lowlands (Woo *et al.* 1997; Phi, 2009; Pijut *et al.* 2012). The *Acacia* species was introduced into Malaysia in the late 1960's for timber production.

Plantation of *Acacia* species gain interest as a major timber species in the 1960's especially in Peninsular Malaysia after failures in introduction of tropical pines. Once the plans for constructing the paper mill were scuttled in Malaysia, pine plantations were left unmanaged and some of the plantation area shifted to meet for more appropriate needs because pines were initially planted for the purpose of pulp production (FAO, 2002). *Acacia mangium* became one of the most extensively planted species in Malaysia along with *Hevea brasiliensis* and some trial plots have been initiated to study their performance under tropical environment. Almost 68.5% of the total plantation areas in Peninsular Malaysia are planted with *Acacia* species followed by *Hevea* species with 20.6% and others (**Figure 1.3**)

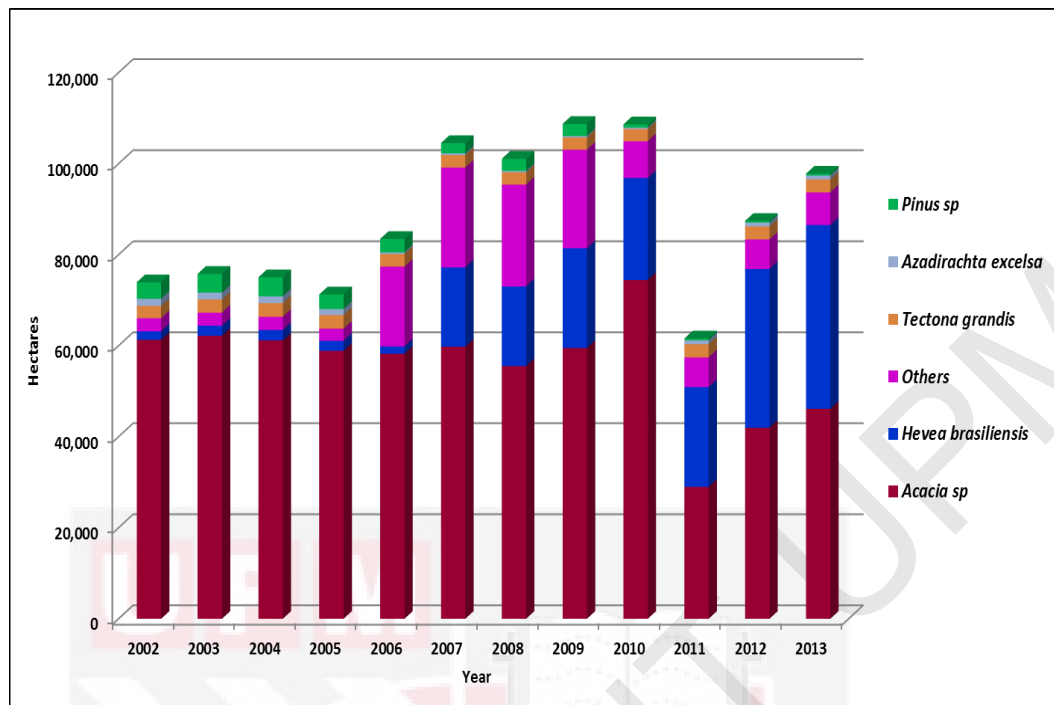


Figure 1.3: Forest plantation establishment according to species distribution in Malaysia from 2002-2013

(Source: Forestry Department of Peninsular Malaysia, 2013)

The results from international trials of *Acacia* species and provenances (Nor Aini *et al.* 1994; Zhang and Yang, 1996; Luangviriyasaeng and Pinyopusarker, 2002; Kha, 2003; Phi, 2009) have shown that *Acacia* species is a multipurpose tree species, being fast growing and suitable for various purpose such as timber production, pulp and paper production and other end products such as furniture (Turnbull *et al.*, 1998). Products and sawntimber often require trees to acquire appropriate characteristics such as straight, single-stemmed and defect-free trunks for maximum utilization (Nor Aini, 2004; Eldoma *et al.* 2015). Production of such quality wood is another big problem in plantation forestry since trees grown in plantation of some high value tropical hardwood species tend to produce low value, short butt logs and bolts due to crooked stems, low fork heights and delayed shedding of lower branches (Phi, 2009). Results from existing *Acacia* plantation in Malaysia indicated that most trees especially of *Acacia auriculiformis* fork very heavily which lead to the formation of multiple leaders and some forks are so close to the ground that they will produce little to no merchantable wood (Woo *et al.* 1997; Nor Aini *et al.* 1994; Mahat, 2007; Phi, 2009; Scowcroft *et al.* 2010).

On the contrary one of the most established *Acacia*, i.e. *A.mangiu*, is commonly assumed as the miracle tree, which suffers badly due to the incidence of root rot, heartrot and phyllode rust. Until now there are still no solutions for these diseases eventhough many researches have been done on this species. Unlike agricultural products, which can be produced for sale within a short period of time (from a few months to one to two years), most forest trees can only be harvested after 15 years. Rotations depend on the management objective and future markets are inherently

uncertain. Despite the high economic, ecological and cultural value of this multipurpose tree species, there are still no documented examples of planted stands of *Acacia* species in Malaysia that have been through a full silvicultural rotation or tree breeding, i.e. establishment, stand improvement, harvest and re-establishment of superior trees, etc.

In order to have a good productive plantation management programme for a long term, there is an urgent need to have sufficient supply of genetically improved trees for timber production. The possibilities of successful application of traditional plant breeding methods to woody trees species have their limitations due to long regeneration or reproductive cycles (Fillatti *et al.* 1987). There are few plantation trees with desirable stem form, especially if they originate from bulk seed lots collected with or without regard to stem form or timber quality. Even though there is a genetic component to good stem form and volume yield, other factors which are commonly associated with environmental factors often hinder their expression. Progeny via seed sources as a product of open pollination failed to develop better height growth of clear bole, then their parents due to wide variation of many characters from one sexual generation to another. For example, Brewbaker (1997) found that almost 90% of *Acacia Koa* trees established from 500 seed accessions produced only multi-stem progeny which is non-profitable if the objective of the plantation is to produce sawntimber. In addition, despite having maternal parents with good stem form, 55-71% trees of the same species grown in plantation developed major forking problems in the trunk within 0.3 m of the ground or at diameter of breast height (Daehler *et al.* 1999).

Management interventions in terms of traditional tree breeding alone do not ensure the establishment of quality sawntimber plantation trees without introduction of modern plant breeding. Plantation established through seeds shows a wide variation among and within species in the field due to heterozygous nature of the parents. In addition, traditional vegetative propagations such as cuttings, air layering, grafting, etc. of superior selected genotypes are very poor or in many cases not possible in any advanced tree improvement programmes as age of these elite trees become a limiting factor (Gupta, 1988; Haliza *et al.* 2012).

Taking these factors into consideration, vegetative propagation in the form of micropropagation offers great potential for mass propagation of superior clone. Modern plant breeding through tissue culture has been introduced as early as 1920 for mass production of commercially important timber trees (Krikorian, 1982; Dixon, 1985; Torrey, 1985). Optimized micropropagation protocol along with controlled culture growth is believed to produce the next generation of propagules containing high frequency of genes of favouring characters (provided the gene effects are additive) compared to the one propagated by seeds (Rao and Riley, 1994). The importance of having clonal material to study the heritability of tree form as well as other traits of interest has been stressed by many authors for some economically important tree species including some *Acacia* species (Karnosky, 1981; Glover *et al.* 1991; Vengadesan *et al.* 2003; Giri *et al.* 2003; Merkle and Nairn, 2005; Ishii, 2006; Phi, 2009; Girijashankar, 2011; Haliza *et al.* 2012; Pijut *et al.* 2012). The role and

importance of having tissue culture as a solution for production of quality propagules from superior tree species has also been emphasized by several researches (Yanchuk, 2001; Jain and Ishii, 2003; Varshney and Anis, 2014).

Modern vegetative propagation offers a great impact on advanced tree improvement programmes which includes propagation through existing meristem such as shoot and nodal explants, direct and indirect organogenesis and somatic embryogenesis. In all of these cases, adequate protocol for mass propagation of *Acacia* species has been established and optimized, and in some cases propagated tissue cultured plantlets have even been introduced into the field. (Mittal *et al.*, 1989; Yashoda *et al.*, 2004; Girijashankar, 2011; Haliza *et al.* 2012; Banerjee, 2013). Most micropropagation techniques of tropical hardwood species including of *Acacia* species have been mainly limited to short term studies using juvenile plant sources which is not favourable in advance tree improvement programme and they are limited on seeds as explant sources (Haliza *et al.* 2012; Pijut *et al.* 2012). However production and multiplication of material from matured trees with superior characteristic seems to be problematic (Girijashankar, 2011).

Therefore, taking these limiting factors into consideration, this work has been developed to evaluate the potential of mature explants to be used as initial plant source for optimization of regeneration protocol for some selected *Acacia* species. With the foregoing considerations in mind, the present study was conducted to fulfill the following objectives;

1. To evaluate the growth performance of four *Acacia* species and the selection of plus trees as sources for propagation work.
2. To develop a standard rejuvenation method for mature *Acacia* species as source for micropropagation.
3. To develop standard workable protocols for micropropagation of selected *Acacia* species, viz. *A.aulococarpa*, *A.auriculiformis*, *A.crassicarpa*, *A. hybrid (A. auriculiformis x A. mangium)*, *A. mangium* and *A. mangium* 'Superbulk'.
4. To determine the optimum conditions for the *in vitro* development and the multiplication from the various propagules (development of buds, shoots and callus).
5. To determine the optimum conditions for rooting of shoots and acclimatization.
6. To assess the genetic diversity of selected plus trees using second intron of *Acacia mangium* LEAFY gene.

REFERENCES

- Abbas, H., Qaiser, M. and Naqvi, B. (2010) Rapid *in vitro* multiplication of *Acacia nilotica* subsp. *hemispherica*, a critically endangered endemic taxon. *Pak. J. Bot.* 42: 4087- 4093.
- Abdelrahman, S.A (2000) Micropropagation of *Acacia seyal* Del. *in vitro*. *Journal of Arid Environments* 46(4):425-431.
- Abdelwahd, R., Hakam, H., Labhilili, M. and Udupa S, M. (2008) Use of an adsorbent and antioxidants to reduce the effects of leached phenolics in *in vitro* plantlet regeneration of faba bean. *African Journal of Biotechnology* 7 (8): 997-1002
- Adams, R.M., Koenigsberg, S.S. and Langhans, R.W. (1979) *In vitro* propagation of *Cephalotus follicularis* (Australian pitcher plant). *Hort. Science*, 14: 512-513.
- Ahee, J. and Duhoux, E.(1994) Root culturing of *Faidherbia= Acacia albida* as a source of explants for shoot regeneration. *Plant Cell Tiss.Organ Cult.* 36:219-225.
- Ahirwar, J.R. (2014) The study on Coppicing capacity of *Cordia myxa* (Lasora) . *International Research Journal of Biological Sciences* 3(2): 48-50
- Ahmad, N. and Anis, M. (2007) Rapid clonal multiplication of a woody tree, *Vitex negundo* L. through axillary shoots proliferation. *Agrofor Syst* 71:195–200
- Ahuja, M.R. (1986) Micropropagation of juvenile and mature beech and oak. In: D.A.Somers, B.G. Gengenbach,D.D. Biesboer, W.P. Hackett, C.E. Green (eds).Abstractof VI International Congress of Plant Tissue and Cell Culture, August 3-8, Minnesota, USA, pp. 11
- Akram, M. and Aftab, F. (2009) An efficient method for clonal propagation and *in vitro* establishment of softwood shoots from epicormic buds of teak (*Tectona grandis* L.). *For Stud China* 11:105–110
- Alex, M.D. (1995) Adventitious micropropagation of mature *Larix deciduas* using dormant versus flushed axillary vegetative buds. *New Forest* 9:61-65.
- Al-khayrI, J.M. and Al-bahrany, A.M. (2004) Genotypedependent *in vitro* response of date palm (*Phoenix dactylifera* L.) cultivars to silver nitrate. *Scientia Horticulturae*, 99(2):153-162.
- Al-Sulaiman, M.A. (2010) Clonal Propagation of *Ziziphus Spina-Christi* By Shoot Tip Culture: I. Improved Inorganic And Organic Media Constituents For *In Vitro* Shoot Multiplication *JKAU: Met., Env. and Arid Land Agric. Sci.*,21(2): 3-17.
- Altaf, N., Khan, A.R., Ali, L. and Bhatti, I.A. (2009) Tissue culture of gerbera. *Pak J Bot* 41: 7-10

- Altan, F., Burun, B. and Sahin, N. (2010) Fungal contaminants observed during micropropagation of *Lilium candidum* L. and the effect of chemotherapeutic substances applied after sterilization. *African Journal of Biotechnology* 9 (7): 991-995.
- Alvarez, I. and Wendel, J.F., (2003). Ribosomal ITS sequences and plant phylogenetic inference. *Mol. Phylogenet. Evol.* 29: 417–434.
- Amin, M.N. and Jaiswal, V.S. (1988) Micropropagation as an aid to rapid cloning of guava cultivars. *Sci. Hort.* 36:89-95.
- Anderson, W.C. (1975) Propagation of Rhododendrons by tissue culture. Part I, development of a culture medium for multiplication of shoots. *Combined Proceeding International Plant Propagators' Society*, 25: 129-135.
- Anoop, E.V., Ajayghosh, V., Muhammed Shabab, P. and Aruna, P. (2012) Provenance variation in wood anatomical properties of selected acacia species. *Journal of the Indian Academy of Wood Science* 9(2): 96-100
- Anthony, J.M., Senaratna, T., Dixon, K.W. and Sivasithamparam, K. (2004) The role of antioxidants for initiation of somatic embryos with *Conostephium pendulum* (Ericaceae). *Plant Cell, Tissue and Organ Culture* 78: 247–252, 2004
- Archambault, A. and Bruneau, A. (2004) Phylogenetic Utility of the *LEAFY/FLORICAULA*. Gene in the *Caesalpinioideae* (*Leguminosae*): Gene Duplication and a Novel Insertion. *Systematic Botany*, 29(3), 609–626.
- Arditti, J. and Ernst, R. (1993) Micropropagation of orchids. John Wiley & Sons, New York.
- Arias, A.M.G., Valverde, J.M., Fonseca, P.R. and Melara, M.V. (2010) *In vitro* plant regeneration system for common bean (*Phaseolus vulgaris*): effect of N6-benzylaminopurine and adenine sulphate. 13:1.
- Arnold, R.J. and Cuevas, E. (2003) Genetic variation in early growth, stem straightness and survival in *Acacia crassicaarpa*, *A. mangium* and *Eucalyptus urophylla* in Bukbdnon province, Philippines. *Journal of Tropical Forest Science* 15(2) 332-351
- Arumugam, S., Chu, F. H., Wang, S. Y. and Chang, S. T. (2009) *In Vitro* Plant Regeneration from Immature Leaflets Derived Callus of *Acacia confuse* Merr via Organogenesis. *Journal of Plant Biochemistry and Biotechnology* 18 (2); 197-201
- Arya, I.D., Sharma, S., Chauhan, S. and Arya, S. (2009) Micropropagation of superior eucalyptus hybrids FRI-5 (*Eucalyptus camaldulensis* Dehn x *E. tereticornis* Sm) and FRI-14 (*Eucalyptus torelliana* F.V. Muell x *E. citriodora* Hook): A commercial multiplication and field evaluation. *Afr J Biotechnol* 8(21):5718–5726

- Atwood, C.J., Fox, T.R. and Loftis, D.L. (2009). Effects of alternative silviculture on stump sprouting in the southern Appalachians. *Forest Ecology and Management*, 257: 1305-1313.
- Aziah, M. Y., McKellar, D., Fadhilah, Z., Halilah, A.K. and Haliza, I. (1999) Establishing a protocol for commercial micropropagation of *Acacia mangium* x *Acacia auriculiformis* hybrids. *J. Trop. Fores. Sci.* 11:148-156.
- Aziah, M.Y., Darus, A. and Yusuff, A. L. (1994) Micropropagation of some Tropical Forest Species. Proceeding International Workshop BIO-REFOR, Kangar, Malaysia.
- Badji, S., Mairone, Y., Ndiaye, I., Merlin, G., Danthu, P., Neville, P. and Colonna J. P. (1993) In vitro propagation of the gum Arabic tree (*Acacia Senegal* (L.) Willd) 1. Developing a rapid method for producing plants. *Plant cell Reports* 12:629-633.
- Bailey, C.D. and Doyle, J.J. (1999) Potential phylogenetic utility of the low-copy nuclear gene *pistillata* in dicotyledonous plants: comparison to nrDNA ITS and *trnL* intron in *Sphaerocardamum* and other Brassicaceae. *Molecular Phylogenetics and Evolution*, 13(1), 20–30.
- Bailey, C.D., Carr, T.G., Harris, S.A. and Hughes, C.E. (2003). Characterization of angiosperm nrDNA polymorphism, paralogy, and pseudogenes. *Mol. Phylogenet. Evol.* 29:435–455.
- Bais, H.P., Sudha, G. and Ravishankar, G.A. (2000b) Enhancement of growth and coumarin production in hairy root cultures of witloof chicory (*Cichorium intybus* L. cv. Lucknow local) under the influence of fungal elicitors. *Journal of Bioscience and Bioengineering*, 90: 648-653.
- Bais, H.P., Sudha, G., Suresh, B. and Ravishankar, G.A. (2000a). AgNO₃ influences *in vitro* root formation in *Decalepis hamiltonii* Wight, Arn. *Current Science*, 79: 894-898
- Banerjee, P. (2013) Rapid *in Vitro* Propagation of *Acacia auriculiformis* on Solid and Liquid Media: Role of Organic Additive, Antioxidant and Plant Growth Regulators. *Cibtech Journal of Bio-Protocols* 2 (1): 39-49.
- Banik, R.L. and Islam, S.A.M.N (1997) *In vitro* clonal propagation of hybrid acacia (*A. auriculiformis* × *A. mangium*). *Bangladesh Journal of Forest Science* 25 (1/2):1-7
- Bantawa, P., Roy, O.S., Ghosh, P. and Mondal, T.K. (2009) Effect of Bavistin and Adenin Sulphate on *in vitro* Shoot multiplication of *Picrorhiza scrophulariiflora* Pennell: An Endangered Medicinal Plant of Indo-china Himalayan regions. *Plant Tissue Cult. and Biotech.* 19(2): 237-245.
- Barakat, M.N. and El-Lakany, M.H. (1992) Clonal propagation of *Acacia saligna* by shoot tip culture. *Euphytica* 59: 103-107.

- Barrett, C. and Cassells, A.C. (1994) An evaluation of antibiotics for the elimination of *Xanthomonas campestris pv.pelargonii* (Brown) from *Pelargonium X domesticum* cv. 'Grand Slam'. *Plant Cell Tissue Organ Cult.* 36:169-175.
- Barrett, J.D., Park, Y.S. and Bonga, J.M. (1997) The effectiveness of various nitrogen sources in white spruce (*Picea glauca* (Moench) Voss) somatic embryogenesis. *Plant Cell Rep.* 16:411-415
- Barry, K.M., Irianto, R.S.B., Santoso, E., Turjaman, M., Widyati, E., Sitepu, I., and Mohammed, C.L. (2004) Incidence of heartrot in harvest-age *Acacia mangium* in Indonesia using a rapid survey method. *For. Ecol. Manage.* 190, 273–280.
- Bartels, H. (1971) Genetic control of multiple esterases from needles and macrogametophytes of *Picea abies*. *Planta* 99, 238–289.
- Beck, S., Dunlop, R. and Van Staden, J. (1998a) Micropropagation of *Acacia mearnsii* from ex vitro material. *Plant Growth Reg.* 26:143-148
- Beck, S., Dunlop, R. and Van Staden, J. (1998b) Rejuvenation and micropropagation of adult *Acacia mearnsii* using coppice material. *Plant Growth Reg.* 26:149-153
- Beck, S., Dunlop, R. and Van Staden, J. (2000) Meristem culture of *Acacia mearnsii*. *Plant Growth Reg.* 32:49-58
- Belarmino, M.M and Gonzales, J. (2008) Somatic embryogenesis and plant regeneration in purple food yam (*Dioscorea alata* L.) *Annals of Tropical Research* 30(2):22-33
- Belide, S., Sajjalaguddam, R.R. and Paladugu, A. (2010) Cytokinin preconditioning enhances multiple shoot regeneration in *Pongamia pinnata* (L.) Pierre - a potential, non-edible tree seed oil source for biodiesel. *Electronic Journal of Biotechnology.* 13(6): 1-9
- Bellingham, P.J. and Sparrow, A.D. (2000) Resprouting as a life history strategy in woody plant communities. *Oikos* 89: 409-416.
- Benson, E.E. (2000) *In vitro* plant recalcitrance: an introduction. *In Vitro Cell Dev Biol* 36: 141-148
- Beruto, M., Curir, P. and Debergh, P. (1999) Influence of agar on *in vitro* cultures: Biological performance of *Ranunculus* on media solidified with three different agar brands. *In vitro cell.Dev. Biol. Plant* 3 :94-101.
- Beyer, E.M. (1976a) A potent inhibitor of ethylene action in plants. *Plant Physiology*, 58(3): 268-271.
- Beyer, E.M. (1976b) Silver ion: a potent anti-ethylene agent in cucumber and tomato. *HortScience*, 11(3)175-196.

- Bhaskar, P., Subhash, K. (1995) Micropropagation of *Acacia mangium* Willd through nodal bud culture. *Indian J. Exp. Biol.* 34:590-591
- Bhatt, I.D. and Dhar, U. (2004) Factors controlling micropropagation of *Myrica esculenta* buch. – Ham. ex D. Don: a high value wild edible of Kumaun Himalaya *African Journal of Biotechnology* 3 (10): 534-540
- Black, D.K. (1973) Influences of shoot origin and certain pre and post severance treatments on the rooting and growth characteristic of Douglas fir (*Pseudotsuga menziesii* (Mirb.) Franco) by stem cuttings. Doctoral thesis. Oregon State University
- Blackburn, D.P., Hamilton, M.G., Hardwood, C.E., Innes, T.C., Potts, B.M. and Williams, D. (2011) Genetic variation in traits affecting sawn timber recovery in plantation-grown *Eucalyptus nitens*. *Annals of Forest Sciences* 68: 1187-1195.
- Boland, D. J. and Turnbull, J. W. (1981) Selection of Australian trees other than eucalypts for trials as fuelwood species in developing countries. *Australian Forestry* 44: 235-246.
- Bon, M.C., Bonal, D., Goa, D.K. and Monteuuis, O. (1998) Influence of different macro and micro solutions and growth regulators on micropropagation of juvenile *Acacia mangium* and *Paraserianthes falcataria* explants. *Plant Cell Tiss. Org. Cult.* 53: 171-177.
- Bonga, J.M. (1982) Vegetative propagation in relation to juvenility, maturity and rejuvenation. In *Tissue culture in forestry. Edited by J.M. Bonga and D.J. Durzan.* Martinus Nijhoff and Dr W. Junk Publishers, The Hague. *Forestry Sciences*, 5: 387–412.
- Bonga, J.M. (1987) Clonal propagation of mature trees: problems and possible solutions. In: Bonga J.M. and Durzan D.J. (Eds) *Cell and Tissue Culture in Forestry, Vol 1 General Principles and Borchert, R. 1976. The concept of juvenility in woody plants. Acta Hort.* 56: 21-36.
- Bonga, J.M. and von Aderkas, P. (1993) Rejuvenation of tissues from mature conifers and its implications for clonal propagation in vitro. In *Clonal Forestry: Genetics, Biotechnology and Application.* Eds. M.R. Ahuja and W.J. Libby. Springer Verlag, New York, pp 182-199.
- Booy, G., Hendriks, R.J.J., Smulders, M.J.M., Van Groenendael, J.M. and Vosman, B., (2000) Genetic diversity and the survival of populations. *Plant. Biol.* 2, 379–395.
- Borchert, R. (1976) The concept of juvenility in woody plants. *Acta Hort.* 56: 21-36.
- Borsch, T., Hilu, K.W., Quandt, D., Wilde, V., Neinhuis, C. and Barthlott, W. (2003). Noncoding plastid trnT-trnF sequences reveal a well resolved phylogeny of basal angiosperms. *J. Evol. Biol.*:16:558-576.

- Boulay, M. (1985) Some practical aspects and applications of the micropropagation of forest trees. International Symposium on *in vitro* Propagation of Forest Tree species. Bologna, Italy
- Boyle, T. (2000) Criteria and indicators for the conservation of genetic diversity. In: Young, A.C., Boshier, D., Boyle, T. (Eds.), *Forest Conservation Genetics: Principles and Practice*. CAB International, New York, USA.
- Brand, M.H. (1993) Agar and ammonium nitrate influence hyperhydricity, tissue nitrate and total nitrogen content of serviceberry (*Amelanchier arborea*) shoots *in vitro Plant Cell, Tissue and Organ Culture* 35: 203-209.
- Brewbaker, J.L. (1997) Genetic Improvement, a sine qua non for the future of Koa. In: Ferentinos L, Evans D.O (eds) *Koa, a decade of growth*. Proceeding of the Hawaii Forest Industry. Association 1996 annual symposium. Honolulu, H.L, pp 24-26.
- Brockwell, J., Searle, S., Jeavons, A. and Waayers, M. (2005) *Nitrogen Fixation in Acacias*. ACIAR, Canberra.
- Brondani, G.E., Dutra, L.F., Wendling, I., Grossi, F., Hansel, F.A. and Araujo. M.A. (2011) Micropropagation of an Eucalyptus hybrid (*Eucalyptus benthamii* x *Eucalyptus dunnii*) *Acta Scientiarum. Agronomy* 33(4): 655-663
- Browne, R.D., Davidson, C.G., Steeves, T.A. and Dunstan, D.I. (1997) Effect of Ortet on adventitious rooting of Jackpine (*Pinus banksiana*) long shoot cuttings. *Canadian Journal of Forest Research*, 27:(1) 91-96
- Buckley, P.M., DeWilde, T.N. and Reed, B.M. (1995) Characterization and identification of bacteria isolated from micropropagated mint plants. *In vitro cell.Dev.Biol.*31:58-64.
- Buiteveld, J., Vendramin, G.G., Leonardi, S., Kamer, K. and Geburek, T. (2007). Genetic diversity and differentiation in European beech (*Fagus sylvatica* L.) stands varying in management history. *Forest Ecology and Management*, 247(1-3), 98–106.
- Burns, R.M. and Honkala, B.H. (1990) *Silvics of North America*. 2 vols. U.S. Forest Serv. Handb. 654.
- Butcher, P. and Southerton, S. (2007) Chapter 15:Marker assisted Selection in forestry species.In 2007.Marker assisted Selection: Current status and future perspectives in crop, livestock, forestry and fish edited by Guimaraes, E.P., Ruane, J., Scherf, B.D., Sonnino, A. and Dargie, J.D. Food and Agriculture Organization of the United Nation (FAO, Rome).
- Cameron, A.D. and H. Sani. (1994) Growth and branching habit of rooted cuttings collected from epicormic shoots of *Betula pendula*. *Tree Physiology* 14: 427-436.

- Canadell, J. and Zedler, P. H. (1994) Underground structures of woody plants in Mediterranean ecosystems of Australia, California, and Chile. Pp. 177-210 in M. T. Kalin Arroya, P. H. Zedler & M. D. Fox (eds.), Ecology and biogeography of Mediterranean ecosystems in Chile, California, and Australia. Springer-Verlag, New York.
- Cappa, E.P., Pablo, S. Pathauer, P.S. and Lopez, G.A. (2010) Provenance variation and genetic parameters of *Eucalyptus viminalis* in Argentina. *Tree Genetics & Genomes*, 6:981-994
- Chalupa, V. (1993) Vegetative propagation of oak (*Quercus robur* and *Q. petraea*) by cutting and tissue culture. *Ann sci For* 50: 295-307.
- Chalupa, V. (2002) *In vitro* propagation of mature trees of *Sorbus aucuparia* L. and field performance of micropropagated trees. *Journal of Forest Science*, 48(12):529-535.
- Chang, S.H. Donald, D.G.M. and Jacobs, G. (1992) Micropropagation of *Eucalyptus radiata* ssp. *Radiata* using Explants from Mature and Coppice Material. *South African Forestry Journal*. 162(1): 43-47
- Chen, Z., Kolb, T.E., Clancy, K.M., Hipkins, V.D. and DeWald, L.E. (2001). Allozyme variation in interior Douglas-fir: association with growth and resistance to western spruce budworm herbivory. *Can. J. For. Res.*, 31:1691–1700
- Choudhary, K., Singh, M., Rathore, S. and Shekhawat, N.S., (2009) Somatic embryogenesis and *in vitro* plant regeneration in moth bean [*Vigna aconitifolia* (Jacq.) Marechal]: a recalcitrant grain legume. *Plant Biotechnol Rep*, 3: 205–211
- Chudnoff, M. (1984) Tropical timbers of the world. Agriculture Handbook 607. United states Department of Agriculture, Forest Service, Washington DC, 464 pp.
- Collignon, A.M. and Favre, J.M. (2000) Contribution to the postglacial history at the western margin of *Picea abies*' natural area using RAPD markers. *Ann. Bot.* 85, 713–722.
- Cook, J.E. and Sharik, T.L. (1998) Oak regeneration in the southern Appalachians: Potential, problems, and possible solutions. *Southern J. Appl. Forest.* 22:11-18.
- Copes, D.L (1992) Effects of long term pruning, meristem origin and branch order on the rooting of Douglas-fir stem cuttings. *Canadian Journal of Forest Research* (22): 1888-1894.
- Cornelius J. (1994) The effectiveness of plus-tree selection for yield. *Forest Ecology and Management* 67 23-34

- Correia, I. Alía, R., Yan, W., David, T., Aguiar, A. and Almeida, M.H. (2010) Genotype × Environment interactions in *Pinus pinaster* at age 10 in a multienvironment trial in Portugal: a maximum likelihood approach. *Annals of Forest Science* 67(6): 612
- Cotterill, P.P and Dean, C.A. (1990) Successful tree breeding with index selection. CSIRO Australia, 88p.
- Creer, S. (2007). Choosing and using introns in molecular phylogenetics. *Evolutionary Bioinformatics Online*, 3(0), 99–108.
- Creer, S., Malhotra, A., Thorpe, R.S., et al. (2005) Targeting Optimal Introns for Phylogenetic Analyses in Non-model Taxa: Experimental Results in Asian Pitvipers. *Cladistics*. 21:390–395.
- Cronk, Q.C.B. and Fuller, J.L. (1995) Plant invaders. London, Chapman & Hall.
- Cunningham, P., Nicholson, C., Yaou, S., Rinaudo, A. and Harwood, C. (2008). Utilisation of Australian Acacias for improving food security and environmental sustainability in the Sahel, West Africa. In: Underutilized Plant Species for Food, Nutrition, Income and Sustainable Development, 2008 Arusha, Tanzania.
- Cunningham, P.J. and Abasse, T. (2004) Domestication of Australian acacias for the Sahelian zone of West Africa. p. 64-74. In A. Kalinganire, A. Niang and K. Brehima (eds) Domestication des especes agroforestieres au Sahel: situation actuelle et perspectives ICRAF Working paper No. 5. World Agroforestry Centre, Nairobi.
- Daehler, C.C, Yorkston, M., Sun, W. and Dudley, N. (1999) Genetic variation in morphology and growth characters of *Acacia Koa* in the Hawaiian Island. *Int J Plant Sci* 160(4): 767-773
- Darus, A. (1991) Micropropagation techniques for *Acacia mangium* x *Acacia auriculiformis*. In: Carron, L.T., Aken, K.M., (Eds). Breeding Technologies for Tropical Acacias. Proceedings of an International Workshop held in Tawau, Sabah, Malaysia, 1-4 July 1991. ACIAR Proc. No.37:119-121.
- Darus, H. A. (1991) Multiplication of *Acacia mangium* by stem cuttings and tissue culture techniques. Adv. Trop. Acacia Res. *Proceedings Series* 35:32–35
- Darus, H.A. (1991) Micropropagation of *Acacia mangium* from aseptically germinated seedlings. *J. Trop. For. Sci.* 3:204-208.
- Darus, H.A. Darus, H.A., Carron, L.T. and Aken, K.M. (1992) Micropropagation techniques for *Acacia mangium* x *A. auriculiformis*. In: Breeding technologies for tropical Acacias. Proceedings of an international workshop, ACIAR Proceedings Series, Tawau, Sabah, Malaysia, 37:119–121

- Das, P. K., Chakravarti, V. and Maity, S. (1993) Plantlet formation in tissue culture from cotyledon of *Acacia auriculiformis* A. Cunn.ex Benth. *Indian J. For.* 16:189-192
- Daud, N.H., Jayaraman, S. and Mohamed, R. (2012) Methods Paper: An improved surface sterilization technique for introducing leaf, nodal and seed explants of *Aquilaria malaccensis* from field sources into tissue culture. *AsPac J. Mol. Biol. Biotechnol.* 20 (2) : 55-58
- Davidse, L.C. (1973) Antimitotic activity of methyl benzimidazole - 2 - yl carbamate (MBC) in *Aspergillus nidulans*. *Pesticide Biochem. Physiol.* 3: 317 - 325.
- Davies, M.E. (1972) Polyphenol synthesis in cell suspension cultures of Paul's scarlet rose. *Planta*, 104: 50-65.
- Debergh, P., Aitken, C.J., Cohen, D., Grout, B., Von, A. S., Zimmerman, R. and Ziv M. (1992) Reconsideration of the term 'vitrification' as used in micropropagation. *Plant Cell, Tissue and Organ Culture* 30: 135-140
- Deepa, V.S., Rajaram, K., Kumar, M.A., Das, S. and Kumar, P.S. (2011) High frequency regeneration and shoot multiplication in *Andrographis lineata* wall. ex. nees: an endemic medicinal plant of south India. *Journal of Medicinal Plants Research* 5(20): 5044-5049
- Del Tredici, P. (2001) Sprouting in Temperate Trees: A Morphological and Ecological Review *The Botanical Review* 67 (2):121-140
- Deng, B., Fang, S., Yang, W., Tian, Y., Shang, X. (2014) Provenance variation in growth and wood properties of juvenile *Cyclocarya paliurus*. *New Forests* 45:625-639
- Detrez, C. (1994) Shoot production through cutting culture and micrografting from mature tree explants in *Acacia tortilis* (Forsk.) Hayne subsp. *raddiana* (Savi) Brenan. *Agroforestry Systems* 25: 171-179.
- Dewan, A., Nanda, K. and Gupta S.C. (1992) *In vitro* micropropagation of *Acacia nilotica* Subsp. India Brenan via cotyledonary nodes, *Plant Cell Rep.* 12 :18-21.
- Dhabhai, K., Sharma, M. M. and Batra, A. (2010) *In vitro* clonal propagation of *Acacia nilotica* (L.) - A nitrogen fixing tree. *Res.* 2:7-11.
- Diaz-sala, C., Rey, M. and Rodríguez, R. (1990) *In vitro* establishment of a cycloclonal chain from nodal segments and apical buds of adult hazel (*Corylus avellana* L.). *Plant Cell Tissue and Organ Culture*, 23: 151-157.
- Dibax, R., Eisfeld, C. de L., Cuquel, F., Koehler, H., Quoirin, M. (2005), Plant regeneration from cotyledonary explants of *Eucalyptus camaldulensis*. *Scient. Agríc.* 62: 406-412.

- Diller, O.D. and Marshall, E. D. (1937) The relation of stump height to the sprouting of *Ostrya virginiana* in northern Indiana. *J. Forest.* 35:1116-1119.
- Diner, A.M. (1995) Adventitious micropropagation of mature *Larix decidua* using dormant versus flushed axillary vegetative buds. *New Forests* 9: 61-65
- Dixon, R.A., ed. (1985) Plant cell culture: A practical approach. IRL Press, Oxford
- Doran, J.C. and Gunn, B.V. (1987) Treatments to promote seed germination in Australian acacias. In: Turnbull JW, ed. Australian Acacias in Developing Countries. Proceedings of an International Workshop, Gympie, Qld., Australia, 4-7 August 1986. ACIAR Proceedings No 16:57-63
- Douglas, G.C. and McNamara, J. (2000) Shoot regeneration from seedling explants of *Acacia mangium* Willd, *In vitro Cell Dev. Biol.* 36 (5):412-415.
- Doyle, J.J. and Doyle, J.L. (1990). Isolation of plant DNA from fresh tissue. *Focus* 12: 13-15
- Druart, C.I., Kevers, Boxus and Gaspar. (1982) *In vitro* promotion of root formation by apple shoots through darkness effect on endogenous phenol and peroxidases. *Z. Pflanzenphysiol.* 108:429-436.
- Ducrey, M. and Turrel, M. (1992) Influence of cutting methods and dates on stump sprouting in Holm oak (*Quercus ilex* L) coppice. *Ann Sci For* 49: 449-464.
- Duhoux, E. and Davies, D. (1985) Shoot production from cotyledonary buds of *Acacia albida* and influence of sucrose on rhizogenesis. *Plant Physiol.* 121:175-180
- Dumas, E. and Monteuis, O. (1995) *In vitro* rooting of micropropagated shoots from juvenile and mature *Pinus pinaster* explants: influence of activated charcoal. *Plant Cell, Tissue and Organ Culture* 40: 231-235
- Duncan, D.R., Williams, M.E., Zehr, B. and Widholm, J.M. (1985) The production of callus capable of plant regeneration from immatured embryos of numerous *Zea mays* genotypes, *Biologia Plantarum*, 165: 322-332.
- Dunlop, R.W., Resende, M.D.V and Beck, S.L. (2005) Early assessment of first year height data from five *Acacia mearnsii* (black wattle) sub-populations in south africa using reml/blup. *Silvae genetica* 54: 4-5.
- Dvorak, W.S., Donahue, J.K. Vasquez, J.A (1996) Provenance and progeny results for the tropical white pine, *Pinus chiapensis*, at five and eight years of age. *New Forests* 12(2) :125-140
- Ebert, A. F., Taylor and Blake, J. (1993) Changes of 6-benzylaminopurine and 2,4-dichlorophenoxyacetic acid concentrations in plant tissue culture media in the presence of activated charcoal *Plant Cell, Tissue and Organ Culture* 33: 157-162.

- Economou, A.S. and Read, P.E. (1980) Effect of benzyladenine pretreatments on shoot proliferation from petunia leaf segments cultured *in vitro* . *Proc Plant Growth Reg Working Group* 7:96-103
- El Bar, O.H.A. and Dawayati, M.M.E. (2014) Histological changes on regeneration *in vitro* culture of date palm (*Phoenix dactylifera*) leaf explants. *Australian journal of crop science* 8(6):848-855
- Eldoma, A.M.A., Kumar, S. M., and Nor Aini, A.S (2015) Effects of Site Burning on Multiple Leader Formation and Growth Performance of Selected Acacia Genotypes. *American Journal of Plant Sciences* 6 :777-784.
- Eldridge, K.G., Davidson, J., Harwood, C.E and van Wyk, G. (1993). Eucalypt Domestication and Breeding. Oxford, Clarendon Press.
- Elmore, H.W., Samples, B., Sharma, S. and Harrison, M. (1990) Influence of cultural and physiochemical factors on ascorbate stability in plant tissue culture media. *Plant Cell, Tissue and Organ Culture* 20:131-135.
- Esmailnia, E. and Dehestani, A. (2015) *In Vitro* Plant Regeneration from Matured Tissues of Thomson Navel Sweet Orange (*Citrus Sinensis* L. Osbeck.) *Biharean Biologist* 9 (1): 9-14
- Evers, P. (1984) Growth and morphogenesis of shoot initials of Douglas fir, *Pseudotsuga menziesii* (Mirb.) Franco, *in vitro* 1. Plant nutrition and physical factors. *Uitvoerig Verslag* 16(1): 1-47.
- Faizuddin. M. and Dalmacio. R.V. (1996) Provenance-site interaction in mangium (*Acacia mangium* Willd.) in Philippines. *Bangl. J.For.Sci.* 25 (1-2): 53- 58.
- Falkner, F.R. (1988a) Strategy for the selection of antibiotics for use against common bacterial pathogens and endophytes of plants. *Acta Hort.* 225:53-56.
- Falkner, F.R. (1988b) The criteria for choosing an antibiotic for control of bacteria in plant tissue culture. Newsletter, international association for plant tissue culture 60:13-23.
- Falkner, F.R. (2007) The consequences of antibiotic use in horticulture. Leading article.
- FAO (2002). Case study of tropical forest plantations in Malaysia by D.B.A Krishnapillay. Forest Plantations Working Paper 23. Forest Resources Development Service, Forest Resources Division. Food and Agriculture Organization of the United Nations (FAO), Rome
- Fazal, R., Mussarrat, J., Ihsan, I. (2003) Mass propagation in *Eucalyptus camaldulensis* Dehn. *Asian J Plant Sci.* 2:184–187.
- Fernandez, S., Michaux-Ferriere, N. and Coumans, M. (1999) The embryogenic response of immatured embryo culture of durum wheat (*Triticum durum*

- Desf.): Histology and improvement by AgNO₃. *Plant Growth Regul.* 28:147–155
- Fillatti, J.J., Sellmer, J., McCown B., Haissig, B., Comai, L. (1987) *Agrobacterium* mediated transformation and regeneration of *Populus*. *Mol. Gen. Genet.* 206: 192–199
- Fontanler, E. J. and Jonkers. H. (1976) Juvenility and maturity of plants as influenced by their ontogenetical and physiological aging. *Acta Hort.* 56: 37-44.
- Forestry Department of Malaysia (2014) Annual Report. Government Printer, Kuala Lumpur.
- Francko, (1986) Studies on *Nelumbo lutea* (Willd.) Pers. I. Techniques for axenic liquid seed culture. *Aquatic Botany* 26:113-117.
- Francllet, A. (1991) Biotechnology in 'rejuvenation': hope for the micropropagation of difficult woody plants. *Acta Hort.* 289: 273- 282
- Francllet, A., Boulay, M., Bekkaoui, F., Fouret, Y., Verschoore-Martouzet, B. and Walker, N. (1987) Rejuvenation. In: Bonga JM, Durzan DJ (eds) Cell and tissue culture in forestry, general principles in biotechnology, vol 1. Martinus Nijhoff, Dordrecht
- Franklin, C.I., Dixon, R.A. (1994) Initiation and maintenance of callus and cell suspension cultures. In: Dixon RA, Gonzales RA (eds), Plant Cell Culture – a practical approach, pp.1-29. 2nd edn, Oxford University Press Inc., New York, USA.
- Franklin, C.I., Trieu, T.N., Gonzales, R.A. and Dixon, R.A. (1991) Plant regeneration from seedling explants of green bean (*Phaseolus vulgaris* L) via organogenesis. *Plant Cell Tissue Organ Cult.* 24:199-206.
- Franklin, C.I., Trieu, T.N., Gonzales, R.A., Dixon, R.A. (1991) Plant regeneration from seedling explants of green bean (*Phaseolus vulgaris* L) via organogenesis. *Plant Cell Tissue Organ Cult.* 24:199-206.
- Freeman, A.B., Duong, K.K., Shi, T.L., Hughes, C.F. and Perlin, M.H. (2002). Isolates of *Microbotryum violaceum* from North American host species are phylogenetically distinct from their European host-derived counterparts. *Mol Phylogenet Evol.* 23(2):158-70.
- Friesen, V. (2000) Introns. In: Baker A.J., editor. Molecular Methods in Ecology. Blackwell Science Ltd; pp 274–294.
- Friesen, V.L., Congdon, B.C., Walsh, H.E. and Birt, T.P. (1997) Intron variation in marbled murrelets detected using analyses of single-stranded conformational polymorphisms. *Mol. Ecol.* 6:1047–1058.

- Frohlich, M.W. and Meyerowitz, E.M. (1997) The Search for Flower Homeotic Gene Homologs in Basal Angiosperms and Gnetales: A Potential New Source of Data on the Evolutionary Origin of Flowers. *International Journal of Plant Sciences*, 158(6):131-S142
- Frohlich, M.W. and Parker, D.S.. (2000) The Mostly Male Theory of Flower Evolutionary Origins: From Genes to Fossils *Systematic Botany* 25(2): 155-170
- Froslev, T.G., Matheny, P.B. and Hibbet,t D.S. (2005) Lower level relationships in the mushroom genus *Cortinarius* (Basidiomycota, Agaricales): A comparison of RPB1, RPB2, and ITS phylogenies. *Mol. Phyl. Evol.*37:602–618.
- Frydman, V.M. and Wareing, P.F. (1973a) Phase change in *Hedera helix* L.: I. Gibberellin-like substances in the two growth phases. *J Exp Bot* 25:420–429
- Frydman, V.M. and Wareing, P.F. (1973b) Phase change in *Hedera helix* L.: II. The possible role of roots as a source of shoot gibberellin-like substances. *J Exp Bot* 24:1139–1145
- Frydman, V.M. and Wareing, P.F. (1974) Phase change in *Hedera helix* L.: III. The effects of gibberellins, abscisic acid and growth retardants on juvenile and adult ivy. *J Exp Bot* 25:420-429
- Fuentes, S.R.L., Calheiros, M.B.P., Manetti-Filho, J. and Vieira, L.G.E. (2000) The effects of silver nitrate and different carbohydrate sources on somatic embryogenesis in *Coffea canephora*. *Plant Cell Tiss. Organ Cult.* 60:5-13
- Gabriela, V. (2011) Effect of Adenine Sulfate (Adso4) On the *in Vitro* Evolution of White Clover Variety (*Trifolium Repens* L.) Analele Universitatii Din Oradea, Fascicula Protectia Mediului Vol. Xvii
- Gadow V.K. and Bredenkamp, B.V. (1992) Forest management. Pretoria: Academica
- Gales, A., (2002) Heartrot in plantations- significance to the wood processing industry. In: Barry, K., ed., Heartrots in plantation hardwoods in Indonesia and Australia. Canberra, ACIAR Technical Reports No. 51e, 18–21.
- Galiana, A., Goh, D., Chevallier, M.H., Gidiman, J., Moo, H., Hattah, M. and Japarudin, Y. (2003). Micropropagation of *Acacia mangium* x *A. auriculiformis* hybrids in Sabah. *Bois Et Forets Des Tropique* 275(1):77–82.
- Galiana, A., Tibok, A. and Duhoux, E. (1991) *In vitro* propagation of the nitrogen fixing tree-legume *Acacia mangium* Willd. *Plant Soil* 135:151-159.
- Ganesh, D. and Sreenath, H.L. (2008) Micropropagation of *Coffea arabica* using apical buds of matured field grown plants. *Crop Breeding and Applied Biotechnology*, 36: 1-7.

- Gapare, W.J., Ivkovic, M., Dutkowski, G.W., Spencer, D.J., Buxton, P. and Wu, H.X. (2012) Genetic parameters and provenance variation of *Pinus radiata* D. Don. 'Eldridge collection' in Australia 1: growth and form traits. *Tree Genetics & Genomes* 8:391-407
- Gardiner, E.S. and Helmig, L.M. (1997). Development of water oak stump sprouts under a partial overstory. *New Forests*, 14, 55-62.
- Garg, L., Bhandari, N.N., Rani, V. and Bhojwani, S.S. (1996) Somatic embryogenesis and regeneration of triploid plants of *Acacia nilotica*. *Plant Cell Rep.* 15:855-858
- Gaspar, T., Kevers, C., Penel, C., Greppin, H., Reid, D.M., Trevor and Thorpe. A. (1996) Plant hormones and plant growth regulators in plant tissue culture. *In Vitro Cellular & Developmental Biology - Plant*, 32(4): 272-289
- Gautheret, R. J. (1934) Culture du tissu cambial. *C. R. Hebd. Seanc. Acad., Sci., Paris* 198: 2195-2196.
- Geburek, Th., (1999) Genetic Variation of Norway Spruce (*Picea abies* [L.] Karst.) populations in Austria. III. Macrospatial allozyme pattern of high elevation populations. *For. Genet.* 6, 201–211.
- George, E.F (1993) Plant Propagation by Tissue Culture, Part 1, Technology, England: Exegetics Ltd., pp. 121-145.
- George, E.F. and Sherrington, P.D. (1984) Plant Propagation by Tissue Culture- Handbook and Directory of Commercial Laboratories. Eversley: Exegetics Ltd, England, 709 pp.
- George, M.W. and Tripepi, R.R. (2001) Plant Preservative Mixture™ Can Affect Shoot Regeneration from Leaf Explants of Chrysanthemum, European Birch, and Rhododendron. *Hortscience* 36(4):768-769.
- Georgieva-Todorova, J., Bohorova, N. and Atanassov, A. (1980) Proceedings of the 9th International sunflower conference, Torremolinos, Spain, 122–128
- Ghosh, M., Babu, S.S.P. and Sukul, N.C. (1993). Antifilarial effect of two triterpenoid saponins isolated from *Acacia auriculiformis*. *Indian Journal of Experimental Biology* 31: 604-606
- Giri, C.C., Shyamkumar, B., Anjaneyulu, C. (2003) Progress in Tissue culture, genetic transformation and application of biotechnology to trees: an overview. *Trees* 18(2): 115-135
- Giridhar, P., Indu, E.P., Vijaya, R.D., Ravishankar, G.A. (2003) Effect of silver nitrate on *in vitro* shoot growth of Coffee. *Tropical Science*, 43:144-146.

- Giridhar, P., Obul reddy, B. and Ravishankar, G.A. (2001) Silver nitrate influences *in vitro* shoot multiplication and root formation in *Vanilla planifolia* Andr. *Current Science*, 81(9): 1166-1170.
- Girijashankar, V. (2011) Micropropagation of multipurpose medicinal tree *Acacia auriculiformis*. *Journal of Medicinal Plant Research* 5: 462-466.
- Glover, N., Brewbaker, J.L., Conkle, T., Conrad, C.E., Shebata, S. (1991) Strategies for genetic improvement. In: Brewbaker J.L., Glover N, Moore E (eds) Improvement of *Acacia Koa*: resource documents. Proceeding of a workshop, April 18-20, 1991. Kamuela, H.L., pp27-30.
- Godefroy, I. (2008) Micropropagation by *in vitro* axillary budding of juvenile *Acacia auriculiformis* clones Influence of the genotype. *Bois et Forêts des Tropiques* (297): 27-34
- Goel, V.L. and Behl, H.M. (2001) Genetic selection and improvement of hard wood tree species for fuelwood production on sodic soil with particular reference to *Prosopis juliflora*. *Biomass and Bioenergy* 20(1): 9-15.
- Gould, J.H. and Murashige, T., (1985) Morphogenic substance released by plant tissue cultures. Identification of barberine in *Nandina* culture medium, morphogenesis, and factors influencing accumulation. *Plant Cell Tissue and Organ Culture*, 4: 29-42.
- Green, B., Tabone, T. and Felker, P. (1990) A comparison of amide and ureide nitrogen sources in tissue culture of tree legume *Prosopis alba* clone B2 V50. *Plant Cell Tissue Org Cult* 21:83–86
- Greenwood, M.S. (1987) Rejuvenation of forest trees. *Plant Growth Regul.* 6:1-12
- Greenwood, M.S. (1995) Juvenility and maturation in conifers: current concepts. *Tree Physiol.* 15: 433-438
- Griffin, A. and Nor Aini, A. S. (1998) Callus induction of *Acacia crassicarpa*. *Malays For.* 61:190-195.
- Griffin, A., Kumar, S.M. and Shukor, N.A.A. (2014) *In vitro* regeneration of *Acacia crassicarpa* A. Cunn Ex Benth through organogenesis from juvenile sources. *Journal of Food, Agriculture and Environment* 12: 375-382
- Grob, G.B.J., Gravendeel, B. and Eurlings, M.C.M. (2004). Potential phylogenetic utility of the nuclear *LEAFY/FLORICAULA* second intron: Comparison with three chloroplast DNA regions in *Amorphophallus* (Araceae). *Molecular Phylogenetics and Evolution*, 30(1), 13-23
- GTWPTN. Global Timber and Wood Product Trade Network (2011). Access: <http://www.globalwood.org/>

- Guanih, V.S., Mahali, A. and Tuyok, M. (2004). Seed sterilization of *Dryobalanops lanceolata* Burck. *Sepilok Bulletin* 1: 59-62
- Gugerli, F., Sperisen, C., Buchler, U., Magni, F., Geburek, T., Jeandroz, S. and Senn, J. (2001) Haplotype variation in a mitochondrial tandem repeat of Norway spruce (*Picea abies*) populations suggests a serious founder effect during postglacial re-colonization of the western Alps. *Mol. Ecol.* 10, 1255–1263.
- Guimaraes, E.P., Ruane, J., Scherf, B.D., Sonnino, A. and Dargie, J.D. (2007) Marker assisted Selection: Current status and future perspectives in crop, livestock, forestry and fish. Food and Agriculture Organization of the United Nation (FAO, Rome).
- Guo, Y.L. and Ge, S. (2004). The utility of mitochondrial nad1 intron in phylogenetic study of *Oryzae* with reference to the systematic position of *Porteresia*. *Acta Phytotaxon. Sin.* 42:333–344.
- Gupta, P., Patni, V., Kant, U. and Arya, H.C. (1994) *In vitro* multiple shoot formation from mature trees of *Acacia senegal* (Linn.) Willd. *J. Indian Bot. Soc.* 73: 331-332.
- Gupta, P.K. (1988) Advances in biotechnology of conifers. *Current Science* 57(12): 629-637.
- Gupta, P.K., Nadgir, A.L., Mascarenhas, A.F. and Jagannathan, V. (1980) Tissue culture of forest trees: clonal multiplication of *Tectona grandis* L. by tissue culture. *Plant Sci. Lett.* 17: 259-268.
- Gupta, S.C. and Agrawal, V.P. (1992) Micropropagation of woody taxa and plant productivity. In: Prasad, B. N., Ghimire, G.P.S., Agrawal, V.P., (eds.) Role of biotechnology in agriculture. New York: International Science Publisher :37-52.
- Haines, R.J. (1992) Mass Production Technology for Genetically Improved, Fast Growing Forest Tree Species. Mass Propagation by Cuttings, Biotechnologies, and the Capture of Genetic Gain. Paper Presented At IUFRO Symposium. Bordeaux, France.
- Haldeman, J.H., Thomas, R.L. and McKamy, D.L. (1987). Use of benomyl and rifampicin for *in vitro* shoot tip culture of *Camellia sinensis* and *C. japonica*. *Hort. Sci.* 22: 306-307.
- Halfield-Vieira, B.D., Mourao, M., Tonini, H., Nechet, K.D., (2006) Heartrot in homogeneous stands of *Acacia mangium*. *Pesq. Agropec. Brasil* 41, 709 - 711.
- Haliza, I., Nor Aini, A.S., Aziah, M. Y., Nor, H. H., Fadhilah, Z., Nazirah, A. and Siti, S. A. 2012. *In vitro* shoot induction of *Acacia auriculiformis* from juvenile and mature sources. *J. Biotechnol. Pharm. Res.* 3:88-93.

- Hamami, M. and Semsolbahri (2003) Wood samples and wood properties relationship in planted *Acacia*. Malaysia samples. Proceeding. International symposium on sustainable utilization of *Acacia*, Kyoto. Pp 24-34.
- Hansen, J.K. and Larsen, J.B. (2004) European silver fir (*Abies alba* Mill.) provenances from Calabria, southern Italy: 15-year results from Danish provenance field trials. *European Journal of Forest Research* 123 (2):127-138
- Harikrisna, K., Lee, W.W., Ong, C.H. and Siti, S. (2002). Section 2: Basic Tools and methodologies supportive of Biotechnology. Edited by Krishnapillay in Basic Principles of biotechnology and their application in forestry. Asia pacific Association of forestry research Institutions(APAFRI)
- Harmer, R. (1988) Production and use of epicormic shoots for the vegetative propagation of mature oak. *Forestry* 61: 305-316.
- Harry, I.S. and Thorpe, T.A. (1994) *In vitro* cultures of forest trees. In Vasil, I. K. and Thorpe, T. A. (eds.). Plant Cell and Tissue Culture. Kluwer Academic Publishers, the Netherlands, pp. 539-560.
- Hartmann, H.T., Kester, D.E., Davies, F.T., and Geneve, R.L.(1997) Plant propagation: principles and practices. 6th edition. Prentice-Hall, Inc., Englewood Cliffs, New Jersey, USA.
- Harvey, R.G., Muzik, T.J. and Warner, R.L. (1973) Differences in nitrogen requirements of two clones of *Convolvulus arvensis* *in vitro*. *Amer J Bot* 60:76-79
- Harwood, C. (2011) Strengthening the tropical acacia plantation value chain: the role of research. *Journal of Tropical Forest Science* 23(1):1-3
- Harwood, C.E., Alloysius, D., Pomroy, P., Robson, K.W. and Haines N.W. (1997) Early growth and survival of *Eucalyptus pellita* provenances in a range of tropical environments, compared with *E. grandis*, *E. urophylla* and *Acacia mangium*. *New Forests* 14(3): 203-219
- Harwood, C.E. and Williams, E.R. (1992) A review of provenance variation in growth of *Acacia mangium* In: L.T. Carron and K.M.Aken (Eds.) Breeding Technologies for Tropical Acacias. ACIAR Proc. No.37. 22- 30.
- Henry, P.H. and Preece, J.E. (1997) Production and rooting of shoots generated from dormant stem section of maple species. *Hortscience* 32(7): 1274-1275
- Hodge, G.R. and Dvorak W.S. (2004) The CAMCORE international provenance/progeny trials of *Gmelina arborea*: genetic parameters and potential gain. *New Forests* 28 (2-3):147-166
- Hoot, S.B., and Taylor, W.C. (2001) The utility of nuclear ITS, a *LEAFY* homolog intron, and Chloroplast *atpB-rbcL* spacer region data in phylogenetics analyses and species delimitation in *Isoetes*. *American Fern Journal* 91: 166-177

- Huang, F.H., Al-Khayri, J.M., Gbur, E.E. (1994) Micropropagation of *Acacia mearnsii*. *In Vitro Cell Dev. Biol.* 30:70-74.
- Huang, L., Lius, S., Huang, B., Murashige, T., Mahdi, E.M. and Gundy, R.V. (1992) Rejuvenation of *Sequoia sempervirens* by Repeated Grafting of Shoot Tips onto Juvenile Rootstocks in Vitro. *Plant Physiol* 98: 166-173
- Huccius, B. (1978) Question of unicellular origin of nonzygotic embryos in callus cultures. *Phytomorphology* 28:74-81.
- Huetteman, C.A. and Preece, J.E. (1993) Thidiazuron: a potent cytokinin for woody plant tissue culture. *Plant Cell Tissue Org Cult* 33:105-119.
- Hughes, C.E., Eastwood, R.J. and Bailey, C.D. (2006) From famine to feast? Selecting nuclear DNA sequence loci for plant species-level phylogeny reconstruction. *Phil. Trans. Roy. Soc. B.*, 361:211–225.
- Husen, A. and Pal, M. (2003) Effect of serial bud grafting and etiolation on rejuvenation and rooting cuttings of matured trees of *Tectona grandis* Linn. f. *Silvae Genet.* 52: 84-88
- Hustache, G., Barnoue, F. and Joseleau, J.P. (1986) Callus formation and induction of a cell suspension culture from *Acacia senegal*. *Plant Cell Rep.* 5:365-367
- Ibanez, M.R., Amo-marco, J.B. (1998) Promotion by phoroglucinol of micropropagation of *Minuartia valentine*-an endangered, endemic Spanish plant. *Plant Growth Reg* 26:49-56
- Ide, Y., Wantanabe, Y. and Ikeda, H. (1994) Tissue culture of *Acacia auriculiformis* using the aseptically germinated seedlings. *J. Jap. For. Soc.* 76:576-583
- Indira, E.P. (1999) Provenance trials in *Acacia*. Kerala Forest Research Institute (KFRI) Research Report 171. p21
- Indra, D.B and Dhar, U. (2004). Factors controlling micropropagation of *Myrica esculenta* buch. – Ham. ex D. Don: a high value wild edible of Kumaun Himalaya. *African Journal of Biotechnology* 3 (10): 534-540.
- Inoka, K.P.I. and Dahanayake, N. (2015). Effect of plant growth regulators on micro-propagation of Sunflower (*Helianthus annuus* L.). *International Journal of Scientific and Research Publications*, 5(1): 1-5
- Iqbal, J., Dutt, V., Ahmad, H., Bhat, G.M. and Khan, P.A. (2014) Propagation of *Quercus robur* L. (English oak) by stem cuttings in Western Himalayas (Kashmir). *Environment Conservation Journal* (15)1/2: 185-189
- Isabel, A., Toni, M. and Juan, S., (1991) Micropropagation of juvenile and adult *Sorbus domestica* L. *Plant cell, Tissue and Organ Culture.* 27:341-348.

- Ishii, K. (2006) Recent developments in vegetative propagation techniques and their application for tropical forest trees. pp, 31-39 in Kazuo Suzuki, Katsuaki Ishii, shobu Sakurai and Satohiko Sasaki (eds) in Plantation technology in tropical forest science, 2006. Springer verlag Tokyo.
- Ito, S. (2002) The incidence of heartrot and disease severity on several *Acacia* species in SAFODA plantations. Study Report of SAFODA-JICA project, 16 pp.
- Ito, S. and Nanis, L.H. (1994) Heartrot of *Acacia mangium* in SAFODA plantations. Sabah Reafforestation Technical Development and Training Project, Study Report. Chin Chi Printing Works, Sabah, 52 pp.
- ITTO. International Tropical Timber Organization (2011). Statistics database. Access: http://itto.int/annual_review_output/
- Jahan, M.S., Sabina, R. and Rubaiyat, A. (2008a). Alkaline pulping and bleaching of *Acacia auriculiformis* grown in Bangladesh. *Turkish Journal of Agricultural Forum* 32 (4), 339–347.
- Jain, S. M., and Ishii, K. (2003) Micropropagation of woody trees and fruits. Springer Forestry Sciences (Vol. 75). p. 852
- Jansen, k., Soht, J., Kohnle, U., Ensminger, I. and Gessler, A. (2013) Tree ring isotopic composition, radial increment and height growth reveal provenance-specific reactions of Douglas-fir towards environmental parameters. *Trees* 27:37–52
- Jha, T.H., Mukerjee, P. and Datta, M.M. (2007). Somatic embryogenesis in *Jatropha curcas* Linn., an important biofuel plant. *Plant Biotechnol Rep* 1:135-140
- Johansson, T. (1992). Sprouting of 10- to 50-year-old *Betula pubescens* in relation to felling time. *Forest Ecology and Management*, 53: 283-296.
- Johnson, K.P., and Clayton, D.H. (2000) Nuclear and mitochondrial genes contain similar phylogenetic signal for pigeons and doves (Aves: Columbiformes) *Mol. Phyl. Evol.* 14:141–151.
- Johnson, P.S. (1975) Growth and structural development of red oak sprout clumps. *Forest Sci.* 21: 413-418.
- Johnson, P.S. (1978) Predicting oak stump sprouting and sprout development in the Missouri Ozarks. U.S. Forest Serv. Res. Pap. NC-149.
- Jones, C. and Smith, D. (1988) Effect of 6-benzylaminopurine and 1-naphthlacetic acid on *in vitro* axillary bud development of mature *Acacia melanoxylon*. *Proc. Int. Plant Prop. Soc.* 38:389-393
- Jones, T.C., Batchelor, C.A. and Harris, P.J.C. (1990) *In vitro* culture and propagation of *Acacia* species (*Acacia bivenosa*, *Acacia holosericea*, *Acacia salicina*, *Acacia saligna* and *Acacia sclerosperma*). *Int. Tree Crops J.* 6:183-192.

- Joshi, A.G., Pathak, A.R., Sharma, A.M. and Singh, S. (2010). High frequency of shoot regeneration on leaf explants of *Bacopa monnieri*. *Environmental and Experimental Biology* 8: 81-84
- Joshi, P., Bisht, V.K., Sharma and Uniyal, D.P. (2003) *In vitro* Clonal Propagation of Matured Eucalyptus F1 Hybrid (*Eucalyptus tereticornis* SM. x *E. grandis* HILL ex. MAIDEN) *Silvae Genetica* 52:3-4
- Juddy, E.J. (1997) Micropropagation of sentang (*Azadirachta excels* Jack.). Unpublished Masters of Science dissertation, Universiti Putra Malaysia, Serdang, Malaysia, 182 p.
- Kabir, S.N., Ray, H.N., Pal, B.C. and Mitra, D. (2008) Pharmaceutical composition having virucidal and spermicidal activity. USPTO Application #:20080300197.
- Kamis A., Nor Aini, A.S, Adjers, G., Bhumibhamon S., Pan, F.J and Venkateswarlu, P. (1994) Performance of *Acacia auriculiformis* provenance at 18 months of four sites. *Journal of Tropical Forest Science* 7(2): 251-261.
- Kamis A., Nor Aini, A.S. and Senin A.L. (1995) Two year performance of *Acacia crassicarpa* Provenance at Serdang, Malaysia. *Journal of Tropical Agricultural Sciences* 18(3):177-181.
- Kanada, H. and Harada, J. (1971) Studies on the organogenesis in tissue culture . In: Effects of Growth Regulators on Somatic Embryogenesis and plant formation, *Zpflanzenphysiol* 91: 255-260.
- Karnosky, D.F, (1981) Potential for forest tree Improvement via Tissue Culture. *Bioscience* 31(2); 114-120
- Kaschula, S.A., Twine, W.C. and Scholes, M.C. (2005) The effect of catena position and stump characteristics on the coppice response of three savannah fuelwood species. *Environmental Conservation*, 1:76-84
- Kataeva, N.V., Alexandrova, I.G., Butenko, R.G. and Dragavtceva, E.V. (1991). Effect of applied and internal hormones on vitrification and apical necrosis of different plants cultured in vitro. *Plant Cell, Tissue and Organ Culture* 27:149-154.
- Katakya, A. and Handique, P.J. (2010) Standardization of sterilization techniques prior to in vitro propagation of *Andrographis paniculata* (Burm.f) Nees. *Asian Journal of Science and Technology* 6: 119-122.
- Kaur K., Verma B. and Kant U. (1998) Plants obtained from the khair tree (*Acacia catechu* Willd) using mature nodal explants. *Plant Cell Rep.* 17(5):421-429.
- Kha, L.D. (2003) Chon Tao going va nhan going cho mot so loai cay trong rung chu yeu o Viet Nam. Ha Noi: Agriculture Publishing House, 292 p. (in Vietnamese).

- Kha, L.D., Hai, N.D. and Vinh, H.Q. (1998) Clonal test and propagation options for natural hybrid between *Acacia mangium* and *A.auriculiformis*. In J.W. Turnbull, H.R Crompton and K. Pinyopusarek, eds. Recent development in acacia planting. Pp 203-210.ACIAR Proc No 82, Canberra,ACIAR.
- Khalafalla, M.M. and Daffalla, H.M. (2008) *In Vitro* Micropropagation and Micrografting of Gum Arabic Tree (*Acacia senegal* (L.) Wild). *Int. J. Sustain. Crop Prod.* 3(1):19-27
- Khasa, P.D, Li, P., Vallee, G., Magnussen, S. and Bousquet, J. (1995) Early evaluation of *Racosperma auriculiforme* and *R. mangium* provenance trials on four sites in Zaire. *Forest Ecology and Management* 78:99-103
- Khayri, J.M. and Gbur, E.E. (1994) Micropropagation of *Acacia mearnsii*. *In Vitro Cell Dev. Pl.* 30:70-74.
- Kneifel, W. and Leonhardt, W. (1992) Testing different antibiotics against Gram-positive and gram-negative bacteria isolated from plant tissue culture. *Plant Cell Tiss. Org. Cult.* 29:139-144.
- Ko, W.H., Su, C.C., Chen, C.L. and. Chao, C.P. (2009) Control of lethal browning of tissue culture plantlets of Cavendish banana cv. Formosana with ascorbic acid. *Plant Cell Tiss Organ Cult* 96:137-141
- Kocher, T.D., Thomas, W.K., Meyer, A., Edwards, S.V., Paabo, S., Villablanca, F.X., Wilson, A.C. (1989). Dynamics of mitochondrial DNA evolution in animals: amplification and sequencing with conserved primers. *Proc. Natl. Acad. Sci.*, 86:6196-6200
- Kossuth, S.V. and Ross S.D. (1989) Hormonal Control of Tree Growth: Proceedings of the Physiology Working Group Technical Session, Society of American Foresters National Convention, Birmingham, Alabama, USA, October 6–9, 1986 *Volume 28 of Forestry Sciences* Springer Science & Business Media 243 pages
- Krikorian, A. D. (1982) Cloning Higher Plants from Aseptically Cultured Tissues And Cells. *Biological Reviews* 57(2) : 151–218.
- Krikorian, A.D., Kann, R.P., O'Connor, S.A and Fitter, M.S. (1983) Chromosomal stability in cultured plant cells and tissues. Pp. 541-581. In: Handbook of Plant Cell Culture. Vol. 1. Ed by D.A. Evans, P.V.Ammirato, and Y. Yamada. Macmillan Publications, New York.
- Krutovskii, K.V. and Bergmann, F., (1995) Introgressive hybridization and phylogenetics relationships between Norway, *Picea abies* (L.) Krast., and Siberian, *P. obovata* Ledeb., spruce species studied by isoenzyme loci. *Heredity* 74, 464–480.
- Kulkarni, A.A., and Krishnamurthy, K.V. (2009) Contamination control and enhance axillary budding from mature explants of *Taxus Baccata ssp.Wallichiana*

Chapter 1(1-28) in *Plant tissue culture and Molecular Markers*.edited by Ashwani Kumar and H.C.Arya. I. K. International Publishing house Pvt Ltd 2009.New Delhi.

- Kumar, A. (1992). Micropropagation of a mature leguminous tree *Bauhinia purpurea*. *Plant Cell Tiss Org Cult* 31:257- 259
- Laine, E. and David, A. (1994) Regeneration of plants from leaf explants of micropropagated clonal *Eucalyptus grandis*. *Plant Cell Rep.*, 13: 473-476.
- Lamichhane, D. and Thapa, H.B. (2011) International provenance trial of Neem (*Azadirachta indica*) in the terai region of Nepal. *Agroforest Syst* 81:37-43
- Langner, W. (1953) Eine Mendelspaltung bei Aurea-Formen von *Picea abies* (L.) Karst. als Mittel zur Klärung der Befruchtungsverhältnisse im Walde. *Zeitschrift für Forstgenetik und Forstpflanzenzüchtung*, 2 : 49–51
- Larwanou, M., Abdoulaye, M. and Reij, C. (2006). Etude de la Regeneration naturelle assistee dans la region de Zinder (Niger). United States Agency for International Development (USAID/EGAT). International Resources Group, Washington DC.
- Lau, Oi-Lim and Yang, Shang F. (1976) Inhibition of ethylene production by cobaltous ion. *Plant Physiology*. 58:1. 114-117.
- Laukkanen, H., Haggman, H., Kontunen-Soppela, S. and Hohtola, A. (1999) Tissue browning of *in vitro* cultures of Scots pine: Role of peroxidase and polyphenol oxidase. *Physiol. Plant*. 106: 337-343.
- Lavanya, A.R., Muthukrishnan, S., MuthuKumar, M., Benjamin, J.H.F., Kumar, S T., Kumaresan, V., and Rao, M.V. (2014) Indirect organogenesis from various explants of *Hildegardia populifolia* (Roxb.) Schott and Endl.- A threatened tree species from Eastern Ghats of Tamil Nadu, India..*Journal of Genetic Engineering and Biotechnology* 12: 95-101
- Le Roux, J.J., and Van Staden, J. (1991) Micropropagation of Eucalyptus species. *Journal of Hortscience*, 26(5): 199-200.
- Lee, S.S. (2002) Overview of the heartrot problem in Acacia—gap analysis and research options. In: Barry, K. (Ed.), Heartrots in Plantation Hardwoods in Indonesia and Australia. ACIAR Technical Report 51e. CSIRO Publishing, Canberra, pp. 26-34.
- Lee, S.S., Teng, S.Y., Lim, M.T., Kader, R.A., (1988) Discoloration and heartrot of *Acacia mangium* Willd.- some preliminary results. *J. Trop. For. Sci.* 1: 170-177.
- Leifert, C. and Cassells, A.C. (2001) Microbial hazards in plant tissue and cell cultures *In Vitro Cell. Dev. Biol.-Plant* 37:133-138.

- Leifert, C., Ritchie, J.Y. and Waites, W.M. (1991) Contaminants of plant-tissue and cell cultures. *World Journal of Microbiology and Biotechnology*, 7(4):452-469.
- Leifert, C., Waites, B., Keetley, J.W., Wright, S.M., Nicholas, J.R. and Waites, W.M. (1994) Effect of medium acidification on filamentous fungi, yeasts and bacterial contaminants in Delphinium tissue cultures. *Plant Cell Tissue Organ Cult.*36:149-155
- Lessa, E.P. (1992). Rapid surveying of DNA sequence variation in natural populations. *Mol. Biol. Evol.*9:323–330.
- Linington, I.M. (1991) *In vitro* propagation of *Dipterocarpus intricatus*. *Plant Cell Tissue Org Cult* 27: 81-88
- Liu, Y., Tonga, X., Huib, W., Liu, T., Chen, X., Lia, J., Zhuanga, C., Yanga, Y. and Liua, Z. (2015) Efficient culture protocol for plant regeneration from petiole explants of physiologically matured trees of *Jatropha curcas* L. *Biotechnology & Biotechnological Equipment*, 29(3) :479-488
- Logan, A.F. (1987) Australian Acacias for pulpwood. in: J.W.Turnbull (Ed.) Australian Acacias in developing countries. ACIAR Proc. No. 16. 89- 94.
- Lopez-Upton, J.J.K. Donahue, F.O., Plascencia-Escalante, C. and Ramirez-Herrera. (2005) Provenance variation in growth characters of four subtropical pine species planted in Mexico. *New Forests* 29 (1): 1-13
- Luangviriyasaeng, V. and Pinyopusarek, K., (2002) Genetic variation in second generation progeny trial of *Acacia auriculiformis* in Thailand. *J. Trop. Forest Sci.* 14, 131–144.
- Luoga, J.E., Witkowski, E.T.F. and Balkwill, K. (2004) Regeneration by coppicing (resprouting) of miombo (African savanna) trees in relation to land use. *Forest Ecology and Management*, 189(1-3): 23-35
- Macarenhas, A.F., Kundurkar, S.V. and Khupse, S.S. (1993) Micropropagation of Teak, In *Micropropagation of woody plants*, M.R Ahuja (Ed.). Academic Publishers, Dordrecht: 247-262.
- MacDonald, J.E. and Powell, G.R. (1983) Relationship between stump sprouting and parent-tree diameter in sugar maple in the first year following clear-cutting. *Canad. J. Forest Res.* 13: 390-394.
- Maghuly, F., Pinsker, W., Praznik, W. and Fluch, S. (2006). Genetic diversity in managed subpopulations of Norway spruce [*Picea abies* (L.) Karst.]. *Forest Ecology and Management*, 222(1-3), 266–271.
- Mahat, M. N. (2007) Growth Performance and genetic variation of four *Acacia* species planted in Pahang, Malaysia. Doctoral Thesis, Universiti Putra Malaysia. 298pp.

- Mallika, V.K., Mathew, J.P., Chacko, D., Vijaya Kumar, N.K. and Nair, R.V. (1996) Induction of multiple shoots in nodal explants of fully grown trees of cocoa. *J Plantn Crops*. 24: 503-510.
- Mandal, P, Babu, S.S.P. and Mandal, N.C. (2005) Antimicrobial activity of saponins from *Acacia auriculiformis*. *Fitoterapia* 76(5): 462-465
- Manzanera, J.A. and Pardos, J.A. (1990) Micropropagation of juvenile and adult *Quercus suber* L. *Plant Cell Tissue and Organ Culture*, 21: 1-8.
- Marga, F., Vebret, L. and Morvan, H. (1997) Agar fractions could protect apple shoots cultured in liquid media against hyperhydricity. *Plant Cell Tiss Org Cult* 49: 1-5
- Marthur, I., Chandra, N. (1983) Induced regeneration in stem explants of *Acacia nilotica*. *Curr. Sci.* 52:882-883
- Marton, L. and Browse, J., (1991) Facile transformation of *Arabidopsis*. *Plant Cell Rep* 10:235-239
- Maslin, B.R. and McDonald, M.W. (1996) A key to useful Australian acacias for the seasonally dry tropics. Canberra, CSIRO Forestry and Forest Products.
- Maslin, B.R., Miller, J.T. and Seigler, D.S. (2003). Overview of the generic status of *Acacia* (Leguminosae: Mimosoideae). *Australian Systematic Botany* 16, 1-18
- Mason-Gamer, R.J. (2001) Origin of North American *Elymus* (Poaceae: Triticeae) allotetraploids based on granule-bound starch synthase gene sequences. *Syst. Bot* 26: 757-768
- Mathias, P.J., Alderson, P.G. and Leakey, R.R.B. (1987) Bacterial contamination in tropical hardwood cultures. *Acta Hort.* 212:43-48.
- Mathur, A., Mathur, A.K., Verma, P. Yadav, S. Gupta, M.L. and Darokar, M.P. (2008). Biological hardening and genetic fidelity testing of micro-cloned progeny of *Chlorophytum borivillianum* Sant. et Fernand. *Afr. J. Biotechnol.* 7(8): 1046-1053.
- McCown, B.H., Sellmer, J.C. (1987). General media and vessels suitable for woody plant cultures. In: Bonga, J.M., Durzan, D.J. (Eds.), *Tissue culture in forestry - General principles and biotechnology*, Vol. 2. Martinus Nijhoff Publ., Dordrecht, Boston pp. 4-6.
- McDonald, M.W. (2003). Revision of *Acacia tumida* (Leguminosae: Mimosoideae) and close allies, including the description of three rare taxa. *Aust. Syst. Bot.* 16: 139-164
- McNamara, S., Duong Viet Tinh, Erskine, P.D., Lamb, D., Yates, D., Brown, S., (2006) Rehabilitating degraded forest land in central Vietnam with mixed 495 native species plantings. *Forest Ecology and Management* 233 (2-3):358-365.

- Mead, D.J. and Speechly, H.T. (1991) Growing *Acacia mangium* for high quality sawlogs in Peninsular Malaysia. In: Sheikh Ali Abod et al., (ed.), Recent developments in tree plantations of humid/sub-humid tropics of Asia. Serdang, Selangor, Universiti Pertanian Malaysia, 54–69
- Meeks, J.C., Wolk, C.P., Schilling, N., Shaffer, P.W., Avissar, Y. and Chien, W. (1978) Initial organic products of fixation of [J¹⁵N] dinitrogen by root nodules of soybean *Glycine max*. *Plant Physiol* 61:980-983
- Meier, K. and Reuther, G. (1994) Factors controlling micropropagation of mature *Fagus sylvatica*. *Plant Cell Issue and Organ Culture* 39:231-238
- Menalled, F.D., Kelty, M.J. and Ewel, J.J., (1998) Canopy development in tropical tree plantations: a comparison of species mixtures and monocultures. *For. Ecol. Manage.* 104, 249-263.
- Mercier, H. and Kerbauy, G.B (1998) Endogenous IAA and cytokinin levels in bromeliad shoots as influenced by glutamine and ammonium nitrate treatments. *Rev. Bras. Fisiol. Veg.* 10:225-228
- Merkle, S. A. and Nairn, C.J. (2005) Hardwood Tree Biotechnology. *In vitro Cell. Dev. Biol. Plant* 41: 602-619
- Meyer, H.J. and Van Staden, J. (1987) Regeneration of *Acacia melanoxylon* plantlets in vitro. *S. Afr. J. Bot.* 53:206-209.
- Midgley, S.J. (2000) *Acacia crassicarpa*: A tree in the domestication fast lane. Australian Tree Resources News No. 6, Australian Tree Seed Centre, CSIRO Forestry and Forest Products, Canberra, Australian Tree Resources News 6:1-2.
- Midgley, S.J. and Turnbull, J.W. (2003) Domestication and use of Australian acacias: case studies of five important species. *Australian Systematic Botany* 16(1):89 - 102
- Minquan, Y. and Yutian, Z. (1991). Results from a four-year- old tropical *Acacia* species/provenance trial on Hainan Island, China. pp. 170-172 in Turnbull, J.W. ed. *Advances in Tropical Acacia Research*. ACIAR. Proceedings No. 35.
- Mishra, D.K. (2009) Selection of candidate plus phenotypes of *Jatropha curcas* L. using method of paired comparisons. *Biomass and Bioenergy* 33:542-545
- Mittal, A., Agarwal, R. and Gupta, S. (1989) In vitro development of plantlets from axillary buds of *Acacia auriculiformis*-a leguminous tree. *Plant Cell Tiss. Organ Cult.* 19:65-70
- Mng'omba, S.A., du Toit, E.S., Akinnifesi, F.K. and Venter, H.M. (2007) Effective preconditioning methods for *in vitro* propagation of *Uapaca kirkiana* Muell Arg. tree species. *African Journal of Biotechnology* 6 (14):1670-1676

- Mohammed, C., Barry, K., Battaglia, M., Beadle, C., Eyles, A., Mollon, A., Pinkard, E., (2000) Pruning-associated stem defects in plantation *E. nitens* and *E. globulus* grown for sawlog and veneer in Tasmania. In: The Future of Eucalypts for Wood Production. Proceedings of IUFRO Conference, Launceston, Australia, pp. 357–364.
- Montes, C.S., Vidaurre, H. and Weber, J. (2003) Variation in stem-growth and branch-wood traits among provenances of *Calycophyllum spruceanum* Benth. from the Peruvian Amazon. *New Forests* 26 (1):1-16
- Monteuuis, O. (1989) Maturation concept and possible rejuvenation of arborescent species. Limits and promises of shoot apical meristems to ensure successful cloning. In *Breeding Tropical Trees: Population Structure and Genetic Improvement Strategies in Clonal and Seedling Forestry*. Proceedings of the IUFRO Conference, Pattaya, Thailand, 28 November–3 December 1988. pp. 106–118.
- Monteuuis, O. (1996) *In vitro* shoot apex micrografting of mature *Acacia mangium* *Agroforestry systems*, 34:213-217.
- Monteuuis, O. and Bon, M.C. (2000) Influence of auxin and darkness on *in vitro* rooting of micropropagated shoots from mature and juvenile *Acacia mangium*. *Plant Cell Tiss. Org. Cult.* 63:173-177.
- Monteuuis, O., (1996) *In vitro* shoot apex micrografting of mature *Acacia mangium*. *Agroforestry Systems* 34:213-217.
- Monteuuis, O., (2004) *In vitro* micropropagation and rooting of *Acacia mangium* microshoot from juvenile and mature origins. *In Vitro Cell.Dev.Biol-Plant* 40:102-107
- Monteuuis, O., Galiana, A. and Goh, D. (2013). *In vitro* propagation of *Acacia mangium* and *Acacia mangium* x *Acacia auriculiformis*. In Lambardi, M. et al. (eds) (2013) Protocol for micropropagation of economically-important horticulture plants, *Methods in Molecular Biology* Vol(994). Springer science + Business media New York.
- Monteuuis, O., Goh, D. (2015) Field growth performances of teak genotypes of different ages clonally produced by rooted cuttings, *in vitro* microcuttings, and meristem culture. *Canadian Journal of Forest Research*, 45 (1): 9-14
- Monteuuis, O., Galiana, A. and Goh, D. (2013). *In vitro* propagation of *Acacia mangium* and *A. mangium* × *A. auriculiformis* In: Lambardi M., Ozudogru, E.A. and Jain, S. M. (eds) *Protocols for Micropropagation of Selected Economically-Important Horticultural Plants*. Volume 994 of the series *Methods in Molecular Biology* :199-211
- Morgante, M. and Vendramin, G.G. (1991) Genetic variation in Italian populations of *Picea abies* (L.) Karst. and *Pinus leucodermis* Ant. In: Muller-Starck, G.,

Ziehe, M. (Eds.), Genetic Variation in European Populations of Forest Trees. J.D. Sauerlander's Verlag, Frankfurt am Main, pp. 205–227

MTIB (2007). Eight Selected Species for Forest Plantation Programme in Malaysia. Handbook of Malaysian Timber Industry Board.

Muller-Starck, G. (1995). Genetic variation in high-elevated populations of Norway spruce (*Picea abies* (L.) Karst.) in Switzerland. *Silvae Genet.*, 44 : 356–362

Muller-Starck, G., Baradat, P. and Bergmann, F. (1992) Genetic variation within European tree species. *New Forests* 6: 23-47.

Mullins, K.V., Llewellyn, D.J., Hartney, V.J., Strauss, S. and Dennis, S.E. (1997) Regeneration and transformation of *Eucalyptus camaldulensis*. *Plant Cell Rep.* 16: 787-791.

Muruganantham, M., Ganapathi, A., Selvaraj, N., Prem R., Vasudevan, A. and Vengadesan, G. (2002) Adenine Sulphate and L-Glutamine Enhance Multiple Shoot Induction from Cotyledon Explants of Melon (*Cucumis melo* L. cv. Swarna). *Cucurbit Genetics Cooperative Report* 25: 22-24.

Naaz, A. and Shahzad, A. and Anis M. (2014) Effect Of Adenine Sulphate Interaction on Growth And Development of Shoot Regeneration and Inhibition of Shoot Tip Necrosis Under *in Vitro* Condition in Adult *Syzygium Cumini* L.- A Multipurpose Tree. *Appl Biochem Biotechnol* 173:90–102

Nagy, J.K., Sule, S. and Sampaio, J.P. (2005) Apple tissue culture contamination by *Rhodotorula* spp.: Identification and prevention. *In Vitro Cell. Dev. Biol.-Plant* 41:520-524.

Naiem, M., Widiyatno and Al-Fauzi, M.Z. (2014) Progeny Test of *Shorea Leprosula* as Key Point to Increase Productivity of Secondary Forest in Pt Balik Papan Forest Industries, East Kalimantan, Indonesia. *Procedia Environmental Sciences* 20: 816-822

Nakamura (2006) Chapter 13. K. Micropropagation of *Shorea roxburghii* and *Gmelina arborea* by shoot-tip Apex culture. Susuki K, Ishii K, Sakurai S, Sasaki S, (eds). Plantation Technology in tropical Forest Science. Springer Verlag, Tokyo.

Nakamura, (2006) Chapter 13. K. Micropropagation of *Shorea roxburghii* and *Gmelina arborea* by shoot-tip Apex culture. Susuki K, Ishii K, Sakurai S, Sasaki S, (eds). Plantation Technology in tropical Forest Science. Springer Verlag, Tokyo.

Nanda, R.M. and Rout, G.R. (2003) *In vitro* somatic embryogenesis and plant regeneration in *Acacia Arabica*. *Plant Cell, Tissue and Organ Culture* 73(2):131-135

- Nandagopal, S. and Ranjitha, K.B.D. (2006) Adenne sulphate induced high frequency shoot organogenesis in callus and in vitro flowering of *Cichorium intybus* L. cv. Focus-a potential medicinal plant. *Acta Agriculturae Slovenica*, 87-2: 415-425.
- Nandwani, D. (1995) *In vitro* micropropagation of a tree legume adapted to arid lands: *Acacia tortilis* subsp. *Radiana*. *Ann. Sc. Forest.* 52:183-189.
- Nandwani, D., Ramawat, K.G. (1993). *In vitro* plantlet formation through juvenile and mature explants in *Prosopis cineraria*. *Ind J Exp Biol* 31:156-160.
- Nangia, S. and Singh, R. (1996) Micropropagation of *Acacia tortilis* hayne (Umbrella thorn) through cotyledonary node culture. *Indian J. Exp. Physiol.* 1(2):77-79.
- National Research Council. (1979) Tropical Legumes: Resources for the Future. National Academy of Sciences, Washington, D.C., USA (*Acacia mangium*, p. 194; *Acacia auriculiformis*, p. 165).
- Nei, M. and Kumar, S. (2000) Molecular Evolution and Phylogenetics. Oxford University Press, New York.
- Nghia, N.H. (2003). Phat trien cac loai Keo Acacia o Viet Nam. Ha Noi: Agriculture Publishing House, 121 p. (in Vietnamese)
- Nichols, J.D., Bristow, M. and Vanclay, J.K. (2006) Mixed species plantations: prospects and challenges. *Forest Ecology and Management*, 233(2-3): 383-390.
- Nicholson, D. 1. (1965) A note on *Acacia auriculaeformis* A. Cunn. ex Benth. in Sabah. *Malayan Forester* 28(3):243-244.
- Nickles, D.G. (1996) The first 50 years of the evolution of forest tree improvement in Queensland. In M.J Dieters, A.C. Matheson, D.G. Nickles, C.E Harwood and S.M Walker, eds. Tree Improvement for sustainable tropical forestry. pp51-64. Proc. QFRI-IUFRO Conference, Columbia, Queensland. Australia
- Niedz, R.P. (1998) Isothiazolone biocides to control microbial and fungal contaminants in plant tissue cultures. *HortTechnol.* 8:598- 601
- Niedz, R.P. and Bausher, M.G. (2002) Control of in vitro contamination of explants from greenhouse- and field-grown trees. *In vitro cell. Dev. Biol.-plant* 38:468-471.
- Nisha, M.C., Rajeshkumar, S., Selvaraj, T. and Subramanian, M.S. (2009) A valued Indian medicinal plant – *Begonia malabarica* Lam.: Successful plant regeneration through various explants and field performance. *Maejo Int. J. Sci. Technol.* 3(02): 261-268
- Nishimoto, Y., Ohnishi, O. and Hasegawa, M. (2003) Topological incongruence between nuclear and chloroplast DNA trees suggesting hybridization in the

urophyllum group of the genus *Fagopyrum* (Polygonaceae). *Genes Genet. Syst.* 78,139–153.

- Nor Aini, A.S., Kamis, A., Mansor, M.R., Abd, L.S., (1994) Provenance trial of *Acacia auriculiformis* in Peninsular Malaysia: 12-month performance. *Journal of Tropical Forest Science* 6, 249 -256.
- Nor Aini, A.S., Nang, A.N. and Awang, K. (1998) Selected wood properties of *Acacia auriculiformis* and *A. crassicarpa* provenances in Malaysia. In Turnbull, J. W., Crompton, H. R. & Pinyopusarek, K. (eds.), *Recent Developments in Acacia planting. Proceedings of an international workshop held in Hanoi, Vietnam, 27-30 October 1997*. ACIAR Proceedings No. 82, Canberra.
- Nurhasyibi, Sudrajat, D.J, Diatna, K. (2009) Identification of *Acacia mangium* provenances for solid wood forest plantation. *Journal of Forestry Research* 6(1):1-16
- Oakley, T.H. and Phillips, R.B. (1999) Phylogeny of Salmonine fishes based on growth hormone introns: Atlantic (*Salmo*) and Pacific (*Oncorhynchus*) salmon are not sister taxa. *Mol. Phyl. Evol.* : 11:381–393
- Ogita, S., Sasamoto, H., Yeung, E.C. and Thorpe, T.A. (2001) The effects of glutamine on the maintenance of embryogenic cultures of *Cryptomeria japonica*. *In Vitro Cell. Dev. Biol. - Plant* 37:268-273.
- Oh, S. and Potter, D. (2003) Phylogenetic utility of the second intron of *LEAFY* in *Neillia* and *Stephanandra* (Rosaceae) and implications for the origin of *Stephanandra*. *Molecular Phylogenetics and Evolution* 29: 203-215.
- Oh, S. and Potter, D. (2005) Molecular phylogenetic systematics and biogeography of Tribe *Neillieae* (Rosaceae) using DNA sequences of CPDNa, RDNa, and *LEAFY*. *Amer. J. Botany* 92(1):179-192.
- Olanaceae, I.O.S. and Aum, D.A.A.B. (2006) Phylogenetics of the Florally Diverse a Ndean, 93(8), 1140–1153.
- Oleksyn, J., Prus-Glowacki, W., Giertych, M., Reich, P.B. (1994) Relation between genetic diversity and pollution impact in a 1912 experiment with East European *Pinus sylvestris* provenances. *Can. J. For. Res.* 24: 2390-2394.
- Orchard, A.E. and Maslin, B.R. (2003). Proposal to conserve the name *Acacia* (Leguminosae: Mimosoideae) with a conserved type. *Taxon* 52: 362-363
- Ortiz, B.O.C., Reyes, M.E.P. and Balch, E.P.M. (2000) Somatic embryogenesis and plant regeneration in *Acacia farnesiana* and *Acacia schaffneri*. *In vitro Cell Dev. Biol. Plant*, 36: 268–272
- Orwa, C., Mutua, A., Kindt, R., Jamnadass, R. and Anthony, S. (2009) Agroforestry Database: a tree reference and selection guide version 4.0. World agroforestry Centre, Kenya.

- Osorio, L.F. (2004) Provenance results of *Gmelina arborea* in southwest Colombia at three years of age. *New Forests* 28 (2-3): 179-185
- Otsamo, A.O., Nikels, D.G. and Vuokko, R.H.O. (1996) Species and provenance variation of candidate acacias for afforestation of Imperata cylindrica grasslands in South Kalimantan. Indonesia. In: M.J.Dieters, A.C.Matheson. C.E.Harwood and S.M.Walker (Eds.). Tree improvement for sustainable tropical forestry. Proc. QFRI IUFRO Conf.. Queensland. Australia. 46- 50.
- Oyebanji, O.B., Nweke, O., Odebunmi, O., Galadima, N.B., Idris, M.S., Nnodi, U.N., Afolabi, A.S. and Ogbadu, G.H. (2009) Simple, effective and economical explant-surface sterilization protocol for cowpea, rice and sorghum seeds. *African Journal of Biotechnology* 8 (20): 5395-5399.
- Pakrashi, A., Ray, H., Pal, B.C. and Mahato, S.B. (1991) Sperm immobilizing effect of triterpene saponins from *Acacia auriculiformis*. *Contraception* 43:475-483.
- Pal, D., Chakraborty, P., Ray, H.N., Pal, B.C., Mitra, D. and Kabir, S.N. (2009) Acaciaside-B-enriched fraction of *Acacia auriculiformis* is a prospective spermicide with no mutagenic property. *Reproduction* 138: 453-462.
- Palma, B., Vogt, G. and Neville, P. (1996) A combined *in vitro/in vivo* method for improved grafting of *Acacia senegal* (L.) Willd. *J. Hort. Sci.* 71:379-381.
- Palmberg, C. (1981) A vital fuelwood gene pool in danger. *Unasylva* 33: 22–30.
- Palumbi, S.R., Baker, C.S. (1994) Contrasting population structure from nuclear intron sequences and mtDNA of humpback whales. *Mol. Biol. Evol.* 11:426–435
- Pan, M.J. and Van Staden, J. (1998) The use of charcoal in *in vitro* culture - A review. *Plant Growth Regulation* 26: 155-163.
- Pan, Z., Lu, P. and Yang, M.C.Y. (1988) Preliminary report on the 3-year-old provenance tests of tropical acacias in China. In: Report on Forest Research, Chinese Academy of Forestry. 553- 558.
- Pan, Z.G. and You, Y.T. (1994) Introduction and provenance test of *Acacia crassicarpa*. *For. Res.* 7: 498-505.
- Park, Y.G. and Son, S.H. (1998) *In vitro* organogenesis and somatic embryogenesis from punctured leaf of *Populus nigra* × *P. maximowiczii*. *Plant Cell Tissue Organ Cult.* 15: 95-105.
- Parker, P.G., Snow, A.A., Schug, M.D., Booton, G.C. and Fuerst, P.A. (1998) What molecules can tell us about populations: choosing and using a molecular marker. *Ecology* 79:361–382.

- Pasqualetto, P.L., Zimmerman, R.H. and Fordham, I. (1988) The influence of cation and gelling agent concentration on vitrification of apple cultivars *in vitro*. *Plant Cell Tiss. Org. Cult.* 14:31-40.
- Pate, J.S., Froend, R.H., Bowen, B.J., Hansen A. and Kuo. J. (1990) Seedling growth and storage characteristics of seeder and resprouter species of mediterranean-type ecosystems of S.W. Australia. *Ann. Bot.* 65: 585-601.
- Patricio, H.P., Castaneto, Y.T., Vallesteros, A.P. and Castaneto E.T. (2006) Macropropagation of *Shorea Guiso* using Stem Cuttings. *Journal of Tropical Forest Science* 18(3): 198-201.
- Pedley, L. (1986). Derivation and dispersal of *Acacia* (Leguminosae), with particular reference to Australia, and the recognition of *Senegalia* and *Racosperma*. *Botanical Journal of the Linnean Society* 92(3) :219-254
- Pence, V.C. (2005) *In vitro* Collecting (IVC). I. The Effect of Collecting Method and Antimicrobial Agents on Contamination in Temperate and Tropical Collections. *In Vitro Cell. Dev. Biol. Plant* 41:324–332.
- Perez-Tornero, O., Egea, J., Olmos, E. and Burgos, L. (2001) Control of hyperhydricity in micropropagated apricot cultivars. *In Vitro Cell. Dev. Biol. Plant* 37: 250-254
- Perrando, E.R. and Corder, M.P.M (2006) Resprouting of *Acacia mearnsii* stumps under different ages, seasons and cut heights. *Pesquisa Agropecuária Brasileira* 41(4): 555-562.
- Phi, H.H. (2009) Genetic improvement of plantation grown *Acacia auriculiformis* for sawn timber production. Doctoral Thesis, Swedish University Agricultural Sciences. 54pp
- Phi, H.H., Jansson, G., Harwood, C., Hannrup, B., Thinh, H.H. (2008) Genetic variation in growth, stem straightness and branch thickness in clonal trials of *Acacia auriculiformis* at three contrasting sites in Vietnam. *Forest Ecology and Management* 255: 156-167
- Picoli, E.A.T., Alfenas, A.C., Gonçalves, R.C., Dias, L.L.C., Neves, D.A., Otoni, W.C. and Romeiro, R.S. (2005) Detection and antibiotic treatment of *Herbaspirillum huttiense* isolated from *in vitro* explants of *Eucalyptus* sp. *Crop Breeding and Applied Biotechnology* 5:191-198
- Pijut, P.M., Beasley, R.R., Lawson, S.S., Palla, K.J., Stevens, M.E. and Wang Y. (2012) *In Vitro* propagation of tropical hardwood tree species-A review (2001-2011). *Propagation of Ornamental Plants* 12: 25-51
- Pinto, G., Silva, S., Park, Y.S., Neves, L., Coutinho, J., Araujo, C. and Santos, C. (2008) Factors influencing somatic embryogenesis induction in *Eucalyptus globulus* Labill.: basal medium and anti-browning agents. *Plant Cell Tiss Organ Cult* 95:79-88

- Pinyopusarerk, K. (1990) *Acacia auriculiformis: an annotated bibliography*. Winrock International and ACIAR, Canberra.
- Poethig, R.S. (1990) Phase change and the regulation of shoot morphogenesis in plants. *Science* 250:923-930.
- Pohlman, C.L., Nicotra, A.B., and Murray, B.R. (2005) Geographic range size, seedling ecophysiology and phenotypic plasticity in Australian *Acacia* species. *Journal of Biogeography* 32(2): 341–351
- Potter, D., Luby, J.J. and Harrison, R.E. (2000) Phylogenetic relationships among species of *Fragaria* (Rosaceae) inferred from non-coding nuclear and chloroplast DNA sequences. *Syst. Bot.* 25:337–348
- Poupard, C., Chauviere, M. and Monteuis, O. (1994) Rooting *Acacia mangium* cuttings: effects of age, within-shoot position and auxin treatment. *Silvae Genet.* 43(4): 226-231
- Prakash, M.G. and Gurumurthi, K. (2010) Effects of type of explant and age, plant growth regulators and medium strength on somatic embryogenesis and plant regeneration in *Eucalyptus camaldulensis*. *Plant Cell Tissue Org Cult.*100(1):13-20.
- Premkumar, G., Sankaranarayanan, R., Jeeva, S. and Rajarathinam, K. (2011) Cytokinin induced shoot regeneration and flowering of *Scoparia dulcis* L.(Scrophulariaceae)-an ethnomedicinal herb. *Asian Pacific Journal of Tropical Biomedicine* 169-172
- Priyanka, S. and Anita, R.G. (2011) Enhanced shoot multiplication in *Ficus religiosa* L. in the presence of adenine sulphate, glutamine and phloroglucinol. *Physiol Mol Biol Plants* 17(3):271-280
- Prychitko, T.M. and Moore, W.S. (1997) The utility of DNA sequences of an intron from the β -fibrinogen gene in phylogenetic analysis of woodpeckers (Aves:Picidae) *Mol. Phyl. Evol.*8:193–204.
- Pua, E.C, Sim, G.E., Chi, G.L., Kong, L.F. (1996) Synergistic effect of ethylene inhibitors and putrescine on shoot regeneration from hypocotyls explants of Chinese radish (*Raphanus sativa* L. var *logipinatus* Bailey) *in vitro*. *Plant Cell Rep.* 15: 685-690.
- Purnhauser, L., Medgyesy, P., Czako, M., Dix, P.I. and Martin, L. (1987) Stimulation of shoot regeneration in *Triticum aestivum* and *Nicotiana plumbaginifolia* tissue cultures assaying the ethylene inhibitors AgNO₃. *Plant Cell Rep.* 6:1-4.
- Qader, N.A., Kumar S.M. and Nor Aini A.S (2014) Multiple shoot induction of selected genotypes of *Eucalyptus camaldulensis* DEHN. *The Malaysian forester* 77 (2): 73-86

- Quoirin, M. da Silva, M.C Martins, K.G. de Olivera. D.E. (2002) Multiplication of juvenile black wattle by microcutting. *Plant Cell Tiss. Org. Cult.*, 66 :199-205
- Rajadurai, D., Rao, A.N. and Loh, C.S. (1987) *In vitro* culture studies on three leguminous species. In Rao, A. N. and Aziah, M. Y. (eds). Proceeding of Culture of Forest Species. Forest Research Institute of Malaysia, Kuala Lumpur, pp. 104-128.
- Rajora, O., Rahman, M.H., Buchert, G.P. and Dancik, B.P. (2000) Microsatellite DNA analysis of genetic effects of harvesting in old-growth eastern white pine (*Pinus strobus*) in Ontario, Canada. *Mol. Ecol.* 9: 339–348.
- Rajora, O.P. (1999) Genetic biodiversity impacts of silvicultural practices and phenotypic selection in white spruce. *Theor. Appl. Genet.* 99: 954–961.
- Ramesh, M., Saravanakumar, R.M. and Pandian, K.S. (2005) Benzyl amino purine and adenine sulphate induced multiple shoot and root induction from nodal explants of Brahmi, *Bacopa monnieri* (Linn.) Penn. *Natural Product Radiance* 5(1): 44-51.
- Randall, C.K., Duryea, M.L., Vince, S.W. and English, R.J. (2005). Factors influencing stump sprouting by pondcypress (*Taxodium distichum* var. *nutans* (Ait.) Sweet). *New Forests*, 29, 245-260.
- Rao, R. V. and Riley. K.W. (1994) The use of biotechnology for the conservation and utilization of plant genetic resources. *Plant Genetic Resources Newsletter* 97:1-17.
- Rao, R.G. V. and Prasad, M.N.V. (1991) Plantlet regeneration from the hypocotyl callus of *Acacia auriculiformis*-multipurpose tree legume. *Plant Physiol.* 137:625-627
- Ravindra, B., Malabadi, and Van Staden, J. (2005) Role of antioxidants and amino acids on somatic embryogenesis of *Pinus patula*. *In Vitro Cell. Dev. Biol. - Plant* 41:181-186.
- Raymond, C.A. (2000) Tree breeding issues for solid wood production. The future of *Eucalyptus* for wood products, IUFRO Conference. Launceston, Australia. 11p.
- Raymond, C.A. (2002) Genetics of *Eucalyptus* wood properties. *Annals of Forest Science* 59 (5-6):525-531
- Read, P.E. and Yang, G. (1991) Influence of plant growth regulators and season on growth responses of woody plant stems. *Acta Horticulture* 300: 173-176.
- Read, P.E., Fellman, C.D., Economou, A.S. and Yang, Q.G. (1986) Programming stock plants for propagation success. *Proc Int Plant Propag Soc* 35:84-91

- Read, P.E., Garton, S., Carlson, C., Conviser, L. and Preece, J. (1979) Programming stock plants for tissue culture success. *Proc Plant Growth Reg Working Group* 6:197-204
- Reddy, B.O., Giridhar, P. and Ravishankar, G.A. (2001) *In vitro* rooting of *Decalepis hamiltonii* Wight and Arn. an endangered shrub by auxins and root-promoting agents. *Current Science* 81(11):1479- 1481.
- Reddy, K.V. and Rockwood, D.L. (1989) Breeding strategies for coppice production in a *Eucalyptus grandis* base population with four generation of selection. *Silvae Genetic* 38: 3-4.
- Reddy, P.C., Veeranagouda, P., Prasad, T .G., Padmak, K., Udayakumar, M. and Patil, V. (1995) *In vitro* axillary bud break and multiple shoot production in *Acacia auriculiformis* by tissue culture technique. *Curr. Sci.* 69:495-496
- Reed, B.M. and Tanprasert, P. (1995) Detection and control of bacterial contaminants of plant tissue cultures. A review of recent literature. *PI. Tissue Culture Biotech.* 1:137-142
- Reed, B.M., Mentzer, J., Tanprasert, P. and Yu, X. (1998). Internal bacterial contamination of micropropagated hazelnut : identification and antibiotic treatment. *Plant Cell Tissue Organ Cult.* 52(1-2): 67-70.
- Regina, M., Eduardo, P. and Helenice, M. (2005) Glutamine enhances competence for organogenesis in pineapple leaves cultivated *in vitro*. *Braz. J. Plant Physiol.* 17(4):383-389
- Reinert, J., Bajaj, Y.P.S. and Zbeli, B. (1977) Aspect of organization organogenesis, embryogenesis, cotydifferentiation, In: H.E. Street (Ed.), *Plant Tissue and Cell culture*, University of California Press, Berkely, pp. 389-429.
- Reinert, J.M., Tazawa, R. and Semeroff, S. (1967) Nitrogen compounds as factors of embryogenesis *in vitro*. *Nature* 216: 1215-1216
- Renuka, C. (2001) Maintenance Of Seed Stands And Species Trial Plots Of Rattans. KPRI Research Report NO. 222.
- Reynolds, P.H.S., Boland, M.J., Blevins, D.G., Schubert, K.R. and Randall, D.D. (1982) Enzymes of amide and ureide biogenesis in developing soybean nodule. *Physiol Plant* 69: 1334-1338
- Richardson, D.M. (1998) Forestry trees as invasive aliens. *Conservation Biol.* 12(1): 18-26.
- Richardson, D.M., Pysek, P., Rejmánek, M., Barbour, M.G., Panetta, F.D. and West, C.J. (2000) Naturalization and invasion of alien plants: concepts and definitions. *Diversity and Distributions* 6(2): 93-107.

- Rinaudo, T. (2001). Utilizing the underground forest. Farmer Managed Natural Regeneration of trees. p. 325-336. In: D. Pasternak and A. Schlissel (eds.), Combating Desertification with Plants. Kluwer Academic/Plenum Publishers, New York.
- Roalson, E.H., Columbus, J.T. and Friar, E.A. (2001) Phylogenetic relationships in Cariceae (Cyperaceae) based on ITS (nrDNA) and *trnT-L* (cpDNA) region sequences: assessment of subgeneric and sectional relationships in *Carex* with emphasis on section *Acrocystis* *Syst. Bot.* 26 : 318–341
- Robbins, W.J. (1960). Further observation of juvenile and adult *Hedera*. *American journal of Botany* 47: 485-491.
- Rockwood, D.L. and Geary, T.F. (1982) Genetic variation in biomass productivity and coppicing of intensively grown *Eucalyptus grandis* in southern Florida. Proc. N. Amer. For. Biol Workshop. pp 400-405.
- Rogler, C.E. and Hackett, W.P. (1975a) Phase change in *Hedera helix* : induction of the mature to juvenile phase change by gibberellin A 3 . *Physiol Plant* 34:141-147
- Rogler, C.E. and Hackett, W.P. (1975b) Phase change in *Hedera helix* : stabilization of the mature form with abscisic acid and growth retardants. *Physiol Plant* 34:148-152
- Romano and Martins-Loucou, M.A. (1992) Micropropagation of Mature Cork-Oak (*Quercus Suber* L.) : Establishment Problems. *Scientia Genmdensis*, 18: 17-27
- Romano, A. and Martins-Loucao, M.A. (1992) Micropropagation of mature cork-oak (*Quercus suber* l.): establishment problems. *Scientia Genmdensis* 18: 17-27.
- Rosier, C.L., Frampton, J., Goldfarb, B., Wise, F.C. and Blazich, F.A. (2005) Stumping height, Crown position and age of parent tree influence rooting of stem cuttings of Fraser Fir. *Hortscience*, 40(3):771-777
- Roth, E.R. and Hepting, G.H. (1943) Origin and development of oak stump sprouts as affecting their likelihood to decay. *J. Forest.* 41: 27-36.
- Roth, E.R. and Hepting, G.H. (1969) Prediction of butt rot in newly regenerated sprout oak stands. *J. Forest.* 67: 756-760.
- Rout, G.R., Senapati, S.K., Aparajeta, S. (2008) Micropropagation of *Acacia chundra* (Roxb.) DC. *Hort. Sci.* (Prague) 35(1): 22-26
- Rout, G.R., Samantaray, S. and Das, P. (1995) Somatic embryogenesis and plant regeneration from callus culture of *Acacia catechu*-a multipurpose leguminous tree. *Plant Cell Tiss. Organ Cult.* 42:283-285.
- Rout, G.R., Senapati, S.K. and Aparajeta, S. (2008) Micropropagation of *Acacia chundra* (Roxb.) DC. *Hort. Sci.* 35(1): 22-26

- Ruredzo, T. J. and Hanson, J. (1993) Plant recovery from seedling derived shoot tips of *Faidherbia albida* grown *in vitro*. *Agrofor. Syst.* 22:59-65
- Ryan, P.A. and Bell, R.E. (1989) Growth, coppicing and flowering of Australian tree species in trials in southeast Queensland, Australia. In trees for the tropics (ed. D.J Boland), pp. 49-68. ACIAR Canberra.
- Saher, S., Piqueras, A., Hellin, E. and Olmos, E. (2004) Hyperhydricity in micropropagated carnation shoots: the role of oxidative stress. *Physiol Plant* 120: 152–161
- Sakai, A. and Sakai, S. (1998) A test for the resource remobilization hypothesis: Tree sprouting using carbohydrates from above-ground parts. *Ann.Bot* 82(2): 213-216
- Sakai, A., Sakai, S. and Akiyama, F. (1997) Do sprouting tree species on erosion-prone sites carry large reserves of resources? *Ann. Bot.* 79: 625-630.
- Sakai, A., Sakai, S. Ohsawa, T. and Ohsawa, M. (1995) Adaptive significance of sprouting of *Eupteleapolyandra*, a deciduous tree growing on steep slopes with shallow soil. *J. Pl. Res.* 108: 377-386.
- Salin, M.L. and Bridges, S.M. (1981) Chemiluminescence in wounded root tissue. *Plant Physiol.* 67: 43-46
- Samake, G., Folega, F., Senou, H. and Wang, H.F. (2011) *In vitro* regeneration of *Acacia nilotica* (L.) Willd. ex Del from nodes on B5 medium. *Journal of Agricultural Biotechnology and Sustainable Development* 3(5): 85-89.
- Sanchez, A. and Kron, K.A. (2008) Phylogenetics of *Polygonaceae* with an Emphasis on the Evolution of *Eriogonoideae*. *Systematic Botany*, 33(1): 87–96.
- Sanchez, M.C., San-Jose, M.C., Ballester, A. and Vieitez, A.M.(1996) Requirements for *in vitro* rooting of *Quercus robur* and *Q. rubra* shoots derived from mature trees *Tree Physiology* 16: 673-680
- Sander, I.L. (1971) Height growth of new oak sprouts depends on size of advance reproduction. *J. Forest.* 69: 809-811.
- San-Jose, M.C., Ballester, A. and Vieitez, A.M. (1988) Factors affecting *in vitro* propagation of *Quercus robur* L. *Tree Physiol.* 4:281-290.
- Sanker, A., Libin mary, S., Vijaykumar, A., Karthi, Rani R., Raja Selvam, J., Kohila, R., Liby, I., Vadivukarasi, S., and Ganesh, D. (2008). Phloroglucinol enhances shoot proliferation in nodal explants of *Vanilla planifolia* Andr. *J Plantn Crops.* 36: 127-131
- Santana, J.R.F.D., Paiva, Renato, Aloufa, M.A.I. and Lemos, E.E.P.D. (2003) Efficiency of ampicillin and benomyl at controlling contamination of

Annonaceae leaf segments cultured *in vitro*. Research article ,*Fruits* 58 (6):357-361

- Sanyal, I., Singh, A.K., Kaushik, M. and Amla, D.V. (2005) *Agrobacterium* mediated transformation of chickpea (*Cicer arietinum* L.) with *Bacillus thuringiensis* cry1Ac gene for resistance against pod borer insect *Helicoverpa armigera*. *Plant Sci.* 168: 1135-1146.
- Sastroamidjojo, J.S. (1964) *Acacia auriculiformis* A. Cunn. *Rimba Indonesia* 9(3):214-225.
- Schlotterer, C. (2004) The evolution of molecular markers—just a matter of fashion? *Nature Reviews Genetics* 5:63–69.
- Schluter, P.M., Kohl, G., Stuessy, T.F. and Paulus, H.F. (2007) A screen of low-copy nuclear genes reveals the, 56: 493–504.
- Schmidt, E.D.L., Guzzo, F., Toonen, M.A.J. and Devries, S.C. (1997) A leucine-rich repeat containing receptor-like kinase marks somatic plant cells competent to form embryos. *Development* 124: 2049-2062
- Scholten, H.J. and Pierik, R.L.M. (1998) Agar as a gelling agent: chemical and physical analysis. *Plant Cell Rep.* 17: 230-235
- Scotti, I., Vendramin, G.G., Matteotti, L.S., Scarponi, C., Sari-Gorla, M. and Binelli, G. (2000) Postglacial recolonization routes for *Picea abies* K. in Italy as suggested by the analysis of sequence-characterized amplified region (SCAR) markers. *Mol. Ecol.* 9:699-708.
- Scowcroft, P.G., Friday, J. B., Haraguchi J., Idol, T. and Dudley, N.S. (2010) Poor Stem Form as a Potential Limitation to Private Investment in Koa Plantation Forestry in Hawaii. Small-scale Forestry, Research Article
- Sein, C.C. and Mitlohner, R. (2011) *Acacia* hybrid: ecology and silviculture. CIFOR, Bogor, Indonesia.
- Selby, C., Watson, S. and Harvey, B.M.R. (2005) Morphogenesis in Sitka spruce (*Picea sitchensis* (Bong.) Carr.) bud cultures – tree maturation and explants from epicormic shoots. *Plant Cell, Tissue and Organ Culture*, 83: 279-285
- Semsuntud, N. and Nitiwattanachai, W. (1991) Tissue culture of *Acacia auriculiformis*. *Adv. Trop. Acacia Res. Proceedings Series* 35:39-42.
- Senin, A.L., Hamid, H.A., Kusno, M.A., Abdu, A. and Ismail, M.K. (2011) Comparative growth of 11 year old *Acacia aulocarpa* A. CUNN.EX. Benth from four provenances. *Research Journal of Forestry.* 5:154-161
- Shackleton, C.M. (2001) Managing regrowth of an indigenous savanna tree species (*Terminalia sericea*) for fuelwood: the influence of stump dimensions and post-harvest coppice pruning. *Biomass and Bioenergy* 20(4): 261–270

- Shahana, S. and Gupta. S.C. (2000) Somatic embryogenesis in a leguminous tree *Acacia senegal* (L) Willd, In: S.M. Jain, P.K.Gupta, R.J.Marton (Eds.), Somatic embryogenesis in woody plants 6: 539-552.
- Sharma, S., Shahzad, A., Ahmad, A. and Anjum, L. (2014) *In vitro* propagation and the acclimatization effect on the synthesis of 2-hydroxy-4-methoxy benzaldehyde in *Decalepis hamiltonii* Wight and Arn. *Acta Physiol Plant* 36:2331-2344
- Shaw, A.J., Cox, C.J. and Boles, B. (2003) Polarity of Peatmoss (*Sphagnum*) evolution: Who says Bryophyte have no roots? *American Journal of Botany* 90: 1777-1787.
- Shaw, J, Lickey E.B., Beck,, J.T., Farmer, S.B., Liu, W., Miller, J., Siripun, K.C., Winder, C.T., Schilling, E.E. and Small, R.L. (2005) The tortoise and the hare II: Relative utility of 21 noncoding chloroplast DNA sequences for phylogenetic analysis. *American. J. Bot.* 92:142–166.
- Shetty, K., Asano, Y. and Oosaka, K. (1992) Stimulation of *in vitro* shoot organogenesis in *Glycine max* (Merrill.) by allantoin and amides. *Plant Sci.* 81:245-251.
- Shields, R, Robinson, S.J., Anslow, P.A. (1984) Use of fungicides in plant tissue culture. *Plant Cell Rep.* 3: 33-36.
- Shrivastava, S. and Banerjee, M. (2008) *In vitro* clonal propagation of physic nut (*Jatropha curcas* L.): influence of additives. *International Journal of Intergerity Biology* 3:73 - 79.
- Shu, G.P., Amaral, W., Hileman, L.C. and Baum, D.A., (2000) *LEAFY* and the evolution of rosette flowering in violet cress (*Jonopsidium acaule*, Brassicaceae). *Am. J. Bot.* 87: 634-641.
- Shukla, S.R., Rao, R.V., Sharma, S.K., Kumar, P., Sudheendra, R., Shashikala, S. (2007) Physical and mechanical properties of plantation-grown *Acacia auriculiformis* of three different ages. *Australian Forestry* 70: 86-92.
- Simpson, J. and Osborne, D. (2006) Performance of seven hardwood species underplanted to *Pinus elliottii* in south-east Queensland. *Forest Ecology and Management* 233(2 / 3): 303- 308.
- Singh, H.P., Singh, S., Saxena, R.P., Singh, R.K. (1993) *In vitro* bud break in axillary nodal segments of mature trees of *Acacia nilotica*. *Indian J. Plant Physiol.* 36:21-24
- Singh, P. and Patel, R.M. (2014). Factors Influencing *in Vitro* Growth and Shoot Multiplication of Pomegranate. *The bioscan*, 9(3): 1031-1035

- Singha, S., Townsend, E.C. and Oberly, G.H. (1990) Relationship between calcium and agar on vitrification and shoot-tip necrosis of quince (*Cydonia oblonga* Mill.) shoots in vitro. *Plant Cell, Tissue and Organ Culture* 23: 135-142.
- Singhal, P.C and Joshi, L.D. (1984) Role of Gum Arabica and Gum catechu in glycemia and Cholesterolemia. *Curr.Sci.*53:91.
- Sivanesan, I. and Jeong, B.R. (2007) Micropropagation and *in vitro* flowering in *Pentanema indicum* Ling. *Plant Biotechnology* 24:527-532
- Sivanesan, I. and Jeong, B.R. (2009) Micropropagation of *Plumbago zeylanica* L. *African Journal of Biotechnology* 8 (16): 3761-3768.
- Siwach P. and Gill, A.R. (2011). Enhanced shoot multiplication in *Ficus religiosa* L. in the presence of adenine sulphate, glutamine and phloroglucinol. *Physiol Mol Biol Plants* 17(3): 271-280.
- Siwach, P., Chanana, S., Gill, A.R., Dhanda, P., Rani, J., Sharma, K., Rani, H. and Kumari, D. (2012) Effects of adenine sulphate, glutamine and casein hydrolysate on *in vitro* shoot multiplication and rooting of Kinnow mandarin (*Citrus reticulata* Blanco) *African Journal of Biotechnology*, 11(92): 15852-15862
- Skirvin, R.M., Motoike, S., Norton, M.A., Ozgur, M., Al-Juboory, K. and Mcmeans, O.M. (1999) Workshop on micropropagation – establishment of contaminant-free perennial plants in vitro. *In Vitro Cellular & Developmental Biology-Plant* 35: 278-280
- Skolmen, R.G. (1986) *Acacia* (*Acacia koa* Gray), In: Y.P.S. Bajaj (Ed.), *Biotechnology in Agriculture and Forestry*, vol. Tree 1, Springer, Berlin, pp. 375-383.
- Skolmen, R.G. and Mapes, O.M. (1976) *Acacia koa* Gray plantlets from somatic callus tissue. *J. Hered.* 67:114-115
- Slade, R.W., Moritz, C., Heideman, A., Hale, P.T. (1993) Rapid assessment of single-copy nuclear DNA variation in diverse species. *Mol. Ecol.* 2:359–373.
- Smith, S.D. and Baum, D.A (2006) Phylogenetics of the florally diverse andean clade Iochrominae (solanaceae). *American Journal of Botany* 93(8): 1140–1153.
- Solomon, D.S. and Blum, B.M. (1967) Stump sprouting of four northern hardwoods. U.S. Forest Serv. Res. Pap. NE-59.
- Songstad, D.D, Duncan, D.R., Widholm, J.M. (1988) :Effect of l-aminocyclopropane, l-carboxylase acid, silver nitrate and norbornadine on plant regeneration from maize callus cultures *Plant Cell Rep.* 7:262-265
- Sperisen, C., Buchler, U., Gugerli, F., Matyas, G., Geburek, T. and Vendramin, G.G. (2001) Tandem repeats in plant mitochondrial genomes: application to the

- analysis of population differentiation in the conifer Norway spruce. *Mol. Ecol.* 10: 257–263.
- Spooner, D., van Treuren, R. and de Vicente, M.C. (2005) Molecular Markers for Genebank Management, IPGRI Technical Bulletin No. 10, 2005.
- Sprent, J.I. (1980) Root nodule anatomy, type of export product and evolutionary origin of some Leguminosae. *Plant Cell Environ* 3:35-43
- Srivastava, P.B.L. (1993) Silvicultural practices. In: Awang, K. and Taylor, D. ed., *Acacia mangium, growing and utilization*. FAO, Bangkok, Thailand, and Winrock International, MPTS Monograph Series No. 3, 113–147
- Staden, J.V., Zazimalova, E. and George, E.F. (2008) Plant growth regulators II, Plant Propagation by Tissue Culture: Cytokinins, their analogues and antagonist. In George E F, Hall M, De Kleck GJ (Eds) Academic press, Netherland, pp. 205-226.
- Steele, P.H. (1984) Factors determining lumber recovery in sawmilling. Madison, Wisconsin: United States Department of Agriculture, Forest Service, Forest Products Laboratory. General Technical Report- FPL-39, 10p.
- Stephen, M., Nagarajan, S. and Ganesh, D. (2010) Phoroglucinol and silver nitrate enhances axillary shoot proliferation in nodal explants of *Vitex negundo* L.- an aromatic medicinal plant. *Iran J Biotechnol* 8(2):82–89
- Steinmacher, D.A., Cangahuala-inocente, G.C., Clement, C.R. and Guerra, M.P. (2007) Somatic embryogenesis from peach palm zygotic embryos. *Plant Cell Tissue and Organ Culture*, 43(2):124-132.
- Stewart, F.C., Mapes M.O. and Mears. K.(1958) Growth and organized development of cultured cells. *Am. J. Bot.* 45: 705.
- Stoutmayer, V.T., Britt, O.K. and Doogin, J.K. (1961). The influenced of chemical treatment understocks and environment on growth phase changes and propagation of *Hedera canariensis*. *Proceeding of American Society of Horticulture Science* 77: 552-557
- Strauss, S.H., Lande, R. and Namkoong, G. (1992) Limitation of Molecular Marker-aided selection in forest tree breeding. *Can.J.For.Res*, 22:1050-1061.
- Strosse, H., Van den Houwe I., and Panis, B. (2004) Banana cell and tissue culture – review. In: Jain SM, Swennen R (eds.) *Banana Improvement: Cellular, Molecular Biology, and Induced Mutations*, Science Publishers, Inc., Enfield, NH, USA. pp. 1-12
- Sudarsana, R.G.V., Chandra, R. and Polisetty, R. (2001) Role of amino acids in evolution of ethylene and methane, and development of microshoots in *Cajanus cajan*. *Biol. Plant.* 44(1):13-18.

- Sukartiningsih, Nakamura, K. and Ide, Y. (1999) Clonal propagation of *Gmelina arborea* Roxb. by *in vitro* culture. *Journal of Forest Research* 4: 47-51.
- Sunandakumar, C., Martin, K.P., Chithra, M. and Madhusoodanan, P.V. (2004) Silver nitrate induced rooting and flowering *in vitro* on rare rheophytic woody medicinal plant, *Rotula aquatica* Lour. *Indian Journal of Biotechnology*, 3(3):418-421
- Susanto, M., Prayitno, T.A. and Fujisawa, Y. (2008) Wood Genetic variation *Acacia auriculiformis* at Wonogiri trial in Indonesia. *Journal of Forestry Research* 5(2): 135-145.
- Takeda, T., Mizukami, M. and Matsuoka, H. (2008) Characterization of two-step direct somatic embryogenesis in carrot. *Biochemical Engineering Journal* 38:206-211.
- Tamura, K. and Nei, M. (1993) Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Molecular Biology and Evolution* 10:512-526.
- Tamura, K., Stecher, G., Peterson, D., Filipski, A. and Kumar, S. (2013) MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Molecular Biology and Evolution* 30: 2725-2729.
- Tank, S.D.C. and Sang, T. (2001) Phylogenetic utility of the glycerol-3-phosphate acyltransferase gene: evolution and implications in *Paeonia* (Paeoniaceae). *Mol. Phylogenet. Evol.* 19: 421-429
- Thimmappaiah, S.R.A. and Sadhana, P.H. (2002) *In vitro* propagation of cashew from young trees. *In Vitro Cell Dev Boil- Plant* 38:152-156
- Titon, M., Xavier, A., and Otoni, W.C. (2006) Clonal propagation of *Eucalyptus grandis* using the mini-cutting and micro-cutting techniques. *Cientia Forestalis* 71, 109-117.
- Tiwari, V., Tiwari, K.N. and Sing, B.D. (2006) Shoot bud regeneration from different explants of *Bacopa monniera* (L.) Wettst. by trimethoprim and bavistin. *Plant Cell Reports* 25: 629 - 635.
- Toda, T., Tajima, M. and Brini, P.B. (1995) Tissue culture of *Acacia mangium* and *Acacia auriculiformis* and *Acacia* hybrid. *Bull. Natl Tree Breed. Centre* 13:81-94
- Torrey, J.G. (1985). The development of plant biotechnology. *Anter Scientist* 73: 354-363
- Toth, K., Haapala, T., and Hohtola, A. (1994) Alleviation of browning in oak explants by chemical pretreatments. *Biol. Plant.* 36: 511-517.

- Trindade, H. and Pais, M.S. (1997). *In vitro* studies on *Eucalyptus globulus* rooting ability. *In Vitro Cellular & Developmental Biology - Plant* 33(1):1-5
- Tripathi, R.K. and Ram S. (1982) Induction of growth and differentiation of carrot callus cultures by carbendazim and benzimidazole. *Indian Journal of Experimental Biology*, 20:674-677
- Tripathi, R.K., Tandon, K., Schlosser, E. and Hess, W.M. (1982) Effect of fungicides on the physiology of plants, IV: Protection of cellular organelles of senescent wheat leaves by carbendazim. *Pest Sci.* 13: 395 - 400
- Tsay, H.S., Lee, C.Y., Agrawal, D.C. and Basker, S. (2006) Influence of ventilation closure, gelling agent and explant type on shoot bud proliferation and hyperhydricity in *Scrophularia yoshimurae*- a medicinal plant. *In vitro cell. Dev. Biol.-plant* 42:445-449
- Tu, T., Dillon, M.O., Sun, H. and Wen, J. (2008) Phylogeny of *Nolana* (Solanaceae) of the Atacama and Peruvian deserts inferred from sequences of four plastid markers and the nuclear *LEAFY* second intron. *Molecular Phylogenetics and Evolution*, 49(2): 561–573.
- Tumbull, J.W. (1988) Australian acacias in world forestry. Paper at the Bicentennial Forestry Conference. Albury. Australia 25 April- 1 May. 1988. 15p.
- Turnbull, J.W, Martensz, P.N. and Hall. N. (1986) Notes on lesser known Australian trees and shrubs with potential for fuelwood and agroforestry. In Turnbull J.W (ed) Multipurpose Australian trees and shrubs; lesser known Australian trees and shrubs with potential for fuelwood and agroforestry ACIAR Monograph No. 1: 81-313.
- Turnbull, J.W. (1989) Australian trees for reforestation of wastelands. In Proceedings of an FAD Regional Workshop on Wasteland Development for Fuelwood and Other Rural Needs. Vadodara, Gujarat, India, November 1988.
- Turnbull, J.W., Midgley, S. J., Cossalter, C. (1998) Tropical acacias planted in Asia: an overview. In: Turnbull, J. W., Crompton, H. R. and Pinyopusarerk, K. (eds.). Recent developments in acacia planting: proceedings of an international workshop held in Hanoi, Vietnam, 27 - 30 October 1997. :14-28.
- Turvey, N.D. (1996) Growth at age 30 months of Acacia and Eucalyptus species planted in Imperata grasslands in Kalimantan Selatan. Indonesia. *For. Ecol and Mange.* 82 (1- 3): 185- 195.
- Tyagi, A.K., Rashid, A. and Maheshiwari, S.C. (1981) Promotive effect of polyvinylpyrrolidone on pollen embryogenesis in *Datura innoxia*. *Physiol. Plant.* 53: 405-406.
- USDA FPL. (2011). Center for Wood Anatomy Research, Madison, Wisconsin. Access: [http:// www.fpl.fs.fed.us/research/centers/woodanatomy/index.php](http://www.fpl.fs.fed.us/research/centers/woodanatomy/index.php)

- Valdez-ortiz, A., Medina-godoy, S., Valverde, M.E. and Paredes-lopez, O. (2007) A transgenic tropical maize line generated by the direct transformation of the embryo-scutellum by *A. tumefaciens*. *Plant Cell Tissue and Organ Culture*, 91(3): 201-214
- Van Oppen, M.J., Willis, B.L., Vugt, H.W., Miller, D.J. (2000). Examination of species boundaries in the *Acropora cervicornis* group (Scleractina, Cnidaria) using nuclear DNA sequence analyses. *Mol Ecol*.9:1363–1373
- Van Stedan, J., Zazimalova, E., George, E.F. (2008) Cytokinins, their analogues and antagonist. In: George E.F., Hall M, Delckle G.J. (eds) *Plant Propagation by tissue culture. The background. Plant growth regulators II*, vol 1. Springer, The Netherlands, pp 205–226
- Vanderschaeghe, A.M. and Debergh, P.C. (1987) Technical aspects of the control of the relative humidity in tissue culture containers. In: Ducate G, Jacob M, Simeon A (eds) *Plant Micropropagation in Horticultural Industries* (pp 68-76). Presses Universitaires, Liege
- Vargas, C.F., Lopez, A., Sanchez, H. and Rodriguez, B. (2002) Allozyme analysis of host selection by bark beetles in central Mexico. *Can. J. For. Res.* 32: 24–30.
- Varshney, A. and Anis, M. (2014) *Trees: Propagation and Conservation: Biotechnological Approaches for Propagation of a Multipurpose Tree, Balanites aegyptiaca Del.* Springer India, New Delhi
- Vasudevan, A., Selveraj, N., Ganapathi, A., Kasthurienga, A.R.V. and Manickavasagam, M. (2004) Glutamine: a suitable nitrogen source for enhanced shoot multiplication in *Cucumis sativus* L. *Biol Plant* 48(1):125-128
- Vendramin, G.G., Anzidei, M., Madaghiele, A., Sperisen, C., Bucci, G., (2000) Chloroplast microsatellite analysis reveals the presence of population subdivision in Norway spruce (*Picea abies* K.) *Genome* 43: 68–78.
- Vengadesan, G. Ganapathi, A. Ramesh Anbazhagan, V. Prem Anand R. (2002a). Somatic embryogenesis in cell suspension cultures of *Acacia sinuata* (Lour.) Merr. *In vitro Cell Dev. Biol. Plant* 38: 52–57
- Vengadesan, G., Ganapathi, A., Pream, A.R. and Selvaraj, N. (2003) *In vitro* propagation of *Acacia sinuata* (LOUR) MERR. from nodal segments of a 10-year-old tree. *In vitro Cell. Dev. Biol-Plant* 39:409-414.
- Vengadesan, G., Ganapathi, A., Prem Anand, R. and Anbazhagan R.V. (2000) *In vitro* organogenesis and plant formation in *Acacia sinuata*. *Plant Cell Tiss. Org. Cult.* 61: 1–6
- Vengadesan, G., Ganapathi, A., Prem Anand, R., Anbazhagan, R.V. (2002b) *In vitro* propagation of *Acacia sinuata* (Lour.) Merr. via cotyledonary node. *Agroforestry Syst.* 55:9–15

- Verma, M. and Bansal, Y.K. (2013) Effect of Additives on Plant Regeneration of *Hedychium coronarium* In J. Koenig an Endangered Aromatic and Medicinal Herb. *Int. J. Pharm. Sci. Rev. Res.* 23(1); 105-110
- Vieitez, A.M., Corredoira, E., Ballester, A., Munoz, F., Duran, J. and Ibarra, M.(2009) *In vitro* regeneration of the important North American oak species *Quercus alba*, *Quercus bicolor* and *Quercus rubra*. *Plant Cell Tiss Organ Cult* 98:135-145
- Vollmer, S.V. and Palumbi, S.R. (2004). Testing the utility of internally transcribed spacer sequences in coral phylogenetics. *Mol. Ecol.* 14:2763–2772
- Von, A.S and Eriksson, T. (1984) Effect of agar concentration on growth and anatomy of adventitious shoots of *Picea abies* (L) Karst. *Plant Cell Tiss. Org. Cult.* 3: 257-264
- Wachira, F. (1997) *In vitro* shoot multiplication of *Eucalyptus grandis*. *African Crop Science Journal*, 5(3): 239-251
- Wann, S.R., Johnson, M.A., Noland, T.L. and Carlson, J.A. (1987) Biochemical differences between embryogenesis and non-embryogenesis cells of *Picea abies* (L) Karnst, *Plant Cell Rep.* 6.39-42.
- Wantanabe, Y., Ide, Y. and Ikeda, A. (1994). Plant regeneration from axillary bud culture of one year old seedling of *Acacia auriculiformis* grown in green house. *Bull. Tokyo Univ. For.* 92:29-35
- Warrag. E.I., Lesney, M.S., Rockwood, D.L. (1990) Micropropagation of field tested superior *Eucalyptus grandis* hybrids. *New Forests* 4:67-79
- Waugh, G., (1996) Properties of plantation grown Eucalypts. In: Farm Forestry and Plantations: Investing in Future Wood Supply. Australian Forest Growers Conference, Mt. Gambier, Australia, pp. 83–93.
- WCED, (1988) Our future. World Commission on Environment and Development. Oxford University Press.400p.
- Weber J C., Carmen Sotelo Montes 2008. Geographic variation in tree growth and wood density of *Guazuma crinita* Mart. in the Peruvian Amazon. *New Forests* 36, Issue 1, pp 29-52
- Webster, S., Mitchell, S.A. and Ahmad, M.H. (2003) A novel surface sterilization method for reducing microbial contamination of field grown medicinal explants intended for *in vitro* culture. Biotechnology Centre, U.W.I, Mona, Kingston 7, Jamaica, West Indies. Paper in: Proceedings of 17th SRC conference entitled ‘Science and Technology for Economic Development: Technology Driven Agriculture and Agro-Processing’ SRC, Jamaica.
- Wei, X.X., Yang, Z.Y., Li,Y. and Wang, X.Q. (2010) Molecular phylogeny and biogeography of *Pseudotsuga* (*Pinaceae*): Insights into the floristic

relationship between Taiwan and its adjacent areas. *Molecular Phylogenetics and Evolution*, 55(3): 776–785.

Weinland, G. and Zuhaidi, A.Y. (1991) Management of *Acacia mangium* stands: tending issues. In: Appanah, S., Ng, F.S.P., Ismail, R. (Eds.), Malaysian Forestry and Forest Products Research Proc

Wendel, G.W. (1975) Stump sprout growth and quality of several Appalachian hardwood species after clear cutting. U.S. Forest Sere. Res. Pap. NE-329.

Whitehouse, A.B., Marks, T.R. and Edwards, G.A. (2002) Control of hyperhydricity in *Eucalyptus* axillary shoot cultures grown in liquid medium. *Plant Cell Tiss. Org. Cult.* 71: 245-252

Widiarti, A. and Alrasjid, H. (1987). Introduction of fuelwood trees species on degraded lands in Paseh and Kadipaten areas (Indonesia). *Buletin Penelitian Hutan* (488):1-17.

Widiyatno, Naiem, M., Purnoma, S. and Jatmoko, (2014) Evaluation of Four year old progeny test of *Shorea macrophylla* in PT Sari Bumi Kusuma, Central Kalimantan. *Procedia Environmental Sciences* 20: 809-815.

Williams, E.G., Maheswaran, G. and Hutchinson, J.F. (1987) Use of embryo culture in plant breeding *Plant Breeding Reviews* 181-236.

Williams, E.R. and Matheson. (1994) Experimental design and analysis for use in tree improvement. Australia:CSIRO Publication Service.174pp

Wittenbach, V.A. and Bukovac, M.J. (1980) *In vitro* culture of sour cherry fruits. *J Am Soc Hortic Sci* 105: 277-279

Won, H. and Renner, S.S. (2005) The internal transcribed spacer of nuclear ribosomal DNA in the gymnosperm Gnetum. *Molecular Phylogenetics and Evolution*, 36(3):581–597.

Wong, C.Y. and Wijoyo, F.S. (2005) The Breeding strategy of *Acacia mangium* and *Acacia crassiparva* at PT Riau Andalan Pulp and Paper. *Acacia research in Malaysia*. Proceeding on a seminar on current updates on *Acacia* Genomics and Breeding. 12 July 2005. Marriott Hotel, Putrajaya, Malaysia. 50-54.

Woo, K.C., Montagu, K.D., Metcalfe, A.J., Puangchit, L., Luangviriyasaeng, V., Jiwarawa, P. and Changtragoon, S. (1997) Genetic Improvement and Physiology of *Acacia auriculiformis*. In: The third international *Acacia* Workshop. Hanoi, Vietnam, 26-30 October 1997. pp.292-298.

Wright, J.W. (1976) Introduction to forest genetics. Academic, New York, p 463

Wroblewska, K. (2012). The Influence of Adenine and Benzyladenine on Rooting and Development of *Fuchsia Hybrida* Cuttings. *Acta Agrobotanica* 65 (4): 101–108

- Wu, L., Shinzato, T., Chen, C. and Aramoto, M. (2008). Sprouting characteristics of a subtropical evergreen broad-leaved forest following clear-cutting in Okinawa, Japan. *New Forests*, 36: 239-246.
- Xie, D.Y. and Hong, Y. (2001a) *In vitro* regeneration of *Acacia mangium* via organogenesis. *Plant Cell Tiss. Org.* 66:167-173.
- Xie, D.Y. and Hong, Y. (2001b) Regeneration of *Acacia mangium* through somatic embryogenesis. *Plant Cell Rep.* 20:34-40.
- Xue, Y., Zhang, W., Zhou, J., Ma, C. and Ma, L. (2013) Effects of stump diameter, stump height, and cutting season on *Quercus variabilis* stump sprouting. *Scandinavian Journal of Forest Research* 28(3): 223-231
- Yanchuk, A.D. (2001) The role and implication of biotechnological tools in Forestry. *Unasylva* 204(52): 53-61.
- Yang, J., Chung, J. and Chen, Z. (1995) Vegetative propagation of adult *Eucalyptus grandis* X *urophylla* and comparison of growth between micropropagated plantlets and rooted cuttings. *Plant Cell Reports* 15:170-173
- Yang, J., Ho, C., Chang, S. and Chen, Z. (1996) Micropropagation of Mature Trees of *Eucalyptus Camaldulensis* with Fast-Growing Phenotype . *Taiwan Journal of Forest Science* 11(4) : 421-431
- Yang, J., Yoon, H.S. and Pak, J.H. (2012) Phylogeny of Korean *Rubus* (Rosaceae) based on the second intron of the *LEAFY* gene. *Canadian Journal of Plant Science*, 92(3): 461–472.
- Yang, M., Xie, X., He, X. and Zhang, F. (2006) Plant regeneration from phyllode explants of *Acacia crassicarpa* via organogenesis. *Plant Cell Tiss. Org.* 85:241-245.
- Yang, M., Xie, X., He, X., Zheng, F., Zhang, F. and Li, Z. (2008) *Agrobacterium tumefaciens*-mediated genetic transformation of *Acacia crassicarpa* via organogenesis. *Plant Cell Tiss. Org.* 95:141-147.
- Yang, M.Q., Zhang, F.Q., Xue, H.Z., Wu, Z.Y., Lin, K.L. and Luo, W.X. (1995) Study on selection of *Acacia* species/provenances in water and soil serious erosion area. *For. Res.* 8:489-496.
- Yao, H.Y., Lin, E.K., Wang, C.W., Yu, Y.C., Chang, C.H., Yang, Y.C. and Chang, C.Y. (1996) A PIXE study of vitrification of carnation in vitro culture. *Nuclear Instruments and Methods in Physics Research* 109/110:312-317.
- Yasodha, R., Sumathi, R. and Gurumurthi, K. (2004) Micropropagation of quality propagule production in plantation forestry. *Indian J. Biotechnol.* 3:159-170.
- Yingzhi, L., Yunjiang, C., Nengguo, T. and Xiuxin, D. (2007) Phylogenetic Analysis of Mandarin Landraces, Wild Mandarins, and Related Species in China Using

Nuclear *LEAFY* Second Intron and Plastid trnL-trnF Sequence. *J. Japan. Soc. Hort. Sci.*, 132(6): 796–806.

- Yu, Y., Wang, J., Zhu, M.L. and Wei, Z.M. (2008) Optimization of matured embryo-based high frequency callus induction and plant regeneration from elite wheat cultivars grown in China. *Plant Breeding* 127(3):249-255.
- Zakaria, I., Wan Razali, W.H., Hashim, M.N., Lee, S.S., (1994) The incidence of heartrot in *Acacia mangium* plantations in Peninsula Malaysia. Forest Research Institute Malaysia Research Pamphlet No. 114. 15 pp.
- Zapartan, M., (2001) Conservarea florei spontane prin înmulțire *in vitro*, Ed. ALC Media Group, Cluj – Napoca, pp.119 – 122
- Zashimuddin, M., Latif, M.A., Khan, S.A. and Davidson, J. (1983) Performance of different provenances of *Acacia mangium* in Bangladesh. *Bano Biggyan Patrika*, 12(1&2):57-61.
- Zhang, D.H. and Hewitt, G.M. (2003) Nuclear DNA analyses in genetic studies of populations: practice, problems and prospects. *Mol. Ecol.* 12:563–584.
- Zhang, F.Q. and Yang, M.Q. (1996) Comprehensive selection of provenances and families of *Acacia crassicarpa*. In: M.J.Dieters. A.C.Matheson. C.E.Harwood and S.M.Walker (Eds.). Tree improvement for sustainable tropical forestry. Proc. QFRI- IUFRO Conf. Queensland. Australia. 401- 403.
- Zhang, W., Kan, S., Zhao, H., Li, Z. and Wang, X. (2014) Molecular Phylogeny of Tribe *Theaeae* (Theaceae) and Its Implications for Generic Delimitation. *PLoS ONE*, 9(5): 98-133.
- Zhao, Y.X., Yao, D.Y. and Harris, P.J.C. (1990) *In vitro* regeneration of plantlets from explants and callus of *Acacia salicina*. *Nitrogen Fixing Tree Res. Rep.* 8:113-115.
- Zheng, X., Hu, C., Spooner, D., Liu, J., Cao, J. and Teng, Y. (2011) Molecular evolution of *Adh* and *LEAFY* and the phylogenetic utility of their introns in *Pyrus* (Rosaceae). *BMC Evolutionary Biology*, 11(1): 255.
- Zibbu, G. and Batra, A. (2010) Effect of Adenine Sulphate on Organogenesis Via Leaf Culture in An Ornamental Plant: *Thevetia Peruviana* (Pers.) Schum. *International Journal Of Pharma And Bio Sciences* V1(2)2010
- Zimmerman, L.J. (1993) Somatic embryogenesis: a model for early development in higher plants. *Plant Cell* 5: 1411–1423.
- Zimmerman, T.W. and Cobb, B.G. (1989) Vitrification and soluble carbohydrate levels in *Petunia* leaves as influenced by media Gelrite and sucrose concentrations *Plant Cell Reports* 8:358-360

Zobel, B. and Talbert, J. (1984) Applied forest tree improvement. Wiley, Prospect Heights, Ill.

Zobel, B.J. and Talbert, J. (1991) Applied Forest Tree Improvement. Waveland Press, Inc. Illinois. USA

