

Fusarium: more than a node or a foot-shaped basal cell

P.W. Crous^{1,2}, L. Lombard^{1*}, M. Sandoval-Denis^{1,3*}, K.A. Seifert⁴, H.-J. Schroers⁵, P. Chaverri^{6,7}, J. Gené⁸, J. Guarro⁸, Y. Hirooka⁹, K. Bensch¹, G.H.J. Kema², S.C. Lamprecht¹⁰, L. Cai^{11,12}, A.Y. Rossman¹³, M. Stadler¹⁴, R.C. Summerbell^{15,16}, J.W. Taylor¹⁷, S. Ploch¹⁸, C.M. Visagie¹⁹, N. Yilmaz¹⁹, J.C. Frisvad²⁰, A.M. Abdel-Azeem²¹, J. Abdollahzadeh²², A. Abdolrasouli^{23,24}, A. Akulov²⁵, J.F. Alberts²⁶, J.P.M. Araújo²⁷, H.A. Ariyawansa²⁸, M. Bakhshi²⁹, M. Bendiksby^{30,31}, A. Ben Hadj Amor¹, J.D.P. Bezerra³², T. Boekhout¹, M.P.S. Câmara³³, M. Carbia³⁴, G. Cardinali³⁵, R.F. Castañeda-Ruiz³⁶, A. Celis³⁷, V. Chaturvedi³⁸, J. Collemare¹, D. Croll³⁹, U. Damm⁴⁰, C.A. Decock⁴¹, R.P. de Vries¹, C.N. Ezekiel⁴², X.L. Fan⁴³, N.B. Fernández^{44,45}, E. Gaya⁴⁶, C.D. González⁴⁷, D. Gramaje⁴⁸, J.Z. Groenewald¹, M. Grube⁴⁹, M. Guevara-Suarez⁵⁰, V.K. Gupta^{51,52}, V. Guarnaccia⁵³, A. Haddaji⁵⁴, F. Hagen¹, D. Haelewaters^{55,56}, K. Hansen⁵⁷, A. Hashimoto⁵⁸, M. Hernández-Restrepo¹, J. Houbraken¹, V. Hubka⁵⁹, K.D. Hyde⁶⁰, T. Iturriaga⁶¹, R. Jeewon⁶², P.R. Johnston⁶³, Ž. Jurjević⁶⁴, İ. Karalti⁶⁵, L. Korsten⁶⁶, E.E. Kuramae^{3,67}, I. Kušan⁶⁸, R. Labuda⁶⁹, D.P. Lawrence⁷⁰, H.B. Lee⁷¹, C. Lechat⁷², H.Y. Li⁷³, Y.A. Litovka^{74,75}, S.S.N. Maharachchikumbura⁷⁶, Y. Marin-Felix¹⁴, B. Matio Kemkuignou¹⁴, N. Matočec⁶⁸, A.R. McTaggart⁷⁷, P. Mičoch⁷⁸, L. Mugnai⁷⁹, C. Nakashima⁸⁰, R.H. Nilsson⁸¹, S.R. Noumeur⁸², I.N. Pavlov^{74,75}, M.P. Peralta⁸³, A.J.L. Phillips⁸⁴, J.I. Pitt⁸⁵, G. Polizzi⁸⁶, W. Quaedvlieg⁸⁷, K.C. Rajeshkumar⁸⁸, S. Restrepo⁸⁹, A. Rhaïem⁹⁰, J. Robert⁵⁴, V. Robert¹, A.M. Rodrigues⁹¹, C. Salgado-Salazar⁹², R.A. Samson¹, A.C.S. Santos⁹³, R.G. Shivas⁹⁴, C.M. Souza-Motta⁹³, G.Y. Sun⁹⁵, W.J. Swart⁹⁶, S. Szoke⁵⁴, Y.P. Tan^{94,97}, J.E. Taylor⁹⁸, P.W.J. Taylor⁹⁹, P.V. Tiago⁹³, K.Z. Váczy¹⁰⁰, N. van de Wiele⁵⁴, N.A. van der Merwe¹⁹, G.J.M. Verkley¹, W.A.S. Vieira³³, A. Vizzini¹⁰¹, B.S. Weir⁶³, N.N. Wijayawardene¹⁰², J.W. Xia¹⁰³, M.J. Yáñez-Morales¹⁰⁴, A. Yurkov¹⁰⁵, J.C. Zamora¹⁰⁶, R. Zare²⁹, C.L. Zhang¹⁰⁷, and M. Thines^{18,108,109}

¹Westerdijk Fungal Biodiversity Institute, 3508 AD, Utrecht, the Netherlands; ²Wageningen University and Research Centre (WUR), Laboratory of Phytopathology, Droevendaalsesteeg 1, 6708 PB, Wageningen, the Netherlands; ³Netherlands Institute of Ecology (NIOO-KNAW), Department of Microbial Ecology, Droevendaalsesteeg 10, 6708 PB, Wageningen, the Netherlands; ⁴Department of Biology, Carleton University, 1125 Colonel By Drive, Ottawa, Ontario, K1S 5B6, Canada; ⁵Plant Protection Department, Agricultural Institute of Slovenia, Hacquetova ulica 17, 1000, Ljubljana, Slovenia; ⁶Department of Plant Science and Landscape Architecture, University of Maryland, College Park, MD, USA; ⁷Escuela de Biología y Centro de Investigaciones en Productos Naturales, Universidad de Costa Rica, San Pedro, Costa Rica; ⁸Unitat de Micologia, Facultat de Medicina i Ciències de la Salut i Institut d'Investigació Sanitària Pere Virgili (IISPV), Universitat Rovira i Virgili, 43201, Reus, Spain; ⁹Department of Clinical Plant Science, Faculty of Bioscience, Hosei University, 3-7-2 Kajino-cho, Koganei, Tokyo, 184-8584, Japan; ¹⁰ARC-Plant Health and Protection, Private Bag X5017, Stellenbosch, 7599, Western Cape, South Africa; ¹¹State Key Laboratory of Mycology, Institute of Microbiology, Chinese Academy of Sciences, Beijing, 100101, China; ¹²University of Chinese Academy of Sciences, Beijing, 100049, China; ¹³Department of Botany & Plant Pathology, Oregon State University, Corvallis, OR, 97330, USA; ¹⁴Department of Microbial Drugs, Helmholtz Centre for Infection Research GmbH (HZI), Inhoffenstrasse 7, 38124 Braunschweig, Germany; ¹⁵Sporometrics, Toronto, ON, Canada; ¹⁶Dalla Lana School of Public Health, University of Toronto, Toronto, ON, Canada; ¹⁷Plant and Microbial Biology, 111 Koshland Hall, University of California, Berkeley, CA, 94720-3102, USA; ¹⁸Senckenberg Biodiversity and Climate Research Center, Senckenberganlage 25, D-60325, Frankfurt am Main, Germany; ¹⁹Department of Biochemistry, Genetics and Microbiology, Forestry and Agricultural Biotechnology Institute (FABI), Faculty of Natural and Agricultural Sciences, University of Pretoria, P. Bag X20, Hatfield, 0028, Pretoria, South Africa; ²⁰Department of Biotechnology and Biomedicine, DTU-Bioengineering, Technical University of Denmark, 2800, Kongens Lyngby, Denmark; ²¹Systematic Mycology Lab., Botany and Microbiology Department, Faculty of Science, Suez Canal University, Ismailia, 41522, Egypt; ²²Department of Plant Protection, Faculty of Agriculture, University of Kurdistan, P.O. Box 416, Sanandaj, Iran; ²³Department of Medical Microbiology, King's College Hospital, London, UK; ²⁴Department of Infectious Diseases, Imperial College London, London, UK; ²⁵Department of Mycology and Plant Resistance, V. N. Karazin Kharkiv National University, Maidan Svobody 4, 61022, Kharkiv, Ukraine; ²⁶Department of Food Science and Technology, Cape Peninsula University of Technology, P.O. Box 1906, Bellville, 7535, South Africa; ²⁷School of Forest Resources and Conservation, University of Florida, Gainesville, FL, USA; ²⁸Department of Plant Pathology and Microbiology, College of Bio-Resources and Agriculture, National Taiwan University, No.1, Sec.4, Roosevelt Road, Taipei, 106, Taiwan, ROC; ²⁹Iranian Research Institute of Plant Protection, Agricultural Research, Education and Extension Organization (AREEO), P.O. Box 19395-1454, Tehran, Iran; ³⁰Natural History Museum, University of Oslo, Norway; ³¹Department of Natural History, NTNU University Museum, Trondheim, Norway; ³²Setor de Micologia/ Departamento de Biotecnologia e Tecnologia, Instituto de Patologia Tropical e Saúde Pública, Rua 235 - s/n - Setor Universitário - CEP: 74605-050, Universidade Federal de Goiás/Federal University of Goiás, Goiânia, Brazil; ³³Departamento de Agronomia, Universidade Federal Rural de Pernambuco, Recife, 52171-900, PE, Brazil; ³⁴Departamento de Parasitología y Micología, Instituto de Higiene, Facultad de Medicina - Universidad de la República, Av. A. Navarro 3051, Montevideo, Uruguay; ³⁵Department of Pharmaceutical Science, University of Perugia, Via Borgo 20 Giugno, 74 Perugia, Italy; ³⁶Instituto de Investigaciones Fundamentales en Agricultura Tropical Alejandro de Humboldt (INIFAT), Académico Titular de la Academia de Ciencias de Cuba; ³⁷Grupo de Investigación Celular y Molecular de Microorganismos Patógenos (CeMoP), Departamento de Ciencias Biológicas, Universidad de Los Andes, Bogotá, 111711, Colombia; ³⁸Mycology Laboratory, New York State Department of Health Wadsworth Center, Albany, NY, USA; ³⁹Laboratory of Evolutionary Genetics, Institute of Biology, University of Neuchâtel, CH-2000, Neuchâtel, Switzerland; ⁴⁰Senckenberg Museum of Natural History Görlitz, PF 300 154, 02806, Görlitz, Germany; ⁴¹Mycothèque de l'Université catholique de Louvain (MUCL, BCCMTM), Earth and Life Institute - ELIM - Mycology, Université catholique de Louvain, Croix du Sud 2 bte L7.05.06, B-1348, Louvain-la-Neuve, Belgium; ⁴²Department of Microbiology, Babcock University, Ilishan Remo, Ogun State, Nigeria; ⁴³The Key Laboratory for Silviculture and Conservation of Ministry of Education, Beijing Forestry University, Beijing, 100083, China; ⁴⁴Laboratorio de Micología Clínica, Hospital de Clínicas, Universidad de Buenos Aires, Buenos Aires, Argentina; ⁴⁵Facultad de Farmacia y Bioquímica, Universidad de Buenos Aires, Buenos Aires, Argentina; ⁴⁶Royal Botanic Gardens, Kew, Richmond, Surrey, TW9 3DS, UK; ⁴⁷Laboratorio de Salud de Bosques y Ecosistemas, Instituto de Conservación, Biodiversidad y Territorio, Facultad de Ciencias Forestales y Recursos Naturales, Universidad Austral de Chile, casilla 567, Valdivia, Chile; ⁴⁸Institute of Grapevine and Wine Sciences (ICVV), Spanish National Research Council (CSIC)-University of La Rioja-Government of La Rioja, Logroño, 26007, Spain; ⁴⁹Institut für Biologie, Karl-Franzens-Universität Graz, Holteigasse 6, 8010, Graz, Austria; ⁵⁰Applied genomics research group, Universidad de los Andes, Cr 1 # 18 a 12, Bogotá, Colombia; ⁵¹Center for Safe and Improved Food, Scotland's Rural College (SRUC), Kings Buildings, West Mains Road, Edinburgh, EH9 3JG, UK; ⁵²Biorefining and Advanced Materials Research Center, Scotland's Rural College (SRUC), Kings Buildings, West Mains Road, Edinburgh, EH9 3JG, UK; ⁵³Department of Agricultural, Forestry and Food Sciences (DISAFA), University of Torino, Largo P. Braccini 2, 10095, Grugliasco, TO, Italy; ⁵⁴BioAware, Hannut, Belgium; ⁵⁵Research Group Mycology, Department of Biology, Ghent University, 35 K.L. Ledeganckstraat, 9000, Ghent, Belgium; ⁵⁶Faculty of Science, University of South Bohemia, Branišovská 31, 370 05, České Budějovice, Czech Republic; ⁵⁷Department of Botany, Swedish Museum of Natural History, P.O. Box 50007, SE-104 05, Stockholm, Sweden; ⁵⁸Microbe Division/Japan Collection of Microorganisms RIKEN BioResource Research Center, 3-1-1 Koyadai, Tsukuba, Ibaraki, 305-0074, Japan; ⁵⁹Department of Botany, Charles University in Prague,

Prague, Czech Republic; ⁶⁰Center of Excellence in Fungal Research, Mae Fah Luang University, Chaing Rai, 57100, Thailand; ⁶¹Cornell University, 334 Plant Science Building, Ithaca, NY, 14850, USA; ⁶²Department of Health Sciences, Faculty of Medicine and Health Sciences, University of Mauritius, Reduit, Mauritius; ⁶³Manaaki Whenua Landcare Research, Private Bag 92170, Auckland, 1142, New Zealand; ⁶⁴EMSL Analytical, Inc., 200 Route 130 North, Cinnaminson, NJ, 08077, USA; ⁶⁵Department of Nutrition and Dietetics, Faculty of Health Sciences, Yeditepe University, Turkey; ⁶⁶Department of Plant and Soil Sciences, University of Pretoria, P. Bag X20 Hatfield, Pretoria, 0002, South Africa; ⁶⁷Institute of Environmental Biology, Ecology and Biodiversity, Utrecht University, 3584 CH, Utrecht, the Netherlands; ⁶⁸Laboratory for Biological Diversity, Ruder Bošković Institute, Bijenička cesta 54, HR-10000, Zagreb, Croatia; ⁶⁹University of Veterinary Medicine, Vienna (VetMed), Institute of Food Safety, Food Technology and Veterinary Public Health, Veterinärplatz 1, 1210 Vienna and BiMM – Bioactive Microbial Metabolites group, 3430 Tulln a.d. Donau, Austria; ⁷⁰University of California, Davis, One Shields Ave., Davis, CA, 95616, USA; ⁷¹Department of Agricultural Biological Chemistry, College of Agriculture & Life Sciences, Chonnam National University, Yongbong-Dong 300, Buk-Gu, Gwangju, 61186, South Korea; ⁷²Ascofrance, 64 route de Chizé, 79360, Villiers-en-Bois, France; ⁷³The Key Laboratory of Molecular Biology of Crop Pathogens and Insects of Ministry of Agriculture, The Key Laboratory of Biology of Crop Pathogens and Insects of Zhejiang Province, Institute of Biotechnology, Zhejiang University, 866 Yuhangtang Road, Hangzhou, 310058, China; ⁷⁴V.N. Sukachev Institute of Forest SB RAS, Laboratory of Reforestation, Mycology and Plant Pathology, Krasnoyarsk, 660036, Russia; ⁷⁵Reshetnev Siberian State University of Science and Technology, Department of Chemical Technology of Wood and Biotechnology, Krasnoyarsk, 660037, Russia; ⁷⁶School of Life Science and Technology, University of Electronic Science and Technology of China, Chengdu, 611731, China; ⁷⁷Queensland Alliance for Agriculture and Food Innovation, The University of Queensland, Ecosciences Precinct, G.P.O. Box 267, Brisbane, 4001, Australia; ⁷⁸Department of Botany, Faculty of Science, Palacký University Olomouc, Slechtitelů 27, CZ-783 71, Olomouc, Czech Republic; ⁷⁹Department of Agricultural, Food, Environmental and Forestry Science and Technology (DAGRI), Plant Pathology and Entomology section, University of Florence, Ple delle Cascine 28, 50144, Firenze, Italy; ⁸⁰Graduate school of Bioresources, Mie University, Kurima-machiya 1577, Tsu, Mie, 514-8507, Japan; ⁸¹Gothenburg Global Biodiversity Center at the Department of Biological and Environmental Sciences, University of Gothenburg, Box 461, 405 30, Gothenburg, Sweden; ⁸²Department of Microbiology and Biochemistry, Faculty of Natural and Life Sciences, University of Batna 2, Batna, 05000, Algeria; ⁸³Laboratorio de Micodiversidad y Micoprospcción, PROIMI-CONICET, Av. Belgrano y Pje. Caseros, Argentina; ⁸⁴Universidade de Lisboa, Faculdade de Ciências, Biosystems and Integrative Sciences Institute (BioISI), Campo Grande, 1749-016, Lisbon, Portugal; ⁸⁵Microbial Screening Technologies, 28 Percival Rd, Smithfield, NSW, 2164, Australia; ⁸⁶Dipartimento di Agricoltura, Alimentazione e Ambiente, sez. Patologia vegetale, University of Catania, Via S. Sofia 100, 95123 Catania, Italy; ⁸⁷Phytopathology, Van Zanten Breeding B.V., Lavendelweg 15, 1435 EW, Rijssenhou, the Netherlands; ⁸⁸National Fungal Culture Collection of India (NFCCI), Biodiversity and Palaeobiology (Fungi) Group, Agharkar Research Institute, Pune, Maharashtra, 411 004, India; ⁸⁹Laboratory of Mycology and Phytopathology – (LAMFU), Department of Chemical and Food Engineering, Universidad de los Andes, Cr 1 # 18 a 12, Bogotá, Colombia; ⁹⁰Plant Pathology and Population Genetics, Laboratory of Microorganisms, National Gene Bank, Tunisia; ⁹¹Laboratory of Emerging Fungal Pathogens, Department of Microbiology, Immunology, and Parasitology, Discipline of Cellular Biology, Federal University of São Paulo (UNIFESP), São Paulo, 04023062, Brazil; ⁹²USDA-ARS Mycology & Nematology Genetic Diversity & Biology Laboratory, Bldg. 010A, Rm. 212, BARC-West, 10300 Baltimore Ave, Beltsville, MD, 20705, USA; ⁹³Departamento de Micologia Prof. Chaves Batista, Universidade Federal de Pernambuco, Centro de Biociências, Cidade Universitária, Av. Prof. Moraes Rego, s/n, Recife, PE, CEP: 50670-901, Brazil; ⁹⁴Centre for Crop Health, University of Southern Queensland, Toowoomba, 4350, Queensland, Australia; ⁹⁵College of Plant Protection, Northwest A&F University, Yangling, Shaanxi, China; ⁹⁶Faculty of Natural and Agricultural Sciences, Department of Plant Sciences, University of the Free State, P.O. Box 339, Bloemfontein, 9300, South Africa; ⁹⁷Queensland Plant Pathology Herbarium, Department of Agriculture and Fisheries, Dutton Park, Queensland, 4102, Australia; ⁹⁸Royal Botanic Garden Edinburgh, 20A Inverleith Row, Edinburgh, EH3 5LR, United Kingdom; ⁹⁹Faculty of Veterinary and Agricultural Sciences, The University of Melbourne, Parkville, VIC, 3010, Australia; ¹⁰⁰Food and Wine Research Institute, Eszterházy Károly University, 6 Leányka Street, H-3300, Eger, Hungary; ¹⁰¹Department of Life Sciences and Systems Biology, University of Torino and Institute for Sustainable Plant Protection (IPSP-SS Turin), C.N.R., Viale P.A. Mattioli, 25, I-10125, Torino, Italy; ¹⁰²Center for Yunnan Plateau Biological Resources Protection and Utilization, College of Biological Resource and Food Engineering, Qujing Normal University, Qujing, Yunnan, 655011, China; ¹⁰³Shandong Provincial Key Laboratory for Biology of Vegetable Diseases and Insect Pests, College of Plant Protection, Shandong Agricultural University, Tai'an, 271018, China; ¹⁰⁴Fitosanidad, Colegio de Postgraduados-Campus Montecillo, Montecillo-Texcoco, 56230 Edo. de Mexico, Mexico; ¹⁰⁵Leibniz Institute DSMZ-German Collection of Microorganisms and Cell Cultures GmbH, Inhoffenstrasse 7 B, 38124, Braunschweig, Germany; ¹⁰⁶Museum of Evolution, Uppsala University, Norbyvägen 16, SE-752 36, Uppsala, Sweden; ¹⁰⁷Ministry of Agriculture Key Laboratory of Molecular Biology of Crop Pathogens and Insects, Institute of Biotechnology, College of Agriculture and Biotechnology, Zhejiang University, No. 866 Yuhangtang Road, Hangzhou, 310058, China; ¹⁰⁸Goethe-University Frankfurt am Main, Department of Biological Sciences, Institute of Ecology, Evolution and Diversity, Max-von-Laue Str. 13, D-60438, Frankfurt am Main, Germany; ¹⁰⁹LOEWE Centre for Translational Biodiversity Genomics, Georg-Voigt-Str. 14-16, D-60325, Frankfurt am Main, Germany

*Correspondence: L. Lombard, l.lombard@wi.knaw.nl; M. Sandoval-Denis, m.sandoval@wi.knaw.nl

Abstract: Recent publications have argued that there are potentially serious consequences for researchers in recognising distinct genera in the terminal fusarioid clade of the family *Nectriaceae*. Thus, an alternate hypothesis, namely a very broad concept of the genus *Fusarium* was proposed. In doing so, however, a significant body of data that supports distinct genera in *Nectriaceae* based on morphology, biology, and phylogeny is disregarded. A DNA phylogeny based on 19 orthologous protein-coding genes was presented to support a very broad concept of *Fusarium* at the F1 node in *Nectriaceae*. Here, we demonstrate that re-analyses of this dataset show that all 19 genes support the F3 node that represents *Fusarium sensu stricto* as defined by *F. sambucinum* (sexual morph synonym *Gibberella pulicaris*). The backbone of the phylogeny is resolved by the concatenated alignment, but only six of the 19 genes fully support the F1 node, representing the broad circumscription of *Fusarium*. Furthermore, a re-analysis of the concatenated dataset revealed alternate topologies in different phylogenetic algorithms, highlighting the deep divergence and unresolved placement of various *Nectriaceae* lineages proposed as members of *Fusarium*. Species of *Fusarium s. str.* are characterised by *Gibberella* sexual morphs, asexual morphs with thin- or thick-walled macroconidia that have variously shaped apical and basal cells, and trichothecene mycotoxin production, which separates them from other fusarioid genera. Here we show that the Wollenweber concept of *Fusarium* presently accounts for 20 segregate genera with clear-cut synapomorphic traits, and that fusarioid macroconidia represent a character that has been gained or lost multiple times throughout *Nectriaceae*. Thus, the very broad circumscription of *Fusarium* is blurry and without apparent synapomorphies, and does not include all genera with fusarium-like macroconidia, which are spread throughout *Nectriaceae* (e.g., *Cosmospora*, *Macroconia*, *Microcera*). In this study four new genera are introduced, along with 18 new species and 16 new combinations. These names convey information about relationships, morphology, and ecological preference that would otherwise be lost in a broader definition of *Fusarium*. To assist users to correctly identify fusarioid genera and species, we introduce a new online identification database, Fusarioid-ID, accessible at www.fusarium.org. The database comprises partial sequences from multiple genes commonly used to identify fusarioid taxa (*act1*, *CaM*, *his3*, *rpb1*, *rpb2*, *tef1*, *tub2*, ITS, and LSU). In this paper, we also present a nomenclator of names that have been introduced in *Fusarium* up to January 2021 as well as their current status, types, and diagnostic DNA barcode data. In this study, researchers from 46 countries, representing taxonomists, plant pathologists, medical mycologists, quarantine officials, regulatory agencies, and students, strongly support the application and use of a more precisely delimited *Fusarium* (= *Gibberella*) concept to accommodate taxa from the robust monophyletic node F3 on the basis of a well-defined and unique combination of morphological and biochemical features. This F3 node includes, among others, species of the *F. fujikuroi*, *F. incarnatum-equiseti*, *F. oxysporum*, and *F. sambucinum* species complexes, but not species of *Bisifusarium* [*F. dimerum* species complex (SC)], *Cyanonectria* [*F. buxicola* SC], *Geejayessia* [*F. staphyleae* SC], *Neocosmospora* [*F. solani* SC] or *Rectifusarium* [*F. ventricosum* SC]. The present study represents the first step to generating a new online monograph of *Fusarium* and allied fusarioid genera (www.fusarium.org).

Key words: Multi-gene phylogeny, Mycotoxins, *Nectriaceae*, *Neocosmospora*, Novel taxa, Pathogen, Taxonomy.

Taxonomic novelties: New genera: *Luteonectria* Sand.-Den., L. Lombard, Schroers & Rossman, *Nothofusarium* Crous, Sand.-Den. & L. Lombard, *Scolecopusarium* L. Lombard, Sand.-Den. & Crous, *Setofusarium* (Nirenberg & Samuels) Crous & Sand.-Den.; **New species:** *Fusarium echinatum* Sand.-Den. & G.J. Marais, *Fusarium lyamte* J.L. Walsh, Sangal., L.W. Burgess, E.C.Y. Liew & Summerell, *Fusarium palustre* W.H. Elmer & Marra, *Fusarium prieskaense* G.J. Marais & Sand.-Den., *Fusarium werrikimbe* J.L. Walsh, L.W. Burgess, E.C.Y. Liew & B.A. Summerell, *Fusicolla quarantena* J.D.P. Bezerra, Sand.-Den., Crous & Souza-Motta, *Fusicolla meniscoidea* L. Lombard & Sand.-Den., *Fusicolla sporellula* Sand.-Den. & L. Lombard, *Macroconia bulbipes* Crous & Sand.-Den., *Macroconia phlogioides* Sand.-Den. & Crous, *Neocosmospora epipeda* Quaedvl. & Sand.-Den., *Neocosmospora merckiana* Quaedvl. & Sand.-Den., *Neocosmospora neerlandica* Crous & Sand.-Den., *Neocosmospora nelsonii* Crous & Sand.-Den., *Neocosmospora pseudopisi* Sand.-Den. & L. Lombard, *Nothofusarium devonianum* L. Lombard, Crous & Sand.-Den., *Stylonectria comiculata* Gräfenhan, Crous & Sand.-Den., *Stylonectria hetmanica* Akulov, Crous & Sand.-Den.; **New combinations:** *Apiognomonium platani* (Lév.) L. Lombard, *Calloria tremelloides* (Grev.) L. Lombard, *Cosmosporella cavisperma* (Corda) Sand.-Den., L. Lombard & Crous, *Cylindrodendrum orthosporum* (Sacc. & P. Syd.) L. Lombard, *Dialonectria volutella* (Ellis & Everh.) L. Lombard & Sand.-Den., *Fusarium armeniacum* (G.A. Forbes et al.) L.W. Burgess & Summerell, *Hymenella aurea* (Corda) L. Lombard, *Hymenella spermogoniopsis* (Jul. Müll.) L. Lombard & Sand.-Den., *Luteonectria albida* (Rossman) Sand.-Den. & L. Lombard, *Luteonectria nematophila* (Nirenberg & Hagedorn) Sand.-Den. & L. Lombard, *Neocosmospora floridana* (T. Aoki et al.) L. Lombard & Sand.-Den., *Neocosmospora obliquiseptata* (T. Aoki et al.) L. Lombard & Sand.-Den., *Neocosmospora rekana* (Lynn & Marinc.) L. Lombard & Sand.-Den., *Neocosmospora tuaranensis* (T. Aoki et al.) L. Lombard & Sand.-Den., *Scolecopusarium ciliatum* (Link) L. Lombard, Sand.-Den. & Crous, *Setofusarium setosum* (Samuels & Nirenberg) Sand.-Den. & Crous.; **Epitypes (basionyms):** *Fusarium buharicum* Jacz. ex Babajan & Teterev.-Babajan, *Fusarium cavispermum* Corda, *Fusarium flocciferum* Corda, *Fusarium graminearum* Schwabe, *Fusarium heterosporum* Nees & T. Nees, *Fusarium redolens* Wollenw., *Fusarium reticulatum* Mont., *Fusarium scirpi* Lambotte & Fautrey, *Fusarium stilboides* Wollenw., *Fusarium xylarioides* Steyaert, *Fusisporium culmorum* Wm.G. Sm., *Fusisporium incarnatum* Roberge ex Desm., *Selenosporium equiseti* Corda, *Sphaeria sanguinea* var. *cicatricum* Berk., *Sporotrichum poae* Peck.; **Lectotypes (basionyms):** *Atractium pallidum* Bonord., *Cephalosporium sacchari* E.J. Butler, *Fusarium aeruginosum* Delacr., *Fusarium agaricorum* Sarrazin, *Fusarium albidoviolaceum* Dasz., *Fusarium aleyrodis* Petch, *Fusarium amentorum* Lacroix, *Fusarium annuum* Leonian, *Fusarium arcuatum* Berk. & M.A. Curtis, *Fusarium aridum* O.A. Pratt, *Fusarium arthrosporioides* Sherb., *Fusarium asparagi* Delacr., *Fusarium batatas* Wollenw., *Fusarium bifforme* Sherb., *Fusarium hactacearum* Pasin. & Buzz.-Trav., *Fusarium cacti-maxonii* Pasin. & Buzz.-Trav., *Fusarium caudatum* Wollenw., *Fusarium cavispermum* Corda, *Fusarium cepae* Hanzawa, *Fusarium cesatii* Rabenh., *Fusarium citrifforme* Jamal., *Fusarium citrinum* Wollenw., *Fusarium citrulli* Taubenh., *Fusarium clavatum* Sherb., *Fusarium coccinellum* Kalchbr., *Fusarium cromyophthoron* Sideris, *Fusarium cucurbitae* Taubenh., *Fusarium cuneiforme* Sherb., *Fusarium delacroixii* Sacc., *Fusarium dimerum* var. *nectrioides* Wollenw., *Fusarium epicoccum* McAlpine, *Fusarium eucheliae* Sartory, R. Sartory & J. Mey., *Fusarium fissum* Peyl, *Fusarium flocciferum* Corda, *Fusarium gemmiperda* Aderh., *Fusarium genevense* Dasz., *Fusarium graminearum* Schwabe, *Fusarium graminum* Corda, *Fusarium heterosporioides* Fautrey, *Fusarium heterosporum* Nees & T. Nees, *Fusarium idahoanum* O.A. Pratt, *Fusarium juruanum* Henn., *Fusarium lanceolatum* O.A. Pratt, *Fusarium lateritium* Nees, *Fusarium loncheceras* Sideris, *Fusarium malvacearum* Taubenh., *Fusarium martii* f. *phaseoli* Burkh., *Fusarium muentzii* Delacr., *Fusarium nigrum* O.A. Pratt, *Fusarium oxysporum* var. *asclerotium* Sherb., *Fusarium palczewskii* Jacz., *Fusarium polymorphum* Matr., *Fusarium poolense* Taubenh., *Fusarium prunorum* McAlpine, *Fusarium pusillum* Wollenw., *Fusarium putrefaciens* Osterw., *Fusarium redolens* Wollenw., *Fusarium reticulatum* Mont., *Fusarium rhizochromatistes* Sideris, *Fusarium rhizophilum* Corda, *Fusarium rhodellum* McAlpine, *Fusarium roesleri* Thüm., *Fusarium rostratum* Appel & Wollenw., *Fusarium rubiginosum* Appel & Wollenw., *Fusarium rubrum* Parav., *Fusarium samoense* Gehrm., *Fusarium scirpi* Lambotte & Fautrey, *Fusarium secalis* Jacz., *Fusarium spinaciae* Hungerf., *Fusarium sporotrichioides* Sherb., *Fusarium stercoris* Fuckel, *Fusarium stilboides* Wollenw., *Fusarium stillatum* De Not. ex Sacc., *Fusarium sublnatum* Reinking, *Fusarium succisae* Schröt. ex Sacc., *Fusarium tabacivorum* Delacr., *Fusarium trichothecioides* Wollenw., *Fusarium tritici* Liebman, *Fusarium tuberivorum* Wilcox & G.K. Link, *Fusarium tumidum* var. *humii* Reinking, *Fusarium ustilaginis* Kellerm. & Swingle, *Fusarium viticola* Thüm., *Fusarium willkommii* Lindau, *Fusarium xylarioides* Steyaert, *Fusarium zygopetali* Delacr., *Fusisporium andropogonis* Cooke ex Thüm., *Fusisporium anthophilum* A. Braun, *Fusisporium arundinis* Corda, *Fusisporium clypeaster* Corda, *Fusisporium culmorum* Wm.G. Sm., *Fusisporium didymum* Harting, *Fusisporium elasticae* Thüm., *Fusisporium episphaericum* Cooke & Ellis, *Fusisporium flavidum* Bonord., *Fusisporium hordei* Wm.G. Sm., *Fusisporium incarnatum* Roberge ex Desm., *Fusisporium lolii* Wm.G. Sm., *Fusisporium pandani* Corda, *Gibberella phyllostachydicola* W. Yamam., *Menispora penicillata* Harz, *Selenosporium equiseti* Corda, *Selenosporium hippocastani* Corda, *Selenosporium urticarum* Corda., *Sphaeria sanguinea* var. *cicatricum* Berk.; **Neotypes (basionyms):** *Atractium ciliatum* Link, *Fusarium longipes* Wollenw. & Reinking, *Fusisporium avenaceum* Fr., *Selenosporium sarcochroum* Desm.

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INTRODUCTION

The relevance and impact of *Fusarium* (Ascomycota, Hypocreales, Nectriaceae) to humankind is substantial. Over the past 100 years, it has attracted considerable attention from scientists as the extent of species diversity and the impact on agriculture and human health became clear. After an initial period of discovery and cataloguing by 19th century naturalists, its taxonomy became the target of research from a broad range of scientists, that resulted in the emergence of distinct “schools” that promoted different taxonomic approaches to fusarium-like organisms. With the advent of an objective and reproducible framework for phylogenetic relationships inferred from molecular phylogenetics, it might have been expected that controversies would melt away, and a stable, universally accepted taxonomy of *Fusarium* and its species would emerge, but this does not yet appear to be the case (Fig. 1). However, all scientists working with *Fusarium* desire a stable taxonomic system, and all agree that taxonomic changes should be made with the aim of promoting stability.

Recently, Geiser et al. (2021), largely in response to papers published by Gräfenhan et al. (2011), Schroers et al. (2011), Lombard et al. (2015), and Sandoval-Denis et al. (2019), proposed a cladistic solution to redelimit a generic concept for

Fusarium. The generic treatment of *Fusarium* by Geiser et al. (2013, 2021), produced an ill-delimited genus without clear synapomorphies, as fusarium-like macroconidia are strongly polyphyletic within Nectriaceae and also occur outside their very broadly circumscribed *Fusarium* concept. We argue that a narrower concept of genera with a clear, unique combination of features is needed for the majority of fusarioid species.

Dual nomenclature and consensus on the use of the generic name *Fusarium*

In accordance with the single-name system for fungi, that was adopted at the International Botanical Congress, Melbourne (IBCML) in 2011, we are in full agreement with Geiser et al. (2013, 2021) and O'Donnell et al. (2020) that the name *Fusarium* applies to any genus with a delimitation that includes the conserved lectotype of the type species, *F. sambucinum* (sexual morph synonym *Gibberella pulicaris*), as stated by Rossman et al. (2013). Unfortunately, a single joint paper explaining the choice of this name supported by the entire *Fusarium* community was planned but failed because of the insistence of a subset of authors to adopt a broad generic concept.



Fig. 1. Timeline summarising important events in the taxonomy and nomenclature of *Fusarium* and related taxa.

Taxonomy and nomenclature are different concepts, although they are frequently confused, leading to misinterpretations. Support for dual nomenclature ended at the IBCM in August 2011. The significance of 1 January 2013 was to ensure the formal nomenclatural validity of newly proposed *dual* names (new species or new combinations) that were in press or part of studies about to be submitted for publication. These dates have no significance for names proposed in a single name system, which can be done at any time. Despite these technicalities, virtually all members of the *Fusarium* community accept that *Fusarium* must be used over the sexually-typified name *Gibberella* in the single name system, a recommendation included in the proposed list of Protected Names submitted to the Nomenclature Committee for Fungi, the body with the authority to recommend its formal acceptance (Kirk *et al.* 2013). However, statements in Geiser *et al.* (2013) seem to reflect a confusion about how the nomenclatural decision affected taxonomic concepts.

The name *Fusarium* has never been at risk during the nomenclatural transition, and the community support for its use in a single name system is unanimous. We fully agree with Geiser *et al.* (2013, 2021) and Rossman *et al.* (2013) that *Fusarium* equals *Gibberella*. *Fusarium* will always be applied to the clade that includes the type species of *Fusarium*, *F. sambucinum*, which is the same fungus that also typifies *Gibberella*. In this study, we show that the clade defined as *Fusarium s. str.* (O'Donnell *et al.* 2013, as *Gibberella*; Geiser *et al.* 2013, as Clade B) combines monophyly, morphology of sexual and asexual morphs, and biochemical data in a coherent way that can logically be recognised at the generic rank. Expanding the concept of *Fusarium* to node F1 *sensu* Geiser *et al.* (2013, 2021)

results in the combination of several distinct genera and does not resolve the issue of fusarium-like macroconidia in genera outside their broad circumscription of *Fusarium*.

Phylogenetic structure and distribution of fusarioid asexual morphs in *Nectriaceae* (*Hypocreales*)

Gräfenhan *et al.* (2011) and Schroers *et al.* (2011) presented a phylogenetic overview of selected *Nectriaceae* based on combined analyses of two different genes, namely the commonly employed and phylogenetically informative RNA polymerase II second largest subunit (*rpb2*) and exon regions of the larger subunit of ATP citrate lyase (*ac1*). The two papers were the first to apply a single name system to fusarioid fungi (*i.e.*, genera with fusarium-like macroconidia), and were written along with others (see Rossman & Seifert 2011) to promote discussions that eventually led to changes to the International Code of Nomenclature for algae, fungi, and plants (ICNafp) (Turland *et al.* 2018).

The main focus of the Gräfenhan *et al.* (2011) paper was to deal with extraneous elements that had long been included in *Fusarium*. These fungi had distinct phenotypic characters, such as thin, collapsing perithecial walls, slow growing agar colonies lacking aerial mycelium, or sparsely septate macroconidia. Users of the Gerlach & Nirenberg (1982) and Nelson *et al.* (1983) identification manuals may be familiar with some of these species, then called *Fusarium aquaeductuum*, *F. coccophilum* and *F. merismoides*. There was evidence in the first papers on the molecular phylogeny of *Fusarium* that these species did not belong to *Fusarium* (*e.g.*, see O'Donnell 1993). It was not until

the study by Gräfenhan *et al.* (2011) that other genera in the family, such as members of the *Cylindrocarpon* generic complex (Chaverri *et al.* 2011), *Calonectria* (Liu *et al.* 2020), *Tubercularia* (Hirooka *et al.* 2012), and minor genera such as *Mariannaea*, *Pseudonectria*, and *Volutella* (also see Lombard *et al.* 2015) were adequately sampled to yield generic-level resolution. The phylograms showed the division of fusarioid taxa into two large groups, which Gräfenhan *et al.* (2011) called the Terminal Fusarium Clade (abbreviated TFC by Geiser *et al.* 2013) and the ill-delineated Basal Fusarium Clade (BFC) that contained several of the genera noted above. A single-genus recognition for the BFC was not feasible because of the great morphological, genetic, and ecological divergence among the sampled species. The BFC included seven genera, each with their monophyly strongly supported and more or less ecologically coherent. Species with fusarioid conidia were reclassified in the phylogenetically redefined but previously described genera *Atractium*, *Cosmospora*, *Dialonectria*, *Fusicolla*, *Macroconia*, *Microcera*, and *Stylonectria* (Gräfenhan *et al.* 2011, Schroers *et al.* 2011). Geiser *et al.* (2013) accepted these segregate genera in the BFC as distinct from the TFC, while correctly pointing out the weak support values obtained for the phylogenetic backbone of the tree. One consequence of the widespread occurrence of macroconidia in the taxon sampling (fusarioid genera, cylindrocarpon-like genera, and *Calonectria*) was the suggestion that especially the fusarioid macroconidium is a plesiomorphic character (that is, an ancestral character) and had been lost in some lineages in *Nectriaceae* (Gräfenhan *et al.* 2011).

The second paper by Schroers *et al.* (2011) recovered similar phylogenies as Gräfenhan *et al.* (2011), but focused on the TFC, supplementing this with a five-gene analysis of a particular subclade within the TFC intended to delimit phylogenetic genera and a few species. This demonstrated the monophyly of the treated genera and resulted in the acceptance of the previously described *Cyanonectria* (Samuels *et al.* 2009), as well as the description of the genus *Geejayessia*. Again, Geiser *et al.* (2013) correctly criticised the weakness of the backbone of the tree, especially in the BFC. About 75 % of the phylogenetic signal in the analysis came from one gene, *rpb2*. Schroers *et al.* (2011) did not discuss the taxonomic fate of *Neocosmospora* (the *Fusarium solani* species complex, FSSC), which was represented by only two species in their analysis, but was excluded from *Fusarium s. str.*

The call for more genetic markers and even genome analysis by Geiser *et al.* (2013), to better resolve the phylogenetic backbone of the TFC was justified, but the increased number of markers should have been matched by increased taxon sampling of all known genera of *Nectriaceae*, as taxon sampling is equally important for inferring robust and meaningful phylogenies (Zwickl & Hillis 2002, Heath *et al.* 2008).

Lombard *et al.* (2015) greatly expanded both the number of genetic markers and the taxon sampling in order to explore the generic boundaries across the *Nectriaceae*, including all genera known from culture and many genera for which no DNA data was previously available. A 10-gene phylogeny was inferred including all the markers previously used by Gräfenhan *et al.* (2011), Schroers *et al.* (2011), Geiser *et al.* (2013), and O'Donnell *et al.* (2013), plus nrDNA sequences and other markers of known phylogenetic utility, namely actin (*act1*), beta-tubulin (*tub2*), calmodulin (*CaM*), histone (*his3*), and the translation elongation factor 1- α (*tef1*). From this, a phylogeny of the TFC overall congruent to that presented by Gräfenhan *et al.* (2011) and

Geiser *et al.* (2013) was obtained. Importantly, the monophyly of *Albonectria*, *Cyanonectria*, *Geejayessia*, *Fusarium*, and *Neocosmospora* was reaffirmed and a few early diverging lineages previously included in the TFC were segregated into new fusarioid genera *i.e.*, *Bisifusarium* (formerly the *F. dimerum* species complex) and *Rectifusarium* (formerly the *F. ventricosum* species complex) (Lombard *et al.* 2015).

After nearly a hundred years of quandary, a modern revision was published for *Neocosmospora* (Sandoval-Denis *et al.* 2019). In this study, many unnamed phylogenetic species were morphologically characterised and given Latin binomials, while old names were resurrected, epitypified, and linked to DNA barcodes.

Two recent publications by O'Donnell *et al.* (2020) and Geiser *et al.* (2021) argued for the broad *Fusarium* concept of Geiser *et al.* (2013). Both papers present very similar phylogenetic analyses, relying on 19 genes, including 12 newly sampled markers, namely: cytochrome P450 reductase (*cpr1*), ATP-dependent DNA helicase II (*ku70*), sphinganine palmitoyl transferase subunit 2 (*lcb2*), DNA replication licensing factor (*mcm7*), phosphoglycerate kinase (*pgk1*), topoisomerase (*top1*), two subunits each of the DNA polymerase (*dpa1* and *dpe1*), the fatty acid synthase (*fas1*, *fas2*), alpha-tubulin (*tub1*), and *tub2*. The previously employed marker *his3* was not included, nor were nrDNA markers. The results are in essence the same as those of the previously published phylogenies, but with stronger support for the backbone in the combined analyses (see Cummings & Meyer 2005). Geiser *et al.* (2021) claimed that the F1 node was supported by 12, and the F2 node by 14 of the individual genes, but did not mention that all 19 genes supported the F3 node (*Fusarium s. str.* = the *Gibberella* clade).

In this study we re-investigated the Geiser *et al.* (2021) dataset using several different high-resolution phylogenetic approaches, and we found that their evaluations of concordance were based on an inadequate interpretation of Ultra-Fast bootstrap results (only values ≥ 95 % are to be deemed significant, see Minh *et al.* 2013, Hoang *et al.* 2018). In addition to the topological incongruences among six genes (*act1*, *CaM*, DNA polymerase epsilon subunit *dpe1*, *ku70*, *pgk1*, *tef1*, and *tub2*), only six and 11 genes actually support the F1 and F2 nodes, respectively, while all 19 genes support the F3 node. The low internode certainty (IC) and IC All (ICA) values obtained for F1 (0.19 and 0.33, respectively) were misinterpreted by Geiser *et al.* (2021) as IC values close to 0 indicate conflict between the partitions (Salichos *et al.* 2014). The F3 node was well supported with IC and ICA values at 1 (Geiser *et al.* 2021, Supplementary Table. S1), which indicates the absence of conflict.

While the effort by O'Donnell *et al.* (2020) and Geiser *et al.* (2021) to include a high diversity of DNA markers is commendable, it is undermined by an imbalanced selection of taxa for their analyses. Specifically, there is a marked overrepresentation of node F1 species, while sampling and taxon selection across the *Nectriaceae* is almost absent. Excluding any of the major genus-level clades, especially those relevant to the recognition of *Bisifusarium*, *Neocosmospora* and *Rectifusarium*, introduces taxon sampling biases in a way that reduce the reliability of phylogenetic inferences and support values with respect to the backbone of the *Nectriaceae*. Furthermore, neither O'Donnell *et al.* (2020) nor Geiser *et al.* (2021) give full consideration to morphological and ecological evidence. In principle, a genus should always be delimited as monophyletic, supported by derived traits. In addition, its circumscription should

depend on the systematic (phylogenetic and biological) structure of the family it belongs to, in this case, the *Nectriaceae*.

Phylogenetics has rapidly advanced from a powerful adjunct tool for understanding evolutionary relationships to the dominant principle for classification, especially for delimitation of taxa at all ranks. However, the resulting analyses and phylogenies are compromised if they are not reconciled with other biological data. The call for additional genomic data in the *Fusarium* clade (Geiser et al. 2013, Aoki et al. 2019) may improve backbone node support values, but the phylogenetic structure is unlikely to change; it is the translation of that data into practicable taxonomy. The broad *Fusarium* concept of Aoki et al. (2019), O'Donnell et al. (2020) and Geiser et al. (2021) is phylogenetically possible, but it does not offer a generic definition based on a combination of available genetic, morphological, biochemical and ecological data. It is, thus, impractical in that it is so broad that the genus would not have any synapomorphies when compared to other genera of the *Nectriaceae* outside their broad circumscription of *Fusarium*.

The arguments presented by Aoki et al. (2019), O'Donnell et al. (2020) and Geiser et al. (2021) are centred around the phylogenetic support of some nodes, which have never been a key subject of the discussion, as the made observations generally match the interpretations made by many authors. While the very broad circumscription of *Fusarium* reflects as a monophyletic group in DNA phylogenetic analyses, the TFC is a conglomerate of several monophyletic genera that has a common ancestor (node F1 in Geiser et al. 2013). Each of these genera has a distinctive combination of morphological features. An analogous situation was observed in the monophyletic sister clade that was originally classified as *Cylindrocarpon s. lat.*, but that is currently viewed as composed of several monophyletic genera i.e., *Cinnamomeonectria*, *Corinectria*, *Cylindrodendrum*, *Dactyloectria*, *Ilyonectria*, *Macronectria*, *Neonectria*, *Pleioctarpon*, *Rugonectria*, *Thelonectria* and *Tumenectria* (Chaverri et al. 2011, Gräfenhan et al. 2011, Lombard et al. 2014, Salgado-Salazar et al. 2016, González & Chaverri 2017).

What is a genus?

Taxonomically, a genus is a group that is defined by a type species, and that often includes additional species considered to belong to the same group (Vellinga et al. 2015). The observations or category of data involved in delineating genera have varied over time, and in many cases, the characters used to delimit well accepted genera have proven to be homoplasious and the genera polyphyletic (Crous et al. 2009). However, it is a fundamental principle that taxonomic entities should reflect evolutionary relationships.

This has led to inevitable splitting of well-known fungal taxa, both genera and species, into smaller groups, but sometimes also genera were merged with others based on the reappraisal or discovery of derived characters (e.g., Voglmayr & Thines 2007). This proceeds with each technological revolution providing ever deeper insight into the biological/evolutionary relationships of organisms, and has accelerated again since molecular phylogenetics came into widespread use. There is a prevailing notion that nature made species, but that humans made all other taxonomic ranks for their own convenience. However, it is increasingly recognised that all taxonomic ranks, including the species level, do not have solid boundaries but are more like a steam cloud with fuzzy margins. At the genus level, these

boundaries are often even more obscure, but is a genus just an arbitrary (but statistically well-supported) monophyletic convenience, a consensus accepted by a self-appointed committee? Or is a genus a meaningful, definable unit resulting from evolutionary processes, which can be recognised by patterns of biological structure, biochemistry, behaviour, and adaptation to specific niches? We believe that the latter should be the case. While we recognise that generic delimitations will always depend on a subjective choice, we believe that generic concepts should always be guided in a phylogenetic context by morphological, biochemical, or ecological characters that can both be used for practical recognition and convey evolutionary information.

The generic concept for *Fusarium* proposed by Geiser et al. (2013, 2021) is a rejection of this concept, as it merges lineages with divergent characters that were accepted and applied not only throughout the family *Nectriaceae* for the delimitation of genera but also in other fungal families and orders. The very broad genus *Fusarium* that it gives rise to does not have clear-cut features, as the diversity of characters shared with the rest of the *Nectriaceae* is so high that it could be extended almost arbitrarily to the entire family. It would, in fact be as if the concept of cryptic species was expanded to genera, that is, genera that can only be recognised as a well-supported node on a phylogram, which is, in our view, in disagreement with fundamental principles of practical classification. The node F1 selected by Geiser et al. (2013, 2021) for defining *Fusarium* is devoid of phenotypic support and includes several genera with distinct evolutionary traits. Indeed, the Geiser et al. (2013, 2021) concept of *Fusarium* is strictly phylogenetically defined and essentially amounts to a list of the species bound within a selected clade. Their morphological circumscription does not admit the existence of synapomorphies (i.e., unique diagnostic characters possessed by all included species), and it extends beyond their chosen node to other groups in *Nectriaceae*. In this very wide definition of *Fusarium*, phenotypic characters and ecological patterns that correlate with well-supported monophyletic groups within the larger, poorly supported TFC are disregarded as basis for generic delineation.

Admittedly, phenotypic characters in the TFC are tricky to interpret. The fusarioid macroconidium with or without a well-developed foot-shaped basal cell (i.e., basal conidial cell showing an asymmetrical papillum, delimited from the rest of the cell and forming a distinct notch) occurs in the majority but not all of the species in the traditional generic concept, but is also a feature present in a significant proportion of other members of the *Nectriaceae*, or even of the unrelated genus *Microdochium* (*Amphisphaeriaceae*). It is, therefore, not a unique feature for generic delineation (Gräfenhan et al. 2011).

Perithecial pigmentation has been used to delimit genera in *Nectriaceae*. The orange/red perithecium is an ancestral character in the family and common also to members of the BFC and early diverging lineages of the TFC, including all *Neocosmospora* species known to reproduce sexually, *Setofusarium*, and some species of *Cyanonectria* and *Geejayessia*. These structures are easily distinguished from the homogeneously bluish/black perithecia of true *Fusarium s. str.* species in the *Gibberella* clade sensu O'Donnell et al. (2013). Contrary to what was suggested by Geiser et al. (2021), it is not *Neocosmospora* which represents an interesting but morphologically aberrant lineage, since neither its type nor the members of its modern morphological circumscription (Nalim et al. 2011) exhibit aberrant characteristics. It is the dark-coloured perithecia typical of

Fusarium s. str. (= *Gibberella* clade) that are aberrant and unusual within *Nectriaceae*.

The dark purple to black perithecium formerly used to characterise *Fusarium s. str.* (= *Gibberella*), represents a synapomorphic state. Ascospores with similar colours have evolved independently in some, but not all, species of *Geejayessia*, while heterogeneously coloured bluish black or bicoloured perithecia can be observed in several species of *Cyanonectria*, which often appears as a sister genus to *Fusarium*. However, *Cyanonectria* and *Geejayessia* differ from *Fusarium* and *Neocosmospora* by their typically well-developed stromata as well as their thinner and smooth perithecial walls. Notably, pale yellowish perithecia occur in several clades and are a derived character as well, and one genus that we accept, *Albonectria*, was initially defined by white perithecia (Rossman *et al.* 1999). Also, in terms of its ascospores, *Fusarium* shows a derived state. With the exception of *Albonectria*, which includes species with hyaline, ellipsoidal to fusoid, 3-septate, smooth to finely striated ascospores, the genera mentioned above present mostly pale yellow-brown ascospores. Ascospores of *Fusarium s. str.* are more often subhyaline, ellipsoidal to fusoid, 1–3-septate, and smooth-walled when viewed with light microscopy. Ascospores of *Neocosmospora* are easily distinguished from those of *Fusarium* by being ovoid to ellipsoidal, (0–)1-septate, pigmented, conspicuously striate or more rarely cerebriform or spinulose. It is worth noting that most of the above-mentioned characters and differences are the same applied to define genera across the whole *Nectriaceae* (Rossman *et al.* 1999, Lombard *et al.* 2015), where they correlate well with phylogenetic inferences. Ascospores showing similarly many septa as in *Fusarium s. str.* have independently evolved in *Nectria diploa* (now *Microcera*), as well as in *N. glabra*, and *N. decora* (now *Flammocladiella*). The fact that none of these species is a member of the TFC supports the interpretation that multiseptate ascospores might be apomorphic for *Fusarium s. str.*, separating it clearly from other phylogenetically related genera.

Behaviour and other adaptations, determine how an organism operates and survives in nature and are the ultimate determinants and products of natural selection. They may be difficult to translate into nodes and other results of phylogenetic analyses such as phylogenetic distance. Despite this, similarities in adaptive traits are frequently used to calibrate phylogenetic delimitations of genera. For example, all known species of *Microcera* are pathogens of scale insects. It is easy to understand the hypothesis that the ancestor of this clade jumped to these hosts, followed by subsequent radiation and speciation (Thines 2019). This resulted in considerable micromorphological diversity, while a core of adaptation resulting from the parasitic life style remained conserved. Similarly, several of the genus-level clades include mostly mycoparasitic species or pathogens of plants. If we apply this kind of thinking to the well-supported clades of the TFC, as noted by Schroers *et al.* (2011), species of *Cyanonectria* and *Geejayessia* occur only on woody hosts (mostly species of *Buxus*, *Celtis* and *Staphylea*) and would typically not occur as soil-borne plant pathogens or pathogens of grasses. They are also not known to produce trichothecene mycotoxins. This is in stark contrast with the prevailing ecological concept of *Fusarium s. str.* as a genus of primarily soil-borne fungi, of which many are in a firm biological association with grasses and herbs. Importantly, the vast majority of *Fusarium s. str.* species produce trichothecene mycotoxins as a chemical synapomorphy. Most of the strongly supported clades within the

TFC can be supported by these kinds of morphological, chemical, and biological traits, allowing the possibility of non-arbitrary recognition of biologically meaningful genera. One such clade is *Neocosmospora*.

Arguments for and the practicality of recognising *Neocosmospora* (the *F. solani* species complex) as a genus

In the days of dual nomenclature, the distinction between the red perithecia of *Neocosmospora*, as amended by Nalim *et al.* (2011), and the typically purple or blackish perithecia of the trichothecene-producing *Gibberella* species was generally accepted by *Fusarium* taxonomists. The ecological distinctiveness of *Neocosmospora* as a group of soil fungi, often associated with roots and causing root rot and vascular wilt diseases, was also generally acknowledged. In addition to the dissimilar sexual characters mentioned above, the asexual morphs of this group are also distinctive. The macroconidia are usually thick-walled, with blunt, rounded apical cells, and they usually have inconspicuous foot-shaped basal cells. Microconidia are produced on very long, narrow phialides. Cultures of a vast majority of species of this group can easily be recognised morphologically, even with a dissecting microscope.

The ecological similarities of the members of *Neocosmospora* with *F. oxysporum* have to be acknowledged, as noted by Geiser *et al.* (2013, 2021). However, these two groups of species are morphologically distinct, even as asexual morphs. *Fusarium oxysporum* produces macroconidia with acutely pointed apical cells, and microconidia from phialides that are usually 5–10 times shorter than those of *Neocosmospora* species.

Geiser *et al.* (2013, 2021) have pointed out that microchromosomes or conditionally dispensable chromosomes occur in *Neocosmospora* and members of their F3 clade, namely *F. oxysporum*. Microchromosomes have been observed, however, also in phylogenetically distinct taxa such as *Magnaporthe oryzae* (Yoshida *et al.* 2009, now *Pyricularia oryzae*), *Mycosphaerella graminicola* (Stukenbrock *et al.* 2010, now *Zymoseptoria tritici*), and *Alternaria arborescens* (Hu *et al.* 2012) and might occur sporadically as a result of horizontal gene transfer. They are thought to increase the ability of a pathogen to adapt to the host's defence mechanisms. The ability to acquire conditionally dispensable chromosomes might thus be seen as a general genetic tool allowing organisms to gain ecologically advantageous genes. Similarly, they could present a general driving force in co-evolutionary processes, but the per se occurrence of conditionally dispensable chromosomes in two taxa can hardly be used as a criterion for drawing conclusions on or imply generic relatedness.

In the Nelson *et al.* (1983) manual and in one of the last vestiges of the ultra-reductionist Snyder & Hansen (1941) system, *F. solani* was recognised as the only species of section *Martiella*, even though the existence of several distinct mating populations was known. The European system (exemplified by Gerlach & Nirenberg 1982) accepted several more species, derived from the classic Wollenweber & Reinking (1935) treatment. When molecular phylogenetic studies of this group began in earnest, *Neocosmospora* included three major clades and many species (O'Donnell 1993, 2000, O'Donnell *et al.* 2008a). To date, 86 species are formally described in this group (Aoki *et al.* 2019, Sandoval-Denis *et al.* 2019, Guarnaccia *et al.* 2021), but

additional novel phylogenetic lineages are recognised and await formal description.

Thus, in *Neocosmospora* we have a group of species that can easily be recognised morphologically by both sexual and asexual morphs, exhibit generally consistent ecological behaviour, lack trichothecene mycotoxins, and form a strongly supported monophyletic group. This sounds like a biologically meaningful calibration of a genus, but what about the practicality of doing this? Presently, the data supporting the recognition of *Neocosmospora* (and equally, also *Fusarium s. str.*, the F3 clade) is stronger than the data supporting either of the nodes favoured for designating a broader concept of *Fusarium*. If there are 100 plus species in *Neocosmospora*, and hundreds of species in the trichothecene-producing, *Poaceae*-loving *Fusarium s. str.* clade, it will be useful for students, plant pathologists, clinical microbiologists, and other scientists to have different generic names for each group. Those names will convey information about relationships and behaviour that are lost in a broader definition of *Fusarium* with much greater diversity of ecological and biochemical behaviours. Geiser *et al.* (2013) raised concerns that grant evaluators, government regulators and medical practitioners who now believe they know what *Fusarium* means will be confused by the segregation of these fusarioid fungi into different genera, and that confusion could lead to unpredictable consequences. However, in our experience these end users continuously familiarise themselves with up-to-date, informative taxonomic and nomenclatural concepts for socio-economically important fungal groups, thus allowing them to predict the possible real-world effects of reliably identified fungi with increased precision. To them, the segregation of a heterogeneous concept of *Fusarium* into biologically and biochemically predictive genera will be helpful.

With *Neocosmospora* accepted as a different genus, *Albonectria*, *Cyanonectria*, and *Geejayessia*, as defined by Schroers *et al.* (2011), as well as *Bisifusarium* and *Rectifusarium* as defined in Lombard *et al.* (2015) must also be accepted as separate genera. As previously said, these are all monophyletic groups, also characterised by distinctive ecological and morphological traits.

The end consequence of our strategy is a series of phylogenetically well-supported genera, each with a recognisable suite of morphological characters, and ecological, pathological, and biochemical behaviour. Indeed, the results of such splitting activities applied to what we called the Wollenweber concept of *Fusarium s. lat.* accounts for 20 segregate genera. Most importantly, both *Fusarium* and *Neocosmospora* will have generic names to indicate their important but distinct significance. The extraneous species, with different ecology and generally much lower economic or agricultural significance can now justifiably be classified elsewhere, where they can be appreciated for their own features without the need for the uncertainty inherent in a broad concept of the generic name *Fusarium*.

The generic concept of *Fusarium* proposed by Geiser *et al.* (2013, 2021) functions well as a phylogenetic concept only if taxonomists turn their eyes away from all other kinds of data and observations applied to the family *Nectriaceae*. It is a political generic concept, meant to assuage the concerns of plant pathologists and other applied scientists, many of whom are already upset by the proliferation of cryptic phylogenetic species. Ironically, this late-blooming alleged pragmatism seems to betray

the cladistic ideals that many of its authors profess to adhere to (Taylor 2014).

All authors agree on the use of the single name *Fusarium*, have a common understanding of a phylogenetic structure of the family *Nectriaceae*, and agree that removing *Neocosmospora* from the main *Fusarium* core is the critical point of discussion. Sequencing additional markers may lead to increased phylogenetic support, but it is a false comfort if the taxon sampling does not include as many genera of *Nectriaceae* as possible. Expanded representation of the TFC in the dataset will not solve the controversy, and the resulting phylogenies will remain unbalanced. The segregation of *Neocosmospora* from *Fusarium* certainly needs to be done efficiently by those who have the most comprehensive expertise on the relevant species, which include several of the co-authors of the Geiser *et al.* (2013, 2021) and O'Donnell *et al.* (2020) papers as well as the present one.

Fusarium taxonomy has long been confused because of the nine-species system of Snyder & Hansen (1940, 1941), the misleading overlaps caused by convergent evolution and character loss, the difficulty in characterising perithecia, the phenomenon of cultural degeneration, and rigid opinions of the taxonomists and plant pathologists who have worked on them. To arrive at a stable taxonomy for *Fusarium*, the generic concept needs to be fixed in a practical and evolutionary reasonable manner so that future technologies and applications will not disrupt it.

SECONDARY METABOLITES OF FUSARIOID GENERA

The phylogenetic distribution of the fusarioid genera presented here is further corroborated by their ability to produce genus-specific secondary metabolites. The commercial database Dictionary of Natural Products (DNP; <http://dnp.chemnetbase.com>), was used to search for secondary metabolites produced by the genera and species treated here. The database contained (as of March 6, 2021) over 720 entries on metabolites from *Fusarium s. lat.*, even though some plant metabolites, discovered during studies on the elicitation of phytoalexins by challenging plant cells with a *Fusarium* strain, are included. The number of metabolites from *Fusarium s. lat.* is therefore estimated to be around 680, which is still behind *Aspergillus s. lat.* (over 3 000 entries) and *Penicillium s. lat.* (over 2 700 entries). Hits that were retrieved were confirmed by consulting the original literature. The reported structures were corroborated, with a selection of these compounds presented here (Figs 2–4).

It remains uncertain if the reported taxonomy is reliable, since the producer strains may have been misidentified or determined using one of many outdated taxonomic concepts. However, several compound classes have been encountered multiple times from the same species or species complex, and in some instances, the strains were identified by experts and/or sequenced later in phylogenetic studies (O'Donnell *et al.* 2018). The situation is further complicated by the fact that certain secondary metabolites have been given similar names, but represent different molecules. The name solaniol has been given to both a trichothecene (*Fusarium s. str.*) and a naphthoquinone (*Neocosmospora*), and the fusariumins represent four different secondary metabolites.

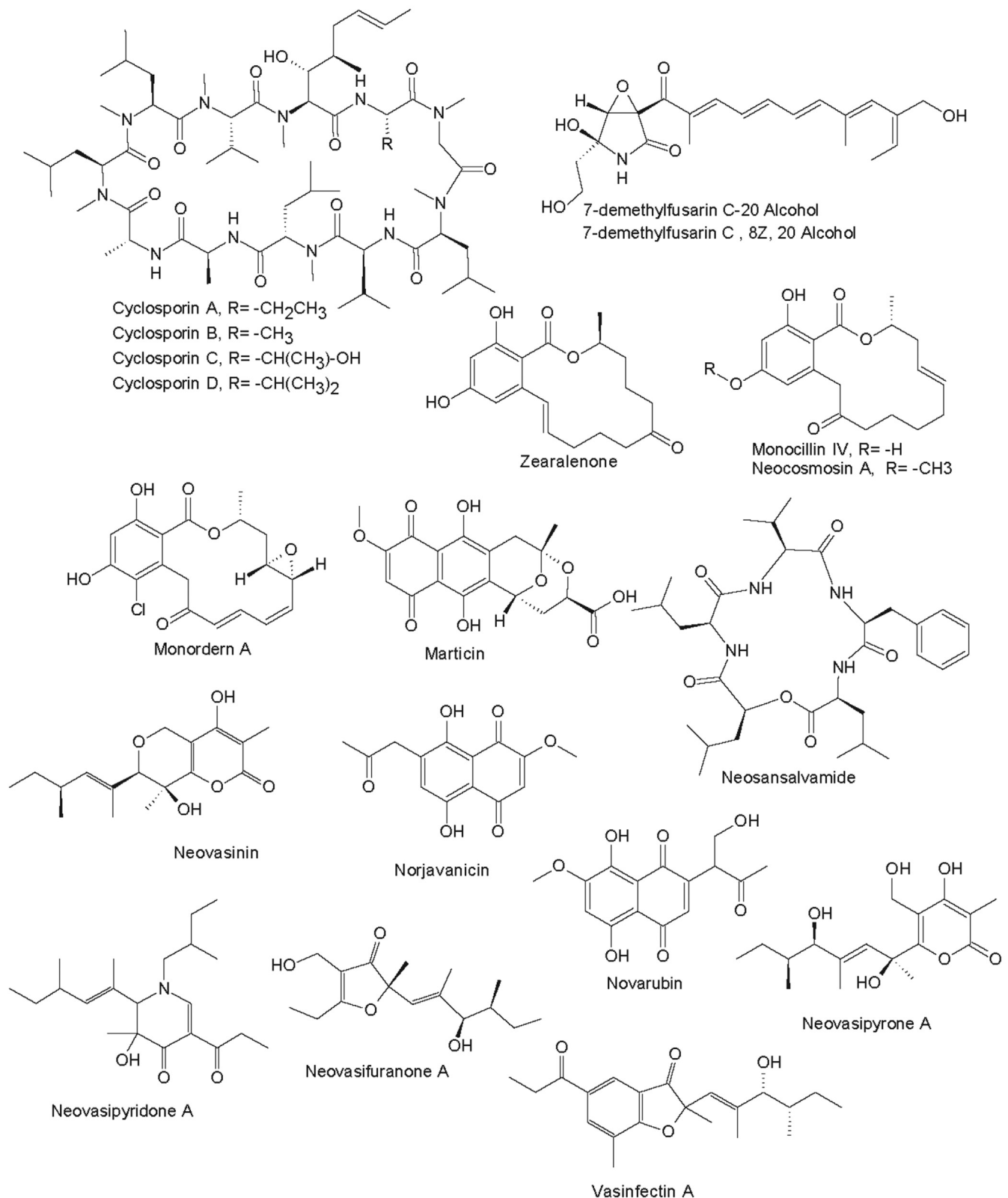


Fig. 2. Secondary metabolites from *Fusarium* spp. / *Neocosmospora* spp.

Typical metabolites of *Fusarium s. str.*

Fusarium sambucinum, the type species of the genus, has not been studied in much detail, but among the 20 metabolites known from this species, several metabolites are ranked in the classes trichothecenes and enniatins. The trichothecenes represents a well-known and notoriously dangerous class of mycotoxins belonging to the scirpene terpenoid type. These

compounds are widely distributed within the genus *Fusarium s. str.*, including familiar plant pathogenic species such as, *F. culmorum*, *F. graminearum*, *F. sporotrichioides* and *F. tricinctum* (Bamburg *et al.* 1968, Tatsuno *et al.* 1968, Yoshizawa & Morooka, 1973, Jiménez *et al.* 1997). The enniatins, known from 17 *Fusarium s. str.* species (Munkvold 2017, O'Donnell *et al.* 2018), are cyclic depsipeptides that have strong antibiotic activities (Plattner *et al.* 1948, German-Fattal 2001,

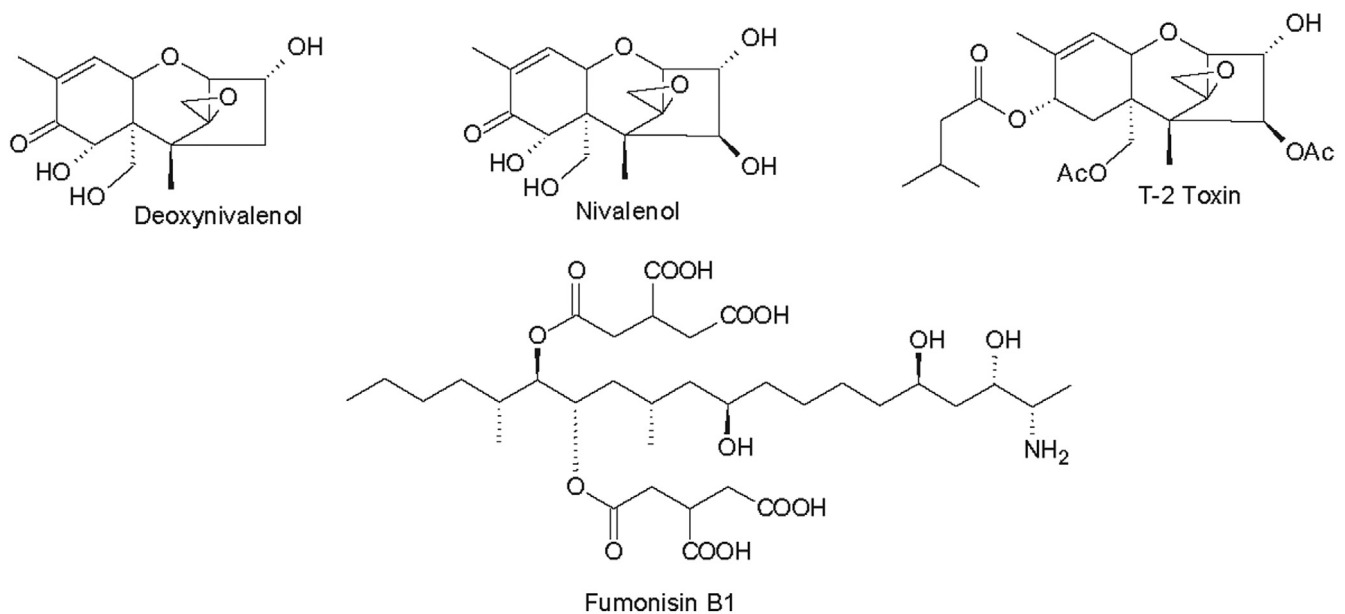


Fig. 3. Some of the most important mycotoxins from *Fusarium* spp.

Bills & Gloer 2017). Similar to trichothecenes, they are only known from *Fusarium* s. str. in the current taxonomic concept, although *Trichoderma* and *Beauveria*, which belong to different families of the *Hypocreales*, also produce trichothecenes or enniatin-like beauvericins, respectively. However, trichothecenes have not been reported from *Neocosmospora* or “*F. solani*” except from two isolates misidentified as “*F. solani*” (Ueno *et al.* 1972, Sugimoto *et al.* 2002) (Supplementary Table S2)

Two other well-known classes of mycotoxins, the fumonisins (Bezuidenhout *et al.* 1988) and zearalenone (Urry *et al.* 1966), are also found frequently among species of *Fusarium* s. str. Similarly, equisetin, also considered a “mycotoxin” and originally found from a *Fusarium* sp. strain (NRRL 5537) in the FIESC (Vesonder *et al.* 1979, Xia *et al.* 2019) is actually a strong antibiotic. A more complex derivative known as fusarisetin A was reported from an unidentified *Fusarium* sp. (Jang *et al.* 2011). Some rather unique compounds only known from *Fusarium* s. str., include wortmannin (Abbas & Mirocha, 1988) and oxysporizoline (Nenkep *et al.* 2016), which have interesting biological activities and may be species or even strain-specific.

Among the compounds that are not regarded as mycotoxins, the antimicrobial sesquiterpenes of the fusarielin type (Sørensen *et al.* 2013) and the antiparasitic and cytostatic cyclopeptides of the apicidin type (Jiang *et al.* 2002, Von Bargen *et al.* 2013) have been respectively isolated from *Fusarium* s. str. Additionally, aurofusarin (Munkvold 2017, O'Donnell *et al.* 2018), chlamydo-sporol (Munkvold 2017, O'Donnell *et al.* 2018), fusapyrone (Evidente *et al.* 1994), fusaric acid (Munkvold 2017, O'Donnell *et al.* 2018), fusoxysporone (Abraham & Hannsen 1992), fusaproliferin, moniliformin (Munkvold 2017, O'Donnell *et al.* 2018) and the terpestacins (Liu *et al.* 2013) are other examples of secondary metabolites found only in *Fusarium* s. str. Thus far, only one report has indicated that a *Neocosmospora* species can produce fusaric acid (Zhou *et al.* 2019). Both aurofusarin and bikaverin produced by *Fusarium* s. str. and other bis-naphthoquinone and bis-naphthopyrone pigments protect fungi from predation (Xu *et al.*, 2019), while *Neocosmospora* species produce other naphthoquinones such as javanicin (Arnstein & Cook 1947, Kimura *et al.* 1981) as potential predator protectors. Some unique compounds have been reported from marine strains of

certain *Fusarium* species, which include the mangicols, rare sesterterpenes produced by a strain tentatively classified as *F. heterosporum* (Renner *et al.* 2000).

Typical metabolites of *Neocosmospora* and other fusarioid genera

Neocosmospora species and other fusarioid genera apparently have a different secondary metabolism, or have not been intensively studied in the past. A striking example are the cyclosporins, which are immunosuppressive peptides. Originally, these were obtained from *Tolypocladium inflatum*, but later also found to be produced by species of *Neocosmospora* (Sawai *et al.* 1981, Nakajima *et al.* 1989). However, they have not been reported from *Fusarium* s. str. Other unique compounds only known from *Neocosmospora* species, include dihydrofusarin (Kurobane *et al.* 1980, Kyekyeku *et al.* 2017), the polyketides neovasipyrone (Furumoto *et al.* 1995, Nakajima *et al.* 1995) and vasinfectin A (Furumoto *et al.* 1997). The rare cyclopeptides of the neosansalvamide type (Lee & Lee 2012) and the resorcylic acid lactones of the monorden/monocillin type (Cutler *et al.* 1987, Gao *et al.* 2013) are also known from *Neocosmospora* and other fungi, but not from *Fusarium* s. str., even though the latter compounds bear a high structural resemblance to zearalenone. Several *Neocosmospora* species produce a range of naphthoquinones that are members of a widespread class of polyketides (Roos 1977).

The fusarioid genus *Bisifusarium* is known to produce the PKS/NRPS hybrid siderophore, dimerumic acid (= dimerum acid) (Diekmann 1970), and indole acetic acid (Reddy & Reddy 1992, Kulkarni *et al.* 2011, 2013). The parnafungins, which are under development as antimycotics, are only known from *Microcera larvarum* (Parish *et al.* 2008). Additionally, *Microcera larvarum* is also known to produce monocerin and fusarentins, which are not known from any other fungi (Grove & Pople 1979), except a *Colletotrichum* species (Tianpanich *et al.* 2011). The anticancer agent balanol (azepinostat) (Ohshima *et al.* 1994) is known to be produced by two *Fusicolla* species, which might be applied as a taxonomic marker for this genus, although it has also been

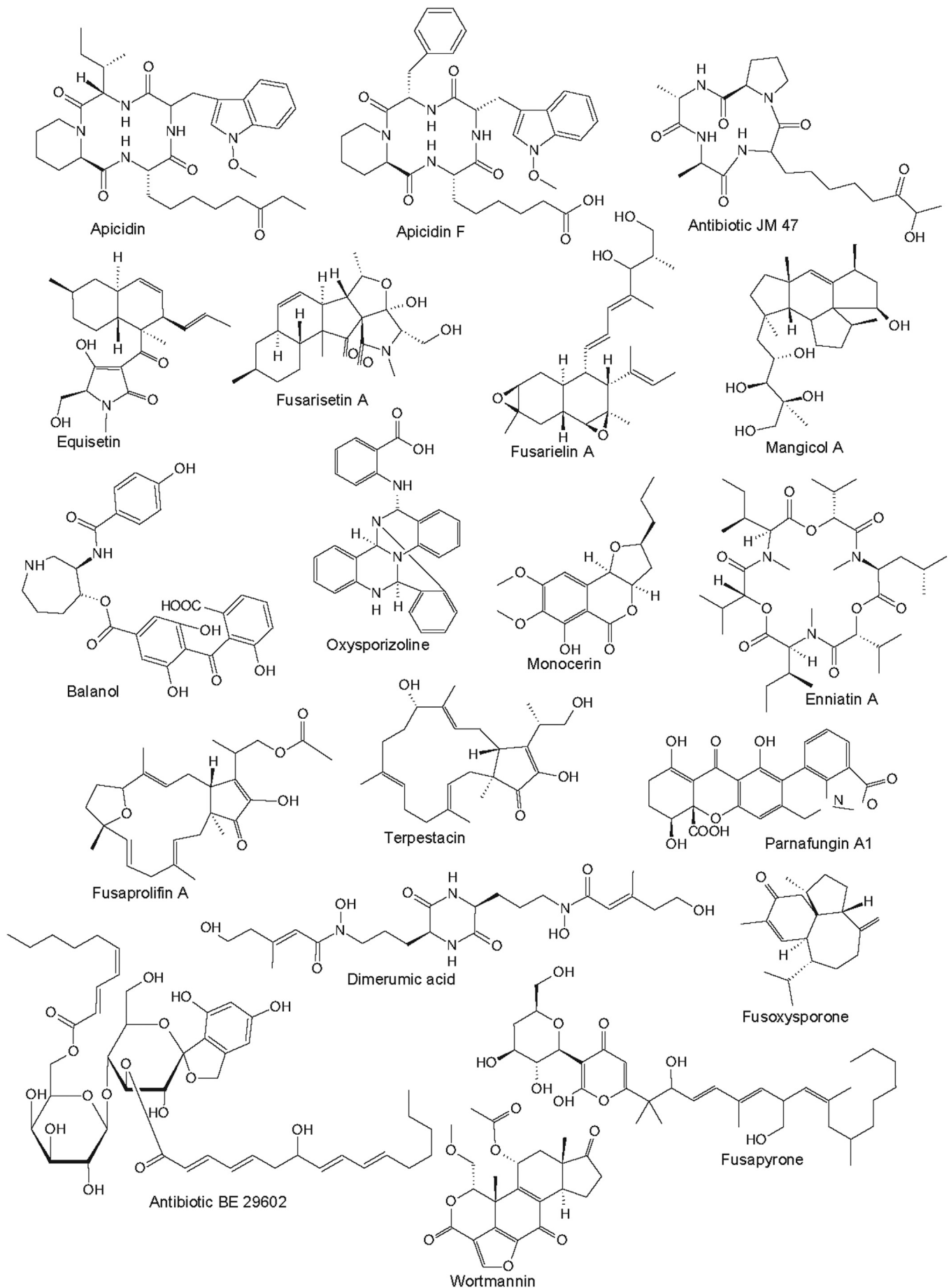


Fig. 4. Secondary metabolites from fusarioid *Hypocreales*.

found in species of the *Ophiocordycipitaceae*. Unfortunately, there is no available information on secondary metabolites for the other fusarioid genera treated here. However, secondary

metabolite studies of these missing genera will facilitate for the discovery of novel molecules and help to elucidate the functional biodiversity of these fungi.

RECOMMENDED METHODS FOR THE IDENTIFICATION AND CHARACTERISATION OF *FUSARIUM* AND ALLIED GENERA

The following part of this study presents an overview of the morphological and phylogenetic characters of *Fusarium* and related genera as well as an account of recommended methods for the identification and characterisation of these taxa. In addition, novel genera and species are described and, in view of the recent taxonomic data, a list of names that are applied to the genus *Fusarium* s. lat. with their current scientific names is presented.

Morphology

Current *Fusarium* taxonomy is dominated by molecular phylogenetic studies. Nonetheless, morphology is a fundamental component of the generic and species concepts of fungi and must not be overlooked. Key morphological features for generic circumscription include characteristics of sexual morphs such as perithecial colour, wall thickness and anatomy, surface structures and the presence and nature of a basal stroma, ascospore shape, septation, colour and surface ornamentation (Rossman et al. 1999). Classification of taxa solely based on their asexual morphs can be trickier than integrated systems using sexual and asexual characters. However, the general shapes, different types and combinations of conidiogenous structures and conidia present in culture can be sufficient to allow a preliminary identification (Fig. 5), especially if host data are also available (Leslie & Summerell 2006). For species-level characterisation, a number of morphological traits must be carefully studied, particularly those of the asexual morph, while sexual morphs are generally less suitable, especially as they are typically not produced in culture. Diagnostic characters for species identification include colony characters such as colony morphology, pigmentation, and type of aerial mycelium. Also included are the dimensions and characteristics of aerial conidiophores and conidiogenous cells (mono- vs polyphialides), presence/absence and characteristics of sporodochia, the types of conidia produced, e.g., aerial microconidia, mesoconidia, and aerial and sporodochial macroconidia. In examining conidia themselves, consideration is given to the overall shape, septation and curvature of the macroconidia, as well as characteristics of their apical and basal cells; with aerial microconidia, their dimensions, shape, septation and spatial organisation (forming slimy heads, chains or a combination of both) are noted. Finally, the presence or absence of chlamydospores may be important.

Culture media and incubation

Vigorous growth, sporulation, and pigment production of fusarioid fungi can be achieved on numerous agar formulations. The morphology of fungal structures will vary dramatically depending on the selection of media and growth conditions which may compromise the identification process. In addition, it is also common for fusaria to degenerate and lose viability in culture, particularly when they are grown on nutrient-rich media (Nelson et al. 1983, Nirenberg 1990, Summerell et al. 2003, Leslie & Summerell 2006). Culture conditions and media have been extensively summarised in the literature (Booth 1971, Nirenberg

1990, Nelson et al. 1994, Summerell et al. 2003, Leslie & Summerell 2006). Consequently, we recommend the agar formulations listed in Table 1 to be employed for the isolation and description of fusaria. A summary of the procedures and conditions suitable for work with fusarioid fungi is shown in Fig. 6.

An important condition that must be stressed is that the identification must always be made on the basis of a monosporic culture (a culture produced from a single sporulating conidium, ascospore, or hyphal tip), as multiple species are commonly found to co-occur in the same substrate tissue. A freshly isolated fusarioid strain should be sub-cultured onto at least two different culture media, a relatively rich one suitable for examination of gross morphology, and a nutrient-poor one for micromorphological examination and for further culture propagation. The standard culture setup for initial assessment of growth rates and colony characters i.e., colony pigmentation, diffusible pigments, and colour of sporodochia, is to use potato dextrose agar (PDA) incubated for 1–2 wk. *Fusarium* and related genera will also grow and sporulate well on malt extract agar (MEA, recipe in Crous et al. 2019a), which can be a suitable alternative for initial isolation and monosporic cultivation. However, MEA should not be used to assess colony or morphological characters. Standard incubation is commonly made in total darkness; however, exposure to light will normally result in a faster and more intense pigmentation. We have observed better colour formation using in-house prepared media rather than commercial formulae. While colony colour cannot be employed as a primary criterion for species identification, it can provide useful means to grossly distinguish related groups and to direct the identification process towards determining genera or species complexes. The high nutrient content of these agar media strongly affects sporulation, commonly resulting in the development of atypical structures. Therefore, we strongly discourage the use of PDA for micromorphological assessment or culture propagation of *Fusarium* spp. (Nelson et al. 1994, Summerell et al. 2003). Oatmeal agar (OA) is a suitable alternative for strain sub-culturing, allowing for good sporulation with reduced strain degeneration; however, it is not recommended for micromorphological studies.

Carnation leaf agar (CLA), synthetic nutrient-poor agar (SNA), and water agar (WA) are the standard culture media for micromorphological analyses. Also, by reducing culture degeneration, they allow for prolonged storage of actively growing cultures (Nirenberg 1976, Nelson et al. 1983, Leslie & Summerell 2006). Subcultures on CLA will normally produce abundant sporodochia and macroconidia on the surface or around the carnation leaf pieces with consistent morphological features. Incubation at room temperature (20–25 °C) for 1–2 wk under a 12/12 h near-UV light (wavelength 320–400 nm)/dark or near-UV light/cool fluorescent light cycles results in stronger sporulation and good development of sporodochial pigmentation (Nirenberg 1990, Seifert 1996, Summerell et al. 2003, Leslie & Summerell 2006). The use of continuous near-UV light (also commonly termed "blacklight" or UV-A light) is also suitable although it often results in the formation of unusually long macroconidia (Nirenberg 1990), and it can suppress the development of useful morphological characters such as the globose microconidia of *Fusarium globosum*. Nevertheless, incubation under near-UV light is fundamental since isolates of some species such as *Fusarium poae* and *F. sacchari* are known to lack macroconidia or to produce them in only small quantities unless they are stimulated by incubation under a near-UV light source (Leslie et al. 2005, Leslie & Summerell 2006). *Fusarium* cultures also need

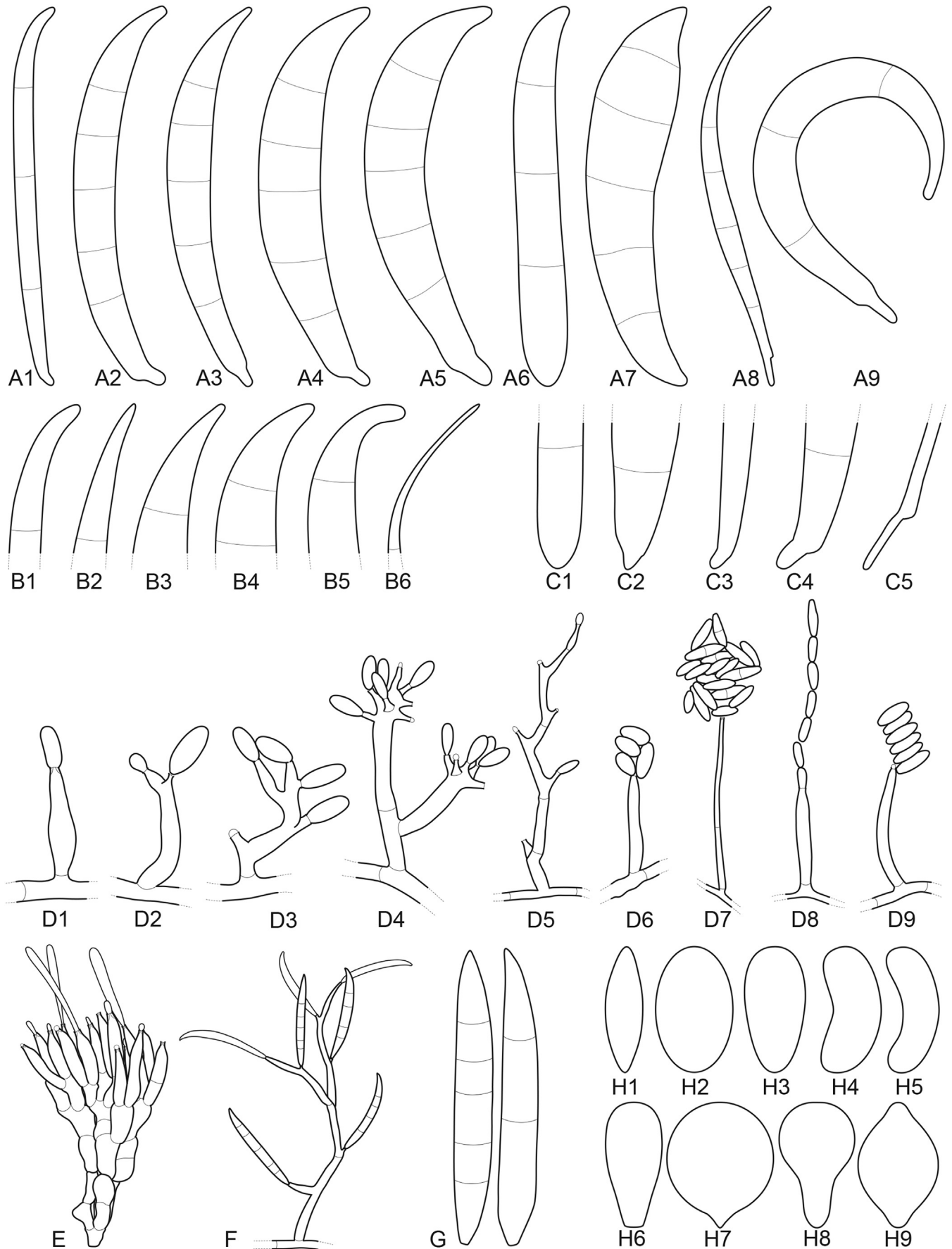


Fig. 5. Basic morphological features of fusarioid fungi. **A.** Macroconidial shapes. A1. Slender with no significant curvature. A2. Curved with parallel walls. A3. Unequally curved. A4. Widest at the middle portion. A5. Widest at the apical third, wedge-shaped. A6. Widest at the basal portion. A7. Irregularly clavate and swollen. A8. Elongate, whip-like. A9. Distinctly curved. **B.** Macroconidial apex. B1. Curved. B2. Long and tapered. B3. Pointed. B4. Blunt. B5. Hooked. B6. Elongated. **C.** Macroconidial base. C1. Obtuse, non foot-shaped. C2. Papillate, non foot-shaped. C3. Poorly developed, foot-shaped. C4. Well-developed, foot-shaped. C5. Elongate, foot-shaped. **D.** Aerial phialides and microconidial organization. D1. Monophialide. D2–D5. Polyphialides. D2. Simple polyphialide. D3–D4. Polyphialides with multiple conidiogenous loci. D5. Sympodially proliferating polyphialides. D6, D7. Microconidia forming false heads. D8, D9. Microconidia in chains (D8. Dry chain. D9. Palisade). **E.** Sporodochial conidiophore and conidiogenous cells. **F.** Aerial conidiophore bearing mesoconidia. **G.** Mesoconidia. **H.** Microconidial shapes. H1. Fusiform. H2. Oval. H3. Obovoid. H4. Reniform. H5. Allantoid. H6. Clavate. H7. Napiform. H8. Pyriform. H9. Limoniform.

Table 1. Recommended agar media formulations for the isolation and cultivation of fusaria.

Agar media	Components ¹	Preparation ²	Incubation ³	Application	Reference
Carnation leaf agar (CLA)	Sterilised carnation leaves WA	Carnation leaves are cut into approximately 5 × 5 mm pieces and dried at 60 °C for 24 h; sterilise by gamma radiation or autoclave; place 3–5 pieces on nearly solid 2 % WA surface.	25 °C; 7–14 d under 12 h near-UV-light/dark cycle; 7–14 d under 24 h near-UV-light	Micro-morphological characterisation: formation of sporodochia; sporodochial macroconidia	Fisher et al. (1982) , Crous et al. (2019a)
Selective Fusarium Agar (SFA)	Glucose (Dextrose) 20 g KH ₂ PO ₄ 1 g NaNO ₃ 2 g MgSO ₄ ·7H ₂ O 0.5 g Yeast Extract 1 g 1 % FeSO ₄ ·7H ₂ O (aqueous) 1 ml Streptomycin (5 % w/v) 20 mL Neomycin (1 % w/v) 12 mL Dichloran (50 % w/v in ethanol) 13 mL Agar 20 g Water 1 000 mL	Add all components, except antibiotics, to water and autoclave; cool to 45–50 °C and add antibiotics. Dichloran can be replaced by PCNB (0.75 g).	25 °C; 7–14 d in dark	Selective isolation of fusaria from soil	Tio et al. (1977) , Leslie & Summerell (2006)
Komada's Medium	D-Galactose 20 g L-Asparagine 2 g KH ₂ PO ₄ 1 g KCl 0.5 g MgSO ₄ ·7H ₂ O 0.5 g PCNB 0.75 g Fe ₃ Na EDTA 0.01 g Streptomycin (5 % w/v) 6 mL Oxgall stock solution 0.5 g Na ₂ B ₄ O ₇ ·10H ₂ O (borax) 0.5 g Agar 15–20 g Water 1 000 mL	Add all components, except antibiotics, oxgall and borax; to water and autoclave; cool to 45–50 °C and add the remaining components. Adjust pH to 3.8 ± 0.2 prior to autoclaving.	25 °C; 7–14 d in dark	Selective isolation of fusaria from soil, particularly those belonging to the <i>Fusarium</i> oxysporum species complex. Other fusaria can be inhibited by this medium	Komada (1975) , Leslie & Summerell (2006)
Malachite Green Agar (MGA)	Peptone 15 g KH ₂ PO ₄ 1 g MgSO ₄ ·7H ₂ O 0.5 g Malachite green oxalate 2.5 mg Streptomycin (5 % w/v) 20 mL Penicillin (5 % w/v) 20 mL Agar 20 g Water 1 000 mL	Add all components, except antibiotics, to water and autoclave; cool to 45–50 °C and add antibiotics. Penicillin can be also replaced by chloramphenicol (5 % w/v) or neomycin (1 % w/v).	25 °C; 7–14 d in dark	Selective isolation of fusaria from soil and plant material, with improved inhibition of non-fusarioid contaminants	Castellá et al. (1997) , Leslie & Summerell (2006)
Oatmeal agar (OA)	Oatmeal extract 1 000 mL Agar 15–20 g	Oatmeal flakes (30 g/L) are wrapped in cloth and simmered in water for 2 h; liquid is squeezed and filtered through cloth.	25 °C; 7–14 d in dark	Macro-morphological characterisation, colony characteristics	Crous et al. (2019a)

Table 1. (Continued).

Agar media	Components ¹		Preparation ²	Incubation ³	Application	Reference
Potato dextrose agar (PDA)	Potato extract	230 mL	Potatoes (5 kg; peeled and sliced) are minced; soak in water (300 mL/100 g potato) overnight at 4 °C; filter through cloth; adjust pH to 6.6.	25 °C; 7–14 d in dark; 5–40 °C (5 °C increments for growth curves)	Inoculum preparation, macro-morphological characterisation: colony characteristics; growth curve	Crous et al. (2019a)
	Agar	15–20 g				
	Water	770 mL				
Peptone Pentachloronitrobenzene (PCNB) agar (PPA)	Peptone	15 g	Add all components, except antibiotics, to water and autoclave; cool to 45–50 °C and add antibiotics. Penicillin can be also replaced by chloramphenicol (5 % w/v) or neomycin (1 % w/v).	25 °C; 7–14 d in dark	Selective isolation of fusaria from soil and plant material	Nash & Snyder (1962) , Booth (1971) , Leslie & Summerell (2006)
	KH ₂ PO ₄	1 g				
	MgSO ₄ ·7H ₂ O	0.5 g				
	PCNB	0.75 g				
	Streptomycin	(5 % w/v) 20 mL				
	Penicillin	(5 % w/v) 20 mL				
	Agar	20 g				
	Water	1 000 mL				
Rose Bengal-Glycerine-Urea Medium (RbGU)	Glycerol	10 g	Add all components, except antibiotics, to water and autoclave; cool to 45–50 °C and add antibiotics.	25 °C; 7–14 d in dark	Isolation of fusaria from soil and plant material	van Wyk et al. (1986) , Leslie & Summerell (2006)
	Urea	1 g				
	L-Alaninw	0.5 g				
	PCNB	1 g				
	Rose Bengal	0.5 g				
	Streptomycin	(5 % w/v) 20 mL				
	Agar	15 g				
	Water	1 000 mL				
Synthetic nutrient-poor agar (SNA)	KH ₂ PO ₄	1 g	Add all components to water and autoclave.	25 °C; 7–14 d under 12 h near-UV-light/dark cycle	Inoculum preparation, micro-morphological characterisation: aerial conidiophores and micro- & macroconidia; chlamydospore formation	Nirenberg (1976) , Crous et al. (2019a)
	KNO ₃	1 g				
	MgSO ₄ ·7H ₂ O	0.5 g				
	KCl	0.5 g				
	Glucose	0.2 g				
	Saccharose	0.2 g				
	Water	1 000 mL				
Water agar (WA)	Agar	15–20 g		25 °C; 7–14 d in dark	Inoculum preparation, base agar for CLA	Crous et al. (2019a)
	Water	1 000 mL				

¹ Unless specified differently, antibiotic stock solutions are prepared in distilled water.

² Water refers to distilled water; autoclave = 121 °C for 15 min.

³ Near-UV = near ultraviolet spectrum (wavelength 320–400 nm).

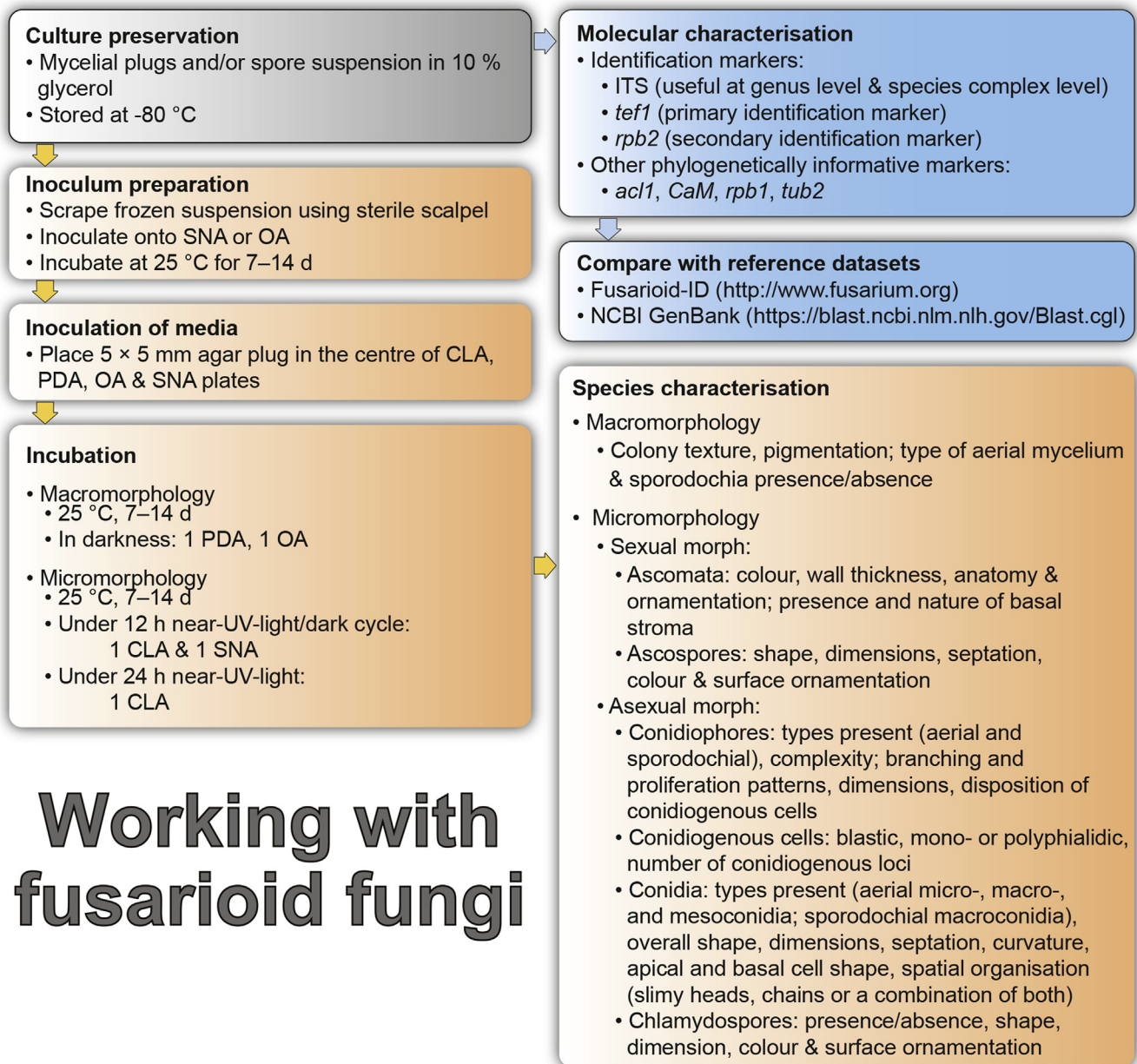


Fig. 6. Flow diagram summarising recommended methods for the preservation, identification, and characterisation of fusarioid fungi.

adequate aeration to produce conidia reliably and to attain stable growth rates, and hence we discourage the incubation of sealed plates. Carnation leaf agar, SNA, and WA are also suitable for the observation of conidiophore disposition and microconidial arrangements such as the formation of false heads, chains or both. These structures can easily be examined under a dissecting microscope or at low magnification under a compound light microscope (Leslie & Summerell 2006). Examination of micromorphological characters must be carried out using slide preparations mounted in water. Lactic acid, lactophenol and Shear's mounting media can cause considerable shrinking of the structures and can alter the appearance of the cell surface; hence we advise against the use of these mountants for examination of morphological characters in *Fusarium* and related genera.

Additional culture media, incubation conditions, and protocols are available for induction of sexual characters in *Fusarium* and related genera (Klittich & Leslie 1988, Leslie & Summerell 2006, Guo *et al.* 2018, Kim *et al.* 2019, Santos *et al.* 2019). Carrot agar (CA) and half-strength CA are the most commonly used media.

The crossing procedures are often variations from the protocol of Klittich & Leslie (1988), in which strains of opposite mating types are paired in all possible combinations as male and female parents, together with crosses made against tester strains from known mating populations (Leslie & Summerell 2006). The process can be shortened by reducing the number of combinations to be crossed by first determining the *MAT* gene alleles carried by each strain by means of specific mating type idiomorph PCR primers (Kerényi *et al.* 1999, 2004, Steenkamp *et al.* 2000).

Molecular studies

Several genes, primer combinations and PCR conditions have been listed in the *Fusarium* literature (O'Donnell *et al.* 1998a, b, 2000a, b, 2007, 2010, 2013, Gräfenhan *et al.* 2011, Lombard *et al.* 2015, 2019a, b), including whole-genome sequencing to mine for the desired genes (O'Donnell *et al.* 2020, Geiser *et al.* 2021). Here we detail those DNA markers that have shown the best results in routine diagnosis (Table 2, Fig. 6).

Table 2. Recommended PCR primers for DNA amplification of *Fusarium* and related genera.

Gene/DNA region		Primer			
Name	Abbreviation	Name	Direction	Sequence (5' → 3')	Reference
28S large subunit of the nrDNA	LSU	LR0R	Forward	ACCCGCTGAACTTAAGC	Vilgalys & Sun (1994)
		LR5	Reverse	ATCCTGAGGGAAACTTC	Vilgalys & Hester (1990)
		NL4 ²	Reverse	GGTCCGTGTTTCAAGACGG	Kurtzman & Robnett (1997)
ATP citrate lyase	<i>acl1</i>	230up	Forward	AGCCCGATCAGCTCATCAAG	Gräfenhan <i>et al.</i> (2011)
		1220low	Reverse	CCTGGCAGCAAGATCVAGGAAGT	Gräfenhan <i>et al.</i> (2011)
Beta-tubulin	<i>tub2</i>	T1	Forward	AACATGCGTGAGATTGTAAGT	O'Donnell & Cigelnik (1997)
		TUB-2Fd ²	Forward	GTBACCTYCARACCGGYCARTG	Woudenberg <i>et al.</i> (2009)
		TUB4RD	Reverse	CCRGAYTGRCCRAARACRAAGTTGTC	Woudenberg <i>et al.</i> (2009)
Calmodulin	<i>CaM</i>	CAL-228f	Forward	GAGTTCAAGGAGGCCTTCTCCC	Carbone & Kohn (1999)
		CAL-CL1 ²	Forward	GARTWCAAGGAGGCCTTCTC	O'Donnell <i>et al.</i> (2000b)
		CAL-CL2A ²	Reverse	TTTTTGCATCATGAGTTGGAC	O'Donnell <i>et al.</i> (2000b)
		CAL-2Rd	Reverse	TGRTCNGCCTCDCGGATCATCTC	Quaedvlieg <i>et al.</i> (2011)
Internal transcribed spacer region of the nrDNA	ITS	ITS5	Forward	GGAAGTAAAAGTCGTAACAAGG	White <i>et al.</i> (1990)
		V9G ²	Forward	TTACGTCCTGCCCTTTGTA	de Hoog & van den Ende (1998)
		ITS4	Reverse	TCCTCCGCTTATTGATATGC	White <i>et al.</i> (1990)
RNA polymerase largest subunit	<i>rpb1</i>	Fa	Forward	CAYAARGARTCYATGATGGGWC	O'Donnell <i>et al.</i> (2010)
		F7	Forward	CRACACAGAAGAGTTTGAAGG	O'Donnell <i>et al.</i> (2010)
		F8 ¹	Forward	TTCTCCACGCCATGGCTGGTCG	O'Donnell <i>et al.</i> (2010)
		F6 ¹	Forward	CTGCTGGTGGTATCATTACAG	O'Donnell <i>et al.</i> (2010)
		R8	Reverse	CAATGAGACCTTCTCGACCAGC	O'Donnell <i>et al.</i> (2010)
		R9	Reverse	TCARGCCCATGCGAGAGTTGTC	O'Donnell <i>et al.</i> (2010)
		G2R ¹	Reverse	GTCATYTGDTGDCDGGYTCDDC	O'Donnell <i>et al.</i> (2010)
RNA polymerase second largest subunit	<i>rpb2</i>	RPB2-5f2	Forward	GGGGWGAYCAGAAGAAGGC	Reeb <i>et al.</i> (2004)
		fRPB2-7cf	Forward	ATGGGYAARCAAGCYATGGG	Liu <i>et al.</i> (1999)
		fRPB2-7cr	Reverse	CCCATRGCTTGYTTRCCCAT	Liu <i>et al.</i> (1999)
		RPB2-11ar	Reverse	GCRTGGATCTTRTCRTCSACC	Liu <i>et al.</i> (1999)
Translation elongation factor 1-alpha	<i>tef1</i>	EF-1	Forward	ATGGGTAAGGARGACAAGAC	O'Donnell <i>et al.</i> (1998b)
		EF-2	Reverse	GGARGTACCAGTSATCATG	O'Donnell <i>et al.</i> (1998b)

¹ Used only for sequencing reactions.

² Alternative primer, not used in this study.

Nuclear ribosomal DNA (nrDNA), including the internal transcribed spacer region cistron (ITS) and the 28S large subunit nrDNA (LSU), are nearly useless for species recognition in *Fusarium* and related genera. Nevertheless, given the ease of amplification and the extensive data available for comparison in public databases (Schoch *et al.* 2012), these markers are useful in the discrimination between the multiple species complexes of *Fusarium*, and for obtaining a confident genus-level identification for *Fusarium* and related genera, allowing further DNA markers to be incorporated in the analyses. The ITS region can still provide valuable information at species level for related genera containing species formerly included in *Fusarium* (*Bisifusarium*, *Cosmospora*, *Fusicolla*, *Macroconia*, *Microcera*, and *Stylonectria*).

Many protein-coding genes have been explored for identification and taxonomic purposes in *Fusarium* and fusarioid fungi. The two main genes used for identification are *tef1* and *rpb2*. Both offer high discriminatory power and are well represented in public databases. Translation elongation factor 1- α is commonly the first-choice identification marker as it has very good resolution power for most species in all the genera treated here, while *rpb2* allows for enhanced discrimination between closely related species. For example, some species in the *Fusarium fujikuroi* species complex (FFSC) and in *Neocosmospora* that are not easily separated by using *tef1* alone (O'Donnell 2000, Nalim *et al.* 2011, Herron *et al.* 2015), can be resolved with *rpb2*. On the other hand, PCR amplification and sequencing success are often better for *tef1* than for *rpb2*. When used for phylogenetic analyses, sequence alignments of *rpb2* sequences are much

more robust and less ambiguous than *tef1* data, given the former gene's advantageously low proportion of introns. An analogous situation has been shown in *Aspergillus* (Samson *et al.* 2014) and *Penicillium* (Visagie *et al.* 2014).

Additional genetic markers, often employed in association with the previously mentioned genes in multigene phylogenetic analyses include *acl1*, *tub2*, *CaM*, and *rpb1*. These markers have variable resolution or applicability depending on the genus or species complex. For example, use of *CaM* data may yield conflicting clade resolutions in the FFSC (O'Donnell 2000, Al-Hatmi *et al.* 2019), while paralogous or xenologous gene copies have been demonstrated for *tub2* in the *F. chlamydosporum* and *F. incarnatum-equiseti* species complexes (O'Donnell *et al.* 2009) as well as in *Neocosmospora* (O'Donnell 2000, O'Donnell *et al.* 2008a).

The most widely used algorithm for fungal identification by means of DNA markers is the Basic Local Alignment Search Tool (BLAST), available at the NCBI's GenBank website. This is a quick and useful method that can convey a great deal of information, but its results must be analysed with care given the presence of a high proportion of misidentified strains and low-quality sequences that must be filtered out (Vilgalys 2003, Nilsson *et al.* 2012). Sequences from type material are present in the GenBank nucleotide database for most fusarioid species known from culture, especially for *rpb2* and *tef1* barcodes, but the ex-type status of these sequences is not always explicitly mentioned. In many cases the names listed do not reflect the current taxonomy, even for sequences derived from ex-type cultures.

Some sequences used in past phylogenetic analyses of O'Donnell *et al.* (2020) and Geiser *et al.* (2021) appear to be linked to incorrect *Fusarium* names, likely due to errors in the database used. For this reason, we recommend the use of our curated database: Fusarioid-ID (<https://www.fusarium.org>). It can also be used for sequence similarity-based analysis of routine isolations and for identifications within several related genera.

MALDI-TOF

A number of studies have thus far demonstrated the utility of mass spectrometry (MS) for species determination of subgroups of *Fusarium*, particularly members of the FFSC (Al-Hatmi *et al.* 2015, 2016, Wiggmann *et al.* 2019). It is also useful for clinically relevant subgroups within several *Fusarium* species complexes (Marinach-Patrice *et al.* 2009, Triest *et al.* 2015, Sleiman *et al.* 2016, Paziani *et al.* 2020) and clinically relevant *Bisifusarium* (Triest *et al.* 2015, Paziani *et al.* 2020) and *Neocosmospora* species (Marinach-Patrice *et al.* 2009, Triest *et al.* 2015, Sleiman *et al.* 2016, Paziani *et al.* 2020). These techniques show highly accurate discriminative power, comparable to what has been shown with bacteria and yeasts. Only a limited number of taxa have thus far been evaluated, and a genus-wide evaluation of applicability of MALDI-TOF to *Fusarium* and related taxa is pending. The main limiting factor is, as usual, the current lack of representation of these taxa in commercial spectrum databases, a matter that can be resolved by constructing in-house, curated reference databases of spectra. Online availability and comparison of MS spectra of *Fusarium* has been proposed by Triest *et al.* (2015).

MATERIALS AND METHODS

Isolates and fungarium specimens

Fungal strains were obtained from the Westerdijk Fungal Biodiversity Institute (WI) collection (CBS), the Belgian Coordinated Collections of Microorganisms (IHEM), the International Mycological Institute (IMI), and the personal collection of Pedro W. Crous (CPC) housed at WI. For the list of names applied to the genus *Fusarium* and related fungarium specimens, the following fungaria were approached for holotype specimens: B, BM, BO, BP, BPI, BR, BRA, C, CBS, CO, DAOM, E, FH, H, HAL, IMI, K(M), L, LEP, M, MASS, MPA, NY, PC, PAD, PARMA, PAV, PH, PRM, ROVP, SIENA, STR, UPS, VPRI, W, and WIR.

DNA amplification and phylogeny

Total genomic DNA was extracted from isolates grown for 7 d on PDA or MEA (recipes in Crous *et al.* 2019a; Table 1) incubated at 24 °C under a 12/12 h photoperiod using the Wizard® Genomic DNA purification Kit (Promega Corporation, Madison, WI, USA), following the manufacturer's instructions. Partial gene sequences were determined for eight DNA markers, *i.e.*, *act1*, *CaM*, ITS, LSU, *rpb1*, *rpb2*, *tef1*, and *tub2* using PCR protocols described elsewhere (O'Donnell *et al.* 1998b, 2007, 2010, Lombard *et al.* 2015). Primer pairs used for amplification and sequencing of

the respective gene regions are summarised in Table 2. Consensus sequences for each marker were assembled in Geneious R11 (Kearse *et al.* 2012) or SeqMan Pro v. 15.3.0 (DNASTAR, Madison, WI, USA). All sequences generated in this study were deposited in GenBank (Table 3; also see Diagnostic DNA Barcodes in list of *Fusarium* names). The multiple sequence alignments and phylogenetic trees were deposited in TreeBASE (study ID 28093).

Sequences of the individual markers, including introns, were aligned using MAFFT v. 7.110 (Kato *et al.* 2019) using default parameters and manually corrected where necessary. Seven multimarker datasets (Table 4) were assembled and analysed using Maximum Likelihood (ML) and Bayesian Inference (BI). For the ML analyses, concatenated phylogenies, where each marker was treated as a separate partition, were determined using IQ-TREE v. 2.1.2 (Nguyen *et al.* 2015, Minh *et al.* 2020b) with ultrafast bootstrapping (UFBoot2; Hoang *et al.* 2018) for estimation of branch support. The most suitable evolutionary model for each partition was estimated using ModelFinder (Kalyaanamoorthy *et al.* 2017; Minh *et al.* 2020b) as implemented in IQ-TREE. To assess whether the individual markers were compatible, genealogical concordance factors (gCF) were calculated using IQ-TREE (Minh *et al.* 2020a, b). Additional ML analyses were performed using RAXML v. 8.2.12 (randomised accelerated (sic) maximum likelihood for high performance computing; Stamatakis 2014) with the system's default modelling options. The robustness of the analysis was evaluated by bootstrap support (BS) with the number of bootstrap replicates automatically determined by the software. The BI analyses were carried out through the CIPRES website (<http://www.phylo.org>) using MrBayes v. 3.2.7a (Ronquist & Huelsenbeck 2003) incorporating the best evolutionary models for each marker as determined by MrModeltest v. 2.3 (Nylander 2004). Two parallel Markov Chain Monte Carlo (MCMC) runs of four incrementally heated chains (temp parameter = 0.2) were run starting from a random tree topology. The MCMC analyses lasted for 5M generations, and convergence of the runs was checked by average standard deviation of split frequencies below 0.01. Trees were saved every 1 000 generations and the first 25 % of saved trees were discarded as the "burn-in" phase. Posterior probabilities (PP) were determined from the remaining trees. Proper mixing of the MCMC runs was further confirmed by checking that all chains converged (minimum and average Estimated Sampled Size [ESS >200], Potential Scale Reduction Factor [PSRF = 1.0]) and by plotting and analysing trace file results using Tracer v.1.7.1 (Rambaut *et al.* 2018).

The phylogenetic re-analysis of the dataset presented by Geiser *et al.* (2021) was first made according to the original exons-only alignment file and procedures as indicated in Geiser *et al.* (2021) (Supplementary Table S1). Additionally, the dataset was split into the 19 genes according to the original partitioning file, and every gene was realigned using the MAFFT webserver (v. 7, Kato *et al.* 2019) applying the G-INS-i algorithm. All other parameters were set to default. Six of the 19 genes exhibited a diverging alignment length. No subsequent changes were done to the alignments. The sequences were merged using BioEdit (v. 7.2.5, Hall 1999), and the phylogenetic trees were calculated using Minimum evolution (ME) and ML algorithms, and BI. The ME tree was calculated using FastTree 2 (Price *et al.* 2010) using standard settings and 1 000 bootstraps (Felsenstein 1985). The ML analysis was done using RAXML (v. 8.2.12, Stamatakis 2014) with the

Table 3. Details of strains included in the phylogenetic analyses.

Species name	Strain ¹	Substrate	Country	GenBank accession number ²							
				<i>acl1</i>	<i>CaM</i>	ITS	LSU	<i>rpb1</i>	<i>rpb2</i>	<i>tef1</i>	<i>tub2</i>
<i>Albonectria albosuccinea</i>	NRRL 20459	Unidentified tree	Venezuela	—	—	JAADYS010000048.1*	JAADYS010000048.1*	JX171471	JX171585	JAADYS010002360.1*	—
<i>A. rigidiuscula</i>	CBS 133754	<i>Bauhinia longicupsis</i>	French Guiana	—	—	MW827602	MW827641	MW834177	MW833995	MW834269	—
<i>Atractium crassum</i>	CBS 180.31 ^T = NRRL 20894	Water tap	Germany	—	—	KM231790	MH866623	MW834178	HQ897722	KM231919	—
<i>At. stilbaster</i>	DAOM 215627	Cut stump	Canada	—	—	—	HQ843769	—	HQ897748	—	—
<i>Bisfusarium delphinoides</i>	CBS 110140 = FRC E-0073 = NRRL 36160	Human eye	USA	—	—	MW827603	MW827642	JX171535	HM347219	EU926302	—
<i>B. dimerum</i>	CBS 108944 ^{ET} = NRRL 36140	Human blood	Netherlands	—	—	JQ434586	JQ434514	—	HM347218	KR673912	—
<i>B. nectrioides</i>	CBS 176.31 ^T = NRRL 20689	Humus	Honduras	—	—	EU926245	EU926245	JX171477	JX171591	EU926312	—
<i>B. penzigi</i>	CBS 116508 = ATCC 15621 = NRRL 20711	Human eye	Sri Lanka	—	—	EU926256	EU926256	JX171482	HM347217	EU926323	—
<i>Corinectria fuckeliana</i>	CBS 239.29 = IMI 039700	<i>Picea sitchensis</i>	Scotland	—	—	MW827604	MW827643	MW834179	MW833996	DQ789728	—
<i>Co. tsugae</i>	CBS 788.69 ^T	<i>Tsuga heterophylla</i>	Canada	—	—	KM231763	KM231763	—	KM231763	MW834270	—
<i>Cosmospora butyri</i>	CBS 301.38 ^T = MUCL 9950	Butter	Denmark	—	—	MW827605	MW827644	MW834180	HQ897729	—	—
<i>Cs. coccinea</i>	CBS 341.70	<i>Inonotus nodulosus</i> on <i>Fagus sylvatica</i>	Germany	—	—	MH859703	KM231692	MW834181	HQ897777	KM231947	—
<i>Cs. khandalensis</i>	CBS 356.65 ^T = ATCC 16091 = IMI 112790 = MUCL 7974	<i>Bambusa</i> sp.	India	—	—	MH858608	NG_069711	—	MW833997	—	—
<i>Cs. lavitskiae</i>	CBS 530.68 ^T = ATCC 18666 = IMI 133984	Plant debris	Ukraine	—	—	KU563624	HQ231997	—	MW833998	MW834271	—
<i>Cs. viridescens</i>	CBS 102433	<i>Tilia</i> sp.	Czech Republic	—	—	KJ676148	KJ676185	MW834182	MW833999	KJ676343	—
<i>Cosmospora cavisperma</i>	CBS 172.31 ^{ET} = NRRL 13996	<i>Pinus sylvestris</i>	Norway	—	—	MW827606	MW827645	JX171465	MW834000	—	—
<i>Cyanonectria buxi</i>	CBS 125551 ^{ET}	Dead terminal branches connected with alive <i>Buxus sempervirens</i> var. <i>elegantissima</i>	Slovenia	—	—	NR_145049	MH875034	MW834183	MW834001	KM231939	—
<i>C. cyanostoma</i>	CBS 101734 ^{ET} = CBS 115512 = GJS 98-127	<i>Buxus sempervirens</i>	France	—	—	FJ474076	MH874353	MW834184	MW834002	HM626647	—
<i>Dialonectria episphaeria</i>	CBS 125494	Old ascomycete ascomata	Canada	—	—	MH863609	MH875085	MW834185	HQ897756	KM231953	—
<i>D. ullevolea</i>	CBS 125493	Ascomycete on <i>Fagus americana</i>	USA	—	—	KM231821	KM231696	—	HQ897782	KM231952	—
<i>Fusarium acutatum</i>	CBS 402.97 ^T = BBA 69580 = FRC O-1117 = NRRL 13309	Unknown	India	—	MW402459	—	—	MW402653	MW402768	MW402125	MW402323
<i>F. agapanthi</i>	NRRL 54463 ^T	<i>Agapanthus</i> sp.	Australia	—	KU900611	—	—	KU900620	KU900625	KU900630	KU900635
<i>F. ananatum</i>	CBS 118516 ^T = CMW 18685 = MRC 8165	<i>Ananas comosus</i> fruit	South Africa	—	LT996175	—	—	LT996188	LT996137	LT996091	LT996112
<i>F. andiyazi</i>	CBS 119857 ^T = NRRL 31727	<i>Sorghum bicolor</i> soil debris	South Africa	—	LT996176	—	—	LT996189	LT996138	LT996092	LT996113
<i>F. anthophilum</i>	CBS 737.97 = DAOM 225119 = FRC M-1355 = IMI 375325 = NRRL 13602	<i>Hippeastrum</i> sp.	Germany	—	LT996177	—	—	LT996190	LT996139	LT996093	LT996114
<i>F. bactridioides</i>	NRRL 20476	<i>Cronartium conigenum</i>	USA	—	AF158343	—	—	Not public	Not public	AF160290	U34434
<i>F. begoniae</i>	CBS 403.97 ^T = BBA 67781 = DAOM 225116 = IMI 375315 = NRRL 25300	<i>Begonia elatior</i> hybrid	Germany	—	AF158346	—	—	LT996191	LT996140	AF160293	U61543
<i>F. beomiforme</i>	CBS 740.97 = BBA 65829 = DAOM 225123 = IMI 375328 = NRRL 25174	Soil	New Caledonia	—	—	U61674	U61648	JX171506	JX171619	PVQB02000800*	—

(continued on next page)

Table 3. (Continued).

Species name	Strain ¹	Substrate	Country	GenBank accession number ²							
				<i>acl1</i>	<i>CaM</i>	ITS	LSU	<i>rpb1</i>	<i>rpb2</i>	<i>tef1</i>	<i>tub2</i>
<i>F. brevicatenuatum</i>	CBS 404.97 ^T = BBA 69197 = DAOM 225122 = IMI 375329 = NRRL 25446	<i>Striga asiatica</i>	Madagascar	—	MW834108	—	—	—	MN534295	MN533995	MN534063
<i>F. buhanicum</i>	CBS 796.70 = ATCC 24135 = BBA 11122 = DSM 62165 = FRC R-4955 = IMI 141195 = NRRL 13371	<i>Hibiscus cannabinus</i>	Iran	—	—	U34581	U34552	JX171449	JX171563	—	—
<i>F. bulbicola</i>	CBS 220.76 ^T = BBA 12293 = BBA 63628 = DAOM 225114 = IMI 202877 = IMI 375322 = NRRL 13618	<i>Nerine bowdenii</i>	Germany	—	KF466327	—	—	KF466394	KF466404	KF466415	KF466437
<i>F. circinatum</i>	CBS 405.97 ^T = BBA 69720 = DAOM 225113 = IMI 375321 = MRC 7541 = NRRL 25331	<i>Pinus radiata</i>	USA	—	KM231393	—	—	JX171510	HM068354	KM231943	KM232080
<i>F. coicis</i>	NRRL 66233 ^T = RBG 5368	<i>Coix gasteenii</i>	Australia	—	LT996178	—	—	KP083269	KP083274	KP083251	LT996115
<i>F. compactum</i>	NRRL 13829	River sediments	Japan	—	—	—	—	JX171460	JX171574	—	—
<i>F. concentricum</i>	CBS 450.97 ^T = BBA 64354 = CBS 833.85 = DAOM 225146 = IMI 375352 = NRRL 25181	<i>Musa sapientum</i>	Costa Rica	—	AF158335	—	—	LT996192	JF741086	AF160282	U61548
<i>F. cugenangense</i>	CBS 130308 = NRRL 25387 = ATCC 26225	Human toe nail	New Zealand	—	—	MW827607	MW827646	JX171512	JX171625	MH485011	—
<i>F. curvatum</i>	CBS 744.97 = IMI 375335 = NRRL 22902	<i>Pseudotsuga menziesii</i>	USA	—	AF158365	—	—	LT996203	LT575065	AF160312	U34424
<i>F. denticulatum</i>	CBS 735.97 = NRRL 25302	<i>Ipomoea batatas</i>	USA	—	AF158322	—	—	LT996195	LT996143	AF160269	U61550
<i>F. dlaminii</i>	CBS 119860 ^T = BBA 69859 = FRC M-1637 = MRC 3032 = NRRL 13164	Soil debris in cornfield	South Africa	—	AF158330	—	—	KU171681	KU171701	AF160277	U34430
<i>F. echinatum</i>	CBS 146496 = CPC 30814 CBS 146497 ^T = CPC 30815	Unidentified tree Unidentified tree	South Africa South Africa	— —	MW834109 MW834110	— —	— —	MW834186 MW834187	MW834003 MW834004	MW834272 MW834273	MW834300 MW834301
<i>F. equiseti</i>	CBS 245.61 = NRRL 20697	<i>Beta vulgaris</i>	Chile	—	—	MH858038	MH869603	JX171481	JX171595	—	—
<i>F. flocciferum</i>	CBS 831.85 = BBA 64346 = NRRL 25473	<i>Triticum aestivum</i>	Germany	—	—	—	MW827647	JX171514	JX171627	—	—
<i>F. fracticaudum</i>	CBS 137234 ^{PT} = CMW 25237	<i>Pinus maximonii</i>	Colombia	—	LT996179	—	—	LT996196	LT996144	KJ541059	KJ541051
<i>F. fractiflexum</i>	NRRL 28852 ^T	<i>Cymbidium</i> sp.	Japan	—	AF158341	—	—	Not public	LT575064	AF160288	AF160315
<i>F. fredkrugeri</i>	CBS 144209 ^T = CPC 33747	<i>Melhanina acuminata</i> rhizosphere	South Africa	—	LT996181	—	—	LT996199	LT996147	LT996097	LT996117
<i>F. fujikuroi</i>	CBS 221.76 ^T = BBA 12428 = BBA 63630 = IHEM 3821 = IMI 196086 = IMI 202879 = NRRL 13620 = NRRL 13998 = NRRL 22174 NRRL 13566 = ATCC 38941 = DAOM 225143 = IMI 300793 = IMI 375349 = NRRL 5538 = NRRL A-26483	<i>Oryza sativa</i> <i>Oryza sativa</i>	Taiwan China	— —	— AF158332	MW827608 —	MW827648 —	MW834188 JX171456	MW834005 JX171570	AF160279	— U34415
<i>F. globosum</i>	CBS 428.97 ^T = DAOM 214966 = FRC M-8014 = IMI 375330 = MRC 6647 = NRRL 26131 = PREM 51878	<i>Zea mays</i>	South Africa	—	KF466329	—	—	KF466396	KF466406	KF466417	KF466439
<i>F. graminearum</i>	CBS 123657 = NRRL 31084	<i>Zea mays</i>	USA	—	—	DQ459823	DQ459823	JX171531	JX171644	AY452957	—
<i>F. heterosporum</i>	CBS 720.79 = NRRL 20693	<i>Claviceps purpurea</i> on <i>Lolium perenne</i>	Netherlands	—	—	MW827609	MW827649	JX171480	JX171594	JAAGWP010000622.1*	—
<i>F. inflexum</i>		<i>Vicia faba</i>	Germany	—	AF158366	—	—	JX171469	JX171583	AF008479	U34435

Table 3. (Continued).

Species name	Strain ¹	Substrate	Country	GenBank accession number ²									
				<i>acl1</i>	<i>CaM</i>	ITS	LSU	<i>rpb1</i>	<i>rpb2</i>	<i>tef1</i>	<i>tub2</i>		
	CBS 716.74 ^T = ATCC 32213 = BBA 63203 = DAOM 225130 = DSM 63203 = IMI 375336 = NRRL 20433												
<i>F. konzum</i>	CBS 119849 ^T = MRC 8427	<i>Sorghastrum nuttans</i>	USA	—	LT996182	—	—	LT996200	LT996148	LT996098	LT996118		
<i>F. lactis</i>	CBS 411.97 ^{ET} = BBA 68590 = DAOM 225145 = IMI 375351 = NRRL 25200	<i>Ficus carica</i>	USA	—	AF158325	—	—	LT996201	LT996149	AF160272	U61551		
<i>F. lateritium</i>	NRRL 13622 = NRRL A-26433	<i>Ulmus</i> sp.	USA	—	—	—	—	JX171457	JX171571	—	—		
<i>F. longipes</i>	NRRL 20723 = IMI 265540	Unknown	England	—	—	—	—	JX171483	JX171596	—	—		
<i>F. mangiferae</i>	NRRL 25226 = BBA 69662 = DAOM 225155 = IMI 304063 = IMI 375361	<i>Mangifera indica</i>	Israel	—	AF158334	—	—	JX171509	HM068353	AF160281	U61561		
" <i>F.</i> " <i>melanochlorum</i>	CBS 202.65 = ATCC 16069 = BBA 9831 = DSM 62248 = NRRL 36353	<i>Fagus sylvatica</i>	Austria	—	—	MH858541	MH870179	JX171537	JX171649	—	—		
<i>F. mexicana</i>	NRRL 47473	<i>Mangifera indica</i>	Mexico	—	GU737389	—	—	LR792579	LR792615	GU737416	GU737308		
<i>F. napiforme</i>	CBS 748.97 ^T = BBA 69861 = DAOM 225147 = FRC M-3563 = IMI 375353 = MRC 4144 = NRRL 13604	<i>Pennisetum typhoides</i>	Namibia	—	AF158319	—	—	HM347136	EF470117	AF160266	U34428		
<i>F. nurragi</i>	CBS 392.96 = NRRL 36452	Soil	Australia	—	—	MW827610	MW827650	JX171538	JX171650	JAALX1010000436.1*	—		
<i>F. nygamai</i>	CBS 749.97 ^T = ATCC 58555 = BBA 69862 = DAOM 225148 = FRC M-1375 = IMI 375354 = NRRL 13448	<i>Sorghum bicolor</i>	Australia	—	AF158326	—	—	LT996202	EF470114	AF160273	U34426		
<i>F. parvisorum</i>	CBS 137236 ^T	<i>Pinus patula</i>	Colombia	—	LT996183	—	—	—	LT996150	KJ541060	KJ541055		
<i>F. phyllophilum</i>	CBS 216.76 ^T = BBA 11730 = BBA 63625 = DAOM 225132 = IMI 202874 = IMI 375338 = NRRL 13617	<i>Dracaena deremensis</i>	Italy	—	KF466333	—	—	KF466399	KF466410	KF466421	KF466443		
<i>F. poae</i>	NRRL 13714 = FRC T-503 = MRC 2181	Overwintered wheat	Canada	—	—	—	—	JX171458	JX171572	—	—		
<i>F. prieskaense</i>	CPC 30825	<i>Aloidendron dichotomum</i>	South Africa	—	MW834111	—	—	MW834189	MW834006	MW834274	MW834302		
	CBS 146498 ^T = CPC 30826	<i>Prunus spinosa</i>	South Africa	—	MW834112	—	—	MW834190	MW834007	MW834275	MW834303		
	CBS 146499 = CPC 30827	<i>Prunus spinosa</i>	South Africa	—	MW834113	—	—	MW834191	MW834008	MW834276	MW834304		
<i>F. phyllophilum</i>	CBS 217.76 = BBA 11341 = BBA 63624 = DAOM 225133 = IMI 202873 = IMI 375339 = NRRL 22944	<i>Cattleya</i> sp.	Germany	—	KF466333	U34558	U34529	JX171504	JX171617	AF160280	KF466443		
<i>F. pseudocircinatum</i>	CBS 449.97 ^T = ATCC 24379 = BBA 69636 = CBS 126.73 = IMI 105384 = NRRL 22946	<i>Solanum</i> sp.	Ghana	—	AF158324	—	—	LT996204	LT996151	AF160271	U34427		
<i>F. pseudograminearum</i>	CBS 109956 ^T = NRRL 28062	<i>Hordeum vulgare</i>	Australia	—	—	DQ459871	DQ459871	JX171524	JX171637	AF212468	—		
<i>F. pseudonygamai</i>	CBS 417.97 ^T = BBA 69552 = FRC M-1166 = IMI 375342 = NRRL 13592	<i>Pennisetum typhoides</i>	Nigeria	—	AF158316	—	—	LT996205	LT996152	AF160263	U34421		
<i>F. ramigenum</i>	CBS 418.98 ^T = BBA 68592 = DAOM 225137 = IMI 375343 = NRRL 25208	<i>Ficus carica</i>	USA	—	KF466335	—	—	KF466401	KF466412	KF466423	KF466445		
<i>F. redolens</i>	CBS 743.97 = DAOM 225128 = IMI 375334 = NRRL 22901	<i>Pseudotsuga menziesii</i>	Canada	—	—	U34565	U34536	JX171503	JX171616	MT409452	—		

(continued on next page)

Table 3. (Continued).

Species name	Strain ¹	Substrate	Country	GenBank accession number ²							
				<i>acl1</i>	<i>CaM</i>	ITS	LSU	<i>rpb1</i>	<i>rpb2</i>	<i>tef1</i>	<i>tub2</i>
<i>F. sacchari</i>	CBS 223.76 ^{ET} = BBA 63340 = DAOM 225138 = IMI 202881 = NRRL 13999	<i>Saccharum officinarum</i>	India	—	AF158331	—	—	JX171466	JX171580	AF160278	U34414
<i>F. sambucinum</i>	CBS 146.95 = BBA 64226 = NRRL 22187 = NRRL 20727	<i>Solanum tuberosum</i>	England	—	—	—	—	JX171493	JX171606	MW834277	—
<i>F. sarcochroum</i>	CBS 745.79 = BBA 63714 = NRRL 20472	<i>Viscum album</i>	Switzerland	—	—	MW827611	MW827651	JX171472	JX171586	MW834278	—
<i>F. scirpi</i>	NRRL 13402	Soil	Australia	—	—	GQ505681	GQ505681	JX171452	JX171566	GQ505592	—
<i>F. sororula</i>	CBS 137242 ^T = CMW 40578	<i>Pinus patula</i>	Colombia	—	LT996184	—	—	LT996206	LT996153	KJ541067	KJ541057
<i>Fusarium</i> sp.	CBS 102163 = GJS 84-426	Bamboo	Venezuela	—	—	KM231812	KM231681	MW834193	MW834009	KM231940	—
<i>F. sterilithyposum</i>	NRRL 25623	Mango	South Africa	—	AF158353	—	—	MW402713	MN193897	AF160300	AF160316
<i>F. stilboides</i>	NRRL 20429 = ATCC 15662	<i>Coffea</i> sp.	Nyasaland	—	—	—	—	JX171468	JX171582	—	—
<i>F. subglutinans</i>	CBS 747.97 ^{ET} = BBA 62451 = DAOM 225141 = FRC M-36 = MRC 8554 = NRRL 22016 = NRRL 22114	<i>Zea mays</i>	USA	—	AF158342	—	—	JX171486	JX171599	AF160289	U34417
<i>F. sublunatum</i>	CBS 189.34 ^T = BBA 62431 = DSM 62431 = NRRL 20840 = NRRL 13384	Soil	Costa Rica	—	—	HQ897830	KM231680	JX171451	JX171565	—	—
<i>F. succisae</i>	CBS 219.76 ^{ET} = BBA 12287 = BBA 63627 = DAOM 225142 = IMI 202876 = IMI 375347 = NRRL 13613	<i>Succisa pratensis</i>	Germany	—	AF158344	—	—	LT996207	LT996154	AF160291	U34419
<i>F. sudanense</i>	CBS 454.97 ^T = BBA 65862 = NRRL 25451 = NRRL 26793	<i>Striga hermonthica</i>	Sudan	—	LT996185	—	—	LT996208	LT996155	KU711697	KU603909
<i>F. temperatum</i>	NRRL 25622 = NRRL 26616	<i>Zea mays</i>	South Africa	—	AF158354	—	—	Not public	Not public	AF160301	AF160317
<i>F. terricola</i>	CBS 483.94 ^T = FRC M-1650	Soil	Australia	—	KU603951	—	—	LT996209	LT996156	KU711698	KU603908
<i>F. thapsinum</i>	CBS 733.97 = DAOM 225109 = IMI 375317 = MRC 6002 = NRRL 22045	<i>Sorghum bicolor</i>	South Africa	—	LT996186	—	—	JX171487	JX171600	AF160270	U34418
<i>F. tjaetaba</i>	CBS 144400 ^T = NRRL 66243 = RBG 5361	<i>Sorghum interjectum</i>	Australia	—	LT996187	—	—	MW834192	KP083275	KP083263	GU737296
<i>F. torreyae</i>	CBS 133858 ^T = NRRL 54151	<i>Torreyia</i> sp.	USA	—	—	HM068344	MW827652	JX171548	JX171660	HM068337	—
<i>F. tricinatum</i>	CBS 393.93 ^{ET} = BBA 64485 = NRRL 25481	Winter wheat culm base	Germany	—	—	HM068317	HM068317	JX171516	JX171629	AB674263	—
<i>F. tupiense</i>	NRRL 53984	<i>Mangifera indica</i>	Brazil	—	GU737377	—	—	LR792583	LR792619	GU737404	GU737296
<i>F. udum</i>	CBS 178.32 = BBA 1813 = DAOM 225111 = IMI 375319 = NRRL 22949	<i>Lactarius pubescens</i>	Germany	—	AF158328	—	—	LT996220	LT996172	AF160275	U34433
<i>F. venenatum</i>	NRRL 22196 = BBA 65031	<i>Zea mays</i>	Germany	—	—	—	—	JX171494	JX171607	—	—
<i>F. verticillioides</i>	CBS 734.97 = BBA 62264 = IMI 375318 = NRRL 22172	<i>Zea mays</i>	Germany	—	AF158315	—	—	LT996221	EF470122	AF160262	U34413
<i>F. xylarioides</i>	CBS 258.52 ^{ET} = NRRL 25486	<i>Coffea</i> sp.	Ivory Coast	—	—	—	—	JX171517	HM068355	AY707136	AY707118
<i>Fusicolla acetilireae</i>		Polluted soil	Japan	—	—	HQ897790	U88108	—	HQ897701	—	—

Table 3. (Continued).

Species name	Strain ¹	Substrate	Country	GenBank accession number ²									
				<i>acl1</i>	<i>CaM</i>	ITS	LSU	<i>rpb1</i>	<i>rpb2</i>	<i>tef1</i>	<i>tub2</i>		
	BBA 63789 ^T = IMI 181488 = NRRL 20827												
<i>Fu. aquaeductum</i>	BBA 63789 = IMI 181488 = NRRL 20827	Polluted soil	Japan	HQ897839	—	HQ897790	U88108	—	HQ897701	—	—	—	—
	CBS 734.79 = BBA 63669 = NRRL 20686	Drinking water	Germany	—	—	MW827612	MW827653	JX171476	HQ897742	MW847905	—	—	—
	CBS 268.53	Rubber tubing	Netherlands	—	—	MH857190	MH868728	—	—	—	—	—	—
	CBS 837.85 ^{ET} = BBA 64559 = NRRL 20865 = NRRL 37595	Plug in water tap	Germany	—	—	KM231823	KM231699	—	—	—	—	—	KM232094
<i>Fu. betae</i>	BBA 64317 ^{ET}	<i>Triticum aestivum</i>	Germany	HQ897917	—	MH855265	MH866717	—	HQ897781	—	—	—	—
<i>Fu. bharatavarshae</i>	NFCCI 4423 ^T	<i>Avicennia marina</i>	India	—	—	MK152510	MK152511	—	MK157022	—	—	—	MK376462
<i>Fu. cassiae-fistulae</i>	MFLUCC 19-0318 ^T	<i>Cassia fistula</i>	Thailand	—	—	MT215497	MT215549	—	—	—	—	—	—
<i>Fu. epistroma</i>	BBA 62201 ^{ET} = ATCC 24369 = IMI 85601 = NRRL 20439 = NRRL 20461	<i>Diatrypella</i> sp., on <i>Betula</i> sp.	England	HQ897901	—	—	AF228352	—	HQ897765	—	—	—	—
<i>Fu. gigantispora</i>	HKAS 101990	<i>Bruguiera</i> sp.	Thailand	—	—	MN047106	MN017870	—	—	—	—	—	—
	MFLU 161206 ^T	<i>Avicennia marina</i>	Thailand	—	—	MN047105	MN017876	—	—	—	—	—	—
<i>Fu. matuoi</i>	CBS 581.78 = ATCC 18694 = MAFF 238445 = NRRL 20427	<i>Albizia julibrissin</i>	Japan	HQ897858	—	KM231822	KM231698	MW834194	HQ897720	KM231954	KM232093	—	—
<i>Fu. melogrammae</i>	CBS 141092 ^T	<i>Melogramma campylosporum</i> on <i>Carpinus</i> sp.	England	—	—	KX897140	KY092489	—	HQ897720	—	—	—	MW834305
<i>Fu. meniscoidea</i>	CBS 110189 = FRC E-0086	Soil	Australia	MW834043	—	MW827613	MW827654	—	MW834010	MW834279	MW834306	—	—
<i>Fu. merismoides</i>	CBS 186.34 = BBA 1867a = NRRL 20895	<i>Acer</i> sp.	Germany	—	—	MH855482	MH866963	—	—	—	—	—	—
<i>Fu. ossicola</i>	CBS 140161 ^T	Bone of wild boar	Belgium	—	—	MF628022	MF628021	—	MW834011	MW834280	MW834307	—	—
<i>Fu. quarantena</i>	URM 8367 ^T = CBS 141541	<i>Melocactus zehntneri</i>	Brazil	—	—	MW553789	MW553788	—	MW556626	MW556625	MW556624	—	—
<i>Fu. septimanifiniscientiae</i>	CBS 144935 ^T	Soil	Netherlands	—	—	MK069422	MK069418	—	—	MK077808	MK069408	—	—
<i>Fu. siamensis</i>	MFLUCC 17-2577 ^T	<i>Cassia fistula</i>	Thailand	—	—	MT215498	MT215550	—	—	—	—	—	—
<i>Fu. sporellula</i>	CBS 110191 = FRC E-0139	Soil	South Africa	MW834044	—	MW827614	MW827655	—	MW834012	MW834281	MW834308	—	—
<i>Fu. violacea</i>	CBS 634.76 ^T = BBA 62461 = NRRL 20896	<i>Quadraspidiotus pemiciosus</i>	Iran	—	—	KM231824	U88112	MW834195	HQ897696	KM231956	KM232095	—	—
<i>Geejeyessia atrofusca</i>	CBS 125482 = DAOM 238117	<i>Staphylea trifolia</i>	Canada	—	—	MH863592	MH875066	MW834196	HQ897775	MW834282	—	—	—
	NRRL 22316	<i>Staphylea trifolia</i>	USA	—	—	AF178423	AF178392	JX171496	EU329502	AF178361	—	—	—
<i>G. cellidicola</i>	CBS 125502 ^T	<i>Celtis occidentalis</i>	Canada	HM626625	—	HM626657	HM626669	MW834197	MW834013	HM626638	KM232074	—	—
<i>G. cicatricum</i>	CBS 125550	Dead twig connected with alive <i>Buxus sempervirens</i> var. <i>elegantissima</i>	Slovenia	—	—	HM626654	HM626666	MW834198	HQ897697	HM626642	—	—	—
	CBS 125552	Dead twig	Slovenia	HQ728171	—	HQ728145	MH875038	—	HQ728153	HM626644	—	—	—
<i>Ilyonectria capensis</i>	CBS 132815 ^T	<i>Protea</i> sp.	South Africa	—	—	NR_152887	NG_070049	MW834199	MW834014	JX231119	—	—	—
<i>I. destructans</i>	CBS 264.65	<i>Cyclamen persicum</i>	Sweden	—	—	MH858563	KM515927	—	MW834015	JF735695	—	—	—
<i>Luteonectria albida</i>	CBS 102683 = GJS 99-73 = GJS 8522A	Tree bark	Costa Rica	—	—	MW827615	MH874402	MW834200	MW834016	MW834283	—	—	—

(continued on next page)

Table 3. (Continued).

Species name	Strain ¹	Substrate	Country	GenBank accession number ²							
				<i>acl1</i>	<i>CaM</i>	ITS	LSU	<i>rpb1</i>	<i>rpb2</i>	<i>tef1</i>	<i>tub2</i>
	NRRL 22152 ^T = NRRL 13950	Woody stem bark	Jamaica	—	—	JABFEP010000142.1*	JABFEP010000142.1*	JX171492	JX171605	JABFEP010002685.1*	—
<i>L. nematophila</i>	NRRL 54600	Unknown	Germany	—	—	JABFFA010000104.1*	JABFFA010000104.1*	JX171552	JX171664	JABFFA010003988.1*	—
<i>Macroconia bulbipes</i>	CBS 146678 = CPC 37137	<i>Erica</i> sp. associated with <i>Dimerosporiopsis engleriana</i>	South Africa	MW834045	MW834114	MW827616	MW827656	MW834201	MW834017	—	MW834309
	CBS 146679 ^T = CPC 37138	<i>Erica</i> sp. associated with <i>Dimerosporiopsis engleriana</i>	South Africa	MW834046	MW834115	MW827617	MW827657	MW834202	MW834018	—	MW834310
<i>Ma. cupularis</i>	HMAS 173240 ^T	<i>Stylobothis</i> sp. on unidentified tree	China	—	—	EF121864	EF121870	—	—	—	—
<i>Ma. gigas</i>	HMAS 173239 ^T	Rotten stem of bamboo associated with other fungi	China	—	—	EF121853	EF121869	—	—	—	—
<i>Ma. leptosphaeriae</i>	CBS 100001	<i>Leptosphaeria</i> on dead stem of <i>Urtica dioica</i>	Netherlands	HQ897891	MW834116	HQ897810	HQ897755	MW834203	HQ728164	KM231959	KM232097
<i>Ma. papilionacearum</i>	CBS 125495	Ascomycete on <i>Fabaceae</i>	USA	HQ897912	MW834117	HQ897826	MH875086	MW834204	HQ897776	—	KM232096
<i>Ma. phlogioides</i>	CBS 125496	<i>Quercus</i> sp., branch in stream	USA	HQ897868	MW834118	MW827618	MW827658	MW834205	HQ897732	MW834284	MW834311
	CBS 146500 = CPC 35388	<i>Encephalartos</i> sp. leaf	South Africa	MW834047	MW834119	MW827619	MW827659	MW834206	MW834019	—	MW834312
	CBS 146501 ^T = CPC 35389	<i>Encephalartos</i> sp. leaf	South Africa	MW834048	MW834120	MW827620	MW827660	MW834207	MW834020	—	MW834313
<i>Ma. sphaeriae</i>	CBS 717.74	Pyrenomycete on <i>Coronilla emerus</i>	France	MW834049	MW834121	MW827621	MW827661	—	KM232390	—	KM232099
	CBS 112770	<i>Cucurbitaria laburni</i> on <i>Laburnum anagyroides</i>	Austria	KM231061	KM231413	MW827622	MW827662	MW834208	MW834021	—	KM232098
<i>Mariannaea elegans</i>	DAOM 226709	<i>Betula</i> sp.	Canada	—	—	—	HQ843768	—	HQ897747	—	—
<i>M. samuelsii</i>	CBS 125515 ^T = DAOM 235814	Soil	Guatemala	—	—	NR_137767	NG_060269	—	HQ897752	—	—
<i>Microcera coccophila</i>	CBS 310.34 = NRRL 13962	Scale insect	Italy	—	—	MH855540	KM231703	JX171462	JX171576	—	—
<i>Mi. diploa</i>	CBS 735.79 = BBA 61173 = NRRL 36545	<i>Quadraspidiotus perniciosus</i>	Iran	—	—	MW827623	MW827663	JX171463	JX171577	—	—
<i>Mi. larvarum</i>	CBS 738.79 = BBA 62239 = DSM 62239 = MUCL 19033 = NRRL 20473	<i>Quadraspidiotus perniciosus</i>	Iran	—	—	KM231825	KM231701	JX171473	JX171587	KM231957	—
<i>Mi. rubra</i>	CBS 638.76 ^T = BBA 62460 = NRRL 20475; NRRL 22111; NRRL 22170	<i>Quadraspidiotus perniciosus</i> on <i>Prunus domestica</i>	Iran	HQ897903	KM231409	MH861019	MH872790	—	HQ897767	—	MW834314
<i>Microcera</i> sp.	NRRL 26790	<i>Parmelia rudecta</i>	USA	—	—	—	—	JX171523	JX171636	—	—
<i>Nectria cinnabarina</i>	CBS 125165 ^{ET}	<i>Aesculus</i> sp.	France	KM231074	—	HM484548	HM484562	—	KM232402	HM484527	—
"Nt." <i>flavoviridis</i>	CBS 124353 = BBA 65542 = NRRL 22093	Decorticated wood	USA	—	—	HQ897791	MW827664	MW834209	HQ897702	—	—
<i>Neocosmospora acutispora</i>	CBS 145461 ^T = NRRL 22574 = BBA 62213	<i>Coffea arabica</i>	Guatemala	MW834050	MW834122	LR583700	LR583908	MW834210	LR583814	LR583593	—
<i>N. addoensis</i>	CBS 146509 = CPC 37127	<i>Citrus sinensis</i>	South Africa	MW218004	MW218051	MW173041	MW173032	MW218097	MW446574	MW248740	—
	CBS 146510 ^T = CPC 37128	<i>Citrus sinensis</i>	South Africa	MW218005	MW218052	MW173042	MW173033	MW218098	MW446575	MW248741	—
<i>N. ambrosia</i>	CBS 571.94 ^{ET} = NRRL 22346 = BBA 65390 = MAFF 246287	<i>Euwallacea fornicatus</i>	India	—	—	EU329669	EU329669	MW834211	EU329503	FJ240350	—
	NRRL 20438 = IMI 296597	<i>Xyleborus fornicatus</i>	India	—	—	AF178397	AF178366	JX171470	JX171584	NIZV01000014.1*	—
<i>N. ampla</i>	CBS 202.32 ^T = BBA 4170	<i>Coffea</i> sp.	German East Africa	MW834051	MW834123	LR583701	LR583909	MW834212	LR583815	LR583594	—
<i>N. bataticola</i>	CBS 144397 = NRRL 22400 = BBA 64683	<i>Ipomoea batatas</i>	USA	MW218006	MW218053	AF178407	AF178376	MW218099	EU329509	AF178343	—
	CBS 144398 ^T = NRRL 22402 = BBA 64954 = FRC S-0567	<i>Ipomoea batatas</i>	USA	MW218007	MW218054	AF178408	AF178377	MW218100	FJ240381	AF178344	—

Table 3. (Continued).

Species name	Strain ¹	Substrate	Country	GenBank accession number ²							
				<i>acl1</i>	<i>CaM</i>	<i>ITS</i>	<i>LSU</i>	<i>rpb1</i>	<i>rpb2</i>	<i>tef1</i>	<i>tub2</i>
<i>N. borneensis</i>	CBS 145462 ^{ET} = NRRL 22579 = BBA 65095 = GJS 85-197	Bark or recently dead tree	Indonesia	MW834052	MW834124	AF178415	AF178384	MW834213	EU329515	AF178352	—
<i>N. bostrycooides</i>	CBS 144.25 ^{NT}	Soil	Honduras	MW218008	MW218055	LR583704	LR583912	MW218101	LR583818	LR583597	—
	CBS 392.66 = NRRL 25325 = BBA 69595	<i>Bertholletia excelsa</i>	Unknown	MW218009	MW218056	LR583705	LR583913	MW218102	LR583819	LR583598	—
<i>N. brevicona</i>	CBS 204.31 ^{ET} = NRRL 22659 = BBA 2123	<i>Gladiolus</i> sp.	Indonesia	MW218010	MW218057	LR583707	LR583915	MW218103	LR583821	LR583600	—
<i>N. brevis</i>	CBS 130326 = NRRL 28009 = CDC B-5543	Human eye	USA	MW834053	MW834125	DQ094351	DQ236393	MW834214	EF470136	DQ246869	—
<i>N. catenata</i>	CBS 143228 = NRRL 54992 = UTHSC 09-1008	<i>Stegostoma fasciatum</i>	USA	MW218011	MW218058	KC808255	KC808255	MW218104	KC808354	KC808213	—
	CBS 143229 ^T = NRRL 54993 = UTHSC 09-1009	<i>Stegostoma fasciatum</i>	USA	MW218012	MW218059	KC808256	KC808256	MW218105	KC808355	KC808214	—
<i>N. citricola</i>	CBS 146512 = CPC 37130	<i>Citrus sinensis</i>	South Africa	MW218014	MW218061	MW173047	MW173035	MW218107	MW446580	MW248746	—
	CBS 146513 ^T = CPC 37131	<i>Citrus sinensis</i>	South Africa	MW218015	MW218062	MW173048	MW173036	MW218108	MW446581	MW248747	—
<i>N. crassa</i>	CBS 144386 ^T = MUCL 11420	Unknown	France	MW218016	MW218063	LR583709	LR583917	MW218109	LR583823	LR583604	—
<i>N. cryptoseptata</i>	CBS 145463 ^T = NRRL 22412 = BBA 65024	Bark	French Guiana	MW834054	MW834126	AF178414	AF178383	MW834215	EU329510	AF178351	—
<i>N. cucurbitae</i>	CBS 410.62 = NRRL 22658 = CECT 2864	<i>Cucurbita viciifolia</i>	Netherlands	MW834055	MW834127	LR583710	LR583918	MW834216	LR583824	DQ247640	—
	CBS 616.66 ^T = NRRL 22399 = BBA 64411	<i>Cucurbita viciifolia</i>	Netherlands	MW834056	MW834128	LR583711	LR583919	MW834217	LR583825	DQ247592	—
<i>N. cyanescens</i>	CBS 518.82 ^T	Human foot	Netherlands	MW218017	MW218064	AB190389	LR583920	MW218110	LR583826	LR583605	—
	CBS 637.82	Human foot	Netherlands	MW218018	MW218065	LR583712	LR583921	MW218111	LR583827	LR583606	—
<i>N. diminuta</i>	CBS 144390 ^T = MUCL 18798	<i>Coelocaryon preusii</i>	Unknown	MW834057	MW834129	LR583713	LR583922	MW834218	LR583828	LR583607	—
<i>N. elegans</i>	CBS 144395 = NRRL 22163 = MAFF 238540 = ATCC 18690	<i>Xanthoxylum piperitum</i>	Japan	MW218019	MW218066	AF178394	AF178363	MW218112	EU329496	AF178328	—
	CBS 144396 ^{ET} = NRRL 22277 = MAFF 238541 = ATCC 42366	<i>Xanthoxylum piperitum</i>	Japan	MW218020	MW218067	AF178401	AF178370	MW218113	FJ240380	AF178336	—
<i>N. epipeda</i>	CBS 146523 ^T = CPC 38310	<i>Bouvardia</i> sp. imported from Uganda	Netherlands	MW834058	MW834130	MW827624	MW827665	MW834219	MW834022	MW834285	—
	CBS 146524 = CPC 38311	<i>Bouvardia</i> sp. imported from Uganda	Netherlands	MW834059	MW834131	MW827625	MW827666	MW834220	MW834023	MW834286	—
<i>N. euwallaceae</i>	CBS 135854 ^T = NRRL 54722	<i>Euwallacea</i> sp.	Israel	—	—	JQ038014	JQ038014	JQ038021	JQ038028	JQ038007	—
<i>N. falciformis</i>	CBS 475.67 ^T = IMI 268681	Human mycetoma	Puerto Rico	MW218021	MW218068	MG189935	MG189915	MW218114	LT960558	LT906669	—
	CBS 121450	Declined grape vine	Syria	MW218022	MW218069	JX435211	JX435211	MW218115	JX435261	JX435161	—
	NRRL 43529 = CDC 2006743575	Human cornea	USA	—	—	EF453117	EF453117	JX171541	JX171653	EF452965	—
<i>N. ferruginea</i>	CBS 109028 ^T = NRRL 32437	Human subcutaneous nodule	Switzerland	MW834060	MW834132	DQ094446	DQ236488	MW834221	EU329581	DQ246979	—
	CPC 28194	<i>Citrus sinensis</i>	Italy	MW834061	MW834133	LT746276	LT746276	MW834222	LT746341	LR583602	—
<i>N. floridana</i>	NRRL 62628 ^T = MAFF 246849	<i>Euwallacea interjectus</i>	USA	—	—	KC691563	KC691563	KC691593	KC691624, KC691653	KC691535	—
<i>N. gamsii</i>	CBS 143207 ^T = NRRL 32323 = UTHSC 99-205	Human bronchoalveolar lavage fluid	USA	MW834062	MW834134	DQ094420	DQ236462	MW834223	EU329622	DQ247103	—
	CBS 143211 = NRRL 32794 = FRC S-1152	Humidifier coolant	USA	MW834063	MW834135	DQ094563	DQ236605	MW834224	EU329576	DQ246951	—
<i>N. gamtoosensis</i>	CBS 146502 ^T = VG16 = CPC 37120	<i>Citrus sinensis</i>	South Africa	MW218023	MW218070	MW173063	MW173038	MW218116	MW446611	MW248762	—
<i>N. haematococca</i>	CBS 119600 ^{ET} = FRC S-1832	Dying tree	Sri Lanka	MW834064	MW834136	KM231797	KM231664	—	LT960561	DQ247510	—

(continued on next page)

Table 3. (Continued).

Species name	Strain ¹	Substrate	Country	GenBank accession number ²							
				<i>act1</i>	<i>CaM</i>	ITS	LSU	<i>rpb1</i>	<i>rpb2</i>	<i>tef1</i>	<i>tub2</i>
<i>N. hypohenemi</i>	CBS 145464 ^T = NRRL 52782 = ARSEF 5878	<i>Hypothenemus hampei</i>	Benin	MW218024	—	LR583715	LR583923	MW218117	JF741176	JF740850	—
	CBS 145466 = NRRL 52783 = ARSEF 5879	<i>Hypothenemus hampei</i>	Uganda	MW218025	MW218071	MW827626	MW827667	MW218118	MW834024	MW834287	—
<i>N. illudens</i>	CBS 147303 = NRRL 22090 = BBA 67606 = GJS 82-98	<i>Beilschmiedia tawa</i>	New Zealand	MW834065	MW834137	AF178393	AF178362	JX171488	JX171601	AF178326	—
<i>N. ipomoeae</i>	CBS 353.87 = NRRL 22657	<i>Gerbera</i> sp.	Netherlands	MW218026	MW218072	LR583717	LR583925	MW218119	LR583831	DQ247639	—
	CBS 833.97	<i>Rosa</i> sp.	Netherlands	MW218027	MW218073	LR583719	LR583927	MW218120	LR583833	LR583611	—
<i>N. keleraja</i>	CBS 125720 ^{PT} = FRC S-1837 = GJS 02-114	Branch of unidentified tree	Sri Lanka	MW834066	MW834138	LR583720	LR583928	MW834225	LR583834	LR583612	—
	CBS 125722 ^{PT} = FRC S-1836 = GJS 02-114	Branch of unidentified tree	Sri Lanka	MW834067	MW834139	JF433039	JF433039	MW834226	LR583835	DQ247515	—
<i>N. keratoplastica</i>	CBS 490.63 ^T	Human	Japan	MW218028	MW218074	LR583721	LR583929	MW218121	LT960562	LT906670	—
	CBS 144389 = MUCL 18301	Greenhouse humic soil	Belgium	MW218029	MW218075	LR583722	LR583930	MW218122	LR583836	LR583613	—
<i>N. kuroshio</i>	CBS 142642 ^T	<i>Euwallacea</i> sp.	USA	MW834068	MW834140	LR583723	LR583931	MW834227	LR583837	KX262216	—
<i>N. kurunegalensis</i>	CBS 119599 ^T = GJS 02-94	Recently cut tree	Sri Lanka	MW834069	MW834141	JF433036	JF433036	MW834228	LR583838	DQ247511	—
<i>N. lerouxii</i>	CBS 146514 ^T = CPC 37132	<i>Citrus sinensis</i>	South Africa	MW218030	MW218076	MW173069	MW173039	MW218123	MW446617	MW248768	—
<i>N. lichenicola</i>	CBS 509.63 = MUCL 8050 = IMUR 410	Air	Brazil	MW834070	MW834142	LR583728	LR583936	MW834229	LR583843	LR583618	—
	CBS 623.92 ^{ET}	Human	Germany	MW834071	MW834143	LR583730	LR583938	—	LR583845	LR583620	—
<i>N. liiodendri</i>	CBS 117481 ^T = NRRL 22389 = BBA 67587 = GJS 91-148	<i>Liiodendron tulipifera</i>	USA	MW218031	MW218077	AF178404	AF178373	MW218124	EU329506	AF178340	—
<i>N. longissima</i>	CBS 126407 ^T = GJS 85-72	Tree bark	New Zealand	MW834072	MW834144	LR583731	LR583939	MW834230	LR583846	LR583621	—
<i>N. macrospora</i>	CBS 142424 ^T = CPC 28191	<i>Citrus sinensis</i>	Italy	MW218032	MW218078	LT746266	LT746281	MW218125	LT746331	LT746218	—
	CPC 28193	<i>Citrus sinensis</i>	Italy	MW218033	MW218079	LT746268	LT746283	MW218126	LT746333	LT746220	—
<i>N. mahasenii</i>	CBS 119594 ^T	Dead branch on live tree	Sri Lanka	MW834073	MW834145	JF433045	JF433045	MW834231	LT960563	DQ247513	—
<i>N. martii</i>	CBS 115659 ^{ET} = FRC S-0679 = MRC 2198	<i>Solanum tuberosum</i>	Germany	MW834074	MW834146	JX435206	JX435206	MW834232	JX435256	JX435156	—
<i>N. merxiana</i>	CBS 146525 ^T	<i>Chrysanthemum</i> sp. imported from Uganda	Netherlands	MW834075	MW834147	MW827627	MW827668	MW834233	MW834025	MW834288	—
	CBS 146526	<i>Chrysanthemum</i> sp. imported from Uganda	Netherlands	MW834076	MW834148	MW827628	MW827669	MW834234	MW834026	MW834289	—
<i>N. metavorans</i>	CBS 135789 ^T	Human pleural effusion	Greece	MW218034	MW218080	LR583738	LR583946	MW218127	LR583849	LR583627	—
	CBS 143219 = NRRL 46708 = FMR 8634	Human foot	Spain	MW218035	MW218081	LR583744	LR583948	MW218128	LR583851	LR583629	—
<i>N. mori</i>	CBS 145467 ^T = NRRL 22230 = MAFF 238539	<i>Morus alba</i>	Japan	MW834077	MW834149	DQ094305	DQ236347	MW834235	EU329499	AF178358	—
	CBS 145468 = NRRL 22157 = MAFF 238538	<i>Morus alba</i>	Japan	MW834078	MW834150	DQ094306	DQ236348	MW834236	EU329493	AF178359	—
<i>N. neerlandica</i>	CBS 232.34 ^T	<i>Pisum sativum</i>	Netherlands	MW834079	MW834151	MW827629	MW827670	MW834237	MW847903	MW847906	—
<i>N. nelsonii</i>	CBS 309.75 ^T	<i>Pisum sativum</i>	Unknown	MW834080	MW834152	MW827630	MW827671	MW834238	MW847904	MW847907	—
<i>N. nirenbergiana</i>	CBS 145469 ^T = NRRL 22387 = BBA 65023 = GJS 87-127	Bark	French Guiana	MW834081	MW834153	AF178403	AF178372	—	EU329505	AF178339	—
<i>N. noneumartii</i>	CBS 115658 ^T = FRC S-0661	<i>Solanum tuberosum</i>	Israel	MW218036	MW218082	LR583745	LR583949	MW218129	MW446618	LR583630	—
<i>N. obliquiseptata</i>	NRRL 62611 = MAFF 246845	<i>Euwallacea</i> sp.	Australia	—	—	KC691576	KC691576	KC691606	KC691637, KC691666	KC691548	—

Table 3. (Continued).

Species name	Strain ¹	Substrate	Country	GenBank accession number ²							
				<i>acl1</i>	<i>CaM</i>	<i>ITS</i>	<i>LSU</i>	<i>rpb1</i>	<i>rpb2</i>	<i>tef1</i>	<i>tub2</i>
<i>N. oblonga</i>	CBS 130325 ^T = NRRL 28008 = CDC B-4701	Human eye	USA	MW834082	MW834154	LR583746	LR583950	MW834239	LR583853	LR583631	—
<i>N. oligoseptata</i>	CBS 143241 ^T = NRRL 62579 = FRC S-2581 = MAFF 246283	<i>Euwallacea validus</i>	USA	MW834083	MW834155	KC691566	KC691566	KC691596	LR583854	KC691538	—
<i>N. paraeumartii</i>	CBS 487.76 ^T = NRRL 13997 = BBA 62215	<i>Solanum tuberosum</i>	Argentina	MW834084	MW834156	LR583747	LR583951	MW834240	LR583855	DQ247549	—
<i>N. parceramosa</i>	CBS 115695 ^T	Soil	South Africa	MW218037	MW218083	JX435199	JX435199	—	JX435249	JX435149	—
<i>N. perseae</i>	CBS 144142 ^T = CPC 26829	<i>Persea americana</i>	Italy	MW218038	MW218084	LT991940	LT991947	MW218130	LT991909	LT991902	—
<i>N. petroliphila</i>	CBS 203.32 = NRRL 13952	<i>Pelargonium</i> sp.	South Africa	MW218039	MW218085	DQ094320	DQ236362	MW218131	LR583857	DQ246835	—
	CBS 224.34 = NRRL 28579	Human toenail	Cuba	MW218040	MW218086	DQ094383	DQ236425	MW218132	LR583858	DQ246910	—
<i>N. phaseoli</i>	CBS 265.50	<i>Phaseolus</i> sp.	USA	MW834085	MW834157	LR583750	LR583954	—	KJ511278	FJ919464	—
	NRRL 22276 = ATCC 38466	<i>Phaseolus vulgaris</i>	USA	—	—	EU329668	EU329668	JX171495	JX171608	AY220186	—
<i>N. piperis</i>	CBS 145470 ^T = NRRL 22570 = GJS 89-14 = CML 1888	<i>Piper nigrum</i>	Brazil	MW834086	MW834158	AF178422	AF178391	MW834241	EU329513	AF178360	—
<i>N. pisi</i>	CBS 123669 ^{ET} = NRRL 45880 = ATCC MYA-4622	Progeny of parentals from <i>Pisum sativum</i> and soil	USA	MW834087	MW834159	LR583753	LR583957	MW834242	LR583862	LR583636	—
	CBS 142372	<i>Trifolium subterraneum</i>	Germany	MW834088	MW834160	LR583755	LR583959	MW834243	LR583864	KY556454	—
<i>N. plagianthi</i>	NRRL 22632 = GJS 83-146	<i>Hoheria glabrata</i>	New Zealand	—	—	AF178417	AF178386	JX171501	JX171614	AF178354	—
<i>N. protoensiformis</i>	CBS 145471 ^T = NRRL 22178 = GJS 90-168	Dicot tree	Venezuela	MW834089	MW834161	AF178399	AF178368	MW834244	EU329498	AF178334	—
<i>N. pseudensiformis</i>	CBS 130.78 = NRRL 22575 = NRRL 22653	<i>Cocos nucifera</i>	Indonesia	MW834090	MW834162	LR583759	LR583963	MW834245	LR583868	DQ247635	—
<i>N. pseudopisi</i>	CBS 266.50	<i>Pisum sativum</i>	Unknown	MW834091	MW834163	MW827631	MW827672	MW834246	MW834027	MW834290	—
<i>N. pseudoradicicola</i>	CBS 145472 ^T = NRRL 25137 = ARSEF 2313	Diseased cocoa pods	Papua New Guinea	MW218041	MW218087	JF740899	JF740899	MW218133	JF741084	JF740757	—
<i>N. quercicola</i>	CBS 141.90 ^T = NRRL 22652	<i>Quercus cerris</i>	Italy	MW834092	MW834164	LR583760	LR583964	MW834247	LR583869	DQ247634	—
<i>N. rectiphora</i>	CBS 125726 = FRC S-1842	Dead tree	Sri Lanka	MW834093	MW834165	JF433043	JF433043	MW834248	MW834028	JF433026	—
	CBS 125727 ^T = GJS 02-89 = FRC S-1831	Dead tree	Sri Lanka	MW834094	MW834166	JF433034	JF433034	MW834249	LR583871	DQ247509	—
<i>N. regularis</i>	CBS 190.35	<i>Phaseolus</i> sp.	USA	MW834095	MW834167	LR583762	LR583966	MW834250	LR583872	LR583642	—
	CBS 230.34 ^T	<i>Pisum sativum</i>	Netherlands	MW834096	MW834168	LR583763	LR583967	—	MW834029	LR583643	—
<i>N. rekana</i>	CMW 52862 ^T	<i>Euwallacea perbrevis</i>	Indonesia	—	—	MN249094	—	—	MN249137, MN249108	MN249151	—
<i>N. robusta</i>	CBS 145473 ^T = NRRL 22395 = BBA 65682	Bark	Venezuela	—	MW834169	AF178405	LR583968	MW834251	EU329507	AF178341	—
<i>N. samuelsii</i>	CBS 114067 ^T = GJS 89-70	Bark	Guyana	MW834097	MW834170	LR583764	LR583969	MW834252	LR583874	LR583644	—
<i>N. silvicola</i>	CBS 119601 = GJS 98-135	<i>Populus nigra</i>	France	MW834098	MW834171	LR583765	LR583970	MW834253	LR583875	LR583645	—
	CBS 123846 ^T = GJS 04-147	<i>Liriodendron tulipifera</i>	USA	MW834099	MW834172	LR583766	LR583971	MW834254	LR583876	LR583646	—
<i>N. solani</i>	CBS 140079 ^{ET} = NRRL 66304 = GJS 09-1466 = FRC S-2364	<i>Solanum tuberosum</i>	Slovenia	MW218042	MW218088	KT313633	KT313633	MW218134	KT313623	KT313611	—
<i>N. spathulata</i>	CBS 145474 ^T = NRRL 28541 = UTHSC 98-1305	Human synovial fluid	USA	MW218045	MW218091	EU329674	EU329674	MW218137	EU329542	DQ246882	—
<i>N. stercicola</i>	CBS 142481 ^T = DSM 106211	Compost yard debris	Germany	MW834100	MW834173	LR583779	LR583984	MW834255	LR583887	LR583658	—

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Table 3. (Continued).

Species name	Strain ¹	Substrate	Country	GenBank accession number ²							
				<i>acl1</i>	<i>CaM</i>	ITS	LSU	<i>rpb1</i>	<i>rpb2</i>	<i>tef1</i>	<i>tub2</i>
	CBS 144388 = MUCL 18299	Greenhouse humic soil	Belgium	MW834101	MW834174	LR583780	LR583985	MW834256	LR583888	LR583659	—
<i>N. suttoniana</i>	CBS 143214 ^T = NRRL 32858	Human wound	USA	MW218046	MW218092	DQ094617	DQ236659	MW218138	EU329630	DQ247163	—
	CBS 143224 = NRRL 54972	Equine eye	USA	MW218047	MW218093	MG189940	MG189925	MW218139	KC808336	KC808197	—
<i>N. tonkinensis</i>	CBS 115.40 ^T	<i>Musa sapientum</i>	Vietnam	MW218048	MW218094	MG189941	MG189926	MW218140	LT960564	LT906672	—
	CBS 118931	<i>Solanum lycopersicum</i>	UK	MW218049	MW218095	LR583784	LR583989	MW218141	LR583891	LR583662	—
<i>N. tuaransensis</i>	NRRL 22231 ^T = ATCC 16563 = MAFF 246842	<i>Hevea brasiliensis</i> damaged by unknown ambrosia beetle	Malaysia	—	—	KC691570	KC691570	KC691600	KC691631, KC691660	KC691542	—
<i>N. vasinfecta</i>	CBS 325.54 = ATCC 16238 = IFO 7591 = IMI 251386 = NRRL 22436	Soil	South Africa	—	—	AF178412	AF178381	JX171497	JX171610	AF178348	—
	CBS 446.93 = IMI 316967 = NHL 2919	Soil	Japan	MW834102	MW834175	LR583791	LR583996	MW834257	LR583898	LR583670	—
	CBS 533.65 = IMI 302625	Unknown	India	MW834103	MW834176	LR583792	LR583997	MW834258	LR583899	LR583671	—
<i>Neonectria coccinea</i>	CBS 125484	<i>Fagus sylvatica</i>	Germany	—	—	HQ897832	MH875068	MW834259	HQ897785	—	—
<i>Ne. ditissima</i>	CBS 125486	<i>Fagus americana</i>	Canada	—	—	HQ897824	MH877864	—	HQ897774	—	—
<i>Nothofusarium devonianum</i>	CBS 147304 ^T = NRRL 22134	<i>Ruscus aculeatus</i>	United Kingdom	—	—	MW827632	MW827673	JX171490	JX171603	MW834291	—
<i>Pseudofusicolla belgica</i>	CBS 147300 = IHEM 5322	Recycled water from air-conditioning humidifier	Belgium	—	—	KJ125590	KJ126478	—	KP835473	KJ126182	—
	CBS 147301 ^T = IHEM 2413	Recycled water, spray humidifier in air-conditioned building	Belgium	—	—	KJ125588	KJ126476	—	KP835474	KJ126180	—
	CBS 147302 = IHEM 2440	Humidifier water from air-conditioning	Belgium	—	—	KJ125589	KJ126477	—	KP835475	KJ126181	—
	IHEM 2105	Recycled humidifier water from airconditioning	Belgium	—	—	KP835478	KP835480	—	KP835476	KP835484	—
<i>Rectifusarium robinianum</i>	CBS 430.91 ^T = NRRL 25729	<i>Robinia pseudoacacia</i>	Germany	—	—	KM231794	NG_058096	JX171520	JX171633	KM231923	—
<i>R. ventricosum</i>	CBS 748.79 ^T = BBA 62452 = NRRL 20846 = NRRL 22113	Wheat field soil	Germany	—	—	HQ897816	KM231658	JX171484	JX171597	KM231924	—
<i>Rugonectria castaneicola</i>	CBS 128360	Bark	China	—	—	MH864901	MH876352	MW834260	MW834030	MW834292	—
<i>Ru. neobalansae</i>	CBS 125120 = GJS 85-219	Dead tree	Indonesia	—	—	KM231750	HM364322	—	MW834031	KM231874	—
<i>Ru. rugulosa</i>	CBS 126565 = GJS 09-1245	Dead wood	Venezuela	—	—	KM231749	MH877897	MW834261	MW834032	KM231873	—
<i>Setofusarium setosum</i>	CBS 574.94 = BBA 65063	Unknown	French Guiana	—	—	MW827633	MW827674	MW834262	MW834033	MW834293	—
	CBS 635.92 ^{ET} = GJS 88-12 = NRRL 36526	Tree bark	French Guiana	—	—	MW827634	MW827675	JX171539	JX171651	MW834294	—
<i>Scolecopusarium ciliatum</i>	CBS 155.86 = NRRL 22284	<i>Hordeum vulgare</i> mouldy grain, associated with scale insects	Denmark	—	—	MW827635	MW827676	MW834263	MW834034	MW834295	—
	CBS 191.65 ^{NT} = ATCC 16068 = ATCC 24137 = BBA 9661 = DSM 62172 = IMI 112499 = NRRL 20431	<i>Fagus sylvatica</i>	Germany	—	—	MW827636	MW827677	MW834264	MW834035	MW834296	—
	CBS 144385 = IHEM 2989	<i>Fagus sylvatica</i>	Belgium	—	—	KJ125591	KJ126479	MW834265	KP835472	MW834297	—
<i>Stylonectria applanata</i>	CBS 125489	Unidentified ascomycete on <i>Betula</i> sp.	Canada	HQ897875	—	HQ897805	KM231689	—	HQ897739	KM231944	—
<i>St. carpini</i>	DAOM 235819	<i>Melanconis spodiarea</i> on <i>Carpinus betulus</i>	Austria	HQ897909	—	HQ897823	—	—	HQ897773	—	—
<i>St. corniculata</i>	CBS 125491 ^T	Unidentified ascomycete on <i>Carpinus</i> sp.	Germany	HQ897915	—	HQ897829	KM231691	—	HQ897779	KM231946	—
<i>St. hetmanica</i>	CBS 147305 ^T = CPC 38725	<i>Diaporthe</i> sp. on <i>Frangula alnus</i>	Ukraine	MW834104	—	MW827637	—	—	MW834036	—	—

Table 3. (Continued).

Species name	Strain ¹	Substrate	Country	GenBank accession number ²							
				<i>acl1</i>	<i>CaM</i>	ITS	LSU	<i>rpb1</i>	<i>rpb2</i>	<i>tef1</i>	<i>tub2</i>
<i>St. norvegica</i>	CBS 147306 = CPC 38848	<i>Dothiorella saementorum</i> on <i>Acer platanoides</i>	Ukraine	MW834105	—	MW827638	—	—	MW834037	—	—
	CBS 139239 ^T	Dead sporodochia of fusarium state on pyrenomycete (presumably <i>Amphiporthe</i> sp.)	Norway	MW834106	—	KR605485	—	—	MW834038	—	—
	CBS 139242	On sporodochia of fusarium-like on unidentified pyrenomycete	Norway	MW834107	—	MW827639	—	—	MW834039	—	—
<i>St. purtonii</i>	DAOM 235818	<i>Picea abies</i>	Germany	HQ897919	—	HQ897831	—	—	HQ897783	—	—
<i>St. qilianshanensis</i>	HMAS 255803 ^T	Unknown ascomycete on <i>Picea asperata</i>	China	MT087289	—	—	—	—	MT087288	—	—
<i>St. wegelianiana</i>	CBS 125490	<i>Hapalycystis bicaudata</i> on <i>Ulmus glabra</i>	Austria	HQ897890	—	KM231817	KM231690	—	HQ897754	KM231945	—
<i>Thelonectria discophora</i>	CBS 125487	<i>Aesculus hippocastanum</i>	Germany	—	—	HQ897789	MW827678	MW834266	HQ897700	MW834298	—
<i>T. olicia</i>	CBS 215.67 ^{MT} = ATCC 16548 = DSM 62520 = IMI 116873	<i>Asparagus officinalis</i>	Germany	—	—	MW827640	MW827679	MW834267	MW834040	MW834299	—
<i>Tumenectria laetidisca</i>	CBS 100284	Bamboo	Japan	—	—	KJ022017	KJ022066	—	MW834041	KJ022400	—
	CBS 101909 ^{ET}	Bamboo	Jamaica	—	—	KJ022018	KJ022067	MW834268	MW834042	KJ022401	—

¹ ARSEF: Collection of entomopathogenic fungal cultures, US Department of Agriculture (USDA), Agricultural Research Service (ARS), Ithaca, NY, USA; ATCC: American Type Culture Collection, Manassas, VA, USA; BBA: Biologische Bundesanstalt für Land- und Forstwirtschaft, Institut für Mikrobiologie, Berlin, Germany; CBS: Westerdijk Fungal Biodiversity Institute (WI), Utrecht, The Netherlands; CDC: Centers for Disease Control and Prevention, Atlanta, GA, USA; CECT: Spanish Type Culture Collection, Universidad de Valencia, Burjassot, Spain; CML: Coleção Micológica de Lavras, Universidade Federal de Lavras, Minas Gerais, Brazil; CMW: Culture collection at the FABI, University of Pretoria, South Africa; CPC: Collection of P.W. Crous, held at WI; DAOM: Canadian National Mycological Herbarium and Culture Collection, AAFC, Ottawa, Ontario, Canada; DSM: DSMZ-Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, Braunschweig, Germany; FMR: Facultad de Medicina i Ciències de la Salut, Reus, Spain; FRC: Fusarium Research Center, Pennsylvania State University, PA, USA; GJS: Collection of G.J. Samuels, USDA-ARS, USA; HKAS: Herbarium of Cryptogams, Kunming Institute of Botany, Kunming, China; HMAS: Herbarium Mycologicum Academiae Sinicae, Chinese Academy of Sciences, Beijing, China; IFO: Institute for Fermentation, Osaka, Yodogawa-ku, Osaka, Japan; IHEM: Biomedical Fungi and Yeasts Collection, Scientific Institute of Public Health, Belgium; IMI: CABI Bioscience, Egham, UK; IMUR: Institute of Mycology, University of Recife, Recife, Brazil; MAFF: Ministry of Agriculture, Forestry and Fisheries, Tsukuba, Ibaraki, Japan; MFLU: Mae Fah Luang University herbarium, Chiang Rai, Thailand; MRC: Microbial Culture Collection, South African Medical Research Council, Tygerberg, South Africa; MUCL: Mycothèque de l'Université Catholique de Louvain, Louvain-la-Neuve, Belgium; NHL: National Institute of Hygienic Sciences, Tokyo, Japan; NRRL: Agricultural Research Service Culture Collection, National Center for Agricultural Utilization Research, USDA, Peoria, IL, USA; RBG: Royal Botanic Gardens Trust, Sydney, New South Wales, Australia; URM: Micoteca do Departamento de Micologia, Universidade Federal de Pernambuco, Recife, Brazil; UTHSC: Fungus Testing Laboratory, Department of Pathology, University of Texas Health Science Center, San Antonio, USA. ET: Ex-epitype; IT: Ex-isotype; NT: Ex-neotype; PT: Ex-paratype; T: Ex-type.

² *acl1* = ATP citrate lyase; *CaM* = Calmodulin; ITS = Internal transcribed spacer region of the nrDNA; LSU = 28S large subunit of the nrDNA; *rpb1* = RNA polymerase largest subunit; *rpb2* = RNA polymerase second largest subunit; *tef1* = translation elongation factor 1-alpha; *tub2* = Beta-tubulin. Sequences generated in this study are shown in bold; Not public = sequences not available at GenBank, obtained from K. O'Donnell's alignment datasets; * = Whole genome sequence contig accession numbers.

Table 4. Summary of phylogenetic information generated in this study.

Analysis	Nuclear region	Length + gap	PI	Var.	BI unique site patterns	Model (AIC)	Model (BIC)	ML -lnL (IQ)
Generic delimitation	ITS	626	249	310	378	GTR+I+G	TIMe+I+G4	-3099.276
	LSU	435	90	109	118	GTR+I+G	TIM2+F+I+G4	-15223.682
	<i>rpb1</i>	1 371	705	755	823	GTR+I+G	TIM3e+I+G4	-27263.487
	<i>rpb2</i>	1 761	834	892	989	GTR+I+G	GTR+F+I+G4	-8493.378
	<i>tef1</i>	699	448	489	551	GTR+I+G	TIM2e+I+G4	-40875.16
	Combined	4 892	2 326	2 555	2 859	n/d	n/d	-94954.982
Ex-type strains	<i>rpb1</i>	1 724	980	550	1 358	GTR+I+G	TIM3e+R4	-37377.092
	<i>rpb2</i>	1 789	788	916	1 056	GTR+I+G	TIM2e+R6	-44286.314
	<i>tef1</i>	859	463	301	700	GTR+I+G	GTR+F+I+G4	-25546.628
	Combined	4 372	2 231	1 767	3 114	n/d	n/d	-113450.62
<i>Fusarium fujikuroi</i> species complex	<i>CaM</i>	545	76	131	150	SYM+G	G4TNe+G4	-4032.663
	<i>rpb1</i>	1 534	201	340	344	SYM+G	TIM2e+G4	-5669.761
	<i>rpb2</i>	1 541	241	362	365	GTR+I+G	TNe+G4	-7415.729
	<i>tef1</i>	676	137	243	305	GTR+I+G	TNe+I+G4	-2062.906
	<i>tub2</i>	488	76	150	182	SYM+G	TNe+G4	-1930.688
	Combined	4 794	731	1 226	1 346	n/d	n/d	-22043.423
<i>Fusicolla</i>	<i>acl1</i>	908	153	346	241	GTR+G	TNe+I	-3238.214
	ITS	518	54	111	128	GTR+I+G	TIM2e+I+G4	-1704.698
	LSU	476	34	69	72	K80+I	K80+R2	-1229.69
	<i>rpb2</i>	1 702	258	447	359	SYM+I+G	TIM2e+G4	-5692.247
	<i>tef1</i>	476	109	216	202	SYM+I	TIM2+F+G4	-2051.471
	<i>tub2</i>	484	83	162	159	GTR+G	K80+G4	-1780.157
Combined	4 564	691	1 351	1 161	n/d	n/d	-16092.82	
<i>Macroconia</i>	<i>acl1</i>	801	207	332	205	SYM+I	K80+I	-1241.031
	<i>CaM</i>	551	150	223	159	K80+I	K80+I	-2092.487
	ITS	540	36	64	94	GTR+I	TNe+G4	-2259.518
	LSU	694	21	37	3	GTR+I	TNe+I	-3097.338
	<i>rpb1</i>	814	116	182	96	SYM+G	TNe+G4	-2620.526
	<i>rpb2</i>	778	160	618	151	SYM+I	TNe+G4	-1784.381
	<i>tub2</i>	519	101	168	142	SYM+G	TNe+G4	-1205.535
	Combined	4 697	791	1 624	850	n/d	n/d	-14388.257
<i>Neocosmospora</i>	<i>acl1</i>	630	173	271	297	K80+I+G	TIM3e+I+G4	-13572.514
	<i>CaM</i>	586	171	231	280	HKY+I+G	TIM2e+R3	-5595.928
	ITS	476	119	357	211	GTR+I+G	TNe+G4	-4164.678
	LSU	482	36	63	76	GTR+I+G	TIM3e+I+G4	-10056.777
	<i>rpb1</i>	1 492	390	506	636	GTR+I+G	TIM2e+R3	-2888.743
	<i>rpb2</i>	1 613	449	564	621	GTR+I+G	TIM2e+I+G4	-1496.116
	<i>tef1</i>	688	230	323	370	GTR+I+G	K80+G4	-4087.046
	Combined	5 967	1 568	2 315	2 491	n/d	n/d	-46528.083
<i>Stylonectria</i>	<i>acl1</i>	897	254	426	416	GTR+G	K80+I	-1022.317
	ITS	544	21	39	47	HKY+I	TNe+G4	-5181.494
	<i>rpb2</i>	1 631	183	442	299	GTR+G	TNe+G4	-4061.543
	Combined	3 072	458	907	762	n/d	n/d	-10441.718

PI = parsimony informative characters; Var. = variable characters; BI = Bayesian inference; Model (AIC) = evolutionary model selected by MrModeltest; Model (BIC) = evolutionary model selected by ModelFinder in IQ-TREE; ML -lnL (R) = best tree score determined using RAXML; ML -lnL(IQ) = best tree score determined in IQ-TREE. F = Empirical base frequencies; G = Rate of discrete Gamma categories; GTR = General time reversible model; HKY = Unequal transition/transversion rates and unequal base frequencies; I = Proportion of invariable sites; K80 = Unequal transition/transversion rates and equal base frequencies; R = FreeRate model; SYM = Symmetric model; TIM2 = Transition model, AC = AT, CG = GT and unequal base frequencies; TIM2e = TIM2 with equal base frequencies; TIM3e = Transition model, AC = CG, AT = GT with equal base frequencies; TNe = Unequal transition/transversion rates with unequal purine/pyrimidine rates and equal base frequencies; TPM2 = AC = AT, AG = CT, CG = GT and equal base frequencies.

generalized time-reversible (GTR) model and applying the partitioning option, which estimates the Gamma-shape parameter and the proportion of invariable sites for every gene separately. Again 1000 bootstraps were calculated to estimate branch support. Bayesian inference was conducted using MrBayes v. 3.2.7 (Ronquist & Huelsenbeck 2003) with the partitioned dataset. The Gamma-shape parameter and proportion of invariable sites were estimated independently for each partition. MrBayes was run for 5 M generations with every 500th tree sampled and a burn-in of 30 % of the sampled trees to ensure sampling from the stationary phase. All other parameters were set to default.

Morphology

Morphological characterisation followed standard procedures as described by Leslie & Summerell (2006) using PDA, SNA (Nirenberg 1976), and CLA (Fisher *et al.* 1982). Colony morphology and pigmentation were evaluated on PDA after 7 to 14 d at 25 °C in darkness. Colour notation was based on the colour charts of Rayner (1970). Fungarium specimens were rehydrated in 3 % aqueous KOH for a few minutes and then rinsed by replacing the KOH solution with sterile distilled water or 100 % lactic acid (Samuels 1976a, b, Samuels *et al.* 1990). Unless otherwise mentioned, micromorphological characters

were examined using water as mounting medium on a Zeiss Axioskop 2 plus or a Nikon Eclipse 80i, both equipped with Differential Interference Contrast (DIC) optics and a Nikon AZ100 dissecting microscope all fitted with Nikon DS-Ri2 high-definition colour digital cameras to photo-document fungal structures. Measurements were taken using the Nikon software NIS-elements D v. 4.50. The dimensions of at least 30 randomly selected elements were recorded for every fungal structure. Average, standard deviation, and maximum–minimum values were determined for elements using five or more individual measurements. To facilitate the comparison of relevant micro- and macroconidial features, composite photo plates were assembled from separate photo micrographs using Adobe Photoshop CC.

RESULTS

DNA phylogeny

The results of DNA evolutionary model selection, alignment length, and composition as well as tree statistics for all the multimarker datasets included in this study are summarised in Table 4.

Re-analysis of the dataset of Geiser et al. (2021): A re-analysis of the dataset of Geiser et al. (2021) revealed no major differences in the ML analysis. However, in ME analysis (Supplementary Fig. S3), we found that the backbone architecture is less solid than previously thought and a large monophyletic clade containing *Neocosmospora*, *Albonectria*, and several other genera formed as sister group to *Fusarium s. str.* with strong support.

Generic delimitation of fusarioid taxa in Nectriaceae: The analyses included nectriaceous taxa historically ascribed to *Fusarium s. lat.*, including several recently segregated fusarioid genera (Gräfenhan et al. 2011, Schroers et al. 2011, Lombard et al. 2015), cylindrocarpon-like taxa (Chaverri et al. 2011), and the closely related – although morphologically distinct – phylogenetic relatives *Cosmospora* and *Mariannaea*. Analyses using ML and BI of the individual genes and combined datasets resulted in phylogenies with congruent topologies. Therefore, only IQ-TREE-ML topologies are presented with RAxML-BS, UFboot2-BS, BI-PP and gCF support values superimposed (Fig. 7).

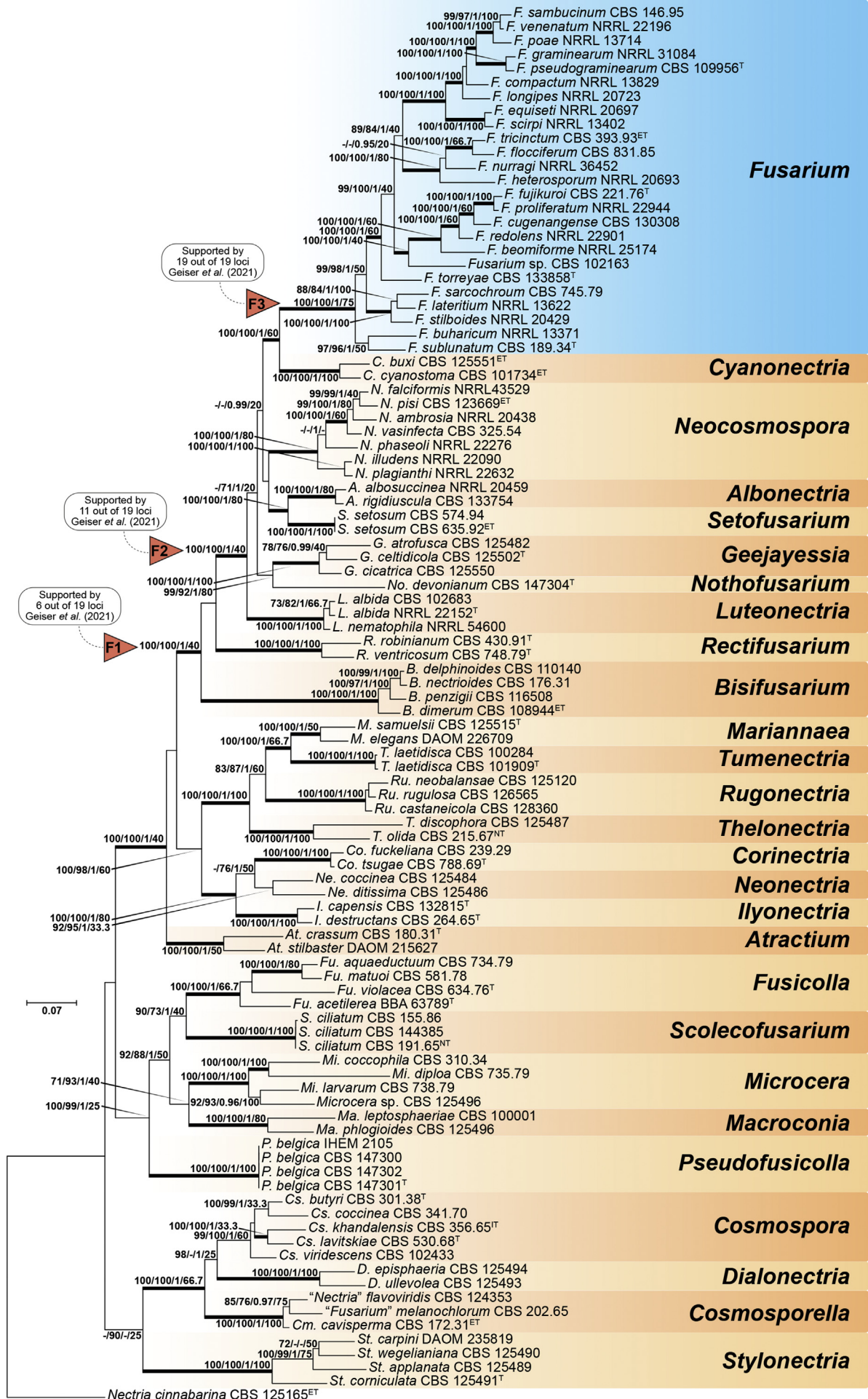
The combined alignment of ITS, LSU, *rpb1*, *rpb2* and *tef1* comprised 100 strains representing 92 species, including the outgroup *Nectria cinnabarina* (CBS 125165). Phylogenetic analyses resolved 27 monophyletic genera, of which 19 contain taxa with fusarioid asexual morphs and nectria- or cosmospora-like sexual morphs. Of these, 15 clades represent currently described genera, namely *Albonectria*, *Atractium*, *Bisifusarium*, *Cosmosporella*, *Cyanonectria*, *Dialonectria*, *Fusarium*, *Fusicolla*, *Geejayessia*, *Macroconia*, *Microcera*, *Neocosmospora*, *Pseudofusicolla*, *Rectifusarium*, and *Stylonectria*. The fusarioid genera *Cosmosporella* and *Dialonectria*, both of which have cosmospora-like sexual morphs, clustered as sister clades to *Cosmospora*; the latter, however, differ by having acremonium-like asexual morphs. The remaining four clades with fusarioid morphology represent undescribed taxa, formally described here as the new genera *Luteonectria*, *Nothofusarium*, *Scolecofusarium*, and *Setofusarium*. A strongly supported clade comprising six cylindrocarpon-like genera (*Corinectria*, *Ilyonectria*, *Neonectria*, *Rugonectria*, *Thelonectria*, and *Tumenectria*)

and the genus *Mariannaea* resolved as successive sister groups to the F1 node.

Twenty-four out of the 27 genera included in the analysis resolved as fully supported clades, including all but one (*Nothofusarium* with RAxML-BS = 99 % / UFboot-BS = 92 % / PP = 1) of the fusarioid genera (Fig. 7). The two remaining clades (*Cosmospora* and *Neonectria*), however, received high statistical support (RAxML-BS = 99 % / UFboot-BS = 100 % / PP = 1 and RAxML-BS = 92 % / UFboot-BS = 95 % / PP = 1, respectively). Similarly, the combined phylogeny resolved most of the internal nodes with high to full bootstrap and Bayesian PP support including the nodes F1, F2, and F3 *sensu* Geiser et al. (2013, 2021) and O'Donnell et al. (2013, 2020). Nevertheless, only F3 was resolved with confidence by all the individual marker phylogenies (Supplementary Fig. S4). Node F2 was resolved with high statistical support in the ITS, *rpb1*, and *tef1* phylogenies, but unsupported in the LSU and *rpb2* trees, while node F1 resolved without bootstrap and PP support in the ITS, *rpb1*, *rpb2*, and *tef1* phylogenies and was not recovered in the LSU tree.

To illustrate shared and differential morphological characters among the different genera recognised here, a tree was constructed based on the phylogeny presented in Fig. 7, and the main morphological features were plotted for each clade/genus (Fig. 8). In addition to the genera recognised above, the recently described aquatic fusarioid genus *Varicosporella* (Lechat & Fournier 2015) is not included in the phylogenetic analyses due to lack of available sequences; however, is accepted here based on its distinct morphology. Non-molecular character variation supports the phylogenetic relationship of fusarioid taxa in *Nectriaceae*. The 20 fusarioid genera in *Nectriaceae* are characterised by phialidic asexual morphs with variously septate, falcate conidia with diverse degrees of foot-shaped basal cell development, formed on aerial or sporodochial conidiophores, with or without additional production of microconidia. Characteristic macroconidial foot-shaped basal cells are found most of the time, but not always (e.g., *Fusarium caeruleum*) in clade F1, i.e., *Albonectria*, *Bisifusarium*, *Cyanonectria*, *Fusarium*, *Geejayessia*, *Luteonectria*, *Neocosmospora*, *Nothofusarium*, *Rectifusarium*, and *Setofusarium*, but are also present in distantly related genera such as *Cosmosporella*, *Dialonectria*, *Macroconia*, and *Microcera*. *Setofusarium* is clearly recognisable by the formation of thick-walled, slightly rugose setae on its sporodochia.

With the exception of *Atractium*, *Bisifusarium*, *Nothofusarium*, and *Pseudofusicolla*, most fusarioid genera have sexual morphs, usually seen as nectria-like or cosmospora-like perithecial ascomata. The ascomata show various colour reactions or no reaction in KOH; the colour reaction correlates with the phylogenetic distribution. Apart from *Albonectria*, with white to pale yellow perithecia, *Luteonectria*, with white to buff coloured perithecia and *Fusarium*, with dark blue-violet to black perithecia, *Fusicolla*, with yellow-orange perithecia and *Varicosporella*, with yellow perithecia, the rest of fusarioid genera all present orange to red perithecial ascomata. Going beyond this prototypical group, perithecia of *Cyanonectria* species are often unequally red to dark blue, while those of *Geejayessia* can be bright red or black. Anatomically, two types of perithecial walls can be distinguished among the known fusarioid genera, based on wall thickness: thin-walled perithecia, in which a single region can be identified, and thick-walled perithecia, on which distinctive inner and outer regions can be recognised (but see Schroers et al. 2011 for differing interpretations). The former is seen in *Cosmosporella*, *Cyanonectria*, *Dialonectria*, *Fusicolla*, *Geejayessia*,



Luteonectria, *Macroconia*, *Microcera*, *Scolecofusarium*, and *Varicosporella*; and the latter is found in *Albonectria*, *Fusarium*, *Neocosmospora*, *Rectifusarium*, *Setofusarium* and *Stylonectria*. With the exception of *Rectifusarium* and *Stylonectria*, the perithecial surface of the thick-walled genera is typically warted; nevertheless, those of *Setofusarium* often present additional scaly protrusions, while smooth perithecia can be rarely found in *Neocosmospora* (i.e., *N. vasinfecta*). Additionally, both *Cyanonectria* and *Geejayessia* most commonly have smooth perithecial walls. The remaining genera, that is *Cosmosporella*, *Dialonectria*, *Fusicolla*, *Luteonectria*, *Macroconia*, *Microcera*, *Rectifusarium*, *Scolecofusarium*, *Stylonectria*, and *Varicosporella*, all form smooth-walled perithecia.

Significant variation also exists among fusarioid genera regarding ascospore characteristics. Most genera consistently form 1-septate ascospores. These are seen in *Cosmosporella*, *Cyanonectria*, *Dialonectria*, *Fusicolla*, *Geejayessia*, *Macroconia*, *Microcera*, *Rectifusarium*, *Scolecofusarium*, *Setofusarium*, *Stylonectria*, and *Varicosporella*. Except for *Cyanonectria*, in which the ascospores remain hyaline and smooth; *Setofusarium*, in which the ascospores surface is finely striated, and *Varicosporella*, in which the ascospore surface is ribbed, ascospores of the above-mentioned genera are often pale yellow to pale brown and smooth at first, becoming finely spinulose or tuberculate. The genus *Neocosmospora* forms (0–)1-septate, yellow-brown ascospores, which are often markedly striate, or more rarely cerebriform (i.e., *N. vasinfecta*) or spiny (i.e., *N. spinulosa*). *Albonectria* and *Luteonectria* form characteristic 3-septate, pale yellow-brown, faintly striate ascospores, while *Fusarium* produces 1–3-septate, hyaline to pale yellow-brown and smooth ascospores.

Based on the morphological variation observed in these taxa, an identification scheme is presented for fusarioid genera of the *Nectriaceae* (Fig. 9).

Ex-type strain phylogeny: The analyses included partial *rpb1*, *rpb2* and *tef1* sequences of only the ex-, epi- and neotype strains as indicated in the nomenclator list of all the names that have been introduced in *Fusarium*. The analyses used both ML inferences and BI of the individual genes and combined datasets, and they resulted in phylogenies with congruent topologies. Therefore, the RAxML topology is presented with RAxML-BS, UFboot2-BS, BI-PP and gCF support values superimposed (Fig. 10).

The combined alignment comprised 325 strains from 309 species of 14 fusarioid genera including *Atractium stilbaster* (CBS 410.67) as the outgroup. A total of 14 fusarioid genera were resolved of which six (*Cosmosporella*, *Microcera*, *Nothofusarium*, *Rectifusarium*, *Scolecofusarium*, and *Setofusarium*) were represented by single lineages, mostly due to a lack of living isolates directly linked to type material available for other species recognised within these genera at present. The genera *Fusarium* (224 strains; 220 accepted species) and *Neocosmospora* (83 strains; 71 accepted species) both represented the largest sampling of living isolates directly linked to type material available. The remaining five genera were represented by two or more strains and include *Bisfusarium* (five species and

strains), *Cyanonectria* (two species and strains), *Fusicolla* (three species and strains), *Geejayessia* (two species and strains), and *Luteonectria* (two species and strains).

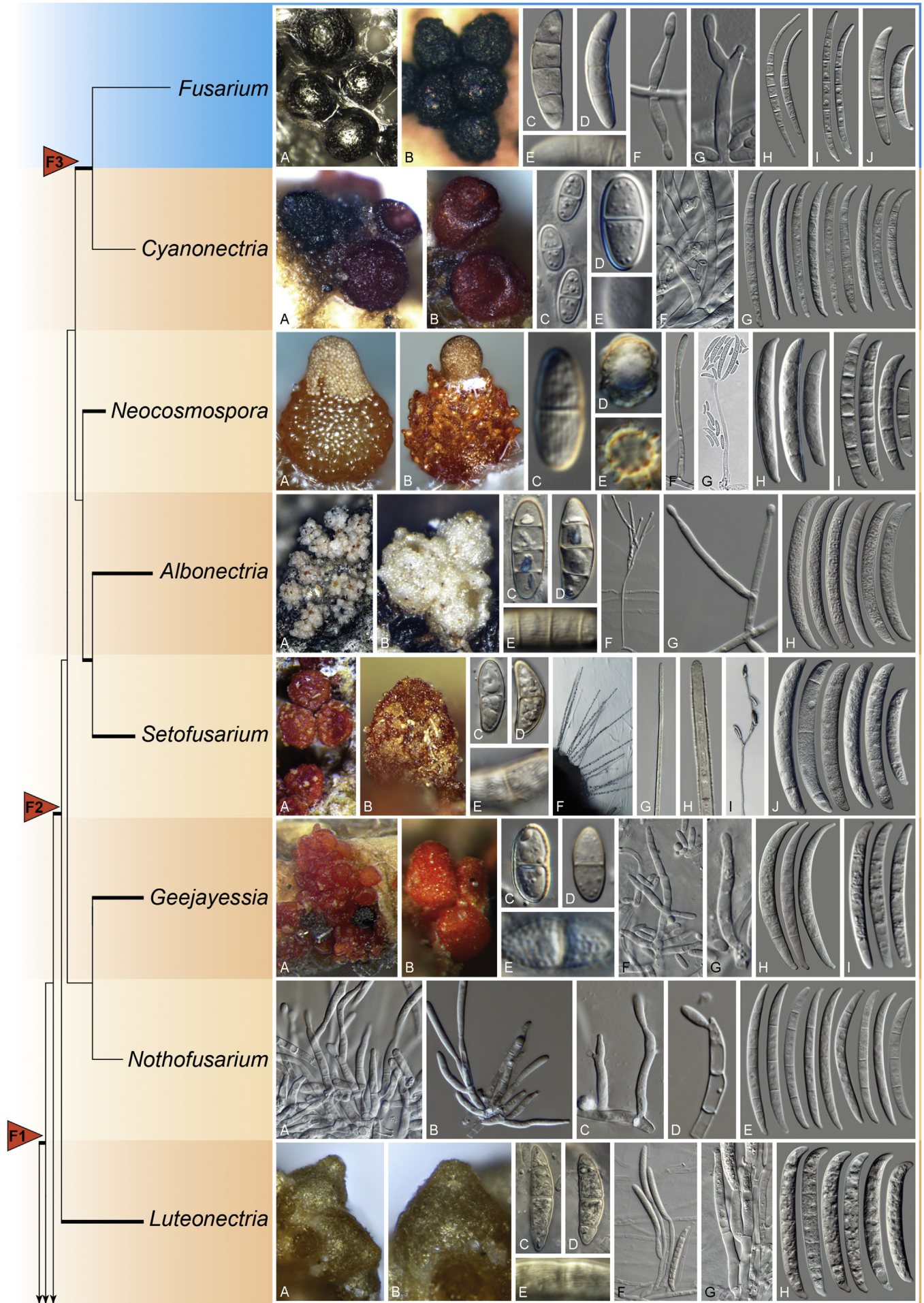
In order to describe novel species found for the genera treated in this study, additional phylogenies were constructed for the *Fusarium fujikuroi* species complex (FFSC), *Fusicolla*, *Macroconia*, *Neocosmospora*, and *Stylonectria*.

Fusarium fujikuroi SC phylogeny: The analyses included partial sequences of five genes (*CaM*, *rpb1*, *rpb2*, *tef1* and *tub2*) from 52 strains representing 46 species of the FFSC, and two outgroup taxa (*F. curvatum* CBS 744.97 and *F. inflexum* CBS 716.74) (Fig. 11). The analysis of the combined dataset fully supported five main clades corresponding to the African, American and Asian clades *sensu* O'Donnell *et al.* (2000b), plus the African B-clade (Sandoval-Denis *et al.* 2018b, Yilmaz *et al.* 2021) and a fifth, monotypic clade, which formed the sister clade to the joint American and African B clades and which is here termed African C. The latter clade included two strains showing a clear genealogical and morphological separation from their closest phylogenetic relatives; both came from an unknown tree species in South Africa. This clade is here described as the novel species *F. echinatum*. Another fully supported novel monophyletic group was found within the main African clade, related to but distinct from *F. brevicatenulatum* and *F. pseudonygamai*. This novel group, represented by isolates of South African origin isolated from *Prunus spinosa* and from the South African indigenous species *Aloidendron dichotomum*, is here recognised as the novel species *F. prieskaense*.

Fusicolla phylogeny: The alignment consisted of partial *acl1*, ITS, LSU, *rpb2*, *tef1*, and *tub2* sequences from 20 type or reference strains, representing 17 species of *Fusicolla* (*Fu.*) plus one outgroup taxon (*Macroconia leptosphaeriae* CBS 100001). The analysis confidently resolved 11 ingroup taxa (Fig. 12), including three novel monotypic lineages, represented by strains URM 8367, CBS 110189, and CBS 110191, described here as the new species *Fu. quarantanae*, *Fu. meniscoidea* and *Fu. sporellula*. Due to a partial lack of sequence data, six species could not be clearly resolved. *Fusicolla cassiae-fistulae* and *Fu. siamensis* did not receive statistical support in the combined analysis but are well-resolved using nrDNA sequence data (data not shown). *Fusicolla acetilerea* and *Fu. bhataravarshae*, while well-delimited in the individual ITS, LSU and *rpb2* analyses (data not shown), were ill-supported in the 6-marker combined analysis. Similarly, *Fu. epistroma* and *Fu. ossicola* were not differentiated in either the multimarker analysis, or in the individual *rpb2* analysis. The lack of sequences available to allow comparison with *Fu. epistroma*, for which only LSU and *rpb2* sequences are available, prevented further analysis, as did a similar problem with *Fu. bhataravarshae*, for which only nrDNA and *rpb2* are available.

Macroconia phylogeny: The analysis consisted of partial *acl1*, *CaM*, ITS, LSU, *rpb1*, *rpb2*, and *tub2* sequences from 12 strains representing seven lineages of *Macroconia* (*Ma.*) plus one outgroup taxon (*Microcera rubra* CBS 638.76) (Fig. 13). Four out of

Fig. 7. Maximum-Likelihood (IQ-TREE-ML) consensus tree inferred from the combined ITS, LSU, *rpb1*, *rpb2* and *tef1* multiple sequence alignment of members of *Nectriaceae*. Numbers at the branches indicate support values (RAxML-BS / UFboot2-BS / BI-PP / gCF) above 70 % / 0.95 with thickened branches indicating full support (RAxML-BS / UFboot2-BS / gCF = 100 %; BI-PP = 1). The scale bar indicates expected changes per site. The tree is rooted to *Nectria cinnabarina* (CBS 125165). Arrows "F1", "F2" and "F3" indicate the three alternative *Fusarium* hypotheses *sensu* Geiser *et al.* (2013). Ex-epitype, ex-isotype, ex-neotype and ex-type strains are indicated with ET, IT, NT, and T, respectively.



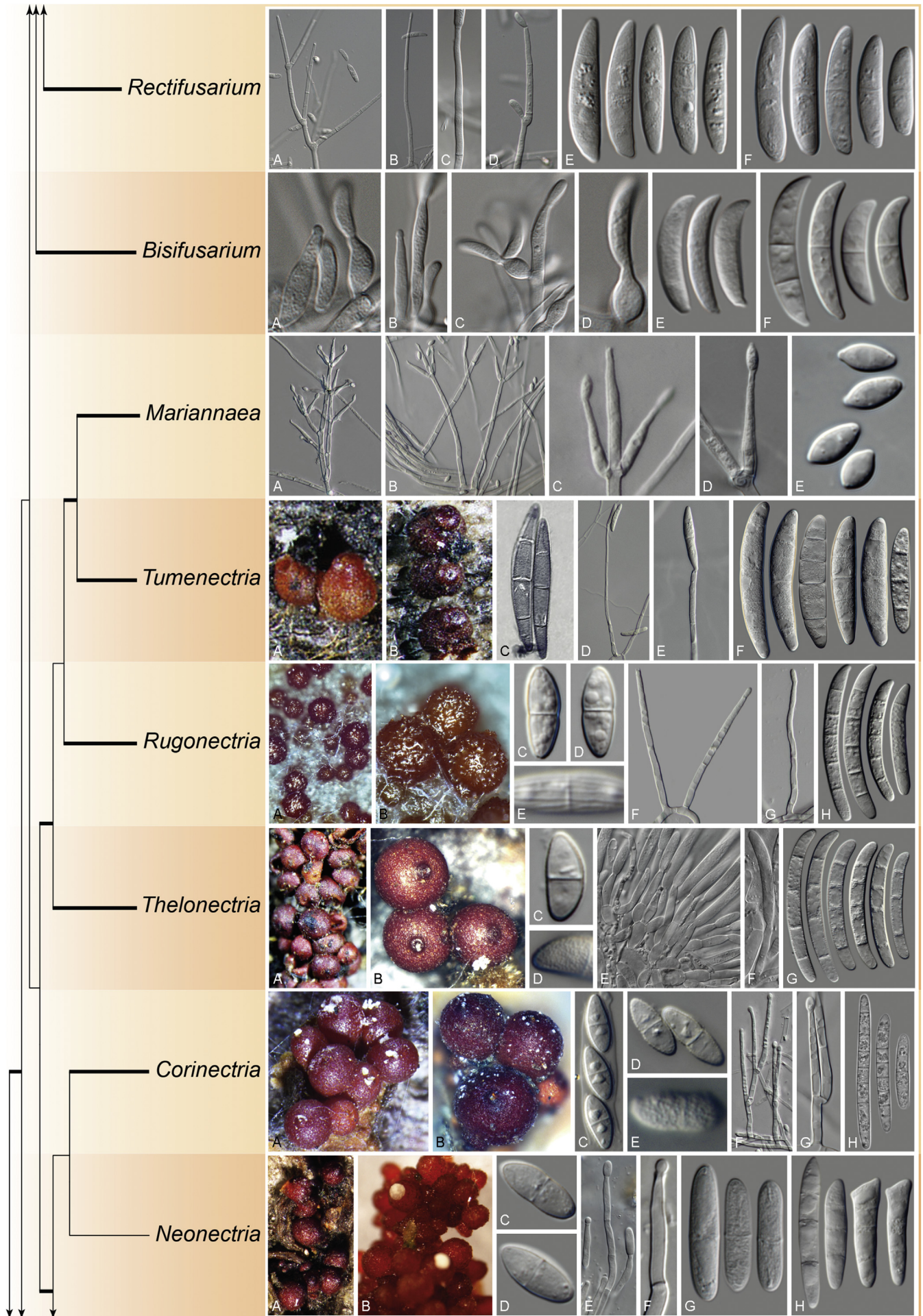


Fig. 8. (Continued).

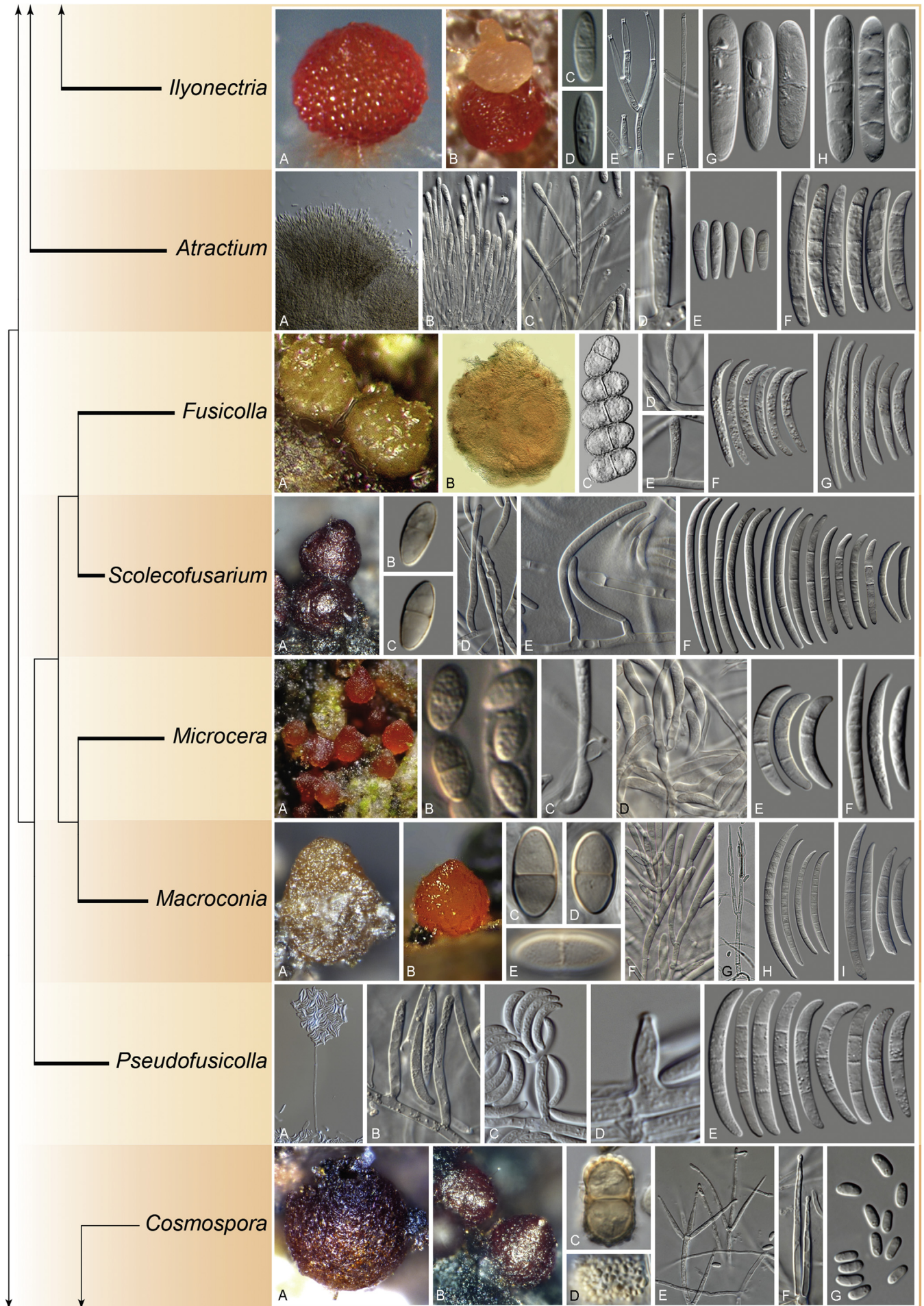


Fig. 8. (Continued).

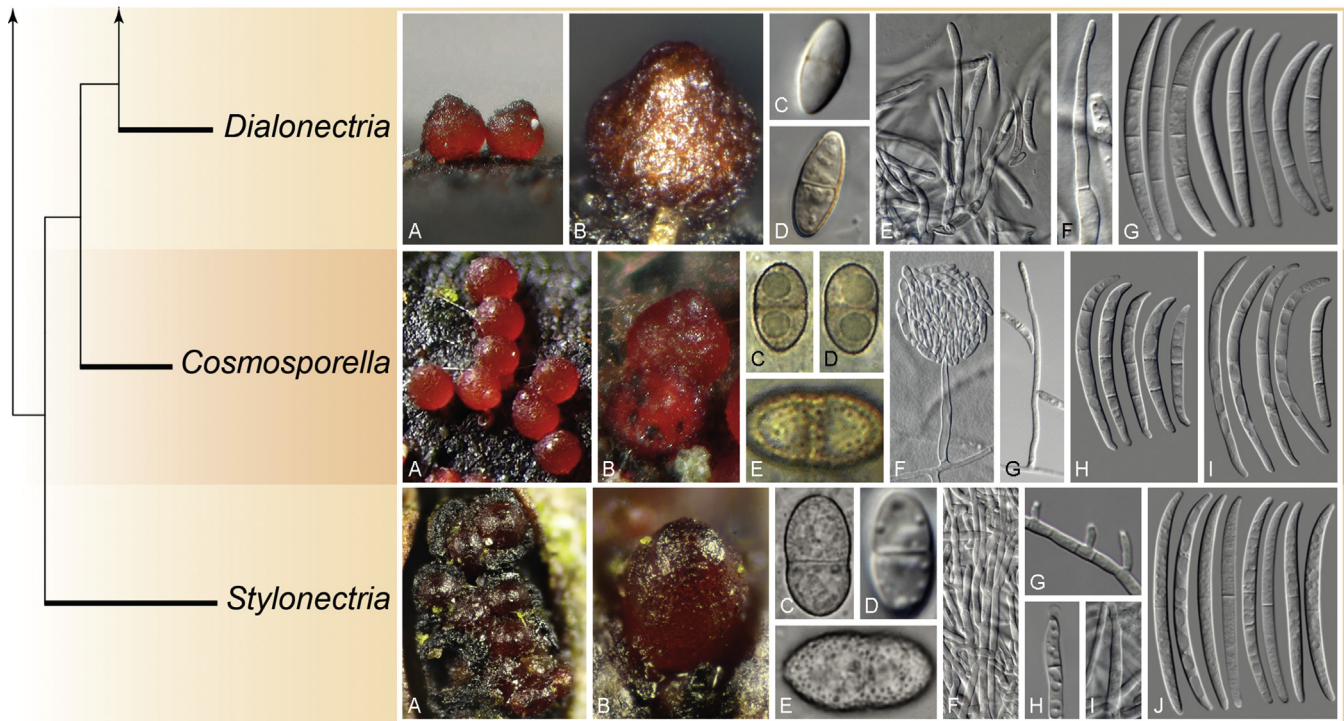


Fig. 8. Morphological features and phylogenetic affinities of fusarioid genera of *Nectriaceae* and close relatives. The tree was delineated based on the phylogeny presented in Fig. 7 and does not indicate phylogenetic distances. Fully supported branches are indicated in **bold**. The genus *Fusarium* is indicated in blue. Arrows "F1", "F2" and "F3" indicate the three alternative *Fusarium* hypotheses *sensu* Geiser *et al.* (2013). *Fusarium*. A, B. Ascomata. C–E. Ascospores. F, G. Conidiogenous cells. H–J. Macroconidia. (B. Adapted from Schroers *et al.* 2011). *Cyanonectria*. A, B. Ascomata. C–E. Ascospores. F. Conidiogenous cells. G. Macroconidia. *Neocosmospora*. A, B. Ascomata. C–E. Ascospores. F, G. Conidiogenous cells. H, I. Macroconidia. [A. Adapted from Sandoval-Denis & Crous (2018). G. Adapted from Sandoval-Denis *et al.* (2019)]. *Albonectria*. A, B. Ascomata. C–E. Ascospores. F, G. Conidiophores and conidiogenous cells. H. Macroconidia. *Setofusarium*. A, B. Ascomata. C–E. Ascospores. F–H. Setae formed on sporodochia. I. Conidiophore. J. Conidia. *Geejayessia*. A, B. Ascomata. C–E. Ascospores. F, G. Conidiophores and conidiogenous cells. H, I. Macroconidia. [A. Adapted from Schroers *et al.* (2011)]. *Nothofusarium*. A–D. Conidiophores and conidiogenous cells. E. Conidia. *Luteonectria*. A, B. Ascomata. C–D. Ascospores. F, G. Conidiophores and conidiogenous cells. H. Conidia. *Rectifusarium*. A–D. Conidiophores and conidiogenous cells. E, F. Conidia. *Bisifusarium*. A–D. Conidiophores and conidiogenous cells. E, F. Conidia. *Mariannaea*. A, B. Conidiophores. C, D. Conidiogenous cells. E. Conidia. *Tumenectria*. A, B. Ascomata. C. Ascospores. D, E. Conidiophores and conidiogenous cells. F. Conidia. [A–C. Adapted from Salgado-Salazar *et al.* (2016)]. *Rugonectria*. A, B. Ascomata. C–E. Ascospores. F, G. Conidiophores and conidiogenous cells. H. Conidia. *Thelonectria*. A, B. Ascomata. C, D. Ascospores. E, F. Conidiophores and conidiogenous cells. G. Conidia. *Corinectria*. A, B. Ascomata. C–E. Ascospores. F, G. Conidiophores and conidiogenous cells. H. Conidia. (H. Picture by C. González). *Neonectria*. A, B. Ascomata. C, D. Ascospores. E, F. Conidiophores and conidiogenous cells. G, H. Conidia. [A. Adapted from Chaverri *et al.* (2011)]. *Ilyonectria*. A, B. Ascomata. C, D. Ascospores. E, F. Conidiophores and conidiogenous cells. G, H. Conidia. *Atractium*. A, B. Conidiophores. C, D. Conidiogenous cells. E, F. Conidia. *Fusicolla*. A, B. Ascomata. C. Ascospores. D, E. Conidiogenous cells. F, G. Conidia. (A–C. Pictures by C. Lechat). *Scolecofusarium*. A. Ascomata. B, C. Ascospores. D, E. Conidiophores and conidiogenous cells. F. Conidia. *Microcera*. A. Ascomata. B. Ascospores. C, D. Conidiogenous cells. E, F. Conidia. (A, B. Pictures by N. Aplin, Fungi of Great Britain and Ireland). *Macroconia*. A, B. Ascomata. C–E. Ascospores. F, G. Conidiophores and conidiogenous cells. H, I. Conidia. (B. Picture by P. Mičoch). *Pseudofusicolla*. A, B. Conidiophores and conidiogenous cells. C, D. Conidia. [A–D. Adapted from Triest *et al.* (2016)]. *Cosmospora*. A, B. Ascomata. C, D. Ascospores. E, F. Conidiophores and conidiogenous cells. G. Conidia. *Dialonectria*. A, B. Ascomata. C–E. Ascospores. F, G. Conidiophores and conidiogenous cells. H. Conidia. (A. Picture by P. Mičoch). *Cosmosporella*. A, B. Ascomata. C–E. Ascospores. F, G. Conidiophores and conidiogenous cells. H, I. Conidia. (A–E. Pictures by P. Mičoch). *Stylonectria*. A, B. Ascomata. C–E. Ascospores. F–I. Conidiophores and conidiogenous cells. J. Conidia. (A–C, E. Pictures by B. Wergen).

the five *Macroconia* spp. previously known from culture, *Ma. gigas*, *Ma. leptosphaeriae*, *Ma. papilionacearum*, and *Ma. sphaeriae*, resolved as highly to fully-supported lineages. The poorly resolved position of the ex-type isolate of *Ma. cupularis* (HMAS 173240) should be interpreted in light of the fact that only nrDNA sequences were available for analysis. However, separate ITS and LSU comparisons demonstrated it as distinct (data not shown). Two distinct and highly supported novel lineages of South African origin were determined and are described here as the novel species, *Ma. bulbipes* and *Ma. phlogioides*.

Neocosmospora phylogeny. The alignment consisted of partial *aci1*, *CaM*, ITS, LSU, *rpb1*, *rpb2*, and *tef1* sequences of 107 ex-type and reference strains, including two outgroup taxa (*Geejayessia atrofusca* NRRL 22316 and *G. cicatricum* CBS 125552). The analysis resolved 76 terminal clades, of which 71 correspond to known species of *Neocosmospora* (Fig. 14).

Seventy of these clades resolved with high support from two or more independent algorithms (RAXML, IQ-TREE-ML, and BI). The position of the ex-type of *N. crassa* (CBS 144386) is poorly resolved and only partially supported by BI. Similarly, except for the types of *N. ambrosia* (CBS 571.94), *N. obliquiseptata* (NRRL 62611), *N. rekana* (CMW 52862), and the reference strain of *N. pseudensiformis* (CBS 130.78), the position of most members of the well-delimited Ambrosia-clade of *Neocosmospora* were only partially supported by the individual analyses (only BI in *N. kuroshio*, *N. oligoseptata*, and *N. tuaranensis*, and only IQ-TREE-ML-BS for *N. euwallaceae* and *N. florida*). All these lineages were represented by single isolates in these analyses. Of the five unnamed phylogenetic clades, one corresponded to a species previously known from phylogenetic analyses (FSSC 41, Cardoso 2015), for which a Latin binomial is lacking; this species is here formally described as *N. merkiana*. The four additional

novel lineages discovered here are proposed as the novel species *N. neerlandica*, *N. nelsonii*, *N. pseudopisi*, and *N. epipeda*.

Stylonectria phylogeny: The alignment consisted of partial *act1*, ITS and *rpb2* sequences of 11 strains, including the out-group (*Nectria cinnabarina* CBS 125165). The analyses (Fig. 15) identified eight species-level clades, of which six represented previously known species of the genus: *St. applanata*, *St. carpini*, *St. norvegica*, *St. purtonii*, *St. qilianshanensis*, and *St. wege-liniana*. One strain, CBS 125491, isolated from an unknown ascomycetous host, corresponded to a previously known unnamed and fully supported monophyletic lineage, which is formally described here as *St. corniculata*. In addition, a fully supported clade formed by two strains, CBS 147305 from *Diaporthe* sp. and CBS 147306 from *Dothiorella sarmentorum*, is here recognised as the novel species *St. hetmanica*.

Taxonomy

Albonectria Rossman & Samuels, Stud. Mycol. 42: 105. 1999. Figs 8, 16.

Type species: *Albonectria rigidiuscula* (Berk. & Broome) Rossman & Samuels, Stud. Mycol. 42: 105. 1999.
(See *F. colorans* in List section for synonyms)

Ascomata perithecial, solitary or gregarious, superficial on a sparse to well-developed, pseudoparenchymatous stroma, globose to subglobose to ellipsoidal or ovoid to obovoid, not collapsing or laterally pinched when dry, off-white to pale yellow to pale ochraceous, not changing in KOH, strongly tuberculate and thick-walled, with or without a small, pointed papilla, lacking hairs or appendages. **Ascomatal wall** of three regions: outer region of thick-walled, *textura angularis* to *textura globulosa*; middle region of elongate thick-walled cells; inner region with thin-walled, hyaline elongated cells. **Asci** narrowly to broadly clavate or ellipsoidal, 4–8-spored, ascospores obliquely uniseriate or biseriate. **Ascospores** ellipsoidal to long-ellipsoidal or fusoid to long-fusoid, 3- to multiseptate, hyaline to yellow-brown, smooth to striate, not to slightly constricted at the septum. **Conidiophores** mononematous (aerial conidiophores) or grouped on sporodochia; **aerial conidiophores** unbranched or irregularly branched, bearing terminal or lateral phialides, often reduced to single phialides; **conidiogenous cells** monophialidic, cylindrical to subcylindrical, smooth- and thin-walled, with periclinal thickening inconspicuous or absent, producing arial micro- and macroconidia. **Microconidia** hyaline, thin-walled, 0- or 1-septate, ovoid to obovoid, with or without a flattened basal papilla, borne in dry chains or small slimy heads. **Macroconidia** falcate, multiseptate, thick-walled, with a blunt to hooked apical cell and well-developed foot-shaped basal cell or distinctly beaked at both ends. **Sporodochia** cream to yellow; **sporodochial conidiophores** verticillately branched and densely packed, consisting of short, smooth- and thin-walled stipes bearing apical whorls of 2–4 monophialides; **sporodochial conidiogenous cells** monophialidic, cylindrical to subulate, smooth- and thin-walled, with reduced or flared collarete. **Sporodochial macroconidia** formed in off-white or creamy slimy masses, falcate, 5–9-septate, thick-walled, gently curved to straight, with a blunt to hooked apical cell and distinct well-developed foot-shaped basal cell. **Chlamydo-spores** absent.
[Description adapted from Rossman et al. (1999), Booth (1971) and Lombard et al. (2015)].

Diagnostic features: Off-white to pale yellow to pale ochraceous perithecia producing narrowly or broadly clavate to ellipsoidal asci containing (long) ellipsoidal to fusoid, 3- to multiseptate ascospores; fusarioid asexual morph characterised by monophialides producing distinctly long, robust, slightly curved to straight multiseptate macroconidia and dry chains or small slimy heads of ovoid microconidia. Chlamydo-spores absent.

Atractium Link, Mag. Ges. Naturf. Freunde Berlin 3: 10. 1809 (Fries, Syst. Mycol. 1: XLI. 1821, *nom. sanct.*). Figs 8, 17.

Type species: *Atractium stilbaster* Link, Mag. Ges. Naturf. Freunde Berlin 3: 10. 1809.

Ascomata unknown. **Conidiophores** aggregated into sporodochia or synnemata, non-stromatic; **synnemata** determinate, pale brown, composed of a stipe of parallel hyphae and a divergent capitulum of conidiophores giving rise to a slimy conidial mass; conidiophore branching once or twice monochasial, 2-level verticillate, monoverticillate or irregularly biverticillate. **Conidiogenous cells** monophialidic, hyaline, subulate with conspicuous periclinal thickening, producing micro- and macroconidia. **Microconidia** hyaline, thin-walled, 0- or 1-septate, ellipsoidal, allantoid, broadly lunate to reniform, straight or slightly curved, tapering towards both apices with rounded base. **Macroconidia** 3–5-septate, falcate, gently curved, with a rounded to blunt apical cell, and obtuse, non foot-shaped basal cell, forming yellow to orange masses.
[Description adapted from Gräfenhan et al. (2011)].

Diagnostic features: Synnematus asexual morph characterised by fusarioid macroconidia lacking foot-shaped basal cells.

Bisifusarium L. Lombard et al., Stud. Mycol. 80: 223. 2015. Figs 8, 18.

Type species: *Bisifusarium dimerum* (Penz.) L. Lombard & Crous, Stud. Mycol. 80: 225. 2015.

(See *F. dimerum* in List section for synonyms)

Ascomata unknown. **Conidiophores** mononematous (aerial conidiophores) or grouped on sporodochia; **aerial conidiophores** simple, unbranched or irregularly branched, mostly reduced to terminal or single lateral conidiogenous cells. **Conidiogenous cells** often formed as (i) lateral phialidic pegs arising from superficial or submerged intercalary hyphal cells or, (ii) cylindrical and slightly tapering towards apex or ampulliform, smooth- and thin-walled monophialides, rarely polyphialides, with inconspicuous or absent periclinal thickening, solitary or aggregated to represent a poorly developed pionnotal sporodochial-like structure, producing micro- and macroconidia. **Microconidia** hyaline, thin-walled, 0- or 1-septate, ellipsoidal, allantoid, broadly lunate to reniform, straight or curved, tapering towards both ends. **Macroconidia** falcate, (0–)1–2(–3)-septate, thick-walled, curved to lunate, with a blunt to hooked apical cell and obtuse to poorly developed, foot-shaped basal cell, typically formed on sporodochia. **Sporodochia** pale yellow to orange; **sporodochial conidiophores** verticillately branched and densely packed, consisting of short, smooth- and thin-walled stipes bearing an apical whorl of 2–3 monophialides; **sporodochial conidiogenous cells** monophialidic, cylindrical to subulate, smooth- and thin-walled, with reduced or flared collarete. **Chlamydo-spores**, if present, globose to subglobose to ellipsoidal, solitary or in chains, sometimes aggregated in sclerotia.

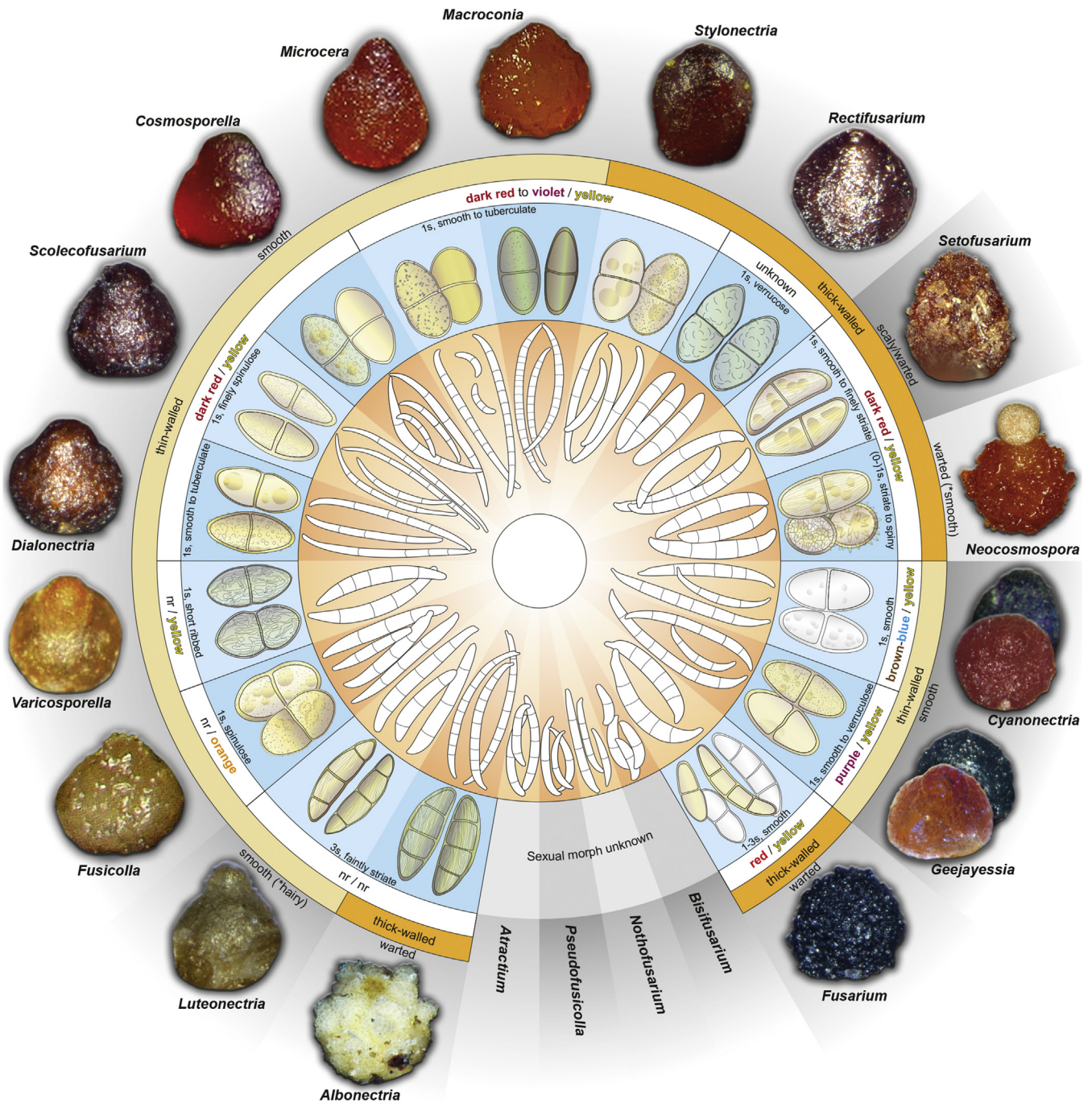


Fig. 9. Characters for morphological identification of fusarioid genera in Nectriaceae. The rings show, from inside to outside: conidial morphology; ascospore morphology, septation and surface; colour reaction of ascospores in 3% KOH/lactic acid (nr = no reaction); ascospore wall thickness; and general colour, appearance and wall surface of ascospores.

[Description adapted from Schroers et al. (2009) and Lombard et al. (2015)].

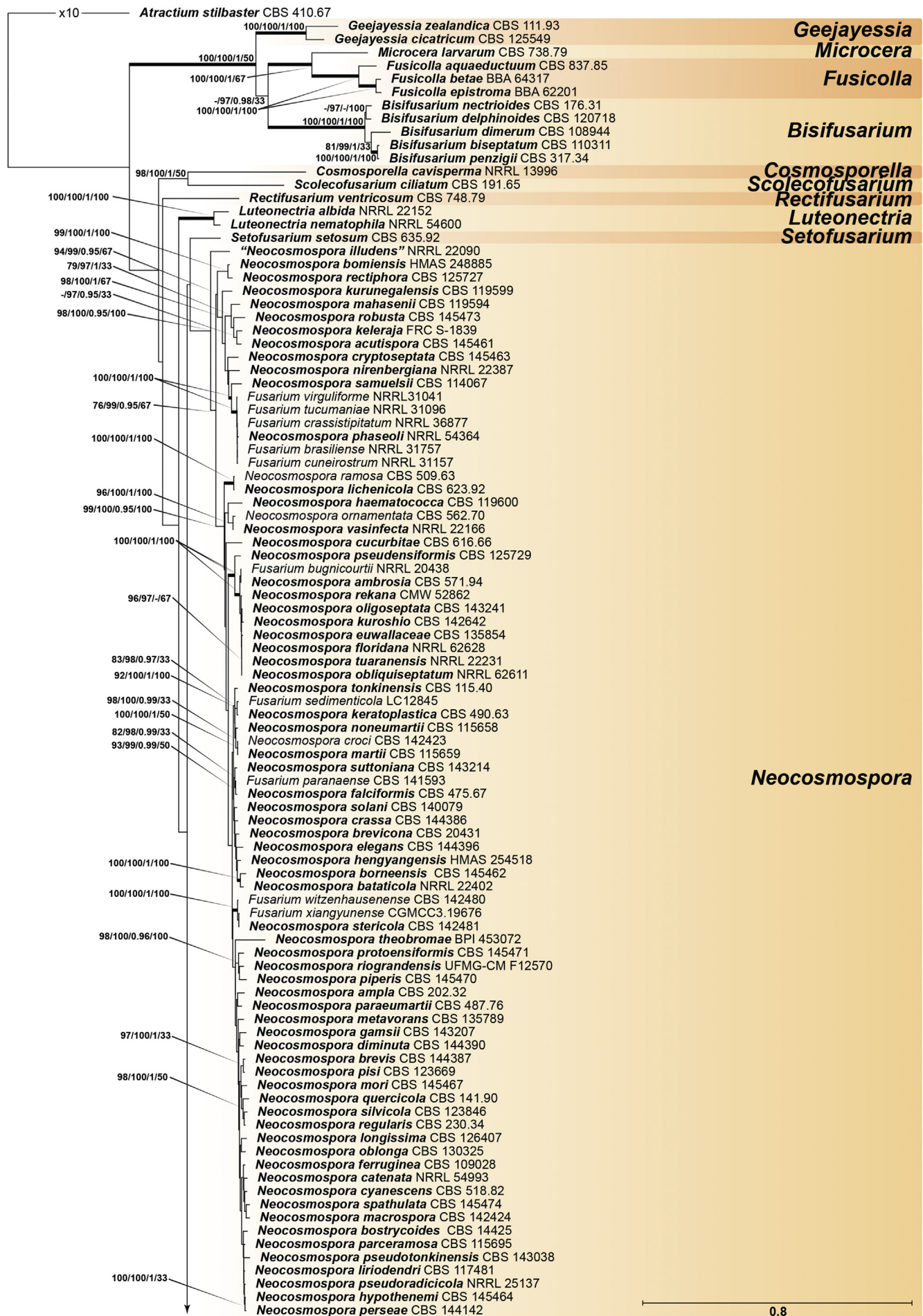
Diagnostic features: Fusarioid asexual morph characterised by lateral phialidic pegs arising from superficial or submerged intercalary hyphal cells or solitarily formed monophialides producing microconidia; distinctly short (< 30 µm long), curved to lunate, (0–)1–2(–3)-septate macroconidia typically formed on sporodochia on plant tissue such as carnation leaf pieces.

Corinectria C. González & P. Chaverri, Mycol. Progr. 16: 1021. 2017. Fig. 8.

Type species: *Corinectria fuckeliana* (C. Booth) C. González & P. Chaverri, Mycol. Progr. 16: 1023. 2017.

Basionym: *Nectria fuckeliana* C. Booth, Mycol. Pap. 73: 56. 1959.
Synonym: *Neonectria fuckeliana* (C. Booth) Castl. & Rossman, Canad. J. Bot. 84: 1428. 2006.

Ascospores perithecial, gregarious, seated on an erumpent stroma, superficial, globose to subglobose, orange, red to dark red darkening around ostiolar region, turning black in KOH, pigment dissolving in lactic acid, not collapsing when dry, slightly papillate to papillate, smooth-walled, lacking hairs or appendages. Ascospore wall of 2–3 regions: outer region of thick-walled, pigmented cells forming a *textura epidermoidea*; middle and inner regions of globose to elongate, hyaline, thin-walled cells, becoming thinner toward the centrum. Asci cylindrical, 8-spored, with an apical ring, uniseriate. Ascospores ellipsoidal to fusoid, 1-



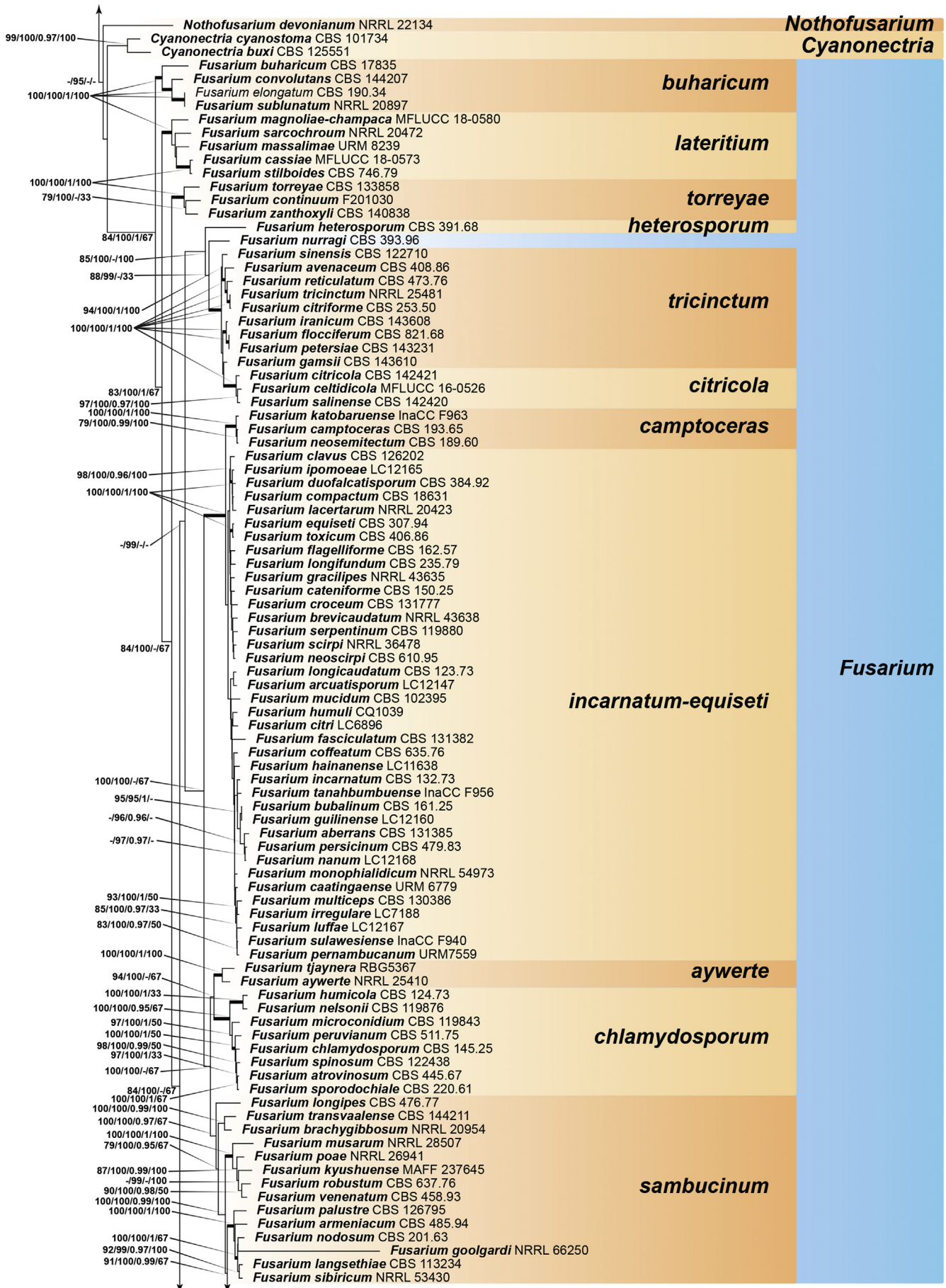


Fig. 10. (Continued).

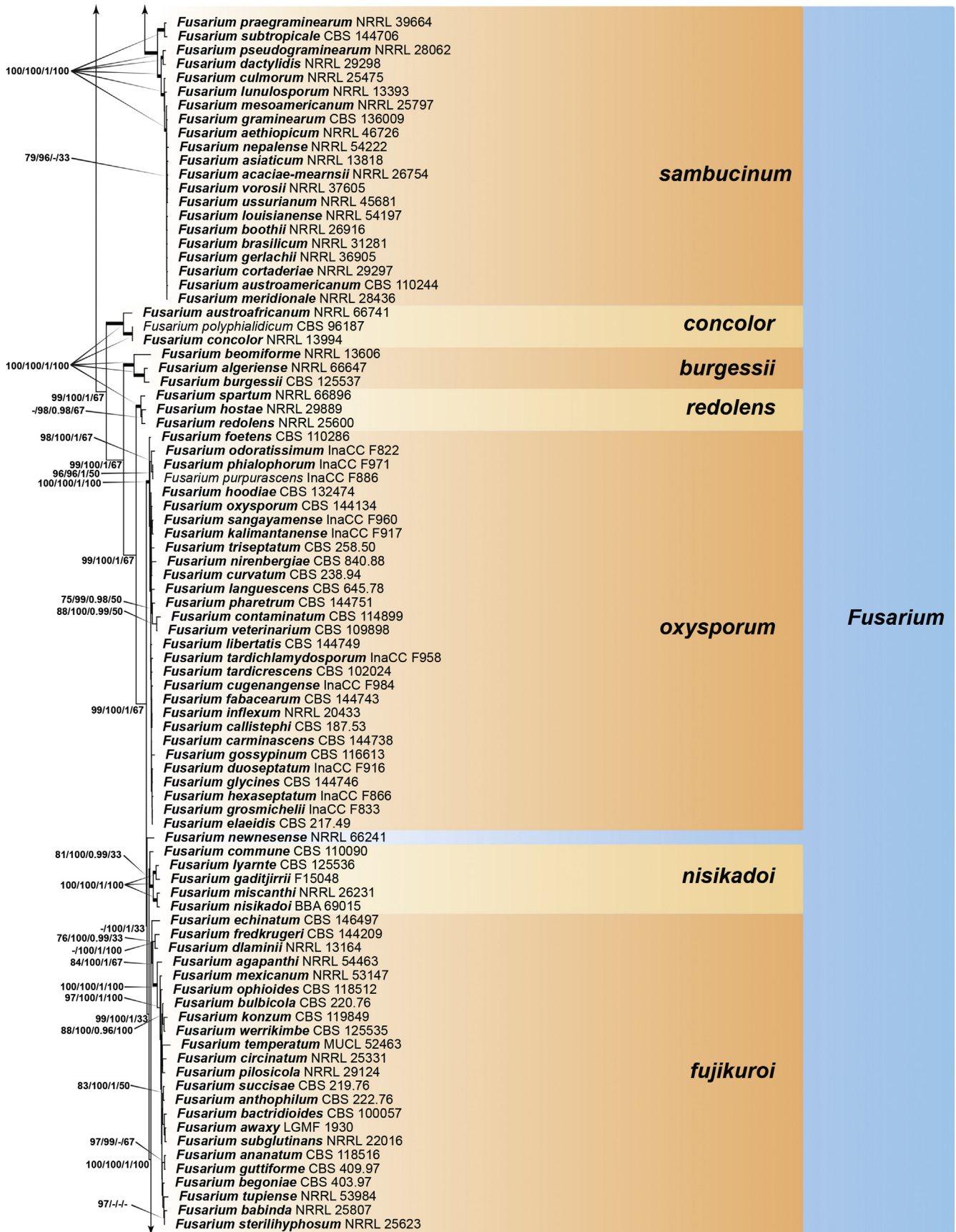




Fig. 10. Maximum-Likelihood (IQ-TREE-ML) consensus tree inferred from the combined *rbp1*, *rbp2* and *tef1* sequence alignment of the living type strains as indicated in the nomenclator list. Numbers at the branches indicate support values (RAxML-BS / UFboot2-BS / BI-PP) above 70 % / 0.95 with thickened branches indicating full support (RAxML-BS / UFboot2-BS = 100 %; BI-PP = 1). The scale bar indicates expected changes per site. The tree is rooted to *Atractium stilbaster* (CBS 410.67). Names indicated in bold are in current use. Subdivision of the *Fusarium* clade (blue block) represent the recognised species complexes.

septate, hyaline, smooth. *Conidiophores* mononematous, hyaline, septate, unbranched or sparsely branched, terminating in 1–2 phialides or reduced to lateral phialides. *Conidiogenous cells* monophialidic, cylindrical, tapering towards the apex, with inconspicuous periclinal thickening and collarettes. *Sporodochia* not formed. *Microconidia* ellipsoidal to obovoid, hyaline, aseptate, sometimes forming false heads on phialides. *Macroconidia* cylindrical, mostly straight, (3–)5–7-septate, with rounded ends. *Chlamydospores* unknown.

[Description adapted from González & Chaverri (2017)].

Diagnostic features: Orange to dark red, smooth-walled perithecia with papilla producing cylindrical asci bearing ellipsoidal to fusoid, 1-septate ascospores and cylindrocarpon-like asexual morph characterised by (3–)5–7-septate macroconidia.

Cosmospora Rabenh., Hedwigia 2: 59. 1862. Fig. 8.

Synonyms: *Cryso-gluten* Briosi & Farneti, Atti Ist. Bot. Univ. Lab. Crittog. Pavia 8: 117. 1904.

Botryocrea Petr., Sydowia 3: 140. 1949.

Type species: *Cosmospora coccinea* Rabenh., Hedwigia 2: 59. 1862 [non *Nectria coccinea* (Pers.) Fr. 1849].

Synonyms: *Nectria cosmariospora* Ces. & De Not., Comment. Soc. Crittog. Ital. 1: 195. 1863.

Dialonectria cosmariospora (Ces. & De Not.) Cooke, Grevillea 12: 110. 1884.

Cucurbitaria cosmariospora (Ces. & De Not.) Kuntze, Revis. Gen. Pl. 3: 461. 1898.

Dialonectria cosmariospora (Ces. & De Not.) Z. Moravec, Česká Mykol. 8: 92. 1954, an isonym, Art. 6.3, Note 2.

Verticillium olivaceum W. Gams, Cephalosporium-artige Schimmelpilze: 129. 1971.

Ascomata perithecial, solitary or gregarious, with inconspicuous or absent stroma, obpyriform with an acute or papillate apex, orange red or bright red, turning dark red in KOH, smooth walled. **Asci** narrowly clavate to cylindrical, with an apical ring, 8-spored. **Ascospores** initially hyaline, becoming yellow brown to reddish brown, 1-septate, becoming tuberculate when mature. **Conidiophores** acremonium-like, consisting of lateral phialides on somatic hyphae, or with one or two levels of monochasial branching, or verticillate, hyaline. **Conidiogenous cells** monophialidic, cylindrical to subulate to subclavate, hyaline. **Microconidia** ellipsoidal, oblong or clavate or slightly allantoid, aseptate, hyaline, forming slimy heads. **Macroconidia** absent or rare, subcylindrical, curved, slightly narrowing towards each end, apical cell often slightly hooked with a more or less pointed apex, basal cell obtuse to poorly developed, foot-shaped, 3–5-septate, hyaline.

[Description adapted from Rossman *et al.* (1999) and Gräfenhan *et al.* (2011)].

Diagnostic features: Orange-red to bright red perithecia with an acute or papillate apex producing cylindrical to narrowly clavate asci, yellow brown to reddish brown, 1-septate, tuberculate ascospores and acremonium-like asexual morph.

Cosmosporella S.K. Huang *et al.*, Cryptog. Mycol. 39: 179. 2018. Figs 8, 19.

Type species: Cosmosporella olivacea S.K. Huang *et al.*, Cryptog. Mycol. 39: 181. 2018.

Ascomata perithecial, solitary to gregarious, superficial, on immersed to erumpent stroma, ovoid, globose to obpyriform, collapsing laterally when dry, orange red, red to pale yellow, not reacting in KOH, with a central ostiole, with hyaline periphyses. *Ascomatal wall* membranous, composed of orange to hyaline cells of *textura angularis*, with septate paraphyses. *Asci* cylindrical to slightly clavate, apically rounded, with evanescent wall, pedicel combined with paraphyses, 8-spored, unitunicate. *Ascospores* hyaline to pale brown, ellipsoidal to ovoid, 0- or 1-septate. *Conidiophores* acremonium-like, mononematous, hyaline, septate, consisting of lateral phialides on somatic hyphae, or with one or two levels of monochasial branching, or irregularly branched. *Conidiogenous cells* monophialidic, cylindrical, producing micro- and macroconidia. *Microconidia* ellipsoidal to obovoid, 0- or 1-septate, hyaline, forming a false head on phialides. *Macroconidia* falcate, almost straight to curved, 1–3-septate, with a blunt to hooked apical cell and poorly to well-developed foot-shaped basal cell. *Chlamydospores* unknown. [Description adapted from Huang *et al.* (2018)].

Diagnostic features: Pale yellow to orange-red perithecia lacking a papilla producing cylindrical to narrowly clavate asci, pale brown, 1-septate ascospores and fusarioid asexual morph characterised by overly long, 1–3-septate macroconidia.

Cosmosporella cavisperma (Corda) Sand.-Den., L. Lombard & Crous, *comb. nov.* MycoBank MB 838659.

Basionym: *Fusarium cavispermum* Corda, Icon. Fung. 1: 3. 1837. *Synonyms:* *Fusarium aquaeductum* var. *cavispermum* (Corda) Raïllo, *Fungi of the Genus Fusarium*: 280. 1950.

Fusarium oxydendri Ellis & Everh., Bull. Torrey Bot. Club 24: 477. 1897.

Fusarium cavispermum var. *minus* Wollenw., *Fusaria Autogr. Delin.* 3: 848. 1930.

Lectotypus: **Czech Republic**, near Carlsstein, on pine resin, A.K.J. Corda, Icon. Fung. 1: tab. I, fig. 58 (MBT 10001322 *hic designatus*). *Epitype* of *Fusarium cavispermum* (CBS 172.31, MBT 10000645 *hic designatus*, a metabolically inactive culture).

Norway, from *Pinus sylvestris*, 1929, H.W. Wollenweber, culture ex-epitype CBS 172.31 = NRRL 13996.

Notes: The genus *Cosmosporella* was erected by Huang *et al.* (2018) to accommodate *Cm. olivacea* and the superfluous taxon *Cm. obscura*, shown to cluster within a subset of taxa pertaining to *Cosmospora s. lat.* (Rossman *et al.* 1999), former members of the *Nectria episphaeria* group *sensu* Booth (1959) and *Nectria* subgenus *Dialonectria* (Samuels *et al.* 1991) characterised by cosmospora-like sexual morphs and fusarioid asexual morphs. More recently, this monophyletic clade had been ascribed to the *Fusarium cavispermum* species complex (O'Donnell *et al.* 2013) and, separated from any of the polyphyletic taxa formerly classified in *Fusarium* section *Eupionnotes* (O'Donnell 1993, Schroers *et al.* 2009, Gräfenhan *et al.* 2011). “*Fusarium*” *melanochlorum*, its purported sexual morph “*Nectria*” *flavoviridis* (Gerlach & Nirenberg 1982), and “*Fusarium*” *cavispermum* have also been resolved as members of this clade (Gräfenhan *et al.* 2011, O'Donnell *et al.* 2013, Huang *et al.* 2018, and Fig. 7 in this paper). Here, the new combination *Cm. cavisperma* is proposed, lectotypified, and an epitype

is designated to stabilise the application of the name based on material studied by Wollenweber [number 849 in Wollenweber (1916–1935) and Gerlach & Nirenberg (1982)]. The suggested conspecificity of “*F.*” *melanochlorum* and “*N.*” *flavoviridis*, however, is questioned given the large phylogenetic distance between the currently available strains. Fresh isolations and a thorough phylogenetic revision of the entire group including additional *Cosmospora s. lat.* taxa having fusarioid asexual morphs are necessary.

Cyanonectria Samuels & Chaverri, Mycol. Progr. 8: 56. 2009. Figs 8, 20.

Type species: Cyanonectria cyanostoma (Sacc. & Flageolet) Samuels & Chaverri, Mycol. Progr. 8: 56. 2009.

Basionym: *Nectria cyanostoma* Sacc. & Flageolet, Rendiconti Congr. Bot. Palermo 1902: 53. 1902.

Synonym: *Fusarium cyanostomum* (Sacc. & Flageolet) O'Donnell & Geiser, *Phytopathology* 103: 404. 2013.

Ascomata perithecial, gregarious or caespitose, with a reduced or well-developed prosenchymatous stroma, smooth- and thin-walled, ampulliform to obpyriform to pyriform, apex dark bluish purple to bluish black and body less intensely dark bluish or red or reddish brown, turning darker in KOH, pigment dissolving in lactic acid to become red to yellow, non-papillate, lacking hairs or appendages. *Ascomatal wall* consisting of a single region, comprising several layers of morphologically similar cells. *Asci* cylindrical to narrowly clavate, with rounded to flattened thickened apex, with or without refractive ring, 8-spored, ascospores overlapping uniseriate or biseriate above and uniseriate below. *Ascospores* ellipsoidal, 1-septate, not or slightly constricted at septum, pale yellow-brown, smooth-walled or finely verrucose. *Conidiophores* mononematous (aerial conidiophores) or grouped on sporodochia; *aerial conidiophores* unbranched or rarely branched, bearing terminal or lateral phialides, often reduced to single phialides. *Conidiogenous cells* monophialidic, cylindrical to subcylindrical, smooth- and thin-walled, with periclinal thickening inconspicuous or absent. *Sporodochia* white to bluish; *sporodochial conidiophores* verticillately branched and densely packed, consisting of short, smooth- and thin-walled stipes bearing apical whorls of 2–3 monophialides; *sporodochial conidiogenous cells* monophialidic, cylindrical to subulate, smooth- and thin-walled, with reduced or flared collarette. *Macroconidia* formed in off-white or creamy or greyish blue slimy masses, falcate, straight to gently curved, with inequilateral fusoid or hooked apical cell and well-developed, foot-shaped basal cell. *Microconidia* unknown. *Chlamydospores* absent or rarely formed from cells of the macroconidia, subglobose. [Description adapted from Samuels *et al.* (2009) and Schroers *et al.* (2011)].

Diagnostic features: Bicoloured or dark bluish purple to bluish black perithecia producing cylindrical to narrowly clavate asci containing ellipsoidal, 1-septate ascospores and fusarioid asexual morph characterised by monophialides producing long, narrow, almost straight macroconidia, lacking microconidia and hyphal-borne chlamydospores.

Dialonectria (Sacc.) Cooke, *Grevillea* 12: 77, 109. 1884. Figs 8, 21. *Basionym:* *Nectria* subgen. *Dialonectria* Sacc., *Syll. Fung.* 2: 490. 1883.

Type species: Dialonectria episphaeria (Tode) Cooke (as “*episphaerica*”), *Grevillea* 12: 82. 1884.

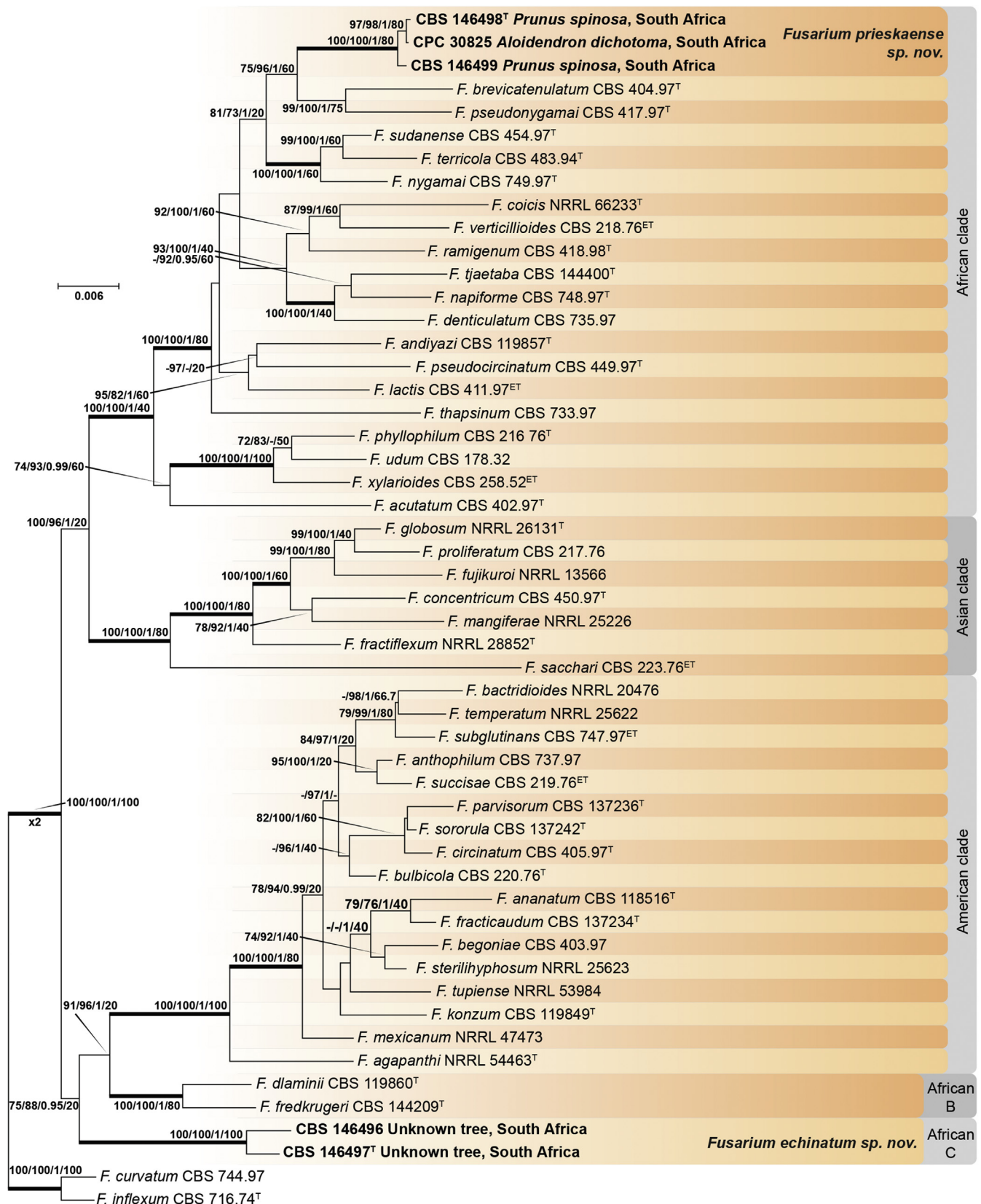


Fig. 11. Maximum-Likelihood (IQ-TREE-ML) consensus tree inferred from the combined *CaM*, *rbp1*, *rbp2*, *tef1*, and *tub2* sequence alignment of members of the *Fusarium fujikuroi* species complex. Numbers at the branches indicate support values (RAxML-BS / UFboot2-BS / BI-PP) above 70 % / 0.95 with thickened branches indicating full support (RAxML-BS / UFboot2-BS = 100 %; BI-PP = 1). Novel taxa are indicated in bold. The scale bar indicates expected changes per site. The tree is rooted to *Fusarium curvatum* CBS 744.97 and *Fusarium inflexum* CBS 716.74. Ex-epitype, ex-neotype and ex-type strains are indicated with ET, NT, and T, respectively.

Basionym: *Sphaeria episphaeria* Tode, Fung. mecklenb. sel. 2: 21. 1791.

Ascomata perithecial, solitary or gregarious, with inconspicuous or absent stroma, obpyriform with an acute or round papilla,

orange red to carmine red, turning dark red in KOH, smooth-walled. *Asci* narrowly clavate to cylindrical, with an apical ring, 8-spored, uniseriate. *Ascospores* initially hyaline, becoming pale brown, 1-septate, becoming tuberculate when mature. *Conidiophores* mononematous, initially as lateral phialides on

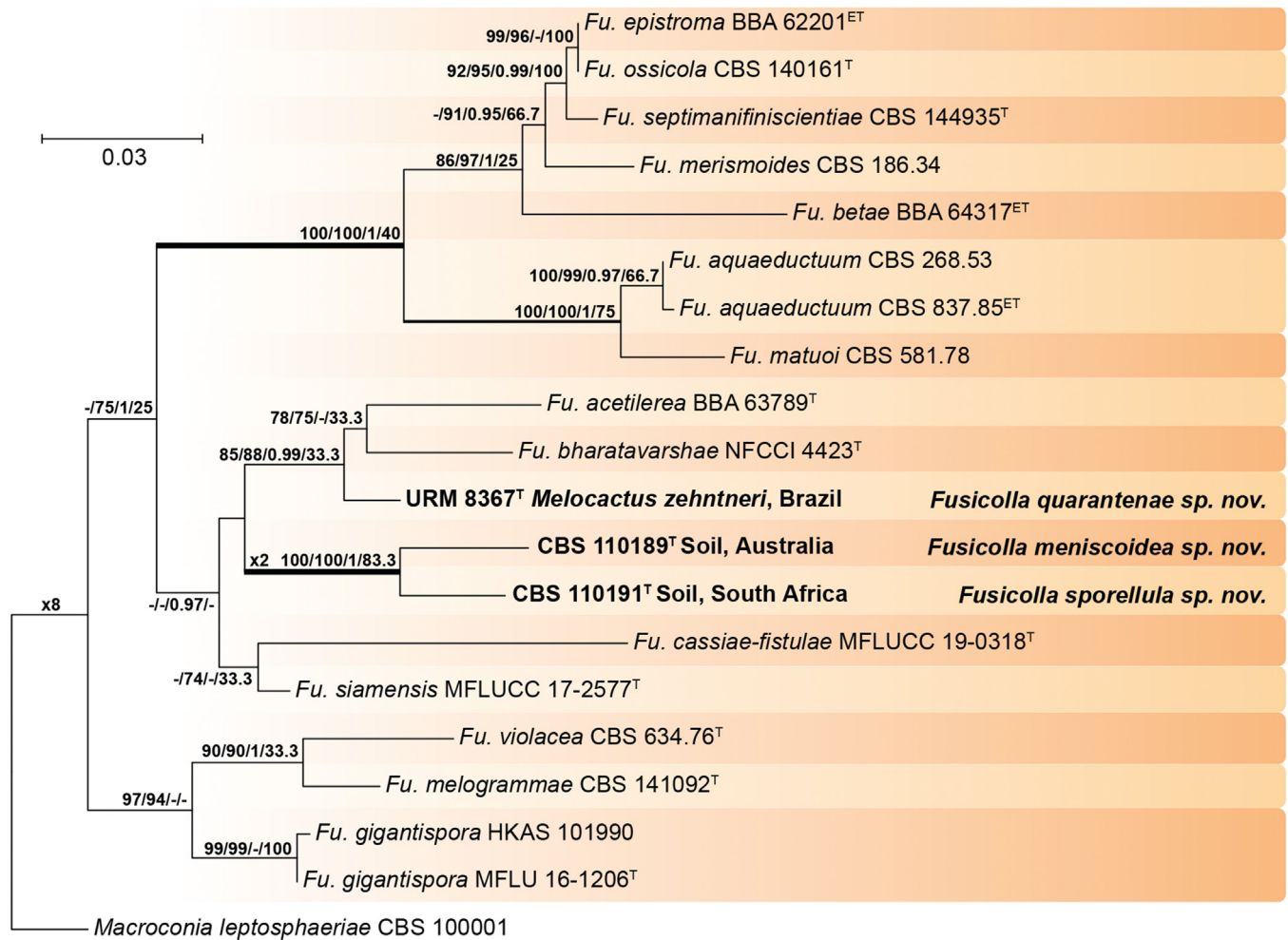


Fig. 12. Maximum-Likelihood (IQ-TREE-ML) consensus tree inferred from the combined *act1*, ITS, LSU, *rpb2*, *tef1*, and *tub2* sequence alignment of members of the genus *Fusicolla*. Numbers at the branches indicate support values (RAxML-BS / UFboot2-BS / BI-PP) above 70 % / 0.95 with thickened branches indicating full support (RAxML-BS / UFboot2-BS = 100 %; BI-PP = 1). Novel taxa are indicated in **bold**. The scale bar indicates expected changes per site. The tree is rooted to *Macroconia leptosphaeriae* CBS 100001. Ex-epitype and ex-type strains are indicated with ET, and T, respectively.

somatic hyphae, sometimes verticillate, hyaline. *Conidiogenous cells* monophialidic, subulate to subclavate, hyaline. *Microconidia* ellipsoidal to clavate, aseptate, hyaline, abundant. *Macroconidia* if present subcylindrical, moderately curved, slightly narrowing towards each end, apical cell often slightly hooked with a more or less pointed tip, basal cell obtuse to poorly developed, foot-shaped, predominantly 3–5-septate, hyaline. *Chlamydospores* unknown.

[Description adapted from [Rossman et al. \(1999\)](#) and [Gräfenhan et al. \(2011\)](#)].

Diagnostic features: Orange-red to carmine-red perithecia with an acute or round papilla producing cylindrical to narrowly clavate asci, pale brown, 1-septate, tuberculate ascospores and asexual morph that rarely produces macroconidia.

Fusarium Link, Mag. Ges. Naturf. Freunde Berlin 3: 10. 1809. [Figs 8, 22](#).

Synonyms: *Fusisporium* Link, Mag. Ges. Naturf. Freunde Berlin 3: 19. 1809.

Selenosporium Corda, Icon. Fung. 1: 7. 1837.

Gibberella Sacc., Michelia 1: 43. 1877.

Lisea Sacc., Michelia 1: 43. 1877.

Sporotrichella P. Karst., Meddel. Soc. Fauna Fl. Fenn. 14: 96. 1887.

Gibberella subgen. *Lisiella* Cooke & Masee, Grevillea 16: 5. 1887.

Lisiella (Cooke & Masee) Sacc., Syll. Fung. 9: 945. 1891.

Septorella Allesch., Hedwigia 36: 241. 1897.

Ustilaginoidella Essed, Ann. Bot. 25: 351. 1911.

Rachisia Linder, Deutsche Essigind. 17: 467. 1913.

Stagonostroma Died., Krypt.-Fl. Mark Brandenb. 9: 561. 1914.

Fusidomus Grove, J. Bot. 67: 201. 1929.

Pseudofusarium Matsush., Microfungi Solomon Isl. Papua-New Guinea: 46. 1971.

Type species: *Fusarium sambucinum* Fuckel, Fungi Rhen. Exs., Fasc. 3, no. 211. 1863, *nom. cons.*

(See List section for synonyms)

Ascomata perithecial, mostly gregarious, non-stromatic or on a thin stroma erumpent through the epidermis, superficial, subglobose to globose to broadly pyriform, not collapsing or laterally pinched when dry, bluish purple to black, turning dark purple in KOH, pigment dissolving in lactic acid, non-papillate, slightly rugose to tuberculate, lacking hairs or appendages. **Ascomatal wall** of two regions: outer region of thick-walled, pigmented cells forming a *textura angularis* or *textura globulosa*; inner region of elongate, hyaline, thin-walled cells, becoming thinner towards the centrum. **Asci** clavate, apex simple, 8-spored often with an apical ring, biseriate to pluriseriate. **Ascospores** ellipsoidal to cylindrical, 1–3-septate, not or slightly constricted at the septa, pale tan, smooth-walled. **Conidiophores** mononematous (aerial conidiophores) or grouped on sporodochia; **aerial conidiophores**, if consistently

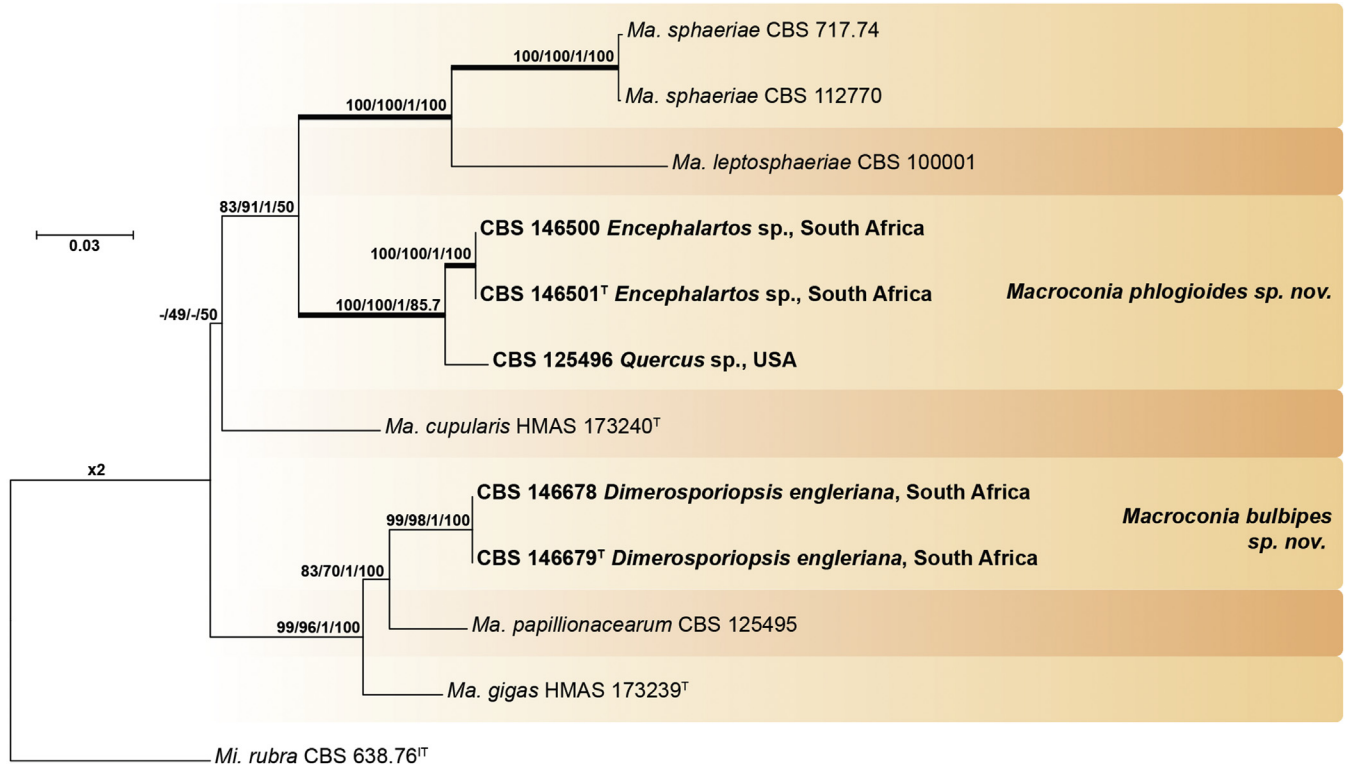


Fig. 13. Maximum-Likelihood (ML) consensus tree inferred from the combined *act1*, *CaM*, ITS, LSU, *rbp1*, *rbp2*, and *tub2* sequence alignment of members of the genus *Macroconia*. Numbers at the branches indicate support values (RAxML-BS / UFboot2-BS / BI-PP / gCF) above 70 % / 0.95 with thickened branches indicating full support (RAxML-BS / UFboot2-BS = 100 %; BI-PP = 1). Novel taxa are indicated in bold. The scale bar indicates expected changes per site. The tree is rooted to *Microcera rubra* CBS 638.76. Ex-type and ex-isotype strains are indicated with T, and IT, respectively.

formed, unbranched, sympodial or irregularly branched, bearing terminal or lateral phialides, often reduced to single phialides. *Conidiogenous cells* mono- or polyphialidic, subulate to subcylindrical, smooth- and thin-walled, sometimes proliferating percurrently, with periclinal thickening inconspicuous or absent. *Aerial conidia* hyaline, smooth- and thin-walled, of three types: *microconidia* ellipsoidal to fusoid to ovoid to obovoid to reniform to allantoid to clavate to napiform to pyriform to limoniform, 0–5-septate, borne in false heads or chains on the phialides; *mesoconidia* (occurring in some species or species complexes) falcate, slender with no significant curvature to curved with parallel walls, 1–5-septate, tapering towards both ends, with a pointed to blunt apical cell and obtuse to flattened basal cell; *macroconidia*, typically formed on sporodochia, falcate, slightly to strongly curved dorsiventrally, 1-septate to multiseptate, with a curved, long and tapering, pointed, blunt, hooked or elongated apical cell and obtuse, poorly developed, well-developed, to elongate, foot-shaped basal cell. *Sporodochia* cream to pale tan to orange to saffron to blue; *sporodochial conidiophores* verticillately branched and densely packed, consisting of short, smooth- and thin-walled stipes bearing an apical whorl of 2–4 monophialides; *sporodochial conidiogenous cells* subulate to subcylindrical, smooth- and thin-walled, with reduced or flared collarete; *sporodochial (macro)conidia* falcate, smooth- and thin-walled, distinctly curved to curved with parallel walls to unequally curved, tapering towards both ends, with pointed, blunt, papillate, hooked, or elongate apical cell and obtuse, poorly developed, well-developed, to elongate, foot-shaped basal cell. *Chlamydospores* globose to subglobose to ovoid to obovoid, hyaline to subhyaline, smooth-walled to slightly verrucose, terminal or intercalary, solitary or in pairs or forming chains or aggregating to form microsclerotia.

[Description adapted from Rossman *et al.* (1999) and Lombard *et al.* (2015)].

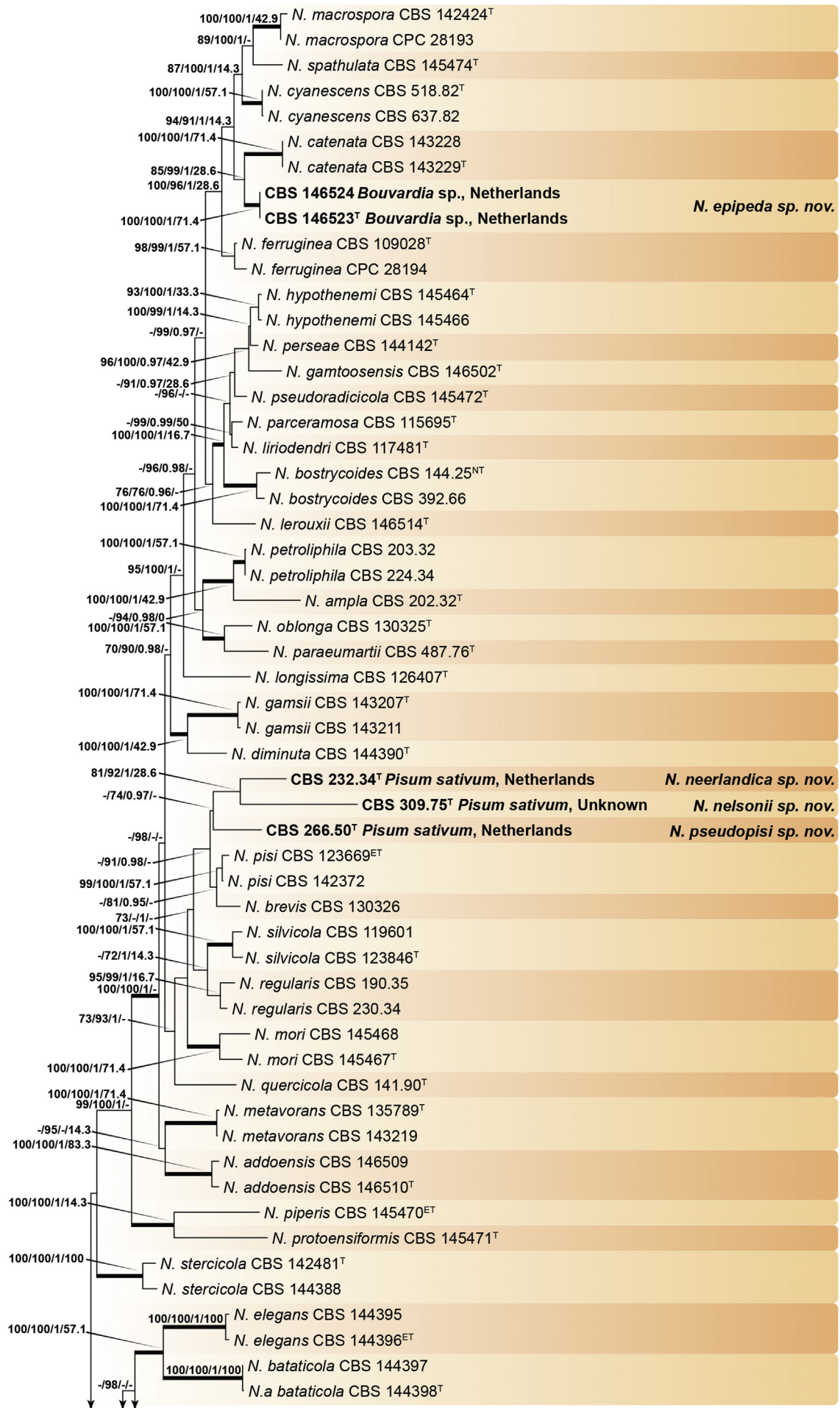
Diagnostic features: Dark blue to black perithecia producing clavate asci bearing ellipsoidal to cylindrical 1- to multiseptate ascospores and asexual morphs producing micro- and macroconidia, and sometimes mesoconidia on aerial conidiophores with mono- and/or polyphialides or only macroconidia in sporodochia. Chlamydospores form in hyphae, rarely in macroconidia.

Fusarium echinatum Sand.-Den. & G.J. Marais, *sp. nov.* MycoBank MB 838660. Fig. 23.

Etymology: From the Latin *echinatus*, prickly, referring to the spiny appearance of its multiloculate, often swollen and rather deformed conidiogenous cells.

Typus: South Africa, unidentified tree species, 2010, A. Lubben (**holotype** CBS H-24658, culture ex-type CBS 146497 = CPC 30815 = CAMS 000733).

Conidiophores on aerial mycelium 10–120 µm tall, unbranched or irregularly laterally branched, bearing lateral and terminal single phialides; *aerial conidiogenous cells* polyphialidic, subulate, subcylindrical or more commonly irregularly shaped, curved, swollen and distorted due to abundant conidiogenous loci, smooth- and thin-walled, 6.5–36.5 × 2–3.5 µm, polyphialides with 2–3 or more commonly 10–18 conidiogenous openings, with inconspicuous to absent periclinal thickening and collarettes. *Aerial microconidia* forming small false heads on tips of phialides, hyaline, smooth, and thin-walled, commonly ovoid to ellipsoidal, 0- or 1-septate, 4–11(–19) × 2–3.5(–4.5) µm (av. 7.5 × 2.7 µm), and more rarely napiform, smooth and thin-walled, 0-septate, (5–)5.5–7 × (3.5–)4.5–5.5 µm (av. 6.4 × 4.5 µm). *Sporodochial conidiophores* 28.5–60(–68.5) µm tall, irregularly branched, bearing terminal solitary monophialides or whorls of up to three monophialides. *Sporodochial conidiogenous*



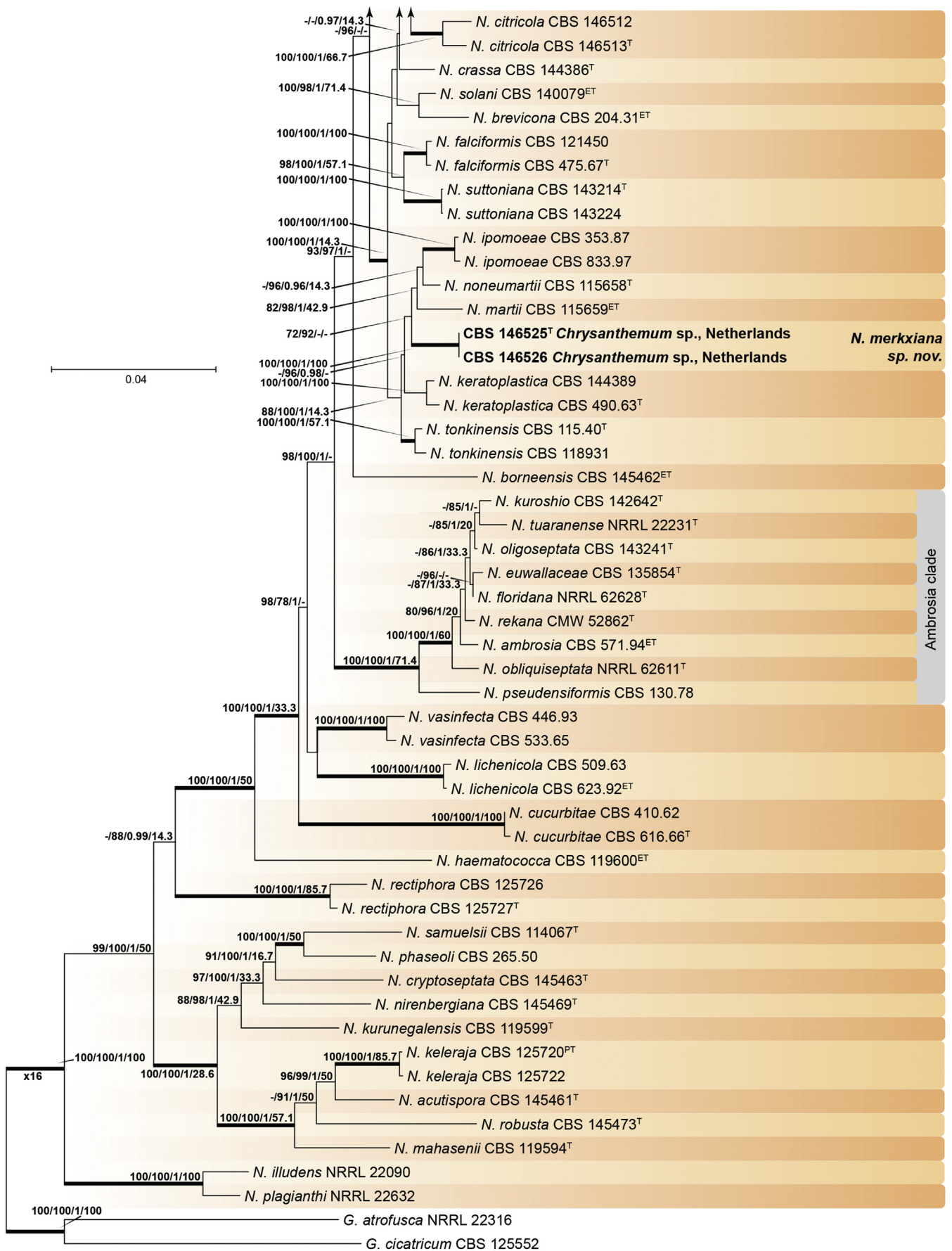


Fig. 14. Maximum-Likelihood (IQ-TREE-ML) consensus tree inferred from the combined *act1*, *CaM*, *ITS*, *LSU*, *rpb1*, *rpb2*, and *tef1* sequence alignment of members of the genus *Neocosmospora*. Numbers at the branches indicate support values (RAxML-BS / UFboot2-BS / I-PP) above 70 % / 0.95 with thickened branches indicating full support (RAxML-BS / UFboot2-BS = 100 %; BI-PP = 1). Novel taxa are indicated in **bold**. The scale bar indicates expected changes per site. The tree is rooted to *Geejayessia atrofusca* NRRL 22316 and *G. cicatricum* CBS 125552. Ex-epitype, ex-neotype, ex-paratype and ex-type strains are indicated with ET, NT, PT, and T, respectively.

cells monophialidic, subulate to subcylindrical, smooth- and thin-walled, (8.5–)11.5–16(–17.5) × (1.5–)2.5–3.5 µm. *Sporodochial macroconidia* moderately curved to wedge-shaped, slender, tapering towards the basal part, apical cell of equal size than the adjacent cell, blunt to slightly hooked; basal cell poorly to well-developed, foot-shaped, (1–)2–3(–4)-septate, hyaline, thin- and smooth-walled: 1-septate conidia: (16.5–)19.5–32.5(–36) × 2.5–3.5 µm (av. 26.1 × 2.9 µm); 2-septate conidia: (19.5–)25–36(–37.5) × 2.5–3.5 µm (av. 30.5 × 3.1 µm); 3-septate conidia: (20.5–)28.5–36(–40) × (2.5–)3–3.5(–4.5) µm (av. 32.5 × 3.2 µm); 4-septate conidia: (27–)30.5–39(–40.5) × 3–4 µm (av. 35.4 × 3.6 µm); overall: (19.5–)28.6–36.5(–40.5) × (2.5–)3–3.5(–4.5) µm (av. 32.4 × 3.2 µm). *Chlamydoconidia* not observed.

Culture characteristics: Colonies on PDA reaching 31–63 mm diam at 25 °C after 7 d. Surface white, pale luteus to sulphur yellow, flat, woolly to cottony with radial patches of white aerial mycelium, margin regular and filiform. Reverse white, sulphur yellow to pure yellow at centre. On OA pale luteus to sulphur yellow, flat, membranous at first, quickly becoming velvety to dusty, margin regular. Reverse sulphur yellow.

Additional material examined: **South Africa**, unidentified tree species, 2010, A. Lubben, culture CBS 146496 = CPC 30814 = CAMS 000730.

Notes: Yilmaz et al. (2021) recently revised the FFSC, including formal descriptions for several species, while fixing the typification of relevant plant pathogenic and toxigenic species. Species in this complex have been traditionally organised according to their biogeographic patterns, which roughly match their phylogenetic distribution. Apart from the monophyletic American and Asian clades, the complex contains a non-monophyletic African clade, which is currently known to cluster into two distinct clades: the speciose core African clade and the African “B” clade encompassing *F. dlamini* and *F. fredkrugeri* (O'Donnell et al. 2000b, Herron et al. 2015, Sandoval-Denis et al. 2018b, Yilmaz et al. 2021). The novel South African species *F. echinatum*, however, formed a fully-supported single lineage that did not belong to any of the currently known biogeographically defined clades (Fig. 11). The most noticeable morphological feature that distinguishes *F. echinatum* is the presence of well-developed polyphialides bearing multiple conidiogenous openings that are often concentrated in large numbers and that cause a deformation of the apical region. Somewhat similar, conspicuous polyphialides can be found in *Fusarium chlamydoconidii* and *F. concolor* (syn. *F. polyphialidicum*); however, these species are not directly related, in that they belong to two different species complexes, the *F. chlamydoconidii* and *F. concolor* species complexes, respectively (Fig. 10). The polyphialides formed by these two species do not show as many conidiogenous loci as do those of *F. echinatum*.

Fusarium prieskaense G.J. Marais & Sand.-Den., *sp. nov.* MycoBank MB 838661. Fig. 24.

Etymology: Referring to Prieska, a town in Northern Cape Province, South Africa, where the type was collected.

Typus: **South Africa**, Northern Cape Province, Prieska, on *Prunus spinosa*, 2010, F.J.J. van der Walt & G.J. Marais (**holotype** CBS H-24660, culture ex-type CBS 146498 = CPC 30826 = CAMS 001176).

Conidiophores on aerial mycelium 12.5–43.5 µm tall, unbranched or rarely irregularly or sympodially branched and proliferating, bearing terminal single phialides or whorls of 2–3 phialides, commonly reduced to solitary conidiogenous cells borne laterally on hyphae; *aerial conidiogenous cells* mono- and polyphialides, subulate to subcylindrical, smooth- and thin-walled, 8–29.5 × 2–5 µm, polyphialides often with 2–3 conidiogenous openings, periclinal thickening and collarettes often inconspicuous or absent. *Aerial microconidia* forming small false heads and short chains on phialide tips, hyaline, obovoid to short clavate, smooth and thin-walled, 0-septate, (4.5–)6–9(–13) × 2–3(–4) µm (av. 7.4 × 2.6 µm). *Sporodochial conidiophores* 24.5–39(–45) µm tall, irregularly branched, bearing terminal solitary or whorls of 2–3 phialides. *Sporodochial conidiogenous cells* monophialidic, doliform, subulate to subcylindrical, smooth- and thin-walled, (8.5–)10–14(–15) × 2–4.5 µm. *Sporodochial conidia* straight to moderately curved and slender, tapering towards the basal part, apical cell more or less equally sized as the adjacent cell, blunt to slightly hooked; basal cell well-developed, foot-shaped, rarely papillate, (1–)3–4-septate, hyaline, thin- and smooth-walled: 1-septate conidia: 23.5 × 3.5 µm; 3-septate conidia: (33.5–)44.5–58(–68.5) × (3–)3.5–4.5(–5) µm (av. 51.1 × 4 µm); 4-septate conidia: (52.5–)55.5–67.5(–71) × 3.5–4.5 µm (av. 61.3 × 4.1 µm); overall: (23–)44–59(–71) × 3–4(–5) µm (av. 51.3 × 4 µm). *Chlamydoconidia* not observed.

Culture characteristics: Colonies on PDA reaching 42–68 mm diam at 25 °C after 7 d. Surface pale luteous, luteous to pale sienna, flat, velvety to felty, sometimes with small white patches of aerial mycelium, margin filiform and regular. Reverse sulphur yellow to amber, pale orange at centre. On OA, sienna to pale umber, flat, membranous to dusty, margin entire and regular; reverse sienna to pale umber.

Additional material examined: **South Africa**, Northern Cape Province, Prieska, on *Prunus spinosa*, 2010, F.J.J. van der Walt & G.J. Marais, culture CBS 146499 = CPC 30827 = CAMS 001177; on *Aloidendron dichotomum*, 2010, F.J.J. van der Walt & G.J. Marais, culture CPC 30825 = CAMS 001175.

Notes: *Fusarium prieskaense* is nested within the core African clade of the FFSC (Fig. 11). Similar to most members of this clade, this species is characterised by forming mostly monophialides and occasional to frequent polyphialides, sometimes proliferating and producing aerial conidia typically organised in a combination of false heads and short to long chains. *Fusarium prieskaense* is morphologically and phylogenetically related to *Fusarium brevicatenuatum* and *F. pseudonygamai* from which it can be differentiated by its pale luteus to yellow colony pigmentation on PDA, versus the orange to dark blue or violet pigments produced by the two latter species (Leslie & Summerell 2006). Additionally, sporodochia and macroconidia are commonly and abundantly produced by *F. prieskaense*, whereas these structures are relatively rare in the two aforementioned species. Moreover, the obovoid to short clavate microconidia of *F. prieskaense* also distinguishes this species from *F. brevicatenuatum*, which is characterised by long oval to obovoid microconidia (Nirenberg et al. 1998).

Fusicolla Bonord., Handb. Allg. Mykol.: 150. 1851. Figs 8, 25.

Type species: *Fusicolla betae* (Desm.) Bonord., Handb. Allg. Mykol.: 150. 1851.

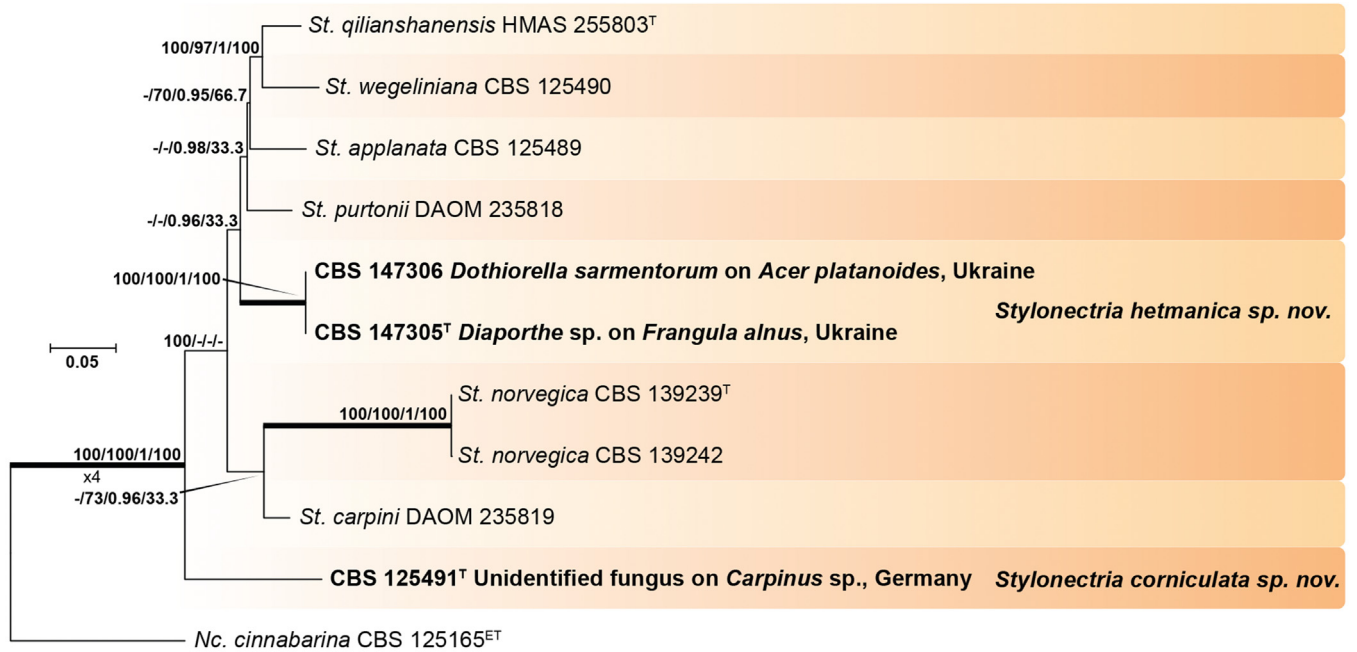


Fig. 15. Maximum-Likelihood (IQ-TREE-ML) consensus tree inferred from the combined *act1*, ITS, and *rpb2* sequence alignment of members of the genus *Styлонectria*. Numbers at the branches indicate support values (RAxML-BS / UFboot2-BS / BI-PP) above 70 % / 0.95 with thickened branches indicating full support (RAxML-BS / UFboot2-BS = 100 %; BI-PP = 1). Novel taxa are indicated in **bold**. The scale bar indicates expected changes per site. The tree is rooted to *Macroconia leptosphaeriae* CBS 100001. Ex-epitype and ex-type strains are indicated with ET and T, respectively.

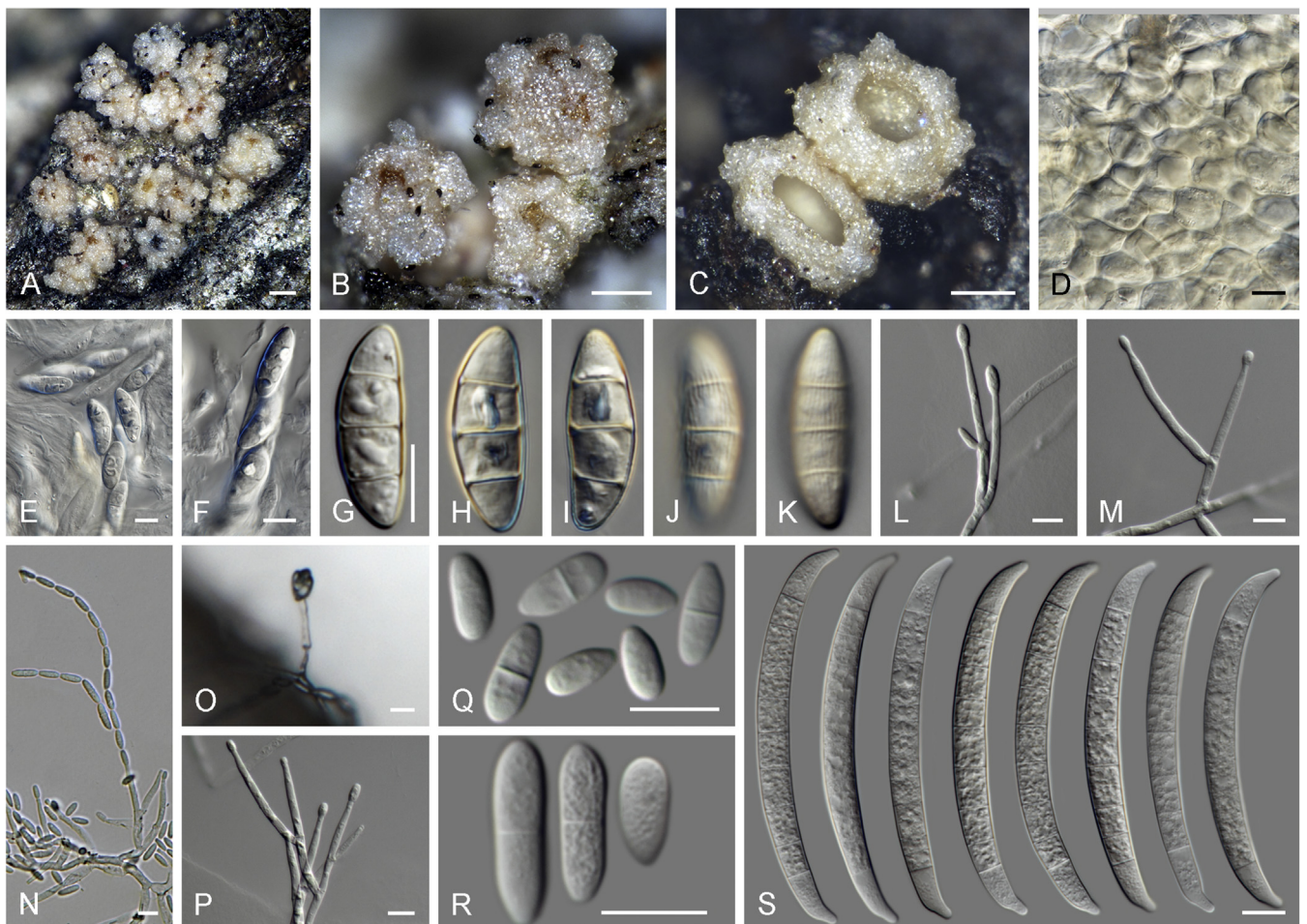


Fig. 16. *Albonectria* spp. **A–C.** Ascomata on natural substrate. **D.** Surface view of perithecial wall in 2% KOH. **E–K.** Asci and ascospores (**J, K.** Surface view). **L–P.** Conidiophores and conidiogenous cells. **Q, R.** Microconidia. **S.** Macroconidia. **A, C–F, H–J.** *Albonectria rigidiuscula* (BPI 553050). **B, G, K.** *Albonectria rigidiuscula* (BPI 1104484). **L, M, P–S.** *Albonectria rigidiuscula* (CBS 122570). **N, O.** *Albonectria rigidiuscula* (CBS 133.25). Scale bars: **A–C** = 100 μm; all others = 10 μm (**G** applies to **H–K**).

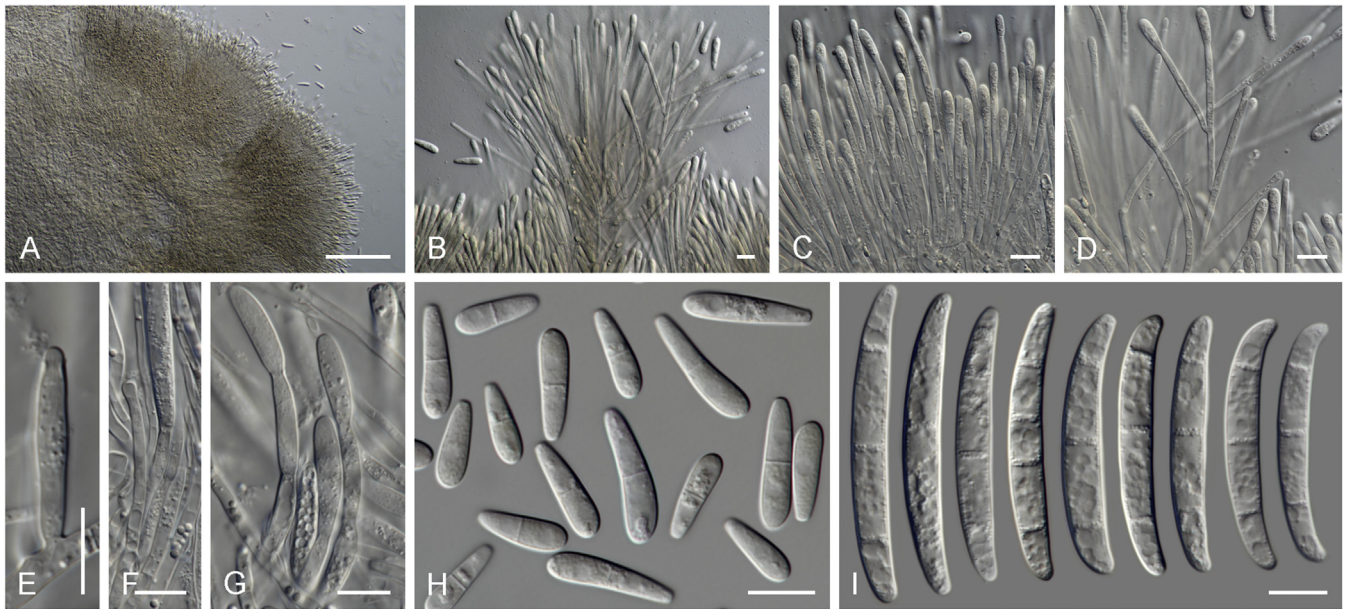


Fig. 17. *Atractium* spp. A, B. Synnemata. C–G. Conidiophores and conidiogenous cells. H. Microconidia. I. Macroconidia. A–D, H. *Atractium stilbaster* (CBS 410.67). E–G, I. *Atractium crassum* (CBS 180.31). Scale bars: A = 100 μ m; all others = 10 μ m.

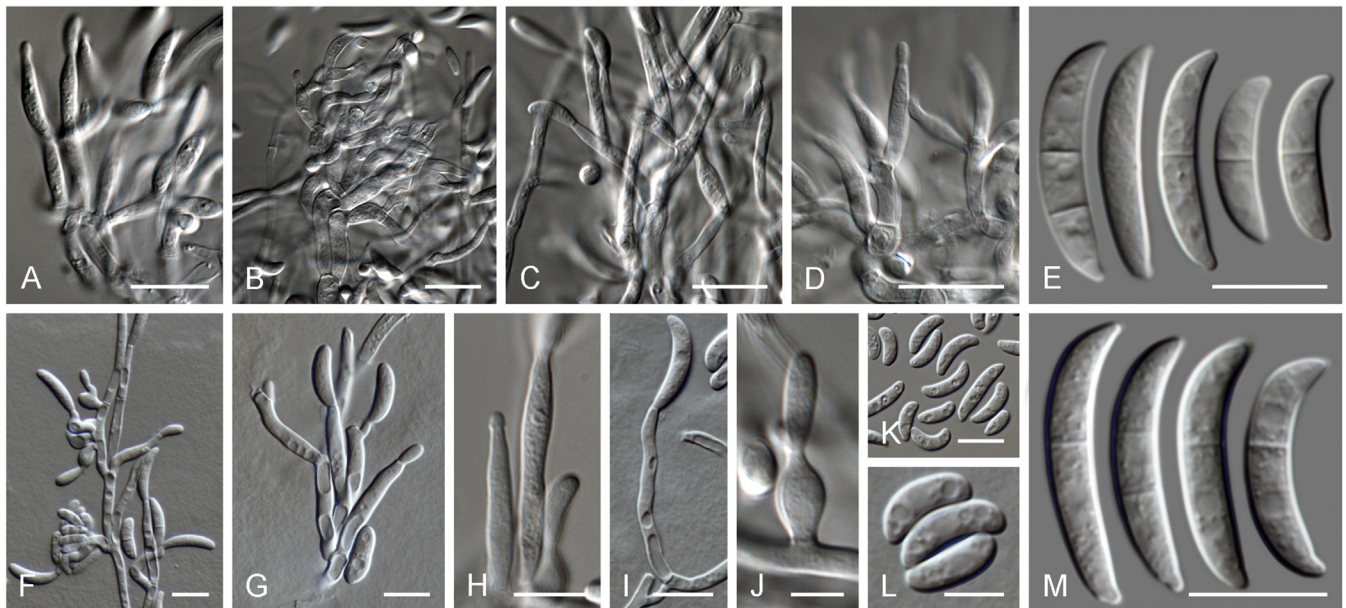


Fig. 18. *Bisufusarium* spp. A–D, F–J. Conidiophores and conidiogenous cells. K, L. Microconidia. E, M. Macroconidia. A–E. *Bisufusarium dimerum* (CBS 108944). F–M. *Bisufusarium delphinoides* (CBS 120718). Scale bars: H, J = 5 μ m; all others = 10 μ m.

(See *F. betae* in List section for synonyms)

Ascomata perithecial, solitary, rarely gregarious, with erumpent stroma, fully or partially immersed in a slimy, pale orange sheet of hyphae over the substrate, globose to pyriform with a short acute or disk-like papilla, not collapsing when dry, yellow, pale buff to orange, not changing colour in KOH, smooth-walled, rarely tuberculate, generally lacking hairs or with short, thick-walled hyphae-like structures. *Asci* cylindrical to narrowly clavate, with an apical ring, 8-spored. *Ascospores* broadly ellipsoidal, 1-septate, slightly constricted at the septum, verrucose, hyaline to pale brown. *Conidiophores* initially as lateral phialides on somatic hyphae, sometimes monochasial, verticillate or penicillate, hyaline. *Conidiogenous cells* monophialidic, cylindrical to subulate, hyaline. *Microconidia* absent or sparse, ellipsoidal to allantoid, aseptate, hyaline. *Macroconidia* falcate, straight to curved, narrowing towards the ends, apical cell often hooked with a pointed tip, basal cell poorly

developed, foot-shaped, 1–3-septate or 3–5-septate or up to 10-septate, hyaline. *Chlamydoconidia* absent to abundant, globose, single, in pairs or chains, sometimes formed in macroconidia.

[Description adapted from Gerlach & Nirenberg (1982) and Gräfenhan *et al.* (2011)].

Diagnostic features: Yellow to orange, mostly smooth-walled perithecia with a short acute or disk-like papilla producing cylindrical to narrowly clavate asci bearing broadly ellipsoidal, 1-septate, verrucose ascospores and fusarioid asexual conidia.

Fusicolla quarantena J.D.P. Bezerra, Sand.-Den., Crous & Souza-Motta, *sp. nov.* MycoBank MB 838692. Fig. 26.

Etymology: The epithet refers to the quarantine period during the 2020–2021 coronavirus pandemic, which killed thousands of people on five continents, and during which this species was described.

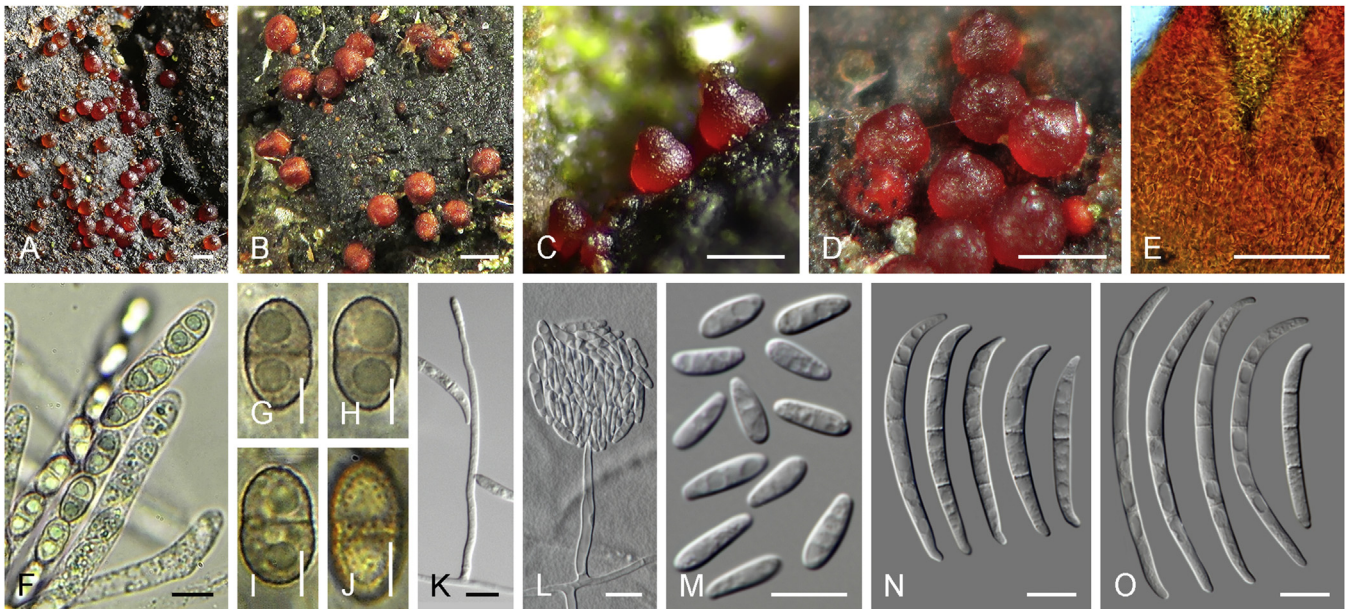


Fig. 19. *Cosmospora* spp. **A–D.** Ascomata on natural substrate. **E.** Surface view of perithecial wall. **F.** Asci. **G–J.** Ascospores. **K, L.** Conidiophores. **M.** Microconidia. **N, O.** Macroconidia. **A–J.** “*Cosmospora flavoviridis*” (photos P. Mičoch). **K–N.** “*Cosmospora flavoviridis*” (CBS 124353). **O.** *Cosmospora cavisperma* (CBS 172.31). Scale bars: **A–D** = 300 µm; **E** = 50 µm; **G–J** = 5 µm; all others = 10 µm.

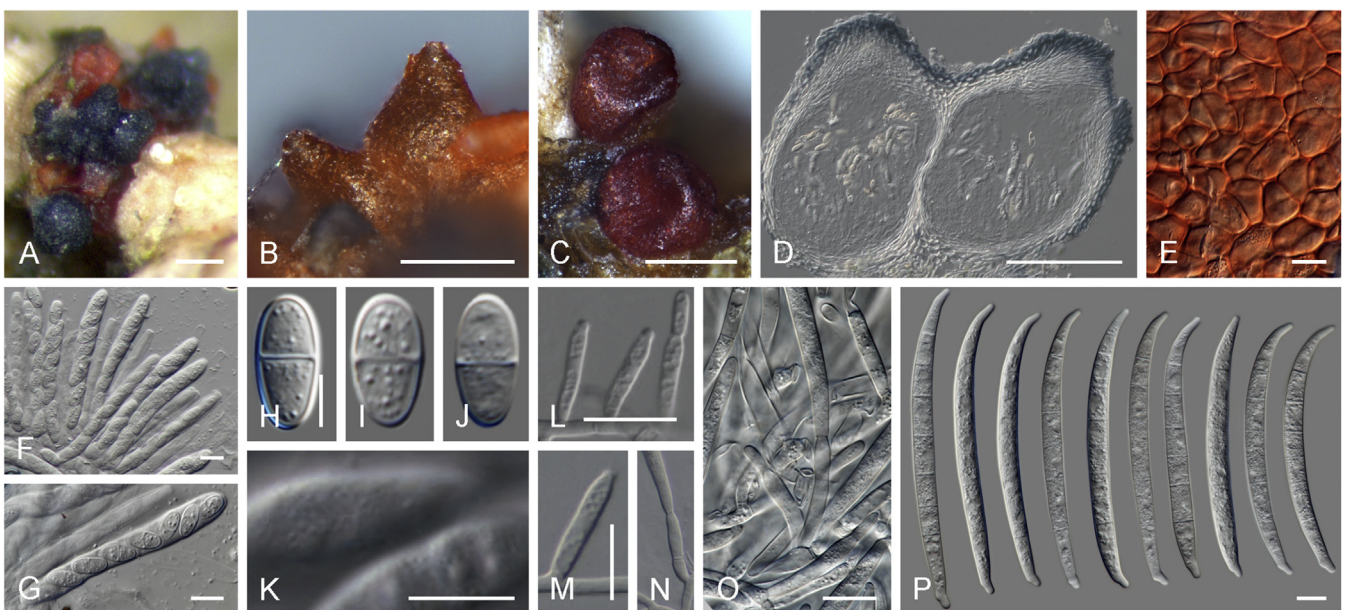


Fig. 20. *Cyanonectria* spp. **A–C.** Ascomata on natural substrate. **D.** Longitudinal section through perithecium in Shears. **E.** Surface view of perithecial wall in 2% KOH. **F, G.** Asci. **H–K.** Ascospores (**K.** Surface view). **L–O.** Conidiogenous cells. **P.** Macroconidia. **A–C, E–J.** *Cyanonectria buxi* (CBS H-20380). **D, K.** *Cyanonectria buxi* (CBS H-20379). **L–N.** *Cyanonectria buxi* (CBS 130.97). **O, P.** *Cyanonectria buxi* (CBS 125551). [**A, D, L.** adapted from [Schroers et al. \(2011\)](#).] Scale bars: **A–D** = 100 µm; **H–K** = 5 µm (**H** applies to **I** and **J**); all others = 10 µm.

Typus: Brazil, Pernambuco state, Itaíba municipality, Curral Velho Farm, 9°08.895 S 37°12.069 W, on cladodes of *Melocactus zehntneri*, Sep. 2013, J.D.P. Bezerra (**holotype** URM 94407, culture ex-type URM 8367 = CBS 141541).

Conidiophores arising laterally from somatic hyphae, simple, straight, hyaline, thin- and smooth-walled, septate, 25–116 × 1.5–2.5 µm, or reduced to solitary conidiogenous cells. **Conidiogenous cells** monophialidic, arising laterally from hyphae, cylindrical to subulate, straight, hyaline, thin- and smooth-walled, 1–22 × 0.5–2 µm, or as short lateral pegs. **Macroconidia** falcate, more or less straight, slightly narrowing towards the ends, apical cell often hooked with a more or less pointed tip, basal cell poorly developed, foot-shaped,

hyaline, thin- and smooth-walled, 3-septate, (21–) 27–35(–38.5) × 2–2.5(–3) µm (av. 29.5 × 2.5 µm, n = 30). **Microconidia**, **chlamydospores** and **sexual morph** not observed.

Culture characteristics: Colonies on PDA reaching 15 mm diam after at 25 °C after 7 d. Surface yellow to apricot in centre, peach to brick in middle, and salmon at margin, flat, aerial mycelium absent, slimy, with entire margin; reverse yellow to brick.

Notes: *Fusicolla quarantanae*, an endophyte of *Melocactus zehntneri*, is morphologically reminiscent of *Fu. betae*, *Fu. epistroma*, and *Fu. septimanifiniscentiae*, all of which produce mainly 3-septate macroconidia. *Fusicolla betae* and *Fu. epistroma* differ by having larger conidia (50–60 µm and 19–45 µm long,

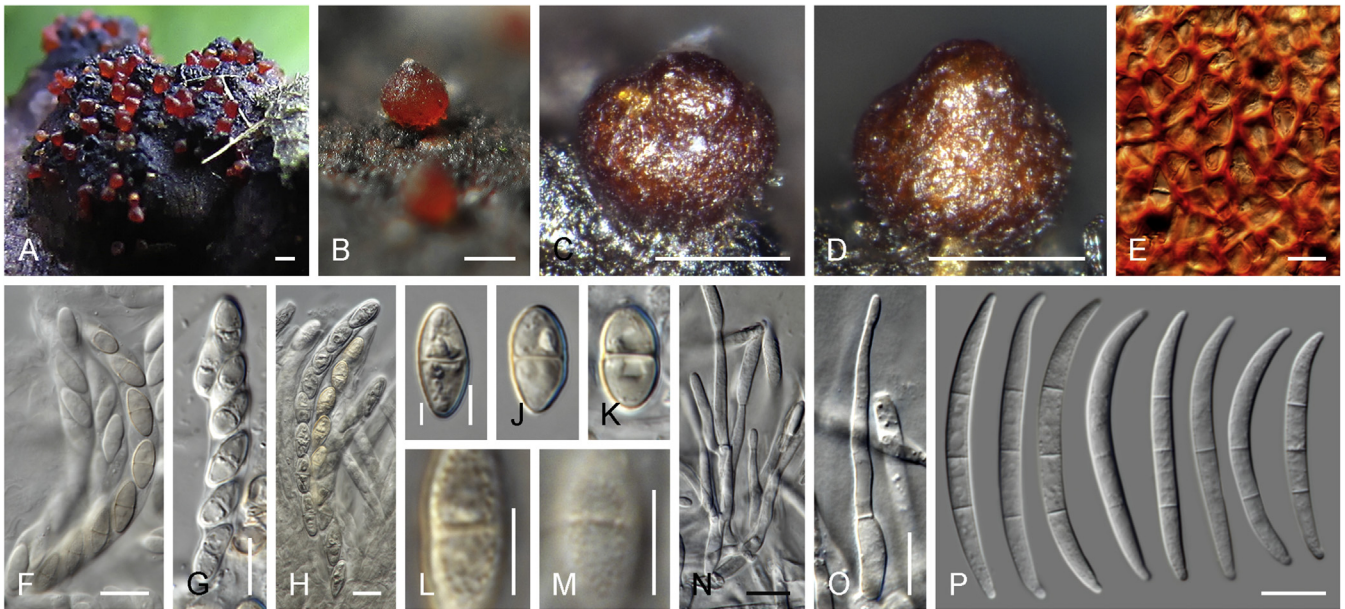


Fig. 21. *Dialonectria* spp. **A–D.** Ascomata on natural substrate. **E.** Surface view of perithecial wall in 2 % KOH. **F–H.** Asci. **I–M.** Ascospores (L, M. Surface view). **N, O.** Conidiophores and conidiogenous cells. **P.** Macroconidia. **A, B.** *Dialonectria episphaeria* (photos P. Mičoch). **C, D, F, M.** *Dialonectria episphaeria* (CBS H-19716). **E, G, K.** *Dialonectria sanguinea* (CBS H-2127). **H–J, L.** *Dialonectria episphaeria* (CBS H-2662). **N–P.** *Dialonectria episphaeria* (CBS 125494). Scale bars: A–D = 100 µm; I, L, M = 5 µm (I applies to J and K); all others = 10 µm.

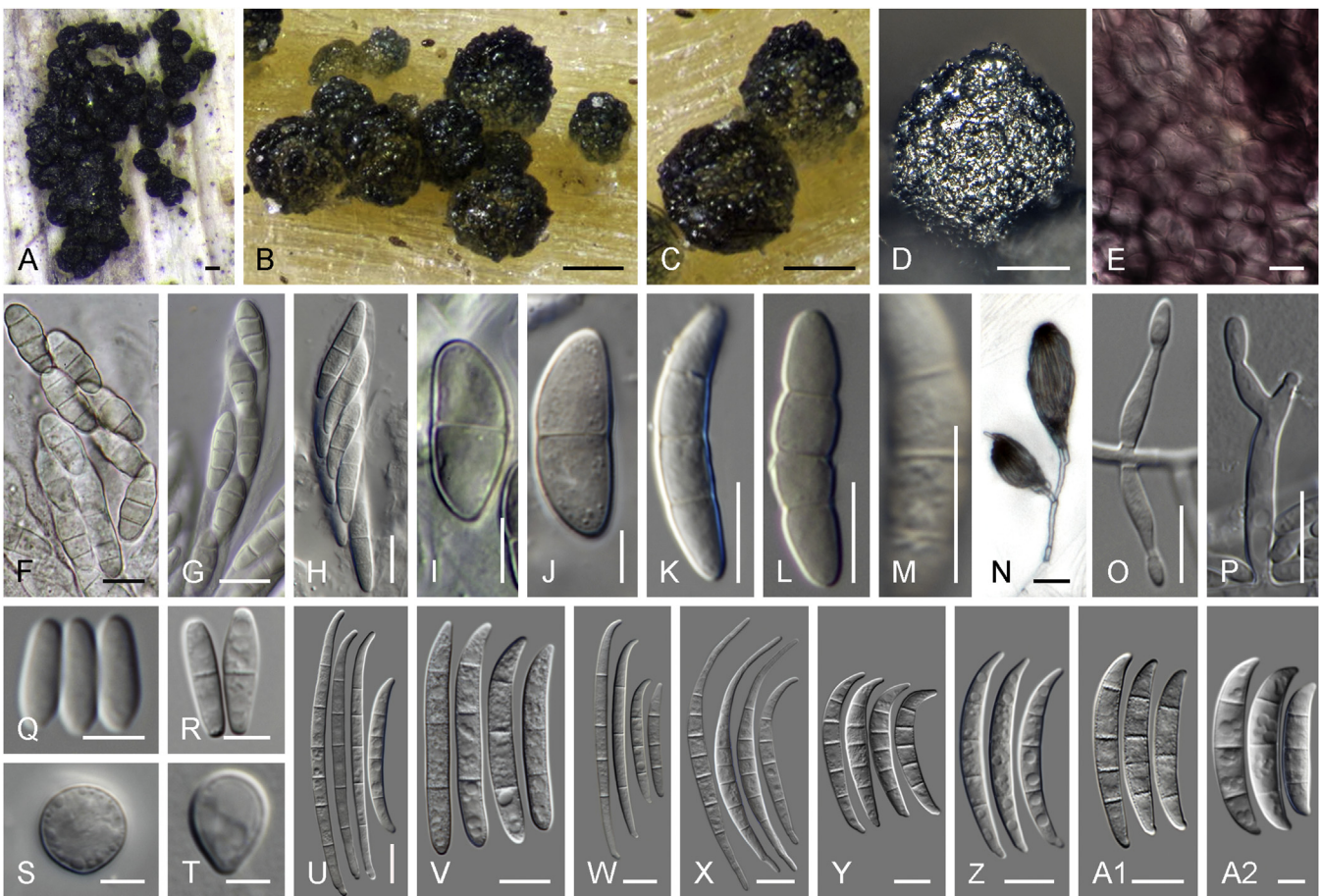


Fig. 22. *Fusarium* spp. **A–D.** Ascomata on natural substrate. **E.** Surface view of perithecial wall in 2 % KOH. **F–H.** Asci. **I–M.** Ascospores (M. Surface view). **N–P.** Conidiophores and conidiogenous cells. **Q–T.** Microconidia. **U–A2.** Macroconidia. **A.** *Fusarium graminearum* (photo P. Cannon). **B, C, F.** *Fusarium sambucinum* [adapted from Wergen (2018)]. **D.** *Fusarium* sp. (HPC 2244). **E.** *Fusarium cf. tricinctum* (CBS H-12819). **G, I.** *Fusarium lateritium* (photo P. Cannon). **H, K.** *Fusarium equiseti* (CBS H-12817). **J.** *Fusarium sambucinum* (BPI 632307). **L, M.** *Fusarium sambucinum* (CBS H-12818). **N.** *Fusarium avenaceum* (CPC 30660). **O, Q.** *Fusarium fredkrugeri* (CBS 144209). **P, W.** *Fusarium prieskaense* (CBS 146498). **R.** *Fusarium madaense* (CBS 146669). **S.** *Fusarium globosum* (CBS 428.97). **T.** *Fusarium echinatum* (CBS 146497). **U.** *Fusarium avenaceum* (CBS 408.86). **V.** *Fusarium caeruleum* (CBS 146590). **X.** *Fusarium longicaudatum* [CBS 123.73, adapted from Xia et al. (2019)]. **Y.** *Fusarium transvaalense* [CBS 144211, adapted from Sandoval-Denis et al. (2018b)]. **Z.** *Fusarium gamsii* (CBS 143610). **A1.** *Fusarium oxysporum* [CBS 144134, adapted from Lombard et al. (2019b)]. **A2.** *Fusarium convolutans* [CBS 144207, adapted from Sandoval-Denis et al. (2018b)]. Scale bars: A–D = 100 µm; I–M, Q–T = 5 µm; all others = 10 µm.

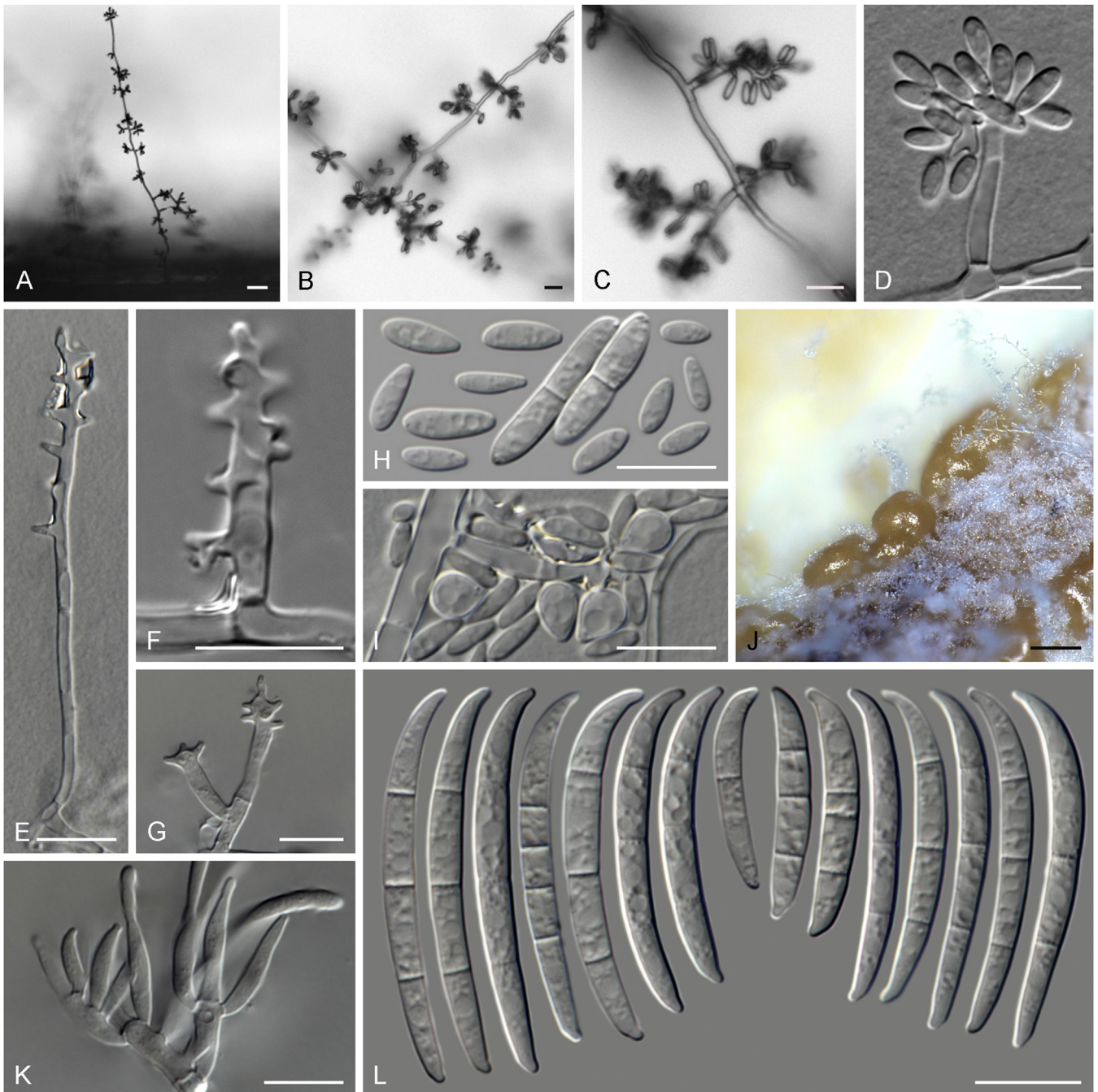


Fig. 23. *Fusarium echinatum* (CBS 146497). **A–D.** Aerial conidiophores. **E–G.** Conidiogenous cells on aerial conidiophores. **H, I.** Microconidia. **J.** Sporodochia formed on the surface of carnation leaves. **K.** Sporodochial conidiophores and conidiogenous cells. **L.** Macroconidia. Scale bars: A = 20 μm ; J = 100 μm ; all others = 10 μm .

respectively, Gerlach & Nirenberg 1982). The absence of chlamydospores in *Fu. quarantena* further differentiates this species from *Fu. epistroma* and *Fu. septimanifiniscentiae* (Gerlach & Nirenberg 1982, Crous *et al.* 2018).

Fusicolla meniscoidea L. Lombard & Sand.-Den., *sp. nov.*
MycoBank MB 838662. Fig. 27.

Etymology: From Greek *mēniskos*, crescent, in reference to the shape of its conidia.

Typus: **Australia**, from soil, unknown collection date (before 1978), unknown collector (**holotype** CBS H-24662, culture ex-type CBS 110189 = FRC E-0086).1

Conidiophores arising laterally or terminally from somatic hyphae 50–70 μm long, simple or sparingly branched laterally, straight,

hyaline, smooth- and thin-walled, bearing terminal and lateral conidiogenous cells, or more commonly reduced to single conidiogenous cells borne laterally on the substrate and aerial hyphae. **Conidiogenous cells** monophialidic, subcylindrical, cylindrical to slightly subulate, 10.5–35 \times 2–3.5 μm , smooth- and thin-walled, without noticeable periclinal thickening, a minute apical collarette can be present. **Macroconidia** falcate, tapering gently towards both ends, apical cell often hooked with a blunt to pointy apex, basal cell obtuse to poorly developed, foot-shaped, 0–2(–3)-septate, predominantly 1-septate, hyaline, smooth- and thin-walled; 0-septate (8–)9–13(–15) \times 2–3.5 μm (av. 11.1 \times 2.9 μm); 1-septate, (9–)11.5–15(–17.5) \times 2.5–3.5 μm (av. 13.1 \times 2.9 μm); 2-septate, 13–17.5(–18) \times 2.5–4 μm (av. 15.4 \times 3 μm); 3-septate, 20–24.5(–25.5) \times 3–3.5 μm (av. 22.6 \times 3.3 μm). **Microconidia**, **chlamydospores** and **sexual morph** not observed.

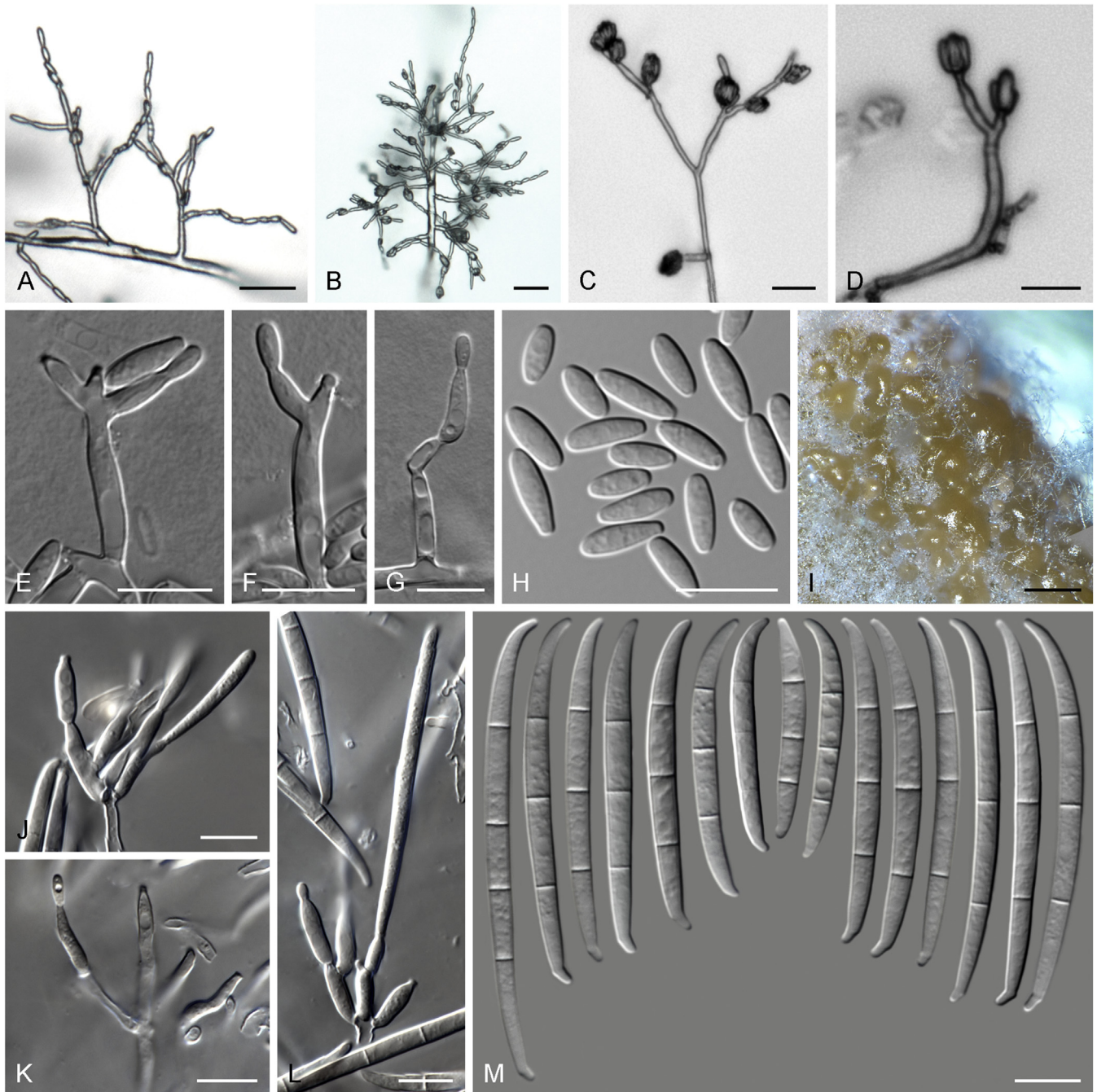


Fig. 24. *Fusarium prieskaense* (CBS 146498). **A–D.** Aerial conidiophores. **E–G.** Conidiogenous cells on aerial conidiophores. **H.** Microconidia. **I.** Sporodochia formed on the surface of carnation leaves. **J–L.** Sporodochial conidiophores and conidiogenous cells. **M.** Macroconidia. Scale bars: A, B = 20 μm ; I = 100 μm ; all others = 10 μm .

Culture characteristics: Colonies on PDA reaching 21–30 mm diam at 25 °C after 7 d. Surface white to pale luteus at periphery, centre salmon to pale orange, flat to slightly radially folded, membranous to slimy, margin entire to slightly undulate; reverse luteous to pale salmon at centre. On OA, pale luteous to pale salmon, flat, membranous, margin entire; reverse pale luteous.

Notes: *Fusicolla meniscoidea* is here introduced based on an isolate originally misidentified as *Bisifusarium dimerum*. Despite the great genetic differences and phylogenetic distance, the two taxa share similar morphological traits, particularly regarding macroscopic aspects of colonial growth, and the shape and size of conidiophores and conidia. However, unlike in *B. dimerum*, conidia of *Fu. meniscoidea* present a much more pronounced curvature involving both conidial planes (somewhat parallel walls), while foot-shaped basal cells are

less evident or absent. *Fusicolla aqueductum*, *Fu. betae*, *Fu. quarantanae*, and *Fu. violacea* are all morphologically related to *Fu. meniscoidea* by showing similar conidial septation ranges and lacking chlamydospores. Conidial size in *Fu. meniscoidea* is, however, markedly reduced and often closer to the lower limits of the conidial size of all the aforementioned species. Another species also described here, *Fusicolla sporellula*, lacks chlamydospores but has similar, although smaller, conidia with a reduced range of septa (0- or 1-septate). It furthermore differs from *Fu. meniscoidea* by its shorter and doliiform conidiogenous cells.

Fusicolla sporellula Sand.-Den. & L. Lombard, **sp. nov.** MycoBank MB 838663. [Fig. 28.](#)

Etymology: From Latin, very small spores, in reference to its very small conidia.

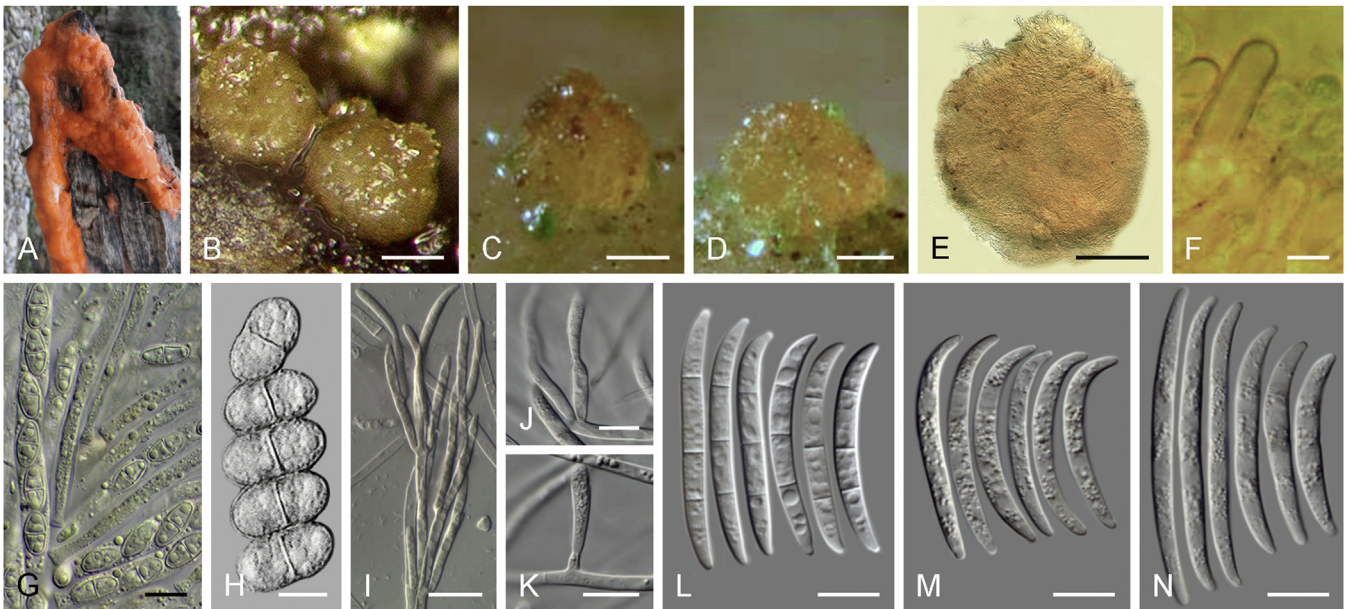


Fig. 25. *Fusicolla* spp. **A.** Slimy macroscopic growth on natural substrate. **B–E.** Ascomata on natural substrate. **F.** Ostiolar hairs. **G.** Asci. **H.** Ascospores. **I–K.** Conidiophores and conidiogenous cells. **L–N.** Macroconidia. **A.** *Fusicolla merismoides* (photo J. Cunningham). **B.** *Fusicolla melogrammae* [CLL 16006, adapted from Crous *et al.* (2016)]. **C–H.** *Fusicolla ossicola* (photos N. Aplin and P. Cannon). **I.** *Fusicolla merismoides* (photo P. Cannon). **J, K, M.** *Fusicolla aquaeductuum* (CBS 734.79). **L.** *Fusicolla violacea* (CPC 38810). **N.** *Fusicolla matuoi* (CBS 581.78). Scale bars: B–E = 100 μ m; F, H, 5 μ m; all others = 10 μ m.

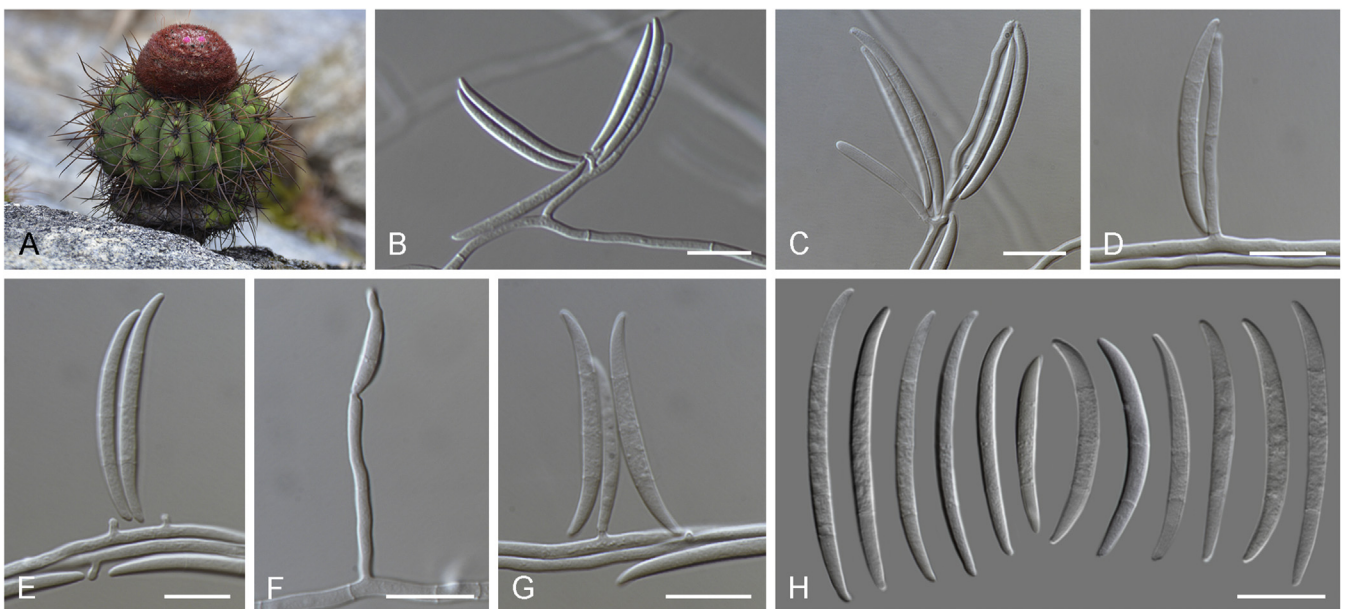


Fig. 26. *Fusicolla quarantena* (URM 8367). **A.** Host. **B–G.** Conidiophores, conidiogenous cells and conidia. **H.** Macroconidia. Scale bars = 10 μ m.

Typus: South Africa, Transkei, from soil, unknown collection date (before 1983), unknown collector (**holotype** CBS H-24663, culture ex-type CBS 110191 = FRC E-0139).

Conidiophores arising laterally from substrate and aerial hyphae 14–35 μ m long, simple or laterally and verticillately branched, straight, hyaline, smooth- and thin-walled, or reduced to single conidiogenous cells. **Conidiogenous cells** monophialidic, doliiform, short lageniform to subulate 7.5–20 \times 2.5–4 μ m, smooth- and thin-walled, with or without inconspicuous periclinal thickening, collarettes absent; or reduced to short phialidic pegs emerging laterally from hyphae, 1–5 \times 1–2.5 μ m, smooth- and thin-walled, with inconspicuous periclinal thickening and an often conspicuously flared collarette. **Macroconidia** lunate to falcate, moderately to strongly

dorsiventrally curved, slightly narrowing towards both ends, apical cell blunt, more or less hooked, basal cell obtuse to poorly developed, foot-shaped, hyaline, thin- and smooth-walled, 0- or 1-septate, predominantly 1-septate, 0-septate: (11–)12–14(–15) \times 2–3(–3.5) μ m (av. 13.2 \times 2.7 μ m), 1-septate: (11.5–)13–16.5(–20) \times 2.5–3.5 μ m (av. 14.6 \times 2.8 μ m). **Microconidia**, **chlamydospores**, and **sexual morph** not observed.

Culture characteristics: Colonies on PDA reaching 24–31 mm diam at 25 $^{\circ}$ C after 7 d. Surface white, luteous to orange, flat to slightly radially folded, membranous to slimy, margin entire; reverse pale luteous to saffron, peach at centre. On OA, pale luteous to peach, flat, membranous with filiform to undulate margins; reverse pale peach to saffron.

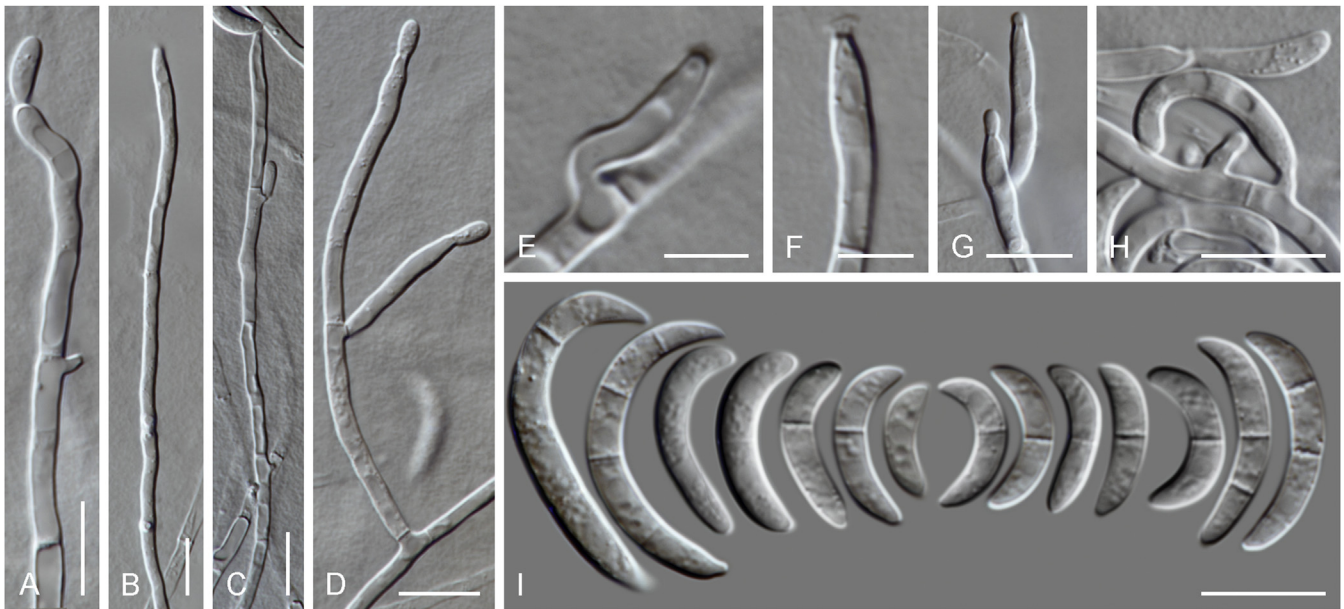


Fig. 27. *Fusicolla meniscoidea* (CBS 110189). A–D. Conidiophores. E–H. Conidiogenous cells. I. Macroconidia. Scale bars: A–D, G–I = 10 μ m; E, F = 5 μ m.

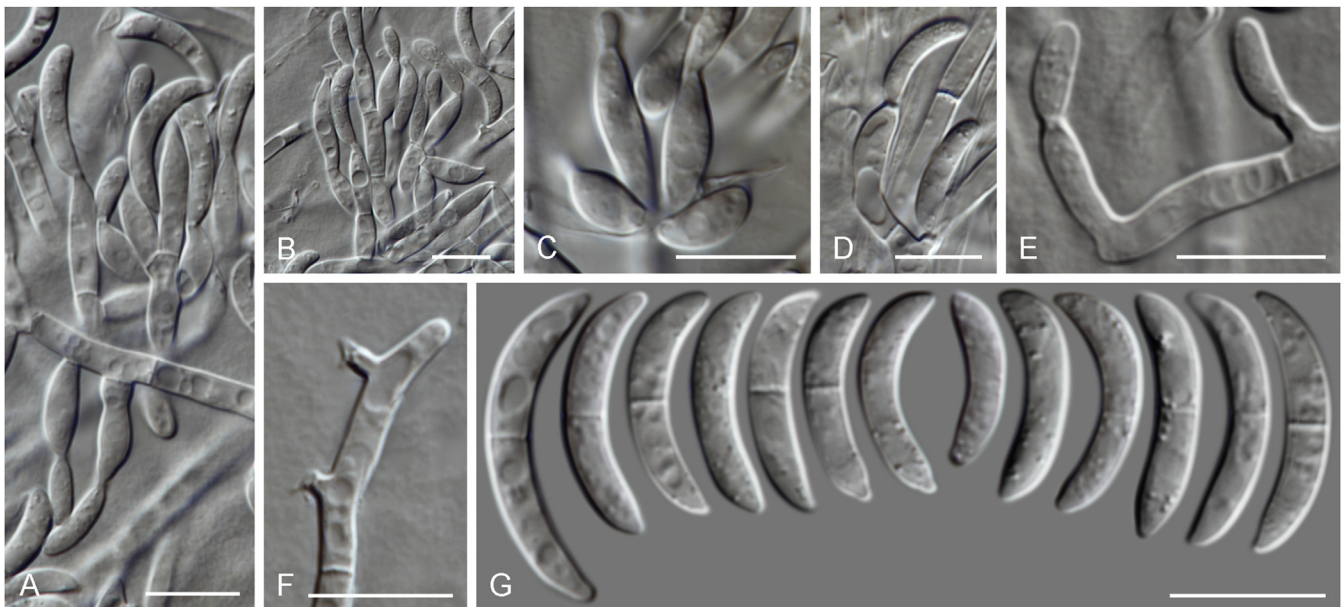


Fig. 28. *Fusicolla sporellula* (CBS 110191). A–C. Conidiophores. D–F. Conidiogenous cells. G. Macroconidia. Scale bars = 10 μ m.

Notes: *Fusicolla sporellula* presents the smallest conidia described to date for any species in this genus. This taxon is phylogenetically and morphologically related to *Fu. meniscoidea*, from which it can be differentiated by its smaller and less septate conidia, and by the characteristic doliiform shape of its conidiogenous cells.

Geejayessia Schroers *et al.*, Stud. Mycol. 68: 124. 2011. Figs 8, 29.

Type species: *Geejayessia cicatricum* (Berk.) Schroers, Stud. Mycol. 68: 124. 2011.

(See *F. cicatricum* in List section for synonyms)

Ascomata perithecial, caespitose, with erumpent, byssoid or densely prosenchymatous stroma, superficial, broadly ampulliform with short ostiolar neck to broadly ellipsoidal, not collapsing when dry, pale orange, brownish to reddish orange, bright, reddish black or black, changing colour in KOH if not black and

becoming purple in lactic acid, mostly smooth-walled, lacking hairs or appendages. **Ascomatal wall** consists of a single region, comprising several layers of morphologically similar cells. **Asci** cylindrical to clavate, with a broadly rounded or flattened apex, with or without a minute refractive ring, 8-spored, mostly overlapping, uniseriate or biseriate above and uniseriate below. **Ascospores** broadly ellipsoidal to ellipsoidal, 1-septate, slightly constricted at the septum, verrucose, hyaline to pale brown. **Conidiophores** mononematous (aerial conidiophores) or grouped on sporodochia. **Aerial conidiophores** unbranched, sympodial or irregularly branched, bearing terminal or lateral phialides, often reduced to single phialides. **Conidiogenous cells** monopialidic, subcylindrical to cylindrical, smooth- and thin-walled, with periclinal thickening inconspicuous or absent. **Aerial conidia** hyaline, smooth- and thin-walled, of two types: **microconidia**, present in some species, ellipsoidal to fusoid, 0- or 1-septate, with rounded ends, straight to slightly curved; **macroconidia** typically formed on sporodochia, falcate, straight to gently curved dorsiventrally,

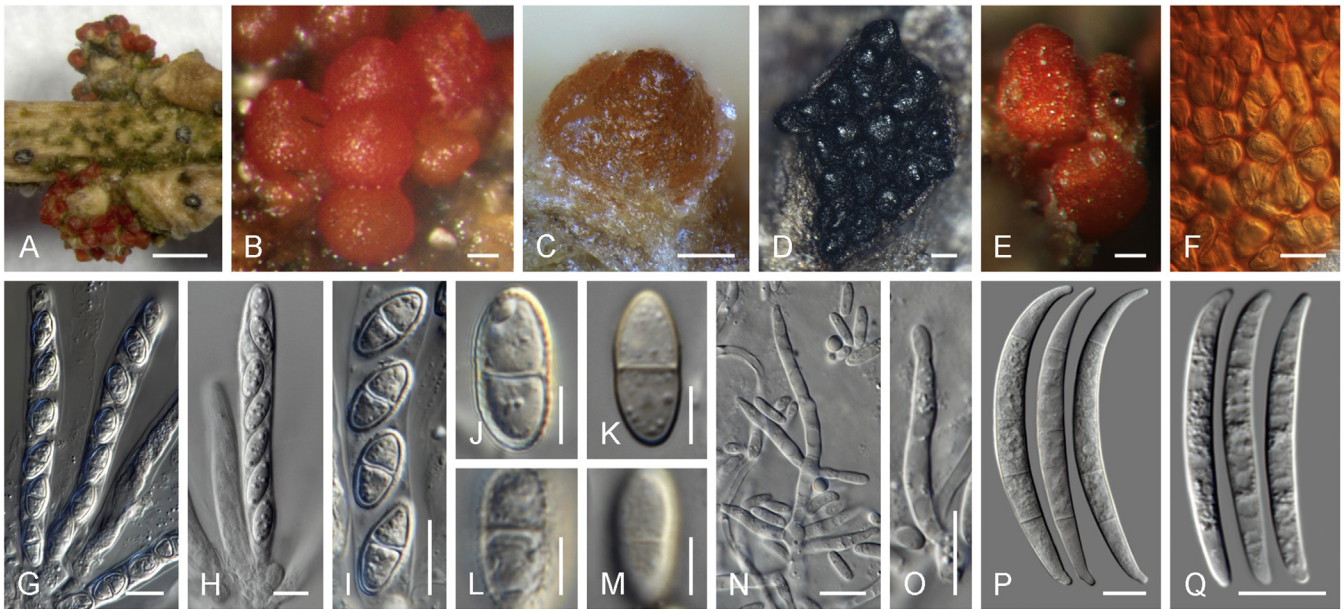


Fig. 29. *Geejayessia* spp. **A–E.** Ascomata on natural substrate. **F.** Surface view of perithecial wall in 2 % KOH. **G–I.** Asci. **J–M.** Ascospores. **N, O.** Conidiophores and conidiogenous cells. **P, Q.** Macroconidia. **A, C.** *Geejayessia cicatricum* [CBS H-20375, adapted from [Schroers et al. \(2011\)](#)]. **C.** *Geejayessia cicatricum* (CBS H-20374). **D, H, K, M.** *Geejayessia atrofusca* (CBS H-20381). **E–G, I, J, L.** *Geejayessia desmazieri* (CBS H-20372). **N, O, Q.** *Geejayessia atrofusca* (CBS 502.94). **P.** *Geejayessia cicatricum* (CBS 125549). Scale bars: A = 500 µm; B, D, E = 200 µm; C = 100 µm; J–M = 5 µm; all others = 10 µm.

3–8-septate, with a blunt apical cell and well-developed foot-shaped basal cell. *Sporodochia* cream to pale yellow; *sporodochial conidiophores* verticillately branched and densely packed, consisting of short, smooth- and thin-walled stipes bearing an apical whorl of 2–3 monophialides; *sporodochial conidiogenous cells* monophialidic, cylindrical to subcylindrical, smooth- and thin-walled, with reduced or flared collarette. *Chlamydospores* unknown.

[Description adapted from [Schroers et al. \(2011\)](#) and [Lombard et al. \(2015\)](#)].

Diagnostic features: Pale orange, brownish to reddish orange, bright red, reddish black to black, mostly smooth-walled perithecia with short ostiolar neck producing clavate to cylindrical asci bearing ellipsoidal, 1-septate, verrucose ascospores and asexual morphs producing only macroconidia on sporodochia or micro- and macroconidia on elongate subulate to subcylindrical aerial conidiophores with monophialides. *Chlamydospores* absent.

Ilyonectria P. Chaverri & C. Salgado, *Stud. Mycol.* 68: 69. 2011. [Fig. 8.](#)

Type species: *Ilyonectria destructans* (Zinssm.) Rossman et al., *Stud. Mycol.* 80: 217. 2015.

(See *F. aderholdii* in List section for synonyms)

Ascomata perithecial, solitary or gregarious, non-stromatic, superficial, globose to subglobose or ovoid to obpyriform, red, turning purple to dark purple in KOH, pigment dissolving in lactic acid, not collapsing when dry, with broadly conical papilla or flattened apex, smooth to slightly rugulose, lacking hairs or appendages. **Ascomatal wall** of two regions: outer region of thick-walled, pigmented cells forming a *textura globosa*; inner region of compressed, flattened cells, becoming thinner towards the centrum. **Asci** narrowly clavate to cylindrical, 8-spored, apex subtruncate, with inconspicuous apical ring, uniseriate. **Ascospores** ellipsoidal, 1-septate, hyaline, smooth. **Conidiophores** simple or complex or sporodochial; *simple conidiophores* arising laterally or terminally from aerial mycelium, solitary or loosely

aggregated, unbranched or sparsely branched, bearing up to three phialides; *complex conidiophores* solitary or aggregated in small sporodochia, repeatedly and irregularly branched. *Conidiogenous cells* monophialidic, cylindrical, tapering towards the apex. *Microconidia* 0- or 1-septate, ovoid to fusoid to ellipsoidal, with a minutely or clearly laterally displaced hilum, formed in heads on solitary conidiophores or as masses on sporodochia. *Macroconidia* straight, cylindrical, 1–3(–4)-septate, with both ends obtusely rounded, base sometimes with a visible, centrally located to laterally displaced hilum, forming flat domes of slimy masses. *Chlamydospores* globose to subglobose, thick-walled, intercalary or solitary, initially hyaline, becoming brown with age. [Description adapted from [Chaverri et al. \(2011\)](#)].

Diagnostic features: Red, mostly smooth-walled perithecia with conical papilla or flattened apex producing cylindrical asci bearing ellipsoidal, 1-septate ascospores and cylindrocarpon-like asexual morph characterised by 1–3(–4)-septate macroconidia with centrally located to laterally displaced hilum.

Luteonectria Sand.-Den., L. Lombard, Schroers & Rossman, **gen. nov.** MycoBank MB 838664. [Figs 8, 30.](#)

Etymology: Name refers to the luteous coloured, nectria-like ascomata characteristic of these fungi.

Type species: *Luteonectria albida* (Rossman) Sand.-Den. & L. Lombard

Ascomata perithecial, gregarious on a well-developed stroma composed of pseudoparenchymatous cells, covered with loose, white hyphae, smooth and thin-walled, globose to pyriform, off-white to pale luteous, becoming ochraceous when dry, with a broadly rounded and papillate apical region, not changing colour in KOH or lactic acid, short setae-like hairs sometimes emerging from perithecial wall. **Asci** clavate with simple apex, 8-spored, ascospores overlapping irregularly uniseriate to biseriate. **Ascospores** fusiform with rounded ends, 3-septate, slightly constricted at septum, hyaline, becoming pale yellow-brown, smooth-walled to finely striate. **Conidiophores** mononematous,

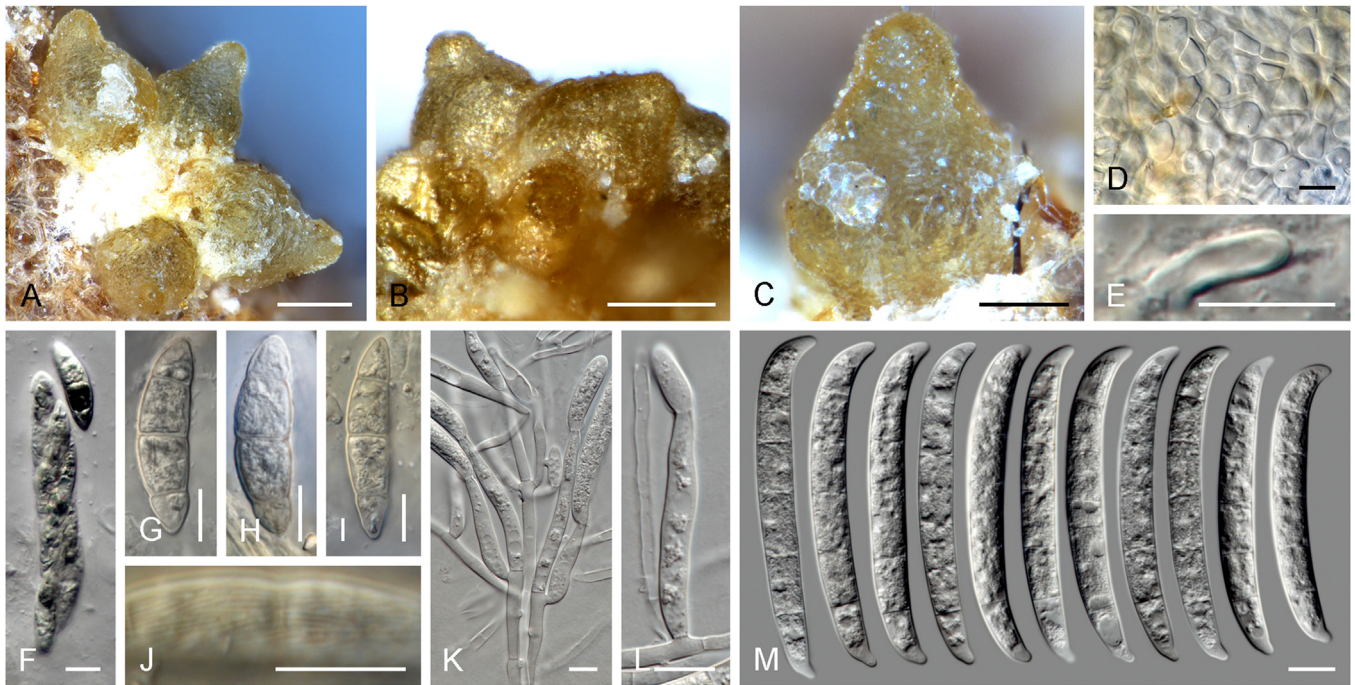


Fig. 30. *Luteonectria albida*. **A–C.** Ascomata on natural substrate. **D.** Surface view of perithecial wall in lactic acid. **E.** Detail of ascomata hair. **F.** Asci. **G–J.** Ascospores (J. Surface view). **K, L.** Conidiophores and conidiogenous cells. **M.** Macroconidia. A, C. BPI 550103. B. BPI 1108874. D–J. BPI 1108875. K–M. CBS 102683. Scale bars: A, B = 100 μ m; C = 50 μ m; all others = 10 μ m.

septate and irregularly branched, bearing terminal phialides. *Conidiogenous cells* monophialidic, cylindrical to subcylindrical, smooth- and thin-walled, with periclinal thickening inconspicuous to absent. *Macroconidia* fusoid and multiseptate, 1–7-septate, curved, hyaline, with a wide, blunt apical cell and a poorly- to well-developed, foot-shaped basal cell. *Micro- and mesoconidia* unknown. *Chlamydospores* unknown.

[Description adapted from Rossman (1983) and Schroers *et al.* (2011)].

Diagnostic features: Off-white to pale luteous perithecia that do not change colour on KOH or lactic acid, formed on well-developed stroma producing clavate asci containing fusiform, 3-septate, finely striate ascospores and fusarioid asexual morph characterised by monophialides producing robust multiseptate conidia from aerial conidiophores, lacking micro- and mesoconidia, and chlamydospores.

Luteonectria albida (Rossman) Sand.-Den. & L. Lombard, **comb. nov.** MycoBank MB 838665.

Basionym: *Nectria albida* Rossman, Mycol. Pap. 150: 79. 1983. **Synonyms:** *Albonectria albida* (Rossman) Guu & Y.M. Ju, Bot. Stud. (Taipei) 48: 189. 2007.

Fusarium albidum (Rossman) O'Donnell & Geiser, Phytopathology 103: 404. 2013.

Typus: **Jamaica**, Hanover Parish, Dolphin Head Mt. near Askenish, on bark of woody stem of unknown host, 22 Jan. 1971, R.P. Korf *et al.* (**holotype** CUP-MJ 942, culture ex-type ATCC 44543 = CTR 71-110 = BBA 67603 = NRRL 13950 = NRRL 22152).

Description and illustration: Rossman (1983), Guu *et al.* (2007), Schroers *et al.* (2011).

Additional material examined: **Costa Rica**, Limón, Central Distrito Valle, Valle del Estrella, Selva Biología Hitoi Caneri, 100–150 m alt, on bark of living tree, 7 Jul.

1999, G.J. Samuels *et al.*, BPI 746587, culture CBS 102683. **Jamaica**, Newcastle, Chesterville Youth Developmental Camp, on undetermined host, 8 Jan. 1971, A.Y. Rossman, BPI 550103. **Venezuela**, Los Venados, El Avila, along Trail 1–2 km above Los Venados, El Avila, on undetermined substrate, 24 Jul. 1972, K.P. Dumont *et al.*, BPI 1108875.

Luteonectria nematophila (Nirenberg & Hagedorn) Sand.-Den. & L. Lombard, **comb. nov.** MycoBank MB 838666.

Basionym: *Fusarium nematophilum* Nirenberg & Hagedorn, Nachrichtenbl. Deutsch. Pflanzenschutzdienstes 60: 214. 2008.

Typus: **Germany**, Berlin, from soil with roots of *Hedera helix*, unknown date and collector (**holotype** BBA 72279 in B, culture ex-type BBA 72279 = NRRL 54600).

Description and illustration: Nirenberg & Hagedorn (2008).

Macroconia (Wollenw.) Gräfenhan *et al.*, Stud. Mycol. 68: 101. 2011. **Figs 8, 31.**

Basionym: *Nectria* sect. *Macroconia* Wollenw., Angew. Bot. 8: 179. 1926.

Type species: *Macroconia leptosphaeriae* (Niessl) Gräfenhan & Schroers, Stud. Mycol. 68: 102. 2011.

Synonyms: *Nectria leptosphaeriae* Niessl, in Krieger, Fungi Saxon. Exs.: no. 165. 1886.

Cucurbitaria leptosphaeriae (Niessl) Kuntze, Revis. Gen. Pl. 3: 461. 1898.

Hypomyces leptosphaeriae (Niessl) Wollenw., Fusaria Autogr. Delin. 1: 57. 1916.

Lasionectria leptosphaeriae (Niessl) Petch, Trans. Brit. Mycol. Soc. 21: 268. 1938.

Cosmospora leptosphaeriae (Niessl) Rossman & Samuels, Stud. Mycol. 42: 122. 1999.

Ascomata perithecial, solitary, with stroma inconspicuous or absent, subglobose with or without a small apical papilla, orange to carmine red, turning dark red to violet in KOH, sometimes with

hyphal hairs arising from the outer wall. *Asci* cylindrical to narrowly clavate, with a simple apex, 8-spored, uniseriate or partially biseriata. *Ascospores* yellowish, 1-septate, smooth, sometimes becoming striate when mature. *Conidiophores* initially as lateral phialides on somatic hyphae, later monochasial to verticillate, hyaline. *Conidiogenous cells* monophialidic, cylindrical to subulate, hyaline. *Microconidia* rare or absent, ellipsoidal to allantoid, hyaline. *Macroconidia* subcylindrical to curved, apical cell conical or hooked, basal cell poorly- to well-developed, foot-shaped, 3–7(–14)-septate, hyaline. *Chlamydoconidia* absent to rare, globose, single, in pairs or chains in hyphae.

[Description adapted from Gräfenhan *et al.* (2011)].

Diagnostic features: Orange-red to carmine-red perithecia with or without a small papilla producing cylindrical to narrowly clavate asci bearing 1-septate ascospores that sometimes become striate when mature, and asexual morphs characterised by verticillate conidiophores producing large, multiseptate fusarioid macroconidia.

Macroconia bulbipes Crous & Sand.-Den., *sp. nov.* MycoBank MB 838667. Fig. 32.

Etymology: Named after the shape of the basal cell, which is commonly swollen, bulbous.

Typus: South Africa, Western Cape Province, Swellendam, Bontebok National Park, from *Erica* sp. associated with *Dimerosporiopsis engleriana*, 24 Sep. 2018, A.R. Wood (**holotype** CBS H-24664, culture ex-type CBS 146679 = CPC 37138).

Conidiophores commonly aggregated into sporodochia, more rarely simple (aerial). *Aerial conidiophores* borne laterally on hyphae and commonly reduced to single conidiogenous cells, hyaline, thin- and smooth-walled, 23.5–39.6 µm long; *conidiogenous cells* monophialidic, subcylindrical to cylindrical, hyaline, (23–) 24–25(–26.5) × 3–4 µm, without discernible periclinal thickening or collarettes. *Sporodochia* abundantly formed on carnation leaves and on the agar surface, pink to pink-brown coloured. *Sporodochia*

light orange-peach, turning dark brick coloured in old cultures; *sporodochial conidiophores* irregularly or verticillately branched, 40–55.5 µm long, irregularly branched, bearing lateral and terminal solitary monophialides. *Sporodochial conidiogenous cells* monophialidic, cylindrical to subcylindrical to subulate, (8–) 14.5–26.5(–30.5) × 3.5–5.5 µm with inconspicuous periclinal thickening, flared collarettes absent. *Microconidia* absent. *Macroconidia* straight to moderately dorsiventrally curved, tapering toward the apex, apical cell conical or hooked, and slightly extended, basal cell well-developed, foot shaped, commonly irregularly swollen at bottom, (2–)3–5(–6)-septate, predominantly 4-septate, hyaline, thick- and smooth-walled: 2-septate conidia: 43–45.5 × 5–5.5 µm (av. 44.2 × 5.1 µm); 3-septate conidia: (38.5–) 41–53(–55) × 5–6 µm (av. 48.1 × 5.4 µm); 4-septate conidia: (45.5–)50–62(–67.5) × 5–6(–7) µm (av. 56.1 × 5.8 µm); 5-septate conidia: (58–)61–77(–80.5) × 5–6.5 µm (av. 68.9 × 5.8 µm); 6-septate conidia: (70–)71–74 × 5.5–6.5(–7) µm (av. 72.1 × 6.4 µm); overall: (38.5–)48–68(–80.5) × 5–6(–7) µm (av. 58 × 5.7 µm). *Chlamydoconidia* commonly formed in the substrate mycelium and conidia, spherical to subspherical, 8.5–11(–12.5) µm diam, hyaline and smooth-walled. *Sexual morph* not observed.

Culture characteristics: Colonies on PDA reaching 21–24 mm diam at 25 °C after 7 d. Surface salmon to buff, flat, membranous to velvety, with scant aerial mycelium and pionnotal, margin white and regular; reverse pale salmon with radial white to pale yellow patches. On OA, salmon to buff, flat, membranous and pionnotal, with regular margin; reverse pale pink to salmon.

Additional material examined: South Africa, Western Cape Province, Swellendam, Bontebok National Park, from *Erica* sp. associated with *Dimerosporiopsis engleriana*, 24 Sep. 2018, A.R. Wood, culture CBS 146678 = CPC 37137.

Notes: *Macroconia bulbipes* resolved as the closest phylogenetic relative to *Ma. gigas* and *Ma. papilionacearum* (Fig. 13). The former is, however, clearly distinguished morphologically by its

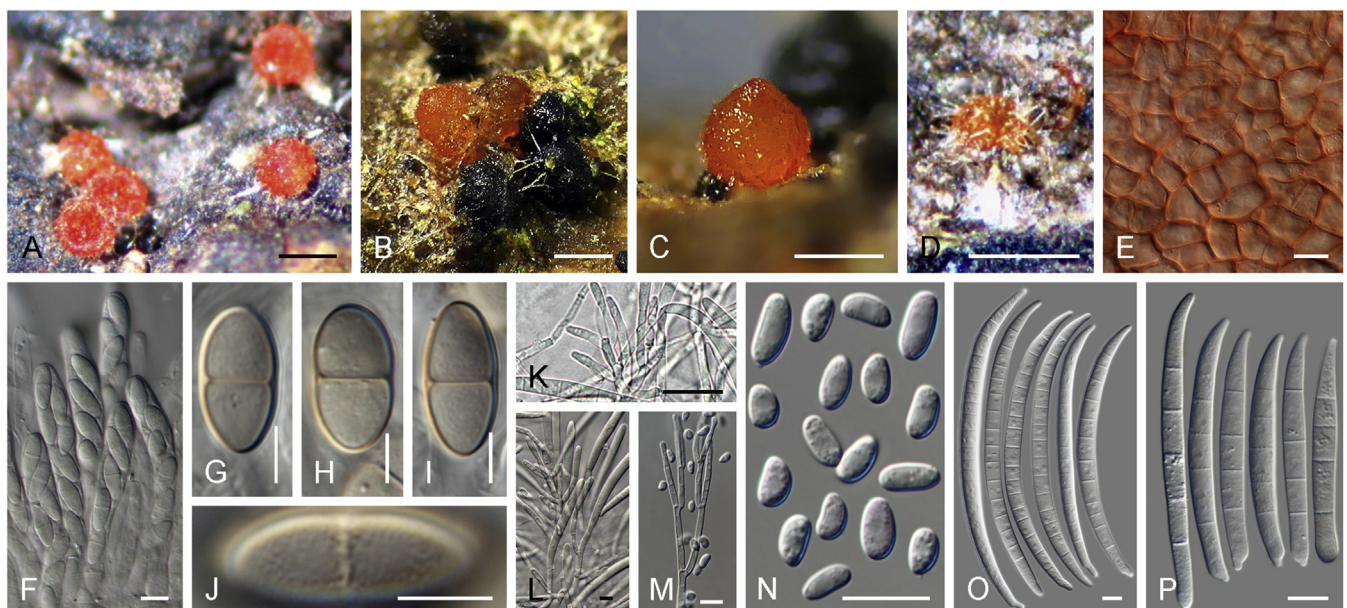


Fig. 31. *Macroconia* spp. **A–D.** Ascomata on natural substrate. **E.** Surface view of perithecial wall in 2% KOH. **F.** Asci. **G–J.** Ascospores (J. Surface view). **K–M.** Conidiophores and conidiogenous cells. **N.** Microconidia. **O, P.** Macroconidia. **A.** *Macroconia cupularis* [HMAS 97514, adapted from Luo & Zhuang (2008)]. **B, C.** *Macroconia leptosphaeriae* (photo P. Mičoch). **D.** *Macroconia gigas* [HMAS 99592, adapted from Luo & Zhuang (2008)]. **E–J.** *Macroconia leptosphaeriae* (CBS H-15051). **K, L.** *Macroconia phlogioides* (CBS 125496). **M, N.** *Macroconia leptosphaeriae* (CBS 10001). **O.** *Macroconia phlogioides* (CBS 146500). **P.** *Macroconia bulbipes* (CBS 146679). Scale bars: A–D = 100 µm; G–J = 5 µm; all others = 10 µm.

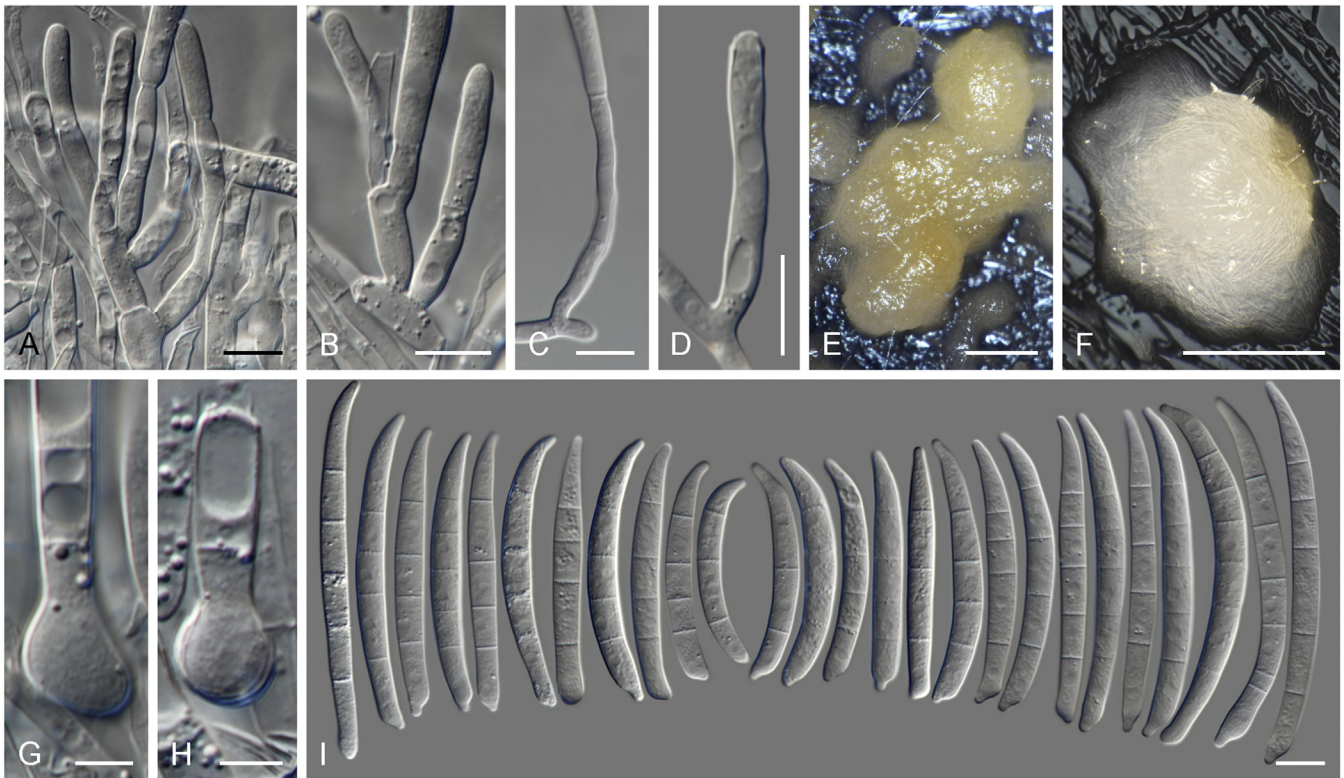


Fig. 32. *Macroconia bulbipes* (CBS 146679). **A–D.** Conidiophores and conidiogenous cells. **E, F.** Sporodochia formed on the agar surface. **G, H.** Detail of macroconidia basal cells. **I.** Macroconidia. Scale bars: E, F = 100 μ m; G, H = 5 μ m; all others = 10 μ m.

smaller and less septate conidia (rarely up to 80.5 μ m long and up to 6-septate vs longer than 100 μ m and more than 10-septate in the latter two species). On the contrary, the asexual morph of *Ma. bulbipes* is closer to that of *Ma. leptosphaeriae* and *Ma. sphaeriae* (recognised as two distinct species in Gräfenhan et al. 2011). The conidia of *Ma. bulbipes*, however, differ by having commonly swollen basal cells.

Macroconia phlogioides Sand.-Den. & Crous, *sp. nov.* MycoBank MB 838668. Fig. 33.

Etymology: From Greek *flōga*. Referring to the flame-like macroscopic semblance of the sporodochia.

Typus: South Africa, Limpopo Province, Tzaneen, on leaf of *Encephalartos* sp., 2019, P.W. Crous (**holotype** CBS H-24665, culture ex-type CBS 146501 = CPC 35389).

Conidiophores simple (aerial) or aggregated into sporodochia. **Aerial conidiophores** often borne laterally on hyphae and reduced to single conidiogenous cells, rarely 1-septate, hyaline, thin- and smooth-walled, 13–17 \times 26–32 μ m; **conidiogenous cells** monophialidic, subcylindrical to cylindrical, hyaline, (13–)16–24(–27.5) \times (3.5–)4–5 μ m conidiogenous opening rather wide, with inconspicuous periclinal thickening and no discernible apical collarettes. **Sporodochia** orange-pink to pink-brown coloured, often acquiring a flame-like, somewhat pointy macroscopic appearance and later merging into pionnotal crusts; **sporodochial conidiophores** irregularly or verticillately branched, 37.5–46 μ m long, often bearing groups of 2–3 conidiogenous cells; **sporodochial conidiogenous cells** monophialidic, subcylindrical to subulate, (10–)18.5–26(–30) \times (2.5–)3.5–5 μ m with inconspicuous periclinal thickening, collarettes absent. **Microconidia** absent. **Macroconidia** robust, often with a nearly

straight central portion and markedly curved and tapering towards both ends, apical cell conical to hooked, basal cell well-developed, foot-shaped, (1–)9–15(–19)-septate, predominantly 11-septate, hyaline, thick- and smooth-walled: 9-septate conidia: (106.5–)119.5–140(–143.5) \times 7.5–8.5(–9) μ m (av. 129 \times 8 μ m); 10-septate conidia: (116–)120–144.5(–164) \times (7–)7.5–9 μ m (av. 132 \times 8 μ m); 11-septate conidia: (122–)127–140(–153.5) \times 7.5–9(–9.5) μ m (av. 134 \times 8.5 μ m); 12-septate conidia: (119–)127.5–146.5(–153) \times 7.5–9.5(–10) μ m (av. 137 \times 8.5 μ m); 13-septate conidia: (128–)132–155(–172) \times (7–)8–9(–10) μ m (av. 143.5 \times 8.5 μ m); 14-septate conidia: (133.5–)136–157(–168) \times 8–9.5 μ m (av. 146.5 \times 9 μ m); 15-septate conidia: 147–163.5(–173.5) \times 8.5–9.5(–10) μ m (av. 155 \times 9 μ m); overall: (86–)123.5–150(–175) \times (7–)8–9(–10) μ m (av. 137 \times 8.5 μ m). **Chlamydospores** and **sexual morph** not observed.

Culture characteristics: Colonies on PDA reaching 17–25 mm diam at 25 °C after 7 d. Surface salmon, buff to rosy buff, flat to slightly raised at centre, glabrous or with central patches of white, dense aerial mycelium; membranous to dusty with regular margin; reverse pale luteous to sulphur yellow, with salmon patches. On OA, salmon, flat, membranous, inconspicuously radially folded with regular margin; reverse pale pink to luteous with more intense salmon-coloured patches.

Additional material examined: South Africa, Limpopo Province, Tzaneen, on leaf of *Encephalartos* sp., 2019, P.W. Crous, culture CBS 146500 = CPC 35388. **USA,** Arizona, Huachuca Mountains, Miller Canyon, on branch of *Quercus* sp. in stream, 1 Oct. 2008, T. Gräfenhan, culture CBS 125496.

Notes: *Macroconia phlogioides* is morphologically related to *Ma. papilionacearum* and *Ma. gigas*. These three species are characterised by producing robust and large (often above 100 μ m

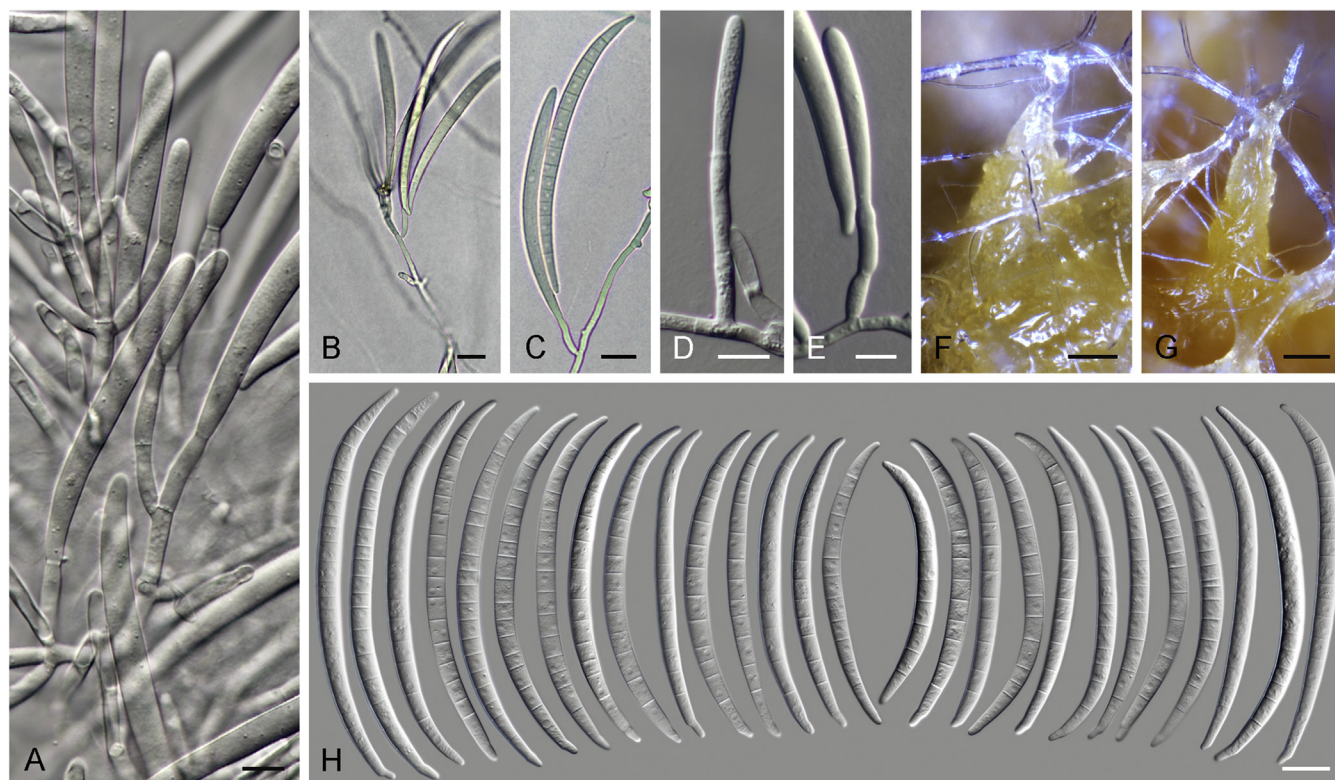


Fig. 33. *Macroconia phlogioides* (CBS 146501). **A–C.** Conidiophores. **D, E.** Conidiogenous cells. **F, G.** Sporodochia formed on the agar surface. **H.** Macroconidia. Scale bars: B, C = 20 μ m; F, G = 50 μ m; all others = 10 μ m.

long) macroconidia. Unlike the above-mentioned species, however, conidia of *Ma. phlogioides* tend to present a higher number of septa (up to 19 vs up to 12 and 14, for *Ma. papilionacearum* and *Ma. gigas*, respectively), with rounder and less tapered apical cells, contrasting with the elongated conidial apices of *Ma. gigas*. Conidia of *Ma. phlogioides* also differ by having a more pronounced and continuous curvature compared to *Ma. gigas* and *Ma. papilionacearum*. These three species are clearly different phylogenetically, clustering in distant monophyletic lineages of the genus (Fig. 13).

Mariannaea G. Arnaud ex Samson, Stud. Mycol. 6: 74. 1974. Fig. 8.

Type species: Mariannaea elegans (Corda) Samson, Stud. Mycol. 6: 75. 1974.

Basionym: Penicillium elegans Corda, Icon. Fung. 2: 17. 1838.

Synonyms: Hormodendron elegans (Corda) Bonorden, Handb. Allg. Mykol.: 76. 1851.

Spicaria elegans (Corda) Harz., Bull. Soc. Imp. Naturalistes Moscou 44: 238. 1871.

Paecilomyces elegans (Corda) Mason & Hughes, Mycol. Pap. 45: 27. 1951.

Ascomata perithecial, solitary, non-stromatic or on inconspicuous stroma, superficial, globose with flat apex, not collapsing or laterally pinched when dry, pale yellow, orange or brown, not reacting in KOH, smooth-walled to slightly rugose, lacking hairs or appendages. *Asci* cylindrical to narrowly clavate, 8-spored sometimes with inconspicuous apical ring, uniseriate to apically biseriate. *Ascospores* 1-septate, hyaline, smooth-walled to spinulose. *Conidiophores* verticillate to penicillate, hyaline, with phialides arising directly from the stipe or forming whorls of metulae on lower parts of stipe; stipe hyaline, becoming yellow-

brown at the base. *Conidiogenous cells* monophialidic, ampulliform, hyaline, usually with obvious periclinal thickening and inconspicuous collarettes. *Conidia* limoniform, aseptate, hyaline, in chains that collapse to form slimy heads. *Chlamydospores* globose to ellipsoidal, hyaline, formed in intercalary chains. [Description adapted from Samson (1974), Gräfenhan *et al.* (2011) and Lombard *et al.* (2015)].

Diagnostic features: Pale yellow, orange to brown perithecia with flattened apex producing cylindrical to narrowly clavate asci bearing 1-septate ascospores and asexual morphs characterised by verticillate to penicillate conidiophores producing small, aseptate, limoniform conidia in chains that collapse into slimy heads.

Microcera Desm., Ann. Sci. Nat. Bot., sér. 3, 10: 359. 1848. Figs 8, 34.

Synonym: Pseudomicrocera Petch, Trans. Brit. Mycol. Soc. 7: 164. 1921.

Type species: Microcera coccophila Desm., Ann. Sci. Nat. Bot., sér. 3, 10: 359. 1848.

(See *F. coccophilum* in List section for synonyms)

Ascomata perithecial, solitary or gregarious, with stroma and/or byssus covering host, globose, with a blunt papilla, orange to dark red, turning dark red or violet in KOH, finely roughened. *Asci* cylindrical to narrowly clavate, with an apical ring, 8-spored. *Ascospores* hyaline to pale yellow-brown, 1(–3)-septate, smooth, sometimes becoming tuberculate when mature. *Conidiophores* as lateral phialides on somatic hyphae, becoming monochasial, verticillate to penicillate, hyaline, forming discrete sporodochia or synnemata on the host. *Conidiogenous cells* monophialidic, cylindrical to subulate to subclavate, hyaline. *Macroconidia* pale, orange, pink or bright red in mass, sub-cylindrical, moderately or conspicuously curved, apical cell often

slightly or conspicuously hooked, basal cell papillate to well-developed, foot-shaped, (0–)3–5(–12)-septate, hyaline. [Description adapted from Gräfenhan *et al.* (2011)].

Diagnostic features: Orange to dark red perithecia with a blunt papilla producing cylindrical to narrowly clavate asci bearing yellow-brown, 1(–3)-septate ascospores; asexual morphs characterised by verticillate to penicillate conidiophores producing small macroconidia; species typically associated with scale insects.

Neocosmospora E.F. Sm., Bull. U.S.D.A. 17: 45. 1899. Figs 8, 35.

Type species: *Neocosmospora vasinfesta* E.F. Sm., Bull. U.S.D.A. 17: 45. 1899.

(See *F. neocosmosporiellum* in List section for synonyms)

Ascomata perithecial, solitary or gregarious, non-stromatic or with reduced basal stroma, superficial, globose to pyriform, not collapsing when dry, orange-brown to bright red, darkening or becoming purple in KOH, papillate or with short ostiolar neck, commonly tuberculate, rarely smooth-walled, lacking hairs or appendages. **Ascomatal wall** of two regions: outer region of thick-walled, pigmented cells forming a *textura angularis*; inner region of elongate, hyaline, thin-walled cells, becoming thinner towards the centrum. **Asci** saccate, clavate to cylindrical, unitunicate, apex simple, rounded or flattened, 8-spored, uniseriate to irregularly biseriate. **Ascospores** globose to ellipsoidal, with or without slightly truncate ends, typically 1-septate, hyaline when young becoming yellow golden-brown at maturity, thick-walled, longitudinally striate; ascospores in some species 0-septate, cerebriform or spinulose. **Conidiophores** mononematous (aerial) or grouped on sporodochia, or somewhat erect, loosely branched sporodochial pustules. **Aerial conidiophores** simple, sparsely to highly branched; aerial **conidiogenous cells** monophialidic, elongate subulate to subcylindrical. **Aerial conidia**

hyaline, smooth- and thick-walled, of two types: *microconidia* subglobose, ellipsoidal to somewhat clavate, 0–2(–4)-septate, borne in false heads on phialides; *macroconidia* falcate, slightly to strongly curved dorsiventrally, 1-septate to multiseptate, with blunt to hooked to slightly pointed apical cell and papillate to well-developed foot-shaped basal cell. **Sporodochia** cream, pale luteous, light green, olivaceous, bluish, hazel to greyish sepia; **sporodochial conidiophores** verticillately or sympodially branched or sparingly branched and densely packed, consisting of short, smooth- and thin-walled stipes bearing apical whorl of 2–4 monophialides; **sporodochial conidiogenous cells** monophialidic, doliiform, short subcylindrical to subulate, smooth- and thin-walled, periclinal thickening and collarettes inconspicuous or absent. **Sporodochial macroconidia** falcate, smooth- and thick-walled, straight or curved with parallel walls to unequally curved, in some species clavate and asymmetrical, tapering towards both ends, with a pointed to blunt to hooked apical cell and papillate to well-developed foot-shaped basal cell. **Chlamydo-spores** globose to subglobose to ovoid to obovoid, hyaline to pale golden brown, smooth-walled to slightly verrucose, terminal or intercalary, solitary or in pairs or forming chains or aggregating in some species to form buff, olive aeruginous or bluish microsclerotia.

[Description adapted from Rossman *et al.* (1999) and Sandoval-Denis *et al.* (2019)].

Diagnostic features: Orange-brown to frequently bright, blood red warted perithecia with papillate or short ostiolar neck producing saccate, clavate to cylindrical asci bearing globose to ellipsoidal, 0- or 1-septate, longitudinally striate, cerebriform or spinulose ascospores and asexual morphs producing micro- and macroconidia on elongate subulate to subcylindrical aerial conidiophores with monophialides or only macroconidia in sporodochia. Chlamydo-spores formed in hyphae, rarely observed in macroconidia.

Neocosmospora epipeda Quaedvl. & Sand.-Den., *sp. nov.* MycoBank MB 838669. Fig. 36.

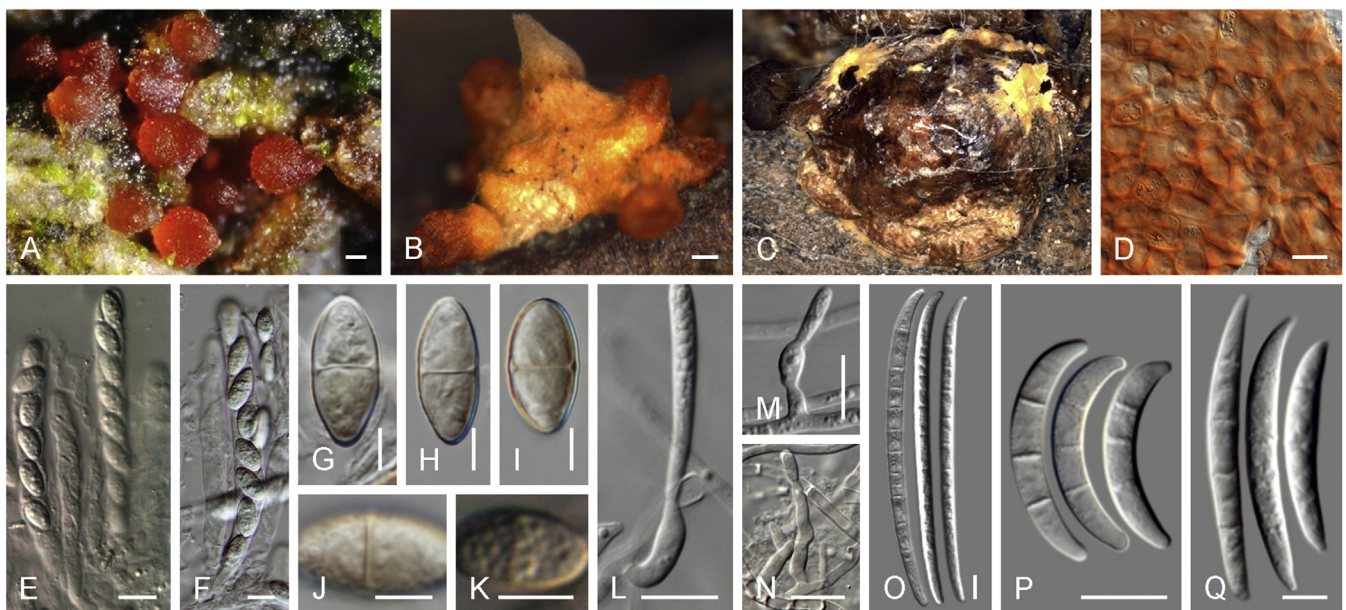


Fig. 34. *Microcera* spp. **A–C.** Ascomata on natural substrate. **D.** Surface view of perithecial wall in 2% KOH. **E, F.** Asci. **G–K.** Ascospores (**J, K.** Surface view). **L–N.** Conidiophores and conidiogenous cells. **O–Q.** Macroconidia. **A.** *Microcera auranticola* (photo N. Aplin). **B, O.** *Microcera coccophila* [adapted from Gräfenhan *et al.* (2011)]. **C.** *Microcera larvarum* [adapted from Gräfenhan *et al.* (2011)]. **D, F–J.** *Microcera coccophila* (K(M) 165807). **E, K.** *Microcera larvarum* (photo P. Cannon). **L, M, Q.** *Microcera rubra* (CBS 638.76). **N, P.** *Microcera larvarum* (CBS 169.30). Scale bars: **A, B** = 100 µm; **G–K** = 5 µm; all others = 10 µm.

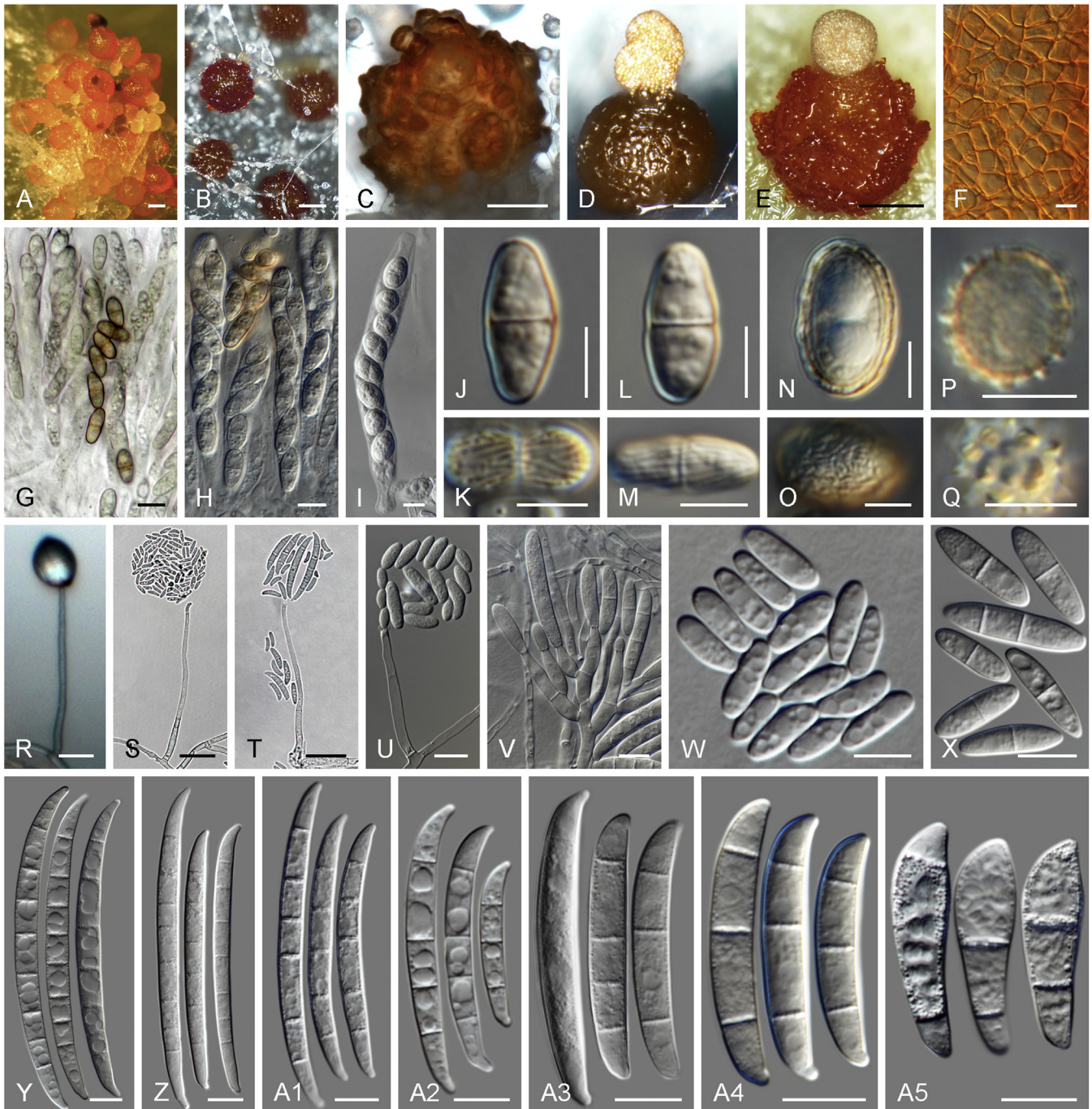


Fig. 35. *Neocosmospora* spp. **A–E.** Ascomata on culture. **F.** Surface view of perithecial wall in 2% KOH. **G–I.** Asci. **J–Q.** Ascospores (K, M, O, Q. Surface view). **R–U.** Aerial conidiophores. **V.** Sporodochial conidiophores. **W, X.** Microconidia. **Y–A5.** Macroconidia. A, I, N, O. *Neocosmospora vasinfecta* (CBS 446.93). B. *Neocosmospora* sp. (CPC 34617). C, S, W, A1. *Neocosmospora elegans* (CBS 144396). D. *Neocosmospora vasinfecta* (CBS 863.70). E. *Neocosmospora bataticola* (CBS 144398). F, L, M. *Neocosmospora ipomoeae* (CBS 833.97). G. *Neocosmospora robiniae* (CBS 119601). H, J, K. *Neocosmospora diminuta* (CBS 144390). O, Q. *Neocosmospora spinulosa* (CBS H-5443). R, V, A3. *Neocosmospora solani* (CBS 140079). T. *Neocosmospora bataticola* (CBS 144398). U. *Neocosmospora suttoniana* (CBS 143214). X. *Neocosmospora tonkinensis* (CBS 115.40). Y. *Neocosmospora longissima* (CBS 126407). Z. *Neocosmospora mori* (CBS 145467). A2. *Neocosmospora pseudoradicicola* (CBS 145472). A4. *Neocosmospora keratoplastica* (CBS 490.63). A5. *Neocosmospora oligoseptata* (CBS 143241). [A, C, S, T, W, Y, Z, A1, A2. Adapted from adapted from [Sandoval-Denis et al. \(2019\)](#). R, V, A3. Adapted from [Crous et al. \(2019a\)](#). U, X, A4. Adapted from [Sandoval-Denis & Crous \(2018\)](#)]. Scale bars: A, B = 200 μ m; C–E 100 μ m; R–T = 20 μ m; J–Q, W, X = 5 μ m; all others = 10 μ m.

Etymology: From the Greek *επιπέδα*, flat; referring to the microconidia of this species commonly being flattened on one side.

Typus: Netherlands, from *Bouvardia* sp. imported from Uganda, 2019, W. Quaedvlieg (**holotype** CBS H-24666, culture ex-type CBS 146523 = CPC 38310).

Conidiophores borne on the agar substrate and aerial mycelium, 78–230 μ m tall, unbranched or more commonly sympodially branched at various levels, bearing terminal single phialides;

aerial *conidiogenous cells* monophialidic, subulate, subcylindrical to acicular, smooth- and thin-walled, 27.5–62 \times 2–3.5 μ m, short apical collarettes and periclinal thickening inconspicuous or absent. **Aerial conidia** microconidial, arranged in false heads on phialide tips, hyaline, broadly ellipsoidal, ellipsoidal to short clavate, commonly asymmetrical with a somewhat flattened side, smooth- and thin-walled, aseptate, (4.5–)6–10(–13.5) \times (2–)3–5 μ m (av. 8 \times 3.5 μ m). **Sporodochia** pale luteous to orange, formed abundantly on the surface of carnation leaves;

sporodochial conidiophores laterally and irregularly branched bearing apical groups of 2–3 monophialides; *sporodochial conidiogenous cells* monophialidic, subulate to subcylindrical, 11–19.5 × 3–4.5 µm, smooth and thin-walled, with short, non-flared collarettes and inconspicuous or absent periclinal thickening. *Sporodochial conidia* falcate, almost straight to slightly curved dorsoventrally, broadest near the half portion or the upper third, tapering towards both ends, with a blunt to somewhat pointy and slightly curved apical cell and an often well-developed foot-shaped basal cell, (3–)4–7(–8)-septate, predominantly 5-septate, hyaline, smooth- and thick-walled; 3-septate conidia: 42.5 × 4.4 µm; 4-septate conidia: (41.5–)44–58(–60) × 4–5 µm (av. 51.1 × 4.4 µm); 5-septate conidia: (53.5–)59–69.5(–76) × 4–6 µm (av. 64.3 × 5 µm); 6-septate conidia: 68–75.5(–79.5) × 4.5–6 µm (av. 71.7 × 5.3 µm); 7-septate conidia: (68–)69–74.5(–77) × 5–6 µm (av. 71.7 × 5.5 µm); 8-septate conidia: 74–75.5 × 5–6 µm (av. 74.7 × 5.3 µm); overall: (42.5–)59–73.5(–79.5) × (4–)5–6 µm (av. 66.3 × 5.1 µm). *Chlamydo-spores* and *sexual morph* not observed.

Culture characteristics: Colonies on PDA reaching 38–53 mm diam at 25 °C after 7 d. Surface white to sulphur yellow with scarce pale ochreous to pale rust patches, flat to slightly raised with abundant white aerial mycelium, cottony to woolly, margin filiform; reverse pale luteous to sulphur yellow, pale apricot to pale rust at centre. On OA, pale luteous, flat, membranous with entire margin; reverse pale luteous.

Additional material examined: Netherlands, from *Bouvardia* sp. imported from Uganda, 2019, W. Quaedvlieg, culture CBS 146524 = GPC 38311.

Notes: The name *N. epipeda* is coined here for a novel phylogenetic lineage discovered on a *Bouvardia* sp. imported from Uganda. The new species clusters as the closest phylogenetic relative of *N. catenata* (Fig. 14), an opportunistic animal-pathogenic species characterised by abundant production of catenate to clustered, pigmented chlamydo-spores, and by the absence (as far as known) of macroconidia (O'Donnell *et al.* 2016, Sandoval-Denis & Crous 2018). These characters form the most notable differences with respect to *N. epipeda*. Additionally, *N. epipeda* can be differentiated from *N. catenata* by its less

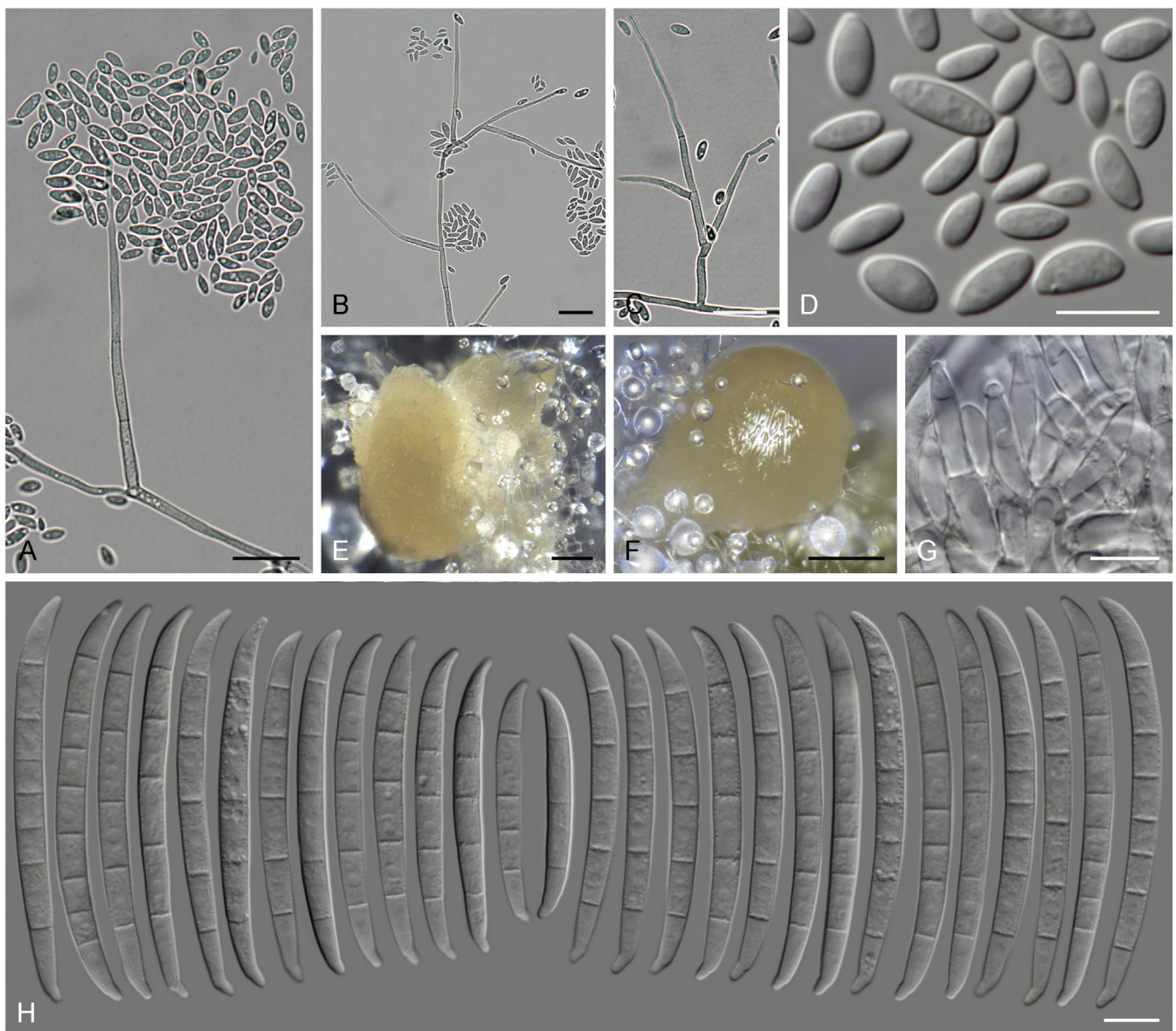


Fig. 36. *Neocosmospora epipeda* (CBS 146524). **A–C.** Aerial conidiophores and conidiogenous cells. **D.** Microconidia. **E, F.** Sporodochia formed on the surface of carnation leaves. **G.** Sporodochial conidiophores and conidiogenous cells. **H.** Macroconidia. Scale bars: A–C = 20 µm; E, F = 200 µm; D, G, H = 10 µm.

septate and shorter microconidia (aseptate and up to 13.5 µm vs up to 1-septate and 11 µm in *N. catenata*). Other species producing macroconidia of similar size and shape to those of *N. epipeda* include *N. quercicola*, *N. robusta*, and *N. silvicola*; however, the three latter species are genetically distant in that they belong to monophyletic lineages of clade 3 (*N. quercicola* and *N. silvicola*) and clade 1 (*N. robusta*) of *Neocosmospora sensu O'Donnell et al. (2008a)*. *Neocosmospora epipeda* can be distinguished morphologically from *N. robusta* by the production of microconidia with absence of aerial macroconidia in the former species. Morphological differentiation of the novel species from *N. quercicola* and *N. silvicola* is difficult because of overlapping features; nevertheless, subtle differences exist in the size and morphology of the microconidia (aseptate in *N. epipeda* vs up to 1-septate in both *N. quercicola* and *N. silvicola*, being also reniform and longer in the latter species) and sporodochial colour (pale luteous to orange in *N. epipeda* vs greenish to citrine in *N. quercicola* and *N. silvicola*, respectively).

Neocosmospora merxiana Quaedvl. & Sand.-Den., **sp. nov.**
MycoBank MB 838670. Fig. 37.

Etymology: Named after Trix Merx, senior technician at the Westerdijk Fungal Biodiversity Institute, in recognition of her career as the foremost link in strain handling between the research groups and the culture collection.

Typus: Netherlands, from *Chrysanthemum* sp. imported from Uganda, unknown date, W. Quaedvlieg (**holotype** CBS H-24669, culture ex-type CBS 146525 = CPC 38701).

Conidiophores borne on the agar substrate and aerial mycelium, 99–205 µm tall, unbranched or rarely laterally branched, bearing terminal single phialides; aerial *conidiogenous cells* monophialidic, subulate to subcylindrical, smooth- and thin-walled, 41.5–77 × 2.5–4.5 µm, with short and flared apical collarettes and inconspicuous periclinal thickening. *Aerial conidia* of two types: *microconidia* oval to broadly ellipsoidal, straight to slightly curved and asymmetrical, smooth- and thin-walled, 0(–1)-aseptate, (8.5–)9–15.5(–18.5) × 3–5.5 µm (av. 12.4 × 4.3 µm), arranged in false heads on phialide tips; *macroconidia* falcate to navicular, smooth- and thin-walled, almost straight to slightly dorsiventrally curved, ventral face almost straight, with a blunt apical cell, basal cell obtuse to poorly-developed, foot-shaped, 1–3-septate, predominantly 1-septate, 1-septate conidia: (17.5–)20.5–27(–30.5) × (4.5–)5–6.5(–7.5) µm (av. 23.8 × 5.8 µm); 2-septate conidia: (25.5–)27–30(–32) × 5.5–7 µm (av. 28.4 × 6 µm); 3-septate conidia: (27–)28.5–33.5(–35.5) × 5–7.5 µm (av. 31.1 × 6.3 µm); overall: (17.5–)22–31(–35.5) × (4.5–)5–6.5(–7.5) µm (av. 26.4 × 6 µm), arranged in false heads at the tip of monophialides and produced intermixed with microconidia. *Sporodochia* pale luteous, formed on aerial and substrate mycelium, uncommon on carnation leaves. *Sporodochial conidiophores* laterally and irregularly branched bearing apical groups of 2–3 monophialides; *sporodochial conidiogenous cells* monophialidic, doliiform, subulate to subcylindrical, (13.5–)15–21.5(–27) × 2.5–5.5 µm, smooth and thin-walled, lacking apical collarettes and with inconspicuous periclinal thickening. *Sporodochial macroconidia* falcate, straight to slightly dorsiventrally curved, broadest at the half portion and tapering towards both ends, apical cell blunt and slightly curved, basal cell poorly- to well-developed, foot-shaped, (1–)3–5-septate, predominantly 4-septate, hyaline, smooth- and thick-walled; 1-septate conidia:

(23.5–)24.5–28.5 × 5–6.5 µm (av. 25.8 × 5.6 µm); 2-septate conidia: 27–29 × 5.5–6.5 µm (av. 28 × 6 µm); 3-septate conidia: (29–)35–45 × (4.5–)5–6 µm (av. 40.1 × 5.3 µm); 4-septate conidia: (41–)44.5–49.5(–51.5) × 4.5–6.5 µm (av. 47 × 5.6 µm); 5-septate conidia: (42–)45.5–51.5(–52.5) × 5–6 µm (av. 48.5 × 5.6 µm); overall: (24.5–)39–51.5(–52.5) × 4.5–6(–6.5) µm (av. 45.2 × 5.6 µm). *Chlamyospores* obovoidal, subspherical to spherical, hyaline to pale yellow brown, smooth-walled to slightly roughened, thick-walled, 5–13.5 µm, single or in chains, terminal, intercalary or produced on short lateral stipes.

Culture characteristics: Colonies on PDA reaching 45–56 mm diam at 25 °C after 7 d. Surface pale luteus to sulphur yellow, becoming buff to honey, flat with abundant aerial mycelium, cottony to woolly with entire to filiform margin; reverse luteous to buff, pale scarlet to bay at centre. On OA pale luteous to peach with sparse white cushions of aerial mycelium, flat, velvety to cottony; reverse pale luteous, peach to pale scarlet.

Additional material examined: Netherlands, from *Chrysanthemum* sp. imported from Uganda, unknown date, W. Quaedvlieg, culture CBS 146526 = CPC 38702.

Notes: *Neocosmospora merxiana* represents the phylogenetic species formerly known as “FSSC 41”, one of the few previously known clades lacking a Latin binomial, originally reported as an agent of collar rot on *Passiflora edulis f. flavicarpa* in Brazil (Cardoso 2015, Sandoval-Denis et al. 2019). Here, this species is reported causing collar and stem rot symptoms in *Chrysanthemum* imported from Uganda.

In the phylogenetic analysis (Fig. 14), *N. merxiana* resolved as the most basal taxon within a lineage containing the morphologically similar species *N. ipomoeae*, *N. martii*, and *N. noneumartii*, all characterised by producing both aerial microconidia and macroconidia, in addition to relatively long sporodochial conidia. Differing from the aforementioned species, *N. merxiana* can be differentiated by its fewer septate and shorter aerial and sporodochial macroconidia formed on pale luteous sporodochia, and its pale luteous colonies on PDA, thus contrasting with the greenish sporodochial colouration observed in both *N. ipomoeae* and *N. noneumartii*, and the red pigmentation on PDA typical of *N. martii*. Sexual morphs were not observed in the isolates studied here; however, this lineage was reported as heterothallic, and fertile perithecial ascomata have been induced *in vitro* (Cardoso 2015), characterised by ascomata measuring 230–355 × 175–290 µm, 57.5–75 × 5 µm asci producing 1-septate, 10–12.5 × 5 µm ascospores.

Neocosmospora neerlandica Crous & Sand.-Den., **sp. nov.**
MycoBank MB 838671. Fig. 38.

Etymology: Named after the country where the type was isolated, the Netherlands.

Typus: Netherlands, Zeeland Province, Zuid-Beveland, near Wolphaartsdijk, from *Pisum sativum*, unknown date, J.C. Went (**holotype** CBS H-24667, culture ex-type CBS 232.34).

Conidiophores borne on agar substrate and aerial mycelium up to 290 µm tall, unbranched or irregularly laterally branched, bearing terminal single monophialides, commonly proliferating percurrently; *aerial conidiogenous cells* monophialidic, subulate to subcylindrical, commonly extended percurrently, smooth- and thin-walled, 21–87 × 1.5–3.5 µm, with short and flared apical

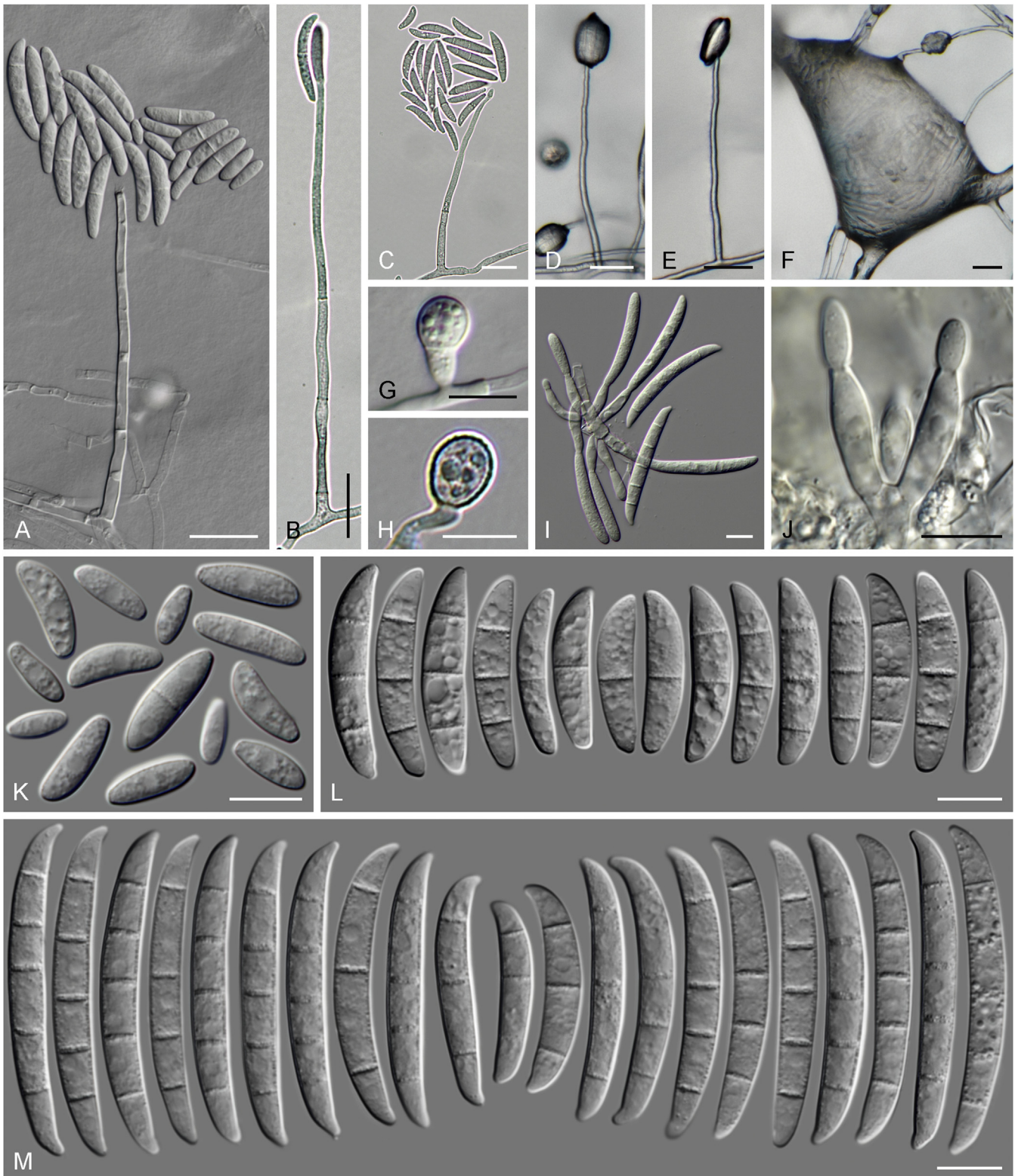


Fig. 37. *Neocosmospora merxiana* (CBS 146525). **A–E.** Aerial conidiophores and conidiogenous cells. **F.** Sporodochium on aerial mycelium. **G, H.** Chlamydospores. **I, J.** Sporodochial conidiophores and conidiogenous cells. **K.** Microconidia. **L.** Aerial macroconidia. **M.** Sporodochial macroconidia. Scale bars: A, E = 100 μm ; C = 20 μm ; all others = 10 μm .

collarettes and rather evident periclinal thickening. *Aerial conidia* of two types: *microconidia* oval to broadly ellipsoidal, smooth- and thin-walled, 0- or 1-septate, (5.5–)8–14(–30) \times (2–)3–4.5(–5.5) μm (av. 11 \times 3.8 μm), arranged in false heads on phialide tips; *macroconidia* fusiform to falcate, smooth- and thick-walled, straight to slightly curved, with a blunt apical cell, basal cell often flattened to obtuse, (1–)2–3-septate, predominantly 3-septate, 1-septate conidia: 22.5–26 \times 4.5–6 μm (av. 24.4 \times 5.1 μm); 2-septate conidia: (22.5–)

23.5–32 \times 3.5–5 μm (av. 27 \times 4.3 μm); 3-septate conidia: (24–)25–32.5(–38.5) \times (3.5–)4.5–5.5(–6) μm (av. 28.7 \times 4.8 μm); overall: (22.5–)24–31.5(–38.5) \times (3.5–)4.5–6 μm (av. 27.7 \times 4.8 μm), arranged in false heads at the tip of monophialides and produced intermixed with microconidia. *Chlamydospores* subspherical to spherical, pale golden brown, smooth- and thick-walled, 6–8 μm , single or in pairs, terminal or more often formed intercalary on hyphae. *Sexual morph* and *sporodochia* unknown.

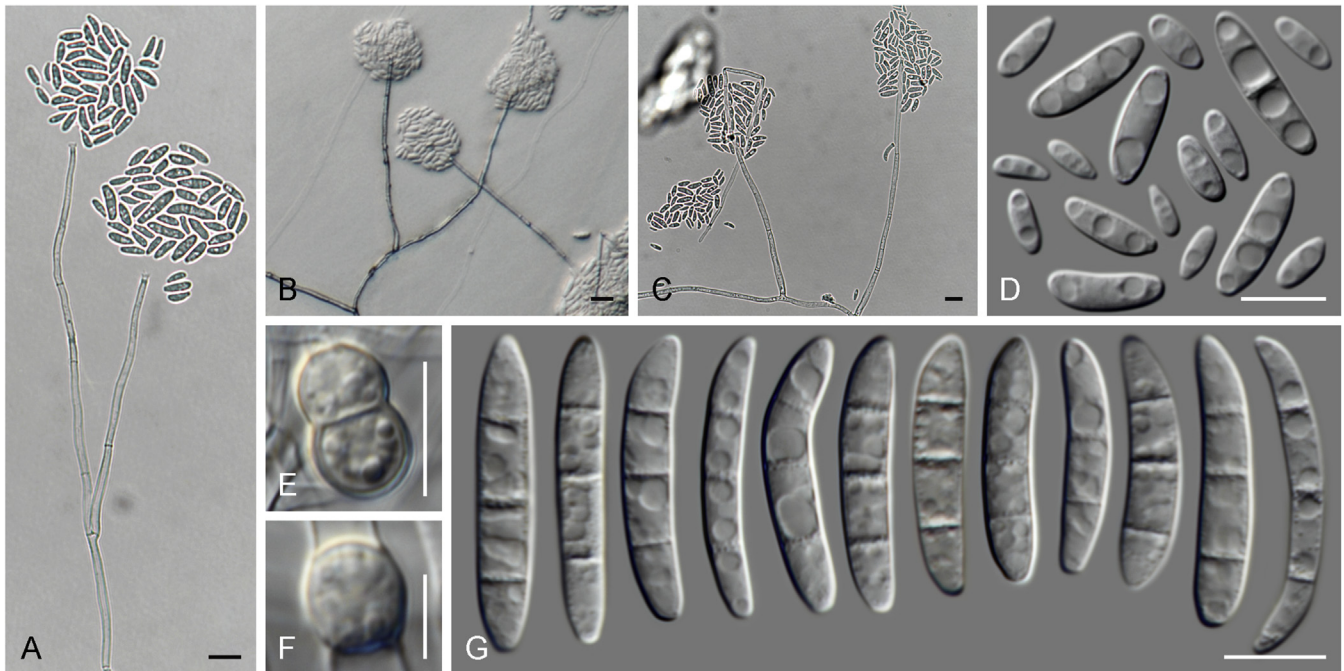


Fig. 38. *Neocosmospora neerlandica* (CBS 232.34). A–C. Conidiophores. D. Microconidia. E, F. Chlamydospores. G. Macroconidia. Scale bars: F = 5 μ m; all others = 10 μ m.

Culture characteristics: Colonies on PDA reaching 42–51 mm diam at 25 °C after 7 d. Surface white to pale luteous, flat with abundant dense aerial mycelium, velvety to cottony, margin regular and filiform; reverse pale luteous to sulphur yellow. On OA white to pale luteous, flat to slightly raised, velvety to cottony, margin regular and filiform; reverse pale luteous.

Notes: The type of *N. neerlandica* was originally deposited as *N. pisi*, an important root pathogen of *Pisum sativum*. Besides sharing the same host association, both species are genetically related, but cluster in distinct phylogenetic lineages and have a different morphology. Although *N. pisi* produces typical wedge-shaped, larger macroconidia (up to 46 μ m long) on abundant sporodochia (Šišić *et al.* 2018b), *N. neerlandica* is characterised by short falcate macroconidia (up to 38.5 μ m long) produced on aerial conidiophores, while sporodochia are not formed. The latter features relate *N. neerlandica* to *N. diminuta*, a phylogenetically distant species that produces the shortest falcate conidia known in *Neocosmospora* (Sandoval-Denis *et al.* 2019). Nevertheless, *N. diminuta* is a homothallic species that conspicuously produces sexual structures, while a sexual morph is not known for *N. neerlandica*. Additionally, macroconidia of *N. neerlandica* differ from those of *N. diminuta* by having less curved apices and poorly developed or non foot-shaped basal cells.

Neocosmospora nelsonii Crous & Sand.-Den., *sp. nov.*
Mycobank MB 838672. Fig. 39.

Etymology: In honour of Paul E. Nelson, prominent *Fusarium* researcher and collector of the ex-type strain of this species.

Typus: **Unknown country**, from *Pisum sativum*, unknown date, P.E. Nelson (**holotype** CBS H-12719, culture ex-type CBS 309.75).

Conidiophores borne on agar substrate and aerial mycelium, 59–330 μ m tall, often simple and reduced to solitary phialides borne laterally from hyphae, or laterally irregularly and sympodially branching one or two times, bearing terminal single phialides; **aerial conidiogenous cells** monophialidic, subulate to

subcylindrical, smooth- and thin-walled, 21–57.5 \times 2–5 μ m, flared apical collarettes and periclinal thickening present. **Aerial microconidia** arranged in false heads on phialide tips, hyaline, broadly ellipsoidal, obovate to broadly clavate, smooth- and thin-walled, 0(–1)-septate, (5–)7–13(–17) \times 2.5–5 μ m (av. 10.1 \times 3.7 μ m). **Sporodochia** (from holotype specimen) pale citrine to olivaceous; **sporodochial conidiophores** copiously branched, laterally, verticillate and irregularly, bearing apical groups of 2–3 monophialides and lateral solitary phialides; **sporodochial conidiogenous cells** monophialidic, doliiform, subulate to subcylindrical, 6–21.5 \times 3–4.5 μ m, smooth and thin-walled, with short, conspicuously flared collarettes and conspicuous periclinal thickening, profusely proliferating percurrently. **Sporodochial macroconidia** falcate, gently and regularly curved dorsoventrally or with an almost straight ventral line, broadest at the middle portion, apical cell blunt and slightly hooked, basal cell papillate to well-developed, foot-shaped, 1–3(–4)-septate, predominantly 3-septate, hyaline, smooth- and thick-walled; 1-septate conidia: (17.5–)19–26(–29.5) \times 4–5 μ m (av. 22.4 \times 4.4 μ m); 2-septate conidia: (26–)27–34 \times 3.5–5.5 μ m (av. 30 \times 4.7 μ m); 3-septate conidia: (25.5–)30.5–38(–42) \times 4–5.5 μ m (av. 34.3 \times 4.8 μ m); 4-septate conidia: 38.5–43.5 \times 4.5–5.5 μ m (av. 40.7 \times 5.0 μ m); overall: (17.5–)27–38(–43.5) \times (3.5–)4–5.5 μ m (av. 32.5 \times 4.7 μ m). **Chlamydospores** subspherical to spherical, pale golden brown, smooth- and thick-walled, 4–11.5 μ m, formed singly and terminally on hyphae. **Sexual morph** not observed.

Culture characteristics: Colonies on PDA reaching 35–49 mm diam at 25 °C after 7 d. Surface pale luteous, pale saffron to sulphur yellow, flat with abundant dense and short aerial mycelium, velvety to woolly, margin filiform; reverse sulphur yellow. On OA pale luteous, flat, membranous to dusty with filiform margin; reverse pale luteous.

Notes: The ex-type of *N. nelsonii*, originally determined as “*F.*” *solani*, currently presents a very simple microconidial morphology with a rather acremoniid touch given its slender, generally simple conidiophores and mostly aseptate

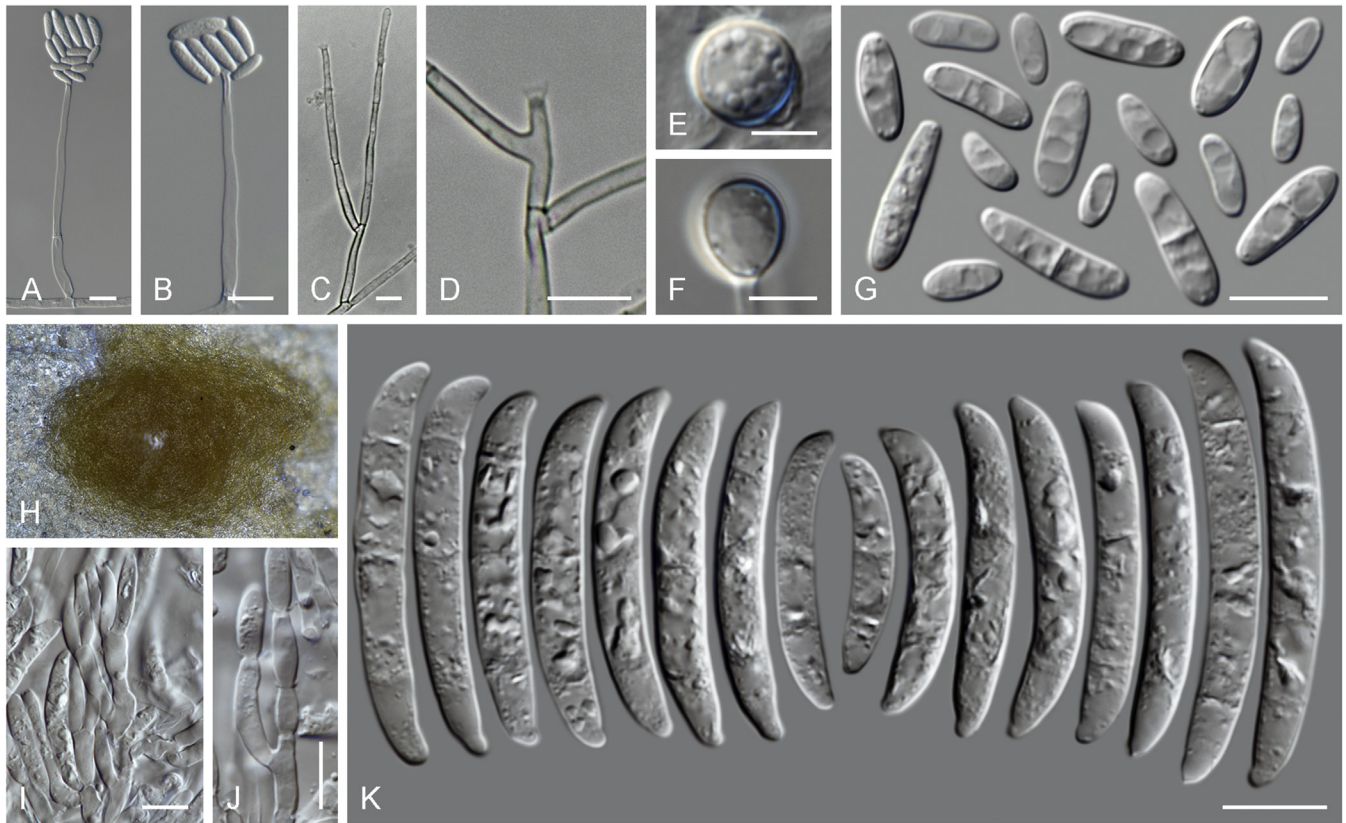


Fig. 39. *Neocosmospora nelsonii* (CBS 309.75). A–D. Conidiophores and conidiogenous cells. E, F. Chlamydospores. G. Microconidia. H. Sporodochium. I, J. Sporodochial conidiophores and conidiogenous cells. K. Macroconidia. Scale bars: E, F = 5 μm ; all others = 10 μm .

microconidia. Hence, there are no clear phenotypic characters to differentiate the species. Failed attempts to induce formation of sporodochia indicate that the ex-type strain may have lost the ability to produce macroconidia *in vitro*. The holotype material, is, however, a dried subculture from the type strain dated from 1982. It still contains a large amount of well-preserved sporodochia and sporodochial conidia, which we describe here. These macroconidia are comparable in size to those observed in closely related species such as *N. brevis*, *N. pisi*, and *N. neerlandica*. However, macroconidia in *N. brevis* and *N. neerlandica* are produced only in the aerial mycelium, while *N. nelsonii* produces only a single type of aerial conidia (microconidia), which also differ from those observed in the aforementioned species by their reduced size. In addition, sporodochial conidia in *N. nelsonii* are shorter and stout, with shorter and rounder apices compared to those of *N. pisi*.

Neocosmospora pseudopisi Sand.-Den. & L. Lombard, *sp. nov.* MycoBank MB 838673. Fig. 40.

Etymology: Named after its morphological, phylogenetic and host affinity with *Neocosmospora pisi*.

Typus: **Unknown country**, from *Pisum sativum*, unknown date and collector (**holotype** CBS H-24668, culture ex-type CBS 266.50).

Conidiophores borne on agar substrate and aerial mycelium, erect and prostrate, up to 340 μm tall, unbranched or irregularly laterally branched, bearing terminal single phialides, rarely proliferating percurrently; *aerial conidiogenous cells* monophialidic, rarely extended percurrently, subulate to subcylindrical, smooth- and thin-walled, 24.5–74 \times 2–4 μm , with cup-shaped, elongated, and flared apical collarettes and

conspicuous periclinal thickening. *Aerial microconidia* arranged in false heads on phialide tips, hyaline, broadly ellipsoidal to clavate, often lightly curved and asymmetrical, smooth- and thin-walled, 0(–1)-septate, (4.5–)6.5–11(–17.5) \times (2–)3–4(–5) μm (av. 8.6 \times 3.2 μm). *Sporodochia* pale luteous to pale sienna coloured, rarely formed on the surface of carnation leaves, agar surface or on aerial mycelium; *sporodochial conidiophores* unbranched or laterally and irregularly branched bearing single monophialides or groups of groups of up to three monophialides; *sporodochial conidiogenous cells* monophialidic, subulate to subcylindrical, 10–25 \times 2–5 μm , smooth and thin-walled, collarettes and periclinal thickening present. *Sporodochial macroconidia* falcate, gently tapering towards both ends, slightly curved dorsoventrally to almost straight, apical cell blunt to inconspicuously papillate, basal cell obtuse to poorly-developed, foot-shaped, 1–4(–5)-septate, predominantly 4-septate, hyaline, smooth- and thick-walled; 1-septate conidia: 21.5–26(–27.5) \times 4–5 μm (av. 24.7 \times 4.3 μm); 2-septate conidia: 28–30 \times 4.5–5 μm ; 3-septate conidia: (28.5–)34–46.5(–50) \times 4–5.5 μm (av. 40.1 \times 4.7 μm); 4-septate conidia: (36–)42.5–54(–56) \times 4–5.5 μm (av. 48 \times 4.9 μm); 5-septate conidia: 50.5 \times 5 μm ; overall: (21.5–)34.5–51.5(–56) \times 4–5.5 μm (av. 42.9 \times 4.8 μm). *Chlamydospores* subspherical to spherical, hyaline to pale yellow, smooth-walled, thick-walled, 5.5–10.5 μm , single or in pairs, terminal or intercalary. *Sexual morph* not observed.

Culture characteristics: Colonies on PDA reaching 35–48 mm diam at 25 $^{\circ}\text{C}$ after 7 d. Surface pale luteous to pale sulphur yellow, flat with abundant short aerial mycelium, velvety to dusty, margin regular entire to filiform; reverse pale luteous to sulphur yellow. On OA pale luteous to pale sulphur yellow,

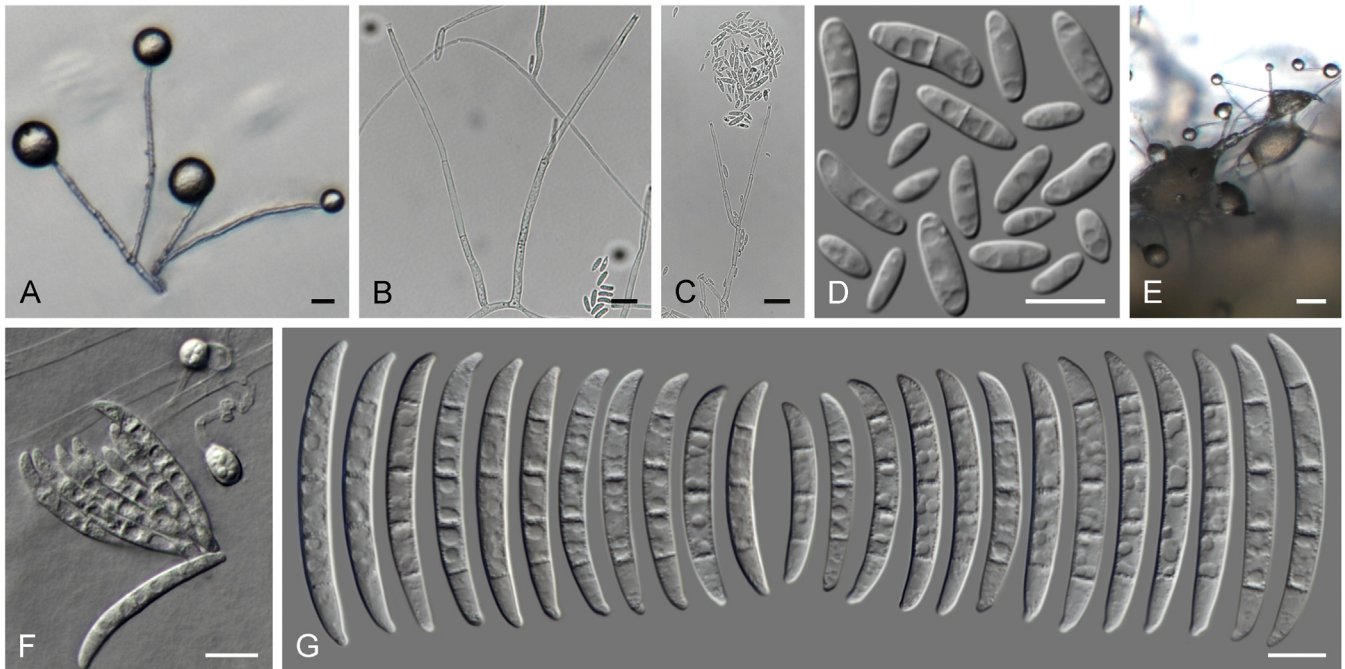


Fig. 40. *Neocosmospora pseudopisi* (CBS 266.50). **A–C.** Conidiophores and conidiogenous cells. **D.** Microconidia. **E.** Sporodochia formed on aerial hyphae. **F.** Macroconidia and chlamydospores. **G.** Macroconidia. Scale bars: C = 20 µm; E = 100 µm; all others = 10 µm.

flat, velvety to dusty, margin entire to filiform; reverse pale luteous.

Notes: The type of *N. pseudopisi* was determined as pathogenic to *Pisum sativum* and deposited in WI by W.C. Snyder. It is phylogenetically and morphologically related to *N. pisi*, a major pathogen of *Pisum sativum* (Šišić *et al.* 2018b). However, both species resolved as very closely related lineages in the seven-marker phylogeny (Fig. 14), as well as on the individual *CaM*, *ITS*, *rpb1*, and *rpb2* phylogenies (data not shown). Morphologically, *N. pseudopisi* can be differentiated from *N. pisi* by its longer sporodochial conidia (up to 56 µm long vs up to 46 µm long in *N. pisi*, Šišić *et al.* 2018b). Based on the features of its macroconidia, *N. pseudopisi* resembles *N. crassa* and *N. pseudotonkinensis*; the two latter species, though, are phylogenetically well-separated. *Neocosmospora pseudopisi*, however, differs from *N. crassa* and *N. pseudotonkinensis* by the absence of aerial macroconidia in the former species, while unlike *N. crassa*, the sporodochial conidia of *N. pseudopisi* are often wider on its apical third (vs wider at its basal part in *N. crassa*).

Neonectria Wollenw., Ann. Mycol. 15: 52. 1917, *nom. cons. prop.* Fig. 8.

Synonym: *Cylindrocarpon* Wollenw., Phytopathology 3: 225. 1913.

(see Chaverri *et al.* 2011 for additional synonyms)

Type species: *Neonectria ramulariae* Wollenw., Ann. Mycol. 15: 52. 1917.

Ascomata perithecial, gregarious, seated on an erumpent stroma, superficial, subglobose to broadly obpyriform, red, turning dark red in KOH, pigment dissolving in lactic acid, not collapsing when dry, with blunt to acute apex, rarely papillate, smooth to slightly rugulose, lacking hairs or appendages. **Ascomatal wall** of two regions: outer region of thick-walled, pigmented cells forming a *textura epidermoidea*; inner region of elongate, hyaline, thin-walled cells, becoming thinner toward the

centrum. **Asci** cylindrical, 8-spored, without an apical ring, uniseriate. **Ascospores** ellipsoidal to fusoid, 1-septate, hyaline, smooth or finely spinulose. **Sporodochia** not formed. **Conidiophores** mononematous, hyaline, septate, unbranched or irregularly branched, terminating in 1–3 phialides or reduced to lateral phialides. **Conidiogenous cells** monopialidic, cylindrical, tapering towards the apex, with inconspicuous periclinal thickening and collarettes. **Microconidia** abundant, ellipsoidal to obovoid, hyaline, aseptate, sometimes forming false heads on phialides. **Macroconidia** cylindrical, mostly straight, 3–7(–9)-septate, with rounded ends. **Chlamydospores** globose to subglobose, hyaline to subhyaline, smooth-walled to slightly verrucose, terminal or intercalary, solitary or in pairs or forming chains. [Description adapted from Chaverri *et al.* (2011)].

Diagnostic features: Red, mostly smooth-walled perithecia lacking papilla producing cylindrical asci bearing ellipsoidal to fusoid, 1-septate ascospores and *Cylindrocarpon* asexual morph.

Nothofusarium Crous, Sand.-Den. & L. Lombard, **gen. nov.** MycoBank MB 838674. Fig. 8.

Etymology: From the Greek prefix notho-, false, illegitimate; and *Fusarium*, in reference to the genetic affinity and morphological resemblance to the genus *Fusarium* s. str.

Type species: *Nothofusarium devonianum* L. Lombard, Crous & Sand.-Den.

Ascomata unknown. **Conidiophores** mononematous (aerial conidiophores) or grouped on sporodochia. **Aerial conidiophores** simple, unbranched or irregularly branched, sometimes reduced to single lateral phialides or phialidic pegs on the hyphae; **conidiogenous cells** monopialidic, cylindrical, tapering towards apex, smooth- and thin-walled, with periclinal thickening inconspicuous or absent, solitary. **Microconidia** not formed. **Aerial macroconidia** falcate, 1–5(–6)-septate, thick-walled, curved to lunate, with a blunt apical cell and often obtuse, poorly- to well-developed foot-shaped basal cell. **Sporodochia** white, pale luteous to pale citrine. **Sporodochial conidiophores** irregularly

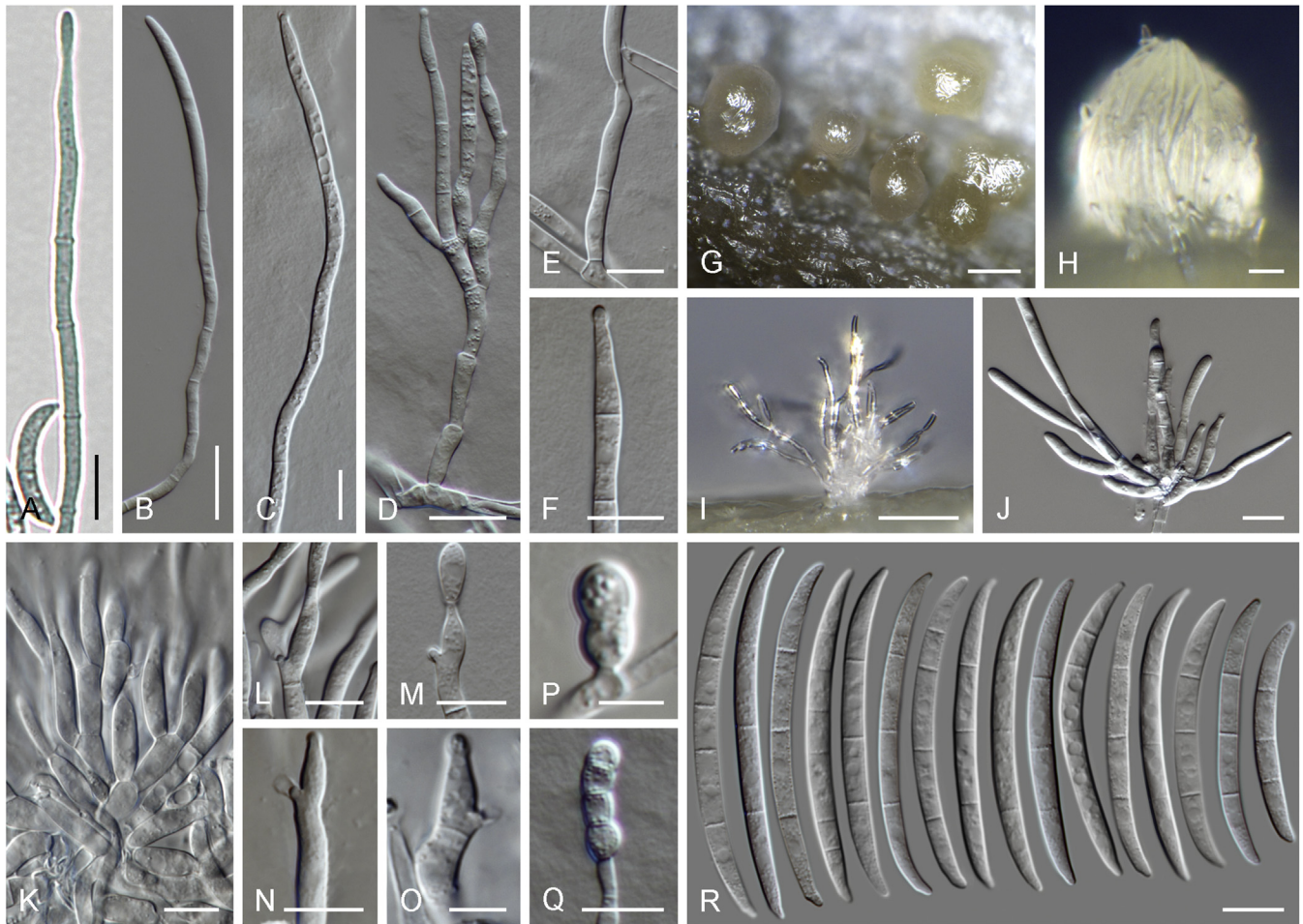


Fig. 41. *Nothofusarium devonianum* (CBS 147304). **A–F.** Aerial conidiophores and conidiogenous cells. **G–I.** Sporodochia formed on the surface of carnation leaves. **J–O.** Sporodochial conidiophores and conidiogenous cells. **P, Q.** Chlamydospores. **R.** Macroconidia. Scale bars: B, D = 20 μm ; G, H = 200 μm ; O, P = 5 μm ; all others = 10 μm .

and verticillately branched, consisting of short, smooth- and thin- to thick-walled stipes bearing apical whorls of mono- and polyphialides. *Sporodochial conidiogenous cells* monopialidic and polyphialidic, doliiform, subulate to subcylindrical, smooth- and thin-walled, with reduced apical collarette. *Sporodochial macroconidia* similar to aerial macroconidia. *Chlamydospores* subglobose to ellipsoidal, solitary or most commonly in chains.

Diagnostic features: Fusarioid asexual morph characterised by aerial monopialides and sporodochial mono- and polyphialides producing slightly curved and slender, mostly 3-septate macroconidia.

Nothofusarium devonianum L. Lombard, Crous & Sand.-Den., **sp. nov.** MycoBank MB 838675. Fig. 41.

Etymology: The epithet refers to Devon, the English county where the type specimen was collected.

Typus: UK, England, Devon, Totnes, Berry Pomeroy, Loventor Manor, on dead cladodes of *Ruscus aculeatus*, 17 Jul. 1983, B.C. Sutton & A.V. Sutton (**holotype** CBS H-24670, culture ex-type CBS 147304 = IMI 279297 = NRRL 22134).

Conidiophores borne on substrate mycelium, prostrate or erect and quickly collapsing to the agar surface, 70–240 μm tall, unbranched or less commonly irregularly laterally branched, bearing terminal single phialides; *aerial conidiogenous cells* monopialidic, subulate to cylindrical, smooth- and thin-walled, 9–34 μm long, 2–5 μm at the widest part, or reduced to short phialidic pegs, 3–6 \times 2–3.5 μm , formed laterally on aerial

hyphae, apical collarettes short or lacking, periclinal thickening absent. *Aerial macroconidia* borne on tips of conidiogenous cells on aerial conidiophores, almost straight or slightly curved, falcate, 1–5(–6)-septate, predominantly 3-septate, hyaline, smooth- and thick-walled, with a blunt apical cell and obtuse, sometimes papillate to poorly-developed, foot-shaped basal cell, 1-septate conidia: (15.5–)19–28(–32) \times 2.5–4 μm (av. 23.5 \times 4.3 μm); 2-septate conidia: (25.5–)27–31 \times 2.5–4 μm (av. 28.8 \times 3.2 μm); 3-septate conidia: (13–)41–57(–63.5) \times 3–4(–4.5) μm (av. 49 \times 3.6 μm); 4-septate conidia: (48.5–)50–60(–61.5) \times 3–4.5 μm (av. 55.1 \times 3.8 μm); 5-septate conidia: (47–)50–64(–71) \times 3.5–4.5 μm (av. 56.9 \times 3.9 μm); 6-septate conidia: (54–)55–71.5 \times 3.5–4 μm (av. 62.3 \times 3.8 μm); overall: (13–)35.5–59(–71.5) \times 2.5–4.5 μm (av. 47.2 \times 3.6 μm). *Sporodochia* pale luteous to pale citrine coloured, small, formed abundantly on the agar surface and less regularly on the surface of carnation leaves; *sporodochial conidiophores* irregularly verticillately branched bearing solitary lateral and terminal phialides or apical groups of 2–3 phialides; *sporodochial conidiogenous cells* mono- and polyphialidic, doliiform, subulate to subcylindrical, 3–25.5 \times 2.5–5 μm , smooth and thin-walled, commonly proliferating sympodially, collarettes and periclinal thickening absent or inconspicuous. *Sporodochial conidia* undifferentiable from aerial conidia. *Chlamydospores* subglobose to ellipsoidal, solitary or most commonly in chains. **Sexual morph** unknown.

Culture characteristics: Colonies on PDA reaching 23–27 mm diam at 25 $^{\circ}\text{C}$ after 7 d. Surface straw-coloured, pale

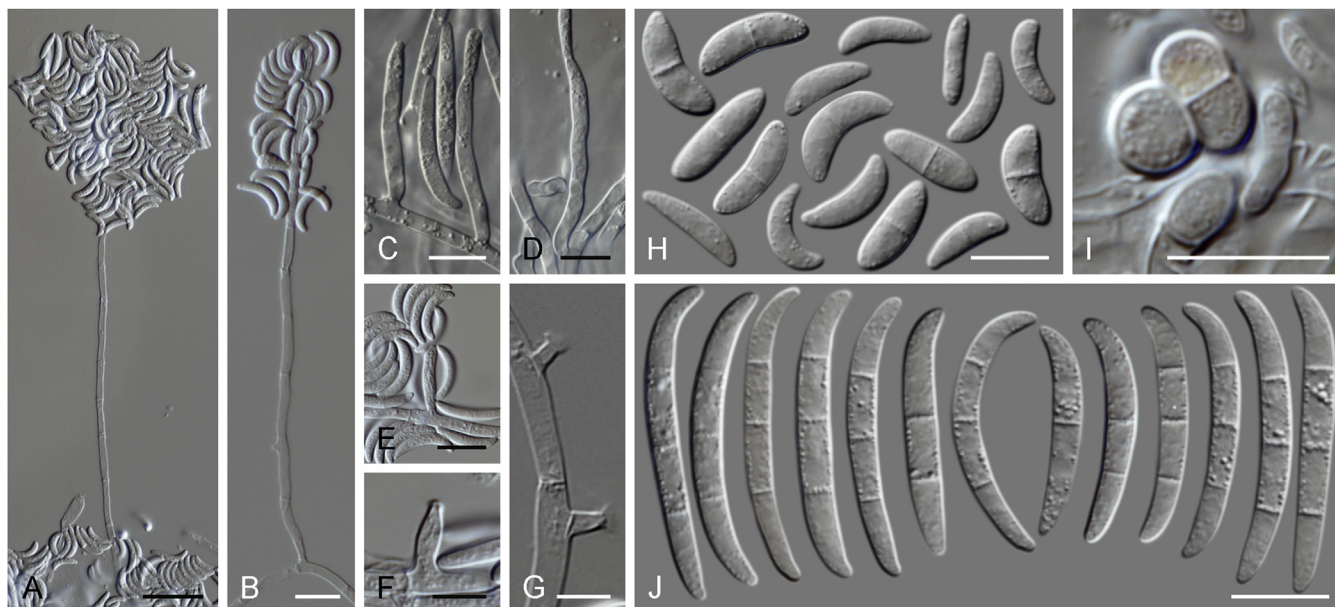


Fig. 42. *Pseudofusicolla belgica*. A, B. Conidiophores. C–G. Conidiogenous cells. H. Microconidia. I. Chlamydospores. J. Macroconidia. A, B, D–J. IHEM 2413. C. IHEM 5322. Scale bars: A = 20 μ m; F, G = 5 μ m; all others = 10 μ m.

luteous to pale ochreous, flat, dusty to velvety; reverse white to pale luteous without diffusible pigments. On OA, grey-white to pale luteous, flat, membranous to dusty, with irregular velvety peripheral patches cottony; reverse pale luteous.

Notes: The type of *No. devonianum* was erroneously assigned to *Trichofusarium rusci* (Sutton, 1986) and recombined in *Fusarium* (*Fusarium rusci*, Geiser *et al.* 2013). Nevertheless, the morphology exhibited by this strain does not match in respect with the original description of the supposed basionym nor its purported synonym *Pycnofusarium rusci*, as confirmed also by examination of authentic material of *T. rusci* (BPI 453152A and IMI 291476). The latter taxon is characterised by a setose sporodochial asexual morph with small, fusoid, aseptate conidia, more reminiscent of the genus *Alfaria* (*Stachybotryaceae*, Crous *et al.* 2014).

Pseudofusicolla D. Triest, *Mycobiology* 44: 127. 2016. Figs 8, 42.

Type species: *Pseudofusicolla belgica* D. Triest, *Mycobiology* 44: 127. 2016.

Ascomata unknown. **Conidiophores** initially as lateral phialides on somatic hyphae, sometimes monochasial, verticillate or penicillate, hyaline. **Conidiogenous cells** monophialidic, cylindrical to subulate, hyaline, producing micro- and macroconidia. **Microconidia** strongly falcate, 0- or 1-septate, hyaline. **Macroconidia** strongly falcate, narrowing towards the ends, apical cell hooked with a pointed tip, basal cell papillate to poorly-developed, foot-shaped, 0–3-septate, hyaline. **Chlamydospores** globose, in terminal pairs or intercalary chains. [Description adapted from Triest *et al.* (2016)].

Diagnostic features: Fusarioid asexual morph that produces strongly curved, 0- or 1-septate microconidia, and 0–3-septate macroconidia.

Rectifusarium L. Lombard *et al.*, *Stud. Mycol.* 80: 229. 2015. Figs 8, 43.

Type species: *Rectifusarium ventricosum* (Appel & Wollenw.) L. Lombard & Crous, *Stud. Mycol.* 80: 229. 2015.

Basionym: *Fusarium ventricosum* Appel & Wollenw., *Phytopathology* 3: 32. 1913.

(See *F. ventricosum* in List section for synonyms)

Ascomata perithecial, mostly gregarious, non-stromatic or on a thin stroma erumpent through the epidermis, superficial, subglobose to globose, laterally pinched when dry, dark red, with short ostiolar neck, smooth-walled, lacking hairs and appendages. **>Ascomatal wall** of two regions: outer region of thick-walled, pigmented cells forming a *textura angularis* or *textura globulosa*; inner region of elongate, hyaline, thin-walled cells, becoming thinner towards the centrum. **Asci** clavate, apex rounded with distinct pore, 8-spored often with an apical ring, uniseriate to biseriate. **Ascospores** ellipsoidal, 1-septate, constricted at the septum, pale tan, verrucose. **Sporodochia** not formed. **Conidiophores** simple, mononematous, straight to flexuous, hyaline, septate, unbranched or rarely branched, terminating in single phialides. **Conidiogenous cells** monophialidic, cylindrical, tapering towards the apex, with periclinal thickening and flared collarettes, usually producing macroconidia. **Microconidia** rarely formed, ellipsoidal to fusoid, 0- or 1-septate, hyaline. **Macroconidia** falcate, straight to slightly curved dorsiventrally, 3-septate, with blunt to slightly pointed apical cell and poorly-developed foot-shaped basal cell. **Chlamydospores** globose to subglobose to ovoid, hyaline to subhyaline, verrucose, terminal or intercalary, solitary or in pairs or forming chains or developing directly from macroconidia.

[Description adapted from Booth (1971), Gerlach & Nirenberg (1982) and Lombard *et al.* (2015)].

Diagnostic features: Dark red, smooth-walled perithecia with short ostiolar neck producing clavate asci bearing ellipsoidal, 1-septate ascospores and asexual morphs producing micro- and macroconidia on elongate cylindrical aerial conidiophores with monophialides, and not forming sporodochia. Chlamydospores formed in hyphae and macroconidia.

Rugonectria P. Chaverri & Samuels, *Stud. Mycol.* 68: 73. 2011. Fig. 8.



Fig. 43. *Rectifusarium* spp. A–F. Conidiophores and conidiogenous cells. G. Microconidia. H. Macroconidia. A–D, H. *Rectifusarium robinianum* (CBS 430.91). E–G. *Rectifusarium ventricosum* (CBS 748.79). Scale bars = 10 µm.

Type species: Rugonectria rugulosa (Pat. & Gaillard) Samuels *et al.*, Stud. Mycol. 68: 73. 2011.

Basionym: Nectria rugulosa Pat. & Gaillard, Bull. Soc. Mycol. France 4: 115. 1889.

Synonyms: Cucurbitaria rugulosa (Pat. & Gaillard) Kuntze, Revis. Gen. Pl. 3: 461. 1898.

Neonectria rugulosa (Pat. & Gaillard) Mantiri & Samuels, Canad. J. Bot. 79: 339. 2001.

Cylindrocarpon rugulosum Brayford & Samuels, Sydowia 46: 148. 1994.

Ascomata perithecial, solitary or gregarious, stromatic, superficial or partly immersed in stroma, subglobose to globose, orange to red, turning dark red in KOH, pigment dissolving in lactic acid, non-papillate, rugose to tuberculate, lacking hairs or appendages. *Ascomatal wall* of two regions: outer region of thick-walled, pigmented cells forming a *textura angularis*; inner region of elongate, hyaline, thin-walled cells, becoming thinner towards the centrum. *Asci* clavate, apex simple, 8-spored. *Ascospores* ellipsoidal to oblong, 1-septate, not to slightly constricted at the septum, pale yellow, striate. *Sporodochia* not formed. *Conidiophores* simple, mononematous, straight to flexuous, hyaline, septate, unbranched or rarely to irregularly branched, terminating in single phialides. *Conidiogenous cells* monophialidic, cylindrical, tapering towards the apex, with periclinal thickening and flared collarettes, producing micro- and macroconidia. *Microconidia* ovoid to cylindrical, 0- or 1-septate, hyaline. *Macroconidia* fusoid, curved, (3–)5–7(–9)-septate, tapering to both ends, basal cell obtuse with inconspicuous hilum. *Chlamydospores* not observed.

[Description adapted from Samuels *et al.* (1990), Samuels & Brayford (1994) and Chaverri *et al.* (2011)].

Diagnostic features: Orange to red, rugose to tuberculate, partially immersed perithecia producing clavate asci bearing fusoid, 1-septate yellowish, striate ascospores and cylindrocarpon-like asexual morph characterised by curved, multi-septate macroconidia with inconspicuous hilum.

Scolecofusarium L. Lombard, Sand.-Den. & Crous, *gen. nov.* MycoBank MB 838676. Figs 8, 44.

Etymology: From Greek *skólēx*, worm, in reference to the worm-like appearance of the macroconidia.

Type species: Scolecofusarium ciliatum (Link) L. Lombard, Sand.-Den. & Crous

Ascomata perithecial, solitary or gregarious, partially immersed on a stroma, smooth- and thin-walled, globose to broadly pyriform, red, with a broad, discoid apical region, turning darker in

KOH, pigment dissolving in lactic acid to become yellow, lacking hairs and warts. *Ascomatal wall* of a single region composed of unevenly thickened cells of *textura epidermoidea*. *Asci* cylindrical, apex with an obscure refractive ring, 8-spored, ascospores uniseriate. *Ascospores* ellipsoidal to fusiform-ellipsoidal, 1-septate, not constricted at septum, yellow-brown, finely spinulose. *Conidiophores* mononematous (aerial) or grouped on sporodochia. *Aerial conidiophores* unbranched to loosely irregularly branched, bearing terminal phialides; *conidiogenous cells* monophialidic, subcylindrical, smooth- and thin-walled, with evident periclinal thickening and a non-flared collarette, producing only macroconidia. *Sporodochia* pink, orange to salmon coloured; *sporodochial conidiophores* irregularly and verticillately branched, consisting of short, often swollen, smooth- and thin-walled stipes bearing single terminal monophialides or apical whorls of 2–3 monophialides; *sporodochial conidiogenous cells* monophialidic, cylindrical to subcylindrical, smooth- and thin-walled, with evident periclinal thickening. *Macroconidia* formed in pink to salmon slimy masses, subcylindrical, (0–)3–7(–10)-septate, straight or slightly curved, with blunt apical cell and obtuse to poorly developed, foot-shaped basal cell. *Microconidia* unknown. *Chlamydospores* unknown.

[Description adapted from Samuels *et al.* (1991) & Gerlach & Nirenberg (1982)].

Diagnostic features: Red perithecia producing cylindrical asci containing ellipsoidal, 1-septate, finely spinulose ascospores and fusarioid asexual morph characterised by monophialides producing slender and delicate, almost cylindrical macroconidia from aerial conidiophores and pink to salmon coloured sporodochia, lacking microconidia as well as chlamydospores.

Scolecofusarium ciliatum (Link) L. Lombard, Sand.-Den. & Crous, *comb. nov.* MycoBank MB 838677.

Basionym: Atractium ciliatum Link, Mag. Ges. Naturf. Freunde Berlin 7: 32. 1816.

Synonyms: Fusarium ciliatum (Link) Link, in Willdenow, Sp. Pl., Ed. 4, 6: 105. 1825.

Microcera ciliata (Link) Wollenw., Fusaria Autogr. Delin. 1: 435. 1916.

Calonectria ciliata (Link) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 32: 664. 1945.

Sphaeria agnina Desm., Ann. Sci. Nat., Bot. sér. 3, 6: 72. 1846.

Calonectria agnina (Desm.) Sacc., Michelia 1: 311. 1878.

Dialonectria agnina (Desm.) Cooke, Grevillea 12: 111. 1884.

Fusarium peltigerae Westend., Herb. Crypt. Belg. 9: no. 414. 1849.

Fusarium parasiticum Westend., Bull. Séances Cl. Sci. Acad. Roy. Sci. Belgique, sér. 2, 11: 652. 1861.

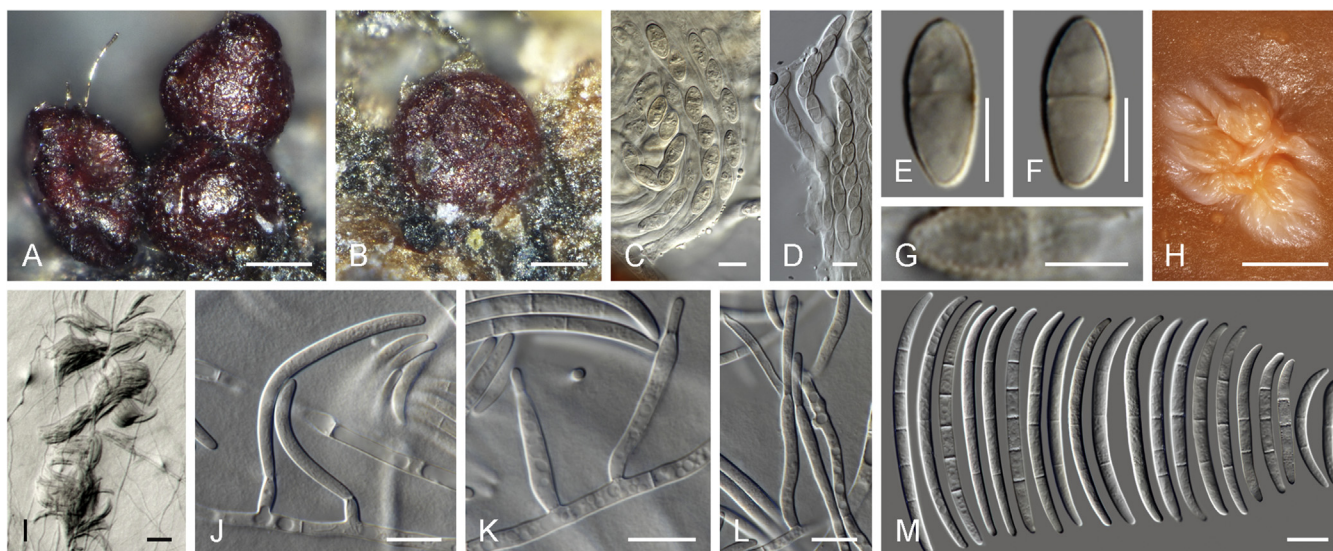


Fig. 44. *Scolecopus ciliatum*. A, B. Ascomata on natural substrate. C, D. Asci. E–G. Ascospores (G. Surface view). H. Pionnote on agar surface. I. Sporodochium. J–L. Conidiophores and conidiogenous cells. M. Macroconidia. A–H, J–L. CBS 146674. I. CBS 146676. M. CBS 144385. Scale bars: A, B = 100 μ m; E–G = 5 μ m; H = 1 mm; I = 20 μ m; all others = 10 μ m.

Nectria massariae Pass., in Rabenhorst, Fungi Eur. Exs. no. 1827. 1874.

Microcera massariae Sacc., Michelia 1: 263. 1878.

Calonectria massariae (Pass.) Sacc., Michelia 1: 312. 1878.

Fusisporium filisporum Cooke, Grevillea 8: 8. 1879.

Fusarium filisporum (Cooke) Sacc., Syll. Fung. 4: 708. 1886.

Fusarium scoleoides Sacc. & Ellis, Atti Reale Ist. Veneto Sci. Lett. Arti, sér. 6, 3: 728. 1885.

Fusarium elongatum Cooke, Grevillea 19: 4. 1890.

Calonectria dearnessii Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 42: 245. 1891.

Typus: **Germany**, on branch canker of *Fagus sylvatica*, 1961, W. Gerlach (**neotype** of *Atractium ciliatum* CBS H-12687 *hic designatus*, MBT 10000646, culture ex-neotype CBS 191.65 = ATCC 16068 = ATCC 24137 = BBA 9661 = DSM 62172 = IMI 112499 = NRRL 20431).

Additional descriptions and illustrations: Wollenweber & Reinking (1935), Doidge (1938), Gerlach & Nirenberg (1982).

Additional material examined: **Belgium**, Mons, Pommeroeul, on leaf of *Fagus sylvatica*, 1984, unknown collector, culture CBS 144385 = IHEM 2989. **Denmark**, on *Hordeum vulgare* mouldy grain, associated with scale insects, 1986, U. Thrane, culture CBS 155.86 = NRRL 22284. **Netherlands**, Noord-Brabant Province, Boxmeer, on *Quercus* sp., Mar. 2016, S. Helleman, cultures CBS 146672 = CPC 30654; CBS 146673 = CPC 30655; CBS 146674 = CPC 30656; CBS 146675 = CPC 30657; CBS 146676 = CPC 30658; CBS 146677 = CPC 30659.

Notes: No existent holotype material was located for *At. ciliatum*. Therefore, a neotype is designated here. The neotype specimen originates from a representative isolate studied by Gerlach & Nirenberg (1982).

Setofusarium (Nirenberg & Samuels) Crous & Sand.-Den., **gen. et stat. nov.** MycoBank MB 838678. Figs 8, 45.

Basionym: *Fusarium* sect. *Setofusarium* Nirenberg & Samuels, Canad. J. Bot. 67: 3376. 1989.

Etymology: The name refers to the presence of setose sporodochia and to its resemblance to the genus *Fusarium*.

Type species: *Fusarium setosum* Nirenberg & Samuels, Canad. J. Bot. 67: 3372. 1989.

Ascomata perithecial, solitary or gregarious on a well-developed immersed stroma composed of pseudoparenchymatous to hyphal cells, scaly to warty and thick-walled, pyriform, dark red with an often darker red-coloured, flattened and non-papillate apical region, turning darker in KOH, pigment dissolving in lactic acid to become yellow, lacking hairs. **Ascumatal wall** of two regions: outer region of thick-walled, pigmented cells of *textura angularis* to *textura globulosa* at warts cells; inner region of elongate, hyaline, thin-walled cells, becoming thinner towards the centrum. **Asci** cylindrical to clavate, with rounded to flattened simple apex, 8-spored, ascospores overlapping uniseriate to biseriate. **Ascospores** ellipsoidal, 1-septate, not constricted at septum, pale yellow-brown, smooth-walled to finely striate. **Conidiophores** mononematous (aerial) or grouped on sporodochia. **Aerial conidiophores** unbranched or rarely branched, bearing terminal phialides; **conidiogenous cells** monophialidic, cylindrical to sub-cylindrical, smooth- and thin-walled, with periclinal thickening inconspicuous to evident, producing only macroconidia. **Sporodochia** grey; **setae** arising between and around sporodochia, stiff, erect, thick-walled with acute tip, at first hyaline later becoming pale golden brown; **sporodochial conidiophores** irregularly and verticillately branched and densely packed, consisting of short, often swollen, smooth- and thin-walled stipes bearing apical whorl of 2–3 monophialides or single, terminal monophialides; **sporodochial conidiogenous cells** monophialidic, cylindrical to sub-cylindrical, smooth- and thin-walled, with inconspicuous to evident periclinal thickening. **Macroconidia** formed in off-white or grey slimy masses, cylindrical, (0–)3–5(–7)-septate, gently curved, with a blunt apical cell and an obtuse to poorly developed foot-shaped basal cell. **Microconidia** unknown. **Chlamydospores** unknown. [Description adapted from Samuels & Nirenberg (1989)].

Diagnostic features: Dark red perithecia producing cylindrical to clavate asci containing ellipsoidal, 1-septate, finely striate ascospores and fusarioid asexual morph characterised by monophialides producing robust, almost cylindrical macroconidia from

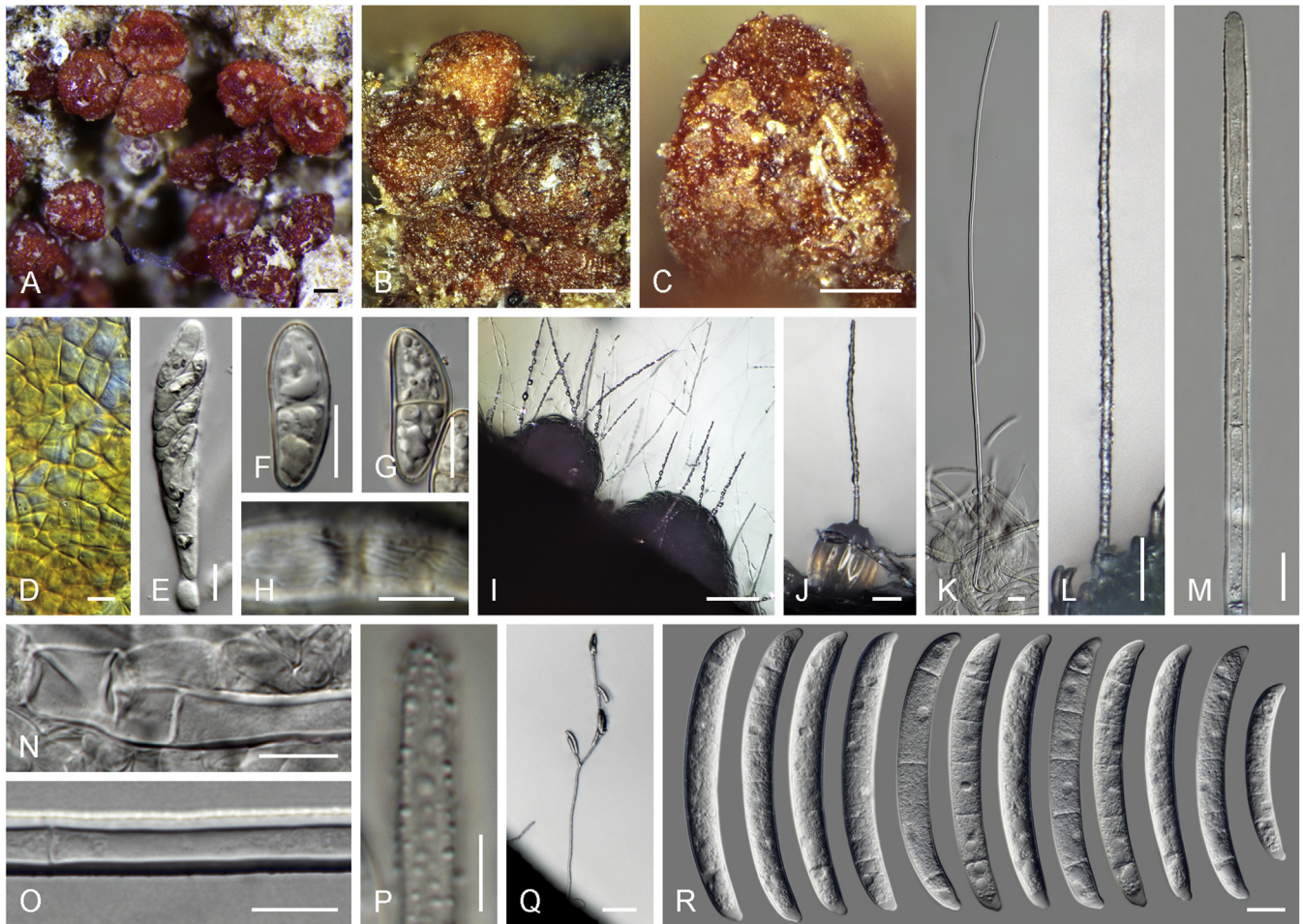


Fig. 45. *Setofusarium setosum*. **A–C.** Ascomata on natural substrate. **D.** Surface view of perithecial wall in lactic acid. **E.** Ascus. **F–H.** Ascospores (**H.** Surface view). **I, J.** Setose sporodochia. **K–M.** Setae. **N–P.** Detail of setae (**N.** Base. **O.** Middle portion wall. **P.** Surface view of apical wall). **Q.** Conidiophore. **R.** Macroconidia. **A–H.** BPI 882043. **I–R.** CBS 635.92. Scale bars: **A–C, I, Q** = 100 μ m; **J–L** = 20 μ m; **H, P** = 5 μ m; all others = 10 μ m.

aerial conidiophores and setose sporodochia, lacking microconidia as well as chlamydospores.

Setofusarium setosum (Nirenberg & Samuels) Sand.-Den. & Crous, **comb. nov.** MycoBank MB 838679.

Basionym: *Fusarium setosum* Nirenberg & Samuels, *Canad. J. Bot.* 67: 3372. 1989.

Synonym: *Nectria setofusarii* Samuels & Nirenberg, *Canad. J. Bot.* 67: 3372. 1989.

Typus: French Guiana, piste de Saint-Elie: km 16 on road between Sinnamary and St. Elie, ORSTOM research area “ECEREX”, on bark of living liana, Mar. 1986, G.J. Samuels, **holotype** NY00927992. **Epitype** of *F. setosum* (CBS H-24723 *hic designatus*, MBT 10000647): French Guiana, Vic. Cayenne, 15 km from Remise, trail to Vidal-old farm, secondary forest, from bark, 25 Feb. 1988, A.Y. Rossman, culture ex-epitype CBS 635.92 = G.J.S. 88-12.

Description and illustrations: Samuels & Nirenberg (1989).

Additional material examined: French Guiana, unknown host and collection date, A.Y. Rossman, culture CBS 574.94; from wood from unknown host, Feb. 1988, A.Y. Rossman, IMI 324476. Ghana, Western Region: Wiawso District, Bia National Park, trail from camp 1, disturbed forest, on living liana, J.G. Samuels & H.C. Evans, BPI 882043.

Notes: The monotypic, former *Fusarium* section *Setofusarium* is here elevated to generic rank to accommodate “*Fusarium setosum*”, a genetically and morphologically divergent taxon

easily differentiated from any known fusarioid taxa by the production of setose sporodochia (Samuels & Nirenberg 1989). No living ex-type culture could be located for this taxon. Isolate CBS 635.92 (as G.J.S. 88-12) is an authentic strain of *Fusarium setosum* (Samuels & Nirenberg 1989). Therefore, a dried culture from this strain is designated as epitype here.

Stylonectria Höhn., *Sitzungsber. Kaiserl. Akad. Wiss. Wien, Math.-Naturwiss. Cl., Abt. 1, 124: 52.* 1915. **Figs 8, 46.**

Type species: *Stylonectria applanata* Höhn., *Sitzungsber. Kaiserl. Akad. Wiss. Wien, Math.-Naturwiss. Kl., Abt. 1, 124: 52.* 1915.

Ascomata perithecial, gregarious in groups of up to 20, on a thin, white to yellow hyphal or subiculum-like stroma, superficial, subglobose, pyriform to subcylindrical, pale yellow, orange-red, orange-brown, or pale to dark red, becoming dark red to purple in KOH, with a rounded or broad, circular, flat disc on a venter-like neck, smooth to slightly rugulose, lacking hairs or appendages. **Ascomatal wall** consisting of two layers; inner layer of hyaline, thin-walled, compressed, elongated cells and outer layer of distinct, isodiametric to oblong, angular or globose, thick-walled cells. **Asci** cylindrical to clavate, 8-spored, with simple apex or apical ring. **Ascospores** cylindrical to allantoid to ellipsoidal, 1-septate, hyaline or yellow to pale brown, smooth or tuberculate. **Conidiophores** initially formed as unbranched phialides on somatic hyphae, sometimes loosely branched, sometimes forming small sporodochia. **Conidiogenous cells** monophialidic,

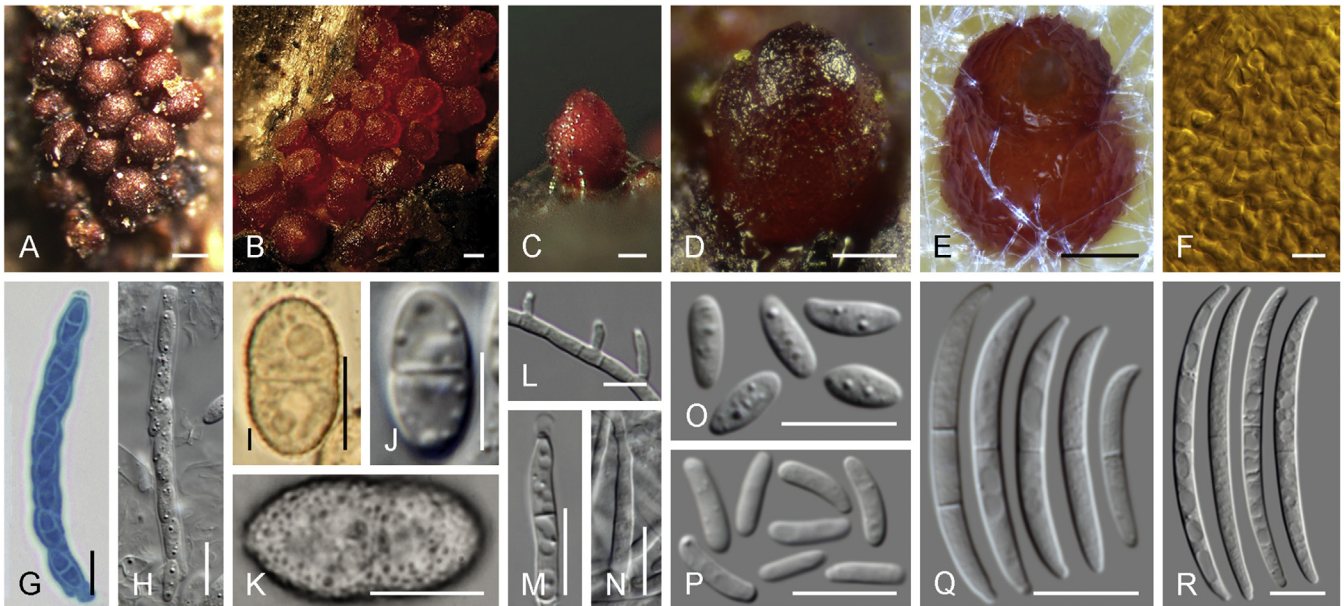


Fig. 46. *Stylonectria* spp. **A–D.** Ascomata on natural substrate. **E.** Ascomata on culture. **F.** Surface view of perithecial wall in lactic acid. **G, H.** Asci. **I–K.** Ascospores. **L–M.** Conidiophores and conidiogenous cells. **O, P.** Microconidia. **Q, R.** Macroconidia. **A, G.** *Stylonectria qilianshanensis* [HMAS 255803, adapted from Zeng *et al.* (2020)]. **B.** *Stylonectria norvegica* [CLL14047, adapted from Lechat *et al.* (2015)]. **C.** *Stylonectria purtonii* (photo P. Mičoch). **D, I, K.** *Stylonectria wegeliana* (photo B. Bergen). **E, F, Q.** *Stylonectria hetmanica* (CBS 147306). **H, J, O.** *Stylonectria* sp. (HPC 2668). **L, M.** *Stylonectria corniculata* (CBS 125491). **N, P, R.** *Stylonectria applanata* (CBS 125489). Scale bars: **A–E** = 100 μ m; **I–K** = 5 μ m; all others = 10 μ m.

cylindrical to subcylindrical, with a distinct collarette. *Microconidia* sparse, allantoid to lunulate, slightly or strongly curved, aseptate, in slimy heads. *Macroconidia* orange in mass, subcylindrical or moderately to strongly curved, falcate, 0- or 1-septate, apex narrower than base, apical cell blunt or hooked, basal cell not or scarcely foot-shaped.

[Description adapted from Höhnel (1915) and Gräfenhan *et al.* (2011)].

Diagnostic features: Pale yellow to dark red, mostly smooth-walled perithecia with rounded or broad, circular, flat disc on a venter-like neck, producing cylindrical to clavate asci bearing cylindrical to allantoid to ellipsoidal, 1-septate hyaline or yellow to pale brown ascospores and fusarioid asexual morph characterised by 0- or 1-septate macroconidia with blunt or hooked apical cell, lacking a foot-shaped basal cell.

Stylonectria corniculata Gräfenhan, Crous & Sand.-Den., *sp. nov.* MycoBank MB 838680. Fig. 47.

Etymology: From Latin *corniculum*, little horn. Referring to the shape of the conidiophores.

Typus: Germany, Brandenburg, Stolpe, near Gellmersdorfer Forst, from unidentified ascomycete on *Carpinus* sp., 1 Mar. 2007, T. Gräfenhan, **holotype** CBS H-24671, culture ex-type CBS 125491.

Conidiophores often as single phialides borne laterally on substrate and aerial hyphae, or irregularly branched and crowded with phialides produced laterally and terminally, hyaline, thin- and smooth-walled, 24–89 μ m long. **Conidiogenous cells** monophialidic, short doliiform, subcylindrical to subulate, 6–28.5 \times 2–3.5 μ m, often with a conspicuous flared collarette, periclinal thickening absent, producing micro- and macroconidia. **Microconidia** cylindrical to allantoid, hyaline, thin- and smooth-walled, 0(–1)-septate, (4.5–)6–13.5(–21) \times (1.5–)2–3 μ m (av. 9.7 \times 2.1 μ m). **Macroconidia** falcate, almost straight or gently

dorsiventrally curved, tapering toward the basal portion, 0(–)1-septate, with a blunt apical cell and obtuse basal cell, (20–)28–47(–56) \times 2–3.5 μ m (av. 37.6 \times 2.5 μ m). **Chlamydospores** and **sexual morph** not observed.

Culture characteristics: Colonies on PDA reaching 16–20 mm diam at 25 °C after 7 d. Surface at first white and membranous, becoming slimy, saffron to orange, to bright orange at the centre, flat, aerial mycelium absent, moisty at the centre, velvety at the margin, margin regular, filiform to undulate; reverse white, pale saffron to orange at centre. On OA, white to pale orange, flat, membranous to slimy, with regular and undulate margin; reverse pale luteous to pale saffron.

Notes: The species is here described based on its morphology *in vitro*, where only the asexual morph was obtained. This prevents further comparisons with known species of this genus. The only known collection, CBS 125491, has been shown to represent the most basal lineage in *Stylonectria* in previous phylogenetic studies (Gräfenhan *et al.* 2011, Lechat *et al.* 2015), which was confirmed here (Fig. 15). Although with neither a clear host association – an important character for species recognition in *Stylonectria* – nor any known sexual morphology, *St. corniculata* shows a distinctive morphology when it comes to its asexual morph, especially regarding the branching pattern and the shape of its mature conidiophores, which can be very elaborate and largely resemble antlers (Fig. 47).

Stylonectria hetmanica Akulov, Crous & Sand.-Den., *sp. nov.* MycoBank MB 838681. Fig. 48.

Etymology: The epithet refers to the Cossack Hetmanate (Ukrainian Hetmanščyna), the name of the former Cossack state territories where the type was collected.

Typus: Ukraine, Sumy, Okhtyrka, vicinities of Klymentove village, Hetmanskyi National Nature Park, on the ascomata of *Diaporthe* sp., associated with *Phomopsis* asexual morph, on

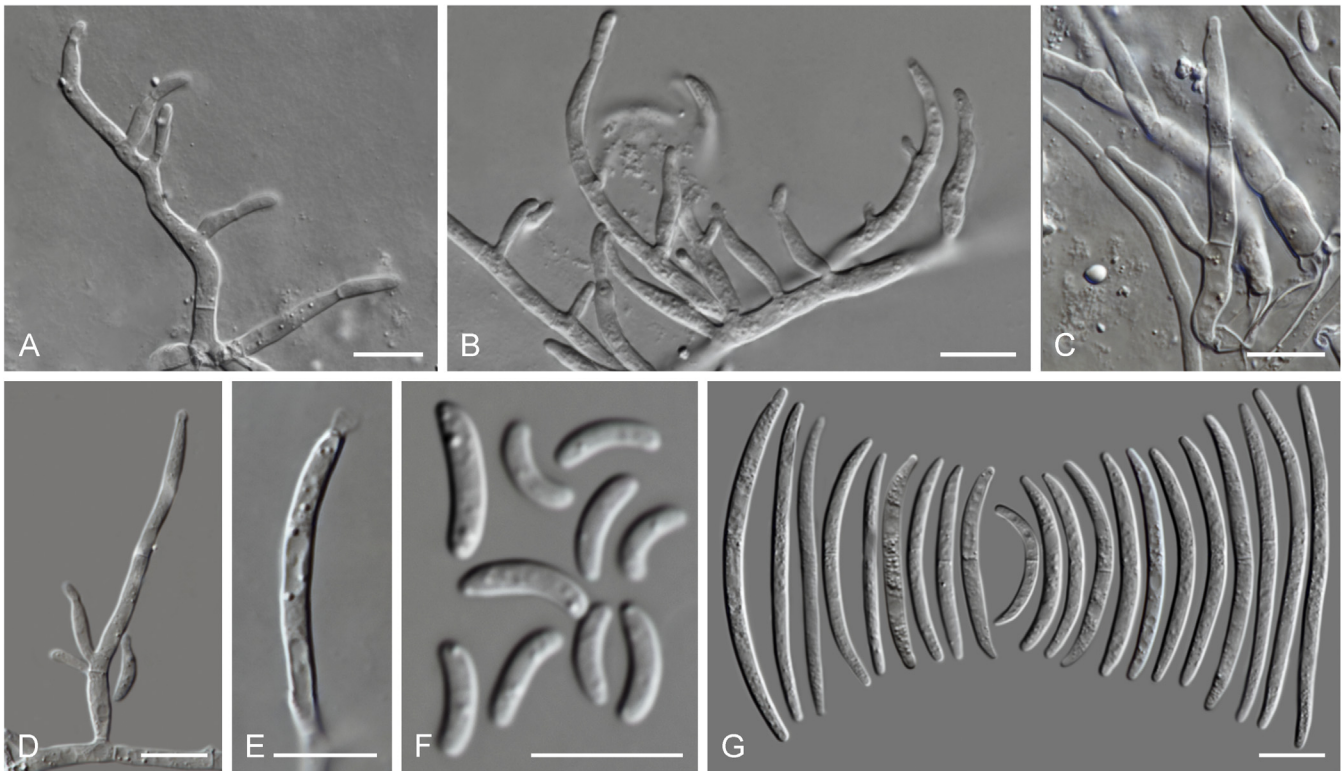


Fig. 47. *Stylonectria corniculata* (CBS 125491). A–E. Conidiophores and conidiogenous cells. F. Microconidia. G. Macroconidia. Scale bars = 10 μ m.

dead branches of *Frangula alnus* still attached to the tree, 13 Oct. 2019, Ya. Mieshkov, CWU (Myc) AS 7177, **holotype** CBS H-24672, culture ex-type CBS 147305 = CPC 38725.

Ascomata perithecial, gregarious or solitary, broadly pyriform, 220–310 μ m wide, with a distinctive flat and discoid papilla, 130–225 μ m wide, dark red, becoming darker in 3% KOH and light yellow in lactic acid. *Ascomatal wall* smooth, 30–45 μ m thick, composed of two regions: outer region 25–40 μ m thick, of irregularly shaped cells of *textura intricata* to *textura epidermoidea*; inner region 5–10 μ m thick of thin-walled, flattened cells of *textura prismatica* to *textura angularis*. *Asci* subcylindrical, 45–72 \times 4–8 μ m, 8-spored, apices rounded and simple, uniseriate or irregularly biseriate. *Ascospores* ellipsoidal, 1-septate, often constricted at septum, (7.5–) 8.5–11(–12.5) \times 3–4.5(–5.5) μ m, smooth to finely spinulose, thick-walled, hyaline at first, becoming pale golden brown at maturity. *Conidiophores* often as single phialides or short phialidic pegs borne laterally on the substrate and aerial hyphae, rarely irregularly to verticillately branched. *Conidiogenous cells* monophialidic, short doliform, subcylindrical to subulate, 4–21(–27.5) \times 2–3.5 μ m, often with a conspicuous flared collarette, periclinal thickening absent, producing micro- and macroconidia. *Microconidia* allantoid, hyaline, smooth- and thin-walled, 0(–1)-septate, (9–)10.5–13.5(–15) \times 2–3 μ m (av. 12 \times 2.4 μ m). *Macroconidia* subcylindrical to falcate, almost straight or moderately dorsiventrally curved, tapering towards both ends, 0–1(–2)-septate, apical cell blunt to slightly hooked, basal cell obtuse to poorly-developed, foot-shaped (11.5–) 16.5–28(–34) \times 2–3 μ m (av. 22.2 \times 2.5 μ m). *Chlamydospores* not observed.

Culture characteristics: Colonies on PDA reaching 2.5–3 mm diam at 25 $^{\circ}$ C after 7 d. Surface straw-coloured to

luteous, pale orange at centre, flat or radially folded, membranous to slimy, margin filiform to undulate; reverse pale luteous to pale orange. On OA orange to pale apricot, flat, membranous to slimy, margin filiform with abundant submerged mycelium; reverse pale orange.

Additional material examined: **Ukraine**, Sumy, Okhtyrka, in the vicinities of the village Klymentove, Hetmanskyi National Nature Park, on the conidiomata of *Dothiorella sarmentorum*, on recently dead branches of *Acer platanoides* still attached to the tree, 13 Oct. 2019, A. Akulov, CWU (Myc) AS 7278, culture CBS 147306 = CPC 38848.

Notes: The morphological description of *St. hetmanica* is based on its growth on OA, where both studied strains showed optimal growth and sporulation. Contrary to most fusarioid genera, *St. hetmanica* grows very poorly and fails to sporulate on SNA and WA. *Stylonectria hetmanica* is morphologically comparable and genetically close to *St. purtonii*, *St. norvegica*, and *St. wegeliiana*. Nevertheless, ascospores of *St. hetmanica* are smaller than those of *St. purtonii* and *St. wegeliiana*. Additionally, macroconidia of *St. hetmanica*, while similar in size to those of *St. purtonii*, are less septate (0- or 1-septate, rarely 2-septate in *St. hetmanica*, and up to 3-septate in *St. purtonii*). The sexual morph of the recently described *St. norvegica* is very similar to that of *St. hetmanica*, although both species are genetically less closely related. The latter species can be distinguished by the production of shorter macroconidia.

Thelonectria P. Chaverri & C. Salgado, Stud. Mycol. 68: 76. 2011. Fig. 8.

Type species: *Thelonectria discophora* (Mont.) P. Chaverri & C. Salgado, Stud. Mycol. 68: 76. 2011.

Basionym: *Sphaeria discophora* Mont., Ann. Sci. Nat. Bot., sér. 2, 3: 353. 1835.

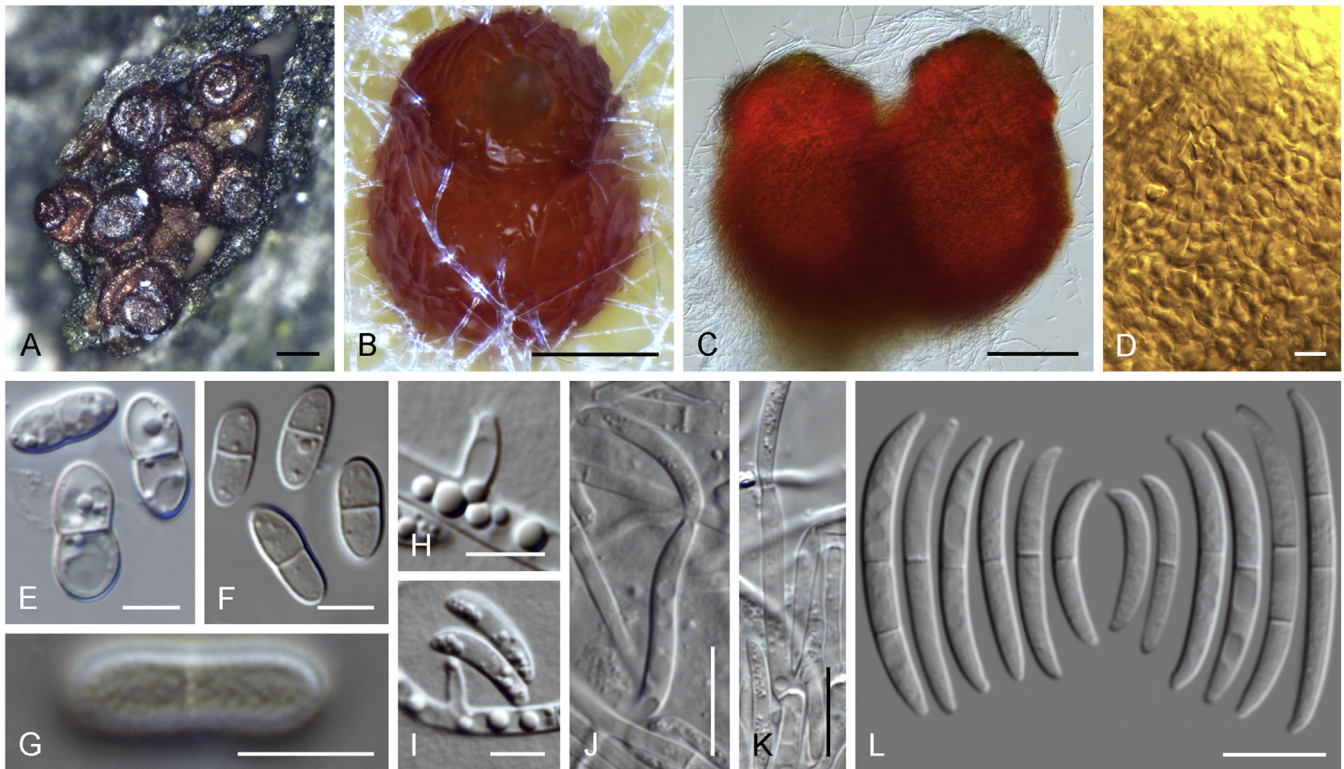


Fig. 48. *Styonectria hetmanica* (CBS 147305). **A–C.** Ascomata (**A.** On natural substrate. **B.** **C.** In culture). **D.** Surface view of perithecial wall in lactic acid. **E–G.** Ascospores (**G.** Surface view). **H–K.** Conidiophores and conidiogenous cells. **L.** Macroconidia. Scale bars: **A–C** = 100 μ m; **E–I** = 5 μ m; all others = 10 μ m.

Synonyms: *Nectria discophora* (Mont.) Mont., Fl. Chil. 7: 454. 1850.

Cucurbitaria discophora (Mont.) Kuntze, Revis. Gen. Pl. 3: 461. 1898.

Neonectria discophora (Mont.) Mantiri & Samuels, Canad. J. Bot. 79: 339. 2001.

Nectria tasmanica Berk., in Hooker, Bot. Antarct. Voy. III, Fl. Tasman. 2: 279. 1860.

Cucurbitaria tasmanica (Berk.) Kuntze, Revis. Gen. Pl. 3: 462. 1898.

Nectria umbilicata Henn., Hedwigia 41: 3. 1902.

Creonectria discostiolata Chardón, Bol. Soc. Venez. Ci. Nat. 5: 341. 1939.

Ascomata perithecial, solitary to gregarious, non-stromatic or sometimes seated on an immersed inconspicuous stroma, superficial, globose to subglobose or pyriform to elongated, orange to red, with prominent areolate papilla or darkly pigmented apex, smooth to slightly rugulose, lacking hairs or appendages. *Ascomatal wall* of two regions: outer region of thick-walled, pigmented cells forming a *textura epidermoidea*; inner region of elongate, hyaline, thin-walled cells, becoming thinner towards the centrum. *Asci* cylindrical to narrowly clavate, 8-spored, with an apical ring, uniseriate. *Ascospores* ellipsoidal to fusoid, 1-septate, hyaline, smooth or finely spinulose or striate. *Sporodochia* not formed. *Conidiophores* mononematous, hyaline, septate, irregularly branched, terminating in 1–3 phialides or reduced to lateral phialides. *Conidiogenous cells* monophialidic, cylindrical or slightly swollen, tapering towards the apex, with periclinal thickening and flared collarettes, producing usually macroconidia. *Microconidia* rarely formed, globose to ovoid, hyaline, aseptate, with displaced inconspicuous hilum. *Macroconidia* subcylindrical to slightly fusoid, curved, broadest at upper third, (3–)5–7(–9)-septate, with rounded ends or flattened at the basal cell. *Chlamydospores* unknown.

[Description adapted from [Chaverri et al. 2011](#)].

Diagnostic features: Orange to red, mostly smooth-walled perithecia with prominent darkened papilla producing cylindrical to narrowly clavate asci bearing ellipsoidal to fusoid, 1-septate ascospores and cylindrocarpon-like asexual morph.

Tumenectria C. Salgado & Rossman, Fungal Diversity 80: 451. 2016. **Fig. 8.**

Type species: *Tumenectria laetidisca* (Rossman) C. Salgado & Rossman, Fungal Diversity 80: 451. 2016.

Basionym: *Nectria laetidisca* Rossman, Mycol. Pap. 150: 36. 1983.

Synonym: *Cylindrocarpon bambusicola* Matsush., Matsush. Mycol. Mem. 5: 9. 1987.

Ascomata perithecial, mostly solitary to gregarious, non-stromatic, superficial, broadly pyriform, not collapsing when dry, orange to sienna, turning blood red in KOH, pigment dissolving in lactic acid, broadly rounded to flattened papilla, smooth-walled, lacking hairs and appendages. *Ascomatal wall* of two regions: outer region of thick-walled, pigmented cells forming a *textura angularis*; inner region of elongate, hyaline, thin-walled cells, becoming thinner towards the centrum. *Asci* narrowly clavate, apex simple, 8-spored, lacking an apical ring, irregularly multiseriate. *Ascospores* fusoid, 3-septate, hyaline, smooth or finely spinulose. *Sporodochia* not formed. *Conidiophores* simple, mononematous, straight to flexuous, hyaline, septate, unbranched or rarely branched, terminating in a single phialide or reduced to lateral phialides. *Conidiogenous cells* monophialidic, cylindrical or slightly swollen, tapering towards the apex, with periclinal thickening and flared collarettes. *Microconidia* not formed. *Macroconidia* cylindrical to slightly fusoid, straight to slightly curved, 3–6-septate, with rounded ends. *Chlamydospores* unknown.

[Description adapted from [Rossman \(1983\)](#) and [Salgado-Salazar et al. \(2016\)](#)].

Diagnostic features: Orange to sienna, smooth-walled perithecia with broadly rounded to flattened papilla producing narrowly clavate asci bearing fusoid, 3-septate phragmo-ascospores and cylindrocarpon-like asexual morph.

FUSARIUM AND ALLIED GENERA: LIST OF ACCEPTED NAMES

The following nomenclator lists names that have been introduced in *Fusarium* up to January 2021, as well as their current status (with accepted names indicated in bold and underlined for easier recognition). Where type specimens have been located, these details, as well as any ex-type cultures and diagnostic DNA barcode data are provided, along with notes regarding potential synonymy. This list will be updated and republished at regular intervals, and will form the basis for a monograph of *Fusarium* and allied genera that will be freely available on www.Fusarium.org.

aberrans Fusarium J.W. Xia et al., *Persoonia* 43: 192. 2019.

Holotypus: CBS H-24050.

Ex-type culture: CBS 131385.

Type locality: **Australia**, Northern Territory, Roper River area.

Type substrate: Stem of *Oryza australiensis*.

Descriptions and illustrations: See [Xia et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb2*: MN170378; *tef1*: MN170445.

acaciae Fusarium Berl. & Voglino, *Syll. Fung., Addit. I–IV*: 201. 1886, *nom. illegit.*, Art. 53.1.

(See *Fusarium acaciae* Cooke & Harkn.)

acaciae Fusarium Cooke & Harkn., *Grevillea* 12: 96. 1884.

Synonyms: ?*Fusarium acaciae* Berl. & Voglino, *Syll. Fung., Addit. I–IV*: 201. 1886, *nom. illegit.*, Art. 53.1.

?*Fusarium acaciae* Sacc., *Syll. Fung.* 9: 958. 1891, *nom. illegit.*, Art. 53.1.

(See ***Fusarium lateritium***)

Holotypus: ?BPI 451718.

Type locality: **USA**, California.

Type substrate: Stem of *Acacia* sp.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

acaciae Fusarium Sacc., *Syll. Fung.* 9: 958. 1891, *nom. illegit.*, Art. 53.1.

(See *Fusarium acaciae* Cooke & Harkn.)

acaciae-mearnsii Fusarium O'Donnell et al., *Fungal Genet. Biol.* 41: 619. 2004.

Holotypus: BPI 843477.

Ex-type culture: CBS 110254 = MRC 5120 = NRRL 25754.

Type locality: **South Africa**, KwaZulu-Natal, Pietermaritzburg.

Type substrate: *Acacia mearnsii*.

Descriptions and illustrations: See [O'Donnell et al. \(2004\)](#).

Diagnostic DNA barcodes: *rpb1*: JAAWUD010000100; *rpb2*: JAAWUD010000080; *tef1*: AF212448.

acicola Fusarium Bres., in Strasser, *Verh. Zool.-Bot. Ges. Wien* 60: 328. 1910.

Holotypus: Not located.

Type locality: **Austria**, Sonntagberg.

Type substrate: Rotting needles of *Pinus* sp.

Descriptions and illustrations: See [Strasser \(1910\)](#).

Note: Status unclear. Not *Fusarium* *fide* [Wollenweber & Reinking \(1935\)](#).

acremoniopsis Fusarium Vincens, *Bull. Soc. Mycol. France* 31: 26. 1915.

(See ***Fusarium larvarum***)

Holotypus: ?PC.

Type locality: **Brazil**, Pará, Belém.

Type substrate: *Agrotis* sp. (cutworm).

Descriptions and illustrations: See [Vincens \(1915\)](#).

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

acidiorum Fusarium (Trab.) Brongn. & Delacr., *Bull. Séances Soc. Natl. Agric. France* 51: 631. 1891.

Trichothecium acidiorum (Trab.) Madelin, *Trans. Brit. Mycol. Soc.* 49: 284. 1966.

Basionym: *Botrytis acidiorum* Trab., *Compt. Rend. Hebd. Séances Acad. Sci.* 112: 1383. 1891.

Synonym: *Lachnidium acidiorum* Giard, *Compt. Rend. Hebd. Séances Acad. Sci.* 112: 1520. 1891.

Holotypus: Not located.

Type locality: **Algeria**.

Type substrate: *Acrididae* (locust).

Description and illustrations: See [Madelin \(1966\)](#).

acuminatum Fusarium Ellis & Everh., *Proc. Acad. Nat. Sci. Philadelphia* 47: 441. 1895.

Synonyms: *Microcera acuminata* (Ellis & Everh.) Höhn., in Weese, *Sitzungsber. Akad. Wiss. Wien, Math.-Naturwiss. Kl., Abt. 1.* 128: 729. 1919.

Fusarium scirpi var. *acuminatum* (Ellis & Everh.) Wollenw., *Fusaria Autogr. Delin.* 3: 930. 1930.

Fusarium scirpi subsp. *acuminatum* (Ellis & Everh.) Raillo, *Fungi of the Genus Fusarium*: 177. 1950.

Fusarium gibbosum var. *acuminatum* (Ellis & Everh.) Bilař, *Mikrobiol. Zhurn.* 49: 6. 1987.

?*Selenosporium hippocastani* Corda, *Icon. Fung.* 2: 7. 1838.

Fusarium hippocastani (Corda) Sacc., *Syll. Fung.* 4: 703. 1886.

Fusarium erubescens Appel & Oven, *Landwirtsch. Jahrb.* 1905, *nom. illegit.*, Art. 53.1.

Fusarium caudatum Wollenw., *J. Agric. Res.* 2: 262. 1914.

Fusarium scirpi var. *caudatum* (Wollenw.) Wollenw., *Fusaria Autogr. Delin.* 3: 934 & 935. 1930.

Fusarium equiseti var. *caudatum* (Wollenw.) Joffe, *Mycopathol. Mycol. Appl.* 53(1–4): 220. 1974.

Fusarium arcuosporum Sherb., *Mem. Cornell Univ. Agric. Exp. Sta.* 6: 186. 1915.

Fusarium ferruginosum Sherb., *Mem. Cornell Univ. Agric. Exp. Sta.* 6: 190. 1915.

Fusarium sanguineum Sherb., *Mem. Cornell Univ. Agric. Exp. Sta.* 6: 193. 1915.

Fusarium lanceolatum O.A. Pratt, *J. Agric. Res.* 13: 83. 1918.

Fusarium pseudoeffusum Murashk., *Arb. Landwirtsch. Akad. Omsk.* 3: 19. 1924.

Fusarium moronei Curzi, *Revista Biol. (Lisbon)* 10: 141. 1928.

Fusarium russianum Manns, *Bull. N. Dakota Agric. Exp. Sta.* 259: 34. 1932.

Gibberella acuminata Wollenw., *Fusarien*: 68. 1935.

Spermospora oryza M. Rao, *Sci. & Cult.* 32: 94. 1966.

Gibberella acuminata C. Booth, *The Genus Fusarium*: 161. 1971, *nom. illegit.*, Art. 53.1.

Holotypus: NY00928689.

Type locality: **USA**, New York, Geneva.

Type substrate: *Solanum tuberosum*.

Descriptions and illustrations: See Sherbakoff (1915), Booth (1971), Gerlach & Nirenberg (1982), Burgess & Summerell (2000) and Leslie & Summerell (2006).

Notes: *Fusarium acuminatum* is an established name in the *Fusarium* literature, but it lacks living type material to confirm its taxonomic position. Although an older epithet, based on *Seleosporium hippocastani*, could be used, we refrain from providing a new combination for this well-known species due to a lack of DNA-based evidence to support this combination. Moreover, Holubová-Jechová *et al.* (1994) could not locate any holotype material for *S. hippocastani*, abstaining from introducing a neotype, which they argued would cause nomenclatural instability, a view we fully support.

acutum Fusarium Nirenberg & O'Donnell, Persoonia 46: 144. 2021.

Synonym: *Fusarium acutum* Nirenberg & O'Donnell, Mycologia 90: 435. 1998, *nom. inval.*, Art. 40.1.

Holotypus: B 70 0001695.

Ex-type culture: BBA 69580 = CBS 402.97 = FRC 0-1117 = IMI 376110 = NRRL 13309.

Type locality: **India**.

Type substrate: Unknown.

Descriptions and illustrations: See Nirenberg & O'Donnell (1998) and Yilmaz *et al.* (2021).

Diagnostic DNA barcodes: *rpb1*: MT010947; *rpb2*: KT154005; *tef1*: KR071754.

acutisporum Fusarium (Sand.-Den. & Crous) O'Donnell *et al.*, Index Fungorum 440: 1. 2020.

Neocosmospora acutispora Sand.-Den. & Crous, Persoonia 43: 108. 2019.

Holotypus: CBS H-23969.

Ex-type culture: BBA 62213 = CBS 145461 = NRRL 22574.

Type locality: **Guatemala**.

Type substrate: *Coffea arabica*

Descriptions and illustrations: See Sandoval-Denis *et al.* (2019).

Diagnostic DNA barcodes: *rpb1*: MW834210; *rpb2*: LR583814; *tef1*: LR583593.

aderholdii Fusarium Osterw., Bericht Schweiz. Versuchsanst. Obst-, Wein- und Gartenbau 1913/14: 519. 1915.

Ilyonectria destructans (Zinssm.) Rossman *et al.*, Stud. Mycol. 80: 217. 2015.

Basionym: *Ramularia destructans* Zinssm., Phytopathology 8: 570. 1918.

Synonyms: *Cylindrocarpon destructans* (Zinssm.) Scholten, Netherlands J. Plant Pathol. 70 suppl. (2): 9. 1964.

Fusarium polymorphum Marchal, Bull. Soc. Roy. Bot. Belgique 34: 145. 1895, *nom. illegit.*, Art. 53.1.

Cylindrocarpon radicolica Wollenw., Fusaria Autogr. Delin. 2: 651. 1924.

Nectria radicolica Gerlach & L. Nilsson, Phytopathol. Z. 48: 255. 1963.

Neonectria radicolica (Gerlach & L. Nilsson) Mantiri & Samuels, Canad. J. Bot. 79: 339. 2001.

Ilyonectria radicolica (Gerlach & L. Nilsson) P. Chaverri & C.G. Salgado, Stud. Mycol. 68: 71. 2011.

?*Fusarium rhizogenum* Aderh., Centralbl. Bacteriol. Parasitenk., 2. Abth., 6: 623. 1900, *nom. illegit.*, Art. 53.1.

?*Septocylindrium radicolica* Aderh., Centralbl. Bacteriol. Parasitenk., 2. Abth., 6: 623, 1900, *nom. illegit.*, Art. 53.1.

?*Septocylindrium aderholdii* Sacc & P. Syd., Syll. Fung. 16: 1048. 1902.

Holotypus: Not located.

Type locality: **Germany**.

Type substrate: Unknown.

Notes: Synonymy *vide* Wollenweber & Reinking (1935). Although older epithets are available for *Ilyonectria destructans*, we refrain from providing a new combination for this well-known species due to a lack of DNA-based evidence to support this combination.

adesmiae Fusarium Henn., Hedwigia 36: 246. 1897.

Synonym: *Ramularia adesmiae* (Henn.) Wollenw., Fusaria Autogr. Delin. 1: 466. 1916.

Holotypus: In B *vide* Hein (1988).

Type locality: **Chile**, Bío-Bío Province.

Type substrate: *Adesmia* sp.

Note: Status unclear, not *Ramularia* *vide* Braun (1998).

aduncisporum Fusarium Weimer & Harter, J. Agric. Res. 32: 312. 1926.

(See *Fusarium solani*)

Lectotypus: BPI 451321, designated in Sandoval-Denis *et al.* (2019).

Lectotype locality: **USA**, California, Ventura.

Lectotype substrate: Stems of *Melilotus alba*.

Note: Synonym *vide* Wollenweber & Reinking (1935).

aecidii-tussilaginis Fusarium Allesch., Ber. Bot. Vereines Landshut 12: 131. 1892.

(See ***Fusarium avenaceum***)

Holotypus: In M.

Type locality: **Germany**, Oberammergau.

Type substrate: *Aecidium tussilaginis*.

Note: Synonym *vide* Wollenweber & Reinking (1935).

aeruginosum Fusarium Delacr., Bull. Soc. Mycol. France 7: 110. 1891.

(See ***Fusarium caeruleum***)

Lectotypus (*hic designatus*, MBT 10000648): **France**, Paris, from *Solanum tuberosum*, April 1891, G. Delacroix, Bull. Soc. Mycol. France 7: pl. VIII, fig. h.

Notes: Synonym *vide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

aethiopicum Fusarium O'Donnell *et al.*, Fungal Genet. Biol. 45: 1521. 2008.

Holotypus: BPI 878409.

Ex-type culture: CBS 122858 = NRRL 46726.

Type locality: **Ethiopia**, Bure district, west Gojjam zone of Amhara region.

Type substrate: *Triticum aestivum*.

Descriptions and illustrations: See O'Donnell *et al.* (2008b).

Diagnostic DNA barcodes: *rpb1*: MW233298; *rpb2*: MW233470; *tef1*: FJ240298.

affine Fusarium Fautrey & Lambotte, Rev. Mycol. (Toulouse) 18: 68. 1896.

Syntypes: ILL00221136 (Roumeguère, Fungi Sel. Gall. Exs. no. 6927) & ILL00221137 (Roumeguère, Fungi Sel. Gall. Exs. no. 6928).

Type locality: **France**.

Type substrate: *Solanum tuberosum*.

Notes: Booth (1971) examined the *exsiccatae* (Fung. Sel. Gall. Exs., No. 6927 & 6928) of *F. affine* and found that one part (no. 6927) is *F. solani* and the other part (no. 6928) represented another fungus that was interpreted as *Hymenula affinis* by Wollenweber (1916–1935). Booth (1971) indicated that *F. affine* might be a possible synonym of *F. tabacinum*, which is now regarded as *Plectosphaerella cucumerina* (Palm et al. 1995, Giraldo & Crous 2019). However, both Gams & Gerlach (1968) and Palm et al. (1995) considered *F. affine* as a misapplied synonym of *P. cucumerina*. Sherbakoff (1915) also treated the fungus as *F. affine*, which was later reinterpreted as *Septomyxa affine* by Wollenweber (1916–1935). Therefore, the current status of *F. affine* is uncertain and requires further investigation.

agapanthi Fusarium O'Donnell et al., Mycologia 108: 987. 2016.

Holotypus: VPRI 41777.

Ex-type culture: NRRL 54463 = VPRI 41777.

Type locality: **Australia**, Victoria, Melbourne, Royal Botanic Gardens.

Type substrate: *Agapanthus praecox*.

Descriptions and illustrations: See Edwards et al. (2016).

Diagnostic DNA barcodes: *rpb1*: KU900620; *rpb2*: KU900625; *tef1*: KU900630.

agaricorum Fusarium Sarrazin, Rev. Mycol. (Toulouse) 9: 170. 1887.

Lectotypus (hic designatus, MBT 10000649): **France**, on the cap of *Psalliota campestris* (syn. *Agaricus campestris*), 1887, F. Sarrazin, ILL00218415 (Roumeguère, Fungi Sel. Gall. Exs. no. 4298).

Notes: Status unclear. Not *Fusarium* fide Wollenweber & Reinking (1935).

ailanthinum Fusarium Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 6: 350. 1899.

(See **Fusarium lateritium**)

Holotypus: In LPS (Fungi Argent. n.v.c. #864).

Type locality: **Argentina**.

Type substrate: Trunk and branches of *Ailanthus glandulosa*.

Note: Synonym fide Wollenweber & Reinking (1935).

alabamense Fusarium Sacc., Syll. Fung. 4: 722. 1886, nom. illegit., Art. 52.1.

Synonym: *Fusarium erubescens* Berk. & M.A. Curtis, Grevillea 3: 98. 1875.

Holotypus: ?K(M).

Type locality: **USA**, Alabama, Beaumont.

Type substrate: Bark.

Notes: Status unclear. Not *Fusarium* fide Wollenweber & Reinking (1935).

albedinis Fusarium Kill. & Maire ex Malençon, Compt. Rend. Hebd. Séances Acad. Sci. 198: 1261. 1934, nom. inval., Art. 6.10.

Synonym: *Cylindrophora albedinis* Kill. & Maire, Bull. Soc. Hist. Nat. Afrique N. 21: 97. 1930, nom. inval., Art. 36.1.

(See **Fusarium oxysporum**)

Authentic material: Not located.

Original locality: Indicated as 'oasis in Sahara'.

Original substrate: Dead trunk and leaf of *Phoenix dactylifera*.

Note: Synonym fide Booth (1971).

albertii Fusarium Roum., Fungi Sel. Gall. Exs., Cent. 19: no. 1867. 1881, nom. inval., Art. 38.1(a).

(See **Fusarium lateritium**)

Authentic material: BR5020140140720.

Original locality: **France**.

Original substrate: Petiole of *Ziziphus volubilis*.

Note: Synonym fide Wollenweber & Reinking (1935).

albidoviolaceum Fusarium Dasz. (as 'albido-violaceum'), Bull. Soc. Bot. Genève, sér. 2, 4: 293. 1912.

(See **Fusarium oxysporum**)

Lectotypus (hic designatus, MBT 10000650): **Switzerland**, Geneva, from soil, 1912, W. Daszewska, Bull. Soc. Bot. Genève, 2 sér. 4: 292, fig. 15.

Notes: Wollenweber (1916–1935; Fusaria Autogr. Delin. 1: 361) indicated that the living ex-type culture was lodged in the laboratory of W.C. Scholten in Amsterdam, which in turn has been accessioned into the CBS. However, no record or culture can be located in the CBS collection. Therefore, an illustration accompanying the original protologue is designated as lectotype here.

albidum Fusarium (Rossman) O'Donnell & Geiser, Phytopathology 103: 404. 2013.

Luteonectria albida (Rossman) Sand.-Den. & L. Lombard, Stud. Mycol. 98 (no. 100116): 60. 2021.

Basionym: *Nectria albida* Rossman, Mycol. Pap. 150: 79. 1983.

Synonym: *Albonectria albida* (Rossman) Guu & Y.M. Ju, Bot. Stud. (Taipei) 48: 189. 2007.

Holotypus: CUP-MJ 942.

Ex-type culture: ATCC 44543 = BBA 67603 = CTR 71-110 = NRRL 13950 = NRRL 22152.

Type locality: **Jamaica**, Hanover Parish, Dolphin Head Mountain, near Askenish.

Type substrate: Erumpent through thin bark of woody stem.

Diagnostic DNA barcode: *rpb1*: JX171492; *rpb2*: HQ897738; *tef1*: MW834283.

albiziae Fusarium Woron., Vestn. Tiflissk. Bot Sada 48: 34. 1920.

(See *Fusarium merismoides*)

Syntypes: BPI 451733, BPI 451734 & CUP-017160.

Type locality: **Georgia**, Batumi, Adjara.

Type substrate: *Albizia julibrissin*.

Notes: Synonym fide Wollenweber & Reinking (1935). Lectotypification requires further investigation of the syntypes.

albobarboneum Fusarium (Cooke & Harkn.) Sacc., Syll. Fung. 4: 720. 1886.

Basionym: *Fusidium albocarboneum* Cooke & Harkn., Grevillea 9: 129. 1881.

Syntype: BPI 408577.

Type locality: **USA**, California, San Francisco, San Francisco Odd Fellows Cemetery.

Type substrate: *Eucalyptus* sp.

Notes: The generic name *Cylindrocarpon* (= *Neonectria*; Rossman et al. 2013) was conserved over *Fusidium*, making the latter generic name a *nom. rej.* (Art. 14.1, 14.6 & 14.7). Therefore, *Fusidium albocarboneum* should be transferred to *Neonectria* after further investigation. Lectotypification requires further investigation of the syntype.

albosuccineum Fusarium (Pat.) O'Donnell & Geiser, Phytopathology 103: 404. 2013.

Albonectria albosuccinea (Pat.) Rossman & Samuels, Stud. Mycol. 42: 107. 1999.

Basionym: *Calonectria albosuccinea* Pat., Bull. Soc. Mycol. France 8: 132. 1892.

Synonyms: *Nectria albosuccinea* (Pat.) Rossman, Mycotaxon 8: 487. 1979.

Calonectria ecuadorica Petrak, Sydowia 4: 463. 1950.

Holotypus: In FH *fide* Rossman (1983).

Type locality: Ecuador, Puente Chimbo.

Type substrate: Bark.

album *Fusarium* Sacc., Michelia 1: 82. 1877.

Neonectria punicea (J.C. Schmidt) Castl. & Rossman, Canad. J. Bot. 84: 1425. 2006.

Basionym: *Sphaeria punicea* J.C. Schmidt, in Schmidt & Kunze, Mykol. Hefte 1: 61. 1817.

Synonyms: *Nectria punicea* (J.C. Schmidt) Fr., Summa Veg. Scand. 2: 387. 1849.

Cucurbitaria punicea (J.C. Schmidt) Kuntze, Revis. Gen. Pl. 3: 461. 1898.

Cylindrocarpon album (Sacc.) Wollenw., Fusaria Autogr. Delin. 1: no. 473. 1916.

Nectria punicea f. ilicicola Rehm, Ascomyceten: no. 337. 1876.

Nectria punicea var. *ilicis* C. Booth, Mycol. Pap. 73: 54. 1959.

Cylindrocarpon album var. *majus* Wollenw., Z. Parasitenk. (Berlin) 1: 154. 1928.

Fusarium album var. *abietinum* Beeli, Bull. Soc. Roy. Bot. Belgique 62: 131. 1930.

Holotypus: Not located.

Type locality: Italy.

Type substrate: Bark of *Pinus* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

aleurinum *Fusarium* Ellis & Everh., Bull. Torrey Bot. Club 24: 476. 1897.

(See ***Fusarium avenaceum***)

Syntypes: In BPI, BRU, CLEMS, CUP, F, FLAS, ILL, ILLS, ISC, MICH, MSC, MU, NCU, NEB, OSC, PH, PUL, UC, WIS & WSP.

Type locality: USA, West Virginia, Fayette County Nuttallburg, south of Edmond.

Type substrate: Wheat flour that had been on the ground for four months.

Notes: Synonym *fide* Wollenweber & Reinking (1935). Lectotypification requires further investigation of the syntypes.

aleyrodis *Fusarium* Petch, Trans. Brit. Mycol. Soc. 7: 164. 1921.

Lectotypus (*hic designatus*, MBT 10000651): USA, Florida, Sutherland, from *Aleyrodes citri*, 13 Sep. 1907, F. Wills, in Petch 1921, Trans. Brit. Mycol. Soc. 7, pl. V, fig. 12.

Notes: Wollenweber & Reinking (1935) considered this species as a synonym of *F. scirpi*. However, based on the descriptions and illustrations provided by Fawcett (1908) and Petch (1920), this species belongs to the genus *Microcera*, which is also in agreement with its aetiology. Therefore, a new combination will presumably be required after further investigation.

algeriense* *Fusarium Laraba & O'Donnell, Mycologia 109: 944. 2017.

Holotypus: BPI 910347.

Ex-type culture: CBS 142638 = NRRL 66647.

Type locality: Algeria, Guelma Province, Djeballah Khemissi.

Type substrate: *Triticum durum*.

Descriptions and illustrations: See Laraba *et al.* (2017).

Diagnostic DNA barcodes: *rpb1*: MF120488; *rpb2*: MF120499; *tef1*: MF120510.

alkanophilum* *Fusarium Palacios-Prü & V. Marcano, Rev. Ecol. Latinoamer. 8: 5. 2001.

Holotypus: EMC, Palacios-Prü, 3 April 1998.

Type locality: Venezuela, Merida State, south of Sierra La Culata, Valle de San Javier, Los Pinos.

Type substrate: Beetle immersed in kerosene.

Descriptions and illustrations: See Marcano *et al.* (2001).

Note: No living type material could be located.

allescheri *Fusarium* Sacc. & P. Syd., Syll. Fung. 14: 1128. 1899.

Replaced synonym: *Fusarium glandicola* Allesch., Ber. Bot. Vereines Landshut 12: 130. 1892, *nom. illegit.*, Art. 53.1; *non* Cooke & W.R. Gerard 1878.

(See *Fusarium melanochlorum*)

Holotypus: In M.

Type locality: Germany, München.

Type substrate: *Quercus pedunculata*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

allescherianum *Fusarium* Henn., Verh. Bot. Vereines Prov. Brandenburg 40: 175. 1899.

Synonyms: *Gloeosporium allescherianum* (Henn.) Wollenw., Fusaria Autogr. Delin. 1: 495. 1916.

?*Fusarium personatum* Cooke, in Harkness, Grevillea 7: 12. 1878.

Holotypus: In B *fide* Hein (1988).

Type locality: Germany.

Type substrate: Leaves of *Ocotea foetens*.

Notes: Status unclear. The taxonomic status of *Gloeosporium allescherianum* is questionable. Furthermore, there is no DNA-based evidence linking *F. allescherianum* to *F. personatum* although Wollenweber & Reinking (1935) considered them both synonyms under *G. allescherianum*.

allii-sativi *Fusarium* Allesch., Ber. Bot. Vereines Landshut 12: 131. 1892.

(See *Fusarium solani*)

Holotypus: In M.

Type locality: Germany, Unterammergau.

Type substrate: *Allium sativum*.

alluviale *Fusarium* Wollenw. & Reinking, Phytopathology 15: 167. 1925.

(See *Fusarium solani*)

Holotypus: Not located.

Type locality: Honduras.

Type substrate: Alluvial soil.

aloes *Fusarium* Kalchbr. & Cooke, Grevillea 9: 23. 1880.

(See ***Fusarium scirpi***)

Holotypus: ?K(M).

Type locality: South Africa, Eastern Cape Province, Somerset East.

Type substrate: *Aloe arborescens*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

ambrosium *Fusarium* (Gadd & Loos) Agnihotr. & Nirenberg, Stud. Mycol. 32: 98. 1990.

Neocosmospora ambrosia (Gadd & Loos) L. Lombard & Crous, Stud. Mycol. 80: 227. 2015.

Basionym: *Monacrosporium ambrosium* Gadd & Loos, Trans. Brit. Mycol. Soc. 31: 17. 1947.

Synonyms: *Dactylella ambrosia* (Gadd & Loos) K.Q. Zhang *et al.*, Mycosystema 7: 112. 1995.

Fusarium bugnicourtii Brayford, Trans. Brit. Mycol. Soc. 89: 350. 1987.

Lectotypus: Trans. Brit. Mycol. Soc. 31: 16, Text-fig. 5. 1947, designated by [Aoki et al. \(2018\)](#).

Lectotype locality: **Sri Lanka**.

Lectotype substrate: Gallery of *Euwallacea fornicatus* infesting *Camellia sinensis*.

Epitypus: BPI 910524, designated by [Aoki et al. \(2018\)](#).

Ex-epitype culture: BBA 65390 = CBS 571.94 = NRRL 22346 = MAFF 246287.

Epitype locality: **India**, Upasi Tea Institute.

Epitype substrate: Gallery of *Euwallacea fornicatus* infesting *Camellia sinensis*.

Diagnostic DNA barcodes: *rpb1*: KC691587; *rpb2*: EU329503; *tef1*: FJ240350

amenti Fusarium Rostr., Bot. Tidsskr. 14: 240. 1885.

(See *Fusarium avenaceum*)

Holotypus: F-604398 in UPS.

Type locality: **Denmark**, Fyn, Holmdrup.

Type substrate: *Salix cinerea*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

amentorum Fusarium Lacroix, Fl. Maine-et-Loire 2 (Suppl.): [1]. 1854.

(See *Fusarium avenaceum*)

Lectotypus (hic designatus, MBT 10000652): **France**, St. Romain-sur-Vienne, from *Salix cinerea*, date unknown, J.B.H.J. Desmazières, BR5020140143752.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

amethysteum Fusarium P. Crouan & H. Crouan, Fl. Finistère: 14. 1867.

Holotypus: Not located.

Type locality: **France**.

Type substrate: Dead stem of *Urtica* sp.

Notes: Status unclear. Not *Fusarium* *fide* [Wollenweber & Reinking \(1935\)](#).

ampelodesmi Fusarium Fautrey & Roum., in Roumeguère, Rev. Mycol. (Toulouse) 13: 82. 1891.

(See *Fusarium reticulatum*)

Syntype: ILL00219841 (Roumeguère, Fungi Sel. Gall. Exs. no. 5687).

Type locality: **France**, Jardin de Noidan.

Type substrate: *Ampelodesmos tenax*

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#). Lectotypification requires further investigation of the syntype.

amplum Fusarium (Sand.-Den. & Crous) O'Donnell et al., Index Fungorum 440: 1. 2020.

Neocosmospora ampla Sand.-Denis & Crous, Persoonia 43: 110. 2019.

Holotypus: CBS H-23970.

Ex-type culture: BBA 4170 = CBS 202.32.

Type locality: **German East Africa**.

Type substrate: *Coffea* sp.

Descriptions and illustrations: See [Sandoval-Denis et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW834212; *rpb2*: LR583815; *tef1*: LR583594.

ananatum Fusarium A. Jacobs et al., Fung. Biol. 114: 522. 2010.

Holotypus: PREM 58713.

Ex-type culture: CBS 118516 = CMW 18685 = FCC 2986 = MRC 8165.

Type locality: **South Africa**, KwaZulu-Natal Province, Hluhluwe.

Type substrate: *Ananas comosus*.

Descriptions and illustrations: See [Jacobs et al. \(2010\)](#).

Diagnostic DNA barcodes: *rpb1*: MT010937; *rpb2*: LT996137; *tef1*: MT010996.

andinum Fusarium Syd., Ann. Mycol. 37: 437. 1939.

Holotypus: S-F 45569.

Type locality: **Ecuador**, Tungurahua.

Type substrate: *Chusquea serrulata*.

Descriptions and illustrations: See [Sydow & Sydow \(1939\)](#).

andiyazi Fusarium Marasas et al., Mycologia 93: 1205. 2001.

Holotypus: BPI 748223.

Ex-type culture: CBS 119857 = IMI 386078 = KSU 4804 = MRC 6122.

Type locality: **South Africa**, KwaZulu-Natal Province, Greytown.

Type substrate: Soil debris of *Sorghum bicolor*.

Descriptions and illustrations: See [Marasas et al. \(2001\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: LT996189; *rpb2*: LT996138; *tef1*: LT996092.

andropogonis Fusarium Cooke ex Sacc., Syll. Fung. 10: 726. 1892.

Synonyms: *Fusisporium andropogonis* Cooke ex Thüm., Mycoth. Univ. 7: no. 676. 1877, nom. inval., Art. 38.1(a).

Ramularia andropogonis (Cooke ex Sacc.) Wollenw., Fusaria Autogr. Delin. 1: 469. 1916.

Lectotypus (hic designatus, MBT 10000653): **USA**, New Jersey, Newfield, from dead stem of *Andropogon virginicus*, Oct. 1874, J.B. Ellis, BR5020081431482 (Thümen, Mycoth. Univ. 7: no. 676).

Notes: Status unclear, not *Ramularia* *fide* [Braun \(1998\)](#). Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

anguioides Fusarium Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 169. 1915.

Typus: ?CUP-007479.

Type locality: **USA**, New York, Castile.

Type substrate: *Solanum tuberosum*.

Descriptions and illustrations: See [Sherbakoff \(1915\)](#), [Gerlach & Nirenberg \(1982\)](#) and [Nelson et al. \(1995\)](#).

Notes: [Nelson et al. \(1995\)](#) designated BPI 72044 as neotype of *F. anguioides*, erroneously stating that no materials were available for epi- and lectotypification. However, [Sherbakoff \(1915\)](#) did provide an illustration with the original protologue of *F. anguioides* and placed material in CUP, as CUP-007479. Furthermore, the neotype (BPI 72044) of [Nelson et al. \(1995\)](#) originated from China and was isolated from soil in a bamboo grove. An isolate from the original locality (USA) and host (*Solanum tuberosum*) needs to be selected. Lectotypification pending study of material lodged in CUP.

angustum Fusarium Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 203. 1915.

(See *Fusarium oxysporum*)

Typus: ?CUP-007435.

Type locality: **USA**, New York, Ithaca.

Type substrate: *Solanum tuberosum*.

Descriptions and illustrations: See [Sherbakoff \(1915\)](#).

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#). Lectotypification pending study of material lodged in CUP.

anisophilum Fusarium Picado, J. Dept. Agric. Porto Rico 16: 391. 1932.

(See *Fusarium lateritium*)

Holotypus: Not located.

Type locality: **Costa Rica**.

Type substrate: Living stem of *Coffea* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

annulatum Fusarium Bugnic., Rev. Gén. Bot. 59: 17. 1952.

Holotypus: IMI 202878.

Ex-type culture: BBA 63629 = CBS 258.54 = IMI 202878 = MUCL 8059 = NRRL 13619.

Type locality: **New Caledonia**.

Type substrate: Grain of *Oryza sativa*.

Descriptions and illustrations: See Bugnicourt (1952), Yilmaz *et al.* (2021).

Diagnostic DNA barcodes: *rpb1*: MT010944; *rpb2*: MT010983; *tef1*: MT010994.

annuum Fusarium Leonian, Bull. New Mex. Coll. Agric. Mech. Arts 121: 9. 1919.

Lectotypus (*hic designatus*, MBT 10000654): **USA**, New Mexico, from *Capsicum annuum*, 1919, L.H. Leonian, In Bull. New Mex. Coll. Agric. Mech. Arts 121: 32, fig. 7.

Notes: No type specimen could be located. Wollenweber & Reinking (1935) mentioned this species but did not study or treat it any further. A new collection is required for epitypification from the type locality and substrate.

anomalum Fusarium Berk. & M.A. Curtis, in Berkeley, Grevillea 3: 99. 1875.

Holotypus: ?K(M).

Type locality: **USA**, the New England region.

Type substrate: *Gleditsia* sp.

Notes: Status unclear. Not *Fusarium* *fide* Wollenweber & Reinking (1935).

anthophilum Fusarium (A. Braun) Wollenw., Ann. Mycol. 15: 14. 1917.

Basionym: *Fusisporium anthophilum* A. Braun, in Rabenhorst, Fungi Eur. Exs.: no. 1964. 1875.

Synonyms: *Fusarium moniliforme* var. *anthophilum* (A. Braun) Wollenw., Fusaria Autogr. Delin. 3: 975. 1930.

Fusarium tricinctum var. *anthophilum* (A. Braun) Bilař, Fusarii (Biologija I sistematika): 251. 1955.

Fusarium sporotrichiella var. *anthophilum* (A. Braun) Bilař, Mikrobiol. Zhurn. 49: 7. 1987.

Fusarium sanguineum var. *pallidius* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 196. 1915.

Fusarium wollenweberi Rallo, Fungi of the Genus *Fusarium*: 189. 1950, *nom. inval.*, Art. 41.1.

Lectotypus: Rabenhorst, Fungi Eur. Exs. no. 1964 in B, designated by Yilmaz *et al.* (2021).

Lectotype locality: **Germany**, Berchtesgaden.

Lectotype substrate: *Succisa pratensis*.

Epitypus: CBS 222.76 (preserved as metabolically inactive culture), designated by Yilmaz *et al.* (2021).

Ex-epitype culture: BBA 63270 = CBS 222.76 = IMI 196084 = IMI 202880 = NRRL 22943 = NRRL 25216.

Epitype locality: **Germany**, Berlin.

Epitype substrate: *Euphorbia pulcherrima*.

Descriptions and illustrations: See Wollenweber & Reinking (1935), Nirenberg (1976), Gerlach & Nirenberg (1982), Nelson *et al.* (1983) and Leslie & Summerell (2006).

Diagnostic DNA barcodes: *rpb1*: MW402641; *rpb2*: MW402811; *tef1*: MW402114.

apii Fusarium P.E. Nelson & Sherb., Techn. Bull. Michigan Agric. Exp. Sta. 155: 42. 1937.

(See *Fusarium oxysporum*)

Holotypus: Not located.

Type locality: **USA**.

Type substrate: *Apium graveolens* var. *dulce*.

apiogenum Fusarium Sacc., Syll. Fung. 4: 717. 1886.

(See *Fusarium lactis*)

Holotypus: Not located.

Type locality: **Germany**.

Type substrate: Rotten fruit.

aquaeductuum Fusarium (Radlk. & Rabenh.) Lagerh. & Rabenh., Centralbl. Bakteriol. Parasitenk. Abth.9: 655. 1891.

Fusicolla aquaeductuum (Radlk. & Rabenh.) Gräfenhan *et al.*, Stud. Mycol. 68: 100. 2011.

Basionym: *Selenosporium aquaeductuum* Radlk. & Rabenh., Kunst- und Gewerbeblatt des Polytechnischen Vereins des Königreichs Bayern 41(1): 10. 1863.

Synonyms: *Fusisporium moschatum* Kitasato, Centralbl. Bakteriol. Parasitenk., 1. Abth. 5: 368. 1889.

Fusarium moschatum (Kitasato) Sacc., Syll. Fung. 10: 729. 1892.

Fusarium magnusianum Allesch., Fungi Bav. no. 400. 1895.

Fusarium aquaeductuum var. *pusillum* Wollenw., Ann. Mycol. 15: 53. 1917.

Fusarium aquaeductuum var. *volutum* Wollenw., Ann. Mycol. 15: 53. 1917.

Fusarium aquaeductuum var. *elongatum* Wollenw., Fusaria Autogr. Delin. 3: 847. 1930.

Fusarium aquaeductuum var. *majus* Wollenw., Fusaria Autogr. Delin. 3: 845. 1930.

Fusarium bicellulare Kirschst., Hedwigia 80: 136. 1941.

Lectotypus: B 700014034, designated in Gräfenhan *et al.* (2011).

Lectotype locality: **Germany**, Bayern, München, Gasteigberg.

Lectotype substrate: Water in water fountain.

Epitypus: BBA 64559, designated in Gräfenhan *et al.* (2011).

Ex-epitype culture: BBA 64559 = CBS 837.85 = NRRL 20865 = NRRL 37595.

Epitype locality: **Germany**.

Epitype substrate: Water from plugged water tap in BBA.

Descriptions and illustrations: See Gerlach & Nirenberg (1982).

Diagnostic DNA barcodes: *rpb1*: KM232250; *rpb2*: HQ897744; *tef1*: KM231955.

arachnoideum Fusarium (Corda) Sacc., Syll. Fung. 4: 721. 1886. *Basionym*: *Fusisporium arachnoideum* Corda, Icon. Fung. 1: 11. 1837.

(See *Fusarium merismoides*)

Typus: In PRM.

Type locality: **Czech Republic**, Prague.

Type substrate: Soil.

Note: Synonym *fide* Wollenweber & Reinking (1935). Lectotypification pending study of material lodged in PRM.

arcuatissporum Fusarium M.M. Wang *et al.*, Persoonia 43: 78. 2019.

Holotypus: HAMS 248034.

Ex-type culture: CGMCC 3.19493 = LC 12147.

Type locality: **China**, Hubei.

Type substrate: Pollen of *Brassica campestris*.

Descriptions and illustrations: See Wang et al. (2019).

Diagnostic DNA barcodes: *rpb1*: MK289799; *rpb2*: MK289739; *tef1*: MK289584.

arcuatum *Fusarium* Berk. & M.A. Curtis, Grevillea 3: 99. 1875.
(See *Fusarium avenaceum*)

Lectotypus (*hic designatus*, MBT 10000655): **USA**, South Carolina, *Malus pumila* (syn. *Pyrus malus*), date unknown, M.A. Curtis, PH00005557.

Note: Synonym *fide* Wollenweber & Reinking (1935).

arcuosporum *Fusarium* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 186. 1915.

(See *Fusarium acuminatum*)

Typus: ?CUP-007477.

Type locality: **USA**, New York, Castile.

Type substrate: *Solanum tuberosum*.

Descriptions and illustrations: See Sherbakoff (1915).

Notes: Synonym *fide* Wollenweber & Reinking (1935). Lectotypification pending study of material lodged in CUP.

argillaceum *Fusarium* (Fr.) Sacc., Syll. Fung. 4: 718. 1886.

Basionym: *Fusisporium argillaceum* Fr., Syst. Mycol. 3: 446. 1832.

Synonyms: *Fusarium solani* var. *argillaceum* (Fr.) Bilař, Mikrobiol. Zhurn. 49: 7. 1987.

Nectria solani Reinke & Berthold, Untersuch. Bot. Lab. Univ. Göttingen 1: 39. 1879.

Dialonectria solani (Reinke & Berthold) Cooke, Grevillea 12: 111. 1884.

Cucurbitaria solani (Reinke & Berthold) Kuntze, Revis. Gen. Pl. 3: 461. 1898.

Holotypus: Not located.

Type locality: **Unknown**.

Type substrate: Periderm of *Cucumis* sp.

Notes: Status unclear. Requires recollection from type locality and substrate.

aridum *Fusarium* O.A. Pratt, J. Agric. Res. 13: 89. 1918.

(See *Fusarium sambucinum*)

Lectotypus (*hic designatus*, MBT 10000656): **USA**, Idaho, from soil, 1918, O.A. Pratt, in J. Agric. Res. 13: 87, fig. 2Q.

Notes: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

armeniicum *Fusarium* (G.A. Forbes et al.) L.W. Burgess & Summerell, **comb. nov.** MycoBank MB 837636.

Basionym: *Fusarium acuminatum* subsp. *armeniicum* G.A. Forbes et al., Mycologia 85: 120. 1993.

Holotypus: DAR 67507.

Ex-type culture: ATCC 90020 = CBS 485.94 = FRC R-9335 = IMI 352099 = MRC 6230 = NRRL 26908 = NRRL 25141 = NRRL 29133.

Type locality: **Australia**, New South Wales, Edgeroi.

Type substrate: *Triticum aestivum*.

Descriptions and illustrations: See Burgess et al. (1993), Burgess & Summerell (2000) and Leslie & Summerell (2006).

Diagnostic DNA barcodes: *rpb1*: KT597715; *rpb2*: GQ915485; *tef1*: GQ915501.

Notes: When proposing *F. armeniicum*, Burgess & Summerell (2000) cited the basionym as *F. acuminatum* subsp. *armeniicum* with reference to the entire pagination of Burgess et al.'s

(1993) paper, rather than the intended basionym alone, rendering the combination invalid (Art. 41.5, Ex. 15). Here we validate the new combination with the correct citation of the basionym.

arthrosporioides *Fusarium* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 175. 1915.

Typus: ?CUP-007467.

Type locality: **USA**, New York, Castile.

Type substrate: *Solanum tuberosum*.

Descriptions and illustrations: See Sherbakoff (1915), Booth (1971) and Gerlach & Nirenberg (1982).

Notes: Synonym *fide* Wollenweber & Reinking (1935). Lecto- and epitypification pending study of material lodged in CUP.

arundinis *Fusarium* (Corda) Sacc., Syll. Fung. 4: 724. 1886.

Trichoderma viride Pers., Neues Mag. Bot. 1: 92. 1794, *nom. sanct.* [Fr., Syst. Mycol. 3: 215. 1829].

Synonyms: *Pyrenium lignorum* Tode, Fung. Mecklenb. Sel. 1: 33, tab. 3, fig. 29. 1790.

Trichoderma lignorum (Tode) Harz, Bull. Soc. Imp. Naturalistes Moscou 44: 116. 1871.

Trichoderma viride Schumach., Enum. Pl. 2: 235. 1803, *nom. illegit.*, Art. 53.1.

Fusisporium arundinis Corda, Icon. Fung. 1: 11. 1837.

Trichoderma glaucum E.V. Abbott, Iowa State Coll. J. Sci. 1: 27. 1927.

Lectotypus (*hic designatus*, MBT 10000657): **Czech Republic**, Prague, rotten leaves of reeds, 1837, A.C.J. Corda, Icon. Fung. 1, tab. II, fig. 163.

Notes: Synonyms *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

arvense *Fusarium* Speg., Anales Soc. Ci. Argent. 10: 60. 1880.
(See *Fusarium merismoides*)

Holotypus: In LPS (Fungi Argent. pug. 2, #153).

Type locality: **Argentina**.

Type substrate: Dried fruits of *Solanum elaeagnifolium*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

asclepiadeum *Fusarium* Fautrey, Rev. Mycol. (Toulouse) 18: 68. 1896.

(See *Fusarium lateritium*)

Syntype: ILL00221138 (Fungi Sel. Gall. Exs. #6929).

Type locality: **France**, Montagne de Bard.

Type substrate: *Vincetoxicum officinale* (syn. *V. hirundinaria*).

Note: Synonym *fide* Wollenweber & Reinking (1935).

asclerotium *Fusarium* (Sherb.) Wollenw., Fusaria Autogr. Delin. 1: 364. 1916.

Basionym: *Fusarium oxysporum* var. *asclerotium* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 222. 1915.

(See *Fusarium oxysporum*)

Lectotypus (*hic designatus*, MBT 10000658): **USA**, New York, Atlanta, rotten tuber of *Solanum tuberosum*, 1915, C.D. Sherbakoff, in Mem. Cornell Univ. Agric. Exp. Sta. 6: 221, fig. 35 B–J.

Notes: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

asiaticum *Fusarium* O'Donnell et al., Fungal Genet. Biol. 41: 619. 2004.

Holotypus: BPI 843478.

Ex-type culture: CBS 110257 = FRC R-5469 = NRRL 13818.

Type locality: **Japan**.

Type substrate: *Hordeum vulgare*.

Descriptions and illustrations: See O'Donnell *et al.* (2004).

Diagnostic DNA barcodes: *rpb1*: JX171459; *rpb2*: JX171573; *tef1*: AF212451.

asparagi Fusarium Briard, Rev. Mycol. (Toulouse) 12: 142. 1890.
(See ***Fusarium incarnatum***)

Holotypus: ?PC.

Type locality: **France**, Aube, Troyes.

Type substrate: *Asparagus* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

asparagi Fusarium Delacr., Bull. Soc. Mycol. France 6: 99. 1890, *nom. illegit.*, Art. 53.1., *non Fusarium asparagi* Briard 1890.
Replacing synonym: *Fusarium delacroixii* Sacc., Syll. Fung. 10: 725. 1892.

(See ***Fusarium sambucinum***)

Notes: Synonym *fide* Wollenweber & Reinking (1935). See *F. delacroixii* for lectotypification.

asperifoliorum Fusarium (Westend.) Sacc., Syll. Fung. 4: 703. 1886.

Basionym: *Selenosporium asperifoliorum* Westend., Bull. Acad. Roy. Sci. Belgique, sér. 2, 11: 652. 1861.

Holotypus: BR5020140146784.

Type locality: **Belgium**, Oudenaarde.

Type substrate: *Borago officinalis*.

Notes: Status unclear. Not *Fusarium* *fide* Wollenweber & Reinking (1935).

aspidioti Fusarium Sawada, Bot. Mag. (Tokyo) 28: 312. 1914.
(See *Fusarium larvarum*)

Holotypus: TNS-F-218710.

Type locality: **Japan**, Shizuoka.

Type substrate: *Quadraspidiotus perniciosus* (= *Aspidiotus perniciosus*) (San Jose scale).

Note: Synonym *fide* Wollenweber & Reinking (1935).

atrovinosum Fusarium L. Lombard & Crous, Fungal Syst. Evol. 4: 190. 2019.

Holotypus: CBS H-24015.

Ex-type culture: CBS 445.67 = BBA 10357 = DSM 62169 = IMI 096270 = NRRL 26852 = NRRL 26913.

Type locality: **Australia**.

Type substrate: *Triticum aestivum*.

Descriptions and illustrations: See Lombard *et al.* (2019a).

Diagnostic DNA barcodes: *rpb1*: MN120713; *rpb2*: MW928822; *tef1*: MN120752.

atrovirens Fusarium (Berk.) Mussat, Syll. Fung. 15: 144. 1901, *nom. inval.*, Arts. 35.1, 36.1(a), (c).

Fusariella atrovirens (Berk.) Sacc., Atti Reale Ist. Veneto Sci. Lett. Arti, ser. 6, 2: 463. 1884.

Basionym: *Fusisporium atrovirens* Berk., in Smith, Engl. Fl. 5 (2): 351. 1836.

Holotypus: ?K(M).

Type locality: **UK**, Northamptonshire, Kings Cliffe.

Type substrate: *Allium* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

aurantiacum Fusarium Corda, in Sturm, Deutschl. Fl., 3 Abt. (Pilze Deutschl.) 2: 19. 1829.

(See ***Fusarium oxysporum***)

Typus: No. 156060 in PRM.

Isotypus: IMI 133948 (slide).

Type locality: **France**.

Type substrate: Dead branch.

Note: Synonym *fide* Wollenweber & Reinking (1935). Lectotypification pending study of material lodged in PRM.

aureum Fusarium Corda, Icon. Fung. 1: 4. 1837.

Hymenella aurea (Corda) L. Lombard, *comb. nov.* MycoBank MB 837637.

Basionym: *Fusarium aureum* Corda, Icon. Fung. 1: 4. 1837.

Synonym: *Hymenula aurea* (Corda) Wollenw., *Fusarien*: 319. 1935.

Typus: In PRM *fide* Pilat (1938).

Type locality: **Czech Republic**, Prague.

Type substrate: Rotten vegetables.

Notes: Wollenweber & Reinking (1935) provided a new combination for *F. aureum* in the genus *Hymenula*. However, the generic name *Hymenella* (1822) predates the generic name *Hymenula* (1828) and therefore we provide a new combination in the latter genus. Lectotypification pending study of material lodged in PRM.

austrorfricanum Fusarium A. Jacobs *et al.*, Mycologia 110: 1197. 2018.

Holotypus: PREM 62137.

Ex-type culture: NRRL 66741 = PPRI 10408.

Type locality: **South Africa**, Eastern Cape Province, Humansdorp.

Type substrate: Endophyte of *Pennisetum clandestinum*.

Descriptions and illustrations: See Jacobs-Venter *et al.* (2018).

Diagnostic DNA barcodes: *rpb1*: MH742537; *rpb2*: MH742616; *tef1*: MH742687.

austroramericanum Fusarium T. Aoki *et al.*, Fungal Genet. Biol. 41: 617. 2004.

Holotypus: BPI 843473.

Ex-type culture: CBS 110244 = NRRL 2903.

Type locality: **Brazil**.

Type substrate: Polypore.

Descriptions and illustrations: See O'Donnell *et al.* (2004).

Diagnostic DNA barcodes: *rpb1*: JAAMOD010000230; *rpb2*: JAAMOD010000315; *tef1*: JAAMOD010000079.

avenaceum Fusarium (Fr.) Sacc., Syll. Fung. 4: 713. 1886.

Basionym: *Fusisporium avenaceum* Fr., Syst. Mycol. 2: 238. 1822, *nom. sanct.* [Fr., l.c.].

Synonyms: *Sarcopodium avenaceum* (Fr.) Fr., Summa Veg. Scand. 2: 472. 1849.

Fusarium herbarum var. *avenaceum* (Fr.) Wollenw., Fusaria Autogr. Delin. 3: 899. 1930.

Fusarium roseum var. *avenaceum* (Fr.) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 32: 663. 1945.

Fusisporium pyrinum Fr., Syst. Mycol. 3: 445. 1832, *nom. sanct.* [Fr., l.c.].

Fusarium pyrinum (Fr.) Sacc., Syll. Fung. 4: 720. 1886, *nom. illegit.*, Art. 53.1.

Fusarium tenue Corda, Icon. Fung. 1: 3. 1837.

Selenosporium tubercularioides Corda, Icon. Fung. 1: 7. 1837.

Fusarium tubercularioides (Corda) Sacc., Syll. Fung. 4: 697. 1886.

Fusarium herbarum var. *tubercularioides* (Corda) Wollenw., Fusaria Autogr. Delin. 3: 892. 1930.

Selenosporium herbarum Corda, Icon. Fung. 3: 34. 1839.

Fusarium herbarum (Corda) Fr., Summa Veg. Scand. 2: 472. 1849.

- Fusarium graminum* var. *herbarum* (Corda) Wollenw., *Fusaria* Autogr. Delin. 3: 941. 1930.
- Fusarium avenaceum* var. *herbarum* (Corda) Bilař, *Fusarii* (Biologija i sistematika): 95. 1955.
- Fusarium tritici* Liebman bis, *Tidsskr. Landoekon.*, n.s., 2: 515. 1840.
- Fusisporium zeae* Westend., *Bull. Acad. Roy. Sci. Belgique* 18: 414. 1851.
- Fusarium zeae* (Westend.) Sacc., *Syll. Fung.* 4: 713. 1886.
- Fusarium amentorum* Lacroix, *Fl. Maine-et-Loire* 2 (Suppl.): [1]. 1854.
- Gloeosporium amentorum* (Lacroix) Lind, *Ann. Mycol.* 3: 431. 1905.
- Calogloeum amentorum* (Lacroix) Nannf., *Svensk Bot. Tidskr.* 25: 25. 1931.
- Platycarpium amentorum* (Lacroix) Petr., *Sydowia* 7: 296. 1953.
- Fusamen amentorum* (Lacroix) Arx, *Verh. Kon. Akad. Wetensch., Afd. Natuurk.* 51: 57. 1957.
- Fusisporium incarcerans* Berk., *Intellectual Observ.* 2: 11. 1863.
- Fusarium incarcerans* (Berk.) Sacc., *Syll. Fung.* 4: 713. 1886.
- Fusarium stercoris* Fuckel, *Fungi Rhen. Exs., Suppl. Fasc.* 5: no. 1921, 1867.
- Menispora penicillata* Harz, *Bull. Soc. Imp. Naturalistes Moscou* 44: 127. 1871.
- Fusarium penicillatum* (Harz) Sacc., *Syll. Fung.* 4: 710. 1886.
- Fusisporium schiedermayeri* Thüm., *Fungi Austr. Exs. Cent.* 1: no. 78. 1871.
- Fusarium schiedermayeri* (Thüm.) Sacc., *Syll. Fung.* 4: 712. 1886.
- Fusarium arcuatum* Berk. & M.A. Curtis, *Grevillea* 3: 99. 1875.
- Fusarium viticola* Thüm., *Pilze Weinst.*: 52. 1878.
- Fusarium herbarum* var. *viticola* (Thüm.) Wollenw., *Fusaria* Autogr. Delin. 3: 898. 1930.
- Fusarium gaudefroyanum* Sacc., *Michelia* 2: 132. 1880.
- Fusisporium cucurbitariae* Pat., *Rev. Mycol. (Toulouse)* 3: 10. 1881.
- Fusarium cucurbitariae* (Pat.) Sacc., *Syll. Fung.* 4: 708. 1886, *nom. illegit.*, Art. 53.1, *non Fusarium cucurbitariae* Peyronel 1918.
- Fusarium amenti* Rostr., *Bot. Tidsskr.* 14: 240. 1885.
- Fusarium urenidicola* Jul. Müll., *Ber. Deutsch. Bot. Ges.* 3: 395. 1885.
- Fusarium diffusum* Carmich., *Grevillea* 16: 81. 1888.
- Fusarium iridis* Oudem., *Ned. Kruidk. Arch.*, ser. 2, 5: 515. 1889.
- Fusarium ustilaginis* Kellerm. & Swingle, *Rep. (Annual) Kansas Agric. Exp. Sta.* 2: 285. 1890.
- Fusarium ruberrimum* Delacr., *Bull. Soc. Mycol. France* 6: 139. 1890.
- Fusarium peckii* Sacc., *Syll. Fung.* 10: 727. 1892, *nom. illegit.*, Art. 53.1 [pro. p. fide [Wollenweber & Reinking \(1935\)](#)].
- Fusarium aecidii-tussilaginis* Allesch., *Ber. Bot. Vereines Landshut* 12: 131. 1892.
- Fusarium subviolaceum* Roum. & Fautrey, *Rev. Mycol. (Toulouse)* 14: 106. 1892.
- Fusarium granulosum* Ellis & Everh., *Proc. Acad. Nat. Sci. Philadelphia* 45: 466. 1894 [1893].
- Fusarium jungiae* Pat., *Bull. Soc. Mycol. France* 11: 234. 1895.
- Fusarium schnablianum* Allesch., *Hedwigia* 34: 289. 1895.
- Fusarium seemenianum* Henn., *Allg. Bot. Z. Syst.* 2: 83. 1896.
- Fusarium aleurinum* Ellis & Everh., *Bull. Torrey Bot. Club* 24: 476. 1897.
- Fusarium pseudonectria* Speg., *Anales Mus. Nac. Hist. Nat. Buenos Aires* 6: 351. 1899.
- Fusarium limosum* Rostr., *Bot. Tidsskr.* 22: 263. 1899.
- Fusarium gracile* McAlpine, *Proc. Linn. Soc. New South Wales* 28: 554. 1903.
- Fusarium putrefaciens* Osterw., *Mitth. Thurgauischen Naturf. Ges.* 16: 123. 1904.
- Fusarium paspali* Henn., *Bot. Jahrb. Syst.* 38: 129. 1905.
- Fusarium sorghi* Henn., *Ann. Mus. Congo Belge, Bot., Sér.* 5, 2: 105. 1907.
- Fusarium speiseri* Lindau, *Rabenh. Krypt.-Fl.*, ed. 2, 1(9): 580. 1909.
- Fusarium palczewskii* Jacz., *Bull. Soc. Mycol. France* 28: 345. 1912.
- Fusarium pseudoheterosporum* Jacz., *Bull. Soc. Mycol. France* 28: 347. 1912.
- Fusarium metachroum* Appel & Wollenw., *Arbeiten Kaiserl. Biol. Anst. Land- Forstw.* 8: 141. 1913.
- Fusarium subulatum* Appel & Wollenw., *Arbeiten Kaiserl. Biol. Anst. Land- Forstw.* 8: 131. 1913.
- Fusarium bifforme* Sherb., *Mem. Cornell Univ. Agric. Exp. Sta.* 6: 166. 1915.
- Fusarium lucidum* Sherb., *Mem. Cornell Univ. Agric. Exp. Sta.* 6: 157. 1915.
- Fusarium metachroum* var. *minus* Sherb., *Mem. Cornell Univ. Agric. Exp. Sta.* 6: 145. 1915.
- Fusarium subulatum* var. *brevius* Sherb., *Mem. Cornell Univ. Agric. Exp. Sta.* 6: 149. 1915.
- Fusarium truncatum* Sherb., *Mem. Cornell Univ. Agric. Exp. Sta.* 6: 155. 1915.
- Fusarium avenaceum* var. *pallens* Wollenw., *Fusaria* Autogr. Delin. 2: 575. 1924.
- Fusarium venerorum* Dounin & Goldmacher, *Actes du premier Congres Internat. des Sylvicult.*: 284–298. 1927.
- Fusarium herbarum* var. *volutum* Wollenw., *Fusaria* Autogr. Delin. 3: 893. 1930.
- Fusarium avenaceum* var. *volutum* (Wollenw.) Wollenw. & Reinking, *Fusarien*: 56. 1935.
- Fusarium avenaceum* subsp. *volutum* (Wollenw.) Raillo, *Fungi of the Genus Fusarium*: 188. 1950.
- Fusarium avenaceum* var. *fabae* T.F. Yu, *Phytopathology* 34: 392. 1944.
- Fusarium avenaceum* f. *fabae* (T.F. Yu) W. Yamam., *Sci. Rep. Hyogo Univ. Agric.*, Ser. Agr. Biol. 2: 60. 1955.
- Gibberella avenacea* R.J. Cook, *Phytopathology* 57: 735. 1967.
- Fusarium avenaceum* f. *fabalis* X.Y. Ruan *et al.*, *Acta Phytopathol. Sin.* 12: 32. 1982, *nom. inval.*, Art. 39.1.
- Fusarium avenaceum* f. *fabarum* X.Y. Ruan *et al.*, *Acta Phytopathol. Sin.* 12: 32. 1982, *nom. inval.*, Art. 39.1.
- Neotypus* (*hic designatus*, MBT 10000659): **Denmark**, *Hordeum vulgare*, 3 Feb. 1986, U. Thrane, CBS 408.86 (preserved as metabolically inactive culture).
- Ex-neotype culture*: CBS 408.86 = FRC R-8510 = IMI 309354 = NRRL 26850 = NRRL 26911.
- Descriptions and illustrations*: See [Wollenweber & Reinking \(1935\)](#), [Booth \(1971\)](#), [Gerlach & Nirenberg \(1982\)](#), [Nelson *et al.* \(1983\)](#) and [Leslie & Summerell \(2006\)](#).
- Diagnostic DNA barcodes*: *rpb1*: MG282372; *rpb2*: MG282401; *tef1*: MW928836.
- Notes*: No type material could be located for this species. Therefore, to provide taxonomic stability to this important cereal-associated *Fusarium* species, CBS 408.86 is designated here as ex-neotype of *Fusisporium avenaceum* (= *Fusarium avenaceum*).
- awaxy *Fusarium*** Petters-Vandresen *et al.*, *Persoonia* 43: 363. 2019.
- Holotypus*: UPCB93138-H.
- Ex-type culture*: CMRP 4013 = LGMF1930.

Type locality: **Brazil**, Paraná, Guarapuava.
 Type substrate: Rotten stalks of *Zea mays*.
 Descriptions and illustrations: See [Crous et al. \(2019b\)](#).
 Diagnostic DNA barcodes: *rpb2*: MK766941; *tef1*: MG839004.

aywerte Fusarium (Sangal. & L.W. Burgess) Benyon & L.W. Burgess, Mycol. Res. 104: 1171. 2000.
 Basionym: *Fusarium avenaceum* subsp. *aywerte* Sangal. & L.W. Burgess, Mycol. Res. 99: 287. 1995.
 Holotypus: DAR 69501.
 Ex-type culture: F10108 = NRRL 25410.
 Type locality: **Australia**, Northern Territory, Deep Well.
 Type substrate: Soil.
 Descriptions and illustrations: See [Sangalang et al. \(1995\)](#), [Benyon et al. \(2000\)](#) and [Leslie & Summerell \(2006\)](#).
 Diagnostic DNA barcodes: *rpb1*: JX171513; *rpb2*: JX171626; *tef1*: JABCQV010000336.

azedarachinum Fusarium (Thüm.) Sacc., Syll. Fung. 4: 704. 1886.
 Basionym: *Fusisporium azedarachinum* Thüm., Mycoth. Univ. 14: no. 1379. 1879.
 (See ***Fusarium lateritium***)
 Syntypes: In BPI, CUP, ILL, NEB, NY, NYS PH & PUL (Mycoteca Universalis no. 1379).
 Type locality: **USA**, South Carolina, Aiken.
 Type substrate: *Melia azedarach*.
 Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

azukiicola Fusarium T. Aoki et al. (as 'azukicola'), Mycologia 104: 1075. 2012.

Neocosmospora phaseoli (Burkh.) L. Lombard & Crous, Stud. Mycol. 80: 227. 2015.
 Basionym: *Fusarium martii* f. *phaseoli* Burkh., Mem. Cornell Univ. Agric. Exp. Sta. 26: 1007. 1919.
 Synonyms: *Fusarium solani* f. *phaseoli* (Burkh.) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 28: 740. 1941.
Fusarium phaseoli (Burkh.) T. Aoki & O'Donnell, Mycologia 95: 671. 2003.
 ?*Fusarium epimyces* Cooke, Grevillea 17: 15. 1888.
 ?*Fusarium pestis* Sorauer, Atlas Pfl.-Krankh. 4: 19, pl. XXV. 1890.
 ?*Fusarium martii* var. *viride* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 247. 1915.
Fusarium solani var. *martii* Appel & Wollenw. f. 3 Snyder, Centralbl. Bakteriolog. Parasitenk., 2. Abth. 91: 179. 1934.
Fusarium solani f. sp. *glycines* K. Roy, Pl. Dis. 81: 264. 1997.
Fusarium tucumaniae T. Aoki et al., Mycologia 95: 664. 2003.
Neocosmospora tucumaniae (T. Aoki et al.) L. Lombard & Crous, Stud. Mycol. 80: 228. 2015.
Fusarium virguliforme O'Donnell & T. Aoki, Mycologia 95: 667. 2003.
Neocosmospora virguliformis (O'Donnell & T. Aoki) L. Lombard & Crous, Stud. Mycol. 80: 228. 2015.
Fusarium brasiliense T. Aoki & O'Donnell, Mycoscience 46: 166. 2005.
Fusarium cuneirostrum O'Donnell & T. Aoki, Mycoscience 46: 170. 2005.
Fusarium crassistipitatum Scandiani et al., Mycoscience 53: 171. 2011.
 Holotypus: BPI 881712.
 Ex-type culture: MAFF 242371 = NRRL 54364.
 Type locality: **Japan**, Hokkaido, Tokachi, Urahoro.
 Type substrate: Roots of *Vigna angularis*.
 Descriptions and illustrations: See [Aoki et al. \(2012b\)](#).

Diagnostic DNA barcodes: *rpb1*: KJ511276; *rpb2*: KJ511287; *tef1*: JQ670137.

babinda Fusarium Summerell et al., Mycol. Res. 99: 1345. 1995.
 Holotypus: DAR 70287.
 Ex-type culture: BBA 69872 = F11217 = NRRL 25807.
 Type locality: **Australia**, Queensland, Mount Lewis.
 Type substrate: Plant material in soil.
 Descriptions and illustrations: See [Summerell et al. \(1995\)](#) and [Leslie & Summerell \(2006\)](#).
 Diagnostic DNA barcode: *rpb2*: MN534245; *tef1*: AF160305.
 Note: The *Fusarium babinda* species complex encompassed strains incorrectly assigned to this taxon, based on reference strains of *F. babinda*, plus one unnamed *Fusarium* species ([O'Donnell et al. 2013](#), [Jacobs-Venter et al. 2019](#), [Geiser et al. 2021](#)). However, DNA sequences from diverse gene regions and phylogenetic analyses made by several authors place the ex-type of *F. babinda* (NRRL 25807) within the *Fusarium fujikuroi* species complex, as confirmed here ([Fig. 8](#)) ([O'Donnell et al. 2000b](#), [Lima et al. 2012](#), [Herron et al. 2015](#), [Crous et al. 2019b](#)). Hence, the species in FBSC need to be reassessed and the species complex renamed accordingly.

baccharidicola Fusarium Henn., Hedwigia 48: 20. 1908.
 (See ***Fusarium coccophilum***)
 Syntype: Puttemans no. 1274 in B (syntype *fide* [Hein \(1988\)](#)).
 Type locality: **Brazil**, São Paulo, Pirutuba.
 Type substrate: *Baccharis dracunculifolia* in association with cochineal (*Dactylopius coccus*)
 Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

bacilligerum Fusarium (Berk. & Broome) Sacc., Syll. Fung. 4: 711. 1886.
Pseudocercospora bacilligera (Berk. & Broome) Y.L. Guo & X.J. Liu, Mycosystema 2: 229. 1989.
 Basionym: *Fusisporium bacilligerum* Berk. & Broome, Ann. Mag. Nat. Hist., ser. 2, 7: 178. 1851.
 Synonyms: *Cercospora bacilligera* (Berk. & Broome) Wollenw., Fusaria Autogr. Delin. 1: 450. 1916.
Fusisporium erubescens Durieu & Mont., Exploration scientifique de l'Algérie 1–9: 351.1848.
Fusarium erubescens (Durieu & Mont.) Sacc., Syll. Fung. 4: 719. 1886, *nom. illegit.*, Art. 53.1.
 Holotypus: ?K(M).
 Type locality: **UK**, Wiltshire, Spye Park.
 Type substrate: Leaves of *Rhamnus alaternus*.
 Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

batruidioides Fusarium Wollenw., Science, N.Y. 79: 572. 1934.
 Lectotypus: NY00936830, designated in [Seifert & Gräfenhan \(2012\)](#).
 Ex-type culture: BBA 4748 = BBA 63602 = CBS 100057 = CBS 177.35 = DAOM 225115 = IMI 375323 = NRRL 22201.
 Type locality: **USA**, Arizona, Chiricahua Mountains.
 Type substrate: Parasitic on *Cronartium conigenum* growing on a mummified cone of *Pinus leiophylla*.
 Descriptions and illustrations: See [Wollenweber \(1934\)](#), [Gerlach & Nirenberg \(1982\)](#) and [Seifert & Gräfenhan \(2012\)](#).
 Diagnostic DNA barcodes: *rpb1*: MT010939; *rpb2*: MT010963; *tef1*: KC514053.

bagnisianum Fusarium Thüm., Nuovo Giorn. Bot. Ital. 8: 252. 1876.
Ascochyta caricis Fuckel, Fungi Rhen. Suppl. Fasc. 2: no. 1697. 1866.

Synonyms: *Phyllosticta caricis* (Fuckel) Sacc., Syll. Fung. 3: 61. 1884.

Ascochyta caricis Lambotte & Fautrey, Rev. Mycol. (Toulouse) 19: 141. 1897, *nom. illegit.*, Art. 53.1.

Syntypes: In BPI, ILL, NEB, NY, PUL & S.

Type locality: **Italy**, Rome, Insugherata.

Type substrate: *Spartium junceum*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

bambusae Fusarium (Teng) Z.Q. Zeng & W.Y. Zhuang, Mycosystema 36: 279. 2017.

Basionym: *Lisea australis* var. *bambusae* Teng, Sinensia 4: 278. 1934.

Synonym: *Gibberella bambusae* (Teng) W.Y. Zhuang & X.M. Zhang, Nova Hedwigia 76: 195. 2003.

Holotypus: BPI 631179.

Type locality: **China**, Anhui, Huang-shan.

Type substrate: *Bambusoideae* culm.

Descriptions and illustrations: See [Zhang & Zhuang \(2003\)](#) and [Zeng & Zhuang \(2017a\)](#).

bambusicola Fusarium Hara, Bot. Mag. (Tokyo) 27: 255. 1913.

Holotypus: Not located.

Type locality: **Japan**, Tokyo.

Type substrate: *Phyllostachys reticulata*.

Note: Type material (specimen(s) and/or living ex-type culture) not located.

baptisiae Fusarium Henn., Notizbl. Bot. Gart. Berlin 2: 383. 1899. (See *Fusarium dimerum*)

Holotypus: In B *fide* [Hein \(1988\)](#).

Type locality: **Germany**, Berlin, Botanical Garden.

Type substrate: *Baptisia tinctoria*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

barbatum Fusarium Ellis & Everh., J. Mycol. 4: 45. 1888.

Raffaelea barbata (Ellis & Everh.) D. Hawksw. (as '*barbatum*'), Bull. Brit. Mus. (Nat. Hist.), Bot. 6: 272. 1979.

Holotypus: NY00928690.

Type locality: **USA**, New Jersey, Newfield.

Type substrate: *Usnea barbata*.

bartholomaei Fusarium Peck, Bull. Torrey Bot. Club 36: 157. 1909.

Septogloeum bartholomaei (Peck) Wollenw., Fusaria Autogr. Delin. 2: 638. 1924.

Synonym: *Trichofusarium bartholomaei* (Peck) Sacc., Syll. Fung. 22: 1473. 1913.

Holotypus: NYS-F-000437.

Type locality: **USA**, Kansas, Stockton.

Type substrate: *Sorghastrum nutans*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

batatas Fusarium Wollenw. (as '*batatae*'), J. Agric. Res. 2: 268. 1914.

(See ***Fusarium oxysporum***)

Lectotypus (*hic designatus*, MBT 10000660): **USA**, Washington, *Ipomoea batatas*, 1914, L.L. Harter & E.C. Field, in Wollenweber, J. Agric. Res. 2: 268, pl. XVI, figs A–E.

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#). As no holotype specimen could be located, an illustration accompanying the original protologue is designated here as lectotype.

bataticola Fusarium (Sand.-Den. & Crous) O'Donnell *et al.*, Index Fungorum 440: 1. 2020.

Neocosmospora bataticola Sand.-Den. & Crous, Persoonia 43: 112. 2019.

Synonym: ?*Fusarium solani* f. *batatas* T.T. McClure, Phytopathology 41: 75. 1951, *nom. inval.*, Art. 39.1.

Holotypus: CBS H-23971.

Ex-type culture: BBA 64954 = CBS 144398 = FRC S-0567 = NRRL 22402.

Type locality: **USA**, North Carolina.

Type substrate: *Ipomoea batatas*.

Descriptions and illustrations: See [Sandoval-Denis *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW218100; *rpb2*: FJ240381; *tef1*: AF178344.

begoniae Fusarium Nirenberg & O'Donnell, Mycologia 90: 437. 1998.

Holotypus: B 70 0001694.

Ex-type culture: BBA 67781 = CBS 403.97 = IMI 375315 = NRRL 25300.

Type locality: **Germany**.

Type substrate: *Begonia elatior*.

Descriptions and illustrations: See [Nirenberg & O'Donnell \(1998\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: JAAOAG010000375; *rpb2*: MN193886; *tef1*: AF160293.

beomiforme Fusarium P.E. Nelson *et al.*, Mycologia 79: 886. 1987.

Holotypus: DAOM 196987.

Ex-type culture: CBS 100160 = DAR 58880 = FRC M-1425 = IMI 316127 = MRC 4593 = NRRL 13606.

Type locality: **Australia**, Queensland, Rockhampton.

Type substrate: Soil.

Descriptions and illustrations: See [Nelson *et al.* \(1987\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: MF120485; *rpb2*: MF120496; *tef1*: MF120507.

berenice Fusarium (Berk. & M.A. Curtis) Sacc., Syll. Fung. 4: 721. 1886.

Ascocalyx berenice (Berk. & M.A. Curtis) Baschien, IMA Fungus 5: 93. 2014.

Basionym: *Fusisporium berenice* Berk. & M.A. Curtis, in Berkeley, Grevillea 3: 147. 1875.

Synonyms: *Bothrodiscus berenice* (Berk. & M.A. Curtis) J.W. Groves, Canad. J. Bot. 46: 1273. 1968.

Holotypus: ?K(M).

Type locality: **USA**, Massachusetts, Boston, Murray.

Type substrate: *Peziza* sp.

berkeleyi Fusarium (Mont.) Berk. & Broome, North Amer. Fung.: 108. 1875.

Basionym: *Gloeosporium berkeleyi* Mont., Ann. Sci. Nat., Bot., sér. 3, 12: 296. 1849.

(See ***Fusarium lateritium***)

Holotypus: Not located.

Type locality: **USA**, Alabama.

Type substrate: Leaves of *Hibiscus syriacus*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

betae Fusarium (Desm.) Sacc., Michelia 2: 132. 1880.

Fusicolla betae (Desm.) Bonord., Handb. Mykol.: 150. 1851.

Basionym: *Fusisporium betae* Desm., Ann. Sci. Nat., Bot., Sér. 1, 19: 436. 1830.

- Synonyms:* *Pionnotes betae* (Desm.) Sacc., Syll. Fung. 4: 726. 1886.
Pionnotes rhizophila var. *betae* (Desm.) De Wild. & Durieu, Prodr. Fl. Belg. 2: 367. 1898.
Lectotypus: K(M) 167520, designated in Gräfenhan *et al.* 2011.
Lectotype locality: **France**.
Lectotype substrate: Tuber of *Beta vulgaris*.
Epitypus: BBA 64317, designated in Gräfenhan *et al.* 2011.
Ex-epitype culture: BBA 64317.
Epitype locality: **Germany**, Schleswig-Holstein, Kiel.
Epitype substrate: *Triticum aestivum*.
Descriptions and illustrations: See Gräfenhan *et al.* (2011).
Diagnostic DNA barcodes: *rpb2*: HQ897781.
- beticola* *Fusarium* A.B. Frank, Kampfbuch gegen die Schädlinge unserer Feldfrüchte: 137. 1897.
(See *Fusarium oxysporum*)
Holotypus: ?NY.
Type locality: **Germany**.
Type substrate: *Beta* sp.
Note: Synonym *vide* Wollenweber & Reinking (1935).
- biasolettianum* *Fusarium* Corda, Icon. Fung. 2: 3. 1838.
(See *Fusarium merismoides*)
Typus: PRM 155487.
Type locality: **Czech Republic**, near Prague.
Type substrate: Young stalks of *Betula* sp.
Notes: Synonym *vide* Wollenweber & Reinking (1935). Synonymy under *Fusicolla merismoides* still questionable (See Gräfenhan *et al.* 2011). Lectotypification pending study of material lodged in PRM.
- bicellulare* *Fusarium* Kirschst., Hedwigia 80: 136. 1941.
(See *Fusarium aquaeductum*)
Holotypus: B 70 0100184.
Type locality: **Germany**.
Type substrate: Parasitic on *Cryptosporrella hypodermia* with *Nectria episphaeria*.
Note: Synonym *vide* Wollenweber & Reinking (1935).
- biforme* *Fusarium* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 166. 1915.
(See *Fusarium avenaceum*)
Lectotypus (*hic designatus*, MBT 10000661): **USA**, Wisconsin, rotten tubers of *Solanum tuberosum*, date unknown, C.D. Sherbakoff, in Mem. Cornell Univ. Agric. Exp. Sta. 6: 166, fig. 17 (1915).
Notes: Synonym *vide* Wollenweber & Reinking (1935). As no holotype specimen could be located, an illustration accompanying the original protologue is designated here as lectotype.
- bipunctatum* *Fusarium* Preuss, Linnaea 25: 741. 1852.
(See *Fusarium tortuosum*)
Holotypus: ?B.
Type locality: **Germany**.
Type substrate: Branches of unknown tree.
Note: Synonym *vide* Wollenweber & Reinking (1935).
- biseptatum* *Fusarium* Sawada, Special Publ. Coll. Agric. Natl. Taiwan Univ. 8: 228. 1959, *nom. inval.*, Art. 39.1.
Authentic material: Not located.
Original locality: **Taiwan**.
Original substrate: Leaves of *Stephania cepharantha*.
biseptatum *Fusarium* Schroers *et al.*, Mycologia 101: 59. 2009. (*non Fusarium biseptatum* Sawada 1959).
Bisifusarium biseptatum (Schroers *et al.*) L. Lombard & Crous, Stud. Mycol. 80: 224. 2015.
Holotypus: CBS H-20126.
Ex-type culture: CBS 110311 = FRC E-0228 = NRRL 36184.
Type locality: **South Africa**, Transkei.
Type substrate: Soil.
Descriptions and illustrations: See Schroers *et al.* (2009).
Diagnostic DNA barcode: *tef1*: EU926319.
- blackmannii* *Fusarium* W. Br. & A.S. Horne (as '*blackmanni*'), Ann. Bot. (London) 38: 379. 1924.
(See *Fusarium lateritium*)
Notes: Name withdrawn by original author (W. Brown), See Brown (1928). Synonym *vide* Wollenweber & Reinking (1935).
- blasticola* *Fusarium* Rostr. (as '*blasticolum*'), Gartn.-Tidende 1895: 122. 1895.
(See *Fusarium oxysporum*)
Holotypus: Not located.
Type locality: **Germany**.
Type substrate: *Pinus sylvestris*.
Note: Synonym *vide* Wollenweber & Reinking (1935).
- bomiense* *Fusarium* (Z.Q. Zeng & W.Y. Zhuang) O'Donnell *et al.*, Index Fungorum 440: 1. 2020.
Neocosmospora bomiensis Z.Q. Zeng & W.Y. Zhuang, Phytotaxa 319: 177. 2017.
Holotypus: HMAS 254519.
Ex-type culture: HMAS 248885.
Type locality: **China**, Tibet Autonomous Region, Bomê County.
Type substrate: Twigs.
Descriptions and illustrations: See Zeng & Zhuang (2017b).
Diagnostic DNA barcode: *tef1*: KY829449.
- bonordenii* *Fusarium* Sacc., Syll. Fung. 4: 699. 1886.
Replaced synonym: *Selenosporium aurantiacum* Bonord., Abh. Naturf. Ges. Halle 8: 97. 1864, *nom. illegit.*, Art. 53.1, *non Fusarium aurantiacum* Corda 1829.
(See *Fusarium dimerum*)
Holotypus: Not preserved *vide* Holubová-Jechová *et al.* (1994).
Type locality: **Germany**.
Type substrate: Branches of unknown tree.
Note: Synonym *vide* Wollenweber & Reinking (1935).
- boothii* *Fusarium* O'Donnell *et al.*, Fungal Genet. Biol. 41: 618. 2004.
Holotypus: BPI 843475.
Ex-type culture: CBS 316.73 = IMI 160243 = NRRL 26916.
Type locality: **South Africa**.
Type substrate: *Zea mays*.
Descriptions and illustrations: See O'Donnell *et al.* (2004).
Diagnostic DNA barcodes: *rpb1*: KM361641; *rpb2*: KM361659; *tef1*: GQ915503.
- borneense* *Fusarium* (Petr.) O'Donnell *et al.*, Index Fungorum 440: 1. 2020.
Neocosmospora borneensis (Petr.) Sand.-Den. & Crous, Persoonia 43: 115. 2019.
Basionym: *Nectria borneensis* Petr., Sydowia 8: 20. 1954.
Holotypus: K(M) 252860.
Epitypus: CBS H-23972, designated in Sandoval-Denis *et al.* (2019).

Ex-epitype culture: BBA 65095 = CBS 145462 = G.J.S. 85-197 = NRRL 22579.

Epitype locality: **Indonesia**, North Sulawesi, Bogani Nani War-tabone National Park.

Epitype substrate: Bark of a recently dead unidentified tree.

Descriptions and illustrations: See [Sandoval-Denis et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW834213; *rpb2*: FJ240381; *tef1*: AF178344.

bostrycoides *Fusarium* Wollenw. & Reinking, *Phytopathology* 15: 166. 1925.

Neocosmospora bostrycoides (Wollenw. & Reinking) Sand.-Den. et al., *Persoonia* 43: 115. 2019.

Neotypus: CBS H-23973, designated in [Sandoval-Denis et al. \(2019\)](#).

Ex-neotype culture: CBS 144.25.

Neotype locality: **Honduras**, Tela.

Neotype substrate: Soil.

Descriptions and illustrations: See [Sandoval-Denis et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW218101; *rpb2*: LR583818; *tef1*: LR583597.

brachiariae* *Fusarium M.M. Costa et al., *Mycol. Progr.* 20: 67. 2021.

Holotypus: UB 24188.

Ex-type culture: CML 3032.

Type locality: **Brazil**, Mato Grosso do Sul, Campo Grande.

Type substrate: Seed of *Brachiaria decumbens*.

Descriptions and illustrations: See [Costa et al. \(2021\)](#).

Diagnostic DNA barcodes: *rpb2*: MT901314; *tef1*: MT901348.

brachygibbosum* *Fusarium Padwick, *Mycol. Pap.* 12: 11. 1945.

Holotypus: IMI 268019.

Ex-type culture: BBA 64691 = NRRL 20954.

Type locality: **India**, Telangana, Hyderabad, Parbhani.

Type substrate: *Sorghum vulgare*.

Descriptions and illustrations: See [Padwick \(1945\)](#).

Diagnostic DNA barcodes: *rpb1*: MW233246; *rpb2*: MW233418; *tef1*: MW233075.

brasiliicum* *Fusarium T. Aoki et al., *Fungal Genet. Biol.* 41: 620. 2004.

Holotypus: BPI 843480.

Ex-type culture: CBS 119180 = NRRL 31281.

Type locality: **Brazil**.

Type substrate: *Avena sativa*.

Descriptions and illustrations: See [O'Donnell et al. \(2004\)](#).

Diagnostic DNA barcodes: *rpb1*: JABCJS010000032; *rpb2*: JABCJS010000357; *tef1*: AY452964.

brasiliense *Fusarium* T. Aoki & O'Donnell, *Mycoscience* 46: 166. 2005.

(See *Fusarium azukiicola*)

Holotypus: BPI 843352.

Ex-type culture: MAFF 239050 = NRRL 31757.

Type locality: **Brazil**, Distrito Federal, Brasilia.

Type substrate: *Glycines max*.

Descriptions and illustrations: See [Aoki et al. \(2005\)](#).

Diagnostic DNA barcodes: *rpb1*: MAEC01003448; *rpb2*: EU329565; *tef1*: MAEC01004196.

brassicae *Fusarium* Lib. ex Cooke, *Grevillea* 8: 83. 1880.

(See *Fusarium candidum* Ehrenb.)

Holotypus: In B, Libert s.n. *vide* Index Fungorum.

Type locality: **France**.

Type substrate: Stem of *Brassica oleracea*.

Note: Synonym *vide* [Wollenweber & Reinking \(1935\)](#).

brassicae *Fusarium* (Thüm.) Sacc., *Syll. Fung.* 4: 701. 1886, *nom. illegit.*, Art. 53.1.

Basionym: *Selenosporium brassicae* Thüm., *Hedwigia* 19: 191. 1880.

(See ***Fusarium avenaceum***)

Holotypus: Not located.

Type locality: **Belgium**.

Type substrate: Stem of *Brassica oleracea*.

Note: Synonym *vide* [Wollenweber & Reinking \(1935\)](#).

breve *Fusarium* (Sand.-Den. & Crous) O'Donnell et al., *Index Fungorum* 440: 1. 2020.

Neocosmospora brevis Sand.-Den. & Crous, *Persoonia* 43: 119. 2019.

Holotypus: CBS H-23975.

Ex-type culture: CBS 144387 = MUCL 16108.

Type locality: **Belgium**, Heverlee.

Type substrate: Soil-water polluted with diethylene glycerol and ethylene glycerol.

Descriptions and illustrations: See [Sandoval-Denis et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW834214; *rpb2*: LR583822; *tef1*: LR583601.

brevicatenulatum* *Fusarium Nirenberg et al., *Mycologia* 90: 460. 1998.

Holotypus: Specimen in B *vide* [Nirenberg et al. \(1998\)](#).

Ex-type culture: BBA 69197 = CBS 404.97 = DAOM 225122 = IMI 375329 = NRRL 25446.

Type locality: **Madagascar**.

Type substrate: *Striga asiatica*.

Descriptions and illustrations: See [Nirenberg et al. \(1998\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: MT010948; *rpb2*: MT010979; *tef1*: MT011005.

brevicaudatum* *Fusarium J.W. Xia et al., *Persoonia* 43: 195. 2019.

Holotypus: CBS H-24051.

Ex-type culture: NRRL 43638 = UTHSC R-3500.

Type locality: **USA**, Florida.

Type substrate: *Trichechus* sp. (manatee).

Descriptions and illustrations: See [Xia et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: KC808322; *rpb2*: GQ505843; *tef1*: GQ505665.

breviconum *Fusarium* (Wollenw.) O'Donnell et al., *Index Fungorum* 440: 1. 2020.

Neocosmospora brevicona (Wollenw.) Sand.-Den. & Crous, *Persoonia* 43: 117. 2019.

Basionym: *Hypomyces haematococcus* var. *breviconus* Wollenw., *Fusaria Autogr. Delin.* 3: no. 828 (1930).

Synonyms: *Fusarium solani* var. *minus* Wollenw., *Fusarien*: 134. 1935.

Nectria haematococca var. *breviconica* (Wollenw.) Gerlach, *Fusarium: Diseases, Biology, and Taxonomy* (Philadelphia): 422. 1981.

Lectotypus: Fig. 828 in Wollenweber (1930), designated in [Sandoval-Denis et al. \(2019\)](#).

Epitypus: CBS H-23974 designated in [Sandoval-Denis et al. \(2019\)](#).

Ex-epitype culture: BBA 2123 = CBS 204.31 = NRRL 22659.

Epitype locality: **Indonesia**, West Java, Bogor.

Epitype substrate: *Gladiolus* sp.

Descriptions and illustrations: See [Sandoval-Denis et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW218103; *rpb2*: LR583821; *tef1*: LR583600.

briosianum *Fusarium* Ferraris, Fl. Ital. Crypt. Hyphales, Fasc. 13: 857. 1914.

(See *Fusarium lateritium*)

Holotypus: Not located.

Type locality: **Italy**, Pavia.

Type substrate: Branches of *Styphnolobium japonicum* (syn. *Sophora japonica*).

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

bulalinum *Fusarium* J.W. Xia et al., *Persoonia* 43: 195. 2019

Holotypus: CBS H-24052.

Ex-type culture: CBS 161.25 = NRRL 26857 = NRRL 26918.

Type locality: **Australia**.

Type substrate: Unknown.

Descriptions and illustrations: See [Xia et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb2*: MN170381; *tef1*: MN170448.

bufonicola *Fusarium* (Speg.) Sacc. & Trotter, *Syll. Fung.* 22: 1486. 1913.

(See *Fusarium graminearum*)

Basionym: *Selenosporium bufonicola* Speg., *Anales Mus. Nac. Buenos Aires*, ser. 3, 13: 458. 1910.

Holotypus: In LPS (Myc. Argent. ser. 5, no. 1166) *fide* [Farr \(1973\)](#).

Type locality: **Argentina**, Buenos Aires.

Type substrate: Decaying body of *Amphibia* (toad).

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

bugnicourtii *Fusarium* Brayford, *Trans. Brit. Mycol. Soc.* 89: 350. 1987.

(See *Fusarium ambrosium*)

Synonym: *Fusarium tumidum* var. *coeruleum* Bugnic., *Encycl. Mycol.* 11: 83. 1939.

Holotypus: IMI 296597.

Ex-type culture: IMI 296597 = NRRL 20438 = MAFF 246291.

Type locality: **India**, Chinchona.

Type substrate: *Euwallacea fornicatus* on *Camellia sinensis*.

Descriptions and illustrations: See [Brayford \(1987\)](#).

Diagnostic DNA barcodes: *rpb1*: JX171470; *rpb2*: JX171584; *tef1*: AF178332.

buharicum *Fusarium* Jacz. ex Babajan & Teteravn.-Babajan, *Mater. Mikol. Fitopatol.* 8: 216. 1929.

Holotypus: LEP 127667.

Epitypus (*hic designatus*, MBT 10000662): **Uzbekistan**, Tashkent, on *Gossypium herbaceum*, 1928, A.I. Raillo, CBS 178.35 (preserved as metabolically inactive culture).

Ex-epitype culture: CBS 178.35 = DSM 62166 = NRRL 25488.

Descriptions and illustrations: See [Gerlach & Nirenberg \(1982\)](#).

Diagnostic DNA barcodes: *rpb1*: KX302920; *rpb2*: KX302928; *tef1*: KX302912.

Notes: [Gerlach & Nirenberg \(1982\)](#) designated CBS 178.35 as neotype of *F. buharicum* as they were unable to locate the type specimen. However, A. Jaczweski did place a specimen in LEP. Therefore, the neotype designation is superseded here (Art. 9.13) and CBS 178.35 is retained as epitype for this species.

bulbicola *Fusarium* Nirenberg & O'Donnell, *Mycologia* 90: 452. 1998.

Replaced synonym: *Fusarium sacchari* var. *elongatum* Nirenberg, *Mitt. Biol. Bundesanst. Land- Forstw. Berlin-Dahlem* 169: 59. 1976, *non* *Fusarium elongatum* Reinking 1934.

Holotypus: IMI 202877.

Ex-type culture: BBA 63628 = CBS 220.76 = DAOM 225114 = IMI 375322 = NRRL 13618.

Type locality: **Germany**.

Type substrate: *Haemanthus* sp.

Descriptions and illustrations: See [Nirenberg \(1976\)](#), [Nirenberg & O'Donnell \(1998\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: KF466394; *rpb2*: KF466404; *tef1*: AF160294.

bulbigenum *Fusarium* Cooke & Masee, *Grevillea* 16: 49. 1887.

(See *Fusarium oxysporum*)

Holotypus: ?K(M).

Type locality: **UK**.

Type substrate: *Narcissus* sp.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

bullatum *Fusarium* Sherb., *Mem. Cornell Univ. Agric. Exp. Sta.* 6: 198. 1915.

(See *Fusarium equiseti*)

Typus: CUP-007455.

Type locality: **USA**, Iowa.

Type substrate: Rotten tuber of *Solanum tuberosum*. Lectotypification pending study of material lodged in CUP.

burgessii *Fusarium* M.H. Laurence et al., *Fungal Diversity* 49: 109. 2011.

Holotypus: CBS 125537 (preserved as metabolically inactive culture).

Ex-type culture: CBS 125537 = NRRL 66654 = RBG 5315.

Type locality: **Australia**, Queensland, Idalia National Park.

Type substrate: Soil.

Descriptions and illustrations: See [Laurence et al. \(2011\)](#).

Diagnostic DNA barcodes: *rpb1*: MT409440; *rpb2*: HQ646393; *tef1*: HQ667148.

butleri *Fusarium* Wollenw., *Phytopathology* 3: 38. 1913, *nom. illegit.*, Art. 52.1.

(See *Fusarium udum*)

Authentic material: ?B.

Original locality: **India**.

Original substrate: *Cajanus cajan*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

butleri *Fusarium* Kr.P. Singh & Edward, *Allahabad Farmer* 49: 94. 1979, *nom. illegit.*, Art. 53.1, *non* *Fusarium butleri* Wollenw. 1913.

Synonym: *Gibberella butleri* Kr.P. Singh & Edward, *Allahabad Farmer* 49: 92. 1979.

Authentic material: Not located.

Original locality: **India**.

Original substrate: *Cajanus cajan*.

Notes: Status unclear. No further records available for this taxon.

buxi *Fusarium* Spreng., *Syst. Veg.*, ed. 16, 4: 565. 1827.

Pseudonectria buxi (DC.) Seifert et al., *Stud. Mycol.* 68: 107. 2011.

Basionym: *Tubercularia buxi* DC., *Fl. Franç.*, ed. 3, 5/6: 110. 1815.

Synonyms: *Fusisporium buxi* (DC.) Fr., *Syst. Mycol.* 3: 447. 1832, *nom. sanct.* [Fr., l.c.]

- Psilonia buxi* (DC.) Fr., Syst. Mycol. 3: 447. 1832, *nom. inval.*, Art. 36.1(c).
- Chaetostroma buxi* (DC.) Corda, Icon. Fung. 2: 31. 1838.
- Volutella buxi* (DC.) Berk. & Broome, Ann. Mag. Nat. Hist., ser. 2, 5: 465. 1850.
- Chaetodochium buxi* (DC.) Höhn., Mitt. Bot. Inst. T. H. Wien 9: 45. 1932.
- Nectria rousseliana* Mont., Ann. Sci. Nat., Bot., sér. 3, 16: 44. 1851.
- Stigmathea rousseliana* (Mont.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 97. 1870.
- Nectriella rousseliana* (Mont.) Sacc., Syll. Fung. 2: 452. 1883.
- Lasionectria rousseliana* (Mont.) Cooke (as '*rousseliana*'), Grevillea 12: 111. 1884.
- Pseudonectria rousseliana* (Mont.) Wollenw., Z. Parasitenk. (Berlin) 3: 489. 1931.
- Notariella rousseliana* (Mont.) Clem. & Shear, The genera of Fungi: 280. 1931.
- Nectria rousseliana* var. *viridis* Berk. & Broome, Ann. Mag. Nat. Hist., ser. 3, 3: 376. 1859.
- Volutella buxi* f. *rusci* Sacc., Michelia 2: 644. 1882.
- Holotypus*: ?PC.
- Type locality*: ?Germany/France.
- Type substrate*: Leaf of *Buxus* sp.
- buxicola* *Fusarium* Sacc., Syll. Fung. 2: 518. 1883.
- Cyanonectria buxi*** (Fuckel) Schroers *et al.*, Stud. Mycol. 68: 120. 2011.
- Basionym*: *Gibbera buxi* Fuckel, Jahrb. Nassauischen Vereins Naturk. 27–28: 32. 1874.
- Synonyms*: *Lisea buxi* (Fuckel) Sacc., Syll. Fung. 2: 518. 1883.
- Gibberella buxi* (Fuckel) Cooke, Grevillea 12: 112. 1884.
- Fusarium subcorticale* Oudem., Ned. Kruidk. Arch., sér. 3, 3: 135. 1898.
- Fusarium dimorphum* J.V. Almeida & Sousa da Câmara, Revista Agron. (Lisbon) 1: 306. 1903.
- Fusarium buxicola* var. *chlamydosporum* Batikyan (as '*chlamydosporae*'), Biol. Zhurn. Armenii 22: 90. 1969.
- Fusarium lateritium* var. *buxi* C. Booth, The Genus *Fusarium*: 113. 1971.
- Lectotypus*: G 00111019, selected in Schroers *et al.* (2011).
- Epitypus*: CBS H-20379, designated in Schroers *et al.* (2011).
- Ex-epitype culture*: CBS 125551.
- Epitype locality*: Slovenia, Arboretum Volčji Potok.
- Epitype substrate*: Decaying twig of *Buxus sempervirens* var. *elegantissima*.
- Descriptions and illustrations*: See Schroers *et al.* (2011).
- Diagnostic DNA barcodes*: *rpb2*: HM626689; *tef1*: HM626648.
- byssinum* *Fusarium* McAlpine, Proc. Linn. Soc. New South Wales 22: 698. 1897.
- Holotypus*: VPRI 2556.
- Type locality*: Australia, New South Wales, Murwillumbah.
- Type substrate*: *Desmodium* sp.
- Notes*: Status unclear. This species was considered a member of *Diymopsis* by Saccardo (1899); *Hymenula* by Wollenweber & Reinking (1935); and *Aschersonia* by Walker (1962), who examined the type specimen and found that the fungus occurred in association with a scale insect on *Desmodium*. It is likely that this species belongs in *Microcera*, which are usually parasites of scale insects.
- caapi* *Fusarium*** M.M. Costa *et al.*, Mycol. Progr. 20: 67. 2021.
- Holotypus*: UB 24189.
- Ex-type culture*: CML 3657.
- Type locality*: Brazil, São Paulo, Guaira.
- Type substrate*: *Brachiaria brizantha*.
- Descriptions and illustrations*: See Costa *et al.* (2021).
- Diagnostic DNA barcodes*: *rpb2*: MT901316; *tef1*: MT901350.
- caatingaense* *Fusarium*** A.C.S. Santos *et al.*, Mycologia 111: 248. 2019.
- Holotypus*: URM 91192.
- Ex-type culture*: MUM 1859 = URM 6779.
- Type locality*: Brazil, Pernambuco, Ibimirim.
- Type substrate*: *Dactylopius opuntiae*.
- Descriptions and illustrations*: See Santos *et al.* (2019).
- Diagnostic DNA barcodes*: *rpb2*: LS398495; *tef1*: LS398466.
- cactacearum* *Fusarium*** Pasin. & Buzz.-Trav., Nuovo Giorn. Bot. Ital. 42: 120. 1935.
- Lectotypus* (*hic designatus*, MBT 10000663): Italy, Milan, *Theolocactus nidulans*, 1935, L. Pasinetti & A. Buzzati-Traverso, in Nuovo Giorn. Bot. Ital. 42: Pl. I, fig. 1.
- Descriptions and illustrations*: See Pasinetti & Buzzati-Traverso (1935).
- Notes*: Based on illustrations by Pasinetti & Buzzati-Traverso (1935), this species could be a synonym of *Neocosmospora solani* but requires further investigation. No holotype specimen could be located and therefore an illustration is designated as lectotype.
- cacti-maxonii* *Fusarium*** Pasin. & Buzz.-Trav., Nuovo Giorn. Bot. Ital. 42: 120. 1935.
- Lectotypus* (*hic designatus*, MBT 10000664): Italy, Milan, *Cactus maxonii*, 1935, L. Pasinetti & A. Buzzati-Traverso, in Nuovo Giorn. Bot. Ital. 42: Pl. I, fig. 4.
- Descriptions and illustrations*: See Pasinetti & Buzzati-Traverso (1935).
- Notes*: Based on illustrations by Pasinetti & Buzzati-Traverso (1935), this species could be a synonym of *Fusarium oxysporum* but requires further investigation. No holotype specimen could be located and therefore an illustration is designated as lectotype.
- caeruleum* *Fusarium* Lib. ex Sacc. (as '*cæruleum*'), Syll. Fung. 4: 705. 1886.
- Synonyms*: *Fusarium solani* var. *caeruleum* (Lib. ex Sacc.) Bilař, Fusarii (Biologija i sistematika): 287. 1955, *nom. inval.*, Art. 41.5.
- Fusarium solani* var. *caeruleum* (Lib. ex Sacc.) C. Booth, The Genus *Fusarium*: 51. 1971.
- ?*Fusarium violaceum* Fuckel, Fungi Rhen. Exs., Fasc. 3: no. 209. 1863.
- Fusarium aeruginosum* Delacr., Bull. Soc. Mycol. France 7: 110. 1891.
- Selenosporium caeruleum* Lib., 1834. (in herb.; *nom. inval.*, Art. 38.1a).
- Fusarium caeruleum* var. *cellulosae* Sartory *et al.*, Papier 38: 43. 1935.
- ?*Hypomyces asclepiadis* Zerova, Zhurn. Inst. Bot. Vseukraïns'k. Akad. Nauk 11: 103. 1937.
- Holotypus*: BR5020140171069.
- Type locality*: Belgium.
- Type substrate*: *Solanum tuberosum*.
- Notes*: Status doubtful. See Sandoval-Denis *et al.* (2019).
- calcareum* *Fusarium* (Thüm.) Sacc., Syll. Fung. 4: 712. 1886.

(See *Fusarium oxysporum*)

Basionym: Fusisporium calcareum Thüm., Inst. Coimbra 28: 262. 1881.

Holotypus: S-F45605.

Type locality: **Portugal**, Coimbra.

Type substrate: *Lagenaria vulgaris*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

?*calidarium* *Fusarium* Sacc., Ann. Mycol. 4: 274. 1906.

Colletotrichum anthurii Delacr., Bull. Soc. Mycol. France 13: 110. 1897.

Synonyms: *Fusoma calidarium* Sacc., Ann. Mycol. 4: 274. 1906.

Fusoma calidarium var. *acanthi* Lindegg, Riv. Patol. Veg. 25: 233. 1935.

Holotypus: In PAD.

Type locality: **Italy**, Padua, botanical garden.

Type substrate: *Anthurium scherzerianum*.

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#). No record could be located for the transfer of this epithet to the genus *Fusarium*. In [Saccardo \(1906\)](#) on p. 274, no new combination is provided and only the new name *Fusoma calidarium* was introduced. Similarly, [Lindegg \(1935\)](#) introduced a new variety as *Fusoma calidarium* var. *acanthi*, not in the genus *Fusarium*. Although [Wollenweber & Reinking \(1935\)](#) did treat this as *Fusoma*, [Booth \(1971\)](#) incorrectly treated the variety *acanthi* in the genus *Fusarium*.

callistephi Fusarium L. Lombard & Crous, Persoonia 43: 15. 2018 [2019].

Holotypus: CBS H-23608.

Ex-type culture: CBS 187.53 = NRRL 36330.

Type locality: **Netherlands**, Oostenbrink.

Type substrate: *Callistephus chinensis*.

Descriptions and illustrations: See [Lombard et al. \(2019b\)](#).

Diagnostic DNA barcodes: *rpb2*: MH484875; *tef1*: MH484966.

callosporium Fusarium Pat., Bull. Soc. Mycol. France 9: 164. 1893.

(See *Fusarium coccophilum*)

Holotypus: Not located.

Type locality: **Ecuador**, Quito.

Type substrate: Parasitic on *Septobasidium pedicellatum*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

camerunense Fusarium Henn., Bot. Jahrb. Syst. 22: 81. 1895.

Gloeosporium camerunense (Henn.) Wollenw., Fusaria Autogr. Delin. 1: 499. 1916.

Holotypus: In B *fide* [Hein \(1988\)](#).

Type locality: **Cameroon**, Itoki.

Type substrate: Bark of unknown tree.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

camptoceras Fusarium Wollenw. & Reinking, Phytopathology 15: 158. 1925.

Neotypus: CBS H-24077, designated in [Xia et al. \(2019\)](#).

Ex-neotype culture: ATCC 16065 = ATCC 24364 = BBA 9810 = CBS 193.65 = DSM 62167 = IMI 112500 = NRRL 20716 = NRRL 36344.

Neotype locality: **Costa Rica**.

Neotype substrate: Cushion gall of *Theobroma cacao*.

Descriptions and illustrations: See [Wollenweber & Reinking \(1935\)](#), [Booth \(1971\)](#), [Gerlach & Nirenberg \(1982\)](#), [Marasas et al. \(1998\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: MW928800; *rpb2*: MN170383; *tef1*: MN170450.

campylopodii Fusarium Weir, Mycologia 60: 374. 1968, *nom. inval.*, Art. 38.1(a).

Authentic material: Not located.

Original locality: **USA**, Washington.

Original substrate: *Arceuthobium* sp.

Note: The name is mentioned but neither a diagnosis nor a description was provided.

candidulum Fusarium Sacc., Ann. Mycol. 6: 567. 1908.

(See *Fusarium oxysporum*)

Holotypus: Not located.

Type locality: **Mexico**.

Type substrate: *Myrtillocactus geometrizans*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

candidum Fusarium Ehrenb., Sylv. Mycol. Berol.: 24. 1818.

Neonectria candida (Ehrenb.) Rossman et al., Stud. Mycol. 80: 217. 2015.

Synonyms: *Ramularia candida* (Ehrenb.) Wollenw., Phytopathology 1: 220. 1913.

Cylindrocarpon ehrenbergii Wollenw., Fusaria Autogr. Delin. 1: 461. 1916.

Sclerotium castaneum Lib., in herb. 1832, *nom. nud.*

Fusarium castaneum Lindau (as "(Lib.) Lindau"), Rabenh. Krypt.-Fl. 1(9): 556. 1909.

?*Fusidium candidum* Willk., Die mikroskopischen Feinde des Waldes 1: 103. 1866, *nom. illegit.*, Art. 53.1.

?*Fusarium candidum* Sacc. & D. Sacc., Syll. Fung. 18: 674. 1906, *nom. illegit.*, Art. 53.1.

?*Fusarium candidum* Dasz., Bull. Soc. Bot. Genève, 2 sér. 4: 293. 1913, *nom. illegit.*, Art. 53.1.

Fusarium brassicae Lib. ex Cooke, Grevillea 8: 83. 1880.

Selenosporium brassicae Thüm., Hedwigia 19: 191. 1880.

Fusarium brassicae (Thüm.) Sacc., Syll. Fung. 4: 701. 1886, *nom. illegit.*, Art. 53.1.

Fusarium obtusiusculum Sacc., Michelia 2: 297. 1881.

Fusarium rhizogenum Pound & Clem., Bot. Surv. Nebraska 3: 12. 1894.

Fusarium oxysporum var. *obtusiusculum* (Sacc.) Cif., Ann. Bot. (Rome) 16: 221. 1924.

Cylindrocarpon obtusiusculum (Sacc.) U. Braun, Cryptog. Bot. 4: 113. 1993.

Fusarium eichleri Bres., Ann. Mycol. 1: 130. 1903.

Neonectria ramulariae Wollenw., Ann. Mycol. 15: 52. 1917.

Nectria ramulariae (Wollenw.) E. Müll., Beitr. Kryptogamenfl. Schweiz 11: 634. 1962.

Cylindrocarpon magnusianum Wollenw., Z. Parasitenk. (Berlin) 1: 172. 1928.

Holotypus: Not located.

Type locality: **Germany**, Berlin.

Type substrate: Unknown.

candidum Fusarium (Link) Sacc., Syll. Fung. 4: 720. 1886, *nom. illegit.*, Art. 53.1.

Neonectria ditissima (Tul. & C. Tul.) Samuels & Rossman, CBS Biodiversity Ser. 4: 134. 2006.

Basionym: *Nectria ditissima* Tul. & C. Tul., Select. Fung. Carpol. 3: 73. 1865.

Synonyms: *Cucurbitaria ditissima* (Tul. & C. Tul.) Kuntze, Revis. Gen. Pl. 3: 461. 1898.

- Fusidium candidum* Link, Mag. Neuesten Entdeck. Gesamnten Naturk. Ges. Naturf. Freunde Berlin 3: 8. 1809, *nom. sanct.* [Fr., Syst. Mycol. 3: 481. 1832].
- Cylindrocarpon candidum* (Link) Wollenw., Fusaria Autogr. Delin. 1: 476. 1916.
- ?*Fusisporium cylindricum* Mont., Ann. Sci. Nat., Bot., sér. 2, 17: 120. 1842.
- ?*Fusarium cylindricum* (Mont.) Sacc., Syll. Fung. 4: 720. 1886.
- Fusarium fissum* Peyl, Lotos 8: 30. 1858.
- ?*Fusarium heteronemum* Berk. & Broome (as 'heteronema'), Ann. Mag. Nat. Hist., Ser. 3, 15: 402. 1865.
- ?*Cylindrocarpon heteronema* (Berk. & Broome) Wollenw. (as 'heteronemum'), Fusaria Autogr. Delin. 1: 460. 1916.
- ?*Ramularia heteronema* (Berk. & Broome) Wollenw. (as 'heteronemum'), Fusaria Autogr. Delin. 1: 460. 1916.
- Fusarium ulmi* P. Crouan & H. Crouan, Fl. Finistère: 14. 1867.
- Fusarium fragrans* P. Crouan & H. Crouan, Fl. Finistère: 14. 1867.
- Fusarium decipiens* Cooke & Masee, in Cooke, Handb. Austral. Fungi: 388. 1892, *nom. inval.*, Art. 39.1.
- Fusarium mali* Allesch., Ber. Bot. Vereines Landshut 12: 130. 1892.
- Fusarium sarcochroum f. mali* (Allesch.) Ferraris, 1910.
- Cylindrocarpon mali* (Allesch.) Wollenw., Phytopathology 18: 225. 1928.
- Sporotrichum amenti* P. Karst., Hedwigia 31: 296. 1892.
- Fusarium fractum* Sacc. & Cavara, Nuovo Giorn. Bot. Ital., n.s. 7: 308. 1900.
- Cylindrocarpon fractum* (Sacc. & Cavara) Wollenw., Fusaria Autogr. Delin. 2: 655. 1924.
- Nectria galligena* Bres., in Strasser, Verh. K. K. Zool.-Bot. Ges. Wien 51: 413. 1901.
- Dialonectria galligena* (Bres.) Petch ex E.W. Mason & Grainger, Cat. Yorkshire Fung.: 32. 1937.
- Neonectria galligena* (Bres.) Rossman & Samuels, Stud. Mycol. 42: 159. 1999.
- Fusarium prunorum* McAlpine, Fungus Diseases of stone-fruit trees in Australia: 91. 1902.
- Fusarium willkommii* Lindau, Rabenh. Krypt.-Fl. ed. 2, 1(9): 551. 1909.
- Cylindrocarpon willkommii* (Lindau) Wollenw., Z. Parasitenk. (Berlin) 1: 150. 1928.
- Fusarium luteum* Parav., Ann. Mycol. 16: 302. 1918, *nom. illegit.*, Art. 53.1.
- Nectria ditissima* var. *arctica* Wollenw., Angew. Bot. 8: 189. 1926.
- Cylindrocarpon candidum* var. *medium* Wollenw., Z. Parasitenk. (Berlin) 1: 158. 1928.
- Cylindrocarpon candidum* var. *majus* Wollenw., Z. Parasitenk. (Berlin) 1: 158. 1928.
- Cylindrocarpon candidum* var. *minus* Wollenw., Z. Parasitenk. (Berlin) 1: 155. 1928.
- Cylindrocarpon mali* var. *flavum* Wollenw., Z. Parasitenk. (Berlin) 1: 150. 1928.
- Cylindrocarpon willkommii* var. *pluriseptatum* Wollenw., Z. Parasitenk. (Berlin) 1: 152. 1928.
- Cylindrocarpon willkommii* var. *minus* Wollenw., Z. Parasitenk. (Berlin) 1: 152. 1928.
- Holotypus*: Not located.
- Type locality*: **Unknown**.
- Type substrate*: Branch.
- Notes*: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#). Several names that include *Fusidium candidum* (1809), *Fusisporium cylindricum* (1842) and *Fusarium fissum* (1858) should take preference for this taxon. However, the epithet "candidum" is already occupied in the genus *Neonectria* and cannot be used. Furthermore, the link between *Fusisporium cylindricum* and *Fusarium fissum* with *Neonectria ditissima* still needs to be established. Therefore, we choose to retain the name *Neonectria ditissima* for this taxon.
- candidum Fusarium* Sacc. & D. Sacc., Syll. Fung. 18: 674. 1906, *nom. illegit.*, Art. 53.1, *non Fusarium candidum* Ehrenb. 1818.
- Basionym*: *Fusidium candidum* Willk., Die mikroskopischen Feinde des Waldes 1: 103. 1866.
- Replacing synonym*: *Fusarium willkommii* Lindau, Rabenh. Krypt.-Fl. ed. 2, 1(9): 551. 1910.
- (See *Fusarium willkommii*)
- capitatum Fusarium* Schwein., Trans. Amer. Philos. Soc., n.s., 4: 302. 1832.
- Synonym*: *Pionnotes capitata* (Schwein.) Fr., Summa Veg. Scand. 2: 481. 1849.
- Holotypus*: PH00081394.
- Type locality*: **USA**, Pennsylvania.
- Type substrate*: *Tsuga canadensis*.
- Notes*: The type material of *Fusarium capitatum*, type species of the genus *Pionnotes*, was re-examined by [Seifert \(2013\)](#). It represents not a hyphomycete but a basidiomycete identical to *Dacrymyces chrysospermus*. Therefore, the generic name *Pionnotes* is a synonym of *Dacrymyces* rather than *Fusarium*. Further evaluations are necessary in future phylogenetic revisions of the *Dacrymycetales*.
- caricis Fusarium* Oudem., Verslagen Meded. Afd. Natuurk. Kon. Akad. Wetensch., ser. 3, 7: 325. 1890.
- (See [Fusarium graminearum](#))
- Holotypus*: ?L.
- Type locality*: **Netherlands**, Zuid-Holland Province, Den Haag.
- Type substrate*: Leaves of *Carex* sp.
- Note*: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).
- caries Fusarium* Nees, Nova Acta Phys.-Med. Acad. Caes. Leop.-Carol. Nat. Cur. 19, Suppl. 1: 478. 1843.
- Holotypus*: ?B, L or STR.
- Type locality*: **China**.
- Type substrate*: *Meoschium lodiculare*.
- Notes*: Status unclear. Not *Fusarium* *fide* [Wollenweber & Reinking \(1935\)](#).
- carminascens Fusarium*** L. Lombard *et al.*, Persoonia 43: 19. 2018 [2019].
- Holotypus*: CBS H-23609.
- Ex-type culture*: CBS 144738 = CPC 25800.
- Type locality*: **South Africa**, KwaZulu-Natal Province.
- Type substrate*: *Zea mays*.
- Descriptions and illustrations*: See [Lombard *et al.* \(2019b\)](#).
- Diagnostic DNA barcodes*: *rpb1*: MW928801; *rpb2*: MH484937; *tef1*: MH485028.
- carneolum Fusarium* P. Karst., Meddeland. Soc. Fauna Fl. Fenn. 16: 35. 1888.
- Vermicularia herbarum*** Westend., Herb. Crypt. Belg. no. 393. 1849.
- Holotypus*: ?H.
- Type locality*: **Finland**, Tammela.
- Type substrate*: *Iris pseudacorus*.
- Note*: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

carneoroseum *Fusarium* Cooke, Grevillea 19: 4. 1890.

(See ***Fusarium lateritium***)

Holotypus: In K(M), Colenso 538 *vide* Index Fungorum.

Type locality: **New Zealand**.

Type substrate: Bark.

Note: Synonym *vide* [Wollenweber & Reinking \(1935\)](#).

carneum *Fusarium* (Mont.) Sacc., Syll. Fung. 4: 724. 1886.

Basionym: *Fusisporium carneum* Mont., Ann. Sci. Nat., Bot., sér. 2, 17: 120. 1842.

Holotypus: ?PC.

Type locality: **Cuba**.

Type substrate: Leaf of monocotyledon.

Notes: Status unclear. Not *Fusarium* *vide* [Wollenweber & Reinking \(1935\)](#).

cariforme *Fusarium* Ellis & Tracy, *nom. inval.*, Art. 38.1(a).

Synonym: *Ramularia cariformis* Sherb., Phytopathology 18: 149. 1928.

Authentic material: NY0093683.

Original locality: **USA**, Mississippi, Starkville.

Original substrate: *Tripsacum dactyloides*.

Notes: Status unclear. [Braun \(1998\)](#) considered this species doubtful as conidia appeared microdochium-like.

carpineum Fusarium Davis, Trans. Wisconsin Acad. Sci. 18: 106. 1915.

Holotypus: BPI 442722.

Type locality: **USA**, Wisconsin, Wyalusing.

Type substrate: *Carpinus caroliniana*.

Notes: This species was not treated by any of [Wollenweber & Reinking \(1935\)](#), [Booth \(1971\)](#), or [Gerlach & Nirenberg \(1982\)](#). A literature search could not find any additional information pertaining to this species.

carpini *Fusarium* Schulzer & Sacc., Hedwigia 23: 128. 1884.

(See ***Fusarium expansum***)

Holotypus: Not located.

Type locality: **Croatia**, Vinkovci.

Type substrate: *Carpinus betulus*.

Note: Synonym *vide* [Wollenweber & Reinking \(1935\)](#).

cassiae Fusarium R.H. Perera *et al.*, Mycosphere 11: 2138. 2020.

Holotypus: MFLU 18-2751.

Ex-type culture: MFLUCC 18-0573.

Type locality: **Thailand**, Phayao Province.

Type substrate: Pods of *Cassia fistula*.

Descriptions and illustrations: See [Perera *et al.* \(2020\)](#).

Diagnostic DNA barcodes: *rpb2*: MT212197; *tef1*: MT212205.

castagnei *Fusarium* Mont., Ann. Sci. Nat., Bot., sér. 3, 12: 296. 1849.

Myxosporium castagnei (Mont.) Wollenw., Fusaria. Autogr. Delin. 1: 489. 1916.

Holotypus: ?PC.

Type locality: **France**, Marseille.

Type substrate: *Psoralea bituminosa*.

Note: Synonym *vide* [Wollenweber & Reinking \(1935\)](#).

castaneicola *Fusarium* W. Yamam., Trans. Mycol. Soc. Japan 3: 114. 1962, *nom. inval.*, Art. 39.1 & 40.1.

Rugonectria castaneicola (W. Yamam. & Oyasu) Hirooka & P. Chaverri, Stud. Mycol. 68: 73. 2011.

Basionym: *Nectria castaneicola* W. Yamam. & Oyasu, Sci. Rep. Hyogo Univ. Agric. 3: 15. 1957.

Synonyms: *Neonectria castaneicola* (W. Yamam. & Oyasu) Tak. Kobay. & Hirooka, J. Gen. Pl. Pathol. 71: 126. 2005, *nom. inval.*, Art. 41.5.

Cylindrocarpon castaneicola Tak. Kobay. & Hirooka, J. Gen. Pl. Pathol. 71: 126. 2005.

Authentic material: Not designated.

Original locality: **Japan**, Hyogo.

Original substrate: *Castanea crenata*.

Note: This *Fusarium* epithet is invalid as neither a Latin diagnosis (Art. 39.1) nor a type designation (Art. 40.1) was included in the original description.

castaneum *Fusarium* Lindau (as "(Lib.) Sacc."), Rabenh. Krypt.-Fl. ed. 2, 1(9): 556. 1909.

Synonym: *Sclerotium castaneum* Lib., in herb. 1832, *nom. nud.* (See *Fusarium candidum* Ehrenb.)

Authentic material: Not located.

Original locality: **Belgium**, Ardennes.

Original substrate: *Brassica oleracea*.

cataleptum *Fusarium* Cooke & Harkn., Grevillea 12: 96. 1884.

(See *Fusarium coccophilum*)

Holotypus: In K(M), Harkness 1981 *vide* Index Fungorum.

Type locality: **USA**, California, San Rafael.

Type substrate: *Acacia* sp.

Note: Synonym *vide* [Wollenweber & Reinking \(1935\)](#).

catenatum *Fusarium* (Sand.-Den. & Crous) O'Donnell *et al.*, Index Fungorum. 440: 1. 2020.

Neocosmospora catenata Sand.-Den. & Crous, Persoonia 41: 115. 2018.

Holotypus: CBS H-23225.

Ex-type culture: CBS 143229 = NRRL 54993 = UTHSC 09-1009.

Type locality: **USA**, Georgia.

Type substrate: *Stegostoma fasciatum*.

Descriptions and illustrations: See [Sandoval-Denis & Crous \(2018\)](#).

Diagnostic DNA barcodes: *rpb1*: KC808292; *rpb2*: KC808355; *tef1*: KC808214.

cateniforme Fusarium J.W. Xia *et al.*, Persoonia 43: 197. 2019

Holotypus: CBS H-24053.

Ex-type culture: ATCC 11853 = CBS 150.25.

Type locality: **Unknown**.

Type substrate: Unknown.

Descriptions and illustrations: See [Xia *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb2*: MN170384; *tef1*: MN170451.

catenulatum Fusarium F.J. Chen, Mycosystema 19: 459. 2000.

Holotypus: HMAS 71749.

Ex-type culture: AS 3.4704.

Type locality: **China**, Shaanxi, Yangling.

Type substrate: *Gossypium hirsutum*.

Descriptions and illustrations: See [Chen \(2000\)](#).

caucasicum *Fusarium* Letov, Mater. Mikol. Fitopatol. 8: 225. 1929.

Holotypus: Not located.

Ex-type culture: CBS 179.35 = IFO 5979 = NRRL 13954.

Type locality: **Republic of Azerbaijan**.

Type substrate: *Gossypium hirsutum*.

Descriptions and illustrations: See [Gerlach & Nirenberg \(1982\)](#).

Notes: Status doubtful/unclear. The ex-type culture (CBS 179.35) accessioned in CBS appears to be either contaminated or transpositioned by another *Fusarium* sp. (Sandoval-Denis et al. 2019). A sequence of the *tef1* gene region (DQ247543) from the copy accessioned at NRRL (NRRL 13954) places this species within the *Neocosmospora falciformis* clade (Sandoval-Denis et al. 2019). The status of the copy accessioned at IFO is not known.

caudatum Fusarium Wollenw., J. Agric. Res. 2: 262. 1914.
(See *Fusarium scirpi*)

Lectotypus (*hic designatus*, MBT 10000665): **USA**, South Carolina, Clemson College, *Ipomoea batatas*, date unknown, Harter & Field, in Wollenweber, J. Agric. Res. 2: 262, pl. 16, fig. M.
Notes: Synonym *vide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

cavispermum Fusarium Corda, Icon. Fung. 1: 3. 1837.

Cosmosporella cavisperma (Corda) Sand.-Den. et al., Stud. Mycol. 98 (no. 100116): 44. 2021.

Synonyms: *Fusarium aquaeductuum* var. *cavispermum* (Corda) Raillo, Fungi of the Genus *Fusarium*: 280. 1950.

Fusarium oxydendri Ellis & Everh., Bull. Torrey Bot. Club 24: 477. 1897.

Fusarium cavispermum var. *minus* Wollenw., Fusaria Autogr. Delin. 3: 848. 1930.

Lectotypus: AKJ. Corda, Icon. Fung. 1: pl. I, fig. 58, designated in this study.

Type locality: **Czech Republic**.

Type substrate: Resin of *Pinus* sp.

Epitypus: CBS 172.31 (metabolic inactive specimen) designated in this study.

Ex-epitype: CBS 172.31 = NRRL 13996.

Epitype locality: **Norway**.

Epitype substrate: *Pinus sylvestris*.

Diagnostic DNA barcodes: *rpb1*: JX171465; *rpb2*: JX171579.

celosiae Fusarium T. Abe, Mem. Coll. Agric. Kyoto Imp. Univ. 7: 51. 1928.

(See *Fusarium fujikuroi*)

Holotypus: Not located.

Type locality: **Japan**.

Type substrate: Living stems and leaves of *Celosia cristata*.

Note: Synonym *vide* Wollenweber & Reinking (1935).

celtidicola Fusarium Q.J. Shang et al., Phytotaxa 361: 255. 2018.

Holotypus: MFLU 15-3646.

Ex-type culture: KUMCC 16-0019 = MFLUCC 16-0526.

Type locality: **Italy**, Forlì-Cesena Province, Forlì, Viale dell'Appennino.

Type substrate: *Celtis australis*.

Descriptions and illustrations: See Shang et al. (2018).

Diagnostic DNA barcodes: *rpb1*: MH576579; *rpb2*: MH576577; *tef1*: MH576581.

celtidis Fusarium Ellis & Tracy, J. Mycol. 6: 76. 1890.

(See *Fusarium lateritium*)

Syntypes: In BPI, ISC & MICH.

Type locality: **USA**, Mississippi, Starkville.

Type substrate: *Celtis occidentalis*.

Note: Synonym *vide* Wollenweber & Reinking (1935).

celtidis Fusarium Pass., Atti Reale Accad. Lincei, Rendiconti Cl. Sci. Fis., 4 sér. 7: 51. 1891, *nom. illegit.*, Art. 53.1.

Replacing synonym: *Fusarium sphaeriiforme* Sacc. (as 'sphaeriaeforme'), Syll. Fung. 10: 723. 1892.

(See *Fusarium melanochlorum*)

Holotypus: ?PARMA.

Type locality: **Italy**, Parma, Vigheffio.

Type substrate: Dead branch of *Celtis australis*.

Note: Synonym *vide* Wollenweber & Reinking (1935).

cepaee Fusarium Hanzawa, Mycol. Centralbl. 5(1): 5. 1914.

(See *Fusarium oxysporum*)

Lectotypus (*hic designatus*, MBT 10000666): **Japan**, Sapporo, *Allium cepa*, 1914, J. Hanzawa, 5(1): 6, fig. 1.

Notes: Synonym *vide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

cerasi Fusarium Rolland & Ferry, in Roumeguère, Rev. Mycol. (Toulouse) 14: 170. 1892.

?*Foveostroma drupacearum* (Lév.) DiCosmo, Canad. J. Bot. 56: 1682. 1978.

Basionym: *Micropera drupacearum* Lév., Ann. Sci. Nat., Bot., sér. 3, 5: 283. 1846.

Synonyms: ?*Peziza cerasi* Pers., Neues Mag. Bot. 1: 115. 1794.

?*Dermea cerasi* (Pers.) Fr., Syst. Orb. Veg. 1: 115. 1825.

Syntype: ILL00220294 (Fungi Sel. Gall. Exs. No. 6119).

Type locality: **France**, Saint-Dié-des-Vosges.

Type substrate: *Prunus* sp. (cherry tree).

Note: This species was excluded from *Fusarium* by Wollenweber (1943). Gerlach & Nirenberg (1982) considered this species as a possible synonym of *Micropera drupacearum* on which the present synonymies are based.

cerealis Fusarium (P. Karst.) Gruyter & J.H.M. Schneid., Jaarb. Plantenziektenk. Dienst 1989/1990, no. 168: 135. 1991, *nom. inval.*, Art. 41.4.

Gliomastix cerealis (P. Karst.) C.H. Dickinson, Mycol. Pap. 115: 19. 1968.

Basionym: *Coniosporium cerealis* P. Karst., Meddeland. Soc. Fauna Fl. Fenn. 14: 109. 1887.

Synonyms: *Acremonium cerealis* (P. Karst.) W. Gams, *Cephalosporium-artige Schimmelpilze* (Stuttgart): 88. 1971.

Gliomastix guttuliformis J.C. Br. & W.B. Kendr., Trans. Brit. Mycol. Soc. 41: 499. 1958.

Holotypus: In herb. P.A. Karsten in H *vide* Dickinson (1968).

Type locality: **Finland**, Mustiala.

Type substrate: *Secale cereale*.

cerealis Fusarium (Cooke) Sacc., Syll. Fung. 4: 713. 1886.

Basionym: *Fusisporium cerealis* Cooke, Grevillea 6: 139. 1878.

Synonym: *Fusarium culmorum* var. *cerealis* (Cooke) Wollenw., Fusaria Autogr. Delin. 3: 946. 1930.

Fusarium roseum f. *cerealis* (Cooke) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 32: 663. 1945.

Gibberella rosea f. *cerealis* (Cooke) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 32: 664. 1945.

Fusarium sambucinum var. *cerealis* (Cooke) Raillo, Fungi of the Genus *Fusarium*: 211. 1950.

Fusarium crookwellense L.W. Burgess et al., Trans. Brit. Mycol. Soc. 79: 498. 1982.

Holotypus: ?K(M) 133541.

Type locality: **USA**, California, Gainesville.

Type substrate: *Zea mays*.

Notes: [Wollenweber & Reinking \(1935\)](#) considered *F. cerealis* as a variety of *F. culmorum*, whereas [Raiilo \(1950\)](#) considered it as a variety of *F. sambucinum*. [Gerlach & Nirenberg \(1982\)](#) applied a broader concept to *F. culmorum* that did not separate this variety in *F. culmorum*. [Nirenberg \(1990\)](#) recognised *F. cerealis* as a species and considered *F. crookwellense* as a synonym of *F. cerealis*. However, [Leslie & Summerell \(2006\)](#) recommend the use of the name *F. crookwellense* over *F. cerealis*, indicating that no type material is available for *F. cerealis*. We choose to follow [Nirenberg \(1990\)](#) to consider *F. crookwellense* a synonym under *F. cerealis*. The material lodged in K(M) requires further investigation to determine whether epi- or neotypification is required.

cesatii *Fusarium* Rabenh., Klotzschii Herb. Viv. Mycol. Cent. 15: no. 1440. 1850.

Hymenula rubella Fr., Elench. Fung. 2: 38. 1828.

Lectotypus (of *Fusarium cesatii*, *hic designatus*, MBT 10000667):

Italy, Vercelli, *Carex* sp., 1849, collector unknown, Rabenh., Klotzschii Herb. Viv. Mycol. Ed. I no. 1440 in HAL.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

cesatii *Fusarium* Thüm., Pilze Weinst.: 49. 1878, *nom. illegit.*, Art. 53.1, *non Fusarium cesatii* Rabenh. 1850.

Elsinoe ampelina (de Bary) Shear, Phytopathology 19: 677. 1929.

Basionym: *Sphaceloma ampelina* de Bary, Ann. Oenol. 4: 165. 1874.

Synonyms: *Manginia ampelina* (de Bary) Viala & Pacottet, C. r. hebdom. Séanc. Acad. Sci., Paris 139: 88. 1904.

Pionnotes cesatii Sacc., Syll. Fung. 4: 726. 1886.

Ramularia ampelophaga Pass., Bol. Comit. Agric. Parmense 9: 125. 1876.

Gloeosporium ampelophagum (Pass.) Sacc., Michelia 1: 217. 1878.

Authentic material: S-F47363.

Original locality: **Italy**, Vercelli.

Original substrate: Decaying stump of *Vitis vinifera*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

chaetomium *Fusarium* Wallr., Fl. Crypt. Germ. 2: 242. 1833.

Colletotrichum chaetomium (Wallr.) S. Hughes, Canad. J. Bot. 36: 753. 1958.

Holotypus: ?STR.

Type locality: **Germany**.

Type substrate: Decaying *Cucurbita*.

chenopodium *Fusarium* (Thüm.) Sacc., Syll. Fung. 4: 701. 1886.

(See ***Fusarium scirpi***)

Basionym: *Fusisporium chenopodium* Thüm., Mycoth. Univ. Cent. 14: no. 1378. 1879.

Syntypes: In BPI, CHR, ILL, NEB, NY, NYS & PUL.

Type locality: **Austria**, Niederösterreich, Klosterneuburg.

Type substrate: *Chenopodium album*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

chilense *Fusarium* (Mont.) Sacc., Syll. Fung. 4: 716. 1886.

Gloeosporium chilense (Mont.) Wollenw., Z. Parasitenk. (Berlin) 3: 496. 1931.

Basionym: *Fusisporium chilense* Mont., in Gay, Fl. Chil. 8: 25. 1852.

Fusisporium argillaceum Mont., Bull. Mass. Agric. Exp. Sta. no. 55. 1842, *nom. illegit.*, Art. 53.1, *non Fusarium argillaceum* Fr. 1832.

Holotypus: In UPS *fide* Wollenweber, Fusaria Autogr. Delin. 2: 658.

Type locality: **Chile**, Juan Fernández Islands.

Type substrate: Bark of *Urtica excelsa*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

chinhoyiense Fusarium Yilmaz & Crous, Persoonia 46: 147. 2021.

Holotypus: PREM 63215.

Ex-type culture: BBA 69031 = DAOM 225149 = Frank 5bCn8 = IMI 375355 = NRRL 25221 = NY007.12.

Type locality: **Zimbabwe**, Chinhoyi.

Type substrate: *Zea mays*.

Descriptions and illustrations: See [Yilmaz et al. \(2021\)](#).

Diagnostic DNA barcodes: *rpb1*: MW402711; *rpb2*: MN534262; *tef1*: MN534050.

chlamydosporum Fusarium Wollenw. & Reinking, Phytopathology 15: 156. 1925.

Synonyms: *Fusarium sporotrichioides* var. *chlamydosporum* (Wollenw. & Reinking) Joffe, Mycopathol. Mycol. Appl. 53: 211. 1974, *nom. inval.*, Art. 41.1.

Dactylium fusarioides Gonz. Frag. & Cif., Bol. Real Soc. Esp. Hist. Nat. 27: 280. 1927.

Fusarium fusarioides (Gonz. Frag. & Cif.) C. Booth, The Genus *Fusarium*: 88. 1971.

Pseudofusarium purpureum Matsush., *Microfungi of the Solomon Islands and Papua New Guinea*: 47. 1971.

Neotypus: CBS 145.25 (preserved as metabolically inactive culture), designated in [Lombard et al. \(2019a\)](#).

Ex-neotype culture: CBS 145.25 = NRRL 26851 = NRRL 26912.

Neotype locality: **Honduras**, Tela.

Neotype substrate: *Musa sapientum*.

Descriptions and illustrations: See [Wollenweber & Reinking \(1925\)](#), [Booth \(1971\)](#), [Gerlach & Nirenberg \(1982\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: MN120715; *rpb2*: MN120735; *tef1*: MN120754.

cicatricum *Fusarium* (Berk.) O'Donnell & Geiser, Phytopathology 103: 404. 2013.

Geejayessia cicatricum (Berk.) Schroers, Stud. Mycol. 68: 124. 2011.

Basionym: *Sphaeria sanguinea* var. *cicatricum* Berk., Mag. Zool. Bot. 1: 48. 1837.

Synonyms: *Nectria cicatricum* (Berk.) Tul. & C. Tul., Select. Fung. Carpol. 3: 77. 1865.

Sphaeria sanguinea var. *cicatricum* Haller, Syst. Nat., ed 13, 1: LII. 1768.

Sphaeria coccinea var. *cicatricum* Desm., Ann. Sci. Nat., Bot., sér. 3, 10: 351. 1848.

Nectria coccinea var. *cicatricum* (Desm.) Sacc., Syll. Fung. 2: 482. 1883.

Cucurbitaria cicatricum (Desm.) Kuntze, Revis. Gen. Pl. 3: 462. 1898.

Nectria gibbera Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 177. 1870.

Fusarium fuckelii Sacc., Syll. Fung. 4: 695. 1886.

Nectria desmazieri Fuckel ex Sacc., Syll. Fung. 4: 695. 1886, *nom. inval.*, Art. 36.1(d).

Lectotypus: K(M) 160064 (MBT 10001323 *hic designatus*).

Epitypus: CBS H-20374 (MBT 10001324 *hic designatus*).

Ex-epitype culture: CBS 125549.

Epitype locality: **Slovenia**, Arboretum Volčji Potok.

Epitype substrate: Decaying twigs of *Buxus sempervirens*.

Descriptions and illustrations: See [Schroers et al. \(2011\)](#).

Diagnostic DNA barcodes: *rpb1*: KM232231; *rpb2*: HM626679; *tef1*: HM626643.

Notes: The epitypification in [Schroers et al. \(2011\)](#) was not Code compliant as neither a supporting holo-, lecto- nor epitype was cited. The specimen in the Kew herbarium was cited as isotype. In the protologue a single gathering is mentioned, but an illustration is also cited so a lectotypification is necessary. The epitypification is validated herein.

ciliatum *Fusarium* (Link) Link, in Willdenow, Sp. Pl., Ed. 4, 6: 105. 1825.

Scolecofusarium ciliatum (Link) L. Lombard et al., Stud. Mycol. 98 (no. 100116): 74. 2021.

Basionym: *Atractium ciliatum* Link, Mag. Neuesten Entdeck. Gesammten Naturk. Ges. Naturf. Freunde Berlin 7: 32. 1816.

Synonyms: *Microcera ciliata* (Link) Wollenw., Fusaria Autogr. Delin. 1: 435. 1916.

Calonectria ciliata (Link) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 32: 664. 1945.

Sphaeria agnina Desm., Ann. Sci. Nat., Bot. sér. 3, 6: 72. 1846.

Calonectria agnina (Desm.) Sacc., Michelia 1(3): 311. 1878.

Dialonectria agnina (Desm.) Cooke, Grevillea 12: 111. 1884.

Fusarium peltigeriae Westend., Herb. Crypt. Belg. 9: no. 414. 1849.

Fusarium parasiticum Westend., Bull. Séances Cl. Sci. Acad. Roy. Sci. Belgique, sér. 2, 11: 652. 1861.

Nectria massariae Pass., in Rabenhorst, Fungi Eur. Exs. no. 1827. 1874.

Microcera massariae Sacc., Michelia 1(2): 262. 1878.

Calonectria massariae (Pass.) Sacc., Michelia 1(3): 312. 1878.

Fusisporium filisporum Cooke, Grevillea 8: 8. 1879.

Fusarium filisporum (Cooke) Sacc., Syll. Fung. 4: 708. 1886.

Fusarium scolecoides Sacc. & Ellis, Atti Reale Ist. Veneto Sci. Lett. Arti, sér. 6, 3: 728. 1885.

Fusarium elongatum Cooke, Grevillea 19: 4. 1890.

Calonectria dearnessii Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 42: 245. 1891.

Neotypus: CBS H-12687 designated in this study.

Ex-neotypus: ATCC 16068 = ATCC 24137 = BBA 9661 = CBS 191.65 = DSM 62172 = IMI 112499 = NRRL 20431.

Neotype locality: **Germany**.

Neotype substrate: Branch canker of *Fagus sylvatica*.

Diagnostic DNA barcodes: *rpb1*: MW834264; *rpb2*: MW834035; *tef1*: MW834296.

cinctum *Fusarium* Corda, Icon. Fung. 5: 80. 1842.

Striaticonidium cinctum (Corda) L. Lombard & Crous, Persoonia 36: 229. 2016.

Synonyms: *Myrothecium cinctum* (Corda) Sacc., Syll. Fung. 4: 751. 1886.

?*Myrothecium ellipsosporum* Fuckel (as '*ellipsisporium*'), Fungi Rhen. Exs. Cent. 16, no. 1529 (1865).

?*Hymenopsis ellipsospora* (as '*ellipsosporum*') (Fuckel) Sacc., Syll. Fung. 4: 745. 1886.

Myrothecium striatisporum N.C. Preston, Trans. Brit. Mycol. Soc. 31: 275. 1948.

Myrothecium longistriatisporum Matsush., Microfungi Solomon Isl. Papua-New Guinea: 39. 1971.

Lectotypus: PR 155489, designated in [Tulloch \(1972\)](#).

Epitypus: CBS H-22471, designated in [Lombard et al. \(2016\)](#).

Ex-epitype culture: CBS 932.69 = IMI 145760.

Epitype locality: **Netherlands**, Eastern Flevoland.

Epitype substrate: Agricultural soil.

Note: The lectotype was cited as holotype in [Lombard et al. \(2016\)](#) but this is correctable according to Art. 9.10 of the Code (see also Ex. 11).

cinnabarinum *Fusarium* (Berk. & M.A. Curtis) Sacc., Syll. Fung. 4: 722. 1886.

(See ***Fusarium lateritium***)

Basionym: *Fusisporium cinnabarinum* Berk. & M.A. Curtis, Grevillea 3: 146. 1875.

Syntypes: In PH, Pul & USCH:Fungi (Ellis, N. Amer. F. 3990).

Type locality: **USA**, Alabama.

Type substrate: *Acer negundo*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

circinatum* *Fusarium Nirenberg & O'Donnell, Mycologia 90: 442. 1998.

Synonyms: *Gibberella circinata* Nirenberg & O'Donnell, Mycologia 90: 440. 1998, *nom. inval.*, Art. 40.3.

Gibberella circinata Nirenberg & O'Donnell ex Britz et al., Sydowia 54: 16. 2002.

Holotypus: B 70 0001693.

Ex-type culture: BBA 69720 = CBS 405.97 = DAOM 225113 = IMI 375321 = NRRL 25331.

Type locality: **USA**, California.

Type substrate: *Pinus radiata*.

Descriptions and illustrations: See [Nirenberg & O'Donnell \(1998\)](#).

Diagnostic DNA barcodes: *rpb1*: JX171510; *rpb2*: JX171623; *tef1*: AF160295.

cirrosom *Fusarium* Höhn., Sitzungsber. Kaiserl. Akad. Wiss. Wien, Math.-Naturwiss. Cl., Abt. 1., 116: 153. 1907.

(See ***Fusarium expansum***)

Holotypus: FH00284266.

Type locality: **Austria**, Niederösterreich, Irenental near Untertullnerbach.

Type substrate: Parasitic in the acervuli of *Steganosporium pyriforme* (syn. *Steganosporium ovatum*).

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

citri* *Fusarium M.M. Wang et al., Persoonia 43: 79. 2019.

Holotypus: HAMS 248036.

Ex-type culture: CGMCC 3.19467 = LC6896.

Type locality: **China**, Hunan Province.

Type substrate: Leaves of *Citrus reticulata*.

Descriptions and illustrations: See [Wang et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MK289828; *rpb2*: MK289771; *tef1*: MK289617.

citricola* *Fusarium Guarnaccia et al., Persoonia 40: 12. 2017. [2018].

Holotypus: CBS H-23020.

Ex-type culture: CBS 142421 = CPC 27805.

Type locality: **Italy**, Cosenza, Rocca Imperiale.

Type substrate: *Citrus reticulata* 'Caffin'.

Descriptions and illustrations: See [Sandoval-Denis et al. \(2018a\)](#).

Diagnostic DNA barcodes: *rpb1*: LT746290; *rpb2*: LT746310; *tef1*: LT746197.

citriforme *Fusarium* Jamal., Valt. Maatalousk. Julk. 123: 11. 1943.

(See *Fusarium tricinctum*)

Lectotypus (*hic designatus*, MBT 10000668): **Finland**, Pyhäjärvi, *Hordeum sativum*, 1938, E. Jamalainen, in Valt. Maatalousk. Julk. 123: 10. 1943, fig. 2.

Ex-type culture: CBS 253.50.

Diagnostic DNA barcodes: *rpb1*: MW928802; *rpb2*: MW928823; *tef1*: KR071775.

Notes: Jamalainen (1943) cited various specimens in the protologue of *F. citriforme*, but failed to indicate a holotype. Therefore, a lectotypification is done here to fix the name. Isolate CBS 253.50 was deposited in the public collection of CBS by E. Jamalainen in 1950. The isolate was indicated as the living ex-type culture of *F. citriforme*.

citrinum *Fusarium* Wollenw., in Lewis, Bull. Maine Agric. Exp. Sta. 219: 256. 1913.

(See *Fusarium oxysporum*)

Lectotypus (*hic designatus*, MBT 10000669): **Germany**, Berlin, Dahlem, rotten fruit of *Solanum lycopersicum*, Oct. 1910, H.W. Wollenweber, B70 0100185.

Notes: Synonym *fide* Wollenweber & Reinking (1935). Only one specimen located at B matches the original collection event, but it is not indicated as the type. Therefore B 70 0100185 is designated as lectotype here.

citrulli *Fusarium* Taubenh., Bull. Texas Agric. Exp. Sta. 260: 27. 1920.

(See *Fusarium oxysporum*)

Lectotypus (*hic designatus*, MBT 10000670): **USA**, Texas, Waller County, seedlings of *Citrullus lanatus*, 1920, J.J. Taubenhous, in Bull. Texas Agric. Exp. Sta. 260: 30, fig. 8h.

Notes: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

citrulli *Fusarium* Sartory, Compt. Rend. Hebd. Séances Acad. Sci. 188: 1434. 1929, *nom. inval.*, Art. 35.2; *nom. illegit.*, Art. 53.1.

Neocosmospora martii (Appel & Wollenw.) Sand.-Den. & Crous, Persoonia 43: 142. 2019.

Basionym: *Fusarium martii* Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land-Forstw. 8: 83. 1910.

Synonyms: *Fusarium solani* var. *martii* (Appel & Wollenw.) Wollenw., Fusaria Autogr. Delin. 3: 1034. 1930.

?*Selenosporium fuscum* Bonord., Handb. Mykol.: 135. 1851.

?*Fusarium fuscum* (Bonord.) Sacc., Syll. Fung. 4: 699. 1886.

Fusarium citrulli Sartory & J. Mey., Compt. Rend. Soc. Biol. 107: 55. 1931, *nom. illegit.*, Art. 53.1, *non Fusarium citrulli* Taubenh. 1920.

Neocosmospora croci Guarnaccia *et al.*, Persoonia 40: 17. 2017 [2018].

Authentic material: Not located.

Original locality: **France**.

Original substrate: *Citrullus vulgaris*.

Note: Synonyms *fide* Wollenweber & Reinking (1935) and Sandoval-Denis *et al.* (2019).

clavatum *Fusarium* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 234. 1915.

(See *Fusarium flocciferum*)

Lectotypus (*hic designatus*, MBT 10000671): **USA**, New York, Castile, rotten tuber of *Solanum tuberosum*, 1915, C.D. Sherbakoff, in Mem. Cornell Univ. Agric. Exp. Sta. 6: 235, fig. 40.

Notes: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

clavus *Fusarium* J.W. Xia *et al.* (as '*clavum*'), Persoonia 43: 199. 2019.

Holotypus: CBS H-24054.

Ex-type culture: CBS 126202 = RMF N 38.

Type locality: **Namibia**, northern Karoo, 30 km west of Maltahöhe.

Type substrate: Desert soil.

Descriptions and illustrations: See Xia *et al.* (2019).

Diagnostic DNA barcodes: *rpb2*: MN170389; *tef1*: MN170456.

clematidis *Fusarium* Rolland & Fautrey, Rev. Mycol. (Toulouse) 16: 72. 1894.

Macroconia sphaeriae (Fuckel) Gräfenhan & Schroers, Stud. Mycol. 68: 103. 2011.

Basionym: *Fusarium sphaeriae* Fuckel, Fungi Rhen. Exs. Fasc. 3: no. 212. 1863.

Synonyms: *Fusarium sphaeriae* var. *robustum* Davis, Trans. Wisconsin Acad. Sci. 19: 714. 1919.

Septogloeum robustum (Davis) Wollenw. & Reinking, *Fusarien*: 336. 1935.

?*Nectria leptosphaeriae* var. *macrospora* Wollenw., Angew. Bot. 8: 187. 1926.

Syntype: ILL00220727 (Roumeguère, Fungi Sel. Gall. Exs. no. 6537).

Type locality: **France**.

Type substrate: *Clematis vitalba*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

clypeaster *Fusarium* (Corda) Sacc., Syll. Fung. 4: 706. 1886.

Septogloeum clypeaster (Corda) Wollenw., *Fusarien*: 321. 1935.

Basionym: *Fusisporium clypeaster* Corda, Icon. Fung. 4: 26. 1840.

Lectotypus (*hic designatus*, MBT 10000672): **Czech Republic**, *Phragmites*, May 1839, A.C.J. Corda, in Icon. Fung. 4, Tab. 6, fig. 82. 1840.

Notes: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

coccidicola *Fusarium* Henn. (as '*coccideicola*'), Bot. Jahrb. Syst. 34: 57. 1904.

Microcera diploa (Berk. & M.A. Curtis) Gräfenhan & Seifert, Stud. Mycol. 68: 106. 2011.

Basionym: *Nectria diploa* Berk. & M.A. Curtis, J. Linn. Soc., Bot. 10: 378. 1868.

Synonyms: *Cucurbitaria diploa* (Berk. & M.A. Curtis) Kuntze, Revis. Gen. Pl. 3: 461. 1898.

Creonectria diploa (Berk. & M.A. Curtis) Seaver, Mycologia 1: 190. 1909.

Calonectria diploa (Berk. & M.A. Curtis) Wollenw., Angew. Bot. 8: 193. 1926.

Cosmospora diploa (Berk. & M.A. Curtis) Rossman & Samuels, Stud. Mycol. 42: 121. 1999.

Fusarium derridis Henn., Beibl. Hedwigia 41: (66). 1902.

Fusarium juruanum Henn., Hedwigia 43: 398. 1904.

- Fusarium pentaclethrae* Henn., Hedwigia 44: 71. 1905.
Aschersonia henningsii Koord., Verh. Kon. Ned. Akad. Wetensch., Afd. Natuurk. 13: 213. 1907.
Microcera henningsii (Koord.) Petch, Ann. Roy. Bot. Gard. (Peradeniya) 5: 533. 1914.
Pseudomicrocera henningsii (Koord.) Petch, Trans. Brit. Mycol. Soc. 7: 100. 1921.
Microcera fujikuroi Miyabe & Sawada, J. Fac. Agric. Hokkaido Imp. Univ. 5: 83. 1913.
Microcera merrillii Syd. & P. Syd., Ann. Mycol. 12(6): 576. 1914.
Pseudomicrocera henningsii var. *longispora* Petch, Trans. Brit. Mycol. Soc. 7: 164. 1921.
Fusarium microcera Bilal, Fusarii (Biologija i sistematika): 292. 1955, *nom. inval.*, Art. 39.1.
Holotypus: Zimmerman no. 26 in B fide [Hein \(1988\)](#).
Type locality: **Tanzania**, East Usambara, Magrotto.
Type substrate: Parasitic on *Coccoidea* sp. on *Camellia sinensis*.
- coccinellum* *Fusarium* Kalchbr., Flora (Regensburg) 59: 426. 1876.
(See *Fusarium coccophilum*)
Syntype: ?NY00899913.
Type locality: **South Africa**, Eastern Cape Province, Somerset-East.
Type substrate: *Acacia horrida*.
Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).
- coccineum* *Fusarium* Schwein., Trans. Amer. Philos. Soc., n.s. 4: 302. 1834.
Holotypus: ?PH00062490.
Type locality: **USA**, Pennsylvania, Northampton, Nazareth.
Type substrate: Bark of *Castanea* sp.
Notes: Status unclear. Not *Fusarium* fide [Wollenweber & Reinking \(1935\)](#).
- coccophilum* *Fusarium* (Desm.) Wollenw. & Reinking, *Fusarien*: 34. 1935.
Microcera coccophila Desm., Ann. Sci. Nat. Bot., sér. 3, 10: 359. 1848.
Synonyms: *Tubercularia coccophila* (Desm.) Bonord., Abh. Naturf. Ges. Halle 8: 96. 1864.
Fusarium episphaeria f. *coccophilum* (Desm.) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 32: 662. 1945.
Nectria episphaeria f. *coccophila* (Desm.) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 32: 662. 1945.
Fusarium coccinellum Kalchbr., Flora (Regensburg) 56: 426. 1876.
Fusisporium coccinellum (Kalchbr.) Kalchbr., in Thümen, Mycoth. Univ. no. 782. 1877.
Fusarium cataleptum Cooke & Harkn., Grevillea 12: 96. 1884.
Microcera pluriseptata Cooke & Masee, Grevillea 17: 43. 1888.
Fusarium callosporium Pat., Bull. Soc. Mycol. France 9: 164. 1893.
Fusarium baccharidicola Henn., Hedwigia 48: 20. 1908.
Microcera coccophila var. *platyspora* Sousa da Câmara, Revista Agron. (Lisbon): 5 (extr.). 1920.
Lectotypus: K(M) 165807, designated in [Gräfenhan et al. \(2011\)](#).
Type locality: **France**, Normandy, near Caen.
Type substrate: Parasitic on *Eulecanium tiliae* on *Salix* sp. and *Fraxinus excelsior*.
Descriptions and illustrations: See [Gräfenhan et al. \(2011\)](#).
Notes: No living type material available. [Gräfenhan et al. \(2011\)](#) designated a lectotype but did not designate an epitype, which is still required.
- coffeatum* *Fusarium*** L. Lombard & Crous, Fungal Syst. Evol. 4: 191. 2019.
Replaced synonym: *Fusarium chlamydosporum* var. *fuscum* Gerlach, Phytopathol. Z. 90: 41. 1977.
Holotypus: BBA 62053.
Isotypus: CBS H-631.
Ex-type culture: BBA 62053 = CBS 635.76 = NRRL 20841.
Type locality: **South Africa**.
Type substrate: *Cynodon lemfuensis*.
Descriptions and illustrations: See [Gerlach \(1977a\)](#), [Gerlach & Nirenberg \(1982\)](#) and [Xia et al. \(2019\)](#).
Diagnostic DNA barcodes: *rpb1*: MN120717; *rpb2*: MN120736; *tef1*: MN120755.
- coffeicola* *Fusarium* Henn., Bot. Jahrb. Syst. 22: 82. 1895.
Synonym: *Gloeosporium coffeicola* (P. Henn.) Wollenw., Fusaria Autogr. Delin. 1: 493. 1916, *nom. illegit.*, Art. 53.1, *non Gloeosporium coffeicola* Tassi 1900.
Holotypus: In B fide [Hein \(1988\)](#).
Type locality: **Cameroon**, Victoria.
Type substrate: *Coffea liberica*.
Notes: Status unclear. Not *Fusarium* fide [Wollenweber & Reinking \(1935\)](#).
- coicis* *Fusarium*** Johanssen et al., Fungal Diversity 77: 356. 2015 [2016].
Holotypus: RBG 5368.
Ex-type culture: FRL 19329 = NRRL 66233 = RBG 5368.
Type locality: **Australia**, Queensland, Mareeba.
Type substrate: *Coix gasteenii*.
Descriptions and illustrations: See [Laurence et al. \(2016\)](#).
Diagnostic DNA barcodes: *rpb1*: KP083269; *rpb2*: KP083274; *tef1*: KP083251.
- colorans* *Fusarium* (De Jonge) Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land-Forstw. 8: 39. 1913.
Albonectria rigidiuscula (Berk. & Broome) Rossman & Samuels, Stud. Mycol. 42: 105. 1999.
Basionym: *Nectria rigidiuscula* Berk. & Broome, J. Linn. Soc., Bot. 14: 116. 1873 [1875].
Synonyms: *Calonectria rigidiuscula* (Berk. & Broome) Sacc., Michelia 1(3): 313. 1878.
Fusarium rigidiusculum (Berk. & Broome) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 32: 664. 1945.
Calonectria eburnea Rehm, Hedwigia 37: 196. 1898.
Calonectria lichenigena Speg., Bol. Acad. Nac. Ci. Republ. Argent. 11: 530. 1889.
Calonectria sulcata Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 25: 29. 1899.
Fusarium decemcellulare Brick, Jahresber. Vereinigung Angew. Bot. 6: 227. 1908.
Spicaria colorans De Jonge, Recueil Trav. Bot. Néerl. 6: 48. 1909.
Scolecnectria tetraspora Seaver, N. Amer. Fl. 3: 27. 1910.
Calonectria tetraspora (Seaver) Sacc. & Trotter, Syll. Fung. 22: 487. 1913.
Nectria rigidiuscula f. *theobromae* E.J. Ford et al., Phytopathology 57: 712. 1967.
Holotypus: Not located.
Type locality: **Surinam**.
Type substrate: *Theobroma cacao*.
Notes: Synonym fide [Wollenweber & Reinking \(1935\)](#). [Wollenweber \(1916–1935\)](#) indicated that cultures and specimens of *Spicaria colorans* (basium of *F. colorans*) were

deposited in the Willie Commelin Scholten collection in Amsterdam. This collection has been accessioned into the CBS collection (CBS & CBS H). However, no cultures and specimens or records could be located at CBS.

commune Fusarium K. Skovg. *et al.*, Mycologia 95: 632. 2003.
Holotypus: BBA 71639 in B.

Ex-type culture: AAS 156 = BBA 71639 = CBS 110090 = NRRL 31076.

Type locality: **Denmark**.

Type substrate: Soil.

Descriptions and illustrations: See [Skovgaard *et al.* \(2003\)](#).

Diagnostic DNA barcodes: *rpb1*: MW928803; *rpb2*: MW934368; *tef1*: AF362263.

commutatam Fusarium Sacc., Syll. Fung. 4: 710. 1886.

(See *Fusarium solani*)

Replaced synonym: *Fusisporium candidum* Bonord., Handb. Allg. Mykol.: 96 (1851), *nom. illegit.*, Art. 53.1, *non Fusisporium candidum* Link 1824.

Holotypus: Not located.

Type locality: **Germany**.

Type substrate: *Solanum tuberosum*.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#) and [Sandoval-Denis *et al.* \(2019\)](#).

compactum Fusarium (Wollenw.) Raillo, Fungi of the Genus Fusarium: 180. 1950.

Basionym: *Fusarium scirpi* var. *compactum* Wollenw., Fusaria Autogr. Delin. 3: no. 924. 1930.

Synonym: *Fusarium compactum* (Wollenw.) Gordon, Canad. J. Bot. 30: 224. 1952, *nom. inval.*, Art. 53.1.

Lectotypus: Illustration in Wollenweber, Fusaria Autogr. Delin. no. 924 (1930), designated in [Xia *et al.* 2019](#).

Epitypus: CBS 186.31 (preserved as metabolically inactive culture), designated in [Xia *et al.* \(2019\)](#).

Ex-epitype culture: CBS 186.31 = NRRL 36323.

Epitype locality: **UK**, Kew.

Epitype substrate: Cotton thread.

Descriptions and illustrations: See [Wollenweber \(1916–1935, no. 924\)](#), [Raillo \(1950\)](#), [Gordon \(1952\)](#), [Gerlach & Nirenberg \(1982\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb2*: GQ505826; *tef1*: GQ505648.

concentricum Fusarium Nirenberg & O'Donnell, Mycologia 90: 442. 1998.

Holotypus: B 70 0001694.

Ex-type culture: BBA 64354 = CBS 450.97 = DAOM 225146 = IMI 375352 = NRRL 25181.

Type locality: **Costa Rica**.

Type substrate: *Musa sapientum*.

Descriptions and illustrations: See [Nirenberg & O'Donnell \(1998\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: LT996192; *rpb2*: LT575063; *tef1*: AF160282.

concolor Fusarium Reinking, Zentralbl. Bakteriol., 2. Abt. 89: 512. 1934.

Synonym: *Fusarium polyphialidicum* Marasas *et al.*, Mycologia 78: 678. 1986.

Holotypus: IMI 112502.

Ex-type culture: BBA 2607 = BBA 63601 = CBS 183.34 = DAOM 225131 = DSM 62179 = IMI 112502 = NRRL 13994.

Type locality: **Uruguay**, Montevideo.

Type substrate: *Hordeum vulgare*.

Descriptions and illustrations: See [Gerlach & Nirenberg \(1982\)](#) and [Marasas *et al.* \(1986\)](#).

Diagnostic DNA barcodes: *rpb1*: MH742492; *rpb2*: MH742569; *tef1*: MH742650.

conglutinans Fusarium Wollenw., Ber. Deutsch. Bot. Ges. 31: 34. 1913.

(See ***Fusarium oxysporum***)

Holotypus: Not located.

Type locality: **USA**, Wisconsin.

Type substrate: *Brassica oleracea* var. *capitata*.

congoense Fusarium Wollenw., Fusaria Autogr. Delin. 1: 307. 1916.

(See ***Fusarium heterosporum***)

Syntype: BPI 451889.

Type locality: **Democratic Republic of the Congo**.

Type substrate: *Bromus willdenowii*.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

coniosporiicola Fusarium Henn., Ann. Mus. Congo Belge, Bot., Sér. 5, 2: 106. 1907.

Dendrodochium coniosporiicola (Henn.) Hansf., Proc. Linn. Soc. London 155: 60. 1943.

Synonym: *Fusidium coniosporiicola* (Henn.) Wollenw., Fusaria Autogr. Delin. 1: 477. 1916.

Syntypes: In BR & S.

Type locality: **Democratic Republic of the Congo**, Gongolo.

Type substrate: *Albizia* aff. *fastigiata*.

constrictum Fusarium Penz., Michelia 2: 486. 1882.

Synonym: *Ramularia constricta* (Penz.) Wollenw., Fusarien: 322. 1935.

Holotypus: Not located; destroyed *fide* U. Braun.

Type locality: **Italy**, Padua.

Type substrate: Leaves of *Citrus* sp.

Notes: Status unclear. Neither *Fusarium* *fide* [Wollenweber & Reinking \(1935\)](#) nor *Ramularia* (pers. comm. U. Braun).

contaminatum Fusarium L. Lombard & Crous, Persoonia 43: 20. 2018 [2019].

Holotypus: CBS H-23610.

Ex-type culture: CBS 114899.

Type locality: **Germany**, Schlüchtern.

Type substrate: Pasteurised chocolate milk.

Descriptions and illustrations: See [Lombard *et al.* \(2019b\)](#).

Diagnostic DNA barcodes: *rpb2*: MH484901; *tef1*: MH484992.

continuum Fusarium X. Zhou *et al.*, Mycologia 108: 677. 2016.

Holotypus: HMNWAFU NX-Fjpl-10-20100851.

Ex-type culture: CBS 140841 = F201030 = NRRL 66286.

Type locality: **China**, Shaanxi, Fuping, Lei village.

Type substrate: *Zanthoxylum bungeanum*.

Descriptions and illustrations: See [Zhou *et al.* \(2016\)](#).

Diagnostic DNA barcodes: *rpb1*: KM520387; *rpb2*: KM236782; *tef1*: KM236722.

convolutans Fusarium Sand.-Den. *et al.*, MycoKeys 34: 77. 2018.

Holotypus: CBS H-23495.

Ex-type culture: CBS 144207 = CPC 33733.

Type locality: **South Africa**, Kruger National Park, Skukuza, Granite Supersite.

Type substrate: Rhizosphere of *Kyphocarpa angustifolia*.

Descriptions and illustrations: See Sandoval-Denis et al. (2018b).

Diagnostic DNA barcodes: *rpb1*: LT996193; *rpb2*: LT996141; *tef1*: LT996094.

corallinum *Fusarium* Mattir., Atti Accad. Sci. Ist. Bologna, Cl. Sci. Fis., Mem. 6: 677. 1897, *nom. illegit.*, Art. 53.1.

(See *Fusarium culmorum*)

Authentic material: Not located.

Type locality: **Italy**.

Type substrate: *Andropogon* sp.

Note: Synonym *fide* Wollenweber (1931).

corallinum *Fusarium* Sacc., Nuovo Giorn. Bot. Ital. 8: 196. 1876. (See ***Fusarium graminum***)

Holotypus: In PAD.

Type locality: **Italy**, Treviso, Selva.

Type substrate: *Cynodon dactylon*.

cordae *Fusarium* Masee, Brit. Fung.-Fl. 3: 481. 1893.

(See ***Fusarium oxysporum***)

Notes: Masee introduced this name to replace *F. aurantiacum* Corda, indicating that *F. aurantiacum* (Link) Sacc., based on *Fusisporium aurantiacum* Link (1809), predates Corda's use of the epithet. However, Corda's use of the epithet in *Fusarium* predates Saccardo's recombination into *Fusarium*.

cortaderiae* *Fusarium O'Donnell et al., Fungal Genet. Biol. 41: 620. 2004.

Holotypus: BPI 843479.

Ex-type culture: CBS 119183 = ICMP 5435 = NRRL 29297.

Type locality: **New Zealand**, Auckland, Henderson.

Type substrate: *Cortaderia selloana*.

Descriptions and illustrations: See O'Donnell et al. (2004).

Diagnostic DNA barcodes: *rpb1*: KM361644; *rpb2*: KM361662; *tef1*: AY225885.

crassistipitatum *Fusarium* Scandiani et al., Mycoscience 53: 171. 2011.

(See *Fusarium azukiicola*)

Holotypus: BPI 871490.

Ex-type culture: MAFF 239757 = NRRL 36877.

Type locality: **Argentina**, Santa Fe, Zavalla.

Type substrate: *Glycine max*.

Descriptions and illustrations: See Aoki et al. (2012a).

Diagnostic DNA barcodes: *rpb2*: FJ240405; *tef1*: FJ240351.

crassum *Fusarium* (Sand.-Den. & Crous) O'Donnell et al., Index Fungorum 440: 1. 2020.

Neocosmospora crassa Sand.-Den. & Crous, Persoonia 43: 122. 2019.

Holotypus: CBS H-23976.

Ex-type culture: CBS 144386 = MUCL 11420.

Type locality: **France**, Paris.

Type substrate: Unknown.

Descriptions and illustrations: See Sandoval-Denis et al. (2019).

Diagnostic DNA barcodes: *rpb1*: MW218109; *rpb2*: LR583823; *tef1*: LR583604.

croceum* *Fusarium J.W. Xia et al., Persoonia 43: 201. 2019.

Holotypus: CBS H-24055.

Ex-type culture: CBS 131777.

Type locality: **Iran**, Golestan Province, Gonbad-e Qabus.

Type substrate: *Triticum* sp.

Descriptions and illustrations: See Xia et al. (2019).

Diagnostic DNA barcodes: *rpb2*: MN170396; *tef1*: MN170463.

croci *Fusarium* (Guarnaccia, Sand.-Den. & Crous) O'Donnell et al., Index Fungorum 440: 1. 2020.

Basionym: *Neocosmospora croci* Guarnaccia, Sand.-Den. & Crous, Persoonia 40: 17. 2017.

(See *Fusarium citrulli* Sartory)

Holotypus: CBS H-23022.

Ex-type culture: CBS 142423 = CPC 27186.

Type locality: **Italy**, Sicily, Catania, Paternó.

Type substrate: *Citrus sinensis*.

Descriptions and illustrations: See Sandoval-Denis et al. (2018a).

Diagnostic DNA barcodes: *rpb2*: LT746329; *tef1*: LT746216.

cromyophthoron *Fusarium* Sideris, Phytopathology 14: 212. 1924.

(See ***Fusarium oxysporum***)

Lectotypus (*hic designatus*, MBT 10000673): **USA**, California, Stockton, roots of *Allium* sp., 1924, C.P. Sideris, in Phytopathology 14, pl. IX.

Notes: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

crookwellense *Fusarium* L.W. Burgess et al., Trans. Brit. Mycol. Soc. 79: 498. 1982.

(See ***Fusarium cerealis* (Cooke) Sacc.**)

Holotypus: FRC R-3090.

Ex-type culture: NRRL 13163.

Type locality: **Australia**, New South Wales, Crookwell.

Type substrate: *Solanum tuberosum* tubers.

Descriptions and illustrations: See Burgess et al. (1982).

Note: See Notes under *F. cerealis*.

cruentum *Fusarium* Teich, Byull. Sredne-Aziatsk. Gosud. Univ. 19: 178. 1934.

Holotypus: Not located.

Type locality: **Uzbekistan**, Tashkent.

Type substrate: Roots and stems of *Vitis vinifera*.

Notes: Status unclear. This species was not treated by any of Wollenweber & Reinking (1935), Raillo (1950), Bilař (1955), Booth (1971), Joffe (1974), or Gerlach & Nirenberg (1982). Furthermore, no additional records could be located.

cryptoseptatum *Fusarium* (Sand.-Den. & Crous) O'Donnell, Index Fungorum 440: 1. 2020.

Neocosmospora cryptoseptata Sand.-Den. & Crous, Persoonia 43: 122. 2019.

Holotypus: CBS H-23977.

Ex-type culture: BBA 65024 = CBS 145463 = NRRL 22412.

Type locality: **French Guiana**.

Type substrate: Bark.

Descriptions and illustrations: See Sandoval-Denis et al. (2019).

Diagnostic DNA barcodes: *rpb1*: MW834215; *rpb2*: EU329510; *tef1*: AF178351.

cryptum *Fusarium* McAlpine, Fungus Diseases of Citrus trees in Australia: 106. 1899.

(See ***Fusarium larvarum***)

Holotypus: VPRI 2557.

Type locality: **Australia**, South Australia.

Type substrate: Twigs of *Citrus limonia*.

Note: Synonym *vide* [Wollenweber & Reinking \(1935\)](#).

cubense *Fusarium* E.F. Sm., Science, N.Y. 31: 754. 1910.

(See [Fusarium oxysporum](#))

Holotypus: Not located.

Type locality: **Cuba**.

Type substrate: *Musa* sp.

Note: Synonym *vide* [Wollenweber & Reinking \(1935\)](#).

cucumerinum *Fusarium* Berk. & Broome, Ann. Mag. Nat. Hist., ser. 4, 17: 141. 1876.

Holotypus: ?K(M).

Type locality: **UK**, Northamptonshire, Daventry, Sibbertoft.

Type substrate: Diseased *Cucumis sativus*.

Notes: Status unclear. [Wollenweber & Reinking \(1935\)](#) synonymised this species under *Septomyxa persicina*. In contrast, Index Fungorum indicates that this species is a synonym under *F. oxysporum*. The original protologue ([Berkeley & Broome 1876](#)) fits neither *S. persicina* nor *F. oxysporum*.

cucurbitae *Fusarium* Taubenh., Bull. Texas Agric. Exp. Sta. 260: 27. 1920.

Lectotypus (*hic designatus*, MBT 10000674): **USA**, Texas, Waller County, from squash, date unknown, J.J. Taubenhous, in Bull. Texas Agric. Exp. Sta. 260: 30, fig. 8j. 1920.

Notes: Based on the description and illustrations provided by [Taubenhous \(1920\)](#), this species could represent *F. oxysporum*. However, recollection and epitypification are required to confirm this. No holotype specimen could be located and therefore an illustration is designated as lectotype.

cucurbitariae *Fusarium* (Pat.) Sacc., Syll. Fung. 4: 708. 1886.

(See [Fusarium avenaceum](#))

Basionym: *Fusisporium cucurbitariae* Pat., Rev. Mycol. (Toulouse) 3: 10. 1881.

Holotypus: ?FH01093588.

Type locality: **France**, Lons-le-Saunier.

Type substrate: Diseased *Cucumis sativus*.

Note: Synonym *vide* [Wollenweber & Reinking \(1935\)](#).

cucurbitariae *Fusarium* Peyronel, Nuovo Giorn. Bot. Ital., n.s. 25: 436. 1918, *nom. illegit.*, Art. 53.1.

Holotypus: ?ROPV.

Type locality: **Italy**, Piemonte, Ricalaretto.

Type substrate: Parasitic on perithecia of *Camarosporidiella laburni* (= *Cucurbitaria laburni*).

Notes: Status unclear. Not treated by any of [Wollenweber & Reinking \(1935\)](#), [Booth \(1971\)](#), or [Gerlach & Nirenberg \(1982\)](#).

cucurbiticola *Fusarium* O'Donnell *et al.*, Index Fungorum 440: 2. 2020.

[Neocosmospora cucurbitae](#) Sand.-Den. *et al.*, Persoonia 43: 125. 2019.

Synonyms: *Fusarium solani* f. *cucurbitae* W.C. Snyder & H.N. Hansen, Amer. J. Bot. 28: 740. 1941.

Fusarium solani f. sp. *cucurbitae* W.C. Snyder & H.N. Hansen, Root rots caused by Phycomycetes 28: 740. 1941.

Hypomyces solani f. *cucurbitae* W.C. Snyder & H.N. Hansen, Amer. J. Bot. 28: 741. 1941.

Nectria haematococca var. *cucurbitae* (W.C. Snyder & H.N. Hansen) Dingley, New Zealand J. Agric. Res. 4: 337. 1961.

Nectria solani f. *cucurbitae* (W.C. Snyder & H.N. Hansen) G.R.W. Arnold, Z. Pflanzk. 37: 193. 1972.

Holotypus: CBS H-23978.

Ex-type culture: BBA 64411 = CBS 616.66 = NRRL 22399.

Type locality: **Netherlands**.

Type substrate: *Cucurbita viciifolia*.

Descriptions and illustrations: See [Sandoval-Denis *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW834217; *rpb2*: LR583825; *tef1*: DQ247592.

[cugenangense Fusarium](#) Maryani *et al.*, Stud. Mycol. 92: 181. 2018 [2019].

Holotypus: InaCC F984 (preserved as metabolically inactive culture).

Ex-type culture: InaCC F984.

Type locality: **Indonesia**, West Java, Cianjur, Cugenang.

Type substrate: Pseudostem of *Musa* var. Pisang Kepok.

Descriptions and illustrations: See [Maryani *et al.* \(2019a\)](#).

Diagnostic DNA barcodes: *rpb1*: LS479560; *rpb2*: LS479308; *tef1*: LS479757.

[culmorum Fusarium](#) (Wm.G. Sm.) Sacc., Syll. Fung. 10: 726. 1892.

Basionym: *Fusisporium culmorum* Wm.G. Sm., Diseases of field and garden crops, chiefly as are caused by fungi: 209. 1884.

Synonyms: *Fusarium schribauxii* Delacr., Bull. Soc. Mycol. France 6: 99. 1890.

Fusarium corallinum Mattir., Atti Accad. Sci. Ist. Bologna, Cl. Sci. Fis., Mem. 6: 677. 1897, *nom. illegit.*, Art. 53.1.

Fusarium versicolor Sacc., Syll. Fung. 16: 1099. 1902.

Fusarium heidelbergense Sacc., Ann. Mycol. 8: 346. 1910.

?*Fusarium neglectum* Jacz., Bull. Trimestriel Soc. Mycol. France 28: 348. 1912.

Fusarium rubiginosum Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land-Forstw. 8: 108. 1910 [1913].

Fusarium culmorum var. *leteius* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 242. 1915.

Fusarium culmorum var. *majus* Wollenw., Fusaria Autogr. Delin. 2: 613. 1924.

Lectotypus (*hic designatus*, MBT 10000675): **UK**, infected ear of *Triticum* sp., 1884, W.G. Smith, in Diseases of field and garden crops, chiefly as are caused by fungi: 210. fig. 92.

Epitypus (*hic designatus*, MBT 10000676): **Denmark**, moldy kernel of *Hordeum vulgare*, 3 Feb. 1986, U. Thrane, CBS 417.86 (preserved as metabolic inactive culture).

Ex-epitype culture: CBS 417.86 = FRC R-8504 = IMI 309344 = NRRL 25475.

Descriptions and illustrations: See [Wollenweber & Reinking \(1935\)](#), [Booth \(1971\)](#), [Gerlach & Nirenberg \(1982\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: JX171515; *rpb2*: JX171628; *tef1*: MW233082.

Notes: No holotype specimen could be located. Therefore, an illustration is designated as lectotype and CBS 417.86 is designated as epitype as this isolate is commonly used as an authentic strain for *F. culmorum* in literature ([Ward *et al.* 2002](#), [O'Donnell *et al.* 2013, 2020](#), [Geiser *et al.* 2021](#)).

cuneiforme *Fusarium* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 129. 1915.

(See [Fusarium ventricosum](#))

Typus: ?CUP-007474.

Type locality: **USA**, New York.

Type substrate: *Solanum tuberosum*.

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#) and [Booth \(1971\)](#). Lectotypification pending study of material lodged in CUP.

cuneirostrum *Fusarium* O'Donnell & T. Aoki, *Mycoscience* 46: 170. 2005.

(See *Fusarium azukiicola*)

Holotypus: BPI 843353.

Ex-type culture: FRC S-1551 = MAFF 239038 = NRRL 31157.

Type locality: **USA**, Michigan, Presque Isle.

Type substrate: *Phaseolus vulgaris*.

Descriptions and illustrations: See [Aoki et al. \(2005\)](#).

Diagnostic DNA barcodes: *rpb1*: KJ511271; *rpb2*: FJ240389; *tef1*: MAEA01003816.

curvatum* *Fusarium L. Lombard & Crous, *Persoonia* 43: 21. 2018 [2019].

Holotypus: CBS H-23611.

Ex-type culture: CBS 238.94 = NRRL 26422 = PD 94/184.

Type locality: **Netherlands**.

Type substrate: *Beaucarnea* sp.

Descriptions and illustrations: See [Lombard et al. \(2019b\)](#).

Diagnostic DNA barcodes: *rpb1*: MW928804; *rpb2*: MH484893; *tef1*: MH484984.

cuticola *Fusarium* (R. Blanch.) Guég., *Champ. Paras. Homme*: 262. 1904.

(See ***Fusarium oxysporum***)

Basionym: *Selenosporium cuticola* R. Blanch., *Compt. Rend. Hebd. Séances Acad. Sci.* 111: 479. 1890.

Holotypus: Not located.

Type locality: **France**.

Type substrate: Skin of *Chamaeleo vulgaris* and *Lacerta viridis* (lizards).

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#). Based on the substrate, this species could belong to the genus *Bisifusarium*. However, the protologue is not definitive, and recollection from type substrate is needed to confirm its taxonomic position.

cyanescens *Fusarium* (G.A. de Vries et al.) O'Donnell et al., *Index Fungorum* 440: 2. 2020.

Neocosmospora cyanescens (G.A. de Vries et al.) Summerb. et al., *Biology of Microfungi* (Cham): 183. 2016.

Basionym: *Phialophora cyanescens* G.A. de Vries et al., *Antonie van Leeuwenhoek* 50: 150. 1984.

Synonyms: *Cylindrocarpon cyanescens* (G.A. de Vries et al.) Sigler, *J. Clin. Microbiol.* 29: 1858. 1991.

Holotypus: CBS 518.82 (maintained as metabolically inactive culture).

Ex-type culture: CBS 518.82.

Type locality: **Netherlands**, Groningen Province, Groningen.

Type substrate: Subcutaneous tissue of the right foot of a male *Homo sapiens*.

Descriptions and illustrations: See [de Vries et al. \(1984\)](#) and [Zoutman & Sigler \(1991\)](#).

Diagnostic DNA barcodes: *rpb1*: MW218110; *rpb2*: LR583826; *tef1*: LR583605.

cyanostomum *Fusarium* (Sacc. & Flageolet) O'Donnell & Geiser, *Phytopathology* 103: 404. 2013.

Cyanonectria cyanostoma (Sacc. & Flageolet) Samuels & P. Chaverri, *Mycol. Progr.* 8: 56. 2009.

Basionym: *Nectria cyanostoma* Sacc. & Flageolet, *Rendiconto Congr. Bot. Palermo* 1902: 53. 1902.

Lectotypus: BPI 551652, designated in [Samuels et al. \(2009\)](#).

Epitypus: BPI 748307, designated in [Samuels et al. \(2009\)](#).

Ex-epitype culture: BBA 70964 = CBS 101734 = G.J.S. 98-127.

Epitype locality: **France**.

Epitype substrate: *Buxus sempervirens*.

Descriptions and illustrations: See [Samuels et al. \(2009\)](#).

Diagnostic DNA barcodes: *rpb1*: JX171546; *rpb2*: HQ897759; *tef1*: HM626647.

cyclogenum *Fusarium* Sacc., *Nuovo Giorn. Bot. Ital.* 8: 197. 1876.

Gloeosporium orbiculare (Berk.) Berk., *Just's Bot. Jahresber.* 4: 1274. 1876.

Basionym: *Cytospora orbicularis* Berk., *Ann. Nat. Hist.* 1: 207. 1838.

Synonyms: *Myxosporium orbiculare* (Berk.) Berk., *Outl. Brit. Fungol.*: 325. 1860.

Colletotrichum orbiculare (Berk.) Arx, *Verh. Kon. Ned. Akad. Wetensch., Afd. Natuurk., Sect. 2*, 51: 112. 1957, *nom. inval.*, Art. 36.2 (Melbourne).

Sirogloea orbicularis (Berk.) Arx, *Verh. Kon. Ned. Akad. Wetensch., Afd. Natuurk., Sect. 2*, 51: 113. 1957, *nom. inval.*, Art. 36.2 (Melbourne).

Syntypes: In BPI & S.

Type locality: **Italy**, Treviso, Selva.

Type substrate: *Citrullus* sp.

Note: *Cytospora orbicularis* is not a *Colletotrichum* nor a *Fusarium* (small ellipsoidal conidia discharged in tendrils) as outlined in [Damm et al. \(2013\)](#).

cydoniae *Fusarium* Allesch., *Ber. Bot. Vereines Landshut* 12: 130. 1892.

(See ***Fusarium lateritium***)

Holotypus: In M.

Type locality: **Germany**, München.

Type substrate: *Cydonia vulgaris*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

cydoniae *Fusarium* Roum. & Fautrey, *Rev. Mycol. (Toulouse)* 14: 170. 1892, *nom. illegit.*, Art. 53.1, *non* Allescher 1892.

(See *Fusarium rollandianum*)

cydoniae *Fusarium* (Schulzer) Sacc. & Traverso, *Syll. Fung.* 19: 724. 1910, *nom. illegit.*, Art. 53.1, *non* Allescher 1892, *nec* Roum. & Fautrey 1892.

Basionym: *Selenosporium cydoniae* Schulzer, *Verhand. K.K. Zool.-Bot. Ges. Wien* 21: 1240. 1871.

(See ***Fusarium lateritium***)

Holotypus: Not located.

Type locality: **Austria**, Vienna.

Type substrate: *Cydonia vulgaris*.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

cylindricum *Fusarium* (Mont.) Sacc., *Syll. Fung.* 4: 720. 1886.

Basionym: *Fusisporium cylindricum* Mont., *Ann. Sci. Nat., Bot., sér. 2*, 17: 120. 1842.

(See *Fusarium candidum* (Link) Sacc.)

Holotypus: ?PC.

Type locality: **Cuba**.

Type substrate: Sarcocarp of unknown fruit.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

cymbiferum *Fusarium* Berk. & M.A. Curtis, in Berkeley, Grevillea 3: 98. 1875.

Colletotrichum coccodes (Wallr.) S. Hughes, Canad. J. Bot. 36: 754. 1958.

Basionym: *Chaetomium coccodes* Wallr., Fl. Crypt. Germ. 2: 265. 1833.

Synonyms: *Fusarium effusum* Schwein., Trans. Amer. Philos. Soc., n.s. 4: 302. 1832 [1834].

Fusarium georginae Corda, Icon. Fung. 2: 4. 1838.

Vermicularia atramentaria Berk. & Broome, Ann. Mag. Nat. Hist. 5: 378. 1850.

Colletotrichum atramentarium (Berk. & Broome) Taubenh., Mem. New York Bot. Gard. 6: 554. 1916.

Acrothecium solani Sacc., Michelia 1(3): 74. 1877.

Fusisporium elasticae Thüm., Boll. Soc. Adriat. Sci. Nat. Trieste 3: 440. 1877.

Fusarium elasticae (Thüm.) Sacc., Syll. Fung. 4: 711. 1886.

Gloeosporium elasticae Cooke & Massee, in Cooke, Grevillea 18: 74. 1890.

Fusarium foliicola Allesch., Hedwigia 34: 289. 1895.

Gloeosporium foliicola (Allesch.) Wollenw., *Fusarien*: 325. 1935, *nom. illegit.*, Art. 53.1.

Colletotrichum solanicola O'Gara, Mycologia 7: 39. 1915.

Colletotrichum biologicum Chaudhuri, Ann. Bot. 38: 735. 1924.

Holotypus: ?K(M).

Type locality: **USA**.

Type substrate: Stems of some herbaceous plants.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

cypericola *Fusarium* Henn., Hedwigia 48: 116. 1908.

Libertella cypericola (Henn.) Wollenw., *Fusaria Autogr. Delin.* 1: 486. 1916.

Syntype: In B *fide* [Hein \(1988\)](#).

Type locality: **Brazil**, Pará.

Type substrate: *Cyperus exaltatus*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

dactylidis* *Fusarium T. Aoki *et al.*, Mycologia 107: 412. 2015.

Holotypus: BPI 892886.

Ex-type culture: CBS 119181 = ICMP 5269 = NRRL 29298.

Type locality: **New Zealand**, Manawatu, Palmerston North.

Type substrate: *Dactylis glomerata*.

Descriptions and illustrations: See [Aoki *et al.* \(2015\)](#).

Diagnostic DNA barcodes: *rpb1*: KM361654; *rpb2*: KM361672; *tef1*: DQ459748.

decemcellulare *Fusarium* Brick, Jahresber. Vereinigung Angew. Bot. 6: 227. 1908.

(See *Fusarium colorans*)

Holotypus: ?HBG.

Type locality: **Cameroon**.

Type substrate: *Theobroma cacao*.

decipiens *Fusarium* Cooke & Massee, in Cooke, Handb. Austral. Fungi: 388. 1892, *nom. inval.*, Art. 39.1.

(See *Fusarium candidum* (Link) Sacc.)

Authentic material: ?K(M).

Original locality: **Australia**, Queensland.

Original substrate: *Ficus aspera*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

deformans *Fusarium* J. Schröt., Jahresber. Schles. Ges. Vaterl. Cult. 61: 179. 1883.

Gloeosporium deformans (J. Schröt.) Lind, Ann. Bot. 7: 19. 1908.

Synonyms: *Fusamen deformans* (J. Schröt.) P. Karst., Bidrag Kännedom Finlands Natur Folk 51: 485. 1892.

Calogloeum deformans (J. Schröt.) Nannf., Svensk Bot. Tidskr. 25: 25. 1931.

Platycarpium deformans (J. Schröt.) Petr., Sydowia 7: 296. 1953.

Holotypus: In B *fide* [Wollenweber \(1916–1935\)](#).

Type locality: **Poland**, Breslau.

Type substrate: *Salix cinerea*.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

delacroixii *Fusarium* Sacc., Syll. Fung. 10: 725. 1892.

(See ***Fusarium sambucinum***)

Replaced synonym: *Fusarium asparagi* Delacr., Bull. Soc. Mycol. France 6: 99. 1890, *nom. illegit.*, Art. 53.1, *non* *Fusarium asparagi* Briard 1890.

Lectotypus (*hic designatus*, MBT 10000677): **France**, Paris, *Asparagus officinalis*, 1890, M.G. Delaroix, in Bull. Soc. Mycol. France 6, pl. XV. fig. III.

Notes: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#). No holotype material is available for the replaced synonym *F. asparagi* Delacr. and therefore, an illustration from the original protologue is designated as lectotype.

delphinoides *Fusarium* Schroers *et al.*, Mycologia 101: 57. 2009.

Bisifusarium delphinoides (Schroers *et al.*) L. Lombard & Crous, Stud. Mycol. 80: 224. 2015.

Holotypus: CBS H-20124.

Ex-type culture: CBS 120718 = NRRL 53290.

Type locality: **South Africa**, Western Cape Province, Clanwilliam.

Type substrate: *Hoodia gordonii* stem lesions.

Descriptions and illustrations: See [Schroers *et al.* \(2009\)](#).

Diagnostic DNA barcodes: *rpb1*: KM232210; *tef1*: EU926296.

denticulatum* *Fusarium Nirenberg & O'Donnell, Mycologia 90: 445. 1998.

Holotypus: B 70 0001691.

Ex-type culture: BBA 67772 = CBS 407.97 = IMI 376115 = NRRL 25311.

Type locality: **USA**, Louisiana.

Type substrate: *Ipomoea batatas*.

Descriptions and illustrations: See [Nirenberg & O'Donnell \(1998\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: MT010953; *rpb2*: MT010970; *tef1*: KR909385.

derridis *Fusarium* Henn., Beibl. Hedwigia 41: (66). 1902.

(See *Fusarium coccidicola*)

Holotypus: In B *fide* [Hein \(1988\)](#).

Type locality: **Papua New Guinea**.

Type substrate: *Derris* sp.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

desaboruense *Fusarium* N. Maryani *et al.*, Persoonia 43: 59. 2019.

(See ***Fusarium sacchari***)

Holotypus: InaCC F951 (preserved as metabolically inactive culture).

Ex-type culture: InaCC F951.

Type locality: **Indonesia**, East Nusa Tenggara, Sikka Flores, Kecamatan Waigate, Desa Boru.

Type substrate: *Musa* var. Pisang Kepok.

Descriptions and illustrations: See [Maryani *et al.* \(2019b\)](#).

Diagnostic DNA barcodes: rpb1: LS479870; rpb2: LS479852.

desciscens Fusarium Oudem., Ned. Kruidk. Arch., 2 sér., 5: 515. 1889.

(See ***Fusarium sarcochrom***)

Holotypus: ?L.

Type locality: **Netherlands**, Zuid-Holland Province, Den Haag, Scheveningen.

Type substrate: *Sarothamnus vulgaris*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

detonianum Fusarium Sacc. (as 'de-tonianum'), Syll. Fung. 4: 708. 1886, *nom. illegit.*, Art. 52.1.

(See *Fusarium miniatum* Sacc.)

Authentic material: Not located.

Original locality: **Italy**.

Original substrate: Sporangium of *Cyathus vernicosa*.

dianthi Fusarium Prill. & Delacr., Compt. Rend. Hebd. Séances Acad. Sci. 129: 745. 1899.

(See ***Fusarium oxysporum***)

Holotypus: Not located.

Type locality: **France**, Antibes.

Type substrate: *Dianthus caryophyllus*.

didymum Fusarium (Harting) Lindau, Rabenh. Krypt.-Fl. Ed. 2, 1(9): 574. 1909.

Basionym: *Fusisporium didymum* Harting, Nieuwe Verh. Eerste Kl. Kon. Ned. Inst. Wetensch. Amsterdam 12: 228. 1846.

(See *Fusarium eichleri*)

Lectotypus (hic designatus, MBT 10000678): **Netherlands**, *Solanum tuberosum*, date unknown, Harting, in Nieuwe Verh. Eerste Kl. Kon. Ned. Inst. Wetensch. Amsterdam 12 (1846), tab. II, figs 2–4.

Notes: Requires recombination into *Neonectria* after further investigation. No preserved specimen could be located and therefore an illustration is designated as lectotype.

diffusum Fusarium Carmich., Grevillea 16: 81. 1888.

(See ***Fusarium avenaceum***)

Holotypus: ?K(M).

Type locality: **UK**, Scotland, Appin.

Type substrate: Stems of *Asteraceae* (thistle).

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

dimerum Fusarium Penz., *Michelia* 2: 484. 1882.

Bisfusarium dimerum (Penz.) L. Lombard & Crous, *Stud. Mycol.* 80: 225. 2015.

Synonyms: *Fusarium aquaeductuum* var. *dimerum* (Penz.) Raïllo, *Fungi of the Genus Fusarium*: 279. 1950.

Microdochium dimerum (Penz.) Arx, *Trans. Brit. Mycol. Soc.* 83: 374. 1984.

?*Fusisporium flavum* Fr., *Syst. Mycol.* 3: 444. 1832.

?*Pionnotes flava* (Fr.) Sacc., *Syll. Fung.* 4: 726. 1886.

?*Fusarium flavum* (Fr.) Wollenw., *Z. Parasitenk.* 3: 305. 1931.

?*Fusarium aquaeductuum* var. *flavum* (Fr.) Raïllo, *Fungi of the Genus Fusarium*: 280. 1950.

Selenosporium aurantiacum Bonord., *Abh. Naturf. Ges. Halle* 8: 97. 1864, *nom. illegit.*, Art. 53.1.

Fusarium bonordenii Sacc., *Syll. Fung.* 4: 699. 1886.

Fusarium baptisiae Henn., *Notizbl. Bot. Gart. Berlin* 2: 383. 1899.

Fusarium subnivale Höhn., in Penther & Zederbauer, *Ann. K.K. Naturhist. Hofmus.* 20: 369. 1905.

Fusarium dimerum var. *majusculum* Wollenw., *Fusaria Autogr. Delin.* 1: 90. 1916.

?*Fusarium pusillum* Wollenw., *Fusaria Autogr. Delin.* 2: 550. 1924.

?*Fusarium dimerum* var. *pusillum* (Wollenw.) Wollenw., *Fusaria Autogr. Delin.* 3: 851. 1930.

Fusarium dimerum var. *violaceum* Wollenw., *Fusaria Autogr. Delin.* 3: 854. 1930.

Lectotypus: Fig. 1212 in [Penzig \(1882\)](#), designated in [Schroers et al. \(2009\)](#).

Epitypus: CBS H-20129, designated in [Schroers et al. \(2009\)](#).

Ex-epitype culture: CBS 108944 = NRRL 36140.

Epitype locality: **Netherlands**.

Epitype substrate: Blood of *Homo sapiens* with acute myeloid leukemia.

Descriptions and illustrations: See [Schroers et al. \(2009\)](#).

Diagnostic DNA barcodes: rpb1: KM232212; rpb2: KM232363; tef1: EU926334.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#) and [Booth \(1971\)](#).

diminutum Fusarium (Sand.-Den. & Crous) O'Donnell et al., *Index Fungorum* 440: 2. 2020.

Neocosmospora diminuta Sand.-Den. & Crous, *Persoonia* 43: 127. 2019.

Holotypus: CBS H-23979.

Ex-type culture: CBS 144390 = MUCL 18798.

Type locality: ?**Ivory Coast**.

Type substrate: Treated wood of *Coelocaryon preussii*.

Descriptions and illustrations: See [Sandoval-Denis et al. \(2019\)](#).

Diagnostic DNA barcodes: rpb1: MW834218; rpb2: LR583828; tef1: LR583607.

dimorphum Fusarium J.V. Almeida & Sousa da Câmara, *Revista Agron. (Lisbon)* 1: 306. 1903.

(See *Fusarium buxicola*)

Holotypus: MA-Funhist:6036-1.

Type locality: **Portugal**.

Type substrate: *Buxus sempervirens*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

diplosporum Fusarium Cooke & Ellis, *Grevillea* 7: 38. 1878.

(See ***Fusarium sarcochrom***)

Holotypus: ?K(M).

Type locality: **USA**, New Jersey.

Type substrate: Stems of *Solanum tuberosum*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

discoideum Fusarium Fautrey & Roum., *Rev. Mycol. (Toulouse)* 13: 173. 1891.

(See ***Fusarium lateritium***)

Syntype: ILL00220061 (Roumeguère, *Fungi Sel. Gall. Exs.* no. 5898).

Type locality: **France**, Noidan.

Type substrate: *Sambucus nigra*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

discolor Fusarium Appel & Wollenw., *Arbeiten Kaiserl. Biol. Anst. Land-Forstw.* 8: 114. 1913.

(See ***Fusarium sambucinum***)

Holotypus: ?S-F45617.

Type locality: **Germany**, Berlin.

Type substrate: *Solanum tuberosum*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

diversisporum Fusarium Sherb., *Mem. Cornell Univ. Agric. Exp. Sta.* 6: 161. 1915.

Typus: ?CUP-007430.

Type locality: USA, New York.

Type substrate: *Solanum tuberosum*

Descriptions and illustrations: See [Sherbakoff \(1915\)](#) and [Gerlach & Nirenberg \(1982\)](#).

Notes: This species is recognised by [Gerlach & Nirenberg \(1982\)](#) who considered isolate CBS 795.70 as authentic for *F. diversisporum*. However, typification of *F. diversisporum* first requires study of the specimen lodged in CUP.

dlaminii Fusarium [Marasas et al.](#), *Mycologia* 77: 971. 1986 [1985].

Holotypus: DAOM 191112.

Ex-type culture: ATCC 58097 = BBA 69859 = CBS 175.88 = DAOM 191112 = FRC M-1637 = IMI 290241 = MRC 3032 = NRRL 13164.

Type locality: South Africa, Eastern Cape Province, Butterworth.

Type substrate: Plant debris in soil.

Descriptions and illustrations: See [Marasas et al. \(1985\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: KU171681; *rpb2*: KU171701; *tef1*: KU171721.

domesticum Fusarium (Fr.) H.P. Bachm., *LWT – Food Sci. Technol.* 38: 405. 2005, *nom. inval.*, Art. 41.5, See Art. 41.7.

Bisifusarium domesticum (Fr.) L. Lombard & Crous, *Stud. Mycol.* 80: 225. 2015.

Basionym: *Trichothecium domesticum* Fr., *Syst. Mycol.* 3: 427. 1832.

Neotypus: CBS 434.34 (preserved as metabolically inactive culture), designated in [Bachmann et al. \(2005\)](#).

Ex-neotype culture: ATCC 13417 = CBS 434.34 = MUCL 9826.

Type locality: Belgium.

Type substrate: Cheese.

Descriptions and illustrations: See [Schroers et al. \(2009\)](#).

dominicanum Fusarium Cif., *Sydowia* 9: 325. 1955

Holotypus: ?PAV.

Type locality: Dominican Republic, Santo Domingo, Villa Altigracia.

Type substrate: *Byrsonima* sp. (between mycelium of *Meliola byrsonimae*).

Descriptions and illustrations: See [Ciferri \(1955\)](#).

Notes: [Ciferri \(1955\)](#) considered this a 'conventional' species as the author indicated that more information based on culture characteristics is required. No living material of this species could be located and recollection from the type locality is required.

duofalcatisporum Fusarium J.W. Xia et al., *Persoonia* 43: 201. 2019.

Holotypus: CBS H-24056.

Ex-type culture: CBS 384.92 = NRRL 36448.

Type locality: Sudan, Nile Province.

Type substrate: Seeds of *Phaseolus vulgaris*.

Descriptions and illustrations: See [Xia et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb2*: GQ505830; *tef1*: GQ505652.

duoseptatum Fusarium [Maryani et al.](#), *Stud. Mycol.* 92: 181. 2018 [2019].

Holotypus: InaCC F916 (preserved as metabolically inactive culture).

Ex-type culture: InaCC F916.

Type locality: Indonesia, Central Kalimantan, Kapuas Timur, Anjir Serapat Tengah.

Type substrate: Pseudostem of *Musa* var. Pisang Kepok.

Descriptions and illustrations: See [Maryani et al. \(2019a\)](#).

Diagnostic DNA barcodes: *rpb1*: LS479495; *rpb2*: LS479239; *tef1*: LS479688.

echinatum Fusarium Sand.-Den. & G.J. Marais, *Stud. Mycol.* 98 (no. 100116): 47. 2021.

Holotypus: CBS H-24658.

Ex-type culture: CAMS 000733 = CBS 146497 = CPC 30815.

Type locality: South Africa.

Type substrate: Unidentified tree.

Descriptions and illustrations: See this study.

Diagnostic DNA barcodes: *rpb1*: MW834187; *rpb2*: MW834004; *tef1*: MW834273.

echinosporum Fusarium [Sibilia](#), *Ann. Reale. Ist. Super. Agrar. Forest.*, ser. 2, 1: 77. 1925.

Holotypus: Not located.

Type locality: Italy.

Type substrate: *Cedrus deodara*.

Descriptions and illustrations: See [Sibilia \(1925\)](#).

Notes: This species is recognised in [Petra's Lists V. 3](#). [Wollenweber & Reinking \(1935\)](#) mention this species, but they did not treat it any further. [Booth \(1971\)](#) considered it a possible synonym of *F. graminearum*. Requires recollection from the type locality and substrate.

effusum Fusarium [Schwein.](#), *Trans. Amer. Philos. Soc.*, n.s., 4: 302. 1832 [1834].

(See *Fusarium cymbiferum*)

Holotypus: PH00062491.

Type locality: USA, Pennsylvania, Northampton, Bethlehem.

Type substrate: *Hypericum frondosum*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

eichleri Fusarium [Bres.](#), *Ann. Mycol.* 1: 130. 1903.

(See *Fusarium candidum* [Ehrenb.](#))

Holotypus: S-F45618.

Type locality: Poland.

Type substrate: *Salix caprea*.

elaedis Fusarium L. Lombard & Crous, *Persoonia* 43: 23. 2018 [2019].

Holotypus: CBS H-23612.

Ex-type culture: CBS 217.49 = NRRL 36358.

Type locality: Zaire.

Type substrate: *Elaeis* sp.

Descriptions and illustrations: See [Lombard et al. \(2019b\)](#).

Diagnostic DNA barcodes: *rpb1*: MW928805; *rpb2*: MH484870; *tef1*: MH484961.

elasticae Fusarium (Thüm.) [Sacc.](#), *Syll. Fung.* 4: 711. 1886.

Basionym: *Fusisporium elasticae* Thüm., in [Bolle & Thümen](#), *Boll. Soc. Adriat. Sci. Nat. Trieste* 3: 440. 1877.

(See *Fusarium cymbiferum*)

Lectotypus (*hic designatus*, MBT 10000679): Italy, Gorizia, *Ficus elastica*, 1877, F. de Thümen, in [Bolle & Thümen](#), *Boll. Soc. Adriat. Sci. Nat. Trieste* 3, tab. I, fig. 13.

Notes: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#). No holotype specimen could be located and therefore an illustration is designated as lectotype.

elegans *Fusarium* Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land-Forstw. 8: 94. 1913, *nom. inval.*, Art. 36.1(a).

(See *Fusarium oxysporum*)

Notes: Appel and Wollenweber (*l.c.*) proposed this name only provisionally under *Fusarium solani*. They added an illustration of conidia on page 38 (fig. 2D).

elegans *Fusarium* W. Yamam. & Maeda, Trans. Mycol. Soc. Japan 3: 115. 1962.

Neocosmospora elegans (W. Yamam. & Maeda) Sand.-Den. & Crous, Persoonia 43: 127. 2019.

Basionym: *Nectria elegans* W. Yamam. & Maeda, Hyogo Univ. Agric. ser. Agric. Biol. 3: 15. 1957.

Synonyms: ?*Fusarium solani* f. *xanthoxyli* Y. Sakurai & Matuo, Ann. Phytopathol. Soc. Japan 26: 117. 1961, *nom. inval.*, Art. 39.1.

?*Hypomyces solani* f. *xanthoxyli* Y. Sakurai & Matuo, Ann. Phytopathol. Soc. Japan 26: 117. 1961, *nom. inval.*, Art. 39.1. *Fusarium yamamotoi* O'Donnell *et al.*, Index Fungorum 440: 5. 2020.

Lectotypus: figs 1–9, p. 16, in Yamamoto *et al.* (1957), designated in Sandoval-Denis *et al.* (2019).

Epitypus: CBS H-23980, designated in Sandoval-Denis *et al.* (2019).

Ex-epitype culture: ATCC 42366 = CBS 144396 = MAFF 238541 = NRRL 22277 = SUF XV-1.

Epitype locality: Japan, Hyōgo.

Epitype substrate: Trunk of *Zanthoxylum piperitum*

Diagnostic DNA barcodes: *rpb1*: MW218113; *rpb2*: FJ240380; *tef1*: AF178336

Note: This is a valid species name that is not a homonym since the name *F. elegans* Appel & Wollenw. is an invalid name.

eleocharidis *Fusarium* Rostr. (as '*heleocharidis*'), in Thümen, Mycoth. Univ., Cent. 22: no. 2185. 1883.

(See *Fusarium heterosporum*)

Syntypes: In BPI, NEB & S (Mycoth. Univ., Cent. 22: no. 2185).

Type locality: Denmark, Fyn, Langeland.

Type substrate: *Eleocharis palustris*.

Notes: Synonym *fide* Wollenweber & Reinking (1935).

elongatum *Fusarium* Cooke, Grevillea 19: 4. 1890.

(See *Fusarium ciliatum*)

Holotypus: In K(M), Colenso 538 *fide* Index Fungorum .

Type locality: New Zealand.

Type substrate: Twigs.

Note: Synonym *fide* Wollenweber & Reinking (1935).

elongatum *Fusarium* De Wild., Ann. Soc. Belge Microsc. 17: 42. 1893, *nom. illegit.*, Art. 53.1.

Replacing synonym: *Fusarium longissimum* Sacc. & P. Syd., Syll. Fung. 14: 1128. 1899.

Amniculicola longissima (Sacc. & P. Syd.) Nadeeshan & K.D. Hyde, IMA Fungus 7: 301. 2016.

Synonyms: *Anguillospora longissima* (Sacc. & P. Syd.) Ingold, Trans. Brit. Mycol. Soc. 25: 402. 1942.

Holotypus: Not located.

Type locality: Belgium, Brussels, Botanical Garden.

Type substrate: Submerged plant material.

Note: Synonyms *fide* Rossman *et al.* (2016).

elongatum *Fusarium* O.A. Pratt, J. Agric. Res. 13: 84. 1918, *nom.*

illegit., Art. 53.1.

(See *Fusarium sambucinum*)

Authentic material: Not located.

Original locality: USA, Idaho.

Original substrate: Soil.

Note: Synonym *fide* Wollenweber & Reinking (1935).

elongatum *Fusarium* Reinking, Zentralbl. Bakteriell. Parasitenk., Abt. 2, 89: 511. 1934, *nom. illegit.*, Art. 53.1.

(See *Fusarium sublunatum*)

Authentic material: B 70 0100189.

Original culture: CBS 190.34 = NRRL 20897.

Original locality: Costa Rica.

Original substrate: Soil from *Musa sapientum* and *Theobroma cacao* plantation.

Diagnostic DNA barcodes: *rpb1*: KX302927; *rpb2*: KX302935; *tef1*: KX302919.

Note: Synonym *fide* Wollenweber & Reinking (1935).

ensiforme *Fusarium* Wollenw. & Reinking, Phytopathology 15: 169. 1925.

Synonym: *Fusarium javanicum* var. *ensiforme* (Wollenw. & Reinking) Wollenw., Z. Parasitenk. 3: 483. 1931.

Fusarium javanicum subsp. *ensiforme* (Wollenw. & Reinking) Raillo, Fungi of the Genus *Fusarium*: 229. 1950.

Holotypus: Not located.

Type locality: Honduras.

Type substrate: Rotten fruit of *Ficus* sp.

Notes: Synonyms *fide* Wollenweber & Reinking (1935). Synonym of *F. javanicum* *fide* Gerlach & Nirenberg (1982). Status unclear [see Sandoval-Denis *et al.* (2019)].

entomophilum *Fusarium* Petch, Trans. Brit. Mycol. Soc. 11: 260. 1926.

(See *Fusarium lateritium*)

Holotypus: ?K(M).

Type locality: Sri Lanka, Suduganga.

Type substrate: *Clitellaria heminopla*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

epicoccum *Fusarium* McAlpine, Fungus Diseases of Citrus trees in Australia: 113. 1899.

(See *Fusarium larvarum*)

Lectotypus (*hic designatus*, MBT 10000680): Australia, Victoria, Melbourne, *Aspidiotus aurantium* on *Citrus deliciosa*, 1899, D. McAlpine, in Fungus Diseases of Citrus trees in Australia, figs 177–180.

Note: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

epimyces *Fusarium* Cooke, Grevillea 17: 15. 1888, *nom. inval.*, Art. 38.1(a).

(See *Fusarium azukiicola*)

Authentic material: In K(M) *fide* Index Fungorum.

Original locality: UK, Reading.

Original substrate: *Scleroderma* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

episphaeria *Fusarium* (Tode) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 32: 662. 1945.

Dialonectria episphaeria (Tode) Cooke (as '*episphaerica*'), Grevillea 12: 82. 1884.

Basionym: *Sphaeria episphaeria* Tode, Fung. Mecklenb. Sel. 2: 21. 1791.

Synonyms: *Nectria episphaeria* (Tode) Fr., Summa Veg. Scand. 2: 388. 1849.

Cucurbitaria episphaeria (Tode) Kuntze, Revis. Gen. Pl. 3: 461. 1898.

Cosmospora episphaeria (Tode) Rossman & Samuels, Stud. Mycol. 42: 121. 1999.

Hypoxylon phoeniceum Bull., Hist. Champ. France 1: 171. 1791.

Sphaeria sanguinea var. *media* Fr., Syst. Mycol. 2: 453. 1823.

Nectria episphaeria var. *media* (Fr.) Sacc., Syll. Fung. 2: 497. 1883.

Dialonectria episphaeria var. *verruculosa* Cooke, Grevillea 12: 82. 1884.

Nectria episphaeria var. *verruculosa* (Cooke) Berl. & Voglino, Syll. Fung., Addit. Vol. 1–4: 203. 1886.

Nectria episphaeria var. *kretzschmariae* Henn., Bot. Jahrb. Syst. 14: 364. 1891.

Nectria episphaeria var. *gregaria* Starbäck, Ark. Bot. 5: 9. 1905.

Lectotypus: L 0112704 (Herb. Lugd. Bat. 910267659 ex Herb. Persoon), selected in Booth (1959).

Type locality: **Unknown**.

Type substrate: Partially decorticated twig of *Diatrype stigma*.

episphaericum *Fusarium* (Cooke & Ellis) Sacc., Syll. Fung. 4: 708. 1886.

Basionym: *Fusisporium episphaericum* Cooke & Ellis, Grevillea 5: 50. 1876.

Cosmospora nothepisphaeria (Samuels) Rossman & Samuels, Stud. Mycol. 42: 123. 1999.

Basionym: *Nectria nothepisphaeria* Samuels, Mycol. Pap. 164: 30. 1991.

Synonyms: *Fusarium ciliatum* var. *episphaericum* (Cooke & Ellis) Wollenw., Fusaria Autogr. Delin. 3: 871. 1930.

Fusarium ciliatum var. *majus* Wollenw., Fusaria Autogr. Delin. 3: 872. 1930.

Lectotypus (of *Fusisporium episphaericum*, *hic designatus*, MBT 10000681): **USA**, New Jersey, parasitic on *Diatrypella* sp. on *Corylus avellana*, 1876, M.C. Cooke & J.B. Ellis, in Grevillea 5, pl. 80, fig. 10.

Note: No holotype specimen could be located and therefore an illustration is designated as lectotype.

epistroma *Fusarium* (Höhn.) C. Booth (as '*epistromum*'), The Genus *Fusarium*: 66. 1971.

Fusicolla epistroma (Höhn.) Gräfenhan & Seifert, Stud. Mycol. 68: 100. 2011.

Basionym: *Dendrodochium epistroma* Höhn., Sitzungsber. Kaiserl. Akad. Wiss. Wien. Math.-Naturwiss. Cl., Abt. 1., 118: 424. 1909.

Lectotypus: B 700014042, designated in Gräfenhan et al. (2011).

Lectotype locality: **Germany**, Brandenburg, "Schmidt's Grund" near Tamsel.

Lectotype substrate: Old stromata of *Diatrypella favacea*.

Epitypus: IMI 85601, designated in Gräfenhan et al. (2011).

Ex-epitype culture: ATCC 24369 = BBA 62201 = NRRL 20439 = NRRL 20461.

Epitype locality: **UK**, Yorkshire.

Epitype substrate: *Diatrypella* on *Betula*.

Diagnostic DNA barcode: *rpb2*: HQ897765.

epithele *Fusarium* McAlpine, Fungus Diseases of Citrus trees in Australia: 80. 1899.

(See ***Fusarium reticulatum***)

Holotypus: VPRI 2563.

Type locality: **Australia**, New South Wales.

Type substrate: Rotten fruit of *Citrus x limon*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

equinum *Fusarium* Növgaard, Science, N.Y. 14: 899. 1901.

Holotypus: Not located.

Type locality: **USA**.

Type substrate: Infected skin of *Equus* sp. (horse).

Notes: Status unclear. Doubtful species *fide* Wollenweber & Reinking (1935). Based on the original substrate, this species might belong to the medically important genus *Neocosmospora*. However, recollection is required to confirm its taxonomic affiliation.

equiseti* *Fusarium (Corda) Sacc., Syll. Fung. 4: 707. 1886.

Basionym: *Selenosporium equiseti* Corda, Icon. Fung. 2: 7. 1838.

Synonyms: *Fusisporium ossicola* Berk. & M.A. Curtis, Grevillea 3: 147. 1875.

Fusarium ossicola (Berk. & M.A. Curtis) Sacc., Syll. Fung. 4: 714. 1886.

Fusarium nectriae-palmicolae Henn., Bot. Jahrb. Syst. 23: 290. 1896.

Fusarium gibbosum Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land-Forstw. 8: 190. 1910.

Fusarium roseum var. *gibbosum* (Appel & Wollenw.) Messiaen & R. Cass., Ann. Inst. Natl. Rech. Agron. Tunisie 19: 435. 1968, *nom. inval.*, Art. 41.5.

Fusarium roseum var. *gibbosum* (Appel & Wollenw.) Messiaen & R. Cass., Agronomie 8: 220. 1988, *nom. inval.*, Art. 41.1.

Fusarium bullatum Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 198. 1915.

Fusarium equiseti var. *bullatum* (Sherb.) Wollenw., Fusaria Autogr. Delin. 3: 916. 1930.

Fusarium gibbosum var. *bullatum* (Sherb.) Bilař, Mikrobiol. Zhurn. 49: 6. 1987.

Fusarium bullatum var. *roseum* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 201. 1915.

Fusarium roseobullatum Wollenw. (as '*roseo-bullatum*'), Fusaria Autogr. Delin. 1: 117. 1916.

Fusarium vasinfectum var. *psi* Schikora, Arbeiten. Biol. Anst. Land-Forstw. 5: 188, pl. 7. 1906, *nom. illegit.*, Art. 53.1.

Fusarium falcatum Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land-Forstw. 8: 184. 1910.

Fusarium falcatum var. *fuscum* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 138. 1915.

Fusarium equiseti var. *crassum* Wollenw., Fusaria Autogr. Delin. 3: 921. 1930.

Fusarium terrestre Manns, Bull. North Dakota Agric. Exp. Sta.: no. 259. 1932.

Gibberella intricans Wollenw., Fusaria Autogr. Delin. 3: 810. 1930.

Fusarium eueheliae Sartory, R. Sartory & J. Mey., Ann. Mycol. 30: 471. 1932.

Fusarium equiseti var. *intermedium* Saccas, Agron. Trop. (Maracay) 10: 49. 1955, *nom. inval.*, Art. 39.1.

Lectotypus: (*hic designatus*, MBT 10001325): **Czech Republic**, Kuchelbad, near Prague, on stems of *Equisetum* sp., 1836, AKJ. Corda. Icon. Fung. 2, tab. IX, fig. 32.

Epitypus (*hic designatus*, MBT 10000682): **Germany**, Braunschweig, Niedersachsen, soil, 3 Aug. 1994, H. I. Nirenberg, CBS H-5570.

Ex-epitype culture: BBA 68556 = CBS 307.94 = NRRL 26419.

Descriptions and illustrations: See Wollenweber & Reinking (1935), Booth (1971), Gerlach & Nirenberg (1982), Holubová-Jechová *et al.* (1994) and Leslie & Summerell (2006).

Diagnostic DNA barcodes: *rpb2*: GQ505777; *tef1*: GQ505599.

Notes: Holubová-Jechová *et al.* (1994) incorrectly designated CBS 307.94 (CBS H-5570) as neotype for *Selenosporium equiseti* even though original material was available in PRM as well as an illustration provided in the protologue. A lectotypification rather than a neotypification was required. Therefore, the original illustration is selected as lectotype and CBS H-5570 (= CBS 307.94) is designated as epitype here, superseding the neotype designation.

equiseticola *Fusarium* Allesch., Hedwigia 34: 289. 1895.

(See *Fusarium scirpi*)

Holotypus: In M.

Type locality: **Germany**, Oberammergau.

Type substrate: Dried stems of *Equisetum limosum*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

equisetorum *Fusarium* Desm., Pl. Crypt. N. France: no. 1546/1846? 1843.

Basionym: *Hymenula equiseti* Lib., Pl. Crypt. Arduenna 3: no. 236. 1834.

(See *Fusarium oxysporum*)

Syntypes: In BPI, BRU, CUP, ISC PH, S & UPS (Pl. Crypt. Arduenna 3: no. 236).

Type locality: **Belgium**.

Type substrate: *Equisetum limosum*.

Notes: Synonym *fide* Wollenweber & Reinking (1935).

ershadii *Fusarium* M. Papizadeh *et al.*, Europ. J. Pl. Pathol. 151: 693. 2018, *nom. illegit.*, Art. 52.1.

Basionym: *Cylindrocarpon tonkinense* Bugnic., Encycl. Mycol. 11: 181. 1939.

(See *Fusarium tonkinense*)

erubescens *Fusarium* Berk. & M.A. Curtis, Grevillea 3: 98. 1875. *Synonym:* *Fusarium alabamense* Sacc., Syll. Fung. 4: 722. 1886, *nom. illegit.*, Art. 52.1.

Holotypus: ?K(M).

Type locality: **USA**, Alabama, Beaumont.

Type substrate: Dead bark.

Notes: Status unclear. Not *Fusarium* *fide* Wollenweber & Reinking (1935).

erubescens *Fusarium* (Durieu & Mont.) Sacc., Syll. Fung. 4: 719. 1886, *nom. illegit.*, Art. 53.1.

Basionym: *Fusisporium erubescens* Durieu & Mont., Exploration scientifique de l'Algérie 1–9: 351. 1848.

(See *Fusarium bacilligerum*)

Holotypus: ?PC.

Type locality: **Algeria**, Béjaïa.

Type substrate: *Rhamnus alaternus*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

erubescens *Fusarium* Appel & Oven, Landwirtsch. Jahrb. 1905, *nom. illegit.*, Art. 53.1.

(See *Fusarium acuminatum*)

Authentic material: Not located.

Original locality: **Germany**.

Original substrate: *Solanum lycopersicum*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

eucalypticola *Fusarium* Henn., Hedwigia 40: 355. 1901.

Holotypus: In B *fide* Hein (1988).

Type locality: **Australia**, Western Australia, Cranbrook.

Type substrate: *Eucalyptus baxteri* (syn. *E. santalifolia*)

Notes: Status unclear. Not *Fusarium* *fide* Wollenweber & Reinking (1935).

eucalyptorum *Fusarium* Cooke & Harkn., Grevillea 9: 128. 1881.

(See *Fusarium oxysporum*)

Syntype: BPI 452103.

Type locality: **USA**, California, San Francisco Masonic Cemetery.

Type substrate: *Eucalyptus* sp.

Note: Synonym *fide* Arya & Jain (1962).

eucheliae *Fusarium* Sartory, R. Sartory & J. Mey., Ann. Mycol. 30: 471. 1932.

(See *Fusarium equiseti*)

Lectotypus (*hic designatus*, MBT 10000683): **France**, digestive track of living caterpillar, 1932, A. Sartory, R. Sartory & J. Meyer, in Ann. Mycol. 30: 473, figs 1–13.

Notes: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

eumartii *Fusarium* C.W. Carp., J. Agric. Res. 5: 204. 1915.

(See *Fusarium solani*)

Lectotypus: Illustration Plate XIV, number 4, in Carpenter (1915), designated in Sandoval-Denis *et al.* (2019).

Type locality: **Unknown**.

Type substrate: *Solanum tuberosum*.

euonymi *Fusarium* Syd., Beibl. Hedwigia 39: (6). 1900.

(See *Fusarium lateritium*)

Syntype: S-F45621 (Sydow, Mycoth. March. no. 4896).

Type locality: **Germany**, Berlin.

Type substrate: *Euonymus bungeanus*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

euonymi-japonici *Fusarium* Henn., Hedwigia 41: 139. 1902.

(See *Fusarium lateritium*)

Holotypus: In B *fide* Hein (1988).

Type locality: **Germany**, Berlin.

Type substrate: *Euonymus japonicus*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

euwallaceae *Fusarium* S. Freeman *et al.*, Mycologia 105: 1599. 2013.

Neocosmospora euwallaceae (S. Freeman *et al.*) Sand.-Den. *et al.*, Persoonia 43: 129. 2019.

Holotypus: BPI 884203.

Ex-type culture: CBS 135854 = NRRL 54722.

Type locality: **Israel**, central coastal region, Kibbutz Gilil Yam.

Type substrate: *Euwallacea* sp. beetle infecting *Persea americana* cv. Hass.

Descriptions and illustrations: See Freeman *et al.* (2013).

Diagnostic DNA barcodes: *rpb1*: JQ038021; *rpb2*: JQ038028; *tef1*: JQ038007.

expansum *Fusarium* Schldl., Fl. Berol. 2: 139. 1824.

Synonym: ?*Fusarium carpini* Schulzer & Sacc., Hedwigia 23: 128. 1884.

Fusarium socium Sacc., Atti Ist. Veneto Sci. Lett. Arti, sér. 6, 2: 450. 1884.

Fusarium cirrosum Höhn., Sitzungsber. Kaiserl. Akad. Wiss. Wien, Math.-Naturwiss. Cl., Abt. 1., 116: 153. 1907.

Fusarium macounii Dearn., Mycologia 9: 363. 1917.

Holotypus: HAL 1614 F.

Type locality: **Germany**, Berlin.

Type substrate: *Carpinus betulus*.

Descriptions and illustrations: See [Wollenweber \(1916–1935\)](#) and [Gerlach & Nirenberg \(1982\)](#).

Notes: Both [Wollenweber & Reinking \(1935\)](#) and [Gerlach & Nirenberg \(1982\)](#) recognised this species. This species requires epitypification from the type locality.

fabacearum Fusarium L. Lombard *et al.*, Persoonia 43: 24. 2018 [2019].

Holotypus: CBS H-23613.

Ex-type culture: CBS 144743 = CPC 25802.

Type locality: **South Africa**, Western Cape Province.

Type substrate: *Glycine max*.

Descriptions and illustrations: See [Lombard *et al.* \(2019b\)](#).

Diagnostic DNA barcodes: *rpb1*: MW928806; *rpb2*: MH484938; *tef1*: MH485029.

falcatum Fusarium Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land-Forstw. 8: 184. 1913.

Replaced synonym: *Fusarium vasinfectum* var. *pisi* Schikora, Arbeiten Biol. Anst. Land-Forstw. 5: 188, pl. 7. 1906, *nom. illegit.*, Art. 53.1

(See ***Fusarium equiseti***)

Holotypus: Not located.

Type locality: **Germany**, Berlin.

Type substrate: *Pisum sativum*.

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

falciforme Fusarium (Carrión) Summerb. & Schroers, J. Clin. Microbiol. 40: 2872. 2002.

Neocosmospora falciformis (Carrión) L. Lombard & Crous, Stud. Mycol. 80: 227. 2015.

Basionym: *Cephalosporium falciforme* Carrión, Mycologia 43: 523. 1951.

Synonyms: *Acremonium falciforme* (Carrión) W. Gams, Cephalosporium-artige Schimmelpilze: 139. 1971.

Fusarium paranaense S.S. Costa *et al.*, Fungal Biology 120: 55. 2015 [2016].

Holotypus: CBS 475.67 (preserved as metabolically inactive culture).

Ex-type culture: CBS 475.67 = IHM 939 = IMI 268681.

Type locality: **Puerto Rico**.

Type substrate: Mycetoma from *Homo sapiens*.

Diagnostic DNA barcodes: *rpb1*: MW218114; *rpb2*: LT960558; *tef1*: LT906669.

fasciculatum Fusarium J.W. Xia *et al.*, Persoonia 43: 203. 2019.

Holotypus: CBS H-24057.

Ex-type culture: CBS 131382.

Type locality: **Australia**, Northern Territories, Roper River area.

Type substrate: Stems of *Oryza australiensis*.

Descriptions and illustrations: See [Xia *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb2*: MN170406; *tef1*: MN170473.

fautreyi Fusarium Sacc., Syll. Fung. 10: 934. 1892.

Replaced synonym: *Fusarium parasiticum* Fautrey, Rev. Mycol. (Toulouse) 11: 153. 1889, *nom. illegit.*, Art. 53.1.

(See ***Fusarium lateritium***)

Typus: BR5020140789424.

Type locality: **France**, Noidan.

Type substrate: *Vitis vinifera*.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

ferrugineum Fusarium (Sand.-Den. & Crous) O'Donnell *et al.*, Index Fungorum 440: 2. 2020.

Neocosmospora ferruginea Sand.-Den. & Crous, Persoonia 43: 130. 2019.

Holotypus: CBS H-23981.

Ex-type culture: CBS 109028 = NRRL 32437.

Type locality: **Switzerland**.

Type substrate: Subcutaneous nodule of *Homo sapiens*.

Descriptions and illustrations: See [Sandoval-Denis *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: HM347157; *rpb2*: EU329581; *tef1*: DQ246979.

ferruginosum Fusarium Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 190. 1915.

(See ***Fusarium acuminatum***)

Typus: ?CUP-007445.

Type locality: **USA**, New York, Long Island

Type substrate: *Solanum tuberosum*

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#). Lectotypification pending study of material lodged in CUP.

ficicrescens Fusarium Al-Hatmi *et al.*, Fungal Biol. 120: 274. 2015 [2016].

Holotypus: CBS H-21815.

Ex-type culture: CBS 125178.

Type locality: **Iran**, Estahban.

Type substrate: Fruit of *Ficus carica*.

Descriptions and illustrations: See [Al-Hatmi *et al.* \(2016\)](#).

Diagnostic DNA barcodes: *rpb1*: MT010950; *rpb2*: MT010977; *tef1*: MT011004.

filiferum Fusarium (Preuss) Wollenw., Fusaria Autogr. Delin. 1: 220. 1916.

Basionym: *Fusoma filiferum* Preuss, Linnaea 25: 73. 1852.

Synonym: *Fusarium scirpi* var. *filiferum* (Preuss) Wollenw., Fusaria Autogr. Delin. 3: 936. 1930.

(See ***Fusarium scirpi***)

Holotypus: Not located.

Type locality: **Germany**.

Type substrate: Bark of *Pinus* sp.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

filisporum Fusarium (Cooke) Sacc., Syll. Fung. 4: 708. 1886.

Basionym: *Fusisporium filisporum* Cooke, Grevillea 8: 8. 1879.

(See *Fusarium ciliatum*)

Holotypus: In K(M), Muller s.n. *fide* Index Fungorum.

Type locality: **UK**, Eastbourne.

Type substrate: *Orthotrichum* sp.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

fissum Fusarium Peyl, Lotos 8: 30. 1858.

(See *Fusarium candidum* (Link.) Sacc.)

Lectotypus (*hic designatus*, MBT 10000684): **Germany**, twigs of *Citrus aurantiacum*, 1858, J. Peyl, in Lotos 8, fig. 17.

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#). No holotype specimen could be located and therefore an illustration is designated as lectotype.

flagelliforme Fusarium J.W. Xia *et al.*, Persoonia 43: 204. 2019.
Holotypus: CBS H-24058.

Ex-type culture: CBS 162.57 = NRRL 36269.

Type locality: **Croatia**, Zagreb.

Type substrate: Seedlings of *Pinus nigra*.

Descriptions and illustrations: See [Xia et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb2*: GQ505823; *tef1*: GQ505645.

flavidum *Fusarium* (Bonord.) Sacc., Syll. Fung. 4: 698. 1886.

Basionym: *Fusisporium flavidum* Bonord., Bot. Zeitung (Berlin) 19: 194. 1861.

(See *Fusarium reticulatum*)

Lectotypus (*hic designatus*, MBT 10000685): **Germany**, rotten tree, 1861, H.F. Bonorden, in Bot. Zeitung (Berlin) 19: tab. VIII, fig. 3.

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#). No holotype specimen could be located and therefore an illustration is designated as lectotype.

flavum *Fusarium* (Fr.) Wollenw., Z. Parasitenk. (Berlin) 3: 305. 1931.

Basionym: *Fusisporium flavum* Fr., Syst. Mycol. 3: 444. 1832.

(See *Fusarium dimerum*)

Holotypus: Not located.

Type locality: **Germany**, Bonn.

Type substrate: *Aster* sp.

Note: Synonym *fide* [Booth \(1971\)](#).

flocciferum* *Fusarium Corda, in Sturm, Deutschl. Fl., Abt. 3, Pilze Deutschl. 2: 17. 1828.

Synonyms: *Fusarium vinosum* Masee, Brit. Fung.-Fl. 3: 479. 1893.

Fusarium clavatum Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 234. 1915.

Fusarium idahoanum O.A. Pratt, J. Agric. Res. 13: 86. 1918.

Fusarium nigrum O.A. Pratt, J. Agric. Res. 13: 90. 1918.

Lectotypus: (*hic designatus*, MBT 10001326) **Germany**, Berlin, on shell of the fruit of *Aesculus hippocastanum*. AKJ. Corda, Sturm, Deutschl. Fl., Abt. 3, Pilze Deutschl. 2, pl. 7.

Epitypus (*hic designatus*, MBT 10000686): **Germany**, greenhouse soil, 1966, D. Bredemeier, CBS 821.68 (preserved as metabolically inactive culture).

Ex-epitype culture: CBS 821.68 = NRRL 28450.

Descriptions and illustrations: See [Booth \(1971\)](#) and [Gerlach & Nirenberg \(1982\)](#).

Diagnostic DNA barcodes: *rpb1*: MW928807; *rpb2*: MW928824; *tef1*: MW928837.

Notes: Corda's original illustration of *Fusarium flocciferum* is here selected as lectotype. [Gerlach & Nirenberg \(1982\)](#) considered isolate CBS 821.68, along with CBS 792.70, as good representatives of *F. flocciferum*. Based on their observations and collection locality, CBS 821.68 is designated as epitype of *F. flocciferum*.

floridanum *Fusarium* T. Aoki et al., Mycologia 111: 922. 2019.

Neocosmospora floridana (T. Aoki et al.) L. Lombard & Sand.-Den., *comb. nov.* MycoBank MB 837664.

Basionym: *Fusarium floridanum* T. Aoki et al., Mycologia 111: 922. 2019.

Holotypus: BPI 910972.

Ex-type culture: MAFF 246849 = NRRL 62628.

Type locality: **USA**, Florida, Gainesville.

Type substrate: Mycangium of *Euwallacea interjectus* infesting *Acer negundo*.

Descriptions and illustrations: See [Aoki et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: KC691593; *rpb2*: KC691624, KC691653; *tef1*: KC691535.

Notes: A new combination is provided in the genus *Neocosmospora* based on the phylogenetic relationship ([Aoki et al. 2019](#)) of this species to other *Neocosmospora* spp. in the ambrosia clade.

foeni *Fusarium* (Berk. & Broome) Sacc., Syll. Fung. 4: 699. 1886.

Basionym: *Fusisporium foeni* Berk. & Broome, Ann. Mag. Nat. Hist., ser. 2, 7: 179. 1851.

(See *Fusarium merismoides*)

Holotypus: ?K(M).

Type locality: **UK**, Northamptonshire, Apethrope.

Type substrate: A hay stalk.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

foetens* *Fusarium Schroers et al., Mycologia 96: 398. 2004.

Holotypus: CBS 110286 (preserved as metabolically inactive culture).

Ex-type culture: CBS 110286 = NRRL 31852 = PD 2001/7244.

Type locality: **Netherlands**, Zuid-Holland Province, Maasland.

Type substrate: *Begonia elatior* hybrid.

Descriptions and illustrations: See [Schroers et al. \(2004\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: MW928808; *rpb2*: MW928825; *tef1*: AY320087.

foliicola *Fusarium* Allesch., Hedwigia 34: 289. 1895.

(See *Fusarium cymbiferum*)

Holotypus: In M.

Type locality: **Germany**, Oberammergau.

Type substrate: *Arabis alpina*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

fracticaudum* *Fusarium Herron et al., Stud. Mycol. 80: 137. 2015.

Holotypus: PREM 60895.

Ex-type culture: CBS 137233 = CMW 25245.

Type locality: **Colombia**, Risaralda, Angela Maria (Santa Rosa).

Type substrate: *Pinus maximinoi*.

Descriptions and illustrations: See [Herron et al. \(2015\)](#).

Notes: Comparisons of recently generated sequences for the living ex-type (CBS 137233 = CMW 25245) of *F. fracticaudum* indicate a strain transposition or contamination by another *Fusarium* species. Therefore, this species needs to be recollected from the type locality and substrate or sequences need to be generated from the holotype specimen.

fractiflexum* *Fusarium T. Aoki et al., Mycoscience 42: 462. 2001.

Holotypus: NIAES 20515.

Ex-type culture: MAFF 237529 = NRRL 28852.

Type locality: **Japan**, Yamanashi, Enzan.

Type substrate: *Cymbidium* sp.

Descriptions and illustrations: See [Aoki et al. \(2001\)](#).

Diagnostic DNA barcodes: *rpb1*: LR792578; *rpb2*: LT575064; *tef1*: AF160288.

fractum *Fusarium* Sacc. & Cavara, Nuovo Giorn. Bot. Ital., n.s., 7: 308. 1900.

(See *Fusarium candidum* (Link) Sacc.)

Holotypus: In PAD.

Type locality: **Italy**.

Type substrate: *Fagus* sp.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

fragens *Fusarium* P. Crouan & H. Crouan, Fl. Finistère: 14. 1867.

(See *Fusarium candidum* (Link) Sacc.)

Holotypus: ?PC.

Type locality: **France**.

Type substrate: *Salix* sp.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

fraxini *Fusarium* Allesch., Ber. Bot. Vereines Landshut 12: 130. 1892.

(See *Fusarium sambucinum*)

Holotypus: In M.

Type locality: **Germany**, München.

Type substrate: *Fraxinus excelsior*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

fraxini *Fusarium* Kabát & Bubák, Fungi Imperf. Exs., no. 900. 1912, *nom. illegit.*, Art. 53.1.

Fusicoccum fraxini Sherb., Phytopathology 18: 148. 1928.

Authentic material: BPI 451324.

Original locality: **Czech Republic**.

Original substrate: *Fraxinus excelsior*.

fredkrugeri *Fusarium* Sand.-Den. *et al.*, MycoKeys 34: 79. 2018.

Holotypus: CBS H-23496.

Ex-type culture: CBS 144209 = CPC 33747.

Type locality: **South Africa**, Kruger National Park, Skukuza, Granite Supersite.

Type substrate: Rhizosphere soil of *Melhanian acuminata*.

Descriptions and illustrations: See [Sandoval-Denis *et al.* \(2018b\)](#).

Diagnostic DNA barcodes: *rpb1*: LT996199; *rpb2*: LT996147; *tef1*: LT996097.

fructigenum *Fusarium* Fr., Syst. Mycol. 3: 471. 1832.

(See *Fusarium lateritium*)

Holotypus: Not located.

Type locality: **Unknown**.

Type substrate: Fruit of *Rosa pomifera*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

fuckelii *Fusarium* Sacc., Syll. Fung. 4: 695. 1886.

Geejayessia desmazieri (Becc. & De Not.) Schroers *et al.*, Stud. Mycol. 68: 130. 2011.

Basionym: *Nectria desmazieri* Becc. & De Not., Schem. di Classif. Sferiacei: 10. 1863.

Synonyms: *Dialonectria desmazieri* (Becc. & De Not.) Petch, Naturalist (London): 281. 1937.

Nectria coccinea var. *cicatricum* Desm., Ann. Sci. Nat., Bot 10: 351. 1848 (*fide* [Wollenweber & Reinking 1935](#) and [Booth 1971](#)).

Nectria gibbera Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 177. 1870.

Lectotypus: G 00110886 (Fuckel, Fungi Rhen. No. 2357), designated in [Schroers *et al.* \(2011\)](#).

Type locality: **Germany**, Rheingau.

Type substrate: *Buxus sempervirens*.

fujikuroi *Fusarium* Nirenberg, Mitt. Biol. Bundesanst. Land-Forstw. Berlin-Dahlem 169: 32. 1976

Synonyms: *Lisea fujikuroi* Sawada, Special Bull. Agric. Exp. Sta. Gov. Formosa 19: 251. 1919.

Gibberella fujikuroi (Sawada) Wollenw., Z. Parasitenk. (Berlin) 3: 514. 1931.

?*Gibberella fujikuroi* var. *subglutinans* E.T. Edwards, Agric. Gaz. New South Wales 44: 895. 1933.

?*Gibberella subglutinans* (E.T. Edwards) P.E. Nelson *et al.*, *Fusarium* species. An illustrated manual for identification (University Park): 135. 1983.

?*Oospora cephalosporioides* Luchetti & Favilli, Annali Fac. Agrar. R. Univ. Pisa 1: 399. 1938.

?*Gibberella fujikuroi* f. *oryzae* Saccas, Rev. Pathol. Veg. Entomol. Agric. France 30: 77. 1951.

?*Gibberella fujikuroi* var. *intermedia* Kuhlman, Mycologia 74: 766. 1982.

Holotypus: IMI 202879.

Ex-type culture: BBA 12428 = BBA 63630 = CBS 221.76 = IHEM 3821 = IMI 196086 = IMI 202879 = NRRL 13620 = NRRL 13998 = NRRL 22174.

Type locality: **Taiwan**.

Type substrate: *Oryza sativa*.

Descriptions and illustrations: See [Nirenberg \(1976\)](#), [Gerlach & Nirenberg \(1982\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: JX171456; *rpb2*: JX171570; *tef1*: AF160279.

fuliginosporum *Fusarium* Sibilina, Ann. Reale. Ist. Super. Agrar. Forest., ser. 2, 1: 77. 1925.

Holotypus: Not located.

Type locality: **Italy**.

Type substrate: Forest containing mostly *Cedrus deodara*.

Note: Mentioned by [Wollenweber & Reinking \(1935\)](#), but no additional records of this species could be located.

fungicola *Fusarium* (Har. & P. Karst.) Sacc., Syll. Fung. 10: 730. 1892.

?***Alysidium hypophleodes*** (Corda) Bonord., Handb. Allg. Mykol.: 35. 1851.

Basionym: *Fusidium hypophleodes* Corda, Icon. Fung. 1: 3, tab. 1, fig. 50. 1837.

Synonym: *Fusamen fungicola* Har. & P. Karst. (as '*fungicolum*'), Rev. Mycol. (Toulouse) 12: 129. 1890.

Holotypus: Not located.

Type locality: **Finland**, Mustiala.

Type substrate: *Lenzites betulina*.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

funicola *Fusarium* Tassi, Bull. Lab. Orto Bot. Reale Univ. Siena 3: 131. 1900.

(See *Fusarium graminearum*)

Holotypus: ?SIENA.

Type locality: **Italy**.

Type substrate: Rotten string.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

fusarioides *Fusarium* (Gonz. Frag. & Cif.) C. Booth, The Genus *Fusarium*: 88. 1971.

Basionym: *Dactylium fusarioides* Gonz. Frag. & Cif., Bol. Real Soc. Esp. Hist. Nat. 27: 280. 1927.

(See *Fusarium chlamydosporum*)

Holotypus: ?MA-Funhist: 7609-1.

Type locality: **Dominican Republic**, Moca.

Type substrate: leaves of *Crotalaria* sp.

Note: Synonym *fide* [Gerlach & Nirenberg \(1982\)](#).

fuscum *Fusarium* (Bonord.) Sacc., Syll. Fung. 4: 699. 1886.

Basionym: *Selenosporium fuscum* Bonord., Handb. Mykol.: 135. 1851.

(See *Fusarium citrulli* Sartory)

Holotypus: Not preserved *vide* Holubová-Jechová *et al.* (1994).

Type locality: **Germany**.

Type substrate: Bark.

Note: Synonym *vide* Wollenweber & Reinking (1935).

gaditjirii Fusarium Phan *et al.*, *Stud. Mycol.* 50: 265. 2004.

Synonym: *Gibberella gaditjirii* Phan *et al.*, *Stud. Mycol.* 50: 264. 2004.

Holotypus: DAR 76663.

Ex-type culture: CBS 116011 = F15048 = NRRL 53678.

Type locality: **Australia**, Queensland, Walkamin Research Station.

Type substrate: *Heteropogon triticeus*.

Descriptions and illustrations: See Phan *et al.* (2004).

Diagnostic DNA barcodes: *rpb2*: HQ662690; *tef1*: AY639636.

gallinaceum Fusarium Cooke & Harkn., *Grevillea* 9: 8. 1880.

(See *Fusarium merismoides*)

Holotypus: BPI 452133.

Type locality: **USA**, California, Sausalito.

Type substrate: Feathers of *Gallus* sp. (chicken).

Note: Synonym *vide* Wollenweber & Reinking (1935).

gamsii Fusarium Torbati *et al.*, *Mycol. Progr.* 18: 127. 2018 [2019].

Holotypus: CBS H-23561.

Ex-type culture: CBS 143610 = CPC 30862.

Type locality: **Iran**, West Azerbaijan Province, Orumieh-Salmas.

Type substrate: *Agaricus bisporus*.

Descriptions and illustrations: See Torbati *et al.* (2019).

Diagnostic DNA barcodes: *rpb2*: LT970760; *tef1*: LT970788.

gauderoyanum Fusarium Sacc., *Michelia* 2: 132. 1880.

(See ***Fusarium avenaceum***)

Holotypus: In PAD.

Type locality: **France**, Paris.

Type substrate: *Cyperaceae*.

Note: Synonym *vide* Wollenweber & Reinking (1935).

gemmiperda Fusarium Aderh., *Z. Pflanzenkrankh.* 11: 70. 1901.

(See ***Fusarium lateritium***)

Lectotypus (*hic designatus*, MBT 10000687): **Germany**, *Prunus cerasus*, 1901, R. Aderhold, in *Z. Pflanzenkrankh.* 11: pl. II, figs 1–4.

Notes: Synonym *vide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

genevense Fusarium Dasz., *Bull. Soc. Bot. Genève*, sér. 2, 4: 305. 1912.

(See ***Fusarium sambucinum***)

Lectotypus (*hic designatus*, MBT 10000688): **Switzerland**, Geneva, from soil, 1912, M. Daszewska, in *Bull. Soc. Bot. Genève*, sér. 2, 4: 306, fig. 27.

Notes: Synonym *vide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

georginae Fusarium Corda, *Icon. Fung.* 2: 4. 1838.

(See *Fusarium cymbiferum*)

Typus: In PRM *vide* Pilat (1938).

Type locality: **Czech Republic**, Prague.

Type substrate: *Dahlia* sp.

Notes: Synonym *vide* Wollenweber & Reinking (1935). Lectotypification pending study of material lodged in PRM.

gerlachii Fusarium T. Aoki *et al.*, *Fungal Genet. Biol.* 44: 1202. 2007.

Holotypus: BPI 871657.

Ex-type culture: LRG 00-551 = NRRL 36905.

Type locality: **USA**, Minnesota, Polk County, Climax.

Type substrate: *Triticum aestivum*.

Descriptions and illustrations: See Starkey *et al.* (2007).

Diagnostic DNA barcodes: *rpb1*: KM361646; *rpb2*: KM361664; *tef1*: DQ459742.

gibbosum Fusarium Appel & Wollenw., *Arbeiten Kaiserl. Biol. Anst. Land- Forstw.* 8: 190. 1910 [1913].

(See ***Fusarium equiseti***)

Holotypus: ?BPI 452135.

Type locality: **Germany**, Berlin

Type substrate: *Solanum tuberosum*

Note: Synonym *vide* Booth (1971).

gigas Fusarium Speg. *Anales Soc. Ci. Argent.* 22: 221. 1886.

Holotypus: In LPS *vide* Farr (1973).

Type locality: **Paraguay**.

Type substrate: *Bambusa* sp.

Descriptions and illustrations: See Wollenweber & Reinking (1935), Booth (1971) and Gerlach & Nirenberg (1982).

Notes: This species requires epitypification. Wollenweber & Reinking (1935), Booth (1971), and Gerlach & Nirenberg (1982) accepted this species, although limited information is available.

glandicola Fusarium Cooke & W.R. Gerard, *Grevillea* 7: 14. 1878.

Tubercularia glandicola (Cooke & W.R. Gerard) Wollenw. & Reinking, *Fusarien*: 325. 1935.

Holotypus: In K(M), Gerard s.n. *vide* Index Fungorum.

Type locality: **USA**, New York.

Type substrate: Acorns of *Quercus* sp.

Note: Synonym *vide* Wollenweber & Reinking (1935).

glandicola Fusarium Allesch., *Ber. Bot. Vereines Landshut* 12: 130. 1892, *nom. illegit.*, Art. 53.1.

Replacing synonym. *Fusarium allescheri* Sacc. & P. Syd., *Syll. Fung.* 14: 1128. 1899.

(See *Fusarium melanochlorum*)

Authentic material: In M.

Original locality: **Germany**, München.

Original substrate: Fruits of *Quercus robur* (syn. *Q. pedunculata*).

Note: Synonyms *vide* Wollenweber & Reinking (1935).

gleditschiae Fusarium Therry (as '*gledrischiae*'), in Roumeguère, *Fungi Sel. Gall. Exs.*: no. 5496. 1890, *nom. nud.*, Art. 38.1(a).

Gloeosporium gleditschiae Therry ex Wollenw., *Z. Parasitenk.* (Berlin) 3: 437. 1931.

Note: Synonym *vide* Wollenweber & Reinking (1935).

gleditschiicola Fusarium Dearn. & Barthol. (as '*gleditsiaecolum*'), *Mycologia* 9: 363. 1917.

(See ***Fusarium lateritium***)

Holotypus: JD 4379 in DAOM.

Type locality: **USA**, Kansas, Stockton.

Type substrate: *Gleditsia triacanthos*.

Note: Synonym *vide* Wollenweber & Reinking (1935).

globosum *Fusarium* Rheeder *et al.*, Mycologia 88: 509. 1996.
Holotypus: BPI 802834.

Ex-type culture: CBS 428.97 = DOAM 214966 = FRC M-8014 = IMI 375330 = MRC 6647 = NRRL 26131 = PREM 51878.

Type locality: **South Africa**, Eastern Cape Province, Butterworth district, Teko Experimental Farm.

Type substrate: *Zea mays*.

Descriptions and illustrations: See Rheeder *et al.* (1996) and Leslie & Summerell (2006).

Diagnostic DNA barcodes: *rpb1*: KF466396; *rpb2*: KF466406; *tef1*: KF466417.

globulosum *Fusarium* Pass., in Rabenhorst, Fungi Eur. Exs. no. 2262. 1877.

Syntypes: In BPI, CUP, ILL & S (Fungi Eur. Exs. # 2262).

Type locality: **Italy**, Parma.

Type substrate: *Salvia verticillata*.

Note: Not *Fusarium* fide Wollenweber & Reinking (1935).

gloeosporioides *Fusarium* Speg. (as 'gloeosporoide'), Anales Mus. Nac. Hist. Nat. Buenos Aires 6: 350. 1898 [1899].

(See ***Fusarium incarnatum***)

Holotypus: In LPS fide Farr (1973).

Type locality: **Argentina**, La Plata.

Type substrate: Fruits of *Passiflora tweediana*.

Note: Synonym fide Wollenweber & Reinking (1935).

gloeosporioides *Fusarium* (Speg.) Sacc. & Trotter, Syll. Fung. 22: 1482. 1913, *nom. illegit.*, Art. 53.1.

Basionym: *Selenosporium gloeosporioides* Speg. (as 'gloeosporioides'), Anales Mus. Nac. Hist. Nat. Buenos Aires 13: 458. 1911.

(See ***Fusarium lateritium***)

Holotypus: In LPS (Myc. Argent. ser. 5, no. 1167) fide Farr (1973).

Type locality: **Argentina**, Buenos Aires.

Type substrate: *Pircunia dioica*.

Note: Synonym fide Wollenweber & Reinking (1935).

glumarum *Fusarium* Sacc., Syll. Fung. 4: 706. 1886.

Replaced synonym: *Fusarium pallens* Berk. & M.A. Curtis, Grevillea 3: 99. 1875, *nom. illegit.*, Art. 53.1, *non Fusarium pallens* Nees & T. Nees 1818.

(See ***Fusarium incarnatum***)

Syntype: CBRU00007755.

Type locality: **USA**.

Type substrate: *Juncus* sp.

Note: Synonym fide Wollenweber & Reinking (1935).

glycines* *Fusarium L. Lombard *et al.*, Persoonia 41: 25. 2018 [2019].

Holotypus: CBS H-23614.

Ex-type culture: CBS 144746 = CPC 25808.

Type locality: **South Africa**, North West Province.

Type substrate: *Glycine max*.

Descriptions and illustrations: See Lombard *et al.* (2019b).

Diagnostic DNA barcodes: *rpb1*: MW928809; *rpb2*: MH484942; *tef1*: MH485033.

goolgardi* *Fusarium D.M. Robinson *et al.*, Fungal Diversity 77: 357. 2015 [2016].

Holotypus: RGB5411.

Ex-type culture: NRRL 66250 = RGB5411.

Type locality: **Australia**, New South Wales, Bungonia State Conservation Area.

Type substrate: *Xanthorrhoea glauca*.

Descriptions and illustrations: See Laurence *et al.* (2016).

Diagnostic DNA barcodes: *rpb1*: KP083270; *rpb2*: KP083280; *tef1*: KP101123.

gossypinum* *Fusarium L. Lombard & Crous, Persoonia 41: 26. 2018 [2019].

Holotypus: CBS H-23615.

Ex-type culture: CBS 116613.

Type locality: **Ivory Coast**, Bouaké.

Type substrate: *Gossypium hirsutum*.

Descriptions and illustrations: See Lombard *et al.* (2019b).

Diagnostic DNA barcodes: *rpb2*: MH484909; *tef1*: MH485000.

gracile *Fusarium* McAlpine, Proc. Linn. Soc. New South Wales 28: 554. 1903.

(See ***Fusarium avenaceum***)

Holotypus: VPRI 2564.

Type locality: **Australia**, Victoria, Sandringham.

Type substrate: Flowering stem of *Lobelia gibbosa*.

Note: Synonym fide Wollenweber & Reinking (1935).

gracilipes* *Fusarium J.W. Xia *et al.*, Persoonia 43: 205. 2019.

Holotypus: CBS H-24059.

Ex-type culture: NRRL 43635.

Type locality: **USA**, Nebraska.

Type substrate: *Equus* sp. (horse).

Descriptions and illustrations: See Xia *et al.* (2019).

Diagnostic DNA barcodes: *rpb1*: HM347188; *rpb2*: GQ505840; *tef1*: GQ505662.

graminearum* *Fusarium Schwabe, Fl. Anhalt. 2: 285. 1839.

Synonyms: *Sphaeria zae* Schwein., Schriften Naturf. Ges. Leipzig 1: 48. 1822, *non Fusarium zae* (Westend.) Sacc. 1886.

Dothidea zae (Schwein.) Schwein., Trans. Amer. Philos. Soc., n.s., 4: 230. 1832.

Hendersoniopsis zae (Schwein.) Woron., Fungal and Bacterial Diseases of Agricultural Plants: 255. 1922.

Gibberella zae (Schwein.) Petch, Ann. Mycol. 34: 260. 1936.

Fusarium stictoides Durieu & Mont., Explor. Sci. Algérie 1: 334. 1848.

Sphaeria saubinetii Durieu & Mont., Explor. Sci. Algérie 1: 479. 1849.

Gibbera saubinetii (Durieu & Mont.) Mont., Syll. Gen. Sp. Crypt.: 252. 1856.

Botryosphaeria saubinetii (Durieu & Mont.) Niessl, Verh. Naturf. Vereins Brünn 10: 195. 1872.

Gibberella pulicaris subsp. *saubinetii* (Durieu & Mont.) Sacc., Michelia 1: 317. 1878.

Gibberella saubinetii (Durieu & Mont.) Sacc., Michelia 1: 513. 1879.

Fusisporium insidiosum Berk., Gard. Chron. 1860: 480. 1860.

Fusarium insidiosum (Berk.) Sacc., Syll. Fung. 4: 707. 1886, *nom. illegit.*, Art. 53.1.

Gibberella saubinetii var. *coronillae* Sacc., Michelia 1: 513. 1879.

Fusarium mollerianum Thüm., Inst. Coimbra 28: 263. 1881.

Gibberella saubinetii subsp. *pachyspora* Sacc., Michelia 2: 74. 1880.

- Gibberella saubinetii* var. *pachyspora* (Sacc.) Sacc., Syll. Fung. 2: 555. 1883.
- Fusarium caricis* Oudem., Verslagen Meded. Afd. Natuurk. Kon. Akad. Wetensch., ser. 3, 7: 325. 1890.
- Fusarium graminearum* var. *caricis* (Oudem.) Wollenw., Z. Parasitenk. (Berlin) 3: 365. 1931.
- ?*Fusarium rhoicola* Fautrey, Rev. Mycol. (Toulouse) 17: 171. 1895.
- Fusarium funicola* Tassi, Bull. Lab. Orto Bot. Reale Univ. Siena 3: 131. 1900.
- Gibberella saubinetii* f. *acuum* Feltgen, Vorstud. Pilzfl. Luxemburg, Nachtr. III: 303. 1903.
- Gibberella saubinetii* var. *acuum* (Feltgen) Sacc. & D. Sacc., Syll. Fung. 17: 813. 1905.
- Gibberella saubinetii* var. *tetraspora* Feltgen, Vorstud. Pilzfl. Luxemburg, Nachtr. III: 302. 1903.
- Gibberella saubinetii* var. *calami* Henn., Beibl. Hedwigia 42: (79). 1903.
- Gibberella saubinetii* var. *mate* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 17: 129. 1908.
- ?*Selenosporium bufonicola* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires, ser. 3, 13: 458. 1910.
- ?*Fusarium bufonicola* (Speg.) Sacc. & Trotter, Syll. Fung. 22: 1486. 1913.
- Fusarium rostratum* Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land- Forstw. 8: 30. 1910 [1913].
- Gibberella saubinetii* var. *flacca* Wollenw., Z. Parasitenk. (Berlin) 3: 433. 1931.
- Lectotypus* (*hic designatus*, MBT 10000689): **Germany**, inflorescence of *Triticum* sp., 1839, S.H. Schwabe, in Flora Anhaltina 2, tab. VI. fig. 7.
- Epitypus* (*hic designatus*, MBT 10000690): **Germany**, *Hordeum vulgare*, 1988, L. Niessen, CBS 136009 (preserved as metabolically inactive culture).
- Ex-epitype culture*: CBS 136009.
- Descriptions and illustrations*: See Booth (1971), Gerlach & Nirenberg (1982) and Leslie & Summerell (2006).
- Diagnostic DNA barcodes*: *rpb1*: MW928810; *rpb2*: MW928826; *tef1*: MW928838.
- Notes*: This well-known and economically important pathogen of gramineous hosts has a global distribution and is accepted as originally circumscribed. However, no type material is available for taxonomic reference. Therefore, a lectotype based on an illustration from the original protologue and an epitype is designated here to provide taxonomic stability for this species.
- graminum Fusarium*** Corda, Icon. Fung. 1: 3. 1837.
- Synonym*: *Fusarium herbarum* var. *graminum* (Corda) Wollenw., Fusaria Autogr. Delin. 3: 891. 1930.
- Fusarium avenaceum* var. *graminum* (Corda) Raillo, Fungi of the Genus *Fusarium*: 188. 1950.
- Fusarium corallinum* Sacc., Nuovo Giorn. Bot. Ital. 8: 196. 1876.
- Lectotypus* (*hic designatus*, MBT 10000691): **Germany**, gramineous plant, 1837, A.C.J. Corda, in Icon. Fung. 1, tab. I, fig. 59.
- Descriptions and illustrations*: See Wollenweber & Reinking (1935) and Gerlach & Nirenberg (1982).
- Notes*: This species is recognised by Wollenweber & Reinking (1935) and Gerlach & Nirenberg (1982). Recollection from the type host and locality is required. No holotype specimen could be located and therefore an illustration is designated as lectotype.
- granulare Fusarium* Kalchbr., Crypt. Austro-Afric., no. 1068. 1874.
- (See ***Fusarium sambucinum***)
- Holotypus*: ?B 70 0100191 (Crypt. Austro-Afric., no. 1068).
- Type locality*: **South Africa**, Eastern Cape Province, Somerset-East.
- Type substrate*: *Datura stramonium* (syn. *Datura tatula*).
- Note*: *Synonym fide* Wollenweber & Reinking (1935).
- granulosum Fusarium* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 45: 466. 1894 [1893].
- (See ***Fusarium avenaceum***)
- Holotypus*: Commons 2091 in NY.
- Type locality*: **USA**, Delaware, New Castle, Mount Cuba.
- Type substrate*: *Smilax hispida*.
- Note*: *Synonym fide* Wollenweber & Reinking (1935).
- grosnichelii Fusarium*** Maryani et al., Stud. Mycol. 92: 176. 2018 [2019].
- Holotypus*: InaCC F833 (preserved as metabolically inactive culture).
- Ex-type culture*: InaCC F833.
- Type locality*: **Indonesia**, West Java, Bogor, Suakarya (Megamendung).
- Type substrate*: Pseudostem of *Musa acuminata* var. Pisang Ambon Lumut.
- Descriptions and illustrations*: See Maryani et al. (2019a).
- Diagnostic DNA barcodes*: *rpb1*: LS479548; *rpb2*: LS479295; *tef1*: LS479744.
- guilinense Fusarium*** M.M. Wang et al., Persoonia 43: 80. 2019.
- Holotypus*: HAMS 248037.
- Ex-type culture*: CGMCC 3.19495 = LC12160.
- Type locality*: **China**, Guangxi Province, Guilin.
- Type substrate*: Leaf of *Musa nana*.
- Descriptions and illustrations*: See Wang et al. (2019).
- Diagnostic DNA barcodes*: *rpb1*: MK289831; *rpb2*: MK289747; *tef1*: MK289594.
- guttiforme Fusarium*** Nirenberg & O'Donnell, Mycologia 90: 446. 1998.
- Holotypus*: B 70 0001690.
- Ex-type culture*: BBA 69661 = CBS 409.97 = IMI 376113 = NRRL 25295.
- Type locality*: **Brazil**.
- Type substrate*: *Ananas comosus*.
- Descriptions and illustrations*: See Nirenberg & O'Donnell (1998).
- Diagnostic DNA barcodes*: *rpb1*: MT010938; *rpb2*: MT010967; *tef1*: KC514066.
- gymnosporangii Fusarium* Jaap, Ann. Mycol. 14: 44. 1916.
- Nectria gymnosporangii*** (Jaap) Rossman, Mycotaxon 8: 515. 1979.
- Basionym*: *Calonectria gymnosporangii* Jaap, Ann. Mycol. 14: 10. 1916.
- Synonyms*: *Bactridium gymnosporangii* (Jaap) Wollenw., Fusaria Autogr. Delin. 1: 458. 1916.
- Cylindrocarpon gymnosporangii* (Jaap) Rossman, Mycol. Pap. 150: 31. 1983.
- Holotypus*: In HBG *fide* Rossman (1979).
- Type locality*: **Croatia**, Dalmatia, Lapad near Ragusa.
- Type substrate*: Parasitic on *Gymnosporangium confusum* on *Juniperus phoenicea* branches.

haematococcum Fusarium Nalim *et al.*, Mycologia 103: 1322. 2011.

Neocosmospora haematococca (Berk. & Broome) Samuels *et al.*, Mycologia 103: 1322. 2011.

Basionym: *Nectria haematococca* Berk. & Broome, J. Linn. Soc., Bot. 14: 116. 1875.

Synonyms: *Dialonectria haematococca* (Berk. & Broome) Cooke, Grevillea 12: 110. 1884.

Cucurbitaria haematococca (Berk. & Broome) Kuntze, Revis. Gen. Pl. 3: 461. 1898.

Hypomyces haematococcus (Berk. & Broome) Wollenw., Angew. Bot. 8: 191. 1926.

Haematonectria haematococca (Berk. & Broome) Samuels & Nirenberg, Stud. Mycol. 42: 135. 1999.

?*Nectria lanata* Pat., Bull. Soc. Mycol. France 8: 52. 1892 (*fide* Samuels 1976).

?*Nectria aurantiella* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 6: 287. 1898.

?*Nectria episphaerioides* Penz. & Sacc., Malpighia 11: 511. 1898 [1897].

?*Nectria cinnabarina* var. *jaraguensis* Höhn., Denkschr. Kaiserl. Akad. Wiss. Wien, Math.-Naturwiss. Kl. 83: 18. 1907.

?*Nectria bogoriensis* C. Bernard, Bull. Dép. Agric. Indes Néerl. 11: 45. 1907.

?*Nectria victoriae* Henn., in Rehm, Ann. Mycol. 5: 81. 1907, *nom. inval.*, Art. 38.1(a).

?*Nectria calonectricola* Henn., Hedwigia 48: 105. 1908.

?*Nectria citri* Henn., Hedwigia 48: 104. 1908.

?*Nectria luteococcinea* Höhn., Sitzungsber. Kaiserl. Akad. Wiss. Wien, Math.-Naturwiss. Cl., Abt. 1. 118: 299. 1909.

?*Nectria bainii* var. *hypoleuca* Sacc., Nuovo Giorn. Bot. Ital. 23: 205. 1916.

?*Nectria confluens* Seaver, Sci. Surv. Porto Rico & Virgin Islands 8: 44. 1926, *nom. illegit.*, Art. 53.1.

Lectotypus: K(M) 252877, designated in Samuels (1976).

Lectotype locality: **Sri Lanka**.

Lectotype substrate: Unknown

Epitypus: BPI 871363, designated in Nalim *et al.* (2011).

Ex-epitype culture: CBS 119600 = FRC S-1832.

Epitype locality: **Sri Lanka**, Sabaragamuwa Province, Sinharaja Man and Biosphere Reserve, Morningside, vicinity Bungalow in forested slope.

Epitype substrate: Dying tree.

Descriptions and illustrations: See Nalim *et al.* (2011).

Diagnostic DNA barcodes: *rpb2*: LT960561; *tef1*: KM231926.

hainanense Fusarium M.M. Wang *et al.*, Persoonia 43: 82. 2019.

Holotypus: HAMS 248038.

Ex-type culture: CGMCC 3.19478 = LC11638.

Type locality: **China**, Hainan Province.

Type substrate: Stem of *Oryza* sp.

Descriptions and illustrations: See Wang *et al.* (2019).

Diagnostic DNA barcodes: *rpb1*: MK289833; *rpb2*: MK289735; *tef1*: MK289581.

hakeae Fusarium Henn., Verh. Bot. Vereins Prov. Brandenburg 40: 175. 1899.

Gloeosporium hakeae (Henn.) Wollenw., Fusaria Autogr. Delin. 1: 494. 1916.

Holotypus: In B *fide* Hein (1988).

Type locality: **Germany**, Berlin.

Type substrate: *Hakea salicifolia*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

heidelbergense Fusarium Sacc., Ann. Mycol. 8: 346. 1910.

(See ***Fusarium culmorum***)

Holotypus: In PAD.

Type locality: **Germany**, Heidelberg.

Type substrate: *Cymbidium* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

helgardnirenbergiae Fusarium O'Donnell *et al.*, Index Fungorum 440: 2. 2020.

Neocosmospora nirenbergiana Sand.-Den. & Crous, Persoonia 43: 143. 2019.

Holotypus: CBS H-23988.

Ex-type culture: BBA 65023 = CBS 145469 = G.J.S. 87-127 = NRRL 22387.

Type locality: **French Guiana**.

Type substrate: Bark of unidentified tree.

Descriptions and illustrations: See Sandoval-Denis *et al.* (2019).

Diagnostic DNA barcodes: *rpb2*: EU329505; *tef1*: AF178339.

helianthi Fusarium (Schwein.) Wollenw., Fusaria Autogr. Delin. 2: 555. 1924.

Basionym: *Vermicularia subeffigurata helianthi* Schwein., Trans. Amer. Philos. Soc., n.s., 4: 228. 1832.

(See ***Fusarium tricinctum***)

Holotypus: PH00078405.

Type locality: **Unknown**.

Type substrate: *Helianthus annuus*.

Note: Synonyms *fide* Wollenweber & Reinking (1935).

helotioides Fusarium Berk. & M.A. Curtis, in Berkeley, Grevillea 3: 98. 1875.

Holotypus: ?K(M).

Type locality: **USA**, Alabama.

Type substrate: *Ilex decidua* (syn. *Ilex prinoides*).

Notes: Status unclear. Not *Fusarium* *fide* Wollenweber & Reinking (1935).

hengyangense Fusarium (Z.Q. Zeng & W.Y. Zhuang) O'Donnell *et al.*, Index Fungorum 440: 2. 2020.

Neocosmospora hengyangensis Z.Q. Zeng & W.Y. Zhuang, Phytotaxa 319: 179. 2017.

Holotypus: HMAS 254518.

Ex-type culture: HMAS 248884.

Type locality: **China**, Hunan, Hengyang, Gouloufeng.

Type substrate: Twigs.

Descriptions and illustrations: See Zeng & Zhuang (2017b).

Diagnostic DNA barcodes: *tef1*: KY829448.

herbarum Fusarium (Corda) Fr., Summa Veg. Scand. 2: 472. 1849.

Basionym: *Selenosporium herbarum* Corda, Icon. Fung. 3: 34, tab. 6, fig. 88. 1839.

(See ***Fusarium avenaceum***)

Typus: PRM 155731.

Type locality: **Czech Republic**, Prague.

Type substrate: Gramineous plant part.

Note: Synonyms *fide* Wollenweber & Reinking (1935). Lectotypification pending study of material lodged in PRM.

heteronemum Fusarium Berk. & Broome (as '*heteronema*'), Ann. Mag. Nat. Hist., ser. 3, 15: 402. 1865.

(See *Fusarium candidum* (Link) Sacc.)

Holotypus: ?K(M).

Type locality: **UK**, Batheaston.

Type substrate: Decaying *Pyrus* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

heterosporioides *Fusarium* Fautrey, in Roumeguère, *Fungi Sel. Gall. Exs. No.* 5399. 1890 and *Rev. Mycol. (Toulouse)* 12: 126. 1890.

Syntype: ILL00219542 (Roumeguère, *Fungi Sel. Gall. Exs. no.* 5399).

Type locality: **France**, Charny

Type substrate: *Sclerotium clavus* on *Glyceria fluitans*.

Notes: Status unclear. Not *Fusarium* *fide* Wollenweber & Reinking (1935).

heterosporum* *Fusarium Nees & T. Nees, *Nova Acta Phys.-Med. Acad. Caes. Leop.-Carol. Nat. Cur.* 9: 235. 1818.

Synonyms: ?*Fusarium leucoconium* Corda, *Icon. Fung.* 1: 4. 1837. [*fide* Booth (1971)].

Sphaeria cyanea Sollm., *Bot. Zeitung (Berlin)* 21: 193. 1863.

Botryosphaeria cyanea (Sollm.) Weese, *Sitzungsber. Kaiserl. Akad. Wiss. Wien, Math.-Naturwiss. Cl., Abt. 1*, 128: 707. 1919.

Gibberella cyanea (Sollm.) Wollenw., *Fusaria Autogr. Delin.* 1: 39. 1919.

Fusarium secalis Fée, *Mém. Soc. Mus. Hist. Nat. Strassbourg* 3: 35. 1843.

Fusarium eleocharidis Rostr. (as '*heleocharidis*'), in *Thümen, Mycoth. Univ., Cent.* 22, no. 2185. 1883.

Fusisporium lolii Wm.G. Sm., *Diseases of field and garden crops, chiefly as are caused by fungi*: 213. 1884.

Fusarium lolii (Wm.G. Sm.) Sacc., *Syll. Fung.* 11: 652. 1895.

Fusarium heterosporum var. *lolii* (Wm.G. Sm.) Wollenw., *Z. Parasitenk. (Berlin)* 3: 349. 1931.

Fusarium heterosporum f. *paspali* Ellis & Everh., in *Ellis, North Amer. Fung., Ser. 2*, no. 2395. 1886.

Fusarium parasiticum Ellis & Kellerm., *J. Mycol.* 3: 127. 1887, *nom. illegit.*, Art. 53.1.

Fusarium pucciniophilum Sacc. & P. Syd., *Syll. Fung.* 14: 1128. 1899.

Fusarium stromaticum Delacr., *Bull. Soc. Mycol. France* 9: 186. 1893.

Fusarium paspalicola Henn., *Monsunia* 1: 38. 1899 [1900].

Fusarium heterosporum var. *paspalicola* (Henn.) Wollenw., *Z. Parasitenk. (Berlin)* 3: 349. 1931.

Fusarium congoense Wollenw., *Fusaria Autogr. Delin.* 1: 307. 1916.

Fusarium heterosporum var. *congoense* (Wollenw.) Wollenw., *Z. Parasitenk. (Berlin)* 3: 350. 1931.

Fusarium heterosporum f. *aleuritis* Saccas & Drouillon (as '*aleuritidis*'), *Agron. Trop.* 6: 251. 1951.

Gibberella gordonii C. Booth, *The Genus Fusarium*: 177. 1971.

Lectotypus (*hic designatus*, MBT 10000692): **Germany**, sclerotium of *Claviceps purpurea* on a spike of *Triticum* sp., 1818, G.C.D. Nees von Esenbeck, in *Nova Acta Phys.-Med. Acad. Caes. Leop.-Carol. Nat. Cur.*, tab. V. fig. 5.

Epitypus (*hic designatus*, MBT 10000693): **Germany**, Rotenburg near Bremen, sclerotium of *Claviceps purpurea* on *Lolium perenne*, Aug. 1967, U.G. Schlösser, CBS 391.68 (preserved as metabolically inactive culture).

Ex-epitype culture: CBS 391.68 = NRRL 25798.

Descriptions and illustrations: See Wollenweber & Reinking (1935), Booth (1971), Gerlach & Nirenberg (1982) and Leslie & Summerell (2006).

Diagnostic DNA barcodes: *rpb1*: MW928811; *rpb2*: MW928827; *tef1*: MW928839.

Notes: This species is recognised by Wollenweber & Reinking (1935), Gerlach & Nirenberg (1982), Booth (1971), and Leslie & Summerell (2006). Index Fungorum indicates that the correct name for this species is *F. lolii*. However, this name is not commonly used and considered as a synonym of *F. heterosporum*. Additionally, the epithet '*heterosporum*' is older than the epithet '*lolii*' and should have priority. No holotype specimen is available and therefore an illustration is designated as lectotype.

heveae *Fusarium* Vincens, *Bull. Soc. Pathol. Vég. France* 2: 19. 1915.

(See ***Fusarium incarnatum***)

Holotypus: ?PC.

Type locality: **Brazil**, Para.

Type substrate: *Hevea brasiliensis*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

hexaseptatum* *Fusarium Maryani *et al.*, *Stud. Mycol.* 92: 183. 2018 [2019].

Holotypus: InaCC F866 (preserved as metabolically inactive culture).

Ex-type culture: InaCC F866.

Type locality: **Indonesia**, West Java, Sukabumi, Parakan Lima.

Type substrate: *Musa acuminata* var. *Pisang Ambon Kuning*.

Descriptions and illustrations: See Maryani *et al.* (2019a).

Diagnostic DNA barcodes: *rpb2*: LS479359; *tef1*: LS479805.

hibermans *Fusarium* Lindau, *Rabenh. Krypt.-Fl.*, ed. 2, 1(9): 542. 1909, *nom. superfl.*, Art. 52.1.

Basionym: *Fusarium nivale* Ces. ex Berl. & Voglino, in Saccardo, *Syll. Fung., Addit. I–IV*: 390. 1886, *non* (Fr.) Sorauer, 1901.

(See *Fusarium nivale*)

Authentic material: Klotzsch, *Herb. Viv. Mycol.* no. 1439 in HAL.

Original locality: **Italy**, Vercelli.

Original substrate: Leaves of overwintered crop.

Note: Synonyms *fide* Wollenweber & Reinking (1935).

hippocastani *Fusarium* (Corda) Sacc., *Syll. Fung.* 4: 703. 1886.

Basionym: *Selenosporium hippocastani* Corda, *Icon. Fung.* 2: 7. 1838.

(See ***Fusarium acuminatum***)

Lectotypus (*hic designatus*, MBT 10000694): **Czech Republic**, Prague, *Aesculus hippocastanum*, 1836, A.C.J. Corda, in *Icon. Fung.* 2: tab. IX. fig. 31.

Notes: According to Pilát (1938) and Holubová-Jechová *et al.* (1994), no material was preserved in PRM. Therefore, an illustration is selected as lectotype.

hoodiae* *Fusarium L. Lombard *et al.*, *Persoonia* 41: 27. 2018 [2019].

Holotypus: CBS H-23616.

Ex-type culture: CBS 132474.

Type locality: **South Africa**, Northern Cape Province, Prieska.

Type substrate: Root of *Hoodia gordonii*.

Descriptions and illustrations: See Lombard *et al.* (2019b).

Diagnostic DNA barcodes: *rpb2*: MH484929; *tef1*: MH485020.

hordearium *Fusarium* Ducomet, *Rech. Dével. Champ. Parasit.* 87. 1907.

Holotypus: ?MPA.

Type locality: **France**.

Type substrate: Unknown.

Notes: Status unclear. Not *Fusarium* fide Wollenweber & Reinking (1935).

hordei *Fusarium* (Wm.G. Sm.) Sacc., Syll. Fung. 11: 652. 1895. Basionym: *Fusisporium hordei* Wm.G. Sm., Diseases of field and garden crops, chiefly as are caused by fungi: 212. 1884.

(See *Fusarium sambucinum*)

Lectotypus (*hic designatus*, MBT 10000695): Denmark, *Hordeum* sp., 1884, W. G. Smith, in Diseases of field and garden crops, chiefly as are caused by fungi: 211, fig. 94.

Notes: Synonym fide Wollenweber & Reinking (1935). No holotype material could be located and therefore an illustration is designated as lectotype.

hostae* *Fusarium Geiser & Juba, Mycologia 93: 672. 2001.

Synonym: *Gibberella hostae* Geiser & Juba, Mycologia 93: 672. 2001.

Holotypus: BPI 748169.

Ex-type culture: FRC O-2074 = NRRL 29889.

Type locality: USA, South Carolina.

Type substrate: *Hosta* sp.

Descriptions and illustrations: See Geiser et al. (2001).

Diagnostic DNA barcodes: *rpb1*: JX171527; *rpb2*: JX171640; *tef1*: AY329034.

humi* *Fusarium (Reinking) Nirenberg & Hagedorn, Nachrichtenbl. Deutsch. Pflanzenschutzdienstes 60: 215. 2008.

Basionym: *Fusarium tumidum* var. *humi* Reinking, Zentralbl. Bakteriol., 2. Abth. 89: 513. 1934.

Lectotypus (*hic designatus*, MBT 10000706): Honduras, soil, 1931, O.A. Reinking, in Wollenweber's Fusaria Autogr. Delin. no. 1152 of type culture 5236.

Notes: This species is recognised by Wollenweber & Reinking (1935), Gerlach & Nirenberg (1982), and Nirenberg & Hagedorn (2008). Recollection from the type host and locality is required. No holotype material could be located and therefore an illustration is designated as lectotype.

humicola* *Fusarium L. Lombard & Crous, Fungal Syst. Evol. 4: 191. 2019.

Holotypus: CBS H-24016.

Ex-type culture: ATCC 24372 = CBS 124.73 = IMI 128101 = NRRL 25535.

Type locality: Pakistan.

Type substrate: Soil.

Descriptions and illustrations: See Lombard et al. (2019a).

Diagnostic DNA barcodes: *rpb1*: MN120718; *rpb2*: MN120738; *tef1*: MN120757.

humuli* *Fusarium M.M. Wang et al., Persoonia 43: 83. 2019.

Holotypus: HAMS 248039.

Ex-type culture: CGMCC 3.19374 = CQ1039.

Type locality: China, Jiangsu Province.

Type substrate: Leaves of *Humulus scandens*.

Descriptions and illustrations: See Wang et al. (2019).

Diagnostic DNA barcodes: *rpb1*: MK289840; *rpb2*: MK289724; *tef1*: MK289570.

hydnicola *Fusarium* Ellis & Everh. (as '*hydnicolum*'), J. Mycol. 4(4–5): 45. 1888.

Alysidium hypophleodes (Corda) Bonord., Handb. Mykol.: 35. 1851.

Basionym: *Fusidium hypophleodes* Corda, Icon. Fung. 1: 3, tab. 1, fig. 50. 1837.

Holotypus: NY (fide Index Fungorum).

Type locality: USA, Missouri, Concordia.

Type substrate: Bark of dead *Hydnum membranaceum*.

Note: Synonym fide Wollenweber & Reinking (1935).

hymenula *Fusarium* Pound & Clem., Bot. Surv. Nebraska 4: 7. 1896.

Gloeosporium intermedium* var. *brevipes Sacc., Syll. Fung. 3: 703. 1884.

Holotypus: NEB0040541.

Type locality: USA, Nebraska.

Type substrate: *Helianthus* sp.

Notes: Synonym fide Wollenweber & Reinking (1935). The name is misspelled as '*lymenula*' in the NEB database.

hyperoxysporum *Fusarium* Wollenw., J. Agric. Res. 2: 268. 1914. (See *Fusarium oxysporum*)

Holotypus: Not located.

Type locality: USA.

Type substrate: *Ipomoea batatas*.

Note: Synonym fide Wollenweber & Reinking (1935).

hypocreoideum *Fusarium* Cooke & Masee, Grevillea 16: 76. 1888.

Aschersonia hypocreoidea (Cooke & Masee) Petch, Ann. Roy. Bot. Gard. (Peradeniya) 7: 255. 1922.

Holotypus: K(M) 127920.

Type locality: Australia, Queensland.

Type substrate: *Ficus aspera*.

hypodermium *Fusarium* (Link) Link, in Willdenow, Sp. Pl., ed. 4, 6: 96. 1825.

Basionym: *Fusidium hypodermium* Link, Mag. Neuesten Entdeck. Gesammten Naturk. Ges. Naturf. Freunde Berlin 8: 31. 1816 [1815].

Marssonina aurantiaca (Link) Magnus, Hedwigia 45: 90. 1906.

Basionym: *Cryptosporium aurantiacum* Link, in Willdenow, Sp. Pl., ed 4, 6: 96. 1825, *nom. sanct.* (Fries, Syst. Mycol. 3: 481. 1832).

Synonyms: *Fusidium aurantiacum* (Link) Fr., Syst. Mycol. 3: 481. 1832.

Gloeosporium aurantiacum (Link) Sacc., Syll. Fung. 3: 717. 1884.

Marssonina aurantiaca (Link) Rostr., Bot. Tidsskr. 19: 217. 1895.

Note: Synonyms fide Wollenweber & Reinking (1935).

hypothernemi *Fusarium* (Sand.-Den. & Crous) O'Donnell et al., Index Fungorum 440: 2. 2020.

Neocosmospora hypothernemi Sand.-Den. & Crous, Persoonia 43: 132. 2019.

Holotypus: CBS H-23982.

Ex-type culture: ARSEF 5878 = CBS 145464 = NRRL 52782.

Type locality: Benin, Niaouli.

Type substrate: Adult *Hypothenemus hampei* (coffee borer beetle).

Descriptions and illustrations: See Sandoval-Denis et al. (2019).

Diagnostic DNA barcodes: *rpb1*: MW218117; *rpb2*: JF741176; *tef1*: JF740850.

idahoanum *Fusarium* O.A. Pratt, J. Agric. Res. 13: 86. 1918. (See *Fusarium flocciferum*)

Lectotypus (hic designatus, MBT 10000707): **USA**, Idaho, soil, 1918, O.A. Pratt, in J. Agric. Res. 13: 87, fig. 2.

Notes: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#). No holotype material could be located and therefore an illustration is designated as lectotype.

illosporioides *Fusarium* Sacc., Harriman Alaska Exped. 5: 15. 1904.

(See *Fusarium lateritium*)

Holotypus: In PAD.

Type locality: **USA**, Alaska, Sitka.

Type substrate: *Ribes* sp.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

illudens *Fusarium* C. Booth, The Genus *Fusarium*: 54. 1971.

Neocosmospora illudens (Berk.) L. Lombard & Crous, Stud. Mycol. 80: 227. 2015.

Basionym: *Nectria illudens* Berk., in Hooker, Bot. Antarct. Voy. II (Fl. Nov.-Zel.): 203. 1855.

Synonyms: *Cucurbitaria illudens* (Berk.) Kuntze, Revis. Gen. Pl. 3: 461. 1898.

Haematonectria illudens (Berk.) Samuels & Nirenberg, Stud. Mycol. 42: 136. 1999.

Neotypus: PAD S00012, designated in [Forin et al. \(2020\)](#)

Neotype locality: **New Zealand**.

Neotype substrate: Bark of unknown host plant,

“*inaequale Fusarium*” Auersw. Bot. Zeitung (Berlin) 8: 439. 1850, typographic error (see *Notes*).

Ramularia rosea (Fuckel) Sacc., Fungi Ital. Del., Tab. 1001. 1881.

Basionym: *Fusidium roseum* Fuckel, Fungi Rhen. Fasc. III, no. 219. 1863.

Synonyms: *Ovularia rosea* (Fuckel) Masee, Brit. Fung.-Fl. 3: 323. 1893.

Cylindrospora rosea (Fuckel) J. Schröt., in Cohn, Krypt.-Fl. Schles., Pilze II: 493. 1897.

Fusidium inaequale Auersw., in Rabenh., Klotzschii Herb. Viv. Mycol., Cent. 14: no. 1383. 1850.

Ramularia lucidae Davis, Trans. Wis. Acad. Sci. Art. Lett. 19: 687. 1919.

Authentic material: Rabenh., Klotzschii Herb. Viv. Mycol. 1383 in HAL.

Original locality: **Germany**, Leipzig.

Original substrate: *Salix amygdalina*.

Notes: Not *Fusarium* *fide* [Wollenweber & Reinking \(1935\)](#). This species was first published as *Fusidium inaequale* Auersw., in Rabenh., Klotzschii Herb. Viv. Mycol., Cent. 14: no. 1383, 1850. The description was repeated in Bot. Zeitung 8: 439, 1850 and Flora 33: 283, 1850 (in the latter publication also under *Fusidium*), so that in the simultaneous publication in “Botanische Zeitung” the “F.” was undoubtedly also meant to be *Fusidium* and not *Fusarium*. Syntype material deposited at HAL has recently been examined, and *Fusidium inaequale* turned out to be a heterotypic synonym of *Ramularia rosea* (Fuckel) Sacc (see [Braun 1998](#)).

incarcerans *Fusarium* (Berk.) Sacc., Syll. Fung. 4: 713. 1886.

Basionym: *Fusisporium incarcerans* Berk., Intellectual Observ. 2: 11. 1863.

(See *Fusarium avenaceum*)

Holotypus: ?K(M).

Type locality: **UK**, Northamptonshire, Fotheringhay Castle.

Type substrate: *Orthotrichum* sp.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

incarnatum Fusarium (Roberge ex Desm.) Sacc., Syll. Fung. 4: 712. 1886.

Basionym: *Fusisporium incarnatum* Roberge ex Desm., Ann. Sci. Nat., Bot., sér. 3, 11: 274. 1849.

Synonyms: *Fusarium semitectum* Berk. & Ravenel, Grevillea 3: 98. 1875.

Pseudofusarium semitectum (Berk. & Ravenel) Matsush., Icon. Microfung. Matsush. Lect. (Kobe): 119. 1975.

Fusarium pallens Berk. & M.A. Curtis, Grevillea 3: 99. 1875, *nom. illegit.*, Art. 53.1, *non Fusarium pallens* Nees & T. Nees 1818.

Fusarium glumarum Sacc., Syll. Fung. 4: 706. 1886 (*nom. nov.* for *F. pallens* Berk. & M.A. Curtis).

Fusisporium pallidoroseum Cooke, Grevillea 6: 139. 1878.

Fusarium pallidoroseum (Cooke) Sacc., Syll. Fung. 4: 720. 1886.

Fusarium asparagi Briard, Rev. Mycol. (Toulouse) 12: 142. 1890.

Fusarium gloeosporioides Speg. (as ‘*gloeosporioides*’), Anales Mus. Nac. Hist. Nat. Buenos Aires 6: 350. 1899.

Fusarium juglandinum Peck, Bull. Torrey Bot. Club 36: 157. 1909.

Fusarium heveae Vincens, Bull. Soc. Pathol. Vég. France 2: 19. 1915.

Fusarium tenuistipes Sacc., Atti Mem. Reale Accad. Sci. Lett. Arti, Padova 33: 195. 1917.

Fusarium semitectum var. *majus* Wollenw., Fusaria Autogr. Delin. 3: 907–910. 1930.

Fusarium semitectum var. *violaceum* Batikyan & Abramyan (as ‘*violaceae*’), Biol. Zhurn. Armenii 22: 58. 1969, *nom. inval.*, Art. 39.1.

Lectotypus: (*hic designatus*, MBT 10001327) **France**, from *Tagetes erecta*, 1848, M. Roberge in Desmazières, Pl. Crypt. N. France, éd 2, No. 1303, in PC.

Epitypus: (*hic designatus*, MBT 10001328) **Malawi**, on *Trichosanthes dioica*, date unknown, H.M. Phiri, CBS H-24060.

Ex-epitype culture: ATCC 24387 = CBS 132.73 = IMI 128222 = NRRL 25478.

Descriptions and illustrations: See [Booth \(1971\)](#), [Gerlach & Nirenberg \(1982\)](#) and [Xia et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb2*: MN170409; *tef1*: MN170476.

Note: The epitypification of *Fusarium incarnatum* by [Xia et al. \(2019\)](#) was not effective as the holo- or lectotype was not correctly indicated (Art. 9.9). Here, a lectotype is selected and the epitypification is validated.

inflexum Fusarium R. Schneid., in Schneider & Dalchow, Phytopathol. Z. 82: 80. 1975.

Holotypus: DSM 63203.

Ex-type culture: ATCC 32213 = BBA 63203 = CBS 716.74 = DAOM 225130 = DSM 63203 = IMI 375336 = NRRL 20433.

Type locality: **Germany**, Hamburg, Vierlanden.

Type substrate: Stem of *Vicia faba*.

Descriptions and illustrations: See [Schneider & Dalchow \(1975\)](#) and [Gerlach & Nirenberg \(1982\)](#).

Diagnostic DNA barcodes: *rpb1*: JX171469; *rpb2*: JX171583; *tef1*: AF008479.

inseptatum Fusarium Schwein., Trans. Amer. Philos. Soc., n.s., 4: 302. 1832 [1834].

Holotypus: PH00062493.

Type locality: **USA**, Pennsylvania, Bethlehem.

Type substrate: *Daphne mezereum*.

Notes: Status unclear. Not *Fusarium* fide Wollenweber & Reinking (1935).

insidiosum *Fusarium* Roum., *Michelia* 2: 132. 1880.

(See *Fusarium lateritium*)

Syntypes: In BR, CUP & ILL (Roum., *Fungi Sel. Gall. Exs. No.* 57).

Type locality: **France**, Pyrénées-Orientales, Environs de Perpignan.

Type substrate: *Phytolacca decandra*.

Note: Synonym fide Wollenweber & Reinking (1935).

insidiosum *Fusarium* (Berk.) Sacc., *Syll. Fung.* 4: 707. 1886, *nom. illegit.*, Art. 53.1.

Basionym: *Fusisporium insidiosum* Berk., *Gard. Chron.* 1860: 480. 1860.

(See *Fusarium graminearum*)

Holotypus: ?K(M).

Type locality: **UK**.

Type substrate: *Agrostis pulchella*.

Note: Synonyms fide Wollenweber & Reinking (1935).

ipomoeae *Fusarium* M.M. Wang *et al.*, *Persoonia* 43: 83. 2019.

Holotypus: HAMS 248040.

Ex-type culture: CGMCC 3. 19496 = LC12165.

Type locality: **China**, Jiangsu Province.

Type substrate: Leaves of *Ipomoea aquatica*.

Descriptions and illustrations: See Wang *et al.* (2019).

Diagnostic DNA barcodes: *rpb1*: MK289859; *rpb2*: MK289752; *tef1*: MK289599.

iranicum *Fusarium* Torbati *et al.*, *Mycol. Progr.* 18: 129. 2018 [2019].

Holotypus: CBS H-23560.

Ex-type culture: CBS 143608 = CPC 30860.

Type locality: **Iran**, West Azerbaijan Province, Orumieh-Salmas.

Type substrate: *Agaricus bisporus*.

Descriptions and illustrations: See Torbati *et al.* (2019).

Diagnostic DNA barcodes: *rpb2*: LT970757; *tef1*: LT970785.

iridis *Fusarium* Oudem., *Ned. Kruidk. Arch.*, 2 sér. 5: 515. 1889.

(See *Fusarium avenaceum*)

Holotypus: ?L.

Type locality: **Netherlands**.

Type substrate: *Iris pseudacorus*.

Note: Synonym fide Wollenweber & Reinking (1935).

irregulare *Fusarium* M.M. Wang *et al.*, *Persoonia* 43: 84. 2019.

Holotypus: HAMS 248041.

Ex-type culture: CGMCC 3.19489 = LC7188.

Type locality: **China**, Guangdong Province.

Type substrate: *Bambusoideae*.

Descriptions and illustrations: See Wang *et al.* (2019).

Diagnostic DNA barcodes: *rpb1*: MK289863; *rpb2*: MK289783; *tef1*: MK289629.

japonicum *Fusarium* Allesch., *Beibl. Hedwigia* 36: (164). 1897.

(See *Fusarium tortuosum*)

Syntype: S-F45631 (Sydow, *Mycoth. March.* no. 4592).

Type locality: **Germany**, Berlin.

Type locality: *Prunus japonica*.

Note: Synonym fide Wollenweber & Reinking (1935).

javanicum *Fusarium* Koord., *Verh. Kon. Akad. Wetensch., Afd. Natuurk.*, Sect. 2, 13: 247. 1907.

Holotypus: Not located.

Type locality: **Indonesia**, Central Java, Purworejo.

Type substrate: *Ficus elastica*.

Note: Status unclear fide Sandoval-Denis *et al.* (2019).

juglandinum *Fusarium* Peck, *Bull. Torrey Bot. Club* 36: 157. 1909.

(See *Fusarium incarnatum*)

Holotypus: NYSf1607.

Type locality: **USA**, Kansas, Rooks, Stockton.

Type substrate: *Juglans nigra*.

Note: Synonym fide Wollenweber & Reinking (1935).

junci *Fusarium* P. Crouan & H. Crouan, *Fl. Finistère*: 14. 1867.

Holotypus: ?CO.

Type locality: **France**, Paris.

Type substrate: *Juncus effusus*.

Note: ?*Fusidium* fide Wollenweber & Reinking (1935).

jungiae *Fusarium* Pat., *Bull. Soc. Mycol. France* 11: 234. 1895.

(See *Fusarium avenaceum*)

Holotypus: FH00965356.

Type locality: **Argentina**, San Jorge.

Type substrate: Parasitic on *Puccinia* sp. on *Jungia* sp.

Note: Synonym fide Wollenweber & Reinking (1935).

juruanum *Fusarium* Henn., *Hedwigia* 43: 398. 1904.

(See *Fusarium coccidicola*)

Holotypus: In B.

Type locality: **Brazil**, Rio Jurua.

Type substrate: *Annonaceae* sp.

Note: Synonym fide Gerlach & Nirenberg (1982).

kalimantanense *Fusarium* Maryani *et al.*, *Stud. Mycol.* 92: 187. 2018 [2019].

Holotypus: InaCC F917 (preserved as metabolically inactive culture).

Ex-type culture: InaCC F917.

Type locality: **Indonesia**, Central Kalimantan, Katingan, Pulau Malan.

Type substrate: *Musa acuminata* var. Pisang Ambon.

Descriptions and illustrations: See Maryani *et al.* (2019a).

Diagnostic DNA barcodes: *rpb1*: LS479497; *rpb2*: LS479241; *tef1*: LS479690.

kelerajum *Fusarium* Samuels *et al.*, *Mycologia* 103: 1326. 2011.

Neocosmospora keleraja Samuels *et al.*, *Mycologia* 103: 1326. 2011.

Holotypus: BPI 871413.

Ex-type culture: FRC S-1839 = G.J.S. 02-122.

Type locality: **Sri Lanka**, Minneriya Natl. Forest.

Type substrate: Trunk of *Yakuda marang*.

Descriptions and illustrations: See Nalim *et al.* (2011).

Diagnostic DNA barcode: *tef1*: DQ247518.

keratoplasticum *Fusarium* Geiser *et al.*, *Fung. Gen. Biol.* 53: 68. 2013.

Neocosmospora keratoplastica (Geiser *et al.*) Sand.-Den. & Crous, *Persoonia* 41: 120. 2018.

Synonyms: *Cephalosporium keratoplasticum* T. Morik., *Mycopathologia* 2: 66. 1939, *nom. inval.*, Art. 39.1.

Hyalopus keratoplasticum T. Morik. ex M.A.J. Barbosa, *Subsidios Para o Estudo Parasitologico do Genero Hyalopus Corda*, 1838: 19. 1941, *nom. inval.*, Art. 39.1.

Fusarium sedimenticola M.M. Wang *et al.*, *Botanica Marina* 63: 174. 2020.

Holotypus: FRC S-2477.

Ex-type culture: CBS 490.63 = FRC S-2477 = NRRL 22661.

Type locality: **USA**, Virginia, Winchester.

Type substrate: Indoor plumbing.

Descriptions and illustrations: See [Nalim et al. \(2011\)](#).

Diagnostic DNA barcodes: *rpb1*: MW218121; *rpb2*: JN235897; *tef1*: JN235712.

konzum Fusarium Zeller et al., Mycologia 95: 947. 2003.

Synonym: *Gibberella konza* Zeller et al., Mycologia 95: 947. 2003.

Holotypus: DAR 76034.

Ex-type culture: CBS 119849 = KSU 10653 = NRRL 53394.

Type locality: **USA**, Kansas, Manhattan, Konza Prairie Biological Station.

Type substrate: *Sorghastrum nutans*.

Descriptions and illustrations: See [Zeller et al. \(2003\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: LT996200; *rpb2*: LT996148; *tef1*: LT996098.

kotabaruense Fusarium Maryani et al., Persoonia 43: 65. 2019.

Holotypus: InaCC F963 (preserved as metabolically inactive culture).

Ex-type culture: InaCC F963.

Type locality: **Indonesia**, South Kalimantan, Kota Baru, Kecamatan Pamukan Barat, Desa Sungai Birah.

Type substrate: *Musa* var. Pisang Hawa.

Descriptions and illustrations: See [Maryani et al. \(2019b\)](#).

Diagnostic DNA barcodes: *rpb1*: LS479875; *rpb2*: LS479859; *tef1*: LS479445.

kuehnii Fusarium (Fuckel) Sacc., Syll. Fung. 4: 714. 1886.

Basionym: *Fusisporium kuehnii* Fuckel, Fungi Rhen. Exs., Suppl., Fasc. 5, no. 1920. 1867.

?***Athelia arachnoidea*** (Berk.) Jülich, Willdenowia 7: 53. 1972. (fide [Gerlach & Nirenberg 1982](#))

Basionym: *Corticium arachnoideum* Berk., Ann. Mag. Nat. Hist., ser. 1, 13: 345. 1844.

Synonym: *Fusisporium devastans* J.G. Kühn, Krankh. Kultur-gew.: 32. 1858, *nom. inval.*, Art. 38.1(a).

Syntype: Fuckel, Fungi Rhen. Exs., Suppl., Fasc. 5, 1920 (e.g., HAL).

Type locality: **Germany**.

Type substrate: Lichens and mosses.

Notes: Status doubtful. Considered a possible synonym of *F. dimerum* by [Booth \(1971\)](#).

kurdicum Fusarium Petr., Sydowia 13: 96. 1959.

Cosmospora kurdica (Petr.) Rossman & Samuels, Stud. Mycol. 42: 122. 1999.

Basionym: *Calonectria kurdica* Petr., Sydowia 13: 95. 1959.

Synonyms: *Nectria kurdica* (Petr.) Rossman, Mycol. Pap. 150: 35. 1983.

?*Stagonopsis sclerotioides* Höhn., Ann. K. K. Naturhist. Hofmus. 20: 368. 1905.

?*Botryocrea sclerotioides* (Höhn.) Petr., Sydowia 3: 141. 1949.

?*Fusarium sclerotioides* (Höhn.) Samuels & Rossman, Mycol. Pap. 164: 23. 1991.

Holotypus: K.H. Rechinger, 31 Jul. 1957, in W.

Type locality: **Iran**, Kurdistan.

Type substrate: *Astragalus* sp.

Note: Synonyms fide [Rossman et al. \(1999\)](#).

kuroshium Fusarium F. Na et al., Plant Disease 102: 1159. 2018, *nom. inval.*, Art. 40.7.

Neocosmospora kuroshio F. Na et al. ex Sand.-Den. & Crous, Persoonia 43: 137. 2019.

Holotypus: BPI 910340.

Ex-type culture: CBS 142642 = UCR 3641.

Type locality: **USA**, California, San Diego, El Cajon.

Type substrate: *Euwallacea* sp. galleries in *Platanus racemosa*.

Descriptions and illustrations: See [Na et al. \(2018\)](#).

Diagnostic DNA barcodes: *rpb1*: KX262236; *rpb2*: KX262256; *tef1*: KX262216.

kurunegalense Fusarium Samuels et al., Mycologia 103: 1323. 2011.

Neocosmospora kurunegalensis Samuels et al., Mycologia 103: 1324. 2011.

Holotypus: BPI 871391.

Ex-type culture: CBS 119599 = G.J.S. 02-94.

Type locality: **Sri Lanka**, Wagamba Province, Kurunegala.

Type substrate: Recently felled tree.

Descriptions and illustrations: See [Nalim et al. \(2011\)](#).

Diagnostic DNA barcodes: *rpb1*: MW834228; *rpb2*: LR583838; *tef1*: DQ247511.

kyushuense Fusarium O'Donnell & T. Aoki, Mycoscience 39: 2. 1998.

Holotypus: NIAES99701.

Ex-type culture: ATCC 56750 = FRC T-346A = MAFF 237645 = MRC 1767 = NRRL 3509.

Type locality: **Japan**, Kumamoto.

Type substrate: Seed of *Triticum aestivum*.

Descriptions and illustrations: See [Aoki & O'Donnell \(1998\)](#).

Diagnostic DNA barcodes: *rpb2*: MH582098; *tef1*: MH582292.

laboulbeniae Fusarium Cépède, Arch. Parasitol. 16: 373. 1914. (See *Fusarium larvarum*)

Holotypus: Not located.

Type locality: **France**, Pas-de-Calais, Wimereux.

Type substrate: *Demetrias unipunctata*.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

lacterarum Fusarium Subrahm. (as '*laceratum*'), Mykosen 26: 478. 1983.

Holotypus: IMI 300797.

Ex-type culture: ATCC 42771 = CBS 130185 = IMI 300797 = NRRL 20423.

Type locality: **India**, Poona, Pimpri.

Type substrate: Skin of lizard.

Descriptions and illustrations: See [Subrahmanyam \(1983\)](#).

Diagnostic DNA barcodes: *rpb1*: JX171467; *rpb2*: JX171581; *tef1*: GQ505593.

lactis Fusarium Pirota, Arch. Lab. Bot. Crittog. Univ. Pavia 2 & 3: 316. 1879.

Synonyms: ?*Fusarium pyrinum* Schwein., Trans. Amer. Philos. Soc., n.s. 4: 302. 1834.

?*Fusarium apiogenum* Sacc., Syll. Fung. 4: 717. 1886.

Fusarium rubrum Parav., Ann. Mycol. 16: 311. 1918.

Lectotypus: Arch. Lab. Bot. Crittog. Univ. Pavia 2 & 3, Tab. 21, figs 1–6, designated by [Yilmaz et al. \(2021\)](#).

Lectotype locality: **Italy**, Pavia.

Lectotype substrate: Clotted milk.

Epitypus: B 70 0001686, designated by [Yilmaz et al. \(2021\)](#).

Ex-epitype culture: BBA 68590 = CBS 411.97 = IMI 375351 = NRRL 25200.

Epitype locality: **USA**, California.

Epitype substrate: *Ficus carica*.

Descriptions and illustrations: See [Nirenberg & O'Donnell \(1998\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: LT996201; *rpb2*: LT996149; *tef1*: AF160272.

lagenariae *Fusarium* (Schwein.) Sacc., Syll. Fung. 4: 724. 1886.
Basionym: *Fusisporium lagenariae* Schwein., Trans. Amer. Philos. Soc., n.s., 4: 275. 1834.

(See *Fusarium oxysporum*)

Holotypus: PH00062516

Type locality: **USA**, Pennsylvania, Bethlehem.

Type substrate: *Lagenaria siceraria*.

lagenarium *Fusarium* Pass., Erb. Critt. Ital., ser. 2: no. 148. 1871.

Synonym: *Gloeosporium lagenarium* (Pass.) Sacc. & Roum., Rev. Mycol., Toulouse 2(8): 201. 1880.

(See *Fusarium cyclogenum*)

Holotypus: In PAD.

Type locality: **Italy**, Parma.

Type substrate: *Lagenaria* sp.

Note: *Synonym fide* [Wollenweber & Reinking \(1935\)](#).

lanceolatum *Fusarium* O.A. Pratt, J. Agric. Res. 13: 83. 1918.

(See *Fusarium acuminatum*)

Lectotypus (*hic designatus*, MBT 10000709): **USA**, Idaho, from soil, 1918, O.A. Pratt, in J. Agric. Res. 13: 82, fig. 1A–E.

Notes: *Synonym fide* [Wollenweber & Reinking \(1935\)](#). No holotype specimen could be located and therefore an illustration is designated as lectotype.

langsethiae *Fusarium* Torp & Nirenberg, Int. J. Food Microbiol. 95: 248. 2004.

Holotypus: B 70 0012234.

Ex-type culture: BBA 70945 = CBS 113234.

Type locality: **Norway**.

Type substrate: Kernal of *Avena sativa*.

Descriptions and illustrations: See [Torp & Nirenberg \(2004\)](#).

Diagnostic DNA barcodes: *rpb1*: MW928812; *rpb2*: MW928828; *tef1*: AB674298.

languescens *Fusarium* L. Lombard & Crous, Persoonia 43: 28. 2018 [2019].

Holotypus: CBS H-23617.

Ex-type culture: CBS 645.78 = NRRL 36531.

Type locality: **Morocco**.

Type substrate: *Solanum lycopersicum*.

Descriptions and illustrations: See [Lombard et al. \(2019b\)](#).

Diagnostic DNA barcodes: *rpb1*: MW928813; *rpb2*: MH484880; *tef1*: MH484971.

laricis *Fusarium* Sawada, Bull. Gov. Forest Exp. Sta., Meguro 46: 130. 1950.

Holotypus: TFM:FPH 00771.

Type locality: **Japan**, Aomori, Kamikita, Noheji

Type substrate: *Larix kaempferi*.

larvarum *Fusarium* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 369. 1870.

Microcera larvarum (Fuckel) Gräfenhan et al., Stud. Mycol. 68: 105. 2011.

Synonyms: *Fusarium nivale* var. *larvarum* (Fuckel) Bilař, Fusarii (Biologija i sistematika): 295. 1955, *nom. inval.*, Art. 41.1

Fusarium cryptum McAlpine, *Fungus Diseases of Citrus trees in Australia*: 106. 1899.

Fusarium epicoccum McAlpine, *Fungus Diseases of Citrus trees in Australia*: 113. 1899.

Microcera parlatoriae Trab., Bull. Agric. Algérie Tunisie 13: 33. 1907.

Microcera curta Sacc., Ann. Mycol. 7: 437. 1909.

Microcera tonduzii Pat., Bull. Soc. Mycol. France 28: 142. 1912.

Fusarium aspidioti Sawada, Bot. Mag. (Tokyo) 28: 312. 1914.

Fusarium laboulbeniae Cépède, Arch. Parasitol. 16: 373. 1914.

Fusarium acremoniopsis Vincens, Bull. Soc. Mycol. France 31: 26. 1915.

?*Fusarium meliolicola* F. Stevens, Bot. Gaz. 65: 245. 1918.

?*Nectria meliolicola* F. Stevens, Bot. Gaz. 65: 231. 1918.

Microcera aurantiicola Petch, Trans. Brit. Mycol. Soc. 7: 158. 1921.

Lectotypus: G 00111015, selected in [Gräfenhan et al. \(2011\)](#).

Lectotype locality: **Germany**, Hessen, Rheingau, near Oestrich-Winkel.

Lectotype substrate: Larva cuticles of insects on *Malus domestica*.

Epitypus: BBA 62239, designated in [Gräfenhan et al. \(2011\)](#).

Ex-epitype culture: BBA 62239 = CBS 738.79 = MUCL 19033 = NRRL 20473.

Epitype locality: **Iran**, Gilan Province, near Rasht.

Epitype substrate: Parasitic on *Quadraspidiotus perniciosus* (scale) on *Prunus domestica*.

Diagnostic DNA barcodes: *rpb1*: KM232252; *rpb2*: KM232387; *tef1*: KM231957.

lateritium *Fusarium* Nees, Syst. Pilze: 31. 1817.

Synonyms: *Selenosporium lateritium* (Nees) Desm., Fl. Cryptog. Flandres 2: 99. 1867.

Fusarium microsporium Schldtl., Fl. Berol. 2: 139. 1824.

Fusarium fructigenum Fr., Syst. Mycol. 3: 471. 1832.

Fusarium lateritium var. *fructigenum* (Fr.) Wollenw., Fusaria Autogr. Delin. 3: 959. 1930.

Sphaeria baccata Wallroth, Fl. Crypt. Germ. 2: 838. 1833.

Gibbera baccata (Wallr.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 167. 1870.

Gibberella pulicaris subsp. *baccata* (Wallr.) Sacc., Michelia 1 (3): 317. 1878.

Gibberella baccata (Wallr.) Sacc., Syll. Fung. 2: 553. 1883.

Fusarium lateritium var. *mori* Desm., Ann. Sci. Nat. Bot., ser. 2, 8: 10. 1837.

Selenosporium urticarum Corda (as '*urticearum*'), Icon. Fung. 2: 7. 1838.

Fusarium urticarum (Corda) Sacc., Syll. Fung. 4: 698. 1886.

?*Fusarium protractum* Lév., Ann. Sci. Nat., Bot., sér. 3, 9: 246. 1848.

Gloeosporium berkeleyi Mont., Ann. Sci. Nat., Bot., sér. 3, 12: 296. 1849.

Fusarium berkeleyi (Mont.) Berk. & Broome, N. Amer. Fung.: 108. 1875.

Botryosphaeria moricola Ces. & De Not., Hedwigia 4: 27. 1865.

Gibberella moricola (Ces. & De Not.) Sacc., Syll. Fung. 2: 553. 1883.

Gibbera euonymi Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 167. 1870.

Gibberella euonymi (Fuckel) Sacc., Michelia 1: 318. 1878.

Hendersonia euonymi (Fuckel) Sacc., Syll. Fung. 2: 556. 1883.

- Selenosporium cydoniae* Schulzer, Verh. K.K. Zool.-Bot. Ges. Wien 21: 1240. 1871.
- Fusarium cydoniae* (Schulzer) Sacc. & Traverso, Syll. Fung. 19: 724. 1910, *nom. illegit.*, Art. 53.1.
- Fusarium sticticum* Berk. & M.A. Curtis, Grevillea 3: 99. 1875.
- Fusisporium zavianum* Sacc., Michelia 1: 83. 1877.
- Fusarium zavianum* (Sacc.) Sacc., Syll. Fung. 4: 709. 1886.
- Fusarium cydoniae* Roum. & Fautrey, Rev. Mycol. (Toulouse) 14: 170. 1892, *nom. illegit.*, Art. 53.1.
- Fusarium salicis* Fuckel, Fungi Rhen. Exs., Suppl., Fasc. 7, no. 2110. 1868.
- Fusarium salicis* var. *minus* Wollenw., Fusaria Autogr. Delin. 2: 582. 1924.
- Fusarium sambucinum* var. *minus* Wollenw., Fusaria Autogr. Delin. 3: 941. 1930.
- Gibbera mori* Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 168. 1870.
- Fusarium semitectum* Berk. & Ravenel, in Berkeley, Grevillea 4: 98. 1875.
- Fusisporium cinnabarinum* Berk. & M.A. Curtis, Grevillea 3: 146. 1875.
- Fusarium cinnabarinum* (Berk. & M.A. Curtis) Sacc., Syll. Fung. 4: 722. 1886.
- Fusisporium miniatum* Berk. & M.A. Curtis, Grevillea 3: 147. 1875.
- Fusarium miniatum* (Berk. & M.A. Curtis) Sacc., Syll. Fung. 4: 722. 1886, *nom. illegit.*, Art. 53.1.
- Fusisporium putaminum* Thüm., Oesterr. Bot. Z. 27: 272. 1877.
- Fusarium putaminum* (Thüm.) Sacc., Syll. Fung. 4: 703. 1886.
- Fusisporium leguminum* Cooke, Grevillea 6: 139. 1878.
- Fusarium leguminum* (Cooke) Sacc., Syll. Fung. 4: 712. 1886.
- Fusarium limonis* Briosi, Att. Staz. Chim. Agrar. Rome. 1878.
- Fusarium yuccae* Cooke, Grevillea 7: 34. 1878, *nom. inval.*, Art. 36.1(a).
- Fusisporium azedarachinum* Thüm., Mycoth. Univ., cent. 14: no. 1379. 1879.
- Fusarium azedarachinum* (Thüm.) Sacc., Syll. Fung. 4: 704. 1886.
- Fusarium insidiosum* Roum., Michelia 2: 132. 1880.
- Fusarium roumegueri* Sacc. (as '*roumegueri*'), Syll. Fung. 4: 702. 1886, *nom. illegit.*, Art. 52.1.
- Fusarium albertii* Roum., Fungi Sel. Gall. Exs., Cent. 19: no. 1867. 1881.
- Fusarium rimicola* Sacc. (as '*rimicolum*'), Michelia 2: 297. 1881.
- Fusarium ziziphinum* Pass., Rev. Mycol. (Toulouse) 4: 22. 1882.
- Fusarium acaciae* Cooke & Harkn., Grevillea 12: 96. 1884.
- Fusarium longisporum* Cooke & Masee, Grevillea 16: 4. 1887.
- Fusarium sphaeroideum* Pass., Atti Reale Accad. Lincei, Rendiconti Cl. Sci. Fis., sér. 4, 4: 105. 1888.
- Fusarium parasiticum* Fautrey, Rev. Mycol. (Toulouse) 11: 153. 1889, *nom. illegit.*, Art. 53.1.
- Fusarium fautreyi* Sacc., Syll. Fung. 10: 934. 1892.
- Fusarium carneoroseum* Cooke, Grevillea 19: 4. 1890.
- Fusarium celtidis* Ellis & Tracy, J. Mycol. 6: 76. 1890.
- Fusarium nucicola* P. Karst. & Har., Rev. Mycol. (Toulouse) 12: 131. 1890.
- Fusarium discoideum* Fautrey & Roum., Rev. Mycol. (Toulouse) 13: 173. 1891.
- Fusarium cydoniae* Allesch., Ber. Bot. Vereines Landshut 12: 130. 1892.
- ?*Fusarium luteum* Clem., Bot. Surv. Nebraska 3: 12. 1894.
- Fusarium asclepiadeum* Fautrey, Rev. Mycol. (Toulouse) 18: 68. 1896.
- Fusarium samararum* Allesch., Ber. Bayer. Bot. Ges. 4: 39. 1896.
- Fusarium sophorae* Allesch., Beibl. Hedwigia 36: (164). 1897.
- Fusarium ailanthinum* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 6: 350. 1899.
- Fusarium euonymi* Syd., Beibl. Hedwigia 39: (6). 1900.
- Fusarium gemmiperda* Aderh., Z. Pflanzenkrankh. 11: 70. 1901.
- Fusarium euonymi-japonici* Henn., Hedwigia 41: 139. 1902.
- Fusarium illosporoides* Sacc., in Saccardo et al., Harriman Alaska Expedition 5: 15. 1904.
- Fusarium schawrowi* Speschnew, Arbeiten Kaukas. Stat. Seidenzucht 10: 30–41. 1906.
- Selenosporium gloeosporioides* Speg. (as '*gloesporioides*'), Anales Mus. Nac. Hist. Nat. Buenos Aires 13: 458. 1911.
- Fusarium gloeosporioides* (Speg.) Sacc. & Trotter, Syll. Fung. 22: 1482. 1913, *nom. illegit.*, Art. 53.1.
- Fusarium briosianum* Ferraris, Fl. Ital. Crypt. Fungi Fasc. 13: 857. 1912.
- Fusarium pseudacaciae* Rapaics, Z. Pflanzenkrankh. 25: 208. 1915.
- Fusarium gleditschiicola* Dearn. & Barthol. (as '*gleditschicola*'), Mycologia 9: 363. 1917.
- Gibberella briosiana* Turconi & Maffei, Atti Ist. Bot. Univ. Pavia, sér. 2, 15: 148. 1918.
- Botryosphaeria briosiana* (Turconi & Maffei) Weese, Sitzungsber. Akad. Wiss. Wien, Math.-Naturwiss. Kl., Abt. 1, 128: 708. 1919.
- Fusarium uncinatum* Wollenw., Ann. Mycol. 15(1/2): 54. 1917.
- Fusarium blackmannii* W. Br. & A.S. Horne (as '*blackmanni*'), Ann. Bot. (London) 38: 379. 1924.
- Fusarium entomophilum* Petch, Trans. Brit. Mycol. Soc. 11: 260. 1925.
- Fusarium lateritium* var. *tenue* Wollenw., Fusaria Autogr. Delin. 3: 955. 1930.
- Gibberella saubinetii* var. *flacca* Wollenw., Z. Parasitenk. (Berlin) 3: 433. 1931.
- Fusarium anisophilum* Picado, J. Dept. Agric. Porto Rico 16: 391. 1932.
- Lectotypus* (*hic designatus*, MBT 10000710): **Germany**, unknown host, 1817, G.C.D. Nees von Esenbeck, in System der Pilze und Schwämme: 31, tab. 2, fig. 26.
- Descriptions and illustrations*: See [Wollenweber & Reinking \(1935\)](#), [Booth \(1971\)](#), [Gerlach & Nirenberg \(1982\)](#), [Nelson et al. \(1983\)](#) and [Leslie & Summerell \(2006\)](#).
- Notes*: Re-collection from the type host and locality is required. No holotype specimen could be located and therefore an illustration was designated as lectotype.
- laxum* *Fusarium* Peck, Bull. New York State Mus. Nat. Hist. 67: 30. 1903.
- (See ***Fusarium oxysporum***)
- Holotypus*: NYS-F-001667.
- Type locality*: **USA**, New York, Albany, Delmar.
- Type substrate*: *Equisetum hyemale*.
- Note*: Synonym *vide* [Wollenweber & Reinking \(1935\)](#).
- leguminum* *Fusarium* (Cooke) Sacc., Syll. Fung. 4: 712. 1886.
- Basionym*: *Fusisporium leguminum* Cooke, Grevillea 6: 139. 1878.
- (See ***Fusarium lateritium***)
- Syntypes*: In CUP, ISC, NEB & PH (Fungi Amer. Exs. no. 298).
- Type locality*: **USA**, South Carolina, Aiken.

Type substrate: *Acacia* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

leucoconium *Fusarium* Corda, Icon. Fung. 1: 4. 1837.

(See *Fusarium heterosporum* and *F. reticulatum*)

Typus: In PRM *fide* Pilat (1938).

Type locality: **Czech Republic**, Prague.

Type substrate: Rotten plants.

Note: Synonym *fide* Wollenweber & Reinking (1935) and Booth (1971). Lectotypification pending study of material lodged in PRM.

libertatis *Fusarium* L. Lombard & Crous, Persoonia 43: 29. 2018 [2019].

Holotypus: CBS H-23618.

Ex-type culture: CBS 144749 = CPC 28465.

Type locality: **South Africa**, Western Cape Province, Robben Island, Van Riebeeck's Quarry.

Type substrate: Rock surface.

Descriptions and illustrations: See Lombard *et al.* (2019b).

Diagnostic DNA barcodes: *rpb2*: MH484944; *tef1*: MH485035.

lichenicola *Fusarium* C. Massal., in Maire & Saccardo, Ann. Mycol. 1: 223. 1903.

Neocosmospora lichenicola (C. Massal.) Sand.-Den. & Crous, Persoonia 41: 120. 2018.

Synonyms: *Bactridium lichenicola* (C. Massal.) Wollenw. ('as *lichenicolum*'), *Fusaria* Autogr. Delin. 1: 456. 1916.

Cylindrocarpon lichenicola (C. Massal.) D. Hawksw., Bull. Brit. Mus. (Nat. Hist.), Bot. 6: 273. 1979.

Selenosporium lichenicola Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires, ser. 3, 13: 459. 1911.

Fusarium lichenicola (Speg.) Sacc. & Trotter, Syll. Fung. 22: 1486. 1913, *nom. illegit.*, Art. 53.1.

Monacrosporium tedeschi A. Agostini (as '*tedeschi*'), Atti Ist. Bot. Univ. Lab. Crittog. Pavia, ser. 3, 4: 195. 1933.

Euricoa dominguesii Bat. & H. Maia, Anais Soc. Biol. Pernambuco 13: 152. 1955.

Hyaloflorea ramosa Bat. & H. Maia, Anais Soc. Biol. Pernambuco 13: 155. 1955.

Neocosmospora ramosa (Bat. & H. Maia) L. Lombard & Crous, Stud. Mycol. 80: 227. 2015.

Mastigosporium heterosporum R.H. Petersen, Mycologia 51: 729. 1959.

Holotypus: In PAD.

Epitypus: CBS H-23983, designated in Sandoval-Denis *et al.* (2019).

Ex-epitype culture: CBS 623.92.

Epitype locality: **Germany**, Göttingen.

Epitype substrate: Necrotic wounds of *Homo sapiens* under chemotherapy.

Descriptions and illustrations: See Sandoval-Denis *et al.* (2019).

Diagnostic DNA barcodes: *rpb2*: LR583845; *tef1*: LR583620.

limonis *Fusarium* Briosi, Ann. R. Staz. Chim.-Agrar. Sper. Roma. 1878.

(See *Fusarium lateritium*)

Holotypus: Not located.

Type locality: **Italy**, Sicily.

Type substrate: *Citrus limon*.

Notes: Synonym *fide* Wollenweber & Reinking (1935). Protologue not located.

limosum *Fusarium* Rostr., Bot. Tidsskr. 22: 263. 1899.

(See *Fusarium avenaceum*)

Holotypus: C-F-111719.

Type locality: **Sweden**.

Type substrate: Mixture of lime and sugar.

Note: Synonym *fide* Wollenweber & Reinking (1935).

lineare *Fusarium* Moesz, Bot. Közlem. 19: 57. 1920.

(See *Fusarium obtusisporum*)

Holotypus: ?BP.

Type locality: **Hungary**.

Type substrate: *Staphylea pinnata*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

lini *Fusarium* Bolley, Proc. Annual Meeting Soc. Promot. Agric. Sci. 22: 42. 1901.

(See *Fusarium oxysporum*)

Holotypus: Not located.

Type locality: **USA**.

Type substrate: *Linum usitatissimum*.

lini *Fusarium* Remer, Jahresber. Schles. Ges. Vaterl. Cult. 80: 25. 1903, *nom. illegit.*, Art. 53.1

Holotypus: Not located.

Type locality: **Poland**.

Type substrate: *Linum* sp.

liriodendri *Fusarium* (Sand.-Den. & Crous) O'Donnell *et al.*, Index Fungorum 440: 2. 2020.

Neocosmospora liriodendri Sand.-Den. & Crous, Persoonia 43: 139. 2019.

Holotypus: CBS H-23984.

Ex-type culture: BBA 67587 = CBS 117481 = G.J.S 91-148 = NRRL 22389.

Type locality: **USA**, Maryland.

Type substrate: *Liriodendron tulipifera*.

Descriptions and illustrations: See Sandoval-Denis *et al.* (2019).

Diagnostic DNA barcodes: *rpb1*: MW218124; *rpb2*: EU329506; *tef1*: AF178340.

loliaceum *Fusarium* Ducomet, Ann. École Natl. Agric. Rennes 2: 14. 1909.

(See *Fusarium nivale*)

Holotypus: ?MPA.

Type locality: **France**.

Type substrate: Unknown.

Note: Synonym *fide* Wollenweber & Reinking (1935).

lolii *Fusarium* (Wm.G. Sm.) Sacc., Syll. Fung. 11: 652. 1895.

Basionym: *Fusisporium lolii* Wm.G. Sm., Diseases of field and garden crops, chiefly as are caused by fungi: 213. 1884.

(See *Fusarium heterosporum*)

Lectotypus (*hic designatus*, MBT 10000711): **UK**, *Lolium perenne*, date unknown, W.G. Smith, in W.G. Smith, Diseases of field and garden crops, chiefly as are caused by fungi: 213, fig. 96.

Notes: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

loncheceras *Fusarium* Sideris, Phytopathology 14: 213. 1924.

(See *Fusarium oxysporum*)

Lectotypus (*hic designatus*, MBT 10000712): **USA**, California, Stockton, roots of *Allium cepa*, 1924, C.P. Sideris, in Phytopathology 14, pl. XI, fig. of *F. loncheceras*.

Notes: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

longicaudatum Fusarium J.W. Xia *et al.*, *Persoonia* 43: 208. 2019.

Holotypus: CBS H-24061.

Ex-type culture: ATCC 24370 = CBS 123.73 = IMI 160825 = NRRL 25477.

Type locality: Tanzania, Tropical Products Research Inst.

Type substrate: Unknown.

Descriptions and illustrations: See Xia *et al.* (2019).

Diagnostic DNA barcodes: *rpb2*: MN170414; *tef1*: MN170481.

longicornicola Fusarium Sand.-Den., *et al.*, *Persoonia* 46: 149. 2021.

Holotypus: CBS H-24661.

Ex-type culture: ARSEF 6455 = CBS 147247 = NRRL 52706.

Type locality: Ethiopia, Kobo, Welo.

Type substrate: *Aiolopus longicornis*.

Descriptions and illustrations: See Yilmaz *et al.* (2021).

Diagnostic DNA barcodes: *rpb2*: JF741114; *tef1*: JF740788.

longifundum Fusarium J.W. Xia *et al.*, *Persoonia* 43: 208. 2019.

Holotypus: CBS H-24062.

Ex-type culture: CBS 235.79 = NRRL 36372.

Type locality: Netherlands Antilles, Curaçao.

Type substrate: Air.

Descriptions and illustrations: See Xia *et al.* (2019).

Diagnostic DNA barcodes: *rpb2*: GQ505827; *tef1*: GQ505649.

longipes Fusarium Wollenw. & Reinking, *Phytopathology* 15: 160. 1925.

Synonyms: *Fusarium scirpi* var. *longipes* (Wollenw. & Reinking) Wollenw., *Z. Parasitenk.* (Berlin) 3: 337. 1931.

Fusarium equiseti var. *longipes* (Wollenw. & Reinking) Joffe, *Mycopathol. Mycol. Appl.* 53: 221. 1974.

Neotypus (*hic designatus*, MBT 10000713): USA, Florida, soil, 1977, W. Gams, CBS 476.77 (preserved as metabolically inactive culture).

Ex-neotype culture: CBS 476.77 = NRRL 20695.

Descriptions and illustrations: See Gerlach & Nirenberg (1982), Nelson *et al.* (1983).

Diagnostic DNA barcodes: *rpb1*: MW233244; *rpb2*: GQ915493; *tef1*: GQ915509.

Notes: This species is recognised by Gerlach & Nirenberg (1982), Nelson *et al.* (1983), and Leslie & Summerell (2006). No holotype specimen could be located and no illustration accompanied the original protologue. Although an illustration of the original culture (O.A. Reinking no. R34) is provided in Wollenweber's *Fusaria Autogr. Delin.* no. 937 (1924), this cannot be used to designate a lectotype as it does not form part of the original protologue. Therefore, isolate CBS 476.77 is designated as neotype here to provide taxonomic stability to this species, as it appears to have a paraphyletic phylogenetic structure (O'Donnell *et al.* 2013).

longisporum Fusarium Cooke & Masee, *Grevillea* 16: 4. 1887. (See ***Fusarium lateritium***)

Holotypus: K(M) 159680.

Type locality: Australia, Queensland, Brisbane.

Type substrate: Twigs of *Passiflora* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

longissimum Fusarium Sacc. & P. Syd., *Syll. Fung.* 14: 1128. 1899. Replaced synonym: *Fusarium elongatum* De Wild., *Ann. Soc. Belge Microscop.* 17: 43. 1893, *nom. illegit.*, Art. 53.1, non *Fusarium elongatum* Cooke 1890.

(See *Fusarium elongatum* De Wild.)

Holotypus: Not located.

Type locality: Belgium, Brussels, Botanical Garden.

Type substrate: Submerged plant material.

Note: Synonymy *fide* Rossman *et al.* (2016).

longum Fusarium (Wallr.) Sacc., *Syll. Fung.* 4: 719. 1886.

Basionym: *Fusisporium longum* Wallr., *Fl. Crypt. Germ.* 2: 283. 1833.

Holotypus: ?STR.

Type locality: Germany, Berlin.

Type substrate: Dead branch.

Notes: Status unclear. Not *Fusarium* *fide* Wollenweber & Reinking (1935).

louisianense Fusarium L.R. Gale *et al.*, *Fungal Genet. Biol.* 48: 1105. 2011.

Holotypus: BPI 881005.

Ex-type culture: CBS 127525 = NRRL 54197.

Type locality: USA, Louisiana.

Type substrate: Seeds of *Triticum* sp.

Descriptions and illustrations: See Sarver *et al.* (2011).

Diagnostic DNA barcodes: *rpb1*: KM889655; *rpb2*: KM889657; *tef1*: KM889633.

lucidum Fusarium Sherb., *Mem. Cornell Univ. Agric. Exp. Sta.* 6: 157. 1915.

(See ***Fusarium avenaceum***)

Typus: ?CUP-007473.

Type locality: USA, New York.

Type substrate: *Solanum tuberosum*.

Notes: Synonym *fide* Wollenweber & Reinking (1935). Lectotypification pending study of material lodged in CUP.

lucumae Fusarium Henn., *Hedwigia* 48: 116. 1908.

Ascochyta lucumae (Henn.) Wollenw., *Fusaria Autogr. Delin.* 1: 504. 1916.

Syntypes: In BPI, ILL, MIN & WIS (Baker 218).

Type locality: Brazil, Pará.

Type substrate: *Lucuma rivicola*

Note: Synonym *fide* Wollenweber & Reinking (1935).

luffae Fusarium M.M. Wang *et al.*, *Persoonia* 43: 85. 2019

Holotypus: HAMS 248042.

Ex-type culture: CGMCC 3.19497 = LC12167.

Type locality: China, Fujian.

Type substrate: *Luffa aegyptiaca*.

Descriptions and illustrations: See Wang *et al.* (2019).

Diagnostic DNA barcodes: *rpb1*: MK289869; *rpb2*: MK289754; *tef1*: MK289601.

lumajangense Fusarium Maryani *et al.*, *Persoonia* 43: 59. 2019.

Holotypus: InaCC F872 (preserved as metabolically inactive culture).

Ex-type culture: InaCC F872.

Type locality: Indonesia, East Java, Lumajang, Kecamatan Senduro, Desa Kandang Kepus.

Type substrate: *Musa acuminata* var. Pisang Mas Kirana.

Descriptions and illustrations: See Maryani *et al.* (2019b).

Diagnostic DNA barcodes: *rpb2*: LS479850; *tef1*: LS479441.

lunatum *Fusarium* (Ellis & Everh.) Arx, Verh. Kon. Ned. Akad. Wetensch., Afd. Natuurk., Sect. 2, 51: 101. 1957.

Bisifusarium lunatum (Ellis & Everh.) L. Lombard & Crous, Stud. Mycol. 80: 225. 2015.

Basionym: *Gloeosporium lunatum* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 43: 82. 1891.

Synonyms: *Microdochium lunatum* (Ellis & Everh.) Arx, Trans. Brit. Mycol. Soc. 83: 374. 1984.

Fusarium dimerum var. *violaceum* Wollenw., Fusaria Autogr. Delin. 3: 854. 1930.

Holotypus: NY00883039.

Type locality: USA, Texas, San Antonio.

Type substrate: Living leaves of *Opuntia* sp.

Notes: This species requires epitypification. Gerlach & Nirenberg (1982) designated CBS 632.76 (= NRRL 20690) as neotype of *F. dimerum* var. *violaceum*, which was originally collected in Germany. However, Schroers *et al.* (2009) showed that *F. lunatum* is paraphyletic and needs further investigation. Therefore, CBS 632.76 cannot be designated as epitype for *B. lunatum* at this time.

lunulosporum* *Fusarium Gerlach, Phytopathol. Z. 88: 283. 1977.

Holotypus: BBA 62459.

Ex-type culture: ATCC 36747 = BBA 62459 = CBS 636.76 = IMI 322097 = NRRL 13393.

Type locality: South Africa.

Type substrate: *Citrus paradisi*.

Descriptions and illustrations: See Gerlach (1977b), Gerlach & Nirenberg (1982) and Nelson *et al.* (1983).

Diagnostic DNA barcodes: *rpb1*: KM361637; *rpb2*: KM361655; *tef1*: AF212467.

luteum *Fusarium* Clem., Bot. Surv. Nebraska 3: 12. 1894.

(See ***Fusarium lateritium***)

Holotypus: NEB00040542.

Type locality: USA, Nebraska, Lincoln.

Type substrate: Decaying wood.

Note: Synonym *fide* Wollenweber & Reinking (1935).

luteum *Fusarium* Parav., Ann. Mycol. 16: 302. 1918, *nom. illegit.*, Art. 53.1.

(See *Fusarium candidum*)

Authentic material: In Ann. Mycol. 16, pl. 4., figs 1–22.

Original locality: Switzerland.

Original substrate: *Pyrus* sp.

Notes: Synonym *fide* Wollenweber & Reinking (1935).

lutulatum *Fusarium* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 209. 1915.

(See ***Fusarium oxysporum***)

Typus: CUP-007458.

Type locality: USA, Iowa.

Type substrate: *Solanum tuberosum*.

Note: Synonym *fide* Wollenweber & Reinking (1935). Lectotypification pending study of material lodged in CUP.

lyarnte* *Fusarium J.L. Walsh, Sangal., L.W. Burgess, E.C.Y. Liew & Summerell, *sp. nov.* MycoBank MB 837697.

Synonym: *Fusarium lyarnte* J.L. Walsh, Sangal., L.W. Burgess, E.C.Y. Liew & Summerell, Fungal Diversity 44: 153. 2010, *nom. inval.*, Art. 40.7.

Etymology. ‘Lyarnte’, meaning circle in eastern and central Arrernte Aboriginal language (Henderson & Dobson 1994), in reference to the conspicuous globose microconidia.

For diagnosis see Walsh *et al.*, Fungal Diversity 44: 153. 2010.

Holotypus: CBS 125536 (preserved as metabolically inactive culture).

Ex-type culture: CBS 125536 = NRRL 54252 = RBG 5331.

Type locality: Australia, Northern Territory, Litchfield.

Type substrate: Soil.

Descriptions and illustrations: See Walsh *et al.* (2010).

Diagnostic DNA barcodes: *rpb1*: JX171549; *rpb2*: JX171661; *tef1*: EF107118.

Notes: Walsh *et al.* (2010) failed to indicate the holotype for *F. lyarnte*, thereby rendering the species name invalid (Art. 40.7). Here we validate the name.

lycopersici *Fusarium* (Sacc.) Mussat, Syll. Fung. 15: 144. 1901, *nom. inval.*, Art. 36.1(a), (c).

Basionym: *Fusarium oxysporum* subsp. *lycopersici* Sacc., Syll. Fung. 4: 705. 1886.

(See ***Fusarium oxysporum***)

Authentic material: Not located.

Original locality: Italy.

Original substrate: *Solanum lycopersicum*.

lycopersici *Fusarium* Bruschi, Atti Reale Accad. Lincei, Rendiconti Cl. Sci. Fis., ser. 5, 21: 298. 1912.

(See ***Fusarium oxysporum***)

Synonym: *Fusarium bulbigenum* var. *lycopersici* (Bruschi) Wollenw. & Reinking, *Fusarien*: nos. 996–997. 1935.

Holotypus: Not located.

Type locality: Italy.

Type substrate: *Solanum lycopersicum*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

lycopersici *Fusarium* (Sacc.) Wollenw., Phytopathology 3: 29. 1913, *nom. illegit.*, Art. 53.1.

Basionym: *Fusarium oxysporum* subsp. *lycopersici* Sacc., Syll. Fung. 4: 705. 1886.

(See ***Fusarium oxysporum***)

Authentic material: Not located.

Original locality: Italy.

Original substrate: *Solanum lycopersicum*.

macounii *Fusarium* Dearn., Mycologia 9: 363. 1917.

(See ***Fusarium expansum***)

Holotypus: DAOM 223428b.

Type locality: Canada, Vancouver Island.

Type substrate: *Acer* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

macroceras* *Fusarium Wollenw. & Reinking, Phytopathology 15: 166. 1925.

Holotypus: CBS 146.25 (preserved as metabolically inactive culture).

Ex-type culture: CBS 146.25 = NRRL 13958.

Type locality: Honduras.

Type substrate: *Phaseolus vulgaris*.

Descriptions and illustrations: See Wollenweber & Reinking (1925, 1935) and Gerlach & Nirenberg (1982).

Notes: Phylogenetic inference (not shown) revealed that the ex-type culture housed at CBS clustered within the *N. petrophila* clade, indicating a possible strain transposition or contamination of the culture in the past. These species are not morphologically

conspicuous based on the original protologue (Wollenweber & Reinking 1925) of *F. macroceras*.

macrosporum *Fusarium* (Sand.-Den. et al.) O'Donnell et al., Index Fungorum 440: 2. 2020.

Neocosmospora macrospora Sand.-Den. et al., Persoonia 40: 21 2017 [2018].

Holotypus: CBS H-23023.

Ex-type culture: CBS 142424 = CPC 28191.

Type locality: **Italy**, Sicily, Catania, Guardia.

Type substrate: *Citrus sinensis*.

Descriptions and illustrations: See Sandoval-Denis et al. (2018a).

Diagnostic DNA barcodes: *rpb1*: MW218125; *rpb2*: LT746331; *tef1*: LT746218.

macroxyposium *Fusarium* Lindf., Meddel. Centralanst. Försöksväs. Jordbruksomr. Avd. Lantbruksbot. 25: 8. 1922.

(See ***Fusarium oxysporum***)

Holotypus: Not located.

Type locality: **Sweden**.

Type substrate: *Pinus sylvestris*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

maculans *Fusarium* Béranger, Atti Riunione Sci. Ital. 6: 474. 1845.

Neophloeospora maculans (Béranger) Videira & Crous, Stud. Mycol. 87: 338. 2017.

Synonyms: *Phloeospora maculans* (Béranger) Allesch., Rabenh. Krypt.-Fl., ed. 2, 1: 935. 1900.

Phloeospora maculans (Béranger) Höhn., Mitt. Bot. Inst. Techn. Hochsch. Wien 4: 77. 1927.

Cercospora maculans (Béranger) F.A. Wolf, J. Elisha Mitchell Sci. Soc. 51: 165. 1935.

Septoria mori Lév., Ann. Sci. Nat., Bot., ser. 3, 5: 279. 1846.

Cheilaria mori (Lév.) Desm., Ann. Sci. Nat., Bot., ser. 3, 8: 27. 1847.

Phloeospora mori (Lév.) Sacc., *Michelia* 1: 175. 1878.

Septogloeum mori (Lév.) Briosi & Cavara, Fung. Paras. Pianta Colt. Util., Fasc. 1: no. 21. 1888.

Cylindrosporium mori (Lév.) Berl., Riv. Patol. Veg. 5: 205. 1896.

Sphaeria mori Nitschke, Fungi Rhen. Exs. no. 1784. 1866, *nom. inval.*, Art. 38.1(a).

Sphaerella mori Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 106. 1870.

Mycosphaerella mori (Fuckel) F.A. Wolf, J. Elisha Mitchell Sci. Soc. 51: 165. 1935.

Sphaerella morifolia Pass., Erb. Critt. Ital., Ser. 2, Fasc. 30, no. 1464. 1885.

Mycosphaerella morifolia (Pass.) Cruchet, Bull. Soc. Vaud. Sci. Nat. 55: 43. 1923.

Cercospora pulvinulata f. angulosa Săvul. & Sandu, Herb. Mycol. Roman. no. 188. 1931.

Holotypus: Not located.

Type locality: **Italy**.

Type substrate: Leaves of *Morus* sp.

madaense* *Fusarium Ezekiel et al., MycoKeys 67: 112. 2020.

Holotypus: CBS H-24346.

Ex-type culture: CBS 146669 = CPC 38344.

Type locality: **Nigeria**, Nasarawa, Mada Station.

Type substrate: *Arachis hypogaea*.

Descriptions and illustrations: See Ezekiel et al. (2020).

Diagnostic DNA barcodes: *rpb1*: LR792575; *rpb2*: LR792589; *tef1*: LR792625.

magnoliae-champaca* *Fusarium R.H. Perera et al., Mycosphere 11: 2140. 2020.

Holotypus: MFLU 18-2736.

Ex-type culture: MFLUCC 18-0580.

Type locality: **Thailand**, Chiang Rai, Mae Fah Luang University garden.

Type substrate: Dried fruits of *Magnolia champaca*.

Descriptions and illustrations: See Perera et al. (2020).

Diagnostic DNA barcode: *rpb2*: MT212198.

magnusianum *Fusarium* Allesch., Fungi Bav. no. 400. 1895. (See *Fusarium aquaeductuum*)

Holotypus: In M.

Type locality: **Germany**, München.

Type substrate: *Salix incana*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

mahasenii *Fusarium* Samuels et al., Mycologia 103: 1325. 2011.

Neocosmospora mahasenii Samuels et al., Mycologia 103: 1325. 2011.

Holotypus: BPI 881228.

Ex-type culture: CBS 119594 = FRC S-1845 = G.J.S. 02-105.

Type locality: **Sri Lanka**, North Central Province, Giritale. Giritale Forest Training Center.

Type substrate: Small branch of live tree.

Descriptions and illustrations: See Nalim et al. (2011).

Diagnostic DNA barcodes: *rpb1*: MW834231; *rpb2*: LT960563; *tef1*: DQ247513.

mali *Fusarium* Allesch., Ber. Bot. Vereines Landshut 12: 130. 1892.

(See *Fusarium candidum*)

Holotypus: In M.

Type locality: **Germany**, München.

Type substrate: *Malus pumila*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

malli *Fusarium* Taubenh., Bull. Texas Agric. Exp. Sta. 273: 25. 1921.

(See *Fusarium solani*)

Holotypus: ?CUP-011254.

Type locality: **USA**, Texas, Brazos, College Station.

Type substrate: *Allium cepa*.

Note: Typification pending study of material lodged in CUP.

malvacearum *Fusarium* Taubenh., Bull. Texas Agric. Exp. Sta. 260: 27. 1920.

(See ***Fusarium oxysporum***)

Lectotypus (*hic designatus*, MBT 10000714): **USA**, Texas, *Abelmoschus esculentus*, 1920, J.J. Taubenhhaus, in Taubenhhaus, Bull. Texas Agric. Exp. Sta. 260: 30, fig. 8g.

Notes: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration was designated as lectotype.

mangiferae* *Fusarium Britz et al., Mycologia 94: 725. 2002.

Holotypus: PREM 57299.

Ex-type culture: CBS 120994 = KSU 11781 = MRC 7559 = MUCL 54671 = NRRL 53980.

Type locality: **Israel**, Bet Dagan, Volcani Center.
Type substrate: *Mangifera indica*.
Descriptions and illustrations: See [Britz et al. \(2002\)](#).
Diagnostic DNA barcodes: *rpb1*: MW402530; *rpb2*: LT575059;
tef1: LT574978.

marasasianum Fusarium Herron et al., Stud. Mycol. 80: 146. 2015.

Holotypus: PREM 60899.
Ex-type culture: CBS 137238 = CMW 25261.
Type locality: **Colombia**, Vivero Peñas Negra, Valle del Cauca.
Type substrate: *Pinus patula*.
Descriptions and illustrations: See [Herron et al. \(2015\)](#).
Notes: Comparisons of recently generated sequences for the living ex-type (CBS 137238 = CMW 25261) of *F. marasasianum* indicate a strain transposition or contamination by another *Fusarium* species. Therefore, this species needs to be recollected from the type locality and substrate or sequences need to be generated from the holotype specimen.

marginatum Fusarium Berk. & M.A. Curtis, Grevillea 3: 97. 1875.
Holotypus: ?K(M).

Type locality: **USA**, Alabama, Beaumont.
Type substrate: *Smilax* sp.
Note: Not *Fusarium* fide [Wollenweber & Reinking \(1935\)](#).

martiellae-discolorioides Fusarium Batikyan, Biol. Zhurn. Armenii 22: 87. 1969, *nom. inval.*, Art. 39.1.

Authentic material: Not located.
Original locality: **Armenia**.
Original substrate: Soil of wheatfield.
Notes: Published without Latin diagnosis fide [Gerlach & Nirenberg \(1982\)](#). Also described in Biol. Zhurn. Armenii 26(2): 73. 1973, but also not in Latin.

martii Fusarium Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land- Forstw. 8: 83. 1913.

Neocosmospora martii (Appel & Wollenw.) Sand.-Den. & Crous, Persoonia 43: 137. 2019.

Synonyms: *Fusarium solani* var. *martii* (Appel & Wollenw.) Wollenw., Fusaria Autogr. Delin. 3: 1034. 1930.

Neocosmospora croci Guarnaccia et al., Persoonia 40: 17. 2017 [2018].

Lectotypus: BPI 452385, selected in [Sandoval-Denis et al. \(2019\)](#).

Epitypus: CBS H-23986, designated in [Sandoval-Denis et al. \(2019\)](#).

Ex-epitype culture: CBS 115659 = FRC S-0679 = MRC 2198.

Lecto- and epitype locality: **Germany**, Berlin.

Lecto- and epitype substrate: *Solanum tuberosum*.

Descriptions and illustrations: See [Sandoval-Denis et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW834232; *rpb2*: JX435256;
tef1: JX435156.

massalimae Fusarium A.D. Cavalcanti et al., Mycol. Progr. 19: 1137. 2020.

Holotypus: URM 94324.

Ex-type culture: URM 8239.

Type locality: **Brazil**, Alagoas, Quebrangulo, Pedra Talhada Biological Reserve.

Type substrate: *Handroanthus chrysotrichus*.

Descriptions and illustrations: See [Cavalcanti et al. \(2020\)](#).

Diagnostic DNA barcodes: *rpb2*: MN939767; *tef1*: MN939763.

matuoi Fusarium Hosoya & Tubaki, Mycoscience 45: 264. 2004.

Fusicolla matuoi (Hosoya & Tubaki) Gräfenhan & Seifert, Stud. Mycol. 68: 101. 2011.

Synonyms: *Fusarium splendens* Matuo & Takah. Kobay., Trans. Mycol. Soc. Japan 2(4): 13. 1960, *nom. inval.*, Art. 39.1.

Cosmospora matuoi Hosoya & Tubaki, Mycoscience 45: 262. 2004.
Holotypus: TNS F-11127.

Ex-type culture: MAFF 410976.

Type locality: **Japan**, Honshu.

Type substrate: Twigs of *Albizia julibrissin*.

Descriptions and illustrations: See [Hosoya & Tubaki \(2004\)](#).

mauroi Fusarium Av.-Saccá, Revista Agric. (Piracicaba) 8: 93. 1933.

Macronectria jungneri (Henn.) Salgado & P. Chaverri, Fungal Diversity 80: 448. 2016.

Basionym: *Nectria jungneri* Henn., Bot. Jahrb. Syst. 22: 75. 1895.

Synonyms: *Cucurbitaria jungneri* (Henn.) Kuntze, Revis. Gen. Pl. 3: 461. 1898.

Neonectria jungneri (Henn.) Samuels & Brayford (as '*Nenectria*'), Mycologia 96: 580. 2004.

Thelonectria jungneri (Henn.) P. Chaverri & Salgado, Stud. Mycol. 68: 76. 2011.

Nectria eustoma Penz. & Sacc., Malpighia 11: 509. 1898 [1897].

Nectria leucocoma Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 25: 28. 1899.

Nectria cinereopapillata Henn. & E. Nyman, in Warburg, Mon- sunia 1: 161. 1900 [1899].

Nectria striatospora Zimm., Centralbl. Bakteriol. II, 7: 105. 1901.

Nectria theobromae Masee, Bull. Misc. Inform. Kew 1908: 218. 1908.

Cylindrocarpon victoriae Wollenw., Z. Parasitenk. (Berlin) 1: 161. 1928.

Nectria azureo-ostiolata Doi, Mem. Nat. Sci. Mus. Tokyo 10: 23. 1977.

Holotypus: Not located.
Type locality: **Brazil**.

Type substrate: *Caconema radicolata*.
Note: *Synonyms* fide [Wollenweber & Reinking \(1935\)](#) and [Salgado-Salazar et al. \(2016\)](#).

maydiperdum Fusarium Bubák, Centralbl. Bakteriol. 2. Abth. 31: 497. 1911.

(See ***Fusarium poae***)

Holotypus: BPI 452399.

Type locality: **Czech Republic**, Tabor.

Type substrate: Seeds of *Zea mays*.

Note: *Synonym* fide [Wollenweber & Reinking \(1935\)](#).

maydis Fusarium Kalchbr., Math. Term. Közlem. 3: 285. 1865.
(See ***Fusarium sambucinum***)

Holotypus: BRACR33140.

Type locality: **Hungary**.

Type substrate: *Zea mays*.

Note: *Synonym* fide [Wollenweber & Reinking \(1935\)](#).

melanochlorum Fusarium (Casp.) Sacc., Syll. Fung. 4: 725. 1886.

Basionym: *Fusisporium melanochlorum* Casp., Ber. Bekanntm. Verh. Königl. Preuss. Akad. Wiss. Berlin 1855: 309, 314. 1855.

Cosmospora flavoviridis (Fuckel) Rossman & Samuels, Stud. Mycol. 42: 121. 1999.

Basionym: *Sphaerostilbe flavoviridis* Fuckel, Jahrb. Nassauischen Vereins Naturk. 25–26: 310. 1871.

Synonyms: *Nectria flavoviridis* (Fuckel) Wollenw. *Angew. Bot.* 8: 186. 1926.

Fusarium celtidis Pass., *Atti Reale Accad. Lincei, Rendiconti Cl. Sci. Fis.*, 4 sér. 7: 51. 1891, *nom. illegit.*, Art. 53.1.

Fusarium sphaeriiforme Sacc. (as '*sphaeriaeforme*'), *Syll. Fung.* 10: 723. 1892.

Fusarium glandicola Allesch., *Ber. Bot. Vereines Landshut* 12: 130. 1892, *nom. illegit.*, Art. 53.1, *non* Cooke & W.R. Gerard, 1878.

Fusarium allescheri Sacc. & P. Syd., *Syll. Fung.* 14: 1128. 1899. *Holotypus:* Not located.

Type locality: **Germany**, Berlin.

Type substrate: Rotten aquatic plants.

meliolicola *Fusarium* F. Stevens (as '*meliolicolum*'), *Bot. Gaz.* 65: 245. 1918.

(See *Fusarium larvarum*)

Holotypus: ILL00011251.

Type locality: **Puerto Rico**, Mayagüez.

Type substrate: Parasitic on *Meliola paullinae* on *Casearia sylvestris*.

Note: *Synonym fide* Wollenweber & Reinking (1935).

meridionale Fusarium T. Aoki *et al.*, *Fungal Genet. Biol.* 41: 618. 2004.

Holotypus: BPI 843474.

Ex-type culture: CBS 110247 = FRC R-5329 = NRRL 28436.

Type locality: **New Caledonia**.

Type substrate: *Citrus sinensis*.

Descriptions and illustrations: See O'Donnell *et al.* (2004).

Diagnostic DNA barcodes: *rpb1:* KM361642; *rpb2:* KM361660; *tef1:* AF212435.

merismoides *Fusarium* Corda, *Icon. Fung.* 2: 4. 1838.

Fusicolla merismoides (Corda) Gräfenhan *et al.*, *Stud. Mycol.* 68: 101. 2011.

Synonyms: *Fusisporium georginae* Klotzsch, *Herb. Viv. Mycol.*, *Cent.* 2: 186. 1832, *nom. nud.*, Art. 38.1 (a).

Fusarium rhizophilum Corda, *Icon. Fung.* 2: 3. 1838.

Pionnotes rhizophila (Corda) Sacc., *Syll. Fung.* 4: 727. 1886.

?*Fusisporium arachnoideum* Corda, *Icon. Fung.* 1: 11. 1837.

?*Fusarium arachnoideum* (Corda) Sacc., *Syll. Fung.* 4: 721. 1886.

?*Fusarium biasolettianum* Corda, *Icon. Fung.* 2: 3. 1838.

?*Fusisporium biasolettianum* (Corda) Sacc., *Mycoth. Ven.* no. 1040. 1877.

?*Pionnotes biasolettiana* (Corda) Sacc., *Syll. Fung.* 4: 725. 1886.

Fusisporium udum Berk., *Ann. Mag. Nat. Hist.* 6: 438. 1841.

Pionnotes uda (Berk.) Sacc., *Syll. Fung.* 4: 726. 1886.

Fusarium udum (Berk.) Wollenw., *Phytopathology* 3: 38. 1913, *nom. illegit.*, Art. 53.1.

Fusidium udum Berk., in Trotter, *Syll. Fung.* 25: 979. 1931, *nom. inval.*, Art. 36.1.

Fusisporium foeni Berk. & Broome, *Ann. Mag. Nat. Hist.*, ser. 2, 7: 179. 1851.

Fusarium foeni (Berk. & Broome) Sacc., *Syll. Fung.* 4: 699. 1886.

Fusisporium roseolum H.O. Stephens ex Berk. & Broome, *Ann. Mag. Nat. Hist.*, ser. 2, 7: 178. 1851.

Fusarium roseolum (H.O. Stephens ex Berk. & Broome) Sacc., *Syll. Fung.* 4: 710. 1886.

Fusisporium rimosum Peck, *Rep. (Annual) New York State Mus. Nat. Hist.* 30: 58. 1878.

Fusarium rimosum (Peck) Sacc., *Syll. Fung.* 4: 713. 1886.

Fusarium roesleri Thüm., *Pilze Weinst.*: 51. 1878.

Fusarium arvense Speg., *Anales Soc. Ci. Argent.* 10: 60. 1880.

Fusarium gallinaceum Cooke & Harkn., *Grevillea* 9: 8. 1880.

Fusarium nicotianae Oudem., *Ned. Kruidk. Arch.*, sér. 3, 2: 777. 1902.

Fusarium udum var. *pusillum* Wollenw., *Phytopathology* 1: 206. 1913, *nom. nud.*

Fusarium udum var. *solani* Sherb., *Mem. Cornell Univ. Agric. Exp. Sta.* 6: 131. 1915.

Fusarium merismoides f. *nicotianae* (Oudem.) Subram., *Hyphomycetes*: 676. 1971.

Fusarium oxysporum f. sp. *nicotianae* (Oudem.) Subram., *Hyphomycetes*: 676. 1971.

Fusarium pelargonii P. Crouan & H. Crouan, *Fl. Finistère*: 14. 1867.

Fusarium albiziae Woron., *Vestn. Tiflissk. Bot Sada* 48: 34. 1920.

Fusarium merismoides var. *majus* Wollenw., *Fusaria Autogr. Delin.* 3: 857a. 1930.

Fusarium merismoides var. *chlamydosporale* Wollenw., *Z. Parasitenk. (Berlin)* 3: 308. 1931.

Fusarium merismoides var. *artocarp* X.H. Fu & Q.T. Chen, *Acta Mycol. Sin.* 8: 42. 1989.

Fusarium merismoides var. *persicicola* X.H. Fu & Q.T. Chen, *Acta Mycol. Sin.* 8: 44. 1989.

Typus: PRM 155493.

Type locality: **Czech Republic**, Prague.

Type substrate: Wet shards of a plant pot.

Note: Lectotypification pending study of material lodged in PRM.

mesentericum *Fusarium* Cooke & Harkn., *Grevillea* 9: 128. 1881. *Holotypus:* ?K(M).

Type locality: **USA**, California, San Francisco Masonic Cemetery.

Type substrate: *Eucalyptus* sp.

Notes: Status unclear. Not *Fusarium fide* Wollenweber & Reinking (1935).

mesoamericanum Fusarium T. Aoki *et al.*, *Fungal Genet. Biol.* 41: 619. 2004.

Holotypus: BPI 843476.

Ex-type culture: CBS 415.86 = FRC R-8506 = IMI 309346 = NRRL 25797.

Type locality: **Honduras**.

Type substrate: *Musa* sp.

Descriptions and illustrations: See O'Donnell *et al.* (2004).

Diagnostic DNA barcodes: *rpb1:* KM361639; *rpb2:* KM361657; *tef1:* AF212441.

metachroum *Fusarium* Appel & Wollenw., *Arbeiten Kaiserl. Biol. Anst. Land- Forstw.* 8: 141. 1910 [1913].

(See ***Fusarium avenaceum***)

Holotypus: BPI 452408.

Type locality: **Poland**, Poznań, Slivno Manor.

Type substrate: *Triticum aestivum*.

Note: *Synonym fide* Wollenweber & Reinking (1935).

metavorans *Fusarium* Al-Hatmi *et al.*, *Medical Mycol.* 56: S147. 2018.

Neocosmospora metavorans (Al-Hatmi *et al.*) Sand.-Den. & Crous, *Persoonia* 41: 121. 2018.

Holotypus: CBS 135789 (preserved as metabolically inactive culture).

Ex-type culture: CBS 135789.

Type locality: **Greece**, Athens.

Type substrate: Pleural effusion of *Homo sapiens*.

Descriptions and illustrations: See [Al-Hatmi et al. \(2018\)](#) and [Sandoval-Denis & Crous \(2018\)](#).

Diagnostic DNA barcodes: *rpb1*: MW218127; *rpb2*: LR583849; *tef1*: LR583627.

mexicanum Fusarium T. Aoki et al., *Phytopathology* 100: 1180. 2010.

Holotypus: BPI 879150.

Ex-type culture: NRRL 53147.

Type locality: **Mexico**, Nueva Italia, Michoacán.

Type substrate: *Mangifera indica*.

Descriptions and illustrations: See [Otero-Colina et al. \(2010\)](#).

Diagnostic DNA barcodes: *rpb1*: MG838088; *rpb2*: MN724973; *tef1*: MG838032.

microcera Fusarium Bilař, *Fusarii (Biologija i sistematika)*: 292. 1955, *nom. inval.*, Art. 39.1.

(See *Fusarium coccidicola*)

Note: This species was invalidly published without Latin diagnosis.

microconidium Fusarium L. Lombard & Crous, *Fungal Syst. Evol.* 4: 192. 2019.

Holotypus: CBS H-24017.

Ex-type culture: CBS 119843 = MRC 8391 = KSU 11396.

Type locality: **Unknown**.

Type substrate: Unknown.

Descriptions and illustrations: See [Lombard et al. \(2019a\)](#).

Diagnostic DNA barcodes: *rpb1*: MN120721; *tef1*: MN120759.

microphlyctis Fusarium Mont., *Ann. Sci. Nat., Bot., sér.* 3, 12: 297. 1849.

Holotypus: ?PC.

Type locality: **France**.

Type substrate: Fruit of *Olea* sp.

Note: *Gloeosporium* fide [Wollenweber & Reinking \(1935\)](#).

micropus Fusarium Sacc., *Philipp. J. Sci.* 18: 605. 1921.

Infracungus micropus (Sacc.) Cif., *Mycopathol. Mycol. Appl.* 6: 26. 1951.

Holotypus: In PAD.

Type locality: **China**, Guangdong Province.

Type substrate: Parasitic on *Cladosporium herbarum* on leaf of *Morus alba*.

microsperrum Fusarium Berk. & M.A. Curtis, *Grevillea* 3: 98. 1875.

Holotypus: ?K(M).

Type locality: **USA**, South Carolina, Santee River.

Type substrate: *Ficus* sp.

Note: *Hymenula* fide [Wollenweber & Reinking \(1935\)](#).

microsporum Fusarium Schldl., *Fl. Berol.* 2: 139. 1824.

(See ***Fusarium lateritium***)

Holotypus: HAL 1615 F.

Type locality: **Germany**, Berlin.

Type substrate: *Robinia pseudoacaciae*.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

mikaniae Fusarium Berk. & M.A. Curtis, *Grevillea* 3: 98. 1875.

Holotypus: ?K(M).

Type locality: **USA**, South Carolina, Santee River.

Type substrate: Stems and leaves of *Mikania scandens*.

Notes: Status unclear. Not *Fusarium* fide [Wollenweber & Reinking \(1935\)](#).

mindoanum Fusarium Petr., *Sydowia* 4: 576. 1950.

Holotypus: In W as no. 03550 (PetraK, Pilzherbarium no. 32229).

Type locality: **Ecuador**, Pichincha, Mindo.

Type substrate: *Dryopteris diplazioides*.

Notes: No living material available to confirm taxonomic status. Requires recollection from type locality and substrate.

miniaturum Fusarium Sacc., *Syll. Fung.* 10: 727. 1892.

Replaced synonym: *Fusarium miniatum* Prill. & Delacr., *Bull. Soc. Mycol. France* 7: 117. 1891, *nom. illegit.*, Art. 53.1

(See *Fusarium nivale*)

Holotypus: Not located.

Type locality: **France**, Paris.

Type substrate: *Secale cereale*.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

miniaturum Fusarium Sacc., *Michelia* 1: 83. 1877.

Synonym: *Fusarium detonianum* Sacc., *Syll. Fung.* 4: 708. 1886, *nom. illegit.*, Art. 52.1.

Holotypus: In PAD.

Type locality: **Italy**.

Type substrate: Sporangium of *Cyathus vernicosa*.

Note: Status unclear. Requires further investigation.

miniaturum Fusarium (Berk. & M.A. Curtis) Sacc., *Syll. Fung.* 4: 722. 1886, *nom. illegit.*, Art. 53.1.

Basionym: *Fusisporium miniaturum* Berk. & M.A. Curtis, *Grevillea* 3: 147. 1875.

(See ***Fusarium lateritium***)

Holotypus: ?K(M).

Type locality: **USA**, North Carolina.

Type substrate: *Cornus florida*.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

miniaturum Fusarium Prill. & Delacr., *Bull. Soc. Mycol. France* 7: 117. 1891, *nom. illegit.*, Art. 53.1.

Replacing synonym: *Fusarium miniaturum* Sacc., *Syll. Fung.* 10: 727. 1892

(See *Fusarium nivale*)

Authentic material: Not located.

Original locality: **France**, Paris.

Original substrate: *Secale cereale*.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

minimum Fusarium Fuckel, *Fungi Rhen. Exs.*, Fasc. 3, no. 213. 1863.

(See *Fusarium nivale*)

Syntypes: In BPI, MICH, MU & S (*Fungi Rhen. Exs.*, Fasc. 3, no. 213).

Type locality: **Germany**, Oestrich, Nassau region.

Type substrate: Dry leaves of *Poaceae* (mainly *Zea mays*)

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

minutissimum Fusarium (Desm.) Sacc., *Syll. Fung.* 4: 703. 1886.

Passalora minutissima (Desm.) U. Braun & Crous, *CBS Biodiversity Ser.* 1: 276. 2003.

Basionym: *Selenosporium minutissimum* Desm., *Pl. Crypt. France*, ed. 3, Fasc. 10: no. 456. 1857.

Phaeoramularia minutissima (Desm.) U. Braun, *Nova Hedwigia* 55: 214. 1992.

Ramularia geranii-sanguinei C. Massal., *Atti Ist. Veneto Sci. Lett. Arti* 59: 688. 1900.

Cercospora geranii-sanguinei Henn., *Nytt Mag. Naturvidensk.* 42: 33. 1904.

Lectotypus: Desm., Pl. Crypt. France, Fasc. X, no. 456 in PC fide Braun (1998).

Lectotype locality: France, Louvigny, Caen.

Lectotype substrate: *Geranium molle*.

minutulum *Fusarium* Corda, Icon. Fung. 2: 4. 1838.

?*Clonostachys solani* (Harting) Schroers & W. Gams, Stud. Mycol. 46: 111. 2001.

Basionym: *Spicaria solani* Harting, Nieuwe Verh. Eerste Kl. Kon. Ned. Inst. Wetensch. Amsterdam, ser. 2, 12: 226. 1846.

Synonyms: ?*Gliocladium solani* (Harting) Petch, Trans. Brit. Mycol. Soc. 27: 149. 1945.

?*Hypomyces solani* Reinke & Berthold, Untersuch. Bot. Lab. Univ. Göttingen 1: 27. 1879.

?*Hypolyssus solani* (Reinke & Berthold) Kuntze, Revis. Gen. Pl. 3: 488. 1898.

?*Hyphonectria solani* (Reinke & Berthold) Petch, Bot. J. (London) 74: 220. 1937 [1936].

?*Nectriopsis solani* (Reinke & Berthold) C. Booth, Mycol. Pap. 74: 8. 1960.

?*Bionectria solani* (Reinke & Berthold) Schroers, Stud. Mycol. 46: 111. 2001.

?*Gliocladium nigrovirens* J.F.H. Beyma, Verh. Kon. Akad. Wetensch., Afd. Natuurk., Sect. 2, 29: 30. 1931.

?*Clonostachys solani* f. *nigrovirens* (J.F.H. Beyma) Schroers, Stud. Mycol. 46: 115. 2001.

Typus: In PRM fide Pilat (1938).

Type locality: Czech Republic, Prague.

Type substrate: Wood splinters of *Corylus* sp.

Notes: Synonym fide Wollenweber & Reinking (1935). Lectotypification pending study of material lodged in PRM.

miscanthi *Fusarium* W. Gams et al., Mycologia 91: 264. 1999. *Holotypus*: CBS H-6063.

Ex-type culture: CBS 577.97 = NRRL 26231.

Type locality: Denmark, Zealand, Højbakkegård Experimental field.

Type substrate: *Miscanthus sinensis*.

Descriptions and illustrations: See Gams et al. (1999).

Diagnostic DNA barcodes: *rpb1*: JX171521; *rpb2*: JX171634; *tef1*: MN193878.

mollerianum *Fusarium* Thüm., Inst. Coimbra 28: 263. 1881.

(See *Fusarium graminearum*)

Holotypus: ?S-F45644.

Type locality: Portugal, Coimbra.

Type substrate: *Melia azedarach*.

Note: Synonym fide Wollenweber & Reinking (1935).

moniliforme *Fusarium* J. Sheld., Annual Rep. Nebraska Agric. Exp. Sta. 17: 23. 1904.

(See *Fusarium verticillioides*)

Syntypes: BPI 452450 & BPI 452452.

Type locality: USA, Nebraska.

Type substrate: *Zea mays*.

Note: Typification pending further study of the syntypes.

monophialidicum *Fusarium* J.W. Xia et al., Persoonia 43: 211. 2019.

Holotypus: CBS H-24063.

Ex-type culture: NRRL 54973 = UTHSC 06-1473.

Type locality: USA, Ohio.

Type substrate: Eye of *Rhinocerotidae* (rhinoceros).

Descriptions and illustrations: See Xia et al. (2019).

Diagnostic DNA barcodes: *rpb1*: KC808299; *rpb2*: KC808362; *tef1*: MN170483.

mori *Fusarium* (Sand.-Den. & Crous) O'Donnell et al., Index Fungorum 440: 2. 2020.

Neocosmospora mori Sand.-Den. & Crous, Persoonia 43: 143. 2019.

Holotypus: CBS H-23987.

Ex-type culture: ATCC 44934 = CBS 145467 = MAFF 238539 = NRRL 22230.

Type locality: Japan, Miyazaki.

Type substrate: Twigs of *Morus alba*.

Descriptions and illustrations: See Sandoval-Denis et al. (2019).

Diagnostic DNA barcodes: *rpb1*: MW834235; *rpb2*: EU329499; *tef1*: AF178358.

moronei *Fusarium* Curzi, Revista Biol. (Lisbon) 10: 141. 1928.

(See *Fusarium acuminatum*)

Holotypus: ?PAV.

Type locality: Italy.

Type substrate: Vesicle on skin of *Canis lupus familiaris* (dog).

Note: Synonym fide Wollenweber & Reinking (1935).

moschatum *Fusarium* (Kitasato) Sacc., Syll. Fung. 10: 729. 1892.

Basionym: *Fusisporium moschatum* Kitasato, Centralbl. Bakteriologie. Parasitenk., 1. Abth. 5: 365. 1889.

(See *Fusarium aquaeductuum*)

Holotypus: Not located.

Type locality: Germany.

Type substrate: Metallic medical equipment.

Note: Synonym fide Wollenweber & Reinking (1935).

mucidum *Fusarium* J.W. Xia et al., Persoonia 43: 211. 2019.

Holotypus: CBS H-24064.

Ex-type culture: CBS 102395.

Type locality: El Salvador, Cooperación Coralama.

Type substrate: Mouldy nut of *Anacardium occidentale*.

Descriptions and illustrations: See Xia et al. (2019).

Diagnostic DNA barcodes: *rpb2*: MN170418; *tef1*: MN170485.

muentzii *Fusarium* Delacr. (as 'müntzi'), Bull. Soc. Mycol. France 8: 192. 1892.

(See *Fusarium tricinctum*)

Lectotypus (*hic designatus*, MBT 10000715): France, Paris, on animal waste, May 1891, G. Delacroix, Bull. Soc. Mycol. France 8, pl. XVII, fig. V.

multiceps *Fusarium* J.W. Xia et al., Persoonia 43: 212. 2019.

Holotypus: CBS H-24065.

Ex-type culture: CBS 130386 = NRRL 43639 = UTHSC 04-135.

Type locality: USA, Florida.

Type substrate: *Trichechus* sp.

Descriptions and illustrations: See Xia et al. (2019).

Diagnostic DNA barcodes: *rpb1*: HM347190; *rpb2*: GQ505844; *tef1*: GQ505666.

mundagurra *Fusarium* M.H. Laurence et al., Fungal Diversity 77: 359. 2015 [2016].

Holotypus: RBG5717.

Ex-type culture: NRRL 66235 = RBG5717.

Type locality: Australia, Queensland, Carnarvon Gorge National Park.

Type substrate: Soil.

Descriptions and illustrations: See Laurence et al. (2016).

Diagnostic DNA barcodes: *rpb1*: KP083272; *rpb2*: KP083276; *tef1*: KP083256.

musae Fusarium Van Hove *et al.*, Mycologia 103: 579. 2011.
Synonym: *Gibberella musae* Van Hove *et al.*, Mycologia 103: 577. 2011.

Holotypus: MUCL 52574.

Ex-type culture: CBS 624.87 = MUCL 52574 = NRRL 25059.

Type locality: **Honduras**.

Type substrate: *Musa* sp.

Descriptions and illustrations: See [Van Hove *et al.* \(2011\)](#).

Diagnostic DNA barcodes: *rpb1*: MW402689; *rpb2*: FN552108; *tef1*: FN552086.

musarum Fusarium Logrieco & Marasas, Mycologia 90: 510. 1998.

Holotypus: BPI 802928.

Ex-type culture: FRC R-9400 = MRC 6240 = NRRL 28507.

Type locality: **Panama**.

Type substrate: *Musa sapientum*.

Descriptions and illustrations: See [Marasas *et al.* \(1998\)](#).

Diagnostic DNA barcodes: *rpb1*: MW233265; *rpb2*: MW928829; *tef1*: MW233094.

mycophilum Fusarium (P. Karst.) Sacc., Syll. Fung. 10: 730. 1892.

Basionym: *Leptosporium mycophilum* P. Karst., Meddel. Soc. Fauna Fl. Fenn. 16: 24. 1888.

Holotypus: ?H.

Type locality: **Finland**, Merimasku.

Type substrate: *Myxogastris*.

Note: *Hymenula* fide [Wollenweber & Reinking \(1935\)](#).

mucophytum Fusarium (W.G. Sm.) Masee, Brit. Fung.-Fl. 3: 483. 1893.

Basionym: *Fusisporium mucophytum* W.G. Sm., Gard. Chron. n.s., 22: 245. 1884.

(See ***Fusarium scirpi***)

Holotypus: ?K(M).

Type locality: **UK**.

Type substrate: *Agaricus arvensis*.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

myosotidis Fusarium Cooke, Grevillea 16: 49. 1887.

(See ***Fusarium oxysporum***)

Holotypus: In K(M).

Type locality: **UK**, Forden.

Type substrate: *Myosotis* sp.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

nanum Fusarium M.M. Wang *et al.*, Persoonia 43: 85. 2019.

Holotypus: HAMS 248043.

Ex-type culture: CGMCC 3.19498 = LC12168.

Type locality: **China**, Guangxi Province, Guilin.

Type substrate: Leaves of *Musa nana*.

Descriptions and illustrations: See [Wang *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MK289871; *rpb2*: MK289755; *tef1*: MK289602.

napiforme Fusarium Marasas *et al.*, Mycologia 79: 910. 1988 [1987].

Holotypus: DAOM 196924.

Ex-type culture: BBA 69861 = CBS 748.97 = DAOM 196924 = DAOM 225147 = FRC M-3563 = IMI 375353 = MRC 4144 = NRRL 13604.

Type locality: **Namibia**, Ovambo.

Type substrate: *Pennisetum typhoides*.

Descriptions and illustrations: See [Marasas *et al.* \(1987\)](#).

Diagnostic DNA barcodes: *rpb1*: HM347136; *rpb2*: EF470117; *tef1*: AF160266.

nectriae-palmicolae Fusarium Henn., Bot. Jahrb. Syst. 23: 290. 1896.

(See ***Fusarium equiseti***)

Holotypus: In B fide [Hein \(1988\)](#).

Type locality: **Samoa**, Upolu.

Type substrate: Leaves of *Areca* sp.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

nectriae-turraeae Fusarium Henn., Bot. Jahrb. Syst. 22: 82. 1895.

(See *Fusarium coccophilum*)

Holotypus: In B fide [Hein \(1988\)](#).

Type locality: **Tanzania**, Marangu.

Type substrate: *Turraea volkensii*.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

nectricreans Fusarium Kirschst., Ann. Mycol. 37: 138. 1939.

Holotypus: B 70 0100202.

Type locality: **Germany**, Berlin.

Type substrate: Rotting stem of garden plant.

Note: No living material was available for confirmation of taxonomic status.

nectrioides Fusarium (Wollenw.) Schroers *et al.*, Mycologia 101: 59. 2009.

Bisifusarium nectrioides (Wollenw.) L. Lombard & Crous, Stud. Mycol. 80: 225. 2015.

Basionym: *Fusarium dimerum* var. *nectrioides* Wollenw., Fusaria Autogr. Delin. 3: 855. 1930.

Lectotypus: No. 855 in Wollenweber, Fusaria Autogr. Delin. (1930), designated in [Schroers *et al.* \(2009\)](#).

Ex-type culture: CBS 176.31 = NRRL 20689.

Lectotype and ex-type locality: **Honduras**.

Lectotype and ex-type substrate: Soil.

Descriptions and illustrations: See [Schroers *et al.* \(2009\)](#).

Diagnostic DNA barcodes: *rpb1*: JX171477; *rpb2*: JX171591; *tef1*: EU926312.

neglectum Fusarium Jacz., Bull. Trimestriel Soc. Mycol. France 28: 348. 1912.

(See *Fusarium culmorum*)

Holotypus: Not located.

Type locality: **Ukraine**, Poltava.

Type substrate: *Zea mays*.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

negundinis Fusarium Sherb., in Hubert, J. Agric. Res. 26: 451. 1923.

(See ***Fusarium reticulatum***)

Holotypus: Not located.

Type locality: **USA**, Wisconsin, Madison.

Type substrate: *Acer negundo*.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

nelsonii Fusarium Marasas & Logrieco, Mycologia 90: 508. 1998.

Holotypus: BPI 802927.

Ex-type culture: CBS 119876 = FRC R-8670 = MRC 4570 = NRRL 28505 = NRRL 53945.

Type locality: **South Africa**, Western Cape Province, Malmesbury.

Type substrate: Plant debris in *Triticum* soil.

Descriptions and illustrations: See [Marasas et al. \(1998\)](#).

Diagnostic DNA barcodes: *rpb1*: MN120722; *rpb2*: GQ505468; *tef1*: GQ505404.

nematophilum *Fusarium* Nirenberg & Hagedorn, Nachrichtenbl. Deutsch. Pflanzenschutzdienstes 60: 214. 2008.

Luteonectria nematophila (Nirenberg & Hagedorn) Sand.-Den. & L. Lombard, Stud. Mycol. 98 (no. 100116): 60. 2021.

Holotypus: BBA 72279 in B.

Ex-type culture: BBA 72279 = NRRL 54600.

Type locality: **Germany**, Berlin.

Type substrate: Isolated from soil with roots of *Hedera helix*.

Descriptions and illustrations: See [Nirenberg & Hagedorn \(2008\)](#).

Diagnostic DNA barcodes: *rpb1*: JX171552; *rpb2*: JX171664; *tef1*: JABFFA010003988.

neoceras *Fusarium* Wollenw. & Reinking, Phytopathology 15: 164. 1925.

(See ***Fusarium sacchari***)

Holotypus: CBS 147.25 (preserved as metabolically inactive culture).

Ex-type culture: BBA 69863 = CBS 147.25 = DAOM 225410 = IMI 375345 = NRRL 20471.

Type locality: **Honduras**.

Type substrate: Rotting *Musa sapientum*.

Descriptions and illustrations: See [Gerlach & Nirenberg \(1982\)](#).

Diagnostic DNA barcodes: *rpb1*: MT010941; *rpb2*: MT010962; *tef1*: MT010988.

neocosmosporiellum *Fusarium* O'Donnell & Geiser, Phytopathology 103: 405. 2013.

Neocosmospora vasinfecta E.F. Sm., Bull. Div. Veg. Physiol. Pathol. U.S.D.A. 17: 45. 1899.

Synonyms: *Nectriella tracheiphila* E.F. Sm., Proc. Amer. Assoc. Advancem. Sci. 44: 190. 1896, *nom. inval. fide* Cannon & Hawksworth 1984.

Neocosmospora vasinfecta var. *nivea* E.F. Sm., Bull. Div. Veg. Physiol. Pathol. U.S.D.A. 17: 45. 1899.

Neocosmospora vasinfecta var. *tracheiphila* E.F. Sm., Bull. Div. Veg. Physiol. Pathol. U.S.D.A. 17: 45. 1899.

Fusarium tracheiphilum (E.F. Sm.) Wollenw., Phytopathology 3: 29. 1913.

Fusarium vasinfectum var. *pisi* C.J.J. Hall, Ber. Deutsch. Bot. Ges. 21: 4. 1903.

Neocosmospora vasinfecta var. *pisi* (C.J.J. Hall) Sacc., Syll. Fung. 20: 192. 1911.

Neocosmospora africana Arx, Antonie van Leeuwenhoek 21: 161. 1955.

Neocosmospora vasinfecta var. *africana* (Arx) P.F. Cannon & D. Hawksw., Trans. Brit. Mycol. Soc. 82: 676. 1984.

?*Pseudonectria ornata* Bat. & Maia, Anais Soc. Biol. Pernambuco 13: 74. 1955 (*fide* Cannon & Hawksworth 1984).

Neocosmospora vasinfecta var. *major* P. Rama Rao, Mycopathol. Mycol. Appl. 21: 218. 1963.

Neocosmospora ornamentata M.A.F. Barbosa, Garcia de Orta 13: 17. 1965.

Fusarium ornamentatum (M.A.F. Barbosa) O'Donnell et al., Index Fungorum 440: 3. 2020.

Neocosmospora vasinfecta f. *conidiifera* Kamyschko, Novosti Sist. Nizsh. Rast. 1965: 115. 1965.

Neocosmospora boninensis Udagawa et al., Sydowia 41: 350. 1989.

Lectotypus: Pl. V, figs 1–2 (Smith, Bull. Div. Veg. Physiol. Pathol. U.S.D.A. 17, 1899), designated in [Sandoval-Denis et al. \(2019\)](#).

Lectotype locality: **USA**.

Lectotype substrate: *Gossypium* sp.

Epitypus: BPI 910920, designated in [Aoki et al. \(2020\)](#).

Ex-epitype culture: ATCC 62199 = NRRL 22166.

Epitype locality: **USA**, Illinois, southern area.

Epitype substrate: A cyst of *Heterodera glycines* in a soil sample from soybean field.

Diagnostic DNA barcodes: *rpb1*: SSHR01002742; *rpb2*: EU329497; *tef1*: AF178350.

neoscirpi* *Fusarium L. Lombard et al., Persoonia 43: 213. 2019.

Holotypus: CBS H-24066.

Ex-type culture: CBS 610.95 = NRRL 26861 = NRRL 26922.

Type locality: **France**.

Type substrate: Soil.

Descriptions and illustrations: See [Xia et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb2*: GQ505779; *tef1*: GQ505601.

neosemitectum* *Fusarium L. Lombard et al., Persoonia 43: 214. 2019.

Holotypus: CBS H-24067.

Ex-type culture: CBS 189.60.

Type locality: **Democratic Republic of the Congo**.

Type substrate: *Musa sapientum*.

Descriptions and illustrations: See [Xia et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb2*: MN170422; *tef1*: MN170489.

nepalense* *Fusarium T. Aoki et al., Fungal Genet. Biol. 48: 1105. 2011.

Holotypus: BPI 881006.

Ex-type culture: CBS 127503 = NRRL 54222.

Type locality: **Nepal**.

Type substrate: *Oryza sativa*.

Descriptions and illustrations: See [Sarver et al. \(2011\)](#).

Diagnostic DNA barcodes: *rpb1*: KM361650; *rpb2*: KM361668; *tef1*: KM889631.

nervisequum *Fusarium* (Fuckel) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 369. 1870.

Basionym: *Labrella nervisequa* Fuckel, Fungi Rhen. Exs., Fasc. 5, no. 427. 1863.

Apiognomonina platani (Lév.) L. Lombard, **comb. nov.** MycoBank MB 837698.

Basionym: *Hymenula platani* Lév., Ann. Sci. Nat., Bot., sér. 3, 9: 128. 1848.

Synonyms: *Fusarium platani* (Lév.) Mont., Ann. Sci. Nat., Bot., sér. 3, 11: 55. 1849.

Fusarium nervisequum f. *platani* (Lév.) Fuckel, Jahrb. Nassauischen Vereins Naturk. 23–24: 369. 1870.

Gloeosporidium platani (Lév.) Höhn., Sitzungsber. Kaiserl. Akad. Wiss. Wien, Math.-Naturwiss. Cl., Abt. 1, 125: 95. 1916.

Myxosporina platani (Lév.) Höhn., Hedwigia 62: 48. 1920, *nom. inval.*, Art. 35.1.

Gloeosporium nervisequum (Fuckel) Sacc., Syll. Fung. 3: 711. 1884.

Discula nervisequa (Fuckel) M. Morelet, Bull. Soc. Sci. Nat. Archéol. Toulon & Var 203: 12. 1973.

Gloeosporium platani Oudem., Ned. Kruidk. Arch., sér. 2, 1: 258. 1873.

Laestadia veneta Sacc. & Speg., Michelia 1: 351. 1878.

Carlia veneta (Sacc. & Speg.) Kuntze, Revis. Gen. Pl. 2: 846. 1891.

- Apiospora veneta* (Sacc. & Speg.) Sacc. ex Kleb., Z. Pflanzenkrankh. 12: 258. 1902.
- Gnomonia veneta* (Sacc. & Speg.) Kleb., Jahrb. Wiss. Bot. 41: 533. 1905, *nom. illegit.*, Art. 53.1.
- Gnomonia platani* Kleb., Verhandl. Deutsch. Bot. Ges. 1: 28. 1914.
- Guignardia veneta* (Sacc. & Speg.) Traverso, Fl. Ital. Crypt. 1: 392. 1907.
- Apiosporopsis veneta* (Sacc. & Speg.) Traverso, Syll. Fung. 22: 78. 1913.
- Apiognomonia veneta* (Sacc. & Speg.) Höhn., Ann. Mycol. 16: 51. 1918.
- Laestadia veneta* var. *cylindrasca* Sacc. & Speg., Michelia 1: 369. 1878.
- Laestadia cylindrasca* (Sacc. & Speg.) Sacc., Syll. Fung. 1: 422. 1882.
- Carlia cylindrasca* (Sacc. & Speg.) Kuntze, Revis. Gen. Pl. 2: 846. 1891.
- Guignardia cylindrasca* (Sacc. & Speg.) Lindau (as '*cylindracea*'), in Engler & Prantl, Nat. Pflanzenfam., Teil. I, 1(1): 422. 1897.
- Diaporthe veneta* Sacc. & Speg., Michelia 1: 383. 1878.
- Discella platani* Peck, Rep. (Annual) New York State Mus. Nat. Hist. 29: 49. 1878, *nom. illegit.*, Art. 53.1.
- Discula platani* Sacc., Syll. Fung. 3: 674. 1884.
- Sporonema platani* Bäumler, Oesterr. Bot. Z. 40: 17. 1890.
- Placosphaeria platani* (Bäumler) Limber, Mycologia 47: 398. 1955.
- Myxosporium platanicola* Ellis & Everh. (as '*platanicum*'), Proc. Acad. Nat. Sci. Philadelphia 46: 372. 1894.
- Cryptosporiopsis platanicola* (Ellis & Everh.) G.F. Laundon, CBS List of Cultures (Baarn): (1). 1975.
- Gloeosporidina platani* Butin & Kehr, Eur. J. Forest Pathol. 28: 299. 1998.
- Lectotypus*: BPI (Fuckel, Fungi Rhen. 427) of *Labrella nervisequa* Fuckel, designated in [Sogonov et al. \(2007\)](#).
- Lectotype locality*: **Germany**, Reichartshausen.
- Lectotype substrate*: *Plantanus orientalis*.
- Epitypus*: BPI 871953, designated in [Sogonov et al. \(2007\)](#).
- Epitype locality*: **Switzerland**, Geneva.
- Epitype substrate*: *Plantanus orientalis*.
- Notes*: Based on priority and synonymies proposed by [Sogonov et al. \(2007\)](#), the name *Hymenula platani* Lév. (1848) takes precedence over *Laestadia veneta* Sacc. & Speg. (1878). Therefore, a new combination is proposed here applying the older name.
- newnesense Fusarium*** M.H. Laurence et al., Fungal Diversity 77: 360. 2015 [2016].
- Holotypus*: RBG 610.
- Ex-type culture*: NRRL 66241 = RBG 610.
- Type locality*: **Australia**, New South Wales, Newnes State Forest.
- Type substrate*: Soil.
- Descriptions and illustrations*: See [Laurence et al. \(2016\)](#).
- Diagnostic DNA barcodes*: *rpb1*: JABCJW010000176; *rpb2*: JABCJW010000963; *tef1*: KP083261.
- ngaiotongaense Fusarium* O'Donnell et al., Index Fungorum 440: 3. 2020.
- Neocosmospora longissima*** Sand.-Den. & Crous, Persoonia 43: 141 (2019).
- Holotypus*: CBS H-23985.
- Ex-type culture*: CBS 126407 = G.J.S. 85-72.
- Type locality*: **New Zealand**, Russell State Forest, Ngaiotonga Scenic Reserve.
- Type substrate*: From tree bark.
- Descriptions and illustrations*: See [Sandoval-Denis et al. \(2019\)](#).
- Diagnostic DNA barcodes*: *rpb1*: MW834230; *rpb2*: LR583846; *tef1*: LR583621.
- nicotianae Fusarium* Oudem., Ned. Kruidk. Arch., sér. 3, 2: 777. 1902.
- (See *Fusarium merismoides*)
- Holotypus*: ?L.
- Type locality*: **Netherlands**, Noord-Holland Province, Bussum.
- Type substrate*: *Nicotiana tabacum*.
- nigrum Fusarium* O.A. Pratt, J. Agric. Res. 13: 90. 1918.
- (See ***Fusarium flocciferum***)
- Lectotypus* (*hic designatus*, MBT 10000716): **USA**, Idaho, from soil, 1918, O.A. Pratt, in J. Agric. Res. 13: 82, fig. 1J–L.
- Notes*: Synonym *fide* [Wollenweber & Reinking \(1935\)](#). As the holotype specimen was not located, an illustration accompanying the original protologue is designated here as lectotype.
- nirenbergiae Fusarium*** L. Lombard & Crous, Persoonia 43: 29. 2018 [2019].
- Holotypus*: CBS H-23619.
- Ex-type culture*: CBS 840.88.
- Type locality*: **Netherlands**, Noord-Holland Province, Aalsmeer.
- Type substrate*: *Dianthus caryophyllus*.
- Descriptions and illustrations*: See [Lombard et al. \(2019b\)](#).
- Diagnostic DNA barcodes*: *rpb2*: MH484887; *tef1*: MH484978.
- nisikadoi Fusarium*** T. Aoki & Nirenberg, Mycoscience 38: 330. 1997.
- Holotypus*: BBA 69015 in B.
- Ex-type culture*: BBA 69015 = CBS 456.97 = MAFF 237506 = NRRL 25205 = NRRL 25308.
- Type locality*: **Japan**, Oita, Hita.
- Type substrate*: *Triticum aestivum*.
- Descriptions and illustrations*: See [Nirenberg & Aoki \(1997\)](#).
- Diagnostic DNA barcodes*: *rpb1*: MG282391; *rpb2*: MG282421; *tef1*: KR909358.
- nitidum Fusarium* Berk. & M.A. Curtis, Grevillea 3: 98. 1875.
- Holotypus*: ?K(M).
- Type locality*: **USA**, Pennsylvania, Michener.
- Type substrate*: *Aralia spinosa*.
- Note*: Doubtful species *fide* [Wollenweber & Reinking \(1935\)](#).
- nivale Fusarium* Ces. ex Berl. & Voglino, Syll. Fung., Addit. I–IV: 390. 1886.
- Microdochium nivale*** (Fr.) Samuels & I.C. Hallett, Trans. Brit. Mycol. Soc. 81: 479. 1983.
- Basionym*: *Lanosa nivalis* Fr., Summa Veg. Scand. 2: 495. 1849.
- Synonyms*: *Fusarium nivale* (Fr.) Sorauer, Z. Pflanzenkrankh. 11: 220. 1901, *nom. illegit.*, Art. 53.1.
- Fusarium hibernans* Lindau, Rabenh. Krypt.-Fl., ed. 2, 1(9): 542. 1909, *nom. superfl.*, Art. 52.1.
- Gerlachia nivalis* (Ces. ex Berl. & Voglino) W. Gams & E. Müll., Netherlands J. Pl. Pathol. 86: 49. 1980.
- Fusarium minimum* Fuckel, Fungi Rhen. Exs., Fasc. 3, no. 213. 1863.

Fusarium ustilaginis Rostr., Bot. Foren. Festschr. 54: 137. 1890, *nom. illegit.*, Art. 53.1.

Fusarium miniatum Prill. & Delacr., Bull. Soc. Mycol. France 7: 117. 1891, *nom. illegit.*, Art. 53.1.

Fusarium tritici Erikss., Fungi Paras. Scand. Exs. no. 400. 1891, *nom. illegit.*, Art. 53.1.

Fusarium miniatulum Sacc., Syll. Fung. 10: 727. 1892.

Nectria pseudograminicola Weese, Ann. Mycol. 8: 466. 1910, *nom. inval.*, Art. 38.1.

Fusarium loliaceum Ducomet, Ann. École Natl. Agric. Rennes 2: 14. 1909.

Fusarium secalis Jacz., Bull. Trimestriel Soc. Mycol. France 28: 346. 1912, *nom. illegit.*, Art. 53.1.

Sphaerulina divergens Rehm, Ann. Mycol. 11: 397. 1913.

Monographella divergens (Rehm) Petr., Ann. Mycol. 22: 144. 1924.

Calonectria nivalis Schaffnit, Mycol. Centralbl. 2: 257. 1913.

Griphosphaeria nivalis (Schaffnit) E. Müll. & Arx, Phytopathol. Z. 24: 356. 1955.

Micronectriella nivalis (Schaffnit) C. Booth, The Genus *Fusarium*: 42. 1971.

Monographella nivalis (Schaffnit) E. Müll., Rev. Mycol. (Paris) 41: 132. 1977.

Calonectria graminicola F. Stevens, Bot. Gaz. 65: 232. 1918, *nom. illegit.*, Art. 53.1.

Melioliphila graminicola Speg., Bol. Acad. Ci. (Córdoba) 26: 344. 1921.

Calonectria graminicola var. *neglecta* Krampe, Angew. Bot. 8: 252. 1926.

Monographella nivalis var. *neglecta* (Krampe) Gerlach, Netherlands J. Pl. Pathol. 86: 49. 1980.

Fusarium nivale var. *oryzae* Zambett., Mitt. Inst. Colombo-Aleman Invest. Ci. 30: 489. 1950, *nom. inval.*, Art. 39.1.

Syntypes: In HAL & ILL [Rabenhorst, Klotzschii Herb. Viv. Mycol. no. 1439 (sub *F. oxysporum*)].

Type locality: **Italy**.

Type substrate: *Poaceae*.

niveum *Fusarium* E.F. Sm., Proc. Amer. Assoc. Advancem. Sci. 43: 289. 1894, *nom. inval.*, Art. 36.1(a).

(See *Fusarium oxysporum*)

Authentic material: Not located.

Original locality: **USA**.

Original substrate: *Citrullus vulgaris*.

niveum *Fusarium* McAlpine, Australas. J. Pharm. 17: 3. 1902.

Note: Unable to locate protologue.

nodosum* *Fusarium L. Lombard & Crous, Fungal Syst. Evol. 4: 193. 2019.

Holotypus: CBS H-24018.

Ex-type culture: CBS 201.63.

Type locality: **Portugal**, Lisbon.

Type substrate: Seed of *Arachis hypogaea*.

Descriptions and illustrations: See Lombard et al. (2019a).

Diagnostic DNA barcodes: *rpb1*: MN120725; *rpb2*: MN120743; *tef1*: MN120763.

noneumartii *Fusarium* (Sand.-Den. & Crous) O'Donnell et al., Index Fungorum 440: 3. 2020.

Neocosmospora noneumartii Sand.-Den. & Crous, Persoonia 43: 145. 2019.

Holotypus: CBS H-23989.

Ex-type culture: CBS 115658 = FRC S-0661.

Type locality: **Israel**, Palestine.

Type substrate: *Solanum tuberosum*.

Descriptions and illustrations: See Sandoval-Denis et al. (2019).

Diagnostic DNA barcodes: *rpb1*: MW218129; *rpb2*: MW446618; *tef1*: LR583630.

nucicola *Fusarium* P. Karst. & Har., Rev. Mycol. (Toulouse) 12: 131. 1890.

(See *Fusarium lateritium*)

Holotypus: ?UPS fide Wollenweber, Fusaria Autogr. Delin. 1: 236. 1916.

Type locality: **France**.

Type substrate: Epicarp of nut.

Note: Synonym fide Wollenweber & Reinking (1935).

nurragi* *Fusarium (Summerell & L.W. Burgess) Benyon et al., Mycol. Res. 104: 1171. 2000.

Basionym: *Fusarium avenaceum* subsp. *nurragi* Summerell & L.W. Burgess, Mycol. Res. 99: 289. 1995.

Holotypus: DAR 69502.

Ex-type culture: CBS 393.96 = DAR 69501 = F10108 = F11121.

Type locality: **Australia**, Victoria, Wilson's Promontory National Park.

Type substrate: Soil.

Descriptions and illustrations: See Sangalang et al. (1995).

Diagnostic DNA barcodes: *rpb1*: MW928814; *rpb2*: MW928830; *tef1*: MW928840.

nygamai* *Fusarium L.W. Burgess & Trimboli, Mycologia 78: 223. 1986.

Synonym: *Gibberella nygamai* Klaasen & P.E. Nelson, Mycologia 88: 967. 1997.

Holotypus: FRC-M-1375.

Ex-type culture: ATCC 58555 = BBA 69862 = CBS 749.97 = FRC M-1375 = IMI 375354 = NRRL 13448.

Type locality: **Australia**, New South Wales, Narrabri.

Type substrate: Necrotic roots of *Sorghum* sp.

Descriptions and illustrations: See Burgess & Trimboli (1986).

Diagnostic DNA barcodes: *rpb1*: LT996202; *rpb2*: KU604262; *tef1*: MT011009.

obliqueseptatum *Fusarium*, T. Aoki et al., Mycologia 111: 929. 2019.

Neocosmospora obliqueseptata (T. Aoki et al.) L. Lombard & Sand.-Den., **comb. nov.** MycoBank MB 837699.

Basionym: *Fusarium obliqueseptatum*, T. Aoki et al., Mycologia 111: 929. 2019.

Holotypus: BPI 910970.

Ex-type culture: MAFF 246845 = NRRL 62611.

Type locality: **Australia**, Queensland, Beerwah.

Type substrate: A gallery wall of an ambrosia beetle (*Euwallacea* sp.) infecting *Persea americana*.

Descriptions and illustrations: See Aoki et al. (2019).

Diagnostic DNA barcodes: *rpb1*: KC691606; *rpb2*: KC691637, KC691666; *tef1*: KC691535.

Note: A new combination is provided in the genus *Neocosmospora* based on the phylogenetic relationship (Aoki et al. 2019) and morphology.

oblongum *Fusarium* (Sand.-Den. & Crous) O'Donnell et al., Index Fungorum 440: 3. 2020.

Neocosmospora oblonga Sand.-Den. & Crous, Persoonia 43: 148. 2019.

Holotypus: CBS H-23990.

Ex-type culture: CBS 130325 = CDC B-4701 = NRRL 28008.

Type locality: **USA**.

Type substrate: Eye of *Homo sapiens*.

Descriptions and illustrations: See [Sandoval-Denis et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW834239; *rpb2*: LR583853; *tef1*: LR583631.

obtusatum *Fusarium* Corda, Icon. Fung. 1: 3. 1837.

(See *Fusarium tortuosum*)

Typus: In PRM fide [Pilát \(1938\)](#).

Type locality: **Czech Republic**, Liberec (Reichenberg).

Type substrate: Branches of trees and shrubs.

Note: Not *Fusarium* fide [Wollenweber & Reinking \(1935\)](#). Lectotypification pending study of material lodged in PRM.

obtusisporum *Fusarium* Cooke & Harkn., Grevillea 12: 97. 1884.

Neonectria obtusispora (Cooke & Harkn.) Rossman et al., Phytopathol. Medit. 53: 529. 2014.

Synonyms: *Cylindrocarpon obtusisporum* (Cooke & Harkn.) Wollenw., Fusaria Autogr. Delin. 1: 465. 1916.

Ramularia obtusispora (Cooke & Harkn.) Wollenw., Fusaria Autogr. Delin. 1: 465. 1916.

Fusarium lineare Moesz, Bot. Közlem. 19: 57. 1920.

Holotypus: K(M) 128869.

Type locality: **USA**, California.

Type substrate: Twigs of *Acacia* sp.

obtusiusculum *Fusarium* Sacc., Michelia 2: 297. 1881.

(See *Fusarium candidum* Ehrenb.)

Holotypus: In PAD.

Type locality: **Italy**, Padua.

Type substrate: *Nelumbium* sp.

obtusum *Fusarium* (Cooke) Sacc., Syll. Fung. 4: 708. 1886.

Basionym: *Fusisporium obtusum* Cooke, Grevillea 5: 58. 1876.

Mycogloea macrospora (Berk. & Broome) McNabb, Trans. Brit. Mycol. Soc. 48: 187. 1965.

Basionym: *Dacrymyces macrosporus* Berk. & Broome, Ann. Mag. Nat. Hist., ser. 4, 11: 343. 1873.

Holotypus: In K(M) fide Index Fungorum.

Type locality: **UK**, Scotland, Forres.

Type substrate: *Diatrype* sp.

ochraceum *Fusarium* (Mont.) Sacc., Syll. Fung. 4: 722. 1886.

Basionym: *Fusisporium ochraceum* Mont., Ann. Sci. Nat., Bot., sér. 2, 3: 355. 1835.

Holotypus: In ?PC.

Type locality: **Chile**, Juan Fernández Islands.

Type substrate: Bark.

Note: Not *Fusarium* fide [Wollenweber & Reinking \(1935\)](#).

odoratissimum* *Fusarium Maryani et al., Stud. Mycol. 92: 159. 2019.

Synonym: *Fusarium purpurascens* Maryani et al., Stud. Mycol. 92: 160. 2018 [2019a].

Holotypus: InaCC F822 (preserved as metabolically inactive culture).

Ex-type culture: InaCC F822.

Type locality: **Indonesia**, East Kalimantan, Kampung Salak Martadinata.

Type substrate: *Musa* sp. cv. Pisang Kepok.

Descriptions and illustrations: See [Maryani et al. \(2019a\)](#).

Diagnostic DNA barcodes: *rpb1*: LS479618; *rpb2*: LS479386; *tef1*: LS479828.

Notes: Re-analysis of the sequence data set of [Maryani et al. \(2019a\)](#) revealed that the ex-type strain of *F. purpurascens*

(InaCC F971) clustered within the *F. odoratissimum* clade. Therefore, we consider *F. purpurascens* a synonym of *F. odoratissimum*.

oidioides *Fusarium* Speg., Rev. Mycol. (Toulouse) 8: 183. 1886. Holotypus: In LPS (Fungi Japon. No. 2) fide [Farr \(1973\)](#).

Type locality: **Japan**, Tokyo.

Type substrate: *Fallopia multiflora*.

Note: Not *Fusarium* fide [Wollenweber & Reinking \(1935\)](#).

oligoseptatum *Fusarium* T. Aoki et al., Fung. Syst. Evol. 1: 29. 2018.

Neocosmospora oligoseptata (T. Aoki et al.) Sand.-Den. & Crous, Persoonia 43: 149. 2019.

Holotypus: BPI 910525.

Ex-type culture: CBS 143241 = FRC S-2581 = MAFF 246283 = NRRL 62579.

Type locality: **USA**, Pennsylvania, Dauphin.

Type substrate: From a live female ambrosia beetle (*Euwallacea validus*), extracted from a gallery in a tree-of-heaven (*Ailanthus altissima*).

Descriptions and illustrations: See [Aoki et al. \(2018\)](#).

Diagnostic DNA barcodes: *rpb1*: KC691596; *rpb2*: KC691627, KC691656; *tef1*: KC691538.

ophioides* *Fusarium A. Jacobs, et al., Persoonia 46: 149. 2021.

Holotypus: CBS H-24659.

Ex-type culture: CBS 118512 = FCC 2979 = FCC 2980 = MRC 6744.

Type locality: **South Africa**, Mpumalanga Province, Ngodwana.

Type substrate: *Panicum maximum*.

Descriptions and illustrations: See [Yilmaz et al. \(2021\)](#).

Diagnostic DNA barcodes: *rpb2*: MN534303; *tef1*: EU921239.

opuli *Fusarium* Oudem., Hedwigia 37: 318. 1898.

Holotypus: ?L.

Type locality: **Netherlands**, Gelderland Province, Nunspeet.

Type substrate: *Viburnum opulus*.

Note: Not *Fusarium* fide [Wollenweber & Reinking \(1935\)](#).

opuntiarum *Fusarium* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 6: 350. 1898 [1899].

(See ***Fusarium oxysporum***)

Holotypus: In LPS (Fungi Argent. n.v.c. no. 866) fide [Farr \(1973\)](#).

Type locality: **Argentina**, La Plata.

Type substrate: Branches of *Opuntia* sp.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

orchidis *Fusarium* Petch, Ann. Roy. Bot. Gard. (Peradeniya) 6: 256. 1917.

(See ***Fusarium reticulatum***)

Holotypus: PDA 4798.

Type locality: **Sri Lanka**.

Type substrate: Leaves of *Orchidaceae*.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

ornamentatum *Fusarium* (M.A.F. Barbosa) O'Donnell et al., Index Fungorum 440: 3. 2020.

(See *Fusarium neocosmosporiellum*)

Holotypus: CBS 562.70 (preserved as metabolically inactive culture).

Ex-type culture: ATCC 32363 = CBS 562.70 = IMI 251387.

Type locality: **Guinea-Bissau**.

Type substrate: Stored nuts of *Arachis hypogaea*.

Descriptions and illustrations: See [Sandoval-Denis et al. \(2019\)](#).

Diagnostic DNA barcodes: rpb2: LR583901; tef1: DQ247606.

Note: Synonym fide Sandoval-Denis et al. (2019).

orobanches Fusarium Jacz., Ezhegodnik Svedeniy Boleznykh i Povrezhdeniyakh Kult'urnykh i Dikorastushchikh Poleznykh Rasteniy. Pertograd. 6: 190. 1910 [1912].

Holotypus: Not located.

Type locality: Russia, Saratov.

Type substrate: Orobanche sp.

Notes: Status unclear. Could be a synonym of *F. oxysporum*.

orthoceras Fusarium Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land- Forstw. 8: 155. 1910.

(See ***Fusarium oxysporum***)

Syntypes: B 70 0100192 & B 70 0100193.

Type locality: Germany, Berlin, Dahlem.

Type substrate: Solanum tuberosum.

Note: Typification pending further study of the syntypes in B.

orthoconium Fusarium Wollenw., Fusaria Autogr. Delin. 2: 637. 1926.

Mycogloea orthospora (Syd.) McNabb ex Dingley, Mem. New York Bot. Gard. 49: 206. 1989.

Basionym: Microcera orthospora Syd., Ann. Mycol. 22: 317. 1924, non *Fusarium orthosporum* Sacc. 1902.

Synonyms: Fusarium microcera var. *orthoconium* (Wollenw.) Bilař, Mikrobiol. Zhurn. 49: 7. 1987, *nom. inval.*, Arts. 35.1, 41.4.

Holotypus: Not located.

Type locality: New Zealand, Wellington, York Bay.

Type substrate: Nothofagus sp.

orthosporum Fusarium Sacc. & P. Syd., Syll. Fung. 16: 1100. 1902.

Cylindrodendrum orthosporum (Sacc. & P. Syd.) L. Lombard, **comb. nov.** MycoBank MB 837700.

Basionym: Fusarium orthosporum Sacc. & P. Syd., Syll. Fung. 16: 1100. 1902.

Synonyms: Cylindrocarpon orthosporum (Sacc. & P. Syd.) Wollenw., Fusaria Autogr. Delin. 1: 462. 1916.

Ramularia orthospora (Sacc. & P. Syd.) Wollenw., Fusaria Autogr. Delin. 1: 462. 1916.

Neonectria hubeiensis W.Y. Zhuang et al., Fungal Diversity 24: 351. 2007.

Ilyonectria hubeiensis (W.Y. Zhuang et al.) Z.Q. Zeng & W.Y. Zhuang, Phytotaxa 85: 17. 2013.

Cylindrodendrum hubeiense (W.Y. Zhuang et al.) L. Lombard & Crous, Phytopathol. Medit. 53: 523. 2014.

Holotypus: In PAD.

Type locality: France.

Type substrate: Juglans nigra.

Descriptions and illustrations: See Zhuang et al. (2007) and Lombard et al. (2014).

Notes: The epithet of *Fusarium orthosporum* Sacc. & P. Syd (1902) predates that of *Neonectria hubeiensis* W.Y. Zhuang et al. (2007). Therefore, a new combination is proposed here with the older epithet.

oryzae Fusarium Vincens, Rev. Pathol. Veg. Entomol. Agric. France 10: 126. 1923.

Holotypus: ?PC.

Type locality: Vietnam.

Type substrate: Oryza sativa.

Notes: Status unclear. Could be a synonym of *F. fujikuroi*.

osiliense Fusarium Bres. & Vesterg., Bot. Not. 1900: 33. 1900.

Septogloeum oxysporum Sacc. et al., Bull. Soc. Roy. Bot. Belgique 29: 294. 1890.

Syntypes: In BPI, NEB, S & UPS.

Type locality: Estonia, Osilia.

Type substrate: Briza media.

Notes: Synonym fide Wollenweber & Reinking (1935). Typification pending further study of the syntypes.

ossicola Fusarium (Berk. & M.A. Curtis) Sacc., Syll. Fung. 4: 714. 1886.

Basionym: Fusisporium ossicola Berk. & M.A. Curtis, Grevillea 3: 147. 1875.

(See ***Fusarium equiseti***)

Holotypus: ?K(M).

Type locality: USA.

Type substrate: Old decaying bones.

Note: Synonyms fide Wollenweber & Reinking (1935).

osteophilum Fusarium Speg., Anales Soc. Ci. Argent. 10: 60. 1880.

(See ***Fusarium scirpi***)

Holotypus: In LPS (Fungi Argent. pag. 2, no. 155) fide Farr (1973).

Type locality: Argentina, Rio de la Plata, La Recoleta.

Type substrate: Decayed bones of Gallus sp. (chicken).

Note: Synonym fide Wollenweber & Reinking (1935).

otomycosis Fusarium Y.N. Ming & T.F. Yu, Acta Microbiol. Sin. 12: 178. 1966.

Holotypus: Not located.

Type locality: China, Beijing.

Type substrate: Ear of Homo sapiens.

Notes: Status unclear. Requires further investigation.

oxydendri Fusarium Ellis & Everh., Bull. Torrey Bot. Club 24: 477. 1897.

(See *Fusarium cavispermum*)

Syntypes: In BPI, BRU, CLEM, CUP, F, FLAS, ILL, ILLS, ISC, MICH, MSC, MU, NEB, OSC, PH, PUL, UC & WIS.

Type locality: USA, West Virginia.

Type substrate: Oxydendrum arboreum.

Notes: Synonym fide Wollenweber & Reinking (1935). Typification pending further study of the syntypes.

oxysporum Fusarium Schltdl., Fl. Berol. 2: 139. 1824.

Synonyms: Fusisporium aurantiacum Link, Mag. Ges. Naturf. Freunde Berlin 3: 19. 1809.

Fusarium aurantiacum (Link) Sacc., Syll. Fung. 4: 720. 1886, *nom. illegit.*, Art. 53.1.

Fusarium aurantiacum Corda, in Sturm, Deutschl. Fl., 3 Abt. (Pilze Deutschl.) 2: 19. 1829.

Fusarium oxysporum var. *aurantiacum* (Corda) Rabenh., Deutschl. Krypt.-Fl., 1: 51. 1844.

Atractium aurantiacum (Corda) Bonord., Abh. Naturf. Ges. Halle 8: 135. 1851.

Fusisporium lagenariae Schwein., Trans. Amer. Philos. Soc., n.s. 4: 275. 1834.

Fusarium lagenariae (Schwein.) Sacc., Syll. Fung. 4: 724. 1886.

Hymenula equiseti Lib., Pl. Crypt. Arduenna Fasc. 3: no. 236. 1834.

Fusarium equisetorum (Lib.) Desm., Pl. Crypt. N. France no. 1546/1846? 1843.

Fusarium parasiticum Thüm., Nuovo Giorn. Bot. Ital. 12: 198. 1880, *nom. illegit.*, Art. 53.1.

- Fusarium thuemenii* Sacc., Syll. Fung. 4: 722. 1886.
Fusisporium calcareum Thüm., Inst. Coimbra 28: 262. 1881.
Fusarium calcareum (Thüm.) Sacc., Syll. Fung. 4: 712. 1886.
Fusarium eucalyptorum Cooke & Harkn., Grevillea 9: 128. 1881.
Fusarium oxysporum f. *eucalypti* (Cooke & Harkn.) Arya & G.L. Jain, Phytopathology 52: 641. 1962.
Fusarium oxysporum f. *lycopersici* Sacc., Syll. Fung. 4: 705. 1886.
Fusarium lycopersici (Sacc.) Mussat, Syll. Fung. 15: 144. 1901, *nom. inval.*, Art. 36.1(a), (c).
Fusarium lycopersici (Sacc.) Wollenw., Phytopathology 3: 29. 1913, *nom. illegit.*, Art. 53.1.
Fusarium bulbigenum Cooke & Masee, Grevillea 16: 49. 1887.
Fusarium myosotidis Cooke, Grevillea 16: 49. 1887.
Leptosporium mycophilum P. Karst., Meddel. Soc. Fauna Fl. Fenn. 16: 24. 1888.
Fusarium mycophilum (P. Karst.) Sacc., Syll. Fung. 10: 730. 1892.
?Selenosporium cuticola R. Blanch., Compt. Rend. Hebd. Séances Acad. Sci., Ser. D. 111: 479. 1890.
?Fusarium cuticola (R. Blanch.) Guég., Champ. Paras. Homme: 262. 1904.
Fusarium sclerodermatis Peck, Rep. (Annual) Regents Univ. State New York New York State Mus. 43: 77. 1890, *nom. illegit.*, Art. 53.1.
Fusarium peckii Sacc., Syll. Fung. 10: 727. 1892, *nom. illegit.*, Art. 53.1 [pro. p. fide [Wollenweber & Reinking \(1935\)](#)].
Fusarium saccardoanum P. Syd., Syll. Fung. 14: 1128. 1899.
Fusarium vasinfectum G.F. Atk., Bull. Alabama Agric. Exp. Sta. 41: 28. 1892.
Fusarium cordae Masee, Brit. Fung.-Fl. 3: 481. 1893.
Fusarium niveum E.F. Sm., Proc. Amer. Assoc. Advancem. Sci. 43: 289. 1894, *nom. inval.*, Art. 36.1(a).
Fusarium bulbigenum var. *niveum* E.F. Sm. ex Wollenw., *Fusarien*: 117. 1931.
Fusarium blasticola Rostr. (as '*blasticolum*'), Gartn.-Tidende 1895: 122. 1895.
Fusoma blasticola (Rostr.) Sacc. & Traverso, Syll. Fung. 20: 1241. 1911.
Fusarium bulbigenum var. *blasticola* (Rostr.) Wollenw., Z. Parasitenk. (Berlin) 3: 412. 1931.
Fusarium beticola A.B. Frank, Kampfbuch gegen die Schädlinge unserer Feldfrüchte: 137. 1897.
Fusarium dianthi Prill. & Delacr., Compt. Rend. Hebd. Séances Acad. Sci. 129: 745. 1899.
Fusarium oxysporum f. *dianthi* (Prill. & Delacr.) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 27: 66. 1940.
Fusarium oxysporum var. *dianthi* (Prill. & Delacr.) Raillo, Fungi of the Genus *Fusarium*: 255. 1950.
Fusarium opuntiarum Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 6: 350. 1898 [1899].
Fusoma pini Hartig, Lehrb. Pflanzenkrankh., Bot., Forstl., Landw. Gärt.: 116. 1900.
Fusarium laxum Peck, Bull. New York State Mus. Nat. Hist. 67: 30. 1903.
Fusarium lini Bolley, Proc. Annual Meeting Soc. Promot. Agric. Sci. 22: 42. 1902.
Fusarium oxysporum f. *lini* (Bolley) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 27: 66. 1940.
Fusarium tabacivorum Delacr., Ann. Inst. Natl. Rech. Agron., ser. 2, 5: 207. 1906.
Fusarium candidulum Sacc., Ann. Mycol. 6: 567. 1908.
Fusarium cubense E.F. Sm., Science, N.Y. 31: 754. 1910.
Fusarium oxysporum var. *cubense* (E.F. Sm.) Wollenw., *Fusarien*: 119. 1935.
Fusarium oxysporum f. *cubense* (E.F. Sm.) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 27: 66. 1940.
Fusarium orthoceras Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land- Forstw. 8: 155. 1910.
Fusarium oxysporum var. *orthoceras* (Appel & Wollenw.) Bilař, Microbiol. Zhurn. 49: 7. 1987.
?Fusarium violae F.A. Wolf, Mycologia 2: 21. 1910.
Fusarium albidoviolaceum Dasz. (as '*albido-violaceum*'), Bull. Soc. Bot. Genève, sér. 2, 4: 293. 1912.
Fusarium orthoceras var. *albidoviolaceum* (Dasz.) Wollenw., *Fusaria* Autogr. Delin. 1: 361. 1916.
Fusarium lycopersici Bruschi, Atti Reale Accad. Lincei, Rendiconti Cl. Sci. Fis., ser. 5, 21: 298. 1912.
Fusarium bulbigenum var. *lycopersici* (Bruschi) Wollenw. & Reinking, *Fusarien*: 114. 1935.
Fusarium citrinum Wollenw., in Lewis, Bull. Maine Agric. Exp. Sta. 219: 256. 1913.
Fusarium conglutinans var. *citrinum* (Wollenw.) Wollenw., Z. Parasitenk. (Berlin) 3: 407. 1931.
Fusarium conglutinans Wollenw., Ber. Deutsch. Bot. Ges. 31: 34. 1913.
Fusarium orthoceras var. *conglutinans* (Wollenw.) Padwick, Indian J. Agric. Sci. 10: 282. 1940.
Fusarium oxysporum f. *conglutinans* (Wollenw.) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 27: 66. 1940.
Fusarium elegans Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land- Forstw. 8: 94. 1913, *nom. inval.*, Art. 36.1(a) (*non Fusarium elegans* W. Yamam. & Maeda 1962).
Fusarium batatas Wollenw. (as '*batatae*'), J. Agric. Res. 2: 268. 1914.
Fusarium bulbigenum var. *batatas* (Wollenw.) Wollenw., Z. Parasitenk. (Berlin) 3: 414. 1931.
Fusarium oxysporum f. *batatas* (Wollenw.) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 27: 66. 1940.
Fusarium cepae Hanzawa, Mycol. Centralbl. 5(1): 5. 1914.
Fusarium oxysporum f. *cepae* (Hanzawa) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 27: 66. 1940.
Fusarium oxysporum var. *cepae* (Hanzawa) Raillo, Fungi of the Genus *Fusarium*: 253. 1950.
Fusarium hyperoxysporum Wollenw., J. Agric. Res. 2: 268. 1914.
Fusarium angustum Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 203. 1915.
Fusarium lutulatum Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 209. 1915.
Fusarium vasinfectum var. *lutulatum* (Sherb.) Wollenw., *Fusaria* Autogr. Delin. 3: 1019. 1930.
Fusarium lutulatum var. *zonatum* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 214. 1915.
Fusarium zonatum (Sherb.) Wollenw., *Fusaria* Autogr. Delin. 1: 392. 1916.
Fusarium vasinfectum var. *zonatum* (Sherb.) Wollenw., *Fusaria* Autogr. Delin. 3: 1020. 1930.
Fusarium oxysporum var. *asclerotium* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 222. 1915.
Fusarium asclerotium (Sherb.) Wollenw., *Fusaria* Autogr. Delin. 1: 364. 1916.
Fusarium sclerotioides Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 214. 1915.
Fusarium sclerotioides var. *brevius* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 218. 1915.

- Fusarium trifolii* Jacz., Jahrb. Pflanzenkrankh. Russl. VII-VIII: Abt. 6. 1917.
- Fusarium citrulli* Taubenh., Bull. Texas Agric. Exp. Sta. 260: 27. 1920.
- Fusarium malvacearum* Taubenh., Bull. Texas Agric. Exp. Sta. 260: 27. 1920.
- Fusarium poolense* Taubenh., Bull. Texas Agric. Exp. Sta. 260: 27. 1920.
- Fusarium macroxysporum* Lindf., Meddel. Centralanst. För söksväs. Jordbruksomr. Avd. Lantbruksbot. 25: 8. 1922.
- Fusarium spinaciae* Hungerf., Phytopathology 13: 209. 1923.
- Fusarium cromyophthoron* Sideris, Phytopathology 14: 212. 1924.
- Fusarium loncheceras* Sideris, Phytopathology 14: 213. 1924.
- Fusarium loncheceras* var. *microsporon* Sideris, Phytopathology 14: 213. 1924.
- Fusarium rhizochromatistes* Sideris, Phytopathology 14: 212. 1924.
- Fusarium sclerostromaton* Sideris, Phytopathology 14: 213. 1924.
- Fusarium zonatum* f. 1 Link & Bailey, J. Agric. Res. 33: 941. 1926.
- Fusarium zonatum* f. 2 Link & Bailey, J. Agric. Res. 33: 941. 1926.
- Fusarium conglutinans* var. *betae* D. Stewart, Phytopathology 21: 67. 1931.
- Fusarium oxysporum* f. *betae* (D. Stewart) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 27: 66. 1940.
- Fusarium oxysporum* f. 7 Wollenw., Fusaria Autogr. Delin. 4: 1176. 1935
- Fusarium apii* P.E. Nelson & Sherb., Techn. Bull. Michigan Agric. Exp. Sta 155: 42. 1937.
- Fusarium orthoceras* var. *apii* (R. Nelson & Sherb.) Wollenw. & Reinking, *Fusarien*: 112. 1935.
- Fusarium oxysporum* f. *apii* (R. Nelson & Sherb.) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 27: 66. 1940.
- Fusarium apii* var. *pallidum* R. Nelson & Sherb., Techn. Bull. Michigan Agric. Exp. Sta. 155: 42. 1937.
- Fusarium bulbigenum* var. *apii* (R. Nelson & Sherb.) Raillo, Fungi of the Genus *Fusarium*: 251. 1950.
- Cylindrophora albedinis* Kill. & Maire, Bull. Soc. Hist. Nat. Afrique N. 21: 97. 1930, *nom. inval.*, Art. 36.1(b).
- Fusarium oxysporum* var. *albedinis* Kill. & Maire ex Malençon, Rev. Mycol. (Paris) 15: 45–60. 1950, *nom. inval.*, Art. 36.1(b).
- Fusarium oxysporum* f. *sp. albedinis* Kill. & Maire ex W.L. Gordon, Canad. J. Bot. 43: 1310. 1965.
- Fusarium perniciosum* Hepting, Circ. U.S.D.A. 535: 7. 1939.
- Fusarium oxysporum* f. *perniciosum* (Hepting) Toole, Phytopathology 31: 599. 1941.
- Fusarium vasinfectum* var. *perniciosum* (Hepting) Carrera, Rev. Fac. Agron. Buenos Aires 13(3): 483 1955
- ?*Fusarium retusum* Wellman, Phytopathology 33: 957. 1943.
- Holotypus*: HAL 1612 F.
- Epitypus*: CBS H-23620, designated in Lombard *et al.* (2019b).
- Ex-epitype culture*: CBS 144134.
- Type locality*: **Germany**, Berlin.
- Type substrate*: *Solanum tuberosum*.
- Descriptions and illustrations*: See Lombard *et al.* (2019b)
- Diagnostic DNA barcodes*: *rpb2*: MH484953; *tef1*: MH485044.
- palczewskii* *Fusarium* Jacz., Bull. Soc. Mycol. France 28: 345. 1912.
- (See ***Fusarium avenaceum***)
- Lectotypus* (*hic designatus*, MBT 10000717): **Russia**, Ussuriysk, Primorsky krai (Far East Territory), grain of *Lolium* sp., 1912, A.A. Jaczewski, in Bull. Soc. Mycol. France 28: 345, fig. 1.
- Notes*: Synonyms *fide* Wollenweber & Reinking (1935). As no holotype specimen could be located; an illustration accompanying the original protologue is designated here as lectotype.
- pallens* *Fusarium* Berk. & M.A. Curtis, Grevillea 3: 99. 1875, *nom. illegit.*, Art. 53.1.
- Replacing synonym*: *Fusarium glumarum* Sacc., Syll. Fung. 4: 706. 1886.
- (See ***Fusarium incarnatum***)
- Authentic material*: Car. Inf. no. 3799, in K(M).
- Original locality*: **USA**.
- Original substrate*: *Juncus* sp.
- Note*: Synonyms *fide* Wollenweber & Reinking (1935).
- pallens* *Fusarium* (Nees & T. Nees) Link, Sp. pl. 6(2): 104. 1825.
- Basionym*: *Atractium pallens* Nees & T. Nees, Nova Acta Phys.-Med. Acad. Caes. Leop.-Carol. Nat. Cur. 9: 237. 1818.
- Synonyms*: *Volutella pallens* (Nees & T. Nees) Fr., Syst. Mycol. 3: 468. 1832.
- Selenosporium pallens* (Nees & T. Nees) Corda, Icon. Fung. 1: 7. 1837.
- Holotypus*: In B.
- Type locality*: **Germany**.
- Type substrate*: Fallen branch.
- Notes*: The type material of *Atractium pallens* is deposited at B and examined by Gräfenhan *et al.* (2011), identifying it as a coelomycete.
- pallidroseum* *Fusarium* (Cooke) Sacc., Syll. Fung. 4: 720. 1886.
- Basionym*: *Fusisporium pallidroseum* Cooke, Grevillea 6: 139. 1878.
- (See ***Fusarium incarnatum***)
- Holotypus*: S. Car. no. 2279 in ?K(M).
- Type locality*: **USA**, South Carolina, Aiken.
- Type substrate*: *Chenopodium anthelminticum*.
- Note*: Synonyms *fide* Wollenweber & Reinking (1935).
- pallidulum* *Fusarium* Sacc. & Trotter, Syll. Fung. 22: 1483. 1913.
- Replaced synonym*: *Atractium pallidum* Bonord., Handb. Mykol.: 135. 1851.
- Synonym*: *Fusarium pallidum* (Bonord.) Sacc. & Traverso, Syll. Fung. 19: 727. 1910, *nom. illegit.*, Art. 53.1.
- Lectotypus* (*hic designatus*, MBT 10000718): **Germany**, decaying bark, 1913, H.F. Bonorden, in Handb. Mykol., tab. 10, fig. 219.
- Notes*: Status unclear. Not *Fusarium* *fide* Wollenweber & Reinking (1935). As no holotype specimen could be located, an illustration accompanying the original protologue is designated here as lectotype.
- pallidum* *Fusarium* Berk. & M.A. Curtis, J. Linn. Soc., Bot. 10: 359. 1869.
- Holotypus*: In K(M).
- Type locality*: **Cuba**.
- Type substrate*: Dead twigs.

Notes: Status unclear. Not *Fusarium* fide [Wollenweber & Reinking \(1935\)](#).

palustre Fusarium W.H. Elmer & Marra, *sp. nov.* MycoBank MB 837702.

Synonym: *Fusarium palustre* W.H. Elmer & Marra, *Mycologia* 103(4): 815. 2011, *nom. inval.*, Art. 40.7.

Etymology: 'palustre', from Latin *palus*, referring to marsh habitat in which this fungus is found.

For diagnosis see Elmer & Marra, *Mycologia* 103(4): 815. 2011.

Holotypus: CBS 126795 (preserved as metabolically inactive culture).

Ex-type culture: CBS 126796 = NRRL 54056.

Type locality: **USA**, Connecticut, Madison, Hammonasset Beach State Park.

Type substrate: *Spartina alterniflora*.

Descriptions and illustrations: See [Elmer & Marra \(2011\)](#).

Diagnostic DNA barcodes: *rpb1*: KT597718; *rpb2*: KT597731; *tef1*: GQ856941.

Notes: [Elmer & Marra \(2011\)](#) failed to indicate the holotype for *F. palustre*, rendering the species name invalid (Art. 40.7). Here we validate the name.

pampini Fusarium Thüm. & Pass., *Pilze Weinst.*: 50. 1878.

Gloeosporium physalosporae Cavara, *Rev. Mycol. (Toulouse)* 10: 99. 1888.

Holotypus: Not located.

Type locality: **Italy**, Parma.

Type substrate: *Vitis vinifera*.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

pandani Fusarium (Corda) Sacc., *Syll. Fung.* 4: 724. 1886.

Basionym: *Fusisporium pandani* Corda, *Icon. Fung.* 1: 11. 1837. *Lectotypus* (*hic designatus*, MBT 10000719): **Czech Republic**, Liberec (Reichenberg), *Pandanus* sp., 1837, A.C.J. Corda, in *Icon. Fung.* 1, tab. 2, fig. 162.

Notes: Status unclear. Not *Fusarium* fide [Wollenweber & Reinking \(1935\)](#). As no holotype specimen could be located, an illustration accompanying the original protologue is designated here as lectotype.

pannosum Fusarium Masee, *Bull. Misc. Inform. Kew* 1898: 117. 1898.

(See ***Fusarium sambucinum***)

Holotypus: K(M) 191093.

Type locality: **India**, Punjab.

Type substrate: *Cornus macrophylla*.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

paraemartii Fusarium (Sand.-Den. & Crous) O'Donnell *et al.*, *Index Fungorum* 440: 3. 2020.

Neocosmospora paraemartii Sand.-Den. & Crous, *Persoonia* 43: 149. 2019.

Holotypus: CBS H-23991.

Ex-type culture: BBA 62215 = CBS 487.76 = NRRL 13997.

Type locality: **Argentina**.

Type substrate: Decaying stem base of *Solanum tuberosum*.

Descriptions and illustrations: See [Sandoval-Denis *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW834240; *rpb2*: LR583855; *tef1*: DQ247549.

paranaense Fusarium S.S. Costa *et al.*, *Fungal Biology* 120: 55. 2015 [2016].

(See *Fusarium falciforme*)

Holotypus: CML 1830.

Ex-type culture: CBS 141593 = CML 1830.

Type locality: **Brazil**, Goiás State, Cristalina.

Type substrate: Diseased tissue of *Glycine max*.

Descriptions and illustrations: See [Costa *et al.* \(2016\)](#).

Diagnostic DNA barcodes: *rpb2*: KF680011; *tef1*: KF597797.

Note: Synonym fide [Sandoval-Denis *et al.* \(2019\)](#).

parasiticum Fusarium Westend., *Bull. Acad. Roy. Sci. Belgique, Cl. Sci., sér. 2*, 11: 652. 1861.

(See *Fusarium ciliatum*)

Holotypus: BR5020140791441.

Type locality: **Belgium**, Louette-Saint-Pierre.

Type substrate: *Sphaeria gigaspora*.

Note: Synonym fide [Wollenweber & Reinking \(1935\)](#).

parasiticum Fusarium Thüm., *Nuovo Giorn. Bot. Ital.* 12: 198. 1880, *nom. illegit.*, Art. 53.1.

Replacing synonym: *Fusarium thuemenii* Sacc., *Syll. Fung.* 4: 722. 1886.

(See ***Fusarium oxysporum***)

Authentic material: Not located.

Original locality: **Russia**, Orenburg.

Original substrate: *Betula pendula*.

Note: Synonyms fide [Wollenweber & Reinking \(1935\)](#).

parasiticum Fusarium Ellis & Kellerm., *J. Mycol.* 3: 127. 1887, *nom. illegit.*, Art. 53.1.

Replacing synonym: *Fusarium pucciniophilum* Sacc. & P. Syd., *Syll. Fung.* 14: 1128. 1899.

(See ***Fusarium heterosporum***)

Authentic material: Kellerman & Swingle 1104 in NY.

Original locality: **USA**, Manhattan.

Original substrate: Parasitic on *Puccinia seymeriae* on *Swietenia macrophylla*.

Note: Synonyms fide [Wollenweber & Reinking \(1935\)](#).

parasiticum Fusarium Fautrey, *Rev. Mycol. (Toulouse)* 11: 153. 1889, *nom. illegit.*, Art. 53.1.

Replacing synonym: *Fusarium fautreyi* Sacc., *Syll. Fung.* 10: 934. 1892.

(See ***Fusarium lateritium***)

Authentic material: BR5020140789424.

Original locality: **France**, Noidan.

Original substrate: *Vitis vinifera*.

Note: Synonyms fide [Wollenweber & Reinking \(1935\)](#).

parceramosum Fusarium (Sand.-Den. & Crous) O'Donnell *et al.*, *Index Fungorum* 440: 3. 2020.

Neocosmospora parceramosa Sand.-Den. & Crous, *Persoonia* 43: 151. 2019.

Holotypus: CBS H-23992.

Ex-type culture: CBS 115695 = CPC 1246.

Type locality: **South Africa**.

Type substrate: Soil.

Descriptions and illustrations: See [Sandoval-Denis *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb2*: JX435249; *tef1*: JX435149.

parvisorum Fusarium Herron *et al.*, *Stud. Mycol.* 80: 146. 2015.

Holotypus: PREM 60897.

Ex-type culture: CBS 137236 = CMW 25267.

Type locality: **Colombia**, Vivero, Peñas Negra, Valle del Cauca.

Type substrate: *Pinus patula*.

Descriptions and illustrations: See [Herron *et al.* \(2015\)](#).

Notes: Comparisons of recently generated sequences for the living ex-type (CBS 137236 = CMW 25267) of *F. parvisorum* indicate a strain transposition or contamination by another *Fusarium* species. Therefore, this species needs to be recollected from the type locality and substrate or sequences need to be generated from the holotype specimen to confirm that it is indeed distinct.

paspali *Fusarium* Henn., Bot. Jahrb. Syst. 38: 129. 1905.

(See *Fusarium avenaceum*)

Syntype: In B as Zenker, Georg August, no. 2152 fide Hein (1988).

Type locality: **Cameroon**, Bipindi.

Type locality: *Paspalum* sp.

Notes: Synonym fide Wollenweber & Reinking (1935). Typification pending further study of the syntype in B.

paspalicola *Fusarium* Henn., in Warburg, Monsunia 1: 38. 1899 [1900].

(See *Fusarium heterosporum*)

Holotypus: In B fide Wollenweber, Fusaria Autogr. Delin. 1: 299. (1916) & Hein (1988).

Type locality: **Philippines**, Mindanao, Davao.

Type substrate: *Paspalum* sp.

Note: Synonym fide Wollenweber & Reinking (1935).

patouillardii *Fusarium* Sacc. (as 'patouillard'), Syll. Fung. 10: 729. 1892.

Replaced synonym: *Fusarium uredinicola* Pat. & Gaillard, Bull. Soc. Mycol. France 4: 127. 1888, nom. illegit., Art. 53.1.

Holotypus: ?PC or FH.

Type locality: **Venezuela**, Caracas.

Type substrate: Parasitic on *Sphaerellopsis filum* on *Puccinia pallidissima*.

Notes: Status unclear. Not *Fusarium* fide Wollenweber & Reinking (1935).

peckii *Fusarium* Sacc., Syll. Fung. 4: 713. 1886.

Replaced synonym: *Fusisporium parasiticum* Peck, Rep. (Annual) New York State Mus. Nat. Hist. 29: 53. 1878, non *Fusarium parasiticum* Westend. 1861.

Holotypus: NYSf2260.

Type locality: **USA**, New York, Albany.

Type substrate: *Sphaeria collinsii*.

Notes: Status unclear. Not treated by Wollenweber & Reinking (1935) or Booth (1971).

peckii *Fusarium* Sacc., Syll. Fung. 10: 727. 1892, nom. illegit., Art. 53.1.

Replaced synonyms: *Fusarium sclerodermatis* Peck, Rep. (Annual) Regents Univ. State New York New York State Mus. 43: 77. 1890, nom. illegit., Art. 53.1, non *Fusarium sclerodermatis* Oudem. 1889.

Fusarium saccardoanum Syd., Syll. Fung. 13: 1130. 1898.

(See *Fusarium oxysporum* pr. p. & *Fusarium avenaceum* pr. p.)

Authentic material: NYSf2731.

Original locality: **USA**, New York, Suffolk.

Original substrate: *Scleroderma vulgare*.

Note: Synonyms fide Wollenweber & Reinking (1935).

pelargonii *Fusarium* P. Crouan & H. Crouan, Fl. Finistère: 14. 1867.

(See *Fusarium merismoides*)

Holotypus: ?PC.

Type locality: **France**, Finistère.

Type substrate: *Pelargonium* sp.

Note: Synonym fide Wollenweber & Reinking (1935).

peltigerae *Fusarium* Westend., Herb. Crypt. Belg. Fasc. 9: no. 414. 1849.

(See *Fusarium ciliatum*)

Syntypes: In BR & PH (Herb. Crypt. Belg. 9: no. 414).

Type locality: **Belgium**.

Type substrate: *Peltigera rufescens*.

Notes: Synonym fide Wollenweber & Reinking (1935). Typification pending further study of the syntypes.

penicillatum *Fusarium* (Harz) Sacc., Syll. Fung. 4: 710. 1886.

Basionym: *Menispora penicillata* Harz, Bull. Soc. Imp. Naturalistes Moscou 44: 127. 1871.

(See *Fusarium avenaceum*)

Lectotypus (*hic designatus*, MBT 10000720): **Germany**, Berlin, decaying *Sclerotium clavus*, 1886, C. Harz, in Bull. Soc. Imp. Naturalistes Moscou 44, tab. 1, fig. 4.

Notes: Synonym fide Wollenweber & Reinking (1935). As no holotype specimen could be located, an illustration accompanying the original protologue is designated here as lectotype.

pentaclethrae *Fusarium* Henn., Hedwigia 44: 71. 1905.

(See *Fusarium coccidicola*)

Syntype: In B (Ule no. 3011) fide Hein (1988).

Type locality: **Brazil**, Manaus, Rio Negro.

Type substrate: Leaves of *Pentaclethra* sp.

Notes: Synonym fide Wollenweber & Reinking (1935). Typification pending further study of the syntype in B.

penzigii *Fusarium* Schroers et al., Mycologia 101: 61. 2009.

Bisifusarium penzigii (Schroers et al.) L. Lombard & Crous, Stud. Mycol. 80: 225. 2015.

Holotypus: CBS H-20125.

Ex-type culture: CBS 317.34 = NRRL 22109.

Type locality: **UK**, Surrey.

Type substrate: Decayed wood of *Fagus sylvatica*.

Descriptions and illustrations: See Schroers et al. (2009).

Diagnostic DNA barcodes: *rpb1*: KM232211; *rpb2*: KM232362; *tef1*: EU926324.

pernambucanum *Fusarium* A.C.S. Santos et al., Mycologia 111: 253. 2019.

Holotypus: URM 91193.

Ex-type culture: MUM 1862 = URM 7559.

Type locality: **Brazil**, Pernambuco, Paudalho.

Type substrate: *Aleurocanthus woglumi*.

Descriptions and illustrations: See Santos et al. (2019).

Diagnostic DNA barcodes: *rpb1*: MH668869; *rpb2*: LS398519; *tef1*: LS398489.

perniciosum *Fusarium* Hepting, Circul. U.S.D.A. 535: 7. 1939.

(See *Fusarium oxysporum*)

Holotypus: Not located.

Type locality: **USA**.

Type substrate: *Albizia julibrissin*.

persicae *Fusarium* (Sacc.) G.F. Atk., J. Elisha Mitchell Sci. Soc. 8: 41. 1892.

Basionym: *Cercospora persicae* Sacc. (as 'persica'), Hedwigia 15: 119. 1876.

Mycosphaerella pruni-persicae Deighton, Trans. Brit. Mycol. Soc. 50: 328. 1967.

Synonyms: *Cercospora persicae* (Sacc.) Sacc. (as 'persica'), *Michelia* 2: 20. 1880.

Clasterosporium persicae (Sacc.) Tsuji, *Ann. Phytopathol. Soc. Japan* 1(2): 33. 1919.

Miuraea persicae (Sacc.) Hara, *Byogaichu-Hoten (Manual of Pests and Diseases)*: 224. 1948.

Mycosphaerella persicae B.B. Higgins & F.A. Wolf (as 'persica'), *Phytopathology* 27: 695. 1937.

Syntype: In HAL, ILL & NEB (Saccardo, *Mycoth. Ven.* no. 598).

Type locality: **Italy**.

Type substrate: *Prunus persica*.

persicinum Fusarium J.W. Xia *et al.*, *Persoonia* 43: 215 2019.

Holotypus: CBS H-24068.

Ex-type culture: CBS 479.83.

Type locality: **Unknown**.

Type substrate: Unknown.

Descriptions and illustrations: See [Xia *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb2*: MN170428; *tef1*: MN170495.

personatum Fusarium Cooke, in *Harkness, Grevillea* 7: 12. 1878. (See *Fusarium allescherianum*)

Holotypus: ?K(M).

Type locality: **USA**, California.

Type substrate: *Oreodaphne californica*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

perseae Fusarium (Sand.-Den. & Guarnaccia) O'Donnell *et al.*, *Index Fungorum* 440: 3. 2020.

Neocosmospora perseae Sand.-Den. & Guarnaccia, *Fungal Syst. Evol.* 1: 136. 2018.

Holotypus: CBS H-23433.

Ex-type culture: CBS 144142 = CPC 26829.

Type locality: **Italy**, Catania, San Leonardello.

Type substrate: Trunk canker lesions on *Persea americana*.

Descriptions and illustrations: See [Guarnaccia *et al.* \(2018\)](#).

Diagnostic DNA barcodes: *rpb1*: MW218130; *rpb2*: LT991909; *tef1*: LT991902.

peruvianum Fusarium L. Lombard & Crous, *Fungal Syst. Evol.* 4: 194. 2019.

Holotypus: CBS H-24019.

Ex-type culture: CBS 511.75.

Type locality: **Peru**.

Type substrate: Seedlings of *Gossypium* sp.

Descriptions and illustrations: See [Lombard *et al.* \(2019a\)](#).

Diagnostic DNA barcodes: *rpb1*: MN120728; *rpb2*: MN120746; *tef1*: MN120767.

pestis Fusarium Sorauer, *Atlas Pfl.-Krankh.* 4: 19, pl. XXV. 1890. (See *Fusarium azukiicola*)

Holotypus: Not located.

Type locality: **Germany**.

Type substrate: *Solanum tuberosum*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

petersiae Fusarium L. Lombard, *Persoonia* 39: 457. 2017.

Holotypus: CBS H-23233.

Ex-type culture: CBS 143231.

Type locality: **Netherlands**, Gelderland Province, Arnhem.

Type substrate: Soil.

Descriptions and illustrations: See [Crous *et al.* \(2017\)](#).

Diagnostic DNA barcodes: *rpb1*: MG386139; *rpb2*: MG386150; *tef1*: MG386160.

petroliphilum Fusarium (Q.T. Chen & X.H. Fu) Geiser *et al.*, *Fungal Genet. Biol.* 53: 69. 2013.

Neocosmospora petroliphila (Q.T. Chen & X.H. Fu) Sand.-Den. & Crous, *Persoonia* 41: 121. 2018.

Basionym: *Fusarium solani* var. *petroliphilum* Q.T. Chen & X.H. Fu, *Acta Mycol. Sin., Suppl.* 1: 330. 1987.

Synonyms: *Fusarium solani* f. sp. *cucurbitae* (Race 2) W.C. Snyder & H.N. Hansen, *Amer. J. Bot.* 28: 740. 1941.

Holotypus: HMAS 43748.

Ex-type culture: FRC S-2176 = NF4475 = NRRL 22268.

Type locality: **China**, Beijing.

Type substrate: Deteriorated petroleum.

Descriptions and illustrations: See [Sandoval-Denis & Crous \(2018\)](#).

peziziforme Fusarium Berk. & M.A. Curtis (as 'pezizaeforme'), *J. Linn. Soc., Bot.* 10: 360. 1869.

Holotypus: In K(M).

Type locality: **Cuba**.

Type substrate: *Poaceae*.

Note: Not *Fusarium* *fide* [Wollenweber & Reinking \(1935\)](#).

pezizoides Fusarium Desm., *Ann. Sci. Nat., Bot., sér.* 3, 18: 373. 1852.

Trochila craterium (DC.) Fr., *Summa Veg. Scand.* 2: 367. 1849. *Basionym:* *Sphaeria craterium* DC., *Fl. Franç., ed.* 3, 2: 298. 1805.

Synonyms: *Phacidium craterium* (DC.) Gillet, *Champ. France Discomyc.* (7): 167. 1886.

Sphaeria punctiformis var. *hederae* Pers., *Syn. Meth. Fung.* 1: 90. 1801.

Myxosporium paradoxum De Not., *Mem. Reale Accad. Sci. Torino, ser.* 2, 3: 81. 1841.

Gloeosporium paradoxum (De Not.) Mont., in *Berkeley & Broome, Ann. Mag. Nat. Hist.* 5: 455. 1850.

Gloeosporidium paradoxum (De Not.) Petr., *Ann. Mycol.* 20: 14. 1922.

Cryptocline paradoxa (De Not.) Arx, *Verh. Kon. Ned. Akad. Wetensch., Afd. Natuurk.* 51: 115. 1957.

Gloeotrochila paradoxa (De Not.) Petr., *Sydowia* 1: 50. 1947.

Trochila craterium var. *nucleata* Rehm, *Ber. Bayer. Bot. Ges.* 13: 125. 1912.

Ceuthospora hederae Grove, *Bull. Misc. Inform. Kew* 1923: 355. 1923.

Holotypus: ?PC.

Type locality: **France**.

Type substrate: *Peziza insidiosa*.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

pezizoideum Fusarium (Berk. & M.A. Curtis) Sacc., *Syll. Fung.* 4: 711. 1886.

Basionym: *Fusisporium pezizoideum* Berk. & M.A. Curtis, *Grevillea* 3: 147. 1875.

(See ***Fusarium sambucinum***)

Holotypus: ?K(M).

Type locality: **USA**, Pennsylvania.

Type substrate: Stems of herbaceous plants.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

phacidioideum Fusarium Dearn., *Mycologia* 21: 331. 1929.

Holotypus: JD 4303 in DAOM.

Type locality: **Canada**, Vancouver, Stanley Park.

Type substrate: Dead branches of *Pseudotsuga taxifolia*.

Note: Status unclear; requires recollection from type locality and substrate.

pharetrum Fusarium L. Lombard & Crous, *Persoonia* 43: 32. 2018 [2019].

Holotypus: CBS H-23621.

Ex-type culture: CBS 144751 = CPC 30824.

Type locality: **South Africa**.

Type substrate: *Aloidendron dichotomum*.

Descriptions and illustrations: See Lombard *et al.* (2019b).

Diagnostic DNA barcodes: *rpb1*: MW928815; *rpb2*: MH484952; *tef1*: MH485043.

phaseoli Fusarium (Burkh.) T. Aoki & O'Donnell, *Mycologia* 95: 671. 2003.

Basionym: *Fusarium martii* f. *phaseoli* Burkh., Mem. Cornell Univ. Agric. Exp. Sta. 26: 1007. 1919.

(See *Fusarium azukiicola*)

Lectotypus (*hic designatus*, MBT 10000721): **USA**, New York, roots of *Phaseolus vulgaris*, 1919, W.H. Burkholder, in Mem. Cornell Univ. Agric. Exp. Sta. 26: 1009, fig. 134.

Notes: Synonym *fide* Sandoval-Denis *et al.* (2019). Although Burkholder deposited several specimens in CUP, none are directly linked to the original protologue (Burkholder 1919). Several of these specimens appear to have been isolated from greenhouse assays undertaken by Burkholder. Therefore, an illustration accompanying the original protologue is designated here as lectotype.

phialophorum Fusarium Maryani *et al.*, *Stud. Mycol.* 92: 169. 2018 [2019].

Holotypus: InaCC F971 (preserved as metabolically inactive culture).

Ex-type culture: InaCC F971.

Type locality: **Indonesia**, South Kalimantan, Tanah Bumbu, Kampung Betung.

Type substrate: *Musa* var. Pisang Awak.

Descriptions and illustrations: See Maryani *et al.* (2019a).

Diagnostic DNA barcodes: *rpb1*: LS479545; *rpb2*: LS479292; *tef1*: LS479741.

phormii Fusarium Henn., Verh. Bot. Vereins Prov. Brandenburg 40: 175. 1898 [1899].

Colletotrichum phormii (Henn.) D.F. Farr & Rossman, *Mycol. Res.* 110: 1403. 2006.

Synonym: *Gloeosporium phormii* (Henn.) Wollenw., *Fusaria* Autogr. Delin. No. 498. 1916, *nom. illegit.*, Art. 53.1, *non Gloeosporium phormii* Sacc. 1915.

Holotypus: B 70 0005220.

Epitypus: CBS H-20720, designated in Damm *et al.* (2012).

Ex-epitype: A.R. 3546 = CBS 118194.

Type locality: **Germany**, Berlin.

Type substrate: *Phormium tenax*.

phragmiticola Fusarium Kirschst., *Ann. Mycol.* 34: 183. 1936, *nom. inval.*, Art. 39.1.

Authentic material: B 70 0100199, B 70 0100200, B 70010020.

Original locality: **Germany**.

Original substrate: *Phragmites communis*.

phragmitis Fusarium Matsush., *Icon. Microfung. Matsush. Lect.*: 72. 1975, *nom. inval.*, Art. 40.1.

Authentic material: Not indicated.

Original locality: **Japan**.

Original substrate: Rotten wood of *Fagus crenata*.

phyllachorae Fusarium Henn., in de Wildeman, *Mission E. Laurent*, Fasc. 4: 363. 1907.

Syntype: Laurent in B *fide* Hein (1988).

Type locality: **Democratic Republic of Congo**, between Kinshasa and Kwamouth.

Type substrate: *Panicum maximum*.

Notes: Not *Fusarium* *fide* Wollenweber & Reinking (1935). Typification pending further study of the syntype in B.

phyllongenium Fusarium (Cooke & Peck) Sacc., *Syll. Fung.* 4: 703. 1886.

Basionym: *Fusisporium phyllogenium* Cooke & Peck, Rep. (Annual) New York State Mus. Nat. Hist. 29: 53. 1878.

Syntype: NYSf2335.

Type locality: **USA**, New York, Albany, Bethlehem.

Type substrate: *Erigeron annuum*.

Notes: Status unclear. Not *Fusarium* *fide* Wollenweber & Reinking (1935). Typification pending further study of the syntype in NYS.

phyllophilum Fusarium Nirenberg & O'Donnell, *Mycologia* 90: 447. 1998.

Holotypus: IMI 202874.

Ex-type culture: BBA 63625 = CBS 216.76 = DAOM 225132 = IMI 375338 = NRRL 13617.

Type locality: **Italy**.

Type substrate: *Dracaena deremensis*.

Descriptions and illustrations: See Nirenberg & O'Donnell (1998).

Diagnostic DNA barcodes: *rpb1*: KF466399; *rpb2*: KF466410; *tef1*: KF466421.

phyllostachydicola Fusarium W. Yamam., *Trans. Mycol. Soc. Japan* 3: 118. 1962.

Basionym: *Gibberella phyllostachydicola* W. Yamam., *Hyogo Univ. Agric. ser. Agric. Biol.* 3: 15. 1957.

Lectotypus (*hic designatus*, MBT 10000722): **Japan**, Tamba, Sasayama-cho, culms of *Phyllostachys bambusoides*, 31 Aug. 1956, W. Yamamoto, in *Hyogo Univ. Agric. ser. Agric. Biol.* 3: 17, figs 16–18.

Descriptions and illustrations: See Yamamoto *et al.* (1957).

Notes: This species requires recollection from the type host and locality. As no holotype specimen could be located, an illustration accompanying the original protologue is designated here as lectotype.

pilosicola Fusarium Yilmaz *et al.*, *Persoonia* 46: 152. 2021.

Holotypus: PREM 63216.

Ex-type culture: CMWF 1183 = NRRL 29124 = NY007.H7.

Type locality: **USA**, Florida.

Type substrate: *Bidens pilosa*.

Descriptions and illustrations: See Yilmaz *et al.* (2021).

Diagnostic DNA barcodes: *rpb2*: MN534248; *tef1*: MN534055.

pininemorale Fusarium Herron *et al.*, *Stud. Mycol.* 80: 146. 2015.

Holotypus: PREM 60901.

Ex-type culture: CBS 137240 = CMW 25243.

Type locality: **Colombia**, Risaralda, Angela Maria (Santa Rosa).

Type substrate: *Pinus tecunumanii*.

Descriptions and illustrations: See Herron *et al.* (2015).

Notes: Comparisons of recently generated sequences from the living ex-type (CBS 137240 = CMW 25243) of *F. pininemorale* indicate a strain transposition or contamination by another *Fusarium* species. Therefore, this species needs to be

recollected from the type locality and substrate or sequences need to be generated from the holotype specimen to confirm its phylogenetic affiliation.

piperis *Fusarium* (F.C. Albuquerque) O'Donnell *et al.*, Index Fungorum 440: 3. 2020.

Neocosmospora piperis (F.C. Albuquerque) Sand.-Den. & Crous, *Persoonia* 43: 152. 2019.

Basionym: *Fusarium solani* f. *piperis* F.C. Albuquerque, *Circ. Inst. Agron. N.* 5: 19. 1961.

Holotypus: IAN 825 in the herbarium of Embrapa Amazônia Oriental.

Epitypus: CBS H-23993, designated in [Sandoval-Denis *et al.* \(2019\)](#).

Ex-epitype culture: CBS 145470 = CML 1888 = G.J.S. 89-14 = NRRL 22570.

Type locality: **Brazil**.

Type substrate: *Piper nigrum*.

Descriptions and illustrations: See [Sandoval-Denis *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW834241; *rpb2*: EU329513; *tef1*: AF178360.

pisi *Fusarium* (F.R. Jones) A. Šišić *et al.*, *Sci. Rep.* 8(no. 1252): 2. 2018, *nom. inval.*, Art. F.5.1.

Neocosmospora pisi (F.R. Jones) Sand.-Den. & Crous, *Persoonia* 43: 154. 2019.

Basionym: *Fusarium martii* var. *pisi* F.R. Jones, *J. Agric. Res.* 26: 459. 1923.

Synonyms: *Fusarium solani* f. *pisi* (F.R. Jones) W.C. Snyder & H.N. Hansen, *Amer. J. Bot.* 28: 740. 1941.

Fusarium vanettenii O'Donnell *et al.*, Index Fungorum 440: 5. 2020.

Fusarium solani var. *martii* 'f 2' Wollenw., *Z. Parasitenk. (Berlin)* 3: 290. 1931.

Hypomyces solani f. sp. *pisi* Reichle, W.C. Snyder & Matuo, *Nature* 203: 664. 1964.

Lectotypus: Jones (1923; fig. 1 on p. 463), designated in [Sandoval-Denis *et al.* \(2019\)](#).

Epitypus: CBS H-23994, designated in [Sandoval-Denis *et al.* \(2019\)](#).

Ex-epitype culture: ATCC MYA-4622 = CBS 123669 = NRRL 45880 = Vanetten 77-13-4.

Type locality: **USA**.

Type substrate: Sexual cross of parents from *Pisum sativum* and soil from a potato field.

Descriptions and illustrations: See [Šišić *et al.* \(2018b\)](#) and [Sandoval-Denis *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: JX171543; *rpb2*: EU329640; *tef1*: FJ240352.

plagianthi *Fusarium* (Dingley) O'Donnell & Geiser, *Phytopathology* 103: 404. 2013.

Neocosmospora plagianthi (Dingley) L. Lombard & Crous, *Stud. Mycol.* 80: 227. 2015.

Basionym: *Nectria plagianthi* Dingley, *Trans. Roy. Soc. New Zealand* 79: 196. 1951.

?*Nectria pulverulenta* Dingley, *Trans. Roy. Soc. New Zealand* 83: 657. 1956.

Holotypus: PDD 10916.

Type locality: **New Zealand**, Fiordland, Hollyford Valley.

Type substrate: *Plagianthus betulinus*.

Descriptions and illustrations: See [Dingley \(1951\)](#) and [Samuels & Brayford \(1994\)](#).

platani *Fusarium* (Lév.) Mont., *Ann. Sci. Nat., Bot., sér. 3*, 11: 55. 1849.

Basionym: *Hymenula platani* Lév., *Ann. Sci. Nat., Bot., sér. 3*, 9: 128. 1848

(See *Fusarium nervisequum*)

Holotypus: ?PC.

Type locality: **France**.

Type substrate: *Platanus orientalis*.

platanoidis *Fusarium* Oudem., *Ned. Kruidk. Arch., sér. 3*, 2: 1131. 1904.

Holotypus: ?L.

Type locality: **Netherlands**, Gelderland Province, Nunspeet.

Type substrate: *Acer platanoides*.

Note: Not *Fusarium* fide [Wollenweber & Reinking \(1935\)](#).

poae* *Fusarium (Peck) Wollenw., in Lewis, *Bull. Maine. Agric. Exp. Sta.* 219: 254. 1913 [1914].

Basionym: *Sporotrichum poae* Peck, *Bull. New York State Mus.* 67: 29. 1904 [1903].

Synonyms: *Fusarium tricinctum* f. *poae* (Peck) W.C. Snyder & H.N. Hansen, *Amer. J. Bot.* 32: 663. 1945.

Fusarium sporotrichiella var. *poae* (Peck) Bilař, *Yadovitye griby na zeme khlebnykh zlakov (Poisonous fungi on cereal seed)*: 86. 1953, *nom. inval.*, Art. 39.1.

Fusarium sporotrichiella var. *poae* (Peck) Bilař, *Microbiol. Zhurn.* 49: 6. 1987, *nom. inval.*, Arts. 35.1, 41.4.

Sporotrichum anthophilum Peck, *Bull. New York State Mus.* 105: 28. 1906.

Fusarium maydiperdum Bubák, *Centralbl. Bakteriol. Parasitenk.*, 2. Abth. 31: 497. 1911.

Holotypus: NYSf2393.

Type locality: **USA**, New York, Geneva.

Type substrate: Sheaths and culms of *Poa pratensis*.

Epitypus (*hic designatus*, MBT 10000723): **USA**, North Dakota, Minot, from infected barley kernel, date and collector unknown, NRRL 26941 (preserved as metabolically inactive culture).

Ex-epitype culture: NRRL 26941.

Descriptions and illustrations: See [Wollenweber & Reinking \(1935\)](#), [Booth \(1971\)](#), [Gerlach & Nirenberg \(1982\)](#) and [Leslie & Summerell \(2006\)](#).

Diagnostic DNA barcodes: *rpb1*: KU171686; *rpb2*: KU171706; *tef1*: JABFFD010000730.1

Note: No living material linked to the holotype is available for this important mycotoxin producing species, and therefore, an epitype is designated here to provide taxonomic stability for this species.

poincianae *Fusarium* Pass., *Atti Reale Accad. Lincei, Rendiconti Cl. Sci. Fis., sér. 4*, 4: 105. 1888.

Holotypus: Not located.

Type locality: **Italy**, Parma.

Type substrate: *Poinciana gilliesii*.

Note: Not *Fusarium* fide [Wollenweber & Reinking \(1935\)](#).

polymorphum *Fusarium* Matr., *Rech. Dével. Mucéd.*: 84. 1892.

(See ***Fusarium sambucinum***)

Lectotypus (*hic designatus*, MBT 10000724): **France**, horse dung, 1892, L. Matruchot, in *Rech. Dével. Mucéd.*, Pl. 7, figs 6–14.

Notes: Synonym fide [Wollenweber & Reinking \(1935\)](#). As no holotype specimen could be located, an illustration accompanying the original protologue is designated here as lectotype.

polymorphum *Fusarium* Marchal, Bull. Soc. Roy. Bot. Belgique 34: 145. 1895, *nom. illegit.*, Art. 53.1.

(See *Fusarium aderholdii*)

Authentic material: Not located.

Original locality: **Belgium**, Brussels.

Original substrate: *Homo sapiens*.

Notes: Synonym *fide* Wollenweber & Reinking (1935).

polyphialidicum *Fusarium* Marasas *et al.*, Mycologia 78: 678. 1986.

(See *Fusarium concolor*)

Holotypus: DAOM 192986.

Ex-type culture: ATCC 60096 = CBS 961.87 = DAR 52851 = FRC M-2405 = MRC 3389 = NRRL 13459.

Type locality: **South Africa**, Mpumalanga Province, Nelspruit.

Type substrate: Plant debris in soil.

Descriptions and illustrations: See Marasas *et al.* (1986).

Diagnostic DNA barcodes: *rpb1*: JX171455; *rpb2*: JX171569; *tef1*: MH742681.

poncetii *Fusarium* Guiart (as '*ponceti*'), Compt.-Rend. Séances Mém. Soc. Biol. 73: 271. 1912, *nom. inval.*, Art. 36.1(a).

Authentic material: Not located.

Original locality: **?France**.

Original substrate: *Homo sapiens* granuloma teleangiectaticum.

Notes: Status unclear. Not treated by any of Wollenweber & Reinking (1935), Booth (1971), or Gerlach & Nirenberg (1982).

poolense *Fusarium* (as '*poolensis*') Taubenh., Bull. Texas Agric. Exp. Sta. 260: 27. 1920.

(See *Fusarium oxysporum*)

Lectotypus (*hic designatus*, MBT 10000725): **USA**, *Citrullus lanatus*, 1920, J.J. Taubenh., in Bull. Texas Agric. Exp. Sta. 260: 30, fig. 8i.

Notes: Synonym *fide* Wollenweber & Reinking (1935). As no holotype specimen could be located, an illustration accompanying the original protologue is designated here as lectotype.

praegraminearum *Fusarium* Gräfenhan & O'Donnell, Mycologia 108: 1232. 2016.

Holotypus: PDD 47563.

Ex-type culture: CBS 141369 = ICMP 8996 = NRRL 39664.

Type locality: **New Zealand**, North Island, Levin (near Wellington).

Type substrate: Litter in maize paddock.

Descriptions and illustrations: See Gräfenhan *et al.* (2016).

Diagnostic DNA barcodes: *rpb1*: KX260125; *rpb2*: KX260126; *tef1*: KX260120.

prieskaense *Fusarium* G.J. Marais & Sand.-Den., Stud. Mycol. 98 (no. 100116): 50. 2021.

Holotypus: CBS H-24660.

Ex-type culture: CAMS 001176 = CBS 146498 = CPC 30826.

Type locality: **South Africa**, Northern Cape Province, Prieska.

Type substrate: *Prunus spinosa*.

Descriptions and illustrations: See this study.

Diagnostic DNA barcodes: *rpb1*: MW834190; *rpb2*: MW834007; *tef1*: MW834275.

proliferatum *Fusarium* (Matsush.) Nirenberg ex Gerlach & Nirenberg, Mitt. Biol. Bundesanst. Land- Forstw. 209: 309. 1982.

Basionym: *Cephalosporium proliferatum* Matsush., Microfungi of the Solomon Islands and Papua-New Guinea: 11. 1971.

Synonyms: *Fusarium proliferatum* (Matsush.) Nirenberg, Mitt. Biol. Bundesanst. Land- Forstw. 169: 38. 1976, *nom. inval.*, Art. 41.3.

Fusarium proliferatum var. *minus* Nirenberg, Mitt. Biol. Bundesanst. Land- Forstw. 169: 43. 1976. *nom. inval.*, Art. 41.3.

Lectotypus: Microfungi of the Solomon Islands and Papua-New Guinea: 11, fig 121.2, designated by Yilmaz *et al.* (2021).

Lectotype locality: **Papua New Guinea**.

Lectotype substrate: Forest soil.

Epitypus: CBS 480.96 (preserved as metabolically inactive culture), designated by Yilmaz *et al.* (2021).

Epitype locality: **Papua New Guinea**, Morobe Province, Bulolo.

Epitype substrate: Forest soil.

Ex-epitype culture: CBS 480.96 = IAM 14682 = NRRL 26427 = NY007.B6.

Descriptions and illustrations: See Matsushima (1971), Yilmaz *et al.* (2021).

Diagnostic DNA barcodes: *rpb2*: MN534272; *tef1*: MN534059.

protoensiforme *Fusarium* (Sand.-Den. & Crous) O'Donnell *et al.*, Index Fungorum 440: 3. 2020.

Neocosmospora protoensiformis Sand.-Den. & Crous, Persoonia 43: 156. 2019.

Holotypus: CBS H-23995.

Ex-type culture: CBS 145471 = G.J.S. 90-168 = NRRL 22178.

Type locality: **Venezuela**.

Type substrate: Bark of dicot tree.

Descriptions and illustrations: See Sandoval-Denis *et al.* (2019).

Diagnostic DNA barcodes: *rpb1*: MW834244; *rpb2*: EU329498; *tef1*: AF178334.

protractum *Fusarium* Lév., Ann. Sci. Nat., Bot., sér. 3, 9: 246. 1848.

(See *Fusarium lateritium*)

Holotypus: ?PC.

Type locality: **France**, Romainville.

Type substrate: Dead shoots of *Solanum dulcamara*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

prunorum *Fusarium* McAlpine, Fungus Diseases of stone-fruit trees in Australia: 91. 1902.

(See *Fusarium candidum* (Link) Sacc.)

Lectotypus (*hic designatus*, MBT 10000726): **Australia**, Victoria, Melbourne, Burnley, from shriveled and blackened apricot fruit, Jun. 1900, D. McAlpine, in Fungus Diseases of stone-fruit trees in Australia (1902), pl. XX, fig. 42.

Notes: Synonym *fide* Wollenweber & Reinking (1935). As no holotype specimen could be located, an illustration accompanying the original protologue is designated here as lectotype.

pseudacaciae *Fusarium* Rapaics, Z. Pflanzenkrankh. 25: 208. 1915.

(See *Fusarium lateritium*)

Holotypus: Not located.

Type locality: **Hungary**, Debrecen.

Type substrate: *Robinia pseudoacaciae*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

pseudensiforme *Fusarium* Samuels *et al.*, Mycologia 103: 1323. 2011.

Neocosmospora pseudensiformis Samuels *et al.*, Mycologia 103: 1323. 2011.

Holotypus: BPI 881226.

Ex-type culture: CBS 125729 = FRC S-1834 = G.J.S 02-95 = G.J.S 9318 = NRRL 46517.

Type locality: **Sri Lanka**, Wagamba, Kurunegala, Arangakele.

Type substrate: Bark of tree.

Descriptions and illustrations: See [Nalim et al. \(2011\)](#).
Diagnostic DNA barcodes: *rpb1*: KC691615; *rpb2*: KC691645;
tef1: KC691555.

pseudoanthophilum Fusarium Nirenberg et al., *Mycologia* 90: 461. 1998.

Holotypus: In B.

Ex-type culture: BBA 69002 = CBS 414.97 = IMI 376112 = NRRL 25211.

Type locality: **Zimbabwe**, Gambiza.

Type substrate: *Zea mays*.

Descriptions and illustrations: See [Nirenberg et al. \(1998\)](#).

Diagnostic DNA barcodes: *rpb1*: MT010949; *rpb2*: MT010980;
tef1: MK639073.

pseudocircinatum Fusarium O'Donnell & Nirenberg, *Mycologia* 90: 448. 1998.

Holotypus: B 70 0001689.

Ex-type culture: BBA 69636 = CBS 126.73 = CBS 449.97 = DAOM 225117 = IMI 375316 = NRRL 22946.

Type locality: **Ghana**.

Type substrate: *Solanum* sp.

Descriptions and illustrations: See [Nirenberg & O'Donnell \(1998\)](#).

Diagnostic DNA barcodes: *rpb1*: MG838070; *rpb2*: MN724939;
tef1: MG838023.

pseudoeffusum Fusarium Murashk., *Proc. Siberian Agric. Acad. Omsk* 3: 106. 1924.

(See ***Fusarium acuminatum***)

Holotypus: Not located.

Type locality: **Russia**, Siberia.

Type substrate: *Triticum polonicum*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

pseudograminearum Fusarium O'Donnell & T. Aoki, *Mycologia* 91: 604. 1999.

Holotypus: BPI 746087.

Ex-type culture: FRC R-5291 = NRRL 28062.

Type locality: **Australia**, New South Wales, Young.

Type substrate: *Hordeum vulgare*.

Descriptions and illustrations: See [Aoki & O'Donnell \(1999\)](#).

Diagnostic DNA barcodes: *rpb1*: JX171524; *rpb2*: JX171637;
tef1: AF212468.

pseudoheterosporum Fusarium Jacz., *Bull. Soc. Mycol. France* 28: 347. 1912.

(See ***Fusarium avenaceum***)

Holotypus: Not located.

Type locality: **France**.

Type substrate: *Lolium* sp. and *Triticum* sp.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

pseudonectria Fusarium Sp., *Anales Mus. Nac. Hist. Nat. Buenos Aires* 6: 351. 1898 [1899].

(See ***Fusarium avenaceum***)

Holotypus: In LPS (Fungi Argent. n.v.c. no. 867) *fide* [Farr \(1973\)](#).

Type locality: **Ecuador**, San Salvador Island.

Type substrate: Dead culms of *Poaceae*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

pseudonygamai Fusarium O'Donnell & Nirenberg, *Mycologia* 90: 449. 1998.

Holotypus: B 70 0001688.

Ex-type culture: BBA 69552 = CBS 417.97 = DAOM 225136 = FRC M-1166 = IMI 375342 = NRRL 13592.

Type locality: **Nigeria**.

Type substrate: *Pennisetum typhoides*.

Descriptions and illustrations: See [Nirenberg & O'Donnell \(1998\)](#).

Diagnostic DNA barcodes: *rpb1*: LT996205; *rpb2*: LT996152;
tef1: AF160263.

pseudoradicicola Fusarium (Sand.-Den. & Crous) O'Donnell et al., *Index Fungorum* 440: 3. 2020.

Neocosmospora pseudoradicicola Sand.-Den. & Crous, *Persoonia* 43: 157. 2019.

Holotypus: CBS H-23996.

Ex-type culture: ARSEF 2313 = CBS 145472 = NRRL 25137.

Type locality: **Papua New Guinea**, East New Britain, Keravat, Lowlands Agricultural Experiment Station.

Type substrate: Diseased pods of *Theobroma cacao*.

Descriptions and illustrations: See [Sandoval-Denis et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW218133; *rpb2*: JF741084;
tef1: JF740757.

pseudotonkinense Fusarium (Sand.-Den. & Crous) O'Donnell et al., *Index Fungorum* 440: 3. 2020.

Neocosmospora pseudotonkinensis Sand.-Den. & Crous, *Persoonia* 43: 159. 2019.

Holotypus: CBS H-23997.

Ex-type culture: CBS 143038.

Type locality: **Netherlands**, Zuid-Holland Province, Leiden.

Type substrate: Cornea of *Homo sapiens*.

Descriptions and illustrations: See [Sandoval-Denis et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb2*: LR583867; *tef1*: LR583640.

pteridis Fusarium Ellis & Everh., *Proc. Acad. Nat. Sci. Philadelphia* 45: 466. 1894.

Septogloeum pteridis (Ellis & Everh.) Wollenw., *Fusaria Autogr. Delin.* 1: 446. 1916.

Syntypes: In BPI, BRU, CUP, FLAS, ILL, ISC, MICH, MSC, MU, NEB, NY, OSC, PH, PUL, WIS & WSP.

Type locality: **USA**, New Jersey, Gloucester, Newfield.

Type substrate: *Phyllachora flabella* on *Pteris aquilina*.

pucciniophilum Fusarium Sacc. & P. Syd., *Syll. Fung.* 14: 1128. 1899.

Replaced synonym: *Fusarium parasiticum* Ellis & Kellerm., *J. Mycol.* 3 (11): 127. 1887, *nom. illegit.*, Art. 53.1.

(See ***Fusarium heterosporum***)

Holotypus: Kellerman & Swingle no. 1104 in NY.

Type locality: **USA**, Kansas, Manhattan.

Type substrate: Parasitic on *Puccinia seymeriae* on leaves of *Solidago macrophylla*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

pulvinatum Fusarium (Link) Nees, *Syst. Pilze*: 32. 1817.

Basionym: *Atractium pulvinatum* Link, *Mag. Ges. Naturf. Freunde Berlin* 8: 32. 1816.

Holotypus: Not located.

Type locality: **Poland**, Wrocław.

Type substrate: Hanging scrub branches.

Notes: Status unclear. Not treated by any of [Wollenweber & Reinking \(1935\)](#), [Booth \(1971\)](#), or [Gerlach & Nirenberg \(1982\)](#).

pulvinatum Fusarium (Berk. & Broome) Sacc., *Syll. Fung.* 4: 699. 1886, *nom. illegit.*, Art. 53.1.

Basionym: *Fusisporium pulvinatum* Berk. & Broome, *J. Linn. Soc., Bot.* 14: 102. 1873 [1875].

(See ***Fusarium sambucinum***)

Holotypus: In K(M).

Type locality: **Sri Lanka**.

Type substrate: Bark.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

punctiforme *Fusarium* Durieu & Mont., Expl. Sci. Algérie 1: 335. 1848.

(See ***Fusarium reticulatum***)

Holotypus: Not located.

Type locality: **Algeria**.

Type substrate: *Citrus aurantium*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

purpurascens *Fusarium* Maryani *et al.*, Stud. Mycol. 92: 160. 2018 [2019].

(see ***Fusarium odoratissimum***)

Holotypus: InaCC F886 (preserved as metabolically inactive culture).

Ex-type culture: InaCC F886.

Type locality: **Indonesia**, East Kalimantan, Kampung Salak Martadinata.

Type substrate: *Musa* var. Pisang Kepok.

Descriptions and illustrations: See [Maryani *et al.* \(2019a\)](#).

Diagnostic DNA barcodes: *rpb2*: LS479385; *tef1*: LS479827.

pusillum *Fusarium* Wollenw., Fusaria Autogr. Delin. 2: 550. 1924. (See *Fusarium dimerum*)

Lectotypus (*hic designatus*, MBT 10000727): **Germany**, *Solanum tuberosum*, 1919, H.W. Wollenweber, in Fusaria Autogr. Delin. 2: 550. 1924.

Note: As no holotype specimen could be located, an illustration accompanying the original protologue is designated here as lectotype.

putaminum *Fusarium* (Thüm.) Sacc., Syll. Fung. 4: 703. 1886. Basionym: *Fusisporium putaminum* Thüm., Oesterr. Bot. Z. 27: 272. 1877.

(See ***Fusarium lateritium***)

Holotypus: Not located.

Type locality: **Austria**, Klosterneuburg.

Type substrate: *Prunus domestica*.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

putrefaciens *Fusarium* Osterw., Mitth. Thurgauischen Naturf. Ges. 16: 123. 1904.

(See ***Fusarium avenaceum***)

Lectotypus (*hic designatus*, MBT 10000728): **Switzerland**, Zürich, fruit and seeds of *Pyrus* sp., 1904, collector unknown, in Osterwalder, Mitth. Thurgauischen Naturf. Ges. 16, tab. 2, figs 10–30.

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#). As no holotype specimen could be located, an illustration accompanying the original protologue is designated here as lectotype.

pyrinum *Fusarium* Schwein., Trans. Amer. Philos. Soc., n.s. 4: 302. 1834, unavailable, see Art. F.3.4.

(See ***Fusarium lactis***)

pyrinum *Fusarium* (Fr.) Sacc., Syll. Fung. 4: 720. 1886.

Basionym: *Fusisporium pyrinum* Fr., Syst. Mycol. 3: 445. 1832, *nom. sanct.*

(See ***Fusarium avenaceum***)

Holotypus: Not located.

Type locality: **Sweden**.

Type substrate: Rotten fruit of *Pyrus communis*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

pyrochroum *Fusarium* (Desm.) Sacc., Michelia 1: 534. 1879.

Calonectria pyrochroa (Desm.) Sacc., Michelia 1: 308. 1878.

Basionym: *Selenosporium pyrochroum* Desm., Ann. Sci. Nat., Bot., sér. 3, 14: 111. 1850.

Synonyms: *Nectria pyrochroa* Desm., Pl. Crypt. N. France, ed. 2: no. 372. 1856.

Calonectria daldiniana De Not., Comment. Soc. Crittog. Ital. 2: 477. 1867.

Fusarium pyrochroum var. *diatrypellicola* P. Syd., Mycoth. March., Cent. 41: no. 4063. 1893.

Nectria abnormis Henn., Hedwigia 36: 219. 1897.

Holotypus: In ?PAD or PC.

Type locality: **France**.

Type substrate: *Sambucus nigra*.

quercicola *Fusarium* Oudem., Ned. Kruidk. Arch., sér. 3, 2: 777. 1902.

Holotypus: ?L.

Type locality: **Netherlands**, Noord-Holland Province, Bussum.

Type substrate: *Quercus rubra*.

Note: Not *Fusarium* *fide* [Wollenweber & Reinking \(1935\)](#).

quercinum *Fusarium* O'Donnell *et al.*, Index Fungorum 440: 4. 2020.

Neocosmospora quercicola Sand.-Den. & Crous, Persoonia 43: 159. 2019.

Holotypus: CBS H-23998.

Ex-type culture: CBS 141.90 = NRRL 22652.

Type locality: **Italy**.

Type substrate: *Quercus cerris*.

Descriptions and illustrations: See [Sandoval-Denis *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW834247; *rpb2*: LR583869; *tef1*: DQ247634.

radicicola *Fusarium* Wollenw., J. Agric. Res. 2: 257. 1914.

(See *Fusarium solani*)

Lectotypus: Plate XVI, fig. K, in [Wollenweber \(1914\)](#), designated in [Sandoval-Denis *et al.* \(2019\)](#).

Lectotype locality: **USA**, Washington.

Lectotype substrate: *Solanum tuberosum*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#) & [Sandoval-Denis *et al.* \(2019\)](#).

ramigenum *Fusarium* O'Donnell & Nirenberg, Mycologia 90: 451. 1998.

Holotypus: B 70 0001687.

Ex-type culture: BBA 68592 = CBS 418.97 = DAOM 225137 = IMI 375343 = NRRL 25208.

Type locality: **USA**, California.

Type substrate: *Ficus carica*.

Descriptions and illustrations: See [Nirenberg & O'Donnell \(1998\)](#).

Diagnostic DNA barcodes: *rpb1*: KF466401; *rpb2*: KF466412; *tef1*: AF160267.

ramosum *Fusarium* (Batista & H. Maia) O'Donnell *et al.*, Index Fungorum 440: 4. 2020.

Basionym: *Hyaloflorea ramosa* Bat. & H. Maia, Anais Soc. Biol. Pernambuco 13: 155. 1955.

Synonyms: *Neocosmospora ramosa* (Bat. & H. Maia) L. Lombard & Crous, Stud. Mycol. 80: 227. 2015.

(See *Fusarium lichenicola* C. Massal.)

Holotypus: IMUR 410.

Ex-type culture: CBS 509.63 = IMUR 410 = MUCL 8050.

Type locality: **Brazil**.

Type substrate: Air.

Diagnostic DNA barcodes: *rpb2*: LR583843; *tef1*: LR583618.

Note: Synonymies *vide* Sandoval-Denis & Crous (2018).

ramulicola *Fusarium* Sawada, Special Publ. Coll. Agric. Natl. Taiwan Univ. 8: 228. 1959, *nom. inval.*, Art. 39.1.

Authentic material: Not located.

Original locality: **Taiwan**.

Original substrate: Branches of *Citrus tankan f. koshotankan*.

Note: This name is invalid because of missing Latin diagnosis.

rectiphorum *Fusarium* Samuels *et al.* (as '*rectiphorus*'), Mycologia 103: 1324. 2011.

Neocosmospora rectiphora Samuels *et al.*, Mycologia 103: 1324. 2011.

Neocosmospora bomiensis Z.Q. Zeng & W.Y. Zhuang, Phytotaxa 319: 177. 2017.

Holotypus: BPI 881229.

Ex-type culture: CBS 125727 = FRC S-1831 = G.J.S. 02-89.

Type locality: **Sri Lanka**, Wagamba Province, vic. Kurunegala, Arangakele.

Type substrate: Bark.

Descriptions and illustrations: See Nalim *et al.* (2011).

Diagnostic DNA barcodes: *rpb1*: MW834249; *rpb2*: LR583871; *tef1*: LR583641.

redolens* *Fusarium Wollenw., Phytopathology 3: 29. 1913 and Ber. Deutsch. Bot. Ges. 31: 31. 1913.

Synonyms: *Fusarium oxysporum* var. *redolens* (Wollenw.) W.L. Gordon, Canad. J. Bot. 30: 238. 1952.

Fusarium solani var. *redolens* (Wollenw.) Bilaĭ, Fusarii (Biologija i sistematika): 288. 1955.

?*Fusarium retusum* Wellman, Phytopathology 33: 957. 1943.

Holotypus: Not located.

Type locality: **Unknown**.

Type substrate: *Pisum sativum*.

Lectotypus (*hic designatus*, MBT 10000729): **Unknown**, *Pisum sativum*, 1913, H.W. Wollenweber, in Phytopathology 3: 31, fig. E.

Epitypus (*hic designatus*, MBT 10000730): **Germany**, Berlin-Dahlem, vascular bundle of *Dianthus caryophyllus*, 16 May 1959, D. Hantschke & W. Gerlach, CBS 360.87 (preserved as metabolically inactive culture).

Ex-epitype culture: ATCC 16067 = BBA 9526 = CBS 248.61 = CBS 360.87 = DSM 62390 = NRRL 20426 = NRRL 25600.

Descriptions and illustrations: See Gerlach & Pag (1961), Gerlach & Nirenberg (1982) and Leslie & Summerell (2006).

Diagnostic DNA barcodes: *rpb1*: MT409433; *rpb2*: MT409443; *tef1*: MT409453.

Notes: As both protologue publications occurred more or less simultaneously for *F. redolens*, we select the illustration provided in Phytopathology as lectotype, since no holotype material could be located. Gerlach & Nirenberg (1983) considered CBS 248.61 (= CBS 360.87) a good representative of *F. redolens*, which was initially designated by Gerlach & Pag (1961) as representative of *F. redolens f. sp. dianthi*. Therefore, an epitype is designated here to provide taxonomic stability for this species.

regulare *Fusarium* (Sand.-Den. & Crous) O'Donnell *et al.*, Index Fungorum 440: 4. 2020.

Neocosmospora regularis Sand.-Den. & Crous, Persoonia 43: 162. 2019.

Holotypus: CBS H-23999.

Ex-type culture: CBS 230.34

Type locality: **Netherlands**, Zeeland Province, Zuid Beveland, near Kloetinge.

Type substrate: *Pisum sativum*.

Descriptions and illustrations: See Sandoval-Denis *et al.* (2019).

Diagnostic DNA barcodes: *rpb2*: LR583873; *tef1*: LR583643.

rekanum *Fusarium* Lynn & Marinc., Antonie van Leeuwenhoek 113: 816. 2020.

Neocosmospora rekana (Lynn & Marinc.) L. Lombard & Sand.-Den., *comb. nov.* MycoBank MB 837706.

Basionym: *Fusarium rekanum* Lynn & Marinc., Antonie van Leeuwenhoek 113: 816. 2020.

Holotypus: PREM 62333.

Ex-type culture: CMW 52862 = PPRI 27163.

Type locality: **Indonesia**, Sumatra, Riau, Pelalawan.

Type substrate: *Acacia crassicaarpa* infested with *Euwallacea perbrevis*.

Descriptions and illustrations: See Lynn *et al.* (2020).

Diagnostic DNA barcodes: *rpb2*: MN249137, MN249108; *tef1*: MN249151.

Note: Based on the phylogenetic position of this species related to the 'ambrosia' clade as illustrated by Lynn *et al.* (2020), we provide a new combination in the genus *Neocosmospora*.

reticulatum* *Fusarium Mont., Ann. Sci. Nat., Bot., sér. 2, 20: 379. 1843.

Synonyms: ?*Fusarium leucoconium* Corda, Icon. Fung. 1: 4. 1837. (*vide* Wollenweber & Reinking 1935).

?*Fusarium punctiforme* Durieu & Mont., Expl. Sci. Algérie 1: 335. 1848.

Fusisporium flavidum Bonord., Bot. Zeitung (Berlin) 19: 194. 1861.

Fusarium flavidum (Bonord.) Sacc., Syll. Fung. 4: 698. 1886.

Fusarium ampelodesmi Fautrey & Roum., in Roumeguère, Rev. Mycol. (Toulouse) 13: 82. 1891.

Fusarium epithele McAlpine, Fungus Diseases of Citrus trees in Australia: 80. 1899.

Fusarium orchidis Petch, Ann. Roy. Bot. Gard. (Peradeniya) 6: 256. 1917.

Fusarium negundinis Sherb., in Hubert, J. Agric. Res. 26: 451. 1923.

Fusarium reticulatum var. *negundinis* (Sherb.) Wollenw., Z. Parasitenk. (Berlin) 3: 351. 1931.

Fusarium heterosporum var. *negundinis* (Sherb.) Raillo, Fungi of the Genus *Fusarium*: 217. 1950.

Fusarium reticulatum var. *medium* Wollenw., Z. Parasitenk. (Berlin) 3: 358. 1931.

Lectotypus (*hic designatus*, MBT 10000731): **France**, Nouvelle-Aquitaine, Saint-Sever, *Citrullus* sp., 1843, L. Dufour, in Montagne, Ann. Sci. Nat., Bot., 2 sér. 20: 379: pl. 16, fig. 3.

Epitypus (*hic designatus*, MBT 10000732): **Germany**, Rellingen/Holstein, bark lesion of *Sophora japonica*, Jun. 1976, R. Schwarz, CBS 473.76 (preserved as metabolically inactive).

Ex-epitype culture: BBA 63657 = CBS 473.76 = NRRL 20684.

Descriptions and illustrations: See Gerlach & Nirenberg (1982).

Diagnostic DNA barcodes: *rpb1*: MW928816; *tef1*: MW928841.

Notes: Gerlach & Nirenberg (1983) considered CBS 473.76 a good representative of *F. reticulatum*. As no holotype specimen could be located, an illustration is designated as lectotype here and an epitype is designated to provide taxonomic stability for this species.

retusum *Fusarium* Wellman, Phytopathology 33: 957. 1943.

(See *Fusarium oxysporum*)

Holotypus: Not located.

Type locality: USA, Indiana.

Type substrate: *Solanum lycopersicum*.

rhabdophorum *Fusarium* Berk. & Broome, Ann. Mag. Nat. Hist., ser. 4, 17: 142. 1876.

Holotypus: In K(M).

Type locality: UK, Scotland, Forres.

Type substrate: Dead sticks.

Notes: Status unclear. Not *Fusarium* fide Wollenweber & Reinking (1935).

rhizochromatistes *Fusarium* Sideris, Phytopathology 14: 212. 1924.

(See *Fusarium oxysporum*)

Lectotypus (*hic designatus*, MBT 10000733): USA, California, Stockton, roots of *Allium cepa*, 1924, C.P. Sideris, in Phytopathology 14, pl. XI.

Notes: Synonym fide Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration was designated as lectotype.

rhizogenum *Fusarium* Pound & Clem., Bot. Surv. Nebraska 3: 12. 1894.

(See *Fusarium candidum* Ehrenb.)

Holotypus: NEB0040548.

Type locality: USA, Lincoln.

Type substrate: Roots of *Malus domestica* seedlings.

Note: Synonym fide Wollenweber & Reinking (1935).

rhizogenum *Fusarium* Aderh., Centralbl. Bacteriol. Parasitenk., 1. Abth., 6: 623. 1900, *nom. illegit.*, Art. 53.1.

(See *Fusarium aderholdii*)

Authentic material: Not located.

Original locality: Germany.

Original substrate: *Malus domestica*.

Notes: Synonym fide Wollenweber & Reinking (1935). The original publication could not be checked but Sorauer (1923) clearly stated that Aderhold only used the name *Fusarium rhizogenum* Pound & Clem. to describe a disease using the latter name.

rhizophilum *Fusarium* Corda, Icon. Fung. 2: 3. 1838.

Synonym: *Fusisporium georginae* Klotzsch, Herb. Viv. Mycol., Cent. 2: 186. 1832, *nom. nud.*, Art. 38.1(a).

(See *Fusarium merismoides*)

Lectotypus (*hic designatus*, MBT 10000734): Czech Republic, Prague, roots of garden plants, 1838, A.C.J. Corda, in Icon. Fung. 2, Tab. VIII, fig. 15.

Notes: Synonym fide Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

rhizophorae *Fusarium* (Dayar.) O'Donnell et al., Index Fungorum 440: 4. 2020.

Neocosmospora rhizophorae Dayar., Mycosphere 11: 112. 2020.

Holotypus: MFLU 17-2588.

Ex-type culture: MFLUCC 17-2461.

Type locality: Thailand, Krabi Province, Phang Nga.

Type substrate: Submerged wood of *Rhizophora*.

Descriptions and illustrations: See Dayarathne et al. (2020)

rhodellum *Fusarium* McAlpine, Proc. Linn. Soc. New South Wales 24: 122. 1899.

Lectotypus (*hic designatus*, MBT 10000735): Kerguelen Islands, *Pringlea antiscorbutica*, 1899, D. McAlpine, in Proc. Linn. Soc. New South Wales 24: Pl. XIII, Fig. 7.

Notes: Not *Fusarium* fide I. Pascoe. No holotype specimen could be located and therefore an illustration is designated as lectotype.

rhoicola *Fusarium* Fautrey, Rev. Mycol. (Toulouse) 17: 171. 1895.

(See *Fusarium graminearum*)

Holotypus: ?PC.

Type locality: France via USA.

Type substrate: *Rhus toxicodendron*.

Note: Synonym fide Wollenweber & Reinking (1935).

ricini *Fusarium* (Bérenger) Bizz., Fl. Ven. Critt. 1: 539. 1885.

Basionym: *Fusisporium ricini* Bérenger, Mem. Accad. Agric. Verona 44: 257. 1866.

(See *Fusarium sambucinum*)

Holotypus: Not located.

Type locality: Italy.

Type substrate: *Ricinus communis*.

Note: Synonym fide Wollenweber & Reinking (1935).

rigidiusculum *Fusarium* (Berk. & Broome) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 32: 664. 1945.

Basionym: *Nectria rigidiuscula* Berk. & Broome, J. Linn. Soc., Bot. 14: 116. 1873 [1875].

(See *Fusarium colorans*)

Holotypus: ?K(M).

Type locality: ?Sri Lanka.

Type substrate: Bark.

Note: Synonym fide Wollenweber & Reinking (1935).

rimicola *Fusarium* Sacc. (as '*rimicolum*'), Michelia 2: 297. 1881.

(See *Fusarium lateritium*)

Holotypus: Not located.

Type locality: Italy, Padua.

Type substrate: *Erythrina crista-galli*.

Note: Synonym fide Wollenweber & Reinking (1935).

rimosum *Fusarium* (Peck) Sacc., Syll. Fung. 4: 713. 1886.

Basionym: *Fusisporium rimosum* Peck, Rep. (Annual) New York State Mus. Nat. Hist. 30: 58. 1878.

(See *Fusarium merismoides*)

Holotypus: NYSf2609.

Type locality: USA, New York, Albany.

Type substrate: Cut ends of stalks of *Zea mays*.

Note: Synonym fide Wollenweber & Reinking (1935).

riograndense *Fusarium* Dallé Rosa et al., J. Mycol. Med. 28: 33. 2018.

Neocosmospora riograndensis (Dallé Rosa et al.) Sand.-Den. & Crous, Persoonia 43: 165. 2019.

Holotypus: UFMG-CM F12570.

Ex-type culture: UFMG-CM F12570 = URM-7361.

Type locality: Brazil, Rio Grande do Sul, Porto Alegre, Hospital de Clínicas de Porto Alegre.

Type substrate: Nasal cavity of *Homo sapiens*.

Descriptions and illustrations: See Dallé Rosa et al. (2018).

Diagnostic DNA barcodes: *rpb2*: KX534003; *tef1*: KX534002.

robiniae *Fusarium* Pass., Atti Reale Accad. Lincei, Rendiconti Cl. Sci. Fis., sér. 4, 7: 51. 1891.

(See *Fusarium sarcochroum*)

Holotypus: ?PARMA.

Type locality: Italy, Padua.

Type substrate: *Robinia pseudoacacia*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

robustum* *Fusarium Gerlach, Phytopathol. Z. 88: 36. 1977.

Holotypus: In B.

Isotypus: CBS H-629.

Ex-type culture: BBA 63667 = CBS 637.76 = FRC R-5821 = IMI 322102 = NRRL 13392.

Type locality: Argentina.

Type substrate: *Araucaria angustifolia*.

Descriptions and illustrations: See Gerlach (1977c).

Diagnostic DNA barcodes: *rpb2*: MW928831; *tef1*: MW928842.

roesleri *Fusarium* Thüm., Pilze Weinst.: 51. 1878.

(See *Fusarium merismoides*)

Lectotypus (*hic designatus*, MBT 10000736): Austria, Klosterneuburg, *Vitis vinifera*, 1878, K.A.E.J. Thümen, in Pilze Weinst. Tab. 3, fig. 7.

Notes: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

rollandianum *Fusarium* Sacc., Syll. Fung. 11: 650. 1895.

Replaced synonym: *Fusarium cydoniae* Roum. & Fautrey, Rev. Mycol. (Toulouse) 14: 170. 1892, *nom. illegit.*, Art. 53.1, *non* Allescher 1892.

Syntype: ILL00220295 (Fautrey, Fungi Sel. Gall. Exs. No. 6120).

Type locality: France.

Type substrate: Fruit of *Cydonia vulgaris*.

Notes: Not *Fusarium* *fide* Wollenweber & Reinking (1935). Typification pending further study of the syntype lodged in ILL.

rosae *Fusarium* (Preuss) Sacc., Syll. Fung. 4: 697. 1886.

Basionym: *Selenosporium rosae* Preuss, Linnaea 24: 150. 1851.

Holotypus: Not located; not preserved in B *fide* Holubová-Jechová *et al.* (1994).

Type locality: Germany, Hoyerswerda.

Type substrate: *Rosa* sp.

Notes: Status unclear. Not treated by any of Wollenweber & Reinking (1935), Booth (1971), or Gerlach & Nirenberg (1982).

roseobullatum *Fusarium* Wollenw. (as '*roseo-bullatum*'), Fusaria Autogr. Delin. 1: 117. 1916.

Basionym: *Fusarium bullatum* var. *roseum* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 201. 1915.

(See *Fusarium equiseti*)

Holotypus: ?CUP-007433.

Type locality: USA, Iowa.

Type substrate: *Solanum tuberosum*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

roseolum *Fusarium* (H.O. Stephens ex Berk. & Broome) Sacc., Syll. Fung. 4: 710. 1886.

Basionym: *Fusisporium roseolum* H.O. Stephens ex Berk. & Broome, Ann. Mag. Nat. Hist., ser. 2, 7: 178. 1851.

(See *Fusarium merismoides*)

Holotypus: ?K(M).

Type locality: UK, Bristol.

Type substrate: Decayed *Solanum tuberosum*.

Note: Synonyms *fide* Wollenweber & Reinking (1935).

roseum *Fusarium* Link, Mag. Ges. Naturf. Freunde Berlin 3: 10. 1809, *nom. rej.*

(See *Fusarium sambucinum*)

Lectotypus: In B, selected in Gams *et al.* (1997).

Type locality: Germany.

Type substrate: Malvaceae.

Notes: Gams *et al.* (1997) proposed that the name, *F. roseum* be rejected due to ambiguity surrounding the type of this species, with *F. sambucinum* taking preference. This proposal was accepted in 1999 (see Gams 1999).

rostratum *Fusarium* Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land- Forstw. 8: 30. 1910 [1913].

(See *Fusarium graminearum*)

Lectotypus (*hic designatus*, MBT 10000737): Germany, Berlin, *Triticum aestivum*, 1913, O.A. Appel & H.W. Wollenweber, in Arbeiten Kaiserl. Biol. Anst. Land- Forstw. 8: 30, Abb. 1, figs E1–E13.

Notes: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

roumegueri *Fusarium* Sacc. (as '*roumegueri*'), Syll. Fung. 4: 702. 1886, *nom. illegit.*, Art. 52.1.

Replaced synonym: *Fusarium insidiosum* Roum., Michelia 2 (6): 132. 1880.

(See *Fusarium lateritium*)

Type material: See *Fusarium insidiosum*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

ruberrimum *Fusarium* Delacr., Bull. Soc. Mycol. France 6: 139. 1890.

(See *Fusarium avenaceum*)

Holotypus: ?PC.

Type locality: France, Paris.

Type substrate: *Onobrychis viciifolia*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

rubi *Fusarium* (G. Winter) Berl. & Voglino, Add. Syll. Fung. 1–4: 391. 1886.

Basionym: *Fusisporium rubi* G. Winter, in Rabenh., Fungi Eur. Extræur Exs., Ed. Nov., Ser. Sec., Cent. 13 (resp. 33): 3280. 1885.

Synonym: *Ramularia rubi* (G. Winter) Wollenw., Fusaria Autogr. Delin. 1: 470. 1916.

Cercospora rubi (G. Winter) Plakidas, J. Agricultural Research 54: 275. 1937.

Syntypes: In BPI, CHRB, CUP, F, HAL, ISC, LSUM, MSC, MU, NEB & PH (Fungi Eur. Extræur. Exs. no. 3280).

Type locality: USA, Illinois, Cobden

Type substrate: *Rubus villosus*

Note: Status unclear *fide* Braun (1998).

rubicolor *Fusarium* Berk. & Broome, Trans. Linn. Soc. London, Bot. 2: 68. 1883.

Holotypus: ?K(M).

Type locality: Australia, Queensland, Brisbane.

Type substrate: Leaves of *Eucalyptus* sp.

Note: Not *Fusarium* fide Wollenweber & Reinking (1935).

rubiginosum *Fusarium* Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land- Forstw. 8: 108. 1910 [1913].

(See *Fusarium culmorum*)

Lectotypus (*hic designatus*, MBT 10000738): **Germany**, *Solanum tuberosum*, 1913, O.A. Appel & H.W. Wollenweber, in Arbeiten Kaiserl. Biol. Anst. Land- Forstw. 8: Tab. I, figs 31–48. Notes: Synonym fide Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

rubrum *Fusarium* Parav., Ann. Mycol. 16: 311. 1918.

(See *Fusarium lactis*)

Lectotypus (*hic designatus*, MBT 10000739): **Germany**, core of *Malus domestica* fruit, 1918, E. Paravicini, in Ann. Mycol. 16, pl. 4, figs 23–33.

Notes: Synonym fide Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

rusci *Fusarium* (Sacc.) O'Donnell & Geiser, Phytopathology 103: 404. 2013.

Basionym: *Fusarium roseum* var. *rusci* Sacc., Michelia 2: 294. 1881.

Synonyms: *Trichofusarium rusci* (Sacc.) Bubák, Bull. Herb. Boissier, sér. 2, 6: 488. 1906.

Pycnofusarium rusci D. Hawksw. & Punith., Trans. Brit. Mycol. Soc. 61: 63. 1973.

Syntype: BPI 453152.

Type locality: **Italy**, Selva.

Type substrate: *Ruscus aculeatus*.

Notes: Examination of the syntype (BPI 453152) revealed that this species does not belong to the genus *Fusarium*, having a myrothecium-like morphology. Also see notes under *Nothofusarium devonianum*.

russianum *Fusarium* Manns, Bull. North Dakota Agric. Exp. Sta. 259: 34. 1932.

(See *Fusarium acuminatum*)

Holotypus: Not located.

Type locality: **USA**, North Dakota.

Type substrate: *Linum usitatissimum*.

Note: Synonym fide Wollenweber & Reinking (1935).

ruticola *Fusarium* Fautrey & Roum. (as '*rutaecola*'), Rev. Mycol. (Toulouse) 13: 82. 1891.

(See *Fusarium avenaceum*)

Syntype: ?PC (Fungi Sel. Gall. Exs. No. 5686).

Type locality: **France**, Noidan.

Type substrate: *Ruta graveolens*.

Note: Synonym fide Wollenweber & Reinking (1935).

saccardoanum *Fusarium* P. Syd., Syll. Fung. 14: 1128. 1899.

Replaced synonym: *Fusarium sclerodermatis* Peck, Rep. (Annual) Regents Univ. State New York New York State Mus. 43: 77. 1890, *nom. illegit.*, Art. 53.1, *non Fusarium sclerodermatis* Oudem. 1889.

(See *Fusarium oxysporum*)

Holotypus: NYSf2731.

Type locality: **USA**, New York, Suffolk, Manor, Long Island.

Type substrate: *Scleroderma vulgare*.

Note: Synonym fide Wollenweber & Reinking (1935).

sacchari *Fusarium* (E.J. Butler) W. Gams, Cephalosporium-artige Schimmelpilze: 218. 1971.

Basionym: *Cephalosporium sacchari* E.J. Butler, Mem. Dept. Agric. India, Bot. Ser. 6: 185. 1913.

Synonyms: *Fusarium neoceras* Wollenw. & Reinking, Phytopathology 15: 164. 1925.

Gibberella sacchari Summerell & J.F. Leslie, Mycologia 97: 719. 2005, *nom. illegit.*, Art. 53.1, *non Gibberella sacchari* Speg. 1896.

Fusarium desaboruense N. Maryani *et al.*, Persoonia 43: 59. 2019.

Lectotypus: In Mem. Dept. Agric. India, Bot. Ser. 6: 185, pl. II, figs 1–13. 1913, designated by Yilmaz *et al.* (2021).

Epitypus: CBS 223.76 (preserved as metabolically inactive culture), designated by Yilmaz *et al.* (2021).

Ex-epitype culture: BBA 63340 = CBS 223.76 = DAOM 225138 = IMI 202881 = NRRL 13999.

Lectotype and epitype locality: **India**.

Lectotype and epitype substrate: *Saccharum officinarum*.

Descriptions and illustrations: See Butler & Khan (1913), Gams (1971), Gerlach & Nirenberg (1982), Leslie *et al.* (2005) and Leslie & Summerell (2006).

Diagnostic DNA barcodes: *rpb1*: JX171466; *rpb2*: JX171580; *tef1*: AF160278.

salicicola *Fusarium* Allesch. (as '*salicolum*'), Ber. Bayer. Bot. Ges. 4: 39. 1896.

(See *Fusarium avenaceum*)

Holotypus: In M.

Type locality: **Germany**, München, forest near Großhesselohe.

Type substrate: Dead branch of *Salix caprea*.

Note: Synonym fide Wollenweber & Reinking (1935).

salicinum *Fusarium* Corda, Icon. Fung. 3: 33. 1839.

Typus: In PRM fide Pilat (1938).

Type locality: **Czech Republic**, near Prague.

Type substrate: Thin branches of *Salix* sp.

Notes: Not *Fusarium* fide Wollenweber & Reinking (1935). Lectotypification pending study of material lodged in PRM.

salicis *Fusarium* Fuckel, Fungi Rhen. Exs., Suppl., Fasc. 7, no. 2110. 1868.

(See *Fusarium lateritium*)

Syntype: S-F267709 (Fungi Rhen. Exs. no. 2110).

Type locality: **Germany**, Hessen, Münchau, near Hattenheim

Type substrate: Dry branches of *Salix triandra*.

Notes: Synonym fide Wollenweber & Reinking (1935). Typification pending further study of the syntype lodged in S.

salinense *Fusarium* Sand.-Den. *et al.*, Persoonia 40: 15. 2017 [2018].

Holotypus: CBS H-23019.

Ex-type culture: CBS 142420 = CPC 26973.

Type locality: **Italy**, Sicily, Messina, Leni.

Type substrate: Twigs of *Citrus sinensis*.

Descriptions and illustrations: See Sandoval-Denis *et al.* (2018a).

Diagnostic DNA barcodes: *rpb1*: LT746286; *rpb2*: LT746306; *tef1*: LT746193.

salmonicolor *Fusarium* Berk. & M.A. Curtis, J. Linn. Soc., Bot. 10: 359. 1868 [1869].

Synonym: *Fusidium salmonicolor* (Berk. & M.A. Curtis) Wollenw., Fusaria Autogr. Delin. 1: 478. 1916.

Holotypus: In K(M).

Type locality: Cuba.

Type substrate: Dead twigs of unknown host.

Notes: Synonym *fide* Wollenweber & Reinking (1935). This taxon needs to be recombined into the genus *Neonectria* but requires further investigation.

samararum *Fusarium* Allesch., Ber. Bayer. Bot. Ges. 4: 39. 1896. (See *Fusarium lateritium*)

Holotypus: In M.

Type locality: Germany, München, Starnberg.

Type substrate: Fallen fruits of *Fraxinus excelsior*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

sambucinum *Fusarium* Fuckel, Fungi Rhen. Exs., Fasc. 3, no. 211. 1863, *nom. cons.*

Synonyms: *Fusarium roseum* Link, Mag. Ges. Naturf. Freunde Berlin 3: 10. 1809, *nom. rej.*

Fusidium roseum (Link) Link, Mag. Ges. Naturf. Freunde Berlin 8: 31. 1815 [1816].

Gibberella rosea (Link) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 32: 664. 1945.

Sphaeria pulicaris Fr., Mykol. Hefte 2: 37. 1823.

Gibbera pulicaris (Fr.) Fr., Summa Veg. Scand. 2: 402. 1849.

Botryosphaeria pulicaris (Fr.) Ces. & De Not., Comment. Soc. Crittog. Ital. 1: 212. 1863.

Nectria pulicaris (Fr.) Tul. & C. Tul., Select. Fung. Carpol. 3: 63. 1865.

Cucurbitaria pulicaris (Fr.) Quéf., Mém. Soc. Émul. Montbéliard, sér. 2, 5: 511. 1875.

Gibberella pulicaris (Fr.) Sacc., Michelia 1: 43. 1877.

Fusarium sulphureum Schltld., Fl. Berol. 2: 139. 1824, *nom. rej.*

Fusidium sulphureum (Schltld.) Link, in Willdenow, Sp. Pl. ed. 4, 6: 98. 1825.

Fusarium discolor var. *sulphureum* (Schltld.) Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land- Forstw. 8: 115. 1910 [1913].

Sphaeria cyanogena Desm., Ann. Sci. Nat., Bot., sér. 3, 10: 352. 1848.

Botryosphaeria cyanogena (Desm.) Niessl, Verh. Naturf. Vereins Brünn 10: 197. 1872.

Gibberella cyanogena (Desm.) Sacc., Syll. Fung. 2: 555. 1883.

Calonectria cyanogena (Desm.) Lar.N. Vassiljeva, Nizshie Rasteniy, Griby i Mokhoobraznye

Dalnego Vostoka Rossii, Griby. Tom 4. Pirenomitsety i Lokuloaskomitsety: 169. 1998.

Fusarium maydis Kalchbr., Math. Term. Közlem. 3: 285. 1865, *nom. rej.*

Fusisporium ricini Bérenger, Mem. Accad. Agric. Verona 44: 257. 1866, *nom. rej.*

Fusarium ricini (Bérenger) Bizz., Fl. Ven. Critt. 1: 539. 1885.

Fusarium subcarneum P. Crouan & H. Crouan, Fl. Finistère: 14. 1867, *nom. rej.*

Fusarium violaceum P. Crouan & H. Crouan, Fl. Finistère: 14. 1867, *nom. illegit.*, Art. 53.1.

Fusisporium pezizoideum Berk. & M.A. Curtis, Grevillea 3: 147. 1875.

Fusarium pezizoideum (Berk. & M.A. Curtis) Sacc., Syll. Fung. 4: 711. 1886.

Fusisporium pulvinatum Berk. & Broome, J. Linn. Soc., Bot. 14: 102. 1873 [1875].

Fusarium pulvinatum (Berk. & Broome) Sacc., Syll. Fung. 4: 699. 1886, *nom. illegit.*, Art. 53.1.

Fusarium roseum var. *buxi* Sacc., Michelia 2: 294. 1881.

Fusarium roseum var. *calystegiae* Sacc., Michelia 2: 294. 1881.

Fusarium roseum var. *cucubali-bacciferi* Sacc., Michelia 2: 295. 1881.

Fusarium roseum var. *dulcamarae* Sacc., Michelia 2: 295. 1881.

Fusarium roseum var. *filicis* Sacc., Michelia 2: 295. 1881.

Fusarium roseum var. *fraxini* Therry, Cryptog. Lyonn.: 5717. 1881.

Fusarium roseum var. *helianti* Sacc., Michelia 2: 295. 1881.

Fusarium roseum var. *maydis* Sacc., Michelia 2: 295. 1881.

Fusarium roseum var. *phytolaccae* Sacc., Michelia 2: 294. 1881.

Fusarium roseum var. *rosae* Sacc., Michelia 2: 295. 1881.

Fusarium roseum var. *vitalbae* Sacc., Michelia 2: 294. 1881.

Fusarium granulare Kalchbr., Crypt. Austro-Afric., no. 1068. 1874.

Fusarium roseum var. *dracaenae* Roum., Fungi Sel. Gall. Exs., Cent. 19: 1869. 1882.

Fusisporium tenuissimum Peck, Rep. (Annual) New York State Mus. Nat. Hist. 34: 48. 1883.

Fusarium tenuissimum (Peck) Sacc., Syll. Fung. 4: 711. 1886.

Fusisporium hordei Wm.G. Sm., Diseases of field and garden crops, chiefly as are caused by fungi: 212. 1884.

Fusarium hordei (Wm.G. Sm.) Sacc., Syll. Fung. 11: 652. 1895.

Gibberella pulicaris f. *robiniae* P. Syd., Mycoth. March., Cent. 14: 1544. 1887.

Fusarium tenellum Sacc. & Briard, Rev. Mycol. (Toulouse) 7: 212. 1885.

Fusarium asparagi Delacr., Bull. Soc. Mycol. France 6: 99. 1890, *nom. illegit.*, Art. 53.1.

Fusarium delacroixii Sacc., Syll. Fung. 10: 725. 1892.

Fusarium fraxini Allesch., Ber. Bot. Vereines Landshut 12: 130. 1892.

Fusarium polymorphum Matr., Rech. Dével. Mucéd.: 84. 1892.

Fusarium roseum var. *loniceriae* Allesch., Ber. Bayer. Bot. Ges. 5: 22. 1897.

Fusarium roseum f. *visci* Brunaud, Actes Soc. Linn. Bordeaux 52: 149. 1897.

Fusarium pannosum Masee, Bull. Misc. Inform. Kew 1898: 117. 1898.

Gibberella pulicaris var. *subtropica* Rehm, in Theissen, Ann. Mycol. 9: 63. 1911.

Gibberella subtropica (Rehm) Wollenw., Fusaria Autogr. Delin. 1: 38. 1916.

Botryosphaeria subtropica (Rehm) Weese, Sitzungsber. Akad. Wiss. Wien, Math.-Naturwiss. Cl., Abt. 1, 128: 708. 1919.

Fusarium genevense Dasz., Bull. Soc. Bot. Genève, sér. 2, 4: 305. 1912.

Fusarium discolor Appel & Wollenw., Arbeiten Kaiserl. Biol. Anst. Land- Forstw. 8: 114. 1913.

Fusarium subpallidum Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 230. 1915.

Fusarium roseum var. *phaseoli* Gonz. Frag., Trab. Mus. Nac. Cienc. Nat., Ser. Bot. 10: 173. 1916.

Fusarium aridum O.A. Pratt, J. Agric. Res. 13: 89. 1918.

Fusarium elongatum O.A. Pratt, J. Agric. Res. 13: 84. 1918, *nom. illegit.*, Art. 53.1.

Fusarium roseum var. *zeae* Cif., Bull. Soc. Bot. Ital. 1921: 73. 1921.

Fusarium sambucinum var. *medium* Wollenw., Z. Parasitenk. (Berlin) 3: 358. 1931.

Fusarium sambucinum f2 Wollenw., Z. Parasitenk. (Berlin) 3: 357. 1931.

Fusarium sambucinum f3 Wollenw., Z. Parasitenk. (Berlin) 3: 357. 1931.

Fusarium sambucinum f4 Wollenw., Z. Parasitenk. (Berlin) 3: 357. 1931.

Fusarium sambucinum f6 Wollenw., Z. Parasitenk. (Berlin) 3: 358. 1931.

Gibberella pulicaris var. *minor* Wollenw., Z. Parasitenk. (Berlin) 3: 356. 1931.

Fusarium roseum f. *phaseoli* N. Barros, Revista Inst. Colomb. Agropecu. 1: 80. 1966.

Fusarium roseum f. *compactum* Tivoli, Agronomie 8: 220. 1988, *nom. inval.*, Arts. 35.1, 39.1.

Fusarium roseum var. *lavaterae-arboreae* Thüm., Mycoth. Univ. Cent. 11: no. 1084. 1878.

Lectotypus: G00266369.

Type locality: **Germany**, Hessen.

Type substrate: Dead branches of *Sambucus nigra*.

Descriptions and illustrations: See Wollenweber & Reinking (1935), Booth (1971), Gerlach & Nirenberg (1982), Nelson et al. (1983), and Leslie & Summerell (2006).

Notes: The taxonomy of *F. sambucinum*, the type species of the genus *Fusarium*, is confusing. Divergent species concepts have been derived from multiple taxonomic systems and the conflicting application of the older name *F. roseum* (Gams et al. 1997, Leslie & Summerell 2006). After examination of the type material, a proposal to conserve *F. sambucinum* against several earlier names was presented (Gams et al. 1997) and unanimously accepted by the committee for fungal taxonomy (Gams 1999). Further older valid synonymous names are in need to be rejected, notably *Sphaeria pulicaris* and *Sphaeria cyanogena*.

samoense *Fusarium* Gehrman, Arbeiten Kaiserl. Biol. Anst. Land-Forstw. 9: 24. 1913.

(See *Fusarium verticillioides*)

Lectotypus (*hic designatus*, MBT 10000740): **Samoa**, cortex of *Theobroma cacao*, 1913, K. Gehrman, in Arbeiten Kaiserl. Biol. Anst. Land-Forstw. 9: Abb. 6, figs 1–3.

Notes: Synonym *vide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotypus.

sampaioi *Fusarium* Gonz. Frag., Bol. Soc. Brot. 2: 50. 1924.

Synonym: *Illosporium corallinum* Roberge, in Desmazières, Pl. Crypt. N. France, ed. 1, Fasc. 32: no. 1551. 1847 (*pr. p. fide* Hawksworth 1979).

Marchandiomyces corallinus (Roberge) Diederich & D. Hawksw., Mycotaxon 37: 312. 1990 (*pr. p. fide* Diederich 1990).

Aegerita physciae Vouaux, Bull. Trimestriel Soc. Mycol. France 30: 314. 1914.

Holotypus: Not indicated. Several syntypes *vide* Hawksworth (1979).

Type locality: **Portugal**, near Gaia, Alto da Bandeira; and near Tabuaço.

Type substrate: Lichen thallus (on *Lasallia pustulata*, *Parmelia saxatilis*, *P. soredians* and *P. exasperata*; *Physcia semipinnata*, *P. tenella*, *Phaeophyscia orbicularis* and *Physconia grisea*).

Notes: Hawksworth (1979), after examination of a syntype, concluded that the *Fusarium* name should be rejected since the studied material was based on discordant elements. Nevertheless, examination of all available syntypes is required to confirm these observations or otherwise, to fix the use of this name by lectotypification.

samuelsii *Fusarium* (Sand.-Den. & Crous) O'Donnell et al., Index Fungorum 440: 4. 2020.

Neocosmospora samuelsii Sand.-Den. & Crous, Persoonia 43: 165. 2019.

Holotypus: CBS H-24001.

Ex-type culture: CBS 114067 = G.J.S. 89-70.

Type locality: **Guyana**, Mount Wokomung, on ridge leading NW toward summit, 0.5–1 h walk from Base Camp.

Type substrate: Bark.

Descriptions and illustrations: See Sandoval-Denis et al. (2019).

Diagnostic DNA barcodes: *rpb1*: MW834252; *rpb2*: LR583874;

tef1: LR583644.

sangayamense *Fusarium* Maryani et al., Stud. Mycol. 92: 187. 2018 [2019].

Holotypus: InaCC F960 (preserved as metabolically inactive culture).

Ex-type culture: InaCC F960.

Type locality: **Indonesia**, South Kalimantan, Kota Baru, Sengayam.

Type substrate: Pseudostem of *Musa* var. Pisang Kepok.

Descriptions and illustrations: See Maryani et al. (2019a).

Diagnostic DNA barcodes: *rpb1*: LS479537; *rpb2*: LS479283;

tef1: LS479732.

sanguineum *Fusarium* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 193. 1915.

(See *Fusarium acuminatum*)

Typus: ?CUP-007444.

Type locality: **USA**, New York, Ithaca

Type substrate: *Solanum tuberosum*.

Notes: Synonym *vide* Wollenweber & Reinking (1935). Lectotypification pending study of material lodged in CUP.

sapindophilum *Fusarium* Speg., Anales Mus. Nac. Hist. Nat. Buenos Aires 6: 351. 1898 [1899].

Synonym: *Cercoseptoria sapindophila* (Speg.) Cif., Mycopathol. Mycol. Appl. 6: 26. 1951.

Holotypus: In LPS (Fungi Argent. n.v.c. no. 868).

Type locality: **Argentina**, near Tucumán.

Type substrate: Living leaves of unknown climbing *Sapindaceae*.

Note: Synonym *vide* Wollenweber & Reinking (1935).

sarcochroum *Fusarium* (Desm.) Sacc., Michelia 1: 534. 1879.

Basionym: *Selenosporium sarcochroum* Desm., Ann. Sci. Nat., Bot., sér. 3, 14: 112. 1850.

Synonyms: *Fusarium diplosporium* Cooke & Ellis, Grevillea 7: 38. 1878.

Fusarium desciscens Oudem., Ned. Kruidk. Arch., sér. 2, 5: 515. 1889.

Fusarium robiniae Pass., Atti Reale Accad. Lincei, Rendiconti Cl. Sci. Fis., sér. 4, 7: 51. 1891.

Fusarium sarcochroum var. *robiniae* (Pass.) Wollenw., Z. Parasitenk. (Berlin) 3: 388. 1931. *Fusarium sarcochroum* f. *polygalae-myrtifoliae* Henn., Verh. Bot. Vereins Prov. Brandenburg 40: 174. 1898 [1899].

Fusarium sarcochroum var. *casei* Loubière, Rech. Mucédinées: 53. 1924.

Gibberella pseudopulicaris Wollenw., Z. Parasitenk. (Berlin) 3: 387. 1931.

Neotypus (*hic designatus*, MBT 10000741): **Switzerland**, *Viscum album*, 1977, W. Gerlach, CBS 745.79 (preserved as metabolically inactive culture).

Ex-neotype culture: BBA 63714 = CBS 745.79 = NRRL 20472.

Descriptions and illustrations: See Wollenweber & Reinking (1935), Raillo (1950), Bilai (1955), Gerlach & Nirenberg (1982).

Diagnostic DNA barcodes: *rpb1*: JX171472; *rpb2*: JX171586; *tef1*: JABEXW01000634.

Notes: No type material could be located. Therefore, CBS 745.79 is designated as neotype here. Both Gerlach & Nirenberg (1982) and O'Donnell *et al.* (2013) considered this isolate an authentic representation of this species.

schawrowii *Fusarium* Speschnew (as 'schawrovi'), Arbeiten Kaukas. Stat. Seidenzucht 10: 1906.

(See *Fusarium lateritium*)

Holotypus: Not located.

Type locality: Turkey, Anatolia.

Type substrate: Branch of *Morus* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

schiedermayeri *Fusarium* (Thüm.) Sacc., Syll. Fung. 4: 712. 1886.

Basionym: *Fusisporium schiedermayeri* Thüm., Fungi Austr. Exs. Cent. 1: no. 78. 1871.

(See *Fusarium avenaceum*)

Syntypus: In HAL.

Type locality: Austria, Linz.

Type substrate: Ovaries of *Luzula pilosa*, in association with *Ustilago luzulae*.

Note: Synonyms *fide* Wollenweber & Reinking (1935).

schnablium *Fusarium* Allesch., Hedwigia 34: 289. 1895.

(See *Fusarium avenaceum*)

Holotypus: In M.

Type locality: Germany, Großhesselohe, near München.

Type substrate: Decorticated branch of *Acer pseudoplatanus*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

schribauxii *Fusarium* Delacr., Bull. Soc. Mycol. France 6: 99. 1890.

(See *Fusarium culmorum*)

Holotypus: ?PC.

Type locality: France.

Type substrate: Seeds of *Triticum sativum*, in association with *Trichothecium roseum*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

schweinitzii *Fusarium* Ellis & Harkn., Bull. Torrey Bot. Club 8: 27. 1881.

Colletotrichum crassipes (Speg.) Arx, Verh. Kon. Akad. Wetensch., Afd. Natuurk., Sect. 2, 51: 77. 1957.

Basionym: *Gloeosporium crassipes* Speg., Rivista Vitic. Enol. 2: 405. 1878.

Syntypes: In CHR, CUP, ILL, MICH, MU, NEB, NYS, PH, PUL & WIS (Ellis, N. Amer. Fungi no. 539).

Type locality: USA, New Jersey, Newfield.

Type substrate: *Vitis* sp. vine

Note: Synonym *fide* Wollenweber & Reinking (1935).

scirpi *Fusarium* Lambotte & Fautrey, Fautrey, Fungi Sel. Gall. Exs. no. 6540. 1893.

Synonyms: ?*Fusoma helminthosporii* Corda, Icon. Fung. 1: 7. 1837.

?*Fusoma filiferum* Preuss, Linnaea 25: 73. 1852.

?*Fusarium filiferum* (Preuss) Wollenw., Fusaria Autogr. Delin. 1: 220. 1916.

?*Fusarium scirpi* var. *filiferum* (Preuss) Wollenw., Fusaria Autogr. Delin. 3: 936. 1930.

?*Fusisporium chenopodium* Thüm., Mycoth. Univ., Cent. 14: no. 1378. 1879.

?*Fusarium chenopodium* (Thüm.) Sacc., Syll. Fung. 4: 701. 1886.

?*Fusarium aloes* Kalchbr. & Cooke (as 'aloès'), Grevillea 9: 23. 1880.

?*Fusarium osteophilum* Speg., Anales Soc. Ci. Argent. 10: 60. 1880.

?*Fusisporium mucophytum* W.G. Sm., Gard. Chron. n.s. 22: 245. 1884.

?*Fusarium mucophytum* (W.G. Sm.) Masee, Brit. Fung.-Fl. 3: 483. 1893.

Fusarium equiseticola Allesch., Hedwigia 34: 289. 1895.

Fusarium sclerotium Wollenw., Ber. Deutsch. Bot. Ges. 31: 301. 1913.

Fusarium caudatum Wollenw., J. Agric. Res. 2: 262. 1914.

Fusarium sclerodermatis var. *lycoperdonis* Picb., Bull. Ecol. Sup. Agron., Brno, R.C.S. Fac. Silvicult. 13: 27. 1929.

Fusarium scirpi var. *comma* Wollenw., Fusaria Autogr. Delin. 3: 922. 1930.

Fusarium scirpi var. *nigrantum* F.T. Benn. (as 'nigrans'), Ann. Appl. Biol. 19: 26. 1932.

Fusarium scirpi var. *pallens* F.T. Benn., Ann. Appl. Biol. 19: 21. 1932.

Lectotypus (*hic designatus*, MBT 10000742): France, *Schoenoplectus lacustris* (= *Scirpus lacustris*), 1893, F. Fautrey, ILL00220730 (Fautrey, Fungi Sel. Gall. Exs. No. 6540).

Epitypus (*hic designatus*, MBT 10000743): Australia, New South Wales, near Broken Hill, pasture soil, 1981, P.E. Nelson, CBS H-24069.

Ex-epitype culture: CBS 447.84 = FRC R-6252 = NRRL 36478.

Descriptions and illustrations: See Wollenweber (1916–1935), Wollenweber & Reinking (1935), Burgess *et al.* (1985) and Leslie & Summerell (2006).

Diagnostic DNA barcodes: *rpb2*: GQ505832; *tef1*: GQ505654.

Notes: The epitypification of *Fusarium scirpi* by Xia *et al.* (2019) was not Code compliant as the holo- or lectotype was not correctly indicated (Art. 9.9). Here, the lectotype is clearly indicated, making the epitypification valid.

sclerodermatis *Fusarium* Oudem., Ned. Kruidk. Arch., sér. 2, 5: 516. 1889.

(See *Fusarium torulosum*)

Holotypus: ?L.

Type locality: Netherlands, Zuid-Holland Province, Scheveningen.

Type substrate: Rotten *Scleroderma vulgaris*.

Note: Synonym *fide* Nirenberg (1995).

sclerodermatis *Fusarium* Peck, Rep. (Annual) Regents Univ. State New York New York State Mus. 43: 77. 1890, *nom. illegit.*, Art. 53.1.

(See *Fusarium oxysporum*)

Authentic material: NYSf2731.

Original locality: USA, New York, Suffolk.

Original substrate: Peridium of *Scleroderma vulgaris*.

Notes: A later homonym of *F. sclerodermatis* Oudem. Saccardo (1892) published *F. peckii* as a replacement name which was again an illegitimate homonym; the taxon was later synonymised with *F. oxysporum* var. *aurantiacum* (Wollenweber & Reinking 1935).

sclerostromaton *Fusarium* Sideris, Phytopathology 14: 213. 1924.

(See *Fusarium oxysporum*)

Holotypus: Not located.

Type locality: **USA**, California, Delta, near Stockton.

Type substrate: Roots of *Allium* sp. with symptoms of pink root disease.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

sclerotoides *Fusarium* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 214. 1915.

(See *Fusarium oxysporum*)

Typus: ?BPI 452971.

Type locality: **USA**, New York, Ithaca.

Type substrate: *Solanum tuberosum*.

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#). Typification pending further study of the specimen lodged in BPI.

sclerotoides *Fusarium* (Höhn.) Samuels & Rossman, Mycological Papers 164: 23. 1991, *nom. illegit.*, Art. 53.1

Basionym: *Stagonopsis sclerotoides* Höhn., in Penther & Zederbauer, Ann. K. K. Naturhist. Hofmus. 20: 368. 1905.

(See *Fusarium kurdicum*)

Holotypus: FH00965353.

Type locality: **Turkey**, near Erciyes Dağı.

Type substrate: Thin twigs of *Astragalus* sp.

sclerotium *Fusarium* Wollenw., Ber. Deutsch. Bot. Ges. 31: 30. 1913.

(See *Fusarium scirpi*)

Holotypus: Not located.

Type locality: **USA**.

Type substrate: *Citrullus vulgaris* and *Lycopersicon esculentum*.

Note: Synonym *fide* [Nirenberg \(1995\)](#).

scolecoides *Fusarium* Sacc. & Ellis, Miscellanea Mycologia 2: 18. 1885.

(See *Fusarium ciliatum*)

Holotypus: In PAD.

Type locality: **USA**, Pennsylvania, Bethlehem.

Type substrate: Branch of *Robinia* sp.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

secalis *Fusarium* Fée, Mém. Soc. Mus. Hist. Nat. Strassbourg 3: 35. 1843.

(See *Fusarium heterosporum*)

Holotypus: Not located.

Type locality: **France**.

Type substrate: Spikes of *Secale cereale*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

secalis *Fusarium* Jacz., Bull. Trimestriel Soc. Mycol. France 28: 346. 1912, *nom. illegit.*, Art. 53.1.

(See *Fusarium nivale*)

Authentic material: Not located.

Original locality: **Russia**, near Moscow.

Original substrate: Grain of *Secale* sp.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

secorum *Fusarium* Secor *et al.*, Fungal Biology 118: 767. 2014.

Holotypus: BPI 892692.

Ex-type culture: NRRL 62593.

Type locality: **USA**, Minnesota, Sabin.

Type substrate: Root of *Beta vulgaris*.

Descriptions and illustrations: See [Secor *et al.* \(2014\)](#).

Diagnostic DNA barcodes: *rpb1*: JABEEM010001657; *rpb2*: JABEEM010001483; *tef1*: KJ189225.

sedimenticola *Fusarium* M.M. Wang *et al.*, Botanica Marina 63: 174. 2020.

(See *Fusarium keratoplasticum*)

Holotypus: HAMS 248044.

Ex-type culture: CGMCC 3.19499 = LC12845.

Type locality: **China**, South-West Indian Ocean.

Type substrate: Deep-sea sediments.

Descriptions and illustrations: See [Jones *et al.* \(2020\)](#).

Diagnostic DNA barcodes: *rpb2*: MK190729; *tef1*: MK190727.

Notes: *Fusarium sedimenticola* was recently introduced by [Jones *et al.* \(2020\)](#) in the FSSC (= *Neocosmospora*) isolated from deep-sea sediment in the Indian Ocean. However, based on comparisons of both protologues and sequences using a larger sampling of *N. keratoplastica* isolates (results not shown), we consider *F. sedimenticola* a synonym under *N. keratoplastica*.

seemenianum *Fusarium* Henn., in Seemen, Allg. Bot. Z. Syst. 2: 83. 1896.

(See *Fusarium avenaceum*)

Holotypus: B 70 0100194.

Type locality: **Germany**, Borkum.

Type substrate: Leaves of *Platanthera bifolia* var. *robusta*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

semitectum *Fusarium* Berk. & Ravenel, Grevillea 3: 98. 1875.

(See *Fusarium incarnatum*)

Holotypus: ?K(M).

Type locality: **USA**, Pennsylvania, Philadelphia.

Type substrate: Petioles of *Musa* sp.

serjaniae *Fusarium* Syd. & P. Syd., Beibl. Hedwigia 40: (2). 1901.

Synonym: *Cercospora serjaniae* (Syd. & P. Syd.) Wollenw., Z. Parasitenk. (Berlin) 3: 496. 1931.

Holotypus: S-F45658.

Type locality: **Mexico**, Puebla, Tehuacán.

Type substrate: Leaves of *Serjania racemosa*.

Notes: Status unclear. Not *Fusarium* *fide* [Wollenweber & Reinking \(1935\)](#) and not *Cercospora* *fide* [Crous & Braun \(2003\)](#).

serpentinum *Fusarium* J.W. Xia *et al.*, Persoonia 43: 217. 2019.

Holotypus: CBS H-24070.

Ex-type culture: BBA 62209 = CBS 119880 = MRC 1813.

Type locality: **Unknown**.

Type substrate: Unknown.

Descriptions and illustrations: See [Xia *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb2*: MN170432; *tef1*: MN170499.

setosum *Fusarium* Nirenberg & Samuels, Canad. J. Bot. 67: 3372. 1989.

Setofusarium setosum (Samuels & Nirenberg) Sand.-Den. & Crous, Stud. Mycol. 98 (no. 100116): 75. 2021.

Synonym: *Nectria setofusarii* Samuels & Nirenberg (as '*setofusariae*'), Canad. J. Bot. 67: 3372. 1989.

Holotypus: NY00927992.

Type locality: **French Guiana**, Piste de Saint-Elie, Km 16 on road between Sinnamary and St. Elie, ORSTOM research area, "ECEREX".

Type substrate: Bark of living liana.

Epitypus: CBS H-24723, designated in this study.

Ex-epitype culture: CBS 635.92 = G.J.S. 88-12 = NRRL 36526.

Epitype locality: **French Guiana**, Cayenne, 15 km from Remise, trail to Vidal-old farm, secondary forest.

Epitype substrate: Bark.

Descriptions and illustrations: See [Samuels & Nirenberg \(1989\)](#).

Diagnostic DNA barcodes: *rpb1*: JX171539; *rpb2*: JX171651; *tef1*: MW834294.

sibiricum Fusarium Gagkaeva *et al.*, Int. J. Food Microbiol. 147: 64. 2011.

Holotypus: LEP 12652.

Ex-type culture: MFG 11013 = NRRL 53430.

Type locality: **Russia**, Khabarovsk.

Type substrate: Grain of *Avena sativa*.

Descriptions and illustrations: See [Yli-Mattila *et al.* \(2011\)](#).

Diagnostic DNA barcodes: *rpb1*: MW233302; *rpb2*: HQ154472; *tef1*: HM744684.

siculi Fusarium Sand.-Den. *et al.*, Persoonia 40: 17. 2017 [2018].

Holotypus: CBS H-23021.

Ex-type culture: CBS 142422 = CPC 27188.

Type locality: **Italy**, Sicily, Catania, Paternó.

Type substrate: *Citrus sinensis*.

Descriptions and illustrations: See [Sandoval-Denis *et al.* \(2018a\)](#).

Diagnostic DNA barcodes: *rpb1*: LT746299; *rpb2*: LT746327; *tef1*: LT746214.

silvicola Fusarium (Sand.-Den. & Crous) O'Donnell *et al.*, Index Fungorum 440: 4. 2020.

Neocosmospora silvicola Sand.-Den. & Crous, Persoonia 43: 167. 2019.

Synonyms: *Fusarium solani f. robiniae* Matuo & Y. Sakurai, Ann. Phytopathol. Soc. Japan 30: 35. 1965.

Hypomyces solani f. robiniae Matuo & Y. Sakurai, Ann. Phytopathol. Soc. Japan 30: 35. 1965.

Nectria solani f. robiniae (Matuo & Y. Sakurai) G.R.W. Arnold, Z. Pflanzk. 37: 193. 1972.

Holotypus: CBS H-24002.

Ex-type culture: CBS 123846 = G.J.S. 04-147.

Type locality: **USA**, Tennessee, Great Smoky Mountains National Park.

Type substrate: Fallen trunk of *Liriodendron tulipifera*.

Descriptions and illustrations: See [Sandoval-Denis *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW834254; *rpb2*: LR583876; *tef1*: LR583646.

sinense Fusarium Z.H. Zhao & G.Z. Lu (as '*sinensis*'), Mycologia 100: 747. 2008.

Holotypus: IBE 000007.

Ex-type culture: CBS 122710.

Type locality: **China**, Shandong Province, Jinan.

Type substrate: Seed of *Triticum aestivum*.

Descriptions and illustrations: See [Zhao & Lu \(2008\)](#).

Diagnostic DNA barcode: *tef1*: EF531235.

socium Fusarium Sacc., Atti Ist. Veneto Sci. Lett. Arti, sér. 6, 2: 450. 1884.

(See ***Fusarium expansum***)

Holotypus: Not located.

Type locality: **France**, Troyes.

Type substrate: Cortex of *Carpinus* sp. in association with *Stilbospora* sp. and *Nectria stilbosporae*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

solani Fusarium (Mart.) Sacc., Michelia 2: 296. 1881.

Neocosmospora solani (Mart.) L. Lombard & Crous, Stud. Mycol. 80: 228. 2015.

Basionym: *Fusisporium solani* Mart., Die Kartoffel-Epidemie der letzten Jahre oder die Stockfäule und Räude der Kartoffeln: 20. 1842.

Synonyms: *Fusisporium solani-tuberosi* Desm., Ann. Sci. Nat., Bot., sér. 3, 3: 359. 1845.

Fusarium solani-tuberosi (Desm.) Sacc., Syll. Fung. 4: 189. 1886.

Pionnotes solani-tuberosi (Desm.) Sacc., Syll. Fung. 4: 727. 1886.

Fusisporium rhizophylum var. *solani-tuberosi* (Desm.) Westend., Bull. Acad. Roy. Sci. Belgique, Cl. Sci. 18(2): 413. 1852.

Fusisporium candidum Bonord., Handb. Allg. Mykol.: 96. 1851, nom. illegit., Art. 53.1, non *Fusarium candidum* (Link) Sacc. 1886

Fusarium commutatum Sacc., Syll. Fung. 4: 710. 1886.

Fusarium allii-sativi Allesch., Ber. Bot. Vereines Landshut 12: 131. 1892.

Hymenula affinis (Fautrey & Lambotte) Wollenw., Fusaria Autogr. Delin. 1: 484. 1916 [*pr. p. fide* [Booth \(1971\)](#)].

Pionnotes viridis Lechmere, Compt. Rend. Hebd. Séances Acad. Sci. 155: 178. 1912.

Fusarium viride (Lechmere) Wollenw., Fusaria Autogr. Delin. 1: 418. 1916.

Fusarium radicolica Wollenw., J. Agric. Res. 2: 257. 1914.

Fusarium javanicum var. *radicolica* (Wollenw.) Wollenw., Z. Parasitenk. (Berlin) 3: 286. 1931.

Fusarium solani f. radicolica (Wollenw.) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 28: 740. 1941.

Fusarium eumartii C.W. Carp., J. Agric. Res. 5: 204. 1915.

Fusarium solani var. *eumartii* (C.W. Carp.) Wollenw., Z. Parasitenk. (Berlin) 3: 452. 1931.

Fusarium solani f. eumartii (C.W. Carp.) W.C. Snyder & H.N. Hansen, Amer. J. Bot. 28: 740. 1941.

Fusarium malli Taubenh., Bull. Texas Agric. Exp. Sta. 273: 25. 1921.

Fusarium alluviale Wollenw. & Reinking, Phytopathology 15: 167. 1925.

Fusarium aduncisporum Weimer & Harter, J. Agric. Res. 32: 312. 1926.

Fusarium solani var. *aduncisporum* (Weimer & Harter) Wollenw., Fusaria Autogr. Delin. 3: 1035. 1930.

Neocosmospora rubicola L. Lombard & Crous, Stud. Mycol. 80: 227. 2015.

Lectotypus: Illustration tab. III, fig. 29 in von Martius (1842), designated in [Schroers *et al.* \(2016\)](#).

Epitypus: CBS H-22335, designated in [Schroers *et al.* \(2016\)](#).

Ex-epitype culture: CBS 140079 = FRC S-2364 = NRRL 66304.

Epitype locality: **Slovenia**, Doljenska, Radohova.

Epitype substrate: Rotten tuber of *Solanum tuberosum*.

Descriptions and illustrations: See [Wollenweber & Reinking \(1935\)](#), [Leslie & Summerell \(2006\)](#), and [Schroers *et al.* \(2016\)](#).

Diagnostic DNA barcodes: *rpb1*: MW218134; *rpb2*: KT313623; *tef1*: KT313611.

solani-melongenae Fusarium O'Donnell *et al.*, Index Fungorum 440: 4. 2020.

Neocosmospora ipomoeae (Halst.) L. Lombard & Crous, Stud. Mycol. 80: 227. 2015.

Basionym: *Nectria ipomoeae* Halst., Rep. (Annual) New Jersey Agric. Exp. Sta. 12: 281. 1891.

Synonyms: *Cucurbitaria ipomoeae* (Halst.) Kuntze, Revis. Gen. Pl. 3: 461. 1898.

Creonectria ipomoeae (Halst.) Seaver, N. Amer. Fl. 3: 22. 1910.

Hypomyces ipomoeae (Halst.) Wollenw., Phytopathology 3: 34. 1913.

Haematonectria ipomoeae (Halst.) Samuels & Nirenberg, Stud. Mycol. 42: 136. 1999.

Nectria ipomoeae f. *ipomoeae* Halst., Rep. (Annual) New Jersey Agric. Exp. Sta. 12: 281. 1891.

Nectria ipomoeae var. *ipomoeae* Halst., Rep. (Annual) New Jersey Agric. Exp. Sta. 12: 281. 1891.

Hypomyces ipomoeae var. *ipomoeae* (Halst.) Wollenw., Phytopathology 3: 34. 1913.

Hypomyces ipomoeae var. *major* Wollenw., Fusaria Autogr. Delin. 3: 826. 1930.

?*Fusarium striatum* Sherb., Cornell Univ. Agric. Exp. Sta. Mem. 6: 255. 1915

?*Fusarium solani* var. *striatum* (Sherb.) Wollenw., Z. Parasitenk. (Berlin) 3: 451. 1931.

Holotypus: BPI 552416.

Type locality: **USA**, New Jersey, Mickleton.

Type substrate: *Solanum melongena*.

Note: This species requires epitypification from the type locality and host.

solani-tuberosi *Fusarium* (Desm.) Sacc., Syll. Fung. 4: 189. 1886.

Basionym: *Fusisporium solani-tuberosi* Desm., Ann. Sci. Nat., Bot., sér. 3, 3: 359. 1845.

(See *Fusarium solani*)

Holotypus: ?PC.

Type locality: **France**.

Type substrate: Rotten tuber of *Solanum tuberosum*.

Note: Synonyms fide Wollenweber & Reinking (1935).

sophorae *Fusarium* Allesch., Beibl. Hedwigia 36: (164). 1897.

(See *Fusarium lateritium*)

Holotypus: In M.

Type locality: **Germany**, Berlin, Späth'sche Baumschulen.

Type substrate: *Sophora japonica*.

Note: Synonym fide Wollenweber & Reinking (1935).

sorghii *Fusarium* Henn., Ann. Mus. Congo Belge, Bot., sér. 5, 2: 105. 1907.

(See *Fusarium avenaceum*)

Syntype: Vanderyst 171 in B fide Hein (1988).

Type locality: **Democratic Republic of the Congo**, Kisantu.

Type substrate: Spikelet of *Sorghum vulgare* (= *Sorghum bicolor*).

Note: Synonym fide Wollenweber & Reinking (1935).

sororula *Fusarium* Herron et al., Stud. Mycol. 80: 146. 2015.

Holotypus: PREM 60903.

Ex-type culture: CBS 137242 = CMW 40578.

Type locality: **Colombia**, Risaralda, Angela Maria (Santa Rosa).

Type substrate: Stem cankers of *Pinus patula*.

Descriptions and illustrations: See Herron et al. (2015).

Notes: Comparisons of recently generated sequences from the living ex-type culture (CBS 137242 = CMW 40578) of *F. sororula* indicate a strain transposition or contamination by another *Fusarium* species. Therefore, this species needs to be recollected from the type locality and substrate or sequences need to be generated from the holotype specimen.

spartinae *Fusarium* Ellis & Everh., J. Mycol. 8: 14. 1902.

Septogloeum spartinae (Ellis & Everh.) Wollenw. & Reinking, Fusarien: 336. 1935.

Holotypus: NY (fide Index Fungorum).

Type locality: **USA**, California, Pacific Grove.

Type substrate: Leaves of *Spartina stricta*.

Note: Synonym fide Wollenweber & Reinking (1935).

spartum Fusarium S. Gargouri et al., Mycologia 112: 799. 2020.
Holotypus: BPI 911207.

Ex-type culture: NRRL 66896.

Type locality: **Tunisia**, Kasserine Governorate.

Type substrate: Rhizosphere of *Macrochloa tenacissima*.

Descriptions and illustrations: See Gargouri et al. (2020).

Diagnostic DNA barcodes: *rpb1*: MT409439; *rpb2*: MT409449; *tef1*: MT409459.

spathulatum *Fusarium* (Sand.-Den. & Crous) O'Donnell et al., Index Fungorum 440: 4. 2020.

Neocosmospora spathulata Sand.-Den. & Crous, Persoonia 43: 171. 2019.

Holotypus: CBS H-24003.

Ex-type culture: CBS 145474 = NRRL 28541 = UTHSC 98-1305.

Type locality: **USA**, New England.

Type substrate: Synovial fluid from *Homo sapiens*.

Descriptions and illustrations: See Sandoval-Denis et al. (2019).

Diagnostic DNA barcodes: *rpb1*: MW218137; *rpb2*: EU329542; *tef1*: DQ246882.

speiranthae *Fusarium* Henn. (as 'speiranthis'), Verh. Bot. Vereins Prov. Brandenburg 40: 174. 1898.

Colletotrichum dematium (Pers.) Grove, J. Bot. 56: 341. 1918.
Basionym: *Sphaeria dematium* Pers., Syn. Meth. Fung.: 88. 1801.

Synonyms: *Exosporium dematium* (Pers.) Link, in Willdenow, Sp. pl., Ed. 4, 6: 122. 1825.

Vermicularia dematium (Pers.) Fr., Syst. Mycol. 3: 255. 1829.

Lasiella dematium (Pers.) Quél., Mém. Soc. Émul. Montbéliard sér. 2, 5: 518. 1875.

Gloeosporium speiranthae (Henn.) Wollenw., Fusaria Autogr. Delin. 1: 500. 1916.

Holotypus: In B fide Hein (1988).

Type locality: **Germany**, Berlin, botanical garden.

Type substrate: Leaves of *Speirantha convallarioides*.

Notes: Wollenweber (1916) studied and illustrated authentic material of this species, recombining it in *Gloeosporium*. The shape of the conidia is similar to species in the *Colletotrichum dematium* species complex. However, the conidia are slightly broader than those of the ex-type strain of *C. dematium* (3.5–4.5 vs 3–4 µm, Damm et al. 2009); the synonymy needs to be confirmed. Publication data cited in name repositories (Allg. Bot. Z. Syst. 2: 83. 1896.) are incorrect and instead refer to the protologue of *F. seemenianum* (syn. *F. avenaceum*), an unrelated taxon.

speiseri *Fusarium* Lindau, Rabenh. Krypt.-Fl. Ed. 2, 1(9): 580. 1909.

(See *Fusarium avenaceum*).

Holotypus: B 70 0100195.

Type locality: **Poland**, Karthaus, Nydek.

Type substrate: Dead *Auchenorrhyncha* (cicada).

Note: Synonym fide Wollenweber & Reinking (1935).

spermogoniopsis *Fusarium* Jul. Müll., Ber. Deutsch. Bot. Ges. 3: 394. 1885.

Hymenella spermogoniopsis (Jul. Müll.) L. Lombard & Sand.-Den., **comb. nov.** MycoBank MB 837721.

Basionym: *Fusarium spermogoniopsis* Jul. Müll., Ber. Deutsch. Bot. Ges. 3: 394. 1885.

Synonym: *Hymenula spermogoniopsis* (Jul. Müll.) Wollenw., Fusaria Autogr. Delin. 1: 483. 1916.

Syntypes: ?B 70 0100196, B 70 0100197 & B 700100198.

Type locality: **Germany**

Type substrate: Sporocarps of *Phragmidium subcorticium* (= *Phragmidium mucronatum*) and on the uredo- and teliospores of *Phragmidium rubi* (= *Phragmidium barclayi*).

Notes: [Wollenweber \(1916\)](#) provided a new combination for *F. spermogoniopsis* in the genus *Hymenula*. However, the generic name *Hymenella* (1822) predates the generic name *Hymenula* (1828) and therefore we provide a new combination in the latter genus.

sphaeriae *Fusarium* Fuckel, Fungi Rhen. Exs., Fasc. 3, no. 212. 1863.

(See *Fusarium clematidis*)

Lectotypus: G00111017, designated in [Gräfenhan et al. \(2011\)](#).

Lectotype locality: **Germany**, Hessen, Reichartshausen near Oestrich-Winkel.

Lectotype substrate: Parasitic on *Leptosphaeria* (*Sphaeria*) *dioica*, on *Urtica dioica*.

sphaeriiforme *Fusarium* Sacc. (as 'sphaeriaeforme'), Syll. Fung. 10: 723. 1892.

Replaced synonym: *Fusarium celtidis* Pass., Atti Reale Accad. Lincei, Rendiconti Cl. Sci. Fis., sér. 4, 7: 51. 1891, *nom. illegit.*, Art. 53.1.

(See *Fusarium melanochlorum*)

Holotypus: ?PARMA.

Type locality: **Italy**, Parma, Vigheffio.

Type substrate: Dead branches of *Celtis australis*.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

sphaeroideum *Fusarium* Pass., Atti Reale Accad. Lincei, Rendiconti Cl. Sci. Fis., sér. 4, 4: 105. 1888.

(See *Fusarium lateritium*)

Holotypus: ?PARMA.

Type locality: **Italy**, Parma.

Type substrate: Branch of *Ficus carica*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

sphaerosporum *Fusarium* Q.T. Chen & X.H. Fu, Acta Mycol. Sin., Suppl. 1: 331. 1987.

Neocosmospora sphaerospora (Q.T. Chen & X.H. Fu) Sand.-Den. & Crous, Persoonia 43: 173. 2019.

Holotypus: HMAS 43749.

Ex-type culture: NF 5840.

Type locality: **China**, Guangdong Province, Maoming.

Type substrate: Water from underground pipes of oilfield.

Descriptions and illustrations: See [Chen et al. \(1987\)](#).

spinaciae *Fusarium* Hungerf., Phytopathology 13: 209. 1923.

(See *Fusarium oxysporum*)

Lectotypus (*hic designatus*, MBT 10000744): **USA**, Idaho, roots of *Spinacia oleracea*, 1923, C.W. Hungerford, in Phytopathology 13: 208, fig. 4.

Notes: Synonym *fide* [Booth \(1971\)](#). No holotype specimen could be located and therefore an illustration was designated as lectotype.

spinosum* *Fusarium L. Lombard et al., Fungal Syst. Evol. 4: 195. 2019.

Holotypus: CBS H-24020.

Ex-type culture: CBS 122438.

Type locality: **Brazil**.

Type substrate: Galia melon imported into the Netherlands.

Descriptions and illustrations: See [Lombard et al. \(2019a\)](#).

Diagnostic DNA barcodes: *rpb1*: MN120729; *rpb2*: MN120747; *tef1*: MN120768.

spinulosum *Fusarium* (Pfenning) O'Donnell et al., Index Fungorum 440: 4. 2020.

Neocosmospora spinulosa Pfenning, Sydowia 47: 66. 1995.

Holotypus: CBS H-5452a.

Ex-type culture: CBS 321.93.

Type locality: **Brazil**, Pará, Capitão Poço.

Type substrate: Soil under *Theobroma cacao*.

Descriptions and illustrations: See [Pfenning \(1995\)](#).

splendens *Fusarium* Matuo & Takah. Kobay., Trans. Mycol. Soc. Japan 2(4): 13. 1960, *nom. inval.*, Art. 39.1.

(See *Fusarium matuoi*)

Authentic material: Not located.

Original locality: **Japan**.

Original substrate: Twigs of *Albizia julibrissin*.

Descriptions and illustrations: See [Matuo & Kobayashi \(1960\)](#) and [Hosoya & Tubaki \(2004\)](#).

sporodochiale* *Fusarium L. Lombard & Crous, Fungal Syst. Evol. 4: 196. 2019.

Holotypus: CBS H-12681.

Ex-type culture: ATCC 14167 = CBS 220.61 = MUCL 8047 = NRRL 20842.

Type locality: **South Africa**, Gauteng Province, Johannesburg.

Type substrate: Soil.

Descriptions and illustrations: See [Lombard et al. \(2019a\)](#).

Diagnostic DNA barcodes: *rpb1*: MN120731; *rpb2*: MN120749; *tef1*: MN120770.

sporotrichiella *Fusarium* Bilař, Yadovitye griby na zerne khlebnykh zlakov: 86. 1953, *nom. inval.*, Art. 39.1.

(See *Fusarium sporotrichioides*)

Authentic material: Not located.

Original locality: **Ukraine**.

Original substrate: Unknown.

Descriptions and illustrations: See [Bilař \(1955\)](#).

Notes: This taxon was published as a new name for all the taxa in section *Sporotrichiella*. However, it is invalid as no type and Latin diagnosis were provided. Synonym *fide* [Gerlach & Nirenberg \(1982\)](#).

sporotrichioides* *Fusarium Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 183. 1915.

Synonyms: *Fusarium sporotrichiella* var. *sporotrichioides* (Sherb.) Bilař, Yadovitye griby na zerne khlebnykh zlakov (*Poisonous fungi on cereal seed*), Kiev: 87. 1953, *nom. inval.*, Art. 39.1.

Fusarium sporotrichiella Bilař, Yadovitye griby na zerne khlebnykh zlakov (*Poisonous fungi on cereal seed*), Kiev: 86. 1953, *nom. inval.*, Art. 39.1.

Fusarium sporotrichioides var. *minus* Wollenw., Fusaria Autogr. Delin. 3: 886. 1930.

Fusarium sporotrichioides subsp. *minus* (Wollenw.) Raillo, Fungi of the Genus *Fusarium*: 196. 1950.

Lectotypus (*hic designatus*, MBT 10000745): **USA**, New York, rotten tubers of *Solanum tuberosum*, together with *F. solani* and *F. oxysporum*, 1915, C.D. Sherbakoff, in Mem. Cornell Univ. Agric. Exp. Sta. 6: 184, fig. 22.

Notes: This economically important species requires epitypification from the type locality and substrate. No holotype

specimen could be located and therefore an illustration was designated as lectotype.

staphyleae *Fusarium* Samuels & Rogerson, Brittonia 36: 84. 1984.

Geejayessia atrofusca (Schwein.) Schroers & Gräfenhan, Stud. Mycol. 68: 126. 2011.

Basionym: *Sphaeria atrofusca* Schwein., Trans. Amer. Philos. Soc., n.s. 4: 206. 1832.

Synonyms: *Valsaria atrofusca* (Schwein.) Cooke ex Sacc., Syll. Fung. 9: 759. 1891.

Nectria atrofusca (Schwein.) Ellis & Everh., N. Amer. Pyren.: 99. 1892.

Pseudodiplodia atrofusca (Schwein.) Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 19: 94. 1894.

Cucurbitaria atrofusca (Schwein.) Kuntze, Revis. Gen. Pl. 3: 460. 1898.

Creonectria atrofusca (Schwein.) Seaver, Mycologia 1: 186. 1909.

Holotypus: In NY.

Ex-type culture: ATCC 66906 = CBS 502.94 = IMI 345891 = NRRL 22120.

Type locality: **USA**, Massachusetts, Berkshire, south of Ashley Falls, Bartholomew's Cobble.

Type substrate: Branches of *Staphylea trifolia*.

Descriptions and illustrations: See Samuels & Rogerson (1984) and Schroers et al. (2011).

stercicola *Fusarium* Šišić et al., Antonie van Leeuwenhoek 111: 1793. 2018.

Neocosmospora stercicola (Šišić et al.) Sand.-Den. & Crous, Persoonia 43: 173. 2019.

Synonyms: *Fusarium martii* var. *viride* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 247. 1915.

Fusarium solani var. *martii* 'f. 1' Wollenw., Z. Parasitenk. (Berlin) 3: 290. 1931.

Fusarium witzenhausenense Šišić et al., Antonie van Leeuwenhoek 111: 1795. 2018.

Fusarium xiangyunense F. Zhang et al. (as '*xiangyunensis*'), Phytotaxa 450: 278. 2020. *nom. inval.*, Art. 40.8.

Holotypus: CBS H-23352.

Ex-type culture: CBS 142481 = DSM 106211 = FS 89.

Type locality: **Germany**, Niedersachsen, Hannover.

Type substrate: Compost yard waste plant debris.

Descriptions and illustrations: See Šišić et al. (2018a).

Diagnostic DNA barcodes: *rpb1*: MW834255; *rpb2*: LR583887; *tef1*: LR583658.

stercorarium *Fusarium* Rostr., Meddel. Grønland 18: 74. 1894.

Holotypus: C-F-92401

Type locality: **Greenland**, Vestfjord.

Type substrate: Dung of *Rangifer tarandus* (reindeer).

Notes: Status unclear. Not *Fusarium* fide Wollenweber & Reinking (1935).

stercoris *Fusarium* Fuckel, Fungi Rhen. Exs., Suppl., Fasc. 5: no. 1921. 1867 [and Jahrb. Nassauischen Vereins Naturk. 23–24: 369. 1870].

(See ***Fusarium avenaceum***)

Lectotypus (*hic designatus*, MBT 10000746): **Germany**, Hessen, Oestrich-Winkel, soil next to *Peziza stercoraria*, date unknown, K.W.G.L. Fuckel, Fungi Rhen. Exs., Suppl., Fasc. 5: no. 1921 in HAL.

Notes: Synonym fide Wollenweber & Reinking (1935). No holotype specimen could be located and therefore the exsiccate lodged in HAL is designated as lectotype.

sterilhyphosum* *Fusarium Britz et al., Mycologia 94: 726. 2002.

Holotypus: PREM 57302.

Ex-type culture: NRRL 25623.

Type locality: **South Africa**, Limpopo Province, Tzaneen, Let-sitele area.

Type substrate: Malformed inflorescence of *Mangifera indica*.

Descriptions and illustrations: See Britz et al. (2002) and Leslie & Summerell (2006).

Diagnostic DNA barcodes: *rpb1*: MN193925; *rpb2*: MN193897; *tef1*: MN193869.

sticticum *Fusarium* Berk. & M.A. Curtis, in Berkeley, Grevillea 3: 99. 1875.

(See ***Fusarium lateritium***)

Holotypus: ?K(M).

Type locality: **USA**, South Carolina.

Type substrate: Twigs of *Prunus persica*.

Note: Synonym fide Wollenweber & Reinking (1935).

stictoides *Fusarium* Durieu & Mont., Explor. Sci. Algérie 1: 334. 1848.

(See ***Fusarium graminearum***)

Holotypus: ?PC.

Type locality: **Algeria**.

Type substrate: Branch of flowering *Agave* sp.

Note: Synonym fide Wollenweber & Reinking (1935).

stilbaster *Fusarium* (Link) Link, Sp. pl., Ed. 4, 6: 106. 1825.

Atractium stilbaster Link, Mag. Ges. Naturf. Freunde, Berlin 3: 10. 1809.

Synonyms: *Atractium fuscum* Sacc., Syll. Fung. 2: 514. 1883.

Stilbella fusca (Sacc.) Seifert, Stud. Mycol. 27: 77. 1985.

Atractium flavoviride Sacc., Syll. Fung. 2: 514. 1883.

Stilbum madidum Peck, Rep. (Annual) New York State Mus. Nat. Hist. 46: 115. 1894.

Didymostilbe eichleriana Bres. & Sacc., Atti Congr. Bot. Palermo: 59. 1903.

Didymostilbe obovoidea Matsush., Icon. Microfung. Matsush. lect.: 60. 1975.

Lectotypus: Illustration published in Mag. Ges. Naturf. Freunde, Berlin 3, tab. I, fig. 11, designated in Gräfenhan et al. (2011).

Epitypus: CBS 410.67 (preserved as metabolically inactive culture), designated in Gräfenhan et al. (2011).

Ex-epitype culture: CBS 410.67.

Epitype locality: **Germany**, Bayerischer Wald, Rachelseewand.

Epitype substrate: Bark.

Descriptions and illustrations: See Seifert (1985) and Gräfenhan et al. (2011).

Diagnostic DNA barcodes: *rpb1*: KM232206; *tef1*: KM231920.

stilboides* *Fusarium Wollenw., Fusaria Autogr. Delin. 2: 615. 1924.

Synonyms: *Fusarium lateritium* var. *stilboides* (Wollenw.) Bilař, Fusarii (Biologija i sistematika): 266. 1955, *nom. inval.*, Art. 41.5.

Fusarium lateritium var. *stilboides* (Wollenw.) Bilař, Mikrobiol. Zhurn. 49: 6. 1987.

Fusarium lateritium var. *longum* Wollenw., Fusaria Autogr. Delin. 1: 385. 1916.

- Fusarium fructigenum* var. *minus* Wollenw., Z. Parasitenk. (Berlin) 3: 386. 1931.
- Fusarium stilboides* var. *minus* (Wollenw.) Wollenw., Z. Parasitenk. (Berlin) 3: 333. 1931.
- Fusarium stilboides* 'f. 1' Raillo, Fungi of the Genus *Fusarium*: 271. 1950.
- Gibberella stilboides* W.L. Gordon ex C. Booth, The Genus *Fusarium*: 119. 1971.
- Lectotypus* (*hic designatus*, MBT 10000747): **Philippines**, Los Baños, living twigs of *Citrus* sp., invaded by coccids, 1917, O.A. Reinking, in *Fusaria Autogr. Delin.* 2: 615.
- Epitypus* (*hic designatus*, MBT 10000748): **Cook Islands**, *Citrus* sp., Sep. 1978, G.F. Laundon, CBS 746.79 (preserved as metabolically inactive culture).
- Ex-epitype culture*: BBA 63887 = CBS 746.79 = ICMP 10624 = NRRL 25485.
- Descriptions and illustrations*: See Wollenweber (1924, 1930), Wollenweber & Reinking (1935), Doidge (1938), Raillo (1950), Booth (1971), and Gerlach & Nirenberg (1982).
- Diagnostic DNA barcodes*: *rpb1*: MW928817; *rpb2*: MW928832; *tef1*: MW928843.
- Note*: No holotype specimen could be located and therefore an illustration was designated as lectotype.
- stillatum* *Fusarium* De Not. ex Sacc., in Berlese & Voglino, Syll. Fung., Addit. I–IV: 390. 1886.
- Myxosporium stillatum*** (De Not. ex Sacc.) Wollenw., *Fusaria Autogr. Delin.* 1: 490. 1916.
- Lectotypus* (*hic designatus*, MBT 10000749): **Italy**, Valle Intrasca, at the bridge on Possaccio, dried stems of *Genista tinctoria*, 1862, G. de Notaris, S-F45664 [Baglietto, Cesati & Notaris, Erb. Critt. Ital. Ser. I no. 148 (1148)].
- Notes*: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore the exsiccate lodged in S is designated as lectotype.
- stoveri* *Fusarium* C. Booth, The Genus *Fusarium*: 37. 1971.
- Microdochium stoveri*** (C. Booth) Samuels & I.C. Hallett, Trans. Brit. Mycol. Soc. 81: 481. 1983.
- Basionym*: *Micronectriella stoveri* C. Booth, Mycol. Pap. 94: 3. 1964.
- Synonym*: *Monographella stoveri* (C. Booth) Samuels & I.C. Hallett, Trans. Brit. Mycol. Soc. 81: 473. 1983.
- Holotypus*: IMI 92905.
- Type locality*: **Honduras**.
- Type substrate*: Leaf of *Musa* sp.
- Descriptions and illustrations*: See Booth (1964, 1971), Gerlach & Nirenberg (1982) and Samuels & Hallett (1983).
- striatum* *Fusarium* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 255. 1915.
- (See *Fusarium solani-melongenae*)
- Typus*: ?CUP-007460.
- Type locality*: **USA**, Colorado
- Type substrate*: *Solanum tuberosum*.
- Notes*: Synonym *fide* Nirenberg & Brielmaiers-Liebetanz, 1996 and Sandoval-Denis *et al.* (2019). Lectotypification pending study of material lodged in CUP.
- strobilinum* *Fusarium* Corda, Icon. Fung. 1: 4. 1837.
- Sirococcus conigenus*** (Pers.) P.F. Cannon & Minter, Taxon 32: 577. 1983.
- Basionym*: *Hysterium conigenum* Pers., Ann. Bot. (Usteri) 15: 30. 1795.
- Synonyms*: *Hypoderma conigenum* (Pers.) DC., Fl. Franç., ed. 3, 2: 305. 1805.
- Hypodermopsis conigena* (Pers.) Kuntze, Revis. Gen. Pl. 3: 487. 1898.
- Discella conigena* (Pers.) Höhn., Mitt. Bot. Inst. T. H. Wien 6: 120. 1929.
- Ascochyta strobilina* (Corda) Wollenw., *Fusaria Autogr. Delin.* 1: 505. 1916.
- Sphaeria strobilina* Holl & J.C. Schmidt, Deutschl. Schwämme, Erste Lieferung: 4. 1815, *nom. inval.*, Art. 38.1(a).
- Sphaeria strobilina* Holle & J.C. Schmidt ex Fr., Syst. Mycol. 2: 495. 1823.
- Dichaena strobilina* (Holle & J.C. Schmidt ex Fr.) Fr., Summa Veg. Scand. 2: 403. 1849.
- Sporonema strobilinum* Desm., Ann. Sci. Nat., Bot., sér. 3, 18: 368. 1852.
- Plenodomus strobilinus* (Desm.) Höhn., Sitzungsber. Kaiserl. Akad. Wiss. Wien, Math.-Naturwiss. Cl., Abt. 1, 119: 647. 1910.
- Discella strobilina* (Desm.) Died., Krypt.-Fl. Brandenburg 9: 752. 1914.
- Sirococcus strobilinus* (Desm.) Petr., Sydowia 1: 155. 1947, *nom. illegit.*, Art. 53.1.
- Sirococcus strobilinus* Preuss, Linnaea 26: 716. 1855.
- Phoma conigena* P. Karst., Rev. Mycol. (Toulouse) 7: 106. 1885.
- Septoria parasitica* R. Hartig, Z. Forst- Jagdwesen 1890: 1. 1890.
- Diplodina parasitica* (R. Hartig) Prill., Maladies des Plantes Agricoles 2: fig. 365. 1897.
- Ascochyta parasitica* Fautrey, Rev. Mycol. (Toulouse) 13: 79. 1891.
- Ascochyta piniperda* Lindau, Nat. Pflanzenfam., Teil. I, 1: 367. 1900.
- Phoma conigena* var. *abieticola* Sacc., Ann. Mycol. 3: 233. 1905.
- Typus*: In PRM *fide* Pilat (1938).
- Type locality*: **Czech Republic**, near Liberec (Reichenberg).
- Type substrate*: Rotten cone scales of *Pinus* sp.
- Note*: Typification pending study of material lodged in PRM.
- stromaticola* *Fusarium* Henn., Bot. Jahrb. Syst. 28: 280. 1900.
- Dialonectria volutella*** (Ellis & Everh.) L. Lombard & Sand.-Den., **comb. nov.** MycoBank MB 837722.
- Basionym*: *Fusarium volutella* Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 43: 93. 1891.
- Synonyms*: *Fusarium aquaeductuum* var. *medium* Wollenw., *Fusaria Autogr. Delin.* 3: 844. 1930.
- Fusarium aquaeductuum* subsp. *medium* (Wollenw.) Raillo, Fungi of the Genus *Fusarium*: 278. 1950.
- Dialonectria ullevolea* Seifert & Gräfenhan, Stud. Mycol. 68: 97. 2011.
- Holotypus*: In B *fide* Hein (1988).
- Type locality*: **Japan**, Tokyo.
- Type substrate*: Old stroma of *Dothideaceae*, on *Bambusa* sp. branches with *Zythia stromaticola*.
- Notes*: Synonym *fide* Wollenweber & Reinking (1935) and Gräfenhan *et al.* (2011). The older epithet '*volutella*' (1891) supersedes the epithet '*ullevolea*' (2011) and, therefore, a new combination is provided.
- stromaticum* *Fusarium* Delacr., Bull. Soc. Mycol. France 9: 186. 1893.
- (See ***Fusarium heterosporum***)
- Holotypus*: ?PC.
- Type locality*: **France**, overseas department of Mayotte, Mayotte islands.

Type substrate: Seeds of unknown *Poaceae* (= *Gramineae*).

Note: Synonym *fide* Wollenweber & Reinking (1935).

subcarneum *Fusarium* P. Crouan & H. Crouan, Fl. Finistère: 14. 1867, *nom. rej.*

(See *Fusarium sambucinum*)

Authentic material: ?PC.

Original locality: **France**, Brittany, Finistère, marshes.

Original substrate: Twigs and dead leaves of *Ulex* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

subcorticale *Fusarium* Oudem., Ned. Kruidk. Arch., sér. 3, 3: 135. 1898.

(See *Fusarium buxicola*)

Holotypus: ?L.

Type locality: **Netherlands**, Zuid-Holland Province, Zorgvliet.

Type substrate: *Buxus sempervirens*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

subglutinans* *Fusarium (Wollenw. & Reinking) P.E. Nelson *et al.*, *Fusarium* species. An illustrated manual for identification: 135. 1983.

Basionym: *Fusarium moniliforme* var. *subglutinans* Wollenw. & Reinking, *Phytopathology* 15: 163. 1925.

Synonyms: *Fusarium moniliforme* f. *subglutinans* (Wollenw. & Reinking) C. Moreau, *Rev. Mycol. (Paris)* 17: 23. 1952.

Fusarium sacchari var. *subglutinans* (Wollenw. & Reinking) Nirenberg, *Mitt. Biol. Bundesanst. Land- Forstw.* 169: 53. 1976.

Gibberella fujikuroi var. *subglutinans* (Wollenw. & Reinking) E.T. Edwards, *Agric. Gaz. New South Wales* 44: 895. 1933 (Art. F.8.1, Note 2, Exs. 2).

Gibberella subglutinans (Wollenw. & Reinking) P.E. Nelson *et al.*, *Fusarium* species. An illustrated manual for identification (University Park): 135. 1983.

Neotypus: CBS 747.97 (preserved as metabolically inactive culture), designated by Yilmaz *et al.* (2021).

Ex-neotype culture: BBA 62451 = CBS 747.97 = DAOM 225141 = FRC M-36 = MRC 8554 = NRRL 22016 = NRRL 22114.

Neotype locality: **USA**, Illinois, Saint Elmo.

Neotype substrate: *Zea mays*.

Descriptions and illustrations: See Booth (1971), Nirenberg (1976, 1981), Nelson *et al.* (1983), Pascoe (1990), Leslie & Summerell (2006).

Diagnostic DNA barcodes: *rpb1*: JX171486; *rpb2*: JX171599; *tef1*: HM057336.

sublunatum* *Fusarium Reinking, *Zentralbl. Bakteriol.*, 2. Abt. 89: 510. 1934.

Synonyms: *Fusarium elongatum* Reinking, *Zentralbl. Bakteriol. Parasitenk.*, Abt. 2, 89: 511. 1934, *nom. illegit.*, Art. 53.1.

Fusarium sambucinum var. *sublunatum* (Reinking) Biláň, *Mikrobiol. Zhurn.* 49: 6. 1987, *nom. inval.*, Art. 41.4, Note 1.

Lectotypus (*hic designatus*, MBT 10000750): **Costa Rica**, Limon, soil in *Musa sapientum* plantation, 1933, O.A. Reinking, CBS 189.34 (preserved as metabolically inactive culture).

Ex-type culture: BBA 62431 = CBS 189.34 = DSM 62431 = NRRL 13384 = NRRL 20840.

Descriptions and illustrations: See Gerlach & Nirenberg (1982).

Diagnostic DNA barcodes: *rpb1*: JX171451; *rpb2*: KX302935; *tef1*: KX302919.

Notes: No holotype specimen could be located for *F. sublunatum* and therefore the metabolically inactive culture CBS 189.34 (=

IMB 5238), which represents the ex-type culture (Gerlach & Nirenberg 1982), is designated as lectotype.

subnivale *Fusarium* Höhn., in Penther & Zederbauer, *Ann. K. K. Naturhist. Hofmus.* 20: 369. 1905.

(See *Fusarium dimerum*)

Holotypus: FH00965354.

Type locality: **Turkey**, Anatolia.

Type substrate: Stems and leaves of decayed *Astragalus* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

subpallidum *Fusarium* Sherb., *Mem. Cornell Univ. Agric. Exp. Sta. Mem.* 6: 230. 1915.

(See *Fusarium sambucinum*)

Typus: ?CUP-007480.

Type locality: **USA**, Louisiana, Edgerton.

Type substrate: *Solanum tuberosum*.

Notes: Synonym *fide* Wollenweber & Reinking (1935). Lectotypification pending study of material lodged in CUP.

subtectum *Fusarium* Roberge ex Desm., *Pl. Crypt. N. France*, ed. 1, Fasc. 29, no. 1428. 1845.

Rhodesia subtecta (Roberge ex Desm.) Grove, *British Stem- and Leaf-Fungi (Coelomycetes)* 2: 205. 1937.

Synonyms: *Myxosporina subtecta* (Roberge ex Desm.) Höhn., in Weese, *Ber. Deutsch. Bot. Ges.* 37: 155. 1919, *nom. inval.*, Art. 35.1.

Myxosporina subtecta (Roberge ex Desm.) Höhn., *Mitt. Bot. Inst. Tech. Hochsch. Wien* 4: 74. 1927.

Hainesia subtecta (Roberge ex Desm.) Grove, *J. Bot.* 70: 4. 1932.

Hymenula psammae Oudem., *Ned. Kruidk. Arch.*, sér. 3, 1: 533. 1898. (*fide* Wollenweber & Reinking 1935).

Syntypes: *Pl. Crypt. N. France* no. 1428 in ?BRU, PC & PH.

Type locality: **France**.

Type substrate: Dead leaves of *Arundo arenaria*.

subtropicale* *Fusarium C. Pereira *et al.*, *Mycologia* 110: 864. 2018.

Holotypus: BPI 910644.

Ex-type culture: CBS 144706 = NRRL 66764.

Type locality: **Brazil**, Paraná State, Guarapuava.

Type substrate: *Hordeum vulgare*.

Descriptions and illustrations: See Pereira *et al.* (2018).

Diagnostic DNA barcodes: *rpb1*: MH706972; *rpb2*: MH706973; *tef1*: MH706974.

subulatum *Fusarium* Appel & Wollenw., *Arbeiten Kaiserl. Biol. Anst. Land- Forstw.* 8: 131. 1913.

Replaced synonym: *Fusarium roseum* var. *lupini-albi* Sacc., *Michelia* 2: 295. 1881.

(See *Fusarium avenaceum*)

Holotypus: Not located.

Type locality: **Italy**, Selva.

Type substrate: *Lupinus albus*.

Note: Synonyms *fide* Wollenweber & Reinking (1935).

subviolaceum *Fusarium* Roum. & Fautrey, *Fungi Sel. Gall. Exs.* no. 6022. 1892.

(See *Fusarium avenaceum*)

Syntype: ILL0020193 (*Fungi Sel. Gall. Exs.* no. 6022).

Type locality: **France**, Jardin de Noidan.

Type substrate: Dry stems of *Asparagus officinalis*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

succisae Fusarium Schröt. ex Sacc., Syll. Fung. 10: 724. 1892.
 Synonym: *Fusisporium succisae* J. Schröt., Hedwigia 13: 180. 1874, *nom. inval.*, Art. 36.1(a).

Lectotypus: ILL00076313 (Thümen, Mycoth. Univ. no. 675), designated by [Yilmaz et al. \(2021\)](#).

Lectotype locality: **Germany**, Bavaria, Borussia.

Lectotype substrate: *Succisa pratensis*.

Epitypus: IMI 202876, designated by [Yilmaz et al. \(2021\)](#).

Ex-epitype culture: BBA 12287 = BBA 63627 = CBS 219.76 = DAOM 225142 = IMI 202876 = IMI 375347 = NRRL 13613.

Epitype locality: **Germany**.

Epitype substrate: *Succisa pratensis*.

Descriptions and illustrations: See [Nirenberg \(1976\)](#), [Gerlach & Nirenberg \(1982\)](#).

Diagnostic DNA barcodes: *rpb1*: LT996207; *rpb2*: LT970764.

sudanense Fusarium S.A. Ahmed et al., Antonie van Leeuwenhoek 110: 826. 2017.

Holotypus: CBS H-22547.

Ex-type culture: CBS 454.97.

Type locality: **Sudan**.

Type substrate: Plant debris of *Striga hermonthica*.

Descriptions and illustrations: See [Moussa et al. \(2017\)](#).

Diagnostic DNA barcodes: *rpb1*: LT996208; *rpb2*: LT996155; *tef1*: KU711697.

sulawesiense Fusarium Maryani et al. (as 'sulawense'), Persoonia 43: 65. 2019.

Holotypus: InaCC F940 (preserved as metabolically inactive culture).

Ex-type culture: InaCC F940.

Type locality: **Indonesia**, South Sulawesi, Bone, Kecamatan Bongo, Desa Selli.

Type substrate: Infected pseudostem of *Musa acuminata* var. Pisang Cere (AAA).

Descriptions and illustrations: See [Maryani et al. \(2019b\)](#).

Diagnostic DNA barcodes: *rpb2*: LS479855; *tef1*: LS479443.

sulphureum Fusarium Schldl., Fl. Berol. 2: 139. 1824, *nom. rej.* (See ***Fusarium sambucinum***)

Holotypus: HAL 1613 F.

Type locality: **Germany**, Berlin.

Type substrate: Rotting tuber of *Solanum tuberosum*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

suttonianum Fusarium (Sand.-Den. & Crous) O'Donnell et al., Index Fungorum 440: 4. 2020.

Neocosmospora suttoniana Sand.-Den. & Crous, Persoonia 41: 123. 2018.

Holotypus: CBS H-23224.

Ex-type culture: CBS 143214 = FRC S-1423 = NRRL 32858.

Type locality: **USA**, Louisiana.

Type substrate: *Homo sapiens*.

Descriptions and illustrations: See [Sandoval-Denis & Crous \(2018\)](#).

Diagnostic DNA barcodes: *rpb1*: MW218138; *rpb2*: EU329630; *tef1*: DQ247163.

tabacinum Fusarium (J.F.H. Beyma) W. Gams, Persoonia 5: 179. 1968.

Basionym: *Cephalosporium tabacinum* J.F.H. Beyma, Zentralbl. Bakteriolog. 2. Abt. 89: 240. 1933.

Plectosphaerella cucumerina (Lindf.) W. Gams, in Domsch & Gams, *Fungi in Agricultural Soils*: 160. 1972.

Basionym: *Venturia cucumerina* Lindf., Meddn Centralanst. Försksv. Jordbruksomr. Bot. Avd. 17: 7. 1919.

Synonyms: *Monographella cucumerina* (Lindf.) Arx, Trans. Brit. Mycol. Soc. 83: 374. 1984.

Microdochium tabacinum (J.F.H. Beyma) Arx, Trans. Brit. Mycol. Soc. 83: 374. 1984.

Plectosporium tabacinum (J.F.H. Beyma) M.E. Palm, W. Gams & Nirenberg, Mycologia 87: 399. 1995.

Plectosphaerella cucumeris Kleb., Phytopathol. Z. 1: 43. 1929.

Micronectriella cucumeris (Kleb.) C. Booth, The Genus Fusarium: 39. 1971.

Cephalosporium ciferrii Verona, *Studio sulle cause microbiche che danneggiano la carta ed i libri*: 30. 1939.

Cephalosporiopsis imperfecta M. Moreau & Moreau, Rev. Mycol. (Paris) 6: 67. 1941, *nom. inval.*, Art. 39.1.

Neotypus: CBS H-7656, designated in [Palm et al. \(1995\)](#).

Ex-neotype culture: CBS 137.33 = MUCL 9701 = NRRL 22455.

Neotype locality: **UK**, England, Bristol.

Neotype substrate: Stems of *Nicotiana tabacum*.

Descriptions and illustrations: See [Domsch et al. \(2007\)](#), [Carlucci et al. \(2012\)](#), [Giraldo & Crous \(2019\)](#).

tabacivorum Fusarium Delacr., Ann. Inst. Natl. Agron., ser. 2, 5: 207. 1906.

(See ***Fusarium oxysporum***)

Holotypus: ?PC.

Type locality: **France**, Périgueux, Razac-sur-l'Isle.

Type substrate: *Nicotiana tabacum*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

tanahbumbuense Fusarium Maryani et al., Persoonia 43: 63. 2019.

Holotypus: InaCC F965 (preserved as metabolically inactive culture).

Ex-type culture: InaCC F965.

Type locality: **Indonesia**, South Kalimantan, Tanah Bumbu, Kecamatan Kusan Hilir, Desa Betung.

Type substrate: Pseudostem of *Musa* var. Pisang Hawa.

Descriptions and illustrations: See [Maryani et al. \(2019b\)](#).

Diagnostic DNA barcodes: *rpb1*: LS479877; *rpb2*: LS479863; *tef1*: LS479448.

tardichlamydosporum Fusarium Maryani et al., Stud. Mycol. 92: 181. 2018 [2019].

Holotypus: InaCC F958 (preserved as metabolically inactive culture).

Ex-type culture: InaCC F958.

Type locality: **Indonesia**, East Nusa Tenggara, Sikka Flores, Desa Kota Uneng Kecamatan Alok.

Type substrate: Pseudostem of *Musa acuminata* var. Pisang Barangan.

Descriptions and illustrations: See [Maryani et al. \(2019a\)](#).

Diagnostic DNA barcodes: *rpb1*: LS479534; *rpb2*: LS479280; *tef1*: LS479729.

tardicrescens Fusarium Maryani et al., Persoonia 43: 69. 2019.

Synonym: *Fusarium tardicrescens* Maryani et al., Stud. Mycol. 92: 185. 2018 [2019], *nom. inval.*, Art. 40.7.

Holotypus: CBS 102024 (preserved as metabolically inactive culture).

Ex-type culture: CBS 102024 = NRRL 36113.

Type locality: **Malawi**, Karonga, Misuku Hills.

Type substrate: *Musa sapientum* cv. Harare.

Descriptions and illustrations: See [Maryani et al. \(2019b\)](#).

Diagnostic DNA barcodes: *rpb1*: LS479474; *rpb2*: LS479217; *tef1*: LS479665.

tasmaniense *Fusarium* (McAlpine) Rossman (as '*tasmanica*'), Mycol. Pap. 150: 54. 1983.

Basionym: *Microcera tasmaniensis* McAlpine, J. Dept. Agric. Victoria 2: 647. 1904.

Synonyms: *Discofusarium tasmaniense* (McAlpine) Petch, Trans. Brit. Mycol. Soc. 7: 143, 165. 1921.

Microcera myrtilaspis McAlpine, J. Dept. Agric. Victoria 2: 647. 1904.

Calonectria coccidophaga Petch, Trans. Brit. Mycol. Soc. 7: 161. 1921.

Nectria coccidophaga (Petch) Rossman, Mycotaxon 8: 499. 1979.

Holotypus: VPRI 2744.

Type locality: **Australia**, Tasmania.

Type substrate: Parasitic on *Aspidiotus* sp. (scale) on *Eucalyptus* bark.

Descriptions and illustrations: See [Rossman \(1983\)](#).

Notes: Status unclear. [Rossman \(1983\)](#) studied the specimen in K(M) and recombined the asexual morph name in *Fusarium*, which is not supported by the features of the sexual-morph. This species most likely belongs to *Microcera* as originally specified by [McAlpine \(1904\)](#).

temperatum* *Fusarium Scaufl. & Munaut, Mycologia 103: 593. 2011.

Holotypus: MUCL 52463-H.

Ex-type culture: MUCL 52463.

Type locality: **Belgium**, Waals-Brabant Province, Chastre.

Type substrate: *Zea mays*.

Descriptions and illustrations: See [Scauflaire et al. \(2011\)](#).

Diagnostic DNA barcode: *tef1*: KM487197.

tenellum *Fusarium* Sacc. & Briard, Rev. Mycol. (Toulouse) 7: 212. 1885.

(See ***Fusarium sambucinum***)

Holotypus: Not located.

Type locality: **France**, Troyes.

Type substrate: Rotten stem of *Brassica oleracea*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

tenue *Fusarium* Corda, Icon. Fung. 1: 3. 1837.

(See ***Fusarium avenaceum***)

Typus: In PRM *fide* [Pilat \(1938\)](#).

Type locality: **Czech Republic**, near Prague.

Type substrate: Rotting stem of an unidentified host.

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#). Lectotypification pending study of material lodged in PRM.

tenuicristatum *Fusarium* (S. Ueda & Udagawa) O'Donnell et al., Index Fungorum 440: 4. 2020.

Basionym: *Neocosmospora tenuicristata* S. Ueda & Udagawa, Mycotaxon 16: 387. 1983.

Synonym: *Acremonium tenuicristatum* S. Ueda & Udagawa, Mycotaxon 16: 387. 1983.

Holotypus: NHL 2911.

Type locality: **Japan**, Nagasaki.

Type substrate: Marine sludge.

Descriptions and illustrations: See [Ueda & Udagawa \(1983\)](#).

Notes: Status unclear. See [Sandoval-Denis et al. \(2019\)](#).

tenuissimum *Fusarium* (Peck) Sacc., Syll. Fung. 4: 711. 1886. *Basionym*: *Fusisporium tenuissimum* Peck, Rep. (Annual) New York State Mus. Nat. Hist. 34: 48. 1883. 1881.

(See ***Fusarium sambucinum***)

Holotypus: NYSf3163.

Type locality: **USA**, New York, Schenectady.

Type substrate: Dead stem of unidentified host.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

tenuistipes *Fusarium* Sacc., Atti Mem. Reale Accad. Sci. Lett. Arti, Padova 33: 195. 1917.

(See ***Fusarium incarnatum***)

Holotypus: In PAD.

Type locality: **Unknown**.

Type substrate: *Pennisetum spicatum*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

terrestre *Fusarium* Manns, Bull. North Dakota Agric. Exp. Sta.: no. 259. 1932.

(See ***Fusarium equiseti***)

Holotypus: Not located.

Type locality: **USA**, North Dakota.

Type substrate: Soil.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

terricola* *Fusarium Al-Hatmi et al., Antonie van Leeuwenhoek 110: 826. 2017.

Holotypus: CBS H-22548.

Ex-type culture: CBS 483.94.

Type locality: **Australia**, Queensland.

Type substrate: Desert soil.

Descriptions and illustrations: See [Moussa et al. \(2017\)](#).

Diagnostic DNA barcodes: *rpb1*: LT996209; *rpb2*: LT996156; *tef1*: KU711698.

thapsinum* *Fusarium Klittich et al., Mycologia 89: 644. 1997.

Synonym: *Gibberella thapsina* Klittich et al., Mycologia 89: 643. 1997.

Holotypus: BPI 737885.

Ex-type culture: ATCC 200522 = CBS 777.96 = FRC M-6564.

Type locality: **USA**, Kansas.

Type substrate: Stalk of *Sorghum* sp.

Descriptions and illustrations: See [Klittich et al. \(1997\)](#).

Diagnostic DNA barcodes: *rpb1*: MW928818; *rpb2*: MW928833; *tef1*: MW928844.

theobromae *Fusarium* Appel & Strunk, Centralbl. Bacteriol., 2. Abth., 11: 635. 1904.

Neocosmospora theobromae (Appel & Strunk) Sand.-Den. & Crous, Persoonia 43: 174. 2019.

Synonyms: *Fusarium javanicum* var. *theobromae* (Appel & Strunk) Wollenw., Z. Parasitenk. (Berlin) 3: 483. 1931.

Neotypus: BPI 453072, designated in [Sandoval-Denis et al. \(2019\)](#).

Type locality: **Cameroon**, Victoria.

Type substrate: Fruits and seeds of *Theobroma cacao*.

Descriptions and illustrations: See [Sandoval-Denis et al. \(2019\)](#).

Diagnostic DNA barcode: *tef1*: LR583660.

Notes: This *Fusarium* name was recently resurrected, neotypified, and transferred to *Neocosmospora* by [Sandoval-Denis et al. \(2019\)](#). DNA barcodes were generated from the neotype

specimen; however, fresh collections are needed for epitypification.

theobromae *Fusarium* M.L. Lutz, Bull. Soc. Bot. France 53: L. 1907 [1906], *nom. illegit.*, Art. 53.1.

Diplocladium theobromae Sacc. & Trotter, Syll. Fung. 22: 1309. 1913.

Authentic material: Not located.

Original locality: **Democratic Republic of São Tomé and Príncipe**.

Original substrate: Fermented beans of *Theobroma cacao*.

Note: Originally erroneously assigned to the genus *Fusarium*.

thevetiae *Fusarium* Tassi, Atti Reale Accad. Fisiocrit. Siena, sér. 4, 8: 238. 1897.

Holotypus: ?SIENA.

Type locality: **India**.

Type substrate: *Thevetia venenifera*.

Notes: Status unclear. A doubtful species *vide* Wollenweber & Reinking (1935).

thuemenii *Fusarium* Sacc., Syll. Fung. 4: 722. 1886.

Replaced synonym: *Fusarium parasiticum* Thüm., Nuovo Giorn. Bot. Ital. 12: 198. 1880, *nom. illegit.*, Art. 53.1.

(See ***Fusarium oxysporum***)

Holotypus: Not located.

Type locality: **Russia**, Orenburg.

Type substrate: Rotten branches of *Betula verrucosa* (= *Betula pendula*).

Note: Synonym *vide* Wollenweber & Reinking (1935).

tjaetaba* *Fusarium T.T.H. Vu *et al.*, Fungal Diversity 77: 361. 2015 [2016].

Holotypus: RBG 5361.

Ex-type culture: FRL14350 = NRRL 66243 = RBG 5361.

Type locality: **Australia**, Northern Territory, Litchfield National Park.

Type substrate: *Sorghum interjectum*.

Descriptions and illustrations: See Laurence *et al.* (2016).

Diagnostic DNA barcodes: *rpb1*: KP083267; *rpb2*: KP083275; *tef1*: KP083263.

tjaynera* *Fusarium J.L. Walsh *et al.*, Fungal Diversity 77: 361. 2015 [2016].

Holotypus: RBG 5367.

Ex-type culture: NRRL 66246 = RBG 5367.

Type locality: **Australia**, Northern Territory, Litchfield National Park.

Type substrate: *Triodia microstachya*.

Descriptions and illustrations: See Laurence *et al.* (2016).

Diagnostic DNA barcodes: *rpb1*: KP083268; *rpb2*: KP083279; *tef1*: EF107152.

tomentosum *Fusarium* Berk. & M.A. Curtis, J. Linn. Soc., Bot. 10: 359. 1868 [1869].

Holotypus: In K(M).

Type locality: **Cuba**.

Type substrate: Dead sticks.

Notes: Status unclear. Not *Fusarium* *vide* Wollenweber & Reinking (1935).

tonkinense *Fusarium* (Bugnic.) O'Donnell *et al.*, Index Fungorum 440: 4. 2020.

Neocosmospora tonkinensis (Bugnic.) Sand.-Den. & Crous, Personia 41: 126. 2018.

Basionym: *Cylindrocarpon tonkinense* Bugnic., Encyclop. Mycol. 11: 181. 1939.

Synonym: *Fusarium ershadii* M. Papizadeh *et al.*, Europ. J. Pl. Pathol. 151: 693. 2018, *nom. illegit.*, Art. 52.1.

Holotypus: IMI 113868.

Ex-type culture: CBS 115.40 = IMI 113868.

Type locality: **Vietnam**, Tonkin.

Type substrate: *Musa sapientum*.

Diagnostic DNA barcodes: *rpb1*: MW218140; *rpb2*: LT960564; *tef1*: LT906672.

torreyae* *Fusarium T. Aoki *et al.*, Mycologia 105: 314. 2013.

Holotypus: BPI 884050.

Ex-type culture: CBS 133858 = MAFF 243468 = NRRL 54151.

Type locality: **USA**, Florida, Liberty County, Torreya State Park, Aspalaga Tract.

Type substrate: Stem tissue of diseased *Torreya taxifolia*.

Descriptions and illustrations: See Aoki *et al.* (2013).

Diagnostic DNA barcodes: *rpb1*: MW928819; *rpb2*: MW928834; *tef1*: MW928845.

tortuosum *Fusarium* Thüm. & Pass., Pilze Weinst.: 51. 1878.

Neofabraea vagabunda (Desm.) P.R. Johnst., IMA Fungus 5: 103. 2014.

Basionym: *Phlyctema vagabunda* Desm., Ann. Sci. Nat., Bot., sér. 3, 8: 16. 1847.

Synonyms: *Rhabdospora vagabunda* (Desm.) Zerov, Vznachnik gribiv Ukraini. T. 3. Nezaversheni gribi: 501. 1971, *nom. inval.*, Art. 41.1.

Rhabdospora vagabunda (Desm.) R.S. Mathur, *Coelomycetes of India*: 234. 1979.

Gloeosporium tortuosum (Thüm. & Pass.) Sacc., *Michelia* 2: 117. 1880.

Myxosporium tortuosum (Thüm. & Pass.) Allesch., Rabenh. Krypt.-Fl., Ed. 2, 1(7): 534. 1903.

?*Fusarium obtusatum* Corda, Icon. Fung. 1: 3. 1837.

Fusarium bipunctatum Preuss, *Linnaea* 25: 741. 1852.

Lituaria riessii Schulzer, Verh. K. K. Zool.-Bot. Ges. Wien 21: 1241. 1871.

Gloeosporium riessii (Schulzer) Schulzer & Sacc., *Hedwigia* 23: 110. 1884.

Gloeosporium tineum Sacc., *Michelia* 1: 219. 1878.

Gloeosporium frigidum Sacc., *Michelia* 2: 168. 1880.

Cylindrosporium frigidum (Sacc.) Vassiljevsky, *Fungi Imperfecti Parasitici* 2: 515. 1950.

Gloeosporium pyrenoides Sacc. & Malbr., in Saccardo, *Michelia* 2: 633. 1882.

Gloeosporium phillyreae Pass., Atti Reale Accad. Lincei, Rendiconti Cl. Sci. Fis., sér. 4, 4: 103. 1888.

Gloeosporium allantosporum Fautrey, Rev. Mycol. (Toulouse) 14: 97. 1892.

Gloeosporium allantoides Peck, Rep. (Annual) Regents Univ. State New York New York State Mus. 45: 81. 1893.

Gloeosporium alutaceum Sacc., *Malpighia* 11: 317. 1897.

Allantozythia alutacea (Sacc.) Höhn., Ann. Mycol. 22: 203. 1924.

Phlyctema alutacea (Sacc.) Petr., Ann. Mycol. 27: 370. 1929.

Fusarium japonicum Allesch., Beibl. Hedwigia 36: (164). 1897.

Gloeosporium unedonis Traverso, R.C. Congr. Bot. Palermo, 1902: 3 (extr.). 1902.

Trichoseptoria fructigena Maubl., Bull. Trimestriel Soc. Mycol. France 21: 95. 1905.

Gloeosporium begunotii Sacc., in Potebnia, Ann. Mycol. 5: 20.

1907.

Cylindrosporium olivae Petri, Ann. Mycol. 5: 324. 1907.*Gloeosporium olivae* (Petri) Foschi, Ann. Sperim. Agrar, n.s. 9: 911. 1955.*Gloeosporium album* Osterw., Centralbl. Bacteriol. Parasitenk., 2. Abth., 18: 826. 1907.*Gloeosporium diervillae* Grove, J. Bot. 60: 145. 1922.*Pezicula alba* E.J. Guthrie, Trans. Brit. Mycol. Soc. 42: 504. 1959.*Neofabraea alba* (E.J. Guthrie) Verkley, Stud. Mycol. 44: 125. 1999.

Holotypus: ?PARMA.

Type locality: **Italy**, Parma.Type substrate: Dry twigs of *Vitis vinifera*.Note: Synonyms fide [Wollenweber & Reinking \(1935\)](#).*torulosum* *Fusarium* (Berk. & M.A. Curtis) Gruyter & J.H.M. Schneid., Jaarb. Plantenziektenkundige Dienst, Wageningen 1989/1990, no. 168: 135. 1991, *nom. inval.*, Art. 41.4.Basionym: *Fusidium torulosum* Berk. & M.A. Curtis, Grevillea 3: 112. 1875.(See *Fusarium torulosum* (Berk. & M.A. Curtis) Nirenberg)*torulosum* *Fusarium* (Berk. & M.A. Curtis) Nirenberg, Mycopathologia 129: 136. 1995.Basionym: *Fusidium torulosum* Berk. & M.A. Curtis, Grevillea 3: 112. 1875.Synonyms: *Fusoma torulosum* (Berk. & M.A. Curtis) Sacc., Syll. Fung. 4: 220. 1886.*Fusarium torulosum* (Berk. & M.A. Curtis) Gruyter & J.H.M. Schneid., Jaarboek. Plantenziektenkundige Dienst. Wageningen 1989/1990 no. 168: 135. 1991, *nom. inval.*, Art. 41.4.*Fusarium sclerodermatis* Oudem., Nederl. Kruidk. Arch. ser. 2, 5: 516. 1889.*Fusarium sambucinum* var. *coeruleum* Wollenw., Ann. Mycol. 15: 55. 1917.? *Gibberella pulicaris* var. *minor* Wollenw., Z. Parasitenk. (Berlin) 3: 356. 1931.

Syntype: ?Car Inf. no. 6034. in K(M).

Type locality: **USA**, Pennsylvania, Michener.Type substrate: Decaying *Brassica* stalks or *Pinus*.Descriptions and illustrations: See [Nirenberg \(1995\)](#).*toxicum* *Fusarium* L. Lombard & J.W. Xia, Persoonia 43: 220. 2019.

Holotypus: CBS H-24071.

Ex-type culture: CBS 406.86 = FRC R-8507 = IMI 309347 = NRRL 25796.

Type locality: **Germany**, Berlin.

Type substrate: Soil.

Descriptions and illustrations: See [Xia et al. \(2019\)](#).Diagnostic DNA barcodes: *rpb2*: MN170441; *tef1*: MN170508.*tracheiphilum* *Fusarium* (E.F. Sm.) Wollenw., Phytopathology 3: 29. 1913.Basionym: *Neocosmospora vasinfecta* var. *tracheiphila* E.F. Sm., Bull. Div. Veg. Physiol. Pathol. U.S.D.A. 17: 45. 1899.(See *Fusarium neocosmosporiellum*)

Syntypes: IN BPI, F, FLAS, ISC, MICH, PUL, UC & WSP.

Type locality: **USA**, South Carolina, James Island.Type substrate: Dead stem of *Vigna sinensis*.Note: Published as a new name for the sporodochial morph found on the authentic material of *N. vasinfecta* var. *tracheiphila*.*translucentum* *Fusarium* Berk. & Broome, Ann. Mag. Nat. Hist., ser. 4, 17: 141. 1876.

Holotypus: ?K(M).

Type locality: **UK**, Scotland, Glamis.

Type substrate: Wood.

Notes: Status unclear. Not *Fusarium* fide [Wollenweber & Reinking \(1935\)](#).*transvaalense* *Fusarium* Sand.-Den. et al., MycoKeys 34: 82. 2018.

Holotypus: CBS H-23497.

Ex-type culture: CBS 144211.

Type locality: **South Africa**, Kruger National Park, Skukuza, Granite Supersite.Type substrate: Rhizosphere of *Sida cordifolia*.Descriptions and illustrations: See [Sandoval-Denis et al. \(2018b\)](#).Diagnostic DNA barcodes: *rpb1*: LT996210; *rpb2*: LT996157; *tef1*: LT996099.*tremelloides* *Fusarium* Grev., Scott. Crypt. Fl. 1: 10. 1822.*Calloria tremelloides* (Grev.) L. Lombard, *comb. nov.* MycoBank MB 837723.Basionym: *Fusarium tremelloides* Grev., Scott. Crypt. Fl. 1: 10. 1822.Synonyms: *Peziza fusarioides* Berk., Mag. Zool. Bot. 1: 46. 1837.*Calloria fusarioides* (Berk.) Fr., Summa Veg. Scand. 2: 359. 1849.*Callorina fusarioides* (Berk.) Korf, Phytologia 21: 203. 1971.*Peziza neglecta* Lib., Pl. Crypt. Arduenna Fasc. 1: no. 29. 1830.*Calloria neglecta* (Lib.) B. Hein, Beih. Willdenowia 9: 54. 1976.

Holotypus: Not located.

Type locality: **UK**, Scotland, near Edinburg.Type substrate: Dead stems of *Urtica dioica*.Notes: Synonyms fide [Wollenweber & Reinking \(1935\)](#). As the epithet of *F. tremelloides* (1822) takes priority above the epithet of *C. neglecta* (1830), a new combination is introduced here.*trichothecioides* *Fusarium* Wollenw., J. Wash. Acad. Sci. 2: 147. 1912.Synonyms: *Fusarium sambucinum* var. *trichothecioides* (Wollenw.) Bilai, Fusarii (Biologija i sistematika): 268. 1955, *nom. inval.*, Art. 41.1.*Fusarium tuberivorum* Wilcox & G.K. Link, Res. Bull. Nebraska Agric. Exp. Sta. 1: 48. 1913.Lectotypus (*hic designatus*, MBT 10000751): **USA**, rotten tuber of *Solanum tuberosum*, Aug. 1912, H.W. Wollenweber, in J. Wash. Acad. Sci. 2: 150, figs A–F.Descriptions and illustrations: See [Booth \(1971\)](#) and [Gerlach & Nirenberg \(1982\)](#).Notes: A putative synonym of *F. sulphureum* ([Gordon 1959](#), [Subramanian 1971](#), [Gerlach & Nirenberg 1982](#)) or *F. sambucinum* ([Nelson et al. 1983](#), [Nirenberg 1995](#)). The taxonomy of this potato pathogen has not yet been resolved. As no holotype specimen was preserved ([Gerlach & Nirenberg 1982](#)), the figures accompanying the original protologue are designated as lectotype here.*tricinctum* *Fusarium* (Corda) Sacc., Syll. Fung. 4: 700. 1886.Basionym: *Selenosporium tricinctum* Corda, Icon. Fung. 2: 7. 1838.Synonyms: *Fusarium sporotrichioides* var. *tricinctum* (Corda) Raillo, Fungi of the Genus *Fusarium*: 197. 1950.

Fusarium sporotrichiella var. *tricinctum* (Corda) Bilař, Yadovitye griby na zerne khibnykh zlakov. Kiev: 87. 1953, *nom. inval.*, Art. 39.1.

Fusarium sporotrichiella var. *tricinctum* (Corda) Bilař, Mikrobiol. Zhurn. 49: 7. 1987, *nom. inval.*, Art. 35.1.

?*Vermicularia subeffigurata* γ *helianthi* Schwein., Trans. Amer. Philos. Soc., n.s. 4: 228. 1832 [1834].

?*Fusarium helianthi* (Schwein.) Wollenw., Fusaria Autogr. Delin. 2: 555. 1924.

Fusarium muentzii Delacr. (as 'müntzii'), Bull. Soc. Mycol. France 8: 192. 1892.

Fusarium citrifforme Jamal., Valt. Maatalousk. Julk. 123: 11. 1943.

Gibberella tricincta El-Gholl *et al.*, Canad. J. Bot. 56: 2206. 1978.

Lectotypus: PRM 155623 (designated in Holubová-Jechová *et al.* 1994).

Type locality: **Czech Republic**, near Prague, Chuchle, Vyskořilka.

Type substrate: Stem of *Umbelliferae*.

Epitypus: In PRM, designated in Holubová-Jechová *et al.* (1994).

Ex-epitype culture: BBA 64485 = CBS 393.93 = NRRL 25481.

Epitype locality: **Germany**, Berlin.

Epitype substrate: Culm base of *Triticum aestivum*.

Descriptions and illustrations: See Holubová-Jechová *et al.* (1994) and Leslie & Summerell (2006).

Diagnostic DNA barcodes: *rpb1*: JX171516; *rpb2*: JX171629; *tef1*: MH582379.

trifolii *Fusarium* Jacz., Jahrb. Pflanzenkrankh. Russlands. VII-VIII: Abt. 6. 1917.

(See *Fusarium oxysporum*)

Holotypus: Not located.

Type locality: **Russia**, St. Petersburg.

Type substrate: Root crown of *Trifolium* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

triseptatum *Fusarium* L. Lombard & Crous, Persoonia 43: 34. 2018 [2019].

Holotypus: CBS H-23622.

Ex-type culture: CBS 258.50 = NRRL 36389.

Type locality: **USA**.

Type substrate: *Ipomoea batatas*.

Descriptions and illustrations: See Lombard *et al.* (2019b).

Diagnostic DNA barcodes: *rpb1*: MW928820; *rpb2*: MH484873; *tef1*: MH484964.

tritici *Fusarium* Liebman bis, Tidsskr. Landoekon., n.s., 2: 515. 1840.

(See *Fusarium avenaceum*)

Lectotypus (*hic designates*, MBT 10000752): **Denmark**, *Triticum* sp., in Tidsskr. Landoekon., n.s., 2: figs B, 1, 2.

Notes: Synonymy *fide* Rostrup (1894). No holotype specimen could be located and therefore an illustration is designated as lectotype.

tritici *Fusarium* Erikss., Fungi Paras. Scand. Exs. no. 400. 1891, *nom. illegit.*, Art. 53.1.

(See *Fusarium nivale*)

Authentic material: CHR-F-0007556.

Original locality: **Sweden**, Stockholm.

Original substrate: *Triticum durum*.

Note: Synonym *fide* Wollenweber & Reinking (1935).

truncatum *Fusarium* Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 155. 1915.

(See *Fusarium avenaceum*)

Typus: ?CUP-007429.

Type locality: **USA**, New York.

Type substrate: *Solanum tuberosum*.

Note: Synonym *fide* Wollenweber & Reinking (1935). Lectotypification pending study of the material lodged in CUP.

tuaranense *Fusarium* T. Aoki *et al.*, Mycologia 111: 926. 2019.

Neocosmospora tuaranensis (T. Aoki *et al.*) L. Lombard & Sand.-Den., *comb. nov.* MycoBank MB 837724.

Basionym: *Fusarium tuaranense* T. Aoki *et al.*, Mycologia 111: 926. 2019.

Holotypus: BPI 910971.

Ex-type culture: ATCC 16563 = MAFF 246842 = NRRL 22231.

Type locality: **Malaysia**, Sabah State, Tuaran.

Type substrate: *Hevea brasiliensis* damaged by an unknown ambrosia beetle.

Descriptions and illustrations: See Aoki *et al.* (2019).

Diagnostic DNA barcodes: *rpb1*: KC691600; *rpb2*: KC691660, KC691631; *tef1*: KC691542.

Note: A new combination is provided in the genus *Neocosmospora* based on the phylogenetic relationship and morphology of this species (Aoki *et al.* 2019).

tubercularioides *Fusarium* (Corda) Sacc., Syll. Fung. 4: 697. 1886.

Basionym: *Selenosporium tubercularioides* Corda, Icon. Fung. 1: 7. 1837.

(See *Fusarium avenaceum*)

Typus: PRM 155625.

Type locality: **Czech Republic**, Liberec, Hamrřtejn (as 'Sudetenland, Reichenberg, Hammerstein').

Type substrate: Dead branches of *Rubus idaeus*.

Descriptions and illustrations: See Holubová-Jechová *et al.* (1994).

Note: Synonym *fide* Wollenweber & Reinking (1935). Lectotypification pending study of the material lodged in PRM.

tuberis *Fusarium* Preuss, Linnaea 24: 148. 1851.

Holotypus: In B *fide* Jülich (1974).

Type locality: **Germany**, Hoyerswerda.

Type substrate: Tuber of *Dahlia* sp.

Note: Status unclear. Not *Fusarium* *fide* Wollenweber & Reinking (1935).

tuberivorum *Fusarium* Wilcox & G.K. Link, Res. Bull. Nebraska Agric. Exp. Sta. 1: 48. 1913.

(See *Fusarium trichothecioides*)

Lectotypus (*hic designates*, MBT 10000753): **USA**, Nebraska, *Solanum tuberosum*, in Res. Bull. Nebraska Agric. Exp. Sta. 1, Pl. 24.

Notes: Synonym *fide* Wollenweber & Reinking (1935). No holotype specimen could be located and therefore an illustration is designated as lectotype.

tucumaniae *Fusarium* T. Aoki *et al.*, Mycologia 95: 664. 2003.

(See *Fusarium azukiicola*)

Holotypus: BPI 841955.

Ex-type culture: MAFF 238418 = MJ-172 = NRRL 31096.

Type locality: **Argentina**, Tucumán, San Agustín.

Type substrate: *Glycine max*.

Descriptions and illustrations: See Aoki *et al.* (2003).

Diagnostic DNA barcodes: *rpb1*: MAED01000445; *rpb2*: EU329557; *tef1*: GU170636.

tumidum *Fusarium* Sherb., *Phytopathology* 18: 148. 1928.

Synonym: *Gibberella tumida* P.G. Broadh. & P.R. Johnst., *Mycol. Res.* 98: 730. 1994.

Syntypes: Krieger, *Fungi Saxon.* Exs. no. 2499 in BPI & HAL.

Type locality: **Germany**.

Type substrate: Heads of *Sarothamnus scoparius*.

Note: Typification pending further study of the syntypes.

tupiense *Fusarium* C.S. Lima *et al.*, *Mycologia* 104: 1414. 2012.

Holotypus: CMB-UB 22068.

Ex-type culture: CML 262 = CMM 3655 = KSU 16195 = NRRL 53984.

Type locality: **Brazil**, Minas Gerais, Lavras.

Type substrate: Diseased tissue of *Mangifera indica*.

Descriptions and illustrations: See Lima *et al.* (2012).

Diagnostic DNA barcodes: *rpb1*: LR792583; *rpb2*: LR792619; *tef1*: GU737404.

udum *Fusarium* E.J. Butler, *Mem. Dept. Agric. India, Bot. Ser.* 2(9): 54. 1910.

Synonyms: *Fusarium oxysporum* f. sp. *udum* (E.J. Butler) W.C. Snyder & H.N. Hansen, *Amer. J. Bot.* 24: 66. 1940.

Fusarium butleri Wollenw., *Phytopathology* 3: 38. 1913, *nom. illegit.*, Art. 52.1.

Fusarium lateritium var. *uncinatum* (Wollenw.) Wollenw., *Z. Parasitenk. (Berlin)* 3: 375. 1931.

Fusarium vasinfectum var. *crotalariae* Kulkarni, *Indian J. Agric. Sci.* 4: 994. 1934.

Fusarium udum f. sp. *crotalariae* (Kulkarni) Subram., *The Genus Fusarium*: 114. 1971.

Fusarium udum var. *cajani* Padwick, *Indian J. Agric. Sci.* 10: 878. 1940.

Fusarium lateritium f. *cajani* (Padwick) W.L. Gordon, *Canad. J. Bot.* 30: 232. 1952.

Fusarium udum var. *crotalariae* Padwick, *Indian J. Agric. Sci.* 10: 877. 1940.

Fusarium lateritium f. *crotalariae* (Padwick) W.L. Gordon, *Canad. J. Bot.* 30: 232. 1952.

Gibberella indica B. Rai & R.S. Upadhyay, *Mycologia* 74: 343. 1982.

Lectotypus: Butler (1910), Pl. IV, fig. 4, designated in Pfenning *et al.* (2019).

Epitypus: UB23905, designated in Pfenning *et al.* (2019).

Ex-epitype culture: BBA 65058 = CML 3238 = NRRL 25199.

Type locality: **India**.

Type substrate: *Cajanus cajan*.

Descriptions and illustrations: See Wollenweber & Reinking (1935), Booth (1971), Subramanian (1971), Booth (1978), Gerlach & Nirenberg (1982) and Pfenning *et al.* (2019).

Diagnostic DNA barcodes: *rpb2*: KY498875; *tef1*: MK639096.

udum *Fusarium* (Berk.) Wollenw., *Phytopathology* 3: 38. 1913, *nom. illegit.*, Art. 53.1.

Basionym: *Fusisporium udum* Berk., *Ann. Mag. Nat. Hist.* 6: 438. 1841.

(See *Fusarium merismoides*)

Holotypus: ?K(M).

Type locality: **UK**, King's Cliffe.

Type substrate: Unidentified tree.

Note: Synonyms *fide* Wollenweber & Reinking (1935).

ulmi *Fusarium* P. Crouan & H. Crouan, *Fl. Finistère*: 14. 1867.

(See *Fusarium candidum* (Link) Sacc.)

Holotypus: ?PC.

Type locality: **France**, Finistère, edge of a stream.

Type substrate: Roots of *Ulmus* sp.

Note: Synonym *fide* Wollenweber & Reinking (1935).

ulmicola *Fusarium* Dearn. & House, *Circ. New York Stat. Mus.* 24: 60. 1940, *nom. inval.*, Art. 39.1.

Authentic material: NYSf3256.

Original locality: **USA**, New York, Albany, Ravena.

Original substrate: Dead branches of *Ulmus thomasii*.

Notes: Lacks a Latin diagnosis. Requires further investigation to confirm its taxonomic affiliation.

uncinatum *Fusarium* Wollenw., *Ann. Mycol.* 15: 54. 1917.

(See ***Fusarium udum***)

Holotypus: Not located.

Type locality: **India**, Dehli, Pusa.

Type substrate: Dried stem of *Cajanus indicus*.

Note: Synonym *fide* Wollenweber & Reinking (1935) and Gerlach & Nirenberg (1982).

uniseptatum *Fusarium* Höhn., *Ann. Mycol.* 1: 409. 1903.

Synonyms: *Cylindrocarpon uniseptatum* (Höhn.) Wollenw., *Fusaria Autogr. Delin.* 2: 646. 1924.

Ramularia uniseptata (Höhn.) Wollenw., *Fusaria Autogr. Delin.* 2: 646. 1924.

Holotypus: Not located.

Type locality: **Austria**, Vienna.

Type substrate: Rotten *Gleditsia triacanthos*.

Notes: Status unclear. Not *Fusarium* *fide* Wollenweber & Reinking (1935) and not *Ramularia* *fide* Braun (1998).

uredinicola *Fusarium* Jul. Müll., *Ber. Deutsch. Bot. Ges.* 3: 395. 1885.

(See ***Fusarium avenaceum***)

Holotypus: Not located.

Type locality: **Germany**.

Type substrate: Aecidium of *Phragmidium subcorticium* (= *Phragmidium mucronatum*) and *Phragmidium rubi* (= *Phragmidium barclayi*).

Note: Synonym *fide* Wollenweber & Reinking (1935).

uredinicola *Fusarium* Pat. & Gaillard, *Bull. Soc. Mycol. France* 4: 127. 1888, *nom. illegit.*, Art. 53.1.

Synonym: *Fusarium patouillardii* Sacc. (as 'patouillardii'), *Syll. Fung.* 10: 729. 1892.

Authentic material: Not located.

Original locality: **Venezuela**, Caracas.

Original substrate: Parasitic on the bottom of spots of *Puccinia pallidissima*, between the perithecia of *Darluca filum* parasitised by the *Puccinia* sp.

Notes: Status unclear. Not *Fusarium* *fide* Wollenweber & Reinking (1935).

uredinicola *Fusarium* Petch, *Ann. Roy. Bot. Gard. (Peradeniya)* 6: 256. 1917, *nom. illegit.*, Art. 53.1.

Authentic material: PDA 4731.

Original locality: **Sri Lanka**, Hakgala.

Original substrate: Parasitic on *Uredo microglossa* on leaves of *Microglossa zeylanica*.

Notes: Status unclear. A probable synonym of *F. solani* var. *minus* (syn. *Neocosmospora brevicona*) according to Wollenweber & Reinking (1935).

uredinophilum *Fusarium* Speg. (as 'urediniphilum'), *Anales Mus. Nac. Hist. Nat. Buenos Aires* 31: 445. 1922.

Holotypus: In LPS (Fungi Parag. pp. 93–94, no. 262).

Type locality: **Paraguay**, near Puerto Sajonia.

Type substrate: Parasitic on the acervuli of *Uredo cyclotrauma*, on leaving leaves of *Pithecellobium cauliflorum*.

Notes: Status unclear. Not treated by any of [Wollenweber & Reinking \(1935\)](#), [Booth \(1971\)](#), or [Gerlach & Nirenberg \(1982\)](#).

uredinum Fusarium Ellis & Everh., N. Amer. Fungi, Ser. II, no. 2799. 1890, *nom. inval.*, Art. 38.1(a).

Ramularia uredinis (W. Voss) Sacc., Syll. Fung. 4: 199. 1886. *Basionym*: *Cylindrosporium uredinis* W. Voss, Verh. Zool.-Bot. Ges. Wien 29: 684. 1879.

Synonym: *Ramularia nambuana* Henn., Hedwigia 43: 146. 1904. *Authentic material*: NY00928692.

Original locality: **USA**, Wisconsin, Racine.

Original substrate: Parasitic on uredinia of *Melampsora salicina*, on leaf of *Salix* sp.

Notes: [Wollenweber & Reinking \(1935\)](#) considered *F. uredinum* a synonym of *Cladosporium herbarum*. It is quite possible that this common saprobic *Cladosporium* species also occurred on uredinia in N. Am. Fungi 2799, but it can be ruled out that Ellis & Everhard confused this dematiaceous hyphomycete characterised by having long conidiophores with thickened and darkened conidiogeneous loci and large catenate conidia with a colourless *Fusarium*. [Davis \(1915\)](#) found *Ramularia uredinis*, a common mucedinaceous hyphomycete on *Melampsora* spp. on *Populus* and *Salix*, in material authentic for this name. This is undoubtedly correct.

urticearum Fusarium (Corda) Sacc., Syll. Fung. 4: 698. 1886.

Basionym: *Selenosporium urticae* Corda, Icon. Fung. 2: 7. 1838.

(See ***Fusarium lateritium***)

Lectotypus (*hic designatus*, MBT 10000754): **Czech Republic**, Prague, dead branches of *Ficus elastica* and *Morus nigra*, 1838. A.C.J. Corda, in Icon. Fung. 2, Tab. 9, fig. 30.

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#). No holotype specimen could be located and therefore an illustration is designated as lectotype.

ussurianum Fusarium T. Aoki *et al.*, Mycologia 101: 847. 2009. *Holotypus*: BPI 878845.

Ex-type culture: CBS 123752 = NRRL 45681 = TG-2662/0.

Type locality: **Russia**, Ussuriysk, Primorsky krai (Far East territory), agricultural field near the city Ussuriysk.

Type substrate: Seed of *Avena sativa*.

Descriptions and illustrations: See [Yli-Mattila *et al.* \(2009\)](#).

Diagnostic DNA barcodes: *rpb1*: KM361648; *rpb2*: KM361666; *tef1*: FJ240301.

ustilaginis Fusarium Kellerm. & Swingle, Rep. (Annual) Kansas Agric. Exp. Sta. 2: 285. 1890 [1889].

(See ***Fusarium avenaceum***)

Lectotypus (*hic designatus*, MBT 10000755): **USA**, Kansas, Manhattan, on *Ustilago avenae*, on *Avena sativa*, 1890, W.A. Kellerman & W.T. Swingle, in Rep. (Annual) Kansas Agric. Exp. Sta. 2, pl. IX, figs 1–13.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

ustilaginis Fusarium Rostr., Bot. Foren. Festschr. 54: 137. 1890, *nom. illegit.*, Art. 53.1.

(See *Fusarium nivale*)

Authentic material: C-F-125286.

Original locality: **Denmark**, Jutland, near Viborg.

Original substrate: Parasitic on *Ustilago grandis* on *Phragmites communis*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

vanettenii Fusarium O'Donnell *et al.*, Index Fungorum 440: 5. 2020.

Basionym: *Fusarium martii* var. *pisi* F.R. Jones, J. Agric. Res. 26: 459. 1923.

(See *Fusarium pisi*)

vasinfectum Fusarium G.F. Atk., Bull. Alabama Agric. Exp. Sta. 41: 28. 1892.

(See ***Fusarium oxysporum***)

Holotypus: ?CUP-A-(0100)#1.

Type locality: **USA**, Alabama, Montgomery, Mathews.

Type substrate: *Gossypium herbaceum*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

venenatum Fusarium Nirenberg, Mycopathologia 129: 136. 1995.

Misapplied names: *Fusarium sambucinum* var. *coeruleum* Wollenw. *sensu* Booth, The Genus *Fusarium*: 171–172. 1971.

Fusarium sambucinum var. *coeruleum* Wollenw. *sensu* Gerlach & Nirenberg, Mitt. Biol. Bundesanst. Land.- Forstw. 209: 213–216. 1982.

Holotypus: CBS 458.93 (preserved as metabolically inactive culture).

Ex-type culture: BBA 64537 = CBS 458.93 = NRRL 26228.

Type locality: **Austria**.

Type substrate: Culm of *Triticum aestivum*.

Descriptions and illustrations: See [Nirenberg \(1995\)](#).

Diagnostic DNA barcodes: *rpb2*: KM232382; *tef1*: KM231942.

venerorum Fusarium Dounin & Goldmacher, Index of the plant diseases in the U.S. 5: 284–298. 1927.

(See ***Fusarium avenaceum***)

Holotypus: Not located.

Type locality: **Unknown**.

Type substrate: Unknown.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

venezuelense Fusarium O'Donnell *et al.*, Index Fungorum 440: 5. 2020.

Neocosmospora robusta Sand.-Den. & Crous, Persoonia 43: 165. 2019, *non Fusarium robustum* Gerlach 1977.

Holotypus: CBS H-24000.

Ex-type culture: BBA 65682 = CBS 145473 = NRRL 22395.

Type locality: **Venezuela**.

Type substrate: Bark.

Descriptions and illustrations: See [Sandoval-Denis *et al.* \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW834251; *rpb2*: EU329507; *tef1*: AF178341.

ventricosum Fusarium Appel & Wollenw., Phytopathology 3: 32. 1913.

Rectifusarium ventricosum (Appel & Wollenw.) L. Lombard & Crous, Stud. Mycol. 80: 229. 2015.

Synonyms: *Fusarium solani* var. *ventricosum* (Appel & Wollenw.) Joffe, Pl. & Soil 38: 440. 1973.

Fusarium cuneiforme Sherb., Mem. Cornell Univ. Agric. Exp. Sta. 6: 129. 1915.

Hypomyces solani Reinke & Berth., Untersuch. Bot. Lab. Univ. Göttingen 1: 27. 1879.

Hyponectria solani (Reinke & Berth.) Petch, J. Bot. 75: 220. 1937.

Nectriopsis solani (Reinke & Berth.) C. Booth, Mycol. Pap. 74: 8. 1960.

Nectria ventricosa Booth, The Genus Fusarium: 55. 1971.

Holotypus: B 70 0021849.

Epitypus: CBS H-21947, designated in Lombard *et al.* (2015).

Ex-epitype culture: BBA 62452 = CBS 748.79 = NRRL 20846 = NRRL 22113.

Type locality: **Germany**, Berlin.

Type substrate: Tuber of *Solanum tuberosum*.

Descriptions and illustrations: See Wollenweber (1917), Booth (1971) and Lombard *et al.* (2015).

Diagnostic DNA barcodes: *rpb1*: JX171484; *rpb2*: JX171597; *tef1*: KM231924.

Notes: Contrary to Wollenweber & Reinking (1935), Booth (1971) considered this species as different from *F. argillaceum*, which was later confirmed by Lombard *et al.* (2015). The same authors designated an epitype for this taxon and transferred it to the genus *Rectifusarium* as *R. ventricosum*.

veratri Fusarium (Allesch.) Höhn., in Kabát & Bubák, Fungi Imperf. Exs. No. 349. 1906.

Gloeosporium veratri (Allesch.) Höhn., Mitt. Bot. Inst. Tech. Hochsch. Wien 4: 112. 1927.

Basionym: *Fusoma veratri* Allesch., Ber. Bayer. Bot. Ges. 2: 19. 1892.

Synonym: *Septogloeum veratri* (Allesch.) Wollenw., Fusaria Autogr. Delin. 1: 439. 1916.

Holotypus: ?M.

Type locality: **Germany**, Bavaria, Oberammergau.

Type substrate: Leaves of *Veratrum lobelianum*.

Notes: This species produces acervuli and 1-septate conidia with truncate basal cells. Therefore, it was transferred to *Gloeosporium* (*Helotiales*, *Dermataceae*).

verrucosum Fusarium (Pat.) O'Donnell & Geiser, Phytopathology 103: 404. 2013.

Albonectria verrucosa (Pat.) Rossman & Samuels, Stud. Mycol. 42: 108. 1999.

Basionym: *Calonectria verrucosa* Pat., Bull. Soc. Mycol. France 11: 228. 1895.

Synonym: *Nectria astromata* Rossman, Mycotaxon 8: 550. 1979, non *N. verrucosa* (Schwein.) Sacc.

Holotypus: In FH *vide* Rossman *et al.* (1999).

Type locality: **Ecuador**, San Jorge.

Type substrate: *Chusquea* sp.

Descriptions and illustrations: See Rossman (1983) and Rossman *et al.* (1999).

Notes: Although recently recombined in *Fusarium* (Geiser *et al.* 2013), the taxonomy of this species is uncertain. With 5–9(–13)-septate ascospores, this species cannot be a member of *Fusarium* s. str., and the identity of the isolates included in recent phylogenetic estimates (CBS 102163, originally identified as *F. concolor* and NRRL 22566) cannot be confirmed at this stage.

versicolor Fusarium Sacc., Syll. Fung. 16: 1099. 1902.

(See *Fusarium culmorum*)

Holotypus: In PAD.

Type locality: **France**, Côte-d'Or.

Type substrate: Cortex of *Cucurbita* sp.

Note: Synonym *vide* Wollenweber & Reinking (1935).

versiforme Fusarium Kabát & Bubák, Hedwigia 44: 358. 1905.

Holotypus: BPI 453128.

Type locality: **Czech Republic**, Bohemia, Turnov.

Type substrate: Living leaves of *Hosta sieboldii* (syn. *Hosta albomarginata*).

Notes: Status unclear. Not *Fusarium* *vide* Wollenweber & Reinking (1935).

verticillioides Fusarium (Sacc.) Nirenberg, Mitt. Biol. Bundesanst. Land- Forstw. 169: 26. 1976.

Basionym: *Oospora verticillioides* Sacc., Fung. Ital., Fasc. 17–28: pl. 879. 1881.

Synonyms: *Alysidium verticillioides* (Sacc.) Kuntze, Revis. Gen. Pl. 3: 442. 1898.

Fusarium moniliforme J. Sheld., Annual Rep. Nebraska Agric. Exp. Sta. 17: 23. 1904.

Gibberella moniliformis Wineland, J. Agric. Res. 28: 909. 1924.

Lectotypus: Pl. 879 in Saccardo, Fung. Ital. (1881), designated by Yilmaz *et al.* (2021).

Epitypus: CBS 218.76 (preserved as metabolically inactive culture), designated by Yilmaz *et al.* (2021).

Ex-epitype culture: BBA 11782 = CBS 218.76 = DSM 62264 = IMI 202875 = NRRL 13993.

Epitype locality: **Germany**.

Epitype substrate: *Zea mays*.

Descriptions and illustrations: See Nirenberg (1976, 1981), Gerlach & Nirenberg (1982) and Leslie & Summerell (2006).

Diagnostic DNA barcodes: *rpb1*: MW402638; *rpb2*: MW928835; *tef1*: KF499582.

veterinarium Fusarium L. Lombard & Crous, Persoonia 43: 35. 2018 [2019].

Holotypus: CBS H-23623.

Ex-type culture: CBS 109898 = NRRL 36153.

Type locality: **Netherlands**.

Type substrate: Peritoneum of *Selachimorpha* (shark).

Descriptions and illustrations: See Lombard *et al.* (2019b).

Diagnostic DNA barcodes: *rpb2*: MH484899; *tef1*: MH484990.

victoriae Fusarium Henn., in herb., *vide* Wollenweber, Fusaria Autogr. Delin. 1: 66. 1916.

Macronectria jungneri (Henn.) C. Salgado & P. Chaverri, Fungal Diversity 80: 448. 2016. *Basionym*: *Nectria jungneri* Henn., Bot. Jahrb. Syst. 22: 75. 1895.

Synonyms: *Nectria eustoma* Penz. & Sacc., Malpighia 11: 509. 1898.

Nectria leucocoma Starbäck, Bih. Kongl. Svenska Vetensk.-Akad. Handl. 25: 28. 1899.

Nectria cinereopapillata Henn. & E. Nyman, Monunia 1: 161. 1900.

Nectria striatospora Zimm., Centralbl. Bakteriolog. Abt. 1, 7: 105. 1901.

Cylindrocarpon victoriae Wollenw., Z. Parasitenk. (Berlin) 1: 161. 1928.

Nectria azureo-ostiolata Yoshim. Doi, Mem. Nat. Sci. Mus. Tokyo 10: 23. 1977.

Authentic material: In B *vide* Wollenweber, Fusaria Autogr. Delin. 1: 66. 1916.

Original locality: **Cameroon**.

Original substrate: Trunk of an unknown tree.

vinosum Fusarium Masee, Brit. Fung.-Fl. 3: 479. 1893.

(See *Fusarium flocciferum*)

Holotypus: ?K(M).

Type locality: **UK**.

Type substrate: Decaying mast manufactured from *Fagus sylvatica*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

vinosum Fusarium Greco, Origine des Tumeurs (Etiologie du Cancer. etc.) et Observations de Mycoses (Blastomycoses. etc.) Argentines (Buenos Aires): 670. 1916, *nom. illegit.*, Art. 53.1.

Authentic material: Not located.

Original locality: **Argentina**.

Original substrate: *Homo sapiens*.

Note: A late homonym of *F. vinosum* Masee.

violaceum Fusarium P. Crouan & H. Crouan, Fl. Finistère: 14. 1867, *nom. illegit.*, Art. 53.1.

(See ***Fusarium sambucinum***)

Authentic material: ?PC.

Original locality: **France**, Brittany, Finistère, marshes.

Original substrate: Bark of unknown tree.

Notes: An illegitimate homonym of *F. violaceum* Fuckel (1863).

Synonym *fide* [Gams et al. \(1997\)](#).

violaceum Fusarium Fuckel, Fungi Rhen. Exs. No. 209. 1863.

(See *Fusarium caeruleum*)

Syntypes: In BPI, F, HAL, MICH, S & WSP (Fuckel, Fungi Rhen. Exs. No. 209).

Type locality: **Germany**, Hessen, Oestrich.

Type substrate: *Solanum tuberosum*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#) and [Booth \(1971\)](#).

violae Fusarium F.A. Wolf, Mycologia 2: 21. 1910.

(See ***Fusarium oxysporum***)

Holotypus: Not located.

Type locality: **USA**, Nebraska, Lincoln.

Type substrate: Stems and roots of *Viola tricolor*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

virguliforme Fusarium O'Donnell & T. Aoki, Mycologia 95: 667. 2003.

(See *Fusarium azukicola*)

Holotypus: BPI 841956.

Ex-type culture: MAFF 238553 = NRRL 31041 = Shuxian Li # 95.

Type locality: **USA**, Illinois.

Type substrate: *Glycine max*.

Descriptions and illustrations: See [Aoki et al. \(2003\)](#).

Diagnostic DNA barcodes: *rpb1*: JX171530; *rpb2*: JX171643; *tef1*: AY220193.

viride Fusarium (Lechmere) Wollenw., Fusaria Autogr. Delin. 1: 418. 1916.

Basionym: *Pionnotes viridis* Lechmere, Compt. Rend. Hebd. Séances Acad. Sci. 155: 178. 1912.

(See *Fusarium solani*)

Holotypus: Not located.

Type locality: **Ivory Coast**.

Type substrate: Undetermined wood.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

viticola Fusarium Thüm. (as '*viticolum*'), Pilze Weinst.: 52. 1878.

Synonym: *Fusarium herbarum* var. *viticola* (Thüm.) Wollenw., Fusaria Autogr. Delin. 3: 898. 1930.

(See ***Fusarium avenaceum***)

Lectotypus (*hic designatus*, MBT 10000756): **Italy**, Liguria, Genoa, Rapallo, dry twigs of *Vitis vinifera*, Jul. 1876, G. Passerini, in Thümen, Pilze Weinst. 1878: pl. 3, fig. 3.

Notes: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#). No holotype specimen could be located and therefore an illustration is designated as lectotype.

vogelii Fusarium Henn., Z. Pflanzenkrankh. 12: 16. 1902.

Synonyms: *Septosporium curvatum* Rabenh. & A. Braun, Krankh. Pfl.: 14. 1854.

Septoria curvata (Rabenh. & A. Braun) Sacc., Syll. Fung. 3: 484. 1884.

Cercospora curvata (Rabenh. & A. Braun) Wollenw., Fusaria Autogr. Delin. 1: 451. 1916.

Holotypus: In B (Kabát & Bubák, Fungi Imp. Exs. 248) *fide* [Hein \(1988\)](#).

Type locality: **Poland**, Dąbroszyn (former Tamsel).

Type substrate: Leaf of *Robinia pseudoacacia*.

Notes: Status unclear. Neither *Fusarium* *fide* [Wollenweber & Reinking \(1935\)](#) nor *Cercospora* *fide* [Chupp \(1954\)](#).

volatile Fusarium Al-Hatmi et al., Fungal Syst. Evol. 4: 174. 2019.

Holotypus: CBS H-24004.

Ex-type culture: CBS 143874.

Type locality: **French Guiana**, Cayenne.

Type substrate: Bronchoalveolar lavage effusion from *Homo sapiens* with lung infection.

Descriptions and illustrations: See [Al-Hatmi et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb2*: LR596006; *tef1*: LR596007.

volutella Fusarium Ellis & Everh., Proc. Acad. Nat. Sci. Philadelphia 43: 93. 1891.

(See *Fusarium stromaticum*)

Holotypus: Langlois 1505 in NY *fide* Index Fungorum.

Type locality: **USA**, Louisiana, Saint Martinsville.

Type substrate: Dead twigs of *Nekemias arborea* (syn. *Ampelopsis arborea*).

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#) and [Gräfenhan et al. \(2011\)](#).

vorosii Fusarium B. Tóth et al., Fungal Genet. Biol. 44: 1202. 2007.

Holotypus: BPI 871658.

Ex-type culture: NRRL 37605.

Type locality: **Hungary**, Pest, Ipolydamásd.

Type substrate: Spikelet of *Triticum aestivum*.

Descriptions and illustrations: See [Starkey et al. \(2007\)](#).

Diagnostic DNA barcodes: *rpb1*: KM361647; *rpb2*: KM361665; *tef1*: DQ459745.

waltergamsii Fusarium O'Donnell et al., Index Fungorum 440: 5. 2020.

Neocosmospora gamsii Sand.-Den. & Crous, Persoonia 41: 116. 2018.

Holotypus: CBS H-23226.

Ex-type culture: CBS 143207 = NRRL 32323 = UTHSC 99-250.

Type locality: **USA**, Pennsylvania.

Type substrate: Bronchoalveolar lavage fluid from *Homo sapiens*.

Descriptions and illustrations: See [Sandoval-Denis & Crous \(2018\)](#).

Diagnostic DNA barcodes: *rpb1*: MW834223; *rpb2*: KM361665; *tef1*: DQ246951.

werrikimbe Fusarium J.L. Walsh, L.W. Burgess, E.C.Y. Liew & B.A. Summerell, *sp. nov.* MycoBank MB 837725.

Synonym: Fusarium werrikimbe J.L. Walsh, L.W. Burgess, E.C.Y. Liew & B.A. Summerell, *Fungal Diversity* 44: 155. 2010, *nom. inval.*, Art. 40.7.

Etymology: In reference to Werrikimbe National Park, the geographic origin of the isolates first recognised as belonging to this species.

For diagnosis see Walsh *et al.*, *Fungal Diversity* 44: 155. 2010. *Holotypus:* CBS 125535 (preserved as metabolically inactive culture).

Ex-type culture: CBS 125535 = F19350 = RBG 5332.

Type locality: **Australia**, New South Wales, Werrikimbe National Park.

Type substrate: *Sorghum leiocladum*.

Descriptions and illustrations: See Walsh *et al.* (2010).

Diagnostic DNA barcodes: *rpb1:* MW928821; *rpb2:* MN534304; *tef1:* MW928846.

Notes: Walsh *et al.* (2010) did not indicate the holotype for *F. werrikimbe*, rendering the name invalid (Art. 40.7). Here we validate the name.

willkommii *Fusarium* Lindau, Rabenh. Krypt.-Fl. ed. 2, 1(9): 551. 1910.

Replaced synonym: Fusarium candidum Sacc. & D. Sacc., *Syll. Fung.* 18: 674. 1906, *nom. illegit.*, Art. 53.1, *non Fusarium candidum* Ehrenb. 1818.

Lectotypus (hic designatus, MBT 10000757): **Germany**, Saxony, *Fagus sylvatica*, 1866, M. Willkomm, in *Die mikroskopischen Feinde des Waldes* 1, Tab. VI, figs 11–12.

Notes: Lindau's description of *F. willkommii* was based on Willkomm's (1866: 103) description and illustration under the name *Fusidium candidum* Link as well as Saccardo's (*l.c.*) description under *Fusarium candidum*. Therefore, the illustration by Willkomm (1866) is designated as lectotype.

witzenhausenense *Fusarium* Šišić *et al.*, *Antonie van Leeuwenhoek* 111: 1795. 2018.

(See *Fusarium stercicola*)

Holotypus: CBS H-23351.

Ex-type culture: CBS 142480 = DSM 106212.

Type locality: **Germany**, Hessen, Witzenhausen, Neu-Eichenberg.

Type substrate: Branch of *Hibiscus* sp.

Descriptions and illustrations: See Šišić *et al.* (2018a).

Diagnostic DNA barcodes: *rpb1:* MG237865; *rpb2:* LR583886; *tef1:* KY556525.

wolgense *Fusarium* Rodigin, *Trudy Bashkir. Sel'. Khoz. Inst.* 3: 101. 1942.

Holotypus: Not located.

Type locality: **Russia**, Volgograd (formerly Stalingrad).

Type substrate: Fruit of *Citrullus lanatus* (syn. *Citrullus vulgaris*).

Notes: Status unclear. Not treated by either of Booth (1971) and Gerlach & Nirenberg (1982).

wollenweberi *Fusarium* Raillo, *Fungi of the Genus Fusarium*: 189. 1950, *nom. illegit.*, Art. 52.1.

(See *Fusarium anthophilum*)

Authentic material: Not located.

Original locality: **Azerbaijan**.

Original substrate: Seeds and stems of *Gossypium* sp.

Descriptions and illustrations: See Raillo (1950).

Notes: *Fusarium wollenweberi* was published as a new combination, but no basionym was indicated. As a *nomen novum*, it can only be based on *F. anthophilum*, the only cited name, which

is a valid name. Therefore, *F. wollenweberi* would be illegitimate (*nom. superfl.*, Art. 52.1.). Additionally, the condition for the introduction of a new species is also not met as a Latin diagnosis, necessary in 1950, is lacking.

xiangyunense *Fusarium* F. Zhang *et al.* (as '*xiangyunensis*'), *Phytotaxa* 450: 278. 2020, *nom. inval.*, Art. 40.8.

(See *Fusarium stercicola*)

Authentic material: DLU11-1, School of Agriculture and Biology, Dali University, China.

Authentic culture: CGMCC 3.19676.

Original locality: **China**, Yunnan, Xiangyun, Dali, Da-bo-na hot-spring.

Original substrate: Waterlogged soil.

Descriptions and illustrations: See Zhang *et al.* (2020).

Diagnostic DNA barcodes: *rpb1:* MH999281; *tef1:* MH992629.

Note: Based on phylogenetic and morphological evidence provided by Zhang *et al.* (2020), this invalid name (Art. 40.8) belongs to the genus *Neocosmospora* and is a synonym of *N. stercicola*.

xylarioides *Fusarium* Steyaert, *Bull. Soc. Roy. Bot. Belgique* 80: 42. 1948.

Synonyms: *Gibberella xylarioides* (Steyaert) R. Heim & Saccas, *Rev. Mycol. (Paris)* 15 (Suppl. Colon.): 97. 1950.

Fusarium oxysporum f. *xylarioides* (Steyaert) Delassus, *Bull. sci. Minist. Colon., Sect. Agric. trop.* 5: 347. 1954.

Lectotypus (hic designatus, MBT 10000758): **Central African Republic**, Bangui, trunk of *Coffea excelsa*, 1939, H. Frédéric, in Steyaert, *Bull. Soc. Roy. Bot. Belgique* 80, pl. I, fig. 8.

Epitypus (hic designatus, MBT 10001275): **Ivory Coast**, on trunk of *Coffea* sp., Feb. 1951, C. & M. Moreau, CBS 258.52 (preserved as metabolically inactive culture).

Ex-epitype culture: CBS 258.52 = NRRL 25486.

Descriptions and illustrations: See Steyaert (1948), Booth (1971), Gerlach & Nirenberg (1982) and Geiser *et al.* (2005).

Diagnostic DNA barcodes: *rpb1:* JX171517; *rpb2:* JX171630; *tef1:* AY707136.

Notes: A lectotype is designated here based on an illustration provided by Steyaert (1948) accompanying the original protologue. All attempts to locate the holotype specimen lodged at the Université de Bangui (BANG), Central African Republic, as indicated by Steyaert (1948), failed. In addition, an epitype (CBS 258.52) is designated here to provide taxonomic stability for this important species.

xyrophilum *Fusarium* I. Laraba *et al.*, *Mycologia* 112: 45. 2019 [2020].

Holotypus: BPI 910919.

Ex-type culture: FRC M-8921 = NRRL 62721.

Type locality: **Guyana**, Cuyuni-Mazaruni, Kamakusa Mountain.

Type substrate: *Xyris surinamensis*.

Descriptions and illustrations: See Laraba *et al.* (2020).

Diagnostic DNA barcodes: *rpb1:* MN193933; *rpb2:* MN193905; *tef1:* MN193877.

yamamotoi *Fusarium* O'Donnell *et al.*, *Index Fungorum* 440: 5. 2020.

Replaced synonym: Nectria elegans W. Yamam. & Maeda, *Hyogo Univ. Agric. ser. Agric. Biol.* 3: 15. 1957, *non Fusarium elegans* Appel & Wollenw. 1910.

Neocosmospora elegans (W. Yamam. & Maeda) Sand.-Den. & Crous, *Persoonia* 43: 127. 2019.

Lectotypus: Figs 1–9, page 16, in Yamamoto *et al.* (1957), designated in Sandoval-Denis *et al.* (2019).

Epitypus: CBS H-23980, designated in [Sandoval-Denis et al. \(2019\)](#).

Ex-epitype culture: ATCC 42366 = CBS 144396 = MAFF 238541 = NRRL 22277 = SUF XV-1.

Type locality: **Japan**.

Type substrate: Twigs and trunks of *Zanthoxylum piperitum*.

Descriptions and illustrations: See [Sandoval-Denis et al. \(2019\)](#).

Diagnostic DNA barcodes: *rpb1*: MW218113; *rpb2*: FJ240380; *tef1*: AF178336.

yuccae *Fusarium* Cooke, *Grevillea* 7: 34. 1878, *nom. inval.*, Art. 36.1(a).

(See [Fusarium lateritium](#))

Authentic material: BPI 453149.

Original locality: **USA**, South Carolina, Aiken.

Original substrate: *Yucca aloifolia*.

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

zanthoxyli* *Fusarium X. Zhou et al., *Mycologia* 108: 675. 2016.

Holotypus: HMNWAFU XZ-Fyzs133-20130408

Ex-type culture: CBS 140838 = NRRL 66285.

Type locality: **China**, Shaanxi, Tongchuan, Yaozhou, Sunyuan.

Type substrate: *Zanthoxylum bungeanum*.

Descriptions and illustrations: See [Zhou et al. \(2016\)](#).

Diagnostic DNA barcodes: *rpb1*: KM520383; *rpb2*: KM236763; *tef1*: KM236703.

zavianum *Fusarium* (Sacc.) Sacc., *Syll. Fung.* 4: 709. 1886.

Basionym: *Fusisporium zavianum* Sacc., *Michelia* 1: 83. 1877.

(See [Fusarium lateritium](#))

Holotypus: In PAD.

Type locality: **Italy**, Vittorio.

Type substrate: *Vitis vinifera*.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

zeae *Fusarium* (Westend.) Sacc., *Syll. Fung.* 4: 713. 1886.

Basionym: *Fusisporium zeae* Westend., *Bull. Acad. Roy. Sci. Belgique, Cl. Sci.* 18: 414. 1852. (*non Fusisporium zeae* Roum., *Rev. Mycol. (Toulouse)* 6: 163. 1884).

(See [Fusarium avenaceum](#))

Holotypus: BR5020141668483.

Type locality: **Belgium**, Kortrijk railway station.

Type substrate: Rotting stalks of *Zea mays*.

Note: Synonyms *fide* [Wollenweber & Reinking \(1935\)](#).

zealandicum *Fusarium* Nirenberg & Samuels, *Canad. J. Bot.* 78: 1483. 2000.

Geejayessia zealandica (Cooke) Schroers, *Stud. Mycol.* 68: 133. 2011.

Basionym: *Nectria zealandica* Cooke, *Grevillea* 8: 65. 1879.

Synonyms: *Cucurbitaria zealandica* (Cooke) Kuntze, *Revis. Gen. Pl.* 3: 462. 1898.

Cosmospora zealandica (Cooke) Samuels & Nirenberg, *Canad. J. Bot.* 78: 1483. 2000.

Holotypus: BPI 747915.

Ex-type culture: BBA 64792 = CBS 111.93.

Type locality: **New Zealand**, Auckland, Waitakere Ranges Regional Park, Cascades Kauri.

Type substrate: Bark of *Hoheria populnea*.

Descriptions and illustrations: See [Nirenberg & Samuels \(2000\)](#).

Diagnostic DNA barcodes: *rpb2*: HM626684; *tef1*: HQ728148.

ziziphinum *Fusarium* Pass., *Erb. Critt. Ital. ser.* 2 no. 1084. 1881.

(See [Fusarium lateritium](#))

Syntype: F 982523 (Erb. Critt. Ital. no. 1048).

Type locality: **Italy**.

Type substrate: Twigs of *Ziziphus sinensis* (syn. *Ziziphus jujuba*).

Note: Synonym *fide* [Wollenweber & Reinking \(1935\)](#).

zonatum *Fusarium* (Sherb.) Wollenw., *Fusaria Autogr. Delin.* 1: 392. 1916.

Basionym: *Fusarium lutulatum* var. *zonatum* Sherb., *Mem. Cornell Univ. Agric. Exp. Sta.* 6: 214. 1915.

(See [Fusarium oxysporum](#))

Typus: ?CUP-007453.

Type locality: **USA**, New York, Ithaca.

Type substrate: *Solanum tuberosum*.

Notes: Synonym *fide* [Wollenweber & Reinking \(1935\)](#). Lectotypification pending study of the material lodged in CUP.

zygopetali *Fusarium* Delacr., *Bull. Soc. Mycol. France* 13: 103. 1897.

Holotypus: ?PC.

Type locality: **France**, Paris, Luxembourg gardens.

Type substrate: Leaves of *Zygopetalum maculatum* (syn. *Zygopetalum mackayi*).

Notes: Status unclear. Not *Fusarium* *fide* [Wollenweber & Reinking \(1935\)](#).

CONCLUSIONS

The present study is the first to provide an up-to-date morphological, biochemical, and phylogenetic overview of the 20 fusarioid genera that are presently recognised in *Nectriaceae*. Morphological species recognition frequently fails to distinguish fusarioid taxa that have been described based on genealogical concordance phylogenetic species recognition (GCPSR *sensu* [Taylor et al. 2000](#)). To address this issue, we have established a new database, Fusarioid-ID, with accurate names for species and genera of fusarioid taxa. Although the phylogenetically most informative genes remain *tef1*, *rpb1* and *rpb2*, additional markers such as *act1*, *CaM*, *tub2*, ITS and LSU are also incorporated. These genetic fragments can be amplified by PCR and sequenced using the primers indicated in [Table 2](#). In the future, new species and other phylogenetically informative orthologous genes, will be added to resolve isolates at species and genus level. Researchers interested in obtaining reference strains should contact the Westerdijk Fungal Biodiversity Institute (<https://wi.knaw.nl/page/Collection>), which houses a large collection of phylogenetically diverse fusarioid taxa.

As we have shown here, the phylogenetically derived argument that species under the node F1 should be considered members of “*Fusarium*” is not practical, as this circumscription would lead to a genus without apparent synapomorphies, as lineages outside the genus would also share its characteristics. However, the F3 node (corresponding to *Fusarium* s. *str.*) is resolved by all genetic markers so far analysed (e.g., see [Geiser et al. 2021](#)) and delineates the morphologically, ecologically, and biochemically well-delineated genus *Fusarium*.

Fusarium s. *str.* does not have different sexual morphs, other than *Gibberella*. Fusarioid genera are not only morphologically distinct, but as we have shown in this study, correlate to different monophyletic groups and also differ in their biology and mycotoxin profiles.

One of the reasons for the desire to classify any species producing conidia with foot-shaped basal cells into a single genus could be that plant pathologists and clinicians typically

isolate conidia or obtain cultures from vegetative mycelium that inhabits their specimens. Also, Wollenweber and his successors may have primarily worked with vegetatively proliferating materials, although it was also Wollenweber (1924, 1926) who produced the first general synopsis of holomorphs in the *Hypocreales*. However, mainly Joan M. Dingley (1951, 1957), Colin Booth (1959), and especially Gary J. Samuels (Samuels 1976a, b, 1978, 1988, Samuels et al. 1991) significantly changed our points of view by systematically isolating ascospores obtained from ascomata, of which a vast majority were not gathered in agricultural fields but from woody or herbaceous substrata in forests of pantropical, species-rich regions. The result of their taxonomic considerations was an infrageneric subgrouping system in *Nectria* that was based on sexual and asexual connections. The classification of species according to morphological similarities in sexual morphs allowed understanding patterns of asexual characteristics that are unique for the sexually defined subgroups and eventually correlating sexual groupings with Wollenweber's section system. The diversity of nectria-like species Samuels looked at is huge and was eventually interpreted on the level of families, within which numerous genera were recognised or newly described (Rossman et al. 1999) with infrageneric, informal species groups of *Nectria* accepted at the genus level (e.g., see Chaverri et al. 2011 and subsequent studies). Applying the generic level to the numerous nectria-like subgroups producing fusarioid conidia is therefore another small but unavoidable step towards a taxonomic system that allows distinguishing natural diversity above the species level based on morphologically and phylogenetically well-defined units.

When Colin Booth delivered his Presidential address to the British Mycological Society in 1977, he chose the title "Do you believe in genera?". He addressed this topic based on his interpretation of *Nectriaceae* (Booth 1978). Booth subsequently showed that several "groups" of species formed fusarioid asexual morphs, namely *Gibberella* (now *Fusarium s. str.*), *Haematonectria* (now *Neocosmospora*), *Nectria episphaeria* (now *Cosmosporella* and *Dialonectria*), and *Calonectria rigidiuscula* (now *Albonectria*). Booth concluded that the "fusarium morphs" reflected "terms of convenience" rather than genealogical relationships. In moving to the one fungus = one name nomenclature (Hawksworth et al. 2011, Wingfield et al. 2012), *Fusarium s. str.* was chosen over *Gibberella* (Gräfenhan et al. 2011, Schroers et al. 2011, Rossman et al. 2013). As the genus *Fusarium* was thus clearly well-defined, other *Nectriaceae* lineages with a fusarium-like morphology were recognised (Gräfenhan et al. 2011, Schroers et al. 2011, Lombard et al. 2015, Lechat & Fournier 2015). As we have shown here, taxa are constantly being newly collected and added to the phylogeny of *Nectriaceae*. The only stable option forward is to apply and use the genus name *Fusarium* (= *Gibberella*) as more precisely defined based on its own monophyletic node as presented here (F3), supported by morphology, biochemistry, and biology.

DISCLAIMER

The present paper represents a separate initiative to Geiser et al. (NSF 1655980): A phylogenetic revisionary monograph of the genus *Fusarium*.

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APPENDIX A. SUPPLEMENTARY DATA

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.simyco.2021.100116>.

REFERENCES

- Abbas HK, Mirocha CJ (1988). Isolation and purification of a hemorrhagic factor (wortmannin) from *Fusarium oxysporum* (N17B). *Applied and Environmental Microbiology* **54**: 1268–1274.
- Abraham WR, Hanssen HP (1992). Fusoxysporone—a new type of diterpene from *Fusarium oxysporum*. *Tetrahedron* **48**: 10559–10562.
- Al-Hatmi AMS, Ahmed SA, van Diepeningen AD, et al. (2018). *Fusarium met-avorans* sp. nov.: The frequent opportunist 'FSSC6'. *Medical Mycology* **56**: S144–S152.
- Al-Hatmi AMS, Mirabolfathy M, Hagen F, et al. (2016). DNA barcoding, MALDI-TOF, and AFLP data support *Fusarium ficicrescens* as a distinct species within the *Fusarium fujikuroi* species complex. *Fungal Biology* **120**: 265–278.
- Al-Hatmi AMS, Normand AC, van Diepeningen AD, et al. (2015). Rapid identification of clinical members of *Fusarium fujikuroi* complex using MALDI-TOF MS. *Future Microbiology* **10**: 1939–1952.
- Al-Hatmi AMS, Sandoval-Denis M, Nabet C, et al. (2019). *Fusarium volatile*: a new potential pathogen from a human respiratory sample. *Fungal Systematics and Evolution* **4**: 171–181.
- Aoki T, Kasson MT, Berger MC, et al. (2018). *Fusarium oligoseptatum* sp. nov., a mycosymbiont of the ambrosia beetle *Euwallacea validus* in the Eastern U.S. and typification of *F. ambrosium*. *Fungal Systematics and Evolution* **1**: 23–39.
- Aoki T, O'Donnell K (1998). *Fusarium kyushuense* sp. nov. from Japan. *Mycoscience* **39**: 1–6.
- Aoki T, O'Donnell K (1999). Morphological and molecular characterization of *Fusarium pseudograminearum* sp. nov., formerly recognized as the Group 1 population of *F. graminearum*. *Mycologia* **91**: 597–609.
- Aoki T, O'Donnell K, Ichikawa K (2001). *Fusarium fractiflexum* sp. nov. and two other species within the *Gibberella fujikuroi* species complex recently discovered in Japan that form aerial conidia in false heads. *Mycoscience* **42**: 461–478.
- Aoki T, O'Donnell K, Scandiani MM (2005). Sudden death syndrome of soybean in South America is caused by four species of *Fusarium*: *Fusarium brasiliense* sp. nov., *F. cuneirostrum* sp. nov., *F. tucumaniae*, and *F. virguliforme*. *Mycoscience* **46**: 162–183.
- Aoki T, Scandiani MM, O'Donnell K (2012). Phenotypic, molecular phylogenetic, and pathogenetic characterization of *Fusarium crassistipitatum* sp. nov., a novel soybean sudden death syndrome pathogen from Argentina and Brazil. *Mycoscience* **53**: 167–186.
- Aoki T, Smith JA, Kasson MT, et al. (2019). Three novel Ambrosia *Fusarium* Clade species producing clavate macroconidia known (*F. floridanum* and *F. obliquiseptatum*) or predicted (*F. tuaranense*) to be farmed by *Euwallacea* spp. (Coleoptera: Scolytinae) on woody hosts. *Mycologia* **111**: 919–935.
- Aoki T, Smith JA, Mount LL, et al. (2013). *Fusarium torreyae* sp. nov., a pathogen causing canker disease of *Florida torreyae* (*Torreya taxifolia*), a critically endangered conifer restricted to northern Florida and southwestern Georgia. *Mycologia* **105**: 312–319.
- Aoki T, Tanaka F, Suga H, et al. (2012). *Fusarium azukicola* sp. nov., an exotic azuki bean root-rot pathogen in Hokkaido, Japan. *Mycologia* **104**: 1068–1084.
- Aoki T, Vaughan MM, McCormick SP, et al. (2015). *Fusarium dactylidis* sp. nov., a novel nivalenol toxin-producing species sister to *F. pseudograminearum* isolated from orchard grass (*Dactylis glomerata*) in Oregon and New Zealand. *Mycologia* **107**: 409–418.

- Aoki T, Geiser DM, Kasson M, *et al.* (2020). Nomenclatural novelties. *Index Fungorum* **440**: 1–5.
- Aoki T, O'Donnell K, Homma Y, *et al.* (2003). Sudden-death syndrome of soybean is caused by two morphologically and phylogenetically distinct species within the *Fusarium solani* species complex—*F. virguliforme* in North America and *F. tucumaniae* in South America. *Mycologia* **95**: 660–684.
- Arnstein HRV, Cook AH (1947). Production of antibiotics by fungi. 3. Javanicin – an antibacterial pigment from *Fusarium javanicum*. *Journal of the Chemical Society* **1947**: 1021–1028.
- Arya HC, Jain GL (1962). Fusarium wilt of *Eucalyptus*. *Phytopathology* **52**: 638–642.
- Bachmann HP, Bobst C, Bütikofer U, *et al.* (2005). Occurrence and significance of *Fusarium domesticum* alias *Anticollanti* on smear-ripened cheeses. *LWT – Food Science and Technology* **38**: 399–407.
- Bamburg JR, Riggs NV, Strong FM (1968). The structures of toxins from two strains of *Fusarium tricinctum*. *Tetrahedron* **24**: 3329–3336.
- Benyon FHL, Burgess LW, Sharp PJ (2000). Molecular genetic investigations and reclassification of *Fusarium* species in sections *Fusarium* and *Roseum*. *Mycological Research* **104**: 1164–1174.
- Berkeley MJ, Broome CE (1876). Notices of British fungi (1501–1630). *Annals and Magazine of Natural History* **17**: 129–145.
- Bezuidenhout SC, Gelderblom WC, Gorst-Allman CP, *et al.* (1988). Structure elucidation of the fumonisins, mycotoxins from *Fusarium moniliforme*. *Journal of the Chemical Society, Chemical Communications* **11**: 743–745.
- Bilal VI (1955). *Fusarii (Biologija i sistematika)*. Isdatelstvo Akademii Nauk Ukrainiskoi SSE, Kiev, Ukraine (in Russian).
- Bills GF, Gloer JB (2017). Biologically active secondary metabolites from the fungi. In: *The Fungal Kingdom* (Heitman J, Howlett BJ, Crous PW, *et al.*, eds). ASM Press, Washington D.C.: 1087–1119.
- Booth C (1959). Studies of Pyrenomycetes. IV. *Nectria* (part 1). *Mycological papers* **73**: 1–115.
- Booth C (1964). Studies on Pyrenomycetes. VII. *Mycological Papers* **94**: 1–16.
- Booth C (1971). *The genus Fusarium*. Commonwealth Mycological Institute, Kew, Surrey, England.
- Booth C (1978). Presidential address: Do you believe in genera? *Transactions of the British Mycological Society* **71**: 1–9.
- Braun U (1998). *A monograph of Cercosporella, Ramularia and allied genera (phytopathogenic Hyphomycetes)*: Vol. 2. IHW-Verlag, Munich, Germany.
- Brayford D (1987). *Fusarium bugnicourtii* sp. nov., and its relationship to *F. tumidum* and *F. tumidum* var. *coeruleum*. *Transactions of the British Mycological Society* **89**: 347–351.
- Britz H, Steenkamp ET, Coutinho TA, *et al.* (2002). Two new species of *Fusarium* section *Liseola* associated with mango malformation. *Mycologia* **94**: 722–730.
- Brown W (1928). Studies in the genus *Fusarium*. VI. General description of strains, together with a discussion of the principles at present adopted in the classification of *Fusarium*. *Annals of Botany* **42**: 285–304.
- Bugnicourt F (1952). Une espèce fusarienne nouvelle, parasite du riz. *Revue Générale de Botanique* **59**: 13–18.
- Burgess LW, Forbes GA, Windels C, *et al.* (1993). Characterization and distribution of *Fusarium acuminatum* subsp. *armenicum* subsp. nov. *Mycologia* **85**: 119–124.
- Burgess LW, Nelson PE, Toussoun TA (1982). Characterization, geographic distribution and ecology of *Fusarium crookwellense* sp. nov. *Transactions of the British Mycological Society* **79**: 497–505.
- Burgess LW, Nelson PE, Toussoun TA, *et al.* (1985). *Fusarium scirpi*: emended description and notes on geographic distribution. *Mycologia* **77**: 212–218.
- Burgess LW, Summerell BA (2000). Taxonomy of *Fusarium*: *Fusarium armenicum* stat. & comb. nov. *Mycotaxon* **75**: 347–348.
- Burgess LW, Trimboli D (1986). Characterization and distribution of *Fusarium nygamai*, sp. nov. *Mycologia* **78**: 223–229.
- Burkholder WH (1919). The dry rot of bean. *Memoirs of the Cornell University Agricultural Experimental Station* **26**: 1003–1033.
- Butler EJ, Khan AH (1913). Some new sugarcane diseases. *Memoirs of the Department of Agriculture in India. Botanical Series* **6**: 181–208.
- Butler EJ (1910). The wilt disease of pigeon pea and the parasitism of *Neocosmospora vasinfecta* Smith. *Memoirs of the Department of Agriculture in India* **9**: 1–64.
- Carbone I, Kohn LM (1999). A method for designing primer sets for speciation studies in filamentous ascomycetes. *Mycologia* **91**: 553–556.
- Cardoso AMS (2015). *Caracterização filogenética, biológica e patogênica de espécies do complexo Fusarium solani associadas à podridão do colo do maracujazeiro*. MSc. dissertation. Departamento de Fitopatologia, Universidade Federal de Lavras, Brazil.
- Carlucci A, Raimondo ML, Santos J, *et al.* (2012). *Plectosphaerella* species associated with root and collar rots of horticultural crops in Southern Italy. *Persoonia* **28**: 34–48.
- Castellá G, Bragulat MR, Rubiales MV, *et al.* (1997). Malachite green agar, a new selective medium for *Fusarium*. *Mycopathologia* **137**: 173–178.
- Cavalcanti AD, da Silva Santos AC, de Oliveira Ferro L, *et al.* (2020). *Fusarium massalimae* sp. nov. (*F. lateritium* species complex) occurs endophytically in leaves of *Handroanthus chrysotrichus*. *Mycological Progress* **19**: 1133–1142.
- Chaverri P, Salgado C, Hirooka Y, *et al.* (2011). Delimitation of *Neonectria* and *Cylindrocarpon* (Nectriaceae, Hypocreales, Ascomycota) and related genera with cylindrocarpon-like anamorphs. *Studies in Mycology* **68**: 57–78.
- Chen FJ (2000). *Fusarium catenulatum* sp. nov. from China. *Mycosystema* **19**: 459–462.
- Chen QT, Fu XH, Chen YD (1987). Two petriophilous taxa of *Fusarium*. *Acta Mycologica Sinica Supplement* **1**: 328–333.
- Chupp C (1954). *A monograph of the fungus genus Cercospora*. Published by the author, Ithaca, New York.
- Ciferri R (1955). Observations on meliocolous Hyphales from Santo Domingo. *Sydowia* **9**: 296–335.
- Costa MM, Melo MP, Carmo FS, *et al.* (2021). *Fusarium* species from tropical grasses in Brazil and description of two new taxa. *Mycological Progress* **20**: 61–72.
- Costa SS, Matos KS, Tessmann DJ, *et al.* (2016). *Fusarium paranaense* sp. nov., a member of the *Fusarium solani* species complex causes root rot on soybean in Brazil. *Fungal Biology* **120**: 51–60.
- Crous PW, Braun U (2003). *Mycosphaerella* and its anamorphs: 1. Names published in *Cercospora* and *Passalora*. *CBS Biodiversity Series* **1**: 1–571.
- Crous PW, Luangsa-ard JJ, Wingfield MJ, *et al.* (2018). Fungal Planet description sheets: 785–867. *Persoonia* **41**: 238–417.
- Crous PW, Wingfield MJ, Burgess TI, *et al.* (2016). Fungal Planet description sheets: 469–557. *Persoonia* **37**: 218–403.
- Crous PW, Shivas RG, Quaedvlieg W, *et al.* (2014). Fungal Planet description sheets: 214–280. *Persoonia* **32**: 184–306.
- Crous PW, Summerell BA, Carnegie AJ, *et al.* (2009). Unravelling *Mycosphaerella*: do you believe in genera? *Persoonia* **23**: 99–118.
- Crous PW, Verkley GJM, Groenewald JZ, *et al.* (2019). *Fungal Biodiversity*. In: *Westerdijk Laboratory Manual Series 1*. Westerdijk Fungal Biodiversity Institute, Utrecht, The Netherlands.
- Crous PW, Wingfield MJ, Burgess TI, *et al.* (2017). Fungal Planet description sheets: 625–715. *Persoonia* **39**: 270–467.
- Crous PW, Wingfield MJ, Lombard L, *et al.* (2019). Fungal Planet description sheets: 951–1041. *Persoonia* **43**: 223–425.
- Cummings MP, Meyer A (2005). Magic bullets and golden rules: Data sampling in molecular phylogenetics. *Zoology* **108**: 329–336.
- Cutler HG, Arendale RF, Springer JP, *et al.* (1987). Monorden from a novel source, *Neocosmospora tenuicristata*: Stereochemistry and plant growth regulatory properties. *Agricultural and Biological Chemistry* **51**: 3331–3338.
- Dallé Rosa P, Ramirez-Castrillon M, Valente P, *et al.* (2018). *Fusarium riograndense* sp. nov., a new species in the *Fusarium solani* species complex causing fungal rhinosinusitis. *Journal de Mycologie Médicale* **28**: 29–35.
- Damm U, Cannon PF, Woudenberg JHC, *et al.* (2012). The *Colletotrichum acutatum* species complex. *Studies in Mycology* **73**: 37–113.
- Damm U, Cannon PF, Liu F, *et al.* (2013). The *Colletotrichum orbiculare* species complex: important plant pathogens and mycoherbicides. *Fungal Diversity* **61**: 29–59.
- Damm U, Woudenberg JHC, Cannon PF, *et al.* (2009). *Colletotrichum* species with curved conidia from herbaceous hosts. *Fungal Diversity* **39**: 45–87.
- Davis JJ (1915). Notes on parasitic fungi in Wisconsin – I. *Transactions of the Wisconsin Academy of Sciences, Arts and Letters* **18**: 78–92.
- Dayarathne MC, Jones EBG, Maharachchikumbura SSN, *et al.* (2020). Morphological characterization of microfungi associated with marine based habitats. *Mycosphere* **11**: 1–188.
- de Hoog GS, Gerrits van den Ende AHG (1998). Molecular diagnostics of clinical strains of filamentous Basidiomycetes. *Mycoses* **41**: 183–189.
- Diekmann H (1970). Metabolic products of microorganisms. 81. Occurrence and structures of coprogen B and dimerum acid. *Archiv für Mikrobiologie* **73**: 65–76.
- de Vries GA, de Hoog GS, de Bruyn HP (1984). *Phialophora cyanescens* sp. nov. with Phaeosclera-like synanamorph, causing white-grain mycetoma in man. *Antonie van Leeuwenhoek* **50**: 149–153.
- Dickinson CH (1968). *Gliomastix* Guéguen. *Mycological Papers* **115**: 1–24.

- Diederich P (1990). New or interesting lichenicolous fungi 1. Species from Luxembourg. *Mycotaxon* **37**: 297–330.
- Dingley JM (1951). The *Hypocreales* of New Zealand II. The genus *Nectria*. *Transactions of the Royal Society of New Zealand* **79**: 177–202.
- Dingley JM (1957). Life-history studies of New Zealand species of *Nectria* Fr. *Transactions of the Royal Society of New Zealand* **83**: 643–662.
- Doidge EM (1938). Some South African fusaria. *Bothalia* **3**: 331–483.
- Domsch KH, Gams W, Anderson T-H (2007). *Compendium of soil fungi*. IHW-Verlag, Eching, Germany.
- Edwards J, Auer D, de Alwis S-K, et al. (2016). *Fusarium agapanthi* sp. nov., a novel bikaverin and fusarubin-producing leaf and stem spot pathogen of *Agapanthus praecox* (African lily) from Australia and Italy. *Mycologia* **108**: 981–992.
- Elmer WH, Marra RE (2011). New species of *Fusarium* associated with dieback of *Spartina alterniflora* in Atlantic salt marshes. *Mycologia* **103**: 806–819.
- Evidente A, Conti L, Altomare C, et al. (1994). Fusapyrone and deoxyfusapyrone, two antifungal α -pyrones from *Fusarium semitectum*. *Natural Toxins* **2**: 4–13.
- Ezekiel CN, Kraak B, Sandoval-Denis M, et al. (2020). Diversity and toxigenicity of fungi and description of *Fusarium madaense* sp. nov. from cereals, legumes and soils in north-central Nigeria. *MycoKeys* **67**: 95–124.
- Farr ML (1973). An annotated list of Spegazzini's fungus taxa. *Bibliotheca Mycologica* **35**: 503–507.
- Fawcett HS (1908). *Fungi parasitic upon Aleyrodes citri*. Master of Science thesis. The University of the State of Florida, USA.
- Felsenstein J (1985). Confidence limits on phylogenies: An approach using the bootstrap. *Evolution* **39**: 783–791.
- Fisher NL, Burgess LW, Toussoun TA, et al. (1982). Carnation leaves as a substrate and for preserving cultures of *Fusarium* species. *Phytopathology* **72**: 151–153.
- Forin N, Vizzini A, Nigris S, et al. (2020). Illuminating type collections of the “nectriaceous” fungi in Saccardo's fungarium. *Persoonia* **45**: 221–249.
- Freeman S, Sharon M, Maymon M, et al. (2013). *Fusarium euwallaceae* sp. nov. – a symbiotic fungus of *Euwallacea* sp., an invasive ambrosia beetle in Israel and California. *Mycologia* **105**: 1595–1606.
- Fuckel KWGL (1863). *Fungi Rhenanae Exsiccatae, Fasc. I–VIII*.
- Furumoto T, Fukuyama K, Hamasaki T, et al. (1995). Neovasipyrone and neovasifuranones: Four new metabolites related to neovasinin, a phytotoxin of the fungus, *Neocosmospora vasinfecta*. *Phytochemistry* **40**: 745–751.
- Furumoto T, Hamasaki T, Nakajima H (1997). Vasinfectins A and B: new phytotoxins from *Neocosmospora vasinfecta*. *Tetrahedron Letters* **38**: 5523–5524.
- Gargouri S, Balmas V, Burgess L, et al. (2020). An endophyte of *Macrochloa tenacissima* (esparto or needle grass) from Tunisia is a novel species in the *Fusarium redolens* species complex. *Mycologia* **112**: 792–807.
- Gams W (1971). *Cephalosporium-artige Schimmelpilze*. Fischer, Stuttgart, Germany.
- Gams W (1999). Report of the Committee for Fungi: 8. *Taxon* **48**: 807–810.
- Gams W, Gerlach M (1968). Beiträge zur Systematic und Biologie von *Plectosphaerella cucumeris* und der zugehörigen Konidienform. *Persoonia* **5**: 177–188.
- Gams W, Klamer M, O'Donnell K (1999). *Fusarium miscanthi* sp. nov. from *Miscanthus* litter. *Mycologia* **91**: 263–268.
- Gams W, Nirenberg HI, Seifert KA, et al. (1997). (1275) Proposal to conserve the name *Fusarium sambucinum* (Hyphomycetes). *Taxon* **46**: 111–113.
- Gao J, Radwan MM, León F, et al. (2013). *Neocosmospora* sp.-derived resorcylic acid lactones with *in vitro* binding affinity for human opioid and cannabinoid receptors. *Journal of Natural Products* **76**: 824–828.
- Geiser DM, Al-Hatmi AMS, Aoki T, et al. (2021). Phylogenomic analysis of a 55.1 kb 19-gene dataset resolves a monophyletic *Fusarium* that includes the *Fusarium solani* species complex. *Phytopathology*. <https://doi.org/10.1094/PHYTO-08-20-0330-LE>.
- Geiser DM, Aoki T, Bacon CW, et al. (2013). One fungus, one name: Defining the genus *Fusarium* in a scientifically robust way that preserves long-standing use. *Phytopathology* **103**: 400–408.
- Geiser DM, Juba JH, Wang B, et al. (2001). *Fusarium hostae* sp. nov., a relative of *F. redolens* with a *Gibberella* teleomorph. *Mycologia* **93**: 670–678.
- Geiser DM, Lewis Ivey ML, Hakiza G, et al. (2005). *Gibberella xyloarioides* (anamorph: *Fusarium xyloarioides*), a causative agent of coffee wilt disease in Africa, is a previously unrecognized member of the *G. fujikuroi* species complex. *Mycologia* **97**: 191–201.
- Gerlach W (1977). Drei neue Varietäten von *Fusarium merismoides*, *F. larvarum* und *F. chlamyosporum*. *Phytopathologische Zeitschrift* **90**: 31–42.
- Gerlach W (1977). *Fusarium lunulosporum* spec. nov. von Grapefruit aus Südafrika, ein Fruchtfäuleerreger. *Phytopathologische Zeitschrift* **88**: 280–284.
- Gerlach W (1977). *Fusarium robustum* spec. nov., der Erreger einer Stammfäule an *Araucaria angustifolia* (Bertol.) O. Kuntze in Argentinien? *Phytopathologische Zeitschrift* **88**: 29–37.
- Gerlach W, Nirenberg H (1982). The genus *Fusarium* – a pictorial atlas. *Mitteilungen aus der Biologischen Bundesanstalt für Land- und Forstwirtschaft Berlin-Dahlem* **209**: 1–406.
- Gerlach W, Pag H (1961). *Fusarium redolens* Wr., seine phytopathologische Bedeutung und eine an *Dianthus*-Arten gefäßparasitäre Form (*F. redolens* Wr. f. *dianthi* Gerlach). *Phytopathologische Zeitschrift* **42**: 349–361.
- German-Fattal M (2001). Fusafungine, an antimicrobial with anti-inflammatory properties in respiratory tract infections. *Clinical Drug Investigation* **21**: 653–670.
- Giraldo A, Crous PW (2019). Inside *Plectosphaerellaceae*. *Studies in Mycology* **92**: 227–286.
- González CD, Chaverri P (2017). *Corinectria*, a new genus to accommodate *Neonectria fuckeliana* and *C. constricta* sp. nov. from *Pinus radiata* in Chile. *Mycological Progress* **16**: 1015–1027.
- Gordon WL (1952). The occurrence of *Fusarium* species in Canada. II. Prevalence and taxonomy of *Fusarium* species in cereal seeds. *Canadian Journal of Botany* **30**: 209–251.
- Gordon WL (1959). The occurrence of *Fusarium* species in Canada. VI. Taxonomy and geographic distribution of *Fusarium* species in plants, insects, and fungi. *Canadian Journal of Botany* **37**: 257–290.
- Gräfenhan T, Johnston PR, Vaughan MM, et al. (2016). *Fusarium praegraminearum* sp. nov., a novel trichothecene mycotoxin-producing pathogen from New Zealand can induce head blight on wheat. *Mycologia* **108**: 1229–1239.
- Gräfenhan T, Schroers H-J, Nirenberg HI, et al. (2011). An overview of the taxonomy, phylogeny, and typification of nectriaceous fungi in *Cosmospora*, *Acremonium*, *Fusarium*, *Stilbella* and *Volutella*. *Studies in Mycology* **68**: 79–113.
- Grove JF, Pople M (1979). Metabolic products of *Fusarium larvarum* Fuckel. The fusarentins and the absolute configuration of monocerin. *Journal of the Chemical Society, Perkin Transactions 1* **1979**: 2048–2051.
- Guarnaccia V, Sandoval-Denis M, Aiello D, et al. (2018). *Neocosmospora perseae* sp. nov., causing trunk cankers on avocado in Italy. *Fungal Systematics and Evolution* **1**: 131–140.
- Guarnaccia V, Van Niekerk J, Crous PW, et al. (2021). *Neocosmospora* spp. associated with dry root rot of citrus in South Africa. *Phytopathologia Mediterranea* **60**: 81–102.
- Guo Y, Zhang D, Tang W (2018). *Fusarium graminearum* double (triple) mutants generation using sexual crosses. *Bio-* **101**: e2961.
- Guu J-R, Ju Y-M, Hsieh H-J (2007). Nectriaceous fungi collected from forests in Taiwan. *Botanical Studies* **48**: 187–203.
- Hall TA (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series* **41**: 95–98.
- Hawksworth DL (1979). The lichenicolous Hyphomycetes. *Bulletin of the British Museum (Natural History), Botany* **6**: 183–300.
- Hawksworth DL, Crous PW, Redhead SA, et al. (2011). The Amsterdam Declaration on Fungal Nomenclature. *IMA Fungus* **2**: 105–112.
- Heath TA, Hedtke SM, Hillis DM (2008). Taxon sampling and the accuracy of phylogenetic analyses. *Journal of Systematics and Evolution* **46**: 239–257.
- Hein B (1988). Liste der Arten und infraspezifischen Taxa von P. Hennings mit Angabe der Typen in den Herbarien des Botanischen Museums Berlin-Dahlem und des Instituts für Allgemeine Botanik in Hamburg. *Englera* **10**: 1–374.
- Henderson J, Dobson V (1994). *Eastern & Central Arrernte to English dictionary*. IAD Press, Alice Springs, Australia.
- Herron DA, Wingfield MJ, Wingfield BD, et al. (2015). Novel taxa in the *Fusarium fujikuroi* species complex from *Pinus* spp. *Studies in Mycology* **80**: 131–150.
- Hirooka Y, Rossman AY, Samuels GJ, et al. (2012). A monograph of *Allantonectria*, *Nectria*, and *Pleonectria* (Nectriaceae, Hypocreales, Ascomycota) and their pycnidial, sporodochial, and synnematosous anamorphs. *Studies in Mycology* **71**: 1–210.
- Hoang DT, Chernomor O, von Haeseler A, et al. (2018). UFBoot2: improving the ultrafast bootstrap approximation. *Molecular Biology and Evolution* **35**: 518–522.
- Holubová-Jechová V, Gams W, Nirenberg HI (1994). Revisiones Generum Obscurorum Hyphomycetum: a revision of the *Selenosporium* species described by A.C.J. Corda. *Sydowia* **46**: 247–256.
- Hosoya T, Tubaki K (2004). *Fusarium matuoi* sp. nov. and its teleomorph *Cosmospora matuoi* sp. nov. *Mycoscience* **45**: 261–270.

- Hu J, Chen C, Peever T, *et al.* (2012). Genomic characterization of the conditionally dispensable chromosome in *Alternaria arborescens* provides evidence for horizontal gene transfer. *BMC Genomics* **13**: 171.
- Huang SK, Jeewon R, Hyde KD, *et al.* (2018). Novel taxa within *Nectriaceae*: *Cosmosporella* *gen. nov.* and *Aquanectria* *sp. nov.* from freshwater habitats in China. *Cryptogamie, Mycologie* **39**: 169–192.
- Jacobs A, van Wyk PS, Marasas WFO, *et al.* (2010). *Fusarium ananatum* *sp. nov.* in the *Gibberella fujikuroi* species complex from pineapples with fruit rot in South Africa. *Fungal Biology* **114**: 515–527.
- Jacobs-Venter A, Laraba I, Geiser D, *et al.* (2018). Molecular systematics of two sister clades, the *Fusarium concolor* and *F. babinda* species complexes, and the discovery of a novel microcycle macroconidium-producing species from South Africa. *Mycologia* **110**: 1189–1204.
- Jamalainien EA (1943). Über die fusarien Finnlands II. *Valtion Maatalouskoe-toiminnan Julkaisuja* **123**: 1–24.
- Jang JH, Asami Y, Jang JP, *et al.* (2011). Fusarisetin A, an acinar morphogenesis inhibitor from a soil fungus, *Fusarium* sp. FN080326. *Journal of the American Chemical Society* **133**: 6865–6867.
- Jiang Z, Barret MO, Boyd KG, *et al.* (2002). JM47, a cyclic tetrapeptide HC-toxin analogue from a marine *Fusarium* species. *Phytochemistry* **60**: 33–38.
- Jiménez M, Huerta T, Mateo R (1997). Mycotoxin production by *Fusarium* species isolated from bananas. *Applied and Environmental Microbiology* **63**: 364–369.
- Joffe AZ (1974). A modern system of *Fusarium* taxonomy. *Mycopathologia et Mycologia Applicata* **53**: 201–228.
- Jones EBG, Devadatha B, Abdel-Wahab MA, *et al.* (2020). Phylogeny of new marine *Dothideomycetes* and *Sordariomycetes* from mangroves and deep-sea sediments. *Botanica Marina* **63**: 155–181.
- Kalyanamoorthy S, Minh BQ, Wong TKF, *et al.* (2017). ModelFinder: Fast model selection for accurate phylogenetic estimates. *Nature Methods* **14**: 587–589.
- Katoh K, Rozewicki J, Yamada KD (2019). MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Briefings in Bioinformatics* **20**: 1160–1166.
- Kearse M, Moir R, Wilson A, *et al.* (2012). Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* **28**: 1647–1649.
- Kerényi Z, Moretti A, Waalwijk C, *et al.* (2004). Mating type sequences in asexually reproducing *Fusarium* species. *Applied and Environmental Microbiology* **70**: 4419–4423.
- Kerényi Z, Zeller KA, Hornok L, *et al.* (1999). Standardization of mating type terminology in the *Gibberella fujikuroi* species complex. *Applied and Environmental Microbiology* **65**: 4071–4076.
- Kim W, Cavinder B, Proctor RH, *et al.* (2019). Comparative genomics and transcriptomics during sexual development gives insight into the life history of the cosmopolitan fungus *Fusarium neocosmosporiellum*. *Frontiers in Microbiology* **10**: 1247.
- Kimura Y, Hamasaki T, Nakajima H (1981). Isolation, identification and biological activities of 8-O-methyl-javanicin produced by *Fusarium solani*. *Agricultural and Biological Chemistry* **45**: 2653–2654.
- Kirk PM, Stalpers JA, Braun U, *et al.* (2013). A without-prejudice list of generic names of fungi for protection under the International Code of Nomenclature for algae, fungi, and plants. *IMA Fungus* **4**: 381–443.
- Klittich C, Leslie JF (1988). Nitrate reduction mutants of *Fusarium moniliforme*. *Genetics* **118**: 417–423.
- Klittich CJR, Leslie JF, Nelson PE, *et al.* (1997). *Fusarium thapsinum* (*Gibberella thapsina*): a new species in section *Liseola* from sorghum. *Mycologia* **89**: 643–652.
- Komada H (1975). Development of a selective medium for quantitative isolation of *Fusarium oxysporum* from natural soil. *Review of Plant Protection Research* **8**: 114–124.
- Kulkarni GB, Sajjan S, Koregrudar JB (2011). Pathogenicity of indole-acetic acid producing fungus *Fusarium delphinoides* strain GPK towards chick pea and pigeon pea. *European Journal of Plant Pathology* **131**: 355–369.
- Kulkarni GB, Sanjeevkumar C, Kirankumar B, *et al.* (2013). Indole-3-acetic acid biosynthesis in *Fusarium delphinoides* strain GBK, a causal agent of wilt in chick pea. *Applied Biochemistry and Biotechnology* **169**: 1292–1305.
- Kurobane I, Vining LC, McInnes AG, *et al.* (1980). Metabolites of *Fusarium solani* related to dihydrofusarubin. *Journal of Antibiotics* **33**: 1376–1379.
- Kurtzman CP, Robnett CJ (1997). Identification of clinically important ascomycetous yeasts based on nucleotide divergence in the 5' end of the large-subunit (26S) ribosomal DNA gene. *Journal of Clinical Microbiology* **35**: 1216–1223.
- Kykykyku JO, Kusari S, Adosraku RK, *et al.* (2017). Antibacterial secondary metabolites from an endophytic fungus. *Fusarium solani* JK10. *Fitoterapia* **119**: 108–114.
- Laraba I, Keddad A, Boureghda H, *et al.* (2017). *Fusarium algeriense*, *sp. nov.*, a novel toxigenic crown rot pathogen of durum wheat from Algeria is nested in the *Fusarium burgessii* species complex. *Mycologia* **109**: 935–950.
- Laraba I, Kim HS, Proctor RH, *et al.* (2020). *Fusarium xyrophilum*, *sp. nov.*, a member of the *Fusarium fujikuroi* species complex recovered from pseudoflowers on yellow-eyed grass (*Xyris* spp.) from Guyana. *Mycologia* **112**: 39–51.
- Laurence MH, Summerell BA, Burgess LW, *et al.* (2011). *Fusarium burgessii* *sp. nov.* representing a novel lineage in the genus *Fusarium*. *Fungal Diversity* **49**: 101–112.
- Laurence MH, Walsh JL, Shuttleworth LA, *et al.* (2016). Six novel species of *Fusarium* from natural ecosystems in Australia. *Fungal Diversity* **77**: 349–366.
- Lechat C, Fournier J (2015). *Varicosporella*, a new aquatic genus in the *Nectriaceae* from France. *Ascomycete.org* **7**: 1–8.
- Lechat C, Fournier J, Nordén B (2015). *Stylonectria norvegica* (*Nectriaceae*), a new species from Norway. *Ascomycete.org* **7**: 220–224.
- Lee HS, Lee C (2012). Structural analysis of a new cytotoxic demethylated analogue of neo-n-methylsalsalvamide with a different peptide sequence produced by *Fusarium solani* isolated from potato. *Journal of Agricultural and Food Chemistry* **60**: 4342–4347.
- Leslie JF, Summerell BA (2006). *The Fusarium laboratory manual*. Blackwell Publishing Professional, USA.
- Leslie JF, Summerell BA, Bullock S, *et al.* (2005). Description of *Gibberella sacchari* and neotypification of its anamorph *Fusarium sacchari*. *Mycologia* **97**: 718–724.
- Lima CS, Pfenning LH, Costa SS, *et al.* (2012). *Fusarium tupiense* *sp. nov.*, a member of the *Gibberella fujikuroi* complex that causes mango malformation in Brazil. *Mycologia* **104**: 1408–1419.
- Lindegg G (1935). Cancro picciolare dell'acanto "*Acanthus mollis*" L. *Rivista di Patologia Vegetale* **25**: 229–235.
- Link HF (1809). Observationes in ordines plantarum naturales. *Dissertatio I Magazin der Gesellschaft Naturforschenden Freunde Berlin* **3**: 3–42.
- Liu D, Li XM, Li CS, *et al.* (2013). Sesterterpenes and 2H-pyran-2-ones (= α -pyrones) from the mangrove-derived endophytic fungus *Fusarium proliferatum* MA-84. *Helvetica Chimica Acta* **96**: 437–444.
- Liu Q, Li JQ, Wingfield MJ, *et al.* (2020). Reconsideration of species boundaries and proposed DNA barcodes for *Calonectria*. *Studies in Mycology* **97**: 100106.
- Liu YJ, Whelen S, Hall BD (1999). Phylogenetic relationships among ascomycetes: evidence from an RNA polymerase II subunit. *Molecular Biology and Evolution* **16**: 1799–1808.
- Lombard L, Houbraken J, Decock C, *et al.* (2016). Generic hyper-diversity in *Stachybotriaceae*. *Persoonia* **36**: 156–246.
- Lombard L, Sandoval-Denis M, Lamprecht SC, *et al.* (2019). Epitypification of *Fusarium oxysporum* – clearing the taxonomic chaos. *Persoonia* **43**: 1–47.
- Lombard L, van der Merwe NA, Groenewald JZ, *et al.* (2014). Lineages in *Nectriaceae*: re-evaluating the generic status of *Ilyonectria* and allied genera. *Phytopathologia Mediterranea* **53**: 515–532.
- Lombard L, van der Merwe NA, Groenewald JZ, *et al.* (2015). Generic concepts in *Nectriaceae*. *Studies in Mycology* **80**: 189–245.
- Lombard L, van Doorn R, Crous PW (2019). Neotypification of *Fusarium chlamydosporum* – a reappraisal of a clinically important species complex. *Fungal Systematics and Evolution* **4**: 183–200.
- Luo J, Zhuang WY (2008). Two new species of *Cosmospora* (*Nectriaceae*, *Hypocreales*) from China. *Fungal Diversity* **31**: 83–93.
- Lynn KMT, Wingfield MJ, Duran A, *et al.* (2020). *Euwallacea perbrevis* (*Coleoptera: Curculionidae: Scolytinae*), a confirmed pest on *Acacia crassicaarpa* in Riau, Indonesia, and a new fungal symbiont; *Fusarium rekanum* *sp. nov.* *Antonie van Leeuwenhoek* **113**: 803–823.
- Madelin MF (1966). *Trichothecium acridiorum* (Trabut) *comb. nov.* on red locusts. *Transactions of the British Mycological Society* **49**: 275–288.
- Marasas WFO, Nelson PE, Toussoun TA (1985). *Fusarium dlamini*, a new species from Southern Africa. *Mycologia* **77**: 971–975.
- Marasas WFO, Nelson PE, Toussoun TA, *et al.* (1986). *Fusarium phialidicum*, a new species from South Africa. *Mycologia* **78**: 678–682.
- Marasas WFO, Rabie CJ, Lübber A, *et al.* (1987). *Fusarium napiforme*, a new species from millet and sorghum in Southern Africa. *Mycologia* **79**: 910–914.
- Marasas WFO, Rheeder JP, Lamprecht SC, *et al.* (2001). *Fusarium andiyazi* *sp. nov.*, a new species from sorghum. *Mycologia* **93**: 1203–1210.

- Marasas WFO, Rheeder JP, Logrieco A, et al. (1998). *Fusarium nelsonii* and *F. musarum*: two new species in section *Arthrosporiella* related to *F. camptoceras*. *Mycologia* **90**: 505–513.
- Marcano V, Benitez P, Palacios-Prü E (2001). Ecophysiological responses of a new *Fusarium* grown in non-aromatic hydrocarbon media $\geq \eta\text{-C}_{12}$. *Revista de Ecología Latinoamericana* **8**: 1–19.
- Marinach-Patrice C, Lethuillier A, Marly A, et al. (2009). Use of mass spectrometry to identify clinical *Fusarium* isolates. *Clinical Microbiology and Infection* **15**: 634–642.
- Maryani N, Lombard L, Poerba YS, et al. (2019). Phylogeny and genetic diversity of the banana *Fusarium* wilt pathogen *Fusarium oxysporum* f. sp. *cubense* in the Indonesian centre of origin. *Studies in Mycology* **92**: 155–194.
- Maryani N, Sandoval-Denis M, Lombard L, et al. (2019). New endemic *Fusarium* species hitch-hiking with pathogenic *Fusarium* strains causing Panama disease in small-holder banana plots in Indonesia. *Persoonia* **43**: 48–69.
- Matsushima T (1971). *Microfungi of the Solomon Islands and Papua-New Guinea*. Published by the author, Kobe, Japan.
- Matuo T, Kobayashi T (1960). A new *Fusarium*, the conidial stage of *Hypocrea splendens* Phil. et Plowr. *Transactions of the Mycological Society of Japan* **2(4)**: 13–15.
- McAlpine D (1904). Two new fungi parasitic on scale insects. *Journal of the Department of Agriculture of Victoria* **2**: 646–648.
- Minh BQ, Nguyen MAT, von Haeseler A (2013). Ultrafast approximation for phylogenetic bootstrap. *Molecular Biology and Evolution* **30**: 1188–1195.
- Minh Q, Hahn MW, Lanfear R (2020). New methods to calculate concordance factors for phylogenomic datasets. *Molecular Biology and Evolution* **37**: 2727–2733.
- Minh Q, Schmidt HA, Chernomor O, et al. (2020). IQ-TREE2: new models and efficient methods for phylogenetic inference in the genomic era. *Molecular Biology and Evolution* **37**: 1530–1534.
- Moussa TAA, Al-Zharani HS, Kadasa NMS, et al. (2017). Two new species of the *Fusarium fujikuroi* species complex isolated from the natural environment. *Antonie van Leeuwenhoek* **110**: 819–832.
- Munkvold GP (2017). *Fusarium* species and their associated mycotoxins. In: *Mycotoxigenic fungi: methods and protocols* (Moretti A, Susca A, eds), *Methods in Molecular Biology*, **1542**. Springer, Amsterdam: 51–106.
- Na F, Carrillo JD, Mayorquin JS, et al. (2018). Two novel fungal symbionts *Fusarium kuroshium* sp. nov. and *Graphium kuroshium* sp. nov. of Kuroshio shot hole borer (*Euwallacea* sp. nr. *fornicatus*) cause fusarium dieback on woody host species in California. *Plant Disease* **102**: 1154–1164.
- Nakajima H, Hamasaki T, Tanaka K, et al. (1989). Production of cyclosporin by fungi belonging to the genus *Neocosmospora*. *Agricultural and Biological Chemistry* **53**: 2291–2292.
- Nakajima H, Shimomura K, Furumoto T, et al. (1995). Neovasipyrindones A, B and C: Metabolites related to neovasinin, a phytotoxin of the fungus, *Neocosmospora vasinfecta*. *Phytochemistry* **40**: 1643–1647.
- Nalim FA, Samuels GJ, Wijesundera RL, et al. (2011). New species from the *Fusarium solani* species complex derived from perithecia and soil in the Old-World tropics. *Mycologia* **103**: 1302–1330.
- Nash S, Snyder WC (1962). Quantitative estimations by plate counts of propagules of the bean root rot *Fusarium* in field soils. *Phytopathology* **52**: 567–572.
- Nelson PE, Dignani MC, Anaissie EJ (1994). Taxonomy, biology, and clinical aspects of *Fusarium* species. *Clinical Microbiology Reviews* **7**: 479–504.
- Nelson PE, Toussoun TA, Burgess LW (1987). Characterization of *Fusarium beomiforme* sp. nov. *Mycologia* **79**: 884–889.
- Nelson PE, Toussoun TA, Marasas WFO (1983). *Fusarium species: An illustrated manual for identification*. Pennsylvania State University Press, USA.
- Nelson PE, Toussoun TA, Marasas WFO (1995). Neotypification and emended description of *Fusarium anguioides*. *Mycologia* **87**: 543–546.
- Nenkép V, Yun K, Son BW (2016). Oxysporizoline, an antibacterial polycyclic quinazoline alkaloid from the marine-mudflat-derived fungus *Fusarium oxysporum*. *Journal of Antibiotics* **69**: 709–711.
- Nguyen LT, Schmidt HA, von Haeseler A, et al. (2015). IQ-TREE: a fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Molecular Biology and Evolution* **32**: 268–274.
- Nilsson RH, Tedersoo L, Abarenkov K, et al. (2012). Five simple guidelines for establishing basic authenticity and reliability of newly generated fungal ITS sequences. *Mycocokeys* **4**: 37–63.
- Nirenberg HI (1976). Untersuchungen über die morphologische und biologische Differenzierung in der *Fusarium*-Sektion *Liseola*. *Mitteilungen aus der Biologischen Bundesanstalt für Land- und Forstwirtschaft Berlin-Dahlem* **169**: 1–117.
- Nirenberg HI (1981). A simplified method for identifying *Fusarium* spp. occurring on wheat. *Canadian Journal of Botany* **59**: 1599–1609.
- Nirenberg HI (1990). Recent advances in the taxonomy of *Fusarium*. *Studies in Mycology* **32**: 91–101.
- Nirenberg HI (1995). Morphological differentiation of *Fusarium sambucinum* Fuckel sensu stricto, *F. torulosum* (Berk. & Curt.) Nirenberg comb. nov. and *F. venenatum* Nirenberg comb. nov. *Mycopathologia* **129**: 131–141.
- Nirenberg HI, Aoki T (1997). *Fusarium nisikadoi*, a new species from Japan. *Mycoscience* **38**: 329–333.
- Nirenberg HI, Brielmairers-Liebetanz U (1996). *Nectria ipomoeae* Halst., anamorph: *Fusarium striatum* Sherb. on *Passiflora edulis* Sims. *Nachrichtenblatt des Deutschen Pflanzenschutzdienstes* **48**: 270–275.
- Nirenberg HI, Hagedorn G (2008). *Fusarium nematophilum* spec. nov. – ein neuer Nematoden-assoziiierter Pilz. *Nachrichtenblatt des Deutschen Pflanzenschutzdienstes* **60**: 213–216.
- Nirenberg HI, O'Donnell K (1998). New *Fusarium* species and combinations within the *Gibberella fujikuroi* species complex. *Mycologia* **90**: 434–458.
- Nirenberg HI, Samuels GJ (2000). *Nectria* and *Fusarium*. II. *Cosmospora zealandica* comb. nov. and its anamorph, *Fusarium zealandicum* sp. nov. *Canadian Journal of Botany* **78**: 1482–1487.
- Nirenberg HI, O'Donnell K, Kroschel J, et al. (1998). Two new species of *Fusarium*: *Fusarium brevicatenulatum* from the noxious weed *Striga asiatica* and *Fusarium pseudoanthophilum* from *Zea mays* in Zimbabwe. *Mycologia* **90**: 459–463.
- Nylander JAA (2004). *MrModeltest v. 2*. Evolutionary Biology Centre. Uppsala University. Programme distributed by the author.
- O'Donnell K (1993). *Fusarium* and its near relatives. In: *The fungal holomorph: mitotic, meiotic and pleomorphic speciation in fungal systematics* (Reynolds DR, Taylor JW, eds). CAB International, UK: 225–233.
- O'Donnell K (2000). Molecular phylogeny of the *Nectria haematococca*–*Fusarium solani* species complex. *Mycologia* **92**: 919–938.
- O'Donnell K, Al-Hatmi AMS, Aoki T, et al. (2020). No to *Neocosmospora*: Phylogenomic and practical reasons for continued inclusion of the *Fusarium solani* species complex in the genus *Fusarium*. *mSphere* **5**: e00810–e00820.
- O'Donnell K, Cigelnik E (1997). Two divergent intragenomic rDNA ITS2 types within a monophyletic lineage of the fungus *Fusarium* are nonorthologous. *Molecular Phylogenetics and Evolution* **7**: 103–116.
- O'Donnell K, Cigelnik E, Nirenberg HI (1998). Molecular systematics and phylogeography of the *Gibberella fujikuroi* species complex. *Mycologia* **90**: 465–493.
- O'Donnell K, Kistler HC, Cigelnik E, et al. (1998). Multiple evolutionary origins of the fungus causing Panama disease of banana: Concordant evidence from nuclear and mitochondrial gene genealogies. *Proceedings of the National Academy of Sciences of the USA* **95**: 2044–2049.
- O'Donnell K, Kistler HC, Tacke BK, et al. (2000). Gene genealogies reveal global phylogeographic structure and reproductive isolation among lineages of *Fusarium graminearum*, the fungus causing wheat scab. *Proceedings of the National Academy of Sciences of the USA* **97**: 7905–7910.
- O'Donnell K, McCormick SP, Busman M, et al. (2018). Marasas et al. 1984 “Toxicigen *Fusarium* species: Identity and mycotoxicology” revisited. *Mycologia* **110**: 1058–1080.
- O'Donnell K, Nirenberg HI, Aoki T, et al. (2000). A multigene phylogeny of the *Gibberella fujikuroi* species complex: detection of additional phylogenetically distinct species. *Mycoscience* **41**: 61–78.
- O'Donnell K, Rooney AP, Proctor RH, et al. (2013). Phylogenetic analyses of *RPB1* and *RPB2* support a middle Cretaceous origin for a clade comprising all agriculturally and medically important fusaria. *Fungal Genetics and Biology* **52**: 20–31.
- O'Donnell K, Sarver BAJ, Brandt M, et al. (2007). Phylogenetic diversity and microsphere array-based genotyping of human pathogenic fusaria, including isolates from the multistate contact lens-associated U.S. keratitis outbreak of 2005 and 2006. *Journal of Clinical Microbiology* **45**: 2235–2248.
- O'Donnell K, Sutton DA, Fothergill A, et al. (2008). Molecular phylogenetic diversity, multilocus haplotype nomenclature, and *in vitro* antifungal resistance within the *Fusarium solani* species complex. *Journal of Clinical Microbiology* **46**: 2477–2490.
- O'Donnell K, Sutton DA, Rinaldi MG, et al. (2009). Novel multilocus sequence typing scheme reveals high genetic diversity of human pathogenic members of the *Fusarium incarnatum*-*F. equiseti* and *F. chlamydosporum* species

- complexes within the United States. *Journal of Clinical Microbiology* **47**: 3851–3861.
- O'Donnell K, Sutton DA, Rinaldi MG, *et al.* (2010). Internet-accessible DNA sequence database for identifying fusaria from human and animal infections. *Journal of Clinical Microbiology* **48**: 3708–3718.
- O'Donnell K, Sutton DA, Wiederhold N, *et al.* (2016). Veterinary fusarioses within the United States. *Journal of Clinical Microbiology* **54**: 2813–2819.
- O'Donnell K, Ward TJ, Aberra D, *et al.* (2008). Multilocus genotyping and molecular phylogenetics resolve a novel head blight pathogen within the *Fusarium graminearum* species complex from Ethiopia. *Fungal Genetics and Biology* **45**: 1514–1522.
- O'Donnell K, Ward TJ, Geiser DM, *et al.* (2004). Genealogical concordance between the mating type locus and seven other nuclear genes supports formal recognition of nine phylogenetically distinct species within the *Fusarium graminearum* clade. *Fungal Genetics and Biology* **41**: 600–623.
- Ohshima S, Yanagisawa M, Katoh A, *et al.* (1994). *Fusarium merismoides* CORDA NR 6356, the source of the protein kinase C inhibitor, azepinostatin. Taxonomy, yield improvement, fermentation and biological activity. *Journal of Antibiotics* **47**: 639–647.
- Otero-Colina G, Rodríguez-Alvarado G, Fernández-Pavía S, *et al.* (2010). Identification and characterization of a novel etiological agent of mango malformation disease in Mexico, *Fusarium mexicanum* sp. nov. *Phytopathology* **100**: 1176–1184.
- Padwick GW (1945). Notes on Indian fungi III. *Mycological Papers* **12**: 1–12.
- Palm ME, Gams W, Nirenberg HI (1995). *Plectosporium*, a new genus for *Fusarium tabacinum*, the anamorph of *Plectosphaerella cucumerina*. *Mycologia* **87**: 397–406.
- Parish CA, Smith SK, Calati K, *et al.* (2008). Isolation and structure elucidation of pamafungins, antifungal natural products that inhibit mRNA polyadenylation. *Journal of the American Chemical Society* **130**: 7060–7066.
- Pascoe IG (1990). *Fusarium* morphology I: Identification and characterization of a third conidial type, the mesoconidium. *Mycotaxon* **37**: 121–160.
- Pasinetti L, Buzzati-Traverso A (1935). Su alcune forme di cancrena delle cactacee dovute a nuovi micromiceti e ad un batterio. *Giornale Botanico Italiano* **42**: 89–123.
- Paziani MH, Carvalho LT, Melhem MSC, *et al.* (2020). First comprehensive report of clinical *Fusarium* strains isolated in the state of Sao Paulo (Brazil) and identified by MALDI-TOF MS and molecular biology. *Microorganisms* **8**: 66.
- Penzig AGO (1882). *Funghi agrumicoli: contribuzione allo studio dei funghi parassiti degli agrumi*. Padova, Italy.
- Pereira CB, Ward TJ, Tessmann DJ, *et al.* (2018). *Fusarium subtropicale*, sp. nov., a novel nivalenol mycotoxin-producing species isolated from barley (*Hordeum vulgare*) in Brazil and sister to *F. praegraminearum*. *Mycologia* **110**: 860–871.
- Perera RH, Hyde KD, Maharachchikumbura SSN, *et al.* (2020). Fungi on wild seeds and fruits. *Mycosphere* **11**: 2108–2480.
- Petch T (1920). Studies in entomogenous fungi. The Nectriae parasitic on scale insects. *Transactions of the British Mycological Society* **7**: 133–166.
- Pfenning L (1995). A new species of *Neocosmospora* from Brazil. *Sydowia* **47**: 65–69.
- Pfenning LH, de Melo MP, Costa MM, *et al.* (2019). *Fusarium udum* revisited: a common, but poorly understood member of the *Fusarium fujikuroi* species complex. *Mycological Progress* **18**: 107–117.
- Phan HT, Burgess LW, Summerell BA, *et al.* (2004). *Gibberella gaditjirii* (*Fusarium gaditjirii*) sp. nov., a new species from tropical grasses in Australia. *Studies in Mycology* **50**: 261–272.
- Pilat A (1938). Liste der von A. C. J. Corda beschriebenen Pilzarten, mit Angabe der Originallexemplare, die im Herbarium des Nationalmuseums in Prag aufbewahrt sind. *Acta Musei Nationalis Pragae, Botanica* no. 3, 1B no. 10.
- Plattner PA, Nager U, Boller A (1948). Welkstoffe und Antibiotika. 7. Mitteilung. Über die Isolierung neuartiger Antibiotika aus Fusarien. *Helvetica Chimica Acta* **31**: 594–602.
- Price MN, Dehal PS, Arkin AP (2010). FastTree 2 – approximately maximum-likelihood trees for large alignments. *PLoS ONE* **5**: e9490.
- Quaedvlieg W, Kema GHJ, Groenewald JZ, *et al.* (2011). *Zymoseptoria* gen. nov.: a new genus to accommodate *Septoria*-like species occurring on graminicolous hosts. *Persoonia* **26**: 57–69.
- Raillo AI (1950). *Griby roda Fusarium*. Gosudarstv. Izd. Sel'skochoz. Lit., Moskva.
- Rambaut A, Drummond AJ, Xie D, *et al.* (2018). Posterior summarization in Bayesian phylogenetics using Tracer 1.7. *Systematic Biology* **67**: 901–904.
- Rayner RW (1970). *A mycological colour chart*. CMI and British Mycological Society, Kew, Surrey, UK.
- Reeb V, Lutzoni F, Roux C (2004). Contribution of RPB2 to multilocus phylogenetic studies of the euascomycetes (Pezizomycotina, Fungi) with special emphasis on the lichen-forming *Acarosporaceae* and evolution of polyspory. *Molecular Phylogenetics and Evolution* **32**: 1036–1060.
- Reddy GL, Reddy SM (1992). Production of IAA by some species of *Fusarium*. *National Academy Science Letters, India* **15**: 247–249.
- Renner MK, Jensen PR, Fenical W (2000). Mangicols: structures and biosynthesis of a new class of sesterterpene polyols from a marine fungus of the genus *Fusarium*. *Journal of Organic Chemistry* **65**: 4843–4852.
- Rheeder JP, Marasas WF, Nelson PE (1996). *Fusarium globosum*, a new species from corn in southern Africa. *Mycologia* **88**: 509–513.
- Ronquist F, Huelsenbeck JP (2003). MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* **19**: 1572–1574.
- Roos A (1977). *Physiology and pathogenicity of Neocosmospora vasinfecta EF Smith*. PhD thesis. ETH Zurich, Switzerland.
- Rossmann AY (1979). A preliminary account of the taxa described in *Calonectria*. *Mycotaxon* **8**: 485–558.
- Rossmann AY (1983). The phragmosporous species of *Nectria* and related genera. *Mycological Papers* **150**: 1–164.
- Rossmann AY, Allen WC, Braun U, *et al.* (2016). Overlooked competing asexual and sexually typified generic names of Ascomycota with recommendations for their use or protection. *IMA Fungus* **7**: 289–308.
- Rossmann AY, Samuels GJ, Rogerson CT, *et al.* (1999). Genera of *Bionectriaceae*, *Hypocreaceae* and *Nectriaceae* (*Hypocreales*, *Ascomycetes*). *Studies in Mycology* **42**: 1–248.
- Rossmann AY, Seifert KA (2011). Phylogenetic revision of taxonomic concepts in the *Hypocreales* and other *Ascomycota*. A tribute to Gary J. Samuels. *Studies in Mycology* volume 68, CBS-KNAW Fungal Biodiversity Centre, Utrecht, The Netherlands.
- Rossmann AY, Seifert KA, Samuels GJ, *et al.* (2013). Genera of *Bionectriaceae*, *Hypocreaceae* and *Nectriaceae* (*Hypocreales*) proposed for acceptance and rejection. *IMA Fungus* **4**: 41–51.
- Rostrup E (1894). Sygdomme hos Landbrugsplanter foraarsagede af Snyltesvampe. *Botanisches Zentralblatt* **15**: 284–285.
- Saccardo PA (1892). Supplementum Universale, Pars II. *Discomyceteae-Hyphomyceteae*. *Sylloge Fungorum* **10**: 1–964.
- Saccardo PA (1899). *Sylloge fungorum* **14**: 725–1316.
- Saccardo PA (1906). *Notae mycologicae*. Series VII. *Annales Mycologici* **4**: 273–278.
- Salgado-Salazar C, Rossmann AY, Chaverri P (2016). The genus *Thelonectria* (*Nectriaceae*, *Hypocreales*, *Ascomycota*) and closely related species with cylindrocarpon-like asexual states. *Fungal Diversity* **80**: 411–455.
- Salichos L, Stamatakis A, Rokas A (2014). Novel information theory-based measures for quantifying incongruence among phylogenetic trees. *Molecular Biology and Evolution* **31**: 1261–1271.
- Samson RA (1974). *Paecilomyces* and some allied hyphomycetes. *Studies in Mycology* **6**: 1–119.
- Samson RA, Visagie CM, Houbaker J, *et al.* (2014). Phylogeny, identification and nomenclature of the genus *Aspergillus*. *Studies in Mycology* **78**: 141–173.
- Samuels GJ (1976). A revision of the fungi formerly classified in *Nectria* subg. *Hyphonectria*. *Memoirs of the New York Botanical Garden* **26**: 1–126.
- Samuels GJ (1976). Perfect states of *Acremonium* the genera *Nectria*, *Actinopsis*, *Iluhya*, *Neohenningsia*, *Ophiodictyon*, and *Peristomialis*. *New Zealand Journal of Botany* **14**: 231–260.
- Samuels GJ (1978). Some species of *Nectria* having *Cylindrocarpon* imperfect states. *New Zealand Journal of Botany* **16**: 73–82.
- Samuels GJ (1988). Fungicolous, Lichenicolous, and Myxomyceticolous Species of *Hypocreopsis*, *Nectriopsis*, *Nectria*, *Peristomialis*, and *Trichonectria*. *New York Botanical Garden* **48**: 1–78.
- Samuels GJ, Brayford D (1994). Species of *Nectria* (*sensu lato*) with red perithecia and striate ascospores. *Sydowia* **46**: 75–161.
- Samuels GJ, Rogerson CT (1984). *Nectria atrofusca* and its anamorph, *Fusarium staphyleae*, a parasite of *Staphylea trifolia* in eastern North America. *Brittonia* **36**: 81–85.
- Samuels GJ, Doi Y, Rogerson CT (1990). *Hypocreales*. In: *Contributions toward a mycobiota of Indonesia: Hypocreales, synnematos Hyphomycetes, Aphyllophorales, Phragmobasidiomycetes, and Myxomycetes* (Samuels GJ, ed). New York Botanical Garden, New York: 6–108.

- Samuels GJ, Hallett IC (1983). *Microdochium stoveri* and *Monographella stoveri* new combinations for *Fusarium stoveri* and *Micronectriella stoveri*. *Transactions of the British Mycological Society* **81**: 473–483.
- Samuels GJ, Lu B, Chaverri P, et al. (2009). *Cyanonectria*, a new genus for *Nectria cyanostoma* and its *Fusarium* anamorph. *Mycological Progress* **8**: 49–58.
- Samuels GJ, Nirenberg H (1989). *Nectria* and *Fusarium*. I. *Nectria setofusariae* and its anamorph *Fusarium setosum*. *Canadian Journal of Botany* **67**: 3372–3377.
- Samuels GJ, Rossman AY, Lowen R, Rogerson CT (1991). A synopsis of *Nectria* subgen. *Dialonectria*. *Mycological Papers* **164**: 1–48.
- Sandoval-Denis M, Crous PW (2018). Removing chaos from confusion: assigning names to common human and animal pathogens in *Neocosmospora*. *Persoonia* **41**: 109–129.
- Sandoval-Denis M, Guarnaccia V, Polizzi G, et al. (2018). Symptomatic *Citrus* trees reveal a new pathogenic lineage in *Fusarium* and two new *Neocosmospora* species. *Persoonia* **40**: 1–25.
- Sandoval-Denis M, Lombard L, Crous PW (2019). Back to the roots: a reappraisal of *Neocosmospora*. *Persoonia* **43**: 90–185.
- Sandoval-Denis M, Swart WJ, Crous PW (2018). New *Fusarium* species from the Kruger National Park, South Africa. *MycKeys* **34**: 63–92.
- Sangalang AE, Summerell BA, Burgess LW, et al. (1995). Taxonomy of *Fusarium*: characterization of *Fusarium avenaceum* subsp. *aywerte* and *Fusarium avenaceum* subsp. *nurragi*. *Mycological Research* **99**: 287–290.
- Santos ACS, Trindade JVC, Lima CS, et al. (2019). Morphology, phylogeny, and sexual stage of *Fusarium caatingaense* and *Fusarium pernambucanum*, new species of the *Fusarium incarnatum-equiseti* species complex associated with insects in Brazil. *Mycologia* **111**: 244–259.
- Sarver BAJ, Ward TJ, Gale LR, et al. (2011). Novel *Fusarium* head blight pathogens from Nepal and Louisiana revealed by multilocus genealogical concordance. *Fungal Genetics and Biology* **48**: 1096–1107.
- Sawai K, Okuno T, Terada Y, et al. (1981). Isolation and properties of two antifungal substances from *Fusarium solani*. *Agricultural and Biological Chemistry* **45**: 1223–1228.
- Scauflaire J, Gourgue M, Munaut F (2011). *Fusarium temperatum* sp. nov. from maize, an emergent species closely related to *Fusarium subglutinans*. *Mycologia* **103**: 586–597.
- Schneider R, Dalchow J (1975). *Fusarium inflexum* spec. nov., als Erreger einer Welkekrankheit an *Vicia faba* L. in Deutschland. *Phytopathologische Zeitschrift* **82**: 70–82.
- Schoch CL, Seifert KA, Huhndorf S, et al. (2012). Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for Fungi. *Proceedings of the National Academy of Sciences of the United States of America* **109**: 6241–6246.
- Schroers H-J, Baayen RP, Meffert JP, et al. (2004). *Fusarium foetens*, a new species pathogenic to begonia elatior hybrids (*Begonia x hiemalis*) and the sister taxon of the *Fusarium oxysporum* species complex. *Mycologia* **96**: 393–406.
- Schroers H-J, Gräfenhan T, Nirenberg HI, et al. (2011). A revision of *Cyanonectria* and *Geejayessis* gen. nov., and related species with fusarium-like anamorphs. *Studies in Mycology* **68**: 115–138.
- Schroers H-J, O'Donnell K, Lamprecht SC, et al. (2009). Taxonomy and phylogeny of the *Fusarium dimerum* species group. *Mycologia* **101**: 44–70.
- Schroers HJ, Samuels GJ, Zhang N, et al. (2016). Epitypification of *Fusisporium* (*Fusarium*) *solani* and its assignment to a common phylogenetic species in the *Fusarium solani* species complex. *Mycologia* **108**: 806–819.
- Secor GA, Rivera-Varas V, Christ DS, et al. (2014). Characterization of *Fusarium secorum*, a new species causing *Fusarium* yellowing decline of sugar beet in north central USA. *Fungal Biology* **118**: 764–775.
- Seifert KA (1985). A monograph of *Stilbella* and some allied Hyphomycetes. *Studies in Mycology* **27**: 1–235.
- Seifert KA (1996). *FusKey: Fusarium interactive key*. Agriculture & Agri-Food Canada. Research Branch, Eastern Cereal & Oilseed Research Centre, Canada.
- Seifert KA (2013). *Pionnotes*, a synonym of *Dacrymyces* rather than *Fusarium*. *Mycotaxon* **123**: 205–211.
- Seifert KA, Gräfenhan T (2012). Lectotypification and characterization of the natural phenotype of *Fusarium bactridioides*. *Mycotaxon* **120**: 415–422.
- Shang QJ, Phookamsak R, Camporesi E, et al. (2018). The holomorph of *Fusarium celtidicola* sp. nov. from *Celtis australis*. *Phytotaxa* **361**: 251–265.
- Sherbakoff CD (1915). *Fusaria* of potatoes. *Memoirs of the Cornell University Agricultural Experimental Station* **6**: 87–270.
- Sibilia C (1925). Due specie di *Fusarium* parasite di piantine di Conifere: *Fusarium fuliginosporum* n. sp. e *F. echinosporum* n. sp. *Annals of the Institute of Forestry (Firenze) II* **1**: 77–97.
- Šišić A, Al-Hatmi AMS, Baćanović-Šišić J, et al. (2018). Two new species of the *Fusarium solani* species complex isolated from compost and hibiscus (*Hibiscus* sp.). *Antonie van Leeuwenhoek* **111**: 1785–1805.
- Šišić A, Baćanović-Šišić J, Al-Hatmi AMS, et al. (2018). The 'forma specialis' issue in *Fusarium*: A case study in *Fusarium solani* f. sp. *psi*. *Scientific Reports* **8**: 1252.
- Skovgaard K, Rosendahl S, O'Donnell K, et al. (2003). *Fusarium commune* is a new species identified by morphological and molecular phylogenetic data. *Mycologia* **95**: 630–636.
- Sleiman S, Halliday CL, Chapman B, et al. (2016). Performance of matrix-assisted laser desorption ionization time of flight mass spectrometry for identification of *Aspergillus*, *Scedosporium*, and *Fusarium* spp. in the Australian clinical setting. *Journal of Clinical Microbiology* **54**: 2182–2186.
- Snyder WC, Hansen HN (1940). The species concept in *Fusarium*. *American Journal of Botany* **27**: 64–67.
- Snyder WC, Hansen HN (1941). The species concept in *Fusarium* with reference to section *Martiella*. *American Journal of Botany* **28**: 738–742.
- Sogonov MV, Castlebury LA, Rossman AY, et al. (2007). The type species of *Apiogmonia*, *A. veneta*, with its *Discula* anamorph is distinct from *A. errabunda*. *Mycological Research* **111**: 693–709.
- Sorauer P (1923). *Handbuch der Pflanzenkrankheiten*. Verlagbuchhandlung Paul Parey, Berlin, Germany.
- Sørensen JL, Akk E, Thrane U, et al. (2013). Production of fusarielins by *Fusarium*. *International Journal of Food Microbiology* **160**: 206–211.
- Stamatakis A (2014). RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* **30**: 1312–1313.
- Starkey DE, Ward TJ, Aoki T, et al. (2007). Global molecular surveillance reveals novel *Fusarium* head blight species and trichothecene toxin diversity. *Fungal Genetics and Biology* **44**: 1191–1204.
- Steenkamp ET, Wingfield BD, Coutinho TA, et al. (2000). PCR-based identification of *MAT-1* and *MAT-2* in the *Gibberella fujikuroi* species complex. *Applied and Environmental Microbiology* **66**: 4378–4382.
- Steyaert RL (1948). Contribution à l'étude des parasites des végétaux du Congo Belge. *Bulletin de la Société Royale de Botanique de Belgique T80, Fasc 1/2*: 11–58.
- Strasser PP (1910). Fünfter Nachtrag zur Pilzflora des Sonntagberges (N.-Ö.), 1909. *Verhandlungen der Zoologisch-Botanischen Gesellschaft Wien* **60**: 303–335.
- Stukenbrock EH, Jørgensen FG, Zala M, et al. (2010). Whole-genome and chromosome evolution associated with host adaptation and speciation of the wheat pathogen *Mycosphaerella graminicola*. *PLoS Genetics* **6**: e1001189.
- Subrahmanyam A (1983). *Fusarium laceratum*. *Mykosen* **26**: 478–480.
- Subramanian CV (1971). *Hyphomycetes. An account of Indian species, except Cercosporae*. Indian Council of Agricultural Research, New Delhi, India.
- Sugimoto Y, Ahmed NE, Yasuda N, et al. (2002). Trichothecene inhibitors of *Striga hemonthica* germination produced by *Fusarium solani*. *Weed Science* **50**: 658–661.
- Summerell BA, Rugg CA, Burgess LW (1995). Characterization of *Fusarium babinda* sp. nov. *Mycological Research* **99**: 1345–1348.
- Summerell BA, Salleh B, Leslie JF (2003). A utilitarian approach to *Fusarium* identification. *Plant Disease* **87**: 117–128.
- Sutton BC (1986). Presidential address: Improvizations on conidial themes. *Transactions of the British Mycological Society* **86**: 1–38.
- Sydow H, Sydow P (1939). *Fungi aequatorienses*. *Annales Mycologici* **37**: 275–438.
- Tatsuno T, Saito M, Enomoto M, et al. (1968). Nivalenol, a toxic principle of *Fusarium nivale*. *Chemical and Pharmaceutical Bulletin* **16**: 2519–2520.
- Taubenhaus JJ (1920). Wilts of the watermelon and related crops (*Fusarium* wilts of cucurbits.). *Bulletin of the Texas Agricultural Experiment Station* **260**: 3–49.
- Taylor JW (2014). Appropriately sized genera and appropriately ranked higher taxa. *IMA Fungus* **5**: 1–2.
- Taylor JW, Jacobson DJ, Kroken S, et al. (2000). Phylogenetic species recognition and species concepts in fungi. *Fungal Genetics and Biology* **31**: 21–32.

- Thines M (2019). An evolutionary framework for host shifts—jumping ships for survival. *New Phytologist* **224**: 605–617.
- Tianpanich K, Prachy S, Wiyakrutta S, et al. (2011). Radical scavenging and antioxidant activities of isocoumarins and a phthalide from the endophyte fungus *Colletotrichum* sp. *Journal of Natural Products* **74**: 79–81.
- Tio ML, Burgess W, Nelson PE, et al. (1977). Techniques for the isolation, culture, and preservation of the Fusaria. *Australasian Plant Pathology Society Newsletter* **6**: 11–13.
- Torbati M, Arzanlou M, Sandoval-Denis M, et al. (2019). Multigene phylogeny reveals new fungicolous species in the *Fusarium tricinctum* species complex and novel hosts in the genus *Fusarium* from Iran. *Mycological Progress* **18**: 119–133.
- Torp M, Nirenberg HI (2004). *Fusarium langsethiae* sp. nov. on cereals in Europe. *International Journal of Food Microbiology* **95**: 247–256.
- Triest D, De Cremer K, Piérard D, et al. (2016). Unique phylogenetic lineage found in the *Fusarium*-like clade after re-examining BCCM/IHEM fungal culture collection material. *Mycobiology* **44**: 121–130.
- Triest D, Stubbe D, de Cremer K, et al. (2015). Use of matrix-assisted laser desorption ionization–time of flight mass spectrometry for identification of molds of the *Fusarium* genus. *Journal of Clinical Microbiology* **53**: 465–476.
- Tulloch M (1972). The genus *Myrothecium* Tode ex Fr. *Mycological Papers* **130**: 1–42.
- Turland NJ, Wiersema JH, Barrie FR, et al. (eds) (2018). *International Code of Nomenclature for algae, fungi, and plants (Shenzhen Code)*. *Regnum Vegetabile* 159. Koeltz Botanical Books, Glashütten, Germany.
- Ueda S, Udagawa S (1983). A new Japanese species of *Neocosmospora* from marine sludges. *Mycotaxon* **16**: 387–395.
- Ueno Y, Ishii K, Sakai K, et al. (1972). Toxicological approaches to the metabolites of fusaria. IV. Microbial survey on "bean hulls poisoning of horses" with the isolation of toxic trichothecenes, neosolaniol and T-2 toxin of *Fusarium solani* M-1-1. *Japanese Journal of Experimental Medicine* **42**: 187–203.
- Urry WH, Wehrmeister HL, Hodge EB, et al. (1966). The structure of zearalenone. *Tetrahedron Letters* **7**: 3109–3114.
- van Hove F, Waalwijk C, Logrieco A, et al. (2011). *Gibberella musae* (*Fusarium musae*) sp. nov., a recently discovered species from banana is sister to *F. verticillioides*. *Mycologia* **103**: 570–585.
- van Wyk PS, Scholtz DJ, Los O (1986). A selective medium for the isolation of *Fusarium* spp. from soil debris. *Phytophylactica* **18**: 67–69.
- Vellinga EC, Kuyper TW, Ammirati J, et al. (2015). Six simple guidelines for introducing new genera of fungi. *IMA fungus* **6**: 65–68.
- Vesonder RF, Tjarks LW, Rohwedder WK, et al. (1979). Equisetin, an antibiotic from *Fusarium equiseti* NRRL 5537, identified as a derivative of n-methyl-2,4-pyrrolidone. *Journal of Antibiotics* **32**: 759–761.
- Vilgalys R (2003). Taxonomic misidentification in public DNA databases. *New Phytologist* **160**: 4–5.
- Vilgalys R, Hester M (1990). Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *Journal of Bacteriology* **172**: 4238–4246.
- Vilgalys R, Sun BL (1994). Ancient and recent patterns of geographic speciation in the oyster mushroom *Pleurotus* revealed by phylogenetic analysis of ribosomal DNA sequences. *Proceedings of the National Academy of Science of the USA* **91**: 4599–4603.
- Vincens F (1915). Deux champignons entomophytes sur Lépidoptères, récoltés au nord du Brésil. *Bulletin de la Société Mycologique de France* **31**: 25–28.
- Visagie CM, Houbraken J, Frisvad JC, et al. (2014). Identification and nomenclature of the genus *Penicillium*. *Studies in Mycology* **78**: 343–371.
- Voglmayr H, Thines M (2007). Phylogenetic relationships and nomenclature of *Bremiella sphaerosperma* (*Chromista*, *Peronosporales*). *Mycotaxon* **100**: 11–20.
- von Bargen KW, Niehaus EM, Bergander K, et al. (2013). Structure elucidation and antimalarial activity of apicidin F: an apicidin-like compound produced by *Fusarium fujikuroi*. *Journal of Natural Products* **76**: 2136–2140.
- von Hönel F (1915). Fragmente zur Mykologie (XVII. Mitteilung, Nr. 876 bis 943). *Sitzungsberichte der mathematisch-naturwissenschaftlichen Klasse der Kaiserlichen Akademie der Wissenschaften, Wien* **124**: 49–159.
- Walker J (1962). Notes on plant parasitic fungi. I. *Proceedings of the Linnean Society of New South Wales* **87**: 162–176.
- Walsh JL, Laurence MH, Liew ECY, et al. (2010). *Fusarium*: two endophytic novel species from tropical grasses of northern Australia. *Fungal Diversity* **44**: 149–159.
- Wang MM, Chen Q, Diao YZ, et al. (2019). *Fusarium incarnatum-equiseti* complex from China. *Persoonia* **43**: 70–89.
- Ward TJ, Bielawski JP, Kistler HC, et al. (2002). Ancestral polymorphism and adaptive evolution in the trichothecene mycotoxin gene cluster of phytopathogenic *Fusarium*. *Proceedings of the National Academy of Sciences of the USA* **99**: 9278–9283.
- Wergen B (2018). *Handbook of Ascomycota. Vol. 1b. Pyrenomyces s.l. mit zweifach septierten bis mauerförmigen Sporen*. Funghiparadise Productions, Germany.
- White TJ, Bruns T, Lee S, et al. (1990). Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: *PCR protocols: a guide to methods and applications* (Innes MA, Gelfand DH, Sninsky JJ, et al., eds). Academic Press, USA: 315–322.
- Wigmann EF, Behr J, Vogel RF, et al. (2019). MALDI-TOF MS fingerprinting for identification and differentiation of species within the *Fusarium fujikuroi* species complex. *Applied Microbiology and Biotechnology* **103**: 5323–5337.
- Willkomm M (1866). *Die mikroskopischen Feinde des Waldes: Naturwissenschaftliche Beiträge zur Kenntniss der Baum- und Holzkrankheiten, für Forstmänner und Botaniker, bearbeitet und in zwanglosen Heften herausgegeben*. G. Schönfeld's Buchhandlung, Dresden, Germany.
- Wingfield MJ, De Beer ZW, Slippers B, et al. (2012). One fungus, one name promotes progressive plant pathology. *Molecular Plant Pathology* **13**: 604–613.
- Wollenweber HW (1914). Identification of species of *Fusarium* occurring on the sweet potato, *Ipomoea batatas*. *Journal of Agricultural Research* **2**: 251–285.
- Wollenweber HW (1916–1935). *Fusaria autographice delineata*. Selbstverlag, Berlin, Germany.
- Wollenweber HW (1924). Pyrenomyceten-Studien. I. *Angewandte Botanik* **6**: 300–313.
- Wollenweber HW (1926). Pyrenomyceten-Studien. II. *Angewandte Botanik* **8**: 168–212.
- Wollenweber HW (1934). *Fusarium bactridioides* sp. nov., associated with *Cronartium*. *Science New York* **79**: 572.
- Wollenweber HW (1943). *Fusarium*-Monographie. II. Fungi parasitici et saprophytici. *Zentralblatt für Bakteriologie Abt. 2* **106**: 104–135.
- Wollenweber HW, Reinking OA (1925). Aliquot *Fusaria tropicalia nova vel revisa*. *Phytopathology* **15**: 155–169.
- Wollenweber HW, Reinking OA (1935). *Die Fusarien*. Verlagsbuchhandlung Paul Parey, Berlin, Germany.
- Woudenberg JHC, Aveskamp MM, de Gruyter J, et al. (2009). Multiple *Didymella* teleomorphs are linked to the *Phoma clematidina* morphotype. *Persoonia* **22**: 56–62.
- Xia JW, Sandoval-Denis M, Crous PW, et al. (2019). Numbers to names – reappraisal of the *Fusarium incarnatum-equiseti* species complex. *Persoonia* **43**: 186–221.
- Xu Y, Vinas M, Alsarraq A, et al. (2019). Bis-naphthopyrone pigments protect filamentous ascomycetes from a wide range of predators. *Nature Communications* **10**: 3579.
- Yamamoto W, Maeda M, Oyasu N (1957). Some *Nectriaceae* and *Elsinoe* species from Japan. *Science Reports of the Hyogo University of Agriculture. Series agricultural biology* **3**: 15–18.
- Yilmaz N, Sandoval-Denis M, Lombard L, et al. (2021). Redefining species limits in the *Fusarium fujikuroi* species complex. *Persoonia* **46**: 129–162.
- Yli-Mattila T, Ward T, O'Donnell K, et al. (2011). *Fusarium sibiricum* sp. nov, a novel type A trichothecene-producing *Fusarium* from northern Asia closely related to *F. sporotrichioides* and *F. langsethiae*. *International Journal of Food Microbiology* **147**: 58–68.
- Yli-Mattila Y, Gagkaeva T, Ward TJ, et al. (2009). A novel Asian clade within the *Fusarium graminearum* species complex includes a newly discovered cereal head blight pathogen from the Far East of Russia. *Mycologia* **101**: 841–852.
- Yoshida K, Saitoh H, Fujisawa S, et al. (2009). Association genetics reveals three novel avirulence genes from the rice blast fungal pathogen *Magnaporthe oryzae*. *The Plant Cell* **21**: 1573–1591.
- Yoshizawa T, Morooka N (1973). Deoxynivalenol and its monoacetate: new mycotoxins from *Fusarium roseum* and moldy barley. *Agricultural and Biological Chemistry* **37**: 2933–2934.
- Zeller KA, Summerell BA, Bullock S, et al. (2003). *Gibberella konza* (*Fusarium konzum*) sp. nov. from prairie grasses, a new species in the *Gibberella fujikuroi* species complex. *Mycologia* **95**: 943–954.

- Zeng ZQ, Zhuang WY (2017). Eight new combinations of *Bionectriaceae* and *Nectriaceae*. *Mycosystema* **36**: 278–281.
- Zeng ZQ, Zhuang WY (2017). Two new species of *Neocosmospora* from China. *Phytotaxa* **319**: 175–183.
- Zeng ZQ, Zheng HD, Wang XC, et al. (2020). Ascomycetes from the Qilian Mountains, China – *Hypocreales*. *MycKeys* **71**: 119–137.
- Zhang FA, Liu S-R, Zhou X-J, et al. (2020). *Fusarium xiangyunensis* (*Nectriaceae*), a remarkable new species of the Nematophagus fungi from Yunnan, China. *Phytotaxa* **450**: 273–284.
- Zhang XM, Zhuang WY (2003). Re-examinations of *Bionectriaceae* and *Nectriaceae* (*Hypocreales*) from temperate China on deposit in HMAS. *Nova Hedwigia* **76**: 191–200.
- Zhao ZH, Lu GZ (2008). *Fusarium sinensis* sp. nov., a new species from wheat in China. *Mycologia* **100**: 746–751.
- Zhou G, Qiao L, Zhang X, et al. (2019). Fusaricates H-K and fusalanones A-B from a mangrove endophytic fungus *Fusarium solani* HDN15-410. *Phytochemistry* **158**: 13–19.
- Zhou X, O'Donnell K, Aoki T, et al. (2016). Two novel *Fusarium* species that cause canker disease of prickly ash (*Zanthoxylum bungeanum*) in northern China form a novel clade with *Fusarium torreyae*. *Mycologia* **108**: 668–681.
- Zhuang WY, Nong Y, Luo J (2007). New species and new Chinese records of *Bionectriaceae* and *Nectriaceae* (*Hypocreales*, *Ascomycetes*) from Hubei, China. *Fungal Diversity* **24**: 347–357.
- Zoutman DE, Sigler L (1991). Mycetoma on the foot caused by *Cylindrocarpon destructans*. *Journal of Clinical Microbiology* **29**: 1839–1855.
- Zwickl DJ, Hillis DM (2002). Increased taxon sampling greatly reduces phylogenetic error. *Systematic Biology* **51**: 588–598.