

Varicosporellopsis, a new aquatic genus from southern France

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Abstract: *Varicosporellopsis aquatilis* gen. and sp. nov. is described and illustrated based on two collections on submerged wood in southern France. An acremonium-like asexual morph was obtained in culture and sequenced. The genus is placed in the *Nectriaceae* based on its asexual morph and phylogenetic comparison of ITS and LSU sequences with 21 genera of *Hypocreales* including 14 genera in the *Nectriaceae* and 7 genera in the *Bionectriaceae*. *Varicosporellopsis* is primarily characterized by obpyriform, pale orange, non-stromatic ascospores, freshwater habitat and an acremonium-like asexual morph.

Keywords: Ascomycota, freshwater habitat, *Hypocreales*, *Nectriaceae*, ribosomal DNA, taxonomy.

Résumé : *Varicosporellopsis aquatilis* gen. et sp. nov. est décrite et illustrée d'après deux récoltes effectuées sur bois immergé dans le sud de la France. Un stade asexué de type acremonium a été obtenu en culture et séquencé. Le placement du nouveau genre dans les Nectriacées repose sur le stade asexué et sur la comparaison phylogénétique de ses séquences ITS et LSU avec celles de 21 genres d'*Hypocreales* incluant 14 genres de Nectriacées et 7 genres de Bionectriacées (fig. 2, table 1). *Varicosporellopsis* est caractérisée par des ascospores orange pâle, non stromatiques, ne changeant pas de couleur dans KOH à 3%, devenant jaune très pâle à presque hyalins dans l'acide lactique, des ascospores variqueuses, un habitat aquatique et un stade asexué acremonium-morphe.

Mots-clés : ADN ribosomal, Ascomycota, habitat aquatique, Hypocréales, Nectriacées, taxinomie.

Introduction

In the continuity of a survey of freshwater pyrenomycetes in southwestern France, a hypocrealean fungus was twice collected on submerged wood, which proved different from known terrestrial species and different from species reported in the literature listed in LECHAT & FOURNIER (2015). Two species of *Hypocreales* occurring on submerged wood were recently described: *Lasionectria fourneri* Lechat (LECHAT, 2008) and *Varicosporella aquatica* Lechat & J. Fourn. (LECHAT & FOURNIER, 2015), but these genera are different from *Varicosporellopsis*. *Varicosporella* differs primarily from *Varicosporellopsis* and *Lasionectria* in having a fusarium-like asexual morph, while *Lasionectria* differs from *Varicosporellopsis* in having flexuous hyphae or long, stiff, erect, thick-walled setae on the ascomatal surface.

Materials and methods

Specimens were examined using the method described by ROSSMAN *et al.* (1999). Microscopic observations and measurements were made in water and the ascospore ornamentation was observed in lactic cotton blue without heating. The holotype specimen and the paratype are deposited in LIP herbarium (University of Lille) and cultures at CBS. Cultures of the living specimen were made on PDA (Potato Dextrose Agar) with 5mg/l of streptomycin in Petri dishes 55 mm diam. A mass of ascospores and asci was removed from a perithecium with a fine needle and placed in a drop of sterile water that was stirred with a needle to distribute the elements on the slide. Part of the drop containing ascospores was placed on PDA using a sterile micropipette, then the Petri dish was incubated at 25°C.

DNA extraction, amplification, and sequencing were performed by ALVALAB (Santander, Spain). Total DNA was extracted from dry specimens blending a portion of them using a micropestle in 600 µl CTAB buffer (CTAB 2%, NaCl 1.4 M, EDTA pH 8.0 20 mM, Tris-HCl pH 8.0 100 mM). The resulting mixture was incubated for 15 min at 65°C. A similar volume of chloroform: isoamylalcohol (24:1) was added and carefully mixed with the samples until their emulsion.

It was then centrifugated for 10 min at 13.000 g, and the DNA in the supernatant was precipitated with a volume of isopropanol. After a new centrifugation of 15 min at the same speed, the pellet was washed in cold 70% ethanol, centrifugated again for 2 min and dried. It was finally resuspended in 200 µl ddH₂O. PCR amplification

was performed with the primers ITS1F and ITS4 (WHITE *et al.*, 1990; GARDES & BRUNS, 1993) for ITS, while LR0R and LR5 (VILGALYS & HESTER, 1990) were used to amplify the 28S nLSU region. PCR reactions were performed using a program consisting of a hot start at 95°C for 5 min, followed by 35 cycles at 94°C, 54°C and 72°C (45, 30 and 45 s respectively) and a final 72°C step 10 min. PCR products were checked in 1% agarose gels, and positive reactions were sequenced with primer ITS4. Chromatograms were reviewed searching for putative reading errors, and these were corrected.

Analyses were performed online at www.phylogeny.lirmm.fr (DEREEPER *et al.*, 2008). Maximum likelihood phylogenetic analyses were performed with PhyML 3.0 aLRT (ZWICKL, 2006), using the GTR + I + Γ model of evolution. Branch support was assessed using the non-parametric version of the approximate likelihood-ratio test, implemented in PhyML (SH-aLRT; ANISIMOVA & GASCUEL, 2006).

Taxonomy

Varicosporellopsis Lechat & J. Fourn., *gen. nov.* – MB 815311

Diagnosis: Differs from the genus *Varicosporella* in having smaller ascospores and ascospores, and an acremonium-like asexual morph.

Type species: *Varicosporellopsis aquatilis* Lechat & J. Fourn.

Etymology: *Varicosporellopsis* refers to the morphological similarity with *Varicosporella* Lechat & J. Fourn.

Varicosporellopsis aquatilis Lechat & J. Fourn., *sp. nov.* – MB 815312 – Fig. 1-2.

Diagnosis: Ascospores on submerged wood, superficial, non-stromatic, obpyriform, pale orange, not changing colour in 3% KOH, becoming very pale yellow to nearly hyaline in lactic acid. Differs from *Varicosporella aquatica* in having smaller ascospores and an acremonium-like asexual morph.

Holotype: FRANCE, Ariège: Rimont, La Maille, La Maille brook, 550 m, 6 May 2015, on submerged wood of *Sambucus nigra*, coll. Jacques Fournier JF15039 (LIP). Ex-type culture CBS 140158, ITS and LSU GenBank sequences KU233187 and KU233189.

Additional material examined: FRANCE, Ariège: Clermont, Le Pujol brook along road D 119, ca. 360 m, 31 Jul. 2009, on submerged twig of *Buxus sempervirens*, coll. Jacques Fournier JF 09213 (LIP).

Etymology: The epithet refers to the freshwater lifestyle of the fungus.

Known distribution: Southern France (Ariège).

Ascomata non-stromatic, solitary, superficial with base slightly immersed in substratum, soft-textured, pale orange, not changing colour in 3% KOH, becoming very pale yellow to nearly hyaline in lactic acid, subglobose, laterally collapsing on drying, 240–280 µm high, 170–230 µm diam, with a translucent, broadly conical to rounded apex 50–80 µm high, 80–100 µm diam., composed of cylindrical

hyaline to pale yellow cells 15–35 µm long, 3–3.5 µm diam, thick-walled, septate, clavate at top. **Ascomatal wall** in vertical section 30–35 µm thick, composed of two regions; outer region 18–22 µm thick, composed of globose, subglobose to ellipsoidal, pale yellow, thick-walled cells 6–14 × 4–7.5 µm with wall 1.5–2 µm thick; inner region 10–15 µm thick, composed of hyaline and more flattened cells 6–10 × 3–5 µm. Ascomatal surface made of subglobose, sub-angular cells, covered by smooth, tan or hyaline hyphae 2.5–3 µm diam. Ostiolar canal periphysate. **Asci** unitunicate, cylindrical, short-stipitate, with eight obliquely uniseriate ascospores, 65–80 × 6.5–7.5 µm, apically truncate to slightly rounded with a faint apical ring-like thickening, interspersed with moniliform early deliquescing paraphyses up to 15 µm wide at base. **Ascospores** (9.5–)10–11(–12) × 4.5–6(–6.5) µm (Me = 10.5 × 5.5 µm, n = 30), ellipsoid with narrowly to broadly rounded ends, equally two-celled, slightly

Table 1 – Genera, species and GenBank accession numbers of sequences used in the phylogenetic analyses

Species	Asexual morph	ITS	LSU
<i>Albonectria rigidiuscula</i>	fusarium-like	HM054158	HM042403
<i>Bionectria ochroleuca</i>	clonostachys-like	KC460538	GQ50600
<i>Calonectria lauri</i>	cyliandrocladium-like	GQ280584	GQ280706
<i>Cosmospora vilioscula</i>	acremonium-like	KC291732	KC291777
<i>Cosmospora viridescens</i>	acremonium-like	KC291731	KC291765
<i>Hydropisphaera fusigera</i>	acremonium-like	GU059594	GU059595
<i>Hypomyces aurantius</i>	cladobotryum-like	KM509060	KC009213
<i>Lanatonectria flocculenta</i>	sarcopodium-like	JF832657	JF832714
<i>Lasionectria mantuana</i>	acremonium-like	HM484858	GQ505994
<i>Lasionectria</i> sp.	acremonium-like	JX306092	–
<i>Microcera larvarum</i>	fusarium-like	KC354705	KC338992
<i>Microcera coccophila</i>	fusarium-like	KC338994	KC338993
<i>Myrothecium inundatum</i>	gliocladium-like	AJ302005	–
<i>Myrothecium inundatum</i>	gliocladium-like	FJ797514	–
<i>Myrothecium roridum</i>	gliocladium-like	JF724154	–
<i>Myrothecium inundatum</i>	gliocladium-like	HQ165763	–
<i>Nalanthamala olivacea</i>	acremonium-like	AY554219	AY554255
<i>Nalanthamala psidii</i>	acremonium-like	AY554208	AY554258
<i>Nectria cinnabarina</i>	tubercularia-like	HM484712	HM484756
<i>Nectria pseudotrichia</i>	tubercularia-like	KF611683	JF832704
<i>Nectriopsis exigua</i>	acremonium-like	HM484865	GQ505986
<i>Neocosmospora vasinfecta</i>	fusarium-like	KF612306	AY381155
<i>Neocosmospora ornamentata</i>	fusarium-like	JX868635	AF178382
<i>Neonectria ditissima</i>	cyliandrocarpon-like	JF735309	KM515936
<i>Pseudocosmospora eutypae</i>	acremonium-like	KC291736	KC291786
<i>Pseudocosmospora rogersonii</i>	acremonium-like	KC291728	KC291780
<i>Pseudonectria pachysandricola</i>	volutella-like	JF832658	JF832715
<i>Pseudonectria rousseliana</i>	volutella-like	JF937563	JF937574
<i>Roumegueriella rufula</i>	acremonium-like	–	DQ518776
<i>Selinia pulchra</i>	acremonium-like	HM484859	GQ505992
<i>Stachybotrys chlorohalonata</i>	acremonium-like	AY489712	JN938870
<i>Stachybotrys chartarum</i>	acremonium-like	AF081468	AY489714
<i>Stachybotrys echinata</i>	acremonium-like	KF626506	AY489736
<i>Stachybotrys</i> sp.	acremonium-like	–	JF14101
<i>Stachybotrys</i> sp.	acremonium-like	–	JF14099
<i>Varicosporella aquatica</i>	fusarium-like	KP192669	KP192671
<i>Varicosporella aquatica</i>	fusarium-like	KP192668	KP192670
<i>Xenoacremonium recifei</i>	acremonium-like	KM231834	KM231714
<i>Viridispora diparietispora</i>	penicilifer-like	JN049838	AY489735

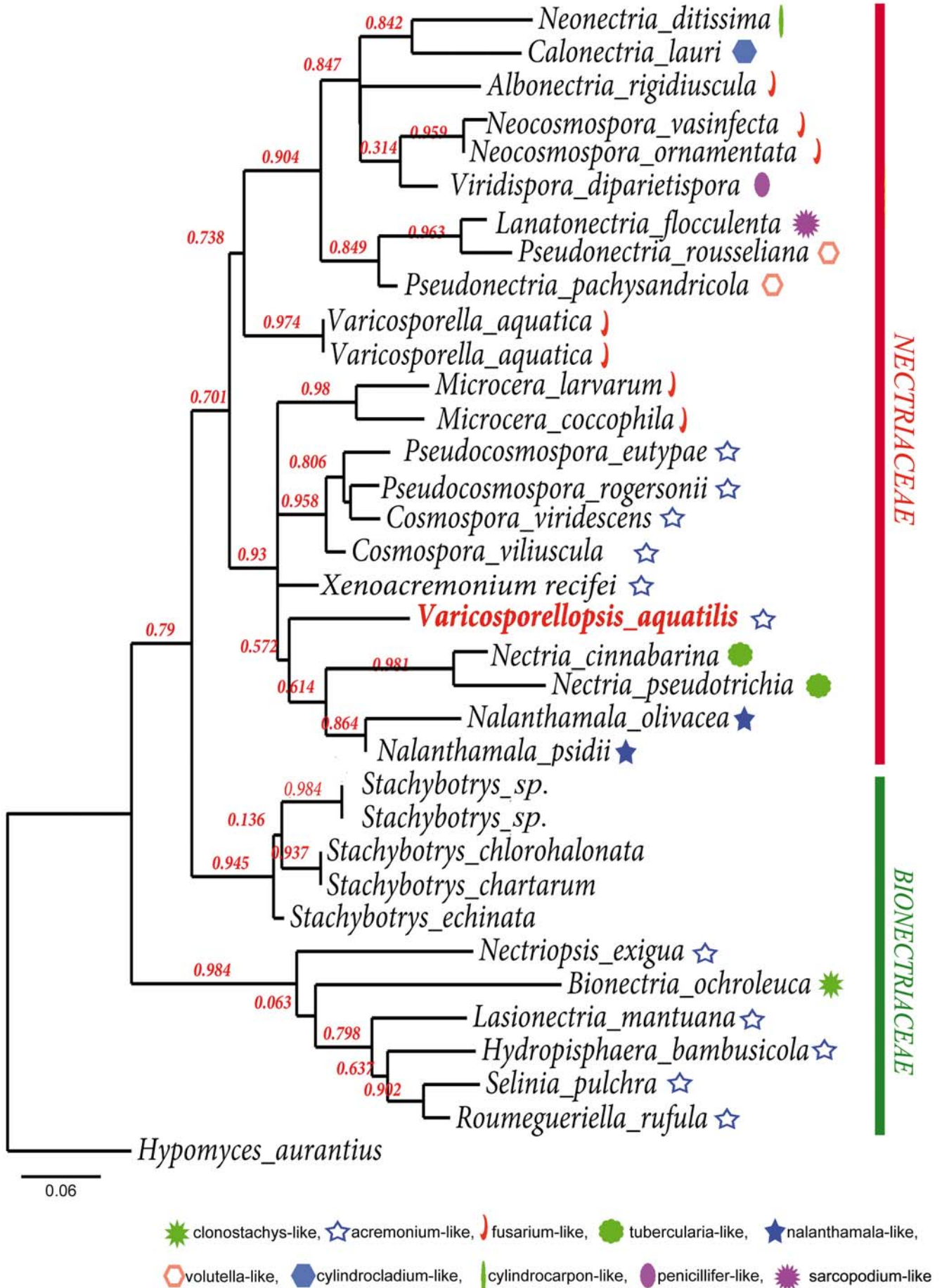


Fig. 1 – Maximum likelihood phylogeny of *Varicosporellopsis aquatilis* based on combined ITS and LSU sequences, rooted with *Hypomyces aurantius*.

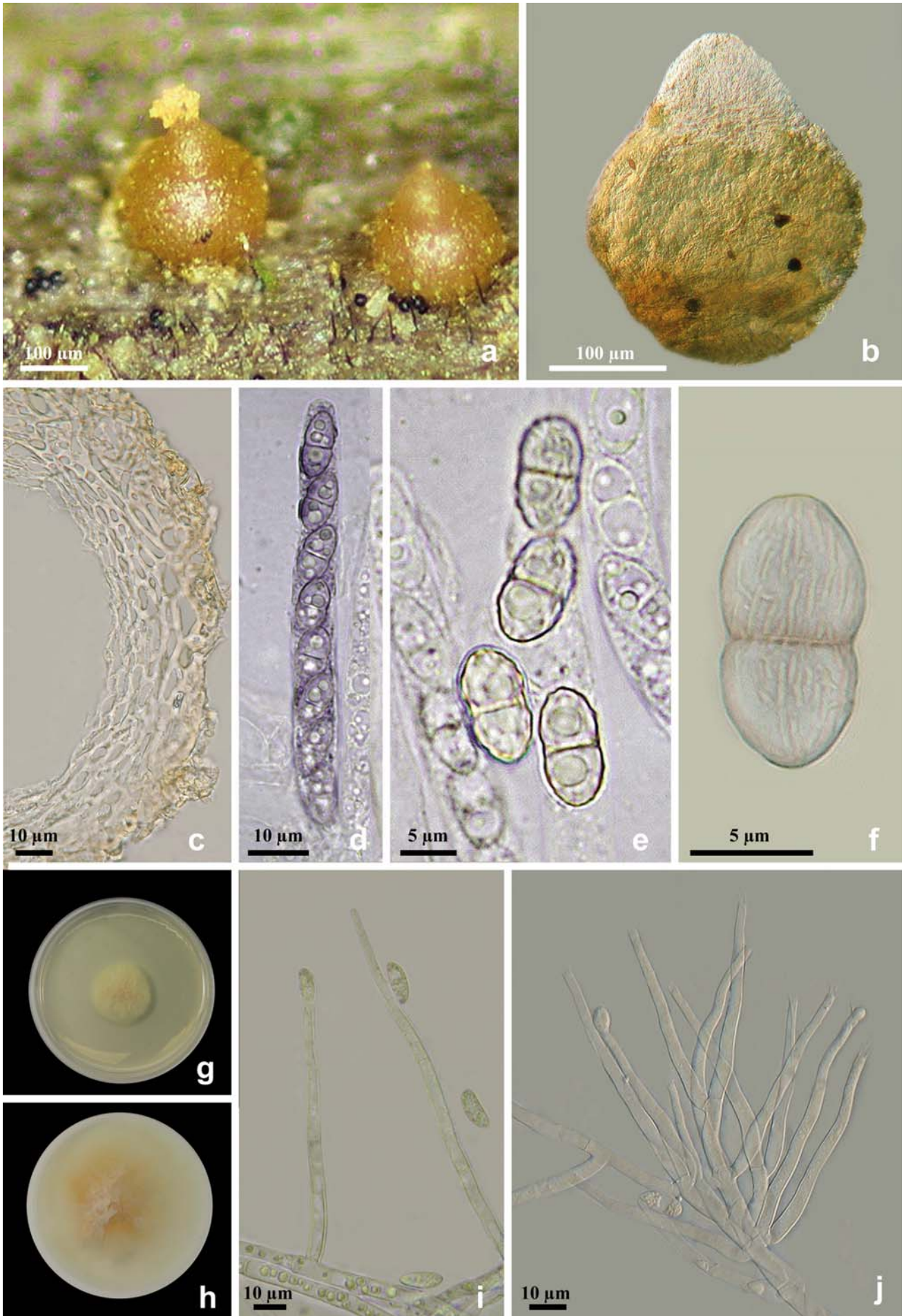


Fig. 2 – a-e: *Varicosporellopsis aquatilis*, Holotype. a: Ascomata on the substratum; b: Close-up of perithecium in water; c: Lateral ascomatal wall in vertical section; d-f: Ascus and ascospores; g: Culture after two weeks; h: Culture after three weeks; i-j: Conidiophores and conidia.

constricted at septum, hyaline to pale yellowish brown, with 2–3 large guttules in each cell, wall roughened by short, sinuous, brown, thick ribs, sometimes anastomosed.

Cultural characteristics: Colony after two weeks on PDA measuring 45–50 mm diam, aerial mycelium white in center, salmon in median area and pale yellowish at margin, abundantly sporulating; reverse pale yellow to pale yellowish brown. Conidiophores macro-nematous, mononematous, unbranched, flexuous, hyaline, smooth; conidiogenous cells monophialidic, 1.5–2 µm wide at apex with a slightly flared collarette, producing narrowly ellipsoidal to subcylindrical conidia 6–11 × 2.8–3.2 µm. After three weeks verticillate conidiophores appear, arising laterally from submerged hyphae. Phialides cylindrical, subulate, smooth, hyaline to pale salmon producing widely ellipsoidal conidia, attenuated at base without a flat abscission scar, 5–8 × 3–4.5 µm, smooth, hyaline.

Discussion

Typically, the ascomata of *Nectriaceae* are brightly coloured, changing colour in 3% KOH or lactic acid, as defined by ROSSMAN *et al.* (1999) and SCHROERS (2011). However, some genera are known to have ascomata not changing colour in 3% KOH but turning yellow in lactic acid: *Albonectria* Rossman & Samuels, *Pseudonectria* Seaver (ROSSMAN *et al.*, 1999) and *Varicosporella* (LECHAT & FOURNIER, 2015). *Varicosporellopsis aquatilis* is primarily characterized by obpyriform, pale orange, non-stromatic ascomata, not changing colour in 3% KOH, becoming very pale yellow to nearly hyaline in lactic acid, short-ribbed ascospores and an acremonium-like asexual morph. Based on the characters of its sexual and asexual morphs as well as phylogenetic analysis (Fig. 1), this taxon clearly belongs to the *Nectriaceae*. Morphologically, *Varicosporellopsis* is similar to *Varicosporella* in having pale orange, non-stromatic ascomata, not changing colour in 3% KOH and freshwater habitat, but differs from it in having smaller ascomata and ascospores, as well as an acremonium-like asexual morph while *Varicosporella* possesses a fusarium-like asexual morph. Molecular analysis carried out in the present study, comparing 14 genera in the *Nectriaceae* including the type species of *Albonectria*, *Pseudonectria* and *Varicosporella* (Fig. 1), shows that *Varicosporellopsis* belongs to the *Nectriaceae* but is placed on a branch distant from these three genera. In the phylogenetic tree, the closest genera to *Varicosporellopsis* are *Nalanthamala* Subram., *Nectria* (Fr.) Fr. and *Xenoacremonium* L. Lombard & Crous.

Only the asexual morph of *Xenoacremonium* is known and although it has acremonium-like morphology, like *Varicosporellopsis*, it differs from the latter in having unbranched or rarely branched conidiophores, and conidia forming slimy heads at tip of the phialide, as defined by LOMBARD *et al.* (2015). *Nalanthamala* and *Nectria* are very different from *Varicosporellopsis* in having brightly coloured ascomata, seated on a pseudoparenchymatous stroma, changing colour in 3% KOH and lactic acid, ascomatal wall over 50 µm thick and a sprodochial asexual morph as defined respectively by LOMBARD *et al.* (2015) and HIROOKA *et al.* (2012). Based on phylogenetic, mor-

phological and ecological divergences, the new genus *Varicosporellopsis* is proposed.

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References

- ANISIMOVA M. & GASCUEL O. 2006. — Approximate likelihood-ratio test for branches: A fast, accurate, and powerful alternative. *Systematic Biology*, 55 (4): 539-552.
- DEREEPER A., GUIGNON V., BLANC G., AUDIC S., BUFFET S., CHEVENET F., DUFAYARD J.F., GUINDON S., LEFORT V., LESCOT M., CLAVERIE J.M. & GASCUEL O. 2008. — Phylogeny.fr: robust phylogenetic analysis for the non-specialist. *Nucleic Acids Research*, 36 (Web Server issue), 2008: W465-W469. doi:10.1093/nar/gkn180
- GARDES M. & BRUNS T.D. 1993. — ITS primers with enhanced specificity for basidiomycetes-application to the identification of mycorrhizae and rusts. *Molecular Ecology*, 2 (2): 113-118.
- HIROOKA Y., ROSSMAN A.Y., SAMUELS G.J., LECHAT C. & CHAVERRI P. 2012. — A monograph of *Allantonectria*, *Nectria* and *Pleonectria* (*Nectriaceae*, *Hypocreales*, *Ascomycota*) and their pycnidial, sporodochial and synnematosus anamorphs. *Studies in Mycology*, 71: 1-210.
- LECHAT C. 2008. — *Lasionectria fournieri* sp. nov. et son anamorphe *Acremonium*. *Bulletin de la Société mycologique de France*, 124 (1-2): 1-5.
- LECHAT C. & FOURNIER J. 2015. — *Varicosporella*, a new aquatic genus in the *Nectriaceae* from France. *Ascomycete.org*, 7 (1): 1-8.
- LOMBARD L., MERWE N.A. (VAN DER), GROENEWALD J.Z. & CROUS P.W. 2015. — Generic concepts in *Nectriaceae*. *Studies in Mycology*, 80: 1-87.
- ROSSMAN A.Y., SAMUELS G.J., ROGERSON C.T. & LOWEN R. 1999. — Genera of *Bionectriaceae*, *Hypocreaceae* and *Nectriaceae* (*Hypocreales*, *Ascomycetes*). *Studies in Mycology*, 42: 1-248.
- SCHROERS H.-J. 2001. — A monograph of *Bionectria* (*Ascomycota*, *Hypocreales*, *Bionectriaceae*) and its *Clonostachys* anamorphs. *Studies in Mycology*, 46: 1-214.
- VILGALYS R. & HESTER M. 1990. — Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *Journal of Bacteriology*, 172 (8): 4238-4246.
- WHITE T.J., BRUNS T., LEE S. & TAYLOR J.W. 1990. — Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: INNIS M.A., GELFAND D.H., SNINSKY J.J. & WHITE T.J. (eds.). *PCR protocols: a guide to methods and applications*. New York, Academic Press: 315-322.
- ZWICKL D.J. 2006. — *Genetic algorithm approaches for the phylogenetic analysis of large biological sequence datasets under the maximum likelihood criterion*. Ph.D. Dissertation. Austin, The University of Texas, i-x + 115 p.



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