



# Fungal diversity notes 491–602: taxonomic and phylogenetic contributions to fungal taxa

Saowaluck Tibpromma<sup>1,2,3,4,5</sup> · Kevin D. Hyde<sup>1,2,3,4,5</sup> · Rajesh Jeewon<sup>29</sup> ·  
Sajeewa S. N. Maharachchikumbura<sup>25</sup> · Jian-Kui Liu<sup>18</sup> · D. Jayarama Bhat<sup>9,10</sup> ·  
E. B. Gareth Jones<sup>11</sup> · Eric H. C. McKenzie<sup>12</sup> · Erio Camporesi<sup>6,7,8</sup> ·  
Timur S. Bulgakov<sup>27</sup> · Mingkwan Doilom<sup>2</sup> · André Luiz Cabral Monteiro de Azevedo Santiago<sup>15</sup> ·  
Kanad Das<sup>34</sup> · Patinjareveettil Manimohan<sup>33</sup> · Tatiana B. Gibertoni<sup>42</sup> ·  
Young Woon Lim<sup>30</sup> · Anusha Hasini Ekanayaka<sup>2</sup> · Benjarong Thongbai<sup>2</sup> ·  
Hyang Burm Lee<sup>17</sup> · Jun-Bo Yang<sup>55</sup> · Paul M. Kirk<sup>60</sup> · Phongeun Sysouphanthong<sup>53</sup> ·  
Sanjay K. Singh<sup>22</sup> · Saranyaphat Boonmee<sup>2</sup> · Wei Dong<sup>20</sup> · K. N. Anil Raj<sup>33</sup> ·  
K. P. Deepna Latha<sup>33</sup> · Rungtiwa Phookamsak<sup>1,2,4</sup> · Chayanard Phukhamsakda<sup>1,2,3,4</sup> ·  
Sirinapa Konta<sup>2,3,5</sup> · Subashini C. Jayasiri<sup>2,3,5</sup> · Chada Norphanphoun<sup>2,3,5</sup> ·  
Danushka S. Tennakoon<sup>2,3,5</sup> · Junfu Li<sup>2,3,5</sup> · Monika C. Dayarathne<sup>2,3,5</sup> ·  
Rekhani H. Perera<sup>2,3,5</sup> · Yuanpin Xiao<sup>2,3,5</sup> · Dhanushka N. Wanasinghe<sup>1,2,3,4,5</sup> ·  
Indunil C. Senanayake<sup>1,2,3,4,5</sup> · Ishani D. Goonasekara<sup>1,2,3,4,5</sup> · N. I. de Silva<sup>1,2,4,13</sup> ·  
Ausana Mapook<sup>2,3</sup> · Ruvishika S. Jayawardena<sup>2,16</sup> · Asha J. Dissanayake<sup>2,16</sup> ·  
Ishara S. Manawasinghe<sup>2,16</sup> · K. W. Thilini Chethana<sup>2,16</sup> · Zong-Long Luo<sup>2,19</sup> ·  
Kalani Kanchana Hapuarachchi<sup>2,3,28</sup> · Abhishek Baghela<sup>22</sup> · Adriene Mayra Soares<sup>42</sup> ·  
Alfredo Vizzini<sup>23,40</sup> · Angelina Meiras-Ottoni<sup>42</sup> · Armin Mešić<sup>46</sup> · Arun Kumar Dutta<sup>31</sup> ·  
Carlos Alberto Fragoso de Souza<sup>15</sup> · Christian Richter<sup>58</sup> · Chuan-Gen Lin<sup>2,3,5,59</sup> ·  
Debasis Chakrabarty<sup>48</sup> · Dinushani A. Daranagama<sup>2,3,5</sup> · Diogo Xavier Lima<sup>15</sup> ·

✉ Samantha C. Karunarathna  
samanthakarunarathna@gmail.com

<sup>1</sup> Key Laboratory for Plant Diversity and Biogeography of East Asia, Kunming Institute of Botany, Chinese Academy of Science, Kunming 650201, Yunnan, People's Republic of China

<sup>2</sup> Center of Excellence in Fungal Research, Mae Fah Luang University, Chiang Rai 57100, Thailand

<sup>3</sup> School of Science, Mae Fah Luang University, Chiang Rai 57100, Thailand

<sup>4</sup> World Agroforestry Centre, East and Central Asia, Kunming 650201, Yunnan, People's Republic of China

<sup>5</sup> Mushroom Research Foundation, 128 M.3 Ban Pa Deng T. Pa Pae, A. Mae Taeng, Chiang Mai 50150, Thailand

<sup>6</sup> A.M.B. Gruppo Micologico Forlivese "Antonio Cicognani", Via Roma 18, Forlì, Italy

<sup>7</sup> A.M.B. Circolo Micologico "Giovanni Carini", C.P. 314, Brescia, Italy

<sup>8</sup> Società per gli Studi Naturalistici della Romagna, C.P. 144, Bagnacavallo, RA, Italy

<sup>9</sup> Formerly Department of Botany, Goa University, Taleigão, Goa, India

<sup>10</sup> No. 128/1-J, Azad Housing Society, Curca, Goa Velha, India

<sup>11</sup> 33 B St. Edwards Road Southsea Hants, Hampshire PO5 3DH, UK

<sup>12</sup> Landcare Research Manaaki Whenua, Private Bag 92170, Auckland, New Zealand

<sup>13</sup> Department of Biology, Faculty of Science, Chiang Mai University, Chiang Mai 50200, Thailand

<sup>14</sup> Division of Applied Science, College of International Education, The Hong Kong Baptist University, Hong Kong SAR, China

<sup>15</sup> PostGraduate Program in Biology of Fungi, Department of Mycology, Federal University of Pernambuco, Av. Nelson Chaves, s/n, Recife, PE 50670-420, Brazil

<sup>16</sup> Institute of Plant and Environment Protection, Beijing Academy of Agriculture and Forestry Sciences, Beijing 100097, People's Republic of China

<sup>17</sup> Environmental Microbiology Lab, Division of Food Technology, Biotechnology & Agrochemistry, College of Agriculture and Life Sciences, Chonnam National University, Yongbong-Dong 300, Buk-Gu, Gwangju 61186, Korea

<sup>18</sup> Guizhou Key Laboratory of Agricultural Biotechnology, Guizhou Academy of Agricultural Sciences, Guiyang 550006, Guizhou, People's Republic of China

<sup>19</sup> College of Agriculture and Biological Sciences, Dali University, Dali 671003, Yunnan, People's Republic of China

Dyutiparna Chakraborty<sup>34</sup> · Enrico Ercole<sup>40</sup> · Fang Wu<sup>26</sup> · Giampaolo Simonini<sup>38</sup> ·  
 Gianrico Vasquez<sup>37</sup> · Gladstone Alves da Silva<sup>15</sup> · Helio Longoni Plautz Jr.<sup>44</sup> ·  
 Hiran A. Ariyawansa<sup>61</sup> · Hyun Lee<sup>30</sup> · Ivana Kušan<sup>46</sup> · Jie Song<sup>26</sup> ·  
 Jingzu Sun<sup>2,3,21</sup> · Joydeep Karmakar<sup>36</sup> · Kaifeng Hu<sup>54</sup> · Kamal C. Semwal<sup>47</sup> ·  
 Kasun M. Thambugala<sup>2,3,5,18</sup> · Kerstin Voigt<sup>59</sup> · Krishnendu Acharya<sup>31</sup> ·  
 Kunhiraman C. Rajeshkumar<sup>22</sup> · Leif Ryvarden<sup>45</sup> · Margita Jadan<sup>46</sup> ·  
 Md. Iqbal Hosen<sup>51</sup> · Michal Mikšík<sup>41</sup> · Milan C. Samarakoon<sup>2,4,5,13</sup> ·  
 Nalin N. Wijayawardene<sup>2</sup> · Nam Kyu Kim<sup>30</sup> · Neven Matočec<sup>46</sup> · Paras Nath Singh<sup>22</sup> ·  
 Qing Tian<sup>2,3,5</sup> · R. P. Bhatt<sup>49</sup> · Rafael José Vilela de Oliveira<sup>15</sup> · Rodham E. Tulloss<sup>50</sup> ·  
 S. Aamir<sup>22</sup> · Saithong Kaewchai<sup>24</sup> · Sayali D. Marathe<sup>22</sup> · Sehroon Khan<sup>1,4</sup> ·  
 Sinang Hongsanan<sup>2</sup> · Sinchan Adhikari<sup>35</sup> · Tahir Mahmood<sup>49</sup> · Tapas Kumar Bandyopadhyay<sup>36</sup> ·  
 Tatyana Yu. Svetasheva<sup>39</sup> · Thi Thuong Thuong Nguyen<sup>17</sup> · Vladimír Antonín<sup>32</sup> ·  
 Wen-Jing Li<sup>1,2,3,5</sup> · Yong Wang<sup>59</sup> · Yuvraj Indoliya<sup>48</sup> · Zdenko Tkalc̆ec<sup>46</sup> ·  
 Abdallah M. Elgorban<sup>57</sup> · Ali H. Bahkali<sup>57</sup> · Alvin M. C. Tang<sup>14</sup> · Hong-Yan Su<sup>19</sup> ·  
 Huang Zhang<sup>20</sup> · Itthayakorn Promputtha<sup>13</sup> · Jennifer Luangsa-ard<sup>56</sup> ·  
 Jianchu Xu<sup>1,4</sup> · Jiye Yan<sup>16</sup> · Kang Ji-Chuan<sup>28</sup> · Marc Stadler<sup>58</sup> · Peter E. Mortimer<sup>1,4</sup> ·  
 Putarak Chomnunti<sup>2,3</sup> · Qi Zhao<sup>52</sup> · Alan J. L. Phillips<sup>43</sup> · Sureeporn Nontachaiyapoom<sup>3</sup> ·  
 Ting-Chi Wen<sup>28</sup> · Samantha C. Karunarathna<sup>1,4</sup>

Received: 6 March 2017 / Accepted: 20 March 2017

© School of Science 2017

**Abstract** This is a continuity of a series of taxonomic and phylogenetic papers on the fungi where materials were collected from many countries, examined and described. In addition to extensive morphological descriptions and appropriate asexual and sexual connections, DNA

sequence data are also analysed from concatenated datasets to infer phylogenetic relationships and substantiate systematic positions of taxa within appropriate ranks. Whenever new species or combinations are proposed, we apply an integrative approach using morphological and molecular

<sup>20</sup> Yunnan Institute of Food Safety, Kunming University of Science & Technology, Kunming 650500, Yunnan, People's Republic of China

<sup>21</sup> State Key Laboratory of Mycology, Institute of Microbiology, Chinese Academy of Sciences (IMCAS), No. 3 Park 1, West Beichen Road, Chaoyang District, Beijing 100101, People's Republic of China

<sup>22</sup> National Fungal Culture Collection of India, Biodiversity and Palaeobiology Group, MACS Agharkar Research Institute, G.G. Agarkar Road, Pune 411 004, India

<sup>23</sup> Institute for Sustainable Plant Protection (IPSP)-CNR, Viale P.A. Mattioli 25, 10125 Turin, Italy

<sup>24</sup> Faculty of Agriculture, Princess of Naradhiwas University, No. 99 Khok Khian, Amphur Muang, Narathiwat Province, Thailand

<sup>25</sup> Department of Crop Sciences, College of Agricultural and Marine Sciences, Sultan Qaboos University, P.O. Box 34, 123 Al-Khod, Oman

<sup>26</sup> Institute of Microbiology, Beijing Forestry University, PO Box 61, Beijing 100083, China

<sup>27</sup> Russian Research Institute of Floriculture and Subtropical Crops, 2/28 Yana Fabritsiusa Street, Sochi, Krasnodar region, Russia 354002

<sup>28</sup> Engineering Research Center of Southwest Bio-Pharmaceutical Resources, Ministry of Education, Guizhou University, Guiyang 550025, Guizhou, People's Republic of China

<sup>29</sup> Department of Health Sciences, Faculty of Science, University of Mauritius, Reduit 80837, Mauritius

<sup>30</sup> School of Biological Sciences, Seoul National University, Seoul 08826, Korea

<sup>31</sup> Molecular and Applied Mycology and Plant Pathology Laboratory, Department of Botany, University of Calcutta, Kolkata, West Bengal 700019, India

<sup>32</sup> Moravian Museum, Department of Botany, Zelný trh 6, 659 37 Brno, Czech Republic

<sup>33</sup> Department of Botany, University of Calicut, Malappuram, Kerala 673 635, India

<sup>34</sup> Botanical Survey of India, Cryptogamic Unit, P.O. Botanic Garden, Howrah 711103, India

<sup>35</sup> Department of Botany, University of Kalyani, Kalyani, Nadia, West Bengal 741235, India

<sup>36</sup> Department of Molecular Biology & Biotechnology, University of Kalyani, Kalyani, Nadia, West Bengal 741235, India

<sup>37</sup> Laboratorio di Isto-Cito-Patologia s.n.c., Via del Bosco, 96, 95125 Catania, Italy

<sup>38</sup> Via Bell'Aria, 8, 42121 Reggio nell'Emilia (RE), Italy

<sup>39</sup> Biology and Technologies of Living Systems Department, Tula State Lev Tolstoy Pedagogical University, 125, Lenin av., Tula, Russia 300026

data as well as ecological features wherever applicable. Notes on 112 fungal taxa are compiled in this paper including *Biatriosporaceae* and *Roussellaceae*, *Didysimulans* gen. nov., 81 new species, 18 new host records and new country records, five reference specimens, two new combinations, and three sexual and asexual morph reports. The new species are *Amanita cornelii*, *A. emodotrygon*, *Angustumassarina alni*, *A. arezzoensis*, *A. italica*, *A. lonicerae*, *A. premilcurensis*, *Ascochyta italicica*, *A. rosae*, *Astroboletus appendiculatus*, *Barriopsis thailandica*, *Berkleasmium ariense*, *Calopoma petasitis*, *Camarosporium laburnicola*, *C. moricola*, *C. grisea*, *C. ossea*, *C. paraincrusted*, *Colletotrichum sambucicola*, *Coprinopsis cerkezii*, *Cytospora gelida*, *Dacrymyces chiangraiensis*, *Didysimulans italicica*, *D. mezzanensis*, *Entodesmium italicica*, *Entoloma magnum*, *Evlachovaea indica*, *Exophiala italicica*, *Favolus gracilisporus*, *Femsjonia monospora*, *Fomitopsis flabellata*, *F. roseoalba*, *Gongronella brasiliensis*, *Helvella crispoides*, *Hermatomyces chiangmaiensis*, *H. chromolaeiae*, *Hysterium centramurum*, *Inflatipsora caryotae*, *Inocybe brunneosquamulosa*, *I. luteobrunnea*, *I. rubrobrunnea*, *Keissleriella cirsii*, *Lepiota cylindrocystidia*, *L. flavocarpa*, *L. maerimensis*, *Lophiotrema guttulata*, *Marasmius luculentus*, *Morenoina calamicola*, *Moelleriella thanathonensis*, *Mucor stercorarius*, *Myrmecridium fluviae*, *Myrothecium*

*septentrionale*, *Neosetophoma garethjonesii*, *Nigrograna cangshanensis*, *Nodulosphaeria guttulatum*, *N. multisep-tata*, *N. sambuci*, *Panus subfasciatus*, *Paraleptosphaeria padi*, *Paraphaeosphaeria viciae*, *Parathyridaria robiniae*, *Penicillium punicae*, *Phaeosphaeria calamicola*, *Phaeosphaeriopsis yuccae*, *Pleurophoma italicica*, *Polyporus brevibasidiosus*, *P. koreanus*, *P. orientivarius*, *P. parvo-varius*, *P. subdictyopus*, *P. ulleungus*, *Pseudoasteromas-saria spadicea*, *Rosellinia mearnsii*, *Rubroboletus demonensis*, *Russula yanheensis*, *Sigarispora muriformis*, *Sillia italicica*, *Stagonosporopsis ailanthicola*, *Strobilomyces longistipitatus*, *Subplenodomus galicola* and *Wolfiporia pseudococos*. The new combinations are *Melanomma popu-lina* and *Rubroboletus eastwoodiae*. The reference specimens are *Cookeina tricholoma*, *Gnomoniopsis sanguisorbae*, *Helvella costifera*, *Polythrincium trifolii* and *Russula virescens*. The new host records and country records are *Ascochyta medicaginicola*, *Boletellus emoden-sis*, *Cryptotrama asprata*, *Cytospora ceratosperma*, *Favo-laschia auriscalpium*, *F. manipularis*, *Hysterobrevium mori*, *Lentinus sajor-caju*, *L. squarrosulus*, *L. velutinus*, *Leucocoprinus cretaceus*, *Lophiotrema vagabundum*, *Nothopoma quercina*, *Platystomum rosae*, *Pseudodidy-mosphaeria phlei*, *Tremella fuciformis*, *Truncatella spartii* and *Vaginatispora appendiculata* and three sexual and

<sup>40</sup> Department of Life Sciences and Systems Biology, University of Turin, Viale P.A. Mattioli, 25, 10125 Turin (TO), Italy

<sup>41</sup> Lečkova 1521/15, 14900 Prague 4, Czech Republic

<sup>42</sup> Universidade Federal de Pernambuco (UFPE), Centro de Biociências (CB), Departamento de Micologia, Avenida da Engenharia, S/N - Cidade Universitária, Recife, PE 50740-600, Brazil

<sup>43</sup> University of Lisbon, Faculty of Sciences, Biosystems and Integrative Sciences Institute (BioISI), Campo Grande, 1749-016 Lisbon, Portugal

<sup>44</sup> Instituto de Ciências Biológicas, Laboratório de Microbiologia, Av. Augusto Corrêa, Belém, Pará 66075-110, Brazil

<sup>45</sup> Department of Botany, University of Oslo, Blindern, P. O. Box 1045, 0316 Oslo, Norway

<sup>46</sup> Ruđer Bošković Institute, Bijenička cesta 54, 10000 Zagreb, Croatia

<sup>47</sup> Department of Biology, College of Sciences, Eritrea Institute of Technology, P. Box 12676, Mai Nafhi, Asmara, Eritrea

<sup>48</sup> Genetics and Molecular Biology Division, CSIR-National Botanical Research Institute, Lucknow 226001, Uttar Pradesh, India

<sup>49</sup> Department of Botany & Microbiology, H.N.B. Garhwal University, Srinagar, Garhwal 246174, Uttarakhand, India

<sup>50</sup> P. O. Box 57, Roosevelt, NJ 08555-0057, USA

<sup>51</sup> State Key Laboratory of Applied Microbiology Southern China, Guangdong Provincial Key Laboratory of Microbial Culture Collection and Application, Guangdong Institute of Microbiology, Guangzhou 510070, People's Republic of China

<sup>52</sup> Key Lab of Southwestern Crop Gene Resources and Germplasm Innovation Ministry of Agriculture, Yunnan Provincial Key Laboratory of Agricultural Biotechnology, Institute of Biotechnology and Germplasm Resources, Yunnan Academy of Agricultural Sciences, Kunming 650223, Yunnan, People's Republic of China

<sup>53</sup> Ecology Division, Biotechnology and Ecology Institute, Ministry of Science and Technology, P.O.Box: 2279, Vientiane Capital, Lao People's Democratic Republic

<sup>54</sup> State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming, People's Republic of China

<sup>55</sup> Germplasm Bank of Wild Species in Southwest China, Kunming Institute of Botany, Chinese Academy of Science, Kunming 650201, Yunnan, People's Republic of China

<sup>56</sup> Microbe Interaction Laboratory (BMIT), BIOTEC, National Science and Technology Development Agency (NSTDA), 113 Thailand Science Park, Thanon Phahonyothin, Tombon Khlong Nueng, Amphoe, Khlong Luang, Pathum Thani 12120, Thailand



**Fig. 126** *Entoloma magnum* (CAL 1385, holotype). Scale bar 20 mm (photo by K. N. Anil Raj)

*Entoloma myochroum* Noordel. & E. Ludw., a species from Germany (Noordeloos 2004), is close to *E. magnum* in having almost similar-sized basidiocarps, a concave and glabrous pileus, adnate lamellae with concolorous edges, a solid stipe with a fibrillose surface, almost similar-sized basidiospores ( $7-10 \times 6.5-8.5 \mu\text{m}$ ) with 4–6 angles, a fertile lamella-edge and clamped hyphae. However, that species has a mouse-grey-coloured pileus, a flexuous or irregularly shaped stipe, a cutis-type pileipellis made up of hyphae with intracellular pigment and lamellar and pileus trama with short inflated elements.

In a BLASTn search using the ITS sequence (628 bp) derived from *E. magnum*, the closest hit was *E. ochreoprunuloides* (GenBank Number KC710092; 92% identity). *Entoloma ochreoprunuloides* Morgado & Noordel., is a species from Germany (Noordeloos 2004; Morgado et al. 2013), resembles *E. magnum* in having a similar-shaped, brownish pileus with radially wrinkled and innately fibrillose surface, isodiametric basidiospores, a fertile-lamella edge, and clamped hyphae. However, *E. ochreoprunuloides* differs from *E. magnum* owing to its smaller-sized and differently-coloured basidiocarps, emarginate lamellae, smaller ( $5.9-7.1 \times 5.7-7.2 \mu\text{m}$ ) basidiospores and hyphae of the pileipellis with intracellular pigment.

In the phylogram (Fig. 126) generated from the ML analysis, *E. magnum* nested within a clade, which represents the subgenus *Entoloma*. Within this clade, *E. magnum*, *E. luteobasis*, *E. ochreoprunuloides*, *E. madidum* and collections of *E. bloxamii* formed a subclade. Within this subclade, *E. magnum* was differentiated as an independent lineage separated from other species with significant (74%) bootstrap support.

### *Inocybaceae* Jülich

The type genus of the Inocybaceae, *Inocybe*, had traditionally been placed within the *Cortinariaceae* family (Kirk et al. 2008, Singer 1986). Despite this, Jülich placed the genus in its own family, the Inocybaceae (Jülich 1982). Members of this family have a widespread distribution in tropical and temperate areas (Cannon & Kirk 2007).

### *Inocybe* (Fr.) Fr.

The genus *Inocybe* (*Inocybaceae*) is a species-rich genus of *Agaricales* and is well-known for their ectomycorrhizal ecology and toxicity of most species (Matheny 2009). Matheny (2009) proposed seven major clades or lineages consisting of *Inocybe* and its allies within the family *Inocybaceae*. Of the seven clades or lineages (*Inocybe* s. str., *Nothocybe*, *Pseudosperma*, *Mallocybe*, *Inosperma*, *Auritella* and *Mallocybella*), *Auritella* and *Mallocybella* were formally recognised as distinct genera, *Auritella* and *Tubariomyces* (Matheny and Bouger 2006a, b; Alvarado et al. 2010). Species of *Inocybe* are characterised by mostly brownish or rarely whitish basidiomata occasionally with a purplish or lilac hue, a fibrillose-rimose or squamulose pileus, brownish lamellae, brown spore-print, a fibrillose-pruinose stipe at times with a distinct marginate-bulbous base, a characteristic odour, smooth, warty, nodulose or spinulose basidiospores and metuloidal cystidia often with crystallloid deposits at their apices. Several species are devoid of metuloidal cystidia and they are characterised by abundant, thin-walled cheilocystidia (Matheny 2005; Larsson et al. 2009). During the course of our studies on this genus in Kerala State, India, we discovered several new species of *Inocybe*. Three of these species belonging to the *Pseudosperma* clade (Fig. 128) are described here.

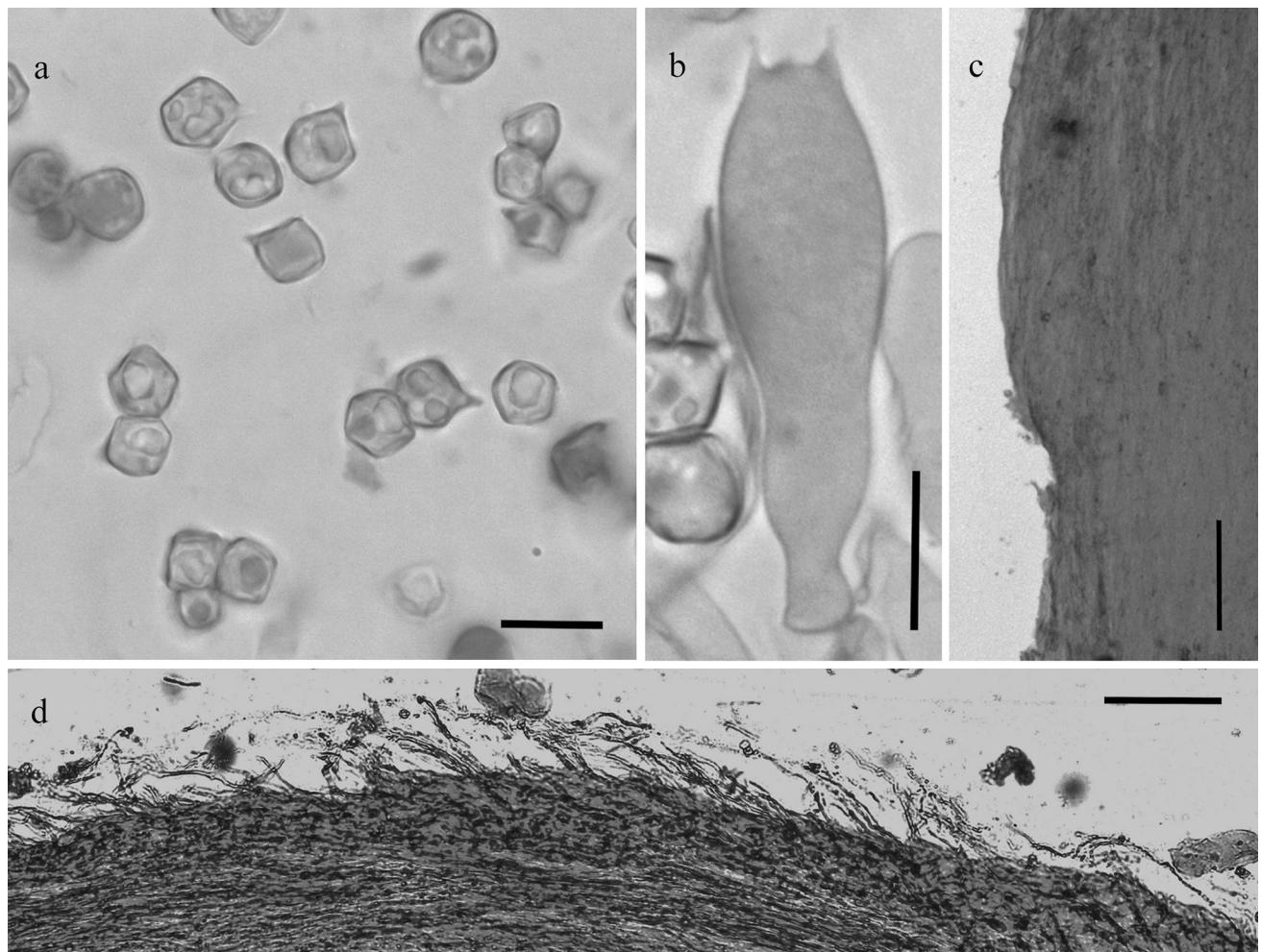
### *Inocybe brunneosquamulosa* K.P.D. Latha & Manim., sp. nov.

*MycoBank number:* MB 816735; *Facesoffungi number:* FoF: 2176, Figs. 129, 130

*Etymology:* referring to the brown squamules on the pileus surface.

*Holotype:* CAL 1308

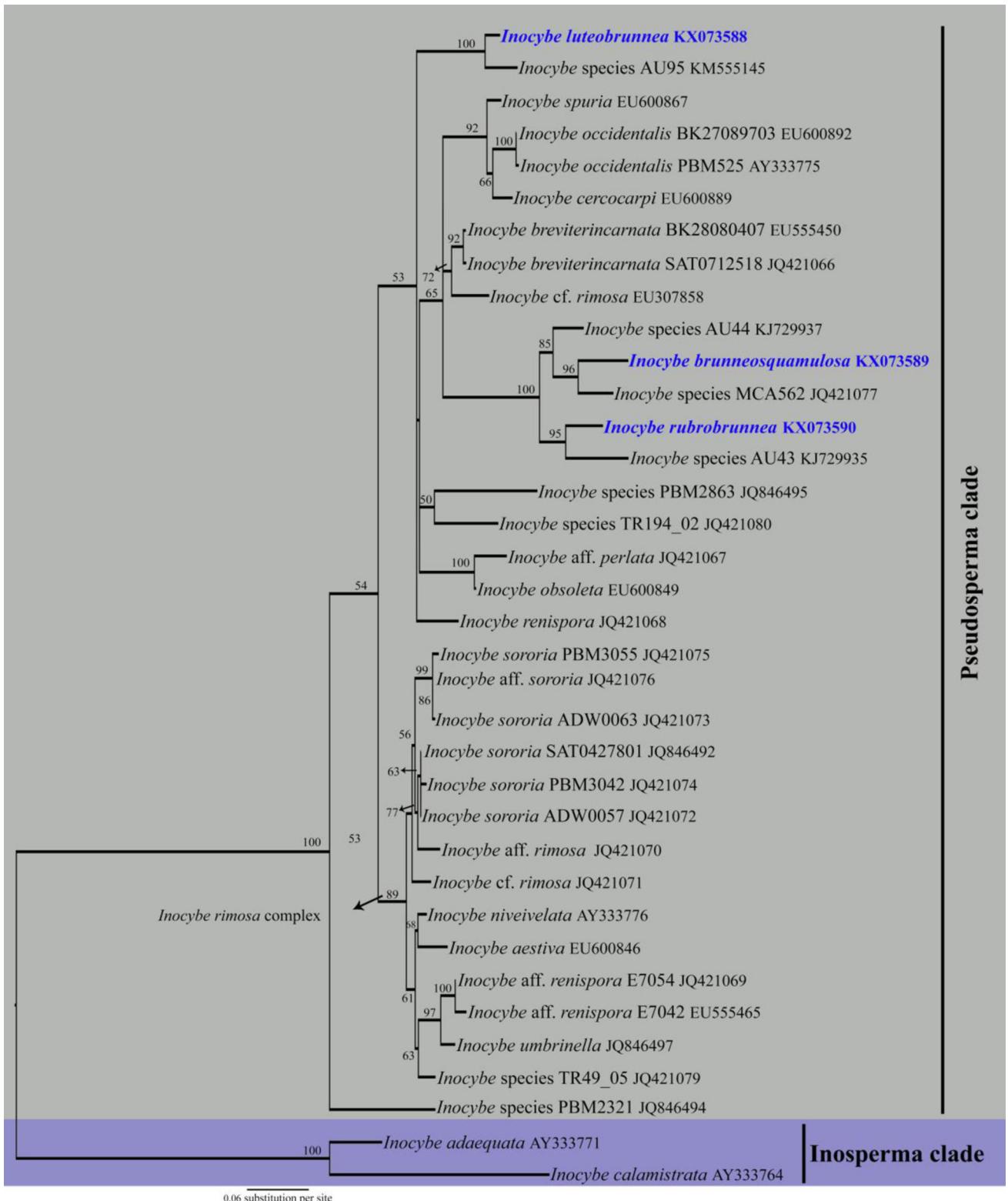
*Basidiocarps* small. *Pileus* 7–11 mm diam., convex with a small umbo when young, becoming broadly convex still with a small umbo at maturity; surface initially dark brown (6F8/OAC636) on the squamules and brownish-orange (6C4/OAC653) elsewhere, becoming dark brown (6F7/OAC637) on the squamules, greyish-orange (6B3/OAC633) on the fibrils and brownish-grey (6C2/OAC662) elsewhere at maturity, appressed- to recurved-squamulose all over when young, becoming appressed- to recurved squamulose on and around the umbo and appressed-fibrillose and rimose towards the margin; margin incurved, becoming decurved, crenate or wavy. *Lamellae* sinuate,



**Fig. 127** *Entoloma magnum* (CAL 1385, holotype). **a** Basidiospores. **b** Basidium. **c** Stipitipellis. **d** Pileipellis. Scale bars a, b = 10 µm; c, d = 100 µm (photos by K. N. Anil Raj)

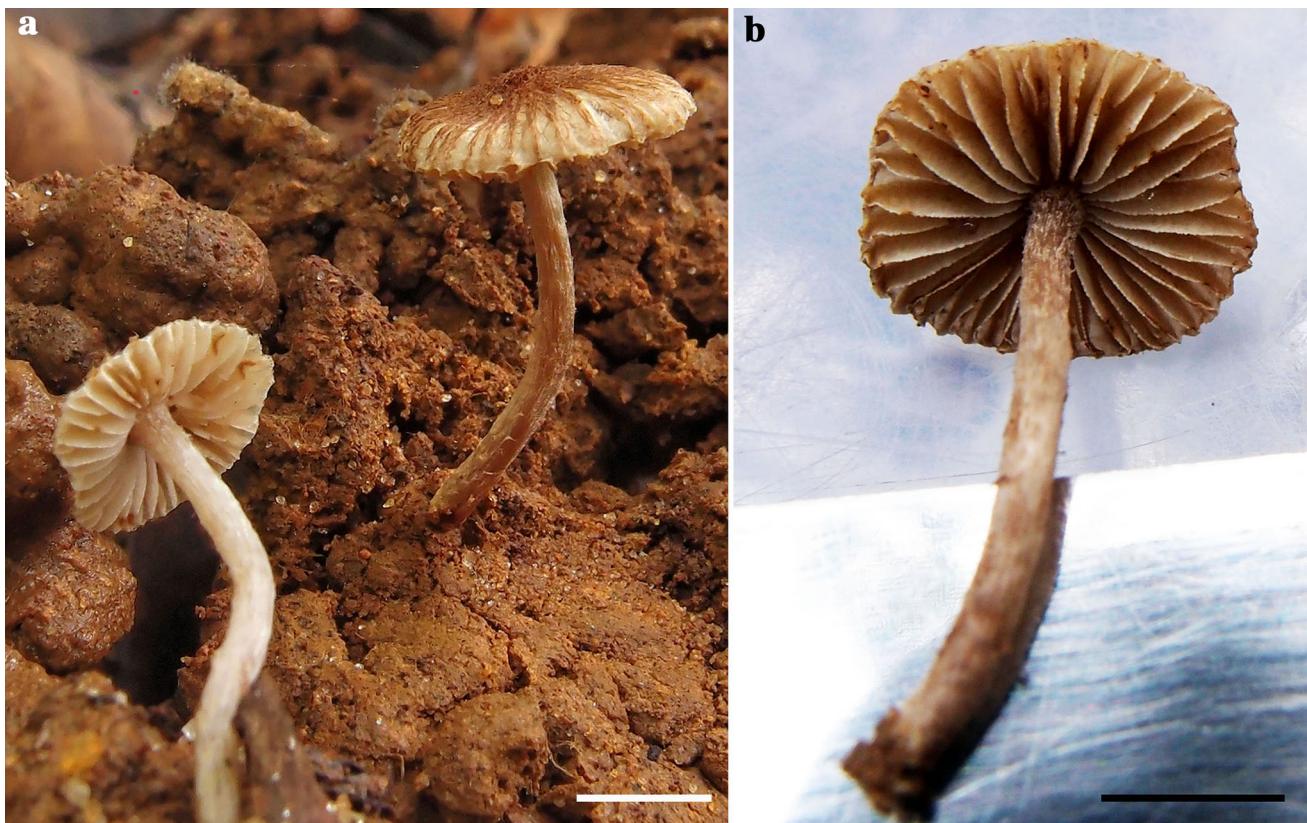
close, initially orange grey (6B2/OAC634), becoming brownish-orange (6C4/OAC655) or light brown (6D4/OAC686) at maturity, up to 1.5 mm wide, with lamellulae of 1 length; edges fimbriate, whitish. *Stipe* 17–19 × 1–2 mm, central, terete, equal, cartilaginous, solid; surface brownish-orange (6C3/OAC633) all over, appressed-fibrillose in most parts, slightly-recurved fibrillose and finely pruinose towards the apex; base not enlarged. *Odour* and *taste* not distinctive. *Basidiospores* 8–10 × 5–6.5 ( $9 \pm 0.6 \times 5.9 \pm 0.4$ ) µm, Q = 1.3–1.9, Qm = 1.5, smooth, ellipsoid to subphaseoliform, slightly thick-walled, yellowish-brown. *Basidia* 21–30 × 11–13 µm, clavate, thin-walled, hyaline, 4-spored; sterigmata up to 4.5 µm long. *Pleurocystidia* absent. *Lamella-edge* heterogeneous. *Cheilocystidia* 17–39 × 11–18 µm, versiform: clavate, utriform, fusiform, cylindrical with an obtuse apex, occasionally subglobose or rarely pedicellate or septate, hyaline with faint hyaline encrustations, thin- to slightly thick-walled. *Lamellar trama* subregular; hyphae 6–15 µm wide, thin-

walled, hyaline or pale yellow. *Subhymenium* pseudo-parenchymatous. *Pileus trama* subregular, composed of both narrow and inflated hyphae; hyphae 3–30 µm wide, pale yellow, thin-walled. *Pileipellis* a cutis frequently disrupted with trichodermal patches, often a perfect trichoderm at the centre; hyphae 8–16 µm wide, thin- to slightly thick-walled, with a brown wall pigment and dense, yellowish-brown or brown spiral encrustations; terminal cells 25–52 × 7–11 µm, clavate or cylindrical with an obtuse apex, thin- to slightly thick-walled. *Stipitipellis* a cutis often disrupted by loose hyphal projections scattered over the entire surface of the stipe and with bunches of caulocystidia confined to the extreme stipe apex; hyphae 5–11 µm wide, thin- to slightly thick-walled, with a pale yellowish-brown wall pigment and faint hyaline encrustations; terminal cells, 21–49 × 5–7 µm, cylindrical or flexuous-cylindric, slightly thick-walled, with a pale yellowish-brown wall pigment and faint hyaline encrustations. *Caulocystidia* 22–50 × 11–14 µm, catenulate,



**Fig. 128** Phylogram generated from maximum likelihood (RAxML) analysis based on RPB2 sequence data matrix for 34 *Inocybe* species. Sequences of *Inocybe* species belonging to the *Pseudosperma* clade used in this study have been selected from a previous analysis of Kropp et al. (2013). Values at nodes indicate bootstrap support. BS

values  $\geq 50\%$  are shown. *Inocybe luteobrunnea*, *I. brunneosquamulosa* and *I. rubrobrunnea* are in pink to highlight its phylogenetic position in the tree. The tree is rooted with *I. adaequata* and *I. calamistrata* of *Inosperma* clade



**Fig. 129** *Inocybe brunneosquamulosa* (CAL 1308, **holotype**). **a, b** Basidiocarps. Scale bars a, b = 5 mm (photos by K. P. Deepna Latha)

clavate, inflated clavate or obovoid, rarely septate, hyaline, occasionally with faint, hyaline encrustations, thin- to slightly thick-walled. *Stipe trama* hyphae with dense, yellowish-brown oleaginous contents. *Clamp connections* seen on all hyphae.

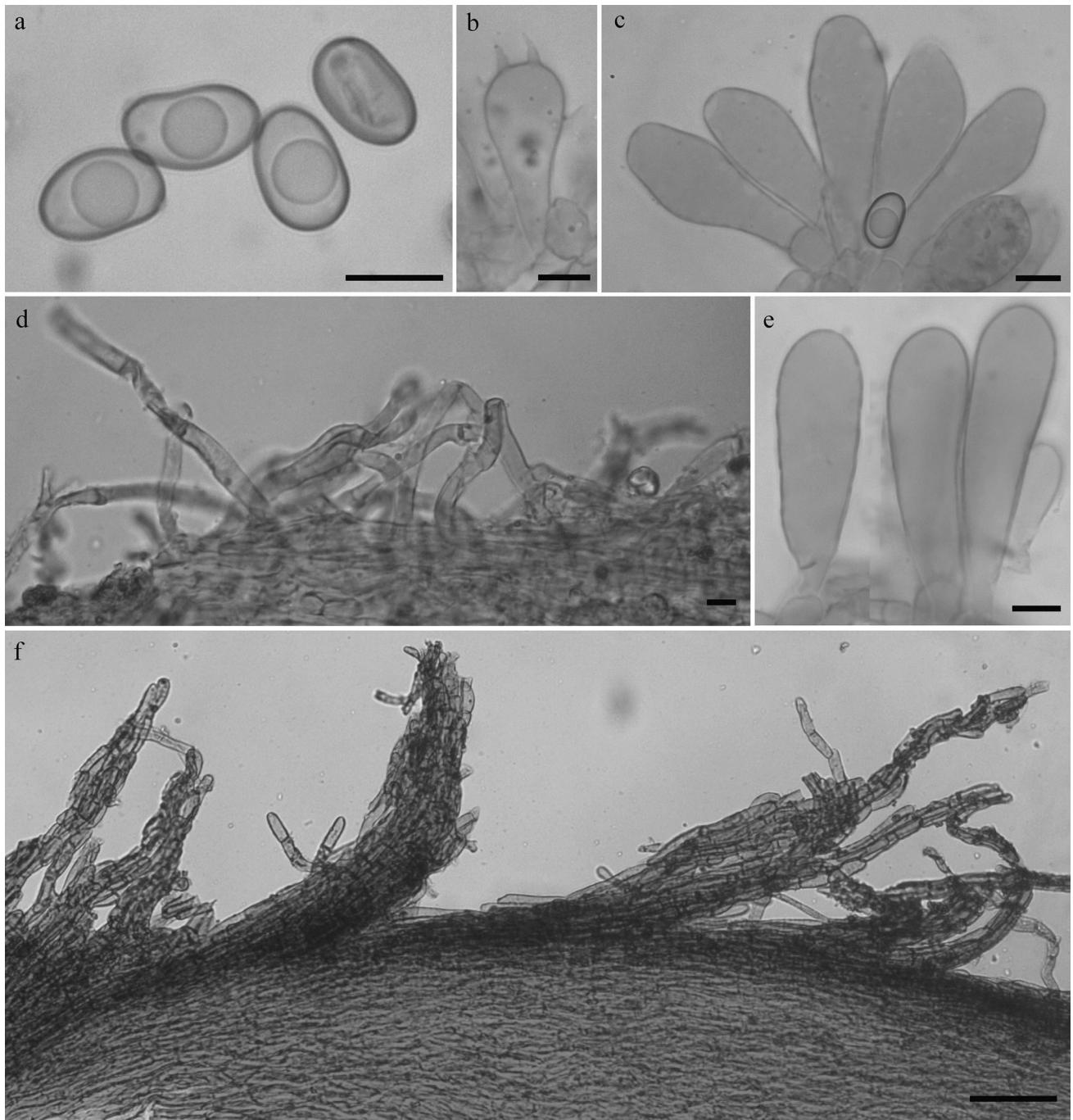
*Habitat:* on the ground, scattered around *Vateria indica* (Dipterocarpaceae) trees.

*Specimen examined:* INDIA, Kerala State, Ernakulam District, Kochi, Thevakkal, Ponnakkudam Kavu sacred grove, 25 August 2014, K.P.D Latha DKP264 (CAL 1308, **holotype**).

GenBank Numbers ITS: KX073582; LSU: KX073586; RPB2: KX073589.

*Notes:* Small basidiocarps with a dark brown, squamulose and fibrillose-rimose pileus; a fibrillose stipe with a finely pruinose apex and an abruptly ending base; smooth, ellipsoid to subphaseoliform basidiospores; a hymenium devoid of pleurocystidia; versiform cheilocystidia with occasional, faint, hyaline encrustations; a cutis-type pileipellis which is disrupted with trichodermal patches and a cutis-type stipitipellis disrupted by loose, hyphal projections and often with caulocystidia at the extreme stipe apex are the salient features of *I. brunneosquamulosa*. *Inocybe fuscospinulosa*, a species originally described from Indonesia (Horak 1980b) and also reported from Sri Lanka

(Pegler 1986), seems to be somewhat similar to *I. brunneosquamulosa* in having a pileus of rather similar colour and surface features, a fimbriate lamella-edge, a fibrillose stipe, a hymenium devoid of pleurocystidia, a trichoderm-type pileipellis and the presence of cheilocystidia. *Inocybe fuscospinulosa*, however, is distinguished from the *I. brunneosquamulosa* in having larger basidiocarps with a densely squamulose pileus, crowded, adnexed, tobacco brown lamellae, a reddish-brown tinted stipe with occasional scales, smaller ( $6.5-8 \times 4-5 \mu\text{m}$ ) and ovoid basidiospores, cylindric to subfusoid cheilocystidia devoid of encrustations and a stipitipellis lacking caulocystidia. *Inocybe brunneosquamulosa* is also somewhat similar to *I. umbrinovirens* E. Horak, a species so far known only from Papua New Guinea (Horak 1980b), in having a somewhat similar-coloured pileus with almost similar surface features, a fibrillose stipe, smooth basidiospores, a hymenium devoid of pleurocystidia, the presence of cheilocystidia with encrusting pigment and a trichoderm-type pileipellis. However, the characters such as the larger basidiocarps with differently-shaped pileus, chocolate brown, adnexed, crowded lamellae, a hollow stipe with a greenish base, larger ( $10-12.5 \times 7-8.5 \mu\text{m}$ ) and ovoid basidiospores, larger basidia, cheilocystidia that are terminal elements of lamellar trama, the absence of



**Fig. 130** *Inocybe brunneosquamulosa* (CAL 1308, holotype). **a** Basidiospores. **b** Basidium. **c** Cheilocystidia. **d** Stipitipellis. **e** Caulocystidia. **f** Pileipellis. Scale bars a–e = 10 µm, f = 100 µm (photos by K. P. Deepna Latha)

caulocystidia and a strong odour make *I. umbrinovirens* different from *I. brunneosquamulosa*.

*Inocybe squamata* J.E. Lange, a species widespread in Europe and USA (Cripps 1997) and also reported from Kerala (Pradeep and Vrinda 2010; Mohanan 2011) is similar to *I. brunneosquamulosa* in having a pileus of somewhat similar texture, rather similarly attached lamellae, a fibrillose stipe, subphaseoliform basidiospores,

a hymenium devoid of pleurocystidia, the presence of cheilo- and caulocystidia and similar type of pileipellis. However, *I. squamata* has larger basidiocarps with a differently-coloured pileus, thick, broad, yellow brown lamellae, a longer, white stipe, larger basidiospores (9–11.5 (13) × (5) 5.5–6.5 µm), occasional 2-spored basidia and narrowly clavate cheilo- and caulocystidia lacking encrustations.

Comparison of the ITS (679 bp), LSU (935 bp) and RPB2 (699 bp) sequences derived from *I. brunneosquamulosa* with the nucleotide sequences available in GenBank Numbers showed that *I. brunneosquamulosa* has distinct sequences. *Inocybe* species MCA562 resulted as the closest hit in megablast searches with ITS (GenBank Numbers JQ408785; Identities = 630/682 (92%)), LSU (GenBank Numbers JN975016; Identities = 921/936 (98%)) and RPB2 (GenBank Numbers JQ421077; Identities = 673/699 (96%)) sequences. *Inocybe* species MCA562 is a collection from Japan, but its morphological features are unavailable for comparison as it remains unpublished.

The phylogenetic placement of *I. brunneosquamulosa* is shown in the phylogram (Fig. 128) generated from the ML analysis of RPB2 sequence data matrix. In the ML analysis, *I. brunneosquamulosa* nested in the Pseudosperma clade with maximum support (100% ML) where it paired with *Inocybe* species MCA562 and had a strong support (96% ML).

***Inocybe luteobrunnea* K.P.D. Latha & Manim., sp. nov.**

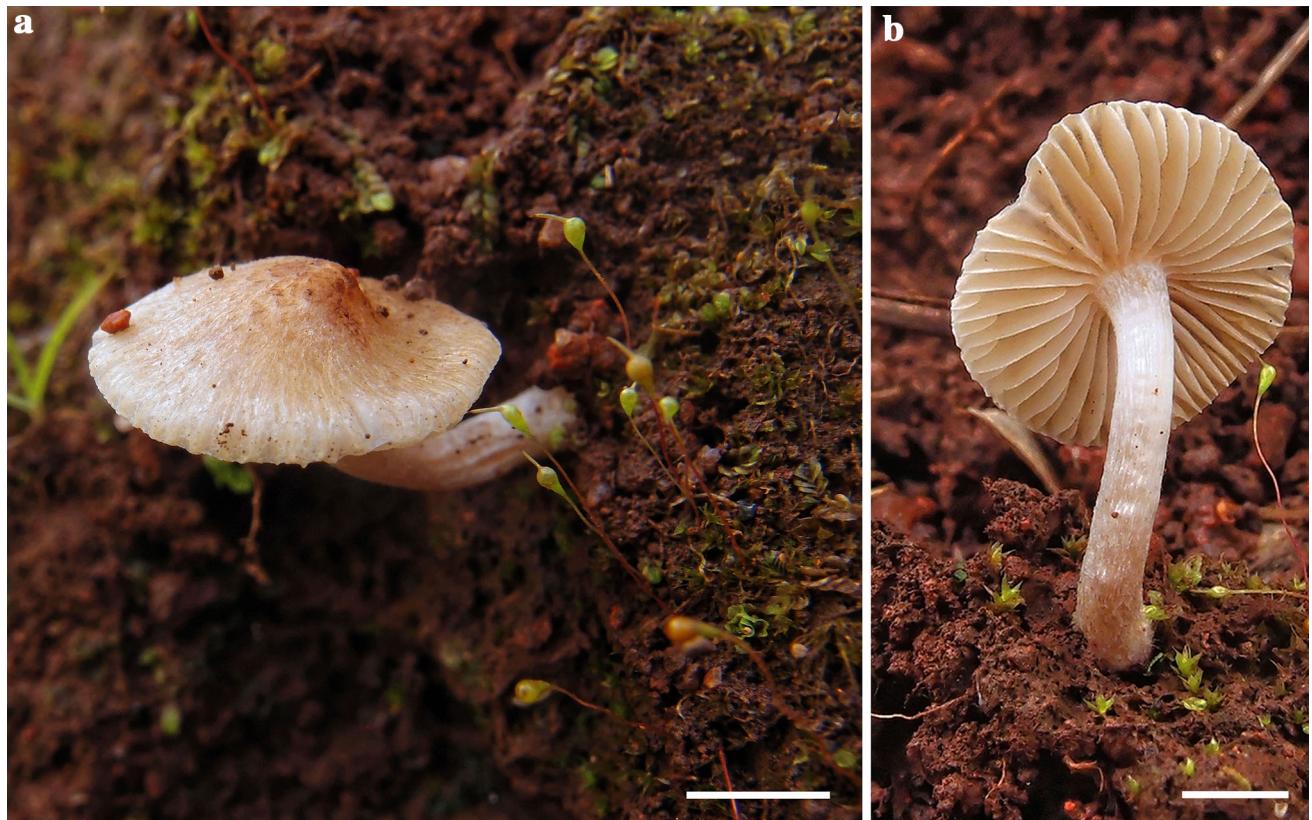
*Mycobank number:* MB 816734; *Facesoffungi number:* FoF 2177; *Figs.* 131, 132

*Etymology:* referring to the yellowish-brown pileus.

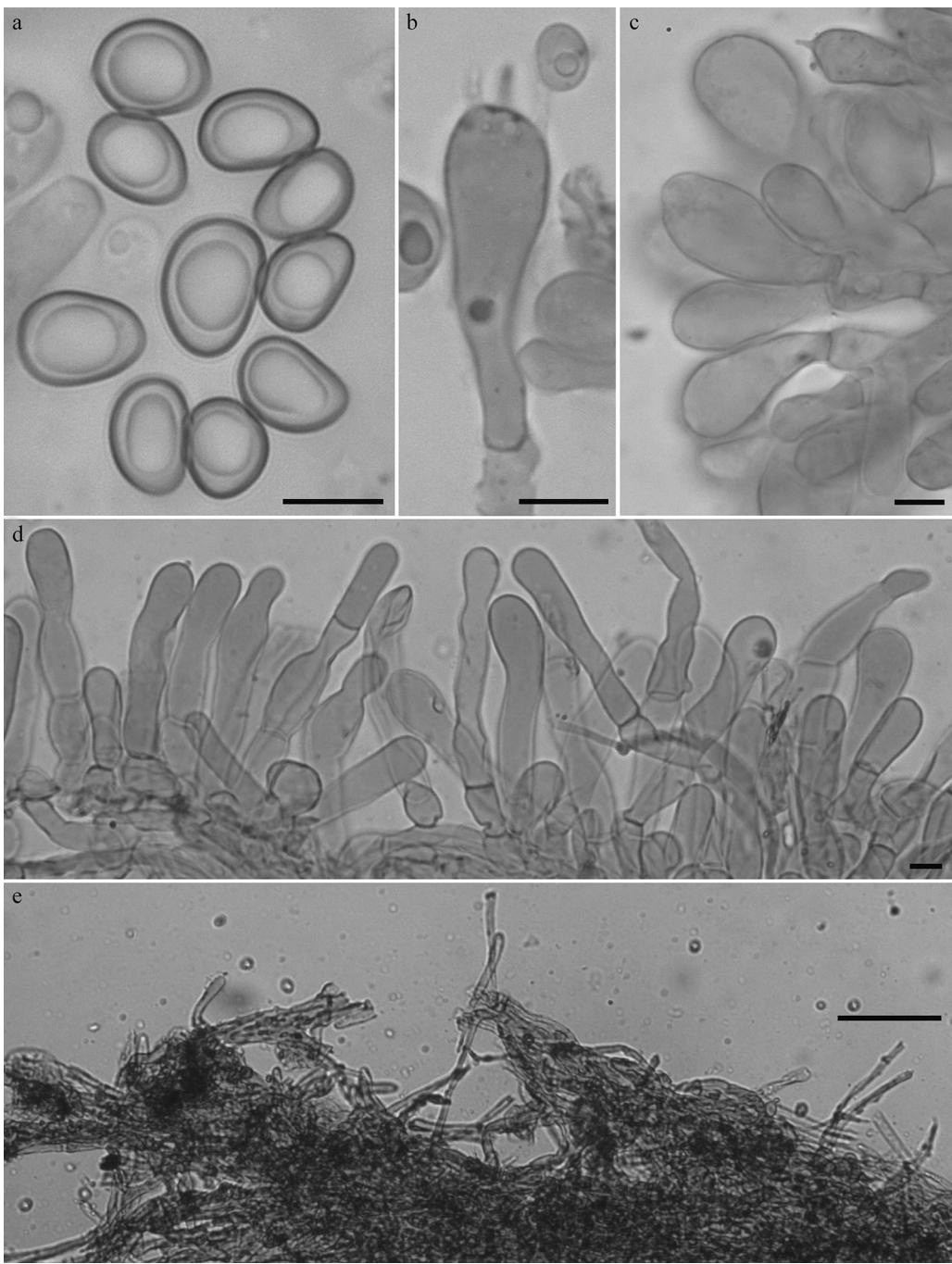
*Holotype:* CAL 1260

*Basidiocarps* small. *Pileus* 6–14 mm diam., narrowly conical when very young, becoming conico-convex and finally convex with a small umbo; surface brown (6F6/OAC636) on the squamules and yellowish-brown (5D8/OAC775) elsewhere when young, becoming dark brown (6F7/OAC639) at the centre and on the squamules and brownish-orange (5C4, 5C5/OAC806) elsewhere at maturity, with appressed- to slightly recurved, minute squamules on and around the umbo, appressed-fibrillose towards the margin; margin incurved when young, becoming decurved to somewhat straight with age, crenate or somewhat wavy, finely fissile. *Lamellae* emarginate, subventricose, rarely furcate, close, greyish-orange (5B3, 5B4/OAC793), up to 2 mm wide, with lamellulae of 3 lengths; edges fimbriate, rather whitish. *Stipe* 13–22 × 2–2.5 mm, central, equal, fistulose; surface initially orange grey (6B2/OAC634), becoming greyish-orange (5B2/OAC675) at maturity, appressed-fibrillose all over, finely pruinose towards the apex; base somewhat bulbous, not marginate-bulbous. *Odour* and *taste* not distinctive.

*Basidiospores* 7–8 (9) × 5–6 (6.5) (7.9 ± 0.7 × 5.9 ± 0.5) µm, Q = 1.2–1.6, Qm = 1.4, smooth, ovoid to amygdaliform, slightly thick-walled, pale yellowish-brown. *Basidia* 23–34 × 8–10 µm, clavate, thin-walled,



**Fig. 131** *Inocybe luteobrunnea* (CAL 1260, holotype). **a, b** Basidiocarp in the field. Scale bars a, b = 5 mm (photos by K. P. Deepna Latha)



**Fig. 132** *Inocybe luteobrunnea* (CAL 1260, holotype). **a** Basidiospores. **b** Basidium. **c** Cheilocystidia. **d** Caulocystidia. **e** Pileipellis. Scale bars a-d = 10 µm, e = 100 µm (photos by K. P. Deepna Latha)

hyaline, 4-spored or rarely 2-spored; sterigmata up to 5 µm long. Mature basidia slightly projecting beyond the hymenial surface. *Pleurocystidia* absent. *Lamella-edge* sterile with abundant cheilocystidia. *Cheilocystidia* 17–52 × 9–15 µm, versiform: clavate, inflated clavate, cylindrical or cylindrical with apical constriction, cylindro-flexuous, cylindrical with irregular constriction, narrowly utriform, utriform with a median constriction, ovoid, fusiform,

submoniliform or short-pedicellate, often septate, thin- to slightly thick-walled, hyaline. *Lamellar trama* subregular, composed of both narrow and inflated hyphae; hyphae 3–25 µm wide, thin- to slightly thick-walled, hyaline, at times with faint, hyaline encrustations especially towards the edge of the hymenium. *Subhymenium* poorly developed. *Pileus trama* subregular; hyphae 9–28 µm wide, pale yellow, thin- to slightly thick-walled. *Pileipellis* a cutis

often transitioning to a trichoderm towards the centre; hyphae 4–11 µm wide, tangled, slightly thick-walled, with a pale-yellow wall pigment and brown spiral encrustations. *Stipitipellis a cutis* frequently disrupted with bunches of caulocystidia towards the apex; hyphae 5–12 µm wide, thin-to slightly thick-walled, with a pale-yellow wall pigment and hyaline encrustations, some hyphae with dense, yellowish-brown, amorphous contents towards the base. *Caulocystidia* 13–74 × 7–13 µm, versiform: clavate, narrowly clavate, moniliform, submoniliform, cylindroflexuous, cylindrical with an obtuse apex, utriform with a median constriction, cylindrical or ovoid with a rostrate apex, often septate, thin- to slightly thick-walled, hyaline. *Clamp connections* observed on all hyphae.

*Habitat:* scattered among bryophytes, on a mud wall.

*Material examined:* INDIA, Kerala State, Idukki District, Munnar, on the way to Mattupetti top hill station, 31 August 2013, K. P. Deepna Latha DKP167 (CAL 1260, **holotype**; GenBank Numbers ITS: KX073580; LSU: KX073584; RPB2: KX073588; INDIA, Kerala State, Idukki District, Munnar, on the way to Mattupetti top hill station, 9 November 2013, K. P. Deepna Latha DKP251 (CAL 1260, **holotype**; CAL 1261, **paratype**).

GenBank Numbers ITS: KX073581; LSU: KX073585.

*Notes:* *Inocybe luteobrunnea* is characterised by a yellowish-brown pileus with a fibrillose to minutely squamulose surface; a fibrillose stipe with a finely pruinose apex and a bulbous base; smooth, ovoid to amygdaliform basidiospores; a hymenium lacking pleurocystidia; abundant, versiform cheilocystidia; subregular lamellar trama with faint hyaline encrustations; a pileipellis that is a cutis with a transition to a trichoderm towards the centre and a cutis-type stipitipellis frequently disrupted with bunches of caulocystidia confined to the stipe apex. *Inocybe palaeotropica* E. Turnbull & Watling, a widespread species reported from Singapore, Malaysia, Sabah, Papua New Guinea, Solomon Islands (Horak 1980b) and also recorded from Kerala by Vrinda et al. (1997, as *I. umbrina* Massee), shares a few characters with *I. luteobrunnea* such as a similar-shaped pileus, similar-sized basidiospores (7.5–10.5 × 4.5–6 µm), a sterile lamella-edge and caulocystidia on the stipe apex. *Inocybe palaeotropica*, however, has larger basidiocarps with dissimilar colour and surface features, adnexed lamellae, a fibrillose stipe devoid of apical pruinosity, ovoid to short ellipsoid basidiospores, only 4-spored basidia, clavate to cylindro-clavate cheilocystidia, hyphae of lamellar trama lacking encrustations, a cutis-type pileipellis and smaller, cylindro-clavate caulocystidia. *Inocybe luteobrunnea* is somewhat similar to *I. fuscospinulosa* Corner & E. Horak, an Indonesian species (Horak 1980b) and also reported from Sri Lanka (Pegler 1986), in having a somewhat similar-shaped pileus, a fibrillose stipe, almost similar-sized basidiospores (6.5–8 ×

4–5 µm), cheilocystidia, a hymenium devoid of pleurocystidia and an almost similar pileipellis structure. However, *I. fuscospinulosa* differs from *I. luteobrunnea* owing to its larger basidiocarps with conspicuous, erect, spiny squamules on the pileus, crowded, adnexed, tobacco-brown lamellae, a red-brown tinged stipe with occasional brown squamules and without a distinct bulbous base, ovoid basidiospores, consistently 4-spored basidia, larger, differently-shaped cheilocystidia and a stipitipellis devoid of caulocystidia.

The distinctive status of (CAL 1260: 675 bp; CAL 1261: 674 bp), LSU (CAL 1260: 838 bp; CAL 1261: 746 bp) and RPB2 (CAL 1260: 651 bp) sequences of *I. luteobrunnea* was confirmed in the BLASTn searches. An unnamed Australian species, *Inocybe* species AU95 was the closest hit in a megablast search for ITS (GenBank Numbers KP636851; Identities = 540/588 (92%)), LSU (GenBank Number KP171053; Identities = 831/839 (99%)) and RPB2 (GenBank Number KM555145; Identities = 638/651 (98%)) sequences.

The phylogram generated from the Maximum Likelihood (ML) analysis (Fig. 129) depicts the relative placement of *I. luteobrunnea*. The ML analysis placed *I. luteobrunnea* in the Pseudosperma clade with full support (100% ML) based on RPB2 sequence data matrix. Within this clade, *I. luteobrunnea* clustered with an unnamed Australian species, *Inocybe* species AU95 (KM555145) with maximum support (100% ML). The macro-morphological and microscopic data of that species are not available for comparison.

#### *Inocybe rubrobrunnea* K.P.D. Latha & Manim., *sp. nov.*

*Mycobank number:* MB 816736; *Facesoffungi number:* FoF: 2175; *Figs.* 133, 134

*Etymology:* referring to the reddish-brown colour of the pileus.

*Holotype:* CAL 1307

*Basidiocarps* small. *Pileus* 6–12 mm diam., somewhat hemispherical or paraboloid when young, becoming convex or plano-convex with a small umbo; surface reddish-brown (8E5, 8E6/OAC609, OAC610) when young, becoming dark brown (7F7/OAC621) at the centre and on the squamules, orange grey (6B2) elsewhere, appressed- to slightly recurved squamulose and appressed-fibrillose all over when young, becoming appressed- to recurved squamulose on and around the centre, appressed-fibrillose and rimulose towards the margin; margin initially incurved, becoming decurved to somewhat straight with age, crenate. *Lamellae* sinuate or emarginate, close, initially greyish-orange (6B3/OAC654), becoming brownish-orange (6C4/OAC695), up to 1.5 mm wide, with lamellulae of 3 lengths; edges fimbriate, whitish. *Stipe* 11–20 × 1–2 mm, central, terete, equal or slightly tapered towards the base,



**Fig. 133** *Inocybe rubrobrunnea* (CAL 1307, holotype). **a, b** Basidiocarp in the field. Scale bars a, b = 5 mm (photos by K. P. Deepna Latha)

cartilaginous, solid; surface brownish-orange (6C3/OAC633), fading towards the apex, appressed-fibrillose all over, slightly pruinose at the apex; base slightly enlarged. *Odour* and *taste* not distinctive.

*Basidiospores*  $7\text{--}9 \times 5\text{--}6$  ( $8.32 \pm 0.63 \times 5.57 \pm 0.43$ )  $\mu\text{m}$ ,  $Q = 1.2\text{--}1.7$ ,  $Qm = 1.4$ , smooth, ellipsoid to slightly phaseoliform, slightly thick-walled, pale yellowish-brown. *Basidia*  $18\text{--}27 \times 9\text{--}12 \mu\text{m}$ , clavate, thin-walled, hyaline, 4-spored; sterigmata up to  $3 \mu\text{m}$  long. *Pleurocystidia* absent. *Lamella-edge* heterogeneous. *Cheilocystidia*  $17\text{--}44 \times 6\text{--}15 \mu\text{m}$ , abundant, versiform: narrowly clavate, clavate, cylindrical, oblong, ellipsoid, fusiform, ovoid, cylindrical with a median constriction or with a subcapitate apex, occasionally septate, exuding some amorphous material at the apex, slightly thick-walled (up to  $1 \mu\text{m}$  thick), hyaline or rarely with pale yellowish-brown, amorphous contents, occasionally with faint, hyaline encrustations. *Lamellar trama* subregular, composed of both narrow and inflated hyphae; hyphae  $3\text{--}23 \mu\text{m}$  wide, thin- to slightly thick-walled, hyaline. *Subhymenium* pseudoparenchymatous. *Pileus trama* subregular; hyphae  $6\text{--}27 \mu\text{m}$  wide, hyaline or pale yellow, thin-walled. *Pileipellis* a cutis often disrupted with bundles of tangled ascending hyphae towards the centre; hyphae  $5\text{--}10 \mu\text{m}$  wide, with a pale brownish-yellow wall

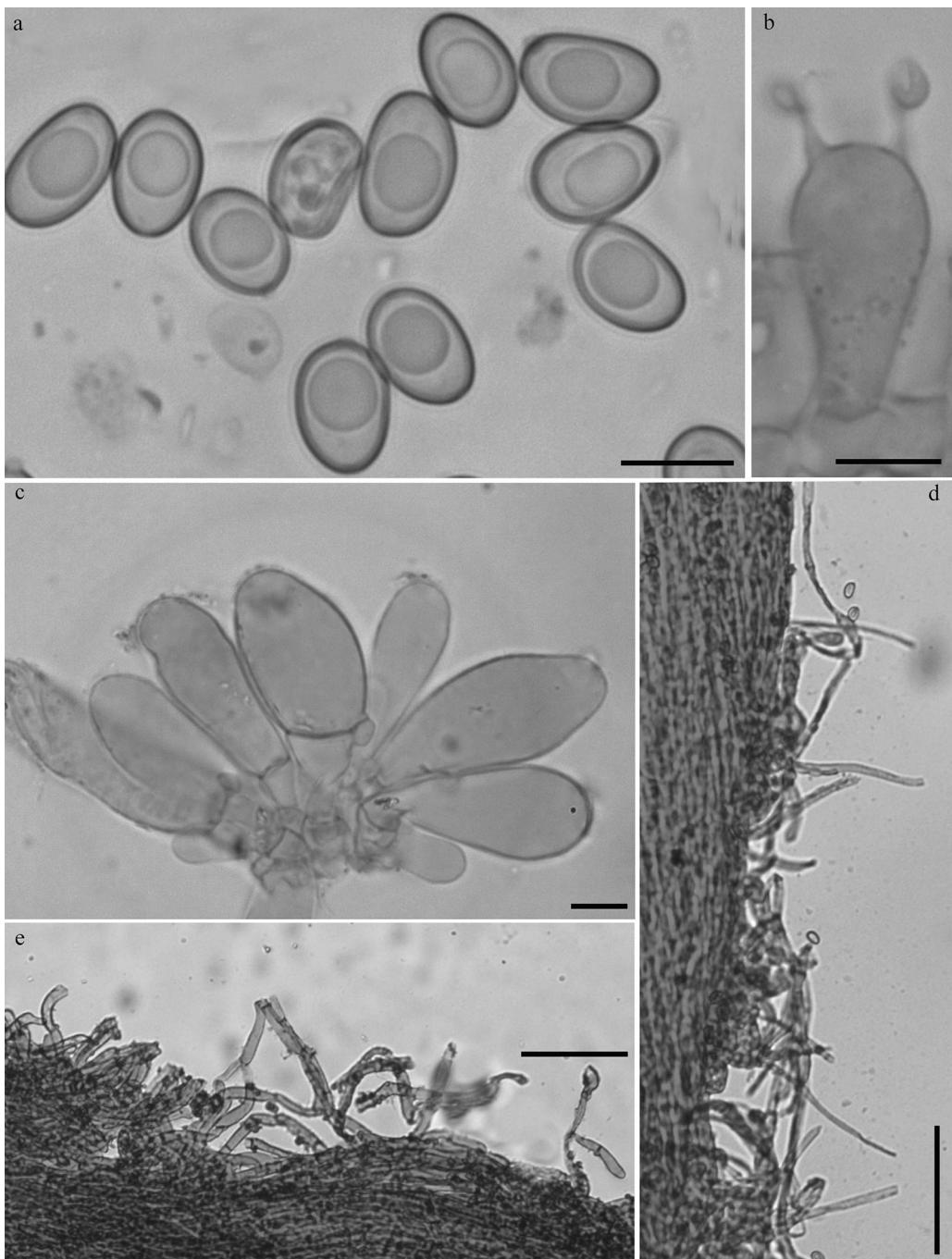
pigment and brown spiral encrustations. *Stipitipellis* a cutis composed of hyaline or pale yellow hyphae ( $3\text{--}10 \mu\text{m}$  wide), devoid of encrustations, frequently disrupted by bundles of loose, tangled hyphae towards the apex ( $2\text{--}6 \mu\text{m}$  wide), with a pale-yellow wall pigment and pale yellow spiral encrustations, thin- to slightly thick-walled, occasionally with yellowish-brown amorphous contents. *Caulocystidia* absent. *Clamp connections* observed on all hyphae.

*Habitat:* on soil, solitary or in small groups, near *Hopea ponga* (Dipterocarpaceae) trees.

*Specimen examined:* INDIA, Kerala State, Wayanad District, Muthanga, Muthanga Wildlife Sanctuary, 21 August 2013, K. P. Deepna Latha DKP142 (CAL 1307, holotype).

GenBank Numbers ITS:KX073583; LSU:KX073587; RPB2:KX073590.

*Notes:* Small basidiocarps with an appressed-fibrillose to squamulose pileus; a stipe with a slightly enlarged base and a fibrillose surface; smooth, ellipsoid to slightly phaseoliform basidiospores; a hymenium devoid of pleurocystidia; versiform cheilocystidia exuding some amorphous material at the apex; a cutis-type pileipellis disrupted with bundles of tangled ascending hyphae; and a cutis-type stipitipellis often disrupted by bundles of loose, tangled



**Fig. 134** *Inocybe rubrobrunnea* (CAL 1307, holotype). **a** Basidiospores. **b** Basidium. **c** Cheilocystidia. **d** Stipitipellis. **e** Caulocystidia. **f** Pileipellis. Scale bars a–c = 10  $\mu\text{m}$ , d, e = 100  $\mu\text{m}$  (photos by K. P. Deepna Latha)

hyphae towards the apex are the diagnostic features of *I. rubrobrunnea*.

Following the key of Kobayashi (2002), *I. rubrobrunnea* keys out close to *I. quercina* Hongo, a species known from Japan, because of its reddish-brown pileus with almost similar size, shape and surface features, a whitish lamella-edge, a solid stipe with a fibrillose surface, almost similar-shaped basidiospores, the absence of pleurocystidia and the

presence of cheilocystidia. However, the characters such as the adnexed lamellae that become red when cut, a longer stipe, larger basidiospores ( $8.2\text{--}10.8 \times 4.8\text{--}5.5 \mu\text{m}$ ) and basidia, smaller cheilocystidia devoid of encrustations, a cutis-type pileipellis lacking encrustations, the presence of caulocystidia and a strong odour of that species make it different from *I. rubrobrunnea*. *Inocybe fuscospinulosa*, a species originally described from Indonesia (Horak 1980b)

and also reported from Sri Lanka (Pegler 1986), has a pileus with dark brown squamules, a whitish lamella-edge, basidiospores of similar size ( $7\text{--}9 \times 4.2\text{--}5.5 \mu\text{m}$ ) and shape and the absence of pleuro- and caulocystidia. However, that species differs in having larger basidiocarps with a campanulate to applanate pileus, a white context discolouring purplish-red on exposure, cheilocystidia devoid of encrustations and a trichoderm-type pileipellis.

Comparison of the ITS (676 bp), LSU (924 bp) and RPB2 (702 bp) sequences of *I. rubrobrunnea* with the nucleotide sequences of taxa available in GenBank Numbers suggests that it has distinct sequences. In a megablast search of the GenBank Numbers database using ITS sequence of *I. rubrobrunnea*, the closest hit was *Inocybe* species MCA562 (GenBank Numbers JQ408785; Identities = 616/668 (92%)) followed by *Inocybe* species AU43 (GenBank Numbers KJ729878; Identities = 605/659 (92%)). An undescribed *Inocybe* species, *Inocybe* species AU44 was the closest hit in BLASTn search with LSU (GenBank Numbers KJ729906; Identities = 896/925 (97%)) sequence. *Inocybe* species AU43 (GenBank Numbers KJ729935; Identities = 670/702 (95%)) resulted as the closest hit in BLASTn search with RPB2 sequences. But, the details of *Inocybe* species MCA562, an unnamed *Inocybe* collection from Japan and *Inocybe* species AU43 and *Inocybe* species AU44, another unnamed collection from Australia, are not available for comparison.

The RPB2-based ML phylogeny (Fig. 128) placed *Inocybe rubrobrunnea* in the Pseudosperma clade with full support (100% ML). Within this clade, *I. rubrobrunnea* clustered with *Inocybe* species AU43 with a significant support (95% ML).

#### ***Marasmiaceae* Roze ex Kühner.**

The family *Marasmiaceae* is characterized by white spores. The members of this family mostly have tough stems and the capability of shrivelling up during a dry period and later recovering. According to Kirk et al. (2008), the family contains 54 genera and 1590 species.

#### ***Marasmius* Fr.**

*Marasmius* is a genus of mushroom-forming fungi in the family *Marasmiaceae*. It contains about 500 species (Kirk et al. 2008) of which a few, such as *Marasmius oreades*, are edible. However, most members of this genus are small, unimpressive, brown mushrooms. Their humble appearance contributes to them not being readily noticeable to the layman, and therefore these mushrooms are seldom collected by mushroom hunters. Quite a few of the species are known to grow in the characteristic fairy ring pattern (Fig. 135).

#### ***Marasmius luculentus* A.K. Dutta, K. Acharya & Antonín, sp. nov.**

*Mycobank number:* MB 816959; *Facesoffungi number:* FoF 2192, Figs. 136, 137

*Etymology:* referring to the beautiful (luculentus) appearance of the pileus.

*Holotype:* CUH AM120

*Pileus* 5–10 mm in diam., conic to hemispherical when very young, becoming convex in age, with a small conic to convex papilla that ranges from reddish-brown (8D6) to violet brown (10E5–6, 11E6–7) when young, but later turning brown (7D7, 7E8) to dark brown (7F7) at maturity, very rarely forming a central dot, often with a depression around the papilla; striate to plicate up to center, margin often crenate; surface dry, glabrous, hygrophanous, brownish-orange (7C4) to light brown (7D5–6) when very young, later reddish-grey (7–8B2) and finally turning white (1A1) to off-white at maturity, greyish-yellow (1–2B4, 4B4–5) on drying; context very thin, white to cream. *Lamellae* adnexed or adnate to a collarium, distant to subdistant (L = 12–14, I = 0), white (1A1), regular, slightly intervenose, edge concolorous. *Stipe* 5–9(–12) × 0.1–0.3 mm, central, glabrous, wiry, pliant, cylindric, equal, simple and insititious on the substratum, dark brownish-black to black overall, often accompanied by black rhizomorphs, 30–42 mm long. *Odour* and *taste* indistinct. *Spore print* white.

*Basidiospores* (9.3–)10–10.3–10.5(–11) × (3.9–)4.3–4.8–5.4(–5.8)  $\mu\text{m}$ , Q = 1.7–2.2–2.6, ellipsoid to ellipsoid-fusoid, smooth, hyaline, IKI-, thin-walled. *Basidia* not observed. *Basidioles* 24–25 × 5.5–7  $\mu\text{m}$ , fusoid to clavate, hyaline, thin-walled. *Lamellae edge* sterile. *Pleurocystidia* absent. *Cheilocystidia* composed of *Siccus*-type broom cells; main body 14–18 × 4–6(–7.5)  $\mu\text{m}$ , sphaeropedunculate to (broadly) clavate or irregular in outline, hyaline, apically thick-walled; apical setulae 2–3.5(–5.5)  $\mu\text{m}$  long, cylindrical or irregular in outline, obtuse, pale yellow to light brown in KOH, thick-walled. *Pileipellis* a hymeniform layer, composed of *Siccus*-type broom cells; main body (17–)19–23(–25) × (7–)8–11(–14.5)  $\mu\text{m}$ , sphaeropedunculate to clavate, broadly clavate, often branched, pale yellow to light brownish in KOH, thick-walled; apical setulae (1.5–)3–4(–5.5)  $\mu\text{m}$  long, cylindrical, obtuse, thick-walled. *Pileus trama* composed of 4.5–7.5  $\mu\text{m}$  broad, interwoven, cylindrical, hyaline, inamyloid, thin-walled hyphae. *Lamellar trama* hyphae 4–6.5  $\mu\text{m}$  broad, interwoven, cylindrical, hyaline, inamyloid, thin-walled. *Stipitipellis* hyphae 5.5–6.5  $\mu\text{m}$  broad, parallel to subparallel, cylindrical, smooth, non-gelatinous, hyaline to pale yellow in KOH, non-dextrinoid to weakly dextrinoid, thick-walled. *Stipe trama* hyphae 6.8–7.5  $\mu\text{m}$  broad, parallel to subparallel, cylindrical,

## References

- Aalto M (1974) *Amanita magnivolvata* sp. nova (Agaricales). Karstenia 14:93–96
- Abbott SP, Currah RS (1997) The *Hevellaceae*: systematic revision and occurrence in northern and northwestern North America. Mycotaxon 62:1–125
- Abdollahzadeh J, Goltapeh EM, Javadi A, Shams-Bakhsh M, Zare R, Phillips AJL (2009) *Barriopsis iraniana* and *Phaeobotryon cupressi*, two new species of the *Botryosphaeriaceae* from trees in Iran. Persoonia 23:1–8
- Adamčík S, Cai L, Chakraborty D, Chen XH, Cotter H, Van T, Dai DQ, Dai YC, Sas K, Deng C, Ghobad-Nejhad M, Hyde KD, Langer E, Latha KPD, Liu F, Liu SL, Liu T, Wei LV, Shu-Xia LV, Machado AR, Pinho DB, Pereira OL, Prasher IB, Rosado AWC, Qin J, Qin WM, Verma RK, Wang Q, Yang ZL, Yu XD, Zhou LW, Buyck B (2015) Fungal biodiversity profiles 1–10. Cryptogam Mycol 36:121–166
- Adams GC, Wingfield MJ, Common R, Roux J (2005) Phylogenetic relationships and morphology of *Cytospora* species and related teleomorphs (Ascomycota, Diaporthales, Valsaceae) from Eucalyptus. Stud Mycol 52:1–144
- Adams GC, Roux J, Wingfield MJ (2006) *Cytospora* species (Ascomycota, Diaporthales, Valsaceae): introduced and native pathogens of trees in South Africa. Australas Plant Pathol 35:521–548
- Addy HD, Piercey MM, Currah RS (2005) Microfungal endophytes in roots. Can J Bot 83:1–13
- Ahmad S (1969) Fungi of West Pakistan, supplement 1. Society at the Biological Laboratories, Government College, Lahore
- Ahmad S, Iqbal SH, Khalid AN (1997) Fungi of Pakistan. Sultan Ahmad Mycological Society of Pakistan, Department of Botany, University of the Punjab, Lahore
- Aime L, Ryvarden L, Henkel TW (2007) Studies in Neotropical polypores 22. Additional new and rare species from Guyana. Synop Fungorum 23:15–31
- Alvarado P, Manjon JL, Matheny PB, Esteve-Raventos F (2010) *Tubariomyces*, a new genus of *Inocybaceae* from the Mediterranean region. Mycologia 102:1389–1397
- Andersson M (2013) A rare bolete, *Boletus legaliae*, in the Royal Garden of Drottningholm, Stockholm. Sven Mykol Tidskr 34(3):9–13
- Antón V, Buyck B (2006) *Marasmius* (Basidiomycota, Marasmiaceae) in Madagascar and the Mascarenes. Fungal Divers 23:17–50
- Antón V (2007) Monograph of *Marasmius*, *Gloiocephala*, *Palaeocephala* and *Setulipes* in tropical Africa. Fungus Flora Trop Afr 1:1–177
- Antón V, Vizzini A, Ercole E, Leonardi M (2015) *Strobilomyces pteroreticulosporus* (Boletales), a new species of the *S. strobilaceus* complex from the Republic of Korea and remarks on the variability of *S. confuses*. Phytotaxa 219(1):78–86
- Ariyawansa HA, Tanaka K, Thambugala KM, Phookamsak R, Tian Q, Camporesi E, Hongsanan S, Monkai J, Wanasinghe DN, Mapook A, Chukeatirote E (2014a) A molecular phylogenetic reappraisal of the *Didymosphaeriaceae* (=Montagnulaceae). Fungal Divers 68:69–104
- Ariyawansa HA, Hawksworth DL, Hyde KD, Jones EG, Maharachchikumbura SS, Manamgoda DS, Thambugala KM, Udayanga D, Camporesi E, Daranagama A, Jayawardena R (2014b) Epitypification and neotypification: guidelines with appropriate and inappropriate examples. Fungal Divers 69(1):57–91
- Ariyawansa HA, Phukhamsakda C, Thambugala KM, Bulgakov TS, Wanasinghe DN, Perera RH, Mapook A, Camporesi E, Kang JC, Jones EG, Bahkali AH (2015a) Revision and phylogeny of *Leptosphaeriaceae*. Fungal Divers 74:19–51
- Ariyawansa HA, Thambugala KM, Manamgoda DS, Jayawardena R, Camporesi E, Boonmee S, Wanasinghe DN, Phookamsak R, Hongsanan S, Singtripop C, Chukeatirote E (2015b) Towards a natural classification and backbone tree for *Pleosporaceae*. Fungal Divers 71(1):85–139
- Ariyawansa HA, Hyde KD, Jayasiri SC, Buyck B, Cheethana KWT, Dai DQ, Dai YC, Daranagama DA, Jayawardena RS, Lücking R, Ghobad-Nejhad M, Niskanen T, Thambugala KM, Voigt K, Zhao RL, Li GJ, Doilom M, Boonmee S, Yang ZL, Cai Q, Cui YY, Bahkali AH, Chen J, Cui BK, Chen JJ, Dayarathne MC, Dissanayake AJ, Ekanayaka AH, Hashimoto A, Hongsanan S, Jones EBG, Larsson E, Li WJ, Li QR, Liu JK, Luo ZL, Maharachchikumbura SSN, Mapook A, McKenzie EHC, Norphanphoun C, Konta S, Pang KL, Perera RH, Phookamsak R, Phukhamsakda C, Pinruan U, Randrianjohany E, Singtripop C, Tanaka K, Tian CM, Tibpromma S, Abdel-Wahab MA, Wanasinghe DN, Wijayawardene NN, Zhang JF, Zhang H, Abdel-Aziz FA, Wedin M, Westberg M, Ammirati JF, Bulgakov TS, Lima DX, Callaghan TM, Callac P, Chang CH, Coca LF, Dal-Forno M, Dollhofer V, Fliegerová K, Greiner K, Griffith GW, Ho HM, Hofstetter V, Jeewon R, Kang JC, Wen TC, Kirk PM, Kyttövuori I, Lawrey JD, Xing J, Li H, Liu ZY, Liu XZ, Liimatainen K, Thorsten Lumbsch H, Matsumura M, Moncada B, Nuankaew S, Parnmen S, Santiago ALCMA, Sommai S, Song Y, de Souza CAF, de Souza-Motta CM, Su HY, Suetrong S, Wang Y, Fong WS YH, Zhou LW, Réblová M, Fournier J, Camporesi E, Luangsa-ard JJ, Tasanathai K, Khonsanit A, Thanakitpipattana D, Somrittipol S, Diederich P, Millanes AM, Common RS, Stadler M, Yan JY, Li XH, Lee HW, Nguyen TTT, Lee HB, Battistin E, Marsico O, Vizzini A, Vila J, Ercole E, Eberhardt U, Simonini G, Wen HA, Chen XH, Miettinen O, Spirin V, Hernawati (2015c) Fungal diversity notes 111–252 taxonomic and phylogenetic contributions to fungal taxa. Fungal Divers 75:27–274
- Arora D, Ferrarese G, Simonini G (1999) *Boletus pulcherrimus* *Boletus rubrosanguineus*: lo stesso taxon? Micol Veget Medit 14(2):116–141
- Arun Kumar TK, Manimohan P (2009) The genus *Lepiota* (Agaricales, Basidiomycota) in Kerala State, India. Mycotaxon 107:105–138
- Arzanlou M, Groenewald JZ, Gams W, Braun U, Shin HD, Crous PW (2007) Phylogenetic and morphotaxonomic revision of *Ramichloridium* and allied genera. Stud Mycol 58:57–93
- Aveskamp MM, de Gruyter J, Crous PW (2008) Biology and recent developments in the systematics of *Phoma*, a complex genus of major quarantine significance. Fungal Divers 31:1–18
- Aveskamp MM, de Gruyter J, Woudenberg JHC, Verkley GJM, Crous PW (2010) Highlights of the *Didymellaceae*: a polyphasic approach to characterise *Phoma* and related pleosporalean genera. Stud Mycol 65:1–60
- Bahl J, Jeewon R, Hyde KD (2005) Phylogeny of *Rosellinia capetribulensis* sp. nov. and its allies (Xylariaceae). Mycologia 97:1102–1110
- Baroni TJ, Hofstetter V, Largent DL, Vilgalys R (2011) *Entocybe* is proposed as a new genus in the *Entolomataceae* (Agaricomycetes, Basidiomycota) based on morphological and molecular evidence. N Am Fungi 6:1–19
- Barr ME (1978) The *Diaporthales* in North America with emphasis on *Gnomonia* and its segregates. Mycol Mem 7:1–232
- Barr ME (1979) A classification of Loculoascomycetes. Mycologia 71:935–957
- Barr ME (1987) Prodomus to class Loculoascomycetes. Published by the Author, Amherst

- Barr ME (1990) *Melanommatales* (Loculoascomycetes). N Am Flora Ser II 13:1–129
- Barr ME (1992) Notes on the *Lophiostomataceae* (*Pleosporales*). Mycotaxon 45:191–221
- Bas C (1969) Morphology and subdivision of *Amanita* and a monograph of its section *Lepidella*. Persoonia 5:285–579
- Bas C (1974) A rare and widespread *Amanita* associated with *Alnus*. Bulletin Mensuel de la Société Linnéenne de Lyon 43:17–23
- Bas C (1984) On the correct name of '*Amanita inaurata* Secr.'. Persoonia Mol Phylogeny Evol Fungi 12:192–193
- Bas C, Kuyper TW, Noordeloos ME, Vellinga EC (1995) Flora Agaricina Nederlandica—3, *Tricholomataceae*. A. A. Balkema, Rotterdam
- Bates ST, Reddy GSN, Garcia-Pichel F (2006) *Exophiala crusticola* anam. nov. (affinity *Herpotrichiellaceae*), a novel black yeast from biological soil crusts in the Western United States. Int J Syst Evol Microbiol 56:2697–2702
- Bau T, Li Y, Irina AG, Eugenia MB, Wasiliy AS (2008) Common wild edible mushroom resource of Russia. Edible Fungi China 27:9–13
- Benny GL (2008) Methods used by Dr. R. K. Benjamin, and other mycologists, to isolate Zygomycetes. Also 26:37–61
- Benny GL, Smith ME, Kirk PM, Tretter ED, White MM (2016) Challenges and Future Perspectives in the Systematics of *Kickxellomycotina*, *Mortierellomycotina*, *Mucoromycotina*, and *Zoopagomycotina*. In: Li D-W (ed) Biology of microfungi, fungal biology. Springer, Basel
- Berkeley MJ (1847) Decades of fungi XII–XIV. Lond J Bot 6:312–326
- Bernicchia A, Gorjón SP (2010) *Corticaceae* s.l. Fungi Europaei, vol 12. Ed Candusso, Italia
- Bessette AE, Roody WC, Bessette AR (2000) North American Boletes. Syracuse University Press, Syracuse
- Bessette AE, Roody WC, Bessette AR (2010) North American Boletes: a color guide to fleshy pored mushrooms. Syracuse University Press, Syracuse
- Bhatt RP, Tulloss RE, Semwal KC, Bhatt VK, Moncalvo JM, Stephenson SL (2003) *Amanitaceae* from India. A critically annotated checklist. Mycotaxon 88:249–270
- Binder M, Hibbett DS (2006) Molecular systematics and biological diversification of Boletales. Mycologia 98:971–981
- Binder M, Justo A, Riley R, Salamov A, Lopez-Giraldez F, Sjökvist E, Copeland A, Foster B, Sun H, Larsson E, Larsson KH, Townsend J, Grigoriev IV, Hibbett D (2013) Phylogenetic and phylogenomic overview of the *Polyporales*. Mycologia 105:1350–1373
- Binion DE, Stephenson SL, Roody WC, Burdsall HH Jr., Vasilyeva LN, Miller OK Jr. (2008) Macrofungi associated with oaks of Eastern North America. West Virginia University Press, Morgantown
- Bisby GR, Buller AHR, Dearness J (1929) The fungi of Manitoba. Longmans, Green and Co, London
- Boehm EW, Schoch CL, Spatafora JW (2009a) On the evolution of the *Hysteriaceae* and *Mytilinidiaceae* (*Pleosporomycetidae*, Dothideomycetes, Ascomycota) using four nuclear genes. Mycol Res 113:461–479
- Boehm EWA, Mugambi G, Miller AN, Huhndorf S, Marinowitz S, Schoch CL, Spatafora JW (2009b) A molecular phylogenetic reappraisal of the *Hysteriaceae*, *Mytilinidiaceae* and *Gloniaceae* (*Pleosporomycetidae*, Dothideomycetes) with key to world species. Stud Mycol 64:49–83
- Bolay A (1971) Contribution à la connaissance de *Gnomonia comari* Karsten (syn. *G. fructicola* [Arnaud] Fall). Etude taxonomique, phytopathologique et recherches sur sa croissance in vitro. Ber Schweizerischen Bot Ges 81:398–482
- Borisov BA, Tarasov KL (1999) Notes on biodiversity of causal agents of invertebrate mycoses in Adjaria (south-western Georgia). I. *Evlachovaea kintrischica* gen. et sp. nov. (Hyphomycetes) from Kintrishi Reservation. Mikologiya i Fitopatologiya 33(4):248–256
- Breitenbach J, Kränzlin F (1991) Fungi of Switzerland, vol. 3, Boletes and Agarics 1st part. Mykologia, Lucerne, Switzerland
- Bresadola J (1896) Fungi Tridentini. 195 plates. 1881–1900. Contains figures of *Hydnaceae*. Fungi Brasilienses. Hedwigia 35:276–302
- Bukovská P, Jelínková M, Hřselová H, Sýkorová Z, Gryndler M (2010) Terminal restriction fragment length measurement errors are affected mainly by fragment length, G+C nucleotide content and secondary structure melting point. J Microbiol Methods 82:223–228
- Bussaban B, Lumyong S, Lumyong P, Mckenzie EHC, Hyde KD (2001) A synopsis of the genus *Berkleasmium* with two new species and new records of *Canalisporium caribense* from *Zingiberaceae* in Thailand. Fungal Divers 8:73–85
- Buyck B, Hofstetter V, Eberhardt U, Verbeken A, Kauff F (2008) Walking the thin line between *Russula* and *Lactarius*: the dilemma of *Russula* subsect. *Ochricompactae*. Fungal Divers 28:15–40
- Buyck B, Hofstetter V, Verbeken A, Walleyn R (2010) (1919) Proposal to conserve *Lactarius* nom. cons. (Basidiomycota) with a conserved type. Taxon 59(1):295–296
- Cai L, Jeewon R, Hyde KD (2006) Phylogenetic investigations of *Sordariaceae* based on multiple gene sequences and morphology. Mycol Res 110:137–150
- Calzada Dominguez A (2007) Guía de los Boletos de España y Portugal. Náyade Editorial, Medina del Campo, Valladolid
- Câmara MPS, Palm ME, van Berkum P, O'Neill NR (2002) Molecular phylogeny of *Leptosphaeria* and *Phaeosphaeria*. Mycologia 94:630–640
- Câmara MP, Ramaley AW, Castlebury LA, Palm ME (2003) *Neophaeosphaeria* and *Phaeosphaeriopsis*, segregates of *Paraphaeosphaeria*. Mycol Res 107:516–522
- Cannon PF, Kirk PM (2007) Fungal families world, 7th edn. CAB International, Wallingford
- Cannon PF, Hawksworth DL, Sherwood-Pike MA (1985) The British Ascomycotina. An annotated checklist. Commonwealth Mycological Institute, Kew, Surrey
- Cannon PF, Damm U, Johnston PR, Weir BS (2012) *Colletotrichum* current status and future directions. Study Mycol 73:181–213
- Carbone I, Kohn LM (1999) A method for designing primer sets for speciation studies in filamentous ascomycetes. Mycologia 91:553–556
- Carmichael JW (1966) Cerebral mycetoma of trout due to a *Phialophora*-like fungus. Sabouraudia 6:120–123
- Carmichael JW (1967) Cerebral mycetoma of trout due to a *Phialophora*-like fungus. Sabouraudia. J Med Vet Mycol 5(2):120–123
- Carson ML (2005) Yield loss potential of *Phaeosphaeria* leaf spot of maize caused by *Phaeosphaeria maydis* in the United States. Plant Dis 89:986–988
- Castañeda RF, Heredia G (2000) Two new dematiaceous hyphomycetes on *Cyathea* from Mexico. Cryptogam Mycol 21:221–228
- Castlebury LA, Rossman AY, Jaklitsch WJ, Vasilyeva LN (2002) A preliminary overview of the *Diaporthales* based on large subunit nuclear ribosomal DNA sequences. Mycologia 94:1017–1031
- Chaverri P, Bischoff JF, Evans HC, Hodge KT (2005) *Regiocrella* a new entomopathogenic genus with a pycnidial anamorph and its phylogenetic placement in the *Clavicipitaceae*. Mycologia 97:1225–1237
- Chaverri P, Liu M, Hodge KT (2008) A monograph of the entomopathogenic genera *Hypocrella*, *Moelleriella* and *Samuelsia* gen. nov. (Ascomycota, Hypocreales, Clavicipitaceae) and their *aschersonia*-like anamorphs in the Neotropics. Stud Mycol 60:1–66

- Checa J, Ramaley AW, Palm-Hernández ME, Câmara MP (2002) *Paraphaeosphaeria barrii*, a new species on *Yucca schidigera* from Mexico. *Mycol Res* 106:375–379
- Cheewangkoon R, Crous PW, Hyde KD, Groenewald JZ, To-Anan C (2008) Species of *Mycosphaerella* and related anamorphs on Eucalyptus leaves from Thailand. *Persoonia* 21:77–91
- Chen Q, Jiang JR, Zhang GZ, Cai L, Crous PW (2015) Resolving the *Phoma enigma*. *Stud Mycol* 82:137–217
- Chen Y, Ran SF, Dai DQ, Wang Y, Hyde KD, Wu YM, Jiang YL (2016) Mycosphere Essays 2. *Myrothecium*. *Mycosphere* 7:64–80
- Chen ZH, Zhang P, Zhang ZG (2013) Investigation and analysis of 102 mushroom poisoning cases in southern China from 1994 to 2012. *Fungal Divers* 64:123–131
- Chethana KWT, Phillips AJL, Zhang W, Chen Z, Hao YY, Hyde KD, Li XH, Yan JY (2016) Mycosphere Essays 5: Is it important to name species of *Botryosphaeriaceae*? *Mycosphere* 7:870–882
- Chevallier FF (1826) Flore Générale des Environs de Paris [General Flora of the Area Around Paris]. Ferra Jeune, France, pp 1–674
- Cheyne JL (1983) *Validation de Boletus rubrosanguineus* (Walty). Doc Mycol 52:53–54
- Chlebicki A, Mukhin VA, Ushakowa N (2003) *Fomitopsis officinalis* on Siberian Larch in Urals. *Mycologist* 17:116–120
- Chomnunti P, Schoch CL, Aguirre-Hudson B, KoKo TW, Hongsanan S, Jones EBG, Kodsub R, Chukeatirote E, Bahkali AH, Hyde KD (2011) Capnodiales. *Fungal Divers* 51:103–134
- Chomnunti P, Hongsanan S, Hudson BA, Tian Q, Peršoh D, Dhami MK, Alias AS, Xu J, Liu X, Stadler M, Hyde KD (2014) The sooty moulds. *Fungal Divers* 66:1–36
- Chouhan JS, Panwar KS (1980) Hyphomycetes of Mount Abu-V. Indian Phytopathol 33:285–291
- Clements FE, Shear CL (1931) Genera of fungi, 2nd edn. H.W. Wilson, New York
- Co-David D, Langeveld D, Noordeloos ME (2009) Molecular phylogeny and spore evolution of *Entolomataceae*. *Persoonia* 23:147–176
- Consiglio G, Simonini G (2005) *Trattamento statistico delle dimensioni sporali*. In: Consiglio G, Antonini D, Antonini M (eds) Il Genere *Cortinarius*, parte III. Centro Studi Micologici AMB, pp 33–44
- Corda ACI (1831) Die Pilze Deutschlands. In: Sturm J (ed) Deutschlands Flora in Abbildungen nach der Natur mit Beschreibungen, vol 12(3). Sturm, Nurnberg, pp 33–64
- Corda ACJ (1837) Icones fungorum hucusque cognitorum, vol 1. J.G. Calve, Prague
- Corlett M (1981) A taxonomic survey of some species of *Didymella* and *Didymella*-like species. *Can J Bot* 59:2016–2042
- Corner EJH (1950) A monograph of *Clavaria* and allied genera. Oxford University Press, Oxford
- Corner EJH (1970) Supplement to a monograph of *Clavaria* and allied genera. Beihefte zur Nova Hedwigia 33:1–299
- Corner EJH (1981) The agaric genera *Lentinus*, *Panus*, and *Pleurotus* with particular reference to Malaysian species. *Nova Hedwigia* 69:1–169
- Corner EJH (1996) The agaric genera *Marasmius*, *Chaetocalathus*, *Crinipellis*, *Heimiomyces*, *Resupinatus*, *Xerula* and *Xerulina* in Malesia. Beih Nova Hedwigia 111:1–175
- Corner EJH, Thind KS, Anand GPS (1956) The *Clavariaceae* of the Mussoorie hills (India). II. *Trans Br Mycol Soc* 39(4):475–484
- Courtecuisse R, Duham B (2011) *Guide des Champignons de France et d'Europe*. Delachaux et Niestlé, Paris
- Cripps CL (1997) The genus *Inocybe* in Montana aspen stands. *Mycologia* 89:670–688
- Crous PW, Braun U (2003) *Mycosphaerella* and its anamorphs: 1. Names published in *Cercospora* and *Passalora*. CBS biodiversity series no Centraalbureau voor Schimmelcultures, Utrecht
- Crous PW, Slippers B, Wingfield MJ, Rheeder J, Marasas WFO, Philips AJL, Alves A, Burgess TI, Barber PA, Groenewald JZ (2006) Phylogenetic lineages in the *Botryosphaeriaceae*. *Stud Mycol* 55:235–253
- Crous PW, Braun U, Groenewald JZ (2007) *Mycosphaerella* is polyphyletic. *Stud Mycol* 58:1–3
- Crous PW, Schoch CL, Hyde KD, Wood AR, Gueidan C, de Hoog GS, Groenewald JZ (2009) Phylogenetic lineages in the *Capnodiales*. *Stud Mycol* 64:17–47
- Crous PW, Summerell BA, Shivas RG, Romberg M, Mel'nik VA, Verleyen GJM, Groenewald JZ (2011) Fungal planet description sheets: 92–106. *Persoonia* 27:130–162
- Crous PW, Quaedvlieg W, Sarpkaya K, Can C, Erkiliç A (2013a) *Septoria*-like pathogens causing leaf and fruit spot of pistachio. *IMA Fungus* 4:187–199
- Crous PW, Braun U, Hunter GC, Wingfield MJ, Verleyen GJ, Shin HD, Nakashima C, Groenewald JZ (2013b) Phylogenetic lineages in *Pseudocercospora*. *Stud Mycol* 75:37–114
- Crous PW, Wingfield MJ, Schumacher RK, Summerell BA, Giraldo A, Gené J, Guarro J, Wanasinghe DN, Hyde KD, Camporesi E, Garethjones EB (2014) Fungal planet description sheets: 281–319. *Persoonia* 33:212–289
- Crous PW, Wingfield MJ, Guarro J, Hernández-Restrepo M, Sutton DA, Acharya K, Barber PA, Boekhout T, Dimitrov RA, Dueñas M, Dutta AK (2015a) Fungal planet description sheets: 320–370. *Persoonia* 34:167–266
- Crous PW, Carris LM, Giraldo A, Groenewald JZ, Hawksworth DL, Hernández-Restrepo M, Jaklitsch WM, Lebrun M-H, Schumacher RK, Stielow JB (2015b) The genera of fungi-fixing the application of the type species of generic names-G 2: *Allantophomopsis*, *Latorua*, *Macrodiplodiopsis*, *Macrohylum*, *Milospium*, *Protostegia*, *Pyricularia*, *Robillarda*, *Rotula*, *Septoriella*, *Torula*, and *Wojnowicia*. *IMA Fungus* 6:163–198
- Dai DQ, Phookamsak R, Wijayawardene NN, Li WJ, Bhat DJ, Xu JC, Taylor JE, Hyde KD, Chukeatirote E (2017) Bambusicolous fungi. *Fungal Divers* 82:1–105
- Dai YC (2012) Polypore diversity in China with an annotated checklist of Chinese polypores. *Mycoscience* 53:49–80
- Dai YC, Yang ZL, Cui BK, Yu CJ, Zhou LW (2009) Species diversity and utilization of medicinal mushrooms and fungi in China (review). *Int J Med Mushrooms* 11:287–302
- Dai YC, Xue HJ, Vlasák J, Rajchenberg M, Wang B, Zhou LW (2014) Phylogeny and global diversity of *Polyporus* group *Melanopus* (*Polyporales*, Basidiomycota). *Fungal Diver* 64:133–144
- Damm U, Woudenberg JHC, Cannon PF, Crous PW (2009) *Colletotrichum* species with curved conidia from herbaceous hosts. *Fungal Divers* 39:45–87
- Daranagama DA, Camporesi E, Tian Q, Liu X, Chamuyang S, Stadler M, Hyde KD (2015) *Anthostomella* is polyphyletic comprising several genera in *Xylariaceae*. *Fungal Divers* 73:203–238
- Daranagama DA, Jones EBG, Liu XZ, To-anun C, Stadler M, Hyde KD (2016) Mycosphere Essays 13—do xylariaceous macro-mycetes make up most of the *Xylariomycetidae*? *Mycosphere* 7:582–601
- Das K, Dentinger BTM (2015) *Austroboletus olivaceoglutinosus*, a new mushroom species from Sikkim, India with a distinctive green, glutinous pileus. *Kew Bull* 70:15
- Das K, Hembrom ME, Parikh A, Mishra D, Sharma JR (2014) *Strobilomyces polypyramis*—rediscovery of a wild mushroom from Sikkim, India. *Indian J Plant Sci* 3(2):13–18
- De Gruyter J, Noordeloos ME (1992) Contributions towards a monograph of *Phoma* (Coelomycetes)-I. 1. Section *Phoma*: taxa with very small conidia in vitro. *Persoonia* 15:71–92
- De Gruyter J, Noordeloos ME, Boerema GH (1998) Contributions towards a monograph of *Phoma* (Coelomycetes)—I. 3.

- Section Phoma: taxa with conidia longer than 7 µm. Persoonia 16:471–490
- De Gruyter J, Aveskamp MM, Woudenberg JHC, Verkley GJM, Groenewald JZ, Crous PW (2009) Molecular phylogeny of Phoma and allied anamorph genera: towards a reclassification of the Phoma complex. Mycol Res 113:508–519
- De Gruyter J, Woudenberg JHC, Aveskamp MM, Verkley GJM, Groenewald JZ, Crous PW (2010) Systematic reappraisal of species in *Phoma* section *Paraphoma*, *Pyrenophaeta* and *Pleurophoma*. Mycologia 102:1066–1081
- De Gruyter J, Woudenberg JHC, Aveskamp MM, Verkley GJM, Groenewald JZ, Crous PW (2013) Redisposition of *phoma*-like anamorphs in *Pleosporales*. Stud Mycol 75:1–36
- de Hoog GS, Hermanides-Nijhof EJ (1977) The black yeasts and allied hyphomycetes. Mycologia 69:1242–1244
- de Hoog GS, Vicente VA, Najafzadeh MJ, Harrak MJ, Badali H, Seyedmousavi S (2011) Waterborne *Exophiala* species causing disease in coldblooded animals. Persoonia 27:46–72
- De Notaris G (1844) Cenno sulla tribù dei pirenomiceti sferiacei e descrizione di alcuni nuovi generi. Giornale Botanico Italiano 1:322–335
- de Souza CAF, Lima DX, Gurgel LMS, Santiago ALCM (2016) Coprophilous *Mucorales* (ex Zygomycota) from three areas in the semi-arid of Pernambuco, Brazil. Braz J Microbiol 48(1):1–8
- Dennis RWG (1951) Some tropical American Agaricaceae referred by Berkeley and Montagne to *Marasmius*, *Collybia* or *Heliomyces*. Kew Bull 6(3):387–410
- Desjardin DE (1989) The Genus *Marasmius* from the southern Appalachian Mountains. Doctoral dissertations, University of Tennessee, Knoxville
- Desjardin DE, Retnowati A, Horak E (2000) Agaricales of Indonesia: 2. A preliminary monograph of *Marasmius* from Java and Bali. Sydowia 52:92–193
- Desjardin DE, Wood MG, Stevens FA (2015) California mushrooms. Timber Press, Portland, Oregon
- Diedicke (1912) Kryptogamen-Flora der Mark Brandenburg 9(5):206
- Dissanayake AJ, Camporesi E, Hyde KD, Phillips AJL, Fu CY, Yan JY, Li XH (2016) *Dothiorella* species associated with woody hosts in Italy. Mycosphere 7:51–63
- Dissing H (1966) genus *Helvella* in Europe with special emphasis on the species found in Norden. Dansk Botanisk Arkiv. 25(1):1–172
- Dissing H (1979) *Helvella papuensis*, a new species from Papua New Guinea. Sydowia Annales Mycologici, Beihefte 8:156–161
- Doilom M, Shuttleworth L, Roux J, Chukeatirote E, Hyde KD (2014) *Barriopsis tectonae* sp. nov. a new species of *Botryosphaeriaceae* from *Tectona grandis* (teak) in Thailand. Phytotaxa 176:081–091
- Doilom M, Dissanayake AJ, Wanasinghe DN, Boonmee S, Liu JK, Bhat DJ, Taylor JE, Bahkali AH, McKenzie EHC, Hyde KD (2017) Microfungi on *Tectona grandis* (teak) in Northern Thailand. Fungal Divers 82:107–182
- Donk MA (1949) New and revised nomina generica conservanda proposed for Basidiomycetes (Fungi). Bull Jard Bot 18:83–168
- Donk MA (1964) A conspectus of the families of *Aphyllophorales*. Persoonia Mol Phylogeny Evol Fungi 3(2):199–324
- Doveri F (2004) Fungi fimicoli italici. A.M.B., Vicenza
- Dudka IO, Heluta VP, Tykhonenko YY, Andrianova TV, Hayova VP, Prydiuk MP, Dzhagan VV, Isikov VP (2004) Fungi of the Crimean peninsula. Institute of Botany, National Academy of Sciences of Ukraine, Ukraine
- Ehrenberg CG (1818) Sylvae mycologiae Berolinenses. Bruschke, Berlin, pp 1–32
- Ellis JP (1980) The genus *Morenoina* in Britain. Trans Br Mycol Soc 74:297–307
- Ellis MB (1971) Dematiaceous Hyphomycetes. Commonwealth Mycological Institute, Kew, Surrey
- Ellis MB (1976) More dematiaceous Hyphomycetes. Commonwealth Mycological Institute, Kew, Surrey
- Ellis MB, Ellis JP (1990) Fungi without gills (Hymenomycetes and Gasteromycetes): an identification handbook. Springer, Berlin
- Ellis NB, Ellis JP (1985) Microfungi on land plants—an identification handbook. Macmillan Publishing, New York
- Erikss OE (1967) Arkiv för Botanik 6(4–5):405
- Eriksson OE, Winka K (1998) Families and higher taxa of Ascomycota. Myconet 1(2):17–24
- Ertz D, Heuchert B, Braun U, Freebury CE, Common RS, Diederich P (2016) Contribution to the phylogeny and taxonomy of the genus *Taeniolaella*, with a focus on lichenicolous taxa. Fungal Biol 120:1416–1447
- Estades A, Lannoy G (2004) Les bolets européens. Bull Mycol Bot Dauphiné-Savoie 44(3):3–79
- Farr DF, Rossman AY (2012) Fungal databases, systematic mycology and microbiology laboratory, ARS, USDA. <https://nt.ars-grin.gov/fungaldatabases/>. Retrieved 24 Nov 2016
- Farr DF, Aime MC, Rossman AY, Palm ME (2006) Species of *Colletotrichum* on Agavaceae. Mycol Res 110:1395–1408
- Ferrari BC, Zhang C, van Dorst J (2011) Recovering greater fungal diversity from pristine and diesel fuel contaminated sub-Antarctic soil through cultivation using both a high and a low nutrient media approach. Front Microbiol 2:217
- Fischer E (1897) Plectacineae. In: Engler A, Prantl K (eds) Die natürlichen Pflanzenfamilien, vol I. Engelmann, Leipzig
- Fotouhiifar KB, Hedjaroude GA, Leuchtmann A (2010) ITS rDNA phylogeny of Iranian strains of *Cytospora* and associated teleomorphs. Mycologia 102:1369–1382
- Frank JL (2015) Nomenclatural novelties. Index Fungorum 248:1
- Fries EM (1821) Systema mycologicum, vol. 1. Ex Officina Berlingiana, Lund & Greifswald
- Fries EM (1849) Summa vegetabilium scandinaviae. Part 2. Bonnier, Leipzig
- Frisvad JC, Samson RA (2004) Polyphasic taxonomy of *Penicillium* subgenus *Penicillium*. A guide to identification of food and airborne terverticillate Penicillia and their mycotoxins. Stud Mycol 49:1–174
- Fuckel L (1870) Symbolae mycologicae. Beiträge zur Kenntniss der Rheinischen Pilze. Jb Nassau Ver Naturk 23–24:212
- Fukami T, Dickie IA, Wilkie JP, Paulus BC, Park D, Roberts A, Buchanan PK, Allen RB (2010) Assembly history dictates ecosystem functioning: evidence from wood decomposer communities. Ecol Lett 13:675–684
- Geiser DM, Gueidan C, Miadlikowska J, Lutzoni F, Kauff F, Hofstetter V, Fraker E, Schoch CL, Tibell L, Untereiner WA, Aptroot A (2006) Eurotiomycetes: Eurotiomycetidae and Chaetothyriomycetidae. Mycologia 98:1053–1064
- Gelardi M, Vizzini A, Ercole E, Voyron S, Wu G, Liu XZ (2012) *Strobilomyces echinocephalus* sp. nov. (*Boletales*) from southwestern China, and a key to the genus *Strobilomyces* worldwide. Mycol Prog 12:575–588
- Giambra S, Piazza G, Alves A, Mondello V, Berbegal M, Armengol J, Burruano S (2016) *Botryosphaeriaceae* species associated with diseased loquat trees in Italy and description of *Diplodia rosacearum* sp. nov. Mycosphere 7:978–989
- Gilbertson RL, Ryvarden L (1986) North American polypores. *Abortiporus-Lindneria*. Fungiflora, Oslo
- Gilbertson RL, Ryvarden L (1987) North American polypores: *Megasporoporia-Wrightoporia*. Fungiflora, Oslo
- Gilliam MS (1976) The genus *Marasmius* in the Northeastern United States and adjacent Canada. Mycotaxon 4:1–144
- Glass NL, Donaldson GC (1995) Development of primer sets designed for use with the PCR to amplify conserved genes from filamentous ascomycetes. Appl Environ Microbiol 61:1323–1330

- Góes-Neto A, Loguercio-Leite C, Guerrero RT (2005) DNA extraction from frozen field collected and dehydrated herbarium fungal basidiomata: performance of SDS and CTAB-based methods. *Biотема* 18:19–32
- Haase G, Sonntag L, Melzer-Krick B, de Hoog GS (1999) Phylogenetic inference by SSU-gene analysis of members of the *Herpotrichiellaceae* with special reference to human pathogenic species. *Stud Mycol* 43:80–97
- Häffner J, Stangl J, Sedlmeier A, Geh G, Kriegsteiner GJ (1987) Di Gattung *Helvella*: morphologie und taxonomie. Deutsche Gesellschaft für Mycologie
- Halama M (2016) *Rubroboletus le-galiae* (*Boletales*, Basidiomycota), a species new for Poland. *Acta Mycol* 50(2):1066
- Hall TA (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucl Acid Symp Ser* 41:95–98
- Halling RE, Osmundson TW, Neves MA (2006) *Austroboletus mutabilis* sp. nov. from northern Queensland. *Muelleria* 24:31–36
- Hawksworth DL, Sutton BC, Ainsworth GC (1983) Ainsworth and Bisby's dictionary of the fungi, 7th edn. Commonwealth Mycological Institute, Kew
- Hawksworth DL, Kirk PM, Sutton BC, Pegler DN (1995) Ainsworth & Bisby's dictionary of the fungi, 8th edn. CAB International, Wallingford
- Helaly SE, Richter C, Thongbai B, Hyde KD, Stadler M (2016) Lentinulactam, a hirsutane sesquiterpene with an unprecedented lactam modification. *Tetrahedron Lett* 57(52):5911–5913
- Henkel TW, Meszaros R, Aime MC, Kennedy A (2005) New *Clavulina* species from the Pakaraima Mountains of Guyana. *Mycol Prog* 4:343–350
- Hennings P (1895) Fungi camerunenses I. Botanische Jahrbücher für Systematik, Pflanzengeschichte und Pflanzengeographie 22:72–111
- Hennings P (1905) Fungi japonici VI. Bot Jahrb Syst 37:156–166
- Hesseltine CW, Ellis JJ (1961) Notes on *Mucorales*, especially *Absidia*. *Mycologia* 53:406–426
- Hesseltine CW, Ellis JJ (1964) The genus *Absidia*: *Gongronella* and cylindrical-spored species of *Absidia*. *Mycologia* 56:568–601
- Hirayama K, Tanaka K (2011) Taxonomic revision of *Lophiostoma* and *Lophiotrema* based on reevaluation of morphological characters and molecular analyses. *Mycoscience* 52:401–412
- Hironaga M, Watanabe S, Nishimura K, Miyaji M (1981) Annellated conidiogenous cells in *Exophiala dermatitidis*, agent of phaeohyphomycosis. *Mycologia* 73:1181–1183
- Hofmann TA, Kirschner R, Piepenbring M (2010) Phylogenetic relationships and new records of *Asterinaceae* (Dothideomycetes) from Panama. *Fungal Divers* 43(1):39–53
- Hoffmann K, Pawłowska J, Walther G, Wrzosek M, de Hoog GS, Benny GL, Kirk PM, Voigt K (2013) The family structure of the *Mucorales*: a synoptic revision based on comprehensive multi-gene-genealogies. *Persoonia* 30:57–76
- Högberg N, Holdenrieder O, Stenlid J (1999) Population structure of the wood decay fungus *Fomitopsis spinicola*. *Heredity* 83:354–360
- Höhnel FXR (1919) Fragmente zur Mykologie (XXIII. Mitteilung, Nr. 1154 bis 1188). *Sitzungsber Akad Wiss Wien Math Naturwiss Kl Abt 1* 128:535–625
- Holm L (1957) Nomenclatural notes on Pyrenomycetes. *Taxon* 24:475–488
- Holm L (1961) Taxonomical notes on Ascomycetes. IV. Notes of *Nodulosphaeria* Rhb. *Sven Bot Tidskr* 55:63–80
- Holubová-Jechová V (1987) Studies on Hyphomycetes from Cuba V. Six new species of Dematiaceous Hyphomycetes from Havana Province. *Ceska Mykologie* 41:29–36
- Hongsanan S, Li YM, Liu JK, Hofmann T, Piepenbring M, Bhat JD, Boonmee S, Doilom M, Singtripop C, Tian Q, Mapook A, Zeng XY, Bahkali AH, Xu JC, Mortimer PE, Wu XH, Yang JB, Hyde KD (2014) Revision of genera in *Asterinaceae*. *Fungal Divers* 68:1–68
- Hopple JS, Vilgalys R (1994) Phylogenetic relationships among coprinoid taxa and allies based on data from restriction site mapping of nuclear rDNA. *Mycologia* 86:96–107
- Hopple JS, Vilgalys R (1999) Phylogenetic relationships in the mushroom genus *Coprinus* and dark-spored allies based on sequence data from the nuclear gene coding for the large ribosomal subunit RNA: divergent domains, outgroups, and monophyly. *Mol Phylogenet Evol* 13:1–19
- Horak E (1980a) Indian *Boletales* and *Agaricales*, revisions and new taxa. *Sydowia* 33:80–110
- Horak E (1980b) *Inocybe* (*Agaricales*) in Indomalaya and Australasia. *Persoonia Mol Phylogeny Evol Fungi* 11:1–37
- Horak E (1980c) Supplementary remarks to *Austroboletus* (Corner) Wolfe (*Boletaceae*). *Sydowia* 33:71–87
- Houbraken J, Samson RA (2011) Phylogeny of *Penicillium* and the segregation of *Trichocomaceae* into three families. *Stud Mycol* 70:1–51
- Houbraken J, Wang L, Lee HB, Frisvad JC (2016) New sections in *Penicillium* containing novel species producing patulin, pyripyropens or other bioactive compounds. *Persoonia* 36:299–314
- Hu H, Jeewon R, Zhou D, Zhou T, Hyde KD (2007) Phylogenetic diversity of endophytic *Pestalotiopsis* species in *Pinus armandii* and *Ribes* spp.: evidence from rDNA and β-tubulin gene phylogenies. *Fungal Divers* 24:1–22
- Hughes SJ (1958) Revisiones hyphomycetum aliquot cum appendice de nominibus rejiciendis. *Can J Bot* 36:727–836
- Hughes SJ (1976) Sooty moulds. *Mycologia* 68:693–820
- Humber RA, Rocha LF, Inglis PW, Kipnis A, Luz C (2013) Morphology and molecular taxonomy of *Evlachovaea* like fungi and the status of this unusual conidial genus. *Fungal Biol* 117:1–12
- Hüseyin E, Selçuk F, Churakov BP (2014) A new species of *Berkleasmium* from Ulyanovsk, Russia. *Mycosphere* 5:462–466
- Hyde KD (1995) Tropical Australasian fungi. IX. *Vaginatispora aquatica* gen. et sp. nov. *Nova Hedwigia* 61:233–241
- Hyde KD, Vrijmoed LLP, Chinnaraj S, Jones EBG (1992) *Massarina armatispora* sp. nov., a new intertidal ascomycete from mangroves. *Bot Mar* 35:325–328
- Hyde KD, Cai L, McKenzie EHC, Yang YL, Zhang JZ, Prihastuti H (2009) *Colletotrichum*: a catalogue of confusion. *Fungal Divers* 39:1–17
- Hyde KD, McKenzie EHC, KoKo TW (2011) Towards incorporating anamorphic fungi in a natural classification—checklist and notes for 2010. *Mycosphere* 2:1–88
- Hyde KD, Jones EBG, Liu JK, Ariyawansa H, Boehm E, Boonmee S, Braun U, Chomnunti P, Crous PW, Dai DQ, Diederich P, Dissanayake A, Doilom M, Doveri F, Hongsanan S, Jayawardena R, Lawrey JD, Li YM, Liu YX, Lücking R, Monka J, Muggia L, Nelsen MP, Pang KL, Phookamsak R, Senanayake IC, Shearer CA, Suetrong S, Tanaka K, Thambugala KM, Wijayawardene NN, Wikee S, Wu HX, Zhang Y, Begofia AH, Alias SA, Aptroot A, Bahkali AH, Bezerra JL, Bhat DJ, Camporesi E, Chukea E, Gueidan C, Hawksworth DL, Hirayama K, Hoog SD, Kang JK, Knudsen K, Li WJ, Li XH, Liu ZY, Mapook A, Mckenzie EHC, Miller AN, Mortimer PE, Phillips AJL, Raja HA, Scheuer C, Schumm F, Taylor JE, Tian Q, Tibpromma S, Wanasinghe DN, Wang Y, Xu JC, Yacharoen S, Yan JY, Zang M (2013) Families of dothideomycetes. *Fungal Divers* 63:1–313
- Hyde KD, Nilsson RH, Alias SA, Ariyawansa HA, Blair JE, Cai L, de Cock AWAM, Dissanayake AJ, Glockling SL, Goonasekara ID, Gorczak M, Hahn M, Jayawardena RS, van Kan JAL, Laurence MH, Lévesque CA, Li XH, Liu JK, Maharachchikumbura SSN,

- Manamgoda DS, Martin FN, McKenzie EHC, McTaggart AR, Mortimer PE, Nair PVR, Pawłowska J, Rintoul TL, Shivas RG, Spies CFJ, Summerell BA, Taylor PWJ, Terhem RB, Udayanga D, Vaghefi N, Walther G, Wilk M, Wrzosek M, Xu JC, Yan JY, Zhou N (2014) One stop shop: backbones trees for important phytopathogenic genera: I. *Fungal Divers* 67:21–125
- Hyde KD, Hongsanan S, Jeewon R, Bhat DJ, McKenzie EHC, Jones EBG, Phookamsak R, Ariyawansa HA, Boonmee S, Zhao Q, Abdel-Aziz FA, Abdel-Wahab MA, Banmai S, Chomnunti P, Cui BK, Daranagama DA, Das K, Dayaratne MC, de Silva NI, Dissanayake AJ, Doilom M, Ekanayaka AH, Gibertoni TB, Góes-Neto A, Huang SK, Jayasiri SC, Jayawardena RS, Konta S, Lee HB, Li WJ, Lin CG, Liu JK, Lu YZ, Luo ZL, Manawasinghe IS, Manimohan P, Mapook A, Niskanen T, Norphanphoun C, Papizadeh M, Perera RH, Phukhamsakda C, Richter C, de Santiago ALCMA, Drechsler-Santos ER, Senanayake IC, Tanaka K, Tennakoon TMDS, Thambugala KM, Tian Q, Tibpromma S, Thongbai B, Vizzini A, Wanasinghe DN, Wijayawardene NN, Wu HX, Yang J, Zeng XY, Zhang H, Zhang JF, Bulgakov TS, Camporesi E, Bahkali AH, Amoozegar AM, Araujo-Neta LS, Ammirati JF, Baghela A, Bhatt RP, Bojantchev S, Buyck B, da Silva GA, de Lima CLF, de Oliveira RV, de Souza CAF, Dai YC, Dima B, Duong TT, Ercole E, Mafalda-Freire F, Ghosh A, Hashimoto A, Kamolhan S, Kang JC, Karunarthna SC, Kirk PM, Kytövuori I, Lantier A, Liimatainen K, Liu ZY, Liu XZ, Lücking R, Medardi G, Mortimer PE, Nguyen TTT, Promputtha I, Raj KNA, Reck MA, Lumyong S, Shahzadeh-Fazeli SA, Stadler M, Soudi MR, Su HY, Takahashi T, Tangthirasunun N, Uniyal P, Wang Y, Wen TC, Xu JC, Zhang ZK, Zhao YC, Zhou JZ, Zhu L (2016) Fungal diversity notes 367–490: taxonomic and phylogenetic contributions to fungal taxa. *Fungal Divers* 80:1–270
- Imler L (1950) Recherches sur les bolets. *Bull. Soc. Mycol. Fr.* 66(4):177–203
- Index Fungorum (2017) <http://www.indexfungorum.org/Names/Names.asp>
- Isola D, Selbmann L, de Hoog GS, Fenice M, Onofri S, Prenafeta-Bold FX, Zucconi L (2013) Isolation and screening of black fungi as degraders of volatile aromatic hydrocarbons. *Mycopathologia* 175:369–379
- Iznova T, Rukseniene J (2012) Ascomycetes species new to Lithuania. *Botanica Lithuanica* 18:35–39
- Jaklitsch WM, Voglmayr H (2016) Hidden diversity in *Thyridaria* and a new circumscription of the *Thyridariaceae*. *Stud Mycol* 85:35–64
- Jayasiri SC, Hyde KD, Abd-Elsalam KA, Abdel-Wahab MA, Ariyawansa HA, Bhat J, Buyck B, Dai YC, Ertz D, Hidayat I, Jeewon R, Jones EBG, Karunarthna SC, Kirk P, Lei C, Liu JK, Maharanachikumbura SSN, McKenzie E, Ghobad-Nejjad M, Nilsson H, Pang KL, Phookamsak R, Rollins AW, Romero AI, Stephenson S, Suetrong S, Tsui CKM, Vizzini A, Wen TC, De Silva NI, Promputtha I, Kang JC (2015) The Facesoffungi database: fungal names linked with morphology, molecular and human attributes. *Fungal Divers* 74:3–18
- Jayawardena RS, Hyde KD, Damm U, Cai L, Liu M, Li XH, Zhang W, Zhao WS, Yan JY (2016a) Notes on currently accepted species of *Colletotrichum*. *Mycosphere* 7:1192–1260
- Jayawardena RS, Hyde KD, Jeewon R, Li XH, Liu M, Yan JY (2016b) Mycosphere essay 6: why is it important to correctly name *Colletotrichum* species? *Mycosphere* 7:1076–1092
- Jeewon R, Hyde KD (2016) Establishing species boundaries and new taxa among fungi: recommendations to resolve taxonomic ambiguities. *Mycosphere* 7:1669–1677
- Jeewon R, Liew ECY, Hyde KD (2002) Phylogenetic relationships of *Pestalotiopsis* and allied genera inferred from ribosomal DNA sequences and morphological characters. *Mol Phylogenet Evol* 25:378–392
- Jeewon R, Liew ECY, Hyde KD (2003a) Molecular systematics of the *Amphisphaeriaceae* based on cladistic analyses of partial LSU rDNA gene sequences. *Mycol Res* 107(12):1392–1402
- Jeewon R, Liew EC, Simpson JA, Hodgkiss JJ, Hyde KD (2003b) Phylogenetic significance of morphological characters in the taxonomy of *Pestalotiopsis* species. *Mol Phylogenet Evol* 27(3):372–383
- Jeewon R, Liew ECY, Hyde KD (2004) Phylogenetic evaluation of species nomenclature of *Pestalotiopsis* in relation to host association. *Fungal Divers* 17:39–55
- Jeewon R, Ittoo J, Mahadeb D, Jaufeerally-Fakim Y, Wang HK, Liu AR (2013) DNA based identification and phylogenetic characterisation of endophytic and saprobic Fungi from *Antidesma madagascariense*, a medicinal plant in Mauritius. *J Mycol*. doi:10.1155/2013/781914
- Jian-Zhe Y (1985) Notes on the genus *Austroboletus* in China. *Agarica* 6(12):80–89
- Jiea CY, Zhoua QX, Zhao WS, Jiang YL, Hyde KD, McKenzie EHC, Wang Y (2013) A new *Myrmecridium* species from Guizhou, China. *Mycotaxon* 124:1–8
- Jones EBG, Suetrong S, Sakayaroj J, Bahkali AH, Abdel-Wahab MA, Boekhout T, Pang KL (2015) Classification of marine *Ascomycota*, *Basidiomycota*, *Blastocladiomycota* and *Chytridiomycota*. *Fungal Divers* 73:1–72
- Julou T, Burghardt B, Gebauer G, Bervellier D, Damesin C, Selosse MA (2005) Mixotrophy in orchids: insights from comparative study of green individuals and nonphotosynthetic individuals of *Cephalanthera damasonium*. *New Phytol* 166:639–653
- Kalb K, Buaruang K, Mongkolsuk P, Boonpragob K (2012) New or otherwise interesting lichens. VI, including a lichenicolous fungus. *Phytotaxa* 42:35–47
- Karsten PA (1873) *Mycologia fennica. Pars secunda. Pyrenomycetes. Bidrag till Kändedom av Finlands Natur och Folk* 23:1–252
- Karunarthna SC, Yang ZL, Zhao RL, Vellinga EC, Bahkali AH, Chukeatirote E, Hyde KD (2011) Three new species of *Lentinus* from northern Thailand. *Mycol Prog* 10:389–398
- Katoh K, Misawa K, Kuma K, Miyata T (2002) MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. *Nucleic Acids Res* 30:3059–3066
- Kennedy LL (1958) The genera of the *Dacrymycetaceae*. *Mycologia* 50:874–895
- Kepler R, Ban S, Nakagiri A, Bischoff J, Hywel-Jones N, Owensby CA, Spatafora JW (2013) The phylogenetic placement of hypocrealean insect pathogens in the genus *Polycephalomyces*: an application of One Fungus One Name. *Fungal Biol* 117:611–622
- Kimura M (1980) A simple method for estimating evolutionary rate of base substitutions through comparative studies of nucleotide sequences. *J Mol Evol* 16:111–120
- Kirk PM, Cannon PF, David JC, Stalpers JA (2001) Ainsworth and Bisby's dictionary of the fungi, 8th edn. CABI Publishing, London
- Kirk PM, Cannon PF, Minter DW, Stalpers JA (2008) Ainsworth & Bisby's dictionary of the fungi, 10th edn. CABI, Wallingford
- Kiyashko AA (2012) *Boletus rubrosanguineus* (Walty) ex Cheype, 1983/Red Data Book of Republic of Adygheya. Rare and threatened representatives of the regional fauna and flora. Maykop (in Russian)
- Knudsen H, Vesterholt J (2012) *Funga nordica*. Nordsvamp, Copenhagen
- Kobayashi T (2002) The Taxonomic studies of the genus *Inocybe*. *Beih Nova Hedwigia* 124:1–246
- Kodsueb R, Dhanasekaran V, Aptroot A, Lumyong S, McKenzie EH, Hyde KD, Jeewon R (2006) The family *Pleosporaceae*:

- intergeneric relationships and phylogenetic perspectives based on sequence analyses of partial 28S rDNA. *Mycologia* 98:571–583
- Konta S, Phillips AJL, Bahkali AH, Jones EBG, Eungwanichayapant DP, Hyde KD, Boonmee S (2016a) *Botryosphaeriaceae* from palms in Thailand—*Barriopsis archontophoenicis* sp. nov. from *Archontophoenix alexandrae*. *Mycosphere* 7:921–932
- Konta S, Hongsanan S, Phillips AJ, Jones EBG, Boonmee S, Hyde KD (2016b) *Botryosphaeriaceae* from palms in Thailand II—two new species of *Neodeightonia*, *N. rattanica* and *N. rattanicola* from Calamus (rattan palm). *Mycosphere* 7:950–961
- Kropp BR, Matheny PB, Hutchison LJ (2013) *Inocybe* section *Rimosae* in Utah: phylogenetic affinities and new species. *Mycologia* 105:728–747
- Krüger D, Petersen RH, Hughes KW (2006) Molecular phylogenies and mating study data in *Polyporus* with special emphasis on group “Melanopus” (Basidiomycota). *Mycol Prog* 5:185–206
- Kunze G, Schmidt JC (1817) Mykologische Hefte 1:1–109
- Kuo M, Methven AS (2014) Mushrooms of the Midwest. University of Illinois Press, Urbana
- Lakhanpal TN (1996) Mushrooms of India: *Boletaceae*, vol I. APH Publishing Corporation, New Delhi
- Lannoy G, Estades A (2001) *Les Bolets*. Flore Mycologique d’Europe 6. CRDP, Amiens
- Largent DL (1974) Rhodophylloid fungi of the Pacific coast (United States) IV: infrageneric concepts in *Entoloma*, *Nolanea*, and *Leptonia*. *Mycologia* 66:987–1021
- Larkin MA, Blackshields G, Brown NP, Chenna R, McGettigan PA, McWilliam H, Valentin F, Wallace IM, Wilm A, Lopez R, Thompson JD, Gibson TJ, Higgins DG (2007) Clustal W and Clustal X version 2.0. *Bioinformatics* 23:2947–2948
- Larsson E, Larsson KH (2003) Phylogenetic relationships of russuloid basidiomycetes with emphasis on aphyllophoralean taxa. *Mycologia* 95:1037–1065
- Larsson E, Örstadius L (2008) Fourteen coprophilous species of *Psathyrella* identified in the Nordic countries using morphology and nuclear rDNA sequence data. *Mycol Res* 112:1165–1185
- Larsson E, Ryberg M, Moreau PA, Mathiesen AD, Jacobsson S (2009) Taxonomy and evolutionary relationships within species of section *Rimosae* (*Inocybe*) based on ITS, LSU and mtSSU sequence data. *Persoonia* 23:86–98
- Leão-Ferreira SM, Gusmão LFP, Castañeda Ruiz RF (2013) Conidial fungi from the semi-arid Caatinga biome of Brazil. Three new species and new records. *Nova Hedwigia* 96:479–494
- Lee S, Crous PW, Wingfield MJ (2006) Pestalotioid fungi from Pestiona in the Cape Floral Kingdom. *Stud Mycol* 55:175–187
- Li DM, de Hoog GS, Lindhardt Saunte DM, Gerrits van den Ende AHG, Chen XR (2008) *Coniosporium epidermidis* sp. nov., a new species from human skin. *Stud Mycol* 61:131–136
- Li DM, Li RY, Hoog GD, Wang YX, Wang DL (2009) *Exophiala asiatica*, a new species from a fatal case in China. *Med Mycol* 47(1):101–109
- Li GJ, Li SF, Wen HA (2010) The *Russula* species resource and its economic values of China. *Acta Edulis Fungi* 17(spl):155–160
- Li GJ, Zhao D, Li SF, Yang HJ, Liu XZ (2013a) *Russula changbaiensis* sp. nov. from northeast China. *Mycotaxon* 124(1):269–278
- Li HJ, Han ML, Cui BK (2013b) Two new *Fomitopsis* species from southern China based on morphological and molecular characters. *Mycol Prog* 12:709–718
- Li GJ, Hyde KD, Zhao RN, Hongsanan S, Abdel-Aziz FA, Abdel-Wahab MA, Alvarado P, Alves-Silva G, Ammirati JF, Ariyawansa HA, Baghela A, Bahkali AH, Beug M, Bhat DJ, Bojantchev D, Boonpratuang T, Bulgakov TS, Camporesi E, Boro MC, Ceska O, Chakraborty D, Chen JJ, Chethana KWT, Chomnunti P, Consiglio G, Cui BK, Dai DQ, Dai YC, Daranagama DA, Das K, Dayarathne MC, Crop ED, De Oliveira RV, de Souza CAF, de Souza JI, Dentinger BTM, Dissanayake AJ, Doilom M, Drechsler-Santos ER, Ghobad-Nejad M, Gilmore SP, Góes-Neto A, Gorczak M, Haitjema GH, Hapuarachchi KK, Hashimoto A, He MQ, Henske JK, Hirayama K, Iribarren MJ, Jayasiri SC, Jayawardena RS, Jeon SJ, Jerónimo GH, Jesus AL, Jones EBG, Kang JC, Karunaratna SC, Kirk PM, Konta S, Kuhnert E, Langer E, Lee HS, Lee HB, Li WJ, Li XH, Liimatainen K, Lima DX, Lin CG, Liu JK, Liu XZ, Liu ZY, Luangsa-ard JJ, Lücking R, Lumbsch HT, Lumyong S, Leaño EM, Marano AV, Matsumura M, McKenzie EHC, Mongkolamrit S, Mortimer PE, Nguyen TTT, Niskanen T, Norphanphoun C, O’Malley MA, Parnmen S, Pawłowska J, Perera RH, Phookamsak R, Phukhamsakda C, Pires-Zottarelli CLA, Raspé O, Reck MA, Rocha SCO, de Santiago ALCMA, Senanayake IC, Setti L, Shang QJ, Singh SK, Sir EB, Solomon KV, Song J, Srikitkulchai P, Stadler M, Suetrong S, Takahashi H, Takahashi T, Tanaka K, Tang LP, Thambugala KM, Thanakittipattana D, Theodorou MK, Thongbai B, Thummarukcharoent, Tian Q, Tibpromma S, Verbenek A, Vizzini A, Vlasák J, Voigt K, Wanasinghe DN, Wang Y, Weerakoon G, Wen HA, Wen TC, Wijayawardene NN, Wongkanoun S, Wrzosek M, Xiao YP, Xu JC, Yan JY, Yang J, Yang SD, Hu Y, Zhang JF, Zhao J, Zhou LW, Persoh D, Phillips AJL, Maharachchikumbura SSN (2016) Fungal divers notes 253–366: taxonomic and phylogenetic contributions to fungal taxa. *Fungal Divers* 78:1–237
- Lib Pl (1830) crypt. Arduenna (Liège), fasc. 1(Praef.):8
- Lin CG, McKenzie EHC, Bhat DJ, Ran SF, Chen Y, Hyde KD, Li DW, Wang Y (2016) *Stachybotrys*-like taxa from karst areas and a checklist of stachybotrys-like species from Thailand. *Mycosphere* 7:1273–1291
- Linaldeddu BT, Deidda A, Scanu B, Franceschini A, Alves A, Abdollahzadeh J, Phillips AJL (2016a) Phylogeny, morphology and pathogenicity of *Botryosphaeriaceae*, *Diatrypaceae* and *Gnomoniaceae* associated with branch diseases of hazelnut in Sardinia (Italy). *J Plant Pathol* 146:259–279
- Linaldeddu BT, Alves A, Phillips AJL (2016b) *Sardinella urbana* gen. et sp. nov., a new member of the *Botryosphaeriaceae* isolated from declining *Celtis australis* trees in Sardinian streetscapes. *Mycosphere* 7:893–905
- Linaldeddu BT, Maddau L, Franceschini A, Alves A, Phillips AJL (2016c) *Botryosphaeriaceae* species associated with lenticel dieback in Italy and description of *Diplodia insularis* sp. nov. *Mycosphere* 7:962–977
- Lindau G (1896) Hysteriineae. In: Engler A, Prantl K (eds) *Naturliche Pflanzenfamilien*. I. Teil, I, vol 1. Abteilung, pp 265–278
- Linnaeus CV (1753) Species plantarum, 2 vols. Laurentii Salvii, Holmiae
- Liu B, Fan L, Tao K (1988) Five new species of *Dacrymycetaceae* from China. *Acta Mycol Sin* 7:1–6
- Liu JK, Phookamsak R, Jones EBG, Zhang Y, Ko-Ko TW, Hu HL, Boonmee S, Doilom M, Chukeatirote E, Bahkali AH, Wang Y, Hyde KD (2011) *Astrosphaeriella* is polyphyletic, with species in *Fissuroma* gen. nov., and *Neoastrosphaeriella* gen. nov. *Fungal Divers* 51:135–154
- Liu JK, Phookamsak R, Doilom M, Wiki S, Mei LY, Ariyawansa HA, Boonmee S, Chomnunti P, Dai DQ, Bhat DJ, Romero AI, Xhuang WY, Monkai J, Jones EBG, Chukeatirote E, KoKo TW, Zhao YC, Wang Y, Hyde KD (2012) Towards a natural classification of *Botryosphaerales*. *Fungal Divers* 57:149–210
- Liu JK, Phookamsak R, Dai DQ, Tanaka K, Jones EG, Xu JC, Chukeatirote E, Hyde KD (2014) *Roussellaceae*, a new pleosporalean family to accommodate the genera *Neoroussella* gen. nov., *Roussella* and *Roussellopsis*. *Phytotaxa* 181:1–33
- Liu JK, Hyde KD, Jones EBG, Ariyawansa HA, Bhat DJ, Boonmee S, Maharachchikumbura SSN, McKenzie EHC, Phookamsak R,

- Phukhamsakda C, Shenoy BD, Abdel-Wahab MA, Buyck B, Chen J, Chethana KWT, Singtripop C, Dai DQ, Dai YC, Daranagama DA, Dissanayake AJ, Doilom M, Dsouza MJ, Fan XL, Goonasekara ID, Hirayama K, Hongsanan S, Jayasiri SC, Jayawardena RS, Karunaratna SC, Li WJ, Mapook A, Norphanphoun C, Pang KL, Perera RH, Peršoh D, Pinruan U, Senanayake IC, Somrithipol S, Suetrong S, Tanaka K, Thambugala KM, Tian Q, Tibpromma S, Udayanga D, Wijayawardene NN, Wanasinghe DN, Wisitrasameewong K, Zeng XY, Abdel-Aziz FA, Adamčík S, Bahkali AH, Boonyuen N, Bulgakov T, Callac P, Chommunti P, Greiner K, Hashimoto A, Hofstetter V, Kang JC, Lewis D, Li XH, Liu XZ, Liu ZY, Matsumura M, Mortimer PE, Rambold G, Randrianjohany E, Sato G, Sri-Indrasutdi V, Tian CM, Verbeken A, von Brackel W, Wang Y, Wen TC, Xu JC, Yan JY, Zhao RL, Camporesi E (2015) Fungal Divers notes 1–110: taxonomic and phylogenetic contributions to fungal species. *Fungal Divers* 72:1–197
- Liu M, Chaverri P, Hodge KT (2006) A taxonomic revision of the insect biocontrol fungus *Aschersonia aleyrodis*, its allies with white stromata and their *Hypocrella* sexual states. *Mycol Res* 110:537–554
- Liu NG, Ariyawansa HA, Hyde KD, Maharachchikumbura SSN, Zhao RL, Phillips AJL, Jayawardena RS, Thambugala KM, Dissanayake AJ, Wijayawardene NN, Liu JK, Liu ZY, Jeewon R, Jones EBG, Jumpatong J (2016) Perspectives into the value of genera, families and orders in classification. *Mycosphere* 7:1649–1668
- Liu Y, Steenkamp ET, Brinkmann H, Forget L, Philippe H, Lang BF (2009) Phylogenomic analyses predict sister group relationship of nucleariids and fungi and paraphyly of zygomycetes with significant support. *BMC Evol Biol* 9:272
- Liu YJ, Whelen S, Hall BD (1999) Phylogenetic relationships among ascomycetes: evidence from an RNA polymerase II subunit. *Mol Biol Evol* 16:1799–1808
- Liu ZY, Liang ZQ, Liu AY, Yao YJ, Hyde KD, Yu ZN (2002) Molecular evidence for teleomorph anamorph connections in *Cordyceps* based on ITS-5.8S rDNA sequences. *Mycol Res* 106:1100–1108
- Locquin M (1984) Mycologie Générale et Structurale. Masson, Paris
- Lombard L, Houbraken J, Decock C, Samson RA, Meijer M, Réblová M, Groenewald JZ, Crous PW (2016) Generic hyper-diversity in *Stachybotriaceae*. *Persoonia* 36:156–246
- Luangsa-ard JJ, Hywel-Jones NL, Manoch L, Samson RA (2005) On the relationships of *Paecilomyces* sect. *Isarioidea* species. *Mycol Res* 109:581–589
- Lumbsch HT, Huhndorf SM (2007) Outline of Ascomycota. *Myconet* 13:1–99
- Lumbsch HT, Huhndorf SM (2010) Myconet volume 14. Part one. Outline of Ascomycota 2009. Part Two. Notes on Ascomycete systematics. Nos. 4751–5113. *Fieldiana Life Earth Sci* 1:1–64
- Maas JL (1998) Compendium of strawberry diseases, 2nd edn. APS Press, St Paul, pp 1–98
- Maharachchikumbura SSN, Guo LD, Cai L, Chukeatirote E, Wu WP, Sun X, Crous PW, Bhat DJ, McKenzie EHC, Bahkali AH, Hyde KD (2012) A multi-locus backbone tree for *Pestalotiopsis*, with a polyphasic characterization of 14 new species. *Fungal Divers* 56:95–129
- Maharachchikumbura SSN, Guo L-D, Chukeatirote E, Hyde KD (2014a) Improving the backbone tree for the genus *Pestalotiopsis*; addition of *P. steyaertii* and *P. magna* sp. nov. *Mycol Prog* 3:617–624
- Maharachchikumbura SSN, Hyde KD, Groenewald JZ, Xu J, Crous PW (2014b) *Pestalotiopsis* revisited. *Stud Mycol* 79:121–186
- Maharachchikumbura SSN, Hyde KD, Jones EBG, McKenzie EHC, Huang S-K, Abdel-Wahab MA, Daranagama DA, Dayarathne M, D'souza MJ, Goonasekara ID, Hongsanan S, Jayawardena RS, Kirk PM, Konta S, Liu J-K, Liu Z-Y, Norphanphoun C, Shenoy BD, Xiao Y, Bahkali AH, Kang J, Somrithipol S, Suetrong S, Wen T, Xu J (2015) Towards a natural classification and backbone tree for Sordariomycetes. *Fungal Divers* 72:199–301
- Maharachchikumbura SSN, Hyde KD, Jones EBG, McKenzie EHC, Bhat DJ, Dayarathne MC, Huang SK, Norphanphoun C, Senanayake IC, Perera RH, Shang QJ, Xiao YP, D'souza MJ, Hongsanan S, Jayawardena RS, Daranagama DA, Konta S, Goonasekara ID, Zhuang WY, Jeewon R, Phillips AJL, Abdel-Wahab MA, Al-Sadi AM, Bahkali AH, Boonmee S, Boonyuen N, Cheewangkoon R, Dissanayake AJ, Kang JC, Li QR, Liu JK, Liu XZ, Liu ZY, Luangsa-ard JJ, Pang KL, Phookamsak R, Promputtha I, Suetrong S, Stadler M, Wen TC, Wijayawardene NN (2016) Families of Sordariomycetes. *Fungal Divers* 79:1–317
- Maharachchikumbura SS, Larignon P, Al-Sadi AM, Zuo-Yi LIU (2017) Characterization of *Neopestalotiopsis*, *Pestalotiopsis* and *Truncatella* species associated with grapevine trunk diseases in France. *Phytopathol Mediterr* 55(3):380–390
- Manawasinghe IS, Phillips AJL, Hyde KD, Chethana KWT, Zhang W, Zhao WS, Yan JY, Li XH (2016) Mycosphere Essays 14: assessing the aggressiveness of plant pathogenic *Botryosphaeriaceae*. *Mycosphere* 7(7):883–892
- Manimohan P, Leelavathy KM (1989) *Marasmius* species new to India. *Sydowia* 41:185–199
- Manimohan P, Joseph AV, Leelavathy KM (1995) The genus *Entoloma* (*Agaricales*) in Kerala State, India. *Mycol Res* 99:1083–1097
- Mantle PG, Hawksworth DL, Pazoutova S, Collinson LM, Rassing BR (2006) *Amorosia littoralis* gen. sp. nov., a new genus and species name for the scorpione and caffeine-producing hyphomycete from the littoral zone in The Bahamas. *Mycol Res* 110(12):1371–1378
- Mapook A, Boonmee S, Ariyawansa HA, Tibpromma S, Campesori E, Jones EG, Bahkali AH, Hyde KD (2016) Taxonomic and phylogenetic placement of *Nodulosphaeria*. *Mycol Prog* 15(4):1–15
- Martin R, Gazis R, Skaltsas D, Chaverri P, Hibbett D (2015) Unexpected diversity of basidiomycetous endophytes in sapwood and leaves of *Hevea*. *Mycologia* 107(2):284–297
- Matheny PB (2005) Improving phylogenetic inference of mushrooms with RPB1 and RPB2 nucleotide sequences (*Inocybe*; *Agaricales*). *Mol Phylogenet Evol* 35:1–20
- Matheny PB (2009) A phylogenetic classification of the *Inocybaceae*. *McIlvainea* 18:11–21
- Matheny PB, Bouger NL (2006a) The new genus *Auritella* from Africa and Australia (*Inocybaceae*, *Agaricales*): molecular systematics, taxonomy and historical biogeography. *Mycol Prog* 5:2–17
- Matheny PB, Bouger NL (2006b) Validation of *Auritella*. *Mycotaxon* 98:231–233
- Matsushima T (1981) Matsushima Mycological Memoirs 2. Matsushima, Kobe, Japan
- Matsushima T (1983) Matsushima Mycological Memoirs 3. Matsushima, Kobe
- Mazzer SJ (1976) A monographic study of the genus *Pouzarella*, a new genus in the *Rhodophyllaceae*, *Agaricales*, Basidiomycetes. *Bibliotheca Mycologica* 46:1–191
- McNabb RFR (1965) Taxonomic studies on the *Dacrymycetaceae*. VI. *Femsjonia Fries*. *NZ J Bot* 3(3):223–228
- McNabb RFR (1973) Taxonomic studies in the *Dacrymycetaceae*. VIII. *Dacrymyces* Nees ex Fries. *NZ J Bot* 11(3):461–524
- Miller MA, Pfeiffer W, Schwartz T (2010) Creating the CIPRES Science Gateway for inference of large phylogenetic trees. In: Proceedings of the gateway computing environments workshop

- (GCE). Institute of Electrical and Electronics Engineers, New Orleans, LA, 14 Nov, pp 1–8
- Miller SL, Buyck B (2002) Molecular phylogeny of the genus *Russula* in Europe with a comparison of modern infrageneric classifications. *Mycol Res* 106:259–276
- Miller SL, Larsson E, Larsson KH, Verbeken A, Nuytinck J (2006) Perspectives in the new Russulales. *Mycologia* 98:960–970
- Minnis AM, Kennedy AH, Grenier DB, Palm ME, Rossman AY (2012) Phylogeny and taxonomic revision of the *Planistromelaceae* including its coelomycetous anamorphs: contributions towards a monograph of the genus *Kellermania*. *Persoonia* 29:11–28
- Mirza F (1968) Taxonomic investigations on the ascomycetous genus *Cucurbitaria*. *Nova Hedwigia* 16:161–213
- Miyake I (1909) Studies on the parasitic fungi of rice in Japan. *Bot Mag Tokyo* 23:85–97
- Mohanam C (2011) Macrofungi of Kerala. KFRI handbook no. 27, Kerala Forest Research Institute, Peechi
- Moncalvo JM, Vilgalys R, Redhead SA, Johnson JE, James TY, Aime CM, Hofsetter V, Verduin SJ, Larsson E, Baroni TJ, Thorn GR, Jacobsson S, Clémencón H, Miller OK (2002) One hundred and seventeen clades of euagarics. *Mol Phylogenetic Evol* 23:357–400
- Moncalvo JM, Nilsson RH, Koster B, Dunham SM, Bernauer T, Matheny PB, McLenon T, Margaritescu S, Weiß M, Garnica S, Danell E, Langer G, Langer E, Larsson E, Larsson KH, Vilgalys R (2006) The cantharelloid clade: dealing with incongruent gene trees and phylogenetic reconstruction methods. *Mycologia* 98:937–948
- Mongkolsamrit S, Luangsa-Ard JJ, Spatafora JW, Sung GH, Hywel-Jones NL (2009) A combined ITS rDNA and β-tubulin phylogeny of Thai species of *Hypocrella* with non-fragmenting ascospores. *Mycol Res* 113(6):684–699
- Monod M (1983) Monographie taxonomique des *Gnomoniaceae* (Ascomycètes de l'ordre des Diaporthales I). Beihefte Sydowia 9:1–315
- Moreau PA, Vila J, Aime MC, Antonín V, Horak E, Pérez-Butrón JL, Richard F, Urban A, Welti S, Vizzini A (2015) *Cibaomyces* and *Cyptotrama*, two new genera for Europe, and an emendation of *Rhizomarasmius* (Basidiomycota, Physalacriaceae). *Mycol Prog* 14(2):1–16
- Moore RT (1958) Deuteromycetes I: the complex. *Mycologia* 50:681–692
- Morgado LN, Noordeloos ME, Lamoureux Y, Geml J (2013) Multi-gene phylogenetic analyses reveal species limits, phyogeographic patterns, and evolutionary histories of key morphological traits in *Entoloma* (Agaricales, Basidiomycota). *Persoonia Mol Phylogeny Evol Fungi* 31:159–178
- Morris EF (1972) Costa Rican Hyphomycetes. *Mycologia* 64:887–896
- Mugambi GK, Huhndorf SM (2009) Molecular phylogenetics of *Pleosporales*: *Melanommataceae* and *Lophiostomataceae* re-circumscribed (Pleosporomycetidae, Dothideomycetes, Ascomycota). *Stud Mycol* 64:103–121
- Mulenko W, Majewski T, Ruszkiewicz-Michalska M (2008) A preliminary checklist of micromycetes in Poland. W. Szafer Institute of Botany, Polish Academy of Sciences, Krakow
- Munk A (1953) The system of the Pyrenomycetes. A contribution to a natural classification of the group *Sphaeriales* sensu Lindau. *Dansk Botanisk Arkiv* 15:1–163
- Munk A (1957) Danish Pyrenomycetes. *Dan Bot Ark* 17:1–491
- Muñoz JA (2005) *Boletus* s.l. (excl. *Xerocomus*). *Fungi Europaei* 2. Edizioni Candusso, Alassio
- Murrill WA (1909) The *Boletaceae* of North America: I. *Mycologia* 1(1):4–18
- MycoBank (2017) <http://www.mycobank.org/quicksearch.aspx>
- Najafzadeh MJ, Dolatabadi S, Keisari MS, Naseri A, Feng P, De Hoog GS (2013) Detection and identification of opportunistic *Exophiala* species using the rolling circle amplification of ribosomal internal transcribed spacers. *J Microbiol Methods* 94:338–342
- Nannf (1932) Nova Acta Regiae Societatis Scientiarum Upsaliensis 8(2):66
- Natarajan K, Manjula B (1982) South Indian Agaricales: XI. *Mycologia* 74:130–137
- Neubert K, Mendgen K, Brinkmann H, Wirsel SGR (2006) Only a few fungal species dominate highly diverse mycofloras associated with the common reed. *Appl Environ Microbiol* 72:1118–1128
- Nguyen TTT, Lee SH, Bae S, Jeong SJ, Mun HY, Lee HB (2016) Characterization of two new records of zygomycete species belonging to undiscovered taxa in Korea. *Mycobiology* 44:29–37
- Noordeloos ME (1981) *Entoloma* subgenera *Entoloma* and *Allocybe* in the Netherlands and adjacent regions with a reconnaissance of its remaining taxa in Europe. *Persoonia* 11:153–256
- Noordeloos ME (1992) *Entoloma* s.l. *Fungi Europaei*, vol. 5. Giovanna Biella, Italy
- Noordeloos ME (2004) *Entoloma* s.l. *Fungi Europaei*, vol 5a. Edizione Candusso, Alassio
- Noordeloos ME (2010) Hoe raak ik thuis in de boleten—8. De roodporieboleten (sectie *Luridi*). *Coolia* 53(2):53–71
- Noordeloos ME, Gates GM (2012) The *Entolomataceae* of Tasmania. Fungal diversity research series, vol 22. Springer, London
- Norphanphon C, Doilom M, Daranagama DA, Phookamsak R, Wen TC, Bulgakov TS, Hyde KD (2017) Revisiting the genus *Cytiospora* and allied species. *Mycosphere* 8:51–97
- Núñez M, Ryvarden L (1995) *Polyporus* (Basidiomycotina) and related genera. *Synop Fungorum* 10:1–85
- Nylander JAA, Wilgenbusch JC, Warren DL, Swofford DL (2008) AWTY (are we there yet?): a system for graphical exploration of MCMC convergence in Bayesian phylogenetics. *Bioinformatics* 24:581–583
- Oberwinkler F (1993) Diversity and phylogenetic importance of tropical heterobasidiomycetes. *Br Mycol Soc Symp* 19:121
- O'Donnell K (1993) *Fusarium* and its near relatives. Wallingford, UK
- Olariaga I, Begoña MJ, García-Etxabarria K, Salcedo I (2009) Species delimitation in the European species of *Clavulina* (*Cantharellales*, Basidiomycota) inferred from phylogenetic analyses of ITS region and morphological data. *Mycol Res* 113:1261–1270
- Oliveira RV, Lima TEF, Cunha IB, Coimbra VRM, Silva GA, Bezerra JL, Cavalcanti MAQ (2014) *Corniculariella brasiliensis*, a new species of coelomycetes in the rhizosphere of *Caesalpinia echinata* (Fabaceae, Caesalpinoideae) in Brazil. *Phytotaxa* 178(3):197–204
- Örstadius L, Ryberg M, Larsson E (2015) Molecular phylogenetics and taxonomy in *Psathyrellaceae* (Agaricales) with focus on psathyelloid species: introduction of three new genera and 18 new species. *Mycol Prog* 14:25
- Ortiz-Santana B, Lindner DL, Miettinen O, Justo A, Hibbett DS (2013) A phylogenetic overview of the *Antrodia* clade (Basidiomycota, *Polyporales*). *Mycologia* 105:1391–1411
- Orton PD, Watling R (1979) *Coprinaceae* 1: *Coprinus*. British Fungus Flora. Agarics and Boleti 2. Royal Botanic Garden, Edinburgh
- Padamsee M, Matheny PB, Dentinger BT, McLaughlin DJ (2008) The mushroom family *Psathyrellaceae*: evidence for large-scale polyphyly of the genus *Psathyrella*. *Mol Phylogenetic Evol* 46:415–429
- Page RDM (2001) Tree View: tree drawing software for Apple Macintosh and Windows. <http://taxonomy.zoology.gla.ac.uk/rod/treeview.html>

- Palazon FL (2006) *Setas para todos*. Pirineo, Huesca
- Pantidou ME (1973) Fungus-host index for Greece. Benaki Phytopathol. Inst., Kiphissia
- Peever TL, Barve MP, Stone LJ, Kaiser WJ (2007) Evolutionary relationships among *Ascochyta* species infecting wild and cultivated hosts in the legume tribes *Ciceraceae* and *Vicieae*. *Mycologia* 99:59–77
- Pegler DN (1972) A revision of the genus *Lepiota* from Ceylon. *Kew Bull* 27:155–202
- Pegler DN (1977) A preliminary agaric flora of East Africa. *Kew Bull Addit Ser* 6:1–615
- Pegler DN (1983) The genus *Lentinus*: a world monograph. *Kew Bull Addit Ser* 10:1–281
- Pegler DN (1986) Agaric flora of Sri Lanka. *Kew Bull Addit Ser* 12:1–519
- Peintner U, Knapp M, Fleischer V, Walch G, Dresch P (2016) *Myrmecridium hiemale* sp nov from snow-covered alpine soil is the first eurypsychrophile in this genus of anamorphic fungi. *Int J Syst Evol Microbiol* 66(7):2592–2598
- Persoon CH (1797) Tentamen dispositionis methodicae fungorum in classes, ordines genera et familias. *Lipsiae*
- Petch T (1948) A revision of the Ceylon Marasmii. *Trans Br Mycol Soc* 31:19–44
- Petersen RH (1983) Notes on clavarioid fungi XVIII. A preliminary outline of *Clavulina* in southeastern Australia. *Nova Hedwigia* 37:19–35
- Petersen RH (1988) The clavarioid fungi of New Zealand. *Bull NZ Dep Ind Res* 236:1–170
- Petersen RH, Hughes KW, Adamčík S, Tkalc̆ec Z, Mešić A (2012) Typification of three European species epithets attributable to *Strobilomyces* (*Boletales*). *Czech Mycol* 64(2):141–163
- Peterson SW (2000a) Phylogenetic analysis of *Penicillium* species based on ITS and LSU-rDNA nucleotide sequences. In: Samson RA, Pitt JI (eds) Integration of modern taxonomic methods for *Penicillium* and *Aspergillus* classification. Plenum Press, New York, pp 163–178
- Peterson SW (2000b) Phylogenetic relationships in *Aspergillus* based on rDNA sequence analysis. In: Samson RA, Pitt JI (eds) Integration of modern taxonomic methods for *Penicillium* and *Aspergillus* classification. Plenum Press, New York, pp 323–355
- Peterson SW (2008) Phylogenetic analysis of *Aspergillus* species using DNA sequences from four loci. *Mycologia* 100:205–226
- Petrak F, Deighton FC (1952) Beiträge zur Pilzflora von Sierra Leone. *Syndowia* 6:309–322
- Petrini LE (2013) *Rosellinia*—a world monograph. *Bibl Mycol* 205:1–410
- Peyronel B, Dal Vesco G (1955) Ricerche sulla microflora di un terreno agrario presso Torino. *Allionia* 2:357–417
- Phillips AJL, Alves A, Pennycook SR, Johnston PR, Ramaley A, Akulov A, Crous PW (2008) Resolving the phylogenetic and taxonomic status of dark-spored teleomorph genera in the *Botryosphaeriaceae*. *Persoonia Mol Phylogeny Evol Fungi* 21:29–55
- Phillips AJL, Alves A, Abdollahzadeh J, Slippers B, Wingfield MJ, Groenewald JZ, Crous PW (2013) The *Botryosphaeriaceae*: genera and species known from culture. *Stud Mycol* 76:51–167
- Phookamsak R, Liu JK, McKenzie EHC, Manamgoda DS, Ariyawansa HA, Thambugala KM, Dai DQ, Camporesi E, Chukeatirote E, Wijayawardene NN, Bahkali AH, Mortimer PE, Xu JC, Hyde KD (2014) Revision of *Phaeosphaeriaceae*. *Fungal Divers* 68:159–238
- Phookamsak R, Norphanphoun C, Tanaka K, Dai DQ, Luo ZL, Liu JK, Su HY, Bhat DJ, Bahkali AH, Mortimer PE, Xu JC, Hyde KD (2015) Towards a natural classification of *Astrophaeliellaceae* like species; introducing *Astrophaeliellaceae* and *Pseudoastrophaeliellaceae* fam. nov and *Astrophaeliellaceopsis*, gen. nov. *Fungal Divers* 74:143–197
- Phukhamsakda C, Hongsanan S, Ryberg M, Ariyawansa HA, Chomnunti P, Bahkali AH, Hyde KD (2016) The evolution of *Massarinae* with *Longipedicellataceae* fam. nov. *Mycosphere* 7:1713–1731
- Pici G (1955) Qualche osservazione sopra due *Mucoraceae*. *Atti Inst Bot Univ Pavia* 13:38–44
- Pinnoli A, Jeewon R, Sakayaroj J, Hyde KD, Jones EG (2007) *Berkleasmium crunisia* sp. nov. and its phylogenetic affinities to the *Pleosporales* based on 18S and 28S rDNA sequence analyses. *Mycologia* 99:378–384
- Pitt JI (1980) The genus *Penicillium* and its teleomorphic states *Eupenicillium* and *Talaromyces*. Academic Press, London
- Pöldmaa K (2011) Tropical species of *Cladobotryum* and *Hypomyces* producing red pigments. *Stud Mycol* 31(68):1–34
- Pradeep CK, Vrinda KB (2010) Ectomycorrhizal fungal diversity in three different forest types and their association with endemic, indigenous and exotic species in the Western Ghats forests of Thiruvananthapuram district, Kerala. *J Mycopathol Res* 48:279–289
- Pratibha J, Prabhugaonkar A, Hyde KD, Bhat DJ (2014) Phylogenetic placement of *Bahusandhika*, *Cancellidium* and *Pseudoepicoccum* (aseexual Ascomycota). *Phytotaxa* 176:68–80
- Promputtha I, Lumyong S, Dhanasekaran V, McKenzie EHC, Hyde KD, Jeewon R (2007) A phylogenetic evaluation of whether endophytes become saprotrophs at host senescence. *Microb Ecol* 53(4):579–590
- Qin J, Yang ZL (2016) *Cyptotrama* (*Physalacriaceae*, *Agaricales*) from Asia. *Fungal Biol* 120:513–529
- Qu C, Yin G, Zhao G, Cui B, Liu X (2014) Three new species of *Berkleasmium* (*Hyphomycetes*) from China. *Nova Hedwigia* 98:151–161
- Quaedvlieg W, Verleyen GJM, Shin H-D, Barreto RW, Alfenas AC, Swart WJ, Groenewald JZ, Crous PW (2013) Sizing up *Septoria*. *Stud Mycol* 75:307–390
- Rabenhorst (1858) Herb. Myc. Ed., 2:725
- Rambaut A (2014) FigTree v1.4: tree figure drawing tool. <http://treebio.ed.ac.uk/software/figtree/>
- Rambaut A, Drummond AJ (2003) Tracer: MCMC trace analysis tool. University of Oxford, Oxford
- Rannala B, Yang Z (1996) Probability distribution of molecular evolutionary trees: a new method of phylogenetic inference. *J Mol Evol* 43:304–311
- Rao PR, Rao D (1963) *Berkleasmium zobel* from India. *Mycopathologia* 22:311–314
- Read SJ, Jones EBG, Moss ST (1997) Ultrastructural observation of asci, ascospores and appendages of *Massarina armatispora* (Ascomycota). *Mycoscience* 38:141–146
- Réblová M, Gams W, Seifert KA (2011) *Monilochaetes* and allied genera of the *Glomerellales*, and a reconsideration of families in the *Microascales*. *Stud Mycol* 68:163–191
- Réblová M, Fournier J, Štěpánek V (2016) Two new lineages of aquatic ascomycetes: *Atractospora* gen. nov. and *Rubellisiaphaeria* gen. et sp. nov., and a sexual morph of *Myrmecridium montsegurinum* sp. nov. *Mycol Prog* 15(3):1–18
- Redhead SA, Ginns J (1980) *Cyptotrama asprata* (Agaricales) from North America and notes on the five other species of *Cyptotrama* sect. *Xerulina*. *Can J Bot* 58(6):731–740
- Redhead SA, Ginns JH (1985) A reappraisal of agaric genera associated with brown rots of wood. *Trans Mycol Soc Jpn* 26:349–381
- Redhead SA, Vilgalys R, Moncalvo JM, Johnson J, Hopple JS Jr (2001) *Coprinus* Pers. and the disposition of *Coprinus* species sensu lato. *Taxon* 50:203–241

- Rehner SA (2001) Primers for elongation factor 1-alpha (EF1-alpha). <http://www.aftol.org/pdfs/EF1primer.pdf>
- Rehner SA, Samuels GJ (1995) Molecular systematics of the *Hypocreales*: a teleomorph gene phylogeny and the status of their anamorphs. *Can J Bot* 31(73):816–823
- Ribaldi MS (1952) Sopra un interessante Zigomicete terricolo: *Gongronella urceolifera* n. gen. et n. sp. *Riv Biol* 49:157–166
- Rogerson CT (1970) The hypocrealean fungi (ascomycetes, *Hypocreales*). *Mycologia* 1:865–910
- Romagnesi H (1967) Les Russules d'Europe et d'Afrique du Nord. Bordas, Paris
- Romagnesi H (1978) les fondements de la taxonomie des rhodophylles et leur classification. *Nova Hedwigia* 59:1–80
- Ronquist F, Huelsenbeck JP (2003) MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19:1572–1574
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Syst Biol* 61:539–542
- Rossmann AY, Farr DF, Castlebury LA (2007) A review of the phylogeny and biology of the *Diaporthales*. *Mycoscience* 48:135–144
- Rossmann AY, Crous PW, Hyde KD, Hawksworth DL, Aptroot A, Bezerra JL, Bhat JD, Boehm E, Braun U, Boonmee S, Camporesi E, Chomnunti P, Dai DQ, D'souza MJ, Dissanayake A, Jones EBG, Groenewald JZ, Hernández-Restrepo M, Hongsanan S, Jaklitsch WM, Jayawardena R, Li WJ, Kirk PM, Lawrey JD, Mapook A, McKenzie EHC, Monkai J, Phillips AJL, Phookamsak R, Raja HA, Seifert KA, Senanayake IC, Slippers B, Suetrong S, Taylor JE, Thambugala KM, Tian Q, Tibpromma S, Wanasinghe DN, Wijayawardene NN, Wikee S, Woudenberg JHC, Wu HX, Yan J, Yang T, Zhang Y (2015) Recommended names for pleomorphic genera in *Dothideomycetes*. *IMA Fungus* 6:507–523
- Ruiz Fernandez JM, Ruiz Pastor E (2006) Guia Micologica Tomo n. 4, Supl. Orden Boletales en España, Imagen. Bilbao: 62
- Rungjindamai N, Pinruan U, Choeyklin R, Hattori T, Jones EBG (2008) Molecular characterization of basidiomycetous endophytes isolated from leaves, rachis and petioles of the oil palm, *Elaeis guineensis*, in Thailand. *Fungal Divers* 33:139–161
- Ryvarden L (1991) Genera of polypores, nomenclature and taxonomy. *Synop Fungorum* 5:1–363
- Ryvarden L, Gilbertson RE (1984) *Mycotaxon* 19:141
- Ryvarden L, Gilbertson RL (1993) European polypores 1. *Abortiporus-Lindneria*. *Synop Fungorum* 6:1–387
- Ryvarden L, Gilvertson RL (1994) European polypores, part 2. Fungiflora, Oslo
- Saccardo PA (1878) *Fungi Veneti novi vel critici vel mycologiae Venetae addendi*. Ser. IX. *Michelia* 1(4):407
- Saccardo PA (1880) *Conspectus generum fungorum italiae inferioriorum nempe ad Sphaeropsideas, Melanconieas et Hyphomyceteas pertinentium, sistente sporologico dispositu*. *Michelia* 2:1–38
- Saccardo PA (1882) *Fungi Gallici lecti a Cl. viris P. Brunaud, C.C. Gillet, Abb. Letendre, A. Malbranche, J. Therry & Dom. Libert*. Series IV. *Michelia* 2(8):583–648
- Saccardo PA (1892) *Supplementum Universale, Pars II. Discomycetaceae-Hyphomycetaceae*. *Sylloge Fungorum* 10:1–964
- Saccardo PA (1899) *Sylloge fungorum* 14:1–1316
- Samson RA, Seifert KA, Kuijpers AFA, Houbraken JAMP, Frisvad JC (2004) Phylogenetic analysis of *Penicillium* subgenus *Penicillium* using partial β-tubulin sequences. *Stud Mycol* 49:175–200
- Samson RA, Houbraken J, Varga J, Frisvad JC (2009) Polyphasic taxonomy of the heat resistant ascomycete genus *Byssochlamys* and its *Paecilomyces* anamorphs. *Persoonia* 22:14–27
- Samson RA, Houbraken J, Thrane U (2010) Food and indoor fungi. CBS KNAW Biodiversity Center, Utrecht
- Santiago ALCMA, Santos PJP, Maia LC (2013) *Mucorales* from the semiarid of Pernambuco, Brazil. *Braz J Microbiol* 44:299–305
- Sarnari M (1998) Monografia illustrate de genere *Russula* in Europa. Tomo Primo. AMB, Centro Studi Micologici, Trento
- Sarnari M (2005) Monografia illustrate de genere *Russula* in Europa. Tomo Secondo. AMB, Centro Studi Micologici, Trento
- Sarycheva LA, Svetasheva TY (2015) *Boletus legaliae* Pilát//Red Data Book of Lipetsk region. Plants, fungi, lichenes, 2nd edn. Lipetsk:536–537 (in Russian)
- Sato H, Hattori T, Kurogi S, Yumoto T (2005) *Strobilomyces mirandus* Corner, a new record from Japan. *Mycoscience* 46:102–105
- Schardl CL, Young CA, Hesse U, Amyotte SG, Andreeva K, Calie PJ, Fleetwood DJ, Haws DC, Moore N, Oeser B, Panaccione DG (2013) Plant-symbiotic fungi as chemical engineers: multi-genome analysis of the *Clavicipitaceae* reveals dynamics of alkaloid loci. *PLoS Genet* 9(2):e1003323
- Schipper MAA, Samson RA (1978) One certain species of *Mucor* with a key to all accepted species. *Stud Mycol* 17:1–52
- Schoch CL, Shoemaker RA, Seifert KA, Hambleton S, Spatafora JW, Crous PW (2006) A multigene phylogeny of the *Dothideomycetes* using four nuclear loci. *Mycologia* 98:1041–1052
- Schoch CL, Crous PW, Groenewald JZ, Boehm EWA, Burgess TI, de Gruyter J, de Hoog GS, Dixon LJ, Grube M, Gueidan C, Harada Y, Hatakeyama S, Hirayama K, Hosoya T, Huhndorf SM, Hyde KD, Jones EBG, Kohlmeyer J, Kruys A, Li YM, Lücking R, Lumbsch HT, Marvanová L, Mbatchou JS, McVay AH, Miller AN, Mugambi GK, Muggia L, Nelsen MP, Nelson P, Owensby CA, Phillips AJL, Phongpaichit S, Pointing SB, Pujade-Renaud V, Raja HA, Rivas Plata E, Robbertse B, Ruibal C, Sakayaroj J, Sano T, Selbmann L, Shearer CA, Shirouzu T, Slippers B, Suetrong S, Tanaka K, Volkmann-Kohlmeyer B, Wingfield MJ, Wood AR, Woudenberg JHC, Yonezawa H, Zhang Y, Spatafora JW (2009) A class-wide phylogenetic assessment of Dothideomycetes. *Stud Mycol* 64:1–15
- Schulzer S (1870) Mykologische Beiträge. *Verhandlungen der Zoologisch-Botanischen Gesellschaft Wien* 20:635–658
- Seelan JSS, Justo A, Nagy LG, Grand EA, Redhead SA, Hibbett D (2015) Phylogenetic relationships and morphological evolution in *Lentinus*, *Polyporellus* and *Neofavolus*, emphasizing southeastern Asian taxa. *Mycologia* 107:460–474
- Seifert K, Morgan-Jones G, Gams W, Kendrick B (2011) The genera of Hyphomycetes. CBS-KNAW Fungal Biodiversity Centre, Utrecht
- Senanayake IC, Maharachchikumbura SSN, Hyde KD, Bhat JD, Jones EBG, McKenzie EHC, Dai DQ, Daranagama DA, Dayarathne MC, Goonasekara ID, Konta S, Li WJ, Shang QJ, Stadler M, Wijayawardene NN, Xiao YP, Norphanphon C, Li QR, Liu XZ, Bahkali AH, Kang JC, Wang Y, Wen TC, Wendt L, Xu JC, Camporesi E (2015) Towards unravelling relationships in Xylariomycetidae (*Sordariomycetes*). *Fungal Divers* 73:73–144
- Senanayake IC, Maharachchikumbura SSN, Jeewon R, Promputtha I, Al-Sadi AM, Camporesi E, Hyde KD (2017) Morphophylogenetic study of *Sydiowellaceae* reveals several new genera. *Mycosphere* 8(1):172–217
- Seyedmousavi S, Badali H, Chlebicki A, Zhao J, Prenafeta-Boldú FX, de Hoog GS (2011) *Exophiala sideris*, a novel black yeast isolated from environments polluted with toxic alkyl benzenes and arsenic. *Fungal Biol* 115:1030–1037
- Shearer CA, Raja HA, Miller AN, Nelson P, Tanaka K, Hirayama K, Marvanova L, Hyde KD, Zhang Y (2009) The molecular phylogeny of freshwater Dothideomycetes. *Stud Mycol* 64:145–153
- Shenoy BD, Jeewon R, Wu WP, Bhat DJ, Hyde KD (2006) Ribosomal and RPB2 DNA sequence analyses suggest that

- Sporidesmium* and morphologically similar genera are polyphyletic. Mycol Res 110:916–928
- Shoemaker RA (1984) Canadian and some extralimital *Nodulophaeria* and *Entodesmium* species. Can J Bot 62:2730–2753
- Shoemaker RA, Babcock CE (1989) *Phaeosphaeria*. Can J Bot 67:1500–1599
- Shirouzu T, Hirose D, Tokumasu S (2009) Taxonomic study of the Japanese Dacrymycetes. Persoonia 23:16
- Silvestro D, Michalak I (2010) raxmlGUI: a graphical front-end for RAxML. <http://sourceforge.net/projects/raxmlgui/>
- Silvestro D, Michalak I (2012) RaxmlGUI: a graphical front-end for RAxML. Org Divers Evol 12:335–337
- Simon UK, Bauer R, Rioux D, Simard M, Oberwinkler F (2005a) The intercellular biotrophic leaf pathogen *Cymadothea trifolii* locally degrades pectins, but not cellulose or xyloglucan in cell walls of *Trifolium repens*. New Phytol 165:243–260
- Simon UK, Bauer R, Rioux D, Simard M, Oberwinkler F (2005b) The vegetative life-cycle of the clover pathogen *Cymadothea trifolii* as revealed by transmission electron microscopy. Mycol Res 109:764–778
- Simon UK, Groenewald JZ, Crous PW (2009) *Cymadothea trifolii*, an obligate biotrophic leaf parasite of *Trifolium*, belongs to *Mycosphaerellaceae* as shown by nuclear ribosomal DNA analyses. Persoonia 22:49–55
- Simonini G (1992) Il valore statistico delle dimensioni sporali nei boleti. Atti riguardanti il V Seminario su *Russulales* e *Boletales*. AMB Reggio Emilia:16–28
- Simonini G (1998) Una metodologia per la descrizione e la rappresentazione delle dimensioni sporali (Applicata al caso delle *Boletaceae*). Pagine di Micologia 9:1–24
- Simonyan SA (1981) Mycoflora of botanical gardens and arboreta in Armenia. Hayka:232
- Singer R (1965) Die Röhrlinge, Teil I. J. Klinkhardt. Bad Heilbrunn:131
- Singer R (1967) Die Röhrlinge, Teil II. J. Klinkhardt. Bad Heilbrunn:151
- Singer R (1975) The Agaricales in modern taxonomy, 3rd edn. J. Cramer, Vaduz
- Singer R (1976) Marasmiaceae (Basidiomycetes-Tricholomataceae). Flora Neotropica Monogr 17:1–347
- Singer R (1986) The Agaricales in modern taxonomy, 4th edn. Sven Koeltz Scientific Books, Königstein
- Sivanesan A (1984) The bitunicate ascomycetes and their anamorphs. J Cramer, Vaduz
- Slippers B, Wingfield MJ (2007) *Botryosphaeriaceae* as endophytes and latent pathogens of woody plants: diversity, ecology and impact. Fungal Biol Rev 21(2):90–106
- Slippers B, Boissin E, Phillips A, Groenewald J, Lombard L, Wingfield M, Postma A, Burgess T, Crous P (2013) Phylogenetic lineages in the *Botryosphaeraiales*: a systematic and evolutionary framework. Stud Mycol 76:31–49
- Sogonov MV, Castlebury LA, Rossman AY, Mejía LC, White JF Jr (2008) Leaf-inhabiting genera of the *Gnomoniaceae*, *Diaporthales*. Stud Mycol 62:1–77
- Sotome K, Hattori T, Kakishima M (2007) *Polyporus phyllostachydis* sp. nov. with notes on other rhizophilic species of *Polyporus* (Basidiomycota, Polyporaceae). Mycoscience 48:42–46
- Sotome K, Hattori T, Ota Y, To-anun C, Salleh B, Kakishima M (2008) Phylogenetic relationships of *Polyporus* and morphologically allied genera. Mycologia 100:603–615
- Sotome K, Hattori T, Ota Y (2011) Taxonomic study on a threatened polypore, *Polyporus pseudobetulinus*, and a morphologically similar species, *P. subvarius*. Mycoscience 52:319–326
- Sotome K, Akagi Y, Lee SS, Ishikawa NK, Hattori T (2013) Taxonomic study of *Favolus* and *Neofavolus* gen. nov. segregated from *Polyporus* (Basidiomycota, Polyporales). Fungal Divers 58:245–266
- Spatafora JW, Chang Y, Benny GL, Lazarus K, Smith ME, Berbee ML, Bonito G, Corradi N, Grigoriev I, Gryganskyi A, James TY (2016) A phylum-level phylogenetic classification of zygomycete fungi based on genome-scale data. Mycologia 108(5):1028–1046
- Spegazzini C (1910) Mycetes Argentinienses (series V). Anales Mus Nac Hist Nat Buenos Aires 20(13):329–467
- Spegazzini C (1911) Hermatomyces Spieg. Anales Mus Nac Hist Nat Buenos Aires 20(13):445
- Spielman LJ (1985) A monograph of *Valsa* on hardwoods in North America. Can J Bot 63(8):1355–1378
- Stadler M, Kuhnert E, Peršoh D, Fournier J (2013) The Xylariaceae as model example for a unified nomenclature following the “One Fungus-One Name” (1F1N) concept. Mycology 4:5–21
- Stamatakis A (2006) RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. Bioinformatics 22:2688–2690
- Stamatakis A (2014) RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics 30(9):1312–1313
- Stevens NE (1926) Two species of *Physalospora* on citrus and other hosts. Mycologia 18:206–217
- Steyaert RL (1949) Contributions à l'étude monographique de Pestalotia de Not. et *Monochaetia* Sacc. (*Truncatella* gen. nov. et *Pestalotiopsis* gen. nov.). Bulletin du Jardin botanique de l'État de Bruxelles 19:285–354
- Stiller JW, Hall BD (1997) The origin of red algae: Implications for plastid evolution. Proc Natl Acad Sci USA 94(9):4520–4525
- Stukenbrock EH, Banke S, McDonald BA (2006) Global migration patterns in the fungal wheat pathogen *Phaeosphaeria nodorum*. Mol Ecol 15:2895–2904
- Sudhaham M, Prakitsin S, Sivichai S, Chaiyarat R, Dorrestein GM, Menken SBJ, De Hoog G (2008) The neurotropic black yeast *Exophiala dermatitidis* has a possible origin in the tropical rain forest. Stud Mycol 61:145–155
- Suetrong S, Schoch CL, Spatafora JW, Kohlmeyer J, Volkmann-Kohlmeyer B, Sakayaroj J, Phongpaichit S, Tanaka K, Hirayama K, Jones EBG (2009) Molecular systematics of the marine Dothideomycetes. Stud Mycol 64:155–173
- Sung G, Hywel-Jones NL, Sung J-M, Luangsa-Ard JJ, Shrestha B, Spatafora JW (2007) Phylogenetic classification of *Cordyceps* and the clavicipitaceous fungi. Stud Mycol 57:5–59
- Sutton BC (1980) The Coelomycetes. Fungi imperfecti with pycnidia, acervuli and stromata. Commonwealth Mycological Institute, Kew
- Svetasheva TY (2010) *Boletus legaliae* Pilát//Red Data Book of Tula region: plants and fungi/Tula Region Administration, Department of the Tula Region on ecology and natural resources—Tula, Grif et K:327 (in Russian)
- Svetasheva TY, Kovalenko AE (2013) *Boletus rubrosanguineus* Cheype//Red Data Book of Karachay-Cherkessia Republic—Cherkessk:201 (in Russian)
- Swofford DL (2003) PAUP: phylogenetic analysis using parsimony, version 4.0 b10. Sinauer Associates, Sunderland, MA. <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3348772>
- Sydow H (1928) Fungi Chilenses a cl. E. Werdermann lecti. Pars prima. Ann Mycol 26(1–2):100–126
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S (2013) MEGA6: molecular evolutionary genetics analysis version 6.0. Mol Biol Evol 30:2725–2729
- Tan YS, Desjardin DE, Perry BA, Vikineswary S, Noorlidah A (2009) *Marasmius* sensu stricto in Peninsular Malaysia. Fungal Divers 37:9–100

- Tanaka K, Hirayama K, Yonezawa H, Sato G, Toriyabe A, Kudo H, Hashimoto A, Matsumura M, Harada Y, Kurihara Y, Shirouzu T (2015) Revision of the *Massarinea* (Pleosporales, Dothideomycetes). *Stud Mycol* 82:75–136
- Tang AMC, Hyde KD, Corlett RT (2003) Diversity of fungi on wild fruits in Hong Kong. *Fungal Divers* 14:165–185
- Tang AMC, Jeewon R, Hyde KD (2009) A re-evaluation of the evolutionary relationships within the Xylariaceae based on ribosomal and protein-coding gene sequences. *Fungal Divers* 34:127–155
- Templeton MD, Rikkerink EHA, Solon SL, Crowhurst RN (1992) Cloning and molecular characterisation of the glyceraldehyde-3-phosphate dehydrogenase-encoding gene and cDNA from the plant pathogenic fungus *Glomerella cingulata*. *Gene* 122:225–230
- Tennakoon DS, Hyde KD, Phookamsak R, Wanasinghe DN, Camporesi E, Promputtha I (2016) Taxonomy and Phylogeny of *Juncaceicola* gen. nov. (Phaeosphaeriaceae, Pleosporinae, Pleosporales). *Cryptogam Mycol* 37:135–156
- Thacker JR, Henkel TW (2004) New species of *Clavulina* from Guyana. *Mycologia* 96:650–657
- Thambugala KM, Daranagama DA, Camporesi E, Singtripop C, Liu ZY, Hyde KD (2014a) Multi-locus phylogeny reveals the sexual state of *Tiarosporella* in Botryosphaeriaceae. *Cryptogam Mycol* 35:359–367
- Thambugala KM, Ariyawansa HA, Li YM, Boonmee S, Hongsanan S, Tian Q, Singtripop C, Bhat DJ, Camporesi E, Jayawardena R, Liu ZY, Xu JC, Chukeatirote E, Hyde KD (2014b) Dothideales. *Fungal Divers* 68:105–158
- Thambugala KM, Hyde KD, Tanaka K, Tian Q, Wanasinghe DN, Ariyawansa HA, Jayasiri SC, Boonmee S, Camporesi E, Hashimoto A, Hirayama K, Schumacher RK, Promputtha I, Liu ZY (2015a) Towards a natural classification and backbone tree for Lophiostomataceae, Floricolaceae, and Amorosiaceae fam. nov. *Fungal Divers* 74:199–266
- Thambugala KM, Chunfang Y, Camporesi E, Bahkali AH, Liu ZY, Hyde KD (2015b) *Pseudodidymosphaeria* gen. nov. in Massariaceae. *Phytotaxa* 231:271–282
- Thambugala KM, Bulgakov TS, Eungwanichayapant PD, Liu ZY, Hyde KD (2016) *Camarosporium uniseriatum* nom. nov., from *Celtis occidentalis* in European Russia. *Stud Fungi* 1:90–98
- Theissen F (1913) Lembosia-Studien. *Ann Mycol* 11(5):424–467
- Thilagam L, Nayak BK, Nanda A (2015) Studies on the diversity of coprophilous microfungi from hybrid cow dung samples. *Int J PharmTech Res* 8:135–138
- Tian Q, Liu JK, Hyde KD, Wanasinghe DN, Boonmee S, Jayasiri SC, Luo ZL, Taylor JE, Phillips AJL, Bhat DJ, Li WJ, Ariyawansa H, Thambugala KM, Jones EBG, Chomnunti P, Bahkali AH, Xu JC, Camporesi E (2015) Phylogenetic relationships and morphological reappraisal of *Melanommataceae* (Pleosporales). *Fungal Divers* 74:267–324
- Tibpromma S, Wijayawardene NN, Manamgoda DS, Boonmee S, Wanasinghe DN, Camporesi E, Yang JB, Hyde KD (2016a) *Camarosporium arezzoensis* on *Cytisus* sp., an addition to sexual state of *Camarosporium* sensu stricto. *Saudi. J Biol Sci* 23(1):1–8
- Tibpromma S, Bhat J, Doilom M, Lumyong S, Nontachaiyapoom S, Yang JB, Hyde KD (2016b) Three new *Hermatomyces* species (Lophiotremataceae) on *Pandanus odorifer* from Southern Thailand. *Phytotaxa* 275(2):127–139
- Tode HJ (1790) Fungi Mecklenburgenses Selecti. Fasc. 1. Nova Fungorum Genera Complectens. i–viii, 1–50, plates:1–7
- Trevisan V (1877) Note sur la tribu des Platystomees de la famille des Hypoxylacées. *Bull Soc R Bot Belg* 16:14–20
- Truszkowska W, Chlebicki A (1983) Pyrenomycetes pogorza cieszyńskiego. I. *Acta Mycol* 19:3–19
- Tulasne R, Tulasne C (1963) Selecta Fungorum Carpologia 2:1–316
- Tulloss RE (1994) Type studies in *Amanita* section *Vaginatae* I: some taxa described in this century (studied 1–23) with notes on description of basidiospores and refractive hyphae in *Amanita*. *Mycotaxon* 52:305–396
- Tulloss RE (2001) *Amanita olivaceogrisea* a little-known species found in Britain. *Field Mycol* 2(3):99–100
- Tulloss RE, Moses E (1995) *Amanita populiphila* a new species from the central United States. *Mycotaxon* 53:455–466
- Tulloss RE, Traverso M (2000) Illustrazioni di una nuova specie di *Amanita* dedicata al dr. Cornelis Bas di Leiden. *Bulletino del Gruppo Micologico G. Bresadola* 43(2):151–153
- Tulloss RE, Traverso M (2001) *Amanita basiana* a new species from pure *Pinus* forest and resembling the *Alnus*-associated species *Amanita friabilis*. *Mycotaxon* 77:47–55
- Tulloss RE, Yang ZL (eds) (2016) Studies in the Amanitaceae. [www.amanitaceae.org](http://www.amanitaceae.org)
- Tulloss RE, Kuyper TW, Vellinga EC, Yang ZL, Halling RE, Geml J, Sánchez-Ramírez S, Gonçalves SC, Hess J, Pringle A (2016) The genus *Amanita* should not be split. *Amanitaceae* 1(3):1–16
- Uehling JK, Henkel TW, Aime MC, Smith ME (2012a) New species of *Clavulina* with effused or resupinate basidiomata from the Guiana Shield. *Mycologia* 104:547–556
- Uehling JK, Henkel TW, Aime MC, Vilgalys R, Smith ME (2012b) New species and distribution records for *Clavulina* (Cantharellales, Basidiomycota) from the Guiana Shield, with a key to the lowland neotropical taxa. *Fungal Biol* 116:1263–1274
- Uljé CB (2005) 1. *Coprinus* Pers. In: Noordeloos ME, Kuyper TW, Vellinga EC (eds) *Flora Agaricina Nederlandica* 6. Taylor & Francis, Boca Raton, pp 22–109
- Untereiner WA, Strauss NA, Malloch D (1995) A molecular-morphotaxonomic approach to the systematics of the *Herpotrichiellaceae* and allied black yeasts. *Mycol Res* 99:897–913
- Upadhyay HP (1969) Soil fungi from north-east and north Brazil-VII. The genus *Gongronella*. *Nova Hedwigia* 17:65–73
- Vasquez G (2012) Indagini micologiche sulle Boletales del territorio siciliano. *Annales Confederationis Europaeae Mycologiae Mediterraneensis* 2009. Alaimo, Palermo, pp 69–81
- Vasquez G (2013) Indagini micologiche sulle Boletales Epigee del territorio siciliano—mappatura e censimento delle specie. Tesi di Dottorato di Ricerca (PhD thesis), XXVI Ciclo, Università degli Studi di Catania
- Vasquez G (2014) Indagini Micologiche sulle Boletales Epigee del Territorio Siciliano-mappatura e censimento delle specie [doctoral thesis]. Univ Studi Catania, Catania
- Vellinga EC (2004) Genera in the family Agaricaceae—evidence from nrITS and nrLSU sequences. *Mycol Res* 108:354–377
- Velmurugan P, Kamala-Kannan S, Balachandar V, Lakshmanaperumalsamy P, Chae JC, Oh BT (2010) Natural pigment extraction from five filamentous fungi for industrial applications and dyeing of leather. *Carbohydr Polym* 79(2):262–268
- Vilgalys R, Hester M (1990) Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *J Bacteriol* 172:4238–4246
- Visagie CM, Houbraken J, Frisvad JC, Hong SB, Klaassen CHW, Perrone G, Seifert KA, Varga J, Yaguchi T, Samson RA (2014) Identification and nomenclature of the genus *Penicillium*. *Stud Mycol* 78:343–371
- Visagie CM, Seifert KA, Houbraken J, Samson RA, Jacobs K (2016) A phylogenetic revision of *Penicillium* sect. *Exilicaulis*, including nine new species from fynbos in South Africa. *IMA Fungus* 7:75–117
- von Arx JA, Müller E (1954) Die Gattungen der amerosporen Pyrenomyceten. Beiträge zur Kryptogamenflora der Schweiz 11(1):1–434

- von Arx JA, Muller E (1975) A re-evaluation of the bitunicate ascomycetes with keys to families and genera. Stud Mycol 9:1–159
- Vrinda KB, Pradeep CK, Abraham TK (1997) Some Inocybes new to India. J Econ Taxon Bot 21(1):41–45
- Walker DM, Castlebury LA, Rossman AY, Sogonov MV, White JF (2010) Systematics of genus *Gnomoniopsis* (*Gnomoniaceae, Diaporthales*) based on a three gene phylogeny, host associations and morphology. Mycologia 102(6):1479–1496
- Walther G, Pawłowska J, Alastraue-Izquierdo WM, Rodriguez-Tudela JL, Dolatabadi S, Chakrabarti A, de Hoog GS (2013) DNA barcoding in *Mucorales*: an inventory of biodiversity. Persoonia 30:11–47
- Walty H (1969) *Tavole Svizzere di funghi*, vol. III, 3rd edn. tav. 39
- Wanasinghe DN, Jones EBG, Camporesi E, Boonmee S, Ariyawansa HA, Wijayawardene NN, Hyde KD (2014) An exciting novel member of *Lentitheciaceae* in Italy from *Clematis vitalba*. Cryptogam Mycol 35:323–337
- Wanasinghe DN, Jones EBG, Camporesi E, Dissanayake AJ, Kamolhan S, Mortimer PE, Xu J, Abd-Elsalam KA, Hyde KD (2016) Taxonomy and phylogeny of *Laburnicola* gen. nov. and *Paramassariosphaeria* gen. nov. (*Didymosphaeriaceae, Massarinae, Pleosporales*). Fungal Biol 120(11):1354–1373
- Wang L, Zhuang WY (2004) Designing primer sets for amplification of partial calmodulin genes from penicillia. Mycosistema 23(4):466–473
- Wang XH, Yang ZL, Li YC, Knudsen H, Liu PG (2009) *Russula griseocarnosa* sp. nov. (*Russulaceae, Russulales*), a commercially important edible mushroom in tropical China: mycorrhiza, phylogenetic position, and taxonomy. Nova Hedwigia 88:269–282
- Wang Y, Hyde KD, McKenzie EHC, Jiang YL, Li DW, Zhao DG (2015) Overview of *Stachybotrys* (*Memnoniella*) and current species status. Fungal Divers 71:17–83
- Wannathes N, Desjardin DE, Hyde KD, Perry BA, Lumyong S (2009) A monograph of *Marasmius* (Basidiomycota) from Northern Thailand based on morphological and molecular (ITS sequences) data. Fungal Divers 37:209–306
- Wartchow F (2012) *Clavulina incrassata*, a new species from Pernambuco, Brazil. Cryptogam Mycol 33(1):105–113
- Watanabe T (1994) Two new species of homothallic *Mucor* in Japan. Mycologia 86:691–695
- Watanabe T, Shitan N, Suzuki S, Umezawa T, Shimada M, Yazaki K, Hattori T (2010) Oxalate efflux transporter from the brown rot fungus *Fomitopsis palustris*. Appl Environ Microbiol 76:7683–7690
- Weber NS (1972) The genus *Helvella* in Michigan. Mich Bot 11:147–201
- Weinstein RN, Pfister DH, Iturriaga T (2002) A phylogenetic study of the genus *Cookeina*. Mycologia 94(4):673–682
- Wen YM, Rajendran RK, Lin YF, Kirschner R, Hu S (2015) Onychomycosis associated with *Exophiala oligosperma* in Taiwan. Mycopathologia 181(1):83–88
- White JF Jr, Bacon CW, Hywel-Jones NL, Spatafora JW (2003) Clavicipitalean fungi: evolutionary biology, chemistry, biocontrol and cultural impacts. CRC Press, Boca Raton
- White TJ, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (eds) PCR protocols: a guide to methods and applications. Academic Press, San Diego, pp 315–322
- Wijayawardene NN, McKenzie EHC, Hyde KD (2012) Towards incorporating anamorphic fungi in a natural classification—checklist and notes for 2011. Mycosphere 3(2):157–228
- Wijayawardene NN, Crous PW, Kirk PM, Hawksworth DL, Boonmee S, Braun U, Dai DQ, Dsouza MJ, Diederich P, Dissanayake A, Doilom M, Hongsanan S, Jones EBG, Groenewald JZ, Jayawardena R, Lawrey JD, Liu JK, Lücking R, Madrid H, Manamgoda DS, Muggia L, Nelsen MP, Phookamsak R, Suetrong S, Tanaka K, Thambugala KM, Wanasinghe DN, Wikee S, Zhang Y, Aptroot A, Ariyawansa HA, Bahkali AH, Bhat DJ, Gueidan C, Chomnunti P, De Hoog GS, Knudsen K, Li WJ, McKenzie EHC, Miller AN, Phillips AJL, Piątek M, Raja HA, Shivas RS, Slippers B, Taylor JE, Tian Q, Wang Y, Woudenberg JHC, Cai L, Jaklitsch WM, Hyde KD (2014a) Naming and outline of Dothideomycetes-2014 including proposals for the protection or suppression of generic names. Fungal Divers 69:1–55
- Wijayawardene NN, Hyde KD, Bhat DJ, Camporesi E, Schumacher RK, Chethana KWT, Wikee S, Bahkali AH, Wang Y (2014b) *Camarosporium*-like species are polyphyletic in *Pleosporales*; introducing *Paracamarosporium* and *Pseudocamarosporium* gen. nov. in *Montagnulaceae*. Cryptogam Mycol 35:177–198
- Wijayawardene NN, Bhat DJ, Hyde KD, Camporesi E, Chethana KWT, Tangthirasunun N, Wang Y (2014c) *Camarosporium* sensu stricto in *Pleosporineae, Pleosporales* with two new species. Phytotaxa 183(1):16–26
- Wijayawardene NN, Hyde KD, Bhat DJ, Goonasekars ID, Nadeeshan D, Camporesi E, Schumacher RK, Wang Y (2015) Additions to brown spored coelomycetous taxa in *Massarinae, Pleosporales*: introducing *Phragmocamarosporium* gen. nov. and *Suttonomyces* gen. nov. Cryptogam Mycol 36:213–224
- Wijayawardene NN, Hyde KD, Wanasinghe DN, Papizadeh M, Goonasekara ID, Camporesi E, Bhat DJ, McKenzie EHC, Phillips AJL, Diederich P, Tanaka K, Li WJ, Tangthirasunun N, Phookamsak R, Dai DQ, Dissanayake AJ, Weerakoon G, Maharachchikumbura SSN, Hashimoto A, Matsumura M, Bahkali AH, Wang Y (2016) Taxonomy and phylogeny of dematiaceous coelomycetes. Fungal Divers 77:1–316
- Wikee S, Lombard L, Nakashima C, Motohashi K, Chukeatirote E, Cheewangkoon R, McKenzie EHC, Hyde KD, Crous PW (2013) A phylogenetic re-evaluation of *Phyllosticta* (*Botryosphaerales*). Stud Mycol 76:1–29
- Winter G (1885) Pilze—Ascomyceten. In: GL Rabenhorst's Kryptogamen-Flora von Deutschland, Oesterreich und der Schweiz 1:65–528
- Winter G (1886) Rabenhorst's Kryptogamen-Flora, Pilze—Ascomyceten 1(2):570
- Wolf FA (1935) Morphology of *Polythrincium*, causing sooty blotch of clover. Mycologia 27(1):58–73
- Wolfe CB (1979) *Austroboletus* and *Tylopilus* subgenus *Porphyrellus* with emphasis on North American taxa. Bibl Mycol 69:1–132
- Wolfe BE, Kuo M, Pringle A (2012a) *Amanita thiersii* is a saprotrophic fungus expanding its range in the United States. Mycologia 104:22–23
- Wolfe BE, Tulloss RE, Pringle A (2012b) The irreversible loss of a decomposition pathway marks the single origin of an ectomycorrhizal symbiosis. PLoS ONE 7(7):539–597
- Woo PC, Ngan AH, Tsang CC, Ling IW, Chan JF, Leung SY, Yuen KY, Lau SK (2013) Clinical spectrum of Exophiala infections and a novel *Exophiala* species, *Exophiala hongkongensis*. J Clin Microbiol 51(1):260–267
- Woronichin NN (1925) Über die Capnodiales. Ann Mycol 23:174–178
- Woudenberg JHC, Aveskamp MM, De Gruyter J, Spiers AG, Crous PW (2009) Multiple *Didymella* teleomorphs are linked to the *Phoma clematidina* morphotype. Persoonia 22(1):56–62
- Wu G, Feng B, Xu J, Zhu XT, Li YC, Zeng NK, Hosen IMD, Yang ZL (2014) Molecular phylogenetic analyses redefine seven major clades and reveal 22 new generic clades in the fungal family *Boletaceae*. Fungal Divers 69:93–115
- Wyka SA, Broders KD (2016) The new family *Septorioideaceae*, within the *Botryosphaerales* and *Septoriooides strobi* as a new

- species associated with needle defoliation of *Pinus strobus* in the United States. *Fungal Biol* 120(8):1030–1040
- Yang T, Groenewald JZ, Cheewangkoon R, Jami F, Abdollahzadeh J, Lombard L, Crous PW (2017) Families, genera and species of *Botryosphaeraiales*. *Fungal Biol.* doi:[10.1016/j.funbio.2016.11.001](https://doi.org/10.1016/j.funbio.2016.11.001)
- Yang YL, Liu ZY, Cai L, Hyde KD, Yu ZN, McKenzie EHC (2009) Colletotrichum anthracnose of Amaryllidaceae. *Fungal Divers* 39:123–146
- Yang ZL (1997) Die *Amanita*-Arten von Südwestchina. *Bibl Mycol* 170:1–240
- Yang ZL (1990) Several noteworthy higher fungi from southern Yunnan, China. *Mycotaxon* 38:407–416
- Yang ZL (2000) Species diversity of the genus *Amanita* (Basidiomycetes) in China. *Acta Bot Yunnan* 22:135–142
- Yip HY (1988) *Berkleasmium correiae* sp. nov. on leaf hairs of *Correa lawrenciana*. *Australas Plant Pathol* 17:31–33
- Yuan HS, Dai YC (2008) *Polypores* from northern and central Yunnan Province, Southwestern China. *Sydowia* 60:147–159
- Zeng JS, Feng PY, Gerrits van den Ende AHG, Xi LY, Harrak MJ, de Hoog GS (2014) Multilocus analysis of the *Exophiala jeanselmei* clade containing black yeasts involved in opportunistic disease in humans. *Fungal Divers* 65:3–16
- Zhang LF, Yang JB, Yang ZL (2004) Molecular phylogeny of eastern Asian species of *Amanita* (Agaricales, Basidiomycota): taxonomic and biogeography implications. *Fungal Divers* 17:219–238
- Zhang N, Castlebury LA, Miller AN, Huhndorf SM, Schoch CL, Seifert KA, Rossman AY, Rogers JD, Kohlmeyer J, Volkmann-Kohlmeyer B, Sung GH (2006) An overview of the systematics of the Sordariomycetes based on four-gene phylogeny. *Mycologia* 98:1076–1087
- Zhang Y, Schoch CL, Fournier J, Crous PW, De Gruyter J, Woudenberg JHC, Hirayama K, Tanaka K, Pointing SB, Spatafora JW, Hyde KD (2009a) Multi-locus phylogeny of the *Pleosporales*: a taxonomic, ecological and evolutionary re-evaluation. *Stud Mycol* 64:85–102
- Zhang Y, Wang HK, Fournier J, Crous PW, Jeewon R, Pointing SB, Hyde KD (2009b) Towards a phylogenetic clarification of *Lophiostoma/Massarina* and morphologically similar genera in the *Pleosporales*. *Fungal Divers* 38:25–51
- Zhang Y, Fournier J, Bahkar AH, Hyde KD (2011) *Inflatispora*, a novel lignicolous genus of *Pleosporales* from France. *Sydowia* 62:287–295
- Zhang XY, Bao J, Wang GH, He F, Xu XY, Qi SH (2012a) Diversity and antimicrobial activity of culturable fungi isolated from six species of the South China Sea gorgonians. *Microb Ecol* 64:617–627
- Zhang Y, Crous PW, Schoch CL, Hyde KD (2012b) *Pleosporales*. *Fungal Divers* 53(1):1–221
- Zhang H, Hyde KD, Zhao Y, McKenzie EH, Zhou D (2014) Freshwater ascomycetes: *Lophiostoma vaginatispora* comb. nov. (Dothideomycetes, *Pleosporales*, *Lophiostomaceae*) based on morphological and molecular data. *Phytotaxa* 176:184–191
- Zhang M, He W, Wu JR, Zhang Y (2017) Two new species of *Spencermartinsia* (*Botryosphaeriaceae*, *Botryosphaeraiales*) from China. *Mycosphere* 7(7):942–949
- Zhao K, Wu G, Jang L (2014) A new genus, *Rubroboletus*, to accommodate *B. sinicus* and its allies. *Phytotaxa* 188(2):61–77
- Zhao Q, Tolgor B, Zhao YC, Yang ZL, Hyde KD (2015) Species diversity within the *Helvella crispa* group (Ascomycota: *Helvellaceae*) in China. *Phytotaxa* 239:130–142
- Zhao Q, Sulayman M, Zhu XT, Zhao YC, Yang ZL, Hyde KD (2016a) Species clarification of the culinary Bachu mushroom in Western China. *Mycologia* 108(4):828–836
- Zhao Q, Zhang XL, Li SH, Chai HM, Bahkali AH, Hyde KD (2016b) New species and records of saddle fungi (*Helvella*, *Helvellaceae*) from Jiuzhaigou Natural Reserve, China. *Mycoscience* 57(6):422–430
- Zhao Q, Brooks S, Zhao YC, Yang ZL, Hyde KD (2016c) Morphology and phylogenetic position of *Wynnelia subalpina* sp. nov. (*Helvellaceae*) from western China. *Phytotaxa* 270(1):41–48
- Zhaxybayeva O, Gogarten JP (2002) Bootstrap, Bayesian probability and maximum likelihood mapping: exploring new tools for comparative genome analyses. *BMC Genom* 3(1):4
- Zhou LW, Wei YL (2011) Changbai wood-rooting fungi 16. A new species of *Fomitopsis* (*Fomitopsidaceae*). *Mycol Prog* 11:435–441
- Zobel (1854) *Icones fungorum hucusque cognitorum* 6:4
- Zogg H (1962) Die *Hysteriaceae* s. str. und *Lophiaceae*. Beiträge zur Kryptogamenflora der Schweiz 11(3):1–190