



Fungal Planet description sheets: 1182–1283

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Key words

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Abstract Novel species of fungi described in this study include those from various countries as follows: **Algeria**, *Phaeoacremonium adelophilialidum* from *Vitis vinifera*. **Antarctica**, *Comoclathris antarctica* from soil. **Australia**, *Coniochaeta salicifolia* as endophyte from healthy leaves of *Geijera salicifolia*, *Eremothecium peggii* in fruit of *Citrus australis*, *Microdochium ratticaudae* from stem of *Sporobolus natalensis*, *Neocelosporium corymbiae* on stems of *Corymbia variegata*, *Phytophthora kelmanii* from rhizosphere soil of *Ptilotus pyramidalis*, *Pseudosydowia backhousiae* on living leaves of *Backhousia citriodora*, *Pseudosydowia indooroopillyensis*, *Pseudosydowia louisecottisae* and *Pseudosydowia queenslandica* on living leaves of *Eucalyptus* sp. **Brazil**, *Absidia montepascoalis* from soil. **Chile**, *Ilyonectria zarorii* from soil under *Maytenus boaria*. **Costa Rica**, *Colletotrichum filicis* from an unidentified fern. **Croatia**, *Mollisia endogranulata* on deteriorated hardwood. **Czech Republic**, *Arcopilus navicularis* from tea bag with fruit tea, *Neosetophoma buxi* as endophyte from *Buxus sempervirens*, *Xerochrysum bohemicum* on surface of biscuits with chocolate glaze and filled with jam. **France**, *Entoloma cyaneobasale* on basic to calcareous soil, *Fusarium aconidiale* from *Triticum aestivum*, *Fusarium juglandicola* from buds of *Juglans regia*. **Germany**, *Tetraploa endophytica* as endophyte from *Microthlaspi perfoliatum* roots. **India**, *Castanediella ambae* on leaves of *Mangifera indica*, *Lactifluus kanadii* on soil under *Castanopsis* sp., *Penicillium uttarakhandense* from soil. **Italy**, *Penicillium ferrianaense* from compost. **Namibia**, *Bezeromyces gobabebensis* on leaves of unidentified succulent, *Cladosporium stipagrostidicola* on leaves of *Stipagrostis* sp., *Cymostachys euphorbiae* on leaves of *Euphorbia* sp., *Deniquelata hypolithi* from hypolith under a rock, *Hysterobrevium walvisbayicola* on leaves of unidentified tree, *Knufia hypolithi* and *Knufia walvisbayicola* from hypolith under a rock, *Lapidomyces stipagrostidicola* on leaves of *Stipagrostis* sp., *Nothophaeothea mirabibensis* (incl. *Nothophaeothea* gen. nov.) on persistent inflorescence remains of *Blepharis obmitrata*, *Paramyothecium salvadorae* on twigs of *Salvadora persica*, *Preussia procaviicola* on dung of *Procapra* sp., *Sordaria equicola* on zebra dung, *Volutella salvadorae* on stems of *Salvadora persica*. **Netherlands**, *Entoloma ammophilum* on sandy soil, *Entoloma pseudocruentatum* on nutrient poor (acid) soil, *Entoloma pudens* on plant debris, amongst grasses. **New Zealand**, *Amorocoelephoma neoregeliae* from leaf spots of *Neoregelia* sp., *Aquilomyces metrosideri* and *Septoriella callistemonis* from stem discoloration and leaf spots of *Metrosideros* sp., *Cadophora neoregeliae* from leaf spots of *Neoregelia* sp., *Flexuomyces asteliae* (incl. *Flexuomyces* gen. nov.) and *Mollisia asteliae* from leaf spots of *Astelia chathamica*, *Ophioceras freycinetiae* from leaf spots of *Freycinetia*

Abstract (cont.)

banksii, *Phaeosphaeria caricis-sectae* from leaf spots of *Carex secta*. **Norway**, *Cuphophyllus flavipesoides* on soil in semi-natural grassland, *Entoloma coracis* on soil in calcareous *Pinus* and *Tilia* forests, *Entoloma cyaneolilacinum* on soil semi-natural grasslands, *Inocybe norvegica* on gravelly soil. **Pakistan**, *Butyriboletus parachinarenensis* on soil in association with *Quercus baloot*. **Poland**, *Hyalodendriella bialowiezensis* on debris beneath fallen bark of Norway spruce *Picea abies*. **Russia**, *Bolbitius sibiricus* on a moss covered rotting trunk of *Populus tremula*, *Crepidotus wasserii* on debris of *Populus tremula*, *Entoloma isborscanum* on soil on calcareous grasslands, *Entoloma subcoracis* on soil in subalpine grasslands, *Hydropus lecythiocystis* on rotted wood of *Betula pendula*, *Meruliopsis faginea* on fallen dead branches of *Fagus orientalis*, *Metschnikowia taurica* from fruits of *Ziziphus jujube*, *Suillus praetermissus* on soil, *Teunia lichenophila* as endophyte from *Cladonia rangiferina*. **Slovakia**, *Hygrocybe fulgens* on mowed grassland, *Pleuroflammula pannonica* from corticated branches of *Quercus* sp. **South Africa**, *Acrodontium burrowsianum* on leaves of unidentified *Poaceae*, *Castanediella senegaliae* on dead pods of *Senegalia ataxacantha*, *Cladophialophora behniae* on leaves of *Behnia* sp., *Colletotrichum cliviigenum* on leaves of *Clivia* sp., *Diatrype dalbergiae* on bark of *Dalbergia armata*, *Falcocladium heteropyxidicola* on leaves of *Heteropyxis canescens*, *Lapidomyces aloidendricola* as epiphyte on brown stem of *Aloidendron dichotomum*, *Lasionectria sansevieriae* and *Phaeosphaeriopsis sansevieriae* on leaves of *Sansevieria hyacinthoides*, *Lylea dalbergiae* on *Diatrype dalbergiae* on bark of *Dalbergia armata*, *Neochaetothyria syzygii* (incl. *Neochaetothyria* gen. nov.) on leaves of *Syzygium chordatum*, *Nothophaeomoniella ekebergiae* (incl. *Nothophaeomoniella* gen. nov.) on leaves of *Ekebergia pterophylla*, *Paracymostachys euphorbiae* (incl. *Paracymostachys* gen. nov.) on leaf litter of *Euphorbia ingens*, *Paramycosphaerella pterocarpis* on leaves of *Pterocarpus angolensis*, *Paramycosphaerella syzygii* on leaf litter of *Syzygium chordatum*, *Parateichospora phoenicicola* (incl. *Parateichospora* gen. nov.) on leaves of *Phoenix reclinata*, *Seiridium syzygii* on twigs of *Syzygium chordatum*, *Setophoma syzygii* on leaves of *Syzygium* sp., *Starmerella xylocopis* from larval feed of an Afrotropical bee *Xylocopa caffra*, *Teratosphaeria combreti* on leaf litter of *Combretum kraussii*, *Teratosphaericola leucadendri* on leaves of *Leucadendron* sp., *Toxicocladosporium pterocarpis* on pods of *Pterocarpus angolensis*. **Spain**, *Cortinarius bonachei* with *Quercus ilex* in calcareous soils, *Cortinarius brunneovolvatus* under *Quercus ilex* subsp. *ballota* in calcareous soil, *Extremopsis radicola* (incl. *Extremopsis* gen. nov.) from root-associated soil in a wet heathland, *Russula quintanensis* on acidic soils, *Tubaria vulcanica* on volcanic lapilli material, *Tuber zambonelliae* in calcareous soil. **Sweden**, *Elaphomyces borealis* on soil under *Pinus sylvestris* and *Betula pubescens*. **Tanzania**, *Curvularia tanzanica* on inflorescence of *Cyperus aromaticus*. **Thailand**, *Simplicillium niveum* on *Ophiocordyceps camponoti-leonardi* on underside of unidentified dicotyledonous leaf. **USA**, *Calonectria californiensis* on leaves of *Umbellularia californica*, *Exophiala spartinae* from surface sterilised roots of *Spartina alterniflora*, *Neophaeococcomyces oklahomensis* from outside wall of alcohol distillery. **Vietnam**, *Fistulinella aurantioflava* on soil. Morphological and culture characteristics are supported by DNA barcodes.

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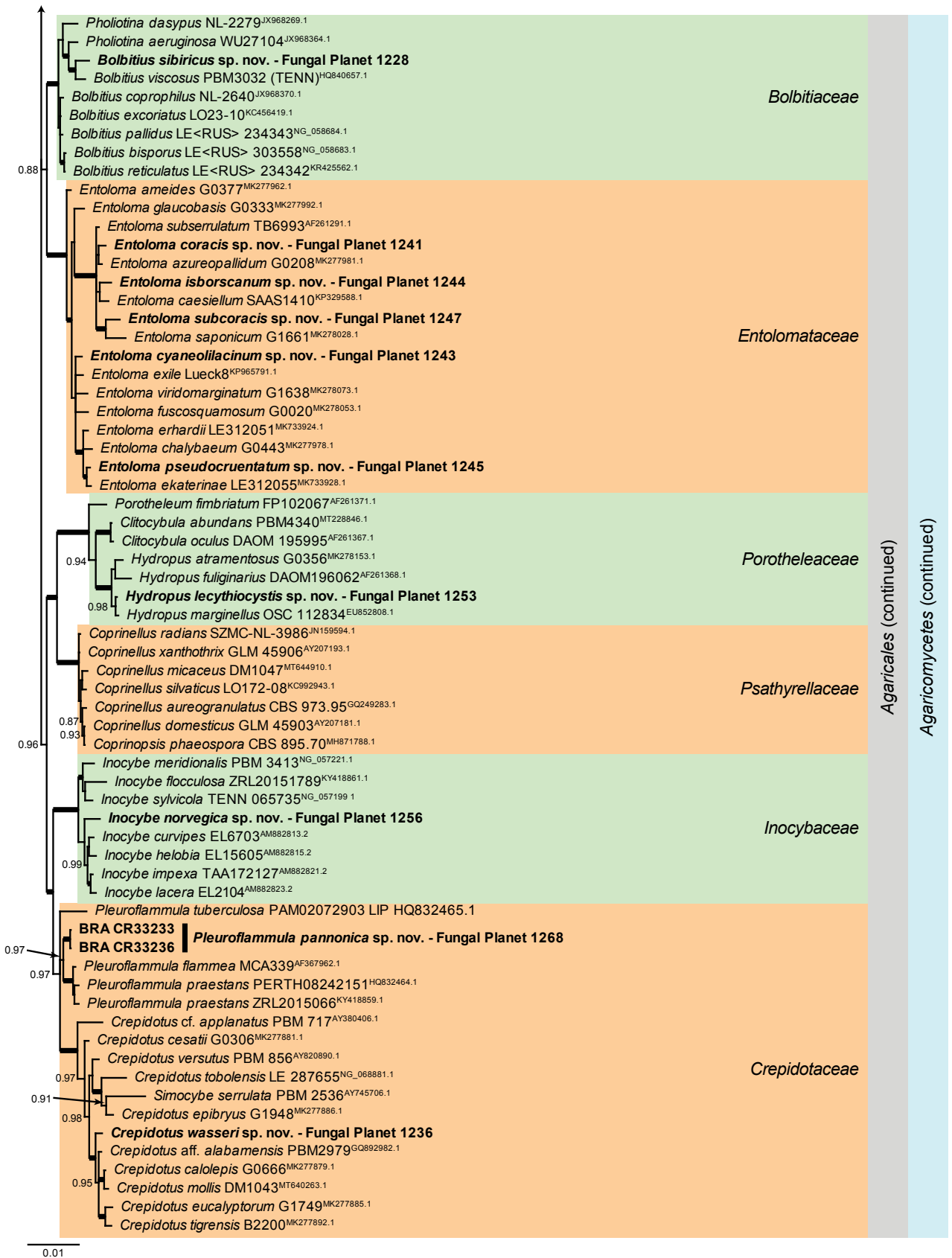
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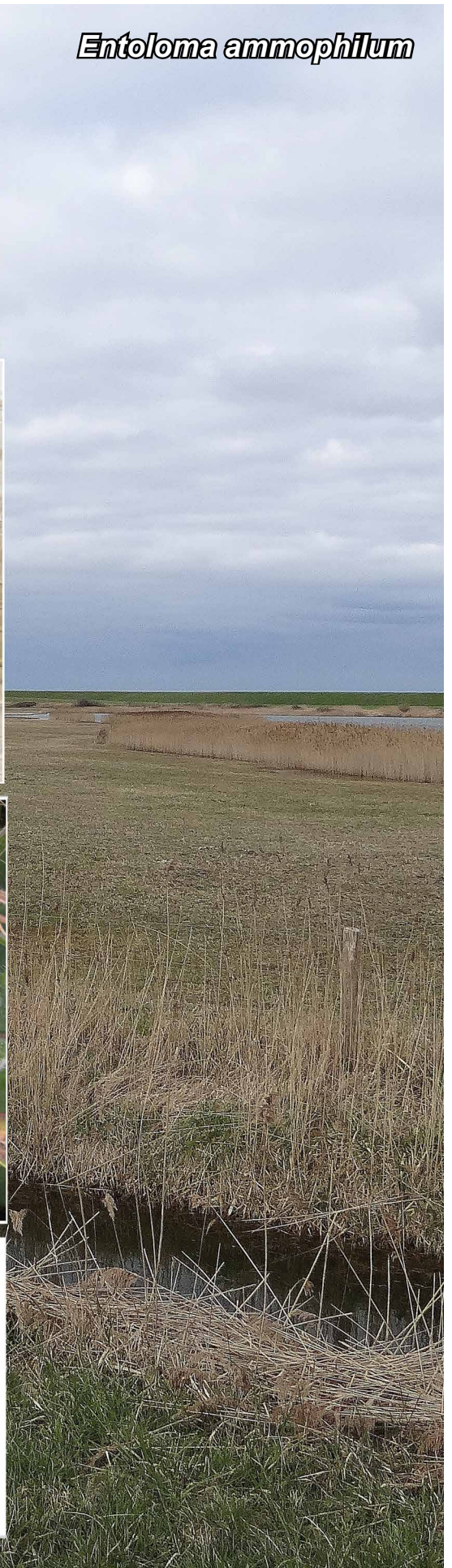
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Overview Agaricomycetes phylogeny (cont.) – part 3

Entoloma ammophilum



Fungal Planet 1240 – 13 July 2021

Entoloma ammophilum G.M. Jansen, Dima, Noordel. & Vila, *sp. nov.*

Etymology. Referring to the habitat on sandy soil (from ἄμμος, Greek, – sand, and φιλέω, Greek, – to like).

Classification — *Entolomataceae*, *Agaricales*, *Agaricomycetes*.

Basidiomata medium-sized, collybioid. *Pileus* 10–20(–35) mm diam, convex to convex flattened, umbilicate, not distinctly hygrophanous, dark brown or grey brown with blackish brown centre, deeply translucently striate up to centre, minutely squamulose at centre, more or less glabrous towards margin. *Lamellae*, L = about 30, l = 3–5, deeply emarginate, ventricose, pink with eroded, concolorous edge. *Stipe* 30–50 × 1–2 mm, cylindrical, bicolored, steel-blue in lower part, brown above, polished. *Smell* and *taste* not noted. *Spores* heterodiametrical, 9.5–11.5 × 7–8.5 μm, av. 10.4 × 7.8 μm, Q = 1.1–1.55, Q_{av} = 1.25, regularly 5–7-angled. *Basidia* 4-spored, clavate, up to 45 × 15 μm, clampless. *Lamella edge* fertile, cystidia absent. *Pileipellis* a cutis at margin, trichoderm-like at centre, made up of clavate terminal elements, up to 20 μm wide. *Pigment* intracellular-granular, brown. *Clamp connections* absent in all tissues.

Habitat & Distribution — Terrestrial on sandy soil, in small groups in moist dune valley with *Salix repens* on calcareous sandy-clayey soil, or in sandy soil of riverbanks, under *Populus nigra* and *Alnus glutinosa*. Known from the Netherlands and Spain.

Typus. THE NETHERLANDS, Prov. Zeeland, Dreischor, 22 Oct. 2016, G.M. Jansen C160-4418 (holotype L0608224, ITS sequence GenBank MW934591, MycoBank MB 839221).

Additional material examined. SPAIN, Barcelona, Vallès Oriental, Can Romegosa, Sant Fost de Campsentelles, alt. 140 m, 24 Oct. 2010, under *Populus nigra* and *Alnus glutinosa*, on sandy soil, F. Caballero & J. Vila, SFC 101024-05 (L0607606, ITS sequence GenBank MW934592).

Colour illustrations. The Netherlands, Prov. Zeeland, Dreischor (type locality). Spores and pileipellis (from holotype); basidiomata *in situ* (holotype). Scale bars = 1 cm (basidiomata), 10 μm (spores and microstructures).

Notes — *Entoloma ammophilum* has a characteristic dark brown or brown grey pileus and a polished, bicoloured stipe. It differs from *E. glaucobasis*, which frequently occurs in the same habitat, by the polished nature of the stipe, and fertile lamellae edge. Furthermore, *E. glaucobasis* is phylogenetically rather distant in the /Griseocyaneum clade. Within the /Sarcitulum clade, this species resembles *E. montanum*, which may superficially look similar, but differs clearly in having a sterile, often brown coloured lamella edge, and alpine or boreal distribution.

Supplementary material

FP1240 Phylogenetic tree derived from Maximum Likelihood analysis based on nrITS1-5.8S-ITS2 data. Analysis was performed in PhyML v. 3.0 (Guindon et al. 2010) using the non-parametric Shimodaira-Hasegawa version of the approximate likelihood-ratio test (SH-aLRT) and the GTR+I+Γ model of evolution. ML bootstrap support values are shown at the nodes (BS > 50 %).

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Entoloma coracis



Fungal Planet 1241 – 13 July 2021

***Entoloma coracis* Brandrud, Dima, Noordel., G.M. Jansen & Vila, sp. nov.**

Etymology. The epithet refers to the dark blackish to violaceous black colour of the basidiomata, like plumage of a raven (*Corvus corax*).

Classification — *Entolomataceae*, *Agaricales*, *Agaricomycetes*.

Basidiomata medium-sized, collybioid. *Pileus* 10–35 mm, hemispherical to convex expanding plano-convex with involute then deflexed margin, with depressed, rarely umbilicate centre, not hygrophanous, not translucently striate, initially very dark blackish to violaceous black, with age the bluish tinges fade away, leaving the pileus very dark brownish black, violaceous black or porphyry brown, uniformly coloured, not or slightly pallescent on maturing, entirely tomentose and staying so during development or breaking up in small squamules. *Lamellae*, L = 20–30, l = 1–3, moderately distant, adnate-emarginate or with decurrent tooth, segmentiform to subventricose, white, then with pale pink tinge, with irregular, usually with concolorous edge; rarely spotted black from the start, or becoming spotted blackish with age. *Stipe* 20–80 × 3–7 mm, relatively long and stout, initially violaceous grey, fading to pale bluish grey, sometimes developing a lilac-pink tinge, much paler than the pileus, not polished, but covered with blue to violaceous longitudinal fibrils, sometimes scaly-flocculose at apex, especially in rainy conditions, fibrils with same colour or contrastingly darker than background, with abundant white basal mycelium. *Context* white. *Smell* insignificant, *taste* not recorded. *Spores* 8.5–12.5 × 5.5–7.5 µm, av. 9.5–11 × 6–6.5 µm, Q = 1.3–1.7, Qav = 1.3–1.4, heterodiametrical, with 5–7 rather pronounced and sharp angles. *Basidia* 4-spored, claviform, 28.5–41 × 8–13.5 µm, clampless. *Lamella edge* sterile, consisting of a strand of hyphae with clustered cheilocystidia (serrulatum-type) with rather pronounced often somewhat tapering cheilocystidia, 5–15 µm wide, usually not pigmented, but occasionally becoming bluish black with age. *Hymenophoral trama* regular, made up of cylindrical to inflated hyphae, 11–25 µm wide. *Pileipellis* a cutis with transitions to a trichoderm, of clavate, septate, terminal elements, 50–110 × 8–19 µm. *Pigment* intracellular, brown. *Brilliant granules* sparse to abundant. *Clamp connections* absent.

Habitat & Distribution — Saprotrophic, calciphilous or acidophilous. In Norway mainly in open, calcareous *Pinus* and *Tilia* forests, but also in naturally open, steppe-like, thermophilous grassland/shrubland on shallow-soil limestone rocks, and once also recorded in grassland and shrub vegetation on limestone. In South Europe in Mediterranean thermophilous areas, under *Quercus ilex*, *Cistus monspeliensis* or *Pinus halepensis*, also known in the Canary Islands, on woods with *Laurus novocanariensis*, *Pinus radiata* and *Cistus symphytifolius*. Known from Norway, France, Spain and Austria, but certainly more widespread in Europe.

Colour illustrations. Norway, Telemark, Porsgrunn, Frierflogene NR, calcareous dry grassland/margin of calcareous pine forest (type locality). Spores, cheilocystidia, pileipellis, stipitipellis (all from holotype); basidiomata *in situ* (holotype). Scale bars = 1 cm (basidiomata), 10 µm (spores and microstructures).

Typus. NORWAY, Telemark, Porsgrunn, Frierflogene NR, near bridge, calcareous, dry grassland/margin of calcareous pine forest, 14 Sept. 2019, T.E. Brandrud, B. Dima & R. Solvang, TEB 381-19 (holotype O-F-256850, ITS and LSU sequences GenBank MW934571 and MW934251, MycoBank MB 839222).

Additional materials examined. AUSTRIA, Tirol, Ehrwald, 28 Aug. 2018, Rainer Wald (L0608002, ITS sequence GenBank MW934578). – FRANCE, Dordogne, Sanilhac, route de Lafaye, on soil with *Mycenella bryophila*, 241 m a.s.l., 6 Nov. 2019, G. Eyssartier (GE 19027, ITS sequence GenBank MW934581). – NORWAY, Nordland, Alstahaug, Altra, 10 m a.s.l., calcareous pasture, 18 Sept. 2004, D. Pettersen, A.B. Stærnes, J.B. Jordal, A. Knutsen & P. Fadnes (O-F-67255, ITS sequence GenBank MW934572); Trøndelag, Snåsa, Bergsåsen Nature Reserve, calcareous pine forest, 2 Sept. 2009, E. Bendiksen & K. Bendiksen KB&EB51/09 (O-F-252053, ITS sequence GenBank MW934574); Steinkjer, Kvam, Aunvolltangen, 60 m a.s.l., old calcareous *Picea* forest, 3 Sept. 2010, H. Holien & T.E. Brandrud, U.-B. Bøe, A. Molia HH 57/10 (O-F-293335, ITS sequence GenBank MW934575); Telemark, Bamble, Baneåsen Nature Reserve, calcareous *Tilia* forest, 7 Sept. 2015, B. Dima & T.E. Brandrud TEB 244-15 (O-F-251952, ITS sequence GenBank MW934573); Bamble, Røsskleiva Nature Reserve SE, in calcareous *Fraxinus-Corylus* forest, 8 Sept. 2015, T.E. Brandrud & B. Dima TEB 279-15 (O-F-254580, ITS sequence GenBank MW934576); Porsgrunn, Blekebakken Nature Reserve, calcareous *Pinus* forest, 25 Sept. 2015, T.E. Brandrud & B. Dima TEB 557-15 (O-F-254614, ITS sequence GenBank MW934577); Vestfold, Larvik, Løvallåsen, calcareous grassland, 9 Oct. 2013, T. Læssøe & A. Molia AM-2450-2013 (O-F-21892, ITS sequence GenBank MW934579). – SPAIN, Girona, Can Cofi, 1 June 2013, P. Carbo 20130601 (L0608020, ITS sequence GenBank MW934580).

Notes — *Entoloma coracis* is one of the *E. corvinum* look-alikes, with its very dark, opaque, tomentose pileus, white lamellae, and fibrous stipe. *Entoloma aranense* is a sister species of *E. coracis*, less robust, paler, with a lilac-bluish tinged pileus when young, later brown and fibrillose, and a typical subalpine-alpine habitat. Microscopically the differences are minimal. *Entoloma porphyrogriseum* is also closely related, but differs, e.g., in smaller spores, and not so persistently dark pileus. Phylogenetically (see the phylogenetic tree for *E. amophilum* in Supplementary material FP1240), these three species are rather distant from *E. corvinum* s.str., as we now interpret it, and they differ from *E. coracis*, morphologically by the narrower, more sharply angled spores, the serrulatum-type lamella edge, and the habitat. *Entoloma corvinum* is an alpine species, like the similar *E. erhardii*, which differs by having smaller spores, and a polished stipe.

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Entoloma cyaneobasale



Fungal Planet 1242 – 13 July 2021

Entoloma cyaneobasale Corriol, Dima & Noordel., *sp. nov.*

Etymology. The epithet refers to the blue colour in the base of the stipe (from 'cyaneus', Greek – dark blue, and 'basis', Greek – base).

Classification — *Entolomataceae*, *Agaricales*, *Agaricomycetes*.

Basidiomata medium-sized, collybioid. *Pileus* 20–40 mm diam, campanulate to convex with incurved margin, then bluntly conico-convex to convex with more or less lobed margin, brown (near 7.5YR3/4) then brown-yellow (10 YR 5.4; Munsell 1954), darker at centre, distinctly hygrophanous (drying to 10 YR 7/3), nearly opaque with only slight striation at margin, entirely fibrillose to sub-squamulose. *Lamellae* rather distant, deeply emarginate, straight to ventricose, whitish, then pink, with slightly eroded, brown edge. *Stipe* up to 45 × 4.5 mm, cylindrical, or quite often compressed with groove, initially blue-grey at base (2.5 / PB), and pale greyish at apex (10 YR 7–6/2), typically bicoloured, quickly fading, nearly polished, but with fine fibrillose striation over whole length, with white mycelial base. *Context* whitish. *Smell* and *taste* not noted. *Spores* (9–)9.5–11(–11.5) × (7–)7.5–8.5(–9) µm, av. 10 × 8 µm, Q = (1.1–)1.15–1.4, Q_{av} = 1.3, shortly heterodiametrical, with 6–8 weak angles, thick-walled, with granular content. *Basidia* 22–30 × 10–12 µm, 4-spored, shortly cylindrico-clavate to ventricose, with 3–4 µm long sterigmata, clampless. *Lamellae edge* sterile, of serrulatum-type, made up of septate cheilocystidia, with terminal elements 35–55 × 9–13 µm, with brown, intracellular pigment. *Pleurocystidia* not observed. *Subhymenium* branched. *Pileipellis* a trichoderm, with clavate terminal elements, often in clusters, with brown, diffuse, intracellular pigment. *Subpellis* with concentrated brown intracellular pigment, with abundant brilliant granules and mixed with refringent lactiferous hyphae. *Clamp connections* absent.

Habitat & Distribution — Terrestrial in alpine snowbed on basic to calcareous soil, together with *Dryas* and *Salix* species. Known from France and Italy.

Typus. FRANCE, Pyrenées-Atlantiques, Eaux-Bonnes, cirque du Plaa Ségouné, Gourette, 2400 m a.s.l., 30 Aug. 2002, G. Corriol (holotype GC02083008 in BBF, ITS sequence GenBank MW934560, MycoBank MB 839223).

Additional materials examined. ITALY, Trentino-Alto Adige, Passo dello Stelvio/Stilfser Joch, near Berghotel Franzenshöhe, alpine grassland with *Dryas* and *Salix* spp., 2200 m a.s.l., 30 July 2018, B. Dima, DB-2018-07-30-1 (ITS sequence GenBank MW934561).

Colour illustrations. France, Pyrenees-Atlantiques, Eaux-Bonnes, cirque du Plaa Ségouné à Gourette, 2400 m a.s.l., calcareous alpine snowbeds on the northern slope of Pyrenees. Spores; cheilocystidia; pileipellis; stiptipellis (all from holotype); basidiomata *in situ* (holotype). Scale bars = 1 cm (basidiomata), 10 µm (spores and microstructures).

Notes — *Entoloma cyaneobasale* falls within the /Mediterranean clade (see the phylogenetic tree for *E. ammophilum* in Supplementary material FP1240). In the field, these collections were readily identified as *E. glaucobasis* on account of the bicoloured stipe. However, the spores are smaller, the stipe is more polished, and it has a lamella edge of the serrulatum-type with brown pigment. *Entoloma glaucobasis* is also phylogenetically distant, and belongs to the /Griseocyaneum clade.

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Entoloma cyaneofilacinum



Fungal Planet 1243 – 13 July 2021

Entoloma cyaneolilacinum Noordel., J.B. Jordal, Brandrud & Dima, *sp. nov.*

Etymology. The epithet refers to the colours of the basidiocarps, from 'cyaneus', Greek, – blue, and 'lilacinus' – lilac.

Classification — *Entolomataceae*, *Agaricales*, *Agaricomycetes*.

Basidiomata medium-sized, collybioid. *Pileus* 10–25 mm, conico-convex or campanulate-conical, slightly expanding, finally plano-convex, with deflexed then straight margin, not distinctly hygrophanous, deep blue then paler lilac-blue with a slightly darker spot at centre, deeply translucently striate, at first finely radially fibrillose to faintly tomentose, breaking up in small squamules in central part, radially fibrillose to almost smooth towards margin. *Lamellae* moderately distant, deeply emarginate, ventricose, white or with a faint bluish tinge, contrasting with blue pileus and stipe, with entire, concolorous edge. *Stipe* 30–50 × 2–3 mm, cylindrical, deep blue then lilac-blue, concolorous with margin of pileus or paler, glabrous, smooth, polished, with some white mycelium at base, once observed with yellow (discoloured?) mycelium. *Smell* and *taste* not indicated. *Spores* (7.5–)8.0–10.0(–11.0) × 6.0–8.5 µm, av. 8.5–9.5 × 6.5–8.0 µm, Q = 1.2–1.6, Qav = 1.4, heterodiametrical, 5–7-angled in side-view. *Basidia* 30–50 × 8–12 µm, 4-spored, clampless. *Lamella* edge fertile. *Cystidia* absent. *Hymenophoral trama* regular, made up of inflated elements, up to 20 µm wide. *Pileipellis* a transition between a cutis and a trichoderm, made up of clavate terminal elements, 22–75 × 10–25 µm with brownish intracellular pigment. *Brilliant granules* present, but not abundant. *Clamp connections* absent.

Habitat & Distribution — In semi-natural grasslands and in deciduous woodlands with *Betula*, *Corylus*, *Fraxinus* and *Quercus*. Verified with sequenced collections from Norway and The Netherlands, also reported from Germany.

Typus. NORWAY, Møre og Romsdal, Stranda, Liabygda, Ansok, N62.3137° E7.0236° (± 7 m), 310 m a.s.l., seminatural grassland (meadow), on the ground, 2 Sept. 2009, J.B. Jordal, JBJ09-E02 (holotype O-F-252009, ITS and LSU sequences GenBank MW934582 and MW934252, MycoBank MB 839224).

Colour illustrations. Norway, Møre og Romsdal, Stranda, Liabygda, Ansok, seminatural grassland, type locality. Spores, cheilocystidia, pileipellis, stipitipellis (all from holotype); Basidiomata *in situ* (holotype). Scale bars = 1 cm (basidiomata), 10 µm (spores and microstructures).

Additional materials examined. NORWAY, Møre og Romsdal, Sunndal, Jordalsgrenda, Kalvhusvøttu, 60 m a.s.l., seminatural grassland (meadow), 14 Sept. 2004, J.B. Jordal, M.E. Noordeloos & G. Gulden (O-F-177981, ITS sequence GenBank MW934584); *ibid.*, 20 Sept. 2019, JBJ19-049 (O-F-256792, ITS sequence GenBank MW934586); Rogaland, Stavanger, Rennesøy, Askje, V-side, c. 60 m a.s.l., in semi-natural pasture, 3 Oct. 2006, J.I. Johnsen & J.B. Jordal (O-F-361225, ITS sequence GenBank MW934587); Vindafjord, Alnåsen west, 129 m a.s.l., west-faced deciduous forest, 5 Sept. 2008, J.B. Jordal, JBJ08-E02 (O-F-252007, ITS sequence GenBank MW934585). — THE NETHERLANDS, Prov. Utrecht, Soesterberg, former airfield, 30 Sept. 2019, M.E. Noordeloos, P.J. Keizer & J. v. Dongen (L0607898, ITS sequence GenBank MW934583).

Notes — The delicate lilac-blue colour of the basidiocarps as well as the small spores and fertile lamella edge are distinctive for *E. cyaneolilacinum*. It was treated as *E. lepiotosme* in Noordeloos (2004). However, there are considerable discrepancies with the protologue, describing a species with a blackish brown, virgate pileus, reminiscent of a species of *Inocybe*, a fibrillose stipe surface, a strong smell like that of *Lepiota cristata*, and larger spores. The lectotype of *Rhodophyllus lepiotosmus* failed for DNA sequencing. Considering the conflict with the protologue and the lack of molecular data, it was decided to describe the present taxon here as a species in its own right. Morphologically, *E. cyaneolilacinum* resembles *E. violaceoviride*, which has a sterile, brown pigmented lamella edge and often some greenish tinges in the basidiocarp, and has a distant phylogenetic position. *Entoloma cruentatum*, also phylogenetically distant (see the phylogenetic tree for *E. ammophilum* in Supplementary material FP1240), has similar spores and fertile lamella edge, but often turns orange-yellow when bruised at the base of the stipe.

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Entoloma isborscanum



Fungal Planet 1244 – 13 July 2021

Entoloma isborscanum O.V. Morozova, Noordel., Dima, G.M. Jansen & Reschke, *sp. nov.*

Etymology. Named after Izborsk (*Isborscum*, Lat.), a village in the Pskov Region of Russia, one of the oldest Russian towns, type locality and *Entoloma* hot spot.

Classification — *Entolomataceae*, *Agaricales*, *Agaricomycetes*.

Basidiomata medium-sized, collybioid. *Pileus* 15–35 mm diam, hemispherical, with slightly depressed centre, then expanding to convex or plano-convex with slightly umbilicate centre, with deflexed then straight margin, not hygrophanous or in the pileus margin only, not translucently striate or up to 1/3 of radius, radially fibrillose-squamulose all over, more densely in the centre, white background is visible between fibrils, brownish yellow, yellowish brown or light brown, darker in centre (5C5–8, 5D5–8; Kornerup & Wanscher 1978), in old and drying specimens radially cracking, showing white underground. *Lamellae* moderately distant, adnate-emarginate, decurrent with short tooth, whitish, pale or greyish, becoming pinkish grey, with irregular, concolorous, whitish or brownish edge. *Stipe* 20–70 × 2–3 mm, cylindrical or slightly broadened towards the base, minutely distinctly longitudinally striate, completely greyish blue (20B3–4, 20C3–4), greyish brown, bluish on the base only or completely yellow-brown, concolorous with the pileus (5C5–8, 5D5–8), white tomentose at the base. *Context* white, brownish under the surface. *Smell* pleasantly sweet or indistinct, *taste* unpleasant, nitrous. *Basidiospores* 8–12 × 6–7.5 µm, av. 9–10.5 × 6–7.5 µm, Q = (1.15–)1.3–1.7, Qav = 1.35–1.5, heterodiametrical, with 5–7 distinct angles in side-view, sometimes with some large spores up to 14 × 7 µm with indistinct angles from 1–2-spored basidia. *Basidia* 32–35.5 × 9.5–10.5 µm, 1–4-spored, narrowly clavate to clavate, sometimes with broadened walls, clampless. *Lamella edge* sterile of the ‘serrulatum’-type. *Cheilocystidia* 40–90 × 9–25 µm, as terminal elements of the hyphae arising from the subhymenium, clavate, broadly clavate or cylindrical, sometimes septate, sometimes with intracellular pigment, brownish in KOH. *Hymenophoral trama* regular, 4–20 µm wide, cylindrical hyphae. *Pileipellis* a cutis with transition to a trichoderm of cylindrical hyphae, 5–12 µm wide with cylindrical or inflated to ellipsoid or ovoid terminal elements, 48–105 × 12–32 µm, and intracellular, sometimes agglutinate pigment, brown in KOH. *Caulocystidia* as ascending bundled, cylindrical to slightly inflated terminal elements of the stiptipellis hyphae. *Clamp connections* absent.

Habit & Distribution — In small groups on soil on calcareous grasslands. Known from Denmark, Germany, the Netherlands, and Russia (European part).

Colour illustrations. RUSSIA, Pskov region, Pechorsky district, Izborsk village, foot of the Truvor hillfort, calcareous grassland (type locality). Spores, cheilocystidia, pileipellis, caulocystidia (all from holotype); basidiomata *in situ* (holotype). Scale bars = 1 cm (basidiomata), 10 µm (spores and microstructures).

Typus. RUSSIA, Pskov region, Pechorsky district, Izborsk village, foot of the Truvor hillfort, on calcareous grassland, N57.717702° E27.854764°, 24 Aug. 2011, O. Morozova (holotype LE 302088, ITS and LSU sequences GenBank MW934566 and MW934253, MycoBank MB 839225).

Additional materials examined. DENMARK, Jylland, Begtrup Røn, 26 Aug. 2011, R. Ejrnæs (C, DMS167798, ITS sequence GenBank MW934570). – GERMANY, Baden-Württemberg, Heimberg, near Schloßböckelheim, 27 Oct. 2017, W. Prüfert (M-0141378, ITS sequence GenBank MW934565). – RUSSIA, Pskov region, Pechorsky district, Izborsk village, foot of the Truvor hillfort, on calcareous grassland, 24 Aug. 2011, O. Morozova, LE 312486, ITS sequence GenBank MW934564; *ibid.*, 12 Sept. 2020, O. Morozova, LE 312679, ITS sequence GenBank MW934569. – THE NETHERLANDS, PROV. Limburg, Nijswiller-Noord, 14 Aug. 2019, F. & R. Salzmänn, L0607743, ITS sequence GenBank MW934568; *ibid.*, 21 Aug. 2019, L0607927, ITS sequence GenBank MW934567; *ibid.*, 2 Oct. 2019, L0607719, ITS sequence GenBank MW934563; *ibid.*, 16 Oct. 2019, L0607718, ITS sequence GenBank MW934562.

Notes — *Entoloma isborscanum* is characterised by a squamulose yellowish brown pileus, a bluish or brownish blue, longitudinally striate stipe, rather small spores with distinct 5–7 angles, and a sterile lamella edge consisting of large clavate cheilocystidia arising from the subhymenium. Superficially it resembles *E. griseocyaneum*, which differs by the absence of cheilocystidia and smaller spores, or *E. glaucobasis*, which possesses larger, almost nodulose spores (Noordeloos 1992). The holotype specimen was published in Morozova et al. (2015) as *E. exile*, but this species is sufficiently more slender with distinct greenish glaucous tinges in the stipe. Also see the phylogenetic tree for *E. ammophilum* in Supplementary material FP1240.

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Entoloma pseudocruentatum



Fungal Planet 1245 – 13 July 2021

Entoloma pseudocruentatum Noordel., Brandrud, G.M. Jansen, Dima & Læssøe, *sp. nov.*

Etymology. The epithet refers to the erroneously applied name *Entoloma cruentatum* for this species.

Classification — *Entolomataceae*, *Agaricales*, *Agaricomycetes*.

Basidiomata small to medium-sized, collybioid. *Pileus* 15–25 mm diam, convex to flattened-convex, later, when maturing, from flattened to somewhat depressed in the centre, never umbonate; slate blue grey with slight violaceous tinge, deeply translucently striate, innately radially fibrillose, not squamulose at centre, margin somewhat crenulate. *Lamellae* adnate, bluish grey when young. *Stipe* 30–40 × 2–4 mm, similar in colour to the pileus or paler, polished, with white to yellow or orange-yellow basal tomentum. *Context* thin, pale grey bluish. *Smell* indistinct. *Taste* not known. *Basidiospores* 8.5–10.0 × 6.0–7.5 µm, av. 9.1 × 6.8 µm, heterodiametrical, with 5–7 angles in side-view. *Basidia* 28–34 × 9.5–12.5 µm, clavate, 4-spored, clampless. *Lamella edge* fertile, cheilocystidia absent. *Pileipellis* a cutis of cylindrical hyphae, 3.5–9 µm wide, with a transition to a trichoderm at centre of clavate elements, 12–30 µm wide. *Pigment* blue to grey-blue, clotted granular and diffusely intracellular. Stipitipellis cylindrical hyphen 3.5–8.5 µm wide with grey blue clotted granular and diffuse intracellular pigment. *Clamp connections* absent in all tissues.

Habitat & Distribution — Saprotrophic, in groups on nutrient poor (acid) soil, in a regularly mown, species-rich grassland of an old airbase (holotype) and on rich grassland (Denmark), herb/grass-rich *Fraxinus-Quercus* forest (Norway).

Typus. THE NETHERLANDS, Prov. Utrecht, Soesterberg, former airfield, 30 Sept. 2019, M.E. Noordeloos, P.J. Keizer & J.V. Dongen (holotype L0607915, ITS and LSU sequences GenBank MW934588 and MW934254, MycoBank MB 839227).

Additional materials examined. DENMARK, Favrholt, in semi-natural grassland, on shady slope with *Plantago lanceolata* and *Succisa pratensis*, 19 Aug. 2008, T. Læssøe, TL-13373 (C, DMS-730741; ITS sequence GenBank MW934590). – NORWAY, Telemark, Drangedal, Maljell S, 31 Aug. 2015, in rich, somewhat calcareous grass-herb vegetation in open *Fraxinus-Quercus* forest, T.E. Brandrud, TEB 188-15 (O-F-251951; ITS sequence GenBank MW934589).

Colour illustrations. The Netherlands, Prov. Utrecht, Soesterberg, former airfield (type locality, photo credit P.J. Keizer). Basidiomata (left from holotype, right from TEB188-15); spores, cheilocystidia, pileipellis, stipitipellis (all from holotype). Scale bars = 1 cm (habit), 10 µm (spores), 5 µm (pileipellis and stipitipellis).

Notes — *Entoloma pseudocruentatum* was interpreted as *E. cruentatum* by Noordeloos (1987, 2004), as similar to *E. chalybaeum*, with a fertile lamella edge, a glaucous-blue stipe, with the base frequently discolouring yellowish or pale orange. However, there are now reasonable doubts as whether the original diagnosis of Quélet (1886) actually refers to the same species. *Entoloma cruentatum* is described as a species with a more or less conical to umbonate, lilac-blue pileus, and a stipe with glaucous bluish stipe with a strong reddening at base. Kühner & Romagnesi (1953) considered it a dubious species, probably in subg. *Nolanea*. Another option could be that *E. cruentatum* represents a form of *E. exile*, a species with rather variable pileus shape and colour, and often a reddening stipe base. The concept of Noordeloos (1984) was based on a collection from Scotland, which has many characters in common with the species described here. For this reason, we describe *Entoloma cruentatum* sensu Noordeloos (1984) as a new species. Also see the phylogenetic tree for *E. ammophilum* in Supplementary material FP1240.

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Entoloma pudens



Fungal Planet 1246 – 13 July 2021

Entoloma pudens Noordel., G.M. Jansen, M.v.d. Vegte & Dima, *sp. nov.*

Etymology. The epithet refers to the modest size of the species.

Classification — *Entolomataceae*, *Agaricales*, *Agaricomycetes*.

Basidiomata small-sized, omphalioid. *Pileus* 12–18 mm diam, convex with straight or slightly crenulate margin, umbilicate, hygrophanous, deeply translucently striate up to centre, dark brown with obscure sepia-brown centre, finely scaly-irrigate, particularly at central part. *Lamellae* distant, L = 12, l = 1–3, arcuate-deeply decurrent, pale brown with pinkish hue, with concolorous, entire edge. *Stipe* 35–30 × 1–1.5 mm, slightly broadened towards apex, pale horn brown, glabrous, polished. *Smell* and *taste* indistinct. Spores 9.5–13 × 6.5–8.5 µm, av. 11.3–11.5 × 7.6–8.2 µm, Q = 1.2–1.7, Q_{av} 1.4–1.5, heterodiametrical, rather regularly 5–7-angled. *Basidia* 28–53 × 11–11.5 µm, 4-spored, clampless. *Lamella* edge fertile, *cystidia* absent. *Pileipellis* a cutis of broad hyphae with transitions to a trichoderm, made up of clavate elements, up to 25 µm wide, with both intracellular and incrusting pigment. *Clamp connections* absent.

Habitat & Distribution — Gregarious on plant debris, amongst grasses and *Sphagnum* in unfertilized hayfield. Known only from the type locality in The Netherlands.

Typus. THE NETHERLANDS, Prov. Gelderland, Groesbeek, de Bruuk, 17 Sept. 2018, Marjon v.d. Vegte & G. Jansen C173-6268 (holotype L0608054, ITS and LSU sequences GenBank MW934594, MycoBank MB 839226).

Additional material examined. THE NETHERLANDS, Prov. Gelderland, Groesbeek, de Bruuk, 2 Sept. 2018, G. Jansen, C173-6195 (L0607607, not sequenced).

Notes — This small omphalioid species was initially identified as *Entoloma nigellum* sensu Orton (1960), a concept accepted by Noordeloos (2004). The Dutch specimens are very likely the same as Orton's with their dark, translucently striate pileus, distant, decurrent lamellae, polished stipe, large spores and clampless hyphae. However, when comparing the original diagnosis of this poorly known species it became clear that Quélet's *Eccilia nigella* (Quélet 1886) strongly deviates in habit, in having a not translucent, almost black pileus, and a dark stipe, which sometimes has a bluish tinge. It is therefore rather questionable whether we can adopt the epithet '*nigellum*' for our taxon. Quélet's species could well stand for another, dark coloured species in the /*Rusticoidea* clade and type material is not existent. For the time being, it therefore seems prudent to consider *Eccilia nigellum* as a *nomen dubium*. *Entoloma subpusillum* is similar, differing in having a non-translucent, not glabrous but uneven-rugulose pileal surface. Two other apparently similar species, viz., *E. pseudonigellum* and *E. rickenelliformis*, differ both in having a dark, not translucent pileus, differently sized and shaped spores, and abundant clamp-connections. The type sequence of *E. pseudonigellum* cluster distantly, in the /*Undati* clade, while DNA sequencing of the type of *E. rickenelliformis* proved unsuccessful. Several questions therefore remain unanswered in this group. *Entoloma pudens* is described as new, based on well-annotated material and molecular sequence data (see the phylogenetic tree for *E. ammophilum* in Supplementary material FP1240).

Colour illustrations. The Netherlands, Groesbeek, de Bruuk, unfertilized hayfield, type locality. Spores, cheilocystidia, pileipellis, stipitipellis (all from holotype); basidiomata *in situ* (holotype). Scale bars = 1 cm (basidiomata), 10 µm (spores and microstructures).

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Entoloma subcoracis



Fungal Planet 1247 – 13 July 2021

Entoloma subcoracis O.V. Morozova, Noordel. & Dima, *sp. nov.*

Etymology. The epithet refers to the resemblance of *Entoloma coracis* due to its black colour, like a raven (*Corvus corax*).

Classification — *Entolomataceae*, *Agaricales*, *Agaricomycetes*.

Basidiomata medium-sized, collybioid. *Pileus* 10–35 mm diam, hemispherical or abruptly conical with central depression, hardly expanding, with deflexed margin, not hygrophanous, translucently striate almost up to the centre, initially uniformly blackish blue (21F5–8; Kernerup & Wanscher 1978), discolouring to bluish grey (21F3–4), minutely radially fibrillose-squamulose all over. *Lamellae* moderately distant, adnate-emarginate, segmentiform to narrowly ventricose, white, contrasting with the pileus surface, becoming pink, with irregular, serrulate concolorous edge. *Stipe* 30–70 × 1.5–3 mm, cylindrical, sometimes twisted, slightly longitudinally striate, minutely squamulose, especially in the upper part, concolorous with pileus or a little paler (up to 21F3–4), white tomentose at base. *Context* white, greyish under the surface. Smell indistinct, taste not reported. *Basidiospores* 9.5–11 × 6.5–8 µm, av. 10.0 × 7.0 µm, Q = 1.3–1.5, Q_{av} = 1.4; heterodiametrical, with 5–7 angles in side-view, relatively simple. *Basidia* 32–38 × 10–12 µm, 4-spored, narrowly clavate to clavate, clampless. *Lamella edge* sterile. *Cheilocystidia* 37–80 × 8.5–13.5 µm, composed of 3–4 elements, terminal cells mostly lageniform or fusiform, sometimes cylindrical or narrowly clavate, colourless. *Pileipellis* cutis with transition to a trichoderm of cylindrical to slightly inflated hyphae 8–20 µm wide with inflated terminal elements and dark intracellular pigment, brownish in KOH. *Caulocystidia* 35–100 × 5.5–10 µm, as chains of cylindrical or inflated elements, usually with tapered terminal cells. Brilliant granules present. *Clamp connections* absent.

Habitat & Distribution — In small groups on soil in subalpine grasslands. Known from Russia (Caucasus).

Typus. RUSSIA, Karachaevo-Cherkesia Republic, Teberda Nature Reserve, Arkhyz site, near the waterfall, N43.558889° E41.301389°, alt. 1 390 m a.s.l., 17 Aug. 2009, O. Morozova (holotype LE312483, ITS and LSU sequences GenBank MW934593 and MW934255, MycoBank MB 839228).

Colour illustrations. Russia, Karachaevo-Cherkesia Republic, Teberda Nature Reserve, Arkhyz site, near the waterfall, type locality (photo credit E. Malysheva). Spores, cheilocystidia, pileipellis, caulocystidia (all from holotype); basidiomata *in situ* (holotype). Scale bars = 1 cm (basidiomata), 10 µm (spores and microstructures).

Notes — *Entoloma subcoracis* belongs to the form-group of *E. corvinum* s. auct. including taxa such as *E. coracis* (also described in present paper) and *E. porphyrogriseum* characterised by blackish blue basidiocarps. *Entoloma subcoracis* is characterised by the voluminous cheilocystidia, and fusiform, septate caulocystidia. The characteristic large cystidia are shared with the North American *E. subcorvinum* (Hesler 1967, Noordeloos 1988). Both species seem, however, to be geographically separated. Unfortunately, no DNA sequence data are available for the holotype of *E. subcorvinum*. Also see the phylogenetic tree for *E. ammophilum* in Supplementary material FP1240.

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REFERENCES

- Abad ZG, Abad JA, Creswell T. 2002. Advances in the integration of morphological and molecular characterization in *Phytophthora* genus: The case of *P. kelmiana* and other putative new species. *Phytopathology* 92 (6 suppl): S1.
- Abad ZG, Burgess T, Bienapfl JC, et al. 2019. IDphy: Molecular and morphological identification of *Phytophthora* based on the types. USDAAPHIS PPQ S&T Beltsville Lab, USDA APHIS PPQ S&T ITP, Centre for *Phytophthora* Science and Management, and World *Phytophthora* Collection. <https://idtools.org/id/phytophthora/index.php> <different dates in 2020>.
- Abdollahzadeh J, Groenewald JZ, Coetzee MPA, et al. 2020. Evolution of lifestyles in Capnodiales. *Studies in Mycology* 95: 381–414.
- Akaike H. 1974. A new look at the statistical model identification. *IEEE Transactions on Automatic Control* 19: 716–723.
- Alcorn JL. 1982. Ovaricolous bipolaris species on *Sporobolus* and other grasses. *Mycotaxon* 15: 20–48.
- Ames LM. 1949. New cellulose destroying fungi isolated from military material and equipment. *Mycologia* 41: 637–648.
- Ariyawansa HA, Maharachchikumbura SS, Karunaratne SC, et al. 2013. *Deniquelata barringtoniae* gen. et sp. nov., associated with leaf spots of *Barringtonia asiatica*. *Phytotaxa* 105: 11–20.
- Ariyawansa HA, Phookamsak R, Tibpromma S, et al. 2014. A molecular and morphological reassessment of *Diademaceae*. *The Scientific World Journal*: 675348.
- Ariyawansa HA, Thambugala KM, Manamgoda DS, et al. 2015. Towards a natural classification and backbone tree for Pleosporaceae. *Fungal Diversity* 71: 85–139.
- Arora D, Frank JL. 2014. Clarifying the butter Boletes: a new genus, *Butyriboletus*, is established to accommodate *Boletus* sect. *Appendiculati*, and six new species are described. *Mycologia* 106: 464–480.
- Badali H, Gueidan C, Najafzadeh MJ, et al. 2008. Biodiversity of the genus *Cladophialophora*. *Studies in Mycology* 61: 175–191.
- Baiswar P, Ngachan S, Rymbai H, et al. 2014. *Simplicillium lanosoniveum*, a hyperparasite on *Aecidium elaeagni-latifoliae* in India. *Australasian Plant Disease Notes* 9: 144–149.
- Ballarà J, Cadiñanos-Aguirre JA, Campos JC, et al. 2009. *Cortinarius ibero-insulares-2*. Fungi non Delineati. Pars XLVIII-XLIX: 33–35. Edizioni Candusso, Alassio (SV).
- Ballarà J, Suárez E, Mahiques R, et al. 2017. *Cortinarius iunii*, una nueva especie de la sección Bovini. *The Journal of the Journées européennes du Cortinaire* 19: 11–27.
- Bandini D, Oertel B, Schüssler C, et al. 2020. Noch mehr Risspilze: Fünfzehn neue und zwei wenig bekannte Arten der Gattung *Inocybe*. *Mycologia Bavarica* 20: 13–101.
- Baral H-O. 1987. Lugol's solution / IKI versus Melzer's reagent: hemiamyloidity, a universal feature of the ascus wall. *Mycotaxon* 29: 399–450.
- Bas C. 1999. *Hydropus Kuhner* ex Singer. In: Bas C, Kuyper TW, Noordeloos ME, et al. (eds), *Flora Agaricina Neerlandica* 4: 166–173. Rotterdam, Balkema.
- Bensch K, Braun U, Groenewald JZ, et al. 2012. The genus *Cladosporium*. *Studies in Mycology* 72: 1–401.
- Bensch K, Groenewald JZ, Braun U, et al. 2015. Common but different: The expanding realm of *Cladosporium*. *Studies in Mycology* 82: 23–74.
- Bensch K, Groenewald JZ, Dijksterhuis J, et al. 2010. Species and ecological diversity within the *Cladosporium* cladosporioides complex (*Davidiellaceae*, *Capnodiales*). *Studies in Mycology* 67: 1–94.
- Bensch K, Groenewald JZ, Meijer M, et al. 2018. *Cladosporium* species in indoor environments. *Studies in Mycology* 89: 177–301.
- Berraf-Tebbal A, Bouznad Z, Santos JM, et al. 2011. *Phaeoacremonium* species associated with *Eutypa dieback* and *esca* of grapevines in Algeria. *Phytopathologia Mediterranea* 50: S86–S97.
- Bezerra JDP, Oliveira RJV, Paiva LM, et al. 2017. *Bezerromycetales* and *Wiesneriomycetales* ord. nov. (class *Dothideomycetes*), with two novel genera to accommodate endophytic fungi from Brazilian cactus. *Mycological Progress* 16: 297–309.
- Bidaud A, Carteret X, Eyssartier G, et al. 2002. Atlas des Cortinaires XII. Éditions Fédération mycologique Dauphiné Savoie, Marlioz, France.
- Bidaud A, Carteret X, Eyssartier G, et al. 2004. Atlas des Cortinaires XIV. Éditions Fédération mycologique Dauphiné Savoie, Marlioz, France.
- Bidaud A, Moëhne-Loccoz P, Reumaux P, et al. 2009. Atlas des Cortinaires XVIII. Éditions Fédération mycologique Dauphiné Savoie, Marlioz, France.
- Bien S, Damm U. 2020. *Arboricolonus simplex* gen. et sp. nov. and novelties in *Cadophora*, *Minutiella* and *Proliferodiscus* from *Prunus* wood in Germany. *MycoKeys* 2020 63: 119–161.
- Bien S, Kraus C, Damm U. 2020. Novel colophorina-like genera and species from *Prunus* trees and vineyards in Germany. *Persoonia* 45: 46–67.
- Boehm EWA, Mugambi GK, Miller AN, et al. 2009. A molecular phylogenetic reappraisal of the *Hysteriaceae*, *Mytiliniaceae* and *Gloniaceae* (Pleosporomycetidae, *Dothideomycetes*) with keys to world species. *Studies in Mycology* 64: 49–83.
- Boertmann D. 2010. The genus *Hygrocybe*, 2nd revised edition. Danish Mycological Society, Copenhagen.
- Bon M. 1992. Clé monographique des especes *Galero-Naucorioides*. *Documents Mycologiques* 84: 1–89.
- Bonito GM, Gryganskyi AP, Trappe JM, et al. 2010. A global meta-analysis of Tuber ITS rDNA sequences: species diversity, host associations and long-distance dispersal. *Molecular Ecology* 19: 4994–5008.
- Bonthond G, Sandoval-Denis M, Groenewald JZ, et al. 2018. *Seiridium* (*Sporocadaceae*): an important genus of plant pathogenic fungi. *Persoonia* 40: 96–118.
- Bragança CAD, Damm U, Baroncelli R, et al. 2016. Species of the *Colletotrichum acutatum* complex associated with anthracnose diseases of fruit in Brazil. *Fungal Biology* 120: 547–561.
- Brandrud TE, Dima B, Liimatainen K, et al. 2017. *Telamonioid Cortinarius* species of the *C. puellaris* group from calcareous *Tilia* forests. *Sydowia* 69: 37–45.
- Brandrud TE, Lindström H, Marklund H, et al. 1989. *Cortinarius Flora Photographica*. Vol. I (Swedish version). *Cortinarius* HB, Matfors, Sweden.
- Brandrud TE, Lindström H, Marklund H, et al. 1992. *Cortinarius Flora Photographica*. Vol. II (Swedish version). *Cortinarius* HB, Matfors, Sweden.
- Brandrud TE, Lindström H, Marklund H, et al. 1994. *Cortinarius Flora Photographica*. Vol. III (Swedish version). *Cortinarius* HB, Matfors, Sweden.
- Brandrud TE, Lindström H, Marklund H, et al. 1998. *Cortinarius Flora Photographica*. Vol. IV (Swedish version). *Cortinarius* HB, Matfors, Sweden.
- Brandrud TE, Lindström H, Marklund H, et al. 2012. *Cortinarius Flora Photographica*. Vol. V (Swedish version). *Cortinarius* HB, Matfors, Sweden.
- Brayford D, Chapman AU. 1987. *Cylindrocladium ilicicola* on cuttings of evergreen ornamental shrubs in the UK. *Plant Pathology* 36: 413–414.
- Cabral A, Groenewald JZ, Rego C, et al. 2012. *Cylindrocarpon* root rot: multi-gene analysis reveals novel species within the *Ilyonectria radicola* species complex. *Mycological Progress* 11: 655–688.
- Câmara MP, Ramaley AW, Castlebury LA, et al. 2003. *Neophaeosphaeria* and *Phaeosphaeriopsis*, segregates of *Paraphaeosphaeria*. *Mycological Research* 107: 516–522.
- Castresana J. 2000. Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. *Molecular Biology and Evolution* 17: 540–552.
- Ceruti A, Fontana A, Nosenzo C. 2003. Le specie europee del genere *Tuber*: una revisione storica. Vol. 37. Museo Regionale di Scienze Naturali, Turin, Italy.
- Chen CC, Chen CY, Lim YW, et al. 2020. Phylogeny and taxonomy of *Ceriperia* and other related taxa and description of three new species. *Mycologia* 112: 64–82.
- Chen WH, Liu C, Han YF, et al. 2019. Three novel insect-associated species of *Simplicillium* (*Cordycipitaceae*, *Hypocreales*) from Southwest China. *MycoKeys* 58: 83–102.
- Clements FE. 1909. The genera of fungi. The HW Wilson Company.
- Consiglio G, Setti L. 2008. Il genere *Crepidotus* in Europa. A.M.B. Fondazione Centro Studi Micologici, Vincenza.
- Cordeiro TRL, Nguyen TTT, Lima DX, et al. 2020. Two new species of the industrially relevant genus *Absidia* (*Mucorales*) from soil of the Brazilian Atlantic Forest. *Acta Botanica Brasiliica* 34: 549–558.
- Crous PW, Braun U, Schubert K, et al. 2007a. Delimiting *Cladosporium* from morphologically similar genera. *Studies in Mycology* 58: 33–56.
- Crous PW, Carnegie AJ, Wingfield MJ, et al. 2019a. Fungal Planet description sheets: 868–950. *Persoonia* 42: 291–473.
- Crous PW, Cowan DA, Maggs-Kölling G, et al. 2020a. Fungal Planet description sheets: 1112–1181. *Persoonia* 45: 251–409.
- Crous PW, Gams W. 2000. *Phaeomoniella chlamydospora* gen. et comb. nov., a causal organism of *Petri* grapevine decline and *esca*. *Phytopathologia Mediterranea* 39: 112–118.
- Crous PW, Groenewald JZ, Himaman W. 2007b. *Falcocladium thailandicum*. In: Crous PW, Seifert KA, Samson RA, et al. (eds), *Fungal Planet – A Global Initiative to promote the Study of Fungal Biodiversity*. CBS, Utrecht, Netherlands. *Fungal Planet* No. 18.
- Crous PW, Groenewald JZ, Summerell B. 2007c. *Exophiala placitae*. In: Crous PW, Seifert KA, Samson RA, et al. (eds), *Fungal Planet – A Global Initiative to promote the Study of Fungal Biodiversity*. CBS, Utrecht, Netherlands. *Fungal Planet* No. 17.
- Crous PW, Groenewald JZ, Wingfield MJ, et al. 2003. The value of ascospore septation in separating *Mycosphaerella* from *Sphaerulina* in the *Dothideales*: a Saccardoan myth? *Sydowia* 55: 136–152.

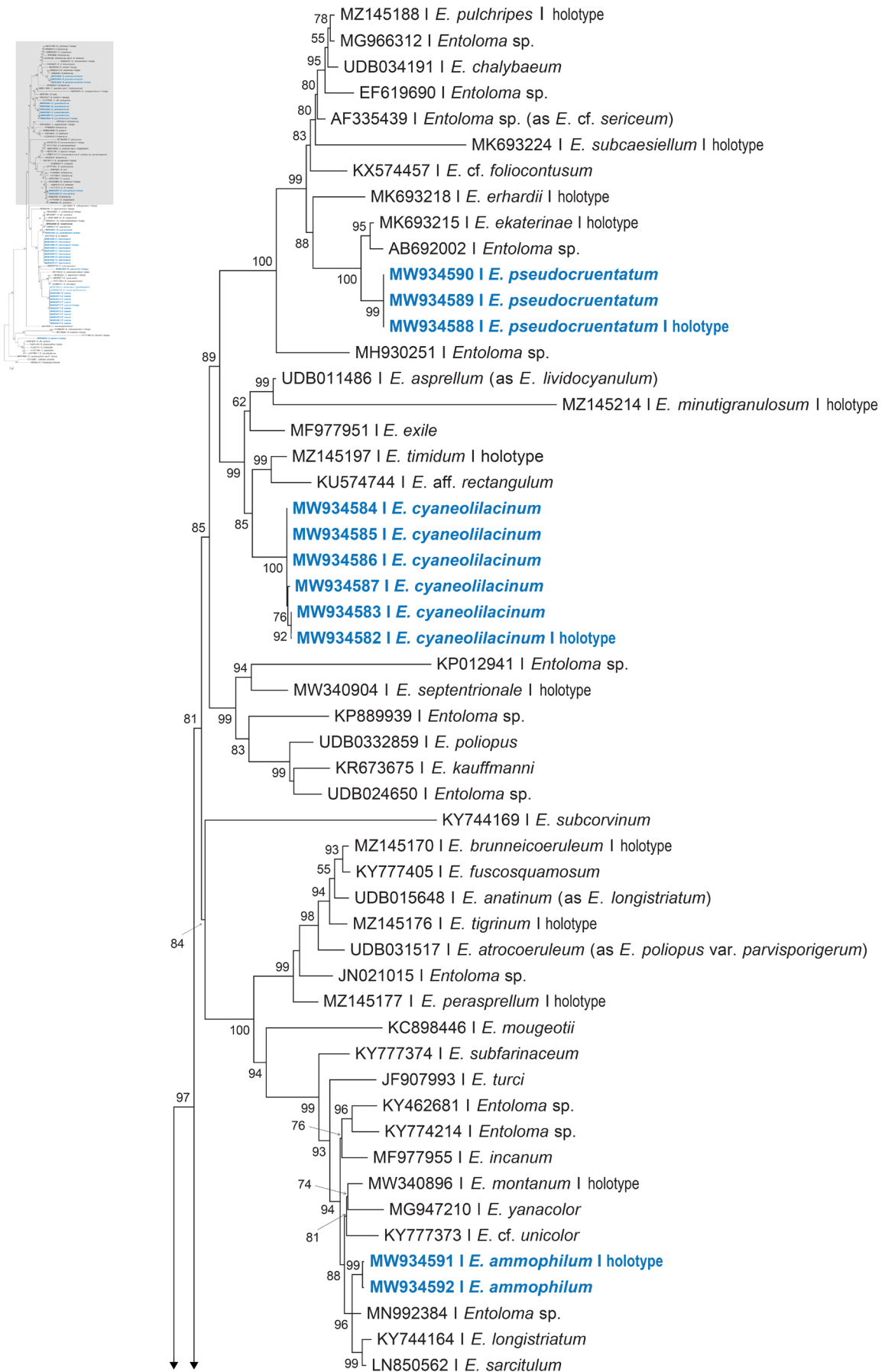
- Crous PW, Hernández-Restrepo M, Schumacher RK, et al. 2021. New and interesting fungi. 4. Fungal Systematics and Evolution 7: 255–343.
- Crous PW, Lennox CL, Sutton BC. 1995. *Selenophoma eucalypti* and *Stigmina robbenensis* spp. nov. from *Eucalyptus* leaves on Robben Island. Mycological Research 99: 648–652.
- Crous PW, Luangsa-ard JJ, Wingfield MJ, et al. 2018a. Fungal Planet description sheets: 785–867. Persoonia 41: 238–417.
- Crous PW, Schubert K, Braun U, et al. 2007d. Opportunistic, human-pathogenic species in the Herpotrichiellaceae are phenotypically similar to saprobic or phytopathogenic species in the Venturiaceae. Studies in Mycology 58: 185–217.
- Crous PW, Schumacher RK, Akulov A, et al. 2019b. New and interesting fungi. 2. Fungal Systematics and Evolution 3: 57–134.
- Crous PW, Schumacher RK, Wingfield MJ, et al. 2018b. New and interesting fungi. 1. Fungal Systematics and Evolution 1: 169–215.
- Crous PW, Shivas RG, Quaedvlieg W, et al. 2014a. Fungal Planet description sheets: 214–280. Persoonia 32: 184–306.
- Crous PW, Tanaka K, Summerell BA, et al. 2011. Additions to the *Mycosphaerella* complex. IMA Fungus 2: 49–64.
- Crous PW, Wingfield MJ, Burgess TI, et al. 2016. Fungal Planet description sheets: 469–557. Persoonia 37: 218–403.
- Crous PW, Wingfield MJ, Burgess TI, et al. 2017. Fungal Planet description sheets: 625–715. Persoonia 39: 270–467.
- Crous PW, Wingfield MJ, Cheewangkoon R, et al. 2019c. Foliar pathogens of eucalypts. Studies in Mycology 94: 125–298.
- Crous PW, Wingfield MJ, Groenewald JZ. 2009. Niche sharing reflects a poorly understood biodiversity phenomenon. Persoonia 22: 83–94.
- Crous PW, Wingfield MJ, Guarro J, et al. 2015a. Fungal Planet description sheets: 320–370. Persoonia 34: 167–266.
- Crous PW, Wingfield MJ, Le Roux JJ, et al. 2015b. Fungal Planet description sheets: 371–399. Persoonia 35: 264–327.
- Crous PW, Wingfield MJ, Lombard L, et al. 2019d. Fungal Planet description sheets: 951–1041. Persoonia 43: 223–425.
- Crous PW, Wingfield MJ, Mansilla JP, et al. 2006. Phylogenetic reassessment of *Mycosphaerella* spp. and their anamorphs occurring on *Eucalyptus*. II. Studies in Mycology 55: 99–131.
- Crous PW, Wingfield MJ, Schumacher RK, et al. 2014b. Fungal Planet description sheets: 281–319. Persoonia 33: 212–289.
- Crous PW, Wingfield MJ, Schumacher RK, et al. 2020b. New and interesting fungi. 3. Fungal Systematics and Evolution 6: 157–231.
- Damm U, Cannon PF, Woudenberg JHC, et al. 2012a. The *Colletotrichum acutatum* species complex. Studies in Mycology 73: 37–113.
- Damm U, Cannon PF, Woudenberg JHC, et al. 2012b. The *Colletotrichum boninense* species complex. Studies in Mycology 73: 1–36.
- Damm U, Fourie P, Crous PW. 2010. *Coniochaeta* (*Lecythophora*), *Collophora* gen. nov. and *Phaeomoniella* species associated with wood necroses of *Prunus* trees. Persoonia 24: 60–80.
- Damm U, Sato T, Alizadeh A, et al. 2019. The *Colletotrichum dracaenophilum*, *C. magnum* and *C. orchidearum* species complexes. Studies in Mycology 92: 1–46.
- Darriba D, Taboada GL, Doallo R, et al. 2012. jModelTest 2: more models, new heuristics and parallel computing. Nature Methods 9: 772.
- Das K, Sharma JR, Verbeken A. 2003. New species of *Lactarius* from Kumaon Himalaya, India. Mycotaxon 88: 333–342.
- De Almeida DAC, Gusmão LFP, Miller AN. 2016. Taxonomy and molecular phylogeny of *Diatrypaceae* (Ascomycota, Xylariales) species from the Brazilian semi-arid region, including four new species. Mycological Progress 15: 53.
- De Crop E, Nuytinck J, Van de Putte K, et al. 2014. *Lactifluus piperatus* (Russulales, Basidiomycota) and allied species in Western Europe and a preliminary overview of the group worldwide. Mycological Progress 13: 493–511.
- De Gruyter J, Woudenberg JHC, Aveskamp MM, et al. 2010. Systematic reappraisal of species in *Phoma* section *Paraphoma*, *Pyrenochaeta* and *Pleurophoma*. Mycologia 102: 1066–1081.
- De Hoog GS. 1972. The genera *Beauveria*, *Isaria*, *Tritirachium* and *Acrodontium* gen. nov. Studies in Mycology 1: 1–41.
- Dennis RWG. 1950. Karsten's species of *Mollisia*. Kew Bulletin 5: 171–187.
- Dubrule G, Pensec F, Picot A, et al. 2020. Phylogenetic diversity and effect of temperature on pathogenicity of *Colletotrichum lupini*. Plant Disease 104: 938–950.
- Edler D, Klein J, Antonelli A, et al. 2021. raxmlGUI 2.0: A graphical interface and toolkit for phylogenetic analyses using RAxML. Methods in Ecology and Evolution 12: 373–377.
- Egidi E, De Hoog GS, Isola D, et al. 2014. Phylogeny and taxonomy of meristematic rock-inhabiting black fungi in the *Dothideomycetes* based on multi-locus phylogenies. Fungal Diversity 65: 127–165.
- Ekanayaka AH, Hyde KD, Gentekaki E, et al. 2019. Preliminary classification of *Leotiomyces*. Mycosphere 10: 310–489.
- Ellis MB. 1971. *Dematiaceae Hyphomycetes*. CABI Publishing, Wallingford.
- Erdoğdu M, Özbek MU. 2017. First record of *Phaeoseptoria* and new species records on *Carex* for Turkey. Plant Pathology & Quarantine 7: 154–158.
- Fassatová O. 1986. Moulds and filamentous fungi in technical microbiology. Elsevier, Amsterdam.
- Garrido-Benavent I, Ballarà J, Mahiques R. 2014. *Cortinarius cad-aguirrei*, un nou tàxon de la secció *Fulvescentes* Melot. The Journal of the Journées européennes du Cortinaire 16: 24–34.
- Gelardi M, Angelini C, Costanzo F, et al. 2021. Outstanding pinkish brown-spored neotropical *Boletes*: *Austroboletus subflavidus* and *Fistulinella gloeocarpa* (*Boletaceae*, *Boletales*) from the Dominican Republic. Mycobiology 49: 24–45.
- Gierczyk B, Kubiński R. 2019. The first report of *Pleuroflammula ragazziana* in Poland. Acta Mycologica 54: 1121.
- Glynnou K, Ali T, Buch A-K, et al. 2016. The local environment determines the assembly of root endophytic fungi at a continental scale. Environmental Microbiology 18: 2418–2434.
- Gorfer M, Blumhoff M, Klaubauf S, et al. 2011. Community profiling and gene expression of fungal assimilatory nitrate reductases in agricultural soil. The ISME Journal 5: 1771–1783.
- Gräfenhan T, Schroers HJ, Nirenberg HI, et al. 2011. An overview of the taxonomy, phylogeny, and typification of nectriaceous fungi in *Cosmospora*, *Acremonium*, *Fusarium*, *Stilbella*, and *Volutella*. Studies in Mycology 68: 79–113.
- Gramaje D, Mostert L, Groenewald JZ, et al. 2015. *Phaeoacremonium*: From esca disease to phaeohyphomycosis. Fungal Biology 119: 759–783.
- Guarro J, Gené J, Stchigel AM, et al. 2012. Atlas of soil ascomycetes. CBS Biodiversity Series no. 10. Westerdijk Fungal Biodiversity Centre, Utrecht, the Netherlands.
- Guindon S, Gascuel O. 2003. A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. Systematic Biology 55: 696–704.
- Guindon S, Lethiec F, Duroux P, et al. 2010. PHYML Online – a web server for fast maximum likelihood-based phylogenetic inference. Nucleic Acids Research 33 (Web Server issue): W557–W559.
- Han JG, Hosoya T, Sung GH, et al. 2014. Phylogenetic reassessment of *Hyaloscyphaceae* sensu lato (*Helotiales*, *Leotiomyces*) based on multi-gene analyses. Fungal Biology 118: 150–167.
- Hansen EM, Maxwell DP. 1991. Species of the *Phytophthora megasperma* complex. Mycologia 83: 376–381.
- Hansen EM, Wilcox WF, Reeser PW, et al. 2009. *Phytophthora rosacearum* and *P. sansomeana*, new species segregated from the *Phytophthora megasperma* 'complex'. Mycologia 101: 129–135.
- Harrington AH, Del Olmo-Ruiz M, U'Ren JM, et al. 2019. *Coniochaeta endophytica* sp. nov., a foliar endophyte associated with healthy photosynthetic tissue of *Platycladus orientalis* (*Cupressaceae*). Plant and Fungal Systematics 64: 65–79.
- He F, Lin B, Sun J, et al. 2013. *Knufia aspidiotus* sp. nov., a new black yeast from scale insects. Phytotaxa 153: 39–50.
- Heilmann-Clausen J, Verbeken A, Vesterholt J. 1998. The genus *Lactarius* (Fungi of Northern Europe, Vol. 2). Danish Mycological Society, Copenhagen.
- Hennings P. 1901. Beiträge zur Flora von Afrika. XXI. *Fungi camerunenses* novi. III. Botanische Jahrbücher für Systematik Pflanzengeschichte und Pflanzengeographie 30: 39–57.
- Hernández-Restrepo M, Gené J, Castañeda-Ruiz RF, et al. 2017. Phylogeny of saprobic microfungi from Southern Europe. Studies in Mycology 86: 53–97.
- Hernández-Restrepo M, Groenewald JZ, Crous PW. 2016. Taxonomic and phylogenetic re-evaluation of *Microdochium*, *Monographella* and *Ildriella*. Persoonia 36: 57–82.
- Hesler LR. 1967. *Entoloma* in southeastern North America. Beihefte Nova Hedwigia 23: 1–245. Cramer, Germany.
- Hesler LR, Smith AH. 1963. North American species of *Hygrophorus*. University of Tennessee Press, Knoxville, Tennessee.
- Hesler LR, Smith AH. 1965. North American species of *Crepidotus*. Hafner Publishing Company, New York.
- Hesler LR, Smith AH. 1979. North American species of *Lactarius*. Ann Arbor, University of Michigan.
- Hesseltine CW, Ellis JJ. 1964. The genus *Absidia*: *Gongronella* and cylindrical-spored species of *Absidia*. Mycologia 56: 568–601.
- Hoang DT, Chernomor O, Von Haeseler A, et al. 2018. UFBBoot2: Improving the ultrafast bootstrap approximation. Molecular Biology and Evolution 35: 518–522.

- Holubová-Jechová V. 1978. Lignicolous hyphomycetes from Czechoslovakia 5. *Septonema*, *Hormiactella*, and *Lylea*. *Folia Geobotanica et Phytotaxonomica* 13: 421–442.
- Hongsanan S, Zhao RL, Hyde KD. 2017. A new species of *Chaetothyria* on branches of mango, and introducing *Phaeothecoidiaceae* fam. nov. *Mycosphere* 8: 137–146.
- Horak E. 1978. *Pleuroflammula*. *Persoonia* 9: 439–451.
- Horak E. 1986. Beiträge zur Systematik und Oekologie von *Pleuroflammula* (Agaricales, Fungi). Veröffentlichungen des Geobotanischen Institutes der Eidgenössische Technische Hochschule 87: 35.
- Horak E. 2018. Fungi of New Zealand. Volume 6. Agaricales (Basidiomycota) of New Zealand. 2. Brown spored genera p.p. *Crepidotus*, *Flammulaster*, *Inocybe*, *Phaeocollybia*, *Phaeomarasmius*, *Pleuroflammula*, *Pyrrhoglossum*, *Simocybe*, *Tubaria* and *Tympanella*. *Westerdijk Biodiversity Series* 16: 1–205.
- Houbraken J, Kocsubé S, Visagie, et al. 2020. Classification of *Aspergillus*, *Penicillium*, *Talaromyces* and related genera (Eurotiales): an overview of families, genera, subgenera, sections, series and species. *Studies in Mycology* 95: 5–169.
- Hubka V, Řeblová M, Řehulka J, et al. 2014. *Bradymyces* gen. nov. (Chaetothyriales, Trichomeriaceae), a new ascomycetous genus accommodating poorly differentiated melanized fungi. *Antonie van Leeuwenhoek* 106: 979–992.
- Huelsensbeck JP, Ronquist F. 2001. MrBayes: Bayesian inference of phylogenetic trees. *Bioinformatics* 17: 754–755.
- Isola D, Zucconi L, Onofri S, et al. 2016. Extremotolerant rock inhabiting black fungi from Italian monumental sites. *Fungal Diversity* 76: 75–96.
- Jančovičová S, Adamčík S. 2012. *Entoloma jahonii* (Fungi, Agaricales) reported from Slovakia and notes on differences with *E. byssisedum*. *Czech Mycology* 64: 209–222.
- Jayasiri SC, Hyde KD, Jones EBG, et al. 2019. Diversity, morphology and molecular phylogeny of Dothideomycetes on decaying wild seed pods and fruits. *Mycosphere* 10: 1–186.
- Johnston PR, Quijada L, Smith CA, et al. 2019. A multigene phylogeny toward a new phylogenetic classification of Leotiomycetes. *IMA Fungus* 10: 1–22.
- Kalyaanamoorthy S, Minh BQ, Wong TKF, et al. 2017. ModelFinder: Fast model selection for accurate phylogenetic estimates. *Nature Methods* 14: 587–589.
- Katoh K, Rozewicki J, Yamada KD. 2019. MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Briefings in Bioinformatics* 20: 1160–1166.
- Katoh K, Standley DM. 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Molecular Biology and Evolution* 30: 772–780.
- Khan Z, Gené J, Ahmad S, et al. 2013. *Coniochaeta polymorpha*, a new species from endotracheal aspirate of a preterm neonate, and transfer of *Lecythophora* species to *Coniochaeta*. *Antonie van Leeuwenhoek* 104: 243–252.
- Knapp DG, Kovács GM, Zajta E, et al. 2015. Dark septate endophytic pleosporalean genera from semiarid areas. *Persoonia* 35: 87–100.
- Kobmoo N, Mongkolsamrit S, Tسانathai K, et al. 2012. Molecular phylogenies reveal host-specific divergence of *Ophiocordyceps unilateralis sensu lato* following its host ants. *Molecular Ecology* 21: 3022–3031.
- Kornerup A, Wanscher JH. 1978. *Methuen handbook of colour*. 3rd ed. London: Eyre Methuen.
- Kozlov AM, Darriba D, Flouri T, et al. 2019. RAXML-NG: a fast, scalable and user-friendly tool for maximum likelihood phylogenetic inference. *Bioinformatics* 35: 4453–4455.
- Kraus C, Damm U, Bien S, et al. 2020. New species of *Phaeomoniellales* from a German vineyard and their potential threat to grapevine (*Vitis vinifera*) health. *Fungal Systematics and Evolution* 6: 139–155.
- Kriegelsteiner L. 2004. Ascomycetenfunde während des Seminars an der Schwarzwälder Pilzlehrschau vom 23. bis 27. Juni 2003. *Zeitschrift für Mykologie* 70: 49–58.
- Kubátová A. 2006. *Chaetomium* in the Czech Republic and notes to three new records. *Czech Mycology* 58: 155–171.
- Kühner R, Romagnesi H. 1953. *Flore analytique des champignons supérieurs* (Agarics, Bolets, Chanterelles). Paris.
- Kumar AM, Vrinda KB, Pradeep CK. 2018a. Two new species of *Crepidotus* (Basidiomycota, Agaricales) from peninsular India. *Phytotaxa* 372: 67–78.
- Kumar S, Stecher G, Li M, et al. 2018b. MEGA X: molecular evolutionary genetics analysis across computing platforms. *Molecular Biology and Evolution* 35: 1547–1549.
- Kumar S, Stecher G, Tamura K. 2016. MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution* 33: 1870–1874.
- Kurtzman CP. 1995. Relationships among the genera *Ashbya*, *Eremothecium*, *Holleya* and *Nematospora* determined from rDNA sequence divergence. *Journal of Industrial Microbiology* 14: 523–530.
- Kurtzman CP, Fell JW, Boekhout T, et al. 2011. Methods for isolation, phenotypic characterization and maintenance of yeasts. In: Kurtzman CP, Fell JW, Boekhout T (eds), *The yeasts* (fifth edition): 87–110. Elsevier.
- Kurtzman CP, Robnett CJ. 2003. Phylogenetic relationships among yeasts of the 'Saccharomyces complex' determined from multigene sequence analyses. *FEMS Yeast Research* 3: 417–432.
- Kurtzman CP, Robnett CJ, Basehoar E, et al. 2018. Four new species of *Metschnikowia* and the transfer of seven *Candida* species to *Metschnikowia* and *Clavispora* as new combinations. *Antonie van Leeuwenhoek* 111: 2017–2035.
- Kušán I, Matočec N, Antonić O, et al. 2014. Biogeographical variability and re-description of an imperfectly known species *Hamatocanthoscypha rotundispora* (Helotiales, Hyaloscyphaceae). *Phytotaxa* 170: 1–12.
- Kuyper TW. 1986. A revision of the genus *Inocybe* in Europe. I. Subgenus *Inosperma* and the smooth-spored species of subgenus *Inocybe*. *Persoonia Supplement* 3: 1–247.
- Kytövuori I, Niskanen T, Liimatainen T, et al. 2005. *Cortinarius sordidemaculatus* and two new related species, *C. anisatus* and *C. neofurvolaeus*, in Fennoscandia (Basidiomycota, Agaricales). *Karstenia* 45: 33–49.
- Lachance MA. 2011. *Starmerella Rosa & Lachance* (1998). In: Kurtzman CP, Fell JW, Boekhout T (eds), *The yeasts, a taxonomic study*, vol II: 811–815. Elsevier, New York.
- Lachance MA. 2016. *Metschnikowia*: half tetrads, a regicide and the fountain of youth. *Yeast* 33: 563–574.
- Læssøe T. 2008. *Hydropus* Singer. In: Knudsen H, Vesterholt J (eds), *Funga Nordica: Agaricoid, boletoid and cyphelloid genera*: 282–285. Nordsvamp, Copenhagen.
- Le Gal M, Mangenot MF. 1958. Contribution à l'étude des Mollisioïdées. II. (1re série). *Revue de Mycologie* 23: 28–86.
- Le Gal M, Mangenot MF. 1961. Contribution à l'étude des Mollisioïdées. IV. (3e série). *Revue de Mycologie* 26: 263–331.
- Lechat C, Crous PW, Groenewald JZ. 2010. The enigma of *Calonectria* species occurring on leaves of *Ilex aquifolium* in Europe. *IMA Fungus* 1: 101–108.
- Li DM, Chen XR. 2010. A new superficial fungal infection caused by *Coniosporium epidermidis*. *Journal of the American Academy of Dermatology* 63: 725–727.
- Li GJ, Hyde KD, Zhao RL, et al. 2016. Fungal diversity notes 253–366: taxonomic and phylogenetic contributions to fungal taxa. *Fungal Diversity* 78: 1–237.
- Liang J, Li G, Zhao M, et al. 2019. A new leaf blight disease of turfgrasses caused by *Microdochium poae*, sp. nov., *Mycologia* 111: 265–273.
- Liimatainen K, Niskanen T, Dima B. 2020. Mission impossible completed: unlocking the nomenclature of the largest and most complicated subgenus of *Cortinarius*, *Telamonia*. *Fungal Diversity* 104: 291–331.
- Lima DX, Cordeiro TR, De Souza CA, et al. 2020. Morphological and molecular evidence for two new species of *Absidia* from Neotropical soil. *Phytotaxa* 446: 61–71.
- Lin CG, Bhat DJ, Liu JK, et al. 2019. The genus *Castanediella*. *MycKeys* 51: 1–14.
- Liu F, Cai L. 2012. Morphological and molecular characterization of a novel species of *Simplicillium* from China. *Cryptogamie Mycologie* 33: 137–144.
- Liu F, Wang J, Li H, et al. 2019. *Setophoma* spp. on *Camellia sinensis*. *Fungal Systematics and Evolution* 4: 43–57.
- Liu H, Li T, Ding Y, et al. 2017. Dark septate endophytes colonizing the roots of 'non-mycorrhizal' plants in a mine tailing pond and in a relatively undisturbed environment, Southwest China. *Journal of Plant Interactions* 12: 264–271.
- Liu Q, Li JQ, Wingfield MJ, et al. 2020. Reconsideration of species boundaries and proposed DNA barcodes for *Calonectria*. *Studies in Mycology* 97: 100106.
- Lodge DJ, Padamsee M, Matheny PB, et al. 2014. Molecular phylogeny, morphology, pigment chemistry and ecology in *Hygrophoraceae* (Agaricales). *Fungal Diversity* 64: 1–99.
- Lombard L, Houbraken J, Decock C, et al. 2016. Generic hyper-diversity in *Stachybotriaceae*. *Persoonia* 36: 156–246.
- Lombard L, Van der Merwe NA, Groenewald JZ, et al. 2015. Generic concepts in *Nectriaceae*. *Studies in Mycology* 80: 189–245.
- Luttrell ES. 1976. Ovarian infection of *Sporobolus poiretii* by *Bipolaris ravenelii*. *Phytopathology* 66: 260–268.
- MacKenzie SJ, Peres NA, Barquero MP, et al. 2009. Host range and genetic relatedness of *Colletotrichum acutatum* isolates from fruit crops and leather-leaf fern in Florida. *Phytopathology* 99: 620–631.
- Magnago AC, Neves MA, Da Silveira BRM. 2017. *Fistulinella ruschii*, sp. nov., and a new record of *Fistulinella campinaranae* var. *scrobiculata* for the Atlantic Forest, Brazil. *Mycologia* 109: 1003–1013.

- Mahiques R, Mateos A, Reyes JD, et al. 2013. Algunos Cortinarioides de Sierra Mágina y Despeñaperros (Jaén). I. *Lactarius* 22: 7–49.
- Marin-Felix Y, Hernández-Restrepo M, Iturrieta-González I, et al. 2019a. Genera of phytopathogenic fungi: GOPHY 3. *Studies in Mycology* 94: 1–124.
- Marin-Felix Y, Hernández-Restrepo M, Wingfield MJ, et al. 2019b. Genera of phytopathogenic fungi: GOPHY 2. *Studies in Mycology* 92: 47–133.
- Melot J. 1990. Une classification du genre *Cortinarius* (Pers.) S.F. Gray. *Documents Mycologiques* 20: 43–59.
- Meng W, Damodara B, Li W, et al. 2017. Molecular phylogeny of *Neodeverisia*, with two new species and several new combinations. *Mycologia* 109: 965–974.
- Miller MA, Pfeiffer W, Schwartz T. 2010. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. In: *Proceedings of the Gateway Computing Environments Workshop (GCE)*, 14 Nov. 2010, New Orleans: 1–8.
- Minh BQ, Nguyen MA, Von Haeseler A. 2013. Ultrafast approximation for phylogenetic bootstrap. *Molecular Biology and Evolution* 30: 1188–1195.
- Mishra B, Thines M. 2014. siMBA – a simple graphical user interface for the Bayesian phylogenetic inference program MrBayes. *Mycological Progress* 13: 1255–1258.
- Molia A, Larsson E, Jeppson M, et al. 2020. *Elaphomyces* section *Elaphomyces* (Eurotiales, Ascomycota) – taxonomy and phylogeny of North European taxa, with the introduction of three new species. *Fungal Systematics and Evolution* 5: 283–300.
- Moreno-Arroyo B, Llistosella J, De la Osa LR. 2002. *Gymnomyces sublevissporus* (Russulales), una nueva especie de la región mediterránea. *Revista Catalana de Micologia* 24: 179–186.
- Morgan-Jones G. 1975. Notes on hyphomycetes. VIII. *Lylea*, a new genus. *Mycotaxon* 3: 129–132.
- Morozova OV, Malysheva EV, Popov ES, et al. 2015. Macromycetes of the Izborsk-Maly Valley, rare and new to the Pskov Region. *Novosti Sistematiki Vysshikh i Nizshikh Rastenii* 49: 186–203.
- Moser M. 1978. Die Röhrlinge und Blätterpilze, 4th edition. In: Gams H (ed), *Kleine Kryptogamenflora* 11b/2. Fischer Verlag, Stuttgart.
- Moser M. 1983. Die Röhrlinge und Blätterpilze. In: Gams H (ed), *Kleine Kryptogamenflora*, Band 11b/2, 5th edn. Fischer Verlag, Stuttgart, Germany.
- Moser M. 2001. Rare, debated and new taxa of the genus *Cortinarius* (Agaricales). *Fungi Delineati* 15: 1–57.
- Moser M, Horak E. 1975. *Cortinarius* Fr. und nahe verwandte Gattungen in Südamerika. *Beihefte Nova Hedwigia* 52: 1–628.
- Moser M, McKnight KH, Ammirati JF. 1995. Studies on North American *Cortinarii* I. New and interesting taxa from the greater Yellowstone area. *Mycotaxon* 55: 301–346.
- Mostert L, Groenewald JZ, Summerbell RC, et al. 2006. Taxonomy and pathology of *Togninia* (Diaporthales) and its *Phaeoacremonium* anamorphs. *Studies in Mycology* 54: 1–115.
- Munsell Soil Color Charts. 1954 edition. Munsell Color, Baltimore, Maryland, USA.
- Nasr S, Bien S, Soudi MR, et al. 2018. Novel *Collophorina* and *Coniochaeta* species from *Euphorbia polycaulis*, an endemic plant in Iran. *Mycological Progress* 17: 755–771.
- Nguyen L-T, Schmidt HA, Von Haeseler A, et al. 2015. IQ-TREE: A fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Molecular Biology and Evolution* 32: 268–274.
- Nguyen NH, Vellinga EC, Bruns TD, et al. 2016. Phylogenetic assessment of global *Suillus* ITS sequences supports morphologically defined species and reveals synonymous and undescribed taxa. *Mycologia* 108: 1216–1228.
- Nirenberg HI, Feiler U, Hagedorn G. 2002. Description of *Colletotrichum lupini* comb. nov. in modern terms. *Mycologia* 94: 307–320.
- Niskanen T, Kytövuori I, Liimatainen K. 2009. *Cortinarius* sect. *Brunnei* (Basidiomycota, Agaricales) in North Europe. *Mycological Research* 113: 182–206.
- Niskanen T, Kytövuori I, Liimatainen K, et al. 2013. *Cortinarius* section *Bovini* (Agaricales, Basidiomycota) in northern Europe, conifer associated species. *Mycologia* 105: 977–993.
- Nonaka K, Kaifuchi S, Omura S, et al. 2013. Five new *Simplicillium* species (Cordycipitaceae) from soils in Tokyo, Japan. *Mycoscience* 54: 42–53.
- Noordeloos ME. 1984. Studies in *Entoloma* 10–13. *Persoonia* 12: 193–122.
- Noordeloos ME. 1987. *Entoloma* (Agaricales) in Europe. Synopsis and keys to all species and a monograph of the subgenera *Trichopilus*, *Inocephalus*, *Alboleptonia*, *Leptonia*, *Paraleptonia*, and *Omphaliopsis*. *Beihefte zur Nova Hedwigia* 91: 1–419.
- Noordeloos ME. 1988. *Entoloma* in North America. The species described by L.R. Hesler, A.H. Smith & S.J. Mazzer: type-species and comments. *Cryptogamic Studies*, Vol. 2. Gustav Fisher Verlag, Stuttgart, Germany.
- Noordeloos ME. 1992. *Entoloma* s.l. *Fungi Europaei*, vol. 5. Giovanna Biella, Saronno, Italy.
- Noordeloos ME. 2004. *Entoloma* s.l. *Fungi Europaei*, vol. 5a. Edizione Candusso, Italy.
- Orton PD. 1960. New check-list of British Agarics and Boleti. Part 3: Notes on genera and species in the list. *Transactions of the British Mycological Society Supplement* 43: 159–439.
- Parmelee JA. 1956. The identification of the *Curvularia* parasite of *Gladiolus*. *Mycologia* 48: 558–567.
- Paz A, Bellanger JM, Lavoise C, et al. 2017. The genus *Elaphomyces* (Ascomycota, Eurotiales): a ribosomal DNA-based phylogeny and revised systematics of European 'deer truffles'. *Persoonia* 38: 197–239.
- Pethybridge GH. 1913. On the rotting of potato tubers by a new species of *Phytophthora* having a method of sexual reproduction hitherto undescribed. *Scientific Proceedings of the Royal Dublin Society* 13: 529–565.
- Pethybridge GH, Lafferty HA. 1919. A disease of tomato and other plants caused by a new species of *Phytophthora*. *Scientific Proceedings of the Royal Dublin Society* 15: 487–503.
- Phukhamsakda C, McKenzie EHC, Phillips AJL, et al. 2020. Microfungi associated with *Clematis* (Ranunculaceae) with an integrated approach to delimiting species boundaries. *Fungal Diversity* 102: 1–203.
- Pitt JI, Lantz H, Pettersson OV, et al. 2013. *Xerochrysum* gen. nov. and *Bettisia*, genera encompassing xerophilic species of *Chrysosporium*. *IMA Fungus* 4: 229–241.
- Quaedvlieg W, Binder M, Groenewald JZ, et al. 2014. Introducing the Consolidated Species Concept to resolve species in the *Teratosphaeriaceae*. *Persoonia* 33: 1–40.
- Quaedvlieg W, Verkley GJM, Shin H-D, et al. 2013. Sizing up *Septoria*. *Studies in Mycology* 75: 307–390.
- Quélet L. 1886. *Les Champignons du Jura et des Vosges*. C. r. Ass. Franc. Av. Sci. (Grenoble, 1885): 446.
- Ramírez C, Martínez AT. 1981. Seven new species of *Penicillium* and a new variety of *Penicillium novae-caledoniae* Smith. *Mycopathologia* 74: 35–49.
- Rashmi M, Kushveer JS, Sarma VV. 2019. A worldwide list of endophytic fungi with notes on ecology and diversity. *Mycosphere* 10: 798–1079.
- Rayner RW. 1970. A mycological colour chart. Commonwealth Mycological Institute, Kew and British Mycological Society.
- Raza M, Zhang Z-F, Hyde KD, et al. 2019. Culturable plant pathogenic fungi associated with sugarcane in southern China. *Fungal Diversity* 99: 1–104.
- Ridgway R. 1912. Color standards and color nomenclature. Ridgway, Washington, DC.
- Ronquist F, Huelsenbeck JP. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19: 1572–1574.
- Ronquist F, Teslenko M, Van der Mark P, et al. 2012. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology* 61: 539–542.
- Safaiefarahani B, Mostowfizadeh-Ghalemlarsa RSTJ, Hardy GSJ, et al. 2015. Re-evaluation of the *Phytophthora* cryptogea species complex and the description of a new species, *Phytophthora pseudocryptogea* sp. nov. *Mycological Progress* 14: 108.
- Salom JC, Esteve-Raventós F. 2011. *Phaeomarasmium siquierii* (Agaricoid clade, Tubariaceae), a new Mediterranean resupinate species found in Formentera (Balearic Islands, Spain). *Micologia e Vegetazione Mediterranea* 26: 29–36.
- Samarakoon BC, Wanasinghe DN, Phookamsak R, et al. 2021. *Stachybotrys musae* sp. nov., *S. microsporus*, and *Memnoniella levispora* (Stachybotryaceae, Hypocreales) Found on Bananas in China and Thailand. *Life* 11: 323.
- Samuels GJ. 1977. *Nectria consors* and its *Volutella* conidial state. *Mycologia* 69: 255–262.
- Sandoval-Denis M, Gené J, Sutton DA, et al. 2016. New species of *Cladosporium* associated with human and animal infections. *Persoonia* 36: 281–298.
- Sandoval-Denis M, Guarnaccia V, Polizzi G, et al. 2018. Symptomatic *Citrus* trees reveal a new pathogen lineage in *Fusarium* and two new *Neocosmospora* species. *Persoonia* 40: 1–25.
- Schiller M, Lübeck M, Sundelin T, et al. 2006. Two subpopulations of *Colletotrichum acutatum* are responsible for anthracnose in strawberry and leatherleaf fern in Costa Rica. *European Journal of Plant Pathology* 116: 107–118.
- Segeth MP, Bonnefoy A, Broenstrup M, et al. 2003. Coniosetin, a novel tetramic acid antibiotic from *Coniochaeta ellipsoidea* DSM 13856. *The Journal of Antibiotics* 56: 114–122.
- Séguy E. 1936. *Encyclopedie Pratique du Naturaliste*, 30. Paul Lechevalier, Paris.
- Shang QJ, Phookamsak R, Camporesi E, et al. 2018. The holomorph of *Fusarium celtidicola* sp. nov. from *Celtis australis*. *Phytotaxa* 361: 251–265.
- Shivas RG, Smith MW, Marney TS, et al. 2005. First record of *Nematospora coryli* in Australia and its association with dry rot of *Citrus*. *Australasian Plant Pathology* 34: 99–101.

- Shoemaker RA, Babcock CE. 1989. Phaeosphaeria. *Canadian Journal of Botany* 67: 1500–1599.
- Shoemaker RA, Babcock CE. 1992. Applanodictyosporous Pleosporales: Clathrospora, Comoclathris, Graphyllum, Macrospora, and Platysporoides. *Canadian Journal of Botany* 70: 1617–1658.
- Singer R. 1947. 'The Boletoidae of Florida with notes on extralimital species III'. *American Midland Naturalist* 37: 1–135.
- Smith AH, Hesler LR. 1968. *The North American species of Pholiotia*. Hafner Publishing Co., New York, USA.
- Smith AH, Thiers HD. 1964. A contribution toward a monograph of the North American species of *Suillus* (Boletaceae). Lubrecht & Cramer, Ann Arbor, Michigan.
- Sousa TF, Dos Santos AO, Da Silva FMA, et al. 2020. *Arcopilus amazonicus* (Chaetomiaceae), a new fungal species from the Amazon rainforest native plant *Paullinia cupana*. *Phytotaxa* 456: 145–156.
- Spies CFJ, Moyo P, Halleen F, et al. 2018. Phaeoacremonium species diversity on woody hosts in the Western Cape Province of South Africa. *Persoonia* 40: 26–62.
- Stamatakis A. 2014. RAxML version 8: A toll for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* 30: 1312–1313.
- Sun W, Su L, Yang S, et al. 2020. Unveiling the hidden diversity of rock-inhabiting fungi: Chaetothiriales from China. *Journal of Fungi* 6: 187.
- Svrček M. 1987. New or less known Discomycetes. XVI. *Česká Mykologie* 41: 88–96.
- Swofford DL. 2003. PAUP*. Phylogenetic Analysis Using Parsimony (*and other methods). Version 4. Sinauer Associates. Sunderland, MA.
- Tamura K, Nei M. 1993. Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Molecular Biology and Evolution* 10: 512–526.
- Tamura K, Stecher G, Peterson D, et al. 2013. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Molecular Biology and Evolution* 30: 2725–2729.
- Tan YP, Crous PW, Shivas RG. 2018. Cryptic species of *Curvularia* in the culture collection of the Queensland Plant Pathology Herbarium. *MycKeys* 35: 1–25.
- Tanaka K, Hirayama K, Yonezawa H, et al. 2009. Molecular taxonomy of bambusicolous fungi: Tetraplosporiaceae, a new pleosporalean family with Tetraploa-like anamorphs. *Studies in Mycology* 64: 175–209.
- Tanaka K, Hirayama K, Yonezawa H, et al. 2015. Revision of the Massariaceae (Pleosporales, Dothideomycetes). *Studies in Mycology* 82: 75–136.
- Tanney JB, Seifert KA. 2020. Mollisiaceae: An overlooked lineage of diverse endophytes. *Studies in Mycology* 95: 293–380.
- Tennakoon DS, Thambugala KM, Wanasinghe DN, et al. 2020. Additions to Phaeosphaeriaceae (Pleosporales): *Elongaticollum* gen. nov., *Ophiophaerella taiwanensis* sp. nov., *Phaeosphaeriopsis beaucarnea* sp. nov. and a new host record of *Neosetophoma poaeicola* from Musaceae. *MycKeys* 70: 59–88.
- Thambugala KM, Ariyawansa HA, Li Y, et al. 2014. Dothideales. *Fungal Diversity* 68: 105–158.
- Trifinopoulos J, Nguyen L-T, Von Haeseler A, et al. 2016. W-IQ-TREE: a fast online phylogenetic tool for maximum likelihood analysis. *Nucleic Acids Research* 44 (W1): W232–W235.
- Tsui CKM, Leung YM, Hyde KD, et al. 2001. Three new *Ophioceras* species (Ascomycetes) from the tropics. *Mycoscience* 42: 321–326.
- Tsuneda A, Currah RS. 2005. Pleomorphic conidiogenesis among strains of *Knufia cryptophialidica*. *Canadian Journal of Botany* 83: 510–517.
- Tsuneda A, Hambleton S, Currah RS. 2011. The anamorph genus *Knufia* and its phylogenetically allied species in *Coniosporium*, *Sarcinomyces*, and *Phaeococcomyces*. *Botany* 89: 523–536.
- Tucker CM. 1931. Taxonomy of the genus *Phytophthora* de Bary. *Research Bulletin of the Missouri Agricultural Experiment Station* 153: 207.
- Tulasne LR, Tulasne C. 1851. *Fungi Hypogaei, Histoire et Monographie des Champignons Hypogés*. F. Klincksieck (ed.), Paris, France.
- Untereiner WA, Gueidan C, Orr MJ, et al. 2011. The phylogenetic position of the lichenicolous ascomycete *Capronia peltigerae*. *Fungal Diversity* 49: 225–233.
- Vasco-Palacios AM, Lopez-Quintero CA, Franco-Molano AE, et al. 2014. *Austroboletus amazonicus* sp. nov. and *Fistulinella campinaranae* var. *scrobiculata*, two commonly occurring boletes from a forest dominated by *Pseudomonotes tropenbosii* (Dipterocarpaceae) in Colombian Amazonia. *Mycologia* 106: 1004–1014.
- Verbeke A, Van de Putte K, De Crop E. 2012. New combinations in *Lactifluus*, 3: L. subgenera *Lactifluus* and *Piperati*. *Mycotaxon* 120: 443–450.
- Verwoerd L, Du Plessis SJ. 1931. Descriptions of some new species of South African fungi and species not previously recorded from South Africa. III. *South African Journal of Science* 28: 290–297.
- Vidal JM, Alvarado P, Loizides M, et al. 2019. A phylogenetic and taxonomic revision of sequestrate Russulaceae in Mediterranean and temperate Europe. *Persoonia* 42: 127–185.
- Videira SIR, Groenewald JZ, Braun U, et al. 2016. All that glitters is not *Ramularia*. *Studies in Mycology* 83: 49–163.
- Videira SIR, Groenewald JZ, Nakashima C, et al. 2017. *Mycosphaerellaceae* – chaos or clarity? *Studies in Mycology* 87: 257–421.
- Vizzini A. 2008. Novitates. *Tubariaceae* fam. nov. *Rivista di Micologia* 51: 174.
- Vizzini A, Consiglio G, Marchetti M. 2019. *Mythicomyetaceae* fam. nov. (Agaricineae, Agaricales) for accommodating the genera *Mythicomyces* and *Stagnicola*, and *Simocybe parvispora* reconsidered. *Fungal Systematics and Evolution* 3: 41–56.
- Voitk A, Saar I, Lodge J, et al. 2020. New species and reports of *Cuphophyllus* from northern North America compared with related Eurasian species. *Mycologia* 112: 438–452.
- Von Arx JA, Guarro J, Figueras MJ. 1986. The ascomycete genus *Chaetomium*. *Beihefte zur Nova Hedwigia* 84: 1–162.
- Walsh E, Luo J, Zhang N. 2014. *Acidomelania panicicola* gen. et sp. nov. from switchgrass roots in acidic New Jersey pine barrens. *Mycologia* 106: 856–864.
- Wanasinghe DN, Phukhamsakda C, Hyde KD, et al. 2018. Fungal diversity notes 709–839: taxonomic and phylogenetic contributions to fungal taxa with an emphasis on fungi on Rosaceae. *Fungal Diversity* 89: 1–236.
- Wang GS, Zhou Y, Xue L, et al. 2020. *Teunia rosae* sp. nov. and *Teunia rudbeckiae* sp. nov. (Cryptococcaceae, Tremellales), two novel basidiomycetous yeast species isolated from flowers. *International Journal of Systematics and Evolutionary Microbiology* 70: 5394–5400.
- Wang H-J, Gloer JB, Scott JA, et al. 1995. *Coniochaetones* A and B: new antifungal benzopyranones from the coprophilous fungus *Coniochaeta saccardoii*. *Tetrahedron Letters* 36: 5847–5850.
- Wang XW, Houbraken J, Groenewald JZ, et al. 2016. Diversity and taxonomy of *Chaetomium* and *chaetomium*-like fungi from indoor environments. *Studies in Mycology* 84: 145–224.
- Watling R. 1975. Observations on the *Bolbitiaceae* 11: A species of *Bolbitius* with ornamented basidiospores. *Notes from the Royal Botanic Garden, Edinburgh* 34: 241–244.
- Watling R. 1987. Observations on the *Bolbitiaceae* – 30. *Agaricus callistus* Peck. *Mycologia* 79: 310–313.
- Wei DP, Wanasinghe DN, Hyde KD, et al. 2019. The genus *Simplicillium*. *MycKeys* 60: 69–92.
- Whitton SR, McKenzie EHC, Hyde KD. 2012. Anamorphic Fungi associated with Pandanaceae. In: Whitton SR, McKenzie EHC, Hyde KD (eds), *Fungi associated with Pandanaceae*: 125–353. Springer, Dordrecht.
- Xia J-W, Ma Y-R, Zhang X-G. 2014. New species of *Corynesporopsis* and *Lylea* from China. *Sydowia* 66: 241–248.
- Xie J, Strobel GA, Feng T, et al. 2015. An endophytic *Coniochaeta velutina* producing broad spectrum antimicrobials. *Journal of Microbiology* 53: 390–397.
- Yang X, Tyler BM, Hong C. 2017. An expanded phylogeny for the genus *Phytophthora*. *IMA Fungus* 8: 355–384.
- Zare R, Gams W. 2001. A revision of *Verticillium* section *Prostrata*. IV. The genera *Lecanicillium* and *Simplicillium* gen. nov. *Nova Hedwigia* 73: 1–50.
- Zhang D, Gao F, Jakovlić I, et al. 2020. *PhyloSuite*: An integrated and scalable desktop platform for streamlined molecular sequence data management and evolutionary phylogenetics studies. *Molecular Ecology Resources* 20: 348–355.
- Zhang ZF, Liu F, Zhou X, et al. 2017. Culturable mycobiota from Karst caves in China, with descriptions of 20 new species. *Persoonia* 39: 1–31.

Fungal Planet 1240 – *Entoloma ammophilum*



FP1240 Phylogenetic tree derived from Maximum Likelihood analysis based on nrITS1-5.8S-ITS2 data. Analysis was performed in PhyML v. 3.0 (Guindon et al. 2010) using the non-parametric Shimodaira-Hasegawa version of the approximate likelihood-ratio test (SH-aLRT) and the GTR+I+ Γ model of evolution. ML bootstrap support values are shown at the nodes (BS > 50 %).

