S672

Asian Pacific Journal of Tropical Biomedicine

journal homepage: www.apjtb.com



Document heading

doi:10.12980/APJTB.4.201414B214

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Antidiarrhoeal and antibacterial activity of aqueous and methanolic leaves extracts of Dissotis thollonii Cogn. (Melastomataceae)

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ARTICLE INFO

Article history: Received 23 May 2014 Received in revised form 6 Jun 2014 Accepted 25 Jun 2014 Available online 9 Jul 2014

Keywords: Antidiarrhoeal Antibacterial Phytochemical test Dissotis thollonii

ABSTRACT

Objective: To investigate the phytochemical test and selected pharmacological activities (antidiarrhoeal and antibacterial activity) of the aqueous and methanolic leaves extracts of Dissotis thollonii Cogn. (Melastomataceae) (D. thollonii).

Methods: The aqueous and methanolic extracts were evaluated for their antibacterial activities on the in vitro growth of 2 clinical isolates (Staphylococcus aureus and Shigella flexneri), and 5 reference bacteria strains [Escherichia coli ATCC 8739 (E. coli), E. coli ATCC 10536, Salmonella typhi ATCC 6539, Enterobacter aerogenes ATCC 13048 and E. coli ATCC 11775] by determining the minimum inhibitory concentrations (MICs) and bactericidal concentrations using broth microdilution method as well as on the infectious, secretory and osmotic induced diarrhoea models in rats.

Results: The aqueous extract inhibited the *in vitro* growth of all bacteria tested (the 05 reference bacteria strains and the 02 clinical isolates), with MICs values comprised between 32 and $512 \mu g/mL$, whereas the methanolic extract has done the same with the MICs values located between 128 and 512 µg/mL. In vivo, the methanolic and aqueous extracts provoqued at all doses, a significant decrease (P<0.001) in the bacterial load in the faces of rats, 6 and 7 d of treatment respectively. Infected animals relatively lost weight when treated with the aqueous extract but it remained constant for those treated with the methanolic extract. The results also showed that, the aqueous and methanolic leaves extracts of *D. thollonii* had, after 6 h of observation, significantly (P<0.001) inhibited in vivo, diarrhoea induced experimentally by castor oil and magnesium sulphate, particularly by prolonging the latency time, reducing the water content of faeces, the frequency of defecation and the number of wet defecation as compared to the negative control which received distilled water and the animals having received 2.5 mg/kg of loperamide used as reference substance. The phytochemical assay revealed the presence of flavonoids, phenols and polyphenols in the leaves extracts of D. thollonii, which are compounds with antidiarrhoeal and antibacterial activities.

Conclusions: The leaves of *D. thollonii* thus have antibacterial and antidiarrhoeal effects, which could result from their activities on blocking the inhibiting effects of the bacterial enzymes, inhibiting the bacterial protein synthesis, allowing the rupture of the lipopolysaccharidic membrane, as well as on inhibiting prostaglandins-E₂ synthesis or increasing the hydroelectrolytic reabsorption. These results attestted the ethnopharmacological use of D. thollonii leaves in the treatment of diarrhoea and gastro-intestinal infections.

1. Introduction

Diarrhoeal diseases represent the second cause of infant mortality after pneumonia in developing countries. In

sub-Saharan Africa, 18% per year is the infant (less than 5 years) mortality rate related to diarrhoea^[1]. According to World Health Organization^[2], diarrhoea is responsible for almost two million deaths of children every year which represent approximately 20% of infant mortality rate. To face with this physiopathology, various products are used, coming from chemotherapy or phytotherapy. In developing countries, the study of the plants can urgently

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lead to the adequate resolution of the medical needs to lower cost, joint with a proven scientific effectiveness and an optimal cultural acceptability. Thus, the World Health Organization encourages the development of the traditional medicine which uses the medicinal plants partly^[3]. Among the many exploited species, Dissotis thollonii (D. thollonii) is traditionally used in the treatment of several evils. Thus it is used by traditional healer in Senegal, on eczematous wounds and in west region of Cameroon^[4]. The leaves are recommended in therapy of the gastrointestinal disorders (obstruction, amoebiasis, diarrhoeas, vomiting, and constipation) and ulcers. The aim of this study was, therefore, to evaluate the antidiarrhoeal activities of aqueous and methanolic leaves extracts of D. thollonii in experimentally induced-diarrhoea in rats and the in vitro antibacterial activities of the same extracts over some enterobacteria strains.

2. Materials and methods

2.1. Collection and identification of plant material

The fresh leaves of *D. thollonii* were collected in April 2012 in the Bafou village, Menoua division, West Region of Cameroon. A sample was identified at the National Herbarium of Cameroon (Yaoundé) and repertoried under code 13292/SRF Cam.

2.2. Preparation of the plant material

The collected plant parts (leaves) were separated from undesirable materials. They were dried under shade, ground and stored in an airtight container prior to extraction.

2.3. Preparation of plant extract

The aqueous extract was prepared by boiling 600 g of powder in 3.6 L distilled water for 15 min as indicated by the traditional healer. The decoction was cooled during 1 h and filtered using filter paper (Whatman No. 1), and the filtrate was evaporated in a regulated drying oven at 35 °C to give 38 g of the aqueous extract corresponding to an extraction yield of 6.33% (w/w). The other portion of leaf powder (200 g) was macerated in 1.5 L of methanol for 72 h, filtered and the solvent removed from the extract under reduced pressure, using a Büchi (R–200) rotary evaporator at 65 °C. This gave 7 g of the methanol extract, corresponding to a yield of 7.00% (w/w).

2.4. Experimental animal

Wistar Albino female rats of 2 to 2.5 months and weighing

on average 140 g were used for infectious induced diarrhoea model whereas both sexes (1.5 to 2 months) of the same rat strain, weighing on average 100 g, were used for the tests of diarrhoea induced by magnesium sulfate and castor oil. They were bred in the animal house of the Department of Animal Biology, University of Dschang, Cameroon, under natural room conditions. Animals were fed with a standard diet and received water *ad libitum*. Prior to experimental protocol, the rats were acclimatized for 48 h to laboratory conditions for minimizing any nonspecific stress. The studies were conducted according to standard protocols for the use of laboratory animals (Registration No. 173/CPCSEA, January 2000, India).

2.5. Chemicals and reagents

All reagents used in the study were of high purity. The chemicals included ethylic alcohol 95°, methanol, distilled water, bleach (Geochim, Cameroon), sodium chloride (Sigma Chemicals Co., UK), iodonitrotetrazolium (Sigma, St Louis), ciprofloxacin (Sinochem Ningbo, P.R., China), tetracycline, Mueller Hinton broth, Salmonella–Shigella agar, loperamide (imodium), castor oil and magnesium sulphate.

2.6. Microorganisms

The microorganisms used in this study included 2 clinical isolates [*Staphylococcus aureus* (*S. aureus*) and *Shigella flexneri* (*S. flexneri*)] and 5 bacterial strains [*Escherichia coli* ATCC 8739 (*E. coli*), *E. coli* ATCC 10536, *Salmonella typhi* ATCC 6539, *Enterobacter aerogenes* ATCC 13048 (*E. aerogenes*) and *E. coli* ATCC 11775]. The strains were maintained at 4 °C on agar plates.

2.7. Phytochemical tests

The extracts were analyzed for the presence of various phytoconstituents. The qualitative determination of these phytochemical compounds was conducted according to the method described by Wahid *et al*[5].

2.8. In vitro antibacterial activity

The *in vitro* antibacterial activity of the extract was performed by determining the minimum inhibitory concentrations (MICs) using broth microdilution method[6]. For that, 100 μ L of Mueller Hinton broth was firstly introduced in 96 wells microplates. The stock solutions of extracts and ciprofloxacin (standard drug) were consequently introduced in triplicate into the first well and a serial two fold dilution was performed to obtain 12 concentrations, ranging from 1024 to 8 μ g/mL for the extracts and from 8 to 1 μ g/mL for the standard drug. Then, 100 μ L of the inoculum

was added into each well for a final volume of 200 µL/well. The microplates were, therefore, carefully covered with a sterile lid, packed and incubated at 37 °C for 24 h after which 20 μL of iodonitrotetrazolium (INT) was added in one of the pairs of column of well. The viable bacteria were observed through a pink colour in the presence of INT. All concentrations at which no visible colour changes were observed were considered as inhibitory concentrations and the lowest of these concentrations was considered as the MIC^[7]. The bactericidal concentrations were determined by adding 50 µL aliquots of the preparations (without INT), which did not show any visible colour change after incubation during MIC assays, into 150 µL of extract-free Mueller Hinton. After 48 h of incubation at 37 °C, all extracts concentrations at which no colour changes were observed were considered as bactericidal concentrations. The smallest of these concentrations was considered as the minimal bactericidal concentration (MBC).

2.9. S. flexneri-induced diarrhoea

The experimental animals were firstly deparasitized through an oral administration of 10 mg/kg of tetracycline for 3 d^[8]. These animals were further acclimatized in fitted cages, where they were receiving standard diet and water ad libitum. Furthermore, animals were left under sterile conditions during 4 d in order to maximize the complete elimination of the deparasitizing substance. At the end of this period, rats were deprived from food for 18 h, but received water ad libitum and then orally inoculated with a S. flexneri suspension, coldly prepared at the 1.0 McFarland standard scale (1.5×10⁸ CFU/mL). Infected rats were set out in groups and submitted to a treatment with three test doses (500, 250 and 125 mg/kg of body weight (b.w) of plant extracts. Group 1 (neutral) was made of non-infected/non-treated rats. The third group (positive control) received ciprofloxacin at 2.5 mg/kg b.w as reference drug and Group 2 (negative control) received distilled water. All the substances (extracts plants and ciprofloxacin) were administered per os until complete elimination of bacterial load in faeces. These faeces were collected each day between 8 h and 9 h. The follow-up of the treatment was carried out by evaluating the load of S. *flexneri* in a culture of 0.5 g of faeces on Salmonella-Shigella agar Petri dishes, incubated at 37 °C for 24 h.

2.10. Castor oil-induced diarrhea in rats

The animals were initially screened by observing stool's aspect. Those not showing diarrheic stools were selected for the final experiment. Thus, forty–eight rats were randomly divided into eight equal group (n=6) divided into controls, standard and test groups. The negative control groups

received distilled water (1 mL/100 g b.w). The positive control group received loperamide at the dose of 2.5 mg/kg orally. The test groups received aqueous and methanolic extracts of *D. thollonii* at doses of 125, 250 and 500 mg/kg orally. Each animal was placed in individual cage, the floor of which was lined with filter paper, changed every hour. Diarrhoea was induced by oral administration of 1 mL/100 g b.w castor oil to each rat, 1 h after the above treatment. During an observation period of 6 h, parameters such as the latency time, the frequency of defecation, the number of wet defecation and the water content of feces were recorded. Water content of feces was expressed in terms of percentages using the formula:

$$Wc (\%) = \left(\frac{Fw - Dw}{Fw}\right) \times 100$$

Where, Wc=Water content of feces; Fw=Fresh weight (g); Dw=Dry weight (g).

2.11. Magnesium sulphate-induced diarrhoea in rats

A similar protocol as for castor oil-induced diarrhoea was followed. Diarrhoea was induced by oral administration of magnesium sulfate at the dose of 3 g/kg to the animals, 1 h after administration of distilled water to the negative control groups, loperamide (2.5 mg/kg) to the positive control group, the plant extracts of *D. thollonii* in doses of 125, 250 and 500 mg/kg b.w to the test groups. All the treatments were administered orally.

2.12. Statistical analysis

Values are presented as mean±SEM (standard error of the mean). The measurement data were analyzed by One–way and Two–way ANOVA, followed by Tukey–Kramer multiple comparison test if the overall differences were significant (P<0.05) for secretory and osmotic induced diarrhoea, whereas the Bonferroni post–test (P<0.05) was used for the infectious diarrhoea.

3. Results

3.1. Phytochemicals tests

Several groups of phytoconstituents compounds were highlighted in the two extracts of *D. thollonii*. The phytochemicals tests showed the presence of tannins, flavonoids, sterols, anthraquinones, phenols and polyphenols in the methanolic extract of the leaves of *D. thollonii* whereas the aqueous extract contained only two types of secondary metabolites (flavonoids and polyphenols).

3.2. Evaluation of antibacterial activity

Both plant expressed antibacterial activities on all the tested microorganisms. However, these activity were more interesting with the aqueous extract, which presented an inhibitory effect at lower concentrations (MIC between 32 and 512 µg/mL) and bactericidal activity on five bacteria (*S. flexneri* and *S. aureus* (clinical isolate); *E. aerogenes* ATCC 13048; *E. coli* ATCC 10536 and *E. coli* ATCC 11775) of the seven tested, with MBC/MIC ratio <4. Whereas methanolic extract did the same with MIC between 128 and 512 µg/mL. Results are showed in Table 1.

Table 1

Antibacterial activities (MIC, MBC and MBC/MIC ratios) of aqueous and methanolic leaves extracts of *D. thollonii*.

Test substances (µg/mL)	Parameters	St	Ec	Ea	Sa	Ec1	Sf	Ec2
Methanolic extract	MIC	128	128	256	128	512	128	128
	MBC	512	512	512	1 0 2 4	512	512	512
	MBC/MIC	4	4	2	8	1	2	4
Aqueous extract	MIC	32	64	128	512	256	128	64
	MBC	256	128	256	>1024	256	256	256
	MBC/MIC	8	2	2	2	1	2	4
Ciprofloxacin	MIC	2.00	0.25	2.00	2.00	0.50	2.00	0.125
	MBC	2.00	0.25	2.00	2.00	0.50	2.00	0.125
	MBC/MIC	1	1	1	1	1	1	1

St: Salmonella typhi ATCC 6539; Ec: E. coli ATCC 11775; Ea: E. aerogenes ATCC 13048; Sa: S. aureus; Ec1: E. coli ATCC 10536; Sf: S. flexneri; Ec2: E. coli ATCC 8739.

3.3. S. flexneri-induced diarrhoea in rats

The aqueous and methanolic extracts of *D. thollonii* leaves, produced significant (P<0.001) decrease in the bacterial load in the feces of rats during 9 and 8 d of treatment, respectively. The doses of 250 and 500 mg/kg of aqueous and methanolic extracts respectively, exhibited a highly significant (P<0.001) effect from Day 7, when compared to negative control group and similarly as the standard drug used. Results are illustrated by Figures 1 and 2.



Figure 1. Evolution of the bacterial load (*S. flexneri*) in the faeces of infected–rats treated with the aqueous extract of *D. thollonii*. C and γ: significant differences (*P*<0.001) compared with negative and positive control, respectively.





control, respectively. 3.4. Castor oil–induced diarrhoea in rats

Both aqueous and methanolic extracts at dose 500 mg/kg, highly significantly (P<0.001) prolonged the latency time, reduced the frequency of defecation, the number of wet defecations and the water content of feces when compared to the negative control groups. In addition, this dose of plant extracts had practically reacted like loperamide, used as standard drug. Generally, the extracts inhibited diarrhoea in a dose–dependent manner (Table 2, Figures 3–8). Table 2

Effects of the aqueous and methanolic leaves extracts of *D. thollonii* on castor oil induced diarrhoea.

Treatment	Dose	Latency time	Defecation Number of v		et Water content	
	(mg/kg)	(min)	frequency	defecations	of faeces (%)	
Distilled water	1 mL/100 g b.w	66.50±2.50	03.66±0.21	03.66±0.21	90.39±3.52	
Standard	2.5	$360.00 \pm 0.00^{\circ}$	$00.00 \pm 0.00^{\circ}$	$00.00 \pm 0.00^{\circ}$	$00.00 \pm 0.00^{\circ}$	
loperamide	125	$77.17 \pm 2.73^{b\gamma}$	$02.00{\pm}0.00^{\mathrm{e}\gamma}$	$02.00 \pm 0.00^{c\gamma}$	84.06 ± 2.13^{b}	
Aqueous	250	$183.00 \pm 1.80^{c\gamma}$	$01.66 \pm 0.20^{c\gamma}$	$01.66 \pm 0.20^{c\gamma}$	$73.64 \pm 1.24^{c\gamma}$	
extract	500	$355.50 \pm 1.20^{\circ}$	$01.33 \pm 0.20^{c\gamma}$	$01.33 \pm 0.20^{c\gamma}$	$51.31 \pm 2.12^{c\gamma}$	
	125	56.83±0.64 ^{cγ}	$03.50 \pm 0.22^{\gamma}$	$03.50 \pm 0.22^{\gamma}$	85.87±1.04 ⁷	
Methanolic	250	$129.00 \pm 1.18^{c\gamma}$	$03.00 \pm 0.00^{\gamma}$	$03.00 \pm 0.00^{\gamma}$	$76.31 \pm 1.20^{e^{\gamma}}$	
extract	500	170.70±1.05 ^{cγ}	02.33±0.20 ^{cγ}	02.33±0.20 ^{cγ}	63.96±1.26 ^{eγ}	

Values of each column represent the mean±ESM (*n*=6); ^{c}P <0.001: significant differences compared to the negative control (distilled-water); $^{\gamma}P$ <0.001: significant differences compared to the positive control (loperamide).







Figure 4. Effect of aqueous leaves extract of *D. thollonii* on water content of faeces in castor oil induced diarrhoea.



Defecation frequency Number of wet defecation

Figure 5. Effect of aqueous leaves extract of *D. thollonii* on defecation frequency and the number of wet defecation in castor oil induced diarrhoea.



Figure 6. Effect of the methanolic leaves extract of *D. thollonii* on latency time in castor oil induced diarrhoea.



Figure 7. Effect of the methanolic leaves extract of *D. thollonii* on water content of faeces in castor oil induced diarrhoea.



Figure 8. Effect of the methanolic leaves extract of *D. thollonii* on defecation frequency and the number of wet defecation in castor oil induced diarrhoea.

3.5. Magnesium sulphate-induced diarrhoea in rats

The treatment with standard drug (loperamide) and with all the doses of aqueous and methanolic leaves extracts of *D. thollonii*, produced a significant inhibition of osmotic diarrhoea induced by the magnesium sulfate. This was made by positively affecting all the diarrhoeal parameters evaluated, that is prolonging the diarrhoeal latency time, reducing the water content of faeces, reducing the defecation frequency and number of wet defecations, compared with negative control rats having received distilled water. Results are more pronounced in Table 3 and Figures 9–14. **Table 3**

Effects of the aqueous and methanolic leaves extracts of *D. thollonii* on magnesium sulfate induced diarrhoea.

Treatment	Dose	Latency time	Defecation Number of w		Water content	
	(mg/kg)	(min)	frequency	defecations	of faeces (%)	
Distilled water	1 mL/100 g b.w	135.90±0.34	04.14±0.14	04.14±0.14	97.61±0.12	
Standard	2.5	$356.70 \pm 0.60^{\circ}$	$01.00 \pm 0.00^{\circ}$	$01.00 \pm 0.00^{\circ}$	$40.69 \pm 2.14^{\circ}$	
loperamide	125	$151.70 \pm 0.56^{c\gamma}$	$03.00 \pm 0.00^{c\gamma}$	$03.00 \pm 0.00^{c\gamma}$	$79.97 \pm 1.00^{e\gamma}$	
Aqueous	250	$209.30 \pm 0.49^{c\gamma}$	$02.50 \pm 0.19^{c\gamma}$	$02.38 \pm 0.19^{c\gamma}$	$63.99 \pm 1.26^{c\gamma}$	
extract	500	$243.00 \pm 1.32^{e\gamma}$	$02.12 \pm 0.12^{e\gamma}$	$01.87 \pm 0.12^{c\gamma}$	$54.95 \pm 1.24^{\circ \gamma}$	
	125	$216.70 \pm 0.64^{c\gamma}$	$02.57 \pm 0.20^{c\gamma}$	$02.57 \pm 0.20^{c\gamma}$	$80.86 \pm 0.74^{c\gamma}$	
Methanolic	250	$234.10 \pm 1.93^{e\gamma}$	$02.28 \pm 0.18^{c\gamma}$	$02.28 \pm 0.18^{c\gamma}$	$71.04 \pm 0.73^{c\gamma}$	
extract	500	$257.30 \pm 2.14^{c\gamma}$	$01.85 \pm 0.14^{c\beta}$	$01.85 \pm 0.14^{c\beta}$	$65.71 \pm 1.02^{e\gamma}$	

Values of each column represent the mean±ESM (*n*=6); ${}^{c}P$ <0.001: significant differences compared to the negative control (distilled-water); ${}^{\beta}P$ <0.01; ${}^{\gamma}P$ <0.001: significant differences compared to the positive control (loperamide).



Figure 9. Effect of the aqueous leaves extract of *D. thollonii* on latency time in magnesium sulfate induced diarrhoea.



Figure 10. Effect of the aqueous leaves extract of *D. thollonii* on water content of faeces in magnesium sulfate induced diarrhoea.



Figure 11. Effect of the aqueous leaves extract of *D. thollonii* on defecation frequency and the number of wet defecations in magnesium sulfate induced diarrhoea.



Figure 12. Effect of the methanolic leaves extract of *D. thollonii* on latency time in magnesium–sulfate induced diarrhoea.



Figure 13. Effect of the methanolic leaves extract of *D. thollonii* on water content of faeces in magnesium–sulfate induced diarrhoea.



Defecation frequency Involves of wet defecation

Figure 14. Effect of the methanolic leaves extract of *D. thollonii* on defecation frequency and the number of wet defecation in magnesium–sulfate induced diarrhoea.

4. Discussion

The results of antimicrobial activities, revealed that the two extracts were active against the the 2 clinical isolates tested and 5 bacterial strains. Nevertheless, aqueous extract presented a greater activity against the bacterial strains and isolates as compared to the methanolic extract. This could be due to secondary metabolites (polyphenols and the flavonoids) present in various extracts, of which several pharmacological and antibacterial activities were proved^[9–12]. It was also demonstrated that these secondary metabolites were equipped with good antibacterial molecule sites, which caused the rupture of the lipopolysaccharidic membrane of the Gram–negative bacteria like *E. coli*, thus facilitating the restoration of protein channels–like structures to support the flow of antimicrobial compounds towards intra bacterial target sites^[13,14]. In another hand, this activity could be done either by inhibiting the bacterial protein synthesis, or by blocking the inhibiting effects of the bacterial enzymes^[15]. It should be noted that the active ingredients which act on the wall of the bacterium must find suitable receptors for their fixation and action, whereas those which act inside the cell must not only cross the membrane, but they must also find target in the cell^[16]. The installation of the diarrhoea, associated to the febrility observed at Day 3 in infected rats, could be explained by the intestinal invasion of S. flexneri, coupled with the action of verotoxin which have led to a generalized tiredness of the organism. This bacterium would have thus penetrated in the epithelial cells of the mucous membrane, where it would have quickly multiplied and provoked the formation of abscesses and ulcerations, leading to the modification of intestinal reabsorption mechanisms[17]. The reduction in the bacterial load observed from the 2nd day in all infected animals faeces, treated with selected dose extracts (500, 250 and 125 mg/kg) could be due to the same presence of polyphenols and the flavonoids which have acted through the bactericidal mechanisms described above, as demonstrated by Yala et al^[18]. The ricinoleic acid, released after digestion by intestinal lipases of the castor oil stimulates the peristaltic activity of the small intestine, thus causing a change of the permeability of the electrolytes of intestinal mucus^[19]. The aqueous and methanolic leaves extract of D. thollonii at different doses could have acted according to anti secretary and spasmolytic mechanisms, through the inhibition of prostaglandins-E₂ biosynthesis and by increasing the reabsorption of electrolytes (Na⁺, K⁺, Cl⁻). Unfortunately, both leaves plant extracts at dose 500 mg/kg could have also acted through the mechanisms comparable to that of loperamide which is known to inhibit the release of the inflammatory mediators, antagonized the histaminic activity when being fixed at the type-1 histaminic receptor (H_1) , thus preventing the excessive contractions of the intestinal smooth muscle. In a healthy person, an excess of magnesium can lead to osmotic diarrhoea associated to distension of the intestine. Magnesium sulfate induced diarrhoea by increasing the secretion of electrolytes which create an osmotic imbalance. The antidiarrhoeal properties of flavonoids rise from their ability to inhibit the intestinal motility and the hydroelectrolytic secretions^[20]. The antidiarrhoeal activity of the aqueous and methanolic extracts against the osmotic diarrhoea, induced by magnesium sulfate could be attributed to their antisecretory actions, coupled with the re-establishment of intestinal osmotic balance. It is thus noticed that the significant prolongment of the latency time, the significant reduction of defecation frequency, the reduction of water content of faeces and number of wet defecations demonstrated the efficacy of leaves extracts of D. thollonii, as effective

antidiarrhoeal agents.

In conclusion, this study showed that *D. thollonii* leaves extracts possessed antibacterial and antidiarrhoeal activities in rats, mediated firstly through bacteriostatic or bactericidal pathway and secondly through the re-establishment of intestinal balance. The study thus provided pharmacological basis to the use of the plant in intestinal disorders like diarrhoea and suggests using other experimental species, in order to illustrate an understandable conclusion which can be allowed to a human approach.

Conflict of interest statement

We declare that we have no conflict of interest.

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