Geosmithia lavendula, a new record for mycobiota of Iran

M. R. Mirzaee [⊠]

H. Mahmoudi

Agricultural and Natural Resources Research Center of Southern Khorasan, Birjand, Iran

R Zare

Iranian Research Institute of Plant Protection, Tehran, Iran

G. R. Hadarbadi

Forests, Rangelands and Watershed Management Organization, Tehran, Iran

A. Ghasemi

Iranian Research Institute of Plant Protection, Tehran, Iran

The genus *Geosmithia* (Ascomycota: Hypocreales) belongs to mitosporic filamentous fungi with a worldwide distribution (Kolařík et al. 2011). This genus is characterized by penicillium-like conidiophores with roughened walls, cylindrical phialides and smooth ellipsoidal to cylindrical conidia, forming long persistent chains (Kolařík & Kirkendall 2010; Sohn et al. 2013). *Geosmithia* species are found in soil, plant debris and wood and can act as true endophytes of healthy trees. Most species are known as exclusive associates of many insects invading phloem or sapwood of various plants (Kolařík et al. 2007; Kolařík & Kirkendall 2010; Moubasher & Soliman 2011).

During July-August 2013, twig samples of pistachio collected from Ghayen, Southern Khorasan province, Iran, with severe die-back symptoms. Symptomatic tissues from diseased twigs were surface-sterilized in 0.5% (v/v) sodium hypochlorite solution for 2 min, rinsed in sterile water and placed onto potato dextrose agar (PDA).

Genomic DNA was extracted using a modified Chelex method with an initial step of grinding the mycelia in liquid nitrogen (Walsh et al. 1991).

Complete internal transcribed spacers (ITS) of ribosomal DNA were amplified using the primer pair ITS4/ITS5 (White et al. 1990). PCR was performed in a 25 µl volume reaction mixture with 4 µl *Taq* master mix (SinaClon, Iran) containing dNTPs, MgCl2, reaction buffer,1µl of each primer (10 pmol) and 5 µl DNA template (equal to 1 ng /µl). PCR amplification was carried out using the following conditions: an initial denaturation at 94°C for 3 min, followed by 33 cycles of denaturation step at 94°C for 1 min, 45 s of annealing at 54°C, 2 min of extension at 72°C and a

final extension of 7 min at 72°C. PCR products were purified using PCR Purification Kit (Bioneer, Korea). Sequencing was performed in a 3730 xl DNA analyzer.

Colonies on malt-extract agar (MA) reached 20-30 mm diam in 7 d at 25°C. They were plane, velutinous; margins white, subsurface to low, narrow; greyish red to violet brown, reverse violet brown; exudate and soluble pigments absent (Fig 1); conidiophores borne from surface hyphae, up to $400 \times 3.5 \mu m$, with walls conspicuously verrucose; penicilli terminal, commonly quaterverticillate with all elements closely appressed and with verrucose walls; phialides in verticils of 4-6, 10- 12.5×2.5 - $3.7 \mu m$ in diameter, cylindrical, abruptly constricting to an apical pore; conidia cylindrical, smooth-walled, 3.7- 3.7×2.5 - $3.7 \mu m$, borne in long disordered chains (Fig 1).

Except for slightly larger conidia, fungal colonies emerging from the plated tissues had morphological characteristics typical of *Geosmithia lavendula* (Raper & Fennell) Pitt, (Pitt 1979; Moubasher & Soliman 2011).

Two fungal isolates were identified as *Geosmithia lavendula* using morphological characteristics and sequence analysis of ITS region. The sequences of a representative isolate (GenBank Accession No. KM396270) displayed 100% homology with sequences AM949861 and AM421126 [*G. lavendula* isolated from *Hypoborus ficus* (Coleoptera: Scolytidae) on *Ficus carica*] from Croatia and Azerbaijan, respectively. A culture of the fungus is preserved at the Iranian Fungal Culture Collection, Tehran, as IRAN 2239C.

Geosmithia lavendula has been reported from clinical materials (USA), soil (Qatar and Venezuela), air (India), phylloplane of citrus (Egypt), elm bark beetle (USA) and two species of ambrosia beetles (Costa Rica) (Pitt 1979; Kolařík et al. 2007; Moubasher & Soliman 2011).

This is the second report of a member of this genus, as well as the first report of *G. lavendula* from Iran. *Geosmithia pallida* has been reported on grapevine from Iran (Hergholi et al. 2013).

REFERENCES

Hergholi N, Javan-Nikkhah M, Ghosta Y, Campisano A, Pancher M. 2013. First report of new endophytic fungi in grapevine trees (*Vitis vinifera* L.) in West Azerbaijan province.1st Iranian Mycological Congress, 3-5 September 2013, University of Guilan, Rasht, Iran.



Fig 1. Geosmithia lavendula IRAN 2239 C. Front (a) and reverse (b) surface of colony of Geosmithia lavendula isolated from pistachio twigs, growing on PDA: conidiophores and conidia from the culture (c). Bar = $20 \mu m$.

Kolařík M, Freeland E, Utley C, Tisserat N. 2011. *Geosmithia morbida* sp. nov., a new phytopathogenic species living in symbiosis with the walnut twig beetle (*Pityophthorus juglandis*) on *Juglans* in USA. Mycologia 103: 325–332.

Kolařík M, Kirkendall LR. 2010. Evidence for a new lineage of primary ambrosia fungi in *Geosmithia* Pitt (Ascomycota: Hypocreales). Fungal Biology 114: 676–689.

Kolařík M, Kostovčík M, Pažoutová S. 2007. Host range and diversity of the genus *Geosmithia* (Ascomycota: Hypocreales) living in association with bark beetles in the Mediterranean area. Mycological Research 111: 1298–1310.

Moubasher AH. Soliman Z. 2011. Contribution to the mycobiota of Egypt: *Geosmithia* Pitt with *G. lavendula*, a new record to Egypt. Journal of Basic and Applied Mycology 2: 91-94.

Pitt JI. 1979. *Geosmithia* gen. nov. for *Penicillium lavendulum* and related species. Canadian Journal of Botany 57: 2021–2030.

Sohn JY, Jang MA, Lee JH, Park KS, Ki CS, Lee NY. 2013. Isolation and identification of *Geosmithia argillacea* from a fungal ball in the lung of a tuberculosis patient. Annals of Laboratory Medicine 33: 136–40.

Walsh PS, Metzger DA, Higuchi R. 1991. Chelex 100 as a medium for simple extraction of DNA for PCR-based typing from forensic material. BioTechniques 10: 506–513.

White TJ, Bruns T, Lee S, Taylor J. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: PCR Protocols: a guide to methods and applications. (Innis MA, Gelfand DH, Sninsky JJ, White TJ, eds). Academic Press, New York, USA: pp. 315–322.