

International Journal of Advances in Pharmacy Medicine and Bioallied Sciences

INTERNATIONAL JOURNAL OF ADVANCES IN PHARMACY MEDICINE AND BIOALUED SCIENCES

The Pharmacy of the Pharmacy of

An International, Multi-Disciplinary, Peer-Reviewed, Open Access, Triannually Published Biomedical Journal | www.biomedjournal.com |

Efficacy of methyl benzimidazole-2-yl carbamate against pathogenic fungi of leafy vegetables

Vishal N Shinde¹, Sandip S Nandre¹, Zafar S. Khan^{2*}

¹Department of Botany, ADMSP's Late Annasaheb R D Deore, Arts & Science College, Mhasdi, Tal. Sakri, Dist. Dhule- 424304, MS, India. ²Department of Botany, SGVSSP's Arts, Commerce and Science College Onde, Tal. Vikramgad, Dist. Thane-401605, MS, India.

SHORT COMMUNICATION

ARTICLE INFORMATION

Article history

Received: 15 March 2014 Revised: 20 March 2014 Accepted: 20 April 2014 Early view: 28 April 2014

 ${}^{\star}Author\, for\, correspondence$

E-mail: vishalshinde1001@gmail.com

Mobile/ Tel.: 0000000000

Keywords:

Methyl benzimidazole-2-yl Carbamate

Mycotoxicity Leafy vegetables

Percent Diseases Reduction (PDR).

ABSTRACT

India has achieved self sufficiency and good degree of stability of vegetable crop production. Leafy vegetables are most essential component of our diet which nourishes with nutrients, minerals and vitamins. The aim of the present work is to evaluate methyl benzimidazole-2-yl carbamate against 10 pathogenic fungi of leafy vegetables in-vitro and in-vivo. The Poisoned food technique was applied for in-vitro fungicides assessment and percent inhibition of mycelia growth over control was calculated. For in-vivo study each leafy vegetable sown in 12×24 M plots in the field. After 10 day of interval, 200 ml spore suspension of each targeted plant pathogenic fungi was mixed in the soil of the respectively field. After 7 days, the diseases symptoms were developed on the leaves of vegetables. Afterwards Minimum Inhibitory Concentration (MIC) in µg/ml of Carbendazim from in vitro results was selected for in-vivo study. The defined concentrations of Methyl benzimidazole-2-yl carbamate was sprayed directly onto the infected leafy vegetables. Efficacy of each fungicide was evaluated by calculating the Percent Diseases Incidence (PDI) and Percent Diseases Reduction (PDR) over control. Invitro Carbendazim revealed 50.99% inhibition of mycelia growth against all of targeted fungal pathogens. MIC of Carbendazim to all fungal pathogens varied from 40-12000 µg/ml. *In vivo* studies, Carbendazim was highly effective in controlling disease incidence. The percent disease reduction (PDR) with Carbendazim treatment was maximum 84.64% in all treated leafy vegetables. The use of Carbendazim is very effective against pathogenic fungi of C. esculanta, S. oleracia and T. foenumgraecum in the field condition as they showed complete percent disease reduction (PDR). Biomedjournal © Copyright 2013, All rights reserved. Biomedjournal Privacy Policy.

INTRODUCTION

India has achieved self sufficiency and good degree of stability of vegetable crop production. Leafy vegetables are most essential component of our diet which nourishes with nutrients, minerals and vitamins. For healthy diet, daily minimum consumption should be about 180 g per head; whereas the present consumption of leafy vegetables is less than 20 g per head. Vegetables being more succulent and rich in nutrients are prone to variety of diseases right from the sowing to marketing, thereby increasing yield losses during pre and post production periods. In India, out of total production, leafy vegetables are prone to several fungal diseases most commonly causing leaf spot and wilting. Due to these diseases annually billions of rupees loss occurs throughout the country (Harlapur et al., 2007). Therefore, there is an urgent need to explore and cultivate leafy vegetables in India; even though India stand second largest producer followed by China. To control these diseases, the Int J Adv Pharmacy Med Bioallied Sci. 2, 1, 2014.

pesticides being widely used throughout the world which on contrary increasing the agricultural production with increasing pesticide concentration. Older pesticides are eliminated from market due to regulatory changes and new pesticides are becoming expensive, so there is a need to find out more potent fungicides for the safe use. The development of new physiological race pathogens to many of the systemic fungicides is gradually becoming ineffective (Wellman, 1977). Thus several broad spectrum fungicides are recommended for controlling fungal diseases. Resulted, the use of fungicides has been increasing steadily at an annual rate of about 14% since the mid 1950s. Therefore, appropriate fungicide should be used to overcome all these major problems. In spite of use of all available means of plant protection, about 1/3rd of yearly harvested leafy vegetable production of the world is destroyed by many diseases and loss due to this is expected to be nearly 600 crore. Different fungicides are tested against various pathogens (Ravishanker

Mamatha, 2005; Harlapur et al., 2007) throughout the world.

In the present investigation, methyl benzimidazole-2-yl carbamate (carbendazim) which is systemic fungicide was tested for their mycotoxicity *in-vitro* and *in-vivo*. Minimum inhibitory concentration (MIC), percent diseases incidence (PDI) and percent diseases reduction (PDR) against ten pathogenic fungi of leafy vegetables were determined.

MATERIAL AND METHODS

In-vitro fungicidal assay

For the assessment of in-vitro fungicidal assay, poisoned food technique was used. The required concentration of fungicide were prepared as parts per million (ppm) in µg/ml with sterilized double distilled water. Out of this standard concentration, 5 ml of methyl benzimidazole-2yl carbamate concentration was taken and added to 45 ml sterilized PDA medium and mixed well. Afterwards medium with fungicide concentration was transferred equally into two sterilized petri plates and media was allowed to solidify. After complete solidification of the medium, 4 mm diameter disk of 5-7 day old culture of targeted fungi was taken and inoculated into the center of petri plates in aseptic condition. The PDA medium containing petri plate without fungicide concentration was served as a control. Then all the petri plates were incubated at 28 ± 2 °C and radial growth of colony was measured after 3rd day up to 7th day constantly. The results of mycelia growth were expressed as mean of triplicates. The concentration of fungicide at which the pathogen showed complete inhibition of its mycelia growth was considered as minimum inhibitory concentration (MIC) of fungicide to respective pathogen and percent inhibition of mycelia growth over control was calculated by the formula given by Vincent (1947).

Where I=Inhibition of mycelia growth C=Mycelia growth in control T=Mycelia growth in treated.

In- vivo fungicidal assay

For the assessment of fungicidal assay at field condition, each leafy vegetable was sown in 12×24 M plot in the field. After 10 days of interval, 200 ml spore suspension of each targeted plant pathogenic fungi was mixed in the soil of the respectively field. After 7 days, the diseases symptoms were developed on the leaves of vegetables. Afterwards minimum inhibitory concentration (MIC) in µg/ml of carbendazim shown in-vitro study considered for in-vivo study. The defined concentration of methyl benzimidazole-2-yl carbamate was sprayed directly on the infected leafy vegetables. The fungicide treatment was applied twice at an interval of 10 days for all leafy vegetables. In all cases, leafy vegetable without fungicide treatment served as control and tagged. Simultaneously all treated leafy vegetables were also tagged with respect to tested concentrations. After 10 days of treatment, among each treated leafy vegetable plants, the total number of leaves on each plant and total number of infected leaves on each plant were counted and average of triplicate was calculated. The effectiveness of each fungicide was evaluated by calculating the percent diseases incidence (PDI) and percent diseases reduction (PDR) over control by using following formula:

PDI= <u>Number of diseased leaves on each plant</u> × 100 Total number of leaves on each plant

PDR= PDI in control - PDI in treatment × 100
PDI in control

RESULTS

Efficacy of carbendazim in-vitro

Methyl benzimidazole-2-yl carbamate (carbendazim) a systemic fungicide was tested for the assessment of fungicidal efficacy and determination of their MIC against ten different pathogenic fungi of leafy vegetables such as Alternaria brassicae, Alternaria carthami, Alternaria humicola, Collectotrichum lindemuthianum, Fusarium moniliforme, Fusarium oxysporum, Fusarium roseum, Helminthosporium sativum, Pullularia pullulans and Stemphylium verruculosum. Percent inhibition of mycelia growth over control was tabulated (Table 2). The lowest concentration which showed complete inhibition of mycelia growth was considered as MIC of fungicide to particular pathogen (Table 1).

The MIC value of carbendazim fungicide against ten pathogenic fungi of leafy vegetables was varied from 40 $\mu g/ml$ to 12,000 $\mu g/ml$ (Table 1). The pathogens *C. lindemuthianum* and *F. moniliforme* were found to be most susceptible and revealed MIC values at 40 $\mu g/ml$ and 80 $\mu g/ml$ respectively. While *H. sativum* and *A. brassicae* were found to be most resistant as showed highest MIC values (12,000 $\mu g/ml$ and 1000 $\mu g/ml$ respectively).

The effect of carbendazim fungicide on the growth rate of mycelium of ten pathogenic fungi of leafy vegetables was most significant (Table 2). The percent inhibition of mycelia growth of *C. lindemuthianum* and *S. verruculosum* were found to be maximum (i.e. 80.07% and 60.18%) respectively to all tested concentrations. The percent inhibition of mycelia growth of *F. roseum*, *F. moniliforme*, *F. oxysporum*, *P. pullulans*, *A. brassicae* and *A. carthami* was significantly inhibited. Lowest percent of mycelia growth was observed in *H. sativum* and *A. humicola*.

Table 1. MIC of carbendazim against plant pathogenic fungi in $\mu g/ml$.

Pathogen	Carbendazim
Alternaria brassicae	*1000
Alternaria carthami	500
Alternaria humicola	700
Collectotrichum lindemuthianum	40
Fusarium moniliforme	80
Fusarium oxysporum	100
Fusarium roseum	90
Helminthosprium sativum	12000
Pullularia pullulans	90
Stemphylium verruculosum	300

^{*}all values expressed in mean of triplicates.

Table 2. Inhibitory effect of carbendazim on the mycelia growth of targeted fungi.

Pat	С												Mean of %					
hog	on				I							III					III	inhibition
en	tr ol	20	40	60	80	90	100	200	300	400	500	600	700	800	900	1000	12000	
A.b.	84	*80	75	72	64	63	62	54	48	42	33	24	13	10	07	-		40.52 <u>+</u> 0.74
		(4.76)	(10.71)	(17.85)	(23.80)	(25)	(26.10)	(37.71)	(42.85)	(50)	(60.71)	(71.42)	(84.52)	(88.09)	(91.66)	(100)		
A.c.	78	74	68	62	55	51	47	33	18	09	-							
		(5.12)	(12.82)	(20.21)	(29.48)	(34.61)	(39.74)	(57.69)	(76.89)	(88.46)	(100)							39.57 <u>+</u> 0.90
A.h.	81	78	74	68	64	61	59	48	35	27	17	09	-					
		(3.70)	(8.06)	(16.04)	(20.28)	(24.69)	(27.16)	(40.74)	(56.79)	(66.66)	(79.01)	(88.88)	(100)					36.41 <u>+</u> 0.79
C.l.	64	24	19	08	-													
		(62.50)	(70.31)	(87.50)	(100)													80.07 <u>+</u> 0.55
F.m	75	59	42	21	-													
		(21.33)	(44)	(72)	(100)													52.99+0.94
F.o.	75	63	48	31	14	10	-											_
		(16)	(36)	(58.66)	(74.66)	(86.66)	(100)											52.26 <u>+</u> 0.89
<i>F.r.</i>	68	46	34	22	11	-												
		(32.35)	(50)	(67.64)	(83.82)	(100)												58.82 <u>+</u> 1.05
H.s.	84										0.0					5 0		
											80					78	(100)	20.24.0.75
D	7.4	60	40	20	10						(4.76)					(7.14)	(100)	38.24 <u>+</u> 0.75
P.p.	74	60	48	29	12	(100)												50.00.0.04
		(18.19)	(35.13)	(60.81)	(83.78)	(100)												50.89 <u>+</u> 0.84
C	72						16	29										
S.v.	12						46		(100)									60.18 <u>+</u> 0.87
							(36.11)	(73.61)	(100)									00.100.07
]																1	

^{*}Mean diameter of mycelia growth in mm at varied concentration ($\mu g/ml$) and figure in parenthesis represents percent inhibition of mycelia growth at varied concentration. Where A.b. = A. Brassicae, A.c. = A. Carthami, A.h. = A. Aumicola, C.l. = C. Color of the concentration of mycelia growth at varied concentration. Where <math>A.b. = A. Carthami, C.l. = C. Carthami, C.l. = C. Carthami, C.l. = C. Carthami, C.l. = C. Carthami, Carthami,

In vivo fungicidal assay

or the assessment of fungicidal efficacy *in-vivo*, MIC in µg/ml of carbendazim shown *in-vitro* study considered and applied for *in-vivo* study the effectiveness of fungicide were recorded as percent disease incidence (PDI) and percent disease reduction (PDR).

The effectiveness of different fungicidal treatments on percent disease incidence (PDI) and percent disease reduction (PDR) revealed that carbendazim was highly effective in controlling the disease incidence. The percent of disease reduction exhibited by carbendazim treatment was significant with an average 84.64% (Table 3). The PDR was occurred from 68-100 percent in selected plants. Hundred percent PDR was observed in *C. esculanta*, *S. oleracia* and *T. foenumgraecum* after spraying of carbendazim.

Table 3. <i>In-vivo</i> effect of carbendazim on leafy vegetable disease reduction

		Control		Treated			DDI :		
Leafy vegetables	Fungicides	No. of infected leaf	Total no. of leaf	No. of Infected leaf	Total no. of leaf	PDI in control	PDI in treated	PDR (%)	
A. gaevolens	Carbendazim	8	16	1	15	50	6.66	86.68	
B. oleraceae	Carbendazim	5	9	1	10	55.55	10	81.99	
C. tinctorius	Carbendazim	6	15	2	16	40	12.50	68.75	
C. esculanta	Carbendazim	3	5	0	5	60	00	100	
C. sativum	Carbendazim	6	19	2	17	31.57	11.76	62.74	
R. vesicarious	Carbendazim	9	29	2	28	31.03	7.14	76.99	
S. oleracia	Carbendazim	5	14	0	13	35.71	00	100	
T. foenum- graecum	Carbendazim	5	12	0	9	41.66	00	100	

All values of mean of triplicate; where PDI = Percent diseases incidence and PDR = Percent diseases reduction.

DISCUSSION

In preliminary assessment, carbendazim was tested for their fungitoxicity against ten fungal pathogens of leafy vegetables namely (Cambell et al. 2000). Among the pathogen *A. carthami*, *F. moniliforme* and *S. verruculosum* were found to be most susceptible against tested fungicides whereas *A. brassicae* and *H. sativum* were most resistant to fungicide. Similar finding was reported by several researchers (Tu and Jarvis, 1979; Ravishanker and Mamatha, 2005; Harlapur et al., 2007). Pandu et al. (1986) and Xiujian et al., (2000) also reported fungicidal efficacy of carbendazim, captan, benomyl, triademefon, propicanzole and suggested that systemic fungicide were more effective than non systemic fungicide against *C. fimbriata* Eillis and Halsted. The fungicide mancozeb and captan being recommended for management of diseases like seedling blight of *A. falcataria*, leaf spot diseases of *Populus deltoids* caused by *Alternaria alternata* (Dey and Debata, 2000); leaf spot and blight of *Syzygium cumini* caused by *Cylindrocladium quinqueseptatum* (Mehrotra and Mehrotra, 2000) followed by Rodomil and Bayleton were effective against *F. solani*.

In the present investigation, it was recorded that there were variation in MIC of carbendazim against ten fungal pathogens of leafy vegetables. Bains and Mohan (1982) reported that heterogeneous population of resistant and sensitive nuclei in the isolate might be responsible for variation in the MIC of fungicides. Similarly variation in sensitivity and resistant of different fungal pathogens to fungicides was reported by several workers (Dekker and Gielink, 1979; Jones and Ehret, 1981; Gangawane and Saler, 1981).

Carbendazim in the field condition reduced the diseases incidence on leafy vegetables. Siddaramaiah et al. (1980) reported similar result as carbendazim is effective against a wide range of fungal pathogen and has been used to control many disease such as leaf spot of *W. tinctoria* caused by *Cercospora wrightii*.

CONCLUSION

The present study demonstrated that the use of carbendazim is very effective against pathogenic fungi of *C. esculanta*, *S. oleracia* and *T. foenumgraecum* in the field condition as they showed complete percent disease reduction (PDR).

CONFLICT OF INTEREST

None declared.

REFERENCES

Bains SS, Mohan C. Location and behavior of *Helminthosporium maydis* isolate, sensitive and tolerant to Diathane M-45. *Indian Phythopath*, 35-589, 1982.

Cambell NA, Mitchell LG, Rece JB. *Biology concepts and connections*. 3rd edition. Addison Wesley Longman: Inc. New York, 2000.

Dekker J, Gielink AJ. Acquired resistance to pimaricin in *Cladosporim cucumerinum* and *Fusarium oxysporum f. sp.* narcicci associated with decreased virulence. Neth J Plant Pathol, 86, 67-73, 1979.

Dey A, Debata DK. Studies on leaf spot disease of *Popules deltoids* Marsh caused by *Alternaria raphani*. *Indian forester*, 126, 1013-1014, 2000.

Gangawane LV, Saler RS. Resistance to fungicide in Aspergillus flavus. Neth J Plant Pathol, 87, 254, 1981.

Harlapur SI, Kulkarni MS, Wali MC, Kulkarni S. Evaluation of plant extracts, Bio-agent and fungicides against *Exserohilum turcicum* (Pass.) Lonard and Suggs. causing Triticum leaf blight of maize. *Karnataka J Agri Sci*, 20, 541-544, 2007.

Jones AL, Ehert GR. Resistance of Coccnyces liemalis to benzimidazole fungicide. *Plant Sis*, 64, 767-769, 1981.

Mehrotra A, Mehrotra MD. Leaf spotting blight, a new disease of Syzygium cumini by two cylindrocladium species from India. Indian J Forestry, 23, 496-500, 2000.

Pandu RS, Rao RSV, Sharma MN, Subbayya J. Effectiveness of non mercurial fungicide for the control of sugarcane. *Indian Phytopath.* 39, 306-308, 1986.

Ravishanker RV, Mamatha T. Seedling disease of some important forest tree species and their management. Working Papers of the Finnish Forest Research Institute 11. 2005.

http://www.metla.fi/julkaisut/workingpapers/2005/mwp01 1.htm

Siddaramaiah Al, Kulkarni S, Basvarajaiah AB. Control of leaf spot diseases of *Wrightia tinctoria* Br. in the forest nursery. *Indian Forester*, 106, 771-774, 1980.

Tu JC, Jarvis WR. Response of *Collectotrichum lindumuthanium* to benomyl. *Canadian J Plant Pathol*, 1, 12-16, 1979.

Vincent JM. Distortion of fungal hyphae in the presence of certain inhibitors. *Nature*, 150, 850-853. 1947.

Wellman RH. Problem in development resistant and use of fungicide. *Annua Rev Phytopathol*, 15, 155-163, 1977.

Xiujian Y, Chen FR, Zhang LXS. Screening of fungicide for control of *Ceratocystis fimbriata* Ellis and Halsted. *Plant Protection*, 26, 38-39, 2000.