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Endophytic *Pestalotiopsis* species associated with plants of Palmae, Rhizophoraceae, Planchonellae and Podocarpaceae in Hainan, China

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A survey of the endophytic *Pestalotiopsis* associated with 27 plant species belonging to four families in Hainan Province was carried out from 2007 to 2008. Colonization frequencies of endophytic *Pestalotiopsis* species varied in the host plant's tissues, sites and natural environmental conditions. Species composition of endophytic *Pestalotiopsis* varied in different families of plants. A total of 43 endophytic *Pestalotiopsis* species were isolated, of which 23, 11, 9 and 8 species were obtained from families of Palmae, Rhizophoraceae, Podocarpaceae and Planchonellae, respectively. The species of *Pestalotiopsis* isolated from different hosts in Palmae family varied from 1 to 7. The colonization frequencies of endophytic *Pestalotiopsis* in dry years were lower than that in usual years. Shannon-Wiener index of endophytic *Pestalotiopsis* in Palmae, Rhizophoraceae and Planchonellae changed from 1.4775 to 2.5013. Evenness index changed from 0.3624 to 1.0431. Richness index of endophytic *Pestalotiopsis* had no correlation with Shannon-Wiener index and evenness index. The coefficient of community of endophytic *Pestalotiopsis* among the four plant families was less than 0.5, showing low similarity.

Key words: Endophytic *Pestalotiopsis*, colonization frequencies, diversity, host preference, richness index.

INTRODUCTION

Fungal endophytes have been characterized by their ability to produce apparently harmless infections in living plant tissues (Carroll and Carroll, 1978; White and Cole, 1985; Tejesvi et al., 2009). Extensive surveys in a wide variety of plants indicated that endophytes are apparently ubiquitous. But the researches of endophytic fungi were mostly concentrated in the northern hemisphere and temperate zones. Recent study of endophyte in tropical region begun with the investigation of endophyte in *Manilkara bidentata* and mangrove wild legume *canavalia cathartica*, which indicated that endophytic fungi were important community among fungi (Lodge et al., 1996; Anita and Sridhar, 2009). Although, investigations of endophytic fungi in some tropical plants were carried out, only several or even one plant species were involved in each study (Ananda and Sridhar, 2002; Kelemu et al., 2003; Maria and Sridhar, 2003; Gao et al., 2005; Suryanayanan et al., 2005; Rodriguez et al., 2009). Extensive surveys with the plants in the tropical area in China have not been done.

Endophytic *Pestalotiopsis* has been considered as a main part of the endophytic fungi community in nature (Strobel et al., 1996, 1997; Okane et al., 1997, 1998;

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Cannon and Simmons, 2002; Toofanee and Dulymamode, 2002; Kumar and Hyde, 2004; Photita et al., 2004; Wei and Xu, 2004; Rodriguez et al., 2009). Diversity analysis of endophytic *Pestalotiopsis* would join the research community composition and species distribution of endophytic fungi.

The regions of tropical forest were considered as the most abundant area of fungal resource (Bills and Polishhook, 1994; Gilbert and Sousa, 2002). Hainan Island lies in South China Sea, latitude 18°16' and 20°23', where the abundant plant, water, and heat resources in tropic rain forest provide basic condition for the development and multiplication of endophytic *Pestalotiopsis*. The aim of this paper was to demonstrate the species diversity of endophytic *Pestalotiopsis* on plants of Palmae, Rhizophoraceae, Podocarpaceae and Planchonellae in the tropical region, of Hainan Province, China.

MATERIALS AND METHODS

Sample collection

The healthy leaves and twigs were collected from the Xinglong Tropical Botanical Garden, the Danzhou Tropical Botanical Garden, the Jianfengling Natural Forest Reserve and the Dongzaigang Mangrove Reserve in Hainan Province in April 2007 and 2008.

Isolation and identification

The leaves and twigs were separated from their branches and washed with running tap water, surface sterilized with 75% ethanol (60 s), 1.3% NaClO (5 min) and 75% ethanol (30 s) (Wei and Xu, 2004). Samples were washed three times with sterilized water, cut into pieces of 1 cm long and placed on PDA medium. The plant tissues were incubated at 25° C for 3-20 days and were checked regularly. Pure fungal cultures were obtained by single spore isolation following the methods outlined by Lacap et al. (2003) and Liu et al. (2007).

Hyphal tip from the colony margin was transferred into new Petridish with PDA medium. When colony grew up to 2 cm in diameter, the autoclaved segment of the carnation leaf (*Dianthi caryophylii* L.) was added to the culture to promote sporulation (Fisher et al., 1982; Strobel et al., 1996). *Pestalotiopsis* species were identified according to the morphological descriptions of Steyaert (1949), Sutton (1980) and Nag Raj (1993).

The living cultures of *Pestalotiopsis* species were deposited in the Institute of Biotechnology, Zhejiang University, Hangzhou and the China General Microbiological Culture Collection Center in Beijing, China.

Data analysis

Colonization frequency was calculated as the number of plant tissue segments colonized by *Pestalotiopsis* species divided by the total number of segments assessed (Liu et al., 2007). The Shannon-Wiener diversity index (H) was used to estimate the species diversity of the fungal assemblages recovered from a particular type of sample (leaf and twig) and from different sampling sites. The H' was calculated according to the following formula:

$$H' = -\sum_{i=1}^{k} pi \times \ln pi$$

Where, k is the total number of fungal species, and pi is the proportion of individuals that species i contributes to the total (Pielou, 1975).

To evaluate the degree of community similarity of endophytic *Pestalotiopsis* species between two sampling sites, Sorenson's coefficient (C_s) was employed and calculated according to the following formula:

 $C_S = 2j/(a+b),$

Where, *j* is the number of endophytic *Pestalotiopsis* species that coexisted in both sampling sites, *a* is the total number of endophytic *Pestalotiopsis* species in one sampling site, and *b* is the total number of endophytic *Pestalotiopsis* species in the other sampling site (Liu et al., 2007; Tejesvi et al., 2008).

Statistical analysis was made by DPS (Data Processing System) version 7.05 professional edition.

RESULTS

Colonization frequencies of endophytic *Pestalotiopsis* in different host plants

The colonization frequencies of endophytic *Pestalotiopsis* species in seven plants of Palmae, nine mangrove plants, five plants of Planchonellae and six plants of Podocarpaceae are shown in Figure 1.

The colonization frequency of endophytic *Pestalotiopsis* species in the leaves of Palmae plants varied from 0.8 to 27.2% (Figure 1), and the colonization of endophytic *Pestalotiopsis* species in leaves of mangrove plants varied from 3.3 to 40.0% (Figure 1). Five plants of Planchonellae were selected to analyze the colonization frequency of endophytic *Pestalotiopsis* (Figure 1). The colonization frequency of the endophytic *Pestalotiopsis* in twigs of *Manilkara zapota* was significantly higher (23.3%) than those of other plants, but there was no significant difference among the other plants. The colonization frequency of Podocarpaceae plants were 1.7, 15, 16.7, 8.3, 20 and 10% in *Podocarpus fleuryi, Podocarpus imbricatus, Dacrydium pierrei, Podocarpus macrophyllus*, and *Nageia nagi*, respectively (Figure 1).

Colonization frequencies of endophytic *Pestalotiopsis* in different years

The colonization frequencies of endophytic *Pestalotiopsis* species in seven common plants of Palmae were analyzed between 2007 and 2008 (Figure 2). The colonization frequencies of endophytic *Pestalotiopsis* species in *Neodypsis decaryi* were 50%. However, the colonization frequencies in 2008 were only 4.4%. The other plants of Palmae behaved in the same trend in varied degree except in *Hyophorbe lagenicaulis* and *Caryota ochlandra*. No endophytic *Pestalotiopsis* species was isolated from *C. ochlandra*, *Zalacca wallichiana* and

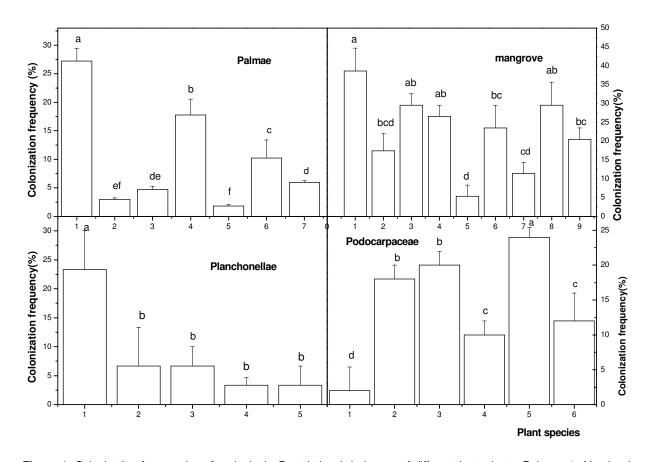


Figure 1. Colonization frequencies of endophytic *Pestalotiopsis* in leaves of different host plants; Palmae: 1. *Neodypsis* decaryi; 2. *Hyophorbe lagenicaulis*; 3. *Wodyetia bifurcate*; 4. *Chamaedorea oblongata*; 5. *Caryota ochlandra*; 6. *Zalacca wallichiana*; 7. *Chrysalidocarpus lutesens*. Mangrove: 1. *Bruguiera gymnorrhiza*; 2. *Kandelia candel*; 3. *Bruguiera sexangula*; 4. *Bruguiera sexangula*; 5. *Ceriops tagal*; 6. *Rhizophora mucronata*; 7. *Hibiscus tiliaceus*; 8. *Aegiceras coniculatum*; 9. *Xylocarpus granatum*. Planchonellae: 1. *Manilkara zapota*; 2. *Madhuca hainanensis*; 3. *Mimusops elengi*; 4. *Synsepalum dulcificum*; 5. *Lucuma nervosa*. Podocarpaceae: 1. *Podocarpus fleuryi*; 2. *Podocarpus imbricatus*; 3. *Dacrydium pierrei*; 4. *Podocarpus macrophyllus*; 5. *Nageia nagi*; 6. *Podocarpus macrophyllus*.

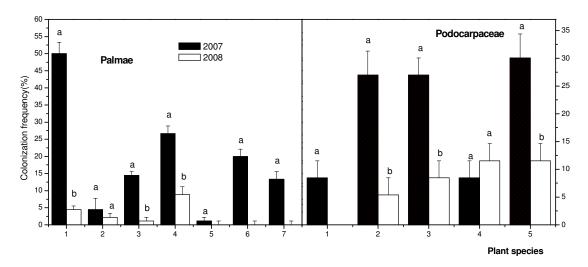


Figure 2. Colonization frequencies of endophytic *Pestalotiopsis* of the leaves in different years. Palmae: 1. *Neodypsis* decaryi; 2. *Hyophorbe lagenicaulis*; 3. *Wodyetia bifurcate*; 4. *Chamaedorea oblongata*; 5. *Caryota ochlandra*; 6. *Zalacca wallichiana*; 7. *Chrysalidocarpus lutesens*. Podocarpaceae: 1. *Podocarpus fleuryi*; 2. *Podocarpus imbricatus*; 3. *Dacrydium pierrei*; 4. *Podocarpus macrophyllus*; 5. *Nageia nagi*.

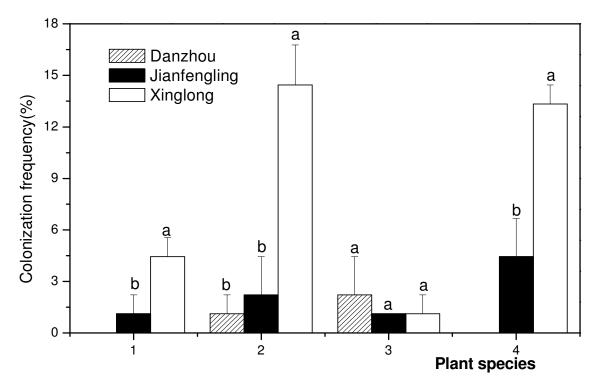


Figure 3. Colonization frequencies of endophytic *Pestalotiopsis* in the leaves of Palmae in three locations. 1. *Hyophorbe lagenicaulis*; 2. *Wodyetia bifurcate*; 3. *Caryota ochlandra*; 4. *Chrysalidocarpus lutesens*.

Chrysalidocarpus lutesens in 2008. The analysis of the colonization frequencies of endophytic *Pestalotiopsis* in five plants of Podocarpaceae between 2007 and 2008 revealed that the colonization frequencies of endophytic *Pestalotiopsis* in 2007 were notably higher than that in 2008, whereas no marked difference in colonization frequencies of *P. macrophyllus* was observed between 2007 and 2008 (Figure 2).

Colonization frequencies of endophytic *Pestalotiopsis* in different locations

Four plants of Palmae were selected to study the influence of sampling site on colonization frequencies of endophytic Pestalotiopsis in Danzhou, Jianfengling and Xinglong (Figure 3). For colonization frequencies on different sites, no significant difference for C. ochlandra were found, however the colonization frequencies of C. lutesens showed distinctly difference in the three locations. Moreover, the colonization frequencies in H. lagenicaulis, Wodyetia bifurcata and C. lutesens in Xinglong were higher than those for the other two positions. It was obviously interesting that the colonization frequencies of endophytic Pestalotiopsis of some plants showed significant difference in different sites, but some plants showed no distinct difference. It indicated that the colonization frequencies of endophytic *Pestalotiopsis* varied as sampling positions and plant species varied.

Colonization frequencies of endophytic *Pestalotiopsis* in different tissue

Nine mangrove plants and five Planchonellae plants were selected to study the influence of different tissues on colonization frequencies of endophytic *Pestalotiopsis* (Figure 4).

The colonization frequencies of endophytic *Pestalotiopsis* species in twigs were significant higher than that in leaves (P<0.05, Figure 4), especially, when there was no endophytic *Pestalotiopsis* isolated from the leaves in some host plants (*Kandelia candel, Bruguiera sexangula, Rhizophora mucronata, Aegiceras coniculatum, Madhuca hainanensis, Mimusops elengi, Synsepalum dulcificum* and *Lucuma nervosa*).

Species composition of endophytic *Pestalotiopsis* species in different host plants

Species composition of endophytic *Pestalotiopsis* varied in different families of plants (Table 1). Among 51 endophytic *Pestalotiopsis* species, 23, 11, 9 and 8 species were obtained from the families of Palmae,

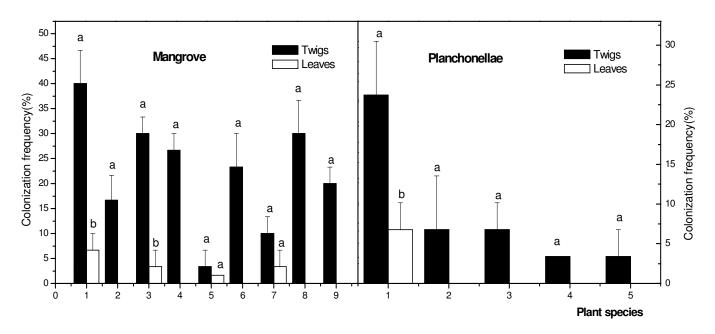


Figure 4. Colonization frequencies of endophytic *Pestalotiopsis* in different organization. Mangrove: 1. *Bruguiera gymnorrhiza*; 2. *Kandelia candel*; 3. *Bruguiera sexangula*; 4. *Bruguiera sexangula*; 5. *Ceriops tagal*; 6. *Rhizophora mucronata*; 7. *Hibiscus tiliaceus*; 8. *Aegiceras coniculatum*; 9. *Xylocarpus granatum*. Planchonellae: 1. *Manilkara zapota*; 2. *Madhuca hainanensis*; 3. *Mimusops elengi*; 4. *Synsepalum dulcificum*; 5. *Lucuma nervosa*.

Rhizophoraceae, Podocarpaceae and Planchonellae, respectively. Species composition of endophytic Pestalotiopsis also displayed great differences in different plants of same family. Among 16 palmaceous plants, 8, 7, 5 and 5 species were isolated from Roystonea regia, Z. wallichiana, N. decaryi and C. lutesens, respectively. However, for the other 12 plants, only 1 to 3 species were obtained from each plant. The average of the species number of endophytic Pestalotiopsis in palmaceous plants was 2.7. The endophytic Pestalotiopsis species number varied from 1 to 8, 1 to 3 and 1 to 5 in different mangrove plants, Podocarpaceae and Planchonellae, with an average of 3.0, 2.2 and 2.6 species per host, respectively.

A total of 180, 52, 59 and 38 *Pestalotiopsis* isolates were obtained from plants of Palmae, Rhizophoraceae, Podocarpaceae and Planchonellae, respectively. The most common species of endophytic *Pestalotiopsis* in plants of Hainan Province were *P. adusta*, *P. clavispora*, *P. paeoniae*, *P. virgatula* and *P. zonata* (Table 2). The most common *Pestalotiopsis* species were similar between Palmae (*P. clavispora*, *P. virgatula* and *P. zonata*) and Podocarpaceae (*P. virgatula* and *P. zonata*) and the most common *Pestalotiopsis* species in plants of Rhizophoraceae and Planchonellae were *P. adusta* and *P. paeoniae*, respectively.

Shannon-Wiener diversity index of endophytic *Pestalotiopsis* species in plants of both Palmae (2.4641) and Rhizophoraceae (2.5013) were much higher than that in Podocarpaceae (1.4775) and in Planchonellae (1.8707) (Table 2). Although, 52 strains were isolated

from Rhizophoraceae, no more than three strains were isolated from Palmae. The Evenness index and Shannon-Wiener index of endophytic *Pestalotiopsis* were very high among the four families.

DISCUSSION

Colonization frequencies of endophytic Pestalotiopsis

Okane et al. (1998) found that endophytic *Pestalotiopsis* species were colonized in seven species of Ericaceae with different colonization frequencies (0.7 to 17.1%) in Kyoto and Japan. Cannon and Simmons (2002) reported the diversity and host preference of leaf endophytic fungi in the Iwokrama Forest Reserve, Guyana. In contrast to studies in temperate ecosystems, no distinct fungal communities were identified for individual plant species, suggesting that the degree of host preference was low. The results of this study showed that colonization frequencies of endophytic *Pestalotiopsis* in Hainan varied with different host plants, but the degrees of host preference were different in different plant families.

The colonization frequencies of endophytic *Pestalotiopsis* species varied with different host tissues in this study. Taylor et al. (1999) investigated the endophytes in *Arenga tremula* from four locations. Quantitative and qualitative differences in endophyte assemblages from old and young tissues were observed, and more isolates were recovered from old tissues

Table 1. Host plants and their endophytic *Pestalotiopsis* species in Hainan, China.

Host plants	Pestalotiopsis species		
Palmae			
Chamaedorea elegans Mart. Liebm	P. clavispora		
Areca catechu L.r	P. photiniae		
<i>Roystonea regia</i> (H.B.K) Cook.	P. briosiana, P. elastica, P. fuchsiae, P. photiniae, P. palmarum, P. pandani, P. virgatula, P. zonata		
Cyrtostachys renda Bl.	P. cinchonae		
Wodyetia bifurcata A.K.Jrvine	P. clavispora, P. menezesiana		
Chamaedorea oblongata Mart.	P. briosiana, P. virgatula, P. zonata		
Hyophorbe lagenicaulis Mart.	P. bicolor, P. clavispora		
Chamaedorea metallica O.F.Cook ex H.E.Moore	P. clavispora		
Livistona chinensis(Jacq.)R.Br.	P. foedans		
Neodypsis decaryi Jum	P. clavispora, P. gracillis, P. lambertiae, P. pauciseta, P. zonata		
Chrysalidocarpus lutesens H. Wendl.	P. adusta, P. clavispora, P. gracillis, P. virgatula, P. zahlbrucknerian		
Arenga engleri Beccari	P. clavispora		
Zalacca wallichiana Salacca.	P. adusta, P. algeriensis, P. briosiana, P. fici, P. palmarum, P. virgatula, P. zonata		
Cocos nucifera L.	P. cinchonae		
Elaeis guineensis Jacq.	P. heterocornis		
Caryota ochlandra Hance	P. coffeae, P. peycnonelii, P. Sorbi		
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Mangrove plants			
Bruguiera sexangula (Lour.) Poir.	P. adusta, P. clavispora, P. foedans, P. Pauciseta		
Lagerstroe miaindica L.	P. clavispora		
Rhizophora mucronata Lam.	P. adusta P. Clavispora		
Hibiscus tiliaceus L.	P. adusta, P. Clavispora		
<i>Bruguiera sexangula</i> Lour	P. adusta, P. heterocornis, P. Clavispora		
Ceriops tagal (Perr.)C.B.Rob.	P. adusta		
<i>Xylocarpus granatum</i> Koenig	P. adusta		
<i>Bruguiera gymnorrhiza</i> (L.) Poir.	P. adusta, P. clavispora, P. gracillis, P. heterocornis, P. neglecta, P. virgatula		
Kandelia candel L.	P. adusta, P. cinchonae, P. gracillis, P. paeoniae, P. pauciseta, P. photiniae, P. vaccinii, P. virgatula		
Aegiceras coniculatum (L.) Blanco	P. foedans, P. Virgatula		
Podocarpaceae			
Podocarpus fleuryi Hickel	P. virgatula		
Podocarpus macrophyllus var. maki	P. photiniae, P. virgatula		
Podocarpus imbricatus BI.	P. clavispora, P. versicolor		
Dacrydium pierrei Hichel	P. alöes, P. vismiae, P. Zonata		
Podocarpus macrophyllus (Thunb.) D. Don.	P. cinchonae, P. Hainanensis		
<i>Nageia nagi</i> (Thunb.) O. Ktze.	P. clavispora, P. virgatula, P. Zonata		
Planchonellae			
Lucuma nervosa A.DC.	P. alöes, P. clavispora, P. theae, P. virgatula, P. Zonata		
<i>Madhuca hainanensis</i> Chun et How	P. paeoniae		
<i>Manilkara zapota</i> (Linn.) Van Royen	P. leucothoes, P. paeoniae, P. subcuticulari, P. Zonata		
Synsepalum dulcificum Denill	P. alöes, P. Theae		
<i>Mimusops elengi</i> Linn	P. zonata		

Table 2. The composition of the endophytic Pestalotiopsis from the four families of the host plants.

Species	Number of the isolates/ Relative abundance (%)				
	Palmae	Rhizophoraceae	Podocarpaceae	Planchonellae	
P. adusta	5/2.78	20/38.46	_	-	
P. algeriensis	1/0.56	-	-	-	
P. alões	-	-	3/5.08	2/5.26	
P. bicolor	2/1.11	-	-	-	
P. briosiana	10/5.56	-	-	-	
P. cinchonae	3/1.67	1/1.95	46.78	-	
P. clavispora	33/18.33	9/17.31	14/16.95	7/18.42	
P. coffeae	2/1.11	-	-	-	
P. elastica	1/0.56	-	-	-	
P. fici	2/1.11	-	-	-	
P. foedans	2/1.11	2/3.85	-	-	
P. fuchsiae	2/1.11	-	-	-	
P. gracillis	21/11.67	5/9.62	-	-	
P. hainanensis	-	-	1/1.69	-	
P. heterocornis	3/1.67	3/5.77	-	-	
P. lambertiae	1/0.56	-	-	-	
P. leucothoes	-	-	-	3/7.89	
P. menezesiana	16/8.89	-	-	-	
P. neglecta	-	3/5.77	-	-	
P. paeoniae	-	1/1.92	-	11/28.94	
, P. palmarum	3/1.67	-	-	-	
P. pandani	1/0.56	-	-	-	
P. pauciseta	2/1.11	3/4.77	-	-	
P. peycnonelii	1/0.56	-	-	-	
P. photiniae	8/4.44	1/1.92	3/5.08	-	
P. sorbi	1/0.56	-	-	-	
P. subcuticulari	-	-	-	3/7.89	
P. theae	-	-	-	4/10.53	
P. vaccinii	-	1/1.92	-	-	
P. versicolor	-	-	3/5.08	-	
P. virgatula	22/12.22	3/5.77	13/22.03	1/2.63	
P. vismiae	-	-	1/1.69	-	
P. zahlbruckneriana	1/0.56	-	-	-	
P. zonata	37/20.56	-	21/35.59	7/18.42	
Total number of isolates	180	52	59	38	
Shannon-Wiener index (H')	2.4641	2.5013	1.4775	1.8707	
Evenness index (J)	0.7859	1.0431	0.3624	0.8996	
Richness index (S)	23	11	9	8	

independent of the age of the palm. The distribution of endophytic fungi might be affected by tissue texture, physiology and chemistry (Petrini and Carroll, 1981; Polishook et al., 1996; Arnold et al., 2001; Cheng et al., 2009). In this study, the colonization frequencies of endophytic *Pestalotiopsis* were different in different tissue of same plant. In general, colonization frequencies in twigs were relatively higher than that in leaves.

The colonization frequencies of endophytic *Pestalotiopsis* in plant were influenced by the natural

condition. Kumaresan and Suryanayanan (2001) investigated the endophytic fungi of *R. apiculata* and *R. mucronata* growing in the Pichavaram mangrove of Tamil Nadu, Southern India. The results showed that more endophytes could be isolated during the rainy months than the dry period. Similar result was obtained in other study (Maria et al., 2003; Karamchand et al., 2009). In this study, the colonization frequencies of endophytic *Pestalotiopsis* in plant samples of Palmae and Podocarpaceae collected in April 2007 were significantly higher than that collected in April 2008. It was rainless in most area of Hainan during September 2007 to May 2008. The drought and high temperature may be the reasons for the differences in 2007 and 2008.

Species composition of endophytic Pestalotiopsis

A total of 43 endophytic *Pestalotiopsis* species were isolated from 37 plant species belonging to 4 families. Species composition of endophytic *Pestalotiopsis* varied in the different families of plants even though, in same family, species composition of endophytic *Pestalotiopsis* also displayed great differences. The number of *Pestalotiopsis* species isolated from Palmae was highest in the four plant families. This was mainly because the numbers of plant species in Palmae which were selected for comparison with the other three families in this study were more. This is also the main reason while the Evenness and Shannon-Wiener indexes of endophytic *Pestalotiopsis* were the highest in Rhizophoraceae and not in Palmae.

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REFERENCES

- Ananda K, Sridhar KR (2002). Diversity of endophytic fungi in the roots of mangrove species on the west of India. Can. J. Microbiol., 48: 871–878.
- Anita DD, Sridhar KR (2009). Assemblage and diversity of fungi associated with mangrove wild legume *canavalia cathartica*. Trop. Subtrop. Agroecosyst., 10: 225–235.
- Arnold AE, Maynard Z, Gilbert GS (2001). Fungal endophytes in dicotyledonous neotropical trees: Patterns of abundance and diversity. Mycol. Res., 105: 1502–1507.
- Bills GF, Polishhook JD (1994). Abundance and diversity of a lowland rain forest in Costa Rica. Mycologia, 86: 187–198.
- Cannon PF, Simmons CM (2002). Diversity and host preference of leaf endophytic fungi in the Iwokrama Forest Reserve, Guyana. Mycologia, 94: 210–220.
- Carroll GC, Carroll FE (1978). Studies on the incidence of coniferous needle endophytes in the Pacific Northwest. Can. J. Bot., 56: 3034–3043.
- Cheng ZS, Pan JH, Tang WC, Chen QJ, Lin YC (2009). Biodiversity and biotechnological potential of mangrove-associated fungi. J. For. Res., 20: 63–72.
- Fisher NL, Burgess LW, Toussoun TA, Nelson PE (1982). Carnation leaves as a substrate and for preserving cultures of *Fusarium* species. Phytopathology, 72: 151–153.

- Gilbert GS, Sousa WP (2002). Host specialization among wood-decay polypore fungi in a caribbean mangrove forest. Biotropica, 34: 396–404.
- Gao XX, Zhou H, Xu DY, Yu CH, Chen YQ, Qu LH (2005). High diversity of endophytic fungi from the pharmaceutical plant, *Heterosmilax japonica* Kunth revealed by cultivation-independent approach. FEMS Microbiol. Lett., 249: 255–266.
- Karamchand KS, Sridhar KR, Bhat R (2009). Diversity of fungi associated with estuarine sedge *Cyperus malaccensis* Lam. J. Agric. Technol., 5: 111-127.
- Kelemu S, Dongyi H, Guixiu H, Takayama Y (2003). Detecting and differentiating *Acremonium implicatum*: Developing a PCR-based method for an endophytic fungus associated with the genus Brachiaria. Mol. Plant Pathol., 4: 115–118.
- Kumar DSS, Hyde KD (2004). Biodiversity and tissue-recurrence of endophytic fungi in *Tripterygium wilfordii*. Fungal Divers., 17: 69–90.
- Kumaresan V, Suryanayanan TS (2001). Occurrence and distribution of endophytic fungi in a mangrove community. Mycol. Res., 105: 1388– 1391.
- Lacap DC, Hyde KD, Liew ECY (2003). An evaluation of the fungal 'morphotype' concept based on ribosomal DNA sequences. Fungal Divers, 12: 53–66.
- Liu AR, Xu T, Guo LD (2007). Molecular and morphological description of *Pestalotiopsis hainanensis* sp. nov., a new endophyte from a tropical region of China. Fungal Divers., 24: 23–36.
- Lodge DJ, Fisher PJ, Sutton BC (1996). Endophytic fungi of *Manilkara bidentata* leaves in Puerto Rico. Mycologia, 88: 733–788.
- Maria GL, Sridhar KR (2003). Diversity of filamentous fungi on woody litter of five mangrove plant species from the southwest coast of India. Fungal Divers., 14: 109–126.
- Nag Raj TR (1993). Coelomycetous Anamorphs with Appendagebearing Conidia. Canada: Mycologue Publications. Waterloo, Ontario, Canada, p. 1101.
- Okane I, Nagagiri A, Ito T (1997). Preliminary study of endophytic fungi in evergreen plants from Ishigaki and Iriomote Islands. The Institute for Fermentation, Osaka Research Communications, 18: 45–51.
- Okane I, Nagagiri A, Ito T (1998). Endophytic fungi in leaves of ericaceous plants. Can. J. Bot., 76: 657–663.
- Petrini O, Carroll GC (1981). Endophytic fungi in foliage of some Cupressaceae in Oregon. Can. J. Bot., 59: 629–636.
- Photita W, Lumyong S, Lumyong P, McKenzie EHC, Hyde KD (2004). Are some endophytes of *Musa acuminata* latent pathogens? Fungal Divers, 16: 131–140.
- Pielou EC (1975). Ecological Diversity. Wiley Inter Science, New York.
- Polishook JD, Bills GF, Lodge DJ (1996). Microfungi from decaying leaves of two rainforest trees in Puerto Rico. J. Ind. Microbiol., 17: 284–294.
- Rodriguez RJ, White Jr JF, Arnold AE, Redman RS (2009). Fungal endophytes: diversity and functional roles. New Phytol., 182: 314– 330.
- Steyaert RL (1949). Contribution to the study of Monographic Pestalotia Not.et Monochaetia Sacc. Bull. Jard. Bot. State Brussels, 19: 285– 354.
- Strobel G, Yang XS, Sears J, Kramer R, Sidhu RS, Hess WM (1996). Taxol from *Pestalotiopsis microspora*, an endophytic fungus of *Taxus wallichiana*. Microbiology, 142: 435–440.
- Suryanarayanan TS, Wittlinger SK, Faeth SH (2005). Endophytic fungi associated with cacti of Arizona. Mycol. Res., 109: 635–639.
- Sutton BC (1980). The Coelomycetes Kew, Surrey, England: Commonwealth Mycological Institute, pp. 263–266.
- Taylor JE, Hyde KD, Jones EBG (1999). Endophytic fungi associated with the temperate palm, *Trachycarpus fortunei*, within and outside its natural geographic range. New Phytol., 142: 335–346.
- Tejesvi MV, Kini KR, Prakash HS, Subbiah V, Shetty HS (2008). Antioxidant, antihypertensive, and antibacterial properties of endophytic *Pestalotiopsis* species from medicinal plants. Can. J. Microbiol., 54: 769–780.
- Tejesvi MV, Tamhankar SA, Kini KR, Rao VS, Prakash HS (2009). Phylogenetic analysis of endophytic *Pestalotiopsis* species from ethnopharmaceutically important medicinal trees. Fungal Divers., 38: 167–183.
- Toofanee BS, Dulymamode R (2002). Fungal endophytes associated

with Cordemoya integrifolia. Fungal Divers, 11: 169–175.

- Wei JG, Xu T (2004). *Pestalotiopsis kunmingensis* sp. nov., an endophyte from *Podocarpus macrophyllus*. Fungal Divers, 15: 247–254.
- White JF, Cole GT (1985). Endophyte-host associations in forage grasses. III. *In vitro* inhibition of fungi by *Acremonium coenophalium*. Mycologia, 77: 487–489.