

BUKTI KORESPONDENSI ARTIKEL

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Berikut adalah proses pengiriman artikel, proses, komentar reviewer, perbaikan dan korespondensi dengan pihak editor jurnal yang dilakukan oleh penulis korespondensi seperti di bawah ini:

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Pengiriman pertama

The image shows two screenshots related to a journal submission. The top screenshot is a browser view of the Biodiversity Journal submission page for user Murdjoko et al. The page title is "6656 / Murdjoko et al. / Heterospecific and conspecific associations of trees in lowland tropical forest of New Guinea". The submission files table is as follows:

Submission Files	Author	Date	Type
28071-1_murdjoko, Heterospecific and conspecific associations of trees in north lowland of inland tropical forest of New Guinea after prothe.doc	amurdjoko	August 22, 2020	Article Text
28071-1_murdjoko, Copy of Certificate of Proofreading (3).pdf	amurdjoko	August 22, 2020	Other

The bottom screenshot is an email titled "[biodiv] Submission Acknowledgement" from Ahmad Dwi Setyawan to Murdjoko, dated Fri, Aug 14, 2020, 11:28 AM. The email text reads:

Thank you for submitting the manuscript "A Heterospecific and conspecific associations of trees in north lowland of inland tropical forest of New Guinea" to Biodiversity Journal of Biological Diversity. With the online journal management system that we are using, you will be able to track its progress through the editorial process by logging in to the journal web site.

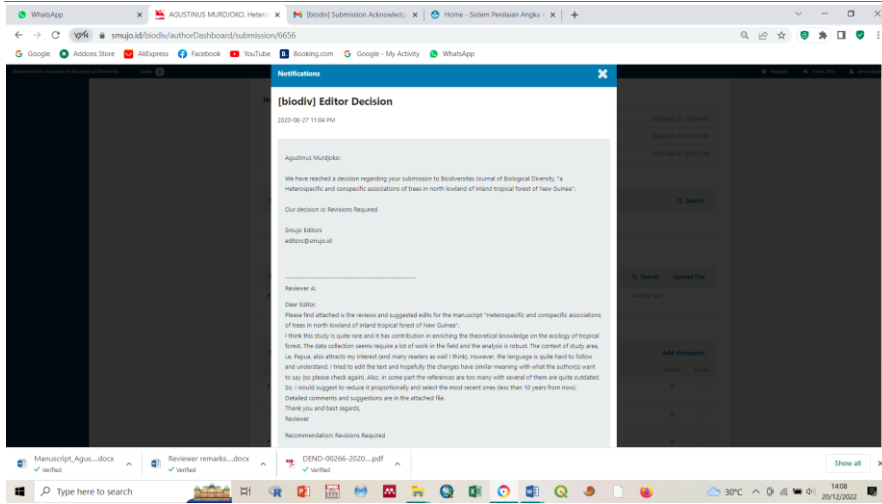
Submission URL: <https://smjoo.id/biodiv/author/Dashboard/submission/6656#>
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If you have any questions, please contact me. Thank you for considering this journal as a venue for your work.

Ahmad Dwi Setyawan

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Keputusan editor



Keputusan adalah revisi mayor dari reviewer

Surat respon terhadap komentar reviewer dan revisi

Heterospecific and conspecific associations of trees in lowland tropical forest of New Guinea

Abstract. The vegetation in the tropical rainforest of New Guinea consists of a large number of species that interact with each other within and among species. While several studies have attempted to reveal the diversity of flora of New Guinea, little is known about plant communities that develop associations. This study aimed to investigate the associations of tree species in lowland tropical forest in New Guinea. The associations depicted in this study were in the form of conspecific associations (among small and large individuals within same species) and heterospecific (among individuals of different species and divided into under and upper story). We established 48 rectangular

plots created in Murkim and Teiraplu as part of Pegunungan Bintang District, Papua Province. Canonical correspondence analysis (CCA) was used to analyze heterospecific and conspecific associations. The results showed that the understory and upper story vegetation had different patterns of heterospecific association. The understory configured three heterospecific associations, consisting of 5, 13, and 90 species, while the upper story formed four heterospecific associations with 4, 8, 11, and 63 species. The analysis of conspecific associations showed of 149 tree species recorded in the study sites, only 66 species that had both small and large individuals, displaying the pattern of conspecific association. Among them 41 species had positive associations while 25 species had negative association. Our findings enrich the knowledge in theoretical ecology of tropical forest, especially in New Guinea.

Keywords: tropical rainforest, Papuaasia, canonical correspondence analysis (CCA), tree community, Vegan package.

Running title: associations of New Guinea tree

INTRODUCTION

Tropical rainforest is a complex ecosystem with many interactions between abiotic and biotic factors, particularly among vegetation (Vitousek, 1984; Thomas & Baltzer, 2002; Hunter et al., 2015). This complexity results in the vegetation that consists of many life forms from vertical and horizontal compositions that interact with each other to obtain sunlight, soil nutrients, and water, and to adapt with microclimatic conditions (Slik et al., 2015, 2018; Murdjoko et al., 2016a). The interactions among vegetation have occurred over a long period due to successional process (Fernández-Lugo et al., 2015). Where the vegetation shares the same ecological condition, the morphological and physiological characters become the driving factors of behavior in the natural tropical rainforest (Gustafsson et al., 2016; Johnson et al., 2017).

The interactions among vegetation elements in the tropical rainforest in some cases represent symbiosis and inter and intra-species relationships (Legendre & Fortin, 1989; Magrach et al., 2014). These interactions can be in the form of competition and association. In old tropical rainforest, the interaction occurs intensively due to the absorption of light and water, where both are the primary growth factors (Yamamoto, 2000; Montgomery & Chazdon, 2001). In secondary forest, canopy gap is very open, leading to more light penetrating the forest floor (Itoh et al., 1997; Angelini et al., 2015; Murdjoko et al., 2017). The association in vegetation communities can be in the form of conspecific or heterospecific and the form of association determines the pattern of the spatial distribution of forest ecosystems either. Conspecific association is the interaction of individuals of similar species while heterospecific occurs among different species of vegetation (Zhu et al., 2015; Wang et al., 2018). Conspecific and heterospecific associations occur during the successional process of the tropical rainforest (Farneda et al., 2018). Some studies explained that the association, either the conspecific or the heterospecific could be in a positive or negative pattern (Castilla et al., 2016).

Vegetation is distributed geographically with the diversity and pattern of plant communities that adapt to particular ecological niche (Brummitt, 2001; Pan et al., 2013). Phytogeographic regions, including mainland New Guinea, have been studied for centuries. The vegetation in New Guinea spreads from coastal to high land areas, containing various types of ecosystems (Cámara-Leret & Dennehy, 2019). As the result, New Guinea contains the highest diversity of flora, such as trees, climbers, shrubs, ferns, rattan, etc. (Murdjoko et al., 2016a) in which about 60% of the species are endemic (Cámara-Leret et al., 2020). For example, a forest area in New Guinea consists of a high diversity of tree species with more than 70 species per hectare that could be found (Robiansyah, 2018; Fatem et al., 2020). While recently more and more studies have attempted to reveal the diversity of flora of New Guinea, little is known about plant communities that develop associations among them.

This study aimed to investigate the association of tree species in the lowland tropical forest in New Guinea. The associations depicted in this study were in the form of conspecific associations (among small and large individuals within same species) and heterospecific (among individuals of different species and

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divided into under and upper story). We hypothesized that the small and large tree species have heterospecific associations within the natural tropical rain forest. This kind of study is important to provide specific contribution of ecological research in the tropical rainforest of Southeast Asia, more specifically the New Guinea region (Brummitt, 2001).

MATERIALS AND METHODS

Study period and area

This study was conducted in the northern part of Pegunungan Bintang District, Papua Province, Indonesia (Figure 1). The study sites were located at Murkim (4°0'0.53"S and 140°49'17.24"E) and Teiraplu (3°59'13.46"S and 140°26'0.06"E) at an altitude of 155 m and 233 m above sea level (m asl), respectively. The ecosystem type of the two study sites are categorized as lowland areas where the southern part is bordered with the mountain range and the northern part is bordered with hills while the western and eastern parts are lowlands. Broadleaves and mixed forests are the dominant vegetation in this area, while the soil is grouped as Ultisols and Inceptisol. The climatic conditions are considered to be very humid with average temperature of 25° C for annual, 20.6° C for daily, and 16.3° C for minimum, and with monthly and annual average rainfall of 448.75 mm and 5385 mm, respectively (Kartikasari et al., 2012).

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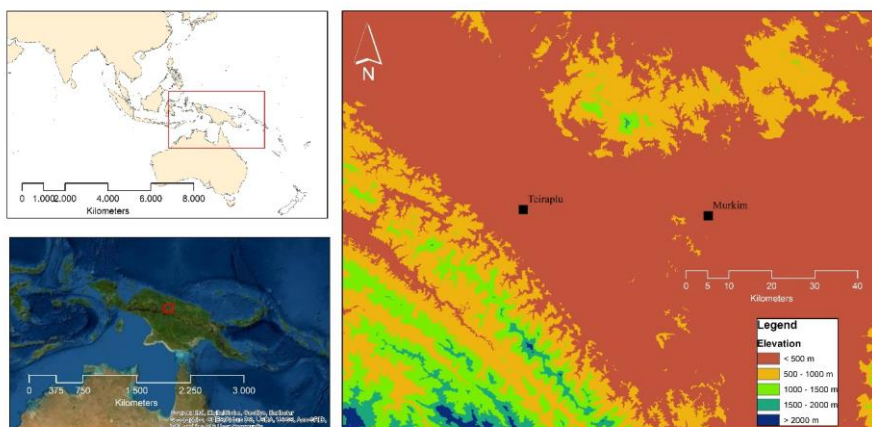


Figure 1. Map of the study sites in Teiraplu and Murkim, Pegunungan Bintang, Papua.

Sampling and data collection

Data were collected using sampling plot method with size of each plot 20 m x 20 m. In total, there were 48 rectangular plots established in which 24 plots were in Teiraplu and 24 plots were in Murkim. In both locations, the plots were placed to north directions at a distance of 100 m away from each other. In the 20 m x 20 m plot (A) we recorded and measured old trees with a diameter of more than 20 cm, and within this plot we established three nested sub-plots with size 10 m x 10 m (B) to record tree with diameter between 10 cm and 20 cm, size 5 m x 5 m (C) to record trees taller than 1.5 m, and size 2 m x 2 m (D) to record the species shorter than 1.5 m. The vegetation in plots A and B were classified as upper story and that in plots C and D were categorized as understory. For the understory vegetation, we recorded data of

taxonomic names of every species and number of individuals, and while for the upper story vegetation we recorded data of taxonomic names of every species, number of individuals and diameter (cm). For identification, we collected the specimens of the plant and sent it to the Herbarium Papuaense of Balai Penelitian dan Pengembangan Lingkungan Hidup dan Kebutuhan (BP2LHK) Manokwari and Herbarium Manokwariense (MAN) Pusat Penelitian Keanekaragaman Hayati Universitas Papua (PPKH-UNIPA), Manokwari. The species name was updated according to The Plant List (TPL) at the website of <http://www.theplantlist.org/>.

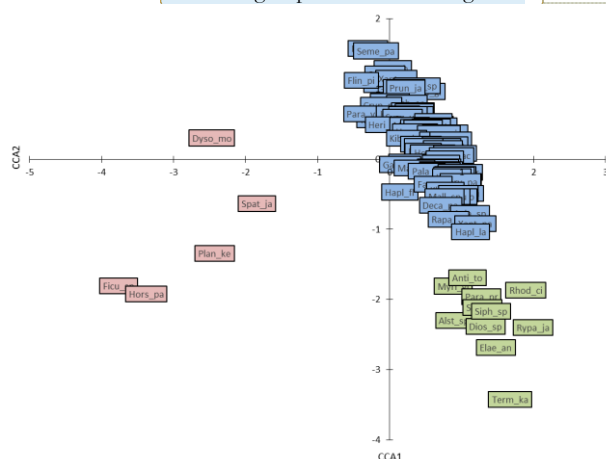
Statistical analysis

The heterospecific and conspecific associations were analyzed using the canonical correspondence analysis (CCA) (Ter Braak, 1986; Caceres & Legendre, 2009), and the chi-square test (χ^2) was implemented to validate the model of CCA (Fatem et al., 2020). Furthermore, this association used the number of each individual (density) as a value in which the columns were the species and the rows were the 48 plots. The conspecific association correlated the under and upper story as small and large individuals. The columns represented the species, while the 48 plots under and upper story represented the rows. The species that did not have under and upper stories were otherwise excluded. The result of CCA displayed species in the graph with the position in the two axes. To investigate the conspecific association whether it was positive or negative, the Euclidean distance between each species as well as the under and upper stories were conducted (Murdjoko et al., 2016b, 2017). If the result of Euclidean distance of species is below the average, then the conspecific association is said to be positive, and vice versa. The vegan package in R version 3.5.3 was used to calculate the statistical analysis (Oksanen et al., 2019).

RESULTS AND DISCUSSION

Heterospecific associations

The heterospecific association was grouped into two: understory and upper story, based on the structure of trees in tropical forests. As such, the analyses of multivariate statistics for the understory and upper story were separated since the natural tropical rainforest is complex with the vegetation structures forming the ecosystem. The structure was also simplified by distinguishing them into two main parts. From the CCA result, the understory and upper story showed different patterns of heterospecific association. The understory configured three groups of communities based on species as the structure of the associations. The three groups are shown in Figure 2.

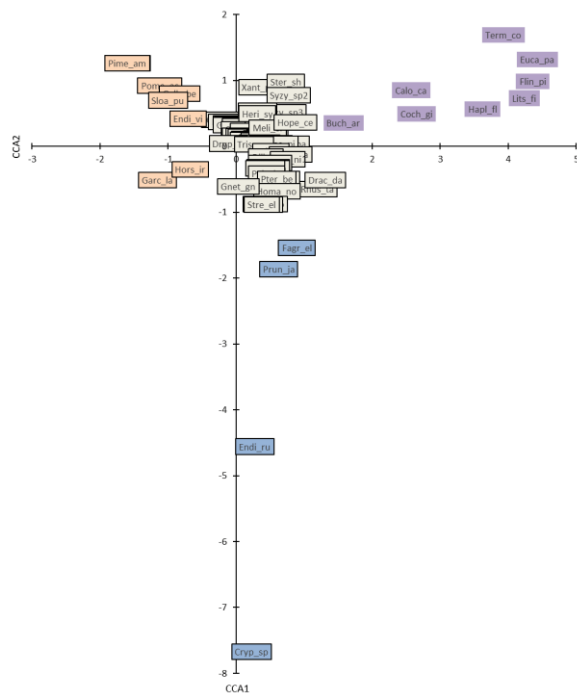


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Figure 2. The result of Canonical Correspondence Analysis (CCA) to analyze the heterospecific associations for understory.

The results showed that 108 species of trees formed the association in natural tropical forests, and was valid statistically as $\chi^2 = 10.686$, $df = 2461$, $p\text{-value} = 1$. The species of understory showed heterospecific associations as tree groups where the first consisted 90 species (blue boxes), the second contained 13 (green boxes), and the third comprised of 5 species (red boxes) (Figure 2). The name of the species in the boxes in Figure 2 was abbreviated and the complete name can be seen in Appendix 1.



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Figure 3. The result of Canonical Correspondence Analysis (CCA) to analyze the heterospecific associations for the upper story.

The CCA result showed that the upper story vegetation community had a pattern of association with a valid result of $\chi^2 = 11.344$, $df = 1955$, $p\text{-value} = 1$. The upper story consisted of 86 species which formed four heterospecific associations, consisting of the first group (63 species) in the grey boxes, second group (11 species) in the red boxes, third group (8 species) in the purple boxes, and fourth group (4 species) in the blues boxes (Figure 3). The complete name of species presented in Figure 3 can be seen in Appendix 2.

The association pattern of the understory and upper story differed from one another even though they grew in the same natural forest. The difference in association has likely resulted from the variation of the vertical structure of the tropical forest. The upper story vegetation has reached the emergent layers of forest canopy, allowing species to benefit by getting more sunlight (Murdjoko et al., 2016a, 2017; Fatem et al., 2020). The formation of understory was caused by competition due to it is below the canopy layers with low solar radiation (Rüger et al., 2011; Laurans et al., 2014; Angelini et al., 2015).

For centuries, the formation of tropical forests has been a sequential process in which large numbers of species compete dynamically each other (Brown et al., 1990; Wright & Muller-Landau, 2006; Liu &

Slik, 2014; Almeida et al., 2019). The heterospecific association can be related with the fact that trees interact each other to form symbiosis with other life forms, such as liana, fern, herb, epiphyte and so on (Johnson et al., 2017; Cirimwami et al., 2019; Steege et al., 2019). The primary factor influencing the pattern of tree communities of understory and upper story during tropical forest succession was probably caused by the abiotic factors, especially to gain nutrients, water, and sunlight as materials to support metabolisms, especially photosynthesis. Nonetheless, many studies showed that the morphological and physiological characters have also affected different responses of species to grow and develop (Goodale et al., 2012; Gustafsson et al., 2016). For example, the nature of shade tolerance species may be a factor that allows small tree species to survive the competition and obtain limited sunlight below the canopy layers (Givnish, 1999; Montesinos-Navarro et al., 2018). Therefore, it is crucial to study the shade-tolerant characters of a species in the rainforest in order to explain forest dynamics in more detail. This study is unable to reveal such characters concerning the light competition because that is beyond the scope of this study.

Conspecific associations

The analysis of conspecific associations was conducted using 149 species that grew in the study sites, but only 66 species that had small and large individuals as understory and upper story. The result of CCA showed statistically valid result as $\chi^2 = 5.8784$, $df = 2904$, $p\text{-value} = 1$ (Figure 4). In addition, it displayed the pattern of conspecific association as 41 species had positive association while 25 species had negative association. In the positive association, the small and large individuals of the 41 species were distributed closely in the same area, representing the tendency of mature trees to reproduce and germinate. Conversely, in the negative association, the small individuals of the 25 species grew mainly far from the large ones that represent the matured trees. The full list of the taxonomic name of the species in Figure 4 is presented in Appendix 3, and the conspecific association can be used to analyze their density dependence since the tropical forest is the place for the high diversity of trees.

Of 149 species, 83 species did not have either small or large individuals, suggesting that the species experienced poor regeneration. Some large individuals act as putative parent trees, even though they have failed to establish seedlings due to many factors (Seidler & Plotkin, 2006; Rahman & Tsukamoto, 2015). One possible factor is caused by the competition of seedlings with other plants in the forest floor, on which many life forms are found. Another rationale is that the seeds and seedlings are eaten by herbivores (Swaine et al., 1987; Houter & Pons, 2014). Many studies have reported that herbivores are found in tropical rainforest since the forest provides a lot of food, for example, during germination, the dicotyledonous tree plants develop shoot from the plumule of the germinating seed (Houter & Pons, 2014; Sawada et al., 2015).

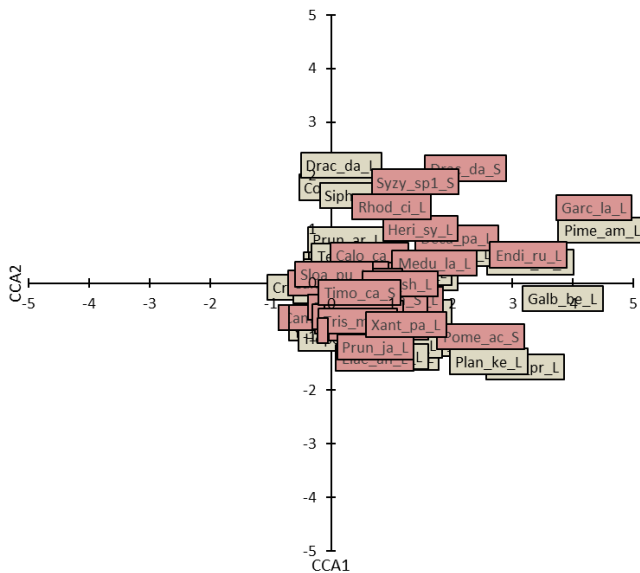


Figure 4. The result of Canonical Correspondence Analysis (CCA) to analyze conspecific associations between the small and the large individuals of the same species.

The distribution of individual trees in tropical forests is influenced by the ability to interact with other species. This pattern of conspecific association should be studied frequently to figure out the method of regeneration and distribution of species. Forest floor encompasses many species with different life forms as a strategy to survive and grow during the competition (Dezzotti et al., 2019). Many lianas and climbers grow fast to occupy the forest canopy and space available for sunlight. These plants suppress a certain seedling establishment (Carreño-Rocabado et al., 2012). The competition to gain sunlight, nutrition, and water is presumed as the limiting factor suffered by some species since they cannot survive below putative parent trees.

Seed dispersal can be the driving force behind the spatial distribution of plants in tropical forests. Moreover, the morphological and anatomical characters of seeds and fruits also influence species regeneration and distribution. For example, small and winged seeds of tree species can spread out by falling around and away from the parent trees (Sebbenn et al., 2008; Lü & Tang, 2010). However, factors such as competition, herbivory, and allelopathy have led to a clear and negative association in natural tropical forests (Padmanaba & Corlett, 2014; Menezes et al., 2019). In contrast, large seeds mostly fall around the parent trees and since they survived germination, they can grow as positive conspecific associations. Therefore, the conspecific association pattern should be studied to know the natural regeneration of certain species in tropical rainforest.

The implication of associations to ecological knowledge for sustainable management of primary forest

The study of conspecific and heterospecific associations in tropical rainforest is extremely important to determine the spatial distribution pattern, especially the conspecific association. In addition, a model of natural regeneration of tree species can be described, and the result can indicate the pattern of recruitment in the population dynamics of tree species (Goodale et al., 2012; Piotta et al., 2019). Tropical rainforest is primarily dominated by flowering plants with their reproduction season is in annual period (Baker et al., 1998; Pan et al., 2013; Cámara-Leret et al., 2020). Furthermore, a suitable area for certain species to grow

has resembled in the conspecific association since the study correlates small individuals with the large ones within the same species. The pattern of conspecific association can also be used to observe natural regeneration. For example, the most appropriate area to plant tree species *in-situ* conservation programs can be decided when artificial regeneration is necessary (Armstrong et al., 2011; Vergara-Rodríguez et al., 2017). The heterospecific association describes the pattern of growth in tropical rainforest since the forest includes the great diversity of tree species. The forest took several decades to develop, and this present study has been able to analyze the pattern of tree species association. Ecological studies on the theme of species association in tropical rainforest need to be replicated in other contexts of region, ecosystem type and forest conditions as tropical forest is very complex as made up of different life forms that interact and create vertical and horizontal structures in the climax phase of the successional process (Chazdon, 2003; Brokaw & Scheiner, 2012).

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Appendix 1. Species name of heterospecific associations for understory for Figure 2.

No.	Code	Species	Group 1	Group 2	Group 3
1	Kiba_co	<i>Kibara coriacea</i> (Blume) Hook. f. & A. Thoms.	√		
2	Rhus_la	<i>Rhus lamprocarpa</i> Merr. & L.M.Perry	√		
3	Dios_pi	<i>Diospyros pilosantha</i> Blanco	√		
4	Lits_le	<i>Litsea ledermannii</i> Tschner	√		
5	Maca_gi	<i>Macaranga gigantea</i> (Rchb.f. & Zoll.) Müll.Arg.	√		
6	Maca_ta	<i>Macaranga tanarius</i> (L.) Müll.Arg.	√		
7	Seme_pa	<i>Semecarpus papuana</i> Lauterb.	√		
8	Ster_sh	<i>Sterculia shillinglawii</i> F.Muell.	√		
9	Case_ca	<i>Casearia carrii</i> Sleumer	√		
10	Xant_no	<i>Wendlandia</i> sp	√		
11	Teij_bo	<i>Teijsmanniodendron bogoriense</i> Koord.	√		
12	Flin_pi	<i>Flindersia pimenteliana</i> F.Muell.	√		
13	Case_sp	<i>Casearia</i> sp	√		
14	Hapl_ce	<i>Haplolobus celebicus</i> H.J.Lam	√		
15	Myri_en	<i>Myristica ensifolia</i> J.Sinclair	√		
16	Octa_in	<i>Octamyrtus insignis</i> Diels	√		
17	Piso_lo	<i>Pisonia longirostris</i> Teijsm. & Binn.	√		
18	Wend_sp	<i>Wendlandia</i> sp	√		
19	Timo_ca	<i>Timonius carrii</i> S.P.Darwin	√		
20	Prun_ja	<i>Prunus javanica</i> (Teijsm. & Binn.) Miq.	√		
21	Hapl_fl	<i>Haplolobus floribundus</i> (K.Schum.) H.J.Lam	√		
22	Anti_de	<i>Alstonia spectabilis</i> R.Br.	√		
23	Buch_ar	<i>Buchanania arborescens</i> (Blume) Blume	√		
24	Cryp_pa	<i>Cryptocarya palmerensis</i> C.K.Allen	√		
25	Gmel_se	<i>Gmelina sessilis</i> C.T.White & W.D.Francis ex Lane-Poole	√		
26	Garc_pi	<i>Garcinia picrobiza</i> Miq.	√		
27	Para_ve	<i>Parastemon versteegii</i> Merr. & L.M.Perry	√		
28	Knem_in	<i>Knema intermedia</i> Warb.	√		
29	Case_mo	<i>Casearia monticola</i> Sleumer	√		
30	Half_ke	<i>Halfordia kendack</i> Guillaumin	√		
31	Syzy_ve	<i>Syzygium versteegii</i> (Lauterb.) Merr. & L.M.Perry	√		
32	Ficu_ro	<i>Ficus robusta</i> Corner	√		
33	Heri_sy	<i>Heritiera sylvatica</i> S.Vidal	√		
34	Cerb_fl	<i>Cerbera floribunda</i> K.Schum.	√		
35	Myri_gl	<i>Myristica globosa</i> Warb.	√		
36	Dryp_gl	<i>Drypetes globosa</i> (Merr.) Pax & K.Hoffm.	√		
37	Phal_ma	<i>Phaleria macrocarpa</i> (Scheff.) Boerl.	√		
38	Cana_ri	<i>Canarium rigidum</i> (Blume) Zipp. ex Miq.	√		
39	Lits_ti	<i>Litsea timoriana</i> Span.	√		
40	Timo_ti	<i>Timonius timon</i> (Spreng.) Merr.	√		
41	Homa_fo	<i>Heritiera sylvatica</i> S.Vidal	√		
42	Endo_me	<i>Endospermum medulosum</i> L.S.Sm.	√		
43	Pome_pi	<i>Pometia pinnata</i> J.R.Forst. & G.Forst.	√		
44	Kiba_bu	<i>Kibara bullata</i> Philipson	√		
45	Drac_da	<i>Dracontomelon dao</i> (Blanco) Merr. & Rolfe	√		
46	Goni_gi	<i>Goniothalamus giganteus</i> Hook.f. & Thomson	√		
47	Nauc_or	<i>Nauclea orientalis</i> (L.) L.	√		
48	Beil_mo	<i>Beilschmedia morobensis</i> Kosterm.	√		
49	Cara_br	<i>Carallia brachiata</i> (Lour.) Merr.	√		
50	Lits_sp	<i>Litsea</i> sp	√		
51	Pime_am	<i>Pimelodendron amboinicum</i> Hassk.	√		
52	Medu_la	<i>Medusanthera laxiflora</i> (Miers) R.A.Howard	√		
53	Myri_fa	<i>Myristica fatua</i> Houtt.	√		
54	Gono_li	<i>Gonocaryum litorale</i> (Blume) Sleumer	√		
55	Lith_ru	<i>Litbocarpus rajavillosus</i> (Markgr.) Rehder	√		
56	Hope_pa	<i>Hopea papuana</i> Diels	√		
57	Dill_pa	<i>Dillenia papuana</i> Martelli	√		
58	Mast_pa	<i>Mastixiodendron pachycladus</i> (K.Schum.) Melch.	√		
59	Pome_ac	<i>Pometia acuminata</i> Radlk.	√		
60	Kiba_el	<i>Kibara elongata</i> A.C.Sm.	√		
61	Clei_pa	<i>Cleistanthus papuanus</i> (Lauterb.) Jabl.	√		

No.	Code	Species	Group 1	Group 2	Group 3
62	Ints_pa	<i>Intsia palembanica</i> Miq.	√		
63	Galb_be	<i>Galbulimima belgraveana</i> (F.Muell.) Sprague	√		
64	Term_co	<i>Terminalia copelandi</i> Elmer	√		
65	Giro_ne	<i>Gironniera nervosa</i> Planch.	√		
66	Mani_pl	<i>Manilloa plurijuga</i> Merr. & L.M.Perry	√		
67	Hope_ce	<i>Hopea celtidifolia</i> Kosterm.	√		
68	Meli_el	<i>Melicope elleryana</i> (F. Muell.) T.G. Hartley	√		
69	Garc_sp	<i>Garcinia</i> sp	√		
70	Pala_lo	<i>Palaquium lobbianum</i> Burck	√		
71	Camp_br	<i>Campnosperma brevipetiolatum</i> Volkens	√		
72	Ster_ma	<i>Sterculia macrophylla</i> Vent.	√		
73	Harp_ca	<i>Harpullia currii</i> Leenh.	√		
74	Hors_la	<i>Horsfieldia laevigata</i> Warb.	√		
75	Arch_pa	<i>Archidendron parviflorum</i> Pulle	√		
76	Chry_pa	<i>Chrysophyllum papuanicum</i> (Pierre ex Dubard) Royen	√		
77	Fagr_ra	<i>Fagraea racemosa</i> Jack	√		
78	Klei_ho	<i>Kleinboria hospita</i> L.	√		
79	Endi_ru	<i>Endiandra rubescens</i> (Blume) Miq.	√		
80	Tris_ma	<i>Tristanopsis macrosperma</i> (F.Muell.) Peter G.Wilson & J.T.Waterh.	√		
81	Cory_la	<i>Corynocarpus laevigatus</i> J.R.Forst. & G.Forst.	√		
82	Cryp_sp	<i>Cryptocarya</i> sp	√		
83	Tabe_au	<i>Tabernaemontana aurantiaca</i> Gaudich.	√		
84	Mall_sp	<i>Mallotus</i> sp	√		
85	Deca_pa	<i>Decaspermum parviflorum</i> (Lam.) A.J.Scott	√		
86	Agla_sp	<i>Aglaia spectabilis</i> (Miq.) S.S.Jain & S.Bennet	√		
87	Rapa_te	<i>Rapanea tempapan</i> P.Royen	√		
88	Calo_ca	<i>Calophyllum caudatum</i> Kaneh. & Hatus.	√		
89	Xant_pa	<i>Xanthophyllum papuanum</i> Whitmore ex Meijden	√		
90	Hapl_la	<i>Haplobolus lanceolatus</i> H.J.Lam ex Leenh.	√		
91	Alst_sp	<i>Alstonia spectabilis</i> R.Br.			√
92	Dios_sp	<i>Diospyros</i> sp			√
93	Elae_an	<i>Elaeocarpus angustifolius</i> Blume			√
94	Rypa_ja	<i>Ryparosa javanica</i> Koord. & Valetton			√
95	Rhod_ci	<i>Rhodammia cinerea</i> Jack			√
96	Myri_gi	<i>Myristica gigantea</i> King			√
97	Anti_to	<i>Antiaris toxicaria</i> Lesch.			√
98	Calo_in	<i>Calophyllum inophyllum</i> L.			√
99	Para_pr	<i>Pararchidendron pruinosum</i> (Benth.) I.C.Nielsen			√
100	Siph_ce	<i>Siphonodon celastrineus</i> Griff.			√
101	Siph_sp	<i>Siphonodon</i> sp			√
102	Maas_gl	<i>Maasia glauca</i> (Hassk.) Mols, Kessler & Rogstad			√
103	Term_ka	<i>Terminalia kaernbachii</i> Warb.			√
104	Spat_ja	<i>Spathiostemon javensis</i> Blume		√	
105	Dyso_mo	<i>Dysoxylum mollissimum</i> Blume		√	
106	Plan_ke	<i>Planchonella keyensis</i> H.J.Lam		√	
107	Ficu_sp	<i>Ficus</i> sp		√	
108	Hors_pa	<i>Horsfieldia parviflora</i> (Roxb.) J.Sinclair		√	

Appendix 2. Species name of heterospecific associations for upper story Figure 3.

No.	Code	Species	Group 1	Group 2	Group 3	Group 4
1	Hors_ir	<i>Horsfieldia irya</i> (Gaertn.) Warb.		√		
2	Garc_la	<i>Garcinia latissima</i> Miq.		√		
3	Alst_sc	<i>Alstonia scholaris</i> (L.) R. Br.		√		
4	Endi_vi	<i>Endiandra virens</i> F.Muell.		√		
5	Sloa_pu	<i>Sloanea pullei</i> O.C.Schmidt ex A.C.Sm.		√		
6	Galb_be	<i>Galbulimima belgraveana</i> (F.Muell.) Sprague		√		
7	Pome_ac	<i>Pometia acuminata</i> Radlk.		√		
8	Chis_ce	<i>Chisocheton ceramicus</i> Miq.		√		
9	Dyso_mo	<i>Dysoxylum mollissimum</i> Blume		√		
10	Endo_me	<i>Endospermum medullosum</i> L.S.Sm.		√		
11	Pime_am	<i>Pimelodendron amboinicum</i> Hassk.		√		
12	Gono_li	<i>Gonocaryum littorale</i> (Blume) Sleumer	√			

No.	Code	Species	Group 1	Group 2	Group 3	Group 4
13	Stre_el	<i>Streblus elongatus</i> (Miq.) Corner	√			
14	Homa_fo	<i>Heritiera sylvatica</i> S.Vidal	√			
15	Gnet_gn	<i>Gnetum gnemon</i> L.	√			
16	Homa_no	<i>Homalanthus novoguineensis</i> (Warb.) K.Schum.	√			
17	Cory_la	<i>Corynocarpus laevigatus</i> J.R.Forst. & G.Forst.	√			
18	Drac_da	<i>Dracontomelon dao</i> (Blanco) Merr. & Rolfe	√			
19	Cana_od	<i>Canarium indicum</i> L.	√			
20	Pter_be	<i>Pterocymbium beccarii</i> K.Schum.	√			
21	Pala_lo	<i>Palaquium lobbianum</i> Burck	√			
22	Cana_in	<i>Canarium indicum</i> L.	√			
23	Cara_br	<i>Carallia brachiata</i> (Lour.) Merr.	√			
24	Timo_ca	<i>Timonius carii</i> S.P.Darwin	√			
25	Hors_sy	<i>Horsfieldia sylvestris</i> Warb.	√			
26	Maas_su	<i>Maasia sumatrana</i> (Miq.) Mols, Kessler & Rogstad	√			
27	Hope_pa	<i>Hopea papuana</i> Diels	√			
28	Hope_ce	<i>Hopea cellidifolia</i> Kosterm.	√			
29	Rhus_ta	<i>Rhus taiensis</i> Guill.	√			
30	Acti_ni	<i>Actinodaphne nitida</i> Teschner	√			
31	Dill_pa	<i>Dillenia papuana</i> Martelli	√			
32	Medu_la	<i>Medusanthura lasziflora</i> (Miers) R.A.Howard	√			
33	Teij_bo	<i>Teijsmanniodendron bogoriense</i> Koord.	√			
34	Tris_ma	<i>Tristaniaopsis macrosperma</i> (F.Muell.) Peter G.Wilson & J.T.Waterh.	√			
35	Call_lo	<i>Calliandra longifolia</i> Lam.	√			
36	Comm_ba	<i>Commersonia bartramia</i> (L.) Merr.	√			
37	Dios_pi	<i>Diospyros pilosanthera</i> Blanco	√			
38	Knem_in	<i>Knema intermedia</i> Warb.	√			
39	Dryp_gl	<i>Drypetes globosa</i> (Merr.) Pax & K.Hoffm.	√			
40	Cryp_pa	<i>Cryptocarya palmerensis</i> C.K.Allen	√			
41	Meli_el	<i>Melicope elleryana</i> (F. Muell.) T.G. Hartley	√			
42	Lits_ti	<i>Litsea timoriana</i> Span.	√			
43	Siph_ce	<i>Siphonodon celastrineus</i> Griff.	√			
44	Siph_sp	<i>Siphonodon</i> sp	√			
45	Vite_pi	<i>Vitex pinnata</i> L.	√			
46	Poly_no	<i>Polyscias nodosa</i> (Blume) Seem.	√			
47	Pome_pi	<i>Pometia pinnata</i> J.R.Forst. & G.Forst.	√			
48	Agla_ar	<i>Aglaia argentea</i> Blume	√			
49	Acro_sp	<i>Acronychia</i> sp	√			
50	Gmel_se	<i>Gmelina sessilis</i> C.T.White & W.D.Francis ex Lane-Poole	√			
51	Mani_br	<i>Maniltoa bronneioides</i> Harms	√			
52	Prun_ar	<i>Prunus arborea</i> (Blume) Kalkman	√			
53	Camp_br	<i>Campnosperma brevipetiolatum</i> Volkens	√			
54	Hors_la	<i>Horsfieldia laevigata</i> Warb.	√			
55	Cana_hi	<i>Campnosperma brevipetiolatum</i> Volkens	√			
56	Deca_pa	<i>Decaspermum parviflorum</i> (Lam.) A.J.Scott	√			
57	Calo_in	<i>Calophyllum inophyllum</i> L.	√			
58	Heri_sy	<i>Heritiera sylvatica</i> S.Vidal	√			
59	Clei_pa	<i>Cleistanthus papuanus</i> (Lauterb.) Jabl.	√			
60	Elae_an	<i>Elaeocarpus angustifolius</i> Blume	√			
61	Gymn_fa	<i>Gymnacranthera farquhariana</i> (Hook.f. & Thomson) Warb.	√			
62	Grew_er	<i>Grewia eriocarpa</i> Juss.	√			
63	Xant_no	<i>Wendlandia</i> sp	√			
64	Rhod_ci	<i>Rhodammia cinerea</i> Jack	√			
65	Arto_al	<i>Artocarpus alilis</i> (Parkinson ex F.A.Zorn) Fosberg	√			
66	Para_pr	<i>Pararchidendron pruinosum</i> (Benth.) I.C.Nielsen	√			
67	Plan_ke	<i>Planchonella keyensis</i> H.J.Lam	√			
68	Dios_pa	<i>Diospyros papuana</i> Valetton ex Bakh.	√			
69	Ochr_gl	<i>Ochrosia glomerata</i> (Blume) F.Muell.	√			
70	Myri_fa	<i>Myristica fatua</i> Houtt.	√			
71	Ster_sh	<i>Sterculia shillinghamii</i> F.Muell.	√			
72	Syzy_sp2	<i>Syzygium</i> sp2	√			
73	Syzy_sp3	<i>Syzygium</i> sp3	√			
74	Xant_pa	<i>Xanthophyllum papuanum</i> Whitmore ex Meijden	√			
75	Euca_pa	<i>Eucalyptopsis papuana</i> C.T.White		√		

No.	Code	Species	Group 1	Group 2	Group 3	Group 4
76	Flin_pi	<i>Flindersia pimenteliana</i> F.Muell.			√	
77	Hapl_fl	<i>Haplolobus floribundus</i> (K.Schum.) H.J.Lam			√	
78	Lits_fi	<i>Litsea firma</i> (Blume) Hook.f.			√	
79	Term_co	<i>Terminalia copelandi</i> Elmer			√	
80	Calo_ca	<i>Calophyllum caudatum</i> Kaneh. & Hatus.			√	
81	Coch_gi	<i>Cochlospermum gillivraei</i> Benth.			√	
82	Buch_ar	<i>Buchanania arborescens</i> (Blume) Blume			√	
83	Fagr_el	<i>Fagraea elliptica</i> Roxb.				√
84	Prun_ja	<i>Prunus javanica</i> (Teijsm. & Binn.) Miq.				√
85	Endi_ru	<i>Endiandra rubescens</i> (Blume) Miq.				√
86	Cryp_sp	<i>Cryptocarya</i> sp				√

Appendix 3. Species name of conspecific associations Figure 4. The S stands for small individuals and L symbolizes large individuals.

No.	Code	Species
1	Agla_ar _L	<i>Aglaia argentea</i> Blume
2	Agla_ar _S	<i>Aglaia argentea</i> Blume
3	Buch_ar _L	<i>Buchanania arborescens</i> (Blume) Blume
4	Buch_ar _S	<i>Buchanania arborescens</i> (Blume) Blume
5	Calo_ca _L	<i>Calophyllum caudatum</i> Kaneh. & Hatus.
6	Calo_ca _S	<i>Calophyllum caudatum</i> Kaneh. & Hatus.
7	Calo_in _L	<i>Calophyllum inophyllum</i> L.
8	Calo_in _S	<i>Calophyllum inophyllum</i> L.
9	Camp_br _L	<i>Campnosperma brevipetiolatum</i> Volkens
10	Camp_br _S	<i>Campnosperma brevipetiolatum</i> Volkens
11	Cana_hi _L	<i>Canarium hirsutum</i> Willd.
12	Cana_hi _S	<i>Canarium hirsutum</i> Willd.
13	Cana_in _L	<i>Canarium indicum</i> L.
14	Cana_in _S	<i>Canarium indicum</i> L.
15	Cara_br _L	<i>Carallia brachiata</i> (Lour.) Merr.
16	Cara_br _S	<i>Carallia brachiata</i> (Lour.) Merr.
17	Clei_pa _L	<i>Cleistanthus papuanus</i> (Lauterb.) Jabl.
18	Clei_pa _S	<i>Cleistanthus papuanus</i> (Lauterb.) Jabl.
19	Cory_la _L	<i>Corynocarpus laevigatus</i> J.R.Forst. & G.Forst.
20	Cory_la _S	<i>Corynocarpus laevigatus</i> J.R.Forst. & G.Forst.
21	Cryp_pa _L	<i>Cryptocarya palmerensis</i> C.K.Allen
22	Cryp_pa _S	<i>Cryptocarya palmerensis</i> C.K.Allen
23	Cryp_sp _L	<i>Cryptocarya</i> sp
24	Cryp_sp _S	<i>Cryptocarya</i> sp
25	Deca_pa _L	<i>Decaspermum parviflorum</i> (Lam.) A.J.Scott
26	Deca_pa _S	<i>Decaspermum parviflorum</i> (Lam.) A.J.Scott
27	Dill_pa _L	<i>Dillenia papuana</i> Martelli
28	Dill_pa _S	<i>Dillenia papuana</i> Martelli
29	Dios_pi _L	<i>Diospyros pilosanthera</i> Blanco
30	Dios_pi _S	<i>Diospyros pilosanthera</i> Blanco
31	Drac_da _L	<i>Dracontomelon dao</i> (Blanco) Merr. & Rolfe
32	Drac_da _S	<i>Dracontomelon dao</i> (Blanco) Merr. & Rolfe
33	Dryp_gl _L	<i>Drypetes globosa</i> (Merr.) Pax & K.Hoffm.
34	Dryp_gl _S	<i>Drypetes globosa</i> (Merr.) Pax & K.Hoffm.
35	Dyso_mo _L	<i>Dysoxylum mollissimum</i> Blume
36	Dyso_mo _S	<i>Dysoxylum mollissimum</i> Blume
37	Elae_an _L	<i>Elaeocarpus angustifolius</i> Blume
38	Elae_an _S	<i>Elaeocarpus angustifolius</i> Blume
39	Endi_ru _L	<i>Endiandra rubescens</i> (Blume) Miq.
40	Endi_ru _S	<i>Endiandra rubescens</i> (Blume) Miq.
41	Endo_me _L	<i>Endospermum medulosum</i> L.S.Sm.
42	Endo_me _S	<i>Endospermum medulosum</i> L.S.Sm.
43	Flin_pi _L	<i>Flindersia pimenteliana</i> F.Muell.
44	Flin_pi _S	<i>Flindersia pimenteliana</i> F.Muell.
45	Galb_be _L	<i>Galbulimima belgraveana</i> (F.Muell.) Sprague
46	Galb_be _S	<i>Galbulimima belgraveana</i> (F.Muell.) Sprague

No.	Code	Species
47	Garc_la _L	<i>Garcinia latissima</i> Miq.
48	Garc_la _S	<i>Garcinia latissima</i> Miq.
49	Giro_ne _L	<i>Gironniera nervosa</i> Planch.
50	Giro_ne _S	<i>Gironniera nervosa</i> Planch.
51	Gmel_se _L	<i>Gmelina sessilis</i> C.T.White & W.D.Francis ex Lane-Poole
52	Gmel_se _S	<i>Gmelina sessilis</i> C.T.White & W.D.Francis ex Lane-Poole
53	Gnet_gn _L	<i>Gnetum gnemon</i> L.
54	Gnet_gn _S	<i>Gnetum gnemon</i> L.
55	Gono_li _L	<i>Gonocaryum litorale</i> (Blume) Sleumer
56	Gono_li _S	<i>Gonocaryum litorale</i> (Blume) Sleumer
57	Gymn_fa _L	<i>Gymnacranthera jarqubariana</i> (Hook.f. & Thomson) Warb.
58	Gymn_fa _S	<i>Gymnacranthera jarqubariana</i> (Hook.f. & Thomson) Warb.
59	Hapl_fl _L	<i>Haplolobus floribundus</i> (K.Schum.) H.J.Lam
60	Hapl_fl _S	<i>Haplolobus floribundus</i> (K.Schum.) H.J.Lam
61	Heri_sy _L	<i>Heritiera sylvatica</i> S.Vidal
62	Heri_sy _S	<i>Heritiera sylvatica</i> S.Vidal
63	Homa_fo _L	<i>Homalium foetidum</i> Benth.
64	Homa_fo _S	<i>Homalium foetidum</i> Benth.
65	Hope_ce _L	<i>Hopea celtidifolia</i> Kosterm.
66	Hope_ce _S	<i>Hopea celtidifolia</i> Kosterm.
67	Hope_no _L	<i>Hopea novoguineensis</i> Slooten
68	Hope_no _S	<i>Hopea novoguineensis</i> Slooten
69	Hope_pa _L	<i>Hopea papuana</i> Diels
70	Hope_pa _S	<i>Hopea papuana</i> Diels
71	Hors_la _L	<i>Horsfieldia laevigata</i> Warb.
72	Hors_la _S	<i>Horsfieldia laevigata</i> Warb.
73	Ints_pa _L	<i>Intsia palembanica</i> Miq.
74	Ints_pa _S	<i>Intsia palembanica</i> Miq.
75	Knem_in _L	<i>Knema intermedia</i> Warb.
76	Knem_in _S	<i>Knema intermedia</i> Warb.
77	Lith_ru _L	<i>Lithocarpus rufovillosus</i> (Markgr.) Rehder
78	Lith_ru _S	<i>Lithocarpus rufovillosus</i> (Markgr.) Rehder
79	Lits_ti _L	<i>Litsea timoriana</i> Span.
80	Lits_ti _S	<i>Litsea timoriana</i> Span.
81	Mani_br _L	<i>Maniltoa brownneoides</i> Harms
82	Mani_br _S	<i>Maniltoa brownneoides</i> Harms
83	Medu_la _L	<i>Medusanthera laxiflora</i> (Miers) R.A.Howard
84	Medu_la _S	<i>Medusanthera laxiflora</i> (Miers) R.A.Howard
85	Meli_el _L	<i>Melicope elleryana</i> (F. Muell.) T.G. Hartley
86	Meli_el _S	<i>Melicope elleryana</i> (F. Muell.) T.G. Hartley
87	Myri_fa _L	<i>Myristica jatua</i> Houtt.
88	Myri_fa _S	<i>Myristica jatua</i> Houtt.
89	Pala_lo _L	<i>Palaquium lobbianum</i> Burck
90	Pala_lo _S	<i>Palaquium lobbianum</i> Burck
91	Para_pr _L	<i>Pararchidendron pruinatum</i> (Benth.) I.C.Nielsen
92	Para_pr _S	<i>Pararchidendron pruinatum</i> (Benth.) I.C.Nielsen
93	Para_ve _L	<i>Parastemon versteeghii</i> Merr. & L.M.Perry
94	Para_ve _S	<i>Parastemon versteeghii</i> Merr. & L.M.Perry
95	Pime_am _L	<i>Pimelodendron amboinicum</i> Hassk.
96	Pime_am _S	<i>Pimelodendron amboinicum</i> Hassk.
97	Plan_ke _L	<i>Planchonella keyensis</i> H.J.Lam
98	Plan_ke _S	<i>Planchonella keyensis</i> H.J.Lam
99	Pome_ac _L	<i>Pometia acuminata</i> Radlk.
100	Pome_ac _S	<i>Pometia acuminata</i> Radlk.
101	Pome_pi _L	<i>Pometia pinnata</i> J.R.Forst. & G.Forst.
102	Pome_pi _S	<i>Pometia pinnata</i> J.R.Forst. & G.Forst.
103	Prun_ar _L	<i>Prunus arborea</i> (Blume) Kalkman
104	Prun_ar _S	<i>Prunus arborea</i> (Blume) Kalkman
105	Prun_ja _L	<i>Prunus javanica</i> (Teijsm. & Binn.) Miq.
106	Prun_ja _S	<i>Prunus javanica</i> (Teijsm. & Binn.) Miq.
107	Rhod_ci _L	<i>Rhodamnia cinerea</i> Jack
108	Rhod_ci _S	<i>Rhodamnia cinerea</i> Jack
109	Siph_ce _L	<i>Siphonodon celastrineus</i> Griff.

No.	Code	Species
110	Siph_ce _S	<i>Siphonodon celastrineus</i> Griff.
111	Siph_sp _L	<i>Siphonodon</i> sp
112	Siph_sp _S	<i>Siphonodon</i> sp
113	Sloa_pu _L	<i>Sloanea pullei</i> O.C.Schmidt ex A.C.Sm.
114	Sloa_pu _S	<i>Sloanea pullei</i> O.C.Schmidt ex A.C.Sm.
115	Ster_sh _L	<i>Sterculia shillinglawii</i> F.Muell.
116	Ster_sh _S	<i>Sterculia shillinglawii</i> F.Muell.
117	Syzy_sp1 _L	<i>Syzygium</i> sp1
118	Syzy_sp1 _S	<i>Syzygium</i> sp1
119	Teij_bo _L	<i>Teijsmanniodendron bogoriense</i> Koord.
120	Teij_bo _S	<i>Teijsmanniodendron bogoriense</i> Koord.
121	Term_co _L	<i>Terminalia copelandi</i> Elmer
122	Term_co _S	<i>Terminalia copelandi</i> Elmer
123	Timo_ca _L	<i>Timonius carii</i> S.P.Darwin
124	Timo_ca _S	<i>Timonius carii</i> S.P.Darwin
125	Tris_ma _L	<i>Tristaniopsis macrosperma</i> (F.Muell.) Peter G.Wilson & J.T.Waterh.
126	Tris_ma _S	<i>Tristaniopsis macrosperma</i> (F.Muell.) Peter G.Wilson & J.T.Waterh.
127	Vati_ra _L	<i>Vatica rassak</i> Blume
128	Vati_ra _S	<i>Vatica rassak</i> Blume
129	Xant_pa _L	<i>Xanthophyllum papuanum</i> Whitmore ex Meijden
130	Xant_pa _S	<i>Xanthophyllum papuanum</i> Whitmore ex Meijden
131	Xant_no _L	<i>Xanthostemon novaguineensis</i> Valetton
132	Xant_no _S	<i>Xanthostemon novaguineensis</i> Valetton

The screenshot shows a web interface for a journal submission. The main content area displays the following information:

- Revised 1**: Submission accepted.
- Notifications**: Three entries from 'Biodiv Editor Decision' dated 2020-08-27 11:04 PM, 2020-08-31 05:11 PM, and 2020-08-31 05:12 PM.
- Reviewer's Attachments**: One attachment titled '07-AUG-2020-Heterospecific and conspecific associations of trees_intrafile.doc' dated August 27, 2020.
- Revisions**: One revision titled 'Article Text_6656-28418-1-5-20200827 REVISION 29 8 2020 OK.docx (2)' dated August 29, 2020.

The taskbar at the bottom shows several open files, including '6656-28418-1-5-2...docx' (verified), '6656-28418-1-5-2...docx' (Failed - network error), '6656-Article Text...docx' (verified), 'Manuscript_Agus...docx' (verified), 'Reviewer remarks...docx' (verified), and 'DEND-00266-2020...pdf' (verified). The system clock shows 14:14 on 20/12/2022.

Bentuk revisi

Heterospecific and conspecific associations of trees in lowland tropical forest of New Guinea

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Abstract. The vegetation in the tropical rainforest of New Guinea consists of a large number of species that interact with each other within and among species. While several studies have attempted to reveal the diversity of flora of New Guinea, little is known about plant communities that develop associations. This study aimed to investigate the associations of tree species in lowland tropical forest in New Guinea. The associations depicted in this study were in the form of conspecific associations (among small and large individuals within same species) and heterospecific (among individuals of different species and divided into under and upper story). We established 48 rectangular plots created in Murkim and Teiraplu as part of Pegunungan Bintang District, Papua Province. Canonical correspondence analysis (CCA) was used to analyze heterospecific and conspecific associations. The results showed that the understory and upper story vegetation had different patterns of heterospecific association. The understory configured three heterospecific associations, consisting of 5, 13, and 90 species, while the upper story formed four heterospecific associations with 4, 8, 11, and 63 species. The analysis of conspecific associations showed of 149 tree species recorded in the study sites, only 66 species that had both small and large individuals, displaying the pattern of conspecific association. Among them 41 species had positive associations while 25 species had negative association. Our findings enrich the knowledge in theoretical ecology of tropical forest, especially in New Guinea.

Keywords: tropical rainforest, Papuasia, canonical correspondence analysis (CCA), tree community, Vegan package.

Running title: associations of New Guinea tree

INTRODUCTION

Tropical rainforest is a complex ecosystem with many interactions between abiotic and biotic factors, particularly among vegetation (Vitousek, 1984; Thomas & Baltzer, 2002; Hunter et al., 2015). This complexity results in the vegetation that consists of many life forms from vertical and horizontal compositions that interact with each other to obtain sunlight, soil nutrients, and water, and to adapt with microclimatic conditions (Slik et al., 2015, 2018; Murdjoko et al., 2016a). The interactions among vegetation have occurred over a long period due to successional process (Fernández-Lugo et al., 2015). Where the vegetation shares the same ecological condition, the morphological and physiological characters become the driving factors of behavior in the natural tropical rainforest (Gustafsson et al., 2016; Johnson et al., 2017).

The interactions among vegetation elements in the tropical rainforest in some cases represent symbiosis and inter and intra-species relationships (Legendre & Fortin, 1989; Magrach et al., 2014). These interactions can be in the form of competition and association. In old tropical rainforest, the interaction occurs intensively due to the absorption of light and water, where both are the primary growth factors (Yamamoto, 2000; Montgomery & Chazdon, 2001). In secondary forest, canopy gap is very open, leading to more light penetrating the forest floor (Itoh et al., 1997; Angelini et al., 2015; Murdjoko et al., 2017).

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The association in vegetation communities can be in the form of conspecific or heterospecific and the form of association determines the pattern of the spatial distribution of forest ecosystems either.

Conspecific association is the interaction of individuals of similar species while heterospecific occurs among different species of vegetation (Zhu et al., 2015; Wang et al., 2018). Conspecific and heterospecific associations occur during the successional process of the tropical rainforest (Farneda et al., 2018). Some studies explained that the association, either the conspecific or the heterospecific could be in a positive or negative pattern (Castilla et al., 2016).

Vegetation is distributed geographically with the diversity and pattern of plant communities that adapt to particular ecological niche (Brummitt, 2001; Pan et al., 2013). Phytogeographic regions, including mainland New Guinea, have been studied for centuries. The vegetation in New Guinea spreads from coastal to high land areas, containing various types of ecosystems (Cámara-Leret & Dennehy, 2019). As the result, New Guinea contains the highest diversity of flora, such as trees, climbers, shrubs, ferns, rattan, etc. (Murdjoko et al., 2016a) in which about 60% of the species are endemic (Cámara-Leret et al., 2020). For example, a forest area in New Guinea consists of a high diversity of tree species with more than 70 species per hectare that could be found (Robiansyah, 2018; Fatem et al., 2020). While recently more and more studies have attempted to reveal the diversity of flora of New Guinea, little is known about plant communities that develop associations among them.

This study aimed to investigate the association of tree species in the lowland tropical forest in New Guinea. The associations depicted in this study were in the form of conspecific associations (among small and large individuals within same species) and heterospecific (among individuals of different species and divided into under and upper story). We hypothesized that the small and large tree species have heterospecific associations within the natural tropical rain forest. This kind of study is important to provide specific contribution of ecological research in the tropical rainforest of Southeast Asia, more specifically the New Guinea region (Brummitt, 2001).

MATERIALS AND METHODS

Study period and area

[This study was conducted] in the northern part of Pegunungan Bintang District, Papua Province, Indonesia (Figure 1). The study sites were located at Murkim (4°0'0.53"S and 140°49'17.24"E) and Teiraplu (3°59'13.46"S and 140°26'0.06"E) at an altitude of 155 m and 233 m above sea level (m asl), respectively. The ecosystem type of the two study sites are categorized as lowland areas where the southern part is bordered with the mountain range and the northern part is bordered with hills while the western and eastern parts are lowlands. Broadleaves and mixed forests are the dominant vegetation in this area, while the soil is grouped as Ultisols and Inceptisol. The climatic conditions are considered to be very humid with average temperature of 25° C for annual, 20.6° C for daily, and 16.3° C for minimum, and with monthly and annual average rainfall of 448.75 mm and 5385 mm, respectively (Kartikasari et al., 2012).

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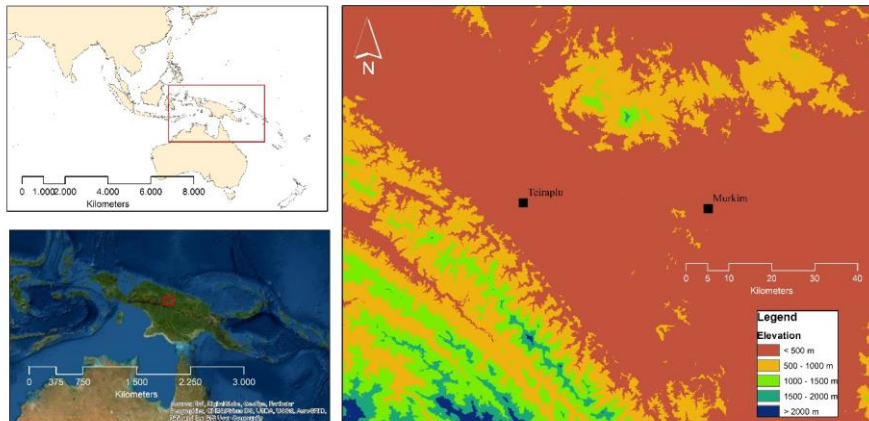


Figure 1. Map of the study sites in Teiraplu and Murkim, Pegunungan Bintang, Papua.

Sampling and data collection

Data were collected using sampling plot method with size of each plot 20 m x 20 m. In total, there were 48 rectangular plots established in which 24 plots were in Teiraplu and 24 plots were in Murkim. In both locations, the plots were placed to north directions at a distance of 100 m away from each other. In the 20 m x 20 m plot (A) we recorded and measured old trees with a diameter of more than 20 cm, and within this plot we established three nested sub-plots with size 10 m x 10 m (B) to record tree with diameter between 10 cm and 20 cm, size 5 m x 5 m (C) to record trees taller than 1.5 m, and size 2 m x 2 m (D) to record the species shorter than 1.5 m. The vegetation in plots A and B were classified as upper story and that in plots C and D were categorized as understory. For the understory vegetation, we recorded data of taxonomic names of every species and number of individuals, and while for the upper story vegetation we recorded data of taxonomic names of every species, number of individuals and diameter (cm).

For identification, we collected the specimens of the plant and sent it to the Herbarium Papuaense of *Balai Penelitian dan Pengembangan Lingkungan Hidup dan Kebutanan (BP2LHK)* Manokwari and Herbarium Manokwariense (MAN) *Pusat Penelitian Keanekaragaman Hayati Universitas Papua (PPKH-UNIPA)*, Manokwari. The species name was updated according to The Plant List (TPL) at the website of <http://www.theplantlist.org/>.

Statistical analysis

The heterospecific and conspecific associations were analyzed using the canonical correspondence analysis (CCA) (Ter Braak, 1986; Caceres & Legendre, 2009), and the chi-square test (χ^2) was implemented to validate the model of CCA (Fatem et al., 2020). Furthermore, this association used the number of each individual (density) as a value in which the columns were the species and the rows were the 48 plots. The conspecific association correlated the under and upper story as small and large individuals. The columns represented the species, while the 48 plots under and upper story represented the rows. The species that did not have under and upper stories were otherwise excluded. The result of CCA displayed species in the graph with the position in the two axes. To investigate the conspecific association whether it was positive or negative, the Euclidean distance between each species as well as the under and upper stories were conducted (Murdjoko et al., 2016b, 2017). If the result of Euclidean distance of species is below the average, then the conspecific association is said to be positive, and vice versa. The vegan package in R version 3.5.3 was used to calculate the statistical analysis (Oksanen et al., 2019).

RESULTS AND DISCUSSION

Heterospecific associations

The heterospecific association was grouped into two: understory and upper story, based on the structure of trees in tropical forests. As such, the analyses of multivariate statistics for the understory and upper story were separated since the natural tropical rainforest is complex with the vegetation structures forming the ecosystem. The structure was also simplified by distinguishing them into two main parts. From the CCA result, the understory and upper story showed different patterns of heterospecific association. The understory configured three groups of communities based on species as the structure of the associations. [The three groups are shown in Figure 2.]

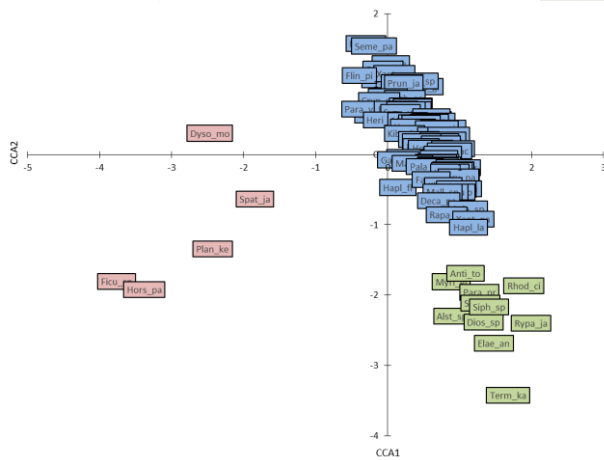


Figure 2. The result of Canonical Correspondence Analysis (CCA) to analyze the heterospecific associations for understory.

The results showed that 108 species of trees formed the association in natural tropical forests, and was valid statistically as $\chi^2 = 10.686$, $df = 2461$, $p\text{-value} = 1$. [The species of understory showed heterospecific associations as tree groups where the first consisted 90 species (blue boxes), the second contained 13 (green boxes), and the third comprised of 5 species (red boxes) (Figure 2).] The name of the species in the boxes in Figure 2 was abbreviated and the complete name can be seen in Appendix 1.

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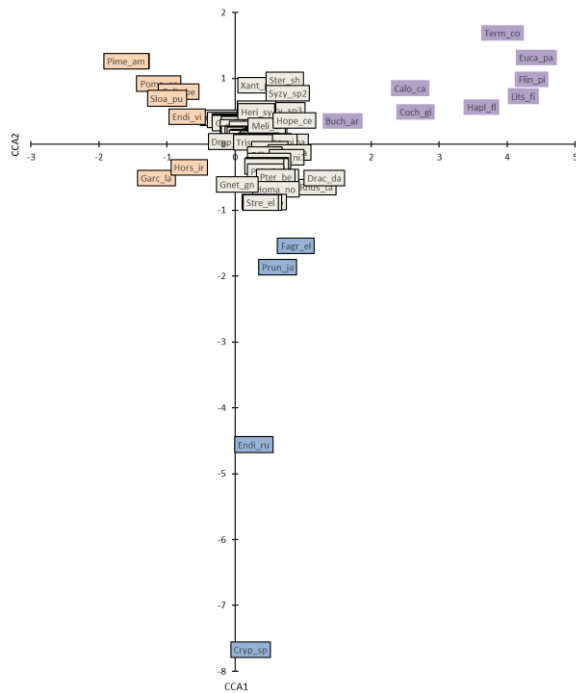


Figure 3. The result of Canonical Correspondence Analysis (CCA) to analyze the heterospecific associations for the upper story.

The CCA result showed that the upper story vegetation community had a pattern of association with a valid result of $\chi^2 = 11.344$, $df = 1955$, $p\text{-value} = 1$. The upper story consisted of 86 species which formed four heterospecific associations, consisting of the first group (63 species) in the grey boxes, second group (11 species) in the red boxes, third group (8 species) in the purple boxes, and fourth group (4 species) in the blues boxes (Figure 3). The complete name of species presented in Figure 3 can be seen in Appendix 2.

The association pattern of the understory and upper story differed from one another even though they grew in the same natural forest. The difference in association has likely resulted from the variation of the vertical structure of the tropical forest. The upper story vegetation has reached the emergent layers of forest canopy, allowing species to benefit by getting more sunlight (Murdjoko et al., 2016a, 2017; Fatem et al., 2020). The formation of understory was caused by competition due to it is below the canopy layers with low solar radiation (Rüger et al., 2011; Laurans et al., 2014; Angelini et al., 2015).

For centuries, the formation of tropical forests has been a sequential process in which large numbers of species compete dynamically each other (Brown et al., 1990; Wright & Muller-Landau, 2006; Liu & Slik, 2014; Almeida et al., 2019). The heterospecific association can be related with the fact that trees interact each other to form symbiosis with other life forms, such as liana, fern, herb, epiphyte and so on (Johnson et al., 2017; Cirimwami et al., 2019; Steege et al., 2019). The primary factor influencing the pattern of tree communities of understory and upper story during tropical forest succession was probably caused by the abiotic factors, especially to gain nutrients, water, and sunlight as materials to support metabolisms, especially photosynthesis. Nonetheless, many studies showed that the morphological and physiological characters have also affected different responses of species to grow and develop (Goodale et al., 2012; Gustafsson et al., 2016). For example, the nature of shade tolerance species may be a factor that

allows small tree species to survive the competition and obtain limited sunlight below the canopy layers (Givnish, 1999; Montesinos-Navarro et al., 2018). Therefore, it is crucial to study the shade-tolerant characters of a species in the rainforest in order to explain forest dynamics in more detail. This study is unable to reveal such characters concerning the light competition because that is beyond the scope of this study.

Conspecific associations

The analysis of conspecific associations was conducted using 149 species that grew in the study sites, but only 66 species that had small and large individuals as understory and upper story. The result of CCA showed statistically valid result as $\chi^2 = 5.8784$, $df = 2904$, $p\text{-value} = 1$ (Figure 4). In addition, it displayed the pattern of conspecific association as 41 species had positive association while 25 species had negative association. In the positive association, the small and large individuals of the 41 species were distributed closely in the same area, representing the tendency of mature trees to reproduce and germinate. Conversely, in the negative association, the small individuals of the 25 species grew mainly far from the large ones that represent the matured trees. The full list of the taxonomic name of the species in Figure 4 is presented in Appendix 3, and the conspecific association can be used to analyze their density dependence since the tropical forest is the place for the high diversity of trees.

Of 149 species, 83 species did not have either small or large individuals, suggesting that the species experienced poor regeneration. Some large individuals act as putative parent trees, even though they have failed to establish seedlings due to many factors (Seidler & Plotkin, 2006; Rahman & Tsukamoto, 2015). One possible factor is caused by the competition of seedlings with other plants in the forest floor, on which many life forms are found. Another rationale is that the seeds and seedlings are eaten by herbivores (Swaine et al., 1987; Houter & Pons, 2014). Many studies have reported that herbivores are found in tropical rainforest since the forest provides a lot of food, for example, during germination, the dicotyledonous tree plants develop shoot from the plumule of the germinating seed (Houter & Pons, 2014; Sawada et al., 2015).

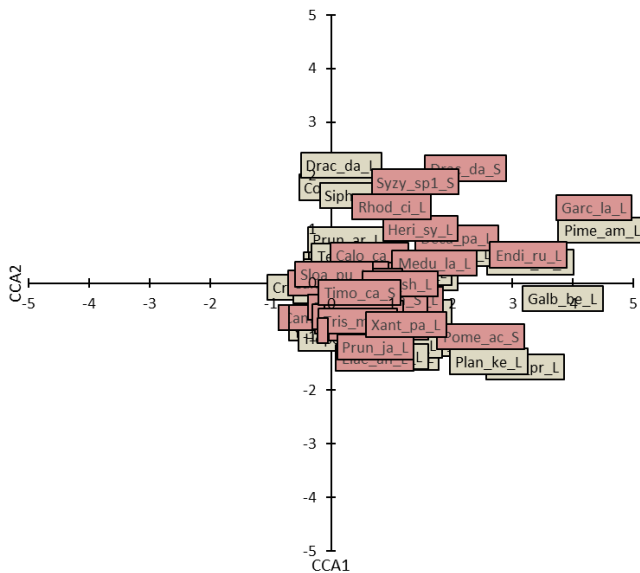


Figure 4. The result of Canonical Correspondence Analysis (CCA) to analyze conspecific associations between the small and the large individuals of the same species.

The distribution of individual trees in tropical forests is influenced by the ability to interact with other species. This pattern of conspecific association should be studied frequently to figure out the method of regeneration and distribution of species. Forest floor encompasses many species with different life forms as a strategy to survive and grow during the competition (Dezzotti et al., 2019). Many lianas and climbers grow fast to occupy the forest canopy and space available for sunlight. These plants suppress a certain seedling establishment (Carreño-Rocabado et al., 2012). The competition to gain sunlight, nutrition, and water is presumed as the limiting factor suffered by some species since they cannot survive below putative parent trees.

Seed dispersal can be the driving force behind the spatial distribution of plants in tropical forests. Moreover, the morphological and anatomical characters of seeds and fruits also influence species regeneration and distribution. For example, small and winged seeds of tree species can spread out by falling around and away from the parent trees (Sebbenn et al., 2008; Lü & Tang, 2010). However, factors such as competition, herbivory, and allelopathy have led to a clear and negative association in natural tropical forests (Padmanaba & Corlett, 2014; Menezes et al., 2019). In contrast, large seeds mostly fall around the parent trees and since they survived germination, they can grow as positive conspecific associations. Therefore, the conspecific association pattern should be studied to know the natural regeneration of certain species in tropical rainforest.

The implication of associations to ecological knowledge for sustainable management of primary forest

The study of conspecific and heterospecific associations in tropical rainforest is extremely important to determine the spatial distribution pattern, especially the conspecific association. In addition, a model of natural regeneration of tree species can be described, and the result can indicate the pattern of recruitment in the population dynamics of tree species (Goodale et al., 2012; Piotta et al., 2019). Tropical rainforest is primarily dominated by flowering plants with their reproduction season is in annual period (Baker et al., 1998; Pan et al., 2013; Cámara-Leret et al., 2020). Furthermore, a suitable area for certain species to grow

has resembled in the conspecific association since the study correlates small individuals with the large ones within the same species. The pattern of conspecific association can also be used to observe natural regeneration. For example, the most appropriate area to plant tree species *in-situ* conservation programs can be decided when artificial regeneration is necessary (Armstrong et al., 2011; Vergara-Rodríguez et al., 2017). The heterospecific association describes the pattern of growth in tropical rainforest since the forest includes the great diversity of tree species. The forest took several decades to develop, and this present study has been able to analyze the pattern of tree species association. Ecological studies on the theme of species association in tropical rainforest need to be replicated in other contexts of region, ecosystem type and forest conditions as tropical forest is very complex as made up of different life forms that interact and create vertical and horizontal structures in the climax phase of the successional process (Chazdon, 2003; Brokaw & Scheiner, 2012).

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Appendix 1. Species name of heterospecific associations for understory for Figure 2.

No.	Code	Species	Group 1	Group 2	Group 3
1	Kiba_co	<i>Kibara coriacea</i> (Blume) Hook. f. & A. Thoms.	√		
2	Rhus_la	<i>Rhus lamprocarpa</i> Merr. & L.M.Perry	√		
3	Dios_pi	<i>Diospyros pilosantha</i> Blanco	√		
4	Lits_le	<i>Litsea ledermannii</i> Tschner	√		
5	Maca_gi	<i>Macaranga gigantea</i> (Rchb.f. & Zoll.) Müll.Arg.	√		
6	Maca_ta	<i>Macaranga tanarius</i> (L.) Müll.Arg.	√		
7	Seme_pa	<i>Semecarpus papuana</i> Lauterb.	√		
8	Ster_sh	<i>Sterculia shillinglawii</i> F.Muell.	√		
9	Case_ca	<i>Casearia carrii</i> Sleumer	√		
10	Xant_no	<i>Wendlandia</i> sp	√		
11	Teij_bo	<i>Teijsmanniodendron bogoriense</i> Koord.	√		
12	Flin_pi	<i>Flindersia pimenteliana</i> F.Muell.	√		
13	Case_sp	<i>Casearia</i> sp	√		
14	Hapl_ce	<i>Haplolobus celebicus</i> H.J.Lam	√		
15	Myri_en	<i>Myristica ensifolia</i> J.Sinclair	√		
16	Octa_in	<i>Octamyrtus insignis</i> Diels	√		
17	Piso_lo	<i>Pisonia longirostris</i> Teijsm. & Binn.	√		
18	Wend_sp	<i>Wendlandia</i> sp	√		
19	Timo_ca	<i>Timonius carrii</i> S.P.Darwin	√		
20	Prun_ja	<i>Prunus javanica</i> (Teijsm. & Binn.) Miq.	√		
21	Hapl_fl	<i>Haplolobus floribundus</i> (K.Schum.) H.J.Lam	√		
22	Anti_de	<i>Alstonia spectabilis</i> R.Br.	√		
23	Buch_ar	<i>Buchanania arborescens</i> (Blume) Blume	√		
24	Cryp_pa	<i>Cryptocarya palmerensis</i> C.K.Allen	√		
25	Gmel_se	<i>Gmelina sessilis</i> C.T.White & W.D.Francis ex Lane-Poole	√		
26	Garc_pi	<i>Garcinia picrobiza</i> Miq.	√		
27	Para_ve	<i>Parastemon versteegii</i> Merr. & L.M.Perry	√		
28	Knem_in	<i>Knema intermedia</i> Warb.	√		
29	Case_mo	<i>Casearia monticola</i> Sleumer	√		
30	Half_ke	<i>Halfordia kendack</i> Guillaumin	√		
31	Syzy_ve	<i>Syzygium versteegii</i> (Lauterb.) Merr. & L.M.Perry	√		
32	Ficu_ro	<i>Ficus robusta</i> Corner	√		
33	Heri_sy	<i>Heritiera sylvatica</i> S.Vidal	√		
34	Cerb_fl	<i>Cerbera floribunda</i> K.Schum.	√		
35	Myri_gl	<i>Myristica globosa</i> Warb.	√		
36	Dryp_gl	<i>Drypetes globosa</i> (Merr.) Pax & K.Hoffm.	√		
37	Phal_ma	<i>Phaleria macrocarpa</i> (Scheff.) Boerl.	√		
38	Cana_ri	<i>Canarium rigidum</i> (Blume) Zipp. ex Miq.	√		
39	Lits_ti	<i>Litsea timoriana</i> Span.	√		
40	Timo_ti	<i>Timonius timon</i> (Spreng.) Merr.	√		
41	Homa_fo	<i>Heritiera sylvatica</i> S.Vidal	√		
42	Endo_me	<i>Endospermum medulosum</i> L.S.Sm.	√		
43	Pome_pi	<i>Pometia pinnata</i> J.R.Forst. & G.Forst.	√		
44	Kiba_bu	<i>Kibara bullata</i> Philipson	√		
45	Drac_da	<i>Dracontomelon dao</i> (Blanco) Merr. & Rolfe	√		
46	Goni_gi	<i>Goniothalamus giganteus</i> Hook.f. & Thomson	√		
47	Nauc_or	<i>Nauclea orientalis</i> (L.) L.	√		
48	Beil_mo	<i>Beilschmedia morobensis</i> Kosterm.	√		
49	Cara_br	<i>Carallia brachiata</i> (Lour.) Merr.	√		
50	Lits_sp	<i>Litsea</i> sp	√		
51	Pime_am	<i>Pimelodendron amboinicum</i> Hassk.	√		
52	Medu_la	<i>Medusanthera laxiflora</i> (Miers) R.A.Howard	√		
53	Myri_fa	<i>Myristica fatua</i> Houtt.	√		
54	Gono_li	<i>Gonocaryum litorale</i> (Blume) Sleumer	√		
55	Lith_ru	<i>Litbocarpus rajavillosus</i> (Markgr.) Rehder	√		
56	Hope_pa	<i>Hopea papuana</i> Diels	√		
57	Dill_pa	<i>Dillenia papuana</i> Martelli	√		
58	Mast_pa	<i>Mastixiodendron pachycladus</i> (K.Schum.) Melch.	√		
59	Pome_ac	<i>Pometia acuminata</i> Radlk.	√		
60	Kiba_el	<i>Kibara elongata</i> A.C.Sm.	√		
61	Clei_pa	<i>Cleistanthus papuanus</i> (Lauterb.) Jabl.	√		

No.	Code	Species	Group 1	Group 2	Group 3
62	Ints_pa	<i>Intsia palembanica</i> Miq.	√		
63	Galb_be	<i>Galbulimima belgraveana</i> (F.Muell.) Sprague	√		
64	Term_co	<i>Terminalia copelandi</i> Elmer	√		
65	Giro_ne	<i>Gironniera nervosa</i> Planch.	√		
66	Mani_pl	<i>Manilloa plurijuga</i> Merr. & L.M.Perry	√		
67	Hope_ce	<i>Hopea celtidifolia</i> Kosterm.	√		
68	Meli_el	<i>Melicope elleryana</i> (F. Muell.) T.G. Hartley	√		
69	Garc_sp	<i>Garcinia</i> sp	√		
70	Pala_lo	<i>Palaquium lobbianum</i> Burck	√		
71	Camp_br	<i>Campnosperma brevipetiolatum</i> Volkens	√		
72	Ster_ma	<i>Sterculia macrophylla</i> Vent.	√		
73	Harp_ca	<i>Harpullia currii</i> Leenh.	√		
74	Hors_la	<i>Horsfieldia laevigata</i> Warb.	√		
75	Arch_pa	<i>Archidendron parviflorum</i> Pulle	√		
76	Chry_pa	<i>Chrysophyllum papuanicum</i> (Pierre ex Dubard) Royen	√		
77	Fagr_ra	<i>Fagraea racemosa</i> Jack	√		
78	Klei_ho	<i>Kleinboria hospita</i> L.	√		
79	Endi_ru	<i>Endiandra rubescens</i> (Blume) Miq.	√		
80	Tris_ma	<i>Tristanopsis macrosperma</i> (F.Muell.) Peter G.Wilson & J.T.Waterh.	√		
81	Cory_la	<i>Corynocarpus laevigatus</i> J.R.Forst. & G.Forst.	√		
82	Cryp_sp	<i>Cryptocarya</i> sp	√		
83	Tabe_au	<i>Tabernaemontana aurantiaca</i> Gaudich.	√		
84	Mall_sp	<i>Mallotus</i> sp	√		
85	Deca_pa	<i>Decaspermum parviflorum</i> (Lam.) A.J.Scott	√		
86	Agla_sp	<i>Aglaia spectabilis</i> (Miq.) S.S.Jain & S.Bennet	√		
87	Rapa_te	<i>Rapanea tempapan</i> P.Royen	√		
88	Calo_ca	<i>Calophyllum caudatum</i> Kaneh. & Hatus.	√		
89	Xant_pa	<i>Xanthophyllum papuanum</i> Whitmore ex Meijden	√		
90	Hapl_la	<i>Haplobolus lanceolatus</i> H.J.Lam ex Leenh.	√		
91	Alst_sp	<i>Alstonia spectabilis</i> R.Br.			√
92	Dios_sp	<i>Diospyros</i> sp			√
93	Elae_an	<i>Elaeocarpus angustifolius</i> Blume			√
94	Rypa_ja	<i>Ryparosa javanica</i> Koord. & Valetton			√
95	Rhod_ci	<i>Rhodammia cinerea</i> Jack			√
96	Myri_gi	<i>Myristica gigantea</i> King			√
97	Anti_to	<i>Antiaris toxicaria</i> Lesch.			√
98	Calo_in	<i>Calophyllum inophyllum</i> L.			√
99	Para_pr	<i>Pararchidendron pruinosum</i> (Benth.) I.C.Nielsen			√
100	Siph_ce	<i>Siphonodon celastrineus</i> Griff.			√
101	Siph_sp	<i>Siphonodon</i> sp			√
102	Maas_gl	<i>Maasia glauca</i> (Hassk.) Mols, Kessler & Rogstad			√
103	Term_ka	<i>Terminalia kaernbachii</i> Warb.			√
104	Spat_ja	<i>Spathiostemon javensis</i> Blume		√	
105	Dyso_mo	<i>Dysoxylum mollissimum</i> Blume		√	
106	Plan_ke	<i>Planchonella keyensis</i> H.J.Lam		√	
107	Ficu_sp	<i>Ficus</i> sp		√	
108	Hors_pa	<i>Horsfieldia parviflora</i> (Roxb.) J.Sinclair		√	

Appendix 2. Species name of heterospecific associations for upper story Figure 3.

No.	Code	Species	Group 1	Group 2	Group 3	Group 4
1	Hors_ir	<i>Horsfieldia irya</i> (Gaertn.) Warb.		√		
2	Garc_la	<i>Garcinia latissima</i> Miq.		√		
3	Alst_sc	<i>Alstonia scholaris</i> (L.) R. Br.		√		
4	Endi_vi	<i>Endiandra virens</i> F.Muell.		√		
5	Sloa_pu	<i>Sloanea pullei</i> O.C.Schmidt ex A.C.Sm.		√		
6	Galb_be	<i>Galbulimima belgraveana</i> (F.Muell.) Sprague		√		
7	Pome_ac	<i>Pometia acuminata</i> Radlk.		√		
8	Chis_ce	<i>Chisocheton ceramicus</i> Miq.		√		
9	Dyso_mo	<i>Dysoxylum mollissimum</i> Blume		√		
10	Endo_me	<i>Endospermum medullosum</i> L.S.Sm.		√		
11	Pime_am	<i>Pimelodendron amboinicum</i> Hassk.		√		
12	Gono_li	<i>Gonocaryum littorale</i> (Blume) Sleumer	√			

No.	Code	Species	Group 1	Group 2	Group 3	Group 4
13	Stre_el	<i>Streblus elongatus</i> (Miq.) Corner	√			
14	Homa_fo	<i>Heritiera sylvatica</i> S.Vidal	√			
15	Gnet_gn	<i>Gnetum gnemon</i> L.	√			
16	Homa_no	<i>Homalanthus novoguineensis</i> (Warb.) K.Schum.	√			
17	Cory_la	<i>Corynocarpus laevigatus</i> J.R.Forst. & G.Forst.	√			
18	Drac_da	<i>Dracontomelon dao</i> (Blanco) Merr. & Rolfe	√			
19	Cana_od	<i>Canarium indicum</i> L.	√			
20	Pter_be	<i>Pterocymbium beccarii</i> K.Schum.	√			
21	Pala_lo	<i>Palaquium lobbianum</i> Burck	√			
22	Cana_in	<i>Canarium indicum</i> L.	√			
23	Cara_br	<i>Carallia brachiata</i> (Lour.) Merr.	√			
24	Timo_ca	<i>Timonius carii</i> S.P.Darwin	√			
25	Hors_sy	<i>Horsfieldia sylvestris</i> Warb.	√			
26	Maas_su	<i>Maasia sumatrana</i> (Miq.) Mols, Kessler & Rogstad	√			
27	Hope_pa	<i>Hopea papuana</i> Diels	√			
28	Hope_ce	<i>Hopea cellidifolia</i> Kosterm.	√			
29	Rhus_ta	<i>Rhus taiensis</i> Guill.	√			
30	Acti_ni	<i>Actinodaphne nitida</i> Teschner	√			
31	Dill_pa	<i>Dillenia papuana</i> Martelli	√			
32	Medu_la	<i>Medusanthura lasziflora</i> (Miers) R.A.Howard	√			
33	Teij_bo	<i>Teijsmanniodendron bogoriense</i> Koord.	√			
34	Tris_ma	<i>Tristaniaopsis macrosperma</i> (F.Muell.) Peter G.Wilson & J.T.Waterh.	√			
35	Call_lo	<i>Callicarpa longifolia</i> Lam.	√			
36	Comm_ba	<i>Commersonia bartramia</i> (L.) Merr.	√			
37	Dios_pi	<i>Diospyros pilosanthera</i> Blanco	√			
38	Knem_in	<i>Knema intermedia</i> Warb.	√			
39	Dryp_gl	<i>Drypetes globosa</i> (Merr.) Pax & K.Hoffm.	√			
40	Cryp_pa	<i>Cryptocarya palmerensis</i> C.K.Allen	√			
41	Meli_el	<i>Melicope elleryana</i> (F. Muell.) T.G. Hartley	√			
42	Lits_ti	<i>Litsea timoriana</i> Span.	√			
43	Siph_ce	<i>Siphonodon celastrineus</i> Griff.	√			
44	Siph_sp	<i>Siphonodon</i> sp	√			
45	Vite_pi	<i>Vitex pinnata</i> L.	√			
46	Poly_no	<i>Polyscias nodosa</i> (Blume) Seem.	√			
47	Pome_pi	<i>Pometia pinnata</i> J.R.Forst. & G.Forst.	√			
48	Agla_ar	<i>Aglaia argentea</i> Blume	√			
49	Acro_sp	<i>Acronychia</i> sp	√			
50	Gmel_se	<i>Gmelina sessilis</i> C.T.White & W.D.Francis ex Lane-Poole	√			
51	Mani_br	<i>Maniltoa bronneoides</i> Harms	√			
52	Prun_ar	<i>Prunus arborea</i> (Blume) Kalkman	√			
53	Camp_br	<i>Campnosperma brevipetiolatum</i> Volkens	√			
54	Hors_la	<i>Horsfieldia laevigata</i> Warb.	√			
55	Cana_hi	<i>Campnosperma brevipetiolatum</i> Volkens	√			
56	Deca_pa	<i>Decaspermum parviflorum</i> (Lam.) A.J.Scott	√			
57	Calo_in	<i>Calophyllum inophyllum</i> L.	√			
58	Heri_sy	<i>Heritiera sylvatica</i> S.Vidal	√			
59	Clei_pa	<i>Cleistanthus papuanus</i> (Lauterb.) Jabl.	√			
60	Elae_an	<i>Elaeocarpus angustifolius</i> Blume	√			
61	Gymn_fa	<i>Gymnacranthera farquhariana</i> (Hook.f. & Thomson) Warb.	√			
62	Grew_er	<i>Grewia eriocarpa</i> Juss.	√			
63	Xant_no	<i>Wendlandia</i> sp	√			
64	Rhod_ci	<i>Rhodamnia cinerea</i> Jack	√			
65	Arto_al	<i>Artocarpus alilis</i> (Parkinson ex F.A.Zorn) Fosberg	√			
66	Para_pr	<i>Pararchidendron pruinosum</i> (Benth.) I.C.Nielsen	√			
67	Plan_ke	<i>Planchonella keyensis</i> H.J.Lam	√			
68	Dios_pa	<i>Diospyros papuana</i> Valetton ex Bakh.	√			
69	Ochr_gl	<i>Ochrosia glomerata</i> (Blume) F.Muell.	√			
70	Myri_fa	<i>Myristica fatua</i> Houtt.	√			
71	Ster_sh	<i>Sterculia shillinghamii</i> F.Muell.	√			
72	Syzy_sp2	<i>Syzygium</i> sp2	√			
73	Syzy_sp3	<i>Syzygium</i> sp3	√			
74	Xant_pa	<i>Xanthophyllum papuanum</i> Whitmore ex Meijden	√			
75	Euca_pa	<i>Eucalyptopsis papuana</i> C.T.White		√		

No.	Code	Species	Group 1	Group 2	Group 3	Group 4
76	Flin_pi	<i>Flindersia pimenteliana</i> F.Muell.			√	
77	Hapl_fl	<i>Haplolobus floribundus</i> (K.Schum.) H.J.Lam			√	
78	Lits_fi	<i>Litsea firma</i> (Blume) Hook.f.			√	
79	Term_co	<i>Terminalia copelandi</i> Elmer			√	
80	Calo_ca	<i>Calophyllum caudatum</i> Kaneh. & Hatus.			√	
81	Coch_gi	<i>Cochlospermum gillivraei</i> Benth.			√	
82	Buch_ar	<i>Buchanania arborescens</i> (Blume) Blume			√	
83	Fagr_el	<i>Fagraea elliptica</i> Roxb.				√
84	Prun_ja	<i>Prunus javanica</i> (Teijsm. & Binn.) Miq.				√
85	Endi_ru	<i>Endiandra rubescens</i> (Blume) Miq.				√
86	Cryp_sp	<i>Cryptocarya</i> sp				√

Appendix 3. Species name of conspecific associations Figure 4. The S stands for small individuals and L symbolizes large individuals.

No.	Code	Species
1	Agla_ar _L	<i>Aglaia argentea</i> Blume
2	Agla_ar _S	<i>Aglaia argentea</i> Blume
3	Buch_ar _L	<i>Buchanania arborescens</i> (Blume) Blume
4	Buch_ar _S	<i>Buchanania arborescens</i> (Blume) Blume
5	Calo_ca _L	<i>Calophyllum caudatum</i> Kaneh. & Hatus.
6	Calo_ca _S	<i>Calophyllum caudatum</i> Kaneh. & Hatus.
7	Calo_in _L	<i>Calophyllum inophyllum</i> L.
8	Calo_in _S	<i>Calophyllum inophyllum</i> L.
9	Camp_br _L	<i>Campnosperma brevipetiolatum</i> Volkens
10	Camp_br _S	<i>Campnosperma brevipetiolatum</i> Volkens
11	Cana_hi _L	<i>Canarium hirsutum</i> Willd.
12	Cana_hi _S	<i>Canarium hirsutum</i> Willd.
13	Cana_in _L	<i>Canarium indicum</i> L.
14	Cana_in _S	<i>Canarium indicum</i> L.
15	Cara_br _L	<i>Carallia brachiata</i> (Lour.) Merr.
16	Cara_br _S	<i>Carallia brachiata</i> (Lour.) Merr.
17	Clei_pa _L	<i>Cleistanthus papuanus</i> (Lauterb.) Jabl.
18	Clei_pa _S	<i>Cleistanthus papuanus</i> (Lauterb.) Jabl.
19	Cory_la _L	<i>Corynocarpus laevigatus</i> J.R.Forst. & G.Forst.
20	Cory_la _S	<i>Corynocarpus laevigatus</i> J.R.Forst. & G.Forst.
21	Cryp_pa _L	<i>Cryptocarya palmerensis</i> C.K.Allen
22	Cryp_pa _S	<i>Cryptocarya palmerensis</i> C.K.Allen
23	Cryp_sp _L	<i>Cryptocarya</i> sp
24	Cryp_sp _S	<i>Cryptocarya</i> sp
25	Deca_pa _L	<i>Decaspermum parviflorum</i> (Lam.) A.J.Scott
26	Deca_pa _S	<i>Decaspermum parviflorum</i> (Lam.) A.J.Scott
27	Dill_pa _L	<i>Dillenia papuana</i> Martelli
28	Dill_pa _S	<i>Dillenia papuana</i> Martelli
29	Dios_pi _L	<i>Diospyros pilosanthera</i> Blanco
30	Dios_pi _S	<i>Diospyros pilosanthera</i> Blanco
31	Drac_da _L	<i>Dracontomelon dao</i> (Blanco) Merr. & Rolfe
32	Drac_da _S	<i>Dracontomelon dao</i> (Blanco) Merr. & Rolfe
33	Dryp_gl _L	<i>Drypetes globosa</i> (Merr.) Pax & K.Hoffm.
34	Dryp_gl _S	<i>Drypetes globosa</i> (Merr.) Pax & K.Hoffm.
35	Dyso_mo _L	<i>Dysoxylum mollissimum</i> Blume
36	Dyso_mo _S	<i>Dysoxylum mollissimum</i> Blume
37	Elae_an _L	<i>Elaeocarpus angustifolius</i> Blume
38	Elae_an _S	<i>Elaeocarpus angustifolius</i> Blume
39	Endi_ru _L	<i>Endiandra rubescens</i> (Blume) Miq.
40	Endi_ru _S	<i>Endiandra rubescens</i> (Blume) Miq.
41	Endo_me _L	<i>Endospermum medulosum</i> L.S.Sm.
42	Endo_me _S	<i>Endospermum medulosum</i> L.S.Sm.
43	Flin_pi _L	<i>Flindersia pimenteliana</i> F.Muell.
44	Flin_pi _S	<i>Flindersia pimenteliana</i> F.Muell.
45	Galb_be _L	<i>Galbulimima belgraveana</i> (F.Muell.) Sprague
46	Galb_be _S	<i>Galbulimima belgraveana</i> (F.Muell.) Sprague

No.	Code	Species
47	Garc_la _L	<i>Garcinia latissima</i> Miq.
48	Garc_la _S	<i>Garcinia latissima</i> Miq.
49	Giro_ne _L	<i>Gironniera nervosa</i> Planch.
50	Giro_ne _S	<i>Gironniera nervosa</i> Planch.
51	Gmel_se _L	<i>Gmelina sessilis</i> C.T.White & W.D.Francis ex Lane-Poole
52	Gmel_se _S	<i>Gmelina sessilis</i> C.T.White & W.D.Francis ex Lane-Poole
53	Gnet_gn _L	<i>Gnetum gnemon</i> L.
54	Gnet_gn _S	<i>Gnetum gnemon</i> L.
55	Gono_li _L	<i>Gonocaryum litorale</i> (Blume) Sleumer
56	Gono_li _S	<i>Gonocaryum litorale</i> (Blume) Sleumer
57	Gymn_fa _L	<i>Gymnacranthera jarqubariana</i> (Hook.f. & Thomson) Warb.
58	Gymn_fa _S	<i>Gymnacranthera jarqubariana</i> (Hook.f. & Thomson) Warb.
59	Hapl_fl _L	<i>Haplolobus floribundus</i> (K.Schum.) H.J.Lam
60	Hapl_fl _S	<i>Haplolobus floribundus</i> (K.Schum.) H.J.Lam
61	Heri_sy _L	<i>Heritiera sylvatica</i> S.Vidal
62	Heri_sy _S	<i>Heritiera sylvatica</i> S.Vidal
63	Homa_fo _L	<i>Homalium foetidum</i> Benth.
64	Homa_fo _S	<i>Homalium foetidum</i> Benth.
65	Hope_ce _L	<i>Hopea celtidifolia</i> Kosterm.
66	Hope_ce _S	<i>Hopea celtidifolia</i> Kosterm.
67	Hope_no _L	<i>Hopea novoguineensis</i> Slooten
68	Hope_no _S	<i>Hopea novoguineensis</i> Slooten
69	Hope_pa _L	<i>Hopea papuana</i> Diels
70	Hope_pa _S	<i>Hopea papuana</i> Diels
71	Hors_la _L	<i>Horsfieldia laevigata</i> Warb.
72	Hors_la _S	<i>Horsfieldia laevigata</i> Warb.
73	Ints_pa _L	<i>Intsia palembanica</i> Miq.
74	Ints_pa _S	<i>Intsia palembanica</i> Miq.
75	Knem_in _L	<i>Knema intermedia</i> Warb.
76	Knem_in _S	<i>Knema intermedia</i> Warb.
77	Lith_ru _L	<i>Lithocarpus rufovillosus</i> (Markgr.) Rehder
78	Lith_ru _S	<i>Lithocarpus rufovillosus</i> (Markgr.) Rehder
79	Lits_ti _L	<i>Litsea timoriana</i> Span.
80	Lits_ti _S	<i>Litsea timoriana</i> Span.
81	Mani_br _L	<i>Maniltoa brownneoides</i> Harms
82	Mani_br _S	<i>Maniltoa brownneoides</i> Harms
83	Medu_la _L	<i>Medusanthera laxiflora</i> (Miers) R.A.Howard
84	Medu_la _S	<i>Medusanthera laxiflora</i> (Miers) R.A.Howard
85	Meli_el _L	<i>Melicope elleryana</i> (F. Muell.) T.G. Hartley
86	Meli_el _S	<i>Melicope elleryana</i> (F. Muell.) T.G. Hartley
87	Myri_fa _L	<i>Myristica jatua</i> Houtt.
88	Myri_fa _S	<i>Myristica jatua</i> Houtt.
89	Pala_lo _L	<i>Palaquium lobbianum</i> Burck
90	Pala_lo _S	<i>Palaquium lobbianum</i> Burck
91	Para_pr _L	<i>Pararchidendron pruinatum</i> (Benth.) I.C.Nielsen
92	Para_pr _S	<i>Pararchidendron pruinatum</i> (Benth.) I.C.Nielsen
93	Para_ve _L	<i>Parastemon versteeghii</i> Merr. & L.M.Perry
94	Para_ve _S	<i>Parastemon versteeghii</i> Merr. & L.M.Perry
95	Pime_am _L	<i>Pimelodendron amboinicum</i> Hassk.
96	Pime_am _S	<i>Pimelodendron amboinicum</i> Hassk.
97	Plan_ke _L	<i>Planchonella keyensis</i> H.J.Lam
98	Plan_ke _S	<i>Planchonella keyensis</i> H.J.Lam
99	Pome_ac _L	<i>Pometia acuminata</i> Radlk.
100	Pome_ac _S	<i>Pometia acuminata</i> Radlk.
101	Pome_pi _L	<i>Pometia pinnata</i> J.R.Forst. & G.Forst.
102	Pome_pi _S	<i>Pometia pinnata</i> J.R.Forst. & G.Forst.
103	Prun_ar _L	<i>Prunus arborea</i> (Blume) Kalkman
104	Prun_ar _S	<i>Prunus arborea</i> (Blume) Kalkman
105	Prun_ja _L	<i>Prunus javanica</i> (Teijsm. & Binn.) Miq.
106	Prun_ja _S	<i>Prunus javanica</i> (Teijsm. & Binn.) Miq.
107	Rhod_ci _L	<i>Rhodamnia cinerea</i> Jack
108	Rhod_ci _S	<i>Rhodamnia cinerea</i> Jack
109	Siph_ce _L	<i>Siphonodon celastrineus</i> Griff.

No.	Code	Species
110	Siph_ce _S	<i>Siphonodon celastrineus</i> Griff.
111	Siph_sp _L	<i>Siphonodon</i> sp
112	Siph_sp _S	<i>Siphonodon</i> sp
113	Sloa_pu _L	<i>Sloanea pullei</i> O.C.Schmidt ex A.C.Sm.
114	Sloa_pu _S	<i>Sloanea pullei</i> O.C.Schmidt ex A.C.Sm.
115	Ster_sh _L	<i>Sterculia shillinglawii</i> F.Muell.
116	Ster_sh _S	<i>Sterculia shillinglawii</i> F.Muell.
117	Syzy_sp1 _L	<i>Syzygium</i> sp1
118	Syzy_sp1 _S	<i>Syzygium</i> sp1
119	Teij_bo _L	<i>Teijsmanniodendron bogoriense</i> Koord.
120	Teij_bo _S	<i>Teijsmanniodendron bogoriense</i> Koord.
121	Term_co _L	<i>Terminalia copelandi</i> Elmer
122	Term_co _S	<i>Terminalia copelandi</i> Elmer
123	Timo_ca _L	<i>Timonius carii</i> S.P.Darwin
124	Timo_ca _S	<i>Timonius carii</i> S.P.Darwin
125	Tris_ma _L	<i>Tristaniopsis macrosperma</i> (F.Muell.) Peter G.Wilson & J.T.Waterh.
126	Tris_ma _S	<i>Tristaniopsis macrosperma</i> (F.Muell.) Peter G.Wilson & J.T.Waterh.
127	Vati_ra _L	<i>Vatica rassak</i> Blume
128	Vati_ra _S	<i>Vatica rassak</i> Blume
129	Xant_pa _L	<i>Xanthophyllum papuanum</i> Whitmore ex Meijden
130	Xant_pa _S	<i>Xanthophyllum papuanum</i> Whitmore ex Meijden
131	Xant_no _L	<i>Xanthostemon novaguineensis</i> Valetton
132	Xant_no _S	<i>Xanthostemon novaguineensis</i> Valetton

Keputusan akhir journal

The screenshot shows a web browser window with a notification pop-up from Biodiversity International. The notification is titled "[biodiv] Editor Decision" and is dated 2020-08-31 05:11 PM. The notification text reads: "AGUSTINUS MURDIOKO, MARTINUS WATHAGE, ITAMAU DONY ARISTONE, ESTIMAU, RIMA HERLINA, SETAWATI, TUBIRAN, ANTONI UDORINJAU, IGNEDEO OTTOY WAMAMA, ZULFIKAR, INDEPRADO, ALEXANDER KUMATORA, MOYIBAN NARHA, NODI, ANTONI SASA SWARI, SERUS MARTIN, PATRI, ESCARDO, WIKABAN, NUNANDI ANAK, MAY, MAX JONKUDASO, THODEE, HERANU, IBRAMEYAL, CHARLY BRAGO, WAKOGAL, JIMMY FRANS WAMAMA, EUSEB, VICTOR ISABE, ZWANA BETY, FERNANDO, ERNI, WANDOK, BELANAN KUMARADI, DEBANA, LINTO, USMA, KHARIT, NITHANIEL, MIRAL, HENDRIK BENU, JUNUS, TAMBING, ANDI, SASTRA BENNY, SARAGIH." Below this, it states: "We have reached a decision regarding your submission to Biodiversity: Journal of Biological Diversity, 'Heterospecific and conspecific associations of trees in lowland tropical forest of New Guinea'." The decision is to "Accept Submission". The editor's name and email address are listed as "Drugi Editor" and "editor@biodiv.int". The notification also includes a "Show all" button and a "Close" button.

Heterospecific and conspecific associations of trees in lowland tropical forest of New Guinea

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Abstract. Murdjoko A, Jitmau MM, Djitmau DA, Siburian R, Ungirwalu A, Wanma AO, Mardiyadi Z, Rumatora A, Mofu WY, Sineri AS, Fatem SM, Worabai D, May NL, Tokede MJ, Warmetan H, Wanggai CB, Wanma JF, Sirami EV, Paemonan JB, Unenor E, Kuswandi R, Lekitoo K, Khayati L, Benu NMH, Tambing J, Saragih ASB. 2020. Heterospecific and conspecific associations of trees in lowland tropical forest of New Guinea. *Biodiversitas* 21: xxxx. The vegetation in the tropical rainforest of New Guinea consists of a large number of species that interact with each other within and among species. While several studies have attempted to reveal the diversity of flora of New Guinea, little is known about plant communities that develop associations. This study aimed to investigate the associations of tree species in lowland tropical forest in New Guinea. The associations depicted in this study were in the form of conspecific associations (among small and large individuals within same species) and heterospecific (among individuals of different species and divided into under and upper story). We established 48 rectangular plots created in Murkim and Teiraplu as part of Pegunungan Bintang Regency, Papua Province. Canonical correspondence analysis (CCA) was used to analyze heterospecific and conspecific associations. The results showed that the understory and upper story vegetation had different patterns of heterospecific association. The understory configured three heterospecific associations, consisting of 5, 13, and 90 species, while the upper story formed four heterospecific associations with 4, 8, 11, and 63 species. The analysis of conspecific associations showed of 149 tree species recorded in the study sites, only 66 species that had both small and large individuals, displaying the pattern of conspecific association. Among them, 41 species had positive associations while 25 species had negative association. Our findings enrich the knowledge in theoretical ecology of tropical forest, especially in New Guinea.

Keywords: canonical correspondence analysis (CCA), Papuaia, tree community, tropical rainforest, vegan package

INTRODUCTION

Tropical rainforest is a complex ecosystem with many interactions between abiotic and biotic factors, particularly among vegetation (Vitousek 1984; Thomas & Baltzer 2002; Hunter et al. 2015). This complexity results in the vegetation that consists of many life forms from vertical and horizontal compositions that interact with each other to obtain sunlight, soil nutrients, and water,

and to adapt with microclimatic conditions (Slik et al. 2015, 2018; Murdjoko et al. 2016a). The interactions among vegetation have occurred over a long period due to successional process (Fernández-Lugo et al. 2015). Where the vegetation shares the same ecological condition, the morphological and physiological characters become the driving factors of behavior in the natural tropical rainforest (Gustafsson et al. 2016; Johnson et al. 2017).

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The interactions among vegetation elements in the tropical rainforest in some cases represent symbiosis and inter and intra-species relationships (Legendre & Fortin 1989; Magrath et al. 2014). These interactions can be in the form of competition and association. In old tropical rainforest, the interaction occurs intensively due to the absorption of light and water, where both are the primary growth factors (Yamamoto 2000; Montgomery & Chazdon 2001). In secondary forest, canopy gap is very open, leading to more light penetrating the forest floor (Itoh et al. 1997; Angelini et al. 2015; Murdjoko et al. 2017). The association in vegetation communities can be in the form of conspecific or heterospecific and the form of association determines the pattern of the spatial distribution of forest ecosystems either. Conspecific association is the interaction of individuals of similar species while heterospecific occurs among different species of vegetation (Zhu et al. 2015; Wang et al. 2018). Conspecific and heterospecific associations occur during the successional process of the tropical rainforest (Farneda et al. 2018). Some studies explained that the association, either the conspecific or the heterospecific could be in a positive or negative pattern (Castilla et al. 2016). Vegetation is distributed geographically with the diversity and pattern of plant communities that adapt to particular ecological niche (Brummitt 2001; Pan et al. 2013). Phytogeographic regions, including mainland New Guinea, have been studied for centuries. The vegetation in New Guinea spreads from coastal to high land areas, containing various types of ecosystems (Cámara-Leret & Dennehy 2019). As the result, New Guinea contains the highest diversity of flora, such as trees, climbers, shrubs, ferns, rattan, etc. (Murdjoko et al. 2016a) in which about 60% of the species are endemic (Cámara-Leret et al. 2020). For example, a forest area in New Guinea consists of a high diversity of tree species with more than 70 species per hectare that could be found (Robiansyah 2018; Fatem et al. 2020). While recently more and more studies have attempted to reveal the diversity of flora of New Guinea, little is known about plant communities that develop associations among them.

This study aimed to investigate the association of tree species in the lowland tropical forest in New Guinea. The associations depicted in this study were in the form of conspecific associations (among small and large individuals within same species) and heterospecific (among individuals of different species and divided into under and upper story). We hypothesized that the small and large tree species have heterospecific associations within the natural tropical rain forest. This kind of study is important to provide specific contribution of ecological research in the tropical rainforest of Southeast Asia, more specifically the New Guinea region (Brummitt 2001).

MATERIALS AND METHODS

Study period and area

This study was conducted in the northern part of Pegunungan Bintang Regency, Papua Province, Indonesia (Figure 1). The study sites were located at Murkim (4°0'0.53"S and 140°49'17.24"E) and Teiraplu (3°59'13.46"S and 140°26'0.06"E) at an altitude of 155 m and 233 m above sea level (m asl), respectively. The ecosystem type of the two study sites are categorized as lowland areas where the southern part is bordered with the mountain range and the northern part is bordered with hills while the western and eastern parts are lowlands. Broadleaves and mixed forests are the dominant vegetation in this area, while the soil is grouped as Ultisols and Inceptisol. The climatic conditions are considered to be very humid with average temperature of 25° C for annual, 20.6° C for daily, and 16.3° C for minimum, and with monthly and annual average rainfall of 448.75 mm and 5385 mm, respectively (Kartikasari et al. 2012).

Sampling and data collection

Data were collected using sampling plot method with size of each plot 20 m x 20 m. In total, there were 48 rectangular plots established in which 24 plots were in Teiraplu and 24 plots were in Murkim. In both locations, the plots were placed to north directions at a distance of 100 m away from each other. In the 20 m x 20 m plot (A) we recorded and measured old trees with a diameter of more than 20 cm, and within this plot we

established three nested sub-plots with size 10 m x 10 m (B) to record tree with diameter between 10 cm and 20 cm, size 5 m x 5 m (C) to record trees taller than 1.5 m, and size 2 m x 2 m (D) to record the species shorter than 1.5 m. The vegetation in plots A and B were classified as upper story and that in plots C and D were categorized as understory. For the understory vegetation, we recorded data of taxonomic names of every species and number of individuals, and while for the upper story vegetation we recorded data of taxonomic names of every species, number of individuals, and diameter (cm).

For identification, we collected the specimens of the plant and sent it to the Herbarium Papuense of *Balai Penelitian dan Pengembangan Lingkungan Hidup dan Kebutuhan (BP2LHK)* Manokwari and Herbarium Manokwariense (MAN) *Pusat Penelitian Keanekaragaman Hayati Universitas Papua (PPKH-UNIPA)*, Manokwari. The species name was updated according to The Plant List (TPL) at the website of <http://www.theplantlist.org/>.

Statistical analysis

The heterospecific and conspecific associations were analyzed using the canonical correspondence

analysis (CCA) (Ter Braak 1986; Caceres & Legendre 2009), and the chi-square test (χ^2) was implemented to validate the model of CCA (Fatem et al. 2020). Furthermore, this association used the number of each individual (density) as a value in which the columns were the species and the rows were the 48 plots. The conspecific association correlated the under and upper story as small and large individuals. The columns represented the species, while the 48 plots under and upper story represented the rows. The species that did not have under and upper stories were otherwise excluded. The result of CCA displayed species in the graph with the position in the two axes. To investigate the conspecific association whether it was positive or negative, the Euclidean distance between each species as well as the under and upper stories were conducted (Murdjoko et al. 2016b, 2017). If the result of Euclidean distance of species is below the average, then the conspecific association is said to be positive, and vice versa. The vegan package in R version 3.5.3 was used to calculate the statistical analysis (Oksanen et al. 2019).

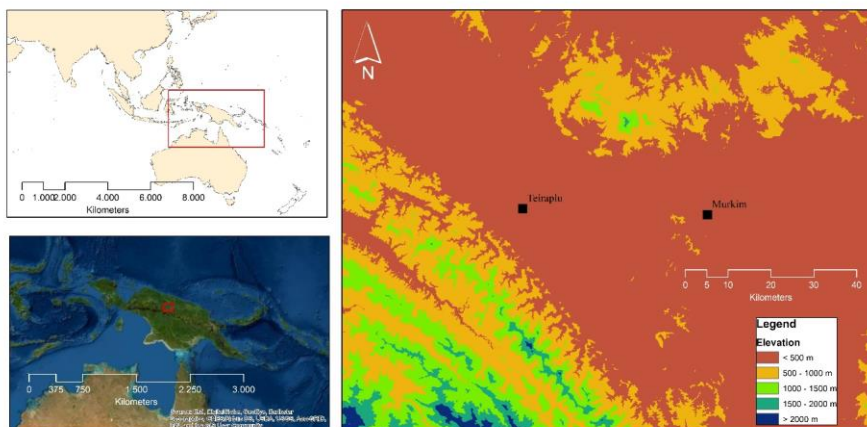


Figure 1. Map of the study sites in Teiraplu and Murkim, Pegunungan Bintang, Papua.

RESULTS AND DISCUSSION

Heterospecific associations

The heterospecific association was grouped into two: understory and upper story, based on the structure of trees in tropical forests. As such, the analyses of multivariate statistics for the understory and upper story were separated since the natural tropical rainforest is complex with the vegetation structures forming the ecosystem. The structure was also simplified by distinguishing them into two main parts.

From the CCA result, the understory and upper story showed different patterns of heterospecific association. The understory configured three groups of communities based on species as the structure of the associations. The three groups are shown in Figure 2.

The results showed that 108 species of trees formed the association in natural tropical forests, and was valid statistically as $\chi^2 = 10.686$, $df = 2461$, $p\text{-value} = 1$. The species of understory showed heterospecific associations as tree groups where the first consisted of 90 species (blue boxes), the second contained 13 (green boxes), and the third comprised of 5 species (red boxes) (Figure 2). The name of the species in the boxes in Figure 2 was abbreviated and the complete name can be seen in Appendix 1.

The CCA result showed that the upper story vegetation community had a pattern of association with a valid result of $\chi^2 = 11.344$, $df = 1955$, $p\text{-value} = 1$. The upper story consisted of 86 species which formed four heterospecific associations, consisting of the first group (63 species) in the grey boxes, second group (11 species) in the red boxes, third group (8 species) in the purple boxes, and fourth group (4 species) in the blue boxes (Figure 3). The complete name of species presented in Figure 3 can be seen in Appendix 2.

The association pattern of the understory and upper story differed from one another even though they grew in the same natural forest. The difference in association has likely resulted from the variation of the vertical structure of the tropical forest. The upper story vegetation has reached the emergent layers of forest canopy, allowing species

to benefit by getting more sunlight (Murdjoko et al. 2016a, 2017; Fatem et al. 2020). The formation of understory was caused by competition due to it is below the canopy layers with low solar radiation (Rüger et al. 2011; Laurans et al. 2014; Angelini et al. 2015).

For centuries, the formation of tropical forests has been a sequential process in which large numbers of species compete dynamically each other (Brown et al. 1990; Wright & Muller-Landau 2006; Liu & Slik 2014; Almeida et al. 2019). The heterospecific association can be related to the fact that trees interact with each other to form symbiosis with other life forms, such as liana, fern, herb, epiphyte and so on (Johnson et al. 2017; Cirimwami et al. 2019; Steege et al. 2019). The primary factor influencing the pattern of tree communities of understory and upper story during tropical forest succession was probably caused by the abiotic factors, especially to gain nutrients, water, and sunlight as materials to support metabolisms, especially photosynthesis. Nonetheless, many studies showed that the morphological and physiological characters have also affected different responses of species to grow and develop (Goodale et al. 2012; Gustafsson et al. 2016). For example, the nature of shade tolerance species may be a factor that allows small tree species to survive the competition and obtain limited sunlight below the canopy layers (Givnish 1999; Montesinos-Navarro et al. 2018). Therefore, it is crucial to study the shade-tolerant characters of a species in the rainforest in order to explain forest dynamics in more detail. This study is unable to reveal such characters concerning the light competition because that is beyond the scope of this study.

Conspecific associations

The analysis of conspecific associations was conducted using 149 species that grew in the study sites, but only 66 species that had small and large individuals as understory and upper story. The result of CCA showed statistically valid result as $\chi^2 = 5.8784$, $df = 2904$, $p\text{-value} = 1$ (Figure 4). In addition, it displayed the pattern of conspecific association as 41 species had positive association

while 25 species had negative association. In the positive association, the small and large individuals of the 41 species were distributed closely in the same area, representing the tendency of mature trees to reproduce and germinate. Conversely, in the negative association, the small individuals of the 25 species grew mainly far from the large ones that represent the matured trees. The full list of the taxonomic name of the species in Figure 4 is presented in Appendix 3, and the conspecific association can be used to analyze their density dependence since the tropical forest is the place for the high diversity of trees.

Of 149 species, 83 species did not have either small or large individuals, suggesting that the species experienced poor regeneration. Some large individuals act as putative parent trees, even though they have failed to establish seedlings due to many factors (Seidler & Plotkin 2006; Rahman & Tsukamoto 2015). One possible factor is caused by the competition of seedlings with other plants on the forest floor, on which many life forms are found. Another rationale is that the seeds and seedlings are eaten by herbivores (Swaine et al. 1987; Houter & Pons 2014). Many studies have reported that herbivores are found in tropical rainforest since the forest provides a lot of food, for example, during germination, the dicotyledonous tree plants develop shoot from the plumule of the germinating seed (Houter & Pons 2014; Sawada et al. 2015).

The distribution of individual trees in tropical forests is influenced by the ability to interact with

other species. This pattern of conspecific association should be studied frequently to figure out the method of regeneration and distribution of species. Forest floor encompasses many species with different life forms as a strategy to survive and grow during the competition (Dezzotti et al. 2019). Many lianas and climbers grow fast to occupy the forest canopy and space available for sunlight. These plants suppress a certain seedling establishment (Carreño-Rocabado et al. 2012). The competition to gain sunlight, nutrition, and water is presumed as the limiting factor suffered by some species since they cannot survive below putative parent trees.

Seed dispersal can be the driving force behind the spatial distribution of plants in tropical forests. Moreover, the morphological and anatomical characters of seeds and fruits also influence species regeneration and distribution. For example, small and winged seeds of tree species can spread out by falling around and away from the parent trees (Sebbenn et al. 2008; Lü & Tang 2010). However, factors such as competition, herbivory, and allelopathy have led to a clear and negative association in natural tropical forests (Padmanaba & Corlett 2014; Menezes et al. 2019). In contrast, large seeds mostly fall around the parent trees and since they survived germination, they can grow as positive conspecific associations. Therefore, the conspecific association pattern should be studied to know the natural regeneration of certain species in tropical rainforest.

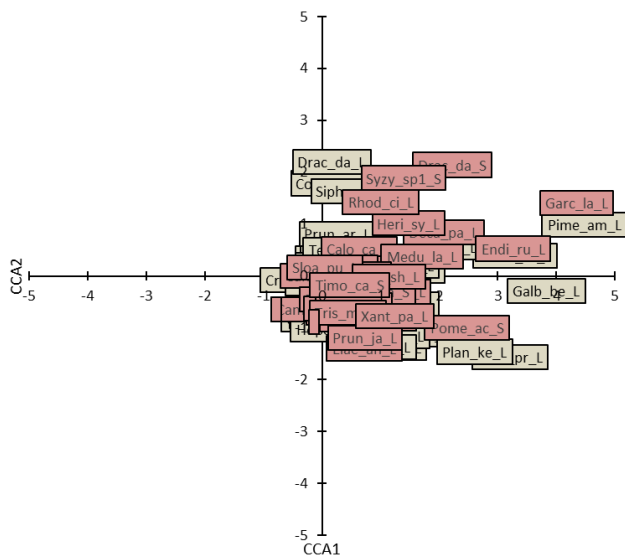


Figure 4. The result of Canonical Correspondence Analysis (CCA) to analyze conspecific associations between the small and the large individuals of the same species.

The implication of associations to ecological knowledge for sustainable management of primary forest

The study of conspecific and heterospecific associations in tropical rainforest is extremely important to determine the spatial distribution pattern, especially the conspecific association. In addition, a model of natural regeneration of tree species can be described, and the result can indicate the pattern of recruitment in the population dynamics of tree species (Goodale et al. 2012; Piotto et al. 2019). Tropical rainforest is primarily dominated by flowering plants with their reproduction season is in annual period (Baker et al. 1998; Pan et al. 2013; Cámara-Leret et al. 2020). Furthermore, a suitable area for certain species to grow has resembled in the conspecific association since the study correlates small individuals with the large ones within the same species. The pattern of conspecific association can also be used to observe natural regeneration. For example, the most appropriate area to plant tree species *in-situ* conservation programs can be decided when artificial regeneration is necessary (Armstrong et al. 2011; Vergara-Rodríguez et al. 2017). The heterospecific association describes the pattern of growth in tropical rainforest since the forest includes the great diversity of tree species. The forest took several decades to develop, and this present study has been able to analyze the pattern of tree species association. Ecological studies on the theme of species association in tropical rainforest need to be replicated in other contexts of region, ecosystem type and forest conditions as tropical forest is very complex as made up of different life forms that interact and create vertical and horizontal structures in the climax phase of the successional process (Chazdon 2003; Brokaw & Scheiner 2012).

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Appendix 1. Species name of heterospecific associations for understory for Figure 2

No.	Code	Species	Group 1	Group 2	Group 3
1	Kiba_co	<i>Kibara coriacea</i> (Blume) Hook. f. & A. Thoms.	√		
2	Rhus_la	<i>Rhus lamprocarpa</i> Merr. & L.M.Perry	√		
3	Dios_pi	<i>Diospyros pilosantha</i> Blanco	√		
4	Lits_le	<i>Litsea ledermannii</i> Teschner	√		
5	Maca_gi	<i>Macaranga gigantea</i> (Rchb.f. & Zoll.) Müll.Arg.	√		
6	Maca_ta	<i>Macaranga tanarius</i> (L.) Müll.Arg.	√		
7	Seme_pa	<i>Semecarpus papuana</i> Lauterb.	√		
8	Ster_sh	<i>Sterculia shillinglawii</i> F.Muell.	√		
9	Case_ca	<i>Casearia carrii</i> Sleumer	√		
10	Xant_no	<i>Wendlandia</i> sp	√		
11	Teij_bo	<i>Teijsmanniodendron bogoriense</i> Koord.	√		
12	Flin_pi	<i>Flindersia pimenteliana</i> F.Muell.	√		
13	Case_sp	<i>Casearia</i> sp	√		
14	Hapl_ce	<i>Haplolobus celebicus</i> H.J.Lam	√		
15	Myri_en	<i>Myristica ensifolia</i> J.Sinclair	√		
16	Octa_in	<i>Octamyrtus insignis</i> Diels	√		
17	Piso_lo	<i>Pisonia longirostris</i> Teijsm. & Binn.	√		
18	Wend_sp	<i>Wendlandia</i> sp	√		
19	Timo_ca	<i>Timonius carii</i> S.P.Darwin	√		
20	Prun_ja	<i>Prunus javanica</i> (Teijsm. & Binn.) Miq.	√		
21	Hapl_fl	<i>Haplolobus floribundus</i> (K.Schum.) H.J.Lam	√		
22	Anti_de	<i>Alstonia spectabilis</i> R.Br.	√		
23	Buch_ar	<i>Buchanania arborescens</i> (Blume) Blume	√		
24	Cryp_pa	<i>Cryptocarya palmerensis</i> C.K.Allen	√		
25	Gmel_se	<i>Gmelina sessilis</i> C.T.White & W.D.Francis ex Lane-Poole	√		
26	Garc_pi	<i>Garcinia picrorhiza</i> Miq.	√		
27	Para_ve	<i>Parastemon versteeghii</i> Merr. & L.M.Perry	√		
28	Knem_in	<i>Knema intermedia</i> Warb.	√		

No.	Code	Species	Group 1	Group 2	Group 3
29	Case_mo	<i>Casearia monticola</i> Sleumer	√		
30	Half_ke	<i>Halfordia kendack</i> Guillaumin	√		
31	Syzy_ve	<i>Syzygium versteegii</i> (Lauterb.) Merr. & L.M.Perry	√		
32	Ficu_ro	<i>Ficus robusta</i> Corner	√		
33	Heri_sy	<i>Heritiera sylvatica</i> S.Vidal	√		
34	Cerb_fl	<i>Cerbera floribunda</i> K.Schum.	√		
35	Myri_gl	<i>Myristica globosa</i> Warb.	√		
36	Dryp_gl	<i>Drypetes globosa</i> (Merr.) Pax & K.Hoffm.	√		
37	Phal_ma	<i>Phaleria macrocarpa</i> (Scheff.) Boerl.	√		
38	Cana_ri	<i>Canarium rigidum</i> (Blume) Zipp. ex Miq.	√		
39	Lits_ti	<i>Litsea timoriana</i> Span.	√		
40	Timo_ti	<i>Timonius timon</i> (Spreng.) Merr.	√		
41	Homa_fo	<i>Heritiera sylvatica</i> S.Vidal	√		
42	Endo_me	<i>Endospermum medulosum</i> L.S.Sm.	√		
43	Pome_pi	<i>Pometia pinnata</i> J.R.Forst. & G.Forst.	√		
44	Kiba_bu	<i>Kibara bullata</i> Philipson	√		
45	Drac_da	<i>Dracontomelon dao</i> (Blanco) Merr. & Rolfe	√		
46	Goni_gi	<i>Goniothalamus giganteus</i> Hook.f. & Thomson	√		
47	Nauc_or	<i>Nauclea orientalis</i> (L.) L.	√		
48	Beil_mo	<i>Beilschmiedia morobensis</i> Kosterm.	√		
49	Cara_br	<i>Carallia brachiata</i> (Lour.) Merr.	√		
50	Lits_sp	<i>Litsea</i> sp	√		
51	Pime_am	<i>Pimelodendron amboinicum</i> Hassk.	√		
52	Medu_la	<i>Medusanthera laxiflora</i> (Miers) R.A.Howard	√		
53	Myri_fa	<i>Myristica fatua</i> Hoult.	√		
54	Gono_li	<i>Gonocaryum litorale</i> (Blume) Sleumer	√		
55	Lith_ru	<i>Lithocarpus rufovillosus</i> (Markgr.) Rehder	√		
56	Hope_pa	<i>Hopea papuana</i> Diels	√		
57	Dill_pa	<i>Dillenia papuana</i> Martelli	√		
58	Mast_pa	<i>Mastiodendron pachyclados</i> (K.Schum.) Melch.	√		
59	Pome_ac	<i>Pometia acuminata</i> Radlk.	√		
60	Kiba_el	<i>Kibara elongata</i> A.C.Sm.	√		
61	Clei_pa	<i>Cleistanthus papuanus</i> (Lauterb.) Jabl.	√		
62	Ints_pa	<i>Intsia palembanica</i> Miq.	√		
63	Galb_be	<i>Galbulimima belgraveana</i> (F.Muell.) Sprague	√		
64	Term_co	<i>Terminalia copelandi</i> Elmer	√		
65	Giro_ne	<i>Gironiera nervosa</i> Planch.	√		
66	Mani_pl	<i>Maniltoa plurijuga</i> Merr. & L.M.Perry	√		
67	Hope_ce	<i>Hopea celtidifolia</i> Kosterm.	√		
68	Meli_el	<i>Melicope elleryana</i> (F. Muell.) T.G. Hartley	√		
69	Garc_sp	<i>Garcinia</i> sp	√		
70	Pala_lo	<i>Palaquium lobbianum</i> Burck	√		
71	Camp_br	<i>Camposperma brevipetiolatum</i> Volkens	√		
72	Ster_ma	<i>Sterculia macrophylla</i> Vent.	√		
73	Harp_ca	<i>Harpullia carrii</i> Leenh.	√		
74	Hors_la	<i>Horsfieldia laevigata</i> Warb.	√		
75	Arch_pa	<i>Archidendron parviflorum</i> Pulle	√		
76	Chry_pa	<i>Chrysophyllum papuanicum</i> (Pierre ex Dubard) Royen	√		
77	Fagr_ra	<i>Fagraea racemosa</i> Jack	√		
78	Klei_ho	<i>Kleinhovia hospita</i> L.	√		
79	Endi_ru	<i>Endiandra rubescens</i> (Blume) Miq.	√		
80	Tris_ma	<i>Tristaniopsis macrosperma</i> (F.Muell.) Peter G.Wilson & J.T.Waterh.	√		
81	Cory_la	<i>Corynocarpus laevigatus</i> J.R.Forst. & G.Forst.	√		
82	Cryp_sp	<i>Cryptocarya</i> sp	√		
83	Taba_au	<i>Tabernaemontana aurantiaca</i> Gaudich.	√		
84	Mall_sp	<i>Mallotus</i> sp	√		
85	Deca_pa	<i>Decaspermum parviflorum</i> (Lam.) A.J.Scott	√		
86	Agla_sp	<i>Aglaia spectabilis</i> (Miq.) S.S.Jain & S.Bennet	√		
87	Rapa_te	<i>Rapanea tempapan</i> P.Royen	√		
88	Calo_ca	<i>Calophyllum caudatum</i> Kaneh. & Hatus.	√		
89	Xant_pa	<i>Xanthophyllum papuanum</i> Whitmore ex Meijden	√		
90	Hapl_la	<i>Haplolobus lanceolatus</i> H.J.Lam ex Leenh.	√		
91	Alst_sp	<i>Alstonia spectabilis</i> R.Br.			√
92	Dios_sp	<i>Diospyros</i> sp			√
93	Elae_an	<i>Elaeocarpus angustifolius</i> Blume			√
94	Rypa_ja	<i>Ryparosa javanica</i> Koord. & Valetton			√

No.	Code	Species	Group 1	Group 2	Group 3
95	Rhod_ci	<i>Rhodammia cinerea</i> Jack			√
96	Myri_gi	<i>Myristica gigantea</i> King			√
97	Anti_to	<i>Antiaris toxicaria</i> Lesch.			√
98	Calo_in	<i>Calophyllum inophyllum</i> L.			√
99	Para_pr	<i>Pararchidendron pruinosa</i> (Benth.) I.C.Nielsen			√
100	Siph_ce	<i>Siphonodon celastrineus</i> Griff.			√
101	Siph_sp	<i>Siphonodon</i> sp			√
102	Maas_gl	<i>Maasia glauca</i> (Hassk.) Mols, Kessler & Rogstad			√
103	Term_ka	<i>Terminalia kaernbachii</i> Warb.			√
104	Spat_ja	<i>Spathiostemon javensis</i> Blume		√	
105	Dyso_mo	<i>Dysoxylum mollissimum</i> Blume		√	
106	Plan_ke	<i>Planchonella keyensis</i> H.J.Lam		√	
107	Ficu_sp	<i>Ficus</i> sp		√	
108	Hors_pa	<i>Horsfieldia parviflora</i> (Roxb.) J.Sinclair		√	

Appendix 2. Species name of heterospecific associations for upper story Figure 3

No	Code	Species	Group 1	Group 2	Group 3	Group 4
1	Hors_ir	<i>Horsfieldia irya</i> (Gaertn.) Warb.		√		
2	Garc_la	<i>Garcinia latissima</i> Miq.		√		
3	Alst_sc	<i>Alstonia scholaris</i> (L.) R. Br.		√		
4	Endi_vi	<i>Endiandra virens</i> F.Muell.		√		
5	Sloa_pu	<i>Sloanea pullei</i> O.C.Schmidt ex A.C.Sm.		√		
6	Galb_be	<i>Galbulimima belgraveana</i> (F.Muell.) Sprague		√		
7	Pome_ac	<i>Pometia acuminata</i> Radlk.		√		
8	Chis_ce	<i>Chisocheton ceramicus</i> Miq.		√		
9	Dyso_mo	<i>Dysoxylum mollissimum</i> Blume		√		
10	Endo_me	<i>Endospermum medullosum</i> L.S.Sm.		√		
11	Pime_am	<i>Pimelodendron amboinicum</i> Hassk.		√		
12	Gono_li	<i>Gonocaryum litorale</i> (Blume) Sleumer	√			
13	Stre_el	<i>Streblus elongatus</i> (Miq.) Corner	√			
14	Homa_fo	<i>Heritiera sylvatica</i> S.Vidal	√			
15	Gnet_gn	<i>Gnetum gnemon</i> L.	√			
16	Homa_no	<i>Homalanthus novoguineensis</i> (Warb.) K.Schum.	√			
17	Cory_la	<i>Corynocarpus laevigatus</i> J.R.Forst. & G.Forst.	√			
18	Drac_da	<i>Dracontomelon dao</i> (Blanco) Merr. & Rolfe	√			
19	Cana_od	<i>Canarium indicum</i> L.	√			
20	Pter_be	<i>Pterocymbium beccarii</i> K.Schum.	√			
21	Pala_lo	<i>Palaquium lobbianum</i> Burck	√			
22	Cana_in	<i>Canarium indicum</i> L.	√			
23	Cara_br	<i>Carallia brachiata</i> (Lour.) Merr.	√			
24	Timo_ca	<i>Timonius carii</i> S.P.Darwin	√			
25	Hors_sy	<i>Horsfieldia sylvestris</i> Warb.	√			
26	Maas_su	<i>Maasia sumatrana</i> (Miq.) Mols, Kessler & Rogstad	√			
27	Hope_pa	<i>Hopea papuana</i> Diels	√			
28	Hope_ce	<i>Hopea celtidifolia</i> Kosterm.	√			
29	Rhus_ta	<i>Rhus taitensis</i> Guill.	√			
30	Acti_ni	<i>Actinodaphne nitida</i> Teschner	√			
31	Dill_pa	<i>Dillenia papuana</i> Martelli	√			
32	Medu_la	<i>Medusanthera laxiflora</i> (Miers) R.A.Howard	√			
33	Teij_bo	<i>Teijsmanniodendron bogoriense</i> Koord.	√			
34	Tris_ma	<i>Tristaniopsis macrosperma</i> (F.Muell.) Peter G.Wilson & J.T.Waterh.	√			
35	Call_lo	<i>Callicarpa longifolia</i> Lam.	√			
36	Comm_b	<i>Commersonia bartramia</i> (L.) Merr.	√			
37	Dios_pi	<i>Diospyros pilosanthera</i> Blanco	√			
38	Knem_in	<i>Knema intermedia</i> Warb.	√			
39	Dryp_gl	<i>Drypetes globosa</i> (Merr.) Pax & K.Hoffm.	√			
40	Cryp_pa	<i>Cryptocarya palmerensis</i> C.K.Allen	√			
41	Meli_el	<i>Melicope elleryana</i> (F. Muell.) T.G. Hartley	√			

No	Code	Species	Group 1	Group 2	Group 3	Group 4
42	Lits_ti	<i>Litsea timoriana</i> Span.	√			
43	Siph_ce	<i>Siphonodon celastrineus</i> Griff.	√			
44	Siph_sp	<i>Siphonodon</i> sp	√			
45	Vite_pi	<i>Vitex pinnata</i> L.	√			
46	Poly_no	<i>Polyscias nodosa</i> (Blume) Seem.	√			
47	Pome_pi	<i>Pometia pinnata</i> J.R.Forst. & G.Forst.	√			
48	Agla_ar	<i>Aglaia argentea</i> Blume	√			
49	Acro_sp	<i>Acronychia</i> sp	√			
50	Gmel_se	<i>Gmelina sessilis</i> C.T.White & W.D.Francis ex Lane-Pool	√			
51	Mani_br	<i>Maniltoa browneoides</i> Harms	√			
52	Prun_ar	<i>Prunus arborea</i> (Blume) Kalkman	√			
53	Camp_br	<i>Camptosperma brevipetiolatum</i> Volkens	√			
54	Hors_la	<i>Horsfieldia laevigata</i> Warb.	√			
55	Cana_hi	<i>Camptosperma brevipetiolatum</i> Volkens	√			
56	Deca_pa	<i>Decaspermum parviflorum</i> (Lam.) A.J.Scott	√			
57	Calo_in	<i>Calophyllum inophyllum</i> L.	√			
58	Heri_sy	<i>Heritiera sylvatica</i> S.Vidal	√			
59	Clei_pa	<i>Cleistanthus papuanus</i> (Lauterb.) Jabl.	√			
60	Elae_an	<i>Elaeocarpus angustifolius</i> Blume	√			
61	Gymn_fa	<i>Gymnacranthera farquhariana</i> (Hook.f. & Thomson) Warb.	√			
62	Grew_er	<i>Grewia eriocarpa</i> Juss.	√			
63	Xant_no	<i>Wendlandia</i> sp	√			
64	Rhod_ci	<i>Rhodamnia cinerea</i> Jack	√			
65	Arto_al	<i>Artocarpus altilis</i> (Parkinson ex F.A.Zorn) Fosberg	√			
66	Para_pr	<i>Pararchidendron pruinosum</i> (Benth.) I.C.Nielsen	√			
67	Plan_ke	<i>Planchonella keyensis</i> H.J.Lam	√			
68	Dios_pa	<i>Diospyros papuana</i> Valetton ex Bakh.	√			
69	Ochr_gl	<i>Ochrosia glomerata</i> (Blume) F.Muell.	√			
70	Myri_fa	<i>Myristica fatua</i> Houtt.	√			
71	Ster_sh	<i>Sterculia shillinglawii</i> F.Muell.	√			
72	Syzy_sp2	<i>Syzygium</i> sp2	√			
73	Syzy_sp3	<i>Syzygium</i> sp3	√			
74	Xant_pa	<i>Xanthophyllum papuanum</i> Whitmore ex Meijden	√			
75	Euca_pa	<i>Eucalyptopsis papuana</i> C.T.White			√	
76	Flin_pi	<i>Flindersia pimenteliana</i> F.Muell.			√	
77	Hapl_fl	<i>Haplotobos floribundus</i> (K.Schum.) H.J.Lam			√	
78	Lits_fi	<i>Litsea firma</i> (Blume) Hook.f.			√	
79	Term_co	<i>Terminalia copelandi</i> Elmer			√	
80	Calo_ca	<i>Calophyllum caudatum</i> Kaneh. & Hatus.			√	
81	Coch_gi	<i>Cochlospermum gillivraei</i> Benth.			√	
82	Buch_ar	<i>Buchanania arborescens</i> (Blume) Blume			√	
83	Fagr_el	<i>Fagraea elliptica</i> Roxb.				√
84	Prun_ja	<i>Prunus javanica</i> (Teijsm. & Binn.) Miq.				√
85	Endi_ru	<i>Endiandra rubescens</i> (Blume) Miq.				√
86	Cryp_sp	<i>Cryptocarya</i> sp				√

Appendix 3. Species name of conspecific associations Figure 4. The S stands for small individuals and L symbolizes large individuals.

No.	Code	Species
1	Agla_ar	_L. <i>Aglaia argentea</i> Blume
2	Agla_ar	_S. <i>Aglaia argentea</i> Blume
3	Buch_ar	_L. <i>Buchanania arborescens</i> (Blume) Blume
4	Buch_ar	_S. <i>Buchanania arborescens</i> (Blume) Blume
5	Calo_ca	_L. <i>Calophyllum caudatum</i> Kaneh. & Hatus.
6	Calo_ca	_S. <i>Calophyllum caudatum</i> Kaneh. & Hatus.
7	Calo_in	_L. <i>Calophyllum inophyllum</i> L.
8	Calo_in	_S. <i>Calophyllum inophyllum</i> L.
9	Camp_br	_L. <i>Camptosperma brevipetiolatum</i> Volkens
10	Camp_br	_S. <i>Camptosperma brevipetiolatum</i> Volkens
11	Cana_hi	_L. <i>Canarium hirsutum</i> Willd.
12	Cana_hi	_S. <i>Canarium hirsutum</i> Willd.

No.	Code	Species
13	Cana_in	_J. <i>Canarium indicum</i> L.
14	Cana_in	_S <i>Canarium indicum</i> L.
15	Cara_br	_J. <i>Carallia brachiata</i> (Lour.) Merr.
16	Cara_br	_S <i>Carallia brachiata</i> (Lour.) Merr.
17	Clei_pa	_J. <i>Cleistanthus papuanus</i> (Lauterb.) Jabl.
18	Clei_pa	_S <i>Cleistanthus papuanus</i> (Lauterb.) Jabl.
19	Cory_la	_J. <i>Corynocarpus laevigatus</i> J.R.Forst. & G.Forst.
20	Cory_la	_S <i>Corynocarpus laevigatus</i> J.R.Forst. & G.Forst.
21	Cryp_pa	_J. <i>Cryptocarya palmerensis</i> C.K.Allen
22	Cryp_pa	_S <i>Cryptocarya palmerensis</i> C.K.Allen
23	Cryp_sp	_J. <i>Cryptocarya</i> sp
24	Cryp_sp	_S <i>Cryptocarya</i> sp
25	Deca_pa	_J. <i>Decaspermum parviflorum</i> (Lam.) A.J.Scott
26	Deca_pa	_S <i>Decaspermum parviflorum</i> (Lam.) A.J.Scott
27	Dill_pa	_J. <i>Dillenia papuana</i> Martelli
28	Dill_pa	_S <i>Dillenia papuana</i> Martelli
29	Dios_pi	_J. <i>Diospyros pilosanthera</i> Blanco
30	Dios_pi	_S <i>Diospyros pilosanthera</i> Blanco
31	Drac_da	_J. <i>Dracontomelon dao</i> (Blanco) Merr. & Rolfe
32	Drac_da	_S <i>Dracontomelon dao</i> (Blanco) Merr. & Rolfe
33	Dryp_gl	_J. <i>Drypetes globosa</i> (Merr.) Pax & K.Hoffm.
34	Dryp_gl	_S <i>Drypetes globosa</i> (Merr.) Pax & K.Hoffm.
35	Dyso_mo	_J. <i>Dysoxylum mollissimum</i> Blume
36	Dyso_mo	_S <i>Dysoxylum mollissimum</i> Blume
37	Elac_an	_J. <i>Elaeocarpus angustifolius</i> Blume
38	Elac_an	_S <i>Elaeocarpus angustifolius</i> Blume
39	Endi_ru	_J. <i>Endiandra rubescens</i> (Blume) Miq.
40	Endi_ru	_S <i>Endiandra rubescens</i> (Blume) Miq.
41	Endo_me	_J. <i>Endospermum medulosum</i> L.S.Sm.
42	Endo_me	_S <i>Endospermum medulosum</i> L.S.Sm.
43	Flin_pi	_J. <i>Flindersia pimenteliana</i> F.Muell.
44	Flin_pi	_S <i>Flindersia pimenteliana</i> F.Muell.
45	Galb_be	_J. <i>Galbulimima begraveana</i> (F.Muell.) Sprague
46	Galb_be	_S <i>Galbulimima begraveana</i> (F.Muell.) Sprague
47	Garc_la	_J. <i>Garcinia latissima</i> Miq.
48	Garc_la	_S <i>Garcinia latissima</i> Miq.
49	Giro_ne	_J. <i>Gironniera nervosa</i> Planch.
50	Giro_ne	_S <i>Gironniera nervosa</i> Planch.
51	Gmel_se	_J. <i>Gmelina sessilis</i> C.T.White & W.D.Francis ex Lane-Poole
52	Gmel_se	_S <i>Gmelina sessilis</i> C.T.White & W.D.Francis ex Lane-Poole
53	Gnet_gn	_J. <i>Gnetum gnemon</i> L.
54	Gnet_gn	_S <i>Gnetum gnemon</i> L.
55	Gono_li	_J. <i>Gonocaryum litorale</i> (Blume) Sleumer
56	Gono_li	_S <i>Gonocaryum litorale</i> (Blume) Sleumer
57	Gymn_fa	_J. <i>Gymnacranthera farquhariana</i> (Hook.f. & Thomson) Warb.
58	Gymn_fa	_S <i>Gymnacranthera farquhariana</i> (Hook.f. & Thomson) Warb.
59	Hapl_fl	_J. <i>Haplolobus floribundus</i> (K.Schum.) H.J.Lam
60	Hapl_fl	_S <i>Haplolobus floribundus</i> (K.Schum.) H.J.Lam
61	Heri_sy	_J. <i>Heritiera sylvatica</i> S.Vidal
62	Heri_sy	_S <i>Heritiera sylvatica</i> S.Vidal
63	Homa_fo	_J. <i>Homalium foetidum</i> Benth.
64	Homa_fo	_S <i>Homalium foetidum</i> Benth.
65	Hope_ce	_J. <i>Hopea celtidifolia</i> Kosterm.
66	Hope_ce	_S <i>Hopea celtidifolia</i> Kosterm.
67	Hope_no	_J. <i>Hopea novoguineensis</i> Slooten
68	Hope_no	_S <i>Hopea novoguineensis</i> Slooten
69	Hope_pa	_J. <i>Hopea papuana</i> Diels
70	Hope_pa	_S <i>Hopea papuana</i> Diels
71	Hors_la	_J. <i>Horsfieldia laevigata</i> Warb.
72	Hors_la	_S <i>Horsfieldia laevigata</i> Warb.
73	Ints_pa	_J. <i>Intsia palembanica</i> Miq.
74	Ints_pa	_S <i>Intsia palembanica</i> Miq.
75	Knem_in	_J. <i>Knema intermedia</i> Warb.

No.	Code	Species
76	Knem_in	_S <i>Knema intermedia</i> Warb.
77	Lith_ru	_L <i>Lithocarpus rufifoliosus</i> (Markgr.) Rehder
78	Lith_ru	_S <i>Lithocarpus rufifoliosus</i> (Markgr.) Rehder
79	Lits_ti	_L <i>Litsea timoriana</i> Span.
80	Lits_ti	_S <i>Litsea timoriana</i> Span.
81	Mani_br	_L <i>Maniltoa brownoides</i> Harms
82	Mani_br	_S <i>Maniltoa brownoides</i> Harms
83	Medu_la	_L <i>Medusanthera laxiflora</i> (Miers) R.A.Howard
84	Medu_la	_S <i>Medusanthera laxiflora</i> (Miers) R.A.Howard
85	Meli_el	_L <i>Melicope elleryana</i> (F. Muell.) T.G. Hartley
86	Meli_el	_S <i>Melicope elleryana</i> (F. Muell.) T.G. Hartley
87	Myri_fa	_L <i>Myristica fatua</i> Houott.
88	Myri_fa	_S <i>Myristica fatua</i> Houott.
89	Pala_lo	_L <i>Palaquium lobbianum</i> Burck
90	Pala_lo	_S <i>Palaquium lobbianum</i> Burck
91	Para_pr	_L <i>Pararchidendron pruinatum</i> (Benth.) I.C.Nielsen
92	Para_pr	_S <i>Pararchidendron pruinatum</i> (Benth.) I.C.Nielsen
93	Para_ve	_L <i>Parastemon versteeghii</i> Merr. & L.M.Perry
94	Para_ve	_S <i>Parastemon versteeghii</i> Merr. & L.M.Perry
95	Pime_am	_L <i>Pimelodendron amboinicum</i> Hassk.
96	Pime_am	_S <i>Pimelodendron amboinicum</i> Hassk.
97	Plan_ke	_L <i>Planchonella keyensis</i> H.J.Lam
98	Plan_ke	_S <i>Planchonella keyensis</i> H.J.Lam
99	Pome_ac	_L <i>Pometia acuminata</i> Radlk.
100	Pome_ac	_S <i>Pometia acuminata</i> Radlk.
101	Pome_pi	_L <i>Pometia pinnata</i> J.R.Forst. & G.Forst.
102	Pome_pi	_S <i>Pometia pinnata</i> J.R.Forst. & G.Forst.
103	Prun_ar	_L <i>Prunus arborea</i> (Blume) Kalkman
104	Prun_ar	_S <i>Prunus arborea</i> (Blume) Kalkman
105	Prun_ja	_L <i>Prunus javanica</i> (Teijsm. & Binn.) Miq.
106	Prun_ja	_S <i>Prunus javanica</i> (Teijsm. & Binn.) Miq.
107	Rhod_ci	_L <i>Rhodammia cinerea</i> Jack
108	Rhod_ci	_S <i>Rhodammia cinerea</i> Jack
109	Siph_ce	_L <i>Siphonodon celastrineus</i> Griff.
110	Siph_ce	_S <i>Siphonodon celastrineus</i> Griff.
111	Siph_sp	_L <i>Siphonodon</i> sp
112	Siph_sp	_S <i>Siphonodon</i> sp
113	Sloa_pu	_L <i>Sloanea pullei</i> O.C.Schmidt ex A.C.Sm.
114	Sloa_pu	_S <i>Sloanea pullei</i> O.C.Schmidt ex A.C.Sm.
115	Ster_sh	_L <i>Sterculia shillinglawii</i> F.Muell.
116	Ster_sh	_S <i>Sterculia shillinglawii</i> F.Muell.
117	Syzy_sp1	_L <i>Syzygium</i> sp1
118	Syzy_sp1	_S <i>Syzygium</i> sp1
119	Teij_bo	_L <i>Teijsmanniodendron bogoriense</i> Koord.
120	Teij_bo	_S <i>Teijsmanniodendron bogoriense</i> Koord.
121	Term_co	_L <i>Terminalia copelandi</i> Elmer
122	Term_co	_S <i>Terminalia copelandi</i> Elmer
123	Timo_ca	_L <i>Timonius carii</i> S.P.Darwin
124	Timo_ca	_S <i>Timonius carii</i> S.P.Darwin
125	Tris_ma	_L <i>Tristaniaopsis macrosperma</i> (F.Muell.) Peter G.Wilson & J.T.Waterh.
126	Tris_ma	_S <i>Tristaniaopsis macrosperma</i> (F.Muell.) Peter G.Wilson & J.T.Waterh.
127	Vati_ra	_L <i>Vatica rassak</i> Blume
128	Vati_ra	_S <i>Vatica rassak</i> Blume
129	Xant_pa	_L <i>Xanthophyllum papuanum</i> Whitmore ex Meijden
130	Xant_pa	_S <i>Xanthophyllum papuanum</i> Whitmore ex Meijden
131	Xant_no	_L <i>Xanthostemon novaguineensis</i> Valetton
132	Xant_no	_S <i>Xanthostemon novaguineensis</i> Valetton

