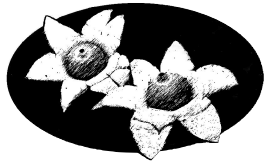


THE QUEENSLAND MYCOLOGIST



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The Queensland Mycological Society

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Society Objectives

The objectives of the Queensland Mycological Society are to:

1. Provide a forum and a network for amateur and professional mycologists to share their common interest in macro-fungi;
2. Stimulate and support the study and research of Queensland macro-fungi through the collection, storage, analysis and dissemination of information about fungi through workshops and fungal forays;
3. Promote, at both the state and federal levels, the identification of Queensland's macrofungal biodiversity through documentation and publication of its macro-fungi;
4. Promote an understanding and appreciation of the roles macro-fungal biodiversity plays in the health of Queensland ecosystems; and
5. Promote the conservation of indigenous macro-fungi and their relevant ecosystems.

Queensland Mycologist

The *Queensland Mycologist* is issued quarterly. Members are invited to submit short articles or photos to the editor for publication. Material can be in any word processor format, but not PDF. The deadline for contributions for the next issue is **1 November 2014**, but earlier submission is appreciated. Late submissions may be held over to the next edition, depending on space, the amount of editing required, and how much time the editor has. Photos should be submitted separately at full-size to allow flexibility in resizing and cropping to fit the space available while minimising loss of quality. Authors who have specific preferences regarding placement of photos should indicate in the text where they want them, bearing in mind that space and formatting limitations may mean that it is not always possible to comply. Material from published sources may be included if that complies with copyright laws and the author and source are properly acknowledged.

Membership

Membership of QMS is \$25 per annum, due at the beginning of each calendar year, and is open to anyone with an interest in Queensland fungi. Membership is **not** restricted to people living in Queensland. Membership forms are available on the website, <http://qldfungi.org.au/>.

Could members please notify the membership secretary (memsec [at] qldfungi.org.au) of changes to their contact details, especially e-mail addresses.

Cover photo: *Mutinus boninensis* photographed at the Linda Garrett Reserve, Mapleton. See page 5 for the foray report. © Frances Guard

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QMS Calendar 2014

2014 Meeting Schedule

Meetings are held in the F.M. Bailey Room at the Queensland Herbarium, Mt Coot-tha, commencing at 7pm on the second Tuesday of the month from February (no January meeting), unless otherwise scheduled. Check the website for details and any changes. There will be 3-4 guest speakers invited during the year and other meetings will be informal. Suggestions from members for topics or names of potential speakers or talks will be welcome at any time. Please contact a member of the executive.

To assist those unable to attend meetings, notes on the

talks are included in the Queensland Mycologist wherever possible. However, the notes never do justice to the topic as they do not reflect the enthusiasm of the speaker or cover the discussion that follows. So remember, where possible it is better to attend the meetings, get the information first hand and participate in the invaluable information sharing opportunity.

Supper. Suppers are provided by volunteers. Check the website for details of the roster and if you are able to assist please contact the secretary.

November 12

Informal-Members' talks

December 10

Christmas Meeting

QMS Forays

The Foray season for 2014 has now concluded.

QMS hold regular forays during the first half of the year. The dates are normally the Saturdays following the QMS meetings of February to July, but additional forays are also held.

Field trip details may change as a result of drought or

other unforeseen circumstances. Check the website for changes.

Members are invited to suggest venues for additional forays. If you have any suggestions (and especially if you are willing to lead a foray), please contact Fran or another member of the executive.

QMS Workshop Program 2014

QMS runs workshops in the second half of the year, when there is little field activity. QMS held a successful workshop on how to study spores as part of the id process on September 27.

Fran is presenting a workshop on the role of fungi in restoring rainforest. It is scheduled for October 31 at Reesville, near Maleny. It is aimed at land managers rather

than QMS, and is more expensive than our usual workshops. Contact Fran for details

Members are invited to suggest topics for workshops. Send your ideas to Fran or Ronda (info@qldfungi.org.au).

Details will be included in future newsletters and on the QMS website.

Editor's Comments

Welcome to new members: Barbara Azevedo de Oliveira, Wayne Boatwright, Thelma Bridle, Bernard Crow, Donna Davis, Ken Goulter, Wayne Hillier, Rod Hobson, Jason Laucher, Mary Mahoney.

Apologies for not welcoming some of you in the last newsletter, and to anybody I missed.

Many thanks to the organisers of the Spores Workshop help on September 27. A group met with our microscopes to look at different kinds of spores and compare notes on their use in identification.

Patrick has once again provided a significant contribution- or rather two of them. Firstly, his report on the Linda Garrett foray, where several hydroid (toothed) fungi were found. These form the subject of a separate article.

We do not have notes from Nigel's talk on fungal biology, but we do have the poem he included, a parody on Dorothea Mackellar's famous work.

There was some hesitation among reviewers about that term, which is often seen as negative these days, but this is in the spirit of the original meaning of "a literary or musical work that copies a serious work in a humorous way". So enjoy!

Theresa Bint has supplied an article on growing oyster and shiitake mushrooms.

Glenda Walter has supplied a note on the beautiful *Hericium* that she photographed in Tasmania, and Vanessa revisits the "white punk" fungus she reported in December 2013.

Finally, Rita de Heer reports a mysterious fungus that appeared after a very wet August in Byron Shire. I must say it reminds me of a fungus that came up during Brisbane's big wet in late 2010, that I thought might be an *Agaricus*, but did not follow up. Lost opportunity!

David Holdom

QMS participation at Rosewood Environment Day

Many thanks to QMS members John Wrench, Bev Miles, Susie Webster, Leesa Baker (photo) and Adrian Harris for operating a QMS stand at the Rosewood Environment Day.

There was quite a bit of interest in the small display and we had specimens that provided a talking point, thanks to Bev and Adrian. The day was enjoyable and we hope to be involved next year.

Photo © Ronda Warhurst



Fungilinx

Two Australian Glossaries of fungi:

Fungi of Australia Glossary

<http://www.environment.gov.au/biodiversity/abrs/online-resources/glossaries/fungi/index.html>

SGSGi - Mycological Glossary

<http://www.sydneyfungalstudies.org.au/glossary.htm>

Some international sites:

New York Botanical Garden <http://www.nybg.org/> Search for "fungi"

Mycological Society of America <http://msafungi.org/>

Introduction to Fungi and the Wisconsin Mycological Society

<http://www.wisconsinmycologicalsociety.org/introtofungiandwms/index.html>

Report on the Linda Garrett Foray 2014

Pat Leonard

The QMS foray at the Linda Garrett Reserve was held in June in 2014, a bit late in the season, but it produced 46 recorded species and was well attended. This site continues to produce species that we have not recorded previously. Only a quarter of the species recorded on this foray had been recorded previously at this site. We managed to key out over half of the collections to a species name and over a third more to genus only.

Number of records	46	
Identified to species	26	56%
Identified to genus only	16	35%
Unidentified	4	8%
Recorded more than twice	12	26%
New to Queensland	3	6%

The find of the foray was the first example of *Multifurca* to be found in Queensland. This is a genus related to *Lactarius*, and recently rediscovered in Tasmania. It remains to be seen whether the species found at LG is the same as the one recorded in Tasmania and Victoria.



Multifurca sp © Patrick Leonard

Multifurca species are often deeply infundibuliform (funnel shaped cap) and the cap is strongly zoned. This one was found growing with *Eucalyptus*. Like *Lactarius* species to which it is related, it exudes milk from the gills when they are cut.

Several clavarioid fungi were seen, *Artomyces austropiperatus* is a small brown candelabra like fungus which is recognised by the very prominent crown like arrangement of short branches that ring the top of each branch. It grows on wood.

Good specimens of *Ramaria filicola* were also found, a white species with crowded fine white branches with a pink tinge.

There were three hydroid fungi found: These are



Artomyces austropiperatus © Susie Webster

reported in a separate article on toothed fungi in this issue.

The most interesting gasteromycete was a fine specimen of *Mutinus boninensis* (front cover) found by Susie Webster whilst searching for truffles. It seems to like growing in damp litter and is thus only found when the litter is disturbed. This was true of the previous collection of this fungus at Mt Cordeaux. It is a slender stinkhorn with a volva like base and strong white rhizoids which appear to link to other specimens and eggs that have not yet opened. The fungus is recognized by its delicate stature, orange gleba and white stipe. There is considerable doubt about the correct name for this

fungus. Cunningham and the interactive catalogue have this as *M. borneensis*, Imazeki and Fuhrer as *Mutinus (Jansia) boninensis*, We adopted the latter name as our specimen matches the images and description presented by Fuhrer and Imazeki.

Four bolete species were found during the foray. Two were *Strobilomyces*, both seen here for the first time, the other two being *Boletus* s.l. The felty brown capped *Boletus dispersus* has been found at other sites in the Blackall Ranges. *Boletus* sp 9 looks very like *Boletus carramarus* until it is cut open to reveal bluing flesh in the cap and upper stipe and beetroot coloured flesh in the lower stipe. The stipe starts almost white but bruises brown on handling.

We were grateful to Fran Guard, our President, for the use of her house and microscopes at the end of the foray. Over lunch and in the hour that followed several specimens were successfully identified.



Boletus sp 9 © Patrick Leonard

Fungi with Teeth

Pat Leonard

The QMS foray at the Linda Garrett Reserve was held in June in 2014, a bit late in the season, but it produced some unusual species of toothed fungi. The 2013 foray at Linda Garrett also recorded an unusual toothed fungus *Sarcodon 'griseoviolaceum'* (Leonard 2013).

Many toothed fungi appear to be rare, or perhaps they are overlooked by mycologists. The latest Linda Garrett finds prompted several questions:

- Is the Linda Garrett Reserve unusual in having several species of toothed fungi?
- are toothed fungi more common in Queensland than elsewhere in Australia?
- what particular characteristics of the reserve favour toothed fungi?

The questions were limited to the species of stipitate hydroid fungi that have a distinct cap and stem and where the fruiting surface is made up of teeth. Non stipitate toothed fungi in Queensland such as *Stecherrinum ochraceum* and *Pseudohydnum gelatinosum* which inhabit wood and form brackets or are resupinate, were excluded, they have very different substrate and habitat requirements.

The number of species of hydnoaceous agarics in Australia is debatable. There seem to be 15 which are published, but only 12 that are confirmed and supported by a collection in a herbarium. Many other names have been used. Only three species are relatively widely distributed or common, where common has been defined as having more than 10 records in two or more states.

Common Stipitate Hydroid Species *Hydnum repandum*



Hydnum repandum © Katie Syme

This species is common in south-east Queensland in early autumn. It is found in rings and quite large groups in a range of habitats, mostly with *Eucalyptus* and has been recorded several times at Linda Garrett. It is a cream to pale apricot coloured fungus with spines (which are usually slightly out of the vertical) and has a pleasant odour. The flesh is white and firm. This is a good edible fungus with a nutty flavour (hazelnuts?). (Pegler 1997).

Hydnaceous agarics recorded in Australia and Queensland.			
Species	Collections in Australia	Collections in Queensland	Notes Collections recorded on ALA
<i>Auriscalpium barbatum</i>	2	0	Only in WA
<i>Beenakia dacostae</i>	132	0	Tas (98) Vic (29) NSW (1)
<i>Hydnellum aurantiacum</i>	0	1	
<i>Hydnellum concrescens</i>	8	0	SA (2) Vic (3) WA (2)
<i>Hydnellum fraudulentum</i>	0	0	In Maas Geesteranus
<i>Hydnellum scrobiculatum</i>	8	0	Vic (7) WA (1)
<i>Hydnum repandum</i>	1199	7	NSW (13) SA (14) Vic (135) Tas (976) WA (42)
<i>Hydnum rufescens</i>	6	0	WA (6)
<i>Phellodon maliensis</i>	10	1	Vic (9)
<i>Phellodon melaleucus</i>	2	0	WA (2)
<i>Phellodon niger</i>	473	2	SA (2) Tas (432) Vic (20) WA (18)
<i>Phellodon plicatus</i>	16	1	Vic (11) WA (5)
<i>Sarcodon fulgineoviolaceus</i>	0	0	
<i>Sarcodon 'griseoviolaceum'</i>	0	1	
<i>Sarcodon laevigatus</i>	0	0	
Source: ALA (2014) Interactive Catalogue			

Phellodon niger

This species is occasionally found in south-east Queensland and the fruiting bodies are often persistent, lasting weeks. It has a zoned cap which is dark brown to black. The teeth are decurrent,



Phellodon niger © Patrick Leonard

grey in colour with a violaceous tinge, becoming ochraceous with age. The flesh is thin and black, which is the diagnostic feature.

Beenakia dacostae

This species has not been recorded in south-east Queensland and appears to be southern in its distribution. It is occasional in Tasmania and occasional in Victoria, New South Wales and New Zealand (Grey & Grey 2005). This is a small fungus, cap less than 25 mm diameter, white to pale yellow brown, with a thin stipe. The teeth are decurrent, beige to pale olive brown. The flesh is thin and yellow brown.

Rare and Uncommon Stipitate Hydroid Species

All the other hydroid fungi appear to be known from a small number of collections. Four of these fungi occur in Queensland:

Hydnellum aurantiacum

The cap is rosette like, irregular, 30 - 50 mm diameter with a silky shiny and radially fibrillose surface. Dark brown to apricot to orange brown at the centre, paler outer third with an undulate (wavy) margin that splits in some specimens. The



Hydnellum aurantiacum © Patrick Leonard

stipe is either central or eccentric, orange brown, with a somewhat swollen base. The spines are usually decurrent, crowded, tapering and about 4 mm long; pale ochre, becoming tawny to purplish brown. It has pale yellowish brown, firm flesh. The spore print is brown and the spores are ellipsoid, 5 - 5.5 × 3.5 - 4 µm, Q = 1.4 and ornamented with flat

topped warts (tubercles). Only a single collection has been made in tall Eucalypt forest growing in litter with characteristic sticks and grass shoots growing through the cap.

There has been some debate over whether the correct name for this fungus is *Hydnellum auratile* or *Hydnellum aurantiacum*. Maas Geesteranus suggested that the former occurred in Australia but that Lloyd had wrongly named the latter on the basis of material that proved to be a polypore. The Queensland collection fits the modern description of *H. aurantiacum* given in Fungi of Switzerland volume 2 (species 257). It is a rare fungus. (Maas Geesteranus 1971).

Phellodon maliensis

The cap is centrally depressed; 25 - 35 mm diameter; radially fibrillose with a dull reflective or shiny buff surface with a few black (olivaceous?) radially arranged spots; faintly concentrically zoned; margin thin, tomentose and appendiculate. The stipe is cylindrical to slightly flattened, tapering towards the base, 25 - 30 × 2 - 3 mm, glabrous and buff. The spines are more or less adnexed, 1 - 5 mm long and dark grey.



Phellodon maliensis © Patrick Leonard

The flesh is thin and dark grey.

The spore print appears to be colourless. The spores are globose to subglobose, 4.1 - 5.6 × 3.8 - 5.3 μm, Q = 1.06 ± 0.05; ornamented with low blunt spines; not amyloid.

Found in wet sclerophyll forest with *Eucalyptus* and *Allocasuarina*. As Maas Geesteranus states, separating *Phellodon* species is very difficult. The

grey flesh, buff pileus with black radial patches and relatively large (for *Phellodon*) spores all suggest that this collection is *Phellodon maliensis*. The spore size and odd pleurocystidia suggested that *Auriscalpium umbella* should also be considered, but that species has amyloid spores. (Maas Geesteranus 1971).

Phellodon plicatus

Caps simple or conrescent (several joined together), 20 - 40 mm diameter, centrally depressed with radial folds, with a silky warm brown surface which is a darker red brown in the centre. Indistinctly zoned. Stipe irregular, mostly short, narrowing towards base and flattened, to 25 mm high, 3 - 7 mm wide. Fibrillose to lanose, dark red brown. Spines decurrent, yellowish grey to very pale brown, 2 mm long. Flesh thin, pale brown. Smell of curry (fenugreek) when dry.



Phellodon plicatus © Patrick Leonard

Spore print not obtained, may be colourless. Spores subglobose to ellipsoid, 3.5 - 5.3 × 3 - 3.5 μm, Q = 1.35, with spines up to 1 μm long.

Growing in wet sclerophyll forest with *Eucalyptus* and *Allocasuarina*.

This species is only known from Australia and New Zealand. (Maas Geesteranus 1971).

Sarcodon 'griseoviolaceum'

The cap is irregularly planoconvex, 35 - 50 mm diameter but some up to 80 mm. The surface is rugulose, concentrically banded with fibrillose brownish bands and velutinous lilac bands. The fibres are more or less hyaline with an overall dull violet (15D3) colour. The margin is distinctly lobed. The stipes are caespitose (joined at the base), irregularly cylindrical or tapering to base, hollow; 25 - 40 × 10 - 15 mm. The surface is minutely velutinate with hyaline fibres, darker lilac (15E4) than cap. The spines are adnate or occasionally adnexed, 4 - 6 mm long, more or less geotropic (pointing straight down) and a bright ochre brown (8E7).

The flesh is firm and relatively thin (1 - 3 mm) in both cap and stipe, pale pink, darkening slightly on exposure to air, with the whole fruit body eventually blackening. Smell: none.



Sarcodon 'griseoviolaceum' © Patrick Leonard

The spore print is a rusty brown, like a *Gymnopilus*. The spores are tuberculate; $7.1 - 8.9 \times 6.5 - 8.4 \mu\text{m}$, $Q = 1.07$. The tubercles are up to $2 \mu\text{m}$ high.

Caespitose in rainforest, growing on a very well rotted mossy log which could not be identified.

Cooke (1892) reports *Sarcodon laevigatum* as present in Queensland on the basis of a collection made by Bailey (1890). The collection is at Kew and Maas Geesteranus (1971) reports that it is not in good condition and is possibly *Hydnum repandum*. This collection was thought at first to be *S. thwaitesii*, but it has different colours and larger spores. It does not match any other species in Maas Geesteranus so we have tagged it '*griseoviolaceum*'.

Discussion

There are very few records of hydroid fungi in Queensland. It is not certain whether this reflects low frequencies or lack of interest in recording this group of fungi. *Hydnum repandum* is much more frequent than the current Queensland records suggest. *Beenakia dacostae* and *Phellodon niger* appear to be a southern species. All the other species are rare both in Queensland and Australia.

Evidence from Britain and Scandinavia suggests the many hydroid species have restricted distributions, and are associated with riverbanks and exposed mineral soils in ancient forests and undisturbed habitats. In these localities it appears that several species occur at the same site, for example in the Caledonian pine forests of the Scottish Highlands (Newton et al 2002).

So it would not be unusual if a similar phenomenon were observed in Queensland. The Linda Garrett forest is a relatively undisturbed site with many ancient trees. There is no evidence of these fungi clustering at other sites, but there is no useful systematic data as yet and there have been no

specialist studies.

There do seem to be differences in the distribution of hydroid fungi between Queensland and the rest of Australia, just as there are for other fungi. For the three more common species there seem to be more records in southern states and *Beenakia* appears to be restricted to the south.

There is as yet too little information to draw firm conclusions on the characteristics that favour these fungi. It may be speculated on the basis of the Linda Garrett records that undisturbed sites which have some bare ground and are occasionally flooded favour the survival of these fungi.

There are so few records of these fungi both in Queensland and elsewhere in Australia that there would appear to be a strong case for placing them on a protected species list to ensure their survival.

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UK Biodiversity Action Plans

<http://www.ukbap.org.uk/ukplans.aspx?ID=338>.

My Fungi!

Compiled by **Nigel Fechner** © 2014

A Parody of "Core of my Heart" (aka "My Country") by Dorothea Mackellar © 1908 [with apologies]

The love of field and coppice,
Of green and shaded lanes.
Of decomposing woodlands
And fruiting after rains,
Strong love of grey-blue cheeses
Brown Swiss and mouldy bread
I know, but cannot eat it,
Lest from my lunch be dead.

I love a fungal bounty,
A land of floating brains,
Of mycorrhizal partners,
Of spores and hyphal strains.
I love her penicillin,
I love her Shiitake,
Her beauty and her terror -
Mycology for me!

A stark white ring-barked forest
Hard brackets do festoon,
Illuminated mountains,
The glowing fungal 'moon'.
Symbiotic dalliance,
Gum tree roots embroil,

And orchids deck the tree-tops
Whilst microbes feed the soil.

Core of my heart, my fungi!
Entoloma - blue as sky,
Puffballs spewing dust clouds,
All of them must die -
But then the grey clouds gather,
And we'll be blessed again
The feeding of an army,
A life reborn from rain.

Core of my heart, my fungi!
With caps of Agarics gold,
Entheogens partaken,
Life's meaning to behold -
Tho' fungi possess mysteries,
Just why are they so named?
Their secrets all revealed to us
Once taxonomy is tamed!

A 'truffle'-laden country,
Harsh unforgiving land -
All you who have not loved them,
You will not understand -
Though earth holds many splendours,
Wherever I may die,
Fair chance that I'll fall victim
To fungi passing by.

Growing oyster and shiitake mushrooms

Theresa Bint

In my experience, growing mushrooms on 'logs' is a lot less tricky than using the mushroom growing kits in boxes. 'Logs' made of compressed sawdust and bran are pre-inoculated with mushroom spawn and matured for some months, so you can usually have your first crop in a couple of weeks. An internet search should bring up some suppliers in your area - I know of one at Coolum and one in Brisbane.

Here's how simple it is:

Log Preparation



Take your plastic wrapped log and cut the covering off each end if it's oyster mushrooms; remove the plastic altogether if you're growing shiitakes. Activate the log by soaking it in water for 24 hours - you may need to hold the log down with something heavy like bricks or pavers.



After soaking, put the log in a large bucket or container with some water in the bottom. Sit it up on a brick so it's out of the water. The idea is to keep the log moist but not sodden, so

have a spray bottle handy to use each day and cover the container with a damp towel. The mushrooms will need a bit of light to develop colour properly - but put them in a cool spot, out of the direct sunlight.

All photos © Theresa Bint



Check your mushroom log each day; soon you'll see little pinheads emerging. They'll quickly develop into button-sized mushrooms then are ready to eat - depending on the temperature - in 3 to 5 days. You can twist them off at the base or snip them with scissors to harvest. Once that flush is finished, rest the log for a couple of days, before re-soaking it for round two!

You'll get four or five harvests from your kit, then put it in the compost or feed it to the worms.

Spore Workshop 27th September 2014

Frances Guard

This Workshop started out as an idea that a few of us, who regularly measure spores, would meet to exchange knowledge and experiences with our microscopes and cameras, and try to learn from each other to improve our results. It then expanded to include members who are just starting to look at spores, and even to those who are checking out microscopes and trying to work out which one to buy. In the end 12 people attended.

We went through a brief run down on the chemicals most commonly used and the care needed when using them. We also looked briefly at spore shapes, colours, and the use of spores in identifying genera. Handouts are available on each of these talks.

We then spent the rest of the Workshop examining different spores, some like *Ganoderma* sp. with classic ornamented double walls, others showing dextrinoid reaction (*Lepiota* sp.), amyloid reaction on ornamentation (*Russula* sp.), some with tails (*Bovista* sp.) etc. We tried measuring spores and comparing our results. (They were comparable!)

Comments from various participants included the following:

"The best part for me was that everyone had something to contribute - whether it be knowledge or enthusiasm. It was also very interesting to see the different kinds of microscopes, cameras and software.

I do hope that we can start building a collection of spore images to add to the FoQ." V.R.

"I found it very useful to be able to compare the DIC microscope with others and see where there were/were not benefits. No benefit with dark, smooth spores I found, but yes it behaved very well with those hyaline spores! Such a pity DIC adds so much to the cost of a microscope. I really do appreciate how lucky I was to get it, even if it is a bit old and temperamental!" D.H.

"I had a great time and learned how to prepare slides from spore prints, gills of fresh specimens and from dried specimens. The discussions about microscope specifications were particularly useful for me. Fran thank you for the organisation. Patrick thank you very much for letting me use your microscope and Megan for stepping me through the preparation of dried specimens and subsequent identification. I'm very much looking forward to getting a microscope and setting up a home lab". D.W.

Another valuable learning experience for all participants!

As a follow on from the Workshop, we felt that it would be useful to go through the setting up and care of our microscopes again with an expert. The Committee will try to organize this for 2015.

Also the suggestion of including a photo of spores with FoQ descriptions was raised. As in some instances spores are unique and diagnostic, we will try to encourage this in future. Further work is needed to refine this suggestion. Other members may have helpful ideas to add. Photos © Vanessa Ryan, Fran Guard



Fungilix

The British Mycological Society has an educational website. It is aimed at teachers but has some useful material and numerous links, including on buying and using microscopes and stains..

<http://www.fungi4schools.org/>

http://www.fungi4schools.org/Microscopy_page.htm

<http://www.fungi4schools.org/Post-16.htm>

Hericium spp. from Tasmania

Glenda Walter

Hericium coralloides is a beautiful fungus, appropriately named for its similarity to marine coral. I saw this one at Liffey Falls, in Tasmania, in August. It grew on the side of a large Myrtle beech, *Nothofagus cunninghamii*, in rainforest, and was about 10 cm in length, although apparently can grow much larger than this. It is described and

pictured in “A Field Guide to Tasmanian Fungi” as having either branched or unbranched spines.

Its close cousin, *Hericium erinaceum*, has a much different growth habit, with shorter spines growing closely in a clump. I saw two groups of *H. erinaceum*, near the Tahune Airwalk and in the forest on the banks of the Gordon River, near Strahan, both about 4cm in length. This species is also found in New Zealand.



Hericium coralloides © Glenda Walter



Hericium erinaceum © Glenda Walter

White Punk - The Truth Revealed!

Vanessa Ryan

If you remember, in a previous article (December 2013), I was scratching my head over the identity of a smallish, white bracket that I had found in Girraween National Park.

At first I thought it might be a baby *Laetiporus portentosus*, commonly known as White Punk. I had done some investigation, but I hadn't come to any happy conclusions. Back then, I had to leave it as a

mystery.

Time, patience and more experience with my microscope and using keys has since revealed the identity of this wonderfully sweet-smelling fungus. Thanks to some suggestions from Tony Young, I discovered it to be trimitic - not monomitic - and this made the process of identification a lot easier. So, at last, with some small confidence I can finally put a name to it... *Trametes lactinea*.



Trametes lactinea © Vanessa Ryan



A Most Mysterious Fungus

Rita de Heer

The Byron Shire, Northern NSW has had a very wet August. About twenty days of cloud. 266 mm of rain in my backyard. Air temperatures for many days around 16 degrees. I'm presuming the soil quite cold too, without the daily dose of sunshine.



Fig 1 © Richard Smith

Yet, after the 'big rain' on the 16th of August, the Leathers in my backyard revived. The *Stereum ostrea* added new golden edges. The various clumps of *Pycnoporus coccineus* in my bush house put out delicate white rims.

At the edge of an old compost heap, the biggest surprise of all ... but let me start at the beginning of that story.

Back in 2013, R Smith, whose property lies on a south-facing slope in local *Eucalyptus* country sent me this [1].

"Came up beside my drive," he said. "I

wasn't home for the early stages of its development. Nothing much to look at now."

I checked my sources. He checked his. *Montagnea arenaria* (Fuhrer, p126) was the fungus most resembling our specimen. Impossible, of course. For one thing, ours wasn't growing on desert sands.

We know we can't ID fungi from pictures in books. We do it to familiarise ourselves with colours and shapes and the language of mycology. Nothing like it, we decided, apart from the naked gills.

This year, after 'big rain' in March, I found these [2, 3, 4] in a pile of grass clippings in one of the parks I frequent. By the way, in Figure 4, I believe the gills are stuck together due to the rain, not that they are forked naturally.



Fig 2 © Rita de Heer

But I made a big jump, I confess, thinking these fungi might be related to R Smith's find. The naked gills, again. These at an earlier stage of disintegration.

A few days ago, after 'big rain' again, in the sodden remains of the old compost heap - soil might be warmer

under there - this one. [5] Similar coloured cap with the wide dark-pinkish rim. Similar texture on the cap with bundles of fibres apparently easily rained off. Gills darkening. White stem.

Unfortunately it stopped raining soon after and a dog walked over it, mashing it into the grass so I wasn't able to study further developments.



Fig 3 © Rita de Heer



Fig 4 © Rita de Heer



Fig 5 © Rita de Heer

Editor's note: Patrick informs me that "*Motangnea* is distinguished by the volva at the base of the stipe. The 'mystery fungus' has chocolate coloured gills and spores, a white ring on the stem and reddening flesh on the cap, all of which suggest that it is an *Agaricus*.." Patrick also notes that a subgenus of *Agaricus*, *Sanguinolenti*, that contains the reddening members of the genus.