# Essential Oils from the Aerial Part and Rhizome of Amomum muricarpum Elmer and Their Antimicrobial Activity

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**Abstract:** Chemical compositions in essential oils of *Amomum muricarpum* Elmer were determined using GC-MS/GC-FID. In the hydro-distilled oils of aerial part and rhizome, twenty-eight and thirty-three constituents were identified, accounting for 98.73 and 97.88%, respectively. *A. muricarpum* essential oils were associated with the presence of monoterpenes, sesquiterpenes, and their oxygenated types. Monoterpenes (> 80.00%) were major compounds, in which the main compound  $\alpha$ -pinene reached the highest percentage of 62.94-74.97%. *A. muricarpum* essential oils showed antimicrobial activity against the positive Gram bacterium *Staphylococcus aureus* and filamentous fungus *Aspergillus niger* with the respective MIC values of 400 and 100 µg/mL. This study provides new information that Vietnamese medicinal plant *A. muricarpum* are a rich resource of monoterpene  $\alpha$ -pinene

#### Keywords: Amomum muricarpum Elmer; Aerial part; Rhizome; Essential oils; Antimicrobial

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#### 1. Introduction

*Zingiberaceae* is a family of essential oil-bearing plants. Genus *Amomum* in this family consisted of about 150-180 species and was native to Asia, Africa, and Australia [1-6]. All parts of *Amomum* plants are notable for their pungency and aromatic features, which have been employed as pharmaceutical and food chemical agents [7-15]. The plants of this genus are also well-known in traditional uses for toothache, dysentery, gastrointestinal, nervous system strengthen, pregnancy-related diseases [16-18]. Phytochemical analyses, mostly based on GC-MS procedures, have identified the presence of a vast amount of terpenoids in *Amomum* essential oils [19-21].

A. muricarpum Elmer is a medicinal plant 2.0-3.0 m tall, which its name was accepted at a high level of confidence [22]. It can be gathered from dense forests in China, the Philippines, and Vietnam [23]. Various phytochemical publications evidently concluded that diarylheptanoids were major components in its plant extracts [23,24]. However, GC-MS analysis aimed to determine chemical components in essential oils of this plant is not available to date. Our current paper deals with the chemical compositions in the essential oils of A. *muricarpum* aerial part and rhizome, collected from Vietnam, together with their antimicrobial activity.

# 2. Materials and Methods

# 2.1. Materials.

The fresh aerial part and rhizome of *A. muricarpum* species were collected from Thua Thien Hue, Vietnam, in 04-2021, around 16°07'23.9"N and 107°36'30.3"E. The plant materials were identified by Dr. Vu Tien Chinh, Vietnam National Museum of Nature, VAST. The voucher specimen AM-2021 was deposited at the Institute of Natural Products Chemistry, VAST.

The positive Gram bacteria *Bacillus subtilis* ATCC 27212 and *Staphylococcus aureus* ATCC 12222, the negative Gram bacteria *Escherichia coli* ATCC 8739 and *Pseudomonas aeruginosa* ATCC 25923, fungi *Aspergillus niger* ATCC 9763 and *Fusarium oxysporum* ATCC 48112, and yeasts *Candida albicans* ATCC 10231 and *Saccharomyces cerevisiae* ATCC9763 were purchased from American Type Culture Collection (ATCC, Manassas, VA, USA).

# 2.1.1. Distillation.

The fresh aerial part (1.0 kg) was chopped and was intermediately hydro-distilled using Clevenger apparatus for 2.5 h, to produce yellow and pleasant smelling oil (0.01% yield, w/w). The oil obtained was dried over anhydrous sodium sulfate and kept in amber-color vials at 4 °C before use. A similar procedure was applied to the fresh rhizome (1.5 kg), to yield the same oil (0.02%, w/w).

# 2.1.2. GC-MS analysis.

An HP7890A model GC (Agilent Technologies, USA) linked to an HP5975C MS detector (MSD), and an HP-5 MS column (dimension: 60 m x 0.25 mm, film thickness: 0.25  $\mu$ m) was used for GC-MS analysis. The injector was set around 250 °C. The temperature program started at 60 °C and increased up to 240 °C with each 4 °C/min. He was used as a carrier gas (1.0 mL/min). The split ratio was 100:1. The MSD circumstance was contingent upon the full scan modes under electron impact ionization (voltage: 70 eV, emission current: 40 mA, mass range scan: 35-450 amu). The GC-FID analysis was performed with the same condition of GC-MS procedure. Each chemical compound in the oil sample was determined by comparing their RI and MS data with those from the NIST standard database and reference books [25], while the relative percentage was calculated based on the peak area in the GC-FID without any correction factor.

# 2.2. Antimicrobial assay.

The antimicrobial procedure was carried out according to our previous publications [26-29]. Briefly, the Gram bacteria were cultured in tryptic soy broth, whereas fungal microorganisms were grown in saboraud-2% dextrose with 150 x 10<sup>6</sup> CFU/mL. The oil sample (12.5-400  $\mu$ g/mL) was loaded into a 96-well plate containing fresh culture and kept at 37 °C and 24 h. The MIC (Minimum inhibitory concentration) denotes the lowest concentration, at which the tested sample suppressed the visible growth of pathogenic microorganisms after 24

h treatment. Streptomycin and tetracycline served as positive controls for the positive and negative Gram bacteria, respectively, whereas nystatin was used as a positive control for fungi and yeasts. DMSO (5%) was used as a negative control. Each experiment was performed in triplicate.

#### **3. Results and Discussion**

The gas chromatography on HP-5 capillary column has identified 28 constituents in the essential oil of the *A. muricarpum* aerial part, which accounted for 98.73% (Table 1). Monoterpenes (11 compounds) were major components with the highest percentage of 87.67%, whereas monoterpenoids (6 compounds), sesquiterpenes (9 compounds), and sesquiterpenoids (2 compounds) reached 7.07, 3.00, and 0.99%, respectively.  $\alpha$ -Pinene with 74.97% was a principal compound in the monoterpene group, in addition to several significant ones such as camphene (2.96%), myrcene (2.74%),  $\beta$ -phellandrene (2.21%), and limonene (2.17%). Linalool (4.37%) and  $\alpha$ -terpineol (1.10%) were major compounds in the monoterpenoid derivatives was less than 1.00%.

No	<sup>a</sup> Rt	<sup>b</sup> RI <sub>E</sub>	° <b>RI</b> L	Constituents	Classification	Aerial part [%]	Rhizome [%]
1	10.21	928	927	Tricyclene	Monoterpene	0.23	0.21
2	10.57	940	939	α-Pinene	Monoterpene	74.97	62.94
3	11.03	955	954	Camphene	Monoterpene	2.96	3.66
4	11.91	984	979	β-Pinene	Monoterpene	0.96	2.63
5	12.13	991	991	Myrcene	Monoterpene	2.74	3.22
6	12.73	1010	1003	α-Phellandrene	Monoterpene	0.38	0.75
7	13.13	1022	1017	α-Terpinene	Monoterpene	-	0.24
8	13.39	1029	1026	<i>O</i> -Cymene	Monoterpene	0.19	0.53
9	13.55	1034	1029	Limonene	Monoterpene	2.17	3.92
10	13.61	1036	1030	β-Phellandrene	Monoterpene	2.21	2.88
11	14.55	1063	1060	γ-Terpinene	Monoterpene	0.28	1.10
12	15.62	1094	1089	Terpinolene	Monoterpene	0.59	0.76
13	15.86	1101	1097	Linalool	Monoterpenoid	4.37	1.98
14	16.57	1121	1122	Endo-fenchol	Monoterpenoid	-	0.18
15	18.46	1175	1168	Endo-borneol	Monoterpenoid	0.63	0.48
16	18.82	1185	1177	Terpinen-4-ol	Monoterpenoid	0.62	0.27
17	19.25	1198	1189	α-Terpineol	Monoterpenoid	1.10	1.02
18	20.30	1228	1220	Fenchyl acetate	Monoterpenoid	0.13	5.10
19	21.25	1256	1253	Geraniol	Monoterpenoid	-	0.18
20	22.59	1294	1289	Bornyl acetate	Monoterpenoid	0.22	1.25
21	23.03	1308	1298	Sabinyl acetate	Monoterpenoid	-	0.40
22	23.88	1333	1327	Myrtenyl acetate	Monoterpenoid	-	0.36
23	25.74	1390	1377	α-Copaene	Sesquiterpene	0.36	0.20
24	26.10	1401	1388	β-Bourbonene	Sesquiterpene	0.22	-
25	27.08	1432	1427	α-Santalene	Sesquiterpene	-	0.48
26	27.98	1460	1452	<i>Epi</i> -β-santalene	Sesquiterpene	-	0.31
27	28.58	1480	1470	9- <i>Epi</i> -( <i>E</i> )- caryophyllene	Sesquiterpene	0.35	0.20
28	28.93	1491	1500	γ-Muurolene	Sesquiterpene	0.17	-

Table 1. Essential oils of A. muricarpum aerial part and rhizome.

No	aRt	<sup>b</sup> RI <sub>E</sub>	° <b>RI</b> L	Constituents	Classification	Aerial part [%]	Rhizome [%]
29	29.17	1498	1485	Germacrene D	Sesquiterpene	0.42	0.26
30	29.37	1505	1490	β-Selinene	Sesquiterpene	-	0.21
31	29.53	1510	1507	(Z)-α-Bisabolene	Sesquiterpene	-	0.18
32	29.58	1512	1506	( <i>E</i> , <i>E</i> )-α-Farnesene Sesquiterpene		0.27	-
33	29.65	1515	1500	Bicyclogermacrene	Sesquiterpene -		0.72
34	29.75	1518	1506	β-Bisabolene	Sesquiterpene 0.80		0.91
35	30.13	1530	1523	γ-Cadinene	Sesquiterpene 0.22		-
36	30.32	1537	1523	δ-Cadinene	Sesquiterpene	0.19	-
37	31.29	1569	1563	(E)-Nerolidol	Sesquiterpenoid	0.26	-
38	32.12	1597	1579	Spathulenol	Sesquiterpenoid	0.73	-
39	38.46	1829	1809	γ- Bicyclohomofarnesal	Sesquiterpenoid	-	0.15
40	46.26	2154	2135	Coronarin E	Diterpenoid	-	0.20
				Total		98.73	97.88
				Monoterpenes		87.67	82.84
				Monoterpenoids		7.07	11.22
				Sesquiterpenes		3.00	3.47
				Sesquiterpenoids		0.99	0.15
				Diterpenoids			0.20

<sup>a</sup>Retention time, <sup>b</sup>Retention indices relative to *n*-alkanes ( $C_7$ - $C_{30}$ ) on HP-5 MS column, <sup>c</sup>Retention indices from NIST standard database

Regarding essential oil from the rhizome, 33 constituents were namely identified in a total amount of 97.88%. Monoterpenes, sesquiterpenes, and their oxygenated derivatives were also characteristic of rhizome oil. Once again, monoterpenes (12 compounds, 82.84%) were found to be the main components. Monoterpenoids (10 compounds, 11.22%) were the second class of remarkable compounds in rhizome oil. It is similar to the essential oil of the aerial part, rhizome oil, accompanied by the appearance of minor compounds type sesquiterpenes (3.47%) and sesquiterpenoids (0.15%).  $\alpha$ -Pinene in rhizome oil still ranked the highest amount of 62.94%, but less than that of aerial part by 12.03%. The percentages of four monoterpenes, camphene, myrcene,  $\beta$ -phellandrene, and limonene, have increased in the rhizome oil. In contrast to the amount of linalool, two minor compounds  $\beta$ -pinene and fenchyl acetate in the aerial part were being become remarkable compounds in rhizome oil (Table 1). Coronarin E (0.20%) was the only diterpenoid found in rhizome oil.

The difference between two oils is that sesquiterpenes  $\beta$ -bourbonene (0.22%),  $\gamma$ muurolene (0.17%),  $\gamma$ -cadinene (0.22%), and  $\delta$ -cadinene (0.19%), sesquiterpenoids (*E*)nerolidol (0.26%) and spathulenol (0.73%) were only detected in the essential oil of aerial part, whereas monoterpene  $\alpha$ -terpinene (0.24%), monoterpenoids *endo*-fenchol (0.18%), geraniol (0.18%), sabinyl acetate (0.40%), myrtenyl acetate (0.36%), sesquiterpenes  $\alpha$ -santalene (0.48%), *epi*- $\beta$ -santalene (0.31%),  $\beta$ -selinene (0.21%), (*Z*)- $\alpha$ -bisabolene (0.18%), and bicyclogermacrene (0.72%), sesquiterpenoid  $\gamma$ -bicyclohomofarnesal (0.15%), and diterpenoid coronarin E were only observed in the rhizome oil.

Significantly, our study demonstrated that monoterpene  $\alpha$ -pinene possessed such a high amount of over 60.00% in *A. muricarpum* species, collected from Vietnam. In comparison with previous reports, this compound was also found in several *Amomum* species, such as essential oils of *A. villosum* leaf and root bark (11.6-22.1%), *A. rubidium* rhizome (7.70%), *A. subulatum* 

seed and rind (4.8-6.4%) [30-32]. Therefore, it can be concluded that *A. muricarpum* is likely to be a rich resource of  $\alpha$ -pinene.

Mic	robial strains	Minimum Inhibitory Concentration (MIC: mg/mL)					
		Aerial part	Rhizome	Streptomycin	Tetracyclin	Nystatin	
Gram (+)	B. subtilis	(-)	(-)	7.20			
	S. aureus	400	400	14.38			
Gram (-)	E. coli	(-)	(-)		5.5		
	P. aeruginosa	(-)	(-)		11.0		
Fungi	A. niger	100	100			23.13	
	F. oxysporum	(-)	(-)			11.57	
Yeasts	C. albicans	(-)	(-)			11.56	
	S. cerevisiae	(-)	(-)			5.78	

Table 2. Antimicrobial activity of tested samples.

Two oil samples have been further to subject biological assay. Both of them showed antimicrobial activity against the positive Gram bacterium *S. aureus* and filamentous fungus *A. niger* with the MIC values of 400 and 100  $\mu$ g/mL, but failed to control the remaining pathogenic microorganisms (Table 2). Amonum essential oils were recognized to be potential agents against pathogenic micro bacteria. For example, Geogre *et al.* (2006) also identified that essential oils of *A. cannicarpum* rhizome were capable of inhibiting the growth of the positive Gram bacteria *B. subtilis* and *S. aureus* and the negative Gram bacterium *E. coli* [33]. In the same manner, terpenoids in *A. tsao-ko* essential oils response for its moderate inhibitory antimicrobial activities against *S. aureus, Staphylococcus epidermidis, B. subtilis, Propionibacterium acnes, E. coli, Pseudomonas aeruginosa*, and *C. albicans* [34].

## 4. Conclusions

For the first time, our study reported chemical compositions of the Vietnamese *A*. *muricarpum* essential oils. Monoterpenes with over 80.0% were a main chemical class of compounds in essential oils of both aerial part and rhizome.  $\alpha$ -Pinene was a principal component in these oils, which ranged from 62.94 to 74.97%. The oils of this plant were also characterized by the presence of monoterpenoids, sesquiterpenes, sesquiterpenoids, and diterpenoids. *A. muricarpum* essential oils successfully inhibited filamentous fungus *A. niger* growth with the MIC value of 100 µg/mL. This paper can be seen as a basic guide for future research.

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# **Conflicts of Interest**

The authors declare no conflict of interest.

#### References

- Huong, L.T.; Hung, N.V.; Chung, M.V.; Dai, D.N.; Ogunwande, I.A. Essential oils Constituents of the leaves of Amomum gagnepainii and Amomum repoense. *Nat. Prod. Res.* 2018, *32*, 316-321, https://doi.org/10.1080/14786419.2017.1346643.
- Boer, H.d.; Newman, M.; Poulsen, A.D.; Droop, A.J.; Fér, T.; Thu Hiền, L.T.; Hlavatá, K.; Lamxay, V.; Richardson, J.E.; Steffen, K.; Leong-Škorničková, J. Convergent morphology in Alpinieae (Zingiberaceae): Recircumscribing Amomum as a monophyletic genus. *Taxon* 2018, 67, 6-36, https://doi.org/10.12705/671.2.
- 3. Valukattil, P.T; Mamiyil, S.; Vettathukattil, A.G.M.N. A New species of *Amomum* Roxb. (Zingiberaceae) from Nagaland, India. *Taiwania* **2019**, *64*, 9-12, https://doi.org/10.6165/tai.2019.64.9.
- Thinh, B.B.; Doudkin, R.V.; Thanh, V.Q. Chemical Composition of Essential Oil of Amomum xanthioides Wall. ex Baker from Northern Vietnam. *Biointerface Res. Appl. Chem.* 2021, 11, 12275-12284 https://doi.org/10.33263/BRIAC114.1227512284.
- Ding, H.-B.; Yang, B.; Maw, M.B.; Win, P.P.; Tan, Y.-H. Taxonomic studies on Amomum Roxburgh sl (Zingiberaceae) in Myanmar II: one new species and five new records for the flora of Myanmar. *PhytoKeys* 2020, 138, 139, https://doi.org/10.3897/phytokeys.138.38736.
- 6. Syazana, S.A.; Meekiong, K.; Afifah, N.; Syauqina, M.Y. *Amomum bungoensis*: A New species of *Amomum* (Zingiberaceae) from Sarawak, Malaysia. *J. Bot.* **2018**, 2018, https://doi.org/10.1155/2018/1978607.
- Liu, H.; Yan, Q.; Zou, D.; Bu, X.; Zhang, B.; Ma, X.; Leng, A.; Zhang, H.; Li, D.; Wang, C. Identification and bioactivity evaluation of ingredients from the fruits of Amomum tsaoko Crevost et Lemaire. *Phytochemistry Letters* 2018, 28, 111-115, https://doi.org/10.1016/j.phytol.2018.10.007.
- Lu, S.; Zhang, T.; Gu, W.; Yang, X.; Lu, J.; Zhao, R.; Yu, J. Volatile oil of *Amomum villosum* inhibits nonalcoholic fatty liver disease via the gut-liver axis, *Biomed Int. Res.* 2018, 2018, https://doi.org/10.1155/2018/3589874.
- Kim, J.G.; Jang, H.; Le, T.P.L.; Hong, H.R.; Lee, M.K.; Hong, J.T.; Lee, D.; Hwang, B.Y. Pyranoflavanones and Pyranochalcones from the Fruits of Amomum tsao-ko. *J. Nat. Prod.* 2019, 82, 1886-1892, https://doi.org/10.1021/acs.jnatprod.9b00155.
- Chen, Z.-y.; Guo, S.-s.; Cao, J.-q.; Pang, X.; Geng, Z.-f.; Wang, Y.; Zhang, Z.; Du, S.-s. Insecticidal and repellent activity of essential oil from Amonum villosum Lour. and its main compounds against two storedproduct insects. *Int. J. Food Prop.* 2018, *21*, 2265-2275, https://doi.org/10.1080/10942912.2018.1508158.
- Lv, H.; Pian, R.; Xing, Y.; Zhou, W.; Yang, F.; Chen, X.; Zhang, Q. Effects of citric acid on fermentation characteristics and bacterial diversity of Amomum villosum silage. *Bioresour. Technol.* 2020, 307, 123290, https://doi.org/10.1016/j.biortech.2020.123290.
- Yuan, H.; Zhang, T.; Huang, S.; Zhou, J.; Park, S. Six Gentlemen Decoction adding Aucklandia and Amomum (Xiangsha Liujunzi Tang) for the treatment of ulcerative colitis: A systematic review and metaanalysis of randomized clinical trials. *European Journal of Integrative Medicine* 2020, 36, 101119, https://doi.org/10.1016/j.eujim.2020.101119.
- He, X.-F.; Chen, J.-J.; Li, T.-Z.; Hu, J.; Zhang, X.-K.; Guo, Y.-Q.; Zhang, X.-M.; Geng, C.-A. Tsaokols A and B, unusual flavanol-monoterpenoid hybrids as α-glucosidase inhibitors from Amomum tsao-ko. *Chin. Chem. Lett.* **2021**, *32*, 1202-1205, https://doi.org/10.1016/j.cclet.2020.08.050.
- 14. He, X.-F.; Chen, J.-J.; Huang, X.-Y.; Hu, J.; Zhang, X.-K.; Guo, Y.-Q.; Zhang, X.-M.; Geng, C.-A. The antidiabetic potency of Amomum tsao-ko and its active flavanols, as PTP1B selective and α-glucosidase dual inhibitors. *Industrial Crops and Products* **2021**, *160*, 112908, https://doi.org/10.1016/j.indcrop.2020.112908.
- He, X.-F.; Wang, H.-M.; Geng, C.-A.; Hu, J.; Zhang, X.-M.; Guo, Y.-Q.; Chen, J.-J. Amomutsaokols A–K, diarylheptanoids from Amomum tsao-ko and their α-glucosidase inhibitory activity. *Phytochemistry* 2020, *177*, 112418, https://doi.org/10.1016/j.phytochem.2020.112418.
- Chau, L.T.M.; Thang, T.D.; Huong, L.T.; Ogunwande, I.A. Constituents of Essential Oils from Amonum longiligulare from Vietnam. *Chem. Nat. Compd.* 2015, *51*, 1181-1183, https://doi.org/10.1007/s10600-015-1525-z.
- 17. Ao, H.; Wang, J.; Chen, L.; Li, S.; Dai, C. Comparison of Volatile Oil between the Fruits of Amomum villosum Lour. and Amomum villosum Lour. var. xanthioides T. L. Wu et Senjen Based on GC-MS and Chemometric Techniques. *Molecules* **2019**, *24*, https://doi.org/10.3390/molecules24091663.
- 18. Yin, H.; Dan, W.-J.; Fan, B.-Y.; Guo, C.; Wu, K.; Li, D.; Xian, K.-F.; Pescitelli, G.; Gao, J.-M. Antiinflammatory and α-Glucosidase Inhibitory Activities of Labdane and Norlabdane Diterpenoids from the

Rhizomes of Amonum villosum. *J. Nat. Prod.* **2019**, *82*, 2963-2971, https://doi.org/10.1021/acs.jnatprod.9b00283.

- 19. Alam, A.; Singh, V. Composition and pharmacological activity of essential oils from two imported Amomum subulatum fruit samples. *Journal of Taibah University Medical Sciences* **2021**, *16*, 231-239, https://doi.org/10.1016/j.jtumed.2020.10.007.
- Kurup, R.; Thomas, V.P.; Jose, J.; Dan, M.; Sabu, M.; Baby, S. Chemical Composition of Rhizome Essential Oils of Amomum agastyamalayanum and Amomum newmanii from South India. *Journal of Essential Oil Bearing Plants* 2018, 21, 803-810, https://doi.org/10.1080/0972060X.2018.1500182.
- Govindarajan, M.; Rajeswary, M.; Senthilmurugan, S.; Vijayan, P.; Alharbi, N.S.; Kadaikunnan, S.; Khaled, J.M.; Benelli, G. Larvicidal activity of the essential oil from Amomum subulatum Roxb. (Zingiberaceae) against Anopheles subpictus, Aedes albopictus and Culex tritaeniorhynchus (Diptera: Culicidae), and non-target impact on four mosquito natural enemies. *Physiol. Mol. Plant Pathol.* 2018, 101, 219-224, https://doi.org/10.1016/j.pmpp.2017.01.003.
- 22. Theplantlist.org. Available online: http://www.theplantlist.org/tpl1.1/record/kew-219398 (accessed on 05 May **2021**).
- 23. Giang, P.M.; Son, P.T.; Matsunami, K.; Otsuka, H. New diarylheptanoids from *Amomum muricarpum* ELMER. *Chem. Pharm. Bull.* **2006**, *54*, 139-140, https://doi.org/10.1248/cpb.54.139.
- Giang, P.M.; Son, P.T.; Matsunami, K.; Otsuka, H. One new and several minor diarylheptanoids from Amomum muricarpum. *Nat. Prod. Res.* 2012, 26, 1195-1200, http://dx.doi.org/10.1080/14786419.2010.545775.
- 25. https://webbook.nist.gov/chemistry; Accessed April 28, 2021
- Son, N.T.; Oda, M.; Hayashi, N.; Yamaguchi, D.; Kawagishi, Y.; Takahashi, F.; Harada, K.; Cuong, N.M.; Fukuyama, Y. Antimicrobial Activity of the Constituents of Dalbergia tonkinensis and Structural-Bioactive Highlights. *Nat. Prod. Commun.* 2018, *13*, 157-161, https://doi.org/10.1177/1934578X1801300212.
- Son, N.T.; Harada, K.; Cuong, N.M.; Fukuyama, Y. Two New Carboxyethylflavanones from the Heartwood of Dalbergia tonkinensis and Their Antimicrobial Activities. *Nat. Prod. Commun.* 2017, *12*, 1721–1723, https://doi.org/10.1177/1934578X1701201115.
- Cuong, N.M.; Nhan, N.T.; Son, N.T.; Nghi, D.H.; Cuong, T.D. Daltonkins A and B, Two New Carboxyethylflavanones from the Heartwood of Dalbergia tonkinensis. *Bull. Korean Chem. Soc.* 2017, *38*, 1511-1514, https://doi.org/10.1002/bkcs.11313.
- Son, N.T.; Tuan, A.L.; Thu, T.D.T.; Dinh, L.N.; Thi, T.T. Essential Oils of Polyalthia suberosa Leaf and Twig and Their Cytotoxic and Antimicrobial Activities. *Chem. Biodivers.* 2021, 18, e2100020, https://doi.org/10.1002/cbdv.202100020.
- 30. Dai, D.N.; Huong, L.T.; Thang, T.D.; Ogunwande, I.A. Chemical composition of essential oils of Amomum villosum Lour. *Am. J. Essent. Oils Nat. Prod* **2016**, *4*, 8-11.
- Huong, L.T.; Sam, L.N.; Giang, C.N.; Dai, D.N.; Ogunwande, I.A. Chemical Composition and Larvicidal Activity of Essential Oil from the Rhizomes of Amomum rubidum Growing in Vietnam. *Journal of Essential Oil Bearing Plants* 2020, 23, 405-413, https://doi.org/10.1080/0972060X.2020.1756425.
- Satyal, P.; Dosoky, N.S.; Kincer, B.L.; Setzer, W.N. Chemical Compositions and Biological Activities of Amomum subulatum Essential Oils from Nepal. *Nat. Prod. Commun.* 2012, 7, 1233-1236, https://doi.org/10.1177/1934578X1200700935.
- George, V.; Mathew, J.; Sabulal, B.; Dan, M.; Shiburaj, S. Chemical composition and antimicrobial activity of essential oil from the rhizomes of Amomum cannicarpum. *Fitoterapia* 2006, 77, 392-394, https://doi.org/10.1016/j.fitote.2006.04.003.
- Cui, Q.; Wang, L.-T.; Liu, J.-Z.; Wang, H.-M.; Guo, N.; Gu, C.-B.; Fu, Y.-J. Rapid extraction of Amomum tsao-ko essential oil and determination of its chemical composition, antioxidant and antimicrobial activities. *J. Chromatogr. B* 2017, *1061-1062*, 364-371, https://doi.org/10.1016/j.jchromb.2017.08.001.