

Multiple colonizations from Madagascar and converged acquisition of dioecy in the Mascarene Dombeyoideae (Malvaceae) as inferred from chloroplast and nuclear DNA sequence analyses

Timothée Le Péchon^{1,5,*}, Jean-Yves Dubuisson¹, Thomas Haevermans², Corinne Cruaud³, Arnaud Couloux³ and Luc D. B. Gigord⁴

¹Université Pierre et Marie Curie, UMR 7207 CNRS-MNHN-UPMC “Centre de Recherche sur la Paléobiodiversité et les Paléoenvironnements”, bâtiment de géologie, CP48, 57 rue Cuvier, F-75005 Paris, France, ²Muséum National d’Histoire Naturelle, UMR 7205 CNRS “Origine, Structure et Evolution de la Biodiversité”, Herbarium National, 57 rue Cuvier, 75231 Paris cedex 05, France, ³Genoscope. Centre National de Séquençage, 2, rue Gaston Crémieux, CP5706, 91057 Evry Cedex, France,

⁴Conservatoire Botanique National de Mascarin, 2, rue du Père Georges, F-97436, Saint Leu, La Réunion and ⁵IREMIA-LIM, Université de La Réunion, Parc Technologique Universitaire, bâtiment 2, 2 rue Joseph Wetzell, 97490 Sainte Clotilde, France

* For correspondence. E-mail tlpechon@snv.jussieu.fr

Received: 1 December 2009 Returned for revision: 1 February 2010 Accepted: 10 May 2010 Published electronically: 18 June 2010

• **Background and Aims** In the Mascarenes, a young oceanic archipelago composed of three main islands, the Dombeyoideae (Malvaceae) have diversified extensively with a high endemism rate. With the exception of the genus *Trochetia*, Mascarene Dombeyoideae are described as dioecious whereas Malagasy and African species are considered to be monoecious, species with individuals bearing hermaphrodite/perfect flowers. In this study, the phylogenetic relationships were reconstructed to clarify the taxonomy, understand the phylogeographic pattern of relationships and infer the evolution of the breeding systems for the Mascarenes Dombeyoideae.

• **Methods** Parsimony and Bayesian analysis of four DNA markers (ITS, *rpl16* intron and two intergenic spacers *trnQ-rsp16* and *psbM-trnD*) was used. The molecular matrix comprised 2985 characters and 48 taxa. The Bayesian phylogeny was used to infer phylogeographical hypotheses and the evolution of breeding systems.

• **Key Results** Parsimony and Bayesian trees produced similar results. The Dombeyoideae from the Mascarenes are polyphyletic and distributed among four clades. Species of *Dombeya*, *Trochetia* and *Ruizia* are nested in the same clade, which implies the paraphyly of *Dombeya*. Additionally, it is shown that each of the four clades has an independent Malagasy origin. Two adaptive radiation events have occurred within two endemic lineages of the Mascarenes. The polyphyly of the Mascarene Dombeyoideae suggests at least three independent acquisitions of dioecy.

• **Conclusions** This molecular phylogeny highlights the taxonomic issues within the Dombeyoideae. Indeed, the limits and distinctions of the genera *Dombeya*, *Trochetia* and *Ruizia* should be reconsidered. The close phylogeographic relationships between the flora of the Mascarenes and Madagascar are confirmed. Despite their independent origins and a distinct evolutionary history, each endemic clade has developed a different breeding systems (dioecy) compared with the Malagasy Dombeyoideae. Sex separation appears as an evolutionary convergence and may be the consequence of selective pressures particular to insular environments.

Key words: Dombeyoideae, Mascarene archipelago, *Dombeya*, *Ruizia*, *Trochetia*, dioecy, Indian Ocean, biogeography, ITS, *rpl16* intron, *psbM-trnD*, *trnQ-rsp16*.

INTRODUCTION

Since the observations of Darwin in the Galapagos, oceanic islands have been considered as ideal natural laboratories for the study of evolutionary processes. This is a direct consequence of their isolation from the continents, high geological dynamics, high habitat diversity, well-established flora and fauna, and the reduced gene flow between insular and continental populations (Emerson, 2002). In particular, oceanic islands and archipelago display the most striking examples of adaptive radiations (Baldwin, 1991; Seung-Chul *et al.*, 1996; Austin *et al.*, 2004). In comparison to Hawaii and the Galapagos, the evolutionary history of the Mascarene archipelago (also named Mascarenes) biodiversity has been overlooked, although it is included in a biodiversity hotspot

(Myers *et al.*, 2000). This archipelago is located 700 km east of Madagascar and comprises three young oceanic islands (Reunion island, 2500 km², 2.1 My; Mauritius, 1800 km², 7.2 My; Rodrigues, 110 km², 8–10 My or 1.2 My for the emerged part of the island; McDougall *et al.*, 1965; Giorgi and Borchiellini, 1998; Hanyu *et al.*, 2001; Sheth *et al.*, 2003). The largest island, La Réunion, possesses an active volcano and a high mountainous formation with the highest point at 3070 m a.s.l. As a consequence, the island shows numerous and diverse ecosystems (Strasberg *et al.*, 2005). Some angiosperm groups contain a high number of endemic species. For instance, the Dombeyoideae, subfamily of Malvaceae (ex-‘Sterculiaceae’) includes 23 species (22 Mascarene endemics, one indigenous but not endemic) distributed in four genera: *Trochetia* DC., *Ruizia* Cav., *Astiria*

Lind. – all restricted to the archipelago – and *Dombeya* Cav. (Friedmann, 1987). The subfamily is mainly distributed in Madagascar (with seven genera and 250 species out of 350 in total), but also in Africa, Australia and tropical Asia (Bayer and Kubitzki, 2003). In addition to their high specific diversity, Dombeyoideae play an important ecological role in Mascarene ecosystems (Cadet, 1980; Blanchard, 2000). The subfamily members occur in most of the habitats found on the islands. The most diverse genus *Dombeya* (14 species) forms the canopy of the tropical mountain cloud forests (hence the local name ‘coloured tree forest’, due to the large range of leaf coloration among different *Dombeya* species; Cadet, 1980). Despite their key ecological role, phylogenetic relationships among the Mascarene Dombeyoideae and species from other regions are poorly understood, and no molecular phylogenetic studies have been conducted on this subfamily so far. Several taxonomic studies have been carried out for this group of Mascarene species (Cavanilles, 1787; Baker, 1877; Jacob de Cordemoy, 1895; Arènes, 1959a; Friedmann, 1987), but major disagreements exist between some of the authors (Table 1). Recently, a phylogenetic study based on morphological characters (Le Péchon et al., 2009) cast doubt on the validity of the genera *Dombeya*, *Astiria* and *Ruizia*.

Molecular phylogenetic relationships could potentially clarify the taxonomy of the Dombeyoideae genera endemic to the Mascarene Archipelago. Furthermore the molecular phylogenies can be used to understand both the phylogeographic history and character evolution of the Mascarene Dombeyoideae. The endemic Mascarene species of *Dombeya*, *Astiria* and *Ruizia* demonstrate specific breeding systems that have not been recorded in the Malagasy and continental African species. While bearing hermaphroditic flowers in these two regions, *Dombeya* species in the Mascarenes were described as dioecious by Friedmann (1987) and Jacob de Cordemoy (1895). These observations were confirmed and detailed by Humeau (1999) and Humeau et al. (1999a, b). Molecular phylogenies can be used to infer the evolution of dioecy acquisition in the Mascarene taxa and to determine whether sex separation within the Dombeyoideae was acquired by evolutionary convergence.

In this study, the first molecular phylogeny of Mascarene Dombeyoideae based on four DNA markers is presented. The main aims were to (a) compare molecular and morphological phylogenetic hypotheses (Le Péchon et al., 2009); (b) clarify the taxonomy of the Dombeyoideae genera endemic to the Mascarenes; (c) construct a phylogeographic pattern for Mascarene Dombeyoideae; and finally (d) infer the evolution of breeding systems.

MATERIALS AND METHODS

Taxonomic sampling

With the purpose of reconstructing the relationships among the Dombeyoideae, taxa were chosen to represent both the morphological and ecological diversity of the subfamily. As the study is focused on the Mascarene species, the majority of the sampling is composed of taxa endemic to this archipelago. Some species are found at several localities in the Indian Ocean (in the Mascarenes as well as in Madagascar; Table 2). In those cases, a specimen from each island was included in the sampling. A total of 26 specimens from the Mascarenes, including one undescribed species from Mauritius (*Dombeya* sp. 143), was selected. To establish whether the Mascarene Dombeyoideae are monophyletic, numerous taxa from outside the three islands were also included. Most Dombeyoideae diversity is found on Madagascar. Several studies (Cadet, 1980; Blanchard, 2000; Malcomber, 2002; Plunkett et al., 2004; Micheneau et al., 2008) show important relationships between the Malagasy and the Mascarene floras. Accordingly, a maximum number (i.e. 16 species) of Malagasy species was included, which also represents the maximum number of subsections of *Dombeya* defined by Arènes (1959b), involving three unidentified species (*Dombeya* sp. 252, *Dombeya* sp. 277 and *Dombeya* sp. 310). Three African species, one Asian species and one species from Saint Helena Island were added. Depending on the authors (Arènes, 1959a; Le Péchon et al., 2009), the taxonomic relationships between the genera *Trochetia*, *Helmiopsis* H.Perrier and *Dombeya* are conflicting. To test the phylogenetic relationships among these three

TABLE 1. Comparison between the classification of Arènes (1959a, b) and Friedmann (1987) for the Mascarene taxa of Dombeyoideae

	Family	Genus	Subgenus	Section	No. of taxa	% of endemism	
Arènes (1959a)	Sterculiaceae	<i>Dombeya</i>	<i>Dombeya</i>	<i>Astrapea</i>	1	0	
				<i>Assonia</i>	1	100	
				<i>Trochetia</i>	6	100	
				<i>Capricornua</i>	3	33	
				<i>Dombeya</i>	17	88	
Friedmann (1987)	Sterculiaceae	<i>Ruizia</i>			1	100	
		<i>Astiria</i>			1	100	
		<i>Trochetia</i>			6	100	
		<i>Dombeya</i>					
			<i>Dombeya</i>			14	92

TABLE 2. List of the *Dombeyoideae* taxa included in the analyses and Genbank accession numbers of the sequences

Taxon	Origin; voucher number*	<i>Trnq-rps16</i>	<i>Rpl16</i>	<i>Psbm-trnD</i>	ITS
<i>Dombeya acutangula</i> Cav.	Madagascar; Labat 3654 (P)	GU937945	–	GU937896	GU938038
<i>Dombeya acutangula</i> subsp. <i>acutangula</i> var. <i>acutangula</i> Arènes	Réunion; Le Péchon 58 (P)	GU937946	GU937994	GU937897	GU938039
<i>Dombeya acutangula</i> subsp. <i>acutangula</i> var. <i>acutangula</i>	Cult. CBNB; Le Péchon 131 (P)	GU937947	GU937995	GU937898	GU938040
<i>Dombeya acutangula</i> subsp. <i>rosea</i> Friedmann aff. var. <i>acutangula</i>	Mauritius; Le Péchon 153 (P)	GU937948	GU937996	GU937899	GU938041
<i>Dombeya acutangula</i> subsp. <i>rosea</i> aff. var. <i>palmata</i>	Mauritius; Le Péchon 154 (P)	GU937949	GU937997	GU937900	GU938042
<i>Dombeya amaniensis</i> Engl.	Tanzania; Phillipson 4834 (P)	GU937950	GU937998	GU937901	GU938043
<i>Dombeya blattiolens</i> Frapp. ex Cordem.	Réunion; Le Péchon 294 (P)	GU937951	GU937999	GU937902	GU938044
<i>Dombeya brevistyla</i> Arènes	Réunion; Labat 2863 (P)	GU937952	GU938000	GU937903	GU938045
<i>Dombeya burgessiae</i> Gerr. ex Harv. and Sond	Tropical greenhouses of MNHN Paris; Specimen 15936	GU937953	GU938001	GU937904	GU938046
<i>Dombeya cacuminum</i> Hochr.	Cult. CBNB; Le Péchon 133 (P)	GU937954	GU938002	GU937905	GU938047
<i>Dombeya ciliata</i> Cordem.	Réunion; Le Péchon 262 (P)	GU937955	GU938003	GU937906	GU938048
<i>Dombeya delislei</i> Arènes	Réunion; Le Péchon 114 (P)	GU937956	GU938004	GU937907	GU938049
<i>Dombeya elegans</i> Cordem. var. <i>elegans</i>	Réunion; Le Péchon 129 (P)	GU937957	GU938005	GU937908	GU938050
<i>Dombeya elegans</i> var. <i>virescens</i> Cordem.	Réunion; Le Péchon 272 (P)	GU937958	GU938006	GU937909	GU938051
<i>Dombeya farafanganica</i> Arènes subsp. <i>endrina</i>	Madagascar; Razakamalala <i>et al.</i> 2114 (MO)	GU937959	GU938007	GU937910	GU938052
<i>Dombeya ferruginea</i> Cav. subsp. <i>borbonica</i> Friedmann	Réunion; Le Péchon 22 (P)	GU937960	GU938008	GU937911	GU938053
<i>Dombeya ferruginea</i> subsp. <i>ferruginea</i>	Mauritius; Le Péchon 155 (P)	GU937961	GU938009	GU937912	GU938054
<i>Dombeya ficulnea</i> Baill.	Réunion; Le Péchon 3 (P)	GU937962	GU938010	GU937913	GU938055
<i>Dombeya lucida</i> Baill.	Madagascar; Ravelonarivo <i>et al.</i> 2011 (MO)	GU937963	GU938011	GU937914	GU938056
<i>Dombeya macrantha</i> Baker	Madagascar; Labat 3671 (P)	GU937964	GU938012	GU937915	GU938057
<i>Dombeya mauritiana</i> Friedmann	Mauritius; Le Péchon 129 (P)	GU937965	GU938013	GU937916	GU938058
<i>Dombeya pilosa</i> Cordem.	Réunion; Le Péchon 65 (P)	GU937966	GU938014	GU937917	GU938059
<i>Dombeya populnea</i> Baker	Mauritius; Le Péchon 141 (P)	GU937967	GU938015	GU937918	GU938060
<i>Dombeya populnea</i>	Réunion; Le Péchon 80 (P)	GU937968	GU938016	GU937919	GU938061
<i>Dombeya punctata</i> Cav.	Réunion; Le Péchon 1 (P)	GU937969	GU938017	GU937920	GU938062
<i>Dombeya reclinata</i> Cordem.	Réunion; Le Péchon 6 (P)	GU937970	GU938018	GU937921	GU938063
<i>Dombeya rodriguesiana</i> Friedmann	Rodriguez; Le Péchon; 160 (P)	GU937971	GU938019	GU937922	GU938064
<i>Dombeya rotleroides</i> Baill.	Madagascar; SW 60080 (P)	GU937972	GU938020	GU937923	GU938065
<i>Dombeya</i> sp. 143	Mauritius; Le Péchon 143 (P)	GU937973	GU938021	GU937924	GU938066
<i>Dombeya</i> sp. 252	Madagascar; Rakotonirina <i>et al.</i> 252 (MO)	GU937974	GU938022	GU937925	GU938067
<i>Dombeya</i> sp. 277	Madagascar; Rakotonirina <i>et al.</i> 277 (MO)	GU937975	GU938023	GU937926	GU938068
<i>Dombeya</i> sp. 310	Madagascar; Rakotonirina <i>et al.</i> 310 (MO)	GU937976	–	GU937927	GU938069
<i>Dombeya superba</i> Arènes	Madagascar; Rakotonirina <i>et al.</i> 293 (MO)	GU937977	GU938024	GU937928	GU938070
<i>Dombeya tiliacea</i> Planch.	South Africa; Phillipson 4834 (P)	GU937978	GU938025	GU937929	GU938071
<i>Dombeya tremula</i> Hochr.	Madagascar; Phillipson 5815 (P)	GU937979	–	GU937930	GU938072
<i>Dombeya umbellata</i> Cav.	Réunion; Le Péchon 115 (P)	GU937980	GU938026	GU937931	GU938073
<i>Dombeya viburniflora</i> Boj.	Madagascar; Labat 3771 (P)	GU937981	GU938027	GU937932	GU938074
<i>Helmiopsis bernieri</i> (Baill.) Arènes	Madagascar; Capuron 20937 (P)	GU937982	GU938028	GU937933	GU938075
<i>Helmiopsis pseudopopulus</i> (Baill.) Capuron	Madagascar; Capuron 27412 (P)	GU937983	–	GU937934	GU938076
<i>Nesogordonia crassipes</i> (Baill.) Capuron	Madagascar; Rakotonirina <i>et al.</i> 385 (MO)	GU937984	–	GU937935	GU938077
<i>Nesogordonia suzannae</i> Labat, Munzinger and O.Pascal	Mayotte; Berthelot 1327 (P)	GU937985	GU938029	GU937936	GU938078
<i>Pterospermum heterophyllum</i> Hance	–	–	–	–	AY083661
<i>Pterospermum suberifolium</i> (L.) Wild	Cult. Tropical greenhouses of MNHN Paris; Specimen 5616 (P)	GU937986	GU938030	GU937937	–
<i>Ruizia cordata</i> Cav.	Réunion; Le Péchon 81 (P)	GU937987	GU938031	GU937938	GU938079
<i>Trochetia blackburniana</i> Bojer ex Baker	Mauritius; Le Péchon 144 (P)	GU937988	GU938032	GU937939	GU938080
<i>Trochetia boutoniana</i> Friedmann	Mauritius; Le Péchon 147 (P)	GU937989	GU938033	GU937940	GU938081
<i>Trochetia granulata</i> Cordem.	Réunion; Le Péchon 101 (P)	GU937990	GU938034	GU937941	GU938082
<i>Trochetia parviflora</i> Boj.	Mauritius; Le Péchon 162 (P)	GU937991	GU938035	GU937942	GU938083
<i>Trochetia triflora</i> DC.	Mauritius; Le Péchon 162bis (P)	GU937992	GU938036	GU937943	GU938084
<i>Trochetiopsis erythroxylo</i> (G.Forst.) W.Marais	Cult. CBNB; Le Péchon 132 (P)	GU937993	GU938037	GU937944	GU938085

*CBNB, Conservatoire Botanique National de Brest.

genera with molecular markers, two species of *Helmiopsis* from Madagascar were included. Also included was *Trochetiopsis erythroxyton* endemic to Saint Helena Island, which appears morphologically close to *Dombeya* and *Trochetia* (*Trochetiopsis* was formerly considered to be a taxonomic synonym of *Trochetia*; Brodie *et al.*, 1998). *Nesogordonia* Baill. was chosen as an outgroup to root the inferred phylogenies according to Barnett (1987) and Nyffeler *et al.* (2005) who showed that this genus diverged earlier among the Dombeyoideae. Thus two species of *Nesogordonia* (i.e. *Nesogordonia crassipes* and *Nesogordonia suzannae*) were selected as outgroup members. To complete the outgroup selection, the Asian Dombeyoideae genus *Pterospermum* Schreb., which is morphologically and genetically distinct from *Dombeya* (Tang, 1992; Nyffeler *et al.*, 2005) was also added. The sample thus included a total of 50 specimens comprising 48 taxa. The sampling and voucher numbers are given in Table 2.

Extraction of DNA, amplification of molecular markers, and sequencing protocols

For DNA extraction, all samples originating from the Mascarene Archipelago were fresh leaf material collected from either wild specimens or from cultivated plants (Table 2), dehydrated with silica gel. The major part of the Malagasy species sampling came from herbarium specimens from which only small samples were taken. From this material, the DNA was extracted using the DNeasy Plant Mini (Qiagen) extraction kit, following the manufacturer's instructions.

To reconstruct the relationships among the Dombeyoideae, four molecular markers were selected: the nuclear ribosomal region of internal transcribed spacers (ITS), the chloroplastic spacers *trnQ-rps16* and *psbM-trnD*, and the chloroplastic intron *rpl16*. The primers used to amplify and sequence these regions are listed in Table 3. For the *rpl16* intron, two new internal primers were designed to facilitate the sequencing, as the presence of poly(T) residues led to sequencing difficulties. The amplifications were carried out in a final volume of 25 μL with 0.1 – 3 μL of genomic DNA of unknown concentration, 2.5 μL of specific buffer for the *REDTaq* at a concentration of $\times 10$ and containing MgCl_2 , 1 μL of dNTPs (a final concentration of 100 μM for each nucleotide), 1.25 μL of DMSO (5 % of the final volume), 1 μL of each primer (at 10 pmol μL^{-1}) and 0.15 μL of *REDTaq* DNA polymerase (Sigma-Aldrich); purified water was added to bring the total

quantity to 25 μL . The polymerase chain reactions (PCR) were carried out with a Biometra T3000 thermal cycler. The PCR conditions for the *rpl16* intron and the *trnQ-rps16* intergenic spacer were: initial denaturation at 80 °C for 5 min followed by 35 cycles of 1 min at 95 °C, 1 min at 50 °C and 5 min at 65 °C, a final extension at 65 °C for 4 min. For the *trnD-psbM* spacer, the PCR conditions were: initial denaturation at 80 °C for 5 min followed by 35 cycles of 1 min at 94 °C, 1 min at 55 °C and 3.5 min at 72 °C, a final extension was carried out at 72 °C for 5 min. For the ITS, the following protocol was used: initial denaturation at 94 °C for 3 min followed by 35 cycles of 30 s at 94 °C, 1 min at 55 °C and 1 min at 72 °C, a final extension was carried out at 72 °C for 10 min. PCR results were checked by migrating 2 μL of the product on a 1.5 % agarose gel with ethidium bromide. The PCR purifications and sequencing were carried out at the Centre national de séquençage (Genoscope; <http://www.genoscope.cns.fr/spip/>).

Sequence alignment

The resulting sequences were edited, assembled and aligned visually using the program Bioedit (Hall, 1999). Proper sequence alignment required the introduction of many insertion/deletion events (indels). In the present analysis, the gaps were treated with ID coding (Barriel, 1994, 2004) which has proven its usefulness in several studies (Delarbre *et al.*, 2000, 2002; Raymunde *et al.*, 2002; Hennequin *et al.*, 2003; Rouhan *et al.*, 2004).

Phylogenetic analyses

Two different methods to reconstruct Dombeyoideae phylogenetic relationships were used: the maximum parsimony (MP) and the Bayesian inference (BI). For the MP analysis, the tree search was performed using PAUP* ver. 4.0b1.0 (Swofford, 2001) with a heuristic search including the following options: 1000 replicates of random addition sequence and tree bisection and reconnection branch swapping. A preliminary MP search that included only plastid markers failed to find all the most-parsimonious trees resulting in an almost completely unresolved strict consensus of the trees (data not shown). A second MP analysis including only ITS sequences was performed. The strict consensus tree (data not shown) resulting from this search was better resolved than the strict consensus of plastid markers. Nevertheless, it presented several important

TABLE 3. Selected molecular markers and their primers used in this study

Molecular marker	Primer name	Sequence (5' → 3')	Reference
<i>trnQ-rps16</i>	trnQ ^(uug)	GCG TGG CCA AGY GGT AAG GC	Shaw <i>et al.</i> , 2007
	rps16x1	GTT GCT TTY TAC CAC ATC GTT T	Shaw <i>et al.</i> , 2007
<i>rpl16</i>	rpl16F71	GCT ATG CTT AGT GTG TGAC TCG TTG	Pfeil <i>et al.</i> , 2002
	rpl16R1516	CCC TTC ATT CTT CCT CTA TG TTG	Pfeil <i>et al.</i> , 2002
	rpl16F310	CCA ATA TGT AAG GTC TAT GAA GC	This study
	rpl16R613	TTT TCA ATA GAA TCC TCT GCA TTC	This study
<i>psbM-trnD</i>	psbMF	AGC AAT AAA TGC RAG AAT ATT TAC TTC CAT	Shaw <i>et al.</i> , 2005
	trnD(guc)R	GGG ATT GTA GYT CAA TTG GT	Shaw <i>et al.</i> , 2005
ITS	ITS.leu	GTC CAC TGA ACC TTA TCA TTT AG	Baum <i>et al.</i> , 1998
	ITS4	TCC TTC CGC TTA TTG ATA TGC	White <i>et al.</i> , 1990

polytomies. These two consensus trees (i.e. chloroplastic and nuclear ribosomal consensus) did not show major topological incongruence. Therefore, it was decided to combine the data from the different DNA regions to increase the resolution and the support values. The node support was estimated with a bootstrap analysis (BS) with the following search options: heuristic search with 1000 000 replicates without branch swapping and one random addition sequence per replicate. Clades having a BS of 85–100 % will be considered here as highly supported, and 75–84 % as moderately supported.

The BI analysis was performed using MrBayes 3 (Ronquist and Huelsenbeck, 2003). For each of the four markers, an appropriate model of evolution was selected with the help of MrModeltest ver. 2 (Nylander, 2004) implementing the Akaike criterion. For the four datasets, the GTR + Γ model selected by MrModeltest was applied. A fifth partition, corresponding to the insertion and deletion coded with the ID coding (Barriel, 1994, 2004), was treated using a binary evolution model.

The molecular data analysis with BI was carried out using the following options. Four Markov chains derived using the Monte-Carlo procedure were utilized, and the trees were sampled every 100 generations for a total of 10 000 000 generations. The number of trees sampled before the $-\ln L$ values stabilized ('burn-in phase') was determined graphically. As a result, the first 10 000 generations that were required to achieve Markov chain convergence were discarded. The remaining trees were then used to calculate the majority consensus tree that provided the posterior probability (PP) on each node. Clades are described here as highly supported when PP is above 0.94, and as moderately supported when the PP value is between 0.85 and 0.94.

Phylogeographic inference

Among the sample, seven different geographic regions were represented: continental Africa, tropical Asia, Madagascar (including the Comoro archipelago), Saint Helena and the Mascarene archipelago represented by Mauritius, La Réunion and Rodrigues. Considered as a non-additive multi-state character, the evolution of geographic distribution was inferred using parsimony on the BI majority consensus tree, using Mesquite (Maddison and Maddison, 2006).

Evolution of breeding systems

The endemic Dombeyoideae of the Mascarenes, apart from *Trochetia*, are described as dioecious (Table 4). The evolutionary history of breeding systems was inferred using the same method employed for the phylogeography. In case of ambiguous optimization, two types were assessed: AccTran optimization (i.e. accelerated transformations, favouring reversions) and DelTran (i.e. delayed transformations, favouring convergences).

RESULTS

Combined data analysis

Altogether, 49 sequences were obtained for each intergenic spacer (*trnQ-rps16* and *psbM-trnD*), 48 sequences for the ITS and 44 sequences for the *rpl16* intron.

TABLE 4. Breeding systems [Malagasy species (Arènes, 1959), African species (Seyani, 1991), Mascarene species (Friedmann, 1987; Gigord et al., 1998; Humeau, 1999, Humeau et al., 1999a)] and geographical distribution of each taxon included in the analyses

Taxon	Breeding system	Distribution
<i>Dombeya acutangula</i> subsp. <i>acutangula</i> var. <i>acutangula</i>	Hermaphroditic	Réunion
<i>D. acutangula</i> subsp. <i>acutangula</i> var. <i>acutangula</i>	Hermaphroditic	Rodrigues
<i>D. acutangula</i> subsp. <i>rosea</i> aff. var. <i>acutangula</i>	Hermaphroditic	Mauritius
<i>D. acutangula</i> subsp. <i>rosea</i> aff. var. <i>palmata</i>	Hermaphroditic	Mauritius
<i>D. acutangula</i>	Hermaphroditic	Madagascar
<i>D. amaniensis</i>	Hermaphroditic	Africa
<i>D. blattiolens</i>	Dioecious	Réunion
<i>D. brevistyla</i>	Hermaphroditic	Madagascar
<i>D. burgessiae</i>	Hermaphroditic	Africa
<i>D. cacuminum</i>	Hermaphroditic	Madagascar
<i>D. ciliata</i>	Dioecious	Réunion
<i>D. delislei</i>	Dioecious	Réunion
<i>D. elegans</i> var. <i>elegans</i>	Dioecious	Réunion
<i>D. elegans</i> var. <i>virescens</i>	Dioecious	Réunion
<i>D. farafanganica</i> subsp. <i>endrina</i>	Hermaphroditic	Madagascar
<i>D. ferruginea</i> subsp. <i>borbonica</i>	Dioecious	Réunion
<i>D. ferruginea</i> subsp. <i>ferruginea</i>	Dioecious	Mauritius
<i>D. ficulnea</i>	Dioecious	Réunion
<i>D. lucida</i>	Hermaphroditic	Madagascar
<i>D. macrantha</i>	Hermaphroditic	Madagascar
<i>D. mauritiana</i>	Dioecious	Réunion
<i>D. pilosa</i>	Dioecious	Réunion
<i>D. populnea</i>	Dioecious	Réunion
<i>D. populnea</i>	Dioecious	Mauritius
<i>D. punctata</i>	Dioecious	Réunion
<i>D. reclinata</i>	Dioecious	Réunion
<i>D. rodriguesiana</i>	Dioecious	Rodrigues
<i>D. rotteroides</i>	Hermaphroditic	Madagascar
<i>D. sp. 143</i>	?	Mauritius
<i>D. sp. 252</i>	?	Madagascar
<i>D. sp. 277</i>	?	Madagascar
<i>D. sp. 310</i>	?	Madagascar
<i>D. superba</i>	Hermaphroditic	Madagascar
<i>D. tiliacea</i>	Hermaphroditic	Africa
<i>D. tremula</i>	Hermaphroditic	Madagascar
<i>D. umbellata</i>	Dioecious	Réunion
<i>D. viburnifolia</i>	Hermaphroditic	Madagascar
<i>Helmiopsis pseudopopulus</i>	Hermaphroditic	Madagascar
<i>H. bernieri</i>	Hermaphroditic	Madagascar
<i>Nesogordonia suzannae</i>	Hermaphroditic	Madagascar
<i>N. crassipes</i>	Hermaphroditic	Madagascar
<i>Pterospermum heterophyllum</i>	Hermaphroditic	South-east Asia
<i>P. suberifolium</i>	Hermaphroditic	South-east Asia
<i>Ruizia cordata</i>	Dioecious	Réunion
<i>Trochetia blackburniana</i>	Hermaphroditic	Mauritius
<i>T. boutoniana</i>	Hermaphroditic	Mauritius
<i>T. granulata</i>	Hermaphroditic	Réunion
<i>T. parviflora</i>	Hermaphroditic	Mauritius
<i>T. triflora</i>	Hermaphroditic	Mauritius
<i>T. erythroxylo</i>	Hermaphroditic	Saint Helena

Despite a good ITS amplification for *Pterospermum suberifolium*, sequencing was unsuccessful. The sequence of the ITS for the closely related species *Pterospermum heterophyllum* was available in Genbank (Table 2). To represent the Asian genus *Pterospermum*, a chimeric sequence combining the

chloroplastic sequences of *P. suberifolium* and the nuclear sequence of *P. heterophyllum* was reconstructed.

After aligning and coding the indels, the number of characters used for the phylogenetic analyses was: 439 for *psbM-trnD* [with eight (1.8%) informative characters for the ingroup and eight characters due to the gap handling], 690 for *trnQ-rsp16* [with 35 (5.07%) informative characters for the ingroup and 40 characters due to the gap handling], 694 for the ITS [with 71 (10.23%) informative characters for the ingroup and 38 characters due to the gap handling]. When aligning the chloroplastic intron *rpl16*, a particular region (position 438–530) was discovered that could not be aligned. The alignment issues in this intron-specific region were noted previously by Baum *et al.* (1998) and Pfeil *et al.* (2002). As a consequence, and to avoid any bias in the phylogeny reconstruction, the region of *rpl16* from position 438 to 530 was excluded. Finally, *rpl16* had a total of 1160 characters that were used for the analysis [with 52 (4.48%)–informative characters for the ingroup and 55 characters due to the gap handling].

Following sequence alignment, exclusion of the ambiguous region of the *rpl16* intron, and treatment of the indels with ID coding, the final matrix contained 50 specimens included within 48 taxa and 2985 molecular characters. The unavailable sequences (due to amplification failure) were coded as missing data.

Phylogenetic analyses

A total of 9402 most-parsimonious trees were obtained. The strict consensus is given in Fig. 1. For the most part, the same relationships were retrieved with the BI. However, the BI majority consensus tree (Fig. 2) appears to be more resolved and shows more better-supported nodes.

In the two consensus trees shown in Figs 1 and 2, Clade A contains the *Dombeya* species, in addition to the taxa belonging to *Trochetia*, *Ruizia* and *Helmiopsis*, and is moderately supported (BS = 78%, PP = 0.8). Within Clade A, all the *Dombeyoideae* from the Mascarenes are distributed into four subclades (referred to here as B, C, E and F).

Dombeya rodriguesiana is included in Clade B, which is weakly supported (BS = 50%, PP = 0.8) and otherwise includes only taxa from Madagascar. The two *Helmiopsis* species (i.e. *H. pseudopopulus* and *H. bernieri*), which are endemic to Madagascar, constitute a subclade that is highly supported in both analyses (BS = 100%, PP = 1). Clade B is the sister-group to a clade that contains all the other members of *Dombeya*, *Trochetia* and *Ruizia* (BS = 83%, PP = 1).

Trochetia and *Ruizia*, in addition to six species of *Dombeya*, are included in Clade C, which is moderately supported by the MP (BS = 76%), but more strongly supported by the BI (PP = 0.95). This clade contains only endemic species of the Mascarene archipelago. Because of the basal polytomy, the MP analysis does not allow the phylogenetic relationships between the species of this clade to be precisely established. Regarding the consensus obtained with BI analysis (Fig. 2), Clade C appears completely resolved and is divided into two monophyletic groups. A first subclade, the moderately supported Clade C₁ (PP = 0.9, Fig. 2), includes species of *Dombeya* in addition to the single known species of *Ruizia*,

which is endemic to La Reunion. Clade C₁ is the sister-group of Clade C₂, which is moderately supported (PP = 0.88, Fig. 2). This monophyletic group includes two subspecies of *Dombeya ferruginea* (PP = 0.99) and the species of *Trochetia*. The latter, endemic to the Mascarenes, appears to be monophyletic (PP = 0.86).

The sister-group of Clade C is the species *Dombeya tremula*, which is endemic to Madagascar. The monophyletic group (*D. tremula* + Clade C) is weakly supported by the MP analysis (BS = 63%) but has a strong support in the BI analysis (PP = 1). This group (*D. tremula* + Clade C) is the sister-group of Clade D that only involves *Dombeya* species. Within Clade D, the endemic species of *Dombeya* from the Mascarenes are distributed between Clades E and F (Figs 1 and 2). Clade E includes all the subspecies of *D. acutangula*, which are endemic to the Mascarenes. However, this group appears weakly supported by the MP (BS = 64%), but better supported by the BI analysis (PP = 0.95). The sister-group of Clade E is *D. acutangula* (BS = 80%, PP = 1) from Madagascar, which supports the monophyly of this widespread species.

The other endemic species from the Mascarene archipelago are included in Clade F which is strongly supported only by the BI (BS = 57%, PP = 1). All the species within this Clade F are endemic to La Réunion.

Phylogeographic inferences

The phylogeographic hypotheses inferred are shown on Figs 3 and 4. Madagascar appears to be the ancestral distribution for each clade containing Mascarene species and four colonization events of the Mascarenes can be inferred. A first colonization event from Madagascar to Rodrigues would have provided the lineage of *D. rodriguesiana* (Fig. 4B). A second event of dispersal from Madagascar to Mauritius explains the distribution of Clade C (Fig. 4A). Subsequently, four independent events of dispersal from Mauritius to La Réunion can explain the different geographic distributions for the lineages of *Ruizia cordata*, *Dombeya populnea*, *D. ferruginea* subsp. *borbonica* and *T. granulata*, respectively.

For Clade E, a dispersal event to the Mascarenes from Madagascar can be inferred (Fig. 4C). Clade F includes only the endemic species of La Réunion. This monophyletic group would also have originated from a dispersal event from Madagascar (Fig. 4B). These phylogeographic hypotheses assume that no extinction event has occurred in the Mascarenes.

Evolution of breeding systems

The evolution of breeding systems is shown on Fig. 5. Regardless of the optimization used, hermaphroditism unambiguously appears as the ancestral state and dioecy as the derived state. With Acctran as well as with Deltran, the gender separation was acquired once in Clade F and once for *D. rodriguesiana* (in Clade B). Within Clade C, two interpretations are possible according to the optimization. For the evolutionary scenario using Acctran, dioecy appears once in Clade C, then a reversion to hermaphroditism for the genus *Trochetia* (Fig. 5A). With Deltran, the acquisition of dioecy appears in Clade C₁, and for *D. ferruginea* in Clade C₂ (Fig. 5B).

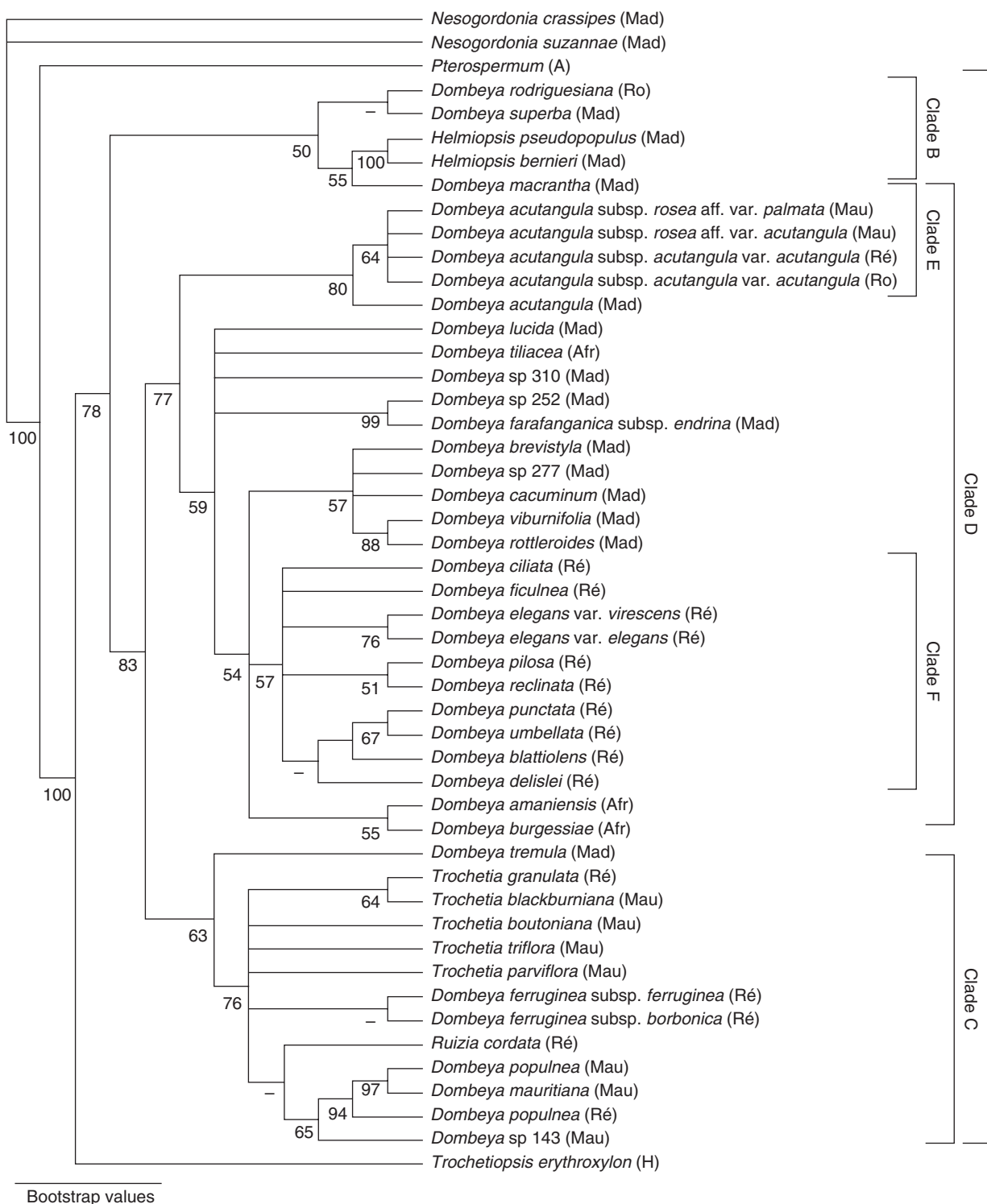


FIG. 1. Strict consensus of the 9402 most-parsimonious trees. Clades A–F correspond to the clades discussed in the text. BS values are indicated below the branches. Dashes indicates nodes that are not supported by the BS analysis (BS < 50%). Mad, Madagascar; Ré, Réunion; Ro, Rodrigues; Mau, Mauritius; As, Asia; Afr, Africa; H, Saint Helena.

DISCUSSION

New insights into the systematics of Dombeyoideae in the Mascarene archipelago

In both types of analysis (MP and BI), all the Mascarene Dombeyoideae are included in Clade A. This monophyletic

group also groups the species of *Dombeya*, *Helmiopsis*, *Trochetia* and *Ruizia*. The presence of all four genera within Clade A implies the paraphyly of the genus *Dombeya*, and supports the results from previous cladistic analyses based on morphological characters (Le Péchon et al., 2009). Despite the existence of morphological homogeneity between the

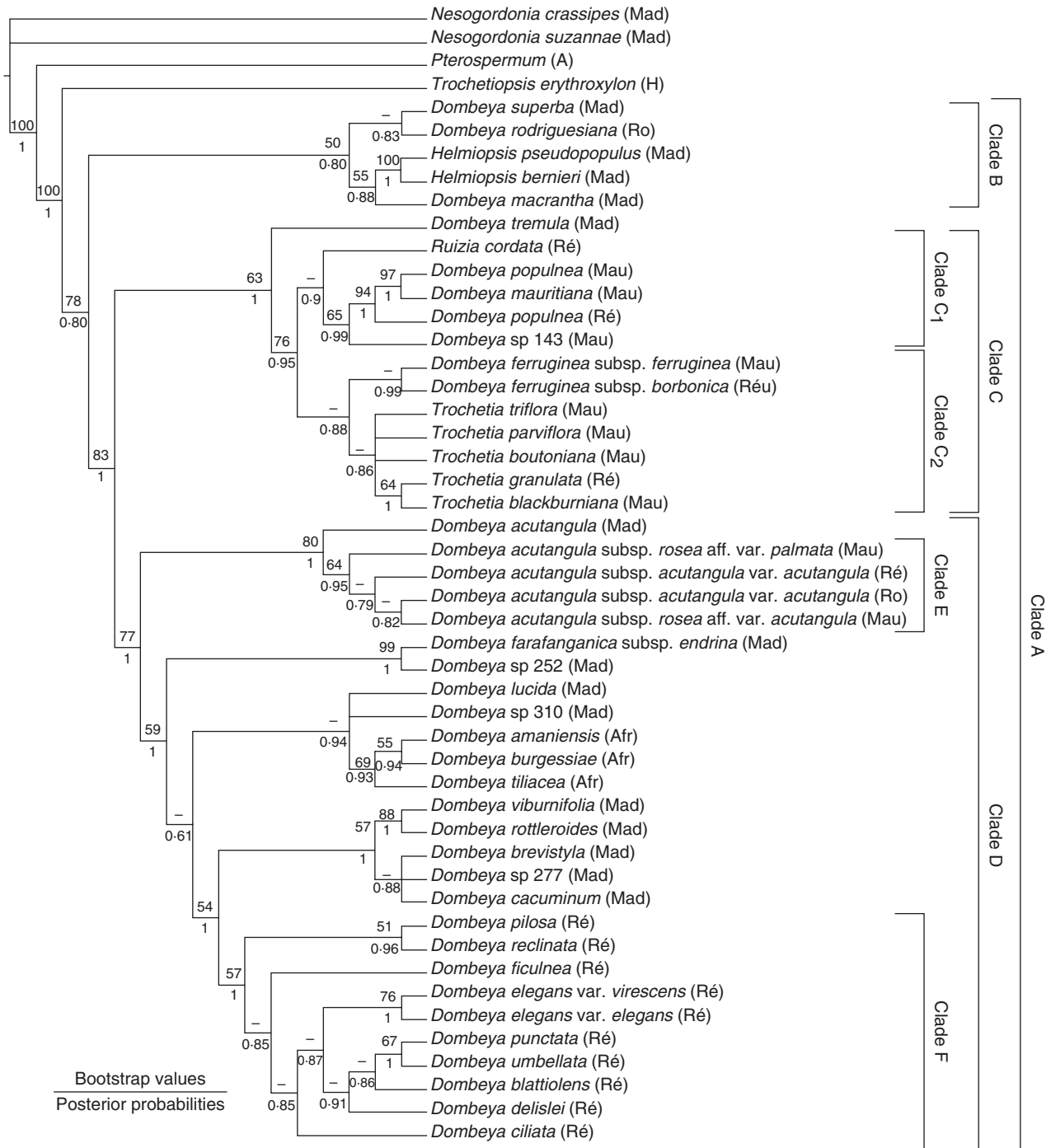
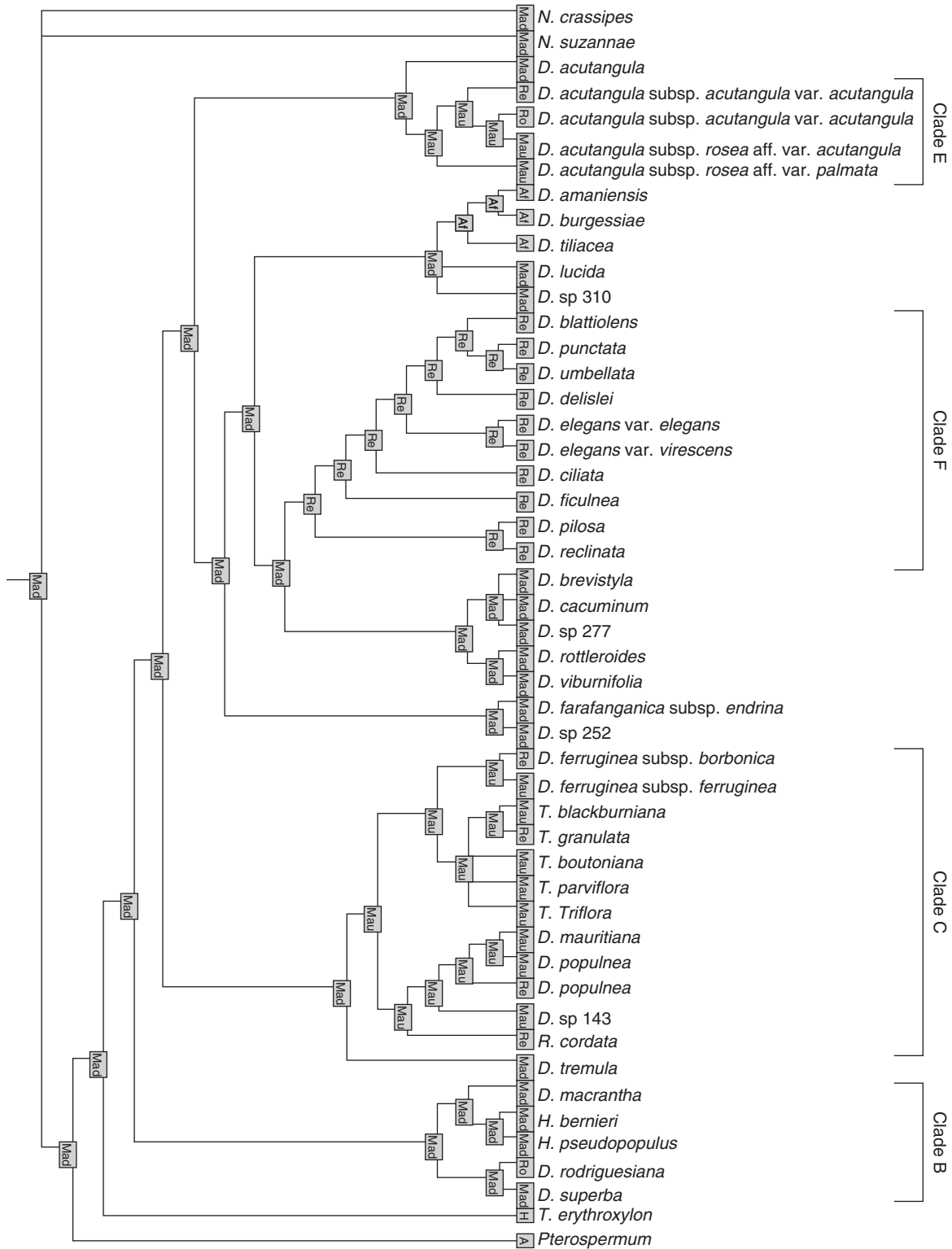


FIG. 2. Bayesian tree with BS (obtained from the MP analysis) and posterior probabilities (PP) values shown above and below the branches respectively. Clades A–F correspond to the clades discussed in the text (see Fig. 1 legend for abbreviations of species geographical distribution).

different taxa, showing their close relationships, it is difficult to find specific characters (i.e. synapomorphies) that define this clade. Within Clade A, the Mascarene Dombeyoideae are polyphyletic and are distributed into four distinct subclades.

Clade B includes the Rodrigues endemic: *D. rodriguesiana*. This species has a biparous pseudo-cyma synflorescence (as defined by Bayer, 1999), as it exhibits reduced staminodes

and short red hairs on all the growing organs. All the other species of this clade are endemic to Madagascar. The general morphology of *D. rodriguesiana* seems radically different from the other species of Clade B, contrasting with *D. macrantha*, *D. superba* and *Helmiopsis bernieri* which are characterized by unifloral synflorescences (as defined by Bayer, 1999) and petaloid staminodes. From a morphological perspective, *D. rodriguesiana* appears closely related to



Downloaded from https://academic.oup.com/iob/article/1/06/2/343/200158 by guest on 23 April 2024

FIG. 3. Biogeographical hypotheses inferred on the Bayesian tree using a parsimony procedure. Abbreviations as in Figs 1 and 2.

another endemic species of Mascarenes, *Dombeya ferruginea* (Le Péchon et al., 2009) from which it has been separated only recently by Friedmann (1981). The molecular phylogenetic position of *D. rodriguesiana* is thus unexpected.

Clade C includes all the species of *Trochetia*, *Ruizia* and four species of *Dombeya*. All these taxa are endemic to the Mascarenes. Except for the presence of *Trochetia*, the relationships among the endemic species of Dombeyoideae from the

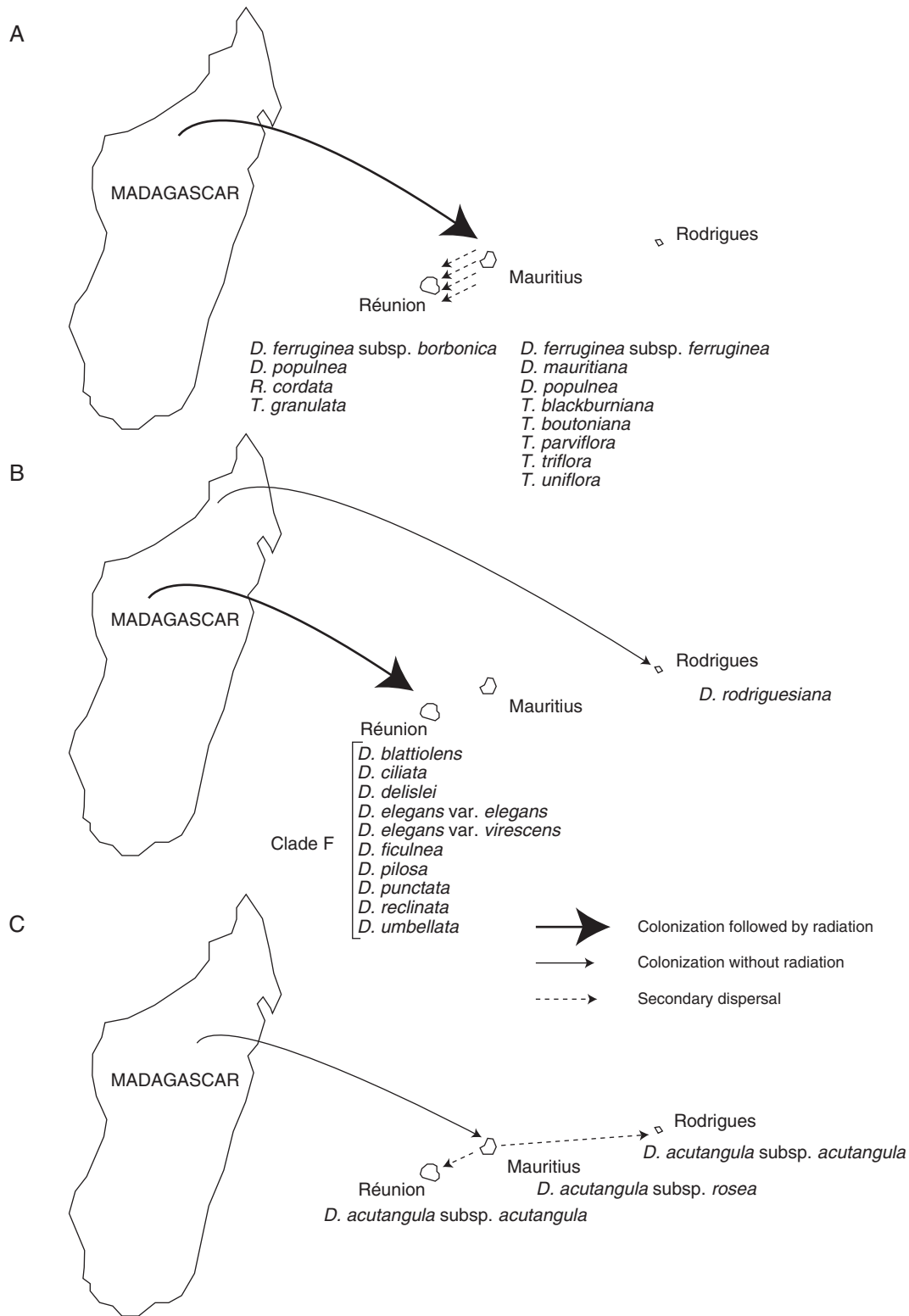


FIG. 4. Colonization hypotheses of the Mascarene archipelago (A) for clade B, (B) for clade F and *D. rodriguesiana* and (C) for clade E.

Mascarenes are consistent with previous morphological analyses (Le Péchon et al., 2009). This monophyletic group is divided into two subclades (Clade C₁ and Clade C₂; see the BI tree in Fig. 2). Clade C₁ includes the genus *Ruizia*,

D. populnea (section *Assonia* Cordem.), *Dombeya mauritiana* and an undescribed Mauritius species (*D. sp.* 143). Using a morphological approach, *D. populnea*, *D. mauritiana* and *Ruizia cordata* can be characterized by their distinctive

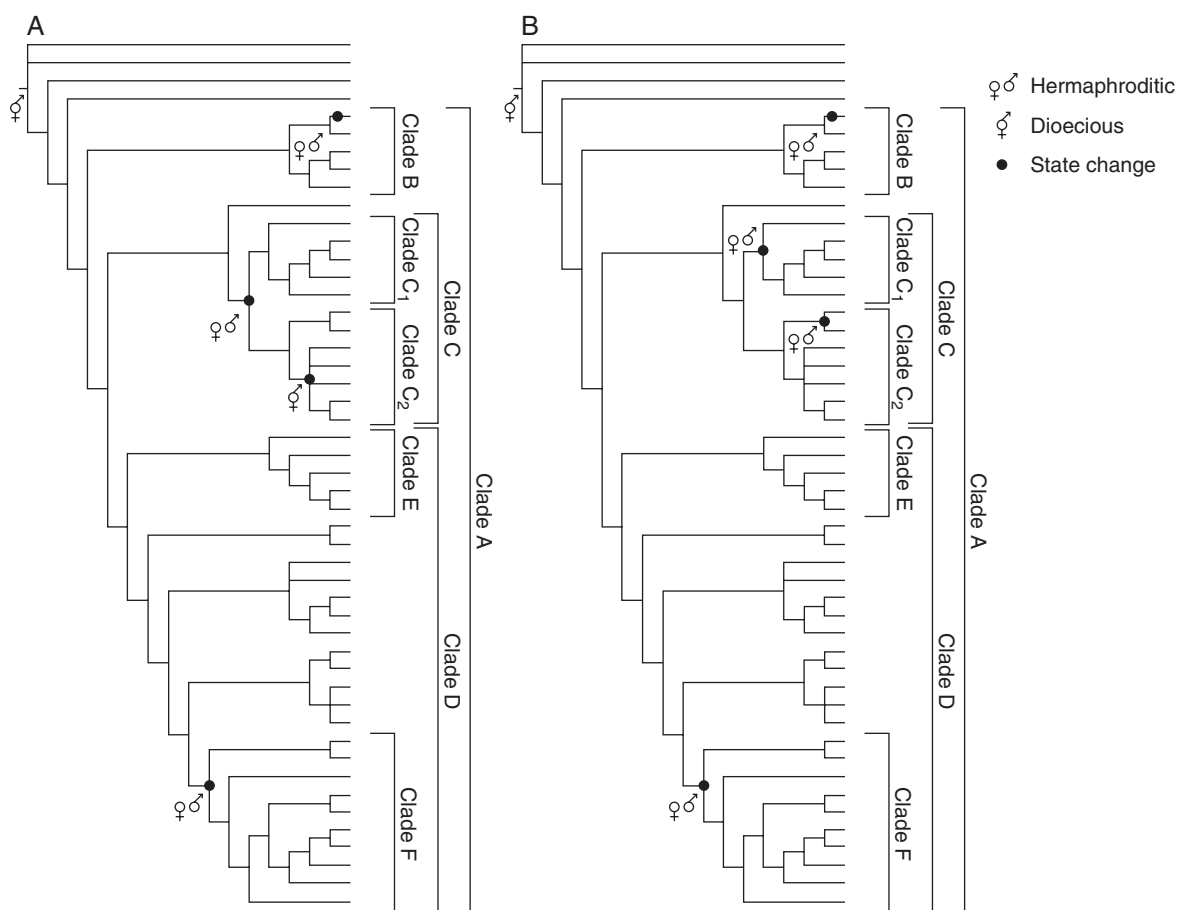


FIG. 5. Evolutionary reconstruction of breeding system pattern inferred on the Bayesian tree using a parsimony procedure: (A) AccTran optimization; (B) DelTran optimization. Clades A–F correspond to the clades discussed in the text.

heterophylly. In addition, the shape and structure of the epicalyx of these two *Dombeya* species are unique among the genus. The floral bracts are fused and the epicalyx comprises a single structure. In *D. populnea*, this structure is persistent and hairless in the specimens from La Réunion, whereas in Mauritius it is deciduous with stellate hairs. In the present phylogenetic analysis, *D. populnea* appears paraphyletic. The Mauritius specimen is more closely related to *D. mauritiana* than the specimen of *D. populnea* from La Réunion. The taxonomic delimitations between *D. mauritiana* and *D. populnea* have not been clearly established (Friedmann, 1987).

The phylogenetic relationships within Clade C₂ confirm the monophyly of *Trochetia* (Le Péchon et al., 2009). Morphologically, *Trochetia* is homogenous and defined by the presence of a spatheform epicalyx, which encloses completely the young floral bud. In addition, the synflorescences (as defined by Bayer, 1999) of *Trochetia* are organized in uni- or tri-floral pseudo-cymes, which are unique in the Mascarenes. *Dombeya ferruginea*, the molecular sister-group of *Trochetia*, is characterized by the presence of biparous pseudo-cymes; an epicalyx composed of three subulate floral bracts; a reduced flower diameter; and sometimes the corolla is absent. Based on morphological characters, *Trochetia* might be considered more closely related to the Malagasy genus *Helmiopsis* (Le Péchon et al., 2009). These two genera share ovoid fruit,

deciduous petals and scaly hairs. However, the latter synapomorphic character is distributed erratically among the Dombeyoideae and is present in several other genera (*Dombeya* Arènes, *Harmsia* K. Schum., *Cheirolaena* Benth. and *Paramelhanina* Arènes; Jenny et al., 1999). This suggests a potentially high level of homoplasy, especially for the highly plastic indument. *Trochetia* and *D. ferruginea* display strongly contrasting characters. Therefore, significant structural modifications could be rapidly acquired (especially in insular context; Baldwin et al., 1991; Emerson, 2002). Further studies using both morphological and molecular data are needed to precisely determine the taxonomic position of *Trochetia* in Dombeyoideae.

Clade E includes all the endemic subspecies and varieties of *D. acutangula* from the Mascarenes. This species is the most widely distributed of the genus *Dombeya* [i.e. Continental Africa (Seyani, 1991), Madagascar (Hochreutiner, 1926; Arènes, 1959b) and Mascarenes (Arènes, 1959a; Friedmann, 1987; Seyani, 1991)] and presents such tremendous morphological variations among populations that Arènes (1959a, b) and Hochreutiner (1926) distinguished four different species on the basis of leaf shape (i.e. entire to deeply incised leaves) and on the length of the floral peduncle. All these species were placed in synonymy by Seyani (1991). In the Mascarenes, Friedmann (1987) separated two subspecies

according to petal coloration. In spite of this high intra-specific variability, the subspecies of *D. acutangula* appear monophyletic. Additionally, *D. acutangula* possesses biparous pseudo-cymes (*sensu* Bayer, 1999) composed of two divergent cincinni. This type of synflorescence is unique within *Dombeya* (Arènes, 1959a, b; Friedmann, 1987; Seyani, 1991).

Clade F is exclusively composed of *Dombeya* from La Réunion. This monophyletic group is also morphologically variable. Only the structure of the synflorescence (an umbellate pseudo-cyme with a single axis of flowering; Bayer, 1999) remains constant. However, this synflorescence architecture is not unique to species from La Réunion because *Dombeya brevistyla* (a species endemic to Madagascar) also exhibits this character. From an ecological point of view, the species of Clade F diversified in habitats at elevations from 800 to 2000 m. These types of ecosystems, which occupy a large area in La Réunion, are absent from Mauritius and Rodrigues islands. The heterogeneity associated with the topographic complexity of the island (i.e. presence of a windward zone and a leeward zone; Strasberg et al., 2005) engendered the creation of multiple environmental conditions, and led to highly diverse ecosystems. The species of Clade F are mainly distributed in the mountain windward rainforests (e.g. *Dombeya pilosa*, *D. ficulnea*, *D. elegans*) but can also be found in the mountain leeward rainforests (e.g. *Dombeya delislei*, *D. punctata*). This diversification, both ecological and taxonomic, probably illustrates an important adaptive radiation event.

Hypotheses of colonization of the Mascarene archipelago

The Mascarenes is a relatively young oceanic archipelago (<10 My; Giorgi and Borchiellini, 1998; Hanyu et al., 2001; Sheth et al., 2003) resulting from a volcanic 'hotspot'. This means that the three islands have never been in contact with any other emerged land. The initial establishment of the fauna and flora could only be explained by long-distance dispersal and colonization events. The number of these events can be assessed from the phylogenetic relationships. Dispersal from Madagascar to Rodrigues Island can be inferred to explain the geographic distribution of *D. rodriguesiana* (Clade B). The phylogenetic history of Clade C is more complex. A single dispersal event from Madagascar to Mauritius can be inferred, followed by four dispersals of Mauritius to La Réunion (for *Ruizia cordata*, *Dombeya populnea*, *D. ferruginea* subsp. *borbonica* and *Trochetia granulata*). The majority of species are restricted to the dry lowland forests of Mauritius and La Réunion (e.g. *Ruizia cordata*, *Dombeya populnea*, *D. mauritiana*, *Trochetia boutoniana*, *T. uniflora*, *T. parviflora*). Although *Trochetia triflora*, *T. blackburniana*, *D. sp. 143* and *D. ferruginea* subsp. *ferruginea* are located in moister environments of Mauritius. *Trochetia granulata* and *D. ferruginea* subsp. *borbonica* are distributed in leeward mountain rainforests, which are present in La Réunion but absent in the other two islands.

For Clade E, an initial colonization of Mauritius by *D. acutangula* can be inferred, followed by two separate dispersal events to La Réunion and Rodrigues.

Clades C and E include taxa distributed on the three islands of the Mascarenes. The ancestral distribution of these two

clades is likely to be Mauritius Island. This phylogenetic inference is congruent with the geological data and the age of the islands.

According to the phylogeographic inference, a unique event of dispersal from Madagascar to La Réunion, followed by a high diversification can explain the diversity within Clade F. The species of this monophyletic group are mainly located in mountain windward rainforest of La Réunion.

Crisp et al. (2009) demonstrated that the distribution of many angiosperm groups in the southern hemisphere is to a large extent conserved phylogenetically and ecologically according to the theory of 'phylogenetic niche conservatism' (Wiens and Graham, 2005). This theory suggests that taxa evolving from a common ancestor are likely to diversify in the same habitat as their ancestor. Because the Malagasy sister-group of Clade F also involves taxa found in mountainous ecosystems, the diversification of the La Réunion species can illustrate such an example of niche conservatism. However, this hypothesis should be tested by further study of Malagasy taxa in order to confirm the observed relationship and the ecology of the retrieved sister-group of Clade F.

In the present study, the three Mascarene clades (Clade C, Clade E and Clade F) and *D. rodriguesiana* (in Clade B), each have a Malagasy origin. This phylogeographic pattern supports the close relationships between the Malagasy and the Mascarene archipelago hypothesized by Cadet (1980). In addition, numerous phylogenetic studies of other organisms obtained similar results. For example, studies within *Polyscias* (Araliaceae; Plunkett et al., 2004), angraecoid orchids (Orchidaceae; Micheneau et al., 2008), geckos (*Phelsuma*; Austin et al., 2004), tree ferns (*Cyathea*; Janssen et al., 2008), *Gaetnera* (Rubiaceae; Malcomber, 2002) demonstrated that all the Mascarene species probably have a Malagasy origin.

Evolution of breeding systems

The majority of *Dombeyoideae* species have been described to bear only hermaphroditic flowers (Bayer and Kubitzki, 2003). Certain genera, such as *Burretiodendron* Rehder and *Excentodendron* Hung T.Chang and R.H.Miao (Bayer and Kubitzki, 2003; Tang et al., 2006, 2007a, b), along with *Astiria*, *Ruizia* and *Dombeya* (Jacob de Cordemoy, 1895; Friedmann, 1987; Humeau, 1999; Humeau et al., 1999a, b; 2003; Bayer and Kubitzki, 2003), involve species described as monoecious or dioecious. Amongst *Dombeya*, only the endemic species from the Mascarenes are considered dioecious (Jacob de Cordemoy, 1895; Friedmann, 1987; Humeau, 1999; Humeau et al., 1999a, b, 2003). This dioecy is cited as cryptic because the flowers appear morphologically bisexual; however, only one of the two sexes is functional (Humeau, 1999; Humeau et al., 1999a, b). In some species (e.g. *Dombeya delislei*, *D. ciliata*, *D. pilosa*), functionally male specimens may carry fruits with a small proportion of fertile seeds. This phenomenon has been referred to 'leaky dioecy' (Humeau et al., 1999a, b). The fact that the flowers are morphologically bisexual implies that this dioecy is derived compared with hermaphroditism (Tang et al., 2006). The evolution of breeding systems inferred from the cladograms (Fig. 3) supports this hypothesis in accordance with Le Péchon et al.

(2009). Regardless of the evolutionary scenario, dioecy appears as homoplastic. In the present study, at least three acquisitions of this breeding system were distinguished in the Mascarenes. Dioecy is a functional feature that can be expressed in different ways. Thus, a number of different processes might explain the separation of sexes from hermaphroditic flowers. For instance, a male flower can be obtained by the reduction (or loss) of the stigmata (*D. ciliata*, *D. pilosa*), by loss (or sterility) of the ovules (*D. ficulnea*), or a combination of the two processes (Humeau, 1999). The same type of differential expression can be observed in female flowers in the absence of pollen (*D. ferruginea*; Friedmann, 1987) or in the presence of sterile pollen (*D. punctata*, *D. elegans*; Humeau, 1999). Therefore dioecy appears as a combination of several characters. The most recent studies of the genus *Dombeya* focused on the expression of dioecy within Clade F (Humeau, 1999; Humeau et al., 1999a, b, 2003). However, despite the taxonomic descriptions of Friedmann (1987) and Jacob de Cordemoy (1895), very little information is currently available on the expression of dioecy within Clade C. The presence of dioecy within each of the endemic Mascarene clades can have emerged via a particular combination of characters.

The independent acquisition of sex separation raises several questions about the origin of the dioecy, particularly in the insular context. The frequency of dioecious taxa in island ecosystems is higher than on continents (Bawa, 1980). The Dombeyoideae from the Mascarene archipelago illustrate this phenomenon well. Several hypotheses might explain the acquisition of dioecy. The classic argument is that this breeding system is a mechanism to avoid selfing (Sakai et al., 1997), thus sex separation could be an adaptive advantage on islands. Indeed, genetic diversity is limited due to long-distance dispersal and loss of contact with the conspecific continental population. The lack of genetic diversity could represent a selective pressure to evolve a breeding system such as dioecy that implies obligatory outcrossing. Other hypotheses (i.e. resource allocation, ecological factors and fruit predation) have also been developed to explain the sex separation (Bawa, 1980; Thomson and Burnet, 1990). However, studies of the Hawaiian flora showed that the high frequency of dioecious taxa is the consequence of a number of dimorphic colonists as well as autochthonous evolution of sex separation (Sakai et al., 1995a, b). Arènes (1959b) described all Malagasy taxa as hermaphroditic but he noticed that some specimens may bear unisexual flowers amongst bisexual flowers. It is worth noting that Arènes (1959a, b) worked exclusively on herbarium specimens, and the cryptic dioecy makes the identification of dioecious species in dried specimens particularly difficult (Le Péchon et al., 2009). Thus, each endemic dioecious clade from the Mascarenes might have a Malagasy dioecious group as sister-group. To conclude on the origin of dioecy in the Mascarene Dombeyoideae (i.e. autochthonous versus phylogenetic origin), further ecological and taxonomic studies are necessary, especially for the Malagasy taxa.

Regardless the origin of dioecy, this breeding system seems to be favoured in an insular environment. The present study suggests that a functional dioecy has differentiated for each endemic clade. For instance, *D. acutangula*, the only hermaphroditic and native species of the archipelago, presents a highly

functional and morphological variability among individuals within and among populations. Some produce more pollen, larger flowers and fewer fruits than others, which produce less pollen, smaller flowers and more fruits (L. D. B. Gigord, University of Lausanne, Switzerland, unpubl. res.), and this could therefore be interpreted as a sexual differentiation toward dioecy. This could be especially selected to avoid autogamy in small and scattered natural populations, which is often the case in an insular context. This hypothesis is especially true for species that possess a late-acting self-incompatibility system such as *D. pilosa*, *D. delislei* and *D. acutangula* (Humeau 1999; Gigord et al., 1998). The expression of this incompatibility system involves a systematic loss of ovules. As a consequence, the reproductive success of such species tends to be extremely low, in particular in small populations. This strong selective pressure might explain the origin of the acquisition or the maintenance of functional dioecy in the Dombeyoideae of the Mascarenes.

Conclusions

This first molecular phylogeny highlights some taxonomic issues within the Dombeyoideae. Indeed, although the sampling was focused on the Mascarenes, the limits and distinctions of the genera *Dombeya*, *Trochetia* and *Ruizia* should be reconsidered. Determining the precise nature of such taxonomic and nomenclatural rearrangements requires stable and well-supported phylogenies. To that end a comprehensive work in phylogenetic systematics including both a more complete sampling of the members of Dombeyoideae and the application of new molecular markers, needs to be conducted in order to validate these results (i.e. the paraphyly of *Dombeya* and the positions of *Ruizia* and *Trochetia*). This first molecular historical framework clarifies the multiple origins of the Mascarene Dombeyoideae. Within the archipelago, this subfamily is divided into four distinct monophyletic groups originating independently by dispersal from Madagascar. The close phylogeographic relationships between the flora of the Mascarenes and Madagascar are confirmed. The phylogenetic pattern reveals two independent adaptive radiations within two main different ecosystems. Despite their independent origins and a distinct evolutionary history, each endemic clade has developed a different breeding system (dioecy) compared with the Malagasy Dombeyoideae which are considered as monoclinal. The present molecular phylogeny suggests at least three distinct acquisitions of this breeding system. This multiple acquisition of dioecy in the Mascarenes appears to be the result of evolutionary convergence, which may be the consequence of selective pressures particular to the insular environments. We think that the Dombeyoideae could be a valuable group for the study of evolutionary processes such as adaptive radiation linked to rapid speciation.

ACKNOWLEDGEMENTS

This work was supported by UMR 7207 “Centre de Recherches sur la Paléobiodiversité et les Paléoenvironnements” (CR2P), IFR 101 CNRS “Institut d’Ecologie, Biodiversité, Evolution, Environnement”, the PPF MNHN ‘Etat et Structure

Phylogénétique de la Biodiversité Actuelle et Fossile’. All the molecular work was supported by the ‘Consortium National de Recherche en Génomique’, and the ‘Service de Systématique Moléculaire’ of the Muséum National d’Histoire Naturelle. This is part of the accord no. 2005/67 between the Genoscope and the Muséum National d’Histoire Naturelle on the project ‘Macrophylogeny of life’ directed by Guillaume Lecointre. We would like to thank the Mauritius National Parks and Conservation Service and the Forestry Service for permission to work in the forests and for general assistance. We are grateful to Annie Tillier for the assistance in the molecular work, to Dr Kurt Liittschwager, Dr Fabienne Audebert-Liittschwager (University Pierre et Marie Curie), Théo Liittschwager, Dr Pascal Antoine Christin (University of Lausanne), Peter B. Phillipson (Missouri Botanical Garden), Isabelle Véa (American Museum of Natural History) and the two anonymous reviewers for the comments on the manuscript, Jean-Bernard Pausé, Dr Christophe Lavergne (CNBM), Edmond and Marie Françoise Grangaud, Jean-Claude Sevathian (Mauritius Wild Life Foundation) for assistance in the field, to the staff of the Paris herbarium and Missouri Botanical Garden for providing dried specimens for this study and to Fanch Le Hir (CBNB) for the collection of material from Conservatoire National Botanique de Brest.

LITERATURE CITED

- Arènes J. 1959a. Les Dombeya des îles des Mascareignes. *Mémoire de l’Institut Scientifique de Madagascar, Série B, Biologie Végétale* 9: 189–216.
- Arènes J. 1959b. 131^e Famille. – Sterculiacées. In: Humbert H. ed. *Flore de Madagascar et des Comores*. Paris: Firmin-Didot et C^{ie}, 1–537.
- Austin JJ, Arnold EN, Jones CG. 2004. Reconstructing an island radiation using ancient and recent DNA: the extinct and living day geckos (*Phelsuma*) of the Mascarene islands. *Molecular Phylogenetics and Evolution* 31: 109–122.
- Baker JB. 1877. *Flora of Mauritius and the Seychelles*. London: L. Reeve & Co.
- Baldwin BG, Kyhos DW, Dvorako J, Carr GD. 1991. Chloroplast DNA evidence for a North American origin of the Hawaiian silversword alliance (Asteraceae). *Proceedings of the National Academy of Sciences of the USA* 88: 1840–1843.
- Barnett LC. 1987. Tribal realignment of certain paleotropical Sterculiaceae. *American Journal of Botany* 74: 724.
- Barriol V. 1994. Phylogénies moléculaires et insertions-délétions de nucléotides. *Compte Rendus de l’Académie des Sciences de Paris, Sciences de la vie, Evolution* 317: 693–701.
- Barriol V. 2004. Les indels, des caractères pas comme les autres? *Biosystema* 22: 35–47.
- Baum DA, Small RL, Wendel JF. 1998. Biogeography and floral evolution of baobabs (*Adansonia*, Bombacaceae) as inferred from multiple data sets. *Systematic Biology* 47: 181–207.
- Bawa KS. 1980. Evolution of dioecy in flowering plants. *Annual Review of Ecology and Systematics* 11: 15–39.
- Bayer C. 1999. The bicolor unit—homology and transformation of an inflorescence structure unique to core Malvales. *Plant Systematics and Evolution* 214: 187–198.
- Bayer C, Kubitzki K. 2003. Malvaceae. In: Kubitzki K. ed. *The families and genera of vascular plants*, Vol. V. Berlin: Springer, 225–314.
- Blanchard F. 2000. *Guide des Milieux Naturels. La Réunion-Maurice-Rodrigues*. Paris: Ulmer.
- Brodie S, Cheek M, Staniforth M. 1998. *Trochetiopsis ebenus*. *Curtis’s Botanical Magazine* 15: 27–36.
- Cadet T. 1980. *La végétation de l’île de la Réunion : étude phytocéologique et phytosociologique*. PhD Thesis, Université Aix-Marseille, France.
- Cavanilles AJ. 1787. *Tertia dissertatio botanica, in Monadelphiae classis dissertationes decem*. Paris: Firmin-Didot et C^{ie}.
- Crisp MD, Arroyo MTK, Cook LG, et al. 2009. Phylogenetic biome conservatism on a global scale. *Nature* 458: 754–756.
- Delarbre C, Escrivá H, Gallut C, et al. 2000. The complete nucleotide sequence of the mitochondrial DNA of the agnathan *Lampetra fluviatilis*: bearings on the phylogeny of cyclostomes. *Molecular Biology and Evolution* 17: 519–529.
- Delarbre C, Gallut C, Barriol V, Janvier P, Gachelin G. 2002. Complete mitochondrial DNA of the hagfish, *Eptatretus burgeri*: the comparative analysis of mitochondrial DNA sequences strongly supports the cyclostome monophyly. *Molecular Phylogenetics and Evolution* 22: 184–192.
- Emerson BC. 2002. Evolution on oceanic islands: molecular phylogenetic approaches to understanding pattern and process. *Molecular Ecology* 11: 951–966.
- Friedmann F. 1981. Sterculiacees et Lecythidaceae nouvelles aux Mascareignes. *Bulletin du Muséum National d’Histoire naturelle, 4e série, Section B, Adansonia* 20: 439–449.
- Friedmann F. 1987. Sterculiacées. In: Bosser J, Cadet T, Guého J, Marais W. eds. *Flore des Mascareignes: La Réunion, Maurice, Rodrigues*, Vol. 53. Port Louis/Paris/London: MSIRI/ORSTOM/Royal Botanical Gardens, Kew, 1–50.
- Gigord LDB, Lavigne C, Shykoff JA. 1998. Partial self-incompatibility and inbreeding depression in a native tree species of La Réunion (Indian Ocean). *Oecologia* 117: 342–352.
- Giorgi L, Borchellini S. 1998. *Carte géologique de l’île Rodrigues au 1:25000. Le schéma hydrogéologique. La notice explicative*. Paris: Ministère Délégué à la Coopération et de la Francophonie and Geolab.
- Hall TA. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series* 41: 95–98.
- Hanyu T, Dunai TJ, Davies GR, Kaneoka I, Nohda S, Uto K. 2001. Noble gas study of the Reunion hotspot: evidence for distinct less-degassed mantle sources. *Earth and Planetary Science Letters* 193: 83–98.
- Hennequin S, Ebihara A, Ito M, Iwatsuki K, Dubuisson J-Y. 2003. Molecular systematics of the fern genus *Hymenophyllum* s.l. (Hymenophyllaceae) based on chloroplast coding and noncoding regions. *Molecular Phylogenetics and Evolution* 27: 283–301.
- Hochreutiner BPG. 1926. Monographie des Dombeya de Madagascar. *Candollea* 3: 5–120.
- Humeau L. 1999. *Ecologie et évolution de la dioécie et du dimorphisme sexuel de la taille des fleurs chez les Dombeya (Sterculiacées) endémiques de la Réunion*. PhD Thesis, Université de La Réunion, France.
- Humeau L, Pailler T, Thompson JD. 1999a. Cryptic dioecy and leaky dioecy in endemic species of *Dombeya* (Sterculiaceae) on La Réunion. *American Journal of Botany* 86: 1437–1447.
- Humeau L, Pailler T, Thompson JD. 1999b. Variation in the breeding system of two sympatric *Dombeya* species on La Réunion island. *Plant Systematics and Evolution* 218: 77–87.
- Humeau L, Pailler T, Thompson JD. 2003. Flower size dimorphism in diclinous plants native to La Réunion Island. *Plant Systematics and Evolution* 240: 163–173.
- Jacob de Cordemoy E. 1895. *Flore de l’île de La Réunion*. Paris: Klincksieck.
- Jassen T, Bystriakova N, Rakotondrainibe F, Coomes D, Labat J-N, Schneider H. 2008. Neoenendism in Madagascan scaly tree ferns results from recent, coincident diversification bursts. *Evolution* 62: 1876–1889.
- Jenny M, Bayer C, Dorr LJ. 1999. *Aethiocarpa* reduced to *Harmsia* (Malvaceae, Dombeyoideae). *Taxon* 48: 3–6.
- Le Péchon T, Cao N, Dubuisson J-Y, Gigord LDB. 2009. Systematics of Dombeyoideae (Malvaceae) in the Mascarene archipelago (Indian Ocean) inferred from morphology. *Taxon* 58: 519–531.
- McDougall I, Upton BGJ, Wadsworth WJ. 1965. A geological reconnaissance of Rodriguez Island, Indian Ocean. *Nature* 206: 26–27.
- Maddison WP, Maddison DR. 2006. *Mesquite: a modular system for evolutionary analysis*, Version 2-6 <http://mesquiteproject.org>.
- Malcomber ST. 2002. Phylogeny of *Gaertnera* Lam. (Rubiaceae) based on multiple DNA markers: evidence of a rapid radiation in a widespread, morphologically diverse genus. *Evolution* 56: 42–57.
- Micheneau C, Carlward BS, Fay MF, Bytbeier B, Pailler T, Chase MW. 2008. Phylogenetics and biogeography of Mascarene angraecoid orchids (Vandaeae, Orchidaceae). *Molecular Phylogenetics and Evolution* 46: 908–922.

- Myers N, Mittermeier RA, Mittermeier CG, da Fonseca GAB, Kent J. 2000. Biodiversity hotspots for conservation priorities. *Nature* **403**: 853–858.
- Nyffeler R, Bayer C, Alverson WS, et al. 2005. Phylogenetic analysis of the Malvaceae clade (Malvaceae s.l.) based on plastid DNA sequences. *Organisms Diversity and Evolution* **5**: 109–123.
- Nylander JAA. 2004. *MrModeltest*. Published by the author, Uppsala University.
- Pfeil BE, Brubaker CL, Craven LA, Crisp MD. 2002. Phylogeny of *Hibiscus* and the tribe Hibisceae (Malvaceae) using chloroplast DNA sequences of ndhF and the rpl16 intron. *Systematic Botany* **27**: 333–350.
- Plunkett GM, Lowry PP, Vu NV. 2004. Phylogenetic relationships among *Polyscias* (Araliaceae) and close relatives from the western Indian Ocean basin. *International Journal of Plant Sciences* **165**: 861–873.
- Raymunde MB, Xena de Enrech N, Mathez J, Dubuisson J-Y. 2002. Coding of insertion-deletion events of the chloroplastic intergene atpB-rbcL for the phylogeny of the Valerianeae tribe (Valerianaceae). *Comptes Rendus Biologies* **325**: 131–139.
- Ronquist F, Huelsenbeck JP. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* **19**: 1572–1574.
- Rouhan G, Dubuisson J-Y, Rakotondrainibe F, et al. 2004. Molecular phylogeny of the fern genus *Elaphoglossum* (Elaphoglossaceae) based on chloroplast non-coding DNA sequences: contributions of species from the Indian Ocean area. *Molecular Phylogenetics and Evolution* **33**: 745–763.
- Sakai AK, Wagner WL, Ferguson DM, Herbst DR. 1995a. Origins of dioecy in the Hawaiian flora. *Ecology* **76**: 2517–2529.
- Sakai AK, Wagner WL, Ferguson DM, Herbst DR. 1995b. Biogeographical and ecological correlates of dioecy in the Hawaiian flora. *Ecology* **76**: 2530–2543.
- Sakai AK, Weller SG, Wagner WL, Soltis PS, Soltis DE. 1997. Phylogenetic perspectives on the evolution of dioecy: adaptive radiation in the endemic Hawaiian genera *Schiedea* and *Alsinidendron* (Caryophyllaceae: Alsinoideae). In: Givnish TJ, Sytsma KJ. eds. *Molecular evolution and adaptive radiation*. New York, NY: Cambridge University Press, 455–473.
- Seung-Chul K, Crawford DJ, Francisco-Ortega J, Santos-Guerra A. 1996. A common origin for woody *Sonchus* and five related genera in the Macaronesian islands: molecular evidence for extensive radiation. *Proceedings of the National Academy of Sciences of the USA* **93**: 7743–7748.
- Seyani JH. 1991. *The genus Dombeya (Sterculiaceae) in continental Africa*. Meise: National Botanic Garden of Belgium.
- Shaw J, Lickey EB, Beck JT, et al. 2005. The tortoise and the hare. II. Relative utility of 21 noncoding chloroplast DNA sequences for phylogenetic analysis. *American Journal of Botany* **92**: 142–166.
- Shaw J, Lickey EB, Schilling EE, Small RL. 2007. Comparison of whole chloroplast genome sequences to choose noncoding regions for phylogenetic studies in Angiosperms: the tortoise and the hare III. *American Journal of Botany* **94**: 275–288.
- Sheth HC, Mahoney JJ, Baxter AN. 2003. Geochemistry of lavas from Mauritius, Indian Ocean: mantle sources and petrogenesis. *International Geology Review* **45**: 780–797.
- Strasberg D, Rouget M, Richardson DM, Baret S, Dupont J, Cowling RM. 2005. An assessment of habitat diversity and transformation on La Réunion Island (Mascarene Islands, Indian Ocean) as a basis for identifying broad-scale conservation priorities. *Biodiversity and Conservation* **14**: 3015–3032.
- Swofford DL. 2001. *PAUP*: phylogenetic analysis using parsimony (* and other methods), version 4.0b10*. Sunderland, MA: Sinauer Associates.
- Tang Y. 1992. On the affinities of *Pterospermum* Schreb. (Sterculiaceae). *Guihaia* **12**: 8–14.
- Tang Y, Gao H, Wang C-M, Chen J-Z. 2006. Microsporogenesis and microgametogenesis of *Excentrodendron hsiennu* (Malvaceae s.l.) and their systematic implications. *Botanical Journal of the Linnean Society* **150**: 447–457.
- Tang Y, Gilbert MG, Dorr LJ. 2007a. *Burretiodendron*. In: Wu ZY, Raven PH, Hong DY. eds. *Flora of China*, Vol. 12. Beijing/St Louis, MO: Science Press/Missouri Botanical Garden Press, 262–263.
- Tang Y, Gilbert MG, Dorr LJ. 2007b. *Excentrodendron*. 12. In: Wu ZY, Raven PH, Hong DY. eds. *Flora of China*, Vol. Beijing/St Louis, MO: Science Press/Missouri Botanical Garden Press, 261–262.
- Thomson JD, Burnet J. 1990. Hypotheses for the evolution of dioecy in seed plants. *Trends in Ecology and Evolution* **1**: 11–16.
- White TJ, Birns T, Lee S, Taylor J. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis M, Gelfand D, Sninsky J, White T. eds. *PCR protocols: a guide to methods and applications*. San Diego, CA: Academic Press, 315–322.
- Wiens JJ, Graham CH. 2005. Niche conservatism: integrating evolution, ecology, and conservation biology. *Annual Review of Ecology, Evolution and Systematics* **36**: 519–539.