# POLLEN MORPHOLOGY OF THE EUPHORBIACEAE WITH SPECIAL REFERENCE TO TAXONOMY

# W. PUNT

(Botanical Museum and Herbarium, Utrecht)

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#### CHAPTER I

#### GENERAL INTRODUCTION

## a. Introduction

Many investigators have stated (e.g. LINDAU 1895, WODEHOUSE 1935, ERDTMAN 1952), that pollen morphology can be of great importance for plant taxonomy, while it was also known that in Euphorbiaceae several types of pollen grains exist (e.g. Erdtman 1952). On the suggestion of Professor Lanjouw, who himself has worked on the Euphorbiaceae of Surinam, the author has investigated the pollen grains of this family of that area. From the result it was apparent that in the Surinam Euphorbiaceae many different pollen types could be distinguished. In the present work the study was therefore extended to all the genera in the Euphorbiaceae, from which one or more species have been examined. Besides a description of the pollen grains examined a drawing of most of the pollen types is given. The pollen morphologic groupings found have been compared with the systems of several authors (Bentham and Hooker 1880, Pax and K. Hoffmann 1931, etc.). The taxonomic conclusions, however, can only have a provisional character. Future investigators of the Euphorbiaceae will have to check the taxonomic notes and comments.

#### b. Acknowledgements

The investigations have been carried out at the "Botanisch Museum en Herbarium", Utrecht. The writer is greatly indebted to the director, Professor Dr. J. Lanjouw, for giving him the opportunity to carry out the investigations at his institute and for his help and guidance in preparing this paper. The author is very much indebted to Professor Dr. F. P. Jonker who advised him in the special problems of pollen morphology. He very gratefully acknowledges the "Ministerie van Onderwijs, Kunsten en Wetenschappen" for sponsoring the present study. The author wishes to express his sincere thanks to the directors of the herbaria at Brussels, Leiden, Kew and Paris for the hospitality and help during his residence in these places. He is also very grateful to the "Miquelfonds" which covered the expenses of his stay in London, Paris and Brussels where so much valuable material could be examined.

#### CHAPTER II

## **HISTORY**

#### a. Pollen morphology

The study of pollen morphology started in 1675, when Malpighi observed, that various types of pollen grains are to be found in different plants. He especially saw variations of colour and shape. Grew (1682) was already aware of a difference in structure, too. He distinguished

smooth and spiny pollen grains. In the next two centuries quite a number of botanists enriched our knowledge of pollen morphology. In 1834 a synopsis of the literature on the subject was published by v. Mohl, which greatly contributed to a better understanding of pollen morphology. He studied representatives of most of the plant families and gave a descriptive classification of their forms. v. Mohl recognized that the apertures of the pollen grains are the most important morphological features. Some years later Fritzsche published his principal work on pollen grains (Ueber den Pollen, 1837). From the descriptions it is clear that, besides a careful study of the shape and apertures of a pollen grain, the structure was accurately analysed. His drawing and description of Jatropha panduraefolia gives an excellent idea of the pollen grain. The description even very accurately reproduces the construction of the croton-pattern (see Pl. I, E4 and Pl. VIII, 1).

In the period after v. Mohl and Fritzsche we come across publications with drawings of pollen grains as additions to analytic drawings of plants (e.g. in Martius, Flora Brasiliensis). Most botanists, however, added but little to pollen morphology. C. A. H. Fischer was the first scientist to give, in 1890, a comprehensive comparative study of the data then available. From the study of 2200 species in 158 families he was able to draw, e.g. the following conclusions:

- 1: Pollen grains of related species are generally similar.
- 2: Some families have more than a single basic form.
- 3: Sometimes unrelated plants have similar pollen grains.

In 1883 RADLKOFER made use of pollen morphology to obtain a better classification of the Acanthaceae. It was done more profoundly by LINDAU (1895) in "Die natürlichen Pflanzenfamiliën" by ENGLER and PRANTL when he stated: "Es ist deshalb nur consequent, wenn auf die Pollenbeschaffenheit die Einteilung der Acanthaceae gegründet wird, wie dies bereits von Radlkofer angedeutet wurde und im folgenden ganz streng durchgeführt werden soll". It is obvious that, with regard to some families, pollen morphology had become a wellknown and indispensable part of taxonomy at the end of the 19th century.

In fact, a renewed general interest in pollen morphology emerged, not from the side of botany, but from the side of geology. It was the geologist v. Post who, in 1919, for the first time published a modern pollen analytic diagram: the primitive pollen analysis had resulted in a refined method in quaternary geology. However, the composition of a pollen diagram requires a sound knowledge of the morphology of pollen grains. Originally a primary acquaintance of the pollen grains of trees and anemophilous plants was considered to be sufficient, but more and more the necessity of a wider knowledge of pollen types became evident. Consequently the number of investigators who made researches into the pollen morphology of special taxonomic groups increased.

POTONIÉ in 1934 and WODEHOUSE in 1935 published works on pollen morphology that formed the basis of our modern terminology.

POTONIÉ as well as WODEHOUSE needed a system of well defined terms for the description of pollen grains. The construction and details of a pollen grain were, however, not well enough known to be arranged in a convenient system. Nevertheless, many terms of these investigators are still in use, although sometimes the circumscription of the terms is more or less altered.

In 1950 FAEGRI and IVERSEN, and IVERSEN and TROELS-SMITH published a system on the morphology of pollen grains. Their terminology excelled in exactitude and simplicity. The well-defined terms made it quite possible to determine pollen grains which occur in

their determination-key of European pollen types.

At the same time ERDTMAN also worked at a system of pollen terminology. He published several papers on this subject, but his main work (Pollen morphology and plant taxonomy, Angiosperms) appeared in 1952. In this book ERDTMAN proposed an extended system of pollen terminology. In many respects, however, the terminology seems too complicated to be of practical use. Only pollen morphologists with a large experience are able to identify all the terms mentioned in the glossary. Moreover, the descriptions of terms are not always as exact as they should be. What, for instance, is the exact boundary between brevicolpate and brevissimicolpate or foramen and foraminoid? Nevertheless, his book is a source of valuable data providing primary information of the pollen types of all the families in the Angiosperms.

## b. Euphorbiaceae

This family is one of the largest in the Angiosperms. It is a difficult one on account of the many different species, so that it is not surprising, that only a few botanists possessed a general knowledge of the whole family.

Since the present report is mainly a pollen morphologic work, the following lines will only briefly comment on the principal authors

on Euphorbiaceae.

In the 19th century A. DE JUSSIEU, J. MUELLER OF ARGAU and H. BAILLON published important papers on the *Euphorbiaceae*. In the Journal of the Linnean Society (Botany) of 1880 BENTHAM reviewed the work of Baillon and Mueller Arg. In the same paper BENTHAM gave extensive notes on his own system.

From 1910–1924 Pax (partly assisted by Käthe Hoffmann) monographed most of the genera in *Euphorbiaceae* in Engler's "Das Pflanzenreich". In the second edition of Engler and Prantl, Die Natürlichen Pflanzenfamilien, Band 19c (1931), a revised synopsis of the *Euphorbiaceae* is given. In the present paper this synopsis of 1931 has been used as the basic classification of the family.

After Pax and K. Hoffmann only incidental remarks have been made on the system of the Euphorbiaceae. Most of them were given in papers concerning floras of certain areas (e.g. Léandri, Flora of Madagascar; Gagnepain, Flora of Indo-Chine; Léonard, Flora of Congo). Important improvements have been made by Croizat on

several genera. Schultes amply discussed the genus Hevea and its allies. A very important work is Webster's publication on Phyllanthus. In his monographic study "The West Indian species of Phyllanthus" he used pollen morphology as one of the main characters to divide the genus Phyllanthus into subgenera and sections.

#### CHAPTER III

#### MATERIAL

Herbarium material obtained from the following herbaria was used in the study:

London (BM): British Museum (Natural History). Jardin Botanique de l'Etat.

Bruxelles (BRLU): Herbier d'Afrique de l'Université de Bruxelles. The Herbarium, Royal Botanic Gardens. Kew (K):

Leiden (L): Rijksherbarium.

Paris (P): Muséum National d'Histoire Naturelle Laboratoire de

Phanérogamie. Botanical Museum and Herbarium. Utrecht (U):

Wageningen (WAG): Laboratory for Plant Taxonomy and Plant Geography.

Generally the use of type-material has been avoided. Only in those cases, where no other specimens were available while flowers were present, type-material has been used for pollen investigation.

The classification of the Euphorbiaceae as given by PAX and K. HOFFMANN in ENGLER and PRANTL, Die natürlichen Pflanzenfamilien ed. 2, Band 19c (1931), was the principal system used for the choice of genera. Besides this system attention has been paid to the new genera published after 1931 and listed in the Index Kewensis. As far as possible all known genera of the Euphorbiaceae were examined. In a number of genera it was, for various reasons, impossible to study the pollen grains. They fall in the following groups:

Male flowers unknown.

Croizatia Steyermark

Lasiochlamys Pax et K. Hoffmann

Kuhlman Paradrypetes Phyllanoa | CROIZAT

The material studied did not contain male flowers, or only a few, or unripe ones.

> Claoxylopsis Léandri Calicopeplus PLANCHON

THOUARS EX BAILLON Cometia

Corythea Watson Ditta GRISEBACH Keyodendron Léandri Mettenia GRISEBACH

Monadenium Pax Neomphalea Pax et K. Hoffmann
Peotrigonostemon Pax et K. Hoffmann

Ramelia BAILLON Riseleya HEMSLEY

Syndyophyllum Schumacher et Lauterbach

Sphyranthera Hooker, J. D. Thelypetalum Gagnepain

3. Genera of which no material has been seen.

Acalyphopsis
Annesijoa
Chlamydojatropha
Clarorivinia
Gitaria
PAX et K. HOFFMANN

Senefelderopsis Stevermark

On account of the large number of genera and species the author has found it impossible to be always sure of the accuracy (both taxonomic and nomenclatural) of the names used in the present paper. Future monographic studies will perhaps prove some names to be incorrect. With all the descriptions of pollen grains, however, the collector's name and number and the herbarium, where the specimen is preserved, are given. In those cases when the name has to be corrected the description of the pollen grain can thus always be placed under the right name.

In the Rijksherbarium at Leiden a large collection of plants of D'Alleizette is present. These plants, however, have not been collected by D'Alleizette himself. In the herbarium at Paris D'Alleizette obtained duplicates from several collectors. These duplicates were sent to the Rijksherbarium without mention of the names and collectioning numbers of the original collectors. As it lies beyond the scope of this work to search for the names of the original collectors, the name of

D'Alleizette has been maintained in the present work.

#### CHAPTER IV

#### **METHODS**

#### a. Flowers

From each genus at least one species and when possible two species were examined. From those genera that can be devided into two or more sections, species were chosen from different sections.

Usually the male inflorescenses of the Euphorbiaceae have many flowers, so that, generally speaking, no important loss occurs if one or two flowers are dissected.

Only those flowers of the Euphorbiaceae are useful that have just opened. When the flowers are younger the pollen has not yet ripened, so that it is impossible to give an accurate description of shape and structure of the pollen grains. It is far better to use old, outpollinated anthers than young and undoubtedly unripe ones. In the corners of their thecas the old anthers always have some pollen grains which, by a special micro-method described below, can be isolated.

#### b. Pollen preparations

For the study of the morphology of pollen grains it is desirable that cytoplasma and intine are dissolved. The microscopic image becomes obscure and the important exine remains indistinct if intine and cytoplasma are not dissolved. For that reason Wodehouse's methylene-green method, the lactic-acid method and any other method, which does not completely dissolve intine and cytoplasma are considered less suitable.

The treatment of the pollen grains with the acetolysis-method of ERDTMAN (1943) produces the best results. Everything inside the exine is dissolved, while, moreover, a colouring of the exine takes place. This method, described by ERDTMAN in 1943 and 1952 for herbarium material, has, in its original form, a great disadvantage. Too much material is required to obtain a usable preparation. In order to reduce this disadvantage a micro-method (see below) has been applied, which leaves the method as such untouched, but requires less material. If necessary even only one anther is sufficient.

#### MICRO-METHOD

The micro-method is a perfection of the method described by WILLRATH in a

publication of Ротоніє (1934).

A flower is boiled in water till it is free of air. Under a preparing-microscope one anther (or more if the material allows) is dissected and laid in the pit of a hollow slide. Some drops of acetolysis mixture (9 parts anhydric acetic acid: 1 part  $H_2SO_4$  conc.) are dropped on the anther till the pit has been filled with the mixture. Now the slide is placed on a heating-apparatus as shown in Fig. 1. The end of the heating-apparatus is heated by a Bunsenburner. The flame should not be too high,

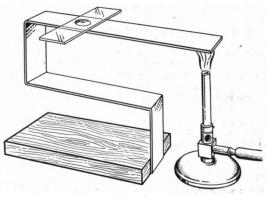


Fig. 1

but not too low either. The slide furthermore has to be removed so far from the flame that the mixture cannot take fire. A part of the mixture will evaporate. When the slide has become almost dry some more drops are added. This is repeated 3 or 4 times till the remaining mixture is turning dark brown. When the preparation once more is nearly "dry", it is quickly cooled down by placing the slide on a stone or iron surface. Under the dissecting microscope the pollen grains are to be seen as brown globules. With anthers which have a solid skin, that cannot easily be dissolved, it is desirable, while heating, to prick this skin in places with a needle to make the acetolyse mixture penetrate deeply enough into the anther.

#### c. Preservation of pollen grains

For the preservation of grains the paraffin-comprising method has been used. A small drop of glycerin (diam, 1 mm.) is laid on a slide. With a brush moisted in glycerin the pollen grains are wiped off the slide or fished out of the liquid. In

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the drop of glycerin the pollen grains come off the brush more or less easily. This is repeated till at least 10 grains are present in the glycerin drop.

Now a little piece of glycerin-gelatin is added. The gelatin is melted and the grains are mixed with it. To prevent the pollen grains from expanding abnormally the whole should not be heated too much. It is of the utmost importance that the grains can be studied from all sides, so that care should be taken that the coverglass does not flatten the grains. Therefore a granule of clay (no synthetic material) is laid next to the glycerin drop. The cover-glass is now supported by the clay when placed on the preparation and the pollen grains cannot be compressed. By lightly pressing the cover-glass the thickness of the preparation can be regulated. By running melted paraffin under the cover-glass the grains are isolated from the air.

Webster (1956) rejects the method of sealing up by paraffin on the ground of the preparations not durable. In Utrecht, however, preparations ten years old are still in excellent condition. It is, however, essential that the layer of paraffin should not be too thin and the drop of glycerin should not approach the edge of

the cover-glass (ERDTMAN and PRAGLOWSKI 1959).

Preparations made by the micro-method have the disadvantage that some acid will enter with the pollen grains. This may cause the colour to fade after some time. After three years, however, form and structure were still unchanged. For that reason the macro-method of Erdtman is always preferable to the micro-method, but the latter has the advantage that very little material is sufficient, so that any rigorous sacrifice of herbarium-material may be prevented. In the case of flowers with large anthers it is even possible that part of a stamen suffices (Campanula, Lilium).

In describing pollen grains it is of great importance that the method used for preparation should be indicated, since the size of the pollen grain varies with it. Though the expansion of a certain pollen grain is constant in either method, only relative value can be attached to the indication of the size of that particular pollen grain. Only with regard to pollen grains that have been treated in precisely the same way the grain measures retain their value. Moreover, the size is influenced by the grade of ripeness of the pollen. When a pollen grain is on the verge of maturing, form and structure may already be fully developed, while the size has not yet reached its maximum. It is, therefore, again emphasized that all absolute numbers which are mentioned in the descriptions have a relative value. For this reason any accurate calculation of average sizes has been relinquished. The figures give an impression of the size in these special circumstances and allow a mutual comparison of the examined pollen grains.

The proportionate numbers, however, are more dependable. Proportions remain the same in either method. Some proportionate numbers are, e.g., Polar Area Index (= P.A.I.) and Polar axis: Equatorial axis (= P:E) (see Pollen Morphology p. 12).

## Microscopes

The pollen grains were studied with a Bausch and Lomb binocular (objective  $97 \times$ , oculair  $10 \times$ ), and an Olympus phase contrast microscope was used. The employment of this microscope was found very useful in detecting the configuration of structure elements. Most of the structure patterns are also visible with the ordinary light-microscope, but, especially in the case of small structure elements, not without difficulty. The phase contrast microscope immediately gives a clear image of the structure.

#### e. Punched cards

The study of large quantities of material requires special methods of compiling the knowledge obtained. A simple card-index is inadequate when there are many genera to be mutually compared; the great number of characters hampers any efficient arrangement. A solution of this problem was found in the use of punched cards. Punched card systems were made for the pollenmorphologic as well as for the taxonomic characters. Thus any particular botanical object could be classified in two independent systems. The groups of genera within those systems are now mutually comparable so that it became possible to recognize correlations

of characters, which, without the punched cards, would have been found at the cost of much time and labour or not at all.

A further advantage is that the investigator is compelled to look over the same characters continually. Even the absence of a special character may be of importance and finds expression in this system.

A disadvantage may be that too much reliance is placed upon the characters noted. Often characters that are indicated in the same way are not mutually comparable. If e.g. stamens are connate, this coalescence can take various forms.

There may be investigators, who are inclined to distrust this system. Nevertheless the use of punched cards and other mechanic systems should undoubtedly be taken into consideration, especially when studying larger taxa. Surveying and working up the continually increasing numbers of characters in a reasonable time is becoming more and more difficult. Loss of time, chance of errors and the risk of being incomplete are apt to compel the taxonomist to utilize some form of mechanic taxonomy. Who would, at present, venture to revise the whole genus *Solanum* or *Euphorbia*? Only through a perfection and especially through an acceleration of our methods the larger taxa will as yet have a chance of being monographically worked on.

#### f. Drawings

The pollen grains have been drawn in such a manner that in one drawing as many characters as possible are to be seen at the same time. Each drawing therefore is midway between a scheme and a photographical reproduction. The pollen grains have been drawn without a camera lucida or other drawing instruments.

Drawing has the advantage that the most important characters can be indicated distinctly, so that the grains are easier to distinguish from others. A disadvantage is the subjective image which the drawing is apt to give.

is the subjective image which the drawing is apt to give.

The other possibility of reproduction is making use of photography. For the following reasons this method has not been applied:

- 1. In most cases at least 4 photographs of perfect quality are required to obtain a distinct picture of the grain.
- 2. In a photograph the structures of the pollen grains can never be pursued distinctly, because the photographic image will only be sharp for a certain optical section.
- 3. Deeper lying layers debase the photographic image and cause an insufficient sharpness of the essential characters.

The plates have been drawn to scale. Each picture is  $1000 \times$  enlarged, except some of the large pollen grains (e.g. *Manihot* and *Croton matourensis*), which are less enlarged. In those cases where the enlargement is not  $1500 \times$  the scale is given in the text to the plates.

#### CHAPTER V

## SOME NOMENCLATURAL REMARKS

## Bischofia Blume

Although Blume dedicated this genus to Bischoff he spelled, in his "Bijdragen" (1826), the name with a single f. Obviously he latinised Bischoff to Bischofius. For this reason the name of the genus should be written in the original spelling.

#### Briedelia Willdenow

In his "Species Plantarum" (4:978. 1805) WILLDENOW deliberately spelled the genus with *i.e.* He commented as follows on the choice of the name: "Genus hoc a Cluytia et Rhamno abunde diversum in honorem Clariss. S. E. Briedel nominavi." Although the name of the botanist in question is spelled Bridel it is not clearly demonstrated that the author made an unintentional error. Under the rules of the international code the genus should be written in the original spelling.

#### Cnesmosa Blume

In his "Bijdragen" (1825) BLUME spelled this genus as Cnesmosa. In the Flora van Java, Praesatio in adnotation, he changed the name, probably for grammatical reasons, in Cenesmon. This is not allowed under the rules of nomenclature. The original spelling should be maintained.

# Koilodepas Hasskarl

Koilodepas was first published in 1855 in the Greek version (Versl. Med. Akad. Wetenschappen Amsterdam 4: 139). Two years later the name was altered in Coelodepas (Flora 40: 531. 1857). As Croizat (1942b) and Airy-Shaw (1960) have stated, the validity of the spelling Koilodepas is unquestionable.

# Omalanthus A. DE JUSSIEU

The first spelling of this genus appeared in A. DE JUSSIEU'S "De Euphorbiacearum" in 1824. In his "Conspectus" (1828) H. G. L. REICHENBACH altered the name in Homalanthus. Of course the first spelling has to be maintained. Carumbium is a synonym of Omalanthus. Reinwardt published this name as a nomen nudum in OKEN'S Isis 1823. So Omalanthus is the name first published and therefore valid.

#### Romanoa Trevisan

In 1824 A. DE JUSSIEU published the genus Anabaena. DE BORY, however, had given the name Anabaina to a new algae genus (Cyanophyta) in 1821, so that the name of A. DE JUSSIEU is a homonym. In 1918 PAX et K. HOFFMANN gave the substitute name Anabaenella to this taxon. They did not know, however, that an older name was available. In 1848 TREVISAN had substituted Anabaena A. DE JUSSIEU by Romanoa (Alghe Coccotale p. 99). If the genus is held separate from Plukenetia, Romanoa is the first correct name for Anabaena A. DE JUSSIEU.

#### CHAPTER VI

#### POLLEN MORPHOLOGY

In describing pollen grains the number of terms employed should be as small as possible. This was the starting point of FAEGRI and IVERSEN in 1950. One of the arguments they advanced was the difficulty of giving an analytic description of a pollen grain in exact terms. The system, however, that resulted from their attempt at

#### PLATE I.

- A. 1. pori; 2. colpi; 3. prolate pollen grain; 4. oblate pollen grain; P = Polar axis;
   E = Equatorial axis.
- B. polar view; 1. triangular; 2. three lobed; 3. circular; 4. convex triangular;
   S = smallest distance between two colpi ends; G = greatest breadth; S:G = Polar Area Index.
- C. 1. porus and colpus nudate; 2. p. and c. with a membrana granulata; 3. p. and c. with an operculum; 4. p. and c. with a margo (thickening a or thinning b of the ekt.); 4c. costae pori and costae colpi; 5a. porus with a vestibulum; 5b and c. composite aperture with an atrium.
- D. 1. colpus transversalis; 2. circular colp. transv.; 3. equatorial colp. transv.; m = meridional axis of the cop. transv.; e. equatorial axis of the colp. transv.; o = os; 4. costa transversalis; 5. costa circularis; 6. costa equatorialis.
- E. 1. tectum; 2. tectum perforatum; 3. reticulum; 4. croton-pattern.
- F. 1. psilate; 2. scabrate; 3. verrucate; 4. gemmate; 5. clavate; 6. baculate; 7. echinate; 8. pila.

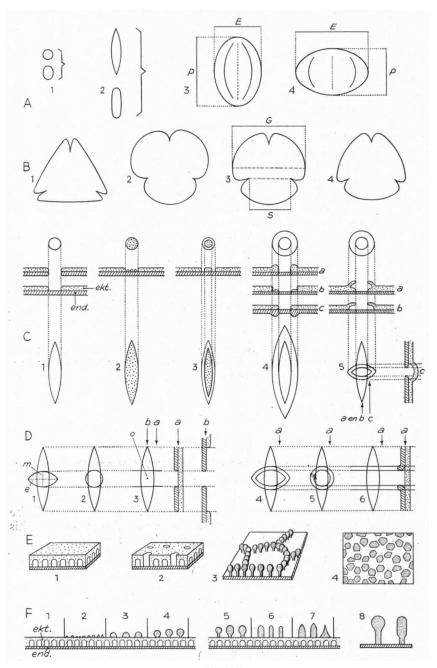


PLATE I.

simplicity was inadequate, and amplification was necessary. Especially when the amount of material to be described is considerable many difficulties are liable to arise. The number of characters of a pollen grain is comparatively small and they often appear in many transitional forms. Yet a precise and exact definition of characters is desirable.

ERDTMAN, who, in his study of the Angiosperms (1952), met with these difficulties, therefore enlarged the terminology considerably. He tried to nominate every transitional form, each nuance, which could be achieved only at the cost of exactitude and convenient arrangement. Nevertheless many pollen morphologists will feel highly attracted towards his method.

For the use of pollen morphology as an additional character in taxonomy this system is too complicated. The more obvious characters will suffice. For that reason a compromise seems to be the right course for the description of diverse pollen grains. Without going as far as ERDTMAN the present author has attempted to use a terminology as simple as was possible without loss of exactitude and completeness.

Pollen grains are composed of three layers. The central part is the cytoplasma. The middle layer is the *intine*; the outer layer the *exine*. The exine of pollen grains consists of a particularly resistant substance of a lipoid character: *sporopollenin*. Sporopollenin is even more resistant to the attack by micro-organisms than cutin. The exine substance is also highly resistant to all sorts of chemicals; it can not, or only slowly, be attacked by fluid KOH, concentrated H<sub>2</sub>SO<sub>4</sub> or even fluoric acid (HF).

The exine which can be devided in two layers, the *endexine* and the *ektexine*, supplies most of the data necessary for pollen description. These data fall into three groups:

- 1. Shape
- 2. Apertures
- 3. Structure

#### 1. Shape

If a pollen grain is not spheroidal we can distinguish two axis:

- a. Polar axis (P) (Pl. I, A3 and A4)
- b. Equatorial axis (E) (Pl. I, A3 and A4)

In equatorial view pollen grains show a shape which will be determined easily now by calculating the P: E. According to this relation we distinguish the following classes (ERDTMAN 1952):

```
P: E > 2
             perprolate
  1,33-2
              prolate (Pl. I, A3)
  1,14-1,33:
              subprolate
      -1,14:
              prolate spheroidal
              spheroidal
      -0,88:
              oblate spheroidal
  0,88-0,75:
              suboblate
  0,75–0,50:
              oblate (Pl. I, A4)
     < 0,50: peroblate
```

In polar view the circumference of a pollen grain can be:

- 1. triangular (Pl. I, B1)
- 2. convex triangular (Pl. I, B4)
- 3. circular (Pl. I, B3)
- 4. three lobed (Pl. I, B2)

## 2. Apertures

In the system of IVERSEN and TROELS-SMITH the principal classification is made according to the occurence of apertures: pori (Pl. I, A1), colpi (Pl. I, A2) and colpi transversales (Pl. I, D1, 2, 3) (in composite apertures only).

Real pori and real colpi are formed by thinnings in the ektexine. A colpus transversalis, on the other hand, by a thinning in the endexine. For that reason apertures of the endexine and the ektexine have to

be described independently of each other.

Pori can be distinguished from colpi by calculating the length: breadth. If length: breadth < 2 the aperture is a porus although the shape is not necessarily a pure circle. If length: breadth > 2 the aperture is a colpus. The endexine below the edges of colpi or pori may be thickened. These thickenings are called costae colpi and costae

pori (Pl. I, C4c).

The colpus transversalis (Pl. I, D) is an easily recognisable character. It is a thinning of the endexine. Yet this aperture has been inaccurately described by many investigators. FAEGRI and IVERSEN (1950) confuse this aperture with a real porus when the colpus transversalis has the shape of a circle. Consequently, when using their key for the determination of European pollen types, it is sometimes difficult to decide if a "porus longitudinal elongated" or a "transversal furrow" is present. Erdtman calls colpi + colpi transversales "composite apertures". For the term colpus transversalis he introduced the new term os (pl. ora). If this term is used in that sense the name will have to be rejected as being a synonym. It may, however, be maintained for that part of the colpus transversalis that crosses the colpus (Pl. I, D3). Colpi transversales stand perpendicular on the colpi. Usually they are boat-shaped. Sometimes they fuse together into one single aperture along the equator of the pollen grain (colpus transversalis equatorialis), but it is also possible that the colpus transversalis becomes isodiametric (colpus transversalis circularis Pl. I, D2). The medial dimension of the colpus transversalis is rarely larger than the equatorial one (*Pleiostemon* type, p. 30). Edges of colpi transversales can also have thickenings; according to the shape of the colpus transversalis they are called: costae circulares, costae transversales and costae equatoriales (Pl. I, D4, 5, 6). To express the proportions of the colpus transversalis in exact figures, the quotient m:e is added to the description of this character. In this formula m is the meridional axis and e the equatorial axis of the colpus transversalis (Pl. I, D1).

IVERSEN and TROELS-SMITH (1950) introduced the term *Polar Area Index (P.A.I.)*. The *P.A.I.* is the ratio between the greatest distance of the ends of two colpi and the greatest breadth of the pollen grain

usually the Equatorial axis. This character is of great value for the description of pollen grains.

#### 3. Structure

The *ektexine* furnishes characters of great diagnostic importance. The description of these characters meets, however, with considerable difficulties, so that it is necessary to use a system as simple as possible and highly desirable to avoid confusing terms (e.g. *sculpture*, see below).

The ektexine is composed of structure elements. Different forms and arrangements of these elements cause several structure types to occur. The most common structure elements occuring on the endexine are pila (Pl. I, F8). Each pilum consists of a swollen apex (caput) and a cylindrical basal part (collum; homologous columella). Pila are strictly found on the endexine. They may occur ordinate or inordinate. Ordinate arranged pila can form regular figures (e.g. reticulum Pl. I, E3) or stand in rows (e.g. striae). Frequently capita of pila are connate or fused. If the fusion is only laterally, and less of the surface is covered by the capita, we speak of tegillate pollen grains. Pollen grains are tectate when the fused capita cover 80 % or more of the surface (Pl. I, E1). In that case the pila form a second membrane outside the endexine. If the tectum is not completely closed (fusion of the capita more than 80 % and less than 100 %) the perforate membrane is a tectum perforatum (Pl. I, E2).

On the top of the tectum we distinguish structure elements of different shape. According to FAEGRI and IVERSEN they are called: baculae, clavae, echinae, gemmae, scabrae and verrucae (Pl. I, F2 t/m 7) (for descriptions see glossary). Psilate pollen grains have no structure elements on the tectum (Pl. I, F1).

Structure elements on the top of the tectum may also be arranged in different structure types (reticulum, striae). If a tectum is present, corresponding structure types of the columellae are called intra-reticulum, intra-striae. Sometimes columellae can give an impression of a reticulum. This is, however, a "pseudo-reticulum" in which each columella represents a "lumen" and the space between the columellae, the "muri". A real reticulum is always formed by several structure elements.

In the present study no difference is made between structure and sculpture. According to Potonie (1934), all elements outside the tectum form the sculpture of a pollen grain. Potonie tried to give sharp definitions of these terms, but admitted at the same time, that structure and sculpture pass into each other. FAEGRI and IVERSEN (1950) also stated, that structure and sculpture are easy to confuse. For that reason it seems better to speak of structure elements and structure only.

#### CHAPTER VII

#### **GLOSSARY**

- Annulus (IVERSEN and TROELS-SMITH 1950) (Pl. I; C 4a en b)
  - Area which surrounds a porus and is distinguished by a thickening or a thinning of the ektexine; thus forming a prominent or depressed margin of the latter.
- Aperture (FAEGRI and IVERSEN 1950)
  - Any weak performed part of the ektexine or endexine of a pollen grain (e.g. colpus, colpus transversalis).
- Atrium (Pl. I; 5a, b en c)
  - A cavity inside a composite aperture caused by a separation between two layers of the exine.
- Bacula pl. Baculae (IVERSEN and TROELS-SMITH 1950) (Pl. I; F 6) Structure elements with at least one dimension 1  $\mu$  or larger in the shape of rods. Occur on the tectum only.
- Baculate (IVERSEN and TROELS-SMITH 1950)
  Pollen grains provided with baculae.
- Caput pl. Capita (ERDTMAN 1952) Upper part of a pilum.
- Clava pl. Clavae (IVERSEN and TROELS-SMITH 1950) (Pl. I; F 5)
  Structure elements with at least one dimension 1 μ or larger in the shape of clubs. Occur on the tectum only.
- Clavate (IVERSEN and TROELS-SMITH 1950)
  Pollen grains provided with clavae.
- Collum (ERDTMAN 1952)

Lower part of a pilum; homologous with columella.

- Columella pl. Columellae (IVERSEN and TROELS-SMITH 1950)
  - Lower part of a pilum. Columellae bear the tectum. In 1956 FAEGRI used the term as a synonym for pilum.
- Colporate (ERDTMAN 1945)

Pollen grains with composite apertures.

- Colpus pl. Colpi (Erdtman 1943) (Pl. I; A2 and C1)

  Longitudinal apertures of the ektexine. Length: breadth > 2.
- Colpus transversalis (ERDTMAN 1943) (Pl. I; D 1, 2, 3)
  - Aperture of the endexine perpendicular to the colpus. Appears in composite apertures only.
- Composite aperture (ERDTMAN 1952) (Pl. I; D 1, 2, 3)

  Ekterine aperture combined with an endevine aperture (aper-
  - Ektexine aperture combined with an endexine aperture (apertures not congruent).
- Costa pl. Costae (FAEGRI and IVERSEN 1950) (Pl. I; C 4 and D 4, 5, 6) Thickenings of the endexine.
  - Costae circulares (D5): Thickened edges of circular colpi transversales.

Costae colpi (C4c): Thickened endexine below the edge of colpi.

Costae equatoriales (D6): Thickened edges of colpi transversales extended along the equator.

Costae pori (C4c): Thickened endexine below the edge of pori. Costae transversales (D5): Thickened edges of colpi transversales.

Croton-pattern (ERDTMAN 1952) (Pl. I; E4)

Structure elements in rings of 5 or 6 around a circular area.

Echina - pl. Echinae (WODEHOUSE 1928) (Pl. I; F7)

Structure elements with at least one dimension 1  $\mu$  or larger in the shape of spines. Occur on the tectum only.

Echinate (WODEHOUSE 1928)

Pollen grains provided with echinae.

Ellipsoid

Inaperturate pollen grains with a long and a short axis.

Endexine (ERDTMAN 1943)

Inner part of the exine.

Ektexine (ERDTMAN 1943)

Outer part of the exine.

Equatorial axis (VAN ZINDEREN BAKKER 1953) (Pl. I; 3E and 4E)

The line perpendicular to the Polar axis in the equatorial plane.

Exine (FRITZSCHE 1837)

The outer, usually resistant, layer of the wall of a pollen grain.

Gemma - pl. Gemmae (IVERSEN and TROELS-SMITH 1950) (Pl. I; F4)

Structure elements with at least one dimension 1  $\mu$  or larger of which the lower parts are constricted. Occur on the tectum only.

Gemmate (IVERSEN and TROELS-SMITH 1950)

Pollen grains provided with gemmae.

Inaperturate (IVERSEN and TROELS-SMITH 1950)

Pollen grains without apertures.

Intectate (IVERSEN and TROELS-SMITH 1950)

Capita of the pila covering less than 80 % of the surface.

Intra-reticulate (FAEGRI and IVERSEN 1950)

Columellae inside the tectum form a network.

Intra-striate (FAEGRI and IVERSEN 1950)

Columellae inside the tectum stand in rows.

Lumen - pl. Lumina (POTONIÉ 1934)

Spaces between the muri of a reticulum.

Margo (Iversen and Troels-Smith 1950)

Area which surrounds a colpus and is distinguished by a thickening or thinning of the ektexine; thus forming a prominent or depressed margin of the latter.

Membrana granulata (ERDTMAN 1952) (Pl. I; C2)

Colpus or porus membrane (= endexine) with some scattered structure elements.

Murus - pl. Muri (Potonié 1934)

Ridges separating the lumina of a reticulum.

Operculum (Wodehouse 1928; FAEGRI and IVERSEN 1950) (Pl. I; C3)

Isolated part of the ektexine which is separated from the edges of a colpus or porus by a narrow zone in which the ektexine is missing or greatly reduced.

Os - pl. Ora (Pl. I; D3, o)

That part of a composite aperture which is formed by the crossing of the colpus and the colpus transversalis.

Periporate (IVERSEN and TROELS-SMITH 1950)

Pori ± uniformly distributed over the surface of a pollen grain.

Pilum - pl. Pila (ERDTMAN 1952) (Pl. I; F8)

Structure elements consisting of a swollen apex (caput) and a cylindrical basal part (collum, columella). Occur on the endexine only.

Polar Area Index = P.A.I. (IVERSEN and TROELS-SMITH 1950) (Pl. I; B3)

The proportion of the greatest distance between the ends of two furrows and the greatest breadth of the pollen grain (usually the Equatorial axis).

Polar axis (ERDTMAN 1943) (Pl. I; A3p and A4p)

The perpendicular line connecting the poles of a pollen grain.

Porus - pl. Pori (Erdtman 1943) (Pl. I; Al and Cl)

Circular apertures of the ektexine. Length: breadth < 2.

Psilate (Wodehouse 1928) (Pl. I; Fl) Surface of tectum smooth.

Reticulum (POTONIÉ 1934) (Pl. I; E3)

Structural pattern in the form of a network in which columel!ae represent the muri of the reticulum.

Scabra - pl. scabrae (Iversen and Troels-Smith 1950) (Pl. I; F2)

Structure elements visible but smaller than 1  $\mu$ . Occur on the tectum only.

Scabrate (IVERSEN and TROELS-SMITH 1950)

Pollen grains provided with scabrae.

Sculpture (POTONIÉ 1934)

A term proposed by Potonié for the joint structure elements outside the tectum (p. 14).

Stephanocolpate (IVERSEN and TROELS-SMITH 1950)

Pollen grains with more than three colpi perpendicular to the equatorial plane.

Stephanocolporate (IVERSEN and TROELS-SMITH 1950)

Pollen grains with more than three composite apertures.

Stephanoporate (IVERSEN and TROELS-SMITH 1950)

Pollen grains with more than three apertures in the equatorial plane.

Stria - pl. Striae (Iversen and Troels-Smith 1950)

Structure elements standing in rows forming narrow grooves (± parallel).

Striate (Iversen and Troels-Smith 1950)

Pollen grains with striae.

Structure

Form and arrangement of the structure elements.

Structure elements

Individual elements either on the endexine or on the tectum.

Syncolpate (Iversen and Troels-Smith 1950)

Colpi anastomosing at the poles.

Tectate (Iversen and Troels-Smith 1950) (Pl. I; E1)

The capita of pila are fused and cover together 80 % or more of the surface of a pollen grain.

Tectum (Iversen and Troels-Smith 1950) (Pl. I; E1)

The membrane outside the endexine formed by fused capita of pila.

Tectum perforatum (IVERSEN and TROELS-SMITH 1950) (Pl. I; E2)

The tectum is not completely closed. Small holes are present.

Tegillate (ERDTMAN 1952)

The capita of the pila are laterally united or fused and cover less than 80 % of the surface of a pollen grain.

Tricolpate (Iversen and Troels-Smith 1950)

Pollen grains with three colpi perpendicular to the equatorial plane.

Tricolporate (IVERSEN and TROELS-SMITH 1950)

Pollen grains with three composite apertures.

Triporate (IVERSEN and TROELS-SMITH 1950)

Pollen grains with three pori in the equatorial plane.

Verruca - pl. Verrucae (IVERSEN and TROELS-SMITH 1950) (Pl. I; F3)

Structure elements with at least one dimension 1  $\mu$  or larger in the shape of warts. Occur on the tectum only.

Verrucate (IVERSEN and TROELS-SMITH 1950)

Pollen grains provided with verrucae.

Vestibulum (Iversen and Troels-Smith 1950) (Pl. I; C5a, b)

Cavity inside a porus, caused by a separation between two layers of the exine.

#### CHAPTER VIII

#### RESULTS

#### A. POLLEN GRAINS OF THE EUPHORBIACEAE

Pollen grains of the Euphorbiaceae show a number of types, which can be distinguished more or less clearly. Generally pollen grains of species inside one genus belong to the same pollen type. The dimensions and proportions may differ, but in principle the pollen grains are identical. In some larger genera different pollen types occur in one genus (Tragia, Phyllanthus). More often we find that different genera possess the same pollen type. Consequently it is not possible, as it is

with the Acanthaceae, to place a plant instantly in a genus by means

of its pollen grains.

For the greater part the pollen types within the subfamilies *Phyllanthoideae* and *Crotonoideae* can be distinguished clearly from each other. Some types, however, show some affinities; e.g. *Clutia* type (p. 77) with *Phyllanthus pentaphyllus* subtype (p. 24), and *Amanoa* type (p. 33) with *Sumbavia* type (p. 69).

According to the system of IVERSEN and TROELS-SMITH (1950) the

following principal types occur:

inaperturate, periporate, triporate, stephanoporate, tricolpate, stephanocolpate, tricolporate, stephanocolporate.

Pollen grains with composite apertures occur most frequently.

Especially the number of tricolporate pollen grains is large.

The shape varies from perprolate (P: E > 2,00) to oblate (P: E = 0,50-0,75). In polar view the pollen grains are mostly convex triangular. In some cases the circumference is triangular (*Clutia* type), circular (*Moultonianthus* type) or three lobed (*Omalanthus nutans* subtype).

The colpi are usually narrow, but sometimes broad (Amanoa type, Plukenetia type). In many types the colpi are accompanied by costae

colpi (Antidesma type, Mallotus type).

The P.A.I. diverge from large (> 0,7) to syncolpate. Syncolpate pollen grains are, however, rarely found in the *Euphorbiaceae* (Amperea type).

Colpi transversales, if present, can be of great diagnostical value. Shape and size are characteristic for several pollen types (Securinega

type, Hippomane type).

Pollen grains of Euphorbiaceae are often tectate. Sometimes the columellae supporting the tectum are so small that they are barely visible (Alchornea type, Ricinus type). If the pollen grains are intectate, they generally possess a reticulum. In a few cases only the pollen grains are intectate, pilate (some Phyllanthus species, Tragia fallax type). On the tectum structure elements of different shape may be found; e.g. clavae, baculae, echinae, etc.

The croton-pattern is a structure type which we meet in many genera of the Crotonoideae. Outside the Euphorbiaceae this structure type

is found in Buxaceae and Thymelaeaceae (ERDTMAN 1952).

In the following treatment of the investigated material the pollen grains are first divided into the two subfamilies:

a. Phyllanthoideae

b. Crotonoideae

Inside these subfamilies the pollen grains are discussed according

to their similarity in type.

Different pollen grains are placed in different pollen types. So the type unit has been chosen as the base of the classification. Descriptions of a type are therefore as extensive as possible. If the differences are

of minor importance, the pollen grains are placed in *subtypes*. Several types can have some characters in common. To express the correspondences, these types are assembled in *configurations*. After the diagnoses of the pollen groups comment is given on the pollen morphologic characters, mostly in their relation to other pollen groupings. Besides this comment in many cases a taxonomic discussion is added, in which the value of pollen morphology in taxonomic problems is discussed.

## B. DISCUSSION OF THE RESULTS

#### a. Phyllanthoideae

#### ANTIDESMA CONFIGURATION

Tricolporate or stephanolcolporate (4). Pollen grains with a Polar axis larger than the Equatorial axis. P. rarely shorter than the E. (e.g. *Phyllanthus acidus* subtype).

Colpus transversalis usually with costae.

Pollen grains reticulate or not reticulate; never echinate.

The Antidesma configuration is characterised by its prolate shape of the pollen grains. The Polar axis is usually larger than the Equatorial axis. In the Phyllanthus acidus subtype (p. 26) only the pollen grains are spheroidal or slightly oblate spheroidal. However, the pollen grains resemble so much the Securinega subtype, that it seems better to place them in the same type.

The three main types of the configuration are the Antidesma type, the Securinega type and the Dicoelia type. The three types are closely related and have several characters in common e.g. shape, costae colpi and costae transversales. The other types resemble the main types but lack one of the important characters e.g. costae (Heywoodia type, p. 30) or have special characters (verrucae in the Zimmermannia type, p. 29).

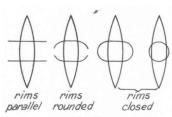


Fig. 2

The differences between the subtypes are rather faint and difficult to define and separate. The most reliable character for determination and separation is found in the colpus transversalis. These apertures are bordered by costae which make them distinctly visible. In some subtypes (e.g. Antidesma subtype) the rims of the costae transversales run parallel and the outer ends of the colpus transversalis are diffuse. Most Phyllanthus species have costae transversales with rounded ends. The outer ends of the colpus transversalis are in that case distinct and frequently even closed.

## Antidesma type

Tricolporate; perprolate to prolate spheroidal. Colpus transversalis small or narrow elongated; costae.

Colpi narrow; costae colpi.

Tectate; psilate. Mostly not reticulate; rarely indistinct reticulate.

The Antidesma type differs from the Dicoelia type (p. 28) by the shape and dimensions of the colpus transversalis. In other characters the two types cannot be differentiated.

From the Securinega type the Antidesma type differs not only by the shape of the colpus transversalis but also by the structure of the exine.

#### Taxonomic discussion

The plants in the Antidesma type can be divided into two groups. The first group combines the Antidesma subtype with the Baccaurea subtype and the second group includes the two Phyllanthus subtypes.

The first group comprises most of the genera which Pax and K. HOFFMANN (1922, 1931) united in the subtribe Antidesminae. The Antidesminae are chiefly characterised by the inflorescense which is catkin-like or in slender spikes. The following characters are of importance too:

- The male flowers have only few (six or less; usually five) stamens round an ovarium rudiment (in Aporosa the ovarium rudiment is small or absent).
  - Usually a disc is present.
- In the female flowers a disc may be present; the styles are usually bifid.
- The plants are nearly almost dioecious. All the genera are found in the Old World except Hyeronima, which is the American representative of Antidesma (Bentham, 1880).

Chonocentrum was first recognised by Mueller as a species of Drypetes. Pax and K. Hoffmann (1922, 1931) separated the plant from this genus and placed it in the vicinity of Discocarpus. Pollen grains of the latter genus are however quite different. Chonocentrum differs from Discocarpus by the following characters:

- Petals are never present.
- The stamens are not connate but free.

Some genera included in the Antidesminae by PAX and K. HOFF-MANN possess pollen grains without an Antidesma type. As for the genera Dicoelia, Richeria and Richeriella they belong to related types. Martretia and Hymenocardia have totally different pollen grains.

The second group of the Antidesma type includes many species of Phyllanthus. Also pollen grains of Reverchonia possess the Phyllanthus pentaphyllus subtype. In many respects the genus is related to Phyllanthus. The branching of the plants, however, is not phyllanthoid. The male disc, always present in *Phyllanthus*, is rather indistinct. The two stamens are inserted on a thick disc-like tissue. As in Phyllanthus there is no ovarium rudiment, while in the female flowers a disc is present.

## Antidesma subtype

Pollen grains tricolporate; perprolate – prolate. Colpus transversalis long; outer ends indistinct and diffuse. Edges of the costae transversales parallel.

The pollen grains are strongly elongated and have narrow elongated

colpi transversales.

Though always referred to the genus Securinega, Securinega congesta has pollen grains largely different from the other pollen grains in the genus. The large P: E and narrow elongated colpus transversalis place the pollen grains straight into the Antidesma subtype.

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Antidesma bunius (LINN.) SPRENG.
Java [U] Pl. II, 2
                                                  Antidesma subtype; perprolate.

P = 37,5 \mu E = 18,5 \mu P:E = 2,04.

m:e < 0,5 P.A.I. = 0,25.
                                                  Antidesma subtype; prolate.

P = 19 \mu E = 13 \mu P:E = 1,50.
Antidesma venosum Tul.
   Stolz 476 [U]
Antidesma agusanense Elmer
                                                  Antidesma subtype.
   Elmer 15881 [U]
                                                     Idem Antidesma venosum.
Antidesma diandrum (ROXB.) ROTH
Hohenacker 167 [U]
Antidesma edule MERR.
                                                  Antidesma subtype.
                                                     Idem Antidesma venosum.
                                                  Antidesma subtype.
   Ramos 1083 [U]
                                                     Idem Antidesma venosum.
Antidesma ghesaembilla GAERTN.
Hort. Calcutta 1897 [U]
                                                  Antidesma subtype.
                                                     Idem Antidesma venosum.
Antidesma membranaceum Muell. Arg.
                                                  Antidesma subtype.
   Bels 126 [U]
                                                     Idem Antidesma venosum.
Antidesma obovatum J. J. SMITH
Versteeg 1789 [U]
                                                   Antidesma subtype.
                                                     Idem Antidesma venosum.
Hyeronima alchorneoides Allem.
                                                   Antidesma subtype.
Krukoff 6110 [U]

Hyeronima laxiflora (Tul..) Muell. Arg.
Sandwith 544 [U] Pl. II, 1
                                                     P = 28.5 \mu E = 12 \mu P:E = 2.20.
                                                   Antidesma subtype.
                                                  Antidesma subtype.
Thecacoris leptobotrya (MUELL. ARG.)
   Brennon
Zenker 23 [U]
Cyathogyne spathulifolia PAX
                                                     P = 36 \mu E = 17.5 \mu P:E = 2.04.
                                                   Antidesma subtype.
                                                     P = 40 \ \mu \ \dot{E} = 20.5 \ \mu \ P:E = 1.94.
   Schlieben [P]
                                                        m:e \pm 0,\overline{5}.
Cyathogyne viridis Muell Arg.
                                                   Antidesma subtype.
   F.H.I. 3936 [K]
                                                     Too young for measurements.
                                                  Antidesma subtype.

P = 30 \mu E = 19 \mu P:E = 1,58.
Apodiscus chevalieri HUTCH.
   Pobéquin 1909 [P]
Protomegabaria macrophylla Нитсн.
                                                   Antidesma subtype. P = 27.5 \mu E = 19 \mu P:E = 1.45.
   Andoh 5474 [K] Pl. II, 3
                                                   Antidesma subtype.

P = 26 \mu E = 18,5 \mu P:E = 1,40.
Spondianthus preussii Engl.
   Versuchsanst, 164 [U]
Maesobotrya floribunda Вентн.
                                                  Antidesma subtype.

P = 21 \mu E = 15 \mu P:E = 1,40.
Lebrun 1039 [U]
Maesobotrya dusenii (PAX) HUTCH.
Zenker 221 [U]
                                                   Antidesma subtype.
Securinega congesta MUELL. ARG.
                                                   Antidesma subtype.
   Black 2432 [U]
                                                     P = 45 \mu \dot{E} = 26 \mu P:E = 1,73.
                                                        m:e < 0.5.
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## Baccaurea subtype

Pollen grains tricolporate; subprolate -- prolate sphaeroidal. Colpus transversalis small. Outer ends indistinct and diffuse. Edges of the costae transversales parallel.

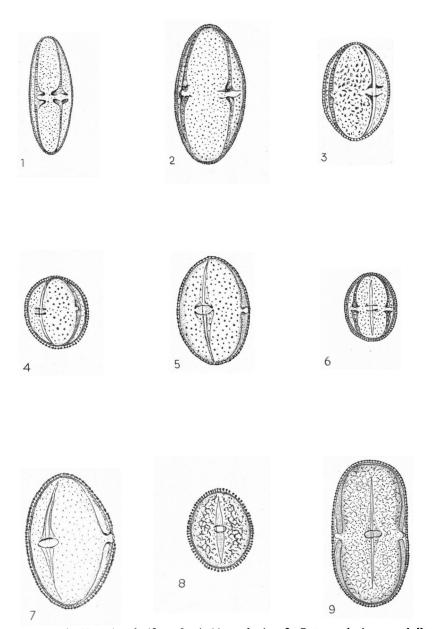


PLATE II. 1. Hyeronima laxiflora; 2. Antidesma bunius; 3. Protomegabaria macrophylla; 4. Aporosa lindleyana; 5. Phyllanthus pentaphyllus; 6. Baccaurea sumatrana; 7. Reverchonia arenaria; 8. Margaritaria nobilis; 9. Phyllanthus niruri.

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In its exine structure this subtype is related to the Antidesma subtype. The pollen grains, however, are less elongated and the colpus transversalis is smaller.

Baccaurea sumatrana (MIQ.) MUELL. ARG. Hort. Bog. VI C. 185 A [U] Pl. II, 6

Baccaurea javanica (BLUME) MUELL. ARG. Hort. Bog. 344 [U] Baccaurea racemosa (Reinw. ex Blume) Muell. Arg. Hort. Bog. 553 [U] Baccaurea oxycarpa GAGNEP. (= Gatnaia annamica GAGNEP.) Eberhardt 3042 [P] Aporosa lindleyana (WIGHT) BAILL. Hohenacker 352 [U] Pl. II, 4

Aporosa dioica (ROXB.) MUELL. ARG. King 298 [Ù] Aporosa frutescens Blume Herb. Utrecht 022876

Collector unknown.

Baccaurea subtype; subprolate. P = 19  $\mu$  E = 15  $\mu$  P:E = 1,33. P.A.I. = 0,35 - 0,3. Baccaurea subtype. Idem Baccaurea sumatrana. Baccaurea subtype.

Idem Baccaurea sumatrana. Baccaurea subtype.

 $P = 21 \mu E = 16 \mu P:E = 1,31.$ Baccaurea subtype.  $P = 19 \mu E = 17,5 \mu P:E = 1,10.$ P.A.I. = 0,2 - 0,25Baccaurea subtype. Idem Aporosa lindleyana. Baccaurea subtype. Idem Aporosa lindleyana.

# Phyllanthus pentaphyllus subtype

Pollen grains tricolporate; prolate – subprolate. Colpus transversalis small; outer ends almost distinct. Edges of the costae transversales rounded at the end.

The difference with the Antidesma subtype and Baccaurea subtype is found in the colpus transversalis.

There is also a relation with the *Phyllanthus niruri* subtype. The latter subtype, however, is 4 — colporate.

Phyllanthus pentaphyllus Wright ex Grisebach Boldingh 7378 [U] Pl. II, 5
Phyllanthus sublanatus Schum. et Thonn. de Wit 1010 [WAG]

Phyllanthus stipulatis (RAF.) WEBSTER Geyskes 13 [U] Phyllanthus amarus Schum. B. W. 10 [U] de Wit 474 [WAG] Reverchonia arenaria A. GRAY Warnock 10723 [U] Pl. II, 7

Savia erytroxyloides GRISEB. Wright 1434 [BM]

Phyllanthus pentaphyllus subtype; prolate.  $P = 32 \mu$   $E = 22.5 \mu$  P:E = 1.43.  $P = 32 \mu$   $E = 22.5 \mu$  m: e = 0.5 P.A.I. = 0.3. Phyllanthus pentaphyllus subtype.  $P = 45 \mu E = 30 \mu \dot{P} : E = 1,50.$ P.A.I. = 0,45.Phyllanthus pentaphyllus subtype.  $P = 24 \mu E = 17.5 \mu P:E = 1.37.$ Phyllanthus pentaphyllus subtype.  $P = 35,5 \mu E = 27 \mu P:E = 1,32.$ 

Phyllanthus pentaphyllus subtype.  $P = 37 \mu E = 30 \mu P:E = 1,60.$  m:e = ca. 0,4. Phyllanthus pentaphyllus subtype.  $P=32~\mu~E=26~\mu~P:E=1,24.$  m:e<0.5~P.A.I.=0,25.

# Phyllanthus niruri subtype

Pollen grains stephanocolporate (4); prolate. Colpi transversales; costae transversales. Edges of the colpus transversalis rounded at the end.

Except for the number of colpi the *Phyllanthus niruri* type is in all its characters identical with the Phyllanthus pentaphyllus subtype.

As in the *Phyllanthus pentaphyllus* subtype the columellae are sometimes fairly coarse. The surface pattern seems to form a reticulum. This is however a "pseudo-reticulum" (p. 14). Only in *Phyllanthus niruri* a real reticulum formed by a regular arrangement of several columellae was seen. This reticulum was only to be seen with a phase-contrast microscope and then very indistinctly.

Phyllanthus niruri LINN. Fuertes 89 [U] Pl. II, 9

Phyllanthus guianensis Klotz. B.W. 4599 [U] Phyllanthus hyssopifolioides H.B.K. Focke 1278 [U] Phyllanthus urinaria Linn. Lanjouw 246 [U]

Chonocentrum cyathoforum (MUELL. ARG.)
PIERRE ex PAX et K. HOFFMANN
Spruce 3781 [P]

Phyllanthus niruri subtype.  $P=39~\mu~E=21~\mu~P:E=1,86.$  P.A.I. > 0,5. Indistinct intra-reticulate. Phyllanthus niruri subtype.  $P=36~\mu.$  Phyllanthus niruri subtype.  $P=30~\mu~E=18~\mu~P:E=1,67.$  Phyllanthus niruri subtype. Too young.

Antidesma type; tricolporate.

Colpus transversalis with costae.

P:E > 1.

Too young to give it a definite place in the Antidesma type.

# Securinega type

Tricolporate or stephanocolporate (4); subprolate – sphaeroidal (rarely oblate sphaeroidal).

Colpus transversalis small, circular or broad elliptic; costae.

Colpi narrow; costae colpi usually developed. Intectate: distinctly reticulate; lumina 1-2  $\mu$ .

Pollen grains small (smaller 30  $\mu$ , usually 15-25  $\mu$ ).

This type has small pollen grains with circular or broad elliptic colpi transversales. The reticulum is distinct and usually fairly coarse in comparison with the small pollen grains.

Taxonomic discussion

The Securinega type comprises, besides the genus Securinega (and Flueggia), many species of Phyllanthus and the small genera Margaritaria and Astrocasia. These genera are all closely related (Webster, 1956).

The genus Aporosella, in Pax's system placed close to Aporosa, should be reduced to a section of Phyllanthus according to Webster (1956). One of the principal reasons which led him to this conclusion is the close relation to Phyllanthus acidus. In fact, the pollen grains of Phyllanthus elsiae, belonging to the section Aporosella, are closely related to those of Phyllanthus acidus (p. 26).

Securinega subtype

Pollen grains subprolate – prolate sphaeroidal. Colpus transversalis circular or broad elliptic.

Pollen grains of the Securinega subtype have always a P: E > 1. The shape of the colpus transversalis is circular to broad elliptic.

Securinega suffruticosa (Pallas) Rehder Securinega subtype. Cantonspark (Baarn) Securinega subtype.  $P = 22,5 \mu E = 21 \mu P:E = 1,08.$  P.A.I. = 0,25 — 0,3. Lumina 1-2  $\mu$ .

Securinega ramiflora (AITON) MUELL. ARG. Linsley Gressitt 229 [U]
Securinega neopellandra (GRISEB.) URBAN ex Pax et K. Hoffmann Shafer 12080 [U]
Securinega virosa (ROXB. ex WILLD.) BAILL. Reichs-Kolon. 506 [U] Pl. III, 2
Margaritaria nobilis LINN. F. Soeprata 2D [U] Pl. II, 8

Astrocasia tremula (GRISEB.) WEBSTER Gaumer et Sens 1261 [K]

Phyllanthus reticulatus Poir. de Wit 128 [WAG] Bakhuizen v. d. Brink 3540 [U] Phyllanthus discoideus MUELL. Arg. Leeuwenberg 2789 [U]

Phyllanthus capillaris Schum. et Thonn. De Wit 229 [WAG] Pl. III, 1

Phyllanthus acuminatissima C. B. Robb. Elmer 15662 [U]

Richeriella gracilis (MERR.) PAX et K. HOFFMANN Hyting 65438 [K] Securinega subtype.  $P=23,5~\mu~E=20~\mu~P:E=1,17.$  Securinega subtype.

P:E = 1,18 Lumina < 1  $\mu$ . Securinega subtype.

Securinega subtype.  $P=22,5~\mu~E=19,5~\mu~P:E=1,15.$  m:e>0,5~P.A.I.=ca.~0,25.Lumina ca.  $1~\mu$ . Securinega subtype.

Securinega subtype.  $P = 29 \mu$  E = 25,5  $\mu$  P:E = 1,14. m:e ca. 1 P.A.I. = 0,3. Lumina 1-2  $\mu$ .

Securinega subtype. P = 18  $\mu$  E = 16,5  $\mu$  P:E = 1,09. P.A.I. = 0,25. Lumina ca. 1  $\mu$ .

Securinega subtype.  $P = 22.5 \mu E = 18 \mu P:E = 1,25.$ Costae colpi. P.A.I. = 0,3. Lumina 1-2  $\mu$ .

Securinega subtype. Stephanocolporate (4) or tricolporate.  $P=21.5~\mu~E=20.5~\mu~P:E=1.05.$ P.A.I. = 0.5.

Securinega subtype.  $P = 22,5 \mu$  E = 18  $\mu$  P:E = 1,25. P.A.I. = 0,25. Lumina ca. 1  $\mu$ . Securinega subtype?

Too young for reliable measurements, but colpus transversalis distinctly circular.

Phyllanthus acidus subtype

Pollen grains tricolporate; sphaeroidal – oblate sphaeroidal. Colpus transversalis circular.

Unlike the Securinega subtype the pollen grains of the Phyllanthus acidus subtype are spheroidal or sometimes slightly oblate spheroidal. The colpus transversalis is always circular. The exine structure is in all respects the same as in the Securinega subtype.

Phyllanthus acidus (LINN.) Skeels Versteeg 566 [U]

Phyllanthus muellerianus (O. KUNTZE) EXELL Zenker 253 [U] Phyllanthus elsiae URBAN

Krug and Urban 4789 [L] Pl. III, 3

Phyllanthis acidus subtype.  $P = E = 17.5 \mu$  P:E = ca. 1. P.A.I. = 0.2 - 0.3.Phyllanthus acidus subtype. Colpi very short. Too young.

Phyllanthus acidus subtype.  $P = E = 20 \mu$ . P.A.I. > 0.5.

# Richeria type

Tricolporate; subprolate. Colpus transversalis narrow; costae. Colpi narrow. Tectate (?); intra-reticulate. Lumina ca. 1 μ.

The Richeria type is an intermediate type between the Antidesma type and the Securinega type. The narrow elongated colpus transversalis

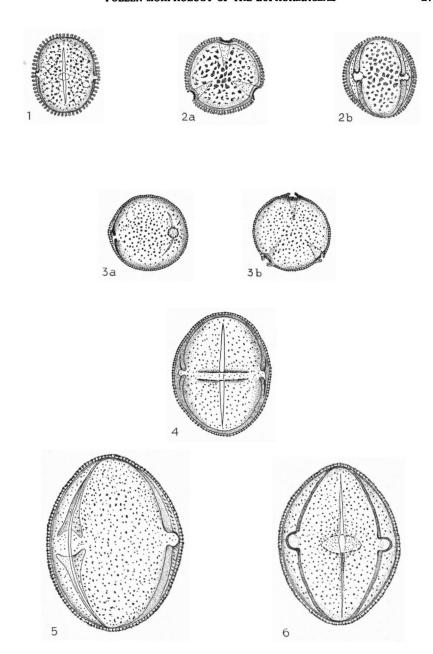


PLATE III. 1. Phyllanthus capillaris; 2. Securinega virosa; 3. Phyllanthus elsiae; 4. Drypetes glauca; 5. Actephila excelsa; 6. Andrachne phyllanthoides.

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is typical of the Antidesma type, but the distinct, rather coarse reticulum is characteristic of the Securinega type.

#### Taxonomic discussion

PAX and K. HOFFMANN (1931) as well as Bentham (1880) placed this genus in the Antidesminae. It occurs in the New World.

Richeria laurifolia BAILL. ex MUELL. ARG. Richeria type; prolate. Krukoff 8776 [U]  $P = 29 \mu \quad E = 24 \mu \quad P:E = 1,21.$ Lumina ca. 1  $\mu$ .

## Dicoelia type

Tricolporate; prolate – subprolate. Colpus transversalis broad and elongated to broadly elliptic; costae. Colpi narrow; costae colpi. Tectate, psilate; sometimes intra-reticulate. Pollen grains of medium size (ca. 25  $\mu$  – 50  $\mu$ ).

The Dicoelia type possesses elements of the Antidesma type as well as of the Securinega type. The pollen grains have always a Polar axis of at least 25  $\mu$ ; usually between 30  $\mu$  and 55  $\mu$ . The colpus transversalis is large; broad and elongated or broadly elliptic, but never circular.

## Taxonomic discussion

PAX and K. Hoffmann (1922, 1931) included the genus Dicoelia in the Antidesminae for its elongated inflorescences. However, the loose, often androgynous clusters along the rachis of the racemes is different from any inflorescence in the Phyllanthoideae (Bentham 1880).

# Dicoelia subtype

Colpus transversalis broad and elongated; m:e < 0,4. Usually not intra-reticulate.

Dicoelia affinis J. J. SMITH Hallier 1255 [U]

Drypetes glauca VAHL Sintenis 5111 [U] Pl. III, 4

Drypetes macrophylla (BLUME) PAX et K. Hoffmann Bogor VIII F. 48 [U]

Drypetes laterifolia (SWARTZ) KRUG et Ūrban Fuertes 824 [U]
Drypetes variabilis UITT.

B.W. 6481 [U]

Dicoelia subtype.  $P = 44 \mu E = 32 \mu P:E = 1,37.$  m:e < 0,4 P.A.I. = 0,3.

Indistinct reticulate. Dicoelia subtype

P = 34  $\mu$  E = 27  $\mu$  P:E = 1,26. m:e < 0,25 P.A.I. = 0,25.

Dicoelia subtype.

 $P = 45 \mu E = 40 \mu P:E = 1,16.$ m:e < 0,4 P.A.I. = 0,35.

Intra-reticulate. Dicoelia subtype.

Dicoelia subtype.

# Andrachne subtype

Colpus transversalis broadly elliptic; m:e = ca. 0.5 or larger. Usually intra-reticulate.

Andrachne phyllanthoides (NUTT.) MUELL. Arg.

Bush 14843 [U] Pl. III, 6

Andrachne telephioides LINN. Dalmatië 1937, 50 [U] Andrachne colchica FISCH. et MEYER Cult. Hort. Canton (Baarn) 4884 [U] Andrachne aspera SPRENG.

Pappi 4892 [U]

Actephila excelsa (DALZ.) MUELL. ARG. Hort. Bog. VII B. 271 [L] Pl. III, 5

Poranthera corymbosa Brong. Constable 30096 [U]

Poranthera microphylla BRONG. Hastings [U]

Andrachne subtype.  $P=45~\mu~E=35,5~\mu~P:E=1,27.$  m:e=0,5~P.A.I.=0,2. Indistinct costae colpi.

Andrachne subtype. Intra-reticulate. Andrachne subtype.

Andrachne subtype.

 $\mu$  E = 31  $\mu$  P:E = 1,40. m:e > 0,5. P.A.I. = 0,3. Longitudinal striation.

Andrachne subtype.

 $P = 48 \mu E = 37 \mu P:E = 1,36.$  P.A.I. = 0,3. Costae colpi.

Andrachne subtype.

P = 34  $\mu$  E = 29,5  $\mu$  P:E = 1,15. m:e ca. 0,5 P.A.I. = 0,25. Tectum perforatum; lumina of the

intra-reticulum 2-3  $\mu$ .

Andrachne subtype.

 $P = 24 \mu \quad E = 19.5 \mu \quad P:E = 1.23.$ Tectate; intra-reticulate; lumina ca. 1 μ.

## Zimmermannia type

Tricolporate; prolate spheroidal. Colpus transversalis small; costae. Colpi narrow. Tectate; verrucate.

The Zimmermannia type is the only type in the Phyllanthoideae with verrucae. Except for this remarkable character the pollen grains belong without doubt in the Antidesma configuration.

Zimmermannia capillepes PAX Fl. Usambarica 5862 [BM] Pl. IV, 1 Zimmermannia type.  $P = 58 \mu$   $E = 51 \mu$  P:E = 1,12. m:e > 0,5 P.A.I. = 0,25.

# Leptonema type

Tricolporate; subprolate.

Colpus transversalis broad and large; costae narrow.

Colpi narrow; costae colpi absent.

Tectate; psilate. The columellae are so small, that they are hardly visible.

The Leptonema type has some relation with the Dicoelia type (p. 28). The type misses, however, the costae colpi. It differs from the Antidesma type (p. 21) by the broad and elongated colpus transversalis.

#### Taxonomic discussion

Pax and K. Hoffmann (1922, 1931) placed in the subtribe Glochidiinae Leptonema together with Glochidion and Breynia. It seems better to keep Leptonema in the vicinity of Phyllanthus and its allied genera, as stated by Bentham (1880).

Leptonema venosa (Poir.) A. DE JUSS. D'Alleizette 6405 Mad. Nov. 1905 [L] Pl. IV, 2

Leptonema type.  $P = 27 \mu E = 22,5 \mu P:E = 1,21.$ m:e < 0,5 P.A.I. = 0,5.

# Pleiostemon type

Tricolporate; prolate spheroidal. Colpus transversalis; costae; m:e larger 1; m = 6-8  $\mu$ . Colpi narrow; costae colpi. P.A.I. small. Pollen grains sometimes syncolpate. Tectate; psilate.

The *Pleiostemon* type belongs undoubtedly to the *Antidesma* configuration. The colpus transversalis however differs from all other types by its meridional elongation. The m:e is larger than 1. Moreover, the colpi are very long. Sometimes the pollen grains are even syncolpate (colpi anastomosing at the poles).

#### Taxonomic discussion

The genera belonging to this type are without doubt related to *Phyllanthus* and its allied genera. Mueller (1866) reduced the genus *Pleiostemon* to a section of *Phyllanthus* and Bentham (1880) referred it to *Securinega*. In the system of Pax and K. Hoffmann (1931) *Pleiostemon* is maintained as a genus close to *Phyllanthus*. On the other hand, *Lingelsheimia* is found in the subtribe Drypetinae in their system. Corresponding characters are:

- 1. In the male flowers an extrastaminal disc present and an ovarium rudiment wanting.
- 2. In the female flowers an annular disc and bifid styles present.
- 3. Calyx is 6-partite (in *Pleiostemon* the male flower is sometimes 4-5-partite. Petals are absent.
- 4. Plants monoecious and only found in Africa and Madagascar (Dangyodrypetes).

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Pleiostemon verrucosus SONDER
Cooper 312 [BM] Pl. IV, 3
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Lingelsheimia frutescens PAX Lebrun 3855 [K] Pl. IV, 4

Dangyodrypetes ambigua Léandri Serv. Eaux et Forests 7675 [P]

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Pleiostemon type. P = 20 \mu E = 19.5 \mu P:E = 1.03 m:e > 1 P.A.I. < 0.2.
Pleiostemon type. P = 22.5 \mu E = 21 \mu P:E = 1.07.
P.A.I. = 0.1.
Pleiostemon type. P = 24 \mu E = 22.5 \mu P:E = 1.07.
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P.A.I. almost syncolpate-syncolpate.

# Heywoodia type

Tricolporate; prolate spheroidal. Colpus transversalis broad; costae absent; m:e ca 1. Colpi narrow; costae colpi absent. Tectate; psilate. Columellae distinct.

As the Polar axis is larger than the Equatorial axis the type is placed in the *Antidesma* configuration. There is, however, a possibility that *Heywoodia* is wrongly placed in this configuration because of the absence of costae.

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Heywoodia lucens SIM
Bally 5133 [K] Pl. IV, 5

Heywoodia type.

P = 27.5 \mu E = 25.5 \mu P:E = 1.08.

m:e ca. 1 P.A.I. = 0.3.
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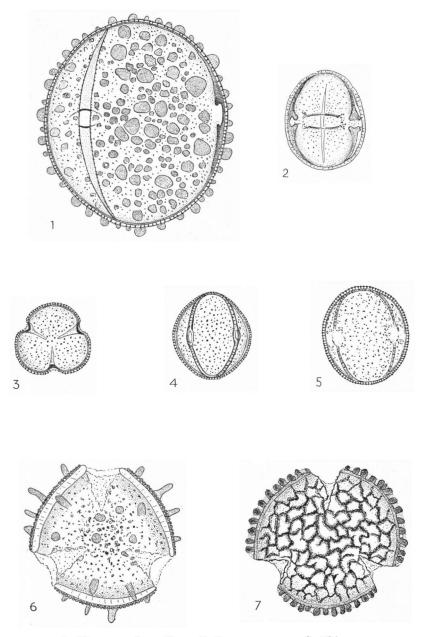


PLATE IV. 1. Zimmermannia capillepes; 2. Leptonema venosa; 3. Pleiostemon verrucosus; 4. Lingelsheimia fruticosa; 5. Heywoodia lucens; 6. Amanoa guianensis; 7. Pseudolachnostylis glauca.

#### AMANOA CONFIGURATION

Tricolporate-stephanocolporate (4-5). Pollen grains usually with a P. shorter than the E.

Colpus transversalis usually with costae.

Pollen grains reticulate. Rarely not reticulate (some species of Amanoa and the Discocarpus type).

The Amanoa configuration includes pollen types with a Polar axis shorter than the Equatorial axis and lesser than six (usually 3-4) colpi. The pollen grains are reticulate except in a few species. These latter species, however, have so many characters in common with related reticulate species, that there is no doubt that they belong to the same type.

As in the Antidesma configuration the types are difficult to separate.

Especially the Amanoa type and Savia type are closely related.

## Taxonomic discussion

The genera belonging to this configuration are all closely related. The division of the pollen grains into four types is not found again in the systems of Mueller, Bentham and Pax and K. Hoffmann.

MUELLER (1866) removed Briedelia and Cleistanthus to a distinct subtribe on account of their valvate calyx (nearly all the other genera in Phyllanthoideae have an imbricate calyx). For this reason PAX and K. HOFFMANN (1931) also placed these genera in a special tribe. Bentham (1880) took less account of the imbricate calyx and found, in the remaining characters, much affinity with Amanoa. So he placed Briedelia and Cleistanthus in close proximity of Amanoa. The pollen grains of Cleistanthus cannot be separated from those of Savia. Except for the longitudinal striation the pollen grains of the Briedelia type are also very close related to those of the Savia type. From these results it is clear that pollen morphology supports the statement of Bentham.

BENTHAM (1880) discussed *Uapaca* in the group of the *Antidesminae*. Pax and K. Hoffmann (1931) established a special subtribe for this genus. Although the pollen grains belong to the *Savia* type it seems better to keep this genus out of the subtribes with a *Savia* type. The involucrum and the absence of a disc in the male flower are characters too divergent for placing *Uapaca* in the vicinity of any other genus with a *Savia* type.

In the system of PAX and K. HOFFMANN (1931) the remaining genera of the Amanoa configuration are divided into four subtribes. 1. Wielandinae. 2. Amanoinae. 3. Discocarpinae. 4. Pseudolachnostylidinae.

In these subtribes the following genera do not have the typical characters of the *Amanoa* configuration.

1. Astrocasia. According to Webster (1956) the genus is related to Securinega. The pollen grains undoubtedly belong to the Securinega type.

2. Actephila. The genus is classified together with Amanoa by all authors. Actephila excelsa, however, possesses pollen grains of the Dicoelia type. Examination of the pollen grains of other species is necessary.

- 3. Chonocentrum. The pollen grains of Chonocentrum belongs without doubt to the Antidesma type (p. 21).
- 4. Savia erytroxyloides. Pollen grains of this Savia species belong to the Phyllanthus pentaphyllus subtype.

The remaining genera have the following characters in common:

1. The calyx is 5-partite.

2. Frequently petals are present.

3. Male as well as female flowers have a disc.

- 4. In the male flowers an ovarium rudiment is present.
- 5. Stamens are often connated into a column.
- 6. Seeds never have a caruncula.

## Amanoa type

Tricolporate-stephanocolporate (4-5); suboblate-oblate spheroidal. Colpus transversalis broad; ora large; costae usually present.

Colpi wide; no costae colpi.

Tectate or intectate; reticulate or not reticulate.

If the pollen grains are reticulate the lumina are at least 3  $\mu$  in diameter. When the pollen grains are not reticulate the structure elements are tall and at least 4  $\mu$ .

Exine very thick, at least 4  $\mu$ .

The Amanoa type is characterised by a coarse reticulum and a thick exine.

The reticulum of the Amanoa type had to be considered as an intrareticulum since the muri are connected by a thin membrane, which covers the lumina.

Some species of Amanoa lack the coarse reticulum. Amanoa guianensis has a tectum provided with long baculae or echinae. The columellae of Amanoa grandiflora are, in optical section, gemma shaped and inordinate arranged. In both Amanoa species the exine is, however, more than  $4 \mu$  thick.

Amanoa oblongifolia Muell. Arg. Krukoff 7015 [U] Pl. V, 1

Amanoa grandiflora Muell. Arg. Murca Pires 835 [U]

Amanoa bracteosa Planch.
D'Alleizette 6392 [L]
Amanoa boiviniana BAILL.
D'Alleizette Mad. Nov. 1906 [L]

Amanoa guianensis AUBL. Bafog S.F. 1192 [U] Wood Herb. 28 [U] Pl. IV, 6 Amanoa type. P = 33,5  $\mu$ . E = 43  $\mu$  P:E = 0,78. Colpi wide. P.A.I. = 0,25. Lumina up to 11,5  $\mu$ .

Amanoa type.  $P = 39 \mu$  E = 45  $\mu$  P:E = 0,87. P.A.I. = 0,5. Not reticulate. Columellae gemma shaped, inordinate arranged.

Amanoa type.  $E = 80 \mu$ . Lumina up to  $10 \mu$ . P.A.I. = 0,4.

Amanoa type.  $P = 37.5 \mu$  E = 45  $\mu$  P:E = 0.83. P.A.I. = 0.4 — 0.5. Lumina up to 4  $\mu$ .

Amanoa type.  $P = 46 \mu$  E = 52  $\mu$  P:E = 0,88. P.A.I. = 0,3. Costae transversalis. Columellae very short. Baculate-echinate; baculae up to 7  $\mu$ . Endexine thick. 34 W. PUNT

Pentabrachion reticulatum (MUELL. ARG.) Jantzon Zenker 2982 [L]

Pseudolachnostylis glauca (HIERN) HUTCH. Lanjouw 1181 [U] Pl. IV, 7

Amanoa type; 3-4 colporate.  $P = 45 \mu$  E = 48  $\mu$  P:E = 0,94. Lumina up to 5  $\mu$ . P.A.I. = 0,4-0,5. Costae transversales. Amanoa type; 2-3 colporate.  $P=31~\mu~E=34~\mu~P:E=0,91.$ Lumina up to 4,5  $\mu$ .

# Savia type

Tricolporate-stephanocolporate (4); subolate-prolate spheroidal.

Colpus transversalis usually narrow; ora small. Rarely colpus transversalis broad and ora large (Cleistanthus ferrugineus).

Colpi narrow; sometimes costae colpi present. Tectate or intectate; reticulate. Lumina of the reticulum 3  $\mu$  or smaller. Exine is medium thick, lesser then 4  $\mu$  but still distinct.

In many respects the Savia type resembles the Amanoa type. The lumina of the reticulum are, however, distinctly smaller. Since the pollen grains are also relatively smaller this difference is perhaps not of fundamental importance, but, besides the smaller lumina, the colpi are narrow and the ora small, so it seems better to separate the pollen grains of the Savia type from those of the Amanoa type.

Some species with a Savia type (Cluytiandra madagascariensis, Lachnostylis hirta) have a Polar axis larger than the Equatorial axis. By this character the pollen grains should be placed in the Antidesma configuration. The exine structure, however, is quite different from the Antidesma configuration and is, on the other hand, in full agreement with the Savia type.

Savia andringitrana LÉANDRI Decary 1926 [L]

Sa ia sessiliflora (Sw.) WILLD. Eggers 1905 [P]

Blotia oblongifolia (BAILL.) LÉANDRI D'Alleizette [L] Mad. 4 Mai. 1918

Wielandia elegans BAILL. D'Alleizette Seych. [L]

Lachnostylis hirta (LINN. F.) MUELL. ARG. [= Discocarpus hirtus (LINN. F.) PAX et K. HOFFMANN] Bolus 2396 [K] Cluytiandra madagascariensis LÉANDRI D'Alleizette Mad. 6402 [L]

Cleistanthus winkleri JABL.
Bogor IX C. 165 A. [U]
Cleistanthus dichotomus J. J. SMITH Gjellerup 142 [U]

Savia type.  $P = 26 \mu$   $E = 29 \mu$  P:E = 0.88. m:e < 0.5. P.A.I. = 0.4. Colpi narrow and short. Lumina 3  $\mu$ . Costae transversales. Savia type. P = 24,5  $\mu$  E = 29  $\mu$  P:E = 0,85. P.A.I. = 0,3.

Savia type.  $P = 25.5 \mu E = 28.5 \mu P:E = 0.89.$  P.A.I. = 0.45.Colpi short and narrow. Lumina 1-2  $\mu$ .

Savia type. 3-4 colporate. P = 37  $\mu$  E = 41,5  $\mu$  P:E = 0,88. P.A.I. = 0,65. Lumina = 1-2  $\mu$ . Colpi narrow and short.

Savia type; prolate spheroidal.  $P = 37 \mu$ .  $E = 35 \mu$  P:E = 1,05. P.A.I. = 0,3. Colpus transversalis with costae. Lumina 1-2 μ.

Savia type; prolate spheroidal.  $P = 32 \mu E = 31 \mu P:E = 1,04.$  P.A.I. = 0,3. Costae transversales. Re-

ticulum indistinct. Lumina 1–2  $\mu$ . Savia type.  $E = 32 \mu$ . Costae colpi.

Savia type.

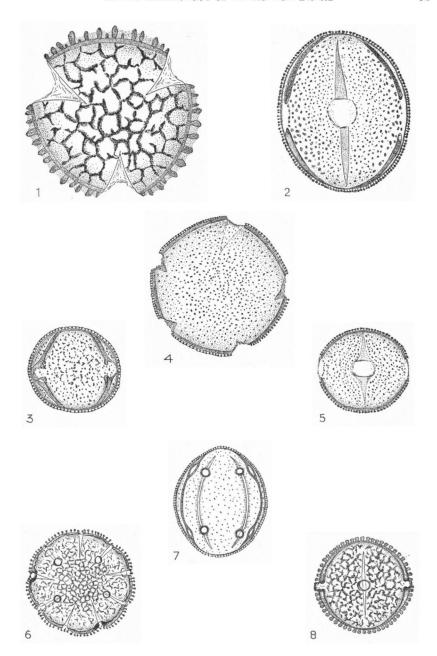


PLATE V. 1. Amanoa oblongifolia; 2. Cleistanthus ferrugineus; 3. Uapaca heudelotii; 4. Discocarpus essequeboensis; 5. Briedelia monoica; 6. Breynia fruticosa; 7. Breyniopsis pierrei; 8. Glochidion sericeum.

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Cleistanthus ferrugineus MUELL. ARG. Hort. Bogor. IX C. 43 A. [U] Pl. V, 2 P = 44 \ \mu \quad E = 33 \ \mu \quad P: E = 1,34. P.A.I. = 0,4 — 0,5. Colpus transversalis isodiametric; no costae; ora large. Lumina 1–2 \mu. Savia type; oblate spheroidal. P = 22 \ \mu \quad E = 25 \ \mu \quad P: E = 0,88. Muell. Arg. Stolz 622 [U] P = 27 \ \mu \quad E = 29,5 \ \mu \quad P: E = 0,93. Tectate.
```

# Briedelia type

Tricolporate; oblate spheroidal. Colpus transversalis broad; costae.

Colpi narrow at the ends, broadened in the equatorial part. Sometimes costae colpi present.

Intectate; reticulate. Lumina small and in chains along the colpi (pseudo-striate). Muri distinct but not tall.

This type is closely related to the Savia type. The greatest difference is found in the longitudinal arrangement of the lumina of the reticulum. The chains of lumina give the pollen grains a striate appearance, which is, however, only a "pseudo-striation": a striation not caused by individual columellae but by the lumina of the reticulum which are, of course, composed of several columellae.

```
Briedelia monoica (Lour.) Merr. Endert E 1066 [U] Pl. V, 5 P = 24 \ \mu \quad E = 25,5 \ \mu \quad P:E = 0,94. m:e > 0,5. \quad P.A.I. = 0,35. \quad Lumina < 1 \ \mu. Briedelia glauca Blume Winckel 865 B [U] Briedelia assamica J. D. Hook. Herb. East Ind. Comp. 4890 [U] Briedelia stipularis (Linn.) Blume v. Leeuwen-Reynvaan [U] Briedelia type. E = 31 \mu. Lumina 1–2 \mu. Briedelia type. E = 42 \mu. m:e < 0,5. \quad P.A.I. = 0,25. Briedelia type. E = 42 \mu. m:e < 0,5. \quad P.A.I. = 0,25. Briedelia type. P = 32 \ \mu \quad E = 33,5 \ \mu \quad P:E = 0,96. P.A.I. = 0,25. Costae colpi.
```

# Discocarpus type

Stephanocolporate (4-5); oblate spheroidal. Colpus transversalis small; no costae present. Colpi narrow. No costae colpi present. Tectate, psilate. Not reticulate.

The Discocarpus type is an intermediate type between the types of the Amanoa configuration and the types of the Antidesma configuration. The small columellae and the absence of a reticulum are characters in favor of placing the type in the Antidesma configuration. On the other hand the oblate shape, the absence of costae and the 4-5 colpi are more in agreement with the Amanoa configuration. For these latter reasons it seems better to discuss the type in the Amanoa configuration.

Discocarpus essequeboensis Klotzsch Schomburgk 659 [U] Pl. V, 4 Discocarpus type. 
$$P = 34.5 \mu$$
 E = 37  $\mu$  P:E = 0.94. Ora small; m:e > 0.5. P.A.I. = 0.6.

#### PHYLLANTHUS NUTANS CONFIGURATION

Pollen grains with an areolate structure.

#### Taxonomic discussion

After a proposal of IVERSEN, ERDTMAN recommended the term areolatus for the structure described under *Phyllanthus nutans* type (1947). The definition given by him is: "Pollen grains with small areas separated by small grooves forming a negative reticulum". Webster (1956) states that the areolate pollen grains are mostly found in the New World. Some species of *Phyllanthus* in the Old World, however, have areolate pollen grains that are superficially very similar to those of the New World.

## Phyllanthus nutans type

Periporate; the surface of the pollen grains is divided into pentagons or hexagons. The pori are situated in the angular points. Pori small.

Intectate. From each aperture two rows of pila lead to the other apertures. In the centre of the penta- and hexagons the pila form an irregular reticulum.

Phyllantus nutans Sw. Hooker, Jamaica [U] Pl. VI, 2

Phyllanthus adianthoides Klotzsch Hulk 21 [U] Phyllanthus speciosus JACQ.

ryllanthus speciosus JACQ. Fresh material, Greenhouse Utrecht Phyllanthus nutans type.

Longer axis 27,5  $\mu$ . Shorter axis 25  $\mu$ .

ca. 25 pori.
Phyllanthus nutans type.
Diam. = 17  $\mu$ .
Phyllanthus nutans type.
Diam. = 20  $\mu$ . Pori ca. 15.

## Dendrophyllanthus type

Tricolporate (?); oblate. Colpus transversalis circular; costae. Colpi very narrow; operculum consisting of one row of pila. Intectate; not reticulate.

Without doubt the pollen grains are related to the *Phyllanthus nutans* type in their exine structure. The surface of the pollen grains is divided into three regions, separated by strings of pila, the "colpi". Each "colpus" consists of three rows of pila. If the two outer rows represent the borders of the colpus, the inner row must be taken as an operculum. However, the space between the rows is so small, that it can hardly be regarded as a weak performed part in the ektexine (see definition aperture: Glossary). For this reason it is possible to defend the opinion, that the pollen grains of Dendrophyllanthus are triporate. In that case the rows of pila would only be an ornamental peculiarity. Nevertheless, the rows look so much like real colpi, that in the author's opinion it is better to maintain the pollen grains in the group of the tricolporate.

Dendrophyllanthus ripicolus Guill. Mackee 3629 [L] Pl. VI, 3 Dendrophyllanthus type.  $P = 19.5 \mu E = 27 \mu P:E = 0.72.$  m:e = ca. 1.

#### Breynia configuration

Stephanocolporate (colpi sometimes with two ora). Colpus transversalis circular rarely somewhat elliptical; costae. Colpi narrow and long. Tectate or intectate.

By its exine structure the *Breynia* configuration is related to the *Antidesma* configuration. Most pollen grains, however, have a Polar axis shorter than the Equatorial axis as in the *Amanoa* configuration. The typical character of the number of colpi with often two circular colpi transversalis per colpus seem to justify this configuration.

## Breynia type

Stephanocolporate (colpi diorate); oblate – subprolate.
Usually two circular colpi transversales per colpus, some colpi have only one colpus transversalis; costae.
Colpi narrow; sometimes costae colpi.
Intectate or tectate; reticulate or not reticulate; lumina small.

The Breynia type is easily recognised by the large number of colpi (six or more) and the double circular colpi transversales. In the species of Breynia one of the two colpi transversales is often reduced. The remaining endexine aperture is never situated in the equatorial

plane but towards the ends of the colpus.

### Taxonomic discussion

Hitherto the genera belonging to this type have never been placed in one group by any author. Mueller (1866) recognised the relationship of Agyneia with Sauropus, which opinion was shared by Bentham (1880) and Pax and K. Hoffmann (1922, 1931). Breynia and Glochidion, on the other hand, are always distinguished as genera closely related to Phyllanthus. Glochidion is even often reduced to a section of Phyllanthus. Pax and K. Hoffmann keep Glochidion, Breynia and Breyniopsis in the subtribe Glochidiinae. In the author's opinion the above genera should all be placed in one group (either subtribe or other taxon). They have the following corresponding characters:

- 1. The number of sepals is usually six (Glochidion sometimes fewer).
- 2. In the male flowers generally three connated stamens form a column. Connectives are frequently appendiculated. Anthers extrors. There is no ovarium rudiment nor a disc. As Bentham states, the scale-like thickenings at the base of the sepals of Agyneia and Sauropus are not to be regarded as disc glands.
- 3. In the female flowers disc glands are wanting.
- 4. The plants are generally monoecious (Some Glochidion species and Arachnodes, however, dioecious) and only occur in the Old World.

# Breynia subtype

Pollen grains oblate spheroidal – oblate. Intectate; reticulate; lumina 1–2  $\mu$ .

Breynia fruticosa (LINN.) BENTH. J. and M. S. Clemens 3182 [U] Beekler 2 [U] Pl. V, 6

Breynia racemosa (Blume) Muell. Arg. Bakhuizen v. d. Brink [U] Breynia nivosa (W. G. SMITH) SMALL Collector unknown, Agyneia bacciformes (LINN.) A. DE JUSSIEU v. Leeuwen-Reynvaan 1978 [U]

Sauropus androgynis (LINN.) MERRILL Docters v. Leeuwen, Depok 7-5-'11 Breynia subtype; oblate.  $P = 21 \mu E = 27 \mu$ P:E = 0.76. Colpi transversales situated almost at the end of the colpi. Colpi not always provided with two colpi transversales. Colpi 8-10. Lumina 1-2  $\mu$ . Breynia subtype; oblate. 5-6 colpi. Breynia subtype. 7-8 colpi.

Breynia subtype; oblate.  $P = 27.5 \mu E = 32.5 \mu P:E = 0.85$ . Always two colpi transversales present. Colpi 5-6. Costae colpi. Breynia subtype; oblate spheroidal.

 $P = 25.5 \mu E = 28 \mu P:E = 0.90.$ 8-9 colpi.

## Breyniopsis subtype

Pollen grains subprolate. Tectate; not reticulate.

The pollen grains of Breyniopsis are related to the pollen grains of Breynia by the number of colpi and the double circular colpi transversales. The shape and structure are, however, quite different.

Breyniopsis pierrei Beille Poilane 19777 [P] Pl. V, 7 Breyniopsis subtype; subprolate.  $P = 32.5 \mu E 28.5 \mu P:E = 1.14$ . Always two colpi transversales per colpus. 6-7 colpi. Costae colpi.

### Glochidion type

Stephanocolporate (4); spheroidal - prolate spheroidal. Colpus transversalis circular or slightly elliptical; costae. Intectate or tectate; reticulate; lumina 1-2  $\mu$ .

In many morphological respects the Glochidion type is closely related to the Breynia type, as Erdtman (1952) and Webster (1956) have stated. The pollen grains lack, however, the typical double colpus transversalis. It is also possible to place the type in the Antidesma configuration. Its single circular, sometimes broad elliptic endexine aperture and the distinct reticulum are characters in favour of a place in the Securinega type (p. 25).

Bakhuizen v. d. Brink 1886 [U] Pl. V, 8 Glochidion concolor MUELL. ARG. Yuncker 15648 [U] Glochidion littorale Blume Bakhuizen v. d. Brink 3182 [U] Glochidion obscurum (WILLD.) BLUME Buysman 295 [U] Glochidion ramiflorum J. R. et G. FORSTER Yuncker 15717 [U] Glochidion superbum BAILLON Dumas 1507 [U]

Glochidion sericeum (Blume) J. D. Hooker Glochidion type; spheroidal. P = E = 24  $\mu$ . Colpi narrow. P.A.I. = 0,3-0,4. Lumina 1-2  $\mu$ . Intectate.

Glochidion type.

Glochidion type.

Glochidion type.

Glochidion type.

Glochidion type.

Glochidion spec.
Teysman 4411 [U]
Arachnodes chevalieri GAGNEP.
Fleury 31755 [P] Pl. VI, 1

Glochidion type.

Glochidion type; prolate spheroidal.  $P=25,5~\mu~E=24~\mu~P:E=1,06.$  P.A.I. > 0,5. Tectate; intra-reticuculate. Colpus transversalis slightly elliptical.

### Aristogeitonia configuration

Stephanocolpate, stephanocolporate, stephanoporate, periporate or inaperturate. The pollen grains are spheroidal or have a Polar axis shorter than the Equatorial axis.

Tectate; usually echinate, sometimes psilate.

The types of the Aristogeitonia configuration form a closely related group which is sharply differentiated from the other configurations in the Phyllanthoideae. They are easily recognised by the number of apertures and structure of the exine. Most types have echinate pollen grains and ca. six small apertures. The shape is spheroidal or oblate spheroidal to oblate.

Taxonomic discussion

The Aristogeitonia configuration includes a typical group of genera which are, in several respects, different from other Phyllanthoideae. Mueller (1866) was acquainted with too few genera of this group to recognise it. Bentham (1880), however, enumerated, in the fourth group of his Phyllantheae, all the genera of the Aristogeitonia configuration then known. Only the genus Bischofia of Bentham's group does not belong to the configuration.

All the genera belonging to the configuration are found in one of the three groups into which Pax divides the Phyllanthoideae (Pax 1924). This group includes his subtribes Drypetinae, Petalostigmatinae, "Toxico-dendrinae" (= Hyaenanchinae), Dissiliarinae, Paivaeusinae and the tribus Caletieae of the Stenolobeae. Only Drypetes, Lingelsheimia and Heywoodia of the Drypetinae differ pollen-morphologically. The genera are small. The largest genus Longetia does not come up to more than 10 species. The male flower lacks an extra staminal disc. The flowers always are apetal, while the number of stamens is seldom constant; there are mostly more than six. In the female flower the styles are undivided, at best emarginate, but never incised. The leaves of this group are often opposite or in whorls. Some genera have, moreover, composite leaves.

### Aristogeitonia type

Stephanocolpate, stephanocolporate or stephanoporate; oblate spherioidal – suboblate.

Colpi, if present, small and very short; costae colpi.

Pori, if present, with costae.

Colpus transversalis, if present, with costae.

Tectate; echinate or psilate.

In the Aristogeitonia type the apertures are always situated in the

equatorial plane. The apertures can have the shape of pori, colpi or composite apertures, but they are always small.

Aristogeitonia subtype

Pollen grains echinate.

Aristogeitonia limoniifolia PRAIN Herb. Kew. Pl. VI, 5

Oldfieldia africana Benth. Šmall 621 [K] Paivaeusa dactylophylla WELW. et OLIVER Shabana 9 [K] Piranhea trifoliata BAILL. Glaziou 14238 [K] Pl. VI, 4

Tetracoccus ilicifolius COVILLE et GILMAY Gilmay Mai. 2A. 1939 [L]

Hyaenanche globosa (GAERTN.) LAMB. et  $\mathbf{V}_{\mathbf{AHL}}$ Splitgerber [L] Pl. VI, 6 Mischodon zeylanicus THWAIT. Hort Bogor. IX A. 125, 80883 [U]

Paragelonium perrieri LÉANDRI Perrier de la Bathie 1178 [P]

Petalostigma quadriloculare F. v. MUELLER Queensland [U]

Longetia malayana (Benth.) Pax et K. Hoffmann D'Alleizette, Penang 6421 [L]

Longetia carunculata (BAILL.) PAX et K. Hoffmann Mackee 4230 [L]

Longetia buxoides subtype

Pollen grains psilate.

Longetia buxoides BAILLON D'Alleizette, Nw. Caled. 6419 [L] Pl. VI, 7

Dissiliaria tricornis BENTH. Brass 5755 [L] Pl. VI, 9 Aristogeitonia subtype; stephanocolpate.  $P=30,5~\mu~E=35~\mu~P:E=0.86$ . Colpi ca. 4,5  $\mu$  long. Echinae ca. 2,5  $\mu$ .

Aristogeitonia subtype; stephanocolpate.  $P = 35,5 \mu$   $E = 43 \mu$  P:E = 0.83.

Aristogeitonia subtype; stephanocolpate.  $P = 43 \mu E = 46,5 \mu P:E = 0,92.$ 

Aristogeitonia subtype; stephanoporate.  $P=27~\mu~E=32~\mu~P:E=0,85.$ Diam. pori 3  $\mu$ .

Aristogeitonia subtype; stephanocolporate.  $P=35,5~\mu~E=40~\mu~P:E=0,91.$  Colpi 6-7. Echinae 3,5  $\mu$ .

Aristogeitonia subtype; stephanocolporate.  $P = 39 \mu E = 46 \mu P:E = 0.85$ . Colpi 6-7. Echinae  $2 \mu$ .

Aristogeitonia subtype; stephanocolpate.  $P = 32 \mu E = 36 \mu P:E = 0.89$ Colpi 5-6. Echinae 3,5  $\mu$ . P:E = 0.89.

Aristogeitonia subtype; stephanoporate.  $P = 32 \mu E = 35 \mu P:E = 0.92.$ Diam. pori 3  $\mu$ . Echinae 2,5-3,5  $\mu$ .

Aristogeitonia subtype; stephanoporate.  $P=28,5~\mu~E=32~\mu~P:E=0,89.$ Pori 4-6; not always exactly in the equatorial plane; diam. 3 μ. Echinae ca. lμ.

Aristogeitonia subtype; stephanoporate.  $P=47~\mu~E=52~\mu~P:E=0.90.$ Pori slightly ellpitical diam. ca. 5  $\mu$ . Echinae up to  $6.5 \mu$ . Aristogeitonia subtype; stephanoporate.

Pori 5(-6).

Longetia buxoides subtype; stephanocolpate.  $P=22~\mu~E=27~\mu~P:E=0.81.$  Colpi (5-) 6 (-7); small and short with thick costae.

Longetia buxoides subtype; stephanoporate.  $P = 19 \mu E = 22.5 \mu P:E = 0.85.$ Pori 6; diam. 2  $\mu$ .

## Stachystemon type

Periporate or inaperturate; spheroidal or ellipsoid (with a longer and a shorter

Pori small; costae pori.

Tectate; echinate.

In the Stachystemon type the apertures are never situated in the equatorial plane, but scattered over the surface.

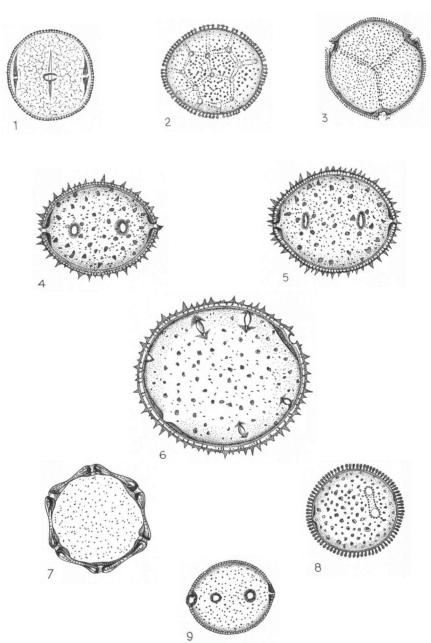


PLATE VI. 1. Arachnodes chevalieri; 2. Phyllanthus nutans; 3. Dendrophyllanthus ripicolus; 4. Piranhea trifoliata; 5. Aristogeitonia limoniifolia; 6. Hyaenanche globosa; 7. Longetia buxoides; 8. Phyllanthus acuminatus; 9. Dissiliaria tricornis.

Stachystemon vermicularis Planch. Pritzel 270 [L] Pl. VII, 2

Micrantheum ericoides. DESF. Buysman 1873; 347 [U]

Micrantheum hexandrum J. D. Hook Tasmania [U] Pseudanthus pimeleoides SIEB. ex SPRENG. Buysman 1890 [U] Pl. VII, 7

Pseudanthus nematophorus F. v. MUELL. Murchinson [U] Neoroepera banksii BENTH. Banks and Solander 1770 [K]

Longetia gynotricha Guill.
Mackee 4191 [L]
Androstachys johnsonii Prain
Pole-Evans, 3-8-1917 [K] Pl. VII, 2

Stachystemon type. Diam. 36 μ. Pori ca. 12; diam. pori 2,5 μ. Echinae 1,5 μ long.

Stachystemon type. Longer axis 45  $\mu$ . Pori ca. 25; diam. 2  $\mu$ . Echinae usually in groups of three around the pori; 3  $\mu$  long.

Stachystemon type. Diam. 45 μ. Pori small; more than 40. Stachystemon type. Diam. 20 μ. Pori ca. 7; diam. 2 μ. Echinae 1 μ long. Stachystemon type. Diam. 20 μ.

Stachystemon type. Diam. 40  $\mu$ . Diam. pori 1,5  $\mu$ ; ca. 30 pori. Echinae 2  $\mu$  long.

Stachystemon type. Diam. 34  $\mu$ . Pori ca. 16. Echinae 2  $\mu$ . Stachystemon type.

Longer axis 46  $\mu$ . Shorter axis 40  $\mu$ . Echinae ca. 1  $\mu$ .

### POLLEN TYPES NOT PLACED IN ONE OF THE ABOVE CONFIGURATIONS

Some types cannot readily be placed in one of the discussed configurations. Since most types have only one representative they are discussed separately and not put in any configuration.

## Phyllanthus acuminatus type

Tricolporate (colpi diorate); spheroidal. Colpi transversales (two in each colpus) circular; costae. Colpi short. Intectate; pilate. Pila ordinate arranged but not in a reticulum.

The presence of two ora in the colpi is also an important character of the *Breynia* type (p. 38). The structure of the exine, however, is quite different. Its remaining characters cannot be compared with any other configuration.

Phyllanthus acuminatus VAHL Hostman 413 [U] Pl. VI, 8 Phyllanthus acuminatus type.  $P = E = 27 \mu$ . P.A.I. = 0.5.

# Bischofia type

Tricolporate; spheroidal, P:E is 1. Colpus transversalis small; costae. Edges of the costae transversales rounded at the ends. Colpi narrow and long; no costae colpi. Tectate; psilate. Columellae short.

This type should be best placed in the Antidesma configuration but differs from other types in this configuration by its shape (P:E=1) and the absence of costae colpi. The columellae are moreover very short.

#### Taxonomic discussion

BENTHAM (1880) discussed Bischofia together with Oldfieldia and Piranhea because of their digitately compound leaves. Pax and K. HOFFMANN (1931) thought it better to put the genus apart in a subtribe. As the pollen grains are certainly not similar to those of Oldfieldia and Piranhea and moreover difficult to place in any other configuration, it seems right to maintain the separate subtribe.

Bischofia javanica Blume
Hohenacker 1573 [U] Pl. VII, 5

Bischofia type.  $P = E = 25 \mu$ . P:E = ca. 1. m:e < 0.5. P.A.I. = 0.2.

## Putranjiva type

Tricolporate; spheroidal. P:E is 1. Colpus transversalis large; costae. Colpi narrow; costae colpi.

Tectate; psilate. Exine thick. Columellae short but endexine and tectum thick.

The *Putranjiva* type has some resemblances with the *Dicoelia* type (p. 28). There is a large colpus transversalis with costae, and also costae colpi. The type differs, however, by its shape (P: E = 1) and the remarkably thick endexine.

In a specimen of *Putranjiva* (Stocks, India), examined by ERDTMAN (1952), the pollen grains are subprolate, reticulate. The specimens Koorders 2156  $\beta$  and Broadway 9249 differ considerably from that description. The typical character of the thick exine is not mentioned by Erdtman and was probably not present in his pollen grains.

### Taxonomic discussion

In the system of Pax and K. Hoffmann (1931) Putranjiva is found in the subtribe Glochidiinae. The genus is put in relatiosnhip with Breynia and Glochidion. Bentham (1880) gives it a place next to Drypetes. It must be admitted, that the pollen grains of Putranjiva are far more similar to those of Drypetes than to the Breynia type.

Putranjiva roxburghii Wall.

Koorders 2156  $\beta$  [U]

Braodway 9249 [U] Pl. VII, 3

Putranjiva type.  $P = E = 40 \mu$ . P:E = ca. 1. m:e < 0.5. P.A.I. = 0.3. Exine thick (4  $\mu$ ).

## Hymenocardia type

Triporate (-diporate); oblate. Costae pori. Tectate; psilate. Columellae short.

The type differs completely from all the other pollen grains in the Phyllanthoideae.

#### Taxonomic discussion

On account of its catkin-like male inflorescenses Pax and K. Hoffmann (1931) placed Hymenocardia in the Antidesminae. The most striking character of the genus is the compressed two-cocced fruit with lateral wings. *Hymenocardia* differs from the other genera in the Antidesminae in some characters:

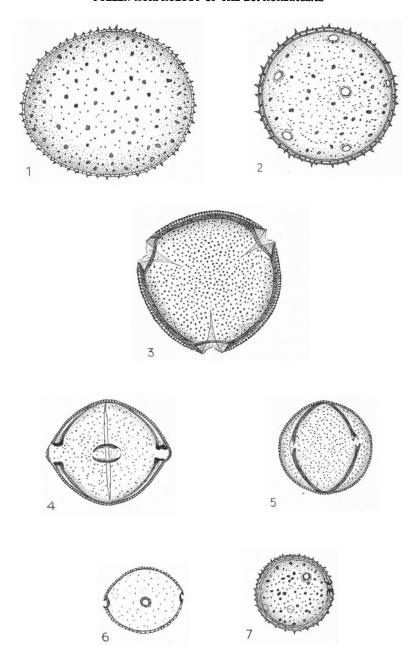


PLATE VII. 1. Androstachys johnsonii; 2. Stachystemon vermicularis; 3. Putranjiva roxburghii; 4. Martretia quadricornis; 5. Bischofia javanica; 6. Hymenocardia ulmoides; 7. Pseudanthus pimeleoides.

- 1. Disc wanting in male as well as in female flowers.
- 2. The styles are long and undivided.

```
Hymenocardia ulmoides OLIV. Koechlin 2371 [U] Pl. VII, 6
Hymenocardia acida Tul.. P = 19 \mu E = 22,5 \mu P:E = 0,85.
Hymenocardia acida type. P = 28,5 \mu E = 31 \mu P:E = 0,92.
Duvigneaud 2422 (BRLU) Diam. pori 3 \mu.
```

## Martretia type

Tricolporate; oblate spheroidal.
Colpus transversalis; costae. Edges of the costae transversales rounded at the ends. Atrium present.
Colpi narrow; no costae colpi present.
Tectate; psilate.

The pollen grains of the Martretia type are oblate spheroidal but do not have any other character in common with the Amanoa or Aristogeitonia configuration. The colpus transversalis has rounded ends. This character is present in some types of the Antidesma configuration. The oblate shape and absence of costae colpi keeps the Martretia type out of the Antidesma configuration. A typical character is the atrium, which is not found in any other type.

## Taxonomic discussion

Martretia has only one species. In many respects this species is related to the Antidesminae (e.g. in its inflorescense). There are, however, some differences.

- 1. No disc is present in the male flower nor in the female one.
- 2. The seeds are provided with a caruncula. No other genus in the Antidesminae has carunculate seeds.
- 3. The only genus in the whole family with a false partition in the fruits (PAX and K. HOFFMANN, 1922).

```
Martretia quadricornis Beille Martretia type. Adames 251 [K] Pl. VII, 4 P = 28.5 \mu E = 31 \mu P:E = 0.92, m:e = 0.3 - 0.4. P.A.I. = 0.25.
```

#### TAXONOMIC COMMENT ON THE PHYLLANTHOIDEAE

The Phyllanthoideae can be divided into three groups of pollen types. These groups, broadly outlined, agree with the three groups Pax (1924) could already distinguish on other grounds, though without nominating them.

The first group, to which, for instance, the subtribes Antidesmineae and Phyllanthinae belong, has pollen grains which mostly have a prolate shape and distinct costae transversales (Antidesma configuration, p. 20). The second group has pollen grains which are mostly reticulate and generally have an oblate shape (Amanoa configuration, p. 32). The subtribes Amanoinae, Sarropodinae, etc., belong to this group. The third group comprises those genera that are to be found in the Aristogeitonia configuration (p. 40).

For the *Phyllanthoideae* (Pax and K. Hoffmann, 1931) composed a highly differentiated system of many subtribes. This system, however, leaves much to be desired. Some subtribes (e.g. *Glochidiinae*) comprise genera which decidedly do not show any relationship with each other. On the contrary, other genera in different subtribes resemble each other so much, that it would seem desirable to unite them (*Wielandiinae* with *Amanoinae* and *Pseudolachnostylidinae*).

In the Phyllanthoideae pollen morphology is obviously exceedingly

useful to arrive at a natural classification.

#### b. Crotonoideae

#### CROTON CONFIGURATION

Croton-pattern present. Structure elements on the tectum are usually clavae but can also be echinae, baculae etc. Sometimes the structure elements are located on ridges (e.g. Croton matourensis, Manihot saxicola). Tectate.

The Croton configuration includes all pollen grains with a croton-pattern. In the Euphorbiaceae the croton-pattern is only found in the subfamily of the Crotonoideae. ERDTMAN (1952) states, that, outside the Euphorbiaceae, pollen grains with a croton-pattern have been found in the Buxaceae and Thymelaeaceae.

Most pollen grains with a croton-pattern belong to the *Croton* type. The *Manihot* type, *Klaineanthus* type and *Hevea* type occur less commonly.

#### Taxonomic discussion

In the subfamily of the Crotonoideae a great number of genera possess

pollen grains with a croton-pattern.

In Pax and K. Hoffmann's system (1931) these genera occur in the following tribes: Crotoneae, Chrozophoreae, Joannesieae, Cluytieae, Manihoteae, Celonieae and Ricinocarpeae. Further also *Neoboutonia* as the only genus out of the Acalypheae. It follows from this enumeration, that the genera possessing a croton-pattern are to be found in widely divergent groupings.

In classifying the Crotonoideae most authors considered the bursting open of the male calyx to be a factor of primary importance for arranging the genera. Thus Mueller (1866) and Pax and K.Hoffmann arrived at a classification, in which genera with an obvious relationship are kept widely apart, e.g. Joannesia and Micrandra (Baldwin et Schultes 1947; Schultes 1955). Bentham (1880) already stated that within the genus Croton the male calyx opens both valvate and imbricate, which is why, in his system, this character plays a less important part. In Bentham's system the genera with a croton-pattern are placed close together. They are to be found in his subtribes Jatrophineae, Eucrotoninae and Chrozophorinae.

In comparing all the taxa that possess a croton-pattern it is seen that several characters correlate with this pollen character. With hardly an exception the male and the female flower contain a disc, while an ovarium rudiment is absent. A less constant but striking

character is the presence of petals in many genera. Genera which form an exception to these characters are:

Sagotia. The disc is wanting in both sexes. The genus, however, is related to *Dodecastigma*, Sandwithia and Garcia (CROIZAT 1948), which, on the other hand, have a disc.

Bertya. This Australian genus has no disc in both sexes either. There is, however, a distinct relation to two other Australian genera, viz. Beyeria and Ricinocarpos, which certainly have a disc.

Tritaxis has no disc in the female flower.

Adenocline and Tetrorchidium have no disc in the male flower.

The genera Pantadenia and Oligoceras, described by Gagnepain, were said to possess an ovarium rudiment in their male flowers. Pantadenia does have a little conical projection in the centre of its receptaculum, but whether this projection represents an ovarium rudiment is not certain. Oligoceras possesses, in its male flower, a column which, according to a drawing by GAGNEPAIN in the Flore Géneral de l'Indochine (1926), was said to be an ovarium rudiment. In studying the type material in Paris, however, the author concluded, that the drawing gives an erroneous picture of the reality. The column is not bottle-shaped but cylindrical. The five stamens are not placed at the foot of the column, but are grown together with it at a little distance of the base. The projections which stand at the end of the column and which, in the drawing, look like stigmas, are three staminodies. These staminodies even contain rudimentary pollen sacs. Therefore there are two whorls of stamens, the topmost of which is sterile.

A more or less distinct ovarium rudiment is found in: Klaineanthus, Cladogelonium, Micrandra and Endospermum, which all belong to the Klaineanthus type.

Genera not possessing a corolla are: Neoboutonia, Benoistia, Elateriospermum, Baliospermum, Eremocarpus, Micrandra (Cunuria), Hevea, Bertya, Beyeria, Adenocline and Tetrorchidium. Suregada mostly has no corolla (Léonard 1958).

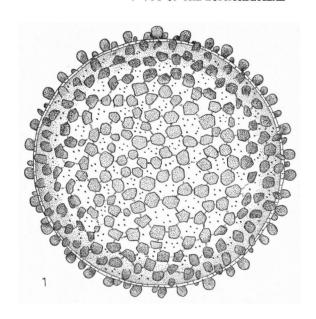
The type occurring most frequently is the Croton type. Less common are the Manihot type, the Klaineanthus type, Hevea type and Adenocline type.

# Croton type

Inaperturate; spheroidal or ellipsoid i.e. with a longer and a shorter axis. Tectate; endexine thin. Croton-pattern. Clavate or echinate.

The Croton type has no apertures and the shape is fairly uniform. There may be a longer and a shorter axis, but usually the pollen grains are spheroidal. The structure elements have more variations. The differences in these elements and the lumina enclosed by the elements, however, are slight and many transitions exist, so that it seems better to omit the formation of groups (subtypes).

The pollen grains of Sagotia racemosa show two shapes. The specimens Y. Mexia and Woob Herb. are echinate, while the specimen For. Dep.



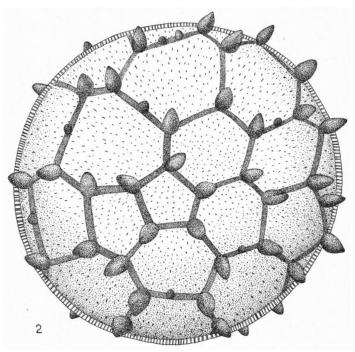


PLATE VIII. 1. Croton cuneatus; 2. Croton matourensis ( $\times$  800).

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Garcia nutans VAHL Lundell 12248 [U]

is clavate. Further examination will be necessary to ascertain whether these two shapes of Sagotia racemosa can be distinguished by other characters besides.

According to Schultes (1952) Cunuria cannot be distinguished from Micrandra. The pollen grains of the Cunuria specimen Ducke 1087 belong to the Croton type, and not to the Micrandra type. In another Cunuria specimen (Ducke 24874) ERDTMAN (1952) described the pollen grains as being 5-colporate. According to Schultes (1952) the two Ducke specimens both belong to Cunuria spruceana var. bracteosa. Micrandra siphonoides and Micrandra brownsbergensis stand pollenmorphologically between the two specimens of Cunuria spruceana var. bracteosa.

Croton cuneatus Klotzsch Croton type. J. et P. Florschütz 1113 [U] Pl. VIII, 1 Diam. longer axis 75  $\mu$ . Clavate; diam. clavae 3,5 μ. Clavae on ridges.

Croton type. Diam. 55 μ.

D. cl. 2,8 μ.

Croton type. Diam. 75 μ.

D. cl. 2,2 μ. Clavae distinctly on ridges. Croton bahamensis MILLSP. Brace 4064 [U] Croton longiradiatus LANJ. B.W. 6711 [U] Croton hirtus L'HERITIER Croton type. Versteeg 504 [U] Croton matourensis AUBL. Croton type. B.W. 5050 [U] Pl. VIII, 2 Clavae acuminate; distinctly on ridges. Diam. 115  $\mu$ . D. cl. 6,5  $\mu$ . Croton pullei LANJ. Croton type.
Diam. 115 μ. D. cl. 6,5 μ. Clavae Rombouts 654 [U] distinctly on ridges. Croton type. Diam. 120  $\mu$ . Julocroton triqueter BAILLON D. cl.  $3,2^{-}\mu$ . Hassler 6016 [U] D. cl. 3,2 μ.

Croton type. Diam. 120 μ.
D. cl. 3,2 μ.

Croton type. Diam. 60 μ.
D. cl. 1,5 μ.

Croton type. Diam. 60 μ. Julocroton argenteus (LINN.) DIDRICHS
C. L. Schulz 1225 [U] Crotonopsis elliptica WILLD. Delzie Demaree 7066 [U] Eremocarpus setigerus (Hook.) BENTH. D. cl. 2  $\mu$ . Croton type. Diam. 48  $\mu$ . L. S. Rose 41457 [U] Grossera major Pax Zenker 469 [U] D. cl. 1,8  $\mu$ . D. cl. 1,8  $\mu$ . Croton type. Diam. 45  $\mu$ . D. cl. 2  $\mu$ . Croton type. Diam. 60  $\mu$ . D. cl. 3,5  $\mu$ . Croton type. Diam. 60  $\mu$ . D. cl. 2  $\mu$ . Croton type. Diam. 50  $\mu$ . D. cl. 2 8  $\mu$ . Holstia tenuifolia PAX Drummond et Hemsley 3480 [K] Cyrtogonone argentea (PAX) PRAIN Zenker 561 [U] Crotonogyne preussii Pax Zenker 4666 [L] Crotonogyne parvifolia PRAIN D. cl. 2,8  $\mu$ . D'Alleizette 6845 [L] Croton type. Diam. 60  $\mu$ . D. cl. 3,5  $\mu$ . Croton type. Diam. 70  $\mu$ . D. cl. 2,0  $\mu$ . Croton type. Diam. 65  $\mu$ . Manniophyton fulvum MUELL. ARG. Zenker 1404 [L]

Aleurites moluccana (LINN.) WILLD.

Bunnesmeyer 8060 [U] Deutzianthus tonkinensis GAGNEP. Croton type. Diam. 03  $\mu$ . D. cl. 2,0  $\mu$ . Croton type. Diam. 50  $\mu$ . D. cl. 1,8  $\mu$ . Croton type. Diam. 40  $\mu$ . D. cl. 2,0  $\mu$ . Croton type. Diam. 60  $\mu$ . D. cl. 2,0  $\mu$ . Balansa 3149 [P] Oligoceras eberhardtii GAGNEP. Eberhardt [P] Alphandia furfuracea BAILLON Balansa 3439 [P]

Croton type. Diam. 70  $\mu$ .

D. cl. 3,0  $\mu$ . Clavae on ridges. Croton type. Diam. 40  $\mu$ .

Joannesia princeps VELLOSO T. W. Brown 35 [K] Neoboutonia macrocalyx PAX Stolz 2043 [U] Pantadenia adenanthera GAGNEP. British Museum Baloghia lucida Endlicher Beckler, Hastings River [U] Ostodes paniculata BLUME Griffith 4790 [U] Ostodes pendulus (Hassk.) Meeuse et Adelbert Sandakan For. Dep. A. 2964 [U] Codiaeum variegatum (LINN.) BLUME Fresh material from greenhouse at Utrecht. Pl. IX, 3 Codiaeum stellingianum WARB. Docters v. Leeuwen 1567 [U] Fontainea pancheri (BAILLON) HECKEL D'Alleizette 6474 [L] Dimorphocalyx murina ELMER Elmer 12773 [U] Benoistia perrieri LEANDRI D'Alleizette Madag. Dec. 1905 [L] Tritaxis gaudichaudi BAILL. Clemens 4351 [U]
Strophioblachia fimbricalyx BOERL.
Hort. Bog. VIII F. 43 [U]
Blachia umbellata (WILLD.) BAILL. D'Alleizette Austr. Nov. 1907 [L] Sagotia racemosa BAILLON For. Dep. 7443 [U] Mexia 6050 [U] Wood Herb. 311 [U] Jatropha multifida LINN. Groll-Meyer, St. Eustatius [U] Jatropha integerrima JACQ Docters v. Leeuwen 19-7-1950 [U] Mildbraedia paniculata PAX Leeuwenberg 3004 [U] Acidocroton adelioides GRISEB. Ekman 16896 [K] Pausandra densiflora LANJ. Krukoff 5464 [U] Pausandra morisiana (CASAR.) RADLK. Ducke et Kuhlmann 16594 [U] Givotia rottleriformis GRIFFITH Wight 2638 [L] Ricinodendron heudelotii (BAILL.) PIERRE ex Pax Zenker 487 [U] Elateriospermum tapos BLUME Hort. Bog. IX. A. 129 [U]
Baliospermum motanum (WILLD.) MUELL. Bengal, J. D. Hooker [U] Cavaçoa aurea (CAVACO) LÉONARD de Winter 2109b [K] Anomalocalyx uleanus (PAX et K. HOFFM.) DUCKE

Ducke 23518 [U]

D. cl. 1,5  $\mu$ . Croton type. Diam. 45  $\mu$ . D. cl. 1,0  $\mu$ . Croton type. Diam. 40 µ. D. cl. 1,0  $\mu$ . Croton type. Diam. 65 μ. D. cl. 3,5  $\mu$ . Croton type. Diam. 53  $\mu$ . D. cl. 2,8  $\mu$ . Croton type. Diam. 50  $\mu$ . D. cl. 2  $\mu$ . Croton type. Diam. 50  $\mu$ . D. cl. 2,0  $\mu$ . Croton type. Diam. 60 µ. D. cl. 2,8  $\mu$ . Croton type. Diam. 80 u. D. cl. 3,5  $\mu$ . Croton type. Diam. 35  $\mu$ . D. cl. 1,8  $\mu$ . Croton type. Diam. 45  $\mu$ . Echinate. Diam. ech. 1,8  $\mu$ . Croton type. Diam. 60  $\mu$ . D. cl. 1,8  $\mu$ . Croton type. Diam. 45  $\mu$ . D. cl. 1,8  $\mu$ . Croton type. Diam. 50  $\mu$ . D. cl. 2,0. Clavae sharp pointed. Echinate. Diam. 40  $\mu$ . D. ech. 1,8  $\mu$ .

Croton type. Diam. 95  $\mu$ . D. cl. 4,0  $\mu$ . Clavae on ridges. Croton type. Idem. Jatropha multifida

Croton type. Diam. 50  $\mu$ . D. cl. 2,5  $\mu$ . Croton type. Diam. 45  $\mu$ . D. cl. 1,5  $\mu$ . Croton type. Diam. 50  $\mu$ . D. cl. 2,0  $\mu$ . Croton type. Diam. 75  $\mu$ . D. cl. 3,5  $\mu$ . Croton type. Diam. 48  $\mu$ . D. cl. 2,8 μ. Croton type. Diam. 50 u. D. cl. 2,5  $\mu$ .

Croton type. Diam. 70  $\mu$ . D. cl. 3,2  $\mu$ . Clavae on ridges. Croton type. Diam. 43  $\mu$ . D. cl. 1,5  $\mu$ .

Croton type. Diam. 75  $\mu$ . D. cl. 2,0  $\mu$ . Croton type. Diam. 55  $\mu$ . D. cl. 3,2  $\mu$ .

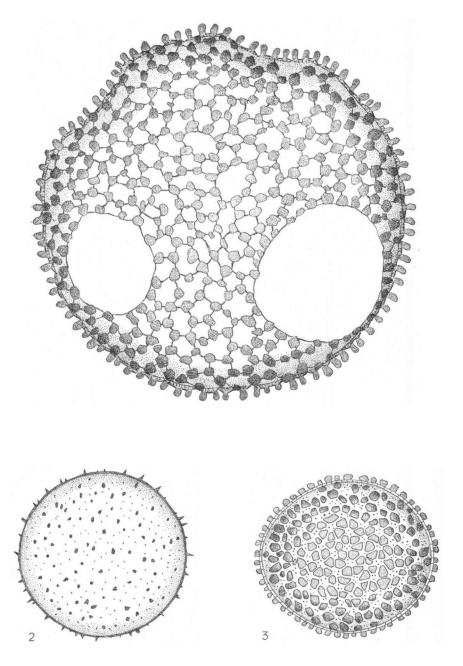


PLATE IX. 1. Manihot saxicola ( $\times$  600); 2. Tannodia cordifolia; 3. Codiaeum variegatum.

Sandwithia guyanensis LANJ.
For. Dep. 2731 [U]
Dodecastigma amazonicum Ducke
Ducke 23543 [U]
Cunuria spruceana Balllon var. bracteosa
(Ducke) R. E. Schultes
Ducke 1087 [K]
Vaupesia cataractarum R. E. Schultes
Schultes 14006 [K]
Ricinocarpos pinifolius Desf.
Austr. felix [U]
Beyeria leschenaultii (Decand.) Balll.
Austr. felix [U]

Bertya gummifera Planch.
Johnson et Constable 16024 [U]

Tannodia cordifolia BAILLON
D'Alleizette, Comoren 6484 [L]
Pl. IX, 2

D'Alleizette 6479 Dec. 1905 [L] Pl. X, 1

Domohinea perrieri LÉANDRI

Croton type. Diam. 55  $\mu$ . D. cl. 3,5  $\mu$ . Croton type. Diam. 80  $\mu$ . D. cl. 4,5  $\mu$ . Croton type. Diam. 50  $\mu$ . D. cl. 1,8  $\mu$ .

Croton type. Diam. 70 μ.
D. cl. 3,0 μ.
Croton type. Diam. 60 μ.
D. cl. 1,8 μ.
Croton type. Diam. 43 μ.
D. cl. 1,5 u.
Clavae sharp pointed.
Croton type. Diam. 55 μ.
Croton-pattern indistinct.
Croton type. Diam. 50 μ.

Croton-pattern on ridges. Echinate; diam. ech. 1,5 μ. Croton type. Diam. 35 μ. Echinate; diam. ech. 1-1,5 μ.

## Manihot type

Periporate; spheroidal. Pori large, borders indistinct. Tectate. Croton-pattern.

The type is closely related to the *Croton* type. The pollen grains, however, have large apertures with indistinct rims. The number of pori varies in the different species.

#### Taxonomic discussion

According to Croizat (1943b) Cnidiscolus does not have a calyx but a corolla. This is also the case with Manihot. In a very definite sense Cnidiscolus connects Jatropha, sensu strictu, with Manihot. Both the sorts of Cnidiscolus examined had pollen grains of the Manihot type, sothat at least in this respect they are closer related to Manihot than to Jatropha.

The pollen-morphological similarity between Manihot and Suregada is probably accidental and has to be considered a parallel development.

Manihot saxicola LANJ. Proefst. Buitenz. 922 Boom 8 [U] Pl. IX, 1

Manihot esculenta CRANTZ
Rombouts 757 [U]
B.W. 2780 [U]
Cnidiscolus urens (LINN.) ARTHUR
Boldingh 4829 [U]
Cnidiscolus stimulosus (MICHAUX) GRAY
Killip 31636 [U]
Suregada glomerulata (BLUME) BAILL.
Griffith 4772 [U]
Suregada subglomerulata (ELMER) CROIZAT
Elmer 12967 [U]

Manihot type.
Diam. 160 μ. Croton-pattern on a distinct reticulum.
Pori ca. 25. Diam. clavae 6,8 μ.
Manihot type.
Idem Manihot saxicola.
Pori ca. 25.
Manihot type. Diam. 75 μ.
D. cl. 2,5 μ. Pori 3-4.
Manihot type. Pori ca. 6.

Manihot type. Diam. 40 μ.
D. cl. 2,5 μ. Pori 3-5.
Manihot type. Diam. 45 μ.
Pori 5-6.

## Klaineanthus type

Tricolpate; oblate spheroidal to oblate. Colpi wide or narrow; no operculum present. Tectate; croton-pattern.

The Klaineanthus type, too, is closely related to the Croton type and the Manihot type. Pollen grains of this type have colpi without an operculum. The colpus membrane is frequently ruptured.

Klaineanthus subtype

Boarders of colpi distinct. Clavate.

The rims of the colpi of this subtype are distinct. The colpus membrane is frequently totally ruptured. In *Cladogelonium* the colpus membrane is not ruptured but granulate. Sometimes a rudiment of the colpus membrane remains at the outer ends of the colpi.

Klaineanthus gaboniae Pierre ex Prain Zenker 583 [U] Pl. X, 3

Cladogelonium madagascariense LÉANDRI Perrier de la Bathie 9696 [P] Pl. X, 4

Endospermum moluccanum (Thysm. et BINNEND.) BECCARI Kornasse 462 [U]

Glycydendron amazonicum Ducke Ducke 17108 [U]

Micrandra brownsbergensis Lanj. v. Emden 9 [U] Micrandra siphonioides BENTH. Lopez 9625 [U] Pl. X, 5 Klaineanthus subtype; suboblate.  $P = 29,5 \mu E = 37 \mu P:E = 0,80.$ Colpi wide. P.A.I. = 0,4. Diam. clavae 1.8  $\mu$ .

Diam. clavae 1,8  $\mu$ . Klaineanthus subtype; spheroidal.  $P = 30 \mu E = 30,5 \mu P:E = 0,98$ . Colpus membrane granulate. P.A.I. = 0,4.

Klaineanthus subtype; suboblate.  $P=33.5~\mu~E=44.5~\mu~P:E=0.80.$  Colpus membrane absent. P.A.I. = 0.4. Diam. clavae 1.8  $\mu$ .

Klaineanthus subtype; oblate.  $P = 29 \mu E = 41,5 \mu P:E = 0,69.$ Colpus membrane absent. P.A.I. = 0,35. Diam. clavae 1,5  $\mu$ .

Klaineanthus subtype.
Colpus membrane absent.

Klaineanthus subtype.  $P = 41,5 \mu E = 46 \mu P:E = 0,92.$ P.A.I. = 0,3. Croton-pattern indistinct. Clavae candelabrum-shaped, in-ordinate arranged.

Adenocline subtype

Borders of the colpi not sharply cut but irrigularly ruptured. Baculate. Baculae small and crowded,

The baculae on the tectum are small and crowded. With low magnification it is not directly obvious that the structure elements form a croton-pattern and not a reticulum.

### Taxonomic discussion

The genus Adenocline was placed in the vicinity of Seidelia and Leidesia in the Acalypheae by Bentham (1880). Pax and K. Hoffmann (1931) could not find the same relation and put the genus in a separate subtribe in the tribe Gelonieae. This tribe also comprises the genus Klaineanthus. The two genera are, however, quite different in other respects.

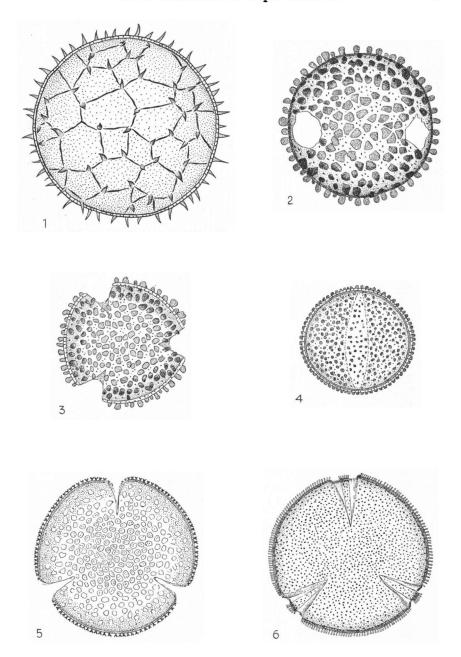


PLATE X. 1. Domohinea perrieri; 2. Suregada subglomerulata; 3. Klaineanthus gaboniae; 4. Cladogelonium madagascariense; 5. Micrandra siphonioides; 6. Hevea brasiliensis.

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Adenocline acuta (THUNB.) BAILL. [=A. mercurialis Turcz.] Lanjouw 613 [U] Pl. XI, 2

Adenocline subtype; oblate spheroidal.  $P = 36 \mu E = 37.5 \mu P:E = 0.96$ . P.A.I. = 0.4.

### Hevea type

Tricolpate; oblate spheroidal.

Colpi wide; operculum present. Tectate; baculate. The baculae are so small and crowded, that the structure is hardly recognised as a croton-pattern.

As in the Adenocline subtype the baculae of the Hevea type are small in diameter and crowded. With low magnification the structure elements seems to form a reticulum, but with high magnification the arrangement of the structure elements is clearly a croton-pattern.

The operculum distinguishes this type from the Klaineanthus type.

```
Hevea type; oblate spheroidal. P = 33.5 \mu E = 35 \mu P:E = 0.96. P.A.I. = 0.3.
Hevea brasiliensis (WILLD. ex A. JUSSIEU)
   Muell. Arg.
   Schultes 8050 [U]
B.W. 752 [U] Pl. X, 6
Hevea guianensis AUBLET
B.W. 41 [U]
                                                      Hevea type. P=35~\mu~E=38~\mu~P:E=0.95.
   Krukoff 5800 [U]
```

## Tetrorchidium type

Tricolporate; oblate spheroidal to prolate spheroidal. Colpus transversalis exceptionally broad and elongated. No costae present. Colpi narrow; rims irregular. Intectate; pilate. Pila coarse.

The Tetrorchidium type is tricolporate and differs by this character from the tricolpate Klaineanthus type. The extremely broad colpus transversalis is elongated to a colpus transversalis equatorialis. The tectum of this type is indistinct and the croton-pattern is not clear in all the pollen grains. In Tetrorchidium didymostemon the structure elements on the tectum are certainly not arranged in a croton-pattern. Tetrorchidium rubrivenium shows the croton-pattern more readily.

Although the most important characters of the Croton configuration are not always present, the structure of the pollen grains and the shape of the colpi are very much like those in the Klaineanthus type. The author therefore prefers to keep the Tetrorchidium type in the vicinity of the Klaineanthus type.

### Taxonomic discussion

PAX and K. HOFFMANN (1931) referred Tetrorchidium into the subtribe Tetrorchidiinae. This subtribe was placed in the tribe Gelonieae, to which also Klaineanthus belongs. Although in habit and character Adenocline and Tetrorchidium differ widely, the flowers have some characters in common:

- In the male flowers disc and ovarium rudiment are absent.
- In the female flowers the disc consists of three free glands.

Tetrorchidium rubrivenium POEPP. Harris and Britton 10746 [K]

Tetrorchidium didymostemon (BAILLON) PAX et K. Hoffmann Zenker 4590 [L] Pl. XI, 3

Tetrorchidium type; oblate spheroidal.  $P=36~\mu$   $E=40~\mu$  P:E=0,90. P.A.I. = 0,35. Bacculate-verrucate. Tetrorchidium type; prolate spheroidal.  $P = 28 \mu$   $E = 27 \mu$  P:E = 1,04. P.A.I. = 0.35.

#### CNESMOSA CONFIGURATION

Inaperturate; no croton-pattern present. Tectate or intectate.

Like the Croton type the Cnesmosa configuration is inaperturate, but a croton-pattern is always absent.

### Cnesmosa type

Inaperturate. Pollen grains ellipsoid, i.e. with a longer and a shorter axis. Tectate; psilate. No reticulum.

The pollen grains of this type are closely related to the Tragia fallax type and the Platygyne type. In the Cnesmosa type the pollen grains are tectate.

#### Taxonomic discussion

The genera belonging to this type are closely related and not easy to separate. Croizat (1941a) discussed the relationship between some genera of the *Plukenetiinae* and came to the following conclusions:

- Cenesmon Gagnepain cannot be separated from Cnesmosa Blume.
- Megistostigma malaccense, reduced to Sphaerostylis by PAX and K. Hoffmann, is the taxonomic type of the genus Megistostigma, which is congeneric with Clavistylus J. J. SMITH.
- Tragiella Pax and K. Hoffmann should be treated as a synonym of Sphaerostylis, of which S. tulasneana is the taxonomic type (p. 63).

The pollen-morphological results are in full agreement with the statements of Croizat.

Cnesmosa javanica Blume Elmer, Borneo 20663 [U] Java 1855 [L] Cnesmosa type; ellipsoid. Longer axis 55  $\mu$ .

Megistostigma malaccense J. D. Hook. Sinclair 40242 [U] Pl. XI, 4

Megistostigma peltatus (J. J. SMITH) CROIZAT Cnesmosa type; ellipsoid.
v. Steenis 1235 [L] Pl. XI, 6 Longer axis 56 μ; shorter axis 53 μ.
Megistostigma malaccense J. D. Hook. Cnesmosa type; ellipsoid.

Acidoton urens SWARTZ Jamaica, Hooker [U] Longer axis 42  $\mu$ ; shorter axis 37  $\mu$ . Columellae in groups.

Cnesmosa type; ellipsoid. Longer axis 37,5  $\mu$ ; shorter axis 35,5  $\mu$ .

# Tragia fallax type

Pollen grains inaperturate; spheroidal or ellipsoid. Intectate, pilate. Pila not in a reticulum. Endexine very thin.

This type differs from the *Cnesmosa* type by its intectate structure. The pila are not arranged in a reticulum.

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#### Taxonomic discussion

The examined species belong to the section Bia of Tragia. Klotzsch and Baillon (1858) separated this section from Tragia. Pax and K. Hoffmann (1919a) also stated: "Die männliche Blüte weckt den Eindruck, als ob es um eine eigene Gattung handelt". If all the species of the section Bia have the inaperturate pollen grains, it is perhaps better to treat this section as a distinct genus, although a great affinity with Tragia cannot be denied.

Tragia fallax Muell. Arg. Klug 4207 [U] Pl. XI, 5 Tragia sellowiana (Klotzsch) Muell. Arg. Smith 3558 [U] Tragia fallax type. Diam. 50  $\mu$ . Pila 1,5  $\mu$ . Tragia fallax type.

Diam. 55  $\mu$ .

## Platygyne type

Inaperturate. Pollen grains ellipsoid. Tectate; intra-reticulate. Endexine with circular thickenings.

The *Platygyne* type is distinguished from the *Cnesmosa* type by its distinct intra-reticulum. Small circular thickenings occur on the endexine. These peculiar "costae" are distributed irregularly over the endexine and certainly not accompagnied by thinnings of the ektexine.

Platygyne hexandra (JACQ.) MUELL. ARG. Rutten-Pekelharing 346 [U] Pl. XI, 1 Platygyne type; ellipsoid. Longer axis 40  $\mu$ . Shorter axis 36  $\mu$ . Lumina 2-3  $\mu$ .

## Trigonostemon verrucosus type

Inaperturate; spheroidal or ellipsoid. Tectate. Gemmate.

Trigonostemon verrucosus J. J. SMITH
Cult. Hort. Bog. VIII. E-16 [U]
Pl. XII, 3
Trigonostemon fungii MERRILL
Linsley Grishitt 1089 [BM]

Trigonostemon verrucosus type; spheroidal. Longer axis 60 μ. Diam. gemmae 2 μ. Gemmae not in a croton-pattern. Trigonostemon verrucosus type. Diam. 60 μ. Diam. gemmae 1,5-2 μ.

# Trigonostemon redioides type

Inaperturate; spheroidal to elliptical. Intectate; reticulate.

Both *Trigonostemon* types are inaperturate without a croton-pattern. They belong for these reasons to the *Cnesmosa* configuration.

The Trigonostemon vertucosa type with the gemmate structure elements on its tectum is perhaps related to the Croton type (p. 48). The gemmae are, however, certainly not arranged in a croton-pattern. Except for the shape and lack of apertures the Trigonostemon redioides type does not have any character in common with the Croton configuration.

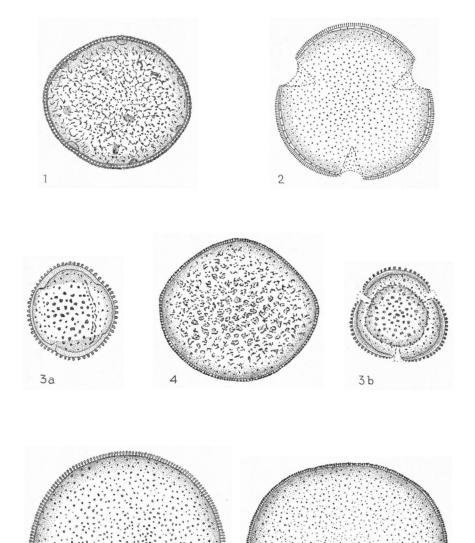


PLATE XI. 1. Platygyne hexandra; 2. Adenocline mercurialis; 3. Tetrorchidium didymostemon; 4. Megistostigma malaccense; 5. Tragia fallax; 6. Megistostigma peltatus.

5

#### Taxonomic discussion

Although the pollen grains of the *Trigonostemon* type are not provided with a croton-pattern, the genus *Trigonostemon* has great affinity with genera in the *Croton* configuration. Corresponding characters are:

- 1. In the male as well in the female flowers a disc is found and an ovarium rudiment is absent.
- 2. Sepals are present.

Actephilopsis malayana RIDL. Ridley 2300 [K] Trigonostemon redioides type. Longer axis 34 μ. Shorter axis 30 μ. Lumina 1-2 μ. Trigonostemon redioides type.
Diam. 55 μ. Lumina 3-5 μ.
Trigonostemon redioides type.
Longer axis 38 μ; snorter axis 35 μ.
Lumina ca. 2 μ.
Trigonostemon redioides type.
Longer axis 45 μ. Lumina small, ca. 1 μ.

### Dysopsis configuration

Inaperturate. Pollen grains quasi tricolpate, distinctly three-lobed. Tectate; no croton-pattern present.

## Dysopsis type

Inaperturate; quasi tricolpate; oblate spheroidal. Tectate; psilate. Columellae fairly coarse.

The most striking character is found in the three-lobed shape of the pollen grains. The declination between the lobes is distinct, but there is not the slightest indication of any part of the exine being weakened. According to the definition (see glossary) the pollen grains are inaperturate.

#### Taxonomic discussion

MUELLER (1866) as well as Pax and K. Hoffmann (1914, 1931) kept Dysopsis in close relationship with Leidesia and Seidelia. These latter African genera are, however, geographically far removed from the Andine genus Dysopsis. The three genera belong undoubtedly to the Acalypheae. The pollen grains of Dysopsis have little affinity with those of Leidesia and Seidelia. Perhaps it should be better to separate Dysopsis in a different subtribe.

Dysopsis glechomoides (RICH.) MUELL. ARG. Dysopsis type. Lechler 956 [K] Pl. XII, 6  $P = 21 \mu$  E = 22,5  $\mu$  P:E = 0,93.

#### PLUKENETIA CONFIGURATION

Tricolpate or triporate. Shape of the pollen grains oblate spheroidal to oblate (rarely prolate spheroidal).

Apertures broad; costae absent.

Colpus or porus membrane ruptured.

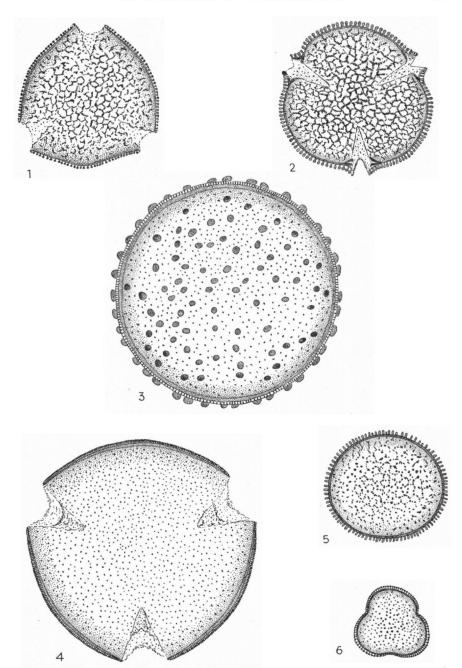


PLATE XII. 1. Plukenetia verrucosa; 2. Fragariopsis scandens; 3. Trigonostemon verrucosus; 4. Plukenetia volubilis; 5. Trigonostemon redioides; 6. Dysopsis glechomoides.

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The types of the *Plukenetia* configuration have simple apertures. In contrast with the *Chiropetalum* configuration the apertures are broad and have a ruptured colpus membrane. In some species a small piece of this membrane persists at the outer ends of the colpi.

## Plukenetia type

Tricolpate; oblate spheroidal to oblate (rarely spheroidal or prolate spheroidal). Colpi broad; colpus membrane ruptured. At the outer ends of the apertures a small piece of the colpus membrane (i.e. endexine) persists.

The ruptured part is sometimes bordered by a margo. Tectate or intectate. Reticulate or not reticulate.

Pollen grains belonging to this type can have very different exine structures. They may be intectate, tectate or tectum perforate, while the columellae are inordinate arranged or form a reticulum. The various subtypes are based on the structure patterns of the exine.

### Taxonomic discussion

From the genus *Plukenetia* new genera have been separated by several authors. Pax and K. Hoffmann (1919a) maintain this division of *Plukenetia*. Bentham (1880), however, has pointed out that *Fragariopsis* and *Romanoa* differ only slightly from the typical *Plukenetia* species. Other authors, too, see little difference. Croizat (1941a): "No characters are available to separate *Pterococcus* and *Tetracarpidium* from *Plukenetia*". Macbride (1951): "Apodandra and Fragariopsis could readily be included in *Plukenetia*". Certainly the pollen grains of the above genera are not easily distinguished from those of *Plukenetia*.

In South-America three tree-shaped, non-climbing genera are to be found. Sandwith (1950) says of them: "Angostyles, Astrococcus and Haematostemon are three closely allied genera of the Plukenetiinae". The pollen is indeed related. They all belong to the Plukenetia volubilis subtype and possess a tectum perforatum. Astrococcus and Haematostemon have moreover a margo.

On account of the imbricate calyx Bentham and Pax and K. Hoffmann placed Omphalea outside the Plukenetiinae. Croizat (1942a, 1948), however, gives as his opinion that the genus shows strong affinity to the Plukenetiinae. The pollen of Omphalea also appears to resemble the Plukenetia volubilis subtype very closely.

## Plukenetia volubilis subtype

Tectate; psilate. Sometimes the tectum is a tectum perforatum.

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Plukenetia volubilis LINN.
Baker 153 [U] Pl. XII, 4

Plukenetia abutifolia (Ducke) Pax et K. Hoffm.
Ducke 20619 [U]
Eleutherostigma lehmannianum Pax et K. Hoffm.
Lehmann 5158 [K] Pl. XIII, 1

Plukenetia volubilis subtype.
P = 30 \mu E = 50 \mu P:E = 0,64.
P.A.I. = 0,35.

Plukenetia volubilis subtype.
P = 50 \mu E = 62 \mu P:E = 0,80.
Tectum perforatum.

Plukenetia volubilis subtype.
P = 37 \mu E = 47,5 \mu P:E = 0,79.
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Tetracarpidium conophorum (MUELL. ARG.) Plukenetia volubilis subtype.  $P = 30.5 \mu E = 44 \mu P:E = 0.69$ . P.A.I. = 0.15 — 0.2. Margo. Pax et K. Hoffm. = Plukenetia conophorum Muell. Arg.] Mann 1739 [U] Versuchsanst. 9 [U] Pterococcus corniculatus (E. SMITH) PAX et Plukenetia volubilis subtype. K. Hoffm.  $P = 32 \mu E = 44 \mu P:E = 0.66$ . [= Plukenetia corniculatus E. SMITH] Koorders 41720 B [U] P.A.I. = 0,2.Plukenetia volubilis subtype.  $P = 43 \mu E = 49,5 \mu P:E = 0,87.$  P.A.I. = 0,2. Margo. Anabaenella tamnoides (Juss.) Pax et K. Hoffm. nom. illeg. [= Romanoa] TREVISAN [= Plukenetia tamnoides (Juss.) Muell. Ärg.] Lutz 1086 [U] Angostyles longifolia BENTH. Plukenetia volubilis subtype.  $P = 45 \mu E = 60 \mu P:E = 0.75$ . P.A.I. = 0.45. Tectum perforatum. Ducke 23528 [U] Plukenetia volubilis subtype. Astrococcus cornutus BENTH.  $P = 36.5 \mu E = 50 \mu P:E = 0.73.$ P.A.I. = 0,4. Margo; tectum per-Spruce 2050 [K] Pl. XIII, 3 foratum. Plukenetia volubilis subtype. Haematostemon guianensis SANDWITH  $P = 29 \mu E = 40 \mu P:E = 0.74.$ P.A.I. = 0.25. Margo; tectum per-Fanshawe 6016 [U] foratum indistinct. Omphalea diandra Linn. Froes (Krukoff) 1893 [U] Pl. XIII, 2 Plukenetia volubilis subtype.  $P = 21.5 \mu E = 30.5 \mu P:E = 0.59.$  P.A.I. = 0.3.Lanjouw and Lindeman 3164 [U] Plukenetia volubilis subtype; spheroidal. Tragia capensis Thunb.  $P = E = 17 \ \mu$ . Hooker Herb. [K] Plukenetia verrucosa subtype Intectate (?); reticulate. Plukenetia verrucosa subtype; suboblate.  $P=28~\mu~E=35~\mu~P:E=0,80.$  P.A.I. = 0,3. Lumina 2-3  $\mu$ . Plukenetia verrucosa Smith B.W. 993 [U] Pl. XII, 1 Apodandra buchtienii (PAX) PAX Plukenetia verrucosa subtype. = Plukenetia buchtienii PAX] Krukoff 10753 [U] Fragariopsis scandens St.-HILL. Plukenetia verrucosa subtype.  $P = 33.5 \ \mu \ E = 41.5 \ \mu \ P:E = 0.81.$ D'Alleizette 6594 [L] Pl. XII, 2 P.A.I. = 0,45.Plukanetia verrucosa subtype.  $P = 29 \mu E = 32 \mu P:E = 0.90.$ Reticulum consisting of separate pila, lumina 1-2  $\mu$ . P.A.I. = 0.6. Tragia stolziana PAX et K. HOFFM. Stolz 1775 [U] Pl. XIII, 4 Plukenetia verrucosa subtype. Sphaerostylis natalensis (SOND.) CROIZAT  $P = 38.5 \mu$   $E = 43 \mu$  P:E = 0.89. P.A.I. = 0.6. Lumina 1-2  $\mu$ . v. Someren 2409 [K] P.A.1. = 0.0. Example Plukenetia verrucosa subtype.  $E = 44 \mu$  P: E = 0.84. Sphaerostylis tulasneana BAILL. D'Alleizette, Nossibé. Nov. 1906 [L] P.A.I. = 0,3. Lumina 1-2  $\mu$ . Tragia tristis subtype

Tragia tristis subtype.

P.A.I. = 0,3.

 $P = 27 \mu \quad E = 40 \mu$ 

P:E = 0.68.

Intectate; pilate. Pila not in a reticulum.

Tragia tristis MUELL. ARG.

Hassler 6645 [BM] Pl. XIII, 5

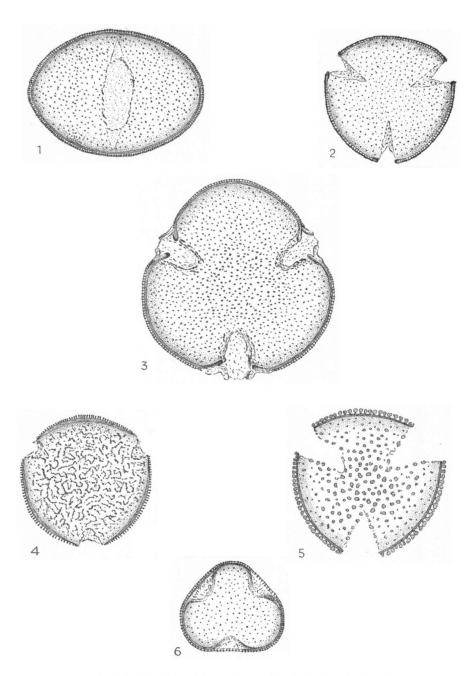


PLATE XIII. 1. Eleutherostigma lehmannianum; 2. Omphalea diandra; 3. Astrococcus cornutus; 4. Tragia stolziana; 5. Tragia tristis; 6. Pachystilidium hirsutum.

Tragia geraniifolia BAILL. Herter 1006 A. [U]

Tragia volubilis LINN.
Focke 992 [U]
Tragia ramosa TORR.
Rogerson 4-6-1954 [U]

Tragia tristis subtype.  $P = 31.5 \mu E = 41.5 \mu P:E = 0.76.$  P.A.I. = 0.3. Pila ca.  $1 \mu$ . Tragia tristis subtype.

Tragia tristis subtype.

## Pachystylidium type

Triporate; spheroidal.

Apertures are circular in shape and sunk down. The borders are indistinct. Tectate; psilate.

The Pachystilidium type has circular apertures, of which the rims are indistinct. It is, however, not clear if the type belongs with certainty to the Plukenetia configuration. The ektexine is only very indistinctly weakened and it is therefore possible to call the type inaperturate. The endexine, however, is clearly thinned. In the author's opinion it is better to keep the type in the Plukenetia configuration and not in the Cnesmosa configuration.

Pachystilidium hirsutum (Blume) Pax et K. Hoffmann Elbert 3340 [L] Pl. XIII, 6 Pachystylidium type.  $P = E = 29 \mu$ . P.A.I. = 0,6-0,7.

#### CHIROPETALUM CONFIGURATION

Tricolpate or stephanocolpate (4); shape variable. Colpi long. No costae colpi. Colpus membrane (endexine) persistent. [Rarely tricolporate, but then pollen grains prolate in shape and an operculum present.]

This configuration is distinguished from the *Plukenetia* configuration by the colpus membrane, which is never ruptured.

#### Taxonomic discussion

The genera of the Chiropetalum type, Ditaxis type and Argythamnia type are closely related. Mueller (1866) and Bentham united them in one taxon, the genus Argythamnia (except Leucocroton). Pax and K. Hoffmann (1912c, 1931) kept the genera separated, and according to pollen morphology this seems the right opinion.

# Chiropetalum type

Tricolpate (or rarely tricolporate); spheroidal to prolate spehroidal. Colpi wide; operculum present; costae colpi absent. Tectate or intectate; finely reticulate. [Rarely colpus transversalis present.]

The Chiropetalum type and the Lasiocroton type are related by the presence of an operculum. Chiropetalum and its allies, however, are finely reticulate.

From the Ekman 16425a sheet ERDTMAN (1952) described *Leucocroton flavicans* as colpate. The Ekman sheet 12343 as examined by the

author from the Kew herbarium has tricolporate pollen grains. These pollen grains should be placed in a different configuration, although other characters (e.g. the operculum and fine reticulum) are consistent with the *Chiropetalum* type. Nothwithstanding the colpus transversalis the author prefers to place the pollen grains of *Leucocroton flavicans* in the *Chiropetalum* type.

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Chiropetalum lanceolatum A. Jussieu Wedermann 34 [U] Pl. XIV, 2
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Chiropetalum canescens PHIL.
Wedermann 773 [U]
Aonikena patagonica SPEGG.
[= Chiropetalum patagonica (SPEGG.)
O'DONN. et LOURTEIG]
Eyerden, Beetle and Grond 24370 [K]
Pl. XIV, 4
Leucoroton flavicans MUELL. ARG.
Ekman 13243 [K] Pl. XIV, 5

Chiropetalum type; spheroidal.  $P = E = 37,5 \mu$  P:E ca. 1. P.A.I. = 0,3. Lumina ca. 1  $\mu$ . Intectate. Chiropetalum type; prolate spheroidal. Lumina 1-2  $\mu$ . Intectate. Chiropetalum type; prolate spheroidal.  $P = 34,5 \mu$  E = 30,5  $\mu$  P:E = 1,12. P.A.I. = 0,3. Lumina 1-2  $\mu$ .

Chiropetalum type; subprolate.  $P = 30.5 \mu$  E = 25.5  $\mu$  P:E = 1.19. Colpus transversalis without costae. P.A.I. = 0,4. Lumina ca. 1  $\mu$ .

## Ditaxis type

Tricolpate; oblate to suboblate. The pollen grains are bi-laterally symmetric. One lobe of the three-lobed pollen grain is smaller than the other two. Colpi long and not very narrow.

Tectate; psilate; not reticulate.

This type has bi-symmetric pollen grains. Of the three lobes one is smaller than the other two. This curious shape was observed in six species of *Ditaxis*. Argythamnia candicans does not possess these bi-symetric pollen grains.

#### Taxonomic discussion

It could not be investigated whether other species of Argythamnia perhaps do show the Ditaxis type.

CROIZAT (1945) did not see any reason to separate *Ditaxis* and *Argythamnia*. The difference in pollen type, however, supports the opinion of PAX and K. HOFFMANN (1931), who maintain *Ditaxis* next to *Argythamnia*.

Ditaxis fendleri (MUELL. ARG.) PAK et K. HOFFM.
Stoffers 1703 [U] Pl. XIV, 1
Ditaxis catamarcensis (GRISEB.) PAK
Brizuela 759 [U]
Ditaxis diversiflora CLOKEY
Clokey 7576 [U]
Ditaxis fasciculata VAHL ex A. DE JUSS.
Boldingh 2733 A [U]

Ditaxis lancifolia Schlechtend. Broadway 9257 [U] Ditaxis salina Pax et K. Hoffm. Pedersen 2638 [U] Ditaxis type; oblate. P = 30,5  $\mu$  E = 51  $\mu$  P:E = 0,60. P.A.I. = 0,15. Ditaxis type. P = 33  $\mu$  E = 42  $\mu$  P:E = 0,79. Ditaxis type. Some pollen grains 4-colpate. Ditaxis type. P = 43  $\mu$  E = 50  $\mu$  P:E = 0,86. P.A.I. = 0,2 - 0,25. Ditaxis type. P.A.I. = 0,2 - 0,25. Ditaxis type.

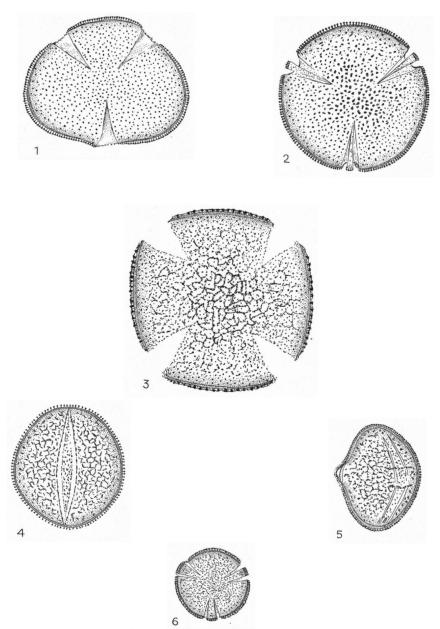


PLATE XIV. 1. Ditaxis fendleri; 2. Chiropetalum lanceolatum; 3. Argythamnia candicans; 4. Aonikena patagonica; 5. Leucocroton flavicans; 6. Lasiocroton macrophyllus.

## Argythamnia type

Stephanocolpate; oblate spheroidal. Colpi long, not very narrow. Tectate; on the top of the tectum a reticulum is superimposed.

Although there are some similarities with the *Ditaxis* type, shape as well as structure differ too much for the pollen grains to be placed in the same type.

Argythamnia candicans Sw. Boldingh 3401 B. [U] Pl. XIV, 3 Argythamnia type; 4 (-3)-colpate. P = 42  $\mu$  E = 45  $\mu$  P:E = 0,93. P.A.I. = 0,5.

### Lasiocroton type

Tricolpate or stephanocolpate; spheroidal to oblate spheroidal. Colpi wide; operculum present; no costae present. Tectate; psilate; no reticulum present.

The difference between the *Chiropetalum* type and *Argythamnia* type is found in the absence of a reticulum.

According to ERDTMAN (1952) the pollen grains of Lasiocroton macrophyllus are reticulate. With the aid of a phase contrast microscope it was possible to conclude, that the pollen grains of the Harris 11868 specimen are certainly not reticulate.

### Taxonomic discussion

Lasiocroton and Adelia, together with Leucocroton (p. 66), formed the series Adeliiformes (Pax and K. Hoffmann 1914, 1931) in the subtribe Mercurialinae. These three genera are certainly related by the following characters:

- 1. Disc present in the male as well as in the female flowers. This disc is usually annular.
- 2. Ovarium rudiment in the male flower absent.
- 3. Petals absent. Number of sepals usually 5.
- 4. Plants usually dioecious and only growing in the New World.

Lasiocroton macrophyllus (SWARTZ) GRISEB. Harris 11868 [K] Pl. XIV, 6 P.A.I. = 0,3. Adelia ricinella Linn. Rutten-Pekelharing [U] Fuertes 833 [U] Lasiocroton type; spheroidal. (3-) 4-colpate.  $P = E = 33 \mu$ . Operculum narrow. P.A.I. = 0,4-0,5.

#### CEPHALOMAPPA CONFIGURATION

Tricolpate. Colpi very short; costae colpi present.

## Cephalomappa type

Tricolpate; oblate spheroidal. Colpi very short; costae colpi present. Tectate; intra-reticulum coarse.

The Cephalomappa type differs completely from all other types in

the Crotonoideae. Besides the very coarse reticulum the colpi are short and bordered by distinct costae colpi.

#### Taxonomic discussion

The genus Cephalomappa is certainly related to those in the Cladogynos type (p. 93).

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Cephalomappa malloticarpa J. J. SMITH Herb. Lugd. Batav. 924.536. 117 [L] Pl. XV, 5 Cephalomappa type. P = 30,5 \mu P:E = 0,94. Colpi length ca. 4 \mu. Lumina 2-3 \mu.
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#### SUMBAVIA CONFIGURATION

Tricolporate - stephanocolporate; usually Polar axis shorter than Equatorial axis.

Colpi wide or narrow.

Tectate or intectate; reticulate. Reticulum coarse.

The most important character of this configuration is the coarse reticulum. Some species in the *Cladogynos* configuration (p. 93) also have a fairly coarse reticulum but in that case they have a Polar axis larger than the Equatorial axis. In the *Sumbavia* configuration the pollen grains usually have a Polar axis shorter than the Equatorial axis.

#### Taxonomic discussion

Most genera in this configuration belong to the Chrozophoreae (Bentham, 1880; Pax and K. Hoffmann 1912c, 1931). These authors split up this tribe in groups. Bentham's third group is comparable with the subtribe Regulares of Pax and K. Hoffmann, in which Sumbavia, Chrozophora and Caperonia are found.

### Sumbavia type

Tricolporate; oblate spheroidal to oblate. Colpus transversalis large; broad or long and narrow. Colpi long; costae colpi present or absent. Intectate; reticulate. Reticulum coarse.

The pollen grains of the *Sumbavia* type are always tricolporate and have a distinct reticulum. They differ from the *Chrozophora* type and *Caperonia* subtype by the number of colpi and from the *Philyra* subtype by the coarse reticulum and elongated colpus transversalis.

In 1952 Erdtman described pollen grains of *Monotaxis grandiflora* which are subprolate and have a membrana granulata. This discription is not in accordance with the data found by the author. The pollen grains examined were suboblate and had no membrana granulata. The exine structure, however, is quite the same.

#### Taxonomic discussion

In the system of Pax and K. Hoffmann (1931) Thyrsanthera and Sumbaviopsis are placed next to Sumbavia. This is in agreement with the close relationship of the pollen grains.

Monotaxis differs from the other genera in the Sumbavia type by the absence of stellate hairs and the presence of a disc and an ovarium rudiment in the male flowers. The flowers have, however, also five

petals as in most genera of the Sumbavia type.

In many respects Melanolepis resembles Sumbavia and Sumbaviopsis. As with these genera there is no disc or ovarium rudiment in the male flower. On the other hand the female flower possesses a disc. The plants have stellate hairs, they are monoecious and show similarity in the form of the leaves. Melanolepis, however, has no petals. Baillon (1858) therefore calls Melanolepis a Sumbavia without petals. Mueller (1866), Bentham (1880) and Pax and K. Hoffmann see Mallotus as nearest relation of Melanolepis. An accurate comparison of the corresponding characters will have to decide whether Melanolepis is closer related to Mallotus than to Sumbavia and Sumbaviopsis.

Sumbavia rottleroides BAILL. Merrill Sp. Blancoanae 993 [L] Pl. XV, 2

Sumbaviopsis albicans J. J. SMITH Backer 19016 [L]

Thyrsanthera suborbicularis Pierre ex Gagnepain Pierre 512 [K] Pl. XV, 3

Melanolepis multiglandulosa (REINW. ex Blume) Reich.F. et Zollinger Hort. Bogor. 2127 [U] Pl. XV, 1

Monotaxis grandiflora ENDL. Pritzel XI, 1900 [U] Pl. XV, 4 Sumbavia type; oblate spheroidal.  $P=43~\mu~E=46~\mu~P:E=0,93.$  m:e=0,6.~P.A.I.=0,35. Costae colpi absent. Lumina 3-4  $\mu$ . Sumbavia type; suboblate.  $P=32~\mu~E=38,5~\mu~P:E=0,80.$  P.A.I. = 0,3. Lumina 1-2  $\mu$ . Costae colpi. Sumbavia type; suboblate.  $P=43,5~\mu~E=52~\mu~P:E=0,84.$  P.A.I. = 0,3. Lumina 1-2  $\mu$ . Costae colpi absent. Sumbavia type; suboblate.  $P=32~\mu~E=38,5~\mu~P:E=0,82.$  P.A.I. = 0,3 — 0,35. Lumina 1-3  $\mu$ . Costae transversales. Sumbavia type; oblate.  $P=30,5~\mu~E=36~\mu~P:E=0,84.$   $P=30,5~\mu~E=36~\mu~P:E=0,84.$   $P=30,5~\mu~E=36~\mu~P:E=0,84.$   $P=30,5~\mu~E=36~\mu~P:E=0,84.$   $P=30,5~\mu~E=36~\mu~P:E=0,84.$   $P=30,5~\mu~E=36~\mu~P:E=0,84.$   $P=30,5~\mu~E=36~\mu~P:E=0,84.$ 

Lumina 1–2  $\mu$ .

# Chrozophora type

Stephanocolporate; suboblate – oblate. Colpus transversalis broad, isodiametric; costae. Colpi short; costae colpi. Intectate (?); reticulate. Reticulum coarse.

The lumina of the reticulum are much larger in the Chrozophora type than in any other type of the Sumbavia configuration. Besides this character the colpi are short and the number of colpi is large (at least six). In the subfamily of the Crotonoideae pollen grains with more than three or four colpi are rather rare. Only the related Caperonia type has also six colpi.

Chrozophora plicata (VAHL) A. DE JUSS. Feinbrun etc. 49169 A. [U] Pl. XV, 6 Chrozophora type; (7-) 8 (-9)-colporate; suboblate - oblate. P = 60  $\mu$  E = 80  $\mu$  P:E = 0,75. m:e = ca. 1. P.A.I. > 0,5. Lumina 4-6  $\mu$ . Chrozophora type. 6-colporate.

Chozophora tinctoria (LINN.) A. DE JUSS. Faure 31126 A. [U]

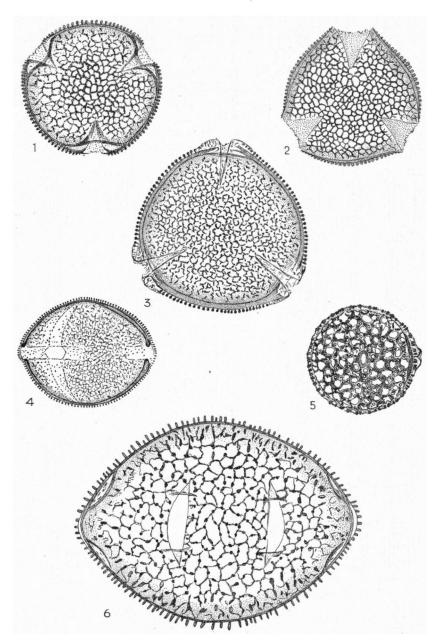


PLATE XV. 1. Melanolepis multiglandulosa; 2. Sumbavia rottleroides; 3. Thyrsanthera suborbiculatus; 4. Monotaxis grandiflora; 5. Cephalomappa malloticarpa; 6. Chrozophora plicata.

## Caperonia type

Stephanocolporate or tricolporate; oblate spheroidal - prolate spheroidal. Colpus transversalis broad, circular - broad elliptic.

Colpi long and narrow; costae colpi absent.

Intectate; reticulate. Lumina fairly small. Columellae elongated.

This type resembles the *Chrozophora* type in its number of colpi (usually six). The colpi are, however, longer, and the lumina of the reticulum much smaller. The *Philyra* subtype differs from the *Sumbavia* type by its large circular colpus transversalis.

#### Taxonomic discussion

MUELLER (1866) and BENTHAM (1880) included *Philyra* in the genus *Argythamnia*. Pax and K. Hoffmann (1912c, 1931) separated *Philyra* from this genus and referred it close to *Caperonia*. Pollen-morphologically the genus is more related to *Caperonia* than to *Argythamnia* and its allies.

## Caperonia subtype

Pollen grains stephanocolporate.

Caperonia palustris (LINN.) St. HILL. Lanjouw 1044 [U] Pl. XVI, 4

Caperonia serrata PRESL de Wit 429 [WAG] Caperonia corchoroides MUELL. ARG. Rombouts 358 [U]

Philyra subtype

Pollen grains tricolporate.

Philyra brasiliensis KLOTZSCH Hassler 12323 [K] Pl. XVI, 2 Caperonia subtype; oblate spheroidal.  $P=32~\mu~E=33~\mu~P:E=0,94.$  m: e ca. 1. P.A.I. = 0,3. Lumina ca. 1  $\mu$ . Caperonia subtype; spheroidal.  $P=E=39~\mu$ . P.A.I. = 0,5. Caperonia subtype.

Philyra subtype; prolate spheroidal.  $P=38,5~\mu~E=35~\mu~P:E=1,10.$   $m:e=0,7.~P.A.I.=0,4.~Lumina~1-2~\mu.$ 

## Caryodendron type

Tricolporate; oblate spheroidal – suboblate. In polar view pollen grains convex triangular. Colpus transversalis broad; no costae present. Colpi narrow; no costae colpi.

Tectate, tectum perforatum; psilate. Intra-reticulum coarse, but lumina small.

The Caryodendron type is closely related to the Sumbavia type. The pollen grains are, however, tectate and the lumina of the intrareticulum are fairly small. The tectum, moreover, is perforate. The holes in the tectum are very small and only visible with high magnification.

#### Taxonomic discussion

Caryodendron is not closely related to the genera in the Sumbavia type. It lacks a stellate indument and petals. Bentham (1880) as well as Pax and K. Hoffmann (1914, 1931) thought of a relationship

with Alchornea. Caryodendron possesses, however, an annular disc in the female flower, a character never to be found in the genera of the Alchornea type, but always present in the genera of the Sumbavia type. This character is also present in genera of the Bernardia type and Speranskia type. On the other hand the authors cited above stated, that in their opinion Adenophaedra is closely related to Caryodendron. Adenophaedra belongs pollen-morphologically to the Bernardia type (p. 75). The pollen grains of Caryodendron show indeed some resemblance with this type, but the differences are too many for a combination of Adenophaedra and Caryodendron in the same type.

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Caryodendron orinocense KARST.
  Krukoff 5553 [U] Pl. XVI, 3
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Caryodendron grandifolius Muell. Arg. Kuhlmann et Ducke 16598 [U]

Caryodendron type; suboblate.  $P=29~\mu~E=34.5~\mu~P:E=0.84.$  m:e=0.4.~P.A.I.=0.3.Lumina ca.  $1 \mu$ ; muri thick. Caryodendron type; oblate spheroidal.  $P=31.5~\mu~E=33~\mu~P:E=0.95.$  P.A.I. = 0,2.

## Nealchornea type

Tricolporate; oblate spheroidal. In polar view the pollen grains circular. Colpus transversalis broad; ora large. Colpi short and wide.

Intectate; reticulate. Reticulum coarse.

The pollen grains have a Polar axis shorter than the Equatorial axis and there is also a reticulum with large lumina. For these reasons the type is placed in the Sumbavia configuration. It differs from the other types by the three short colpi.

#### Taxonomic discussion

Nealchornea is not related to any genus discussed in other types of the Sumbavia configuration. Its nearest allies are perhaps Hamilcoa, Plagiostyles and Pimeleodendron (p. 103). Striking corresponding characters with these genera are the very short to sessile anthers and thickened pedicels.

Pax and K. Hoffmann (1931) put the genus in the subtribe Cluytiinae with e.g. Trigonostemon, Moultonianthus and Trigonopleura. The latter three genera are all placed in different pollen types and even in different configurations.

```
Nealchornea yapurensis Huber
  Krukoff 4967 [U] Pl. XVI, 1
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Nealchornea type.  $P = 27 \mu E = 30 \mu P:E = 0.90.$  m:e = ca. 1. Lumina 3-4  $\mu$ .

#### Bernardia configuration

Tricolporate or stephanolcoporate (except Erismanthus type). Usually Polar axis larger than Equatorial axis.

Colpi narrow; costae colpi.

Tectate or intectate; reticulate. Reticulum fine; lumina  $< 2 \mu$  (Cyttaranthus without reticulum).

The types of the *Bernardia* configuration are finely reticulate and have usually a Polar axis larger than the Equatorial axis. Cyttaranthus

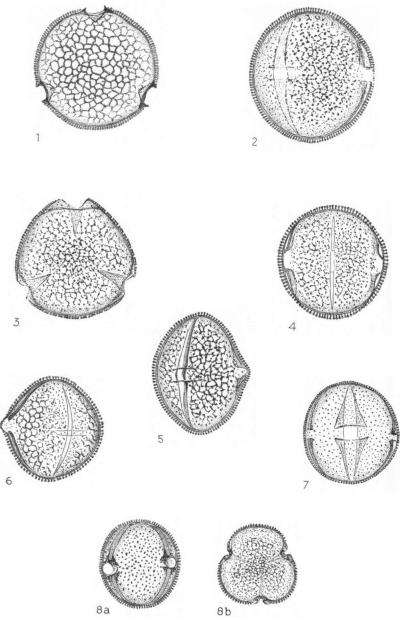


PLATE XVI. 1. Nealchornea yapurensis; 2. Philyra brasiliensis; 3. Caryodendron orinocensis; 4. Caperonia palustris; 5. Pseudagrostistachys africana; 6. Heterocalyx laoticus; 7. Cyttaranthus congolensis; 8. Bernardia pulchella.

only is not reticulate, but in other respects the pollen grains of this genus closely resemble the *Speranskia* type. Although placed in this configuration by their fine reticulum and prolate shape, the *Moultonianthus* type and the *Clutia* type differ largely from the *Bernardia* type and the *Speranskia* type.

## Bernardia type

Tricolporate; spheroidal prolate – subprolate. Pollen grains in polar view distinctly three-lobed.
Colpus transversates splicial RAL graph Colpus transversates splici

Colpi narrow; costae colpi. P.A.I. small < 0.25. Intectate; finely reticulate; lumina ca. 1  $\mu$ .

The Bernardia type differs from the Speranskia type by its distinctly three-lobed pollen grains.

### Taxonomic discussion

MUELLER originally described Adenophaedra as a section of Bernardia. In later publications he raised it to a separate genus. Bentham (1880) and Pax and K. Hoffmann (1931) maintained the two genera and even placed them in a far-away group to which Alchornea belongs. Pollen-morphological study of the two genera showed a close relation, so it seems better to maintain Adenophaedra in the vicinity of Bernardia. Both genera are restricted to the New World.

Bernardia pulchella (BAILLON) MUELL.
Tessman 6078 [U] Pl. XVI, 8
Bernardia lorentzii MUELL. ARG.
Schulz 1554 [U]

Bernardia corensis (JACQ.) MUELL. ARG. Boldingh 665 B. [U]

Adenophaedra minor DUCKE Herb. Rio 10386 [U] Bernardia type; prolate spheroidal.  $P=25,5~\mu~E=24~\mu~P:E=1,07.$  m:e=ca.~0,5.~P.A.I.=0,15. Bernardia type.  $P=29~\mu~E=25~\mu~P:E=1,17.$  P.A.I.=0,2. Bernardia type.  $P=31~\mu~E=25~\mu~P:E=1,24.$  P.A.I.=0,2. Bernardia type.  $P=31~\mu~E=25~\mu~P:E=1,24.$  P.A.I.=0,2. Bernardia type.

P = 28  $\mu$  E = 27  $\mu$  P:E = 1,04. Colpus transversalis narrow; m:e < 0,5. P.A.I. = 0,1-0,15.

# Speranskia type

Tricolporate; spheroidal - prolate spheroidal. Pollen grains in polar view convex triangular.

Colpus transversalis large; costae present or absent.

Colpi narrow; costae colpi. Sometimes costae indistinct.

Tectate; intra-reticulum present or rarely absent (Cyttaranthus).

This type looks somewhat like the *Bernardia* type but differs by its polar view and the shape of the colpus transversalis. *Cyttaranthus* belongs undoubtedly to this type but lacks the fine reticulum.

## Taxonomic discussion

Genera with a Speranskia type are only found in the Old World. They have several characters in common.

- 1. In the female flowers always a disc present.
- 2. Anthers usually pendulous; connectives frequently appendiculated.

3. In the male flowers an ovarium rudiment is wanting except in some Agrostistachys species.

4. The Asiatic genera have petals and an extrastaminal disc. The African representatives frequently lacks petals, while glands are always present on the receptaculum.

5. Stellate hairs never present.

Speranskia pekinensis Pax et K. Hoffmann Bullock 10 Mai. 1888 [K] Pl. XVII, 8

Pseudagrostistachys africana (MUELL. ARG.)
PAX et K. HOFFM.
Moller and Quintus 16 [U] Pl. XVI, 5

Cyttaranthus congolensis LÉONARD Collens 3541 [K] Pl. XVI, 7

Agrostistachys indica DALZ. Stocks [U]

Agrostistachys sessilifolia (Kurz) PAX et K. Hoffmann East Ind. Comp. 4739 [U]

Heterocalyx looticus GAGNEP. Kerr 20895 [K] Pl. XVI, 6

Argomuellera macrophylla PAX de Wit 32 [WAG] Pl. XVII, 3

Swynnerton 119 A [BM]

Discoglypremna caloneura (PAX) PRAIN Zenker 2643 [L] Speranskia type; subprolate. P = 28,5  $\mu$  E = 24  $\mu$  P:E = 1,16. m:e = 0,5-1. P.A.I. = 0,10. Lumina < 1  $\mu$ .

Speranskia type; subprolate.  $P = 33 \mu E = 26,5 \mu P:E = 1,24.$  m:e < 0,5. P.A.I. = 0,3. Lumina ca. 1  $\mu$ .

Speranskia type; prolate spheroidal.  $P = 30 \mu E = 28.5 \mu P:E = 1.05.$ P.A.I. = 0.35. Not reticulate!

Speranskia type; oblate spheroidal.  $P=31,5~\mu~E=33,5~\mu~P:E=0,94.$   $m:e<0,5.~P.A.I.=0,25.~Lumina~1-2~\mu.$  Margo (thinning of the ektexine).

Speranskia type; subprolate.  $P = 26,5 \mu$  E = 23,5  $\mu$  P:E = 1,12. P.A.I. = 0,2-0,25. Costae transversales. Margo (thinning of the ektexine).

Speranskia type; spheroidal.  $P = E = 30 \mu$ . P.A.I. = 0,25. Lumina 1-2  $\mu$ . Margo (thinning of the ektexine).

Speranskia type; spheroidal.  $P = 25,5 \mu$   $E = 25 \mu$  P:E = 1,02. m:e = 0,7-0,8. P.A.I. = 0,4. Lumina ca. 1  $\mu$ . Costae transversales absent.

P = 30  $\mu$  E = 28,5  $\mu$  P:E = 1,08. In other characters as above. Speranskia type; prolate spheroidal. P = 27  $\mu$  E = 25  $\mu$  P:E = 1,09. P.A.I. = 0,25-0,3. Lumina ca. 1  $\mu$ .

# Moultonianthus type

Tricolporate; spheroidal. In polar view the pollen grains circular. Colpus transversalis narrow and small; costae. Colpi narrow and long; costae colpi. Intectate; reticulum fine.

The pollen grains of the *Moultonianthus* type are finely but distinctly reticulate and are therefore placed in the *Bernardia* configuration. The shape, polar view and colpus transversalis of this type differs greatly from the other types in this configuration.

#### Taxonomic discussion

According to v. Steenis (1948) Moultonianthus is related to Erismanthus in several characters. Unfortunately pollen grains of Erismanthus indochinensis do not agree with those of Moultonianthus leem-

bruggianus. Perhaps other species of Erismanthus will show more relationship in their pollen-morphology.

```
Moultonianthus leembruggianus (Boerl. et Koord.) v. Steenis Serawak 464 [L] Pl. XVII, 1 Moultonianthus type. P = E = 23.5 \mu. m:e < 0.5. P.A.I. = 0.4. Lumina 1-2 \mu.
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### Erismanthus type

Tricolpate; oblate spheroidal. In polar view pollen grains circular. Colpi narrow. Rim of the colpi irregularly ruptured. Tectate; psilate. Columellae short.

Pollen grains of *Erismanthus indochinensis* distinctly differ from the *Moultonianthus-type*. There are no composite apertures and the grains are tectate. ERDTMAN (1952) examined *Erismanthus oblique* and on the contrary did find composite apertures in this specimen.

```
Erismanthus indochinensis Gagnep. D'Alleizette 942, 20-4-1908 [L] Pl. XVII, 4 Erismanthus type. P = 24 \mu E = 25,5 \mu P:E = 0,93. P.A.I. = 0,4.
```

### Clutia type

Tricolporate; subprolate. In polar view the pollen grains triangular. Colpus transversalis small; costae. Colpi narrow; costae colpi. Margo present (thinning of the ektexine). Tectate; intra-reticulum indistinct.

The Clutia type strongly recalls the Phyllanthus pentaphyllus subtype in the subfamily of the Phyllanthoideae. The type differs, however, by the shape of the colpus transversalis. The Clutia type differs from the Bernardia type by its polar view. The Bernardia type is three-lobed, the Clutia type triangular.

#### Taxonomical discussion

This African genus stands alone in the Euphorbiaceae. The inflorescence and habit are those of Phyllanthus; but the essential characters are those of the Chrozophorae (BENTHAM, 1880).

```
Clutia type.
Clutia paxii KNAUF
  Crookewit 198 [WAG] Pl. XVII, 2
                                             P = 35 \mu
                                                          E = 27 \mu P:E = 1.30.
                                             m:e < 0.5.
Clutia natalensis BERNH.
                                          Clutia type.
                                             P = 43 \mu
                                                         E=33~\mu
  Wylie Jan. 1935 [U]
                                                                       P:E = 1,30.
                                             Reticulum indistinct.
                                          Clutia type. P = 52 \mu E = 40 \mu
Clutia abyssinica JAUB. et SPACH
  Stolz 1876 [Ŭ]
                                                                       P:E = 1,30.
                                             Reticulum indistinct.
Clutia rubricaulis ECKL. ex SOND.
                                          Clutia type.
                                             P = 54 \mu E = 41 \mu
Reticulum indistinct.
                                                                       P:E = 1,30.
  [= C. alaternoides Linn.]
  Lanjouw 165 [U]
```

### MALLOTUS CONFIGURATION

Tricolporate – stephanocolporate (4). Polar axis usually shorter than Equatorial axis, Colpi narrow.

Tectate (tectum perforate in Blumeodendron type); usually not reticulate; columellae short, capita indistinct.

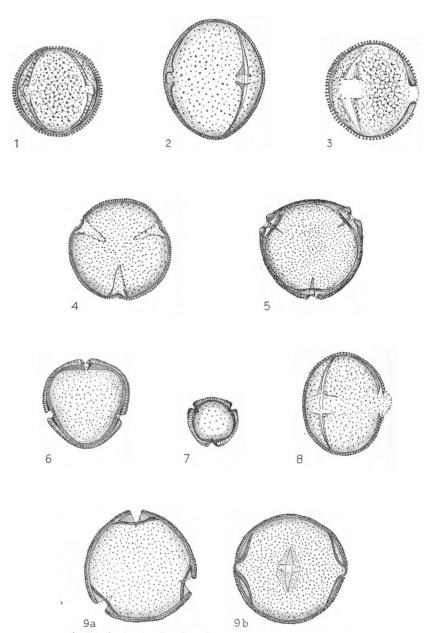


PLATE XVII. 1. Moultonianthus leembruggianus; 2. Clutia paxii; 3. Argomuellera macro-phylla; 4. Erismanthus indochinensis; 5. Mallotus albus; 6. Cleidion javanicum; 7. Macaranga spinosa; 8. Speranskia pekinensis; 9. Trewia nudiflora.

The most important characters of the *Mallotus* configuration are the short columellae and the indistinct capita. Besides these important characters the pollen grains are usually not reticulate.

The Mallotus type, Blumeodendron type, Acalypha type, Alchornea type, Discocleidion type and Mareya type are closely related and differ only

in minor points.

The Ricinus type and Pera type are connected by the very narrow colpi and nearly visible columellae. They certainly belong to the Mallotus configuration although the shape of the pollen grains is often prolate.

The Seidelia type and Leidesia type are related to each other, es-

pecially in their exine structure and polar view.

The Microdesmis type and Cheilosa type are placed in the Mallotus configuration because of their shape and short columellae. The differences with the other types in this configuration, however, are considerable.

## Mallotus type

Tricolporate – stephanocolporate (4); oblate spheroidal – suboblate. Sometimes spheroidal.

Colpus transversalis usually small; costae.

Colpi narrow; costae colpi.

Tectate; psilate or scabrate. Rarely intra-reticulate (Adriana).

The pollen grains are easily distinguished from the other types. The colpi, are short (P.A.I. is at least 0,4) and the shape is usually oblate spheroidal. The columellae are so short, that only with difficulty the tectum can be distinguished from the endexine. Costae colpidistinct.

#### Taxonomic discussion

In the system of Mueller (1866) genera of the Mallotus type are all found in his subtribe Euacalypheae. Bentham (1880) placed them in the subtribe Acalypheae and Pax and K. Hoffmann (1914, 1931) in the subtribe Mercurialinae of the tribe Acalypheae. In all three systems the genera are mixed up with those, closely allied to them in the Acalypha type, Alchornea type, Blumeodendron type, Discocleidion type and Mareya type. Corresponding characters are:

- 1. A disc is wanting in the male as well as in the female flowers.
- 2. An ovarium rudiment is wanting.
- 3. Usually numerous stamens are inserted on a convex receptaculum (Macaranga with few stamens has no receptaculum). Most times the receptaculum is glabrous. Sometimes it is provided with glands (some species of Mallotus and Pycnocoma) or hairs (Wetria).
- 4. The majority of the genera grows in the Old World; only Avellanita is from Chile.

#### Mallotus subtype

Pollen grains psilate.

```
Mallotus subtype; oblate spheroidal.

P = 30 \mu E = 30.5 \mu P:E = 0.98.

m:e < 0.5. P.A.I. = 0.4.
Mallotus albus (ROXB.) MUELL. ARG.
   Thomson [U] Pl. XVII, 5
                                                 Mallotus subtype.
Mallotus claoxyloides (F. v. MUELLER)
                                                   P = 30 \mu \dot{E} = 32,5 \mu P:E = 0,95.
   Muell. Arg.
   Buysman 1874 [U]
                                                   P.A.I. = 0,4.
                                                 Mallotus subtype.
Mallotus miquelianus (Scheff.) Pax et
                                                   P = 20 \mu E = 21 \mu P:E = 0.95.
   K. Hoffm.
                                                   P.A.I. = 0.4.
   Rutten-Kooistra 27 [U]
Mallotus repandus (WILLD.) MUELL. ARG.
                                                 Mallotus subtype. Too young.
   Griffith 4760 [U]
Coelodiscus lanceolatus GAGNEP.
                                                 Mallotus subtype.
                                                   P = 25.5 \mu E = 27 \mu P:E = 0.94.

P.A.I. = 0.4.
   D'Alleizette Jan. 1909 [L]
                                                 Mallotus subtype.
Trewia nudiflora LINN.
                                                   P = 27 \mu \dot{E} = 30.5 \mu P : E = 0.89.
  Hohenacker 453 [U] Pl. XVII, 9
                                                   Colpi very short. P.A.I. > 0.75.
Avellanita bustillosii PHILIPPI
                                                 Mallotus subtype
                                                   P = 25.5 \mu E = 26 \mu P:E = 0.98.
Costae colpi indistinct. P.A.I. = 0.45.
   Philippi [BM]
Cordemoya integrifolia (WILLD.) BAILLON
                                                 Mallotus subtupe.
   Gover. Pires to Kew Nov. 1908 [K]
                                                   P = 20 \mu \quad \hat{E} = 21.5 \mu \quad P:E = 0.93.
                                                   P.A.I. = 0.7.
                                                Mallotus subtype; 3-4 colporate.

P=37,5~\mu~E=39~\mu~P:E=0,96.

Colpi very short. P.A.I. = 0,7.
Coccoceras borneënse J. J. SMITH
   F. H. Endert 2078 [L]
                                                Mallotus subtype.

P = 20 \mu E = 22,5 \mu P:E = 0,88.

Colpi very short. P.A.I. > 0,7.

Mallotus subtype; 4- (3) colporate.
Deuteromallotus acuminatus (BAILLON)
   PAX et K. HOFFM.
   D'Alleizette Mad. Nov. 1908 [L]
Wetria macrophylla (Blume) J. J. Smith
Cult. Hort. Bog. IXA. 124 [U]
                                                   P = E = 40 \mu P.A.I. = 0.5.
                                                 Mallotus subtype.
Adriana quadripartita (LABILL.) GAUDICH.
                                                   P = 33.5 \mu E = 35 \mu P:E = 0.96.
P.A.I. > 0.5. Intra-reticulate.
  F. v. Mueller, Freemantle 1861 [U]
                                                Mallotus subtype. P = 25.5 \mu E = 27 \mu P:E = 0.95. P.A.I. > 0.7.
Cleidion javanicum Blume
   Cult. Hort. Bog. IX A. 38 [U]
   Pl. XVII, 6
                                                 Mallotus subtype.
Cleidion angustifolium Pax et K. HOFFM.
                                                   P = 19 \mu E = 18 \mu P:E = 0.95.
  Le Rat 468 [P]
                                                   P.A.I. > 0,7.
Macaranga spinosa Muell. Arg.
                                                 Mallotus subtype.
   Steiner 1292 [U] Pl. XVII, 7
                                                   P = E = 14.5 \mu P: E = ca. 1.
                                                   P.A.I. > 0.5.
                                                 Mallotus type.
Macaranga harveyana (MUELL. ARG.)
                                                   P = 17 \hat{\mu} E = 17 \mu P:E = ca. 1.
   Muell. Arg.
   Yuncker 15980 [U]
Macaranga densiflora WARB.
                                                Mallotus subtype; P = E = 17 \mu.
```

Pycnocoma subtype
Pollen grains scabrate.

Gjellerup 236 [U]

This subtype differs from the *Mallotus* subtype by the scabrae on the tectum. In *Pycnocoma cornuta* the colpi are longer than those in other pollen grains of the *Mallotus* type. The colpi of *Pycnocoma macrophylla*, however, are as short as the others.

Pycnocoma cornuta Muell. Arg.

Leeuwenberg 3003 [U] Pl. XVIII, 1

Pycnocoma subtype.

P = 33,5  $\mu$  E = 35,5  $\mu$  P:E = 0,94.

m:e = 0,4. P.A.I. = 0,3.

Pycnocoma macrophylla Bentham Zenker 1251 [L] Pycnocoma subtype.  $P = 30.5 \mu E = 32 \mu P:E = 0.95.$ P.A.I. > 0.5.

## Blumeodendron type

Tricolporate; oblate spheroidal. Colpus transversalis small and narrow; costae. Colpi very short and narrow; costae colpi. Tectum perforatum that may be regarded as a coarse reticulum.

This type is closely related with the *Mallotus* type. The tectum, however, is perforated. The holes are rather wide and it is perhaps better to speak of a coarse reticulum. The muri of the reticulum are thick and consist of many small columellae.

#### Taxonomic discussion

The affinities of Blumeodendron with Botryophora are extensively explained by AIRY-SHAW (1960).

Without doubt *Ptychopyxis* has several characters in common with *Blumeodendron* and *Botryophora*.

- 1. In the male flowers a disc and an ovarium rudiment are absent.

  Many stamens inserted on a receptaculum.
- 2. In the female flowers a disc is present. (AIRY-SHAW 1960, states, that in *Ptychopyxis* at least some species certainly have a disc.)

Blumeodendron tokbrai (BLUME) KURZ Achmad 216 [L] Pl. XVIII, 6 Ptychopyxis costata MIQUEL Kings collector 6740 [L] Ptychopyxis kingii RIDLEY de Haviland et Hose 3673K [K] Botryophora geniculata (MIQ.) BEUMÉE ex AIRY-SHAW Docters v. Leeuwen 7707 [L] Blumeodendron type.  $P=21~\mu~E=23~\mu~P:E=0,91.$  Blumeodendron type.  $P=18,5~\mu~E=21~\mu~P:E=0,88.$  Blumeodendron type.  $P=19,5~\mu~E=21~\mu~P:E=0,83.$  Blumeodendron type.  $P=22~\mu~E=23,5~\mu~P:E=0,93.$  P.A.I. = 0,5-0,6. Colpi somewhat longer.

# Acalypha type

Tricolporate or stephanocolporate; oblate spheroidal to suboblate. The pollen grains are quasi porate. With high magnification the apertures are undoubtedly composite apertures.

Colpus transversalis and colpus of the same size; costae. Tectate; psilate.

The Acalypha type is undoubtedly related to the Mallotus type. The dimensions of the colpi and the colpi transversales, however, are so small that the composite apertures resemble pori.

Acalypha indica LINN.
Dewan, 12-9-1957 [U] Pl. XVIII, 4
Acalypha aronioides PAX et K. HOFFM.
Ellenberg 1064 [U]
Acalypha racemosa WALLICH ex BAILLON
Versuchsanst. 410 [U]
Acalypha platyphylla MUELL. ARG.
Killip et Garcia 33764 [U]
Acalypha scandens BENTHAM
Y. Mexia 6386 [U] Pl. XVIII, 5
Acalypha diversifolia JACQ.
Versteeg 466 [U]

Acalypha type.  $P = 14,5 \mu$  E = 16  $\mu$  P:E = 0,90. Acalypha type; 4-5 (col)-porate.  $P = 14 \mu$  E = 16,5  $\mu$  P:E = 0,85. Acalypha type; 3 (col)-porate.  $P = 12 \mu$  E = 13,5  $\mu$  P:E = 0,89. Acalypha type; 3 (col)-porate.  $P = 13,5 \mu$  E = 14,5  $\mu$  P:E = 0,93. Acalypha type; 4-5 (col)-porate.  $P = 17,5 \mu$  E = 19,5  $\mu$  P:E = 0,90. Acalypha type. Too young.

### Alchornea type

Tricolporate; spheroidal to suboblate. Colpus transversalis with costae. Colpi narrow; costae colpi. Operculum present. Tectate; psilate.

In shape and structure the *Alchornea* type is related to the *Mallotus* type. The pollen grains differ from the latter type by the presence of an operculum.

#### Taxonomic discussion

Genera of this type have much resemblance with those of the *Mallotus* type. In the systems of Mueller (1866), Bentham (1880) and Pax and K. Hoffmann (1931) they are mixed up with them.

Indeed the characters of this group correspond well with those described in the *Mallotus* type (p. 79). There are only a few indistinct differences.

- 1. Number of stamens is often less than 20. Neotrewia, Gavarretia and Polyandra have up to 60 stamens (In the Mallotus type up to several hundreds).
- 2. Receptaculum usually flat and peltate (not in species with many stamens). As in the *Mallotus* type glands are absent or scarce (*Alchornea* and *Neotrewia*).

Coelebogyne and Lepidoturus are genera, which nowadays are considered as belonging to Alchornea.

Originally Croizat placed Adenophaedra prealtum (Croizat) Croizat in the genus Cleidion (1943c). This genus has its widest spread in Asia and was said to have some representatives in America. Afterwards Croizat's opinion was that all American Cleidion species belong to Adenophaedra (1946a). Pollen-morphological investigation shows, that pollen grains of the species Adenophaedra prealtum on no account resemble those of Adenophaedra minor Ducke. The diagnosis of Adenophaedra prealtum, however, completely fits into the new genus Polyandra described by Léal (1951). Léal moreover published a photograph of the pollen grains of this genus, from which it is clear that the pollen grains belong to the Alchornea type like those of Adenophaedra prealtum.

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Alchornea villosa (BENTH.) MUELL. ARG.
                                              Alchornea type; oblate spheroidal.
  Bosch Proefst. F. 1152 [U] Pl. XVIII, 2
                                                 P = 25 \mu E = 26.5 \mu P:E = 0.94.
                                                 m:e = 0.6. P.A.I. = 0.4.
                                              Alchornea type. P = 24 \mu E = 26,5 \mu P:E = 0,92. P.A.I. > 0,5.
Alchornea rugosa (Lour.) Muell. Arg.
  Elmer 14353 [U]
                                              Alchornea type.
Alchornea schomburgkii Klotzsch
  Hilleris-Lambers 1922
                                                 P = 24.5 \mu E = 24.5 \mu P:E = 1.
                                               Alchornea type. P = E = 27 \mu.
Alchornea triplinervia (SPRENG.) MUELL.
  Regnell III, 1066 a [U]
                                              Alchornea type. P = 20,5 \mu E = 22 \mu P:E = 0,93. P.A.I. = 0,35–0,4.
Lepidoturus laxiflorus BENTH.

[= Alchornea laxiflora (BENTH.) PAX
  et K. Hoffmann]
  Faulkner 531 [K]
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```
Alchornea type.

P = 20 \mu E = 23 \mu
                                                                               P:E = 0.88.
  ilicifolia (J. Smith) Muell. Arg.]
  F. v. Muell., Rockhampton 1873
[K] Pl. XVIII, 7
                                                  P.A.I. = 0,4-0,45.
                                               Alchornea type. P = 21 \mu E = 22.5 \mu P:E = 0.93. P.A.I. = 0,4. Operculum consists of
Neotrewia cumingii (MUELL. ARG.) PAX
  et K. Hoffm.
  Elmer 15623 [U]
                                                  only one row of pila.
Gavarretia terminalis BAILL.
                                                Alchornea type.
                                                  P = 27, 5\mu E = 30,5 \mu P:E = 0.91.
  Ducke 23511 [U]
                                                   P.A.I. = 0.3.
                                                Alchornea type. P = 19.5 \mu E = 22.5 \mu P:E = 0.88.
Conceveiba simulata STEYERMARK
  Krukoff 6643 [U]
                                                  P.A.I. = 0,4. Scabrate.
                                               Alchornea type. P = 21,5 \mu E = 22,5 \mu P:E = 0,93. P.A.I. = 0,4-0,45.
Conceveibastrum martianum (BAILL.) PAX
  et K. Hoffm.
  I.B.V. 23529 [U]
                                               Alchornea type. P = 25 \mu E = 27 \mu P:E = 0.92. P.A.I. = 0.4.
Veconcibea latifolia (Benth.) Pax et
  K. Hoffm.
   = Conceveiba latifolia BENTHAM]
  Spruce 2826 [BM]
                                                Alchornea type.

P = 24 \mu E = 30 \mu
Aparisthmium cordatum (Juss.) BAILL.
  Y. Mexia 6273 [U] Pl. XVIII, 3
                                                                                P:E = 0.80.
                                                   P.A.I. = 0,4.
                                                Alchornea type. P = 21,5 \mu E = 25 \mu P:E = 0,86. P.A.I. = 0,35.
Lautembergia multispicata BAILL.
  D'Alleizette Madag. Nov. 1906 [L]
Lautembergia coriacea (BAILL.) PAX
                                                Alchornea type; sometimes 4-colporate.
  D'Alleizette Madag. Oct. 1906 [L]
                                                Alchomea type. P = 21.5 \mu E = 24.5 \mu P:E = 0.88.
Bocquillonia sessiliflora BAILL.
  Balansa 274 [L]
                                                  P.A.I. = 0,35.
                                               Alchornea type. P = 25,5 \mu E = 29 \mu P:E = 0,88. P.A.I. = 0,45.
Adenophaedra prealtum (CROIZAT) CROIZAT
    = Polyandra LEAL?]
   Krukoff 6602 [U]
```

## Discocleidion type

Tricolporate; spheroidal to suboblate.

Coelebogyne ilicifolia J. SMITH [ = Alchornea

Colpus transversalis; costae as broad as the colpi.

Colpi narrow; broad margo present (thickening of the ektexine). P.A.I. small, smaller than 0,2.

Tectate; from psilate to coarsely warted. The ornamentation differs strongly.

The pollen type shows some resemblance to the Mareya type as well as to the Alchornea type. It differs from both types by its curious ornamentation. This ornamentation varies in the individual pollen grains. It is not certain whether other specimens of Discocleidion rufescens will show the same variation of ornamentation.

The type differs from the Mareya type by the presence of costae and from the Alchornea type by the absence of an operculum.

#### Taxonomic discussion

Discocleidion differs from the genera in the Alchornea type by the presence of a disc in the female flowers and many glands inserted on the receptaculum of the male flowers. These characters place the genus next to the genera of the Mareya type.

MUELLER first described Discocleidion as a section of Cleidion. PAX

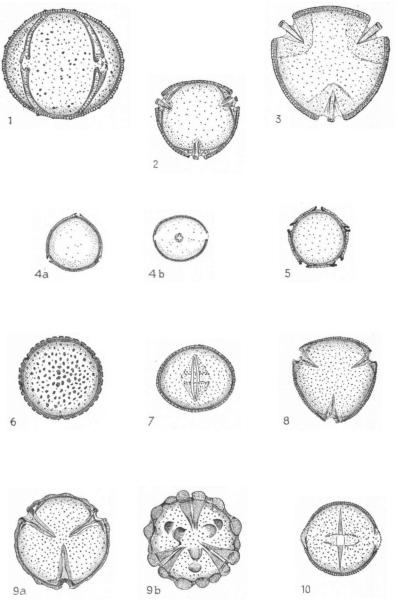


PLATE XVIII. 1. Pycnocoma cornuta; 2. Alchornea villosa; 3. Aparisthmium cordatum; 4. Acalypha indica; 5. Acalypha scandens; 6. Blumeodendron tokbrai; 7. Coelebogyne ilicifolia; 8. Mareya brevipes; 9. Discocleidion rufescens; 10. Kunstlerodendron sublanceolatum.

and K. Hoffmann (1931) thought it better to raise the section to genus level which they place in the Bernardiiformes close to *Bernardia* and *Mareya* with their allies. Hurusawa (1954) at last reduced the genus again to a section, this time of *Alchornea*.

Discocleidion rufescens (Franch.) Pax et K. Hoffmann D'Alleizette China Mai. 1908 [L] Pl. XVIII, 9

Discocleidion type.  $P = 24 \mu E = 27 \mu P:E = 0.89.$ m:e = 0.7. P.A.I. = 0.1-0.15.

## Mareya type

Tricolporate; oblate spheroidal to prolate spheroidal. In polar view pollen grains convex triangular.
Colpus transversalis elongated; no costae.
Colpi narrow; no costae colpi present.
Tectate; psilate.

The pollen grains are small. The largest axis is smaller than 30  $\mu$ . The *Mareya* type is distinguished from the *Mallotus* type by the absence of costae.

#### Taxonomic discussion

In Pax and K. Hoffmann's system (1914, 1931) genera of this type belong to the Bernardiiformes. By the following characters the genera differ from those in the *Mallotus* type and *Alchomea* type:

1. In the female flowers a disc is present.

2. The male flowers have a receptaculum often set with glands. A disc and an ovarium rudiment are wanting. (Mareyopsis is anomalous by the presence of extra-staminal disc glands and the absence of glands in the receptaculum.)

3. Anthers often have appendiculate connectives.

4. Plants usually grow in the Old World. (Alchorneopsis in the New World.)

The American genus Alchorneopsis differs from the above characters by having an ovarium rudiment and the presence of hairs on the receptaculum instead of glands.

Mareya bervipes PAX Zenker 1794 [L] Pl. XVIII, 8

Crotonogynopsis usambarica PAX
Eggeling 3826 [K]
Chondrostylis bancana BOERL.
Cult. Hort. Bogor. II. I 8. [L]
Kunstlerodendron sublanceolatum RIDL.
Sinclair 39882 [L] Pl. XVIII, 10

Mareyopsis longifolia (PAX) PAX et K. HOFFMANN Zenker 4228 [K] Neopalissya castaneifolia (BAILL.) PAX Pervillé 387 [P] Mareya type; oblate spheroidal.  $P=21~\mu~E=23~\mu~P:E=0.91.$  m:e<0.5.~P.A.I.=0.3. Colpus membrane granulate. Mareya type. Too young to give reliable measures.

Mareya type; spheroidal.  $P = E = 25 \mu$ . P.A.I. = 0,3. Mareya type; oblate spheroidal.  $P = 20.5 \mu$  E = 21  $\mu$  P:E = 0,98. P.A.I. = 0,3.

Mareya type; oblate spheroidal. P = 18  $\mu$  E = 19  $\mu$  P:E = 0,96. P.A I. = 0,2. Mareya type; prolate spheroidal. P = 22  $\mu$  E = 21  $\mu$  P:E = 1,05. P.A.I. = 0,25-0,3.

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Necepsia afzelii Prain
Leeuwenberg 2211 [U]

Amyrea humberti Léandri
D'Alleizette Dec. 1905 [L]

Alchorneopsis floribunda (Benth.) Muell.
For. Dept. 2889 [U]

Alchorneopsis trimera Lanj.
Lanjouw 1926; 49 [U]

Mareya type; prolate spheroidal.
P = 24 \mu E = 21,5 \mu P:E = 1,13.
P.A.I. = 0,3.

Mareya type; oblate spheroidal.
P = 18 \mu E = 19 \mu P:E = 0,95.
P.A.I. = 0,2.

Mareya type; oblate spheroidal.
P = 18 \mu E = 19 \mu P:E = 0,95.
P.A.I. = 0,2.
```

## Ricinus type

Tricolporate; shape differs from suboblate to subprolate. Colpus transversalis narrow; no costae. Colpi very narrow; no costae colpi present. Tectate; psilate. Columellae very short, nearly visible in some species.

The pollen grains resemble the *Pera* type, but the *Ricinus* type misses costae as in the *Mareya* type. It differs from the latter type by narrow colpi and nearly visible columellae.

### Taxonomic discussion

The genera in this type fall apart in two groups. The first group includes *Ricinus*, *Homonoia* and *Lasiococca*. These genera are remarkable for the large number of anthers crowded on repeatedly branched filaments.

The second group comprises Koilodepas only, which genus occupies an isolated position in the family (AIRY-SHAW, 1960). CROIZAT (1942b) as well as AIRY-SHAW reduced Nephrostylus GAGNEPAIN and Calpigyne Blume to Koilodepas. According to CROIZAT, Calpigyne hainanense and Nephrostylus poilanei are synonyms. In the opinion of AIRY-SHAW, Nephrostylus poilanei belongs to the species Koilodepas longifolium. Pollen grains of both species are practically identical and show great structural resemblance with Koilodepas bantamense.

```
Ricinus type; spheroidal. P = 32 \mu E = 32 \mu P:E ca. 1. m: e < 0,5. P.A.I. = 0,2.
Ricinus communis LINN.
v. Heerdt 531 [U]
Versteeg 243 [U] Pl. XIX, 3
Lasiococca symphylliifolia (Kurz)
                                                         Ricinus type; spheroidal.

P = E = 23 \mu. P.A.I. = 0,45.
    . D. Ноок.
   J. D. HOOK.
Hort. Bogor. Cult. 6-4-1887 [L]
Homonoia javensis (Blume) Muell. Arg.
                                                         Ricinus type; prolate spheroidal.
                                                            P = 22.5 \mu E = 21 \mu P:E = 1.08.
P.A.I. = 0.5. Costae transversales?
   Hort. Bogor. IX A. 60 [U] Pl. XIX, 2
                                                         Ricinus type; oblate spheroidal-suboblate.

P = 25 \mu E = 29 \mu P:E = 0.88.
Homonoia riparia LOUR.
Helfer 4746 [U]
Koilodepas bantamense HASSK.
                                                         Ricinus type; spheroidal-suboblate.
                                                            P = 20 \mu E = 24 \mu P:E = 0.84.
m:e < 0.5. P.A.I. = 0.3.
   Hort. Bogor. IX C. 36 [U] Pl. XIX, 4
                                                         Ricinus type; subprolate.

P = 21 \mu E = 17,5 \mu P:E = 1,18.
Koilodepas hainanense (MERR.) CROIZAT
   [= Calpigyne hainanense MERR.]
S. K. Lau 3542 [P]
                                                            P.A.I. = 0.3.
                                                            P = 20.5 \mu E = 17.5 \mu P:E = 1.17.
P.A.I. = 0.3.
   [? = Nephrostylus poilanei GAGNEP.
   D'Alleizette, Annam 11-4-1907 [L]
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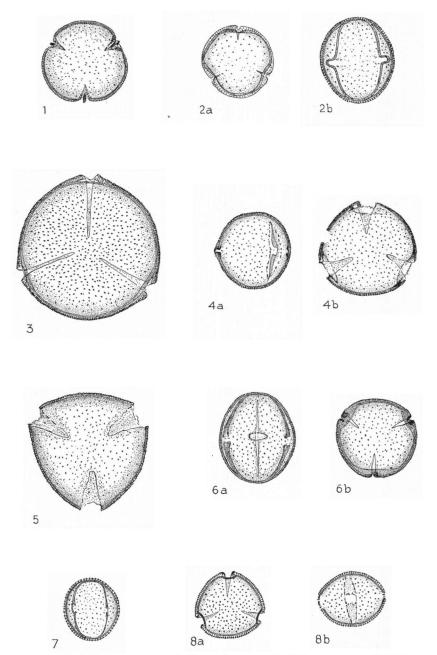


PLATE XIX. 1. Leidesia procumbens; 2. Homonoia javensis; 3. Ricinus communis; 4. Koilodepas bantamense; 5. Chaetocarpus castanocarpus; 6. Pera bicolor; 7. Seidelia triandra; 8. Microdesmis casearifolia.

## Pera type

Tricolporate; prolate spheroidal. Colpus transversalis small; costae. Colpi narrow; costae colpi narrow but distinct. Tectate; psilate. Columellae very short.

The *Pera* type has the same very narrow colpi and nearly visible columellae as the *Ricinus* type, but there are distinct costae colpi and costae transversales present.

 Pera bicolor (KLOTZSCH)
 MUELL. ARG.
 Pera type.

 Ule 8409 [U] Pl. XIX, 6
  $P = 25.5 \mu$  E = 24 μ P:E = 1,07. m:e < 0,5. P.A.I. = 0,3.</td>

 Pera glabrata (SCHOTT)
 BAILL. Pera type.

 Reitz 4548 [U]
  $P = 19 \mu$  E = 18 μ P:E = 1,06. P.A.I. = 0,5.

### Seidelia type

Tricolporate; prolate spheroidal. In polar view the pollen grains distinctly three-lobed.

Colpus transversalis small; costae. Colpi narrow; no costae colpi. Tectate; psilate. Columellae short.

The pollen grains of this type are distinctly three-lobed and have costae transversales.

Seidelia triandra (E. Meyer) Pax Seidelia type. Mac Gregor Mus. 1254 [K] Pl. XIX, 7  $P = 17,5 \mu E = 16,5 \mu P:E = 1,06.$  m:e = 0,6. P.A.I. = 0,25.

## Leidesia type

Tricolporate; oblate spheroidal. In polar view the pollen grains distinctly three-lobed.

Colpus transversalis narrow and long; no costae. Colpi narrow; no costae colpi. Narrow operculum.

Tectate; psilate. Columellae short.

The *Leidesia* type resembles the *Seidelia* type in its polar view and exine structure. It differs, however, by the absence of costae and the addition of a narrow operculum.

#### Taxonomic discussion

Leidesia and Seidelia are Mercurialis-like plants which BENTHAM (1880) as well as PAX and K. HOFFMANN (1914, 1931) enumerated close together with Mercurialis. This relationship, however, is not confirmed by pollen-morphological results.

Leidesia procumbens (Linn.) Prain Splitgerber [L] Pl. XIX, 1 Leidesia type.  $P = 19 \mu E = 20,5 \mu P:E = 0,92.$  m:e < 0,5. P.A.I. = 0,3.

# Microdesmis type

Tricolporate; spheroidal to oblate speroidal. In polar view pollen grains convex triangular.

Colpus transversalis isodiametric; m:e = 1; ora large. Colpi narrow; costae colpi broad and indistinct or absent.

Tectate; psilate. Columellae short.

The *Microdesmis* type is characterised by the isodiametric colpus transversalis and the large ora.

## Taxonomic discussion

Bentham (1880) discussed the genera of this type in two tribes, Galearieae and Gelonieae. Microdesmis, Galearia and Pogonophora occurring in the Galearieae have characters both of the Phyllanthoideae and Crotonoideae. Pollen grains of these genera are, however, not at all comparable with those in the Phyllanthoideae.

Pierre's description being unknown to Pax (1899), he described Centroplacus glaucinus Pierre in Microdesmis as Microdesmis paniculata. Pax, however, did not have female flowers at his disposal. Pierre (1899) and Gilo (1908), on the other hand, studied female material and found 2 ovules per locule, for which reason the plant had to be placed in the subfamily of the Phyllanthoideae. The structure of the male flowers, however, resembles that of Microdesmis to such an extent, that it is by no means strange that Pax reckoned this species to Microdesmis. Since it is evident that the pollen grains of Centroplacus cannot be distinguished from those of Microdesmis, the correctness of placing the genus in the subtribe Antidesminae (Pax 1931) seems questionable. Two conclusions are possible:

- 1. Centroplacus forms a link between the sub-family Phyllanthoideae and Crotonoideae. On one side we find the characters that suggest a close relationship to Microdesmis, while the Phyllanthoideae-character of the two ovula per locule in the ovary stands on the other side.
- 2. Centroplacus has been described on the basis of material belonging to two species. The plant is dioecious, so that the possibility of an erroneous combination of plants should not be overlooked.

A renewed accurate examination of the known material, will possibly remove the doubt.

## Microdesmis subtype

Pollen grains small. Smaller than 20  $\mu$ . Costae colpi absent.

Microdesmis casearifolia Planch. Thorenaar T. 50 [U] Pl. XIX, 8

Microdesmis zenkeri PAX de Wit 4 [WAG]

Centroplacus glaucinus Pierre Tester 1922 [K] Zenker 1761 [K] Galearia dognaiensis Pierre ex GAGNEP. Clemens 3528 [U]

Galearia filiformis (BLUME) PAX Blume, Java [U] Microdesmis subtype; oblate spheroidal.  $P=15.5~\mu~E=17.5~\mu~P:E=0.89.$  P.A.I.=0.3. Microdesmis subtype.  $P=18~\mu~E=19~\mu~P:E=0.95.$  Microdesmis subtype.  $P=14~\mu~E=15.5~\mu~P:E=0.90.$  P.A.I.=0.3. Microdesmis subtype; suboblate.  $P=14.5~\mu~E=17.5~\mu~P:E=0.83.$  P.A.I.=0.35. Microdesmis subtype; suboblate.  $P=14.5~\mu~E=17.5~\mu~P:E=0.83.$  P.A.I.=0.35. Microdesmis subtype

Microdesmis subtype.  $P = 16.5 \mu E = 19 \mu P:E = 0.89.$  P.A.I. = 0.3.

### Chaetocarpus subtype

Pollen grains large. Larger than 30  $\mu$ . Costae colpi broad but indistinct.

Chaetocarpus castanocarpus (ROXB.) Chaetocarpus subtype; spheroidal.  $P = E = 29 \,\mu$  m:e ca. 1. THWAITES P.A.I. = 0.3. Elmer, Borneo 21064 [U] Pl. XIX, 5 Chaetocarpus schomburgkianus (O. Kuntze) Chaetocarpus subtype. Pax et K. Hoffm. For. Dept. 2519 [U] Trigonopleura malayana J. D. HOOK. Chaetocarpus subtype; oblate spheroidal.  $P=35~\mu~E=38~\mu~P:E=0,92.$  P.A.I.=0,2.Bosch Proefst. 83 E. 1P. 1031 [L] Pl. XX, 1 Pogonophora schomburgkiana MIERS B. W. 6015 [U] Chaetocarpus subtype; oblate spheroidal.  $P = 42 \mu E = 45 \mu P : E = 0.93$ . Krukoff 8645 [U] P.A.I. = 0,4-0,45. Tectum perforatum.

#### Cheilosa type

Tricolporate; suboblate to oblate spheroidal. In polar view pollen grains convex triangular.

Colpus transversalis; costae.

Colpi narrow; indistinct costae colpi.

Tectate; echinate or micro-echinate. Columellae short.

The Cheilosa type possesses the most important characters of the Mallotus configuration and is therefore placed in this configuration. The type differs, however, distinctly by the spines on the tectum. In Cheilosa these spines are larger than 1  $\mu$  (echinate); in Neoscortechinia smaller than 1  $\mu$ . According to the nomenclature of FAEGRI and IVERSEN the pollen grains of Neoscortechinia are scabrate. The author would like to call the small spines microechinate according to ERDTMAN (1952). Echinate pollen grains are rarely found in the subfamily of the Crotonoideae, only in the Croton configuration.

### Taxonomic discussion

Bentham (1880) discussed *Cheilosa* in the *Acalypheae*, although the male calyx is divided into slightly imbricate lobes. In the system of Pax and K. Hoffmann (1931) the genus is found in the tribe *Gelonieae*, to which also *Neoscortechinia* belongs. The pollen grains of both genera are undoubtedly related. Unfortunately this is not confirmed by other characters.

Cheilosa montana Blume
Korthals, Java [L] Pl. XX, 9

Korthals, Java [L] Pl. XX, 9

Neoscortechinia arborea (Elmer) Pax et
K. Hoffm.
Hort. Bogor. IX A. 6a. [U] Pl. XX, 8

Cheilosa type; suboblate.

P = 22,5 \mu E = 29 \mu P:E = 0.78.

Echinate; echinate 1-2 \mu.

Cheilosa type; suboblate.

P = 21,5 \mu E = 29,5 \mu P:E = 0.78.

P = 21 \mu E = 22,5 \mu P:E = 0.93.

P.A.I. = 0,3. Scabrate (microechinate).

#### CLAOXYLON CONFIGURATION

Tricolporate or stephanocolporate (4, rarely 5). Colpi narrow.
Tectate; capita of the columellae distinct.
Pollen grains not reticulate.

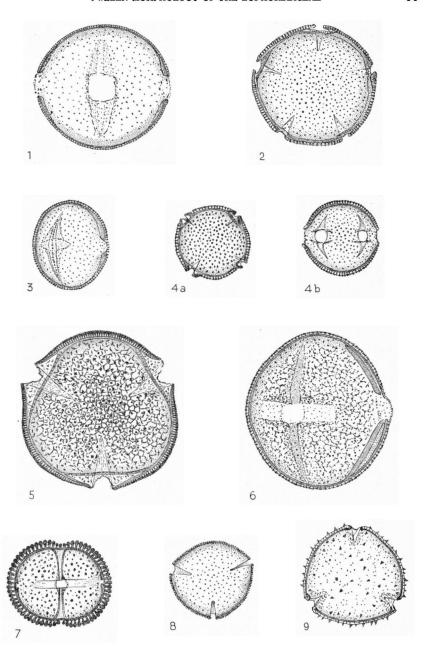


PLATE XX. 1. Trigonopleura malayana; 2. Athroandra mannii; 3. Mercurialis annua; 4. Claoxylon cuneatum; 5. Cladogynos orientalis; 6. Cephalocrotonopsis socotrana; 7. Amperea xiphoclada; 8. Neoscortechinia arborea; 9. Cheilosa montana.

The Claoxylon configuration is related to the Mallotus configuration. The capita of the columellae, however, are distinctly visible.

## Claoxylon type

Tricolporate or stephanocolporate; oblate spheroidal-prolate spheroidal. Colpus transversalis small; costae present but sometimes indistinct. Colpi narrow; costae colpi absent or indistinct.

Tectate; psilate or scabrate. Columellae coarse, capita distinct.

This type is in some characters comparable with types of the *Mallotus* configuration. The small spheroidal pollen grains resemble, in their shape, those of the *Mareya* type (p. 85). As in the *Mallotus* type (p. 79) more than three composite apertures can be present. Placing the type in the *Bernardia* configuration (p. 77) is out of question as a reticulum is always absent.

The Claoxylon type is distinguished by the distinct capita of the columellae, which form a coarse and inordinate pattern.

#### Taxonomic discussion

Genera belonging to this type are all readily recognised by the shape and attachment of the anthers. Other corresponding characters are:

- 1. A disc present in the female flowers.
- 2. In the male flowers an ovarium rudiment is always absent. Usually an extrastaminal disc is also absent, but glands are inserted on the receptaculum. In Discoclaoxylon receptaculum glands fails, but in this genus an extrastaminal disc is present.
- 3. Plants are usually dioecious and grow only in the Old World.

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Claoxylon type; spheroidal.

P = E = 22 \mu. m:e < 0,5.
Erythrococca stolziana PAX et K. HOFFM.
   Stolz 1556 [U]
                                                         P.A.I. > 0.5.
                                                      Claoxylon type; oblate spheroidal-sub-
oblate. 5 (-4) colporate.
Athroandra mannii (J. D. HOOK.) PAX et
   K. Hoffmann
                                                         P = 30 \mu \dot{E} = 33.5 \mu P : E = 0.89.
   [= Erythrococca mannii (J. D. HOOK.)
   PRAIN]
Mann 260 [U] Pl. XX, 2
Athroandra africana (BAILL.) PAX et
                                                      Claoxy!on type; prolate spheroidal. P = 22.5 \mu E = 21 \mu P:E = 1.07.
   K. Hoffmann
   [= Erythrococca africana (BAILL.) PRAIN]
Zenker 453 [U]
Claoxylon cuneatum J. J. SMITH
Versteeg 1752 [U] Pl. XX, 4
Claoxylon tumidum J. J. SMITH
Versteeg 1708 [U]
                                                      Claoxylon type; spheroidal.
                                                         P = E = 21 \ \mu.
                                                      Claoxylon type; spheroidal.
                                                         3- (4) colporate.

P = E = 19 \mu. P.A.I. = 0,4.
                                                         Psilate to scabrate.
Micrococca mercurialis (LINN.) BENTH.
                                                      Claoxylon type; prolate spheroidal.
                                                         3- (4) colporate.

P = 27 \mu E = 25 \mu P:E = 1,08.

P.A.I. = 0,3.
   D'Alleizette Mad. 1906 [L]
                                                      Claoxylon type; spheroidal.

P = E = 21 \mu. P.A.I. = 0,3.
Discoclaoxylon hexandrum (MUELL. ARG.)
   Pax et K. Hoffm.
   Zenker 35 [U]
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### Mercurialis type

Tricolporate; subprolate to prolate.

Colpus transversalis small; costae as long as the costae colpi.

Colpi narrow; costae colpi. Operculum present but very narrow, consisting of only one row of columellae. Tectate; psilate. Columellae distinct.

There is a certain relation between the Mercurialis type and the Claoxylon type. The two types have distinct capita; both are not reticulate and possess small colpi transversales. The Mercurialis type differs by the presence of an operculum, which is narrow and indistinct, and by the distinctly prolate shape of the pollen grains.

#### Taxonomic discussion

Shape and attachement of the anthers show much similarity with the genera of the Claoxylon group. For that reason Bentham (1880) placed Mercurialis in the immediate vicinity of Claoxylon, Erythrococca, etc. Although the Mercurialis type differs somewhat from the Cloxylon type, there is enough affinity for the author to share the opinion of BENTHAM.

Mercurialis annua LINN.

Fresh material, Z.-Limb. Pl. XX, 3

Mercurialis reverchonii Rouy Faure 18-6-1931 [U] Mercurialis tomentosa LINN. Leeuwenberg 1317 [U]

Mercurialis type; subprolate.  $P = 25.5 \mu E = 22.5 \mu P:E = 1.16.$  m:e < 0.5. P.A.I. = 0.4. Operculumindistinct, consisting of some scattered pila on the endexine (membrana

granulata).

Mercurialis type.

 $P = 28.5 \mu$   $E = 22 \mu$  P:E = 1.30.

Mercurialis type.

 $P = 40 \mu E = 28.8 \mu P:E = 1.40.$ 

#### CLADOGYNOS CONFIGURATION

Tricolporate.

Colpi narrow; costae colpi.

Tectate; columellae distinct. Exine thick.

The Cladogynos configuration includes those pollen grains with distinct columellae that bear always a tectum or tectum perforatum. These columellae can be arranged in a reticulum but more frequently they do not form a regular pattern. The pollen grains are usually large; the largest axis is at least 25  $\mu$  and often up to 50  $\mu$ .

# Cladogynos type

Tricolporate; spheroidal prolate (Epiprinus oblate spheroidal).

Colpus transversalis; costae.

Colpi narrow; costae colpi.

Tectate or tectum perforatum. Columellae distinct.

Exine thick. Some species intra-reticulate.

The most striking character of this type is the distinct, sometimes even tall, columellae. The pollen grains are large and generally they have a P: E larger than 1.

#### Taxonomic discussion

By Pax and K. Hoffmann (1914, 1931) most genera of this type are placed in the series *Cladogyniformes* of their subtribe Mercurialinae, to which also *Mallotus* and *Alchornea* belong. The group can be distinguished as follows:

- 1. An ovarium rudiment present in the male flowers while a disc is absent. The few stamens are not inserted on a receptacle.
- 2. Male flowers usually in a capitule.
- 3. Anthers pendulous.
- 4. Stellate hairs always present.
- 5. Plants are monoecious and grow in the Old World.

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Cladogynos orientalis ZIPP ex SPANOGHE
Pierre 6213 [L] Pl. XX, 5

Cephalocroton cordofanus Hochst.
D'Alleizette Nubië, Aug. 1909 [L]

Cephalocrotonopsis socotrana (BALF. F.) PAX
Schweinfurth 594 [K] Pl. XX, 6

Symphyllia siletiana BAILL.
Hort. Bog. Cult. [L]

Adenochlaena leucocephala BAILL.
Hildebrandt 3258b [L]
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Epiprinus poilanei GAGNEP. Clemens 3058 [U] Cladogynos type; prolate spheroidal.  $P=42~\mu~E=40~\mu~P:E=1,05.$  Colpus transversalis narrow. Tectum perforatum. P.A.I. = 0,3. Intra-reticulate; lumina 1-2  $\mu$ . Cladogynos type; subprolate.  $P=49~\mu~E=42~\mu~P:E=1,17.$  Colpus transversalis broad; m:e>0,5. P.A.I. = 0,3. Intra-reticulate. Cladogynos type; prolate spheroidal.  $P=46~\mu~E=42,5~\mu~P:E=1,09.$  P.A.I. = 0,25. Intra-reticulate. Cladogynos type; prolate spheroidal.  $P=27~\mu~E=25~\mu~P:E=1,07.$  P.A.I. = 0,25-0,3. Not reticulate. Cladogynos type; prolate spheroidal.  $P=53~\mu~E=48~\mu~P:E=1,10.$  m:e=0,4. P.A.I. = 0,3. Not reticulate. Cladogynos type; oblate spheroidal.  $P=32~\mu~E=35~\mu~P:E=0,91.$  Results of the color of

## Amperea type

Dicolporate or tricolporate. Shape differs with the number of colpi from oblate spheroidal (dicolporate) to prolate spheroidal (tricolporate). Colpus transversalis narrow; costae. Colpi narrow and long; costae colpi. P.A.I. small to 0. Tectate; psilate. Exine thick. Intrareticulate or not reticulate.

The Amperea-type shows much similarity with the Cladogynos type. Especially the thick exine and the tall columellae are striking. The P.A.I., however, is much smaller. Some pollen grains are even syncolpate.

#### Taxonomic discussion

Amperea belongs to those Australian genera, that were kept separate on account of their narrow cotyles (Stenolobae). Should this group as such be abolished, then, according to the statement of Pax (1924), this genus has to be placed in the Mercurialinae. This group also comprises the genera of the Cladogynos type.

Amperea xiphoclada (Sieb. et Spreng.)
Druce
Constable 26475 [U] Pl. XX, 7

Amperea spartioides Brongn. Tasmania [U] Amperea type. 2-colporate: P:E = 0,90. 3-colporate: P:E = 1,02. P = 27-29  $\mu$  E = 28-30  $\mu$ . P.A.I. = 0,15-0. m:e < 0,5. Not reticulate. Amperea type; spheroidal. P = E = 26  $\mu$ . Intra-reticulate.

## Pseudocroton type

Tricolporate; prolate spheroidal to subprolate. Colpus transversalis; costae? Colpi narrow; costae colpi? Tectate; psilate. Not reticulate.

The pollen grains of the sheet examined were not mature, so it is difficult to put them in the proper type. There is some resemblance with the *Speranskia* type (p. 75). A reticulum, however, is absent. As distinct columellae are present, it seems best to place this type in the *Cladogynos* configuration.

#### Taxonomic discussion

BENTHAM (1880) as well as Pax et K. Hoffmann (1931) discuss *Pseudocroton* in the *Chrozophoreae*. They accept relationship with *Argythamnia* and its allies. The pollen grains have some affinity with the *Speranskia* type. Although the genera of this type belong to the *Chrozophoreae*, they are only found in the Old World, while *Pseudocroton* grows in Central America. Affinities with genera of the *Cladogynos* configuration are few.

Pseudocroton tinctorius Muell. Arg.
O. Kuntze 1808 [K]

Pseudocroton type.

Too young to give reliable measures.

#### HIPPOMANE CONFIGURATION

Tricolporate; pollen grains in polar view circular or three-lobed. Never convex triangular or triangular. Colpus transversalis frequently short; sometimes narrow elongated. Colpi narrow; usually long. A margo is nearly always present. Intectate or more often tectate. Columellae distinct, frequently large.

It is not possible to define this configuration with one or two exceptionless characters. The pollen grains are, however, easily recognised by a combination of several typical characters. The most important ones are: three-lobed polar view; margo; distinct, sometimes large, columellae and short colpi transversales. If one of these striking characters is absent the combination of the others still places the pollen grains immediately in this configuration.

#### Taxonomic discussion

The genera of this configuration can be divided into two groups. The first group comprises genera of the tribe *Hippomaneae* of Pax and K. Hoffmann (1912b, 1931). In 1880, Bentham stated of his subtribe

Hippomaneae: "This is one of the most natural subtribes of Crotoneae". Indeed there is not any doubt of a close relationship between the genera.

Pachystroma is added to the first group. Evidently on technical grounds Bentham added Pachystroma to the group of the Adrianeae with Manihot, Adriana and Cephalocroton, three genera now placed in three different groups of plants. Pax and K. Hoffmann (1919d, 1931) separated the monotypic genus into the tribe Pachystromateae, although they were aware of some affinity with the Hippomaneae. The well-developed calyx of the male flower, which is never present in the Hippomaneae, led them to the decision to separate the genus. There are, however, more characters in favour of a fusion than of a separation. Pollen morphology is one of the data in favour of fusing.

The second group includes all the genera of the Euphorbieae with the addition of Hura. It is remarkable that the typical Euphorbieae, so well characterised by their cyathium, possess pollen grains which can hardly separated from those of the Hippomaneae. Pax (1924) when discussing the phylogenetic relations of the Euphorbiaceae, pointed already to this relationship. He noticed, e.g., the unbranched laticiferous vessels which both taxa possess. Pollen morphology is another

character to support the opinion of Pax.

## Hippomane type

Tricolporate; shape differs from subprolate to suboblate. Pollen grains in polar view circular or three-lobed.

Colpus transversalis small (rarely elongated); costae.

Colpi narrow and long; costae colpi. Usually margo present. P.A.I. is small (smaller 0,3) except in some species of Mabea.

Tectate; psilate Columellae sometimes tall.

The Hippomane type differs from the Euphorbia hirta type by its narrower margo. Usually the margo is narrower than the colpus transversalis. In other respects both types can hardly be separated.

Pollen grains of the *Omalanthus nutans* subtype can be separated from the *Hippomane* subtype by the deeply three-lobed pollen grains.

Some species in the *Hippomane* type have an intra-reticulum. They do not belong altogether to the *Dichostema* type (p. 100) as the upper part of the columellae is fused in all directions, thus forming a tectum.

Hippomane subtype

Pollen grains in polar view circular or slightly three-lobed.

Hippomane mancinella LINN.
Buysman 1856 [U] Pl. XXI, 3

Omalanthus populneus (Geisel) Pax Docters v. Leeuwen 1716 [U]

Mabea piriri AUBL. B. W. 6296 [U]

Mabea taquari AUBL. Pulle 67 [U]

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Hippomane subtype; prolate spheroidal. P = 32,5 \mu E = 30,5 \mu P:E = 1,06. m:e < 0,5. P.A.I. = 0,25. Hippomane subtype. P = 40 \mu E = 34 \mu P:E = 1,17. P.A.I. = 0,15-0,20. Hippomane subtype. P = 50 \mu E = 44 \mu P:E = 1,14. P.A.I. = 0,20-0,25. Not reticulate. Hippomane subtype. P = 43,5 \mu E = 41 \mu P:E = 1,05. P.A.I. = 0,4. Intra-reticulate.
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Mabea caudata PAX et K. HOFFM. v. Emden 18-9-1931 [U]

Mabea fistulifera MART. Y. Mexia 4473 [U]

Actinostemon lanceolatus SALDANHA Y. Mexia 4987 [U]

Gymnanthes lucida SWARTZ Jamaica, Hohsle [U]

Glyphostylus laoticus GAGNEP. Lakshnaka 1352 [K]

Spirostachys venenifera (PAX) PAX [= Excoecaria venenifera PAX] Graham 1464 [K] Pl. XXIII, 3 Excoecaria bicolor HASSK. Hort. Bogor. IX C. 77 [U] Pl. XXII, 4

Stillingia gymnogyna PAX et K. HOFFM. Wiggins 1351 [U]

Sapium longifolium (MUELL. ARG.) HUBER Pedersen 72 [U]

Colliguaya integerrima GILL. et HOOK. Donat 167 [U]

Hippomane subtype.  $P = E = 70^{\circ} \mu$ . P.A.I. = 0,4-0,3.Not reticulate.

Hippomane subtype.  $P = E = 70 \mu$ . P.A.I. = 0,3. Indistinct intra-reticulate.

Tectum perforatum.

Hippomane subtype; oblate spheroidal.  $P=33.5~\mu~E=35~\mu~P:E=0.94.$  P.A.I. = 0.25. Margo absent. Costae indistinct.

Hippomane subtype; oblate spheroidal.  $P=25~\mu~E=26~\mu~P:E=0.96.$  P.A.I. = 0.25. Margo absent. Columellae short.

Hippomane subtype.  $P=36,5~\mu~E=34,5~\mu~P:E=1,06.$  P.A.I. = 0,25. Colpus transversalis elongated. Columellae tall.

Hippomane subtype.  $P = 22 \mu E = 23 \mu$ P:E = 0.96.P.A.I. = 0.25. Margo absent.

Hippomane subtype.  $P = 38 \mu \quad E = 36 \mu$ P:E = 1,05.P.A.I. = 0.25.

Hippomane subtype.  $\hat{P} = 35.5 \ \mu \hat{E} = 33 \ \mu P : E = 1.08.$ P.A.I. = 0.2.

Hippomane subtype. P = 55  $\mu$  E = 43  $\mu$  P:E = 1,26. P.A.I. = 0,25. Colpus transversalis elongated. Costae equatoriales or almost equatorial.

Hippomane subtype.  $P = 35.5 \mu$   $E = 34 \mu$  P:E = 1.04. P.A.I. = 0.2. Margo indistinct; narrow.

### Omalanthus nutans subtype

Pollen grains in polar view distinctly three-lobed.

Omalanthus nutans (FORST.) PAX Yuncker 15356 [U] Pl. XXI, 8

Mabea indorum Sp. Moore Krukoff 1084 [U]

Actinostemon concolor (SPRENG.) MUELL.

Bornmueller 536 [U] Pl. XXII, 3 Gymnanthes jamaicensis (BRITTEN) URBAN Harris 10936 [U] Sebastiania chamaelea (LINN.) MUELL. ARG. Hohenacker 830 [U]

Sebastiania corniculata (VAHL) MUELL. ARG. Omalanthus nutans subtype. Froes 11761 [U]  $P = E = 40 \mu$ . P.A.I. = 0,15-0,2. Macguire and Stahel 22758 [U] Sebastiania scottiana MUELL. ARG. Bertoni 1714 [U]

Omalanthus nutans subtype.  $P = 35 \mu E = 31,5 \mu P:E = 1,11.$ P.A.I. = 0,15.

Omalanthus nutans subtype.  $P = 45 \mu E = 36,5 \mu P:E = 1,22.$ P.A.I. = 0,2-0,25.

Omalanthus nutans subtype.  $P = 30 \mu E = 34 \mu$ P:E = 0.88.Columellae short.

Omalanthus nutans subtype.  $P = E = 22 \mu$ . Margo absent.

Omalanthus nutans subtype.  $P = 33 \ \mu \ E = 29 \ \mu$ P:E = 1,14.P.A.I. = 0,2.

Omalanthus nutans subtype.  $P = E = 27 \mu$ . P.A.I. = 0,2.

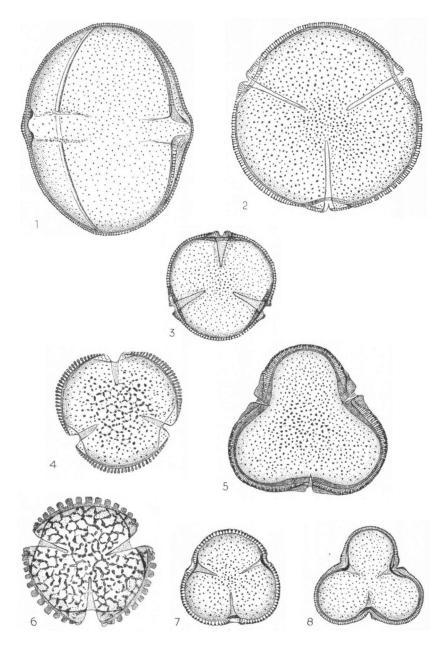


PLATE XXI. 1. Stenadenium spinescens; 2. Elaeophorbia drupifera; 3. Hippomane mancinella; 4. Dichostema glaucescens; 5. Pachystroma longifolium; 6. Senefeldera macrophylla; 7. Tetraplandra gibbosa; 8. Omalanthus nutans.

Omalanthus nutans subtype. Duvigneaudia inopinata (PRAIN) LÉONARD  $P = 23 \mu E = 20^{\circ} \mu$ P.A.I. = 0,2. Zenker 217 [U] P:E = 1,16.Omalanthus nutans subtype. Maprounea brasiliensis ST.-HILL. Williamson-Assis 8047 [U]  $P = 33.5 \mu E = 30.5 \mu P:E = 1.11.$ P.A.I. = 0,2. Maprounea guyanensis AUBL. B. W. 5393 [U] Omalanthus nutans subtype.  $P = 24 \mu E = 20 \mu P:E = 1,20.$ P.A.I. = 0,2.Excoecaria philippinensis MERRILL Elmer, Palawan 13124 [U] Sapium ellipticum (HOCHST.) PAX Croockewit 728 [WAG] Omalanthus nutans subtype.  $P = 32 \mu E = 32 \mu$ . P.A.I. = 0,2-0,25. Omalanthus nutans subtype.  $P = E = 30 \mu$ . P.A.I. = 0,2.  $P = 40 \mu$  E = 34  $\mu$  P:E = 1,18. Stolz 621 [U] Sapium sebiferum (LINN.) ROXB. Omalanthus nutans subtype. Lilnee, Japan [U]  $P=46~\mu$ . Sapium aubletianum (MUELL. ARG.) HUBER Omalanthus nutans subtype.  $P=46~\mu.$ B. W. 2141 [U]  $P=46~\mu.$ Sapium montanum LANJ. Omalanthus nutans subtype. B. W. 5889 [U]  $P = 52 \mu.$ Sapium klotzschianum (MUELL. ARG.) Omalanthus nutans subtype.  $P = 54 \mu$ . Stahel, Oct. 1944 [U] Grimmeodendron eglandulosum (A. RIGH.) Omalanthus nutans subtype.  $P = 28 \mu E = 26' \mu P:E = 1,08.$ P.A.I. = 0,2. Colpus transversalis Urban Curtiss 190 [K] narrow elongated. Omalanthus nutans subtype (?).  $P = 27 \mu E = 25.5 \mu P:E = 1.06.$ Bonania cubana A. RICH. Howard 5774 [U] P.A.I. = 0,2.Omalanthus nutans subtype. Bonania domingensis (URB.) URB. Fuertes 813 [U]  $P = 22 \mu E = 22 \mu P.A.I. = 0,25.$ Columellae small. Omalanthus nutans subtype.  $P = 34.5 \mu E = 33.5 \mu P:E = 1.03.$  P.A.I. = 0.15. Intra-reticulate. Adenopeltis colliguaya BERT. ex Juss. Andreas 834 [U] Omalanthus nutans subtype. Dalembertia populifolia BAILL. Hinton 4353 [K]  $P = 41 \ \mu \ E = 36 \ \mu$ P:E = 1,12.P.A.I. = 0,15-0,2. Intra-reticulate. Omalanthus nutans subtype. Tetraplandra gibbosa PAX et K. HOFFM.  $P = 30 \mu E = 27 \mu P = 1,11.$ Glaziou 5613 [K] Pl. XXI, 7 P.A.I. = 0.15.Omalanthus nutans subtype. Ophthalmoblapton macrophyllum F. Allem. Glaziou 17752 [K]  $P = 42 \mu E = 39 \mu P:E = 1,08.$ P.A.I. = 0,2.Algernonia brasiliensis BAILL. Omalanthus nutans subtype. H. F. Hance 1887 [K] Young.

## Pachystroma type

Tricolporate; spheroidal. Pollen grains slightly three-lobed. Colpus transversalis broad elliptic; costae thick. Colpi narrow and short; costae colpi. No margo present. Tectate; psilate.

Pollen grains of this type are strongly related to those of the *Hippomane* type. The colpus transversalis, however, is broadly elliptic and the P.A.I. is much larger.

Pachystroma longifolium (Nees)
I. M. Johnston
Krug and Urban 1367 [L] Pl. XXI, 5

Pachystroma type.
P = 41  $\mu$  E = 41  $\mu$  P: E ca. 1.
Length of colpi differs; mostly short, sometimes long. Costae thick.

### Euphorbia hirta type

Tricolporate; subprolate to oblate spheroidal. Pollen grains in polar view three-lobed.

Colpus transversalis; costae.

Colpi narrow; margo broad. Sometimes an operculum is present.

Tectate; psilate. Columellae distinct.

The pollen grains belonging to this type can hardly be separated from the *Hippomane* type. The margo is broader; mostly as broad as the colpus transversalis. The colpi are narrow. If in the future more *Euphorbiaceae* species are examined pollen-morphologically, it will perhaps be necessary to combine the two types.

Euphorbia hirta subtype

Pollen grains without an operculum.

Euphorbia hirta Linn. Soeprata 20 H [U]

Euphorbia peplus LINN. Fresh material, Utrecht.

Euphorbia palustris LINN. Fresh material, Hortus Utrecht.

Diplocyathium capitata (REICHB.) Н. Schmidt Baldacci, Aug. 1891 [BR]

Euphorbia esula subtype

Pollen grains with an operculum.

This subtype differs from the other pollen grains in the Euphorbia hirta type by its opercolum.

Euphorbia esula LINN. Fresh material, Utrecht. Euphorbia esula subtype. Oblate spheroidal.  $P=43~\mu~E=40~\mu~P:E=0,93.$  P.A.I. = 0,2.

Euphorbia hirta subtype; subprolate.  $P = 30 \mu E = 23 \mu P:E = 1,30.$ 

 $\dot{P} = 26,5 \ \mu \ E = 28,5 \ \mu \ P:E = 0,93.$ P.A.I. = 0,25.

Euphorbia hirta subtype; oblate spheroidal.  $P = 31,5 \mu E = 34,5 \mu P:E = 0,91$ . P.A.I. = 0,2.

 $E = 45 \mu P: E = 1,10.$ 

Euphorbia hirta subtype. Oblate

Euphorbia hirta subtype. Prolate

P.A.I. = 0,10.

spheroidal.

spheroidal.  $P = 50 \mu$ 

P.A.I. = 0,3.

## Dichostema type

Tricolporate; oblate spheroidal. Pollen grains in polar view circular to three-lobed.

Colpus transversalis; costae.

Colpi narrow; margo small or absent.

Intectate; reticulate. Columellae large. Exine thick.

The Dichostema type is differentiated from the other types by its distinct reticulum and narrow margo. The columellae are fused laterally in their upper part and form a tigillate reticulum. The pollen grains have a thick exine.

Dichostema glaucescens Pierre Klaine 1129 [L] Pl. XXI, 4 Dichostema type; oblate spheroidal-suboblate. P = 31  $\mu$  E = 36  $\mu$  P:E = 0,87. m:e=0,5. P.A.I.=0,3. Lumina 1-3  $\mu$ . Anthostema aubryanum A. DE JUSS. Spuitebanto 1074 [U]

Senefeldera macrophylla Ducke For. Dept. 7549 [U] Pl. XXI, 6

Euphorbia cotinoides MIQUEL Ellenberg 1635 [U] Stahel and Gongryp 816 [U] Dichostema type; oblate spheroidal.  $P=29~\mu~E=31,5~\mu~P:E=0,92.$  Lumina 1-2  $\mu$ .

Dichostema type; spheroidal.  $P = E = 36 \mu$ . P.A.I. = 0,2-0,25. Lumina 2-3  $\mu$ .

Dichostema type; prolate spheroidal.  $P = 28 \mu E = 26 \mu P:E = 1,08.$  m:e = 0.5. P.A.I. = 0,25. Margo absent; Lumina 2-3  $\mu$ .

## Stenadenium type

Tricolporate; subprolate. Pollen grains large. Colpus transversalis broad and elongated; costae.

Colpi narrow and long; costae colpi indistinct. Margo absent.

Tectate; psilate; not reticulate. Columellae distinct but small in diameter. Exine relatively thin.

Pollen grains of this type are large (larger than 40  $\mu$ ) and have a relatively thin exine. The columellae are slender; their diameter is small. In polar view the pollen grains are not distinctly three-lobed. A margo is absent.

The pollen grains of Hura crepitans have a colpus transversalis shorter than those of Stenadenium and Synadenium. The shape and structure, however, are similar.

Stenadenium spinescens PAX Goetze 1099 [BM] Pl. XXI, 1

Elaeophorbia drupifera (THONN.) STAPF
Leeuwenberg 2314 [U] Pl. XXI, 2

Hura crepitans Linn. B. W. 694 [U] B.B.S. 502 [U] Pl. XXII, 2 Synadenium arborescens Boiss. D'Alleizette, Medley 8492 [L] Stenadenium type; subprolate.  $P = 59 \mu E = 47 \mu P:E = 1,26$ . Stenadenium type.  $P = 58 \mu E = 50 \mu P:E = 1,16.$  P.A.I. = 0,2. Tectum perforatum. Stenadenium type.  $P = 75 \mu$   $E = 63 \mu$  P:E = 1,20.

P.A.I. = 0.4.Stenadenium type.

Young.

# Pedilanthus type

Tricolpate; oblate spheroidal to suboblate. Pollen grains slightly three-lobed. Colpi broad. Margo narrow. Membrana granulata. Intectate (tectum perforatum?); psilate. Columellae tall.

Pedilanthus differs from other types in the Hippomane configuration by the absence of a colpus transversalis. Also the P.A.I. is somewhat larger.

Pedilanthus palmeri MILLSP. Hinton [5793 [U] Pl. XXII, 1

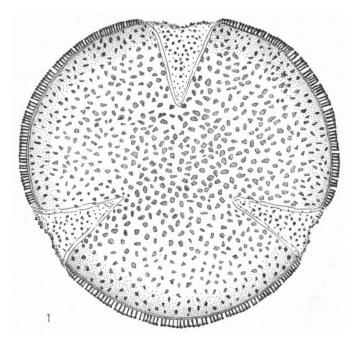
Peailanthus type.  $P = 72 \mu E = 83 \mu P:E = 0.87.$ P.A.I. = 0.3.

POLLEN GRAINS NOT PLACED IN ONE OF THE CROTONOIDEAE CON-**FIGURATIONS** 

# Dalechampia type

Tricolporate; Pollen grains large, usually prolate to prolate spheroidal. Colpus transversalis elongated. Sometimes costae quatoriales.

Colpi short and narrow. No costae colpi present.
Tectate; intra-reticulate. Reticulum very coarse. Between the muri of the reticulum a membrane is present, covering the lumina.



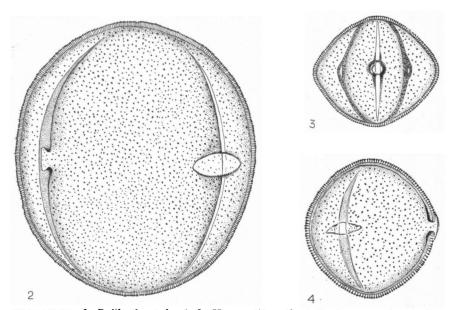


PLATE XXII. 1. Pedilanthus palmeri; 2. Hura crepitans; 3. Actinostemon concolor; 4. Excoecaria bicolor.

The type is characterised by its large pollen grains and wide lumened reticulum. The pollen grains are so typical, that no other type can be compared with them.

### Taxonomic discussion

Dalechampia is easily distinguished from other genera by its typical involucre. For this reason Mueller (1866) placed it in the vicinity of the Euphorbieae. Bentham (1880) pointed to the similar characters in the Plukenetieae: e.g. climbing growth, stinging hairs and thickened style in the female flowers. Pax and K. Hoffmann (1919) thought it better to separate the genus into an apart tribe. Croizat (1940a) and Hurusawa (1954) once again placed the genus near Euphorbia and its allies.

Pollen grains show a typical structure and are neither comparable with the types in the *Plukenetinae* nor with those in the *Euphorbieae*. Pax and K. Hoffmann's classification maintaining the genus in a separate tribus not too far from the *Plukenetinae* seems the best opinion.

Dalichampia dioscoreifolia POEPP. et ENDLICH.

G. Klug 4186 [U] Pl. XXIII, 6 Dalechampia affinis Muell. Arg. B. W. 2183 [U]

Dalechampia brasiliensis LAM. Y. Mexia 5270 [U]

Dalechampia scandens LINN. Serv. Forest. 4328 [U]

Dalechampia spathulata (SCHEIDW.) BAILL.
Fresh material, Hort. Utrecht.
Pl. XXIII, 5

Dalechampia type; prolate.

P = 105  $\mu$  E = 70  $\mu$  P:E = 1,50. Lumina up to 12  $\mu$ . Costae equatoriales. Dalechampia type; prolate spheroidal. P = 105  $\mu$  E = 97  $\mu$  P:E = 1,08. Costae equatoriales.

Dalechampia type; prolate spheroidal.  $P = 57 \mu E = 55 \mu P:E = 1,04$ . Costae equatoriales.

Dalechampia type; oblate spheroidal.  $P = 105 \mu E = 75 \mu P:E = 1,40$ . Costae equatoriales.

Dalechampia type; oblate spheroidal.  $P = 63 \mu E = 65 \mu P:E = 0,97$ . Costae transversales; m:e = 0,5.

# Hamilcoa type

Tricolporate; suboblate. In polar view the pollen grains circular to slightly convex triangular.

Colpus transversalis narrow.

Colpi narrow and short; indistinct costae colpi.

Tectate (?); intra-reticulum fine or not reticulate; columellae distinct but small.

The Hamilcoa type cannot be placed in one of the discussed configurations. The pollen grains have columellae with small but distinct capita and are for that reason excepted from the Mallotus configuration (p. 77). The structure of Hamilcoa would be in better agreement with the Claoxylon configuration, (p. 90), but the related pollen grains of Plagiostyles and Pimeleodendron are reticulate; moreover, the polar view differs. Perhaps the Moultonianthus type in the Bernardia configuration (p. 73) is most related to the Hamilcoa type because of its circular polar view and fine reticulum present in some species.

As the position of this type is still uncertain the author prefers to keep it apart from all other configurations.

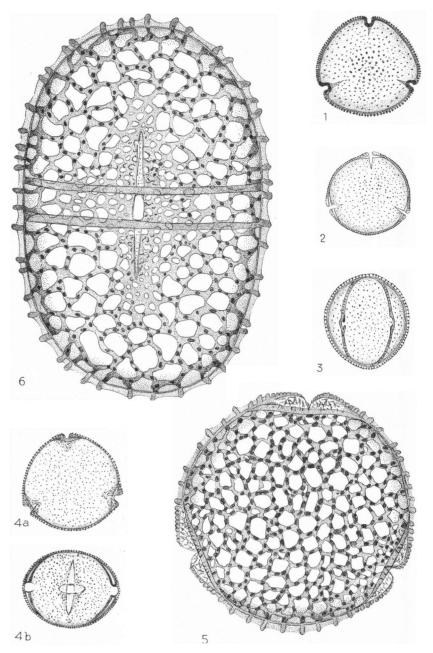


PLATE XXIII. 1. Plagiostyles africana; 2. Pimeleodendron zoanthogyne; 3. Spirostachys venenifera; 4. Hamilcoa zenkeri; 5. Dalechampia spathulata; 6. Dalechampia dioscoreifolia.

#### Taxonomic discussion

Prain (1913) thought Hamilcoa was nearest related to Plagiostyles Pierre and Pimeleodendron Hasskarl. Pax and K. Hoffmann (1912b, 1931) agreed with the relationship of Plagiostyles and Pimeleodendron, which they placed both in the Hippomaneae. Hamilcoa, however, was transferred to the Gelonieae. In doing so they rejected the affinity between Hamilcoa and Plagiostyles.

The three genera have several characters in common:

- 1. Disc absent in male as well as in female flowers.
- 2. In male flowers an ovarium rudiment is absent. Filaments are very short or wanting.
- 3. Pedicels are thickened.
- 4. Styles are undivided.
- 5. Plants dioecious (In Hippomaneae usually monoecious).

Perhaps Nealchornea, in the present paper placed in the Sumbavia configuration (p. 69), is also related to Hamileoa and its allies.

Hamilcoa zenkeri (PAX) PRAIN Zenker 4765 [L] Pl. XXIII, 4

Pimeleodendron zoanthogyne J. J. SMITH Hallier 152a [U] Pl. XXIII, 2

Plagiostyles africana (Muell. Arg.) Prain Zenker 94 [U] Pl. XXIII, 1 Hamilcoa type; suboblate to oblate spheroidal.  $P=23,5~\mu~E=27~\mu~P:E=0,87.$   $m:e<0,5.~P.A.I.>0,5.~Lumina~ca.~1~\mu.$  Hamilcoa type; spheroidal.  $P=E=24~\mu~m:e~ca.~1.~P.A.I.=0,5-0,6.$  Not reticulate. Hamilcoa type; oblate spheroidal.  $P=27~\mu~E=29~\mu~P:E=0,90.$  m:e~ca.~1.~P.A.I.=0,4.~Costae~colpi.

#### TAXONOMIC COMMENT ON THE CROTONOIDEAE

The subfamily of the *Crotonoideae* includes a large number of pollen

The types with a croton-pattern attract most attention. When in the future a revision of the Crotonoideae will be undertaken, the placing of all genera with a croton-pattern into one group will be advisable. This will benefit a natural classification of the subfamily. The relationship of, e.g., Joannesia with Cunuria and Garcia with Sagotia will, in this way, show to better advantage than it does at present in Pax and K. Hoffmann's system.

Two other groups of well-distinguished pollen grains, the *Cnesmosa* configuration and the *Plukenetia* configuration, are found in the subtribe *Plukenetiinae*. Omphalea is added to the *Plukenetia* configuration,

which agrees with the opinion of Croizat (p. 62).

MUELLER ARG., BENTHAM and PAX and K. HOFFMANN include the tribe Chrozophoreae in their systems. This tribe falls into two groups of pollen types. The first group with e.g. Crotonogyne, Aleurites, etc. possesses pollen grains with a croton-pattern. As stated previously these genera should be separated from the other ones and transferred to the group of which Croton is the principal genus. The second group with, e.g., Sumbavia, Chrozophora, Agrostistachys, has aperturate pollen

grains, which belong to several pollen types. The present author would like to retain this second group as the subtribe Chrozophorinae.

The tribe Acalypheae of PAX and K. HOFFMANN includes most genera of the Crotonoideae. There are, however, comparatively few types which are, moreover, closely related (Mallotus configuration, Bernardia configuration, etc.).

The tribes Gelonieae and Cluytieae, too, include genera of which the pollen grains are provided with a croton-pattern mixed up with genera of which the pollen grains lack this structure. These latter genera are divisible into several types. A reclassification of these

tribes is badly needed.

Although Hippomaneae and Euphorbieae are both homogeneous taxa which are taxonomically well-defined, the morphology of the pollen grains shows but little difference between the two groups. Pax (1924) already pointed out the relationship of these groups on other grounds. Since the Hippomaneae are treated by all authors as a tribe of the Crotonoideae, the Euphorbieae should also be treated as a tribe of the Crotonoideae close to the Hippomaneae.

#### **SUMMARY**

In the present study pollen morphology of the Euphorbeaceae is treated as an additional character in taxonomy.

Besides the greater part of the genera occurring in the system of Pax and K. HOFF-MANN (1931), most of the genera published after 1931 are studied.

The pollen grains have been described with the aid of a terminology as simple as possible. In principle the terminology of IVERSEN and TROELS-SMITH has been followed, although in addition, many improvements of ERDTMAN have been used. One of the simplifications is the rejection of POTONIÉ'S term sculpture. All elements occurring on the endexine are called structure elements; all structure elements together form the structure of a pollen grain. For the sake of consequence endexine apertures and extexine apertures are discussed separately.

Different pollen grains are placed in different pollen types. If the differences are of minor importance, the pollen grains are placed in subtypes. Several types can have some characters in common. To express the correspondences, these types are assembled in configurations.

As the pollen types in *Phyllanthoideae* and *Crotonoideae* differ distinctly, the division of the *Euphorbiaceae* in these subfamilies is maintained in the discussion of the results.

The Phyllanthoaieae can be separated in three large groups of pollen types (Antidesma configuration, Amanoa configuration and Aristogeitonia configuration), which agrees with the grouping of PAX in 1924. The remaining small configurations belong in taxonomic respect to the genera of the Antidesma configuration.

In the Crotonoideae many genera possess pollen grains with a croton-pattern. These genera should be treated as a single group.

Besides this natural group, the *Plukenetiinae* possess pollen grains which are clearly distinguished from other genera in the *Crotonoideae*. Pollen grains of *Omphalea* are similar to those in the *Plukenetia* configuration. This pollen-morphological result agrees with the opinion of CROIZAT.

The remaining pollen grains in the Crotonoideae are less easy to differentiate in groups. One of the largest configurations is the Mallotus configuration, which includes most genera of the Acalypheae and several genera or other tribes. The Hippomane configuration is another large one. This configuration comprises the tribes Hippomaneae and Euphorbieae. The pollen grains of both tribes are very similar. The genus Pachystroma is pollen-morphologically as well as taxonomically related to the tribe Hippomaneae.

Pera, treated as a separate tribe by Pax and K. HOFFMANN, is related by its pollen grains to some genera in the Acalypheae.

Dalechampia is habitually related to the genera in the *Plukenetiinae*. Pollenmorphological data, however, do not support this relation. The pollen grains of *Dalechampia* are not similar to any other pollen type.

The morphology of the pollen grains of the Stenolobeae is in agreement with the opinion of Pax, that any separation of these Australian genera is an artificial one.

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