

## THE ASCOMYCETE GENUS GYMNOASCUS

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The ascomycete genus *Gymnoascus* is expanded, comprising all Gymnoascaceae with lenticular or discoid (not bivalvate), pigmented ascospores and with ascomata without long, circinate, arcuate, or comb-like appendages. Several species hitherto classified in separate genera are transferred to *Gymnoascus*. *Gymnoascus udagawae* spec. nov. is described. A key to the 14 accepted species is given. *Gymnoascus durus* Zukal is transferred to *Ascocalvatia*. A checklist of all fungi described in *Gymnoascus* is given.

Within the fungi classified in the Gymnoascaceae, von Arx (1971, 1974, 1977) distinguished three phylogenetic entities, which can be recognized easily by the shape and symmetry of the ascospores. The genera *Myxotrichum* Kunze, *Pseudogymnoascus* Raillo, and *Byssosascus* v. Arx are characterized by ellipsoidal or fusiform ascospores having longitudinal furrows in *Byssosascus striatisporus* (Barron & Booth) v. Arx and being smooth or striate by crests in the species of the other genera (Müller & von Arx, 1982; Currah, 1985). The asci are usually spherical, with a distinct cylindrical stalk. Currah classified the three genera in a new family Myxotrichaceae; von Arx (1986) included them in the Onygenaceae, which were emended comprising all Eurotiales with ellipsoidal or fusiform ascospores.

The species of *Amauroascus* J. Schröt., *Arachnotheca* v. Arx, and *Auxarthron* Orr & Kuehn are characterized by spherical ascospores with an ornamented, reticulate, pitted, or striate, often thick wall. They may be related to *Emmonsiella* Kwon-Chung, *Ajellomyces* McDonough & Levis, *Xylogone* v. Arx & Nilsson, and related genera, which have spherical, but apparently smooth and hyaline ascospores.

The Gymnoascaceae are now restricted to genera characterized by dorsiventrally flattened, bivalvate, lenticular, or discoid ascospores. Several genera are characterized by such ascospores, but von Arx (1974, 1977) accepted only a limited number including *Gymnoascus* Baranetzky (ascospores discoid, pigmented), *Arachniotus* J. Schröt. (ascospores lenticular, pigmented) and *Narasimhella* Thirumalachar & Mathur (ascospores bivalvate, hyaline). *Arachniotus ruber* (Tiegh.) J. Schröt. was selected as type species of *Arachniotus*, which was not accepted by Orr & al. (1977). They reintroduced the name *Gymnascella* for two species with discoid ascospores and 'naked' ascomata. Similar species with lenticular ascospores were classified in *Pseudoarachniotus* Kuehn. Currah (1985) restricted *Arachniotus* to *A. ruber* and transferred several other species to *Gymnascella*.

The present study began, when a specimen growing on a decaying hoof of a cow was collected. The specimen was identified as *Gymnoascus reticulatus* Zukal, which also

was collected on a hoof. Zukal (1887) described the fungus as follows (translated from German).

Ascomata roundish in outline, orange-reddish, about 500 µm in diameter, covered with a network of hyphae, which are about 5 µm broad, distinctly septate, and reddish; ascii botryose, nearly spherical, about 13 µm; ascospores lenticular, yellow, thick-walled, about 6.4 µm. No type specimen in existence.

On the present specimen, the ascomata are pulvinate, 200–400 µm and orange or pale reddish. They are covered by a network of rather thick-walled, 3–4 µm broad, near the septa 4.5–6 µm broad, smooth or verruculose hyphae. The spherical ascii have a diameter of 12–16 µm and a thin but rather persistent wall. The ascospores are lenticular, roundish in face view, elliptical in lateral view, have an equatorial, band-like thickening and measure 6–7 × 4–5 µm. Young ascospores are yellow and become orange or ochraceous when mature.

Cultures on hay infusion agar show a poor growth and remain sterile. Ascomata with mature ascospores were observed only on sterilized pieces of hooves. The fungus apparently is highly keratinolytic. In other characters it represents a typical member of *Gymnoascus*. It differs from *G. reessii* mainly by larger ascospores with an equatorial band and by thicker peridial hyphae without seta-like branches.

The presence of an ascomatal peridium with or without seta-like branches in *Gymnoascus* and *Gymnascella* and its absence in *Arachniotus* is not adequate for the delimitation of genera. In some species a peridium is present when the ascomata develop on the natural substrate or in fresh isolates, but is absent in subcultures. *Arachniotus*, *Pseudoarachniotus*, *Petalosporus*, *Plunkettomyces*, *Disarticulatus*, and *Gymnascella* consequently are synonymized with *Gymnoascus*, which is emended as follows.

### G Y M N O A S C U S Baranetzky

*Gymnoascus* Baranetzky in Bot. Ztg 30: 158. 1872. — Type: *G. reessii* Baranetzky.

*Gymnascella* Peck in Ann. Rep. N. Y. St. Mus. 35: 153. 1884. — Type: *Gymnascella aurantiaca* Peck.

*Arachniotus* J. Schröt. in Krypt.-Fl. Schlesiens 3(2): 210. 1893. — Type: *A. ruber* (Tiegh.) J. Schröt.

*Pseudoarachniotus* Kuehn in Mycologia 49: 694. 1957. — Type: *P. roseus* Kuehn.

*Waldemaria* Batista & al. in Atas Inst. Micol. Recife 1: 5. 1960. — Type: *W. pernambucensis* Batista & al. (= *G. dankaliensis*).

*Petalosporus* Ghosh & al. in Mycopath. Mycol. appl. 21: 36. 1963. — Type: *P. nodulosus* Ghosh & al.

*Plunkettomyces* Orr in Mycotaxon 6: 33. 1977. — Type: *P. littoralis* Orr.

*Gymnoascoides* Orr & al. in Mycotaxon 5: 459. 1977. — Type: *Gymnoascoides petalosporus* Orr & al.

*Disarticulatus* Orr in Mycotaxon 6: 35. 1977. — Type: *D. devroeyi* Orr.

*Acitheca* Currah in Mycotaxon 24: 63. 1985. — Type: *A. purpurea* Currah.

Colonies expanding, lemon yellow, ochraceous, orange, or red; ascomatal initials composed of a clavate or cylindrical ascogonium surrounded by a coiled antheridium, or of two coiled hyphal tips; ascomata embedded in the aerial mycelium, non stipitate,

occasionally pulvinate and covered or surrounded by a peridium composed of a loose network of hyaline or pigmented hyphae which may bear lateral hyphal tips or short setae; ascii irregularly disposed, often botryose, sessile, without croziers, spherical or nearly so, with a thin but rather persistent wall, 8-spored; ascospores discoid or lenticular, round in face view, ellipsoidal or quadrangular in lateral view, aseptate, with or without equatorial thickenings or furrows, yellow, orange, ochraceous, or reddish brown when mature; conidia occasionally present, separated from each other or from the conidiogenous cell by double septa, which may be adjacent or separated by empty parts of the hypha.

*Gymnoascus* species are isolated from soil, dung, and plant debris, occasionally from hooves, nails, feathers, or hairs. On the natural substrate the ascomata are often covered with a network of hyphae, which may be absent or reduced in pure culture or when grown in moist chambers. Only a relatively small number of species includes anamorphs. These belong to the form genera *Malbranchea* and *Chrysosporium*, but are unnamed.

#### KEY TO THE SPECIES

- |   |                        |
|---|------------------------|
| 1 a. Ascospores with an equatorial depression or furrow (Fig. 1a, e) . . . . .  | 2                      |
| b. Ascospores without equatorial furrow or depression . . . . .   | 3                      |
| 2 a. Equatorial depression of the ascospores deep, distinct . . . . .   | <i>G. ruber</i>        |
| b. Equatorial depression of the ascospores shallow . . . . .  | <i>G. desertorum</i>   |
| 3 a. Ascospores 3–4 $\mu\text{m}$ in diameter, ascomata with a distinct peridium . . . . .                                    | 4                      |
| b. Ascospores 4–7 $\mu\text{m}$ in diameter, ascomata with or without peridium . . . . .                                      | 6                      |
| 4 a. Ascospores lenticular, with equatorial thickening . . . . .  | <i>G. alatosporus</i>  |
| b. Ascospores without equatorial thickening, usually discoid (Fig. 1h) . . . . .  | 5                      |
| 5 a. Peridial hyphae stiff, with spine-like, often recurved branches . . . . .  | <i>G. reessii</i>      |
| b. Peridial hyphae thin, without spine-like branches . . . . .  | <i>G. petalosporus</i> |
| 6 a. Ascospores lenticular and with equatorial thickenings (Fig. 1b, c, d) . . . . .  | 7                      |
| b. Ascospores discoid or lenticular and without equatorial thickenings . . . . .  | 10                     |
| 7 a. Colonies lemon-yellow, arthroconidia present, isolated from animals of marine environments . . . . .                     | <i>G. littoralis</i>   |
| b. Colonies not lemon-yellow . . . . .  | 8                      |
| 8 a. Keratinolytic, ascomata with a peridium of thick hyphae . . . . .  | <i>G. reticulatus</i>  |
| b. Not keratinolytic, ascomata without peridium of thick hyphae . . . . .   | 9                      |
| 9 a. Ascospores 5.5–7 $\mu\text{m}$ , with a distinct equatorial thickening and occasionally with polar thickenings . . . . . | <i>G. dankaliensis</i> |
| b. Ascospores 4.5–6 $\mu\text{m}$ , with a broad equatorial band . . . . .  | <i>G. punctatus</i>    |
| 10 a. Ascospores lenticular, with distinct poles; colonies lemon-yellow (Fig. 1f) . . . . .                                   | <i>G. citrinus</i>     |
| b. Above characters not combined . . . . .  | 11                     |
| 11 a. Ascospores 6–7 $\times$ 4–5 $\mu\text{m}$ , thick-walled; colonies expanding, orange . . . . .                          | <i>G. devroeyi</i>     |
| b. Ascospores smaller, usually discoid (Fig. 1g) . . . . .  | 12                     |
| 12 a. Colonies orange or ochraceous, anamorphs absent . . . . .   | <i>G. aurantiacum</i>  |
| b. Colonies lemon-yellow or pale, anamorphs present . . . . .   | 13                     |
| 13 a. Colonies restricted, conidia 2–3.5 $\mu\text{m}$ broad . . . . .  | <i>G. nodulosus</i>    |
| b. Colonies expanding, conidia predominant, 6–15 $\times$ 4–5 $\mu\text{m}$ . . . . .   | <i>G. udagawae</i>     |

Most species are well described and correctly delimited by Currah (1985) as *Arachniotus*, *Gymnascella*, *Gymnoasoides*, and *Gymnoascus*. *Gymnascella* sensu Currah is polyphyletic; two species have to be retained in *Narasimhella* Thirumalachar & Mathur

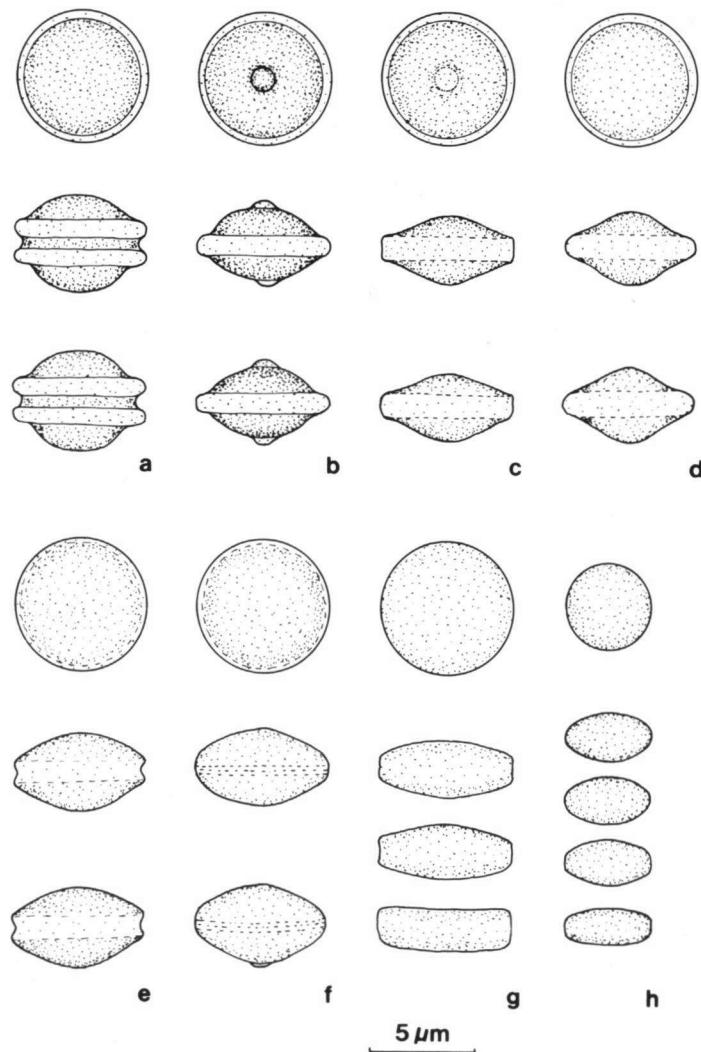


Fig. 1. Ascospores of *Gymnoascus* species in face and lateral view. — a. *G. ruber* (CBS 592.71). — b. *G. dankaliensis* (CBS 294.66). — c. *G. punctatus* (CBS 279.64). — d. *G. littoralis* (CBS 454.73). — e. *G. desertorum* (CBS 634.72). — f. *G. citrinus* (CBS 863.72). — g. *G. aurantiacus* (CBS 655.71) and *G. reessii* (CBS 111.12).

(type species: *N. poonensis* Thirumalachar & Mathur). This genus differs from *Gymnoascus* by unequally bivalve, hyaline ascospores, obovate asci formed from croziers, and by often stipitate ascomata. *Narasimhella* is closely related to *Ctenomyces serratus* Eidam, *Leucosphaera emdenii* v. Arx & Samson and other species with similar, unequally bival-

vate, hyaline ascospores. The mycelial yeast *Saccharomyopsis capsularis* Schiönnig also has bivalvate, smooth, hyaline ascospores and a relationship to the above mentioned Gymnoascaceae has to be considered (von Arx & van der Walt, 1986).

#### CHECKLIST

List of species described as *Gymnoascus* (G.), *Arachniotus* (A.), *Gymnascella*, *Pseudorachniotus* (Ps.), and *Petalosporus* (P.).

Accepted names are printed spaced or in bold-face type (if new).

*P. filamentosus* Orr & Kuehn in Mycologia 64: 62. 1972. — *Gymnascella afilamentosa* (Orr & Kuehn) Currah = *G. nodulosus*.

*G. alatosporus* Natarajan in Proc. Indian Nat. Sci. Acad. 37: 124. 1971.

*A. albicans* Apinis in Mycol. Pap. 96: 45. 1967. — *Arachnotheca albicans* (Apinis) v. Arx in Gen. Fungi, 2nd ed., p. 98. 1974.

*P. anodosus* Kuehn & al. in Mycopath. Mycol. appl. 23: 30. 1964 = *G. nodulosus*.

*G. aurantiacus* (Peck) Sacc., Syll. Fung. 8: 823. 1889 is based on *Gymnascella aurantiaca*, the type species of *Gymnascella* Peck.

*Ps. aurantiacus* Kamyschko in Niv. Sist. Niz. Rast. 4: 224. 1967. — *G. aurantiacus* (Currah, 1985).

*G. aureus* Eidam in Jber. schles. Ges. Kultur 64: 161. 1887. — *Amauroascus aureus* (Eidam) v. Arx in Persoonia 6: 375. 1971.

*G. bifurcatus* (Orr) v. Arx in Gen. fungi, 3rd ed., p. 132. 1981. — *Macronodus bifurcatus* Orr in Mycotaxon 5: 283. 1977 = *Auxarthron conjugatum* (Kuehn) Orr & Kuehn (Currah, 1985).

*G. bourquelotii* Boudier in Bull. Soc. mycol. Fr. 8: 44. 1882 is a nomen dubium. No type specimen in existence.

*G. brevisetosus* Kuehn in Mycologia 48: 813. 1956 = *Auxarthron zuffianum* (Morini) Orr & Kuehn.

*G. californiensis* (Orr & Keuhn) Apinis in Mycol. Pap. 96: 12. 1964. — *Auxarthron californense* Orr & Kuehn.

*G. candidus* Eidam in Jber. Schles. Ges. Kultur 6: 161. 1887 is a nomen dubium. No type specimen in existence.

*G. citrinus* (Massee & Salmon) v. Arx, comb. nov. — *Arachniotus citrinus* Massee & Salmon in Ann. Bot. 16: 62. 1902 (basionym).

*G. confluens* Sartory & Bainier in Bull. Soc. mycol. Fr. 29: 261. 1913 is a nomen dubium. No type specimen in existence. The neotype CBS 352.66 is identical to *G. aurantiacus* (Apinis, 1964).

*G. corniculatus* Orr & Plunkett in Mycopath. Mycol. appl. 21: 11. 1963 = *G. reessii* (Samsno, 1972).

*G. dankaliensis* (Castellani)v. Arx, comb. nov. — *Trichophyton dankaliense* Castellani in J. trop. Med. Hyg. 40: 315. 1937 (basionym). — *A. dankaliensis* (Castellani) v. Beyma in Antonie van Leeuwenhoek 8: 107. 1942.

*G. demonbreunii* Ajello & Cheng in Mycologia 59: 682. 1967. — *Neogymnomycetes demonbreunii* (Ajello & Cheng) Orr (Currah, 1985). Subcultures of the type are sterile.

*G. desertorum* (Moustafa) v. Arx, *comb. nov.* — *A. desertorum* Moustafa in Trans. Br. mycol. Soc. 61: 392. 1973 (basionym).

*G. devroeyi* (Orr) v. Arx, *comb. nov.* — *Disarticulatus devroeyi* Orr in Mycotaxon 6: 35. 1977 (basionym).

*G. dugwayensis* Orr & Kuehn in Mycologia 64: 65. 1972 = *G. reessii* (Currah, 1985). A subculture of the type proved to be sterile.

*G. durus* Zukal in Ber. dt. bot. Ges. 8: 295. 1890 (basionym). — *Keratinophyton durum* (Zukal) Currah in Mycotaxon 24: 156. 1985. — *Ascocalvata dura* (Zukal) v. Arx, *comb. nov.* No type specimen in existence.

Zukal (1890) described the fungus as follows (translated from German): Ascomata spherical, 1–1.5 mm in diameter, white or pale brownish, aggregated in a stroma, hard, with an about 140 µm thick covering composed of thick-walled filaments; ascii in irregular balls, spherical, 8-spored, 6–7 × 5–6 µm, surrounded by branched, tapering filaments; ascospores cylindrical or ellipsoidal, with truncate ends, pale yellow when mature, 3–4.5 × 2.5 µm.

This description agrees with that of *Ascocalvata alveolata* Malloch & Cain (1971). Both species are congeneric, probably conspecific. The genus is a typical Onygenaceae (Malloch & Cain, 1971). The fungus is rare. I observed it about 20 years ago on a cadaver of a salamander. Cultures on agar media from germinating ascospores remained sterile.

The fungus which Currah (1985) identified with *Gymnoascus durus* differs in spherical ascocarps with a dark wall composed of angular cells and in discoid ascospores with a thickened and distinctly pitted margin. It represents a probably undescribed species of *Anxiopsis* = *Aphanoascus*.

*Ps. echinulatus* Dutta & Ghosh in Mycologia 55: 775. 1963. — *Amauroascus echinulatus* (Dutta & Ghosh) v. Arx — *Narasimhella echinulata* (Dutta & Ghosh) v. Arx — *Mallochia echinulata* (Dutta & Ghosh) v. Arx & Samson. This species has to be classified in the Eurotiaceae, because the lenticular, bivalvate ascospores have an equatorial furrow and are echinulate.

*G. eidamii* Cocconi in Mem. Accad. Sci. Inst. Bologna 5: 32. 1891 = *Auxarthron zuffianum* (Currah, 1985).

*A. flavoluteus* Kuehn & Orr in Mycologia 51: 864. 1959 = *G. dankaliensis* (Currah, 1985). Subcultures of the type differ slightly from typical strains of *G. dankaliensis* by paler colonies and the presence of arthroconidia.

*G. flavus* Klöcker in Hedwigia 41: 80. 1902. — *Talaromyces flavus* (Klöcker) Stolk & Samson in Stud. Mycol. 2: 10. 1972 (Onygenaceae).

*A. glomeratus* Müller & Pacha-Aue in Nova Hedwigia 15: 544. 1968. — *Arachnotheca glomerata* (Müller & Pacha-Aue) v. Arx in Persoonia 6: 376. 1971.

*G. gypseus* Nannizzii in Atti Accad. Fisiocr. Sienna 2: 94. 1927. — *Nannizziagypsea* (Nannizzi) Stockdale in Sabouraudia 1: 45. 1961.

*Ps. halophilus* Pawar & al. in Mycopath. Mycol. appl. 40: 100. 1970 = *G. dankaliensis* (von Arx, 1971).

*A. hebridensis* Apinis in Mycol. Pap. 96: 41. 1964. This is a *Chrysosporium* species (von Arx, 1971).

*Ps. hyalinosporus* Kuehn & al. in Mycopath. Mycol. appl. 14: 215. 1961. — *Narasimhella hyalinospora* (Kuehn & al.) v. Arx in Persoonia 6: 374. 1971.

*A. indicus* Chattop. & Das Gupta in Trans. Br. mycol. Soc. 42: 72. 1959 = *Talaromyces flavus* (Stolk & Samson, 1972).

*A. intermedius* Apinis in Mycol. Pap. 96: 45. 1964. — *Talaromyces intermedius* (Apinis) Stolk & Samson in Stud. Mycol. 2: 21. 1972.

*G. intermedius* Orr in Mycotaxon 5: 470. 1977 = *G. reessii* (von Arx, 1981). Currah (1985) accepted this species.

*G. johnstonii* (Massee & Salmon) Orr & Kuehn in Mycopath. Mycol. appl. 21: 8. 1963 is a nomen dubium. No type specimen in existence.

*Gymnascella kamyschkoi* Orr & al. in Mycologia 69: 137. 1977 = *G. aurantiacus* (Currah, 1985).

*A. lanatus* Apinis in Mycol. Pap. 96: 39. 1964 is a nomen dubium. No type specimen in existence.

*A. lectardii* Nicot in Bull. Soc. mycol. Fr. 85: 319. 1969. — *Eleutherascus lectardi* (Nicot) v. Arx in Persoonia 6: 378. 1971.

*G. littoralis* (Orr) v. Arx, comb. nov. — *Plunkettomyces littoralis* Orr in Mycotaxon 6: 33. 1977 (basionym).

*G. longitrichus* Orr & Kuehn in Mycopath. Mycol. appl. 21: 9. 1963 = *G. reessii* (Currah, 1985).

*G. luteus* Sacc., Syll. Fung. 11: 437. 1894. — *Talaromyces luteus* (Sacc.) Stolk & Samson in Stud. Mycol. 2: 23. 1972.

*Ps. marginosporus* Kuehn & Orr in Mycopath. Mycol. appl. 19: 257. 1963 (basionym). — *Narasimhella marginospora* (Kuehn & Orr) v. Arx, comb. nov.

*Narasimhella marginospora* is closely related to *N. poonensis* Thirumalachar & Mathur (1966), but differs by the less distinct equatorial rim of the ascospores and by the absence of stipitate ascomata. *Narasimhella hyalinaspore*, the third species, has ascospores without a distinct rim. In all species the outer (upper) volva of the ascospores has a thicker wall than the inner (lower) volva, which may be covered with some granulae. A strain isolated in 1978 from dung received from India (CBS 125.78) forms distinct, stipitate, orange ascomata and oblate ascospores without rim or brim (Fig. 2).

*G. myriosporus* Rostr. in Meddr. Groenland 18: 12. 1894 is a doubtful species. No type specimen in existence. Probably a *Thelebolus* has been described.

*A. niger* (J. Schröt.) Kuehn & al. in Mycopath. Mycol. appl. 25: 106. 1965. — *Amauroascus niger* J. Schröt.

*G. nodulosus* (Ghosh & al.) v. Arx, comb. nov. — *P. nodulosus* Ghosh & al. in Mycopath. Mycol. appl. 21: 36. 1963 (basionym).

*G. ossicola* Rostr. in Bot. Tidskr. 21: 45. 1897. — *Nannizzia ossicola* (Rostr.) Apinis, but is a doubtful species. No type specimen in existence.

*G. petalosporus* (Orr & al.) v. Arx in Persoonia 9: 397. 1977 is based on *Gymnoascoidea petalosporus* Orr & al. in Mycotaxon 5: 459. 1977.

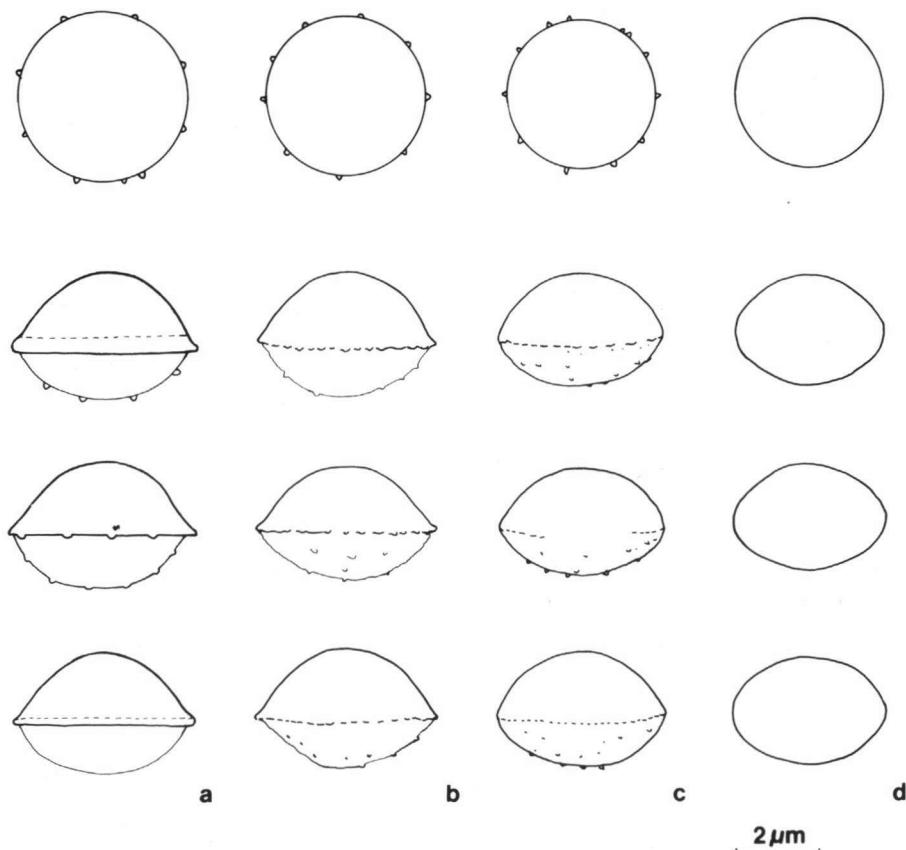


Fig. 2. Ascospores of *Narasimhella* species in face and lateral view. — a. *N. poonensis* (CBS 393.71). — b. *N. marginospora* (CBS 115.54). — c. *N. hyalinospora* (CBS 548.72). — d. *Narasimhella* spec. (CBS 125.78).

*G. punctatus* (Dutta & Ghosh) v. Arx, comb. nov. — *Ps. punctatus* Dutta & Ghosh in Mycologia 56: 153. 1964 (basionym).

*A. purpureus* Müller & Pacha-Aue in Nova Hedwigia 15: 552. 1968. — *Talaromyces purpureus* (Müller & Pacha-Aue) Stolk & Samson in Stud. Mycol. 2: 57. 1972.

*G. reessii* Baranetzky in Bot. Ztg 30: 158. 1872.

*G. reticulatus* Zukal in Verh. zool.-bot. Ges. Wien 37: 40. 1887.

*Ps. reticulatus* Kuehn & Goos in Mycologia 52: 40. 1960. — *Amauroascus reticulatus* (Kuehn & Goos) v. Arx in Persoonia 6: 375. 1971 (but see Currah, 1985).

*A. reticulatus* Kuehn in Mycologia 49: 57. 1957 = *Amauroascus kuehnei* v. Arx in Persoonia 6: 376. 1971.

*G. rhousiogongylinus* Wener & Cain in Can. J. Bot. 48: 325. 1970 = *Pseudogymnoascus roseus* Raillo (Samson, 1972).

*G. roseus* (Raillo) Apinis in Mycol. Pap. 96: 8. 1964. — *Pseudogymnoascus roseus* Raillo. Currah (1985) synonymized *Pseudogymnoascus bhattii* Samson with *Ps. roseus*, but it differs by the absence of a *Geomyces (Chrysosporium)* anamorph and by the colour of the colonies.

*Ps. roseus* Kuehn in Mycologia 49: 695. 1957 = *G. dankaliensis* (Castellani) v. Arx (von Arx, 1971).

*G. ruber* Tiegh. in Bull. Soc. bot. Fr. 24: 159. 1877 is also known as *A. ruber*.

*G. setosus* Eidam in Bot. Zentbl. 10: 107. 1882. — *Myxotrichum setosum* (Eidam) Orr & Plunkett in Can. J. Bot. 41: 1470. 1963.

*G. siglerae* v. Arx in Gen. Fungi, 3rd ed., p. 132. 1981 = *Uncinocarpus reessii* Sigler & Orr in Mycotaxon 4: 461. 1976. The ascospores are similar to those of *G. reessii*: 4–5 µm in diameter, discoid or slightly lenticular, pale brown when mature. *Uncinocarpus* can be accepted as a separate genus of the Gymnoascaceae, when *G. uncinatus* is included in it. Both species form superficial ascromatal structures with long, thick-walled, apically circinate setae and include *Malbranchea* anamorphs (Currah, 1985). In pure culture only the anamorphs develop; asci are formed in moist chambers on hairs mixed with soil.

*G. stipitatus* Lindfors in Svensk bot. Tidskr. 14: 270. 1920. — *Myxotrichum stipitatum* (Lindfors) Orr & Kuehn in Can. J. Bot. 41: 1471. 1963.

*A. striatisporus* Barron & Booth in Can. J. Bot. 44: 1060. 1966. — *Byssodascus striatisporus* (Barron & Booth) v. Arx in Persoonia 6: 377. 1971.

*G. subumbrinus* A. L. Smith in Trans. Br. mycol. Soc. 5: 424. 1917 = *Auxarthron umbrinum* (Boud.) Orr & Plunkett in Can. J. Bot. 41: 1446. 1963.

*G. sudans* Valionis in Vyt. Didziojo Mat. Gamtos Fak. Darbei 11: 115. 1936 = *Byssochlamys nivea* Westling (Stolk & Samson, 1971).

*Ps. terrestris* Thirumalachar & Mathur in Mycopath. Mycol. appl. 40: 102. 1970 = *G. dankaliensis* (von Arx, 1971).

*Ps. thirumalacharii* Mathur in Mycopath. Mycol. appl. 40: 101. 1970 = *G. dankaliensis* (von Arx, 1971).

*A. trachyspermus* Shear in Science 16: 138. 1902. — *Talaromyces trachyspermus* (Shear) Stolk & Samson in Stud. Mycol. 2: 32. 1972. *T. spiculisporus* (Lehman) C. R. Benjamin is a synonym.

*A. trisporus* Hotson in Mycologia 28: 500. 1936 = *Byssochlamys nivea* Westling (Stolk & Samson, 1971).

*Ps. trochleosporus* Kuehn & Orr in Mycologia 64: 58. 1972 = *G. ruber* Tiegh. (Currah, 1985).

*Gymnoascus udagawae* v. Arx, spec. nov. — Fig. 3

Coloniae expandae, citrino-luteae; mycelium ex hyphis septatis hyalinis, 1.5–4 µm in diam. compositum; asci aggregati, sessiles, globosi vel subglobosi, tenui-tunicati, 8-spori, 8–11 µm in diam.; ascosporae dorsiventrali compressae, discoideae, flavae, 4–5 × 2.5–

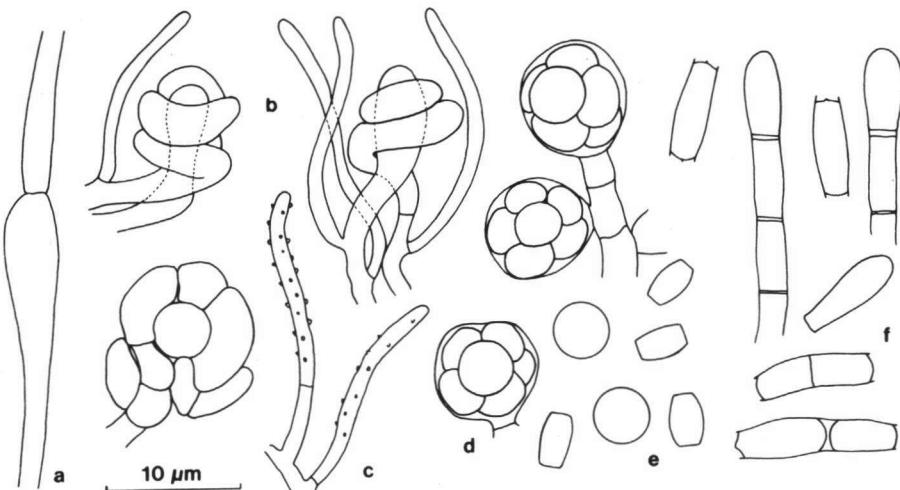


Fig. 3. *Gymnoascus udagawae* (CBS 950.69). — a. Swollen hypha. — b. Initials. — c. Verrucose hyphae surrounding the ascocarps. — d. Ascii. — e. Ascospores in face and lateral view. — f. Conidia.

$3.5 \mu\text{m}$ ; arthro- e aleuroconidia numerosa, cylindracea vel clavata, hyalina, 0- vel 1-septata, 6–15  $\times$  3–5  $\mu\text{m}$ .

Typeus exsiccatus CBS (CBS 950.69, IFO 8921, ATCC 24072).

This species is known by a soil isolate from Japan, described and depicted by Udagawa and Takada (1968) as *A. hebridensis* Apinis. It differs from other species with lemon yellow colonies by the daily growth rate of the aerial mycelium (3–4 mm at  $25^\circ\text{C}$  on hay infusion agar), the relatively small, lenticular ascospores without equatorial thickenings and the abundant formation of relatively large conidia. Young ascocarps are surrounded by delicate, hyaline, often verruculose hyphae extending the ascci (Fig. 3).

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*G. zuffianus* Morini in Mem. Accad. Sci. Inst. Bologna 4: 205. 1889. — *Auxarthron zuffianum* (Morini) Orr & Kuehn in Can. J. Bot. 41: 1445. 1963. Currah (1985) included in *Auxarthron* seven species. Their delimitation, however, is difficult and no key is given.

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