



# Multiple new *Phytophthora* species from ITS Clade 6 associated with natural ecosystems in Australia: evolutionary and ecological implications

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## Key words

aquatic habitat  
breeding systems  
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radiation  
sterility  
survival

**Abstract** During surveys of dying vegetation in natural ecosystems and associated waterways in Australia many new taxa have been identified from *Phytophthora* ITS Clade 6. For representative isolates, the region spanning the internal transcribed spacer region of the ribosomal DNA, the nuclear gene encoding heat shock protein 90 and the mitochondrial *cox1* gene were PCR amplified and sequenced. Based on phylogenetic analysis and morphological and physiological comparison, four species and one informally designated taxon have been described; *Phytophthora gibbosa*, *P. gregata*, *P. littoralis*, *P. thermophila* and *P. taxon paludosa*. *Phytophthora gibbosa*, *P. gregata* and *P. taxon paludosa* form a new cluster and share a common ancestor; they are homothallic and generally associated with dying vegetation in swampy or water-logged areas. *Phytophthora thermophila* and *P. littoralis* are sister species to each other and more distantly to *P. gonapodyides*. Both new species are common in waterways and cause scattered mortality within native vegetation. They are self-sterile and appear well adapted for survival in an aquatic environment and inundated soils, filling the niche occupied by *P. gonapodyides* and *P. taxon salixsoil* in the northern hemisphere. Currently the origin of these new taxa, their pathogenicity and their role in natural ecosystems are unknown. Following the precautionary principle, they should be regarded as a potential threat to native ecosystems and managed to minimise their further spread.

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## INTRODUCTION

During a recent re-evaluation of the *Phytophthora* collection maintained by the Vegetation Health Service of the Department of Environment and Conservation in Western Australia (WA), many new undescribed taxa and unique isolates were identified (Burgess et al. 2009) within what is known as ITS Clade 6 (Cooke et al. 2000). Prior to the advent of molecular systematics in *Phytophthora*, Clade 6 was represented by just three species: *P. gonapodyides*, *P. humicola* and *P. megasperma* (Erwin & Ribeiro 1996). However, an extensive review on the evolution, ecology, reproduction and impact of Clade 6 *Phytophthoras* (Brasier et al. 2003a) introduced eight new informally designated taxa, of which two have subsequently been described as *Phytophthora inundata* (Brasier et al. 2003b) and *P. rosacearum* (Hansen et al. 2009b). Additionally, *P. taxon asparagi* (Saude et al. 2008), a still unnamed pathogen of *Asparagus officinalis*, and *P. pinifolia*, a serious foliar pathogen of *Pinus radiata* in Chile (Durán et al. 2008), have also been described. Recently, further new taxa have been elucidated, but as yet not formally described (i.e. *P. taxon hungarica* and *P. taxon sulawesiensis*).

Clade 6 *Phytophthoras* show a strong association with both forests and riparian ecosystems and, with the exceptions of *P. taxon asparagi*, *P. gonapodyides*, *P. megasperma* and *P. ro-*

*sacearum*, have limited association with agriculture and horticulture. The function of most of these taxa within the ecosystems is very unclear. Brasier et al. (2003a, b) hypothesized a saprotrophic lifestyle for taxa in this clade and their presence, and even dominance, in environmental water surveys is evidence for this (Hansen et al. 2009a, Hwang et al. 2009, Remigi et al. 2009, Hulvey et al. 2010, Reeser et al. 2011). However, some members of ITS Clade 6, such as *P. pinifolia*, *P. inundata*, *P. taxon PgChlamydo* and *P. gonapodyides*, can be opportunistic and sometimes aggressive tree pathogens (Brown & Brasier 2007, Durán et al. 2008, Jung & Nechwatal 2008, Jung 2009).

Based on ITS sequence data, Clade 6 can be divided into three sub-clades. Sub-clade III to date only contains *P. taxon asparagi*, while sub-clade I contains *P. inundata*, *P. humicola*, *P. rosacearum* and some undescribed taxa, separated by relatively long branch lengths. Sub-clade II contains *P. megasperma*, *P. gonapodyides* and the majority of the undescribed taxa, and is characterised by short branch lengths with high support for terminal clades, but weak support for deeper branches suggesting recent radiation from an ancestral type. Within sub-clade II there are several undescribed taxa, including *P. sp. 3*, *P. sp. 7* and *P. sp. 11*, so far found only in Australian natural ecosystems where they are associated with plant mortalities (Burgess et al. 2009).

In this study DNA sequence data from the rDNA internal transcribed spacer regions (ITS) and part of the nuclear heat shock protein 90 (HSP90) and the mitochondrial *cox1* genes were used in combination with morphological and physiological characteristics to describe four new species and a new taxon within sub-clade II: *P. gibbosa*, *P. gregata*, *P. littoralis*, *P. thermophila* and *P. taxon paludosa*.

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Table 1 Identity, host, location, isolation information and GenBank accession numbers for Clade 6 *Phytophthora* isolates used in this study.

Reference collection no. <sup>1,2</sup>	Other collection no.	Identity	Substrate	Host	Location	Isolated by	Date	GenBank Accession No. <sup>3</sup>		HSP90
								ITS	cox1	
VHS21998*		<i>P. gibbosa</i>	Soil	<i>Acacia pycnantha</i>	Australia, WA, Scott River	VHS	2009	HQ012933	HQ012846	HQ012892
CBS127951		<i>P. gibbosa</i>	Soil	<i>Xanthorrhoea gracilis</i>	Australia, WA, Scott River	VHS	2009	HQ012934	HQ012847	HQ012893
VHS21999*		<i>P. gibbosa</i>	Soil	<i>A. pycnantha</i>	Australia, WA, Scott River	VHS	2009	HQ012935	HQ012848	HQ012894
VHS22007*		<i>P. gibbosa</i>	Soil	<i>Grevillea</i> sp.	Australia, WA, Scott River	VHS	2009	HQ012936	HQ012849	HQ012895
VHS22008*		<i>P. gregata</i>	Soil	Road-side (highway)	Australia, WA, Byford		1965	EU301171	HQ012851	HQ012897
DCE68*		<i>P. gregata</i>	Roots	<i>B. prionotes</i>	Australia, WA, Cataby	R Hart	1996	HQ012938	HQ012852	HQ012898
HS42356*		<i>P. gregata</i>	Soil	<i>Pinus radiata</i>	Australia, WA, Nannup	MJC Stukely	1982	EU301172	HQ012853	HQ012899
MJS235*		<i>P. gregata</i>	Root collar	<i>P. radiata</i>	Australia, WA, Nannup	MJC Stukely	1981	EU301173	HQ012854	HQ012900
MJS238*		<i>P. gregata</i>	Soil	<i>Eucalyptus</i> sp.	Australia, VIC, Toolangi North State Forest	WA Dunstan	2008	HQ012939		
MUCC759		<i>P. gregata</i>	Soil	Pasture	Australia, VIC, Devlins Bridge	WA Dunstan	2008	HQ012940		
MUCC760*		<i>P. gregata</i>	Soil	<i>X. preissii</i>	Australia, WA, Lancelin	VHS	2001	EU301174	HQ012856	HQ012902
VHS9854*		<i>P. gregata</i>	Soil	<i>Hakea</i> sp.	Australia, WA, Busseton	VHS	2009	HQ012941	HQ012857	HQ012903
VHS21961		<i>P. gregata</i>	Soil							
VHS21962*		<i>P. gregata</i>	Soil	<i>Patersonia</i> sp.	Australia, WA, Busseton	VHS	2009	HQ012942	HQ012858	HQ012904
CBS127952		<i>P. gregata</i>	Soil	Native forest	Australia, WA, Scott River	VHS	2009	HQ012943	HQ012859	HQ012905
VHS21992*		<i>P. gonapodyides</i>	Soil	<i>Quercus robur</i>	Germany, Rhineland, Bienwald	T Jung	1996	AF541889		
IMI389723	P1047	<i>P. gonapodyides</i>	Soil	Native forest	Australia, TAS, Pine Lake	K Shanahan	1996	AF541888		
IMI389727	P897	<i>P. gonapodyides</i>	Water	<i>E. obliqua</i> forest	Australia, VIC, Toolangi North	WA Dunstan	2008	HQ012937	HQ012850	HQ012896
MUCC761*		<i>P. gonapodyides</i>		<i>Malus sylvestris</i>	USA, New York			AY129175		
IMI302303		<i>P. humicola</i>	Soil	<i>Citrus</i>	Taiwan	PJ Ann & WH Ko	1981	AF266792		EU080172
CBS200.81		<i>P. humicola</i>	Soil			AJ Uddin		AB367496		
VHS16836		<i>P. inundata</i>	Soil	<i>X. preissii</i>	Australia, WA, Boyup Brook	VHS	2007	HQ012944	HQ012860	
VHS19081		<i>P. inundata</i>	Soil	<i>B. attenuata</i>	Australia, WA, Bold Park	VHS	2008	HQ012945	HQ012861	
IMI389750	P210	<i>P. inundata</i>	Roots	<i>Aesculus hippocastanum</i>	UK, Buckinghamshire, Claydon	CM Brasier	1970	AF541912		EU079945
MUCC762*		<i>P. litoralis</i>	Water	Stream baiting	Australia, WA, Borden	D Hüberli	2008	HQ012946	HQ012862	HQ012907
MUCC763*		<i>P. litoralis</i>	Water	Stream baiting	Australia, WA, Borden	D Hüberli	2008	HQ012947	HQ012863	HQ012908
VHS17085*		<i>P. litoralis</i>	Soil	<i>Banksia</i> sp.	Australia, WA, Hopetoun	VHS	2007	EU593262	HQ012864	HQ012909
VHS19173		<i>P. litoralis</i>	Soil	<i>X. preissii</i>	Australia, WA, Wilga	VHS	2008	EU869119	HQ012865	HQ012910
VHS20763*		<i>P. litoralis</i>	Soil	<i>Banksia</i> sp.	Australia, WA, Ravensthorpe	VHS	2008	HQ012948	HQ012866	HQ012911
CBS127953		<i>P. megasperma</i>	Soil	<i>Banksia</i> sp.	Australia, WA, North Dinninup	VHS	1992	HQ012949	HQ012867	HQ012906
DD53432*		<i>P. megasperma</i>	Soil	<i>X. platyphyla</i>	Australia, WA, Esperance	VHS	2007	EU301166	HQ012868	HQ012912
VHS17183		<i>P. megasperma</i>	Soil	<i>M. sylvestris</i>	Australia, WA,	HL Harvey	1968	AF266794	AY129211	
IMI389741	P1278	<i>P. megasperma</i>	Soil	Apricot	USA, California	SM Mircetich		AF514897		EU080062
CMW26667		<i>P. megasperma</i>	Needles	<i>Brassica napus</i>	Australia		1985	EU725805		
CMW26668		<i>P. pinifolia</i>	Needles	<i>P. radiata</i>	Chile, Arauco, Llico plantation	A Durán	2007	EU725806		
IMI389749	P462	<i>P. pinifolia</i>	Water	<i>P. radiata</i>	Chile, Arauco, Llico plantation	A Durán	2007	AF541911		
MUCC764*		<i>P. rosacearum</i>	Water	<i>Malus</i> sp.	USA, California, Sonoma County	SM Mircetich	1979	HQ012950	HQ012869	HQ012913
VHS3655		<i>P. thermophila</i>	Soil	Stream baiting	Australia, WA, Brunswick	GESJ Hardy	2008	HQ012951	HQ012870	HQ012914
VHS7474*		<i>P. thermophila</i>	Soil	Native forest	Australia, WA, Quininup	VHS	1998	HQ012952	HQ012871	HQ012915
		<i>P. thermophila</i>	Soil	Native forest	Australia, WA, Manjimup	VHS	2000			

<b>VHS13530S*</b> <b>CBS127954</b>	<i>P. thermophila</i>	Soil	<i>E. marginata</i>	Australia, WA, Dwellingup	VHS	2004	EUJ01155	HQ012872	HQ012916
VHS13567	<i>P. thermophila</i>	Roots	<i>E. marginata</i>	Australia, WA, Dwellingup	VHS	2004	EUJ01156	HQ012873	HQ012917
VHS13761	<i>P. thermophila</i>	Soil	<i>E. marginata</i>	Australia, WA, Dwellingup	VHS	2004	EUJ01157	HQ012874	HQ012918
<b>VHS16164*</b>	<i>P. thermophila</i>	Soil	<i>B. grandis</i>	Australia, WA, Pemberton	VHS	2006	EUJ01158	HQ012875	HQ012919
VHS17175	<i>P. taxon asparagi</i>	Soil	<i>B. media</i>	Australia, WA, Esperance	VHS	2007	EUJ01167	HQ012844	HQ012890
VHS17644	<i>P. taxon asparagi</i>	Soil	<i>Lomantra sonderi</i>	Australia, WA, Murdoch	VHS	2007	EUJ01168	HQ012845	HQ012891
IMI389747	<i>P. taxon asparagi</i>	Soil	<i>Asparagus officinalis</i>	New Zealand, Whakatane	EM Hansen	1986	FJ801481		EU080568
IMI389747	<i>P. taxon forestisoli</i>	Soil	Native forest	France, Alsace, Illwald forest	EM Hansen	1998	AF541908		
UASWS0315	<i>P. taxon forestisoli</i>	Soil	<i>Alnus glutinosa</i>	Poland, Kolo	L Beibahri	2006	EF522138		
<b>MUCC768</b>	<i>P. taxon humicola-like</i>	Water	Stream baiting	Australia, WA, Esperance	D Hübertli	2008	HQ012959	HQ012883	HQ012927
MUCC769	<i>P. taxon humicola-like</i>	Water	Stream baiting	Australia, WA, Esperance	D Hübertli	2008	HQ012960	HQ012884	HQ012928
UASWS0318	<i>P. taxon hungarica</i>	Soil	<i>A. glutinosa</i>	Poland, Kolo	L Beibahri	2006	EF522141		
UASWS0321	<i>P. taxon hungarica</i>	Soil	<i>A. glutinosa</i>	Poland, Adamowizna	L Beibahri	2006	EF522144		
MUCC770	<i>P. taxon kwongan</i>	Soil	<i>Hibbertia</i> sp.	Australia, WA, Coojarloo	WA Dunstan	2008	HQ012957	HQ012881	HQ012925
IMI329669	<i>P. taxon kwongan</i>	Roots	<i>B. prionotes</i>	Australia, WA, Cervantes	TC Hill	1986	EU593265	HQ012889	HQ012932
HAS2313	<i>P. taxon kwongan</i>	Water	Baiting	Australia, WA, Coojarloo	R Hart	1996	HQ012961	HQ012885	HQ012929
IMI389733	<i>P. taxon oaksoil</i>	Soil	<i>Q. robur</i>	France, Alsace, Illwald Forest	EM Hansen	1998	AF541906		
<b>MUCC765*</b>	<i>P. taxon paludosa</i>	Water	Pond baiting	Australia, VIC, Sugarloaf Reservoir Reserve	WA Dunstan	2008	HQ012953	HQ012876	HQ012920
IMI389730	<i>P. taxon PgChlamydo</i>	Roots	<i>Prunus</i> sp.	UK, Cheltenham	CM Brasier	1982	AF541900		
<b>DD33753</b>	<i>P. taxon PgChlamydo</i>	Soil	Native forest	USA, California	D Ferrin	2002			EU079573
VHS6595	<i>P. taxon PgChlamydo</i>	Soil	Native forest	Australia, WA, Manjimup	VHS	1995	EUJ01160	HQ012878	HQ012922
<b>MUCC766</b>	<i>P. taxon PgChlamydo</i>	Water	Stream baiting	Australia, VIC, Yea Wetlands	VHS	1999	EUJ01159	HQ012879	HQ012923
<b>VHS14801</b>	<i>P. taxon personii</i>	Soil	<i>Nicotiana glauca</i>	USA	WA Dunstan	2008	HQ012955		EU080316
<b>MUCC767</b>	<i>P. taxon personii</i>	Water	Stream baiting	Australia, WA, Busselton	VHS	2005	EUJ01169	HQ012877	HQ012921
IMI389745	<i>P. taxon rasperry</i>	Roots	<i>Rubus idaeus</i>	Australia, VIC, T-Tree Creek, Melba Highway	WA Dunstan	2008	HQ012954		
IMI389744	<i>P. taxon rasperry</i>	Soil	Dying vegetation	Australia, TAS, Pine Lake	G McGregor	1996	AF541904		
IMI389746	<i>P. taxon rasperry</i>	Roots	<i>Tulipa gesneriana</i>	Japan, Chiba, Shirako	K Shanahan	1997	AF541903		
IMI389746	<i>P. taxon rasperry</i>	Soil	<i>R. idaeus</i>	Sweden, Scania	AJ Uddin	1994	AB367377		
92-209C	<i>P. taxon rasperry</i>	Soil	<i>Betula pendula</i>	Germany, Bavaria, Neuburg	CHB Olsson	2006	AF541905	HQ012888	
UASWS0213	<i>P. taxon rasperry</i>	Soil	<i>Ilex aquifolia</i>	Switzerland	T Jung	2006	HQ012964		
2FFL-2008	<i>P. taxon rasperry</i>	Soil	<i>A. glutinosa</i>	Poland	L Beibahri	2008	EUJ06591		
P1044	<i>P. taxon riversoil</i>	Soil	Riparian vegetation	Hungary	I Szabo	1997	DQ396425		
DD32909	<i>P. taxon rosacearum-like</i>	Soil	<i>P. radiata</i>	UK, Worcestershire, Riverbank	J Delcan	1989	EU594600	HQ012958	HQ012926
HAS2529	<i>P. taxon rosacearum-like</i>	Water	Baiting	Australia, WA, Coojarloo	MJC Stukely	1998	AF541907	HQ012882	HQ012930
HS-A2530	<i>P. taxon rosacearum-like</i>	Water	Baiting	Australia, WA, Coojarloo	R Hart	1998	HQ012962	HQ012886	HQ012930
HS-A1959	<i>P. taxon salixsoil</i>	Root	<i>Alnus</i> sp.	Denmark, Funen, Odense	R Hart	1995	HQ012963	HQ012887	HQ012931
IMI389725	<i>P. taxon salixsoil</i>	Water	Road drainage sump baiting	Australia, WA, Weislpool	K Thinggaard	1994	AF541909		
IMI389725	<i>P. taxon sulawesiensis</i>	Root	<i>Salix</i> sp.	UK, Kent, Bexley Heath	R Hart	1972	HQ012956	HQ012880	HQ012924
IMI389735	<i>P. taxon sulawesiensis</i>	Root	<i>Syzygium aromaticum</i>	Indonesia, Sulawesi	CM Brasier	1989	AF266793		EU080534
	<i>P. taxon walnut</i>	Root	<i>Juglans hindsii</i>	USA, California, Merced County,	MD Coffey	1989	FJ801912		
					SM Mircetich	1988	AF541910		

<sup>1</sup> Abbreviations of isolates and culture collections: CBS = Centraalbureau voor Schimmelcultures Utrecht, Netherlands; IMI = CAB International Mycological Institute, UK; VHS = Vegetation Health Service Collection, Department of Environment and Conservation, Perth, Australia; DDS = earlier prefix of VHS Collection; TCH = TC Hill, in VHS Collection; MJS = MJC Stukely, in VHS Collection; HSA = Hart, Simpson and Associates, in VHS Collection; DCE = EM Davison, in VHS Collection; MUCC = Murdoch University Culture Collection.

<sup>2</sup> Numbers in **bold** are isolates used in the morphological studies; numbers with asterisk are isolates used in the growth rate studies; § denotes type isolates.

<sup>3</sup> GenBank numbers in *italics* are from previous studies.

## MATERIAL AND METHODS

### Sampling and *Phytophthora* isolation

Soil and root samples were collected from beneath dying, *Phytophthora*-sensitive 'indicator species' in native ecosystems. Samples were baited with *Eucalyptus sieberi* cotyledons (Marks & Kassaby 1974) that were plated after 5 d and 10 d onto NARPH (Hüberli et al. 2000), or P<sub>10</sub>VPH (Tsao & Guy 1977) selective media, from which pure cultures of *Phytophthora* were then isolated. In some cases plant roots were surface-sterilised in 70 % ethanol for 30 s followed by four rinses in distilled water, and plated directly onto selective media. Some isolates were derived from stream or water baiting. Leaves of several plant species, including *Citrus limon*, *Quercus robur* and *Pittosporum undulatum*, were suspended in mesh bags in streams, rivers or other water bodies for 3–5 d. Small sections of necrotic lesions on leaves were then plated onto selective media. Cultures derived from earlier research and surveys have been incorporated (Table 1).

### DNA isolation, amplification and sequencing

The *Phytophthora* isolates were grown on half-strength potato-dextrose agar (PDA, Becton, Dickinson and Company, Sparks, MD 21152 USA; 19.5 g PDA, 7.5 g of agar and 1 L of distilled water) at 20 °C for 2 wk and the mycelium was harvested by scraping from the agar surface with a sterile blade and placed in a 1.5 mL sterile Eppendorf® tube. Harvested mycelium was frozen in liquid nitrogen, ground to a fine powder and genomic DNA was extracted according to Andjic et al. (2007).

The region spanning the internal transcribed spacer (ITS1–5.8S–ITS2) region of the ribosomal DNA was PCR amplified and sequenced using the primers ITS6 (Cooke et al. 2000) and ITS4 (White et al. 1990). For representative isolates from each species, heat shock protein 90 (HSP90) gene was amplified with HSP90\_F1 and HSP90\_R2 (Blair et al. 2008). Templates were sequenced in both directions with primers HSP90\_F1int, HSP90\_F3, HSP90\_F2, HSP90\_R1 and HSP90\_R2 (Blair et al. 2008). After an initial comparison of sequences the first half of the HSP90 gene was found to be more variable for ITS Clade 6 *Phytophthoras*, and remaining isolates were sequenced with only HSP90\_F1int and HSP90\_R1. The mitochondrial gene *cox1* was amplified with primers FM84 and FM83 (Martin & Tooley 2003). Templates were sequenced in both directions with primers used in amplification, as well as primers FM 85 and FM 50 (Martin & Tooley 2003).

The PCR reaction mixture to amplify the three gene regions, the clean-up of products and sequencing were as described by Andjic et al. (2007). The PCR conditions for ITS, HSP90 and *cox1* amplification were as described by Andjic et al. (2007), Blair et al. (2008) and Martin & Tooley (2003), respectively. All sequences derived in this study were deposited in GenBank and accession numbers are given in Table 1.

### Phylogenetic analysis

The sequence data of *Phytophthora* isolates used in this study were compared with other closely related species (ITS Clade 6) including undescribed taxa available from GenBank (<http://www.ncbi.nlm.nih.gov/>). Sequence data for the ITS region were initially aligned and subsequent manual adjustments made using Geneious Pro v4.8.1 (Drummond et al. 2010).

Using the same aligned datasets, parsimony analysis and partition homogeneity tests were performed in PAUP (Phylogenetic Analysis Using Parsimony) v4.0b10 (Swofford 2003) and Bayesian analysis was conducted with MrBayes v3.1 (Ronquist & Huelsenbeck 2003) using the same constraints and models as described previously (Jung & Burgess 2009). All datasets

and trees derived from parsimony and Bayesian analyses are available from TreeBASE (10764; <http://www.treebase.org/>).

### Morphology of asexual and sexual structures

Sporangia, hyphal swellings, chlamydospores and gametangia of four isolates of *P. gibbosa*, nine isolates of *P. gregata*, five isolates of both *P. litoralis* and *P. thermophila*, two isolates of *P. megasperma* and one isolate of *P. taxon paludosa* (Table 1) were measured on V8 agar (V8A) (16 g agar, 3 g CaCO<sub>3</sub>, 100 mL Campbell's V8 juice, 900 mL distilled water), as described in detail by Jung et al. (1999). Sporangia were produced by flooding 15 × 15 mm agar squares taken from growing margins of 3–5 d old colonies, so that their surfaces were just covered with non-sterile soil extract (100 g of soil from a *Eucalyptus marginata* stand suspended in 1 L distilled water, incubated for 24 h at 20 °C and then filtered through cheesecloth followed by Whatman no. 1 paper) in 9 cm Petri dishes which were incubated at 18–22 °C in natural daylight. The soil extract was decanted and replaced again after 6 and 12 h. After 24–36 h dimensions and characteristic features of 50 mature sporangia and 25 exit pores, and zoospore cysts per isolate, chosen at random, were determined at ×400 magnification (BX51, Olympus). After 5–7 d, 25 hyphal swellings and 50 chlamydospores, if formed, were also measured.

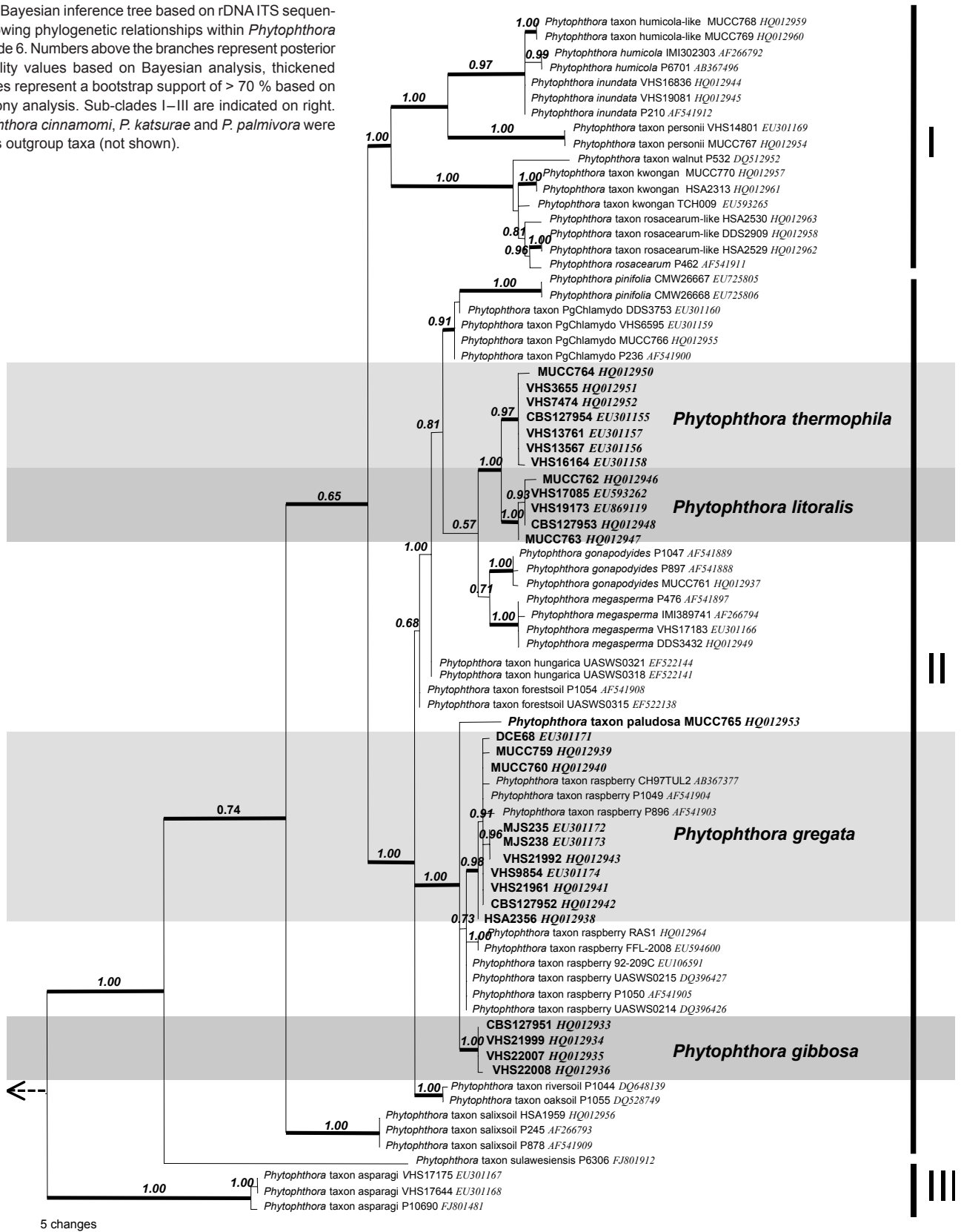
Isolates grown in the dark on V8A plates at 20 °C for 14–21 d were examined for the presence of oogonia. Isolates which had either a high incidence of oogonial abortion or did not produce, or only inconsistently produced oogonia in single culture were paired on V8A with isolates of the same species, with A1 and A2 tester strains of *P. cambivora* (MP45, MP73), *P. cinnamomi* (MP75, DCE60) and *P. cryptogea* (MP21, MP22), and with *Trichoderma reesei* (Brasier 1972). Inoculum plugs (5 mm diam) of the isolate to be tested and the tester isolate were placed on opposite sides of a 9 cm Petri dish, 2 cm from the edge. The plates were incubated at 20 °C in darkness and scored for oogonial formation 30 d after the two colonies had met. For each isolate producing oogonia (either in single culture or when paired), dimensions and characteristic features of 50 mature oogonia, oospores and antheridia chosen at random were measured at ×400. The oospore wall index was calculated as the ratio between the volume of the oospore wall and the volume of the entire oospore (Dick 1990). Descriptions, illustrations and nomenclatural data were deposited in MycoBank ([www.mycobank.org](http://www.mycobank.org); Crous et al. 2004).

### Colony morphology, growth rates and cardinal temperatures

Hyphal morphology and colony growth patterns were described from 7 d old cultures grown at 20 °C in the dark on V8A, malt extract agar (MEA), and half-strength PDA (all from BBL, Becton, Dickinson & Co, Sparks MD 21152 USA). Colony morphologies were described according to patterns observed previously (Erwin & Ribeiro 1996, Brasier et al. 2003a, Jung et al. 2003).

For temperature-growth relationships, representative isolates (Table 1) were sub-cultured onto V8A plates and incubated for 24 h at 20 °C to stimulate onset of growth (Hall 1993). Then three replicate plates per isolate were transferred to 15, 20, 25, 30, 32.5, 35 and 37 °C. Radial growth was recorded after 5–7 d later along two lines intersecting the centre of the inoculum at right angles and the mean growth rates (mm per day) were calculated. Plates showing no growth at 35 and 37 °C were returned to 20 °C to determine isolate viability.

**Fig. 1** Bayesian inference tree based on rDNA ITS sequences showing phylogenetic relationships within *Phytophthora* ITS Clade 6. Numbers above the branches represent posterior probability values based on Bayesian analysis, thickened branches represent a bootstrap support of > 70 % based on parsimony analysis. Sub-clades I–III are indicated on right. *Phytophthora cinnamomi*, *P. katsurae* and *P. palmivora* were used as outgroup taxa (not shown).



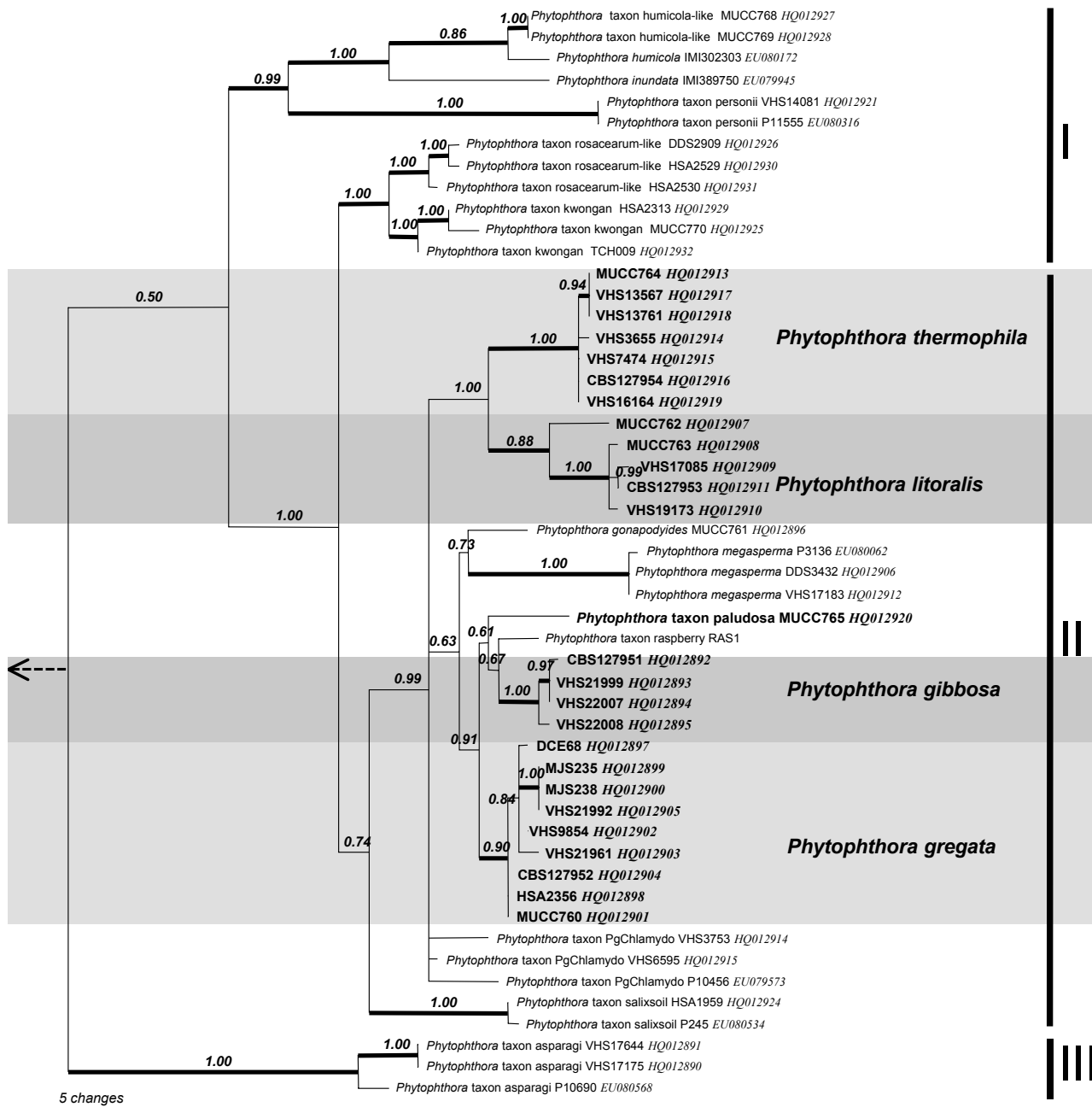
**RESULTS**

**Phylogenetic analysis**

Within the ITS region, the majority of mutations are single base pair mutations and there are occasional small indels of 1–3 bp. There were no gaps in the *cox1* and HSP90 alignments. Excluding outgroups, the aligned datasets for ITS (79 sequences), HSP90 (51 sequences) and *cox1* (49 sequences) consisted of 846, 1 024 and 1 189 characters, respectively. Based on

partition homogeneity tests in PAUP, the ITS and HSP datasets were congruent ( $P = 0.12$ ), but they were not combined due to the lack of HSP90 sequence data for many of the designated taxa. The *cox1* dataset was incongruent with both the nuclear gene regions ( $P < 0.01$ ).

ITS characterisation of 10 species and 15 designated taxa — Including outgroups, the aligned ITS dataset contained 927 characters of which 195 were parsimony informative (158 or 18.67 % of sites are variable within Clade 6 alone) with

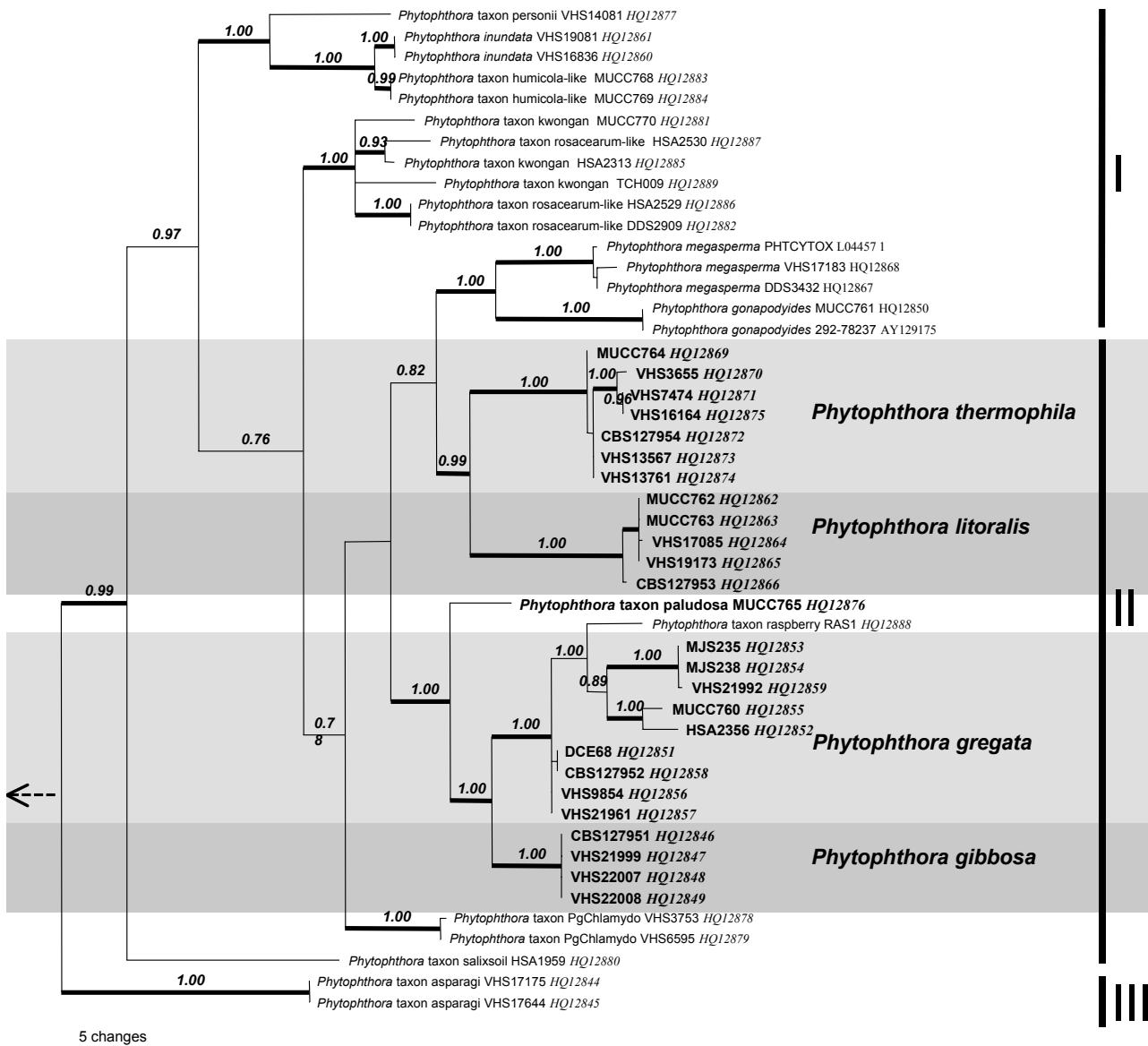


**Fig. 2** Bayesian inference tree based on HSP90 sequences showing phylogenetic relationships within *Phytophthora* ITS Clade 6. Numbers above the branches represent posterior probability values based on Bayesian analysis, thickened branches represent a bootstrap support of > 70 % based on parsimony analysis. Sub-clades I–III are indicated on right. *Phytophthora cinnamomi*, *P. katsurae* and *P. palmivora* were used as outgroup taxa (not shown).

significant ( $P < 0.01$ ,  $g1 = -0.46$ ) phylogenetic signal compared to 1 000 random trees. Heuristic searches resulted in 108 most parsimonious trees of 455 steps (CI = 0.61, RI = 0.89). The Bayesian analysis provided more support for deeper branches. Support for terminal clades and their clustering was equivalent in both analyses and the Bayesian analysis is presented here (Fig. 1, TreeBASE 10764). As previously reported (Brasier et al. 2003a), the analysis resolved three sub-clades. There are 59 variable sites in sub-clade I (7.13 %) and 83 in sub-clade II (10.03 %). Sub-clade I contained two clusters: the first included *P. inundata*, *P. humicola*, *P. humicola*-like and *P. taxon personii*, whilst the second cluster contained *P. rosacearum*, *P. taxon rosacearum*-like, *P. taxon kwongan* and *P. taxon walnut*. Sub-clade III is represented by *P. taxon asparagi*, and *P. taxon sulawesiensis*, for which only a single sequence is available on GenBank.

Sub-clade II is larger and contains 14 discrete lineages corresponding to three described species (*P. gonapodyides*, *P. mega-*

*sperma*, *P. pinifolia*), four new EU species (*P. gibbosa*, *P. gregata*, *P. litoralis*, *P. thermophila*) and seven designated phenotypic taxa (*P. taxon forestsoil*, *P. taxon hungarica*, *P. taxon oaksoil*, *P. taxon paludosa*, *P. taxon PgChlamydo*, *P. taxon riversoil*, *P. taxon salixsoil*). *Phytophthora thermophila* and *P. litoralis* are closely related but differ in the ITS region by 10 steps (changes). At a higher level they cluster with *P. gonapodyides*, *P. megasperma* and *P. taxon PgChlamydo* with distances between lineages of 10–24 steps. These five lineages share a common ancestor. *Phytophthora pinifolia* is loosely associated with this cluster of species. The other two new species, *P. gibbosa* and *P. gregata*, are also closely related to each other, but differ by 7–9 steps. Together with *P. taxon raspberry* and a single isolate designated here as *P. taxon paludosa*, they form a highly supported cluster within sub-clade II. Isolates obtained in Australia and previously designated as *P. taxon raspberry* (Brasier et al. 2003a) have identical ITS sequence to *P. gregata*. Additionally, a single isolate from Japan submitted to GenBank



**Fig. 3** Bayesian inference tree based on mitochondrial gene *cox1* sequences showing phylogenetic relationships within *Phytophthora* ITS Clade 6. Numbers above the branches represent posterior probability values based on Bayesian analysis, thickened branches represent a bootstrap support of > 70 % based on parsimony analysis. Sub-clades I–III are indicated on right. *Phytophthora infestans*, *P. multivora* and *P. nicotianae* were used as outgroup taxa (not shown).

as *P. citricola* also resides within *P. gregata*. *Phytophthora* taxon raspberry isolates from Europe, available on GenBank, form a separate lineage 2–3 bp removed from *P. gregata*. Detailed morphological examination of European isolates is required to determine if they represent a sister species to *P. gregata*.

*Phytophthora* taxon salixsoil is basal to sub-clade II. Lineages corresponding to designated taxa from Europe, *P. taxon forestsoil*, *P. taxon hungarica*, *P. taxon riversoil* and *P. taxon oaksoil*, are excluded from the two species clusters described above.

HSP90 characterisation of eight species and nine designated taxa — The HSP90 dataset contained 142 parsimony informative characters (127 within ITS Clade 6) and significant ( $P < 0.01$ ,  $g_1 = -0.51$ ) phylogenetic signal compared to 1 000 random trees. Heuristic searches resulted in 32 most parsimonious trees of 328 steps (CI = 0.57, RI = 0.83). Bayesian analysis produced trees with similar topology and is presented here (Fig. 2, TreeBASE 10764). Sub-clade I was divided into the same two clusters observed in the ITS analysis, although in the HSP90 analysis these clusters are more distant. Sub-clade II comprises 10 discrete lineages in two main clusters. *Phytophthora thermophila* and *P. litoralis* group together as do *P. gregata*,

*P. gibbosa* and single isolates each of *P. taxon paludosa* and *P. taxon raspberry*. As in the ITS analysis, *P. megasperma* and *P. gonapodyides* group together; however, due to the inclusion of only a single isolate of *P. gonapodyides*, this cluster is not fully resolved. Four sequences submitted to GenBank as *P. gonapodyides* were misidentified and actually correspond to the basal taxa, *P. taxon salixsoil* and *P. taxon PgChlamydo*. Sub-clade III is represented by *P. taxon asparagi*.

*cox1* characterisation of seven species and nine designated taxa — The *cox1* dataset contained 187 parsimony informative characters (172 within ITS Clade 6) and significant ( $P < 0.01$ ,  $g_1 = -0.81$ ) phylogenetic signal compared to 1 000 random trees. Heuristic searches resulted in four most parsimonious trees of 575 steps (CI = 0.49, RI = 0.81). Bayesian analysis produced trees with similar topology and is presented here (Fig. 3; TreeBASE 10764). Sub-clade I was divided into the same two clusters observed in the HSP90 analysis. Sub-clade II comprised 8 lineages. *Phytophthora thermophila* and *P. litoralis* resided in strongly supported terminal clades separated by 54–59 steps. There is greater intraspecific variability in the *cox1* data than in the two nuclear genes with 8 steps variation among isolates of *P. thermophila* and four among isolates of

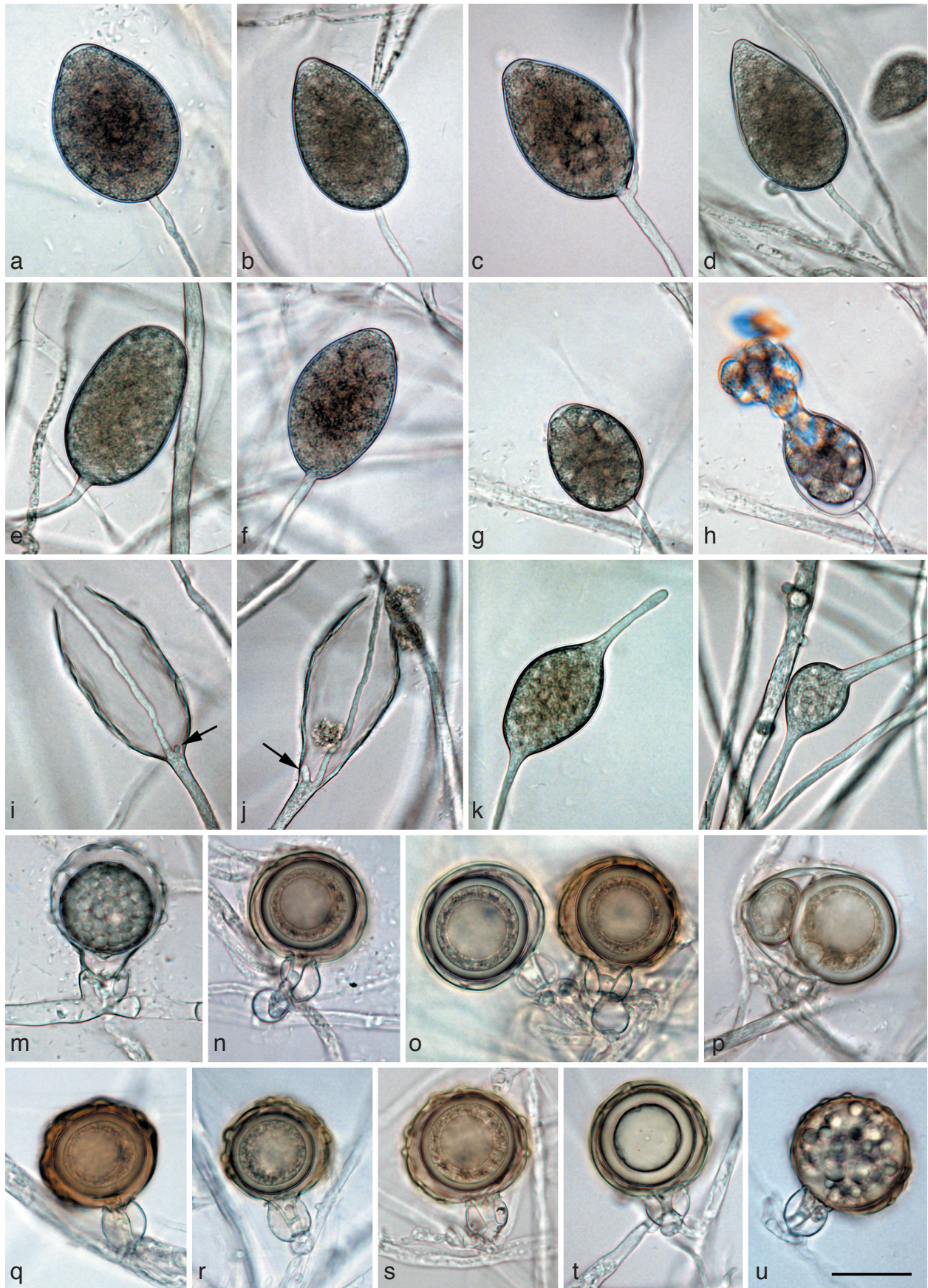
**Table 2** Morphological characters, dimensions and temperature–growth relations of *Phytophthora gibbosa*, *P. gregata*, *P. litoralis*, *P. thermophila*, *P. taxon paludosa*, *P. gonapodyides*, *P. megasperma* and *P. drechsleri*. Characters decisive for species discrimination are highlighted in **bold**.

	<i>P. gibbosa</i>	<i>P. gregata</i>	<i>P. litoralis</i>	<i>P. thermophila</i>	<i>P. taxon paludosa</i>	<i>P. gonapodyides</i>	<i>P. megasperma</i> <sup>1</sup>	<i>P. drechsleri</i>
No. of isolates/source	4	9	5	5	1	1	2	Erwin & Ribeiro (1996)
Sporangia	Ovoid, ellipsoid, limoniform, nonpapillate, <b>some semipapillate</b>	Ovoid, limoniform, obpyriform, nonpapillate	Ovoid, limoniform, nonpapillate	Ovoid, ellipsoid, limoniform, nonpapillate, often with tapering base	Ovoid, limoniform, nonpapillate or <b>semipapillate</b>	Ellipsoid, obpyriform, ovoid, nonpapillate	Ovoid, obpyriform, nonpapillate	Obpyriform, ovoid or elongated, nonpapillate, often with tapering base
l × b mean (µm)	48.8 ± 9.6 × 30.8 ± 5.4	51.0 ± 13.8 × 30.5 ± 5.9	43.6 ± 7.7 × 29.4 ± 5.4	44.8 ± 6.3 × 25.7 ± 3.9	54.7 ± 4.3 × 43.3 ± 3.7	53.7 × 34.3 <sup>2</sup>	59.3 ± 8.8 × 42.8 ± 4.5	52 × 28
Total range (µm)	24.8–71.1 × 17.4–48.0	25.7–102.3 × 14.8–60.7	27.8–76.9 × 16.0–40.4	29.0–64.8 × 15.6–39.3	43.1–64.6 × 31.4–51.7	48–64 × 25–40	37–84 × 35–66	40–71 × 22–34
Isolate means (µm)	44.8–52.2 × 27.9–33.0	37.3–72.7 × 25.6–35.0	39.7–53.4 × 27.1–33.0	44.2–46.8 × 24.1–26.6	–	–	–	–
l/b ratio	1.58 ± 0.15	1.67 ± 0.32	1.51 ± 0.26	1.78 ± 0.26	<b>1.27 ± 0.09</b>	1.43	<b>1.39 ± 0.2</b>	1.9 ± 0.3
Isolate means	1.57–1.60	1.37–2.19	1.35–1.73	1.67–1.86	–	1.48–2.06	1.34–1.40	–
Exit pores								
Width (µm)	12.7 ± 3.5	10.7 ± 2.7	11.9 ± 2.7	13.9 ± 2.9	10.6 ± 2.1	na	12.4 ± 1.2	na
Isolate means (µm)	10.0–14.6	8.4–14.1	10.0–14.4	9.7–16.4	–	na	11.8–12.4	na
Proliferation	Internal extended and external, <b>never nested</b>	Internal extended and nested, <b>never external</b> , <b>sporangiochore partly branching inside empty sporangium</b>	Internal nested and extended and external, <b>secondary lateral sporangia</b>	Internal extended and nested, <b>never external</b> , <b>sporangiochore may branch inside or outside empty sporangium</b>	Internal extended and external, <b>never nested</b>	Internal nested and extended & external	Internal nested and extended, <b>never external</b>	Internal nested and extended
Hypal swellings	Subglobose, elongated <sup>1</sup> , <b>never catenulate</b>	Globose, elongated, angular, partly catenulate	Globose, elongated, angular, partly catenulate	Globose or elongated, partly catenulate	Globose or elongated, partly catenulate	Globose or elongated, only in some isolates	Globose or angular, catenulate or clustered	Globose or angular, catenulate or clustered
Mean diam (µm)	18.7 ± 5.0	14.8 ± 3.8	15.7 ± 4.7	12.6 ± 2.3	16.2 ± 3.1	25.4 ± 1.8	na	na
Hypal aggregations	–	<b>Abundant, up to 170 µm</b>	–	Rare, small	–	–	–	–
Chlamydozoospores	–	–	<b>Globose, radiating hyphae; in 1 isolate</b>	Globose, radiating hyphae; in 3 isolates	–	–	–	Globose, only in some isolates
Mean diam (µm)	–	–	34.3 ± 5.3	41.5 ± 14.7	–	–	–	7.9(4–11)
Sexual system	<b>Homothallic</b>	<b>Homothallic or partially or sporadically self fertile</b>	<b>Sterile or selfsterile silent A1</b>	<b>Sterile or potentially self fertile (1 isolate selfing in soil filtrate)</b>	<b>Homothallic</b>	Sterile, silent A1	Homothallic	Heterothallic
Oogonia	c. 50 % ornamented	Smooth	Smooth	Smooth	Smooth	Smooth	Smooth	Smooth
Mean diam (µm)	<b>38.1 ± 5.4</b>	36.8 ± 4.1	–	31.1 ± 2.5	33.3 ± 3.5	–	41.8 ± 2.4 <sup>3</sup>	33
Total range (µm)	27.0–49.9	23.9–50.9	–	27.2–38.0	24.4–40.7	–	27–52 <sup>3</sup>	28–38
Isolate means (µm)	36.6–39.7	34.0–39.8	–	–	–	–	–	–
Oospores	Always aplerotic	Usually aplerotic	–	Highly aplerotic	Always aplerotic	–	Usually aplerotic	Plerotic
Mean diam (µm)	31.4 ± 4.6	31.6 ± 4.0	–	23.6 ± 2.2	28.1 ± 2.8	–	33.8 ± 2.4 <sup>3</sup>	28
Total range (µm)	18.9–39.4	21.4–45.3	–	20.4–29.7	21.7–34.3	–	23–42 <sup>3</sup>	16–37
Isolate means (µm)	30.0–33.0	27.8–35.5	–	–	–	–	–	–
Wall thickness (µm)	<b>3.17 ± 0.69</b>	2.65 ± 0.81	–	2.3 ± 0.7	2.5 ± 0.4	–	3.31 ± 0.4	3.0
Oospore wall index	0.49 ± 0.06	0.42 ± 0.09	–	0.46 ± 0.09	0.44 ± 0.04	–	0.46 ± 0.06	na
Abortion rate of isolates	16–37 %	<b>52.5–99.8 %</b>	–	< 10 %	< 10 %	–	< 10 %	na
Antheridia	Amphigynous	Predominantly paragynous	Paragynous	Paragynous	Predominantly paragynous	Paragynous and amphigynous	Paragynous and amphigynous	Amphigynous
l × b mean (µm)	13.6 ± 2.4 × 14.0 ± 2.0	17.1 ± 3.0 × 11.0 ± 1.8	–	15.5 ± 2.4 × 9.3 ± 0.9	16.6 ± 3.7 × 13.0 ± 1.5	–	13 ± 1.5 × 10.4 ± 1.3	14–15 × 13
Total range (µm)	10.6–24.9 × 7.6–17.8	10.6–24.9 × 7.6–17.8	–	11.1–20.9 × 7.6–16.6	8.1–23.8 × 10.6–15.0	–	10.7–15.8 × 8.1–13	na
Maximum temperature (°C)	32.5–< 35	32.5–< 35	32.5–< 35	<b>35–&lt; 37</b>	32.5	30–< 35	32.5	35–37
Optimum temperature (°C)	30	<b>25</b> (1 isolate 30)	30	<b>32.5</b>	<b>25</b>	25–30	22.5–25	25–30
Growth rate on V8A at optimum (mm/d)	6.3 ± 0.3	6.5 ± 0.7	4.6 ± 0.3	4.8 ± 0.6	6.3	na	6.7 ± 0.1	na
Growth rate on V8A at 20 °C (mm/d)	5.2 ± 0.1	5.2 ± 0.6	4.0 ± 0.3	<b>3.2 ± 0.5</b>	5.5	na	6.6 ± 0.1	na

<sup>1</sup> measurements were made on isolates available in the current study and the ranges are in agreement with those reported from numerous studies by Erwin & Ribeiro (1996).

<sup>2</sup> data of the British *P. gonapodyides* group. <sup>3</sup> data only from isolate VHS 17183 since isolate DDS 3432 did not produce oogonia in single culture.





**Fig. 4** Morphological structures of *Phytophthora gibbosa*. a–l. Structures formed on V8 agar flooded with soil extract: a, b. ovoid semipapillate sporangia; c. ovoid semipapillate sporangium with external proliferation; d. obpyriform sporangium with nonpapillate pointed apex; e. nonpapillate ellipsoid sporangium; f. ovoid slightly excentric sporangium; g. ovoid sporangium with swollen apex shortly before release of the already differentiated zoospores; h. same sporangium as in g releasing zoospores; i, j. empty elongated ovoid and limoniform sporangium, respectively, showing both internal extended proliferation and formation of an additional basal undeveloped sporangiophore (arrows); k, l. intercalary hyphal swellings originating from undeveloped sporangia that did not form a basal septum and continued to grow at their apex; m. immature ornamented oogonium with aplerotic oospore and amphigynous intercalary antheridium; n–s. mature often bronze-brown oogonia with amphigynous antheridia and thick-walled aplerotic oospores each containing a large ooplast: n. smooth-walled; o, q–s. ornamented gibbose oogonia; p. excentric smooth-walled oogonium with two oospores; t. gibbose oogonium with thickwalled aborted oospore; u. gibbose golden-brown oogonium aborted before oospore formation. — Scale bar = 25  $\mu$ m.

*P. litoralis*. As in the ITS analysis they form a cluster with *P. megasperma* and *P. gonapodyides* and these four species appear to share a common ancestor.

The *cox1* sequence data of all *P. gibbosa* isolates is identical, however large intraspecific variability (22 steps) was observed among isolates designated as *P. gregata* based on nuclear gene sequence. A European isolate of *P. taxon* raspberry clustered with *P. gregata* in the *cox1* analysis. As with the nuclear gene regions, *P. taxon* paludosa together with *P. gibbosa* and *P. gregata* formed a strongly supported cluster indicating a common ancestor. In this analysis, *P. taxon* PgChlamydo is basal to sub-clade II and *P. taxon* salixsoil is basal to sub-clades I and II. Sub-clade III is represented by *P. taxon* asparagi.

## TAXONOMY

Morphological and physiological characters and measurements of the five new *Phytophthora* taxa and related species are given in the comprehensive Table 2.

***Phytophthora gibbosa*** T. Jung, M.J.C. Stukely & T.I. Burgess, *sp. nov.* — MycoBank MB518763; Fig. 4

Systema sexus homothallica; oogonia terminalia vel lateralia, in medio 40 µm (28–48 µm), globosa, subglobosa vel rare excentrica, in medio 31 % abortiva, paries saepe gibbosi et maturitate frequenter pigmentati aureo-fusci ad aerei, rare cum dui oosporis. Oosporae apertoticae, in medio 32 µm (24–38 µm), paries in medio 3.2 µm (1.9–4.3 µm). Antheridia singulares, terminalia, lateralia vel interdum intercalaria, unicellularia, hyalina, globosa ad cylindrica, in medio 14 × 14 µm (11–16 × 11–17 µm), semper amphigynosa. Sporangio-phora simplicia vel rare ramosa sympodiis laxis. Sporangia abundantia in cultura liquida, terminalia, nonpapillata vel interdum semipapillata, ovoidea, ellipsoidea vel limoniformia, in medio 52 × 33 µm (31–71 × 20–45 µm), ratio longitudo ad altitudinem in medio 1.6 (1.3–2.1). Proliferationes sporangiorum semper internae et exterae, numquam niduformis vel externae. Inflationes hypharum subglobosae ad elongatae, numquam catenulatae. Chlamydosporae non observatae. Temperatura crescentiae in agaro 'V8A', optima 30 °C et maxima 33–< 35 °C. Coloniae in agaro 'V8A' uniformes et pubescentes. Regiones 'rDNA ITS', 'cox1' et 'HSP90' cum unica sequentia (GenBank HQ012933, HQ012846, HQ012892).

*Etymology.* Name refers to the gibbous ornamented surface of the oogonia (*gibbosa* Latin = gibbous, knaggy).

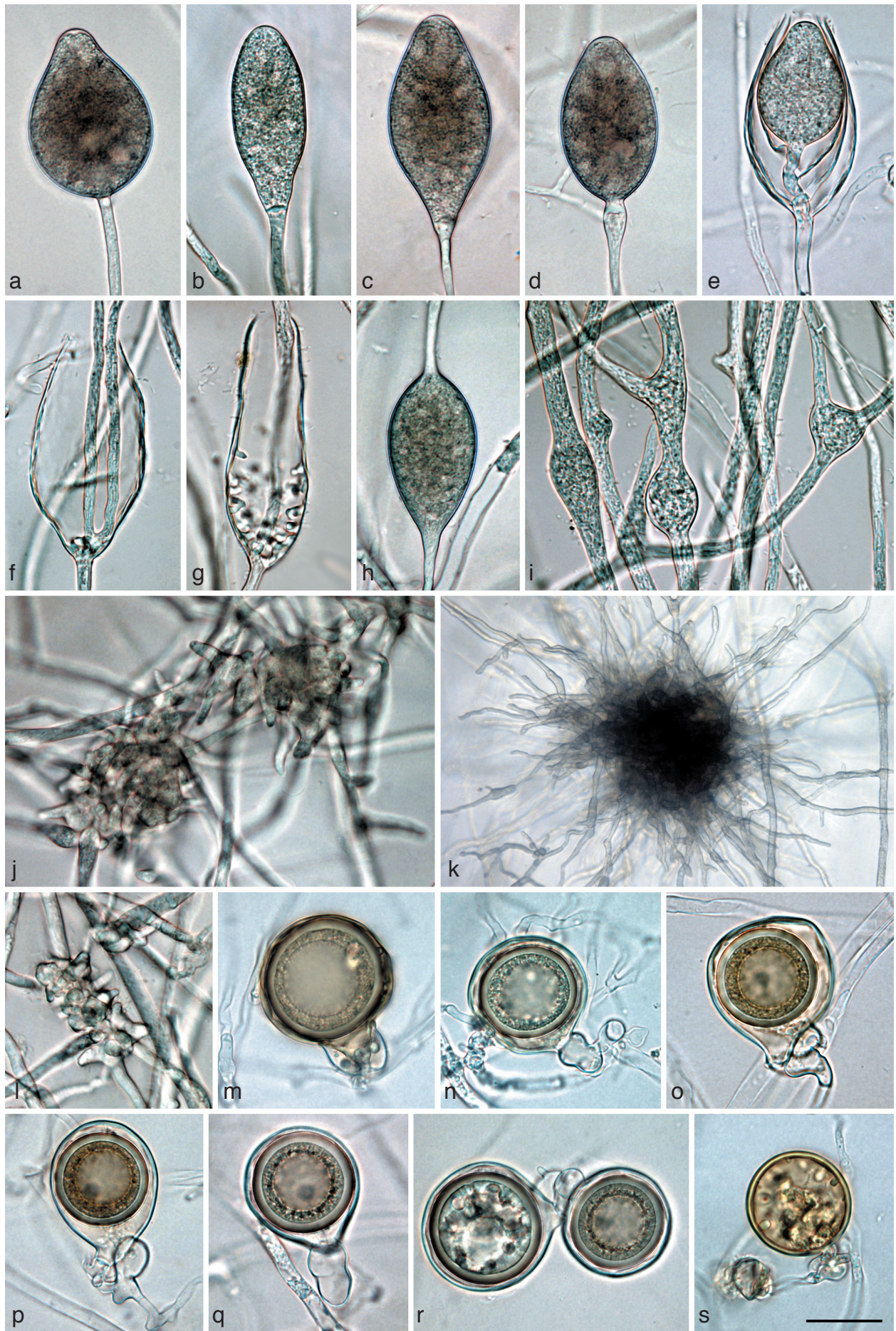
Sporangia and hyphal swellings (Fig. 4a–l) — Sporangia of *P. gibbosa* were not observed on solid agar but were produced abundantly in non-sterile soil extract. Sporangia were typically borne terminally on unbranched sporangiophores, less frequently in lax sympodia. Sporangia were non-caducous, semipapillate (Fig. 4a–c) or more often nonpapillate (Fig. 4d–f), usually with a flat apex (Fig. 4a–c, e–g), sometimes with a pointed apex (Fig. 4d). Sporangial shapes ranged from ovoid (59 %; Fig. 4a–c, f–h) to elongated ovoid (18 %; Fig. 4i), ellipsoid (11 %; Fig. 4e), limoniform (10 %; Fig. 4j) or less frequently pyriform or obpyriform (1 %; Fig. 4d). Sporangia proliferated either externally (Fig. 4c) or internally in an extended way, often with the formation of an

additional basal sporangiophore initial which usually remained short (Fig. 4i, j). Nested proliferation was never observed. Zoospores of *P. gibbosa* were discharged directly through an exit pore 7.7–21.7 µm wide (av. 12.7 ± 3.5 µm) (Fig. 4h). They were limoniform to reniform whilst motile, becoming spherical (av. diam = 12.9 ± 1.6 µm) on encystment. Cysts usually germinated by forming a hypha (98 %). Diplanetism (= germination of cysts by release of a secondary zoospore) or formation of a microsporangium was rarely observed in one isolate (VHS22008). Sporangial dimensions of four isolates of *P. gibbosa* averaged 48.8 ± 9.6 × 30.8 ± 5.4 µm (overall range 24.8–71.1 × 17.4–48.0 µm) with a range of isolate means of 44.8–52.2 × 27.9–33.0 µm. The length/breadth ratio averaged 1.58 ± 0.15. In liquid culture, hyphal swellings were regularly formed which, according to their morphology, were most likely undeveloped sporangia which had failed to form a basal septum and continued to grow at their apex (Fig. 4k, l). Small, globose hyphal swellings along sporangiophores were only rarely observed.

Oogonia, oospores and antheridia (Fig. 4m–u) — Gametangia were readily produced in single culture by all isolates of *P. gibbosa* on V8A within 4 d. Oogonia were borne terminally or laterally, had either wavy edged to ornamented gibbous (32–68 %, on av. 53 %; Fig. 4m, q–u) or smooth walls (Fig. 4n–p) and were usually globose, subglobose or slightly excentric. In all isolates oogonial walls often turned golden-brown to bronze-brown (Fig. 4n, o, q–s, u) while ageing. Oogonial diameters averaged 38.1 ± 5.4 µm (overall range 27.0–49.9 µm and range of isolate means 36.6–39.7 µm). Oospores had a mean diameter of 31.4 ± 4.6 µm (total range 18.9–39.4 µm), were always apertotic, usually globose and contained a large ooplast (Fig. 4m–t). The oospores were relatively thick-walled (3.17 ± 0.69 µm; total range 1.2–5.1 µm), with a mean oospore wall index of 0.49 ± 0.06. On average 30 % (16–37 %) of the oogonia aborted either before (Fig. 4u) or after oospore formation (Fig. 4t). Some oogonia contained two oospores of unequal sizes (Fig. 4p). The antheridia were exclusively amphigynous, averaging 13.6 ± 2.4 × 14.0 ± 2.0 µm, with shapes ranging from subglobose to cylindrical or irregular (Fig. 4m–u). They were usually formed terminally or laterally, and were rarely intercalary (Fig. 4m).

Colony morphology, growth rates and cardinal temperatures (Fig. 9, 11) — All four *P. gibbosa* isolates formed similar uniform colonies on the four different types of media (Fig. 9). Colonies on V8A and MEA had limited aerial mycelium, while colonies on PDA appeared woolly and growth on CMA was mostly submerged with very sparse aerial mycelium. All isolates had identical cardinal temperatures and similar growth rates at all temperatures. The temperature–growth relations are shown in Fig. 11. The maximum growth temperature was between 32.5 and 35 °C. All isolates were unable to grow at 35 °C, and isolates did not resume growth when plates incubated for 7 d at 35 °C were transferred to 20 °C. The average radial growth rate on V8A at the optimum temperature of 30 °C was 6.3 ± 0.3 mm/d. At 20 °C mean growth rates on V8A, MEA, CMA and PDA were 5.2, 5.9, 6.0 and 5.0 mm/d, respectively.

**Fig. 5** Morphological structures of *Phytophthora gregata*. a–l. Structures formed on V8 agar flooded with soil extract: a. obpyriform nonpapillate sporangium; b. elongated-ellipsoid nonpapillate sporangium with a tapering base; c. limoniform nonpapillate sporangium; d. ovoid nonpapillate sporangium with a conspicuous basal plug and a widening of the sporangiophore towards the sporangial base; e. ovoid empty sporangium with internal nested proliferation, and obpyriform young sporangium; f. ovoid empty sporangium with internal extended proliferation via two sporangiophores; g. elongated sporangium with internal extended proliferation and a multitude of wall ingrowths; h. hyphal swellings formed from undeveloped sporangium that did not form a basal septum and continued to grow at the apex; i. elongated and angular hyphal swellings; j–l. hyphal aggregations; m–q. mature oogonia containing a thick-walled oospore with a large ooplast and a nucleus; m, n. oogonia produced by selfing of *P. gregata* isolates MUCC760 (m) and CBS127952 (n) in paired cultures with *P. cinnamomi* mating type A2 isolate DCE60; m. with nearly plerotic oospore and amphigynous antheridium; n. with apertotic oospore and paragynous antheridium; o, p. oogonia produced by selfing in paired cultures of *P. gregata* isolates MJS235 and MUCC760; o. excentric with tapering base, apertotic oospore and paragynous antheridium; p. elongated with apertotic oospore and paragynous antheridium; q. with nearly plerotic oospore and paragynous antheridium, produced by isolate VHS21961 in paired culture with *Trichoderma reesei*; r. mature oogonia with thick-walled aborted (left) and viable oospore (right) produced by isolate VHS21961 in paired culture with *T. reesei*; s. golden-brown aborted oogonium produced by selfing of isolate CBS127952 in paired culture with *P. cinnamomi* mating type A2 isolate DCE60. — Scale bar = 25 µm for all except (k) where scale bar = 50 µm.



*Specimens examined.* WESTERN AUSTRALIA, Scott River ironstones, from rhizosphere soil of dying *Acacia pycnantha*, 2009, VHS, holotype MURU 461 (dried culture on V8A, Herbarium of Murdoch University, Western Australia), cultures ex-type CBS127951 and VHS21998; Scott River ironstones, from rhizosphere soil of dying *Xanthorrhoea gracilis*, 2009, VHS, VHS21999; Scott River ironstones, from rhizosphere soil of dying *Acacia pycnantha*, 2009, VHS, VHS22007; Scott River ironstones, from rhizosphere soil of dying *Grevillea* sp., 2009, VHS, VHS22008.

***Phytophthora gregata* T. Jung, M.J.C. Stukely & T.I. Burgess, sp. nov. — MycoBank MB518764; Fig. 5**

Systema sexus homothallica, solum partim functionalis; oogonia in medio 95 % abortiva, terminalia vel lateral, in medio 38  $\mu\text{m}$  (27–45  $\mu\text{m}$ ), globosa, subglobosa vel rare excentrica, paries semper levigati. Oosporae apertoticae, in medio 32  $\mu\text{m}$  (23–39  $\mu\text{m}$ ), paries in medio 3.0  $\mu\text{m}$  (1.3–4.4  $\mu\text{m}$ ) maturitate frequenter pigmentati lutei ad luteifusci. Antheridia singulares, terminalia vel lateral, unicellularia, hyalina, claviformes, subglobosa vel cylindrica, in medio 19  $\times$  11  $\mu\text{m}$  (14–25  $\times$  9–13  $\mu\text{m}$ ), paragynosa vel interdum amphigynosa. Sporangiohora simplicia. Sporangia abundantia in cultura liquida, terminalia, nonpapillata cum apicibus applanatis, ovoidea, limoniformia vel obpyriformia, frequenter cum basim attenuato, interdum cum multis intrusionibus parierum, in medio 50  $\times$  31  $\mu\text{m}$  (31–68  $\times$  17–43  $\mu\text{m}$ ), ratio longitudo ad altitudinem in medio 1.6 (1.3–2.2). Proliferationes sporangiorum semper internae, niduformes et extantae, numquam externae. Aggregationes hypharum frequenter in agar 'V8A' et in cultura liquida, diameter 15–170  $\mu\text{m}$ . Inflationes hypharum subglobosae, angulares vel elongatae, partim catenulatae. Chlamydosporae non observatae. Temperatura crescentiae in agar 'V8A', optima 25 °C et maxima 33–35 °C. Coloniae in agar 'V8A' striatae cum mycelio aereo restricto. Regiones 'rDNA ITS', 'cox1' et 'HSP' cum unica sequentia (GenBank HQ012942, HQ012858, HQ012904).

*Etymology.* Name refers to the abundant hyphal aggregations regularly formed by all isolates (gregata Latin = aggregated, in clumps).

Sporangia, hyphal swellings and aggregations (Fig. 5a–l) — Sporangia of *P. gregata* were not observed on solid agar but were produced abundantly in non-sterile soil extract. Sporangia were usually borne terminally on unbranched sporangiophores, often in chains of internally proliferating sporangia, or much less frequently in lax sympodia. Sporangia were non-caducous, nonpapillate and usually with a flat apex (Fig. 5a–e). Sporangial shapes ranged from ovoid (81 %; Fig. 5d, e) to elongated ovoid (4 %; Fig. 5f, g), limoniform (10 %; Fig. 5c) or less frequently ellipsoid (Fig. 5b), pyriform or obpyriform (Fig. 5a, e). Sporangia usually proliferated internally in both a nested (Fig. 5e) and extended way (Fig. 5f, g), often with the formation of an additional basal

sporangiophore initial, which usually remained short, but sometimes developed into a second sporangiophore (Fig. 5f). External proliferation was not observed. In all isolates some sporangia formed numerous wall ingrowths which became clearly visible after zoospore release (Fig. 5g). Zoospores were discharged through an exit pore 5.1–18.7  $\mu\text{m}$  wide (av. 10.7  $\pm$  2.7  $\mu\text{m}$ ) (Fig. 5e–g). They were limoniform to reniform whilst motile, becoming spherical (av. diam = 13.1  $\pm$  1.6  $\mu\text{m}$ ) on encystment. Cysts usually germinated by forming a hypha, but both diplanetism and formation of a microsporangium was also observed in all isolates. Sporangial dimensions of nine isolates averaged 51.0  $\pm$  13.8  $\times$  30.5  $\pm$  5.9  $\mu\text{m}$  (overall range 25.7–102.3  $\times$  14.8–50.7  $\mu\text{m}$ ) with a wide range of isolate means of 37.3–72.7  $\times$  25.6–35.0  $\mu\text{m}$ . The length/breadth ratio averaged 1.67  $\pm$  0.32 with a wide range of isolate means of 1.37–2.19. As with *P. gibbosa*, hyphal swellings were formed, which according to their morphology were most likely undeveloped sporangia which, when failing to form a basal septum, continued to grow at their apex (Fig. 5h). In addition, globose, angular or irregular-elongated, often catenulate hyphal swellings were regularly formed (Fig. 5i). Inside the V8 agar and also in liquid culture, all isolates frequently produced dense hyphal aggregations formed from both clusters of lateral hyphae and the twisting of hyphae around each other (Fig. 5j–l). In rare cases such aggregations resulted from sporangia in the agar inside of which the zoospores had directly germinated (not shown). Aggregations had diameters of 15–170  $\mu\text{m}$ .

Oogonia, oospores and antheridia (Fig. 5m–s) — With the exception of HSA2356, all isolates of *P. gregata* produced oogonia in single culture on V8A. In paired cultures with A1 and A2 tester strains of *P. cambivora*, *P. cinnamomi* and *P. cryptogea*, other *P. gregata* isolates, and *T. reesei*, all isolates including HSA2356 produced oogonia within three to four weeks (Table 3). Interestingly, in most isolates, higher numbers of oogonia were produced in paired cultures than in single culture (Table 3). In most pairings of MUCC760 and VHS9854, in single culture and in one pairing of MJS238, and in a few pairing combinations of CBS127952 and VHS21992, oogonia were predominantly produced in chimaeric patches and/or along the distant margin within the *P. gregata* colonies (Table 3). The average abortion rate was very high with 84.9 % in both single culture and over all pairing combinations (Table 3). However, the abortion rate varied considerably between isolates, and in some isolates

**Table 3** Abundance and spatial distribution of oogonial formation and oogonial abortion rates (%) of *Phytophthora gregata* isolates in single culture and in pairings with other *P. gregata* isolates, A1 and A2 tester strains of *P. cambivora*, *P. cinnamomi* and *P. cryptogea*, and *Trichoderma reesei*.

	MJS235	MJS238	VHS21961	CBS127952	VHS21992	VHS9854	MUCC760	HSA2356	Average
Paired with									
MJS235		+++/99	+++/92	+++/96	+++/2	+/e/100	++/p/98	+/e/100	83.9
MJS238	+++/64		+++/64	++/pe/97	+++/4	++/pe/100	++/e/100	–	71.5
VHS21961	+++/70	++/92		+++/90	+/pe/19	+/100	+/92	–	77.2
CBS127952	+++/78	++/94	+++/56		+++/2	100	+/p/100	–	71.7
VHS21992	+++/84	+++/94	+++/70	+++/92		+/p/100	+/p/92	–	88.7
VHS9854	+++/72	+++/96	+++/60	++/e/94	+++/4		+/p/100	–	71.0
MUCC760	+++/70	+++/98	+++/72	+++/96	+++/68	+/100		–	84.0
HSA2356	+++/62	+++/94	+++/70	+++/96	+++/66	++/100	++/100		84.0
<i>P. cambivora</i> A1	+++/92	+++/99	na	+++/96	+++/84	+/99	+++/99	+/99	95.4
<i>P. cambivora</i> A2	+++/90	++/99	na	+++/94	+++/90	+/99	+++/99	+/100	95.9
<i>P. cinnamomi</i> A1	+++/70	+++/99	na	+++/90	+++/92	+/pe/100	+++/99	+/100	92.9
<i>P. cinnamomi</i> A2	+++/60	+++/98	na	+++/88	+++/82	+/pe/100	+++/90	+/99	88.1
<i>P. cryptogea</i> A1	+++/90	+++/99	na	+++/90	+++/88	+/99	+++/99	+/100	95.0
<i>P. cryptogea</i> A2	+++/66	+++/99	na	+++/90	+++/92	+/pe/100	+++/99	+/100	92.3
<i>Trichoderma</i>	+++/72	++/pe/94	+++/44	+++/100	++/p/34	+/pe/100	+/p/100	–	77.7
Average	74.3	96.7	66.0	93.5	51.9	99.8	97.6	99.7	84.9
Single culture	+++/72	+/pe/100	++/68	++/95	++/60	+/100	+++/99	–	84.9

–, no oogonia formed; +, rare; ++, occasional; +++, frequent; p, chimaeric patches of oogonia; e, oogonia formed along the distant margin of the Petri dish.

also between different pairing combinations. While VHS21992, VHS21961 and MJS235 showed average abortion rates of 60.0–72.0 % in single culture and 51.9–74.3 % over all pairings, the other five isolates produced hardly any viable oospores (Table 3). Within-isolate variation was highest in VHS21992 which had mean abortion rates of 2 and 4 % in paired cultures with MJS235 or CBS127952 and MJS238 or VHS9854, respectively, 60 % in single culture and 82–92 % in pairings with the A1 and A2 tester strains. None of the *P. gregata* isolates induced oogonia formation in the A1 and A2 tester isolates. In conclusion, the breeding system of *P. gregata* is homothallic or partially or sporadically self fertile. Oogonia were borne terminally or laterally and had globose, subglobose to slightly excentric or elongated shapes, often with a tapering base (Fig. 5m, o–q). Oogonial walls sometimes turned golden-brown (Fig. 5m, o–p, s) while ageing. Oogonial diameters averaged  $36.8 \pm 4.1 \mu\text{m}$  (overall range 23.9–50.9  $\mu\text{m}$  and range of isolate means 34.0–39.8  $\mu\text{m}$ ). Oospores were usually aplerotic (Fig. 5n–r) although some plerotic oospores could be observed in all isolates (Fig. 5m). Oospores were globose and contained a large ooplast (Fig. 5m–s). They had a mean diameter of  $31.6 \pm 4.0 \mu\text{m}$  (total range 21.4–45.3  $\mu\text{m}$ ), thick walls (av.  $2.65 \pm 0.81 \mu\text{m}$ , total range 1.0–5.3  $\mu\text{m}$ ) and a mean oospore wall index of  $0.42 \pm 0.09$ . In all isolates, the majority of oogonia aborted, either prior to (Fig. 5s) or after forming an oospore (Fig. 5r). Antheridia were formed terminally or laterally (Fig. 5m) and were predominantly paragynous (Fig. 5n–r), averaging  $17.1 \pm 3.0 \times 11.0 \pm 1.8 \mu\text{m}$ , with shapes ranging from clavate, subglobose to irregular. Amphigynous antheridia (Fig. 5m) could also be observed in most isolates.

Colony morphology, growth rates and cardinal temperatures (Fig. 9, 11) — Colony growth patterns of different isolates of *P. gregata* showed some variation. On V8A and MEA faintly striate, stellate or uniform colonies with sparse to limited aerial mycelium were formed. Colonies on CMA had no distinctive growth pattern and were mostly submerged with no, or very sparse, aerial mycelium while colonies on PDA appeared woolly, sometimes dome-shaped, with a uniform or faintly stellate pattern. Temperature-growth relations are shown in Fig. 11. With the exception of HSA2356, which was consistently slower growing at all temperatures, all eight isolates included in the growth test had similar growth rates at their optimum temperature of 25 °C. All isolates except MUCC760 were unable to grow at 35 °C, and isolates did not resume growth when plates incubated for 7 d at 35 °C were transferred to 20 °C. The maximum growth temperature was between 32.5 and 35 °C. The average radial growth rate at the optimum temperature of 25 °C was  $6.5 \pm 0.7 \text{ mm/d}$ . With mean radial growth rates at 20 °C of 5.2, 5.9, 5.6 and 5.1 mm/d on V8A, MEA, CMA and PDA, respectively, *P. gregata* showed almost no agar media preferences.

*Specimens examined.* WESTERN AUSTRALIA, Busselton, from rhizosphere soil of dying *Patersonia* sp., 2009, VHS, holotype MURU 462 (dried culture on V8A, Herbarium of Murdoch University, Western Australia), cultures ex-type CBS127952 and VHS21962; Scott River ironstones, from rhizosphere soil of dying plants in native forest, 2009, VHS, VHS21992; Lancelin, from rhizosphere soil of dying *X. preissii*, 2001, VHS, VHS9854; Busselton, from rhizosphere soil of dying *Hakea* sp., 2009, VHS, VHS21961; Busselton, from rhizosphere soil of dying *Patersonia* sp., 2009, VHS, VHS21962; Nannup, from rhizosphere soil of dying *Pinus radiata*, 1982, M.J.C. Stukely, MJS235; Nannup, from root collar of dying *P. radiata*, 1981, M.J.C. Stukely, MJS238; Byford, from soil, 1965, *not known*, DCE68; Cataby, from root of *B. prionotes*, 1996, R. Hart, HSA 2356. — VICTORIA, Devlin's Bridge, soil from pasture, 2008, W.A. Dunstan, MUCC760.

***Phytophthora litoralis*** T. Jung, M.J.C. Stukely & T.I. Burgess, *sp. nov.* — MycoBank MB518765; Fig. 6

Systema sexus sterilis. Sporangiophora simplicia vel ramosa in sympodiis laxis. Sporangia abundantia in cultura liquida, terminalia vel lateralia, non-

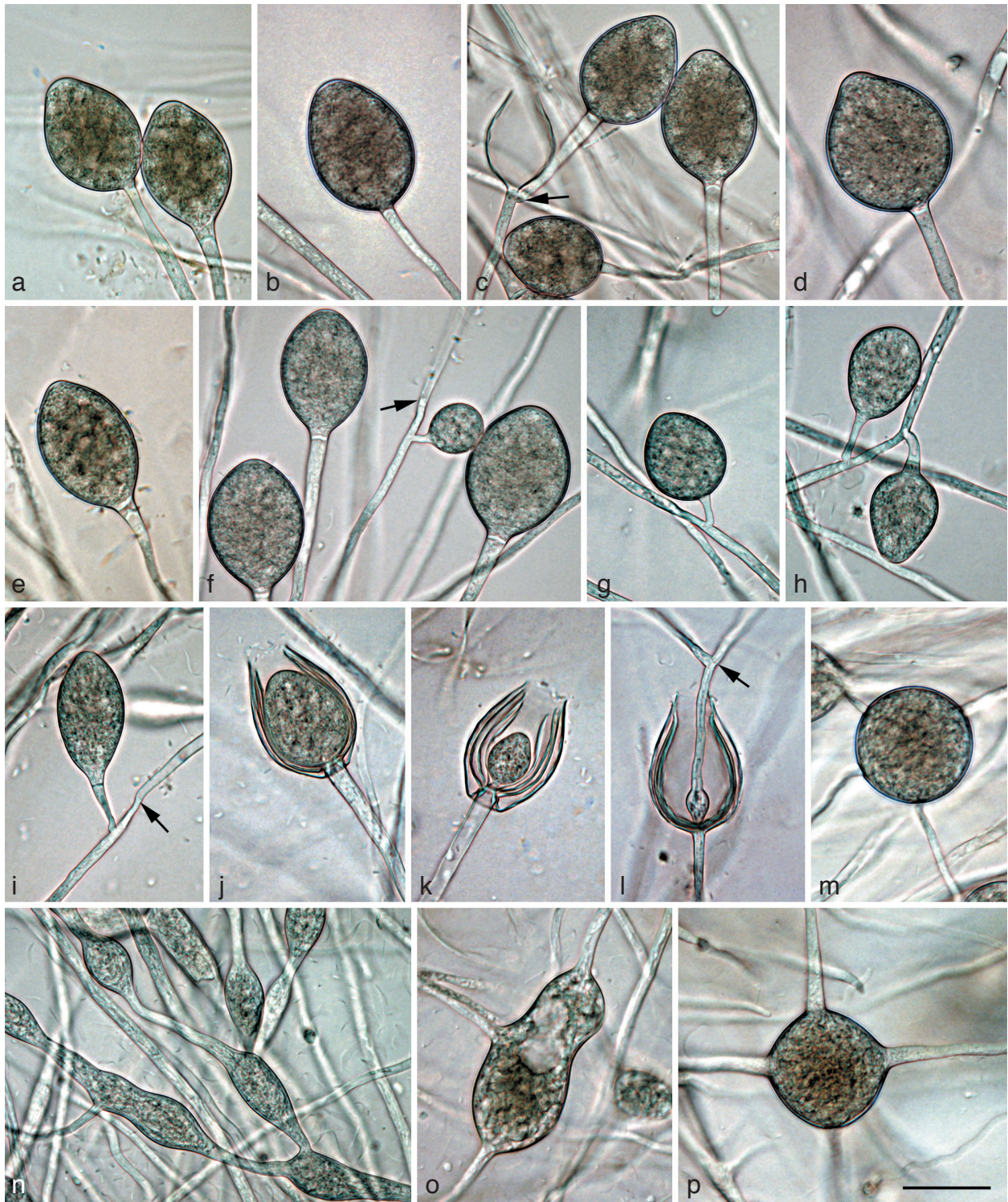
papillata, ovoidea vel limoniformia, in medio  $41 \times 27 \mu\text{m}$  ( $30\text{--}49 \times 16\text{--}34 \mu\text{m}$ ), ratio longitudo ad altitudinem in medio 1.5 (1.2–2.2). Proliferationes sporangiorum internae, niduformae et extensae, et externae. Inflationes hypharum globosae, angulares, irregulares vel elongatae, partim catenulatae et partim cum hyphis radiatis. Chlamydosporae globosae, partim cum hyphis radiatis observatae. Temperaturaes crescentiae in agaro 'V8A', optima 30 °C et maxima 33–35 °C. Coloniae in agaro 'V8A' stellatae cum mycelio aereo restricto. Regiones 'rDNAITS', 'cox1' et 'HSP' cum unica sequentia (GenBank HQ012948, HQ012866, HQ012911).

*Etymology.* Name refers to the frequent association of this species with coastal and riparian vegetation and the littoral zone of water bodies (litus Latin = coast and bank).

Sporangia and hyphal swellings (Fig. 6a–l, n–p) — Sporangia of *P. litoralis* were not formed on solid agar but were produced abundantly in non-sterile soil extract. Sporangia were non-caducous and nonpapillate (Fig. 6a–k). Sporangial shapes ranged from ovoid and elongated ovoid (85.6 %; Fig. 6a–d, g, h, j–l), limoniform (10 %; Fig. 6a, c, e, f, i) to less frequently ellipsoid, pyriform, obpyriform, obovoid (Fig. 6h) or ampulliform. Mean sporangial dimensions of five isolates were  $43.6 \pm 7.7 \times 29.4 \pm 5.4 \mu\text{m}$  (overall range 27.8–76.9  $\times$  16.0–40.4  $\mu\text{m}$ ) with a range of isolate means of 39.7–53.4  $\times$  27.1–33.0  $\mu\text{m}$ . The length/breadth ratio averaged  $1.51 \pm 0.26$  with a range of isolate means of 1.35–1.73. Sporangia were borne terminally on unbranched sporangiophores, often in chains of internally proliferating sporangia, or in lax sympodia. In addition, secondary lateral sporangia were sometimes formed from the cytoplasm remaining in the sporangiophore after the formation of the primary terminal sporangium (Fig. 6f–i). These lateral sporangia were usually considerably smaller than the primary terminal sporangia (Fig. 6f–h). Sporangia usually proliferated internally in both a nested (Fig. 6j–l) and extended way (Fig. 6l). Sometimes, internally proliferating sporangiophores branched just after having passed through the exit pore of the empty primary sporangium (Fig. 6l). The formation of a second basal sporangiophore initial inside internally proliferating sporangia was much less frequent than in *P. gregata*, and these initials always remained short. External proliferation was also common (Fig. 6c). Zoospores of *P. litoralis* were discharged through an exit pore 6.2–19.5  $\mu\text{m}$  wide (av.  $11.9 \pm 2.7 \mu\text{m}$ ). They were limoniform to reniform whilst motile, becoming spherical (av. diam =  $10.8 \pm 1.3 \mu\text{m}$ ) on encystment. Cysts usually germinated by forming a hypha but diplanetism or formation of a microsporangium was also common in all isolates. Only one of the five isolates (VHS17085) produced globose chlamydospores, which averaged  $34.3 \pm 5.3 \mu\text{m}$  diam and sometimes had radiating hyphae (Fig. 6m). Globose, angular or irregular-elongated, often catenulate hyphal swellings with an average diameter of  $15.7 \pm 4.7 \mu\text{m}$  were regularly formed (Fig. 6n–p); often forming branching points (Fig. 6n–o), sometimes with radiating hyphae (Fig. 6p).

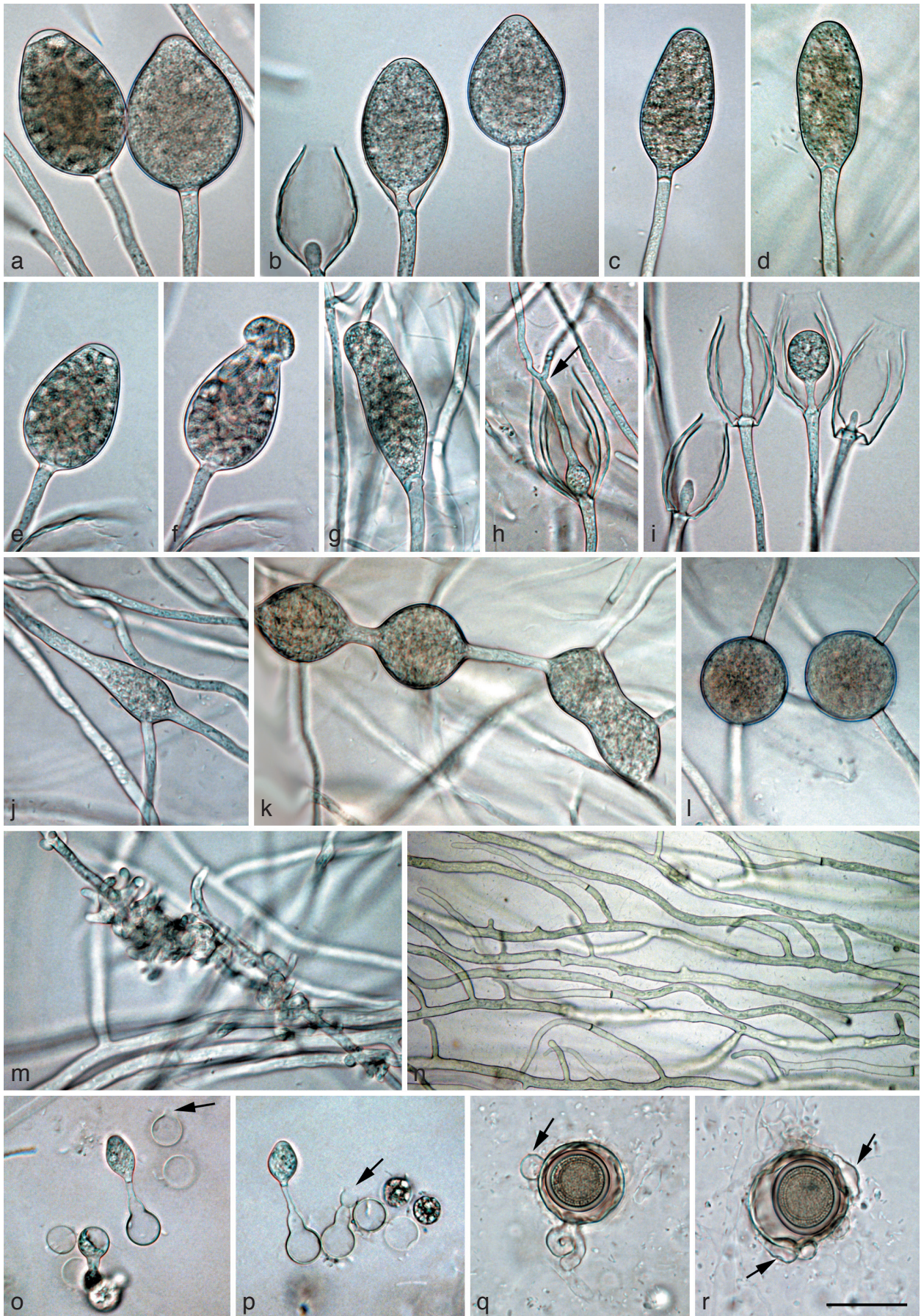
Oogonia, oospores and antheridia — None of the five *P. litoralis* isolates tested produced gametangia in single culture or when paired with other *P. litoralis* isolates, with A1 and A2 tester strains of *P. cambivora*, *P. cinnamomi* and *P. cryptogea* or with *T. reesei*. However, two of the isolates (MUCC762 and VHS17085) stimulated the formation of oogonia in the A2 isolate of *P. cinnamomi*. These oogonia were clearly formed by selfing of *P. cinnamomi* because they were found within the *P. cinnamomi* colony close to the distant margin of the petridish and sometimes had 2–7 paragynous antheridia attached (Hüberli et al. 2001). In conclusion, the sexual system of *P. litoralis* can be either fully sterile or self-sterile silent A1.

Colony morphology, growth rates and cardinal temperatures (Fig. 10, 11) — All four *P. litoralis* isolates examined formed stellate colonies with sparse aerial mycelium on V8A; petaloid, adpressed to submerged colonies on MEA and submerged uniform colonies on CMA (Fig. 10). Colonies on PDA were either petaloid with limited aerial mycelium or irregular and



**Fig. 6** Morphological structures of *Phytophthora litoralis* formed on V8 agar flooded with soil extract. a–l. Nonpapillate sporangia: a. ovoid (left) and limoniform (right); b. ovoid; c. ovoid sporangia and external proliferation (arrow); d. broadly-ovoid; e. limoniform with a conspicuous basal plug; f. ovoid to limoniform terminal sporangia and secondary lateral sporangium formed with cytoplasm remaining after the formation of the terminal primary sporangium (arrow); g. secondary, lateral ovoid sporangium; h. secondary, lateral, obovoid (left) and ovoid (right) sporangia; i. secondary, lateral limoniform sporangium formed just below the empty upper section of the sporangiophore (arrow); j, k. empty sporangia with internal nested proliferation; l. empty sporangium with internal nested and extended proliferation and branching of the sporangiophore outside the sporangium (arrow); m. chlamydospore with radiating hyphae; n. irregular catenulate hyphal swellings; o. irregular hyphal swelling with radiating hyphae; p. globose hyphal swelling with radiating hyphae. — Scale bar = 25  $\mu$ m.

**Fig. 7** Morphological structures of *Phytophthora thermophila* formed on V8 agar flooded with soil extract. a–g. Nonpapillate mature sporangia: a. ovoid, the left one with swollen apex shortly before release of the already differentiated zoospores; b. ovoid (right), ellipsoid formed by nested proliferation (centre), and ovoid empty with internal nested proliferation beginning (left); c. elongated-ovoid with a tapering base and a conspicuous basal plug; d. elongated-ellipsoid with a tapering base; e. ovoid sporangium with swollen apex shortly before release of the already differentiated zoospores; f. same sporangium as in e releasing zoospores; g. elongated, cigar-like with a conspicuous basal plug; h. empty sporangium with nested and extended proliferation, and branching of the sporangiophore (arrow) after leaving the sporangium; i. empty sporangia with internal nested and extended proliferation; j. elongated hyphal swelling at a branching point; k. catenulate globose and irregular hyphal swellings; the right one forming a branching point; l. intercalary globose chlamydospores, distinguished from



hyphal swellings by having thicker walls and septa to the bearing hyphae; m. hyphal aggregation; n. undulating main hyphae; o, p. encysted zoospores having geminated by forming a secondary microsporangium or by having released a secondary zoospore (diplanetism; arrows); q. mature oogonium with a twisted oogonial stalk, a paragynous antheridium (arrow) and an aplerotic oospore containing a large ooplast; r. mature oogonium with two paragynous antheridia (arrows) and an aplerotic oospore containing a large ooplast. — Scale bar = 25 µm.

dense-felty (Fig. 10). The temperature–growth relations are shown in Fig. 11. All isolates had similar growth rates at all temperatures except MUCC763 which was markedly slower at 32.5 °C than the other isolates, and MUCC762 which showed some growth at 35 °C while all other isolates failed to grow at 35 °C. All isolates resumed growth when plates incubated for 7 d at 35 °C were transferred to 20 °C. However, 37 °C was lethal to all isolates when tested in the same way. Thus, the maximum growth temperature was between 32.5 and 35 °C. The average radial growth rate at the optimum temperature of 30 °C was  $4.6 \pm 0.3$  mm/d. At 20 °C mean radial growth rates on V8A, MEA, CMA and PDA were 4.0, 3.1, 4.1 and 2.4 mm/d, respectively.

*Specimens examined.* WESTERN AUSTRALIA, Ravensthorpe, from rhizosphere soil of dying *Banksia* sp., 2008, VHS, holotype MURU 463 (dried culture on V8A, Herbarium of Murdoch University, Western Australia), cultures ex-type CBS127953 and VHS20763; Hopetoun, from rhizosphere soil of dying *Banksia* sp., 2007, VHS, VHS17085; Wilga, from rhizosphere soil of dying *X. preissii*, 2008, VHS, VHS19173; Borden, stream baiting, 2008, *D. Hüberli*, MUCC762; Borden, stream baiting, 2008, *D. Hüberli*, MUCC763.

***Phytophthora thermophila* T. Jung, M.J.C. Stukely & T.I. Burgess, sp. nov. — MycoBank MB518766; Fig. 7**

Systema sexus sterilis. Sporangiophora simplicia. Sporangia abundantia in cultura liquida, terminalia, nonpapillata, ovoidea, ellipsoidea vel limoniformia, in medio  $46 \times 25$  µm ( $33\text{--}65 \times 17\text{--}39$  µm), ratio longitudo ad altitudinem in medio 1.9 (1.4–2.9). Proliferationes sporangiorum solum internae, niduiformes et extantae. Cystae plerumque germinantes cum una zoospora secundaria vel una microsporangia. Hyphae saepe undulatae. Aggregationes hypharum parvae et solum rare observatae. Inflationes hypharum globosae, angulares, irregulares vel elongatae, partim catenulatae et partim cum hyphis radiatis. Chlamydozporae globosae, partim cum hyphis radiatis, in medio 33 ( $24\text{--}33$  µm). Temperatura crescentiae in agar 'V8A', optima 33 °C et maxima 35 °C. Coloniae in agar 'V8A' stellatae cum mycelio aereo restricto. Regiones 'rDNA ITS', 'cox1' et 'HSP' cum unica sequentia (GenBank EU301155, HQ012872, HQ012916).

*Etymology.* Name refers to the high optimum and maximum temperatures for growth (thermophila Latin = thermophilic).

Sporangia, hyphal swellings and chlamydozporae (Fig. 7a–l) — Sporangia of *P. thermophila* were not observed on solid agar but were produced abundantly in non-sterile soil extract. Sporangia were borne terminally on unbranched sporangiophores, often in chains of internally proliferating sporangia. Due to the lack of external proliferation no sympodia are formed. Sporangia were non-caducous and nonpapillate (Fig. 7a–g). Sporangial shapes ranged from ovoid and elongated ovoid (72.8 %; Fig. 7a–c, e, f, h, i) to limoniform (13.2 %), ellipsoid (12 %; Fig. 7b, d) and, less frequently, pyriform, obpyriform or cylindrical. Features such as a tapering base or a conspicuous basal plug were quite common (Fig. 7c, d, g). Sporangia usually proliferated internally in both a nested and extended way (Fig. 7b, h, i). Branching of internally proliferating sporangiophores just after having passed through the exit pore of the empty primary sporangium occurred regularly (Fig. 7h). The formation of a second basal sporangiophore initial inside internally proliferating sporangia was much less common than in *P. gregata*, and these initials always remained short. External proliferation was never observed. The formation of secondary lateral sporangia was very rare and much less frequent than in *P. litoralis*. Sporangial dimensions of five isolates of *P. thermophila* averaged  $44.8 \pm 6.3 \times 25.7 \pm 3.9$  µm (overall range 29.0–64.8 × 15.6–39.3 µm) with rather similar isolate means ( $44.2\text{--}46.8 \times 24.1\text{--}26.6$  µm). The length/breadth ratio of the sporangia averaged  $1.78 \pm 0.26$  with a range of isolate means of 1.67–1.86. Zoospores were discharged through an exit pore 5.5–20.9 µm wide (av.  $13.9 \pm 2.9$  µm) (Fig. 7f). They were limoniform to reniform whilst motile, becoming spherical (av. diam =  $10.9 \pm 1.1$  µm) on encystment. Cysts germinated more often indirectly by releasing a secondary zoospore (diplanetism) or by

forming a microsporangium (Fig. 7o, p) than directly by forming a hypha. In liquid culture globose, angular or irregular-elongated hyphal swellings, sometimes catenulate and with radiating hyphae or forming branching points, were regularly formed (Fig. 7j, k). Hyphal swellings had a mean diameter of  $12.6 \pm 2.3$  µm. Globose, intercalary or terminal chlamydozporae (Fig. 7l) with a mean diameter of  $41.5 \pm 14.7$  µm were produced in liquid culture by three of the five isolates. Some isolates produced hyphal aggregations much smaller and much less frequent than those of *P. gregata* (Fig. 7m). Main hyphae often showed undulating growth (Fig. 7n).

Oogonia, oospores and antheridia — None of the five *P. thermophila* isolates tested produced gametangia in single culture or when paired with other *P. thermophila* isolates, A1 and A2 tester strains of *P. cambivora*, *P. cinnamomi* and *P. cryptogea* or with *T. reesei*. Also, none of the isolates stimulated the formation of oogonia in the A1 or A2 tester strains. Interestingly, VHS3655 formed oogonia abundantly in single culture when flooded with non-sterile soil filtrate. Shapes of oogonia ranged from globose and subglobose to excentric (Fig. 7q, r). Oogonial stalks were sometimes twisted (Fig. 7q). Oogonial diameters averaged  $31.1 \pm 2.5$  µm with a total range of 27.2–38.0 µm. Most oogonia looked viable, containing oospores with a large ooplast (abortion rate = 8 %) (Fig. 7q, r). Oospores were highly aplerotic and averaged  $23.6 \pm 2.2$  µm diam with relatively thick oospore walls (on av.  $2.3 \pm 0.7$  µm) and a high mean oospore wall index of  $0.46 \pm 0.09$ . The antheridia were exclusively paragynous (Fig. 7q, r) and measured  $15.5 \pm 2.4 \times 9.3 \pm 0.9$  µm. Sometimes two or three antheridia were attached to one oogonium (Fig. 7r). In conclusion, the sexual system of *P. thermophila* is sterile or potentially self fertile when induced by certain stimuli.

Colony morphology, growth rates and cardinal temperatures (Fig. 10, 11) — All four *P. thermophila* isolates examined formed stellate to petaloid colonies with sparse to limited aerial mycelium on V8A and MEA, and uniform submerged colonies on CMA (Fig. 10). Colonies on PDA were irregular and dense-felty (Fig. 10). *Phytophthora thermophila* proved to be a high temperature species with an optimum temperature for growth of 32.5 °C and a maximum temperature for growth of 35–< 37 °C (Fig. 11). All isolates failed to grow at 37 °C, but isolate MUCC764 resumed growth when plates incubated for 7 d at 37 °C were transferred to 20 °C. Of the five newly described species and taxa, *P. thermophila* showed the slowest growth on all four agar media. The average radial growth rate on V8A at the optimum temperature of 32.5 °C was  $4.8 \pm 0.6$  mm/d. With a mean radial growth rate of 3.2 mm/d *P. thermophila* showed markedly faster growth at 20 °C on V8A and CMA than on MEA (2.0 mm/d) and PDA (1.3 mm/d).

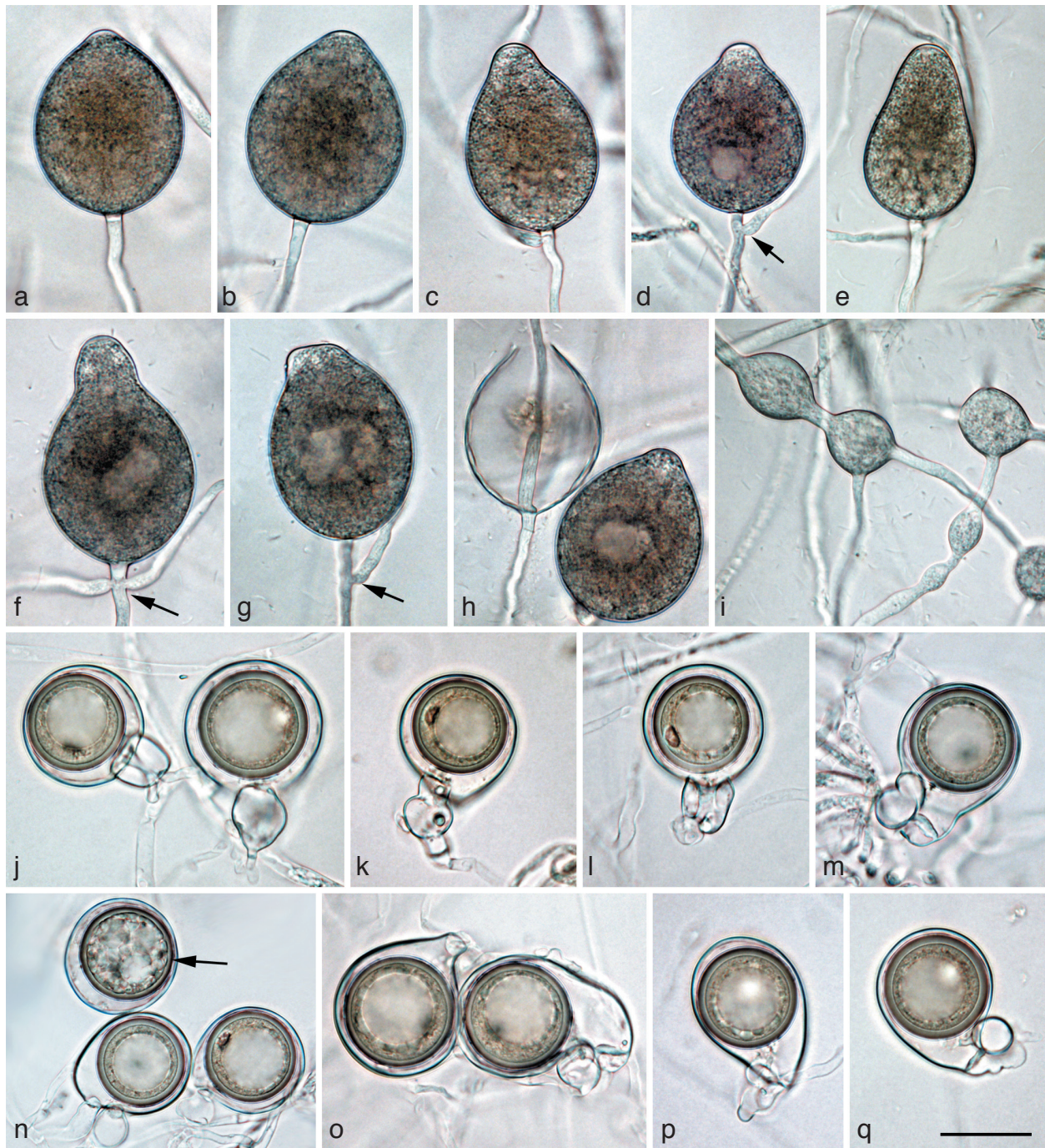
*Specimens examined.* WESTERN AUSTRALIA, Dwellingup, from rhizosphere soil of dying *Eucalyptus marginata*, 2004, VHS, holotype MURU 464 (dried culture on V8A, Herbarium of Murdoch University, Western Australia), cultures ex-type CBS127954 and VHS13530; Quinjinup, from rhizosphere soil of dying plants in native forest, 1998, VHS, VHS3655; Manjimup, from rhizosphere soil of dying plants in native forest, 2000, VHS, VHS7474; Pemberton, from rhizosphere soil of dying *Banksia grandis*, 2006, VHS, VHS16164; Brunswick, baiting from Lunenburg river, 2008, *G.E. St.J. Hardy*, MUCC764.

***Phytophthora taxon paludosa* — Fig. 8**

*Etymology.* The provisional epithet relates to the occurrence of the only known isolate SLPA 166 in a swamp in the Sugarloaf Reservoir Reserve in Victoria (paludosa Latin = swampy).

Sporangia and hyphal swellings (Fig. 8a–h) — Sporangia of *P. taxon paludosa* were not formed on solid agar but were produced abundantly in non-sterile soil extract. Sporangia were typically borne terminally on unbranched sporangiophores or in lax sympodia. Sporangia were non-caducous, semipapillate (Fig. 8a, d, f, g) or nonpapillate (Fig. 8b, c, e, h). Sporangial shapes



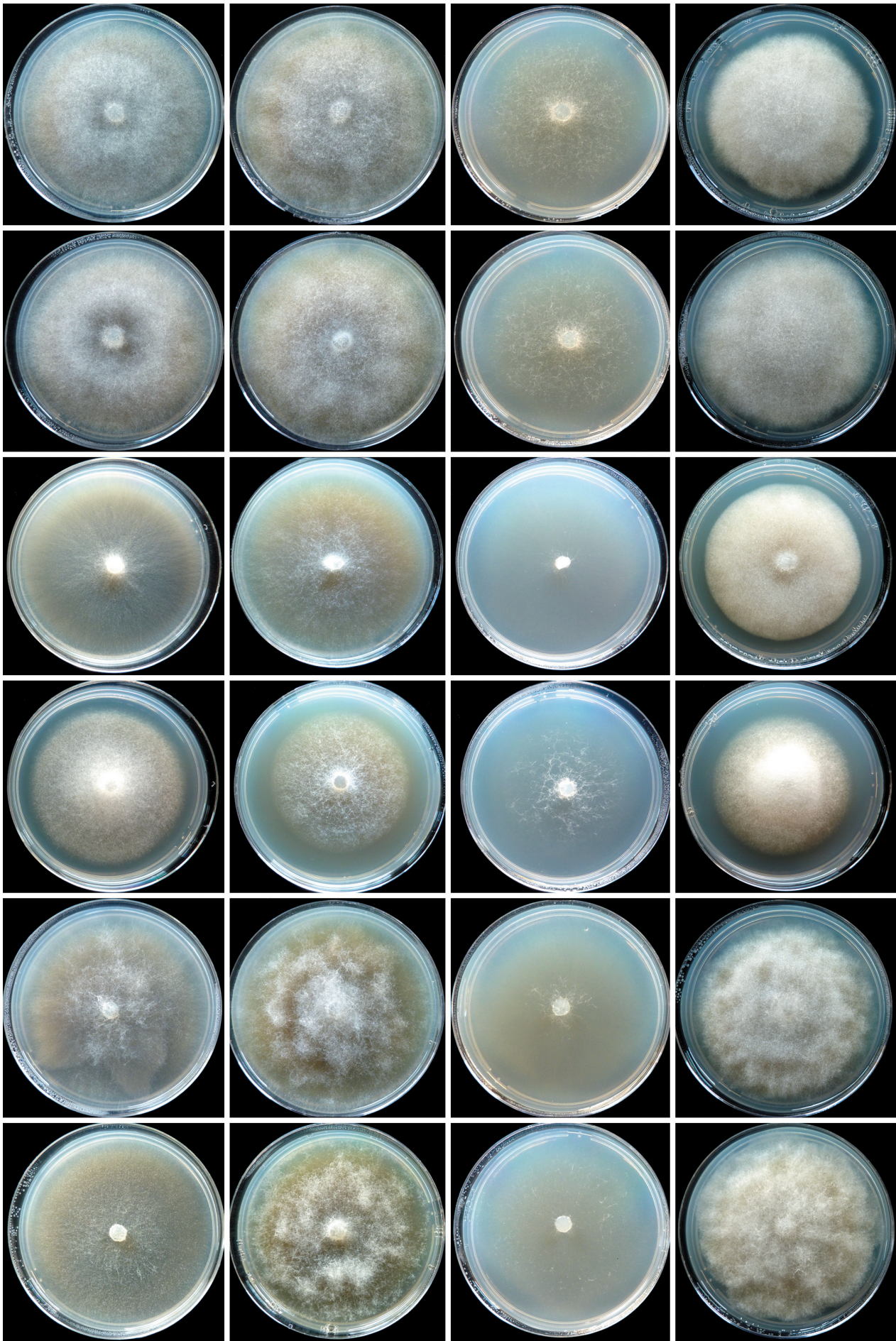


**Fig. 8** Morphological structures of *Phytophthora* taxon *paludosa* formed on V8 agar. a–h. Mature sporangia produced in soil extract: a, b. ovoid semipapillate; c. ovoid nonpapillate with external proliferation; d. ovoid semipapillate with a large vacuole and external proliferation (arrow); e. nonpapillate obpyriform with external monochasial proliferation; f. semipapillate obpyriform with a large vacuole and external dichasial proliferation (arrow); g. semipapillate ovoid with a large vacuole, laterally displaced apex and external monochasial proliferation (arrow); h. two ovoid sporangia with internal extended proliferation (left) and a large vacuole (right); i. intercalary catenulate hyphal swellings produced in soil extract; j–q. mature smooth-walled oogonia with thickwalled aplerotic oospores containing a large ooplast and a nucleus: j. two globose oogonia with amphigynous (left) and paragynous antheridium (right); k. subglobose with paragynous antheridium; l. globose with amphigynous antheridium; m. subglobose with tapering elongated base and paragynous antheridium; n. two subglobose to elongated oogonia with paragynous antheridia and viable oospores and a subglobose oogonium with aborted oospore (arrow); o. subglobose tapering (left) and elongated with curved tapering base (right); p. elongated excentric with paragynous antheridium; q. subglobose with curved tapering base and paragynous antheridium with finger-like hyphal projection. — Scale bar = 25  $\mu$ m.

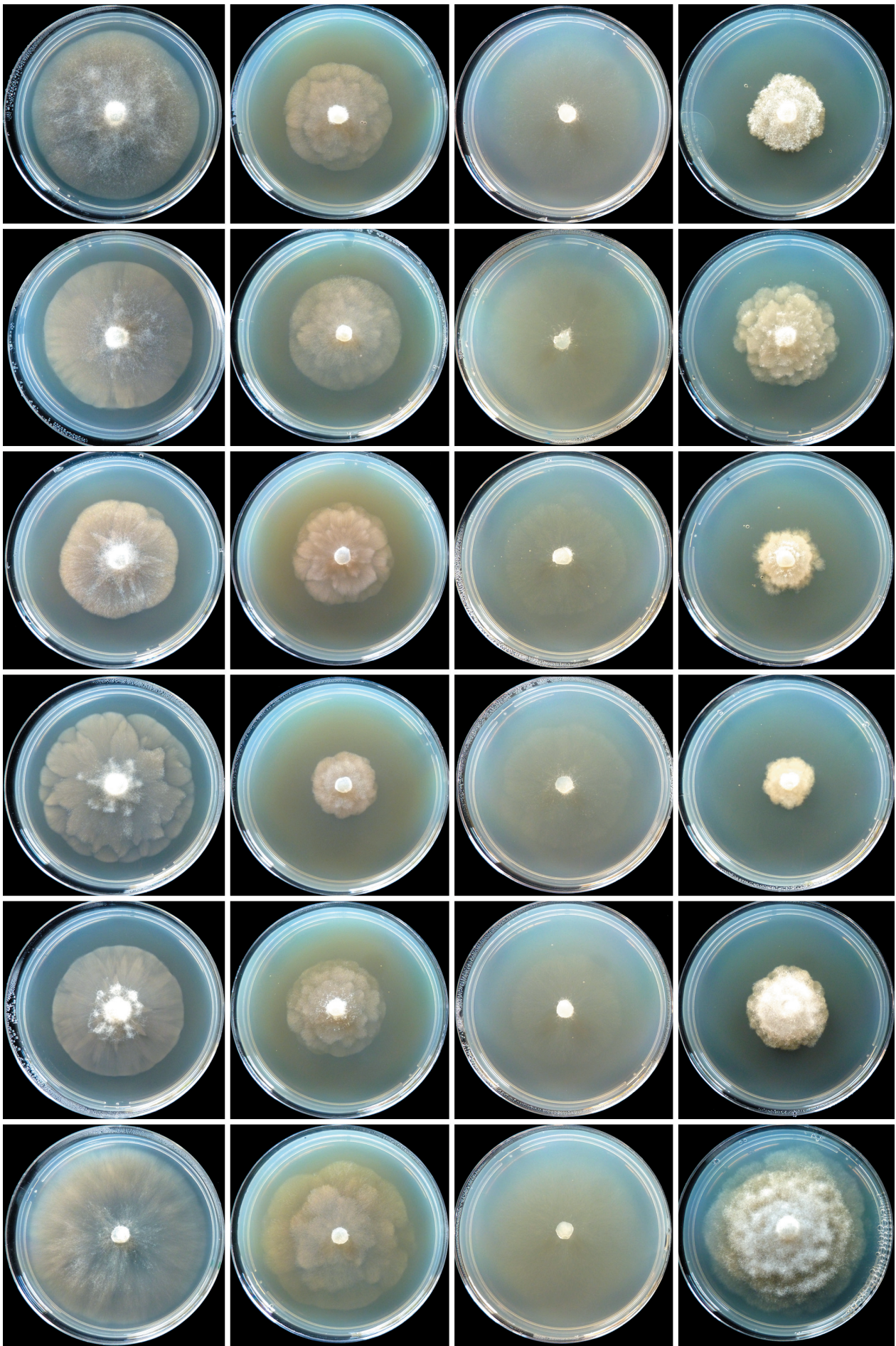
were usually ovoid to broad-ovoid (90 %; Fig. 8a–d, g, h), and less frequently limoniform (8 %) or obpyriform (2 %, Fig. 8e, f). In semipapillate sporangia the apex was sometimes laterally displaced (Fig. 8g). Sporangia were quite large, averaging  $54.7 \pm 4.3 \times 43.3 \pm 3.7 \mu\text{m}$  (overall range  $43.1\text{--}64.6 \times 31.4\text{--}51.7 \mu\text{m}$ ) and often contained a large vacuole (Fig. 8d, f–h). Their mostly broad-ovoid shape is reflected by the low mean length/breadth ratio of  $1.27 \pm 0.09$ . Sporangia proliferated externally (Fig. 8c–g) with one (Fig. 8c, d, g) or sometimes two sporangiophores (Fig. 8f), and internally in an extended way (Fig. 8h). Nested prolifera-

tion was never observed. Zoospores were discharged through an exit pore  $7.5\text{--}14.8 \mu\text{m}$  wide (av.  $10.6 \pm 2.1 \mu\text{m}$ ) (Fig. 8h). They were limoniform to reniform whilst motile, becoming spherical (av. diam =  $11.7 \pm 0.4 \mu\text{m}$ ) on encystment. Cysts germinated by forming a hypha. Diplanetism was never observed. In liquid culture catenulate, globose, ellipsoid or angular hyphal swellings (av. diam =  $16.2 \pm 3.1 \mu\text{m}$ ) were abundant (Fig. 8i).

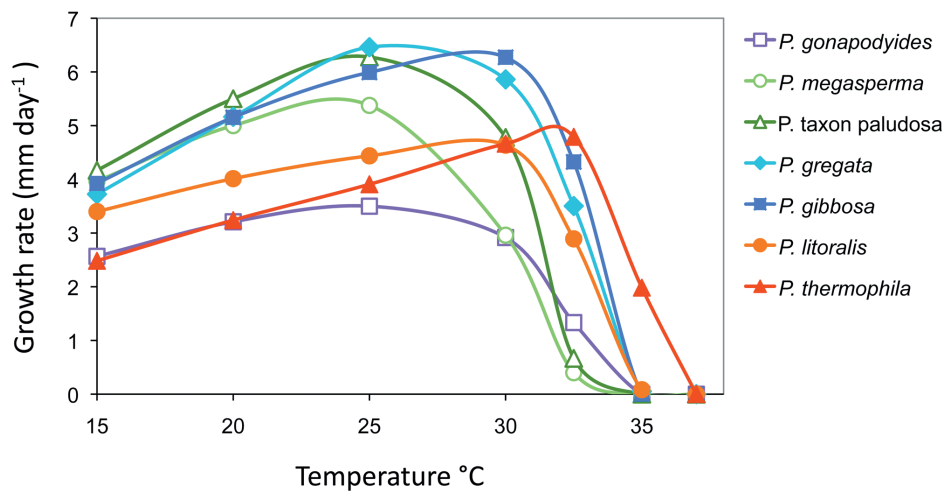
Oogonia, oospores and antheridia (Fig. 8j–q) — Gametangia were readily produced by *P. taxon paludosa* in single culture on V8A within 4 d. Oogonia were borne terminally or laterally, had



**Fig. 9** Colony morphology of *Phytophthora gibbosa* isolates CBS127951 (ex-type) and VHS22007, *P. gregata* isolates CBS127952 (ex-type), MJS235 and VHS9854, and *P. taxon paludosa* isolate MUC765 (from top to bottom) after 7 d growth at 20 °C on V8 agar, malt extract agar, corn meal agar and potato-dextrose agar (from left to right).



**Fig. 10** Colony morphology of *Phytophthora litoralis* isolates CBS127953 (ex-type) and MUC762, *P. thermophila* isolates CBS127954 (ex-type) and MUC764, *P. gonapodyides* isolate MUC761 and *P. megasperma* isolate DDS3432 (from top to bottom) after 7 d growth at 20 °C on V8 agar, malt extract agar, corn meal agar and potato-dextrose agar (from left to right).



**Fig. 11** Mean radial growth rates of *Phytophthora gibbosa* (four isolates), *P. gregata* (eight isolates), *P. litoralis* and *P. thermophila* (each four isolates), *P. taxon paludosa*, *P. gonapodyides* and *P. megasperma* (each one isolate) on V8 agar at different temperatures.

smooth walls and were globose to subglobose (Fig. 8j–o, q) or elongated (Fig. 8n–p), often with a tapering base (Fig. 8m, o–q). Oogonial diameters averaged  $33.3 \pm 3.5 \mu\text{m}$  (overall range 24.4–40.7  $\mu\text{m}$ ). Oospores were always aplerotic, had a mean diameter of  $28.1 \pm 2.8 \mu\text{m}$  (range 21.7–34.3  $\mu\text{m}$ ) and were usually globose, containing a large ooplast and a nucleus (Fig. 8j–q). With a mean thickness of  $2.5 \pm 0.4 \mu\text{m}$  (total range 1.6–3.4  $\mu\text{m}$ ) and a mean oospore wall index of  $0.44 \pm 0.04$  the oospore walls were relatively thick. The abortion rate of oospores was less than 10%. The antheridia were predominantly paragynous (Fig. 8j, k, m–q) but some amphigynous antheridia were also observed (Fig. 8j, l). Antheridia averaged  $16.6 \pm 3.7 \times 13.0 \pm 1.5 \mu\text{m}$ , with shapes ranging from club-shaped, subglobose to cylindrical or irregular (Fig. 8j–q). They were usually formed terminally or laterally, and were rarely intercalary.

Colony morphology, growth rates and cardinal temperatures (Fig. 9, 11) — *Phytophthora taxon paludosa* produced uniform, mostly submerged colonies on V8A and CMA, and faintly stellate colonies on MEA and PDA with limited and fluffy aerial mycelium, respectively (Fig. 9). Growth was fast (6.3 mm/d) at the optimum temperature of 25 °C, and there was only slow growth at 32.5 °C (Fig. 11); 35 °C was lethal since cultures did not resume growth when plates incubated for 7 d at 35 °C were transferred to 20 °C. With mean radial growth rates at 20 °C of 5.5, 5.7, 5.5 and 5.4 mm/d on V8A, MEA, CMA and PDA, respectively, *P. taxon paludosa* showed no agar media preferences.

*Specimen examined.* VICTORIA, Sugarloaf Reservoir Reserve, baiting from an artificial swamp in pasture, 2008, W.A. Dunstan, MUC765.

*Notes* — *Phytophthora gibbosa*, *P. gregata*, *P. litoralis*, *P. thermophila* and *P. taxon paludosa* can easily be separated from each other and from other species of ITS Clade 6 and from morphologically similar *Phytophthora* species by their ITS, *cox1* and HSP90 sequences, and by a combination of morphological and physiological characters of which the most decisive are highlighted in bold in Table 2. In all gene trees, *P. thermophila* and *P. litoralis* are sister species which share a common ancestor with *P. gonapodyides* and *P. megasperma*, while *P. gibbosa*, *P. gregata*, *P. taxon paludosa* and *P. taxon raspberry* are also sister taxa.

*Phytophthora gibbosa* can be easily differentiated from all other species from ITS Clade 6 by the production of ornamented (gibbous) oogonia in single culture. In addition, it can be separated from *P. gregata* and *P. taxon raspberry* by the lack of nested proliferation of sporangia. More distantly related species, which due to the production of ornamented oogonia are morphologi-

cally similar to *P. gibbosa*, can also be easily differentiated by a combination of morphological and physiological features: *P. cambivora* from ITS Clade 7 is heterothallic, has larger two-celled antheridia and shows nested sporangial proliferation (Erwin & Ribeiro 1996). Both, *P. alni* ssp. *alni* and *P. alni* ssp. *multiformis* from ITS Clade 7 have a varying proportion of distorted, often comma-shaped oogonia, usually larger two-celled antheridia and show nested sporangial proliferation (Brasier et al. 2004). *Phytophthora katsurae* from ITS Clade 5 has smaller oogonia with funnel-shaped stalks, smaller papillate sporangia, forms chlamydospores and does not produce hyphal swellings (Erwin & Ribeiro 1996). *Phytophthora cyperi-bulbosi* produces paragynous antheridia and caducous sporangia, and is not culturable (Erwin & Ribeiro 1996). *Phytophthora verrucosa* produces both amphigynous and paragynous antheridia, shows nested sporangial proliferation and is not culturable (Stamps et al. 1990, Erwin & Ribeiro 1996).

In previous studies *P. gregata* is referred to as *P. taxon raspberry* (Brasier et al. 2003a) and *P. sp. 7* (Burgess et al. 2009). It has been previously misidentified in WA as *P. megasperma* or *P. megasperma* var. *sojiae* (Burgess et al. 2009). *Phytophthora gregata* can be separated from *P. megasperma* by the high rate of oogonial abortion, markedly smaller oogonia sizes, the production of hyphal aggregations and different colony growth patterns on MEA; from *P. gibbosa* by the absence of ornamented oogonia, the production of hyphal aggregations and catenulate hyphal swellings and by having nested sporangial proliferation; from *P. taxon paludosa* by high oogonial abortion rates, the production of hyphal aggregations and by nested sporangial proliferation. *Phytophthora gregata* also comprises Australian isolates designated as *P. taxon raspberry*. Brasier et al. (2003a) examined one European isolate (P1050) of *P. taxon raspberry* in detail. It had slightly higher maximum temperature for growth and markedly higher oogonial diameters than *P. gregata* in our study, and an even ratio of amphigynous and paragynous antheridia while *P. gregata* has predominantly paragynous antheridia. However, more European isolates have to be investigated to clarify whether or not these differences are consistent.

*Phytophthora litoralis* is referred to as *P. sp. 11* (Burgess et al. 2009). It is distinguished from *P. thermophila* by being partly silent A1, by the occurrence of external proliferation and secondary lateral sporangia, by having lower optimum and maximum temperatures for growth, and by faster growth on PDA. Characters present in *P. litoralis*, that differentiate it from *P. gonapodyides* include the frequent production of hyphal swellings which were often catenulate and sometimes had ra-

diating hyphae, the production of secondary lateral sporangia, the occurrence of chlamydospores in some isolates, and the predominance of ovoid sporangia (*P. gonapodyides* has mostly obpyriform to ellipsoid sporangia). In addition, most isolates of *P. litoralis* were sterile while all isolates of *P. gonapodyides* are silent A1 (Brasier et al. 1993, 2003a, Erwin & Ribeiro 1996). *Phytophthora litoralis* can be easily separated from *P. megasperma* by the absence of oogonia, the occurrence of external sporangial proliferation and secondary lateral sporangia, the production of chlamydospores by some isolates and higher optimum temperature for growth.

*Phytophthora thermophila* has previously been misidentified as *P. drechsleri* and is referred to as *P. sp. 3* (Burgess et al. 2009). This species can be distinguished from *P. drechsleri* mainly by being either sterile or sporadically self fertile with paragynous antheridia, but also by the branching of internally proliferating sporangiophores outside the empty primary sporangium and the regular occurrence of diplanetism; from *P. gonapodyides* by its markedly higher optimum temperature for growth, a higher length/breadth ratio of the sporangia, the absence of external sporangial proliferation, the frequent production of hyphal swellings which were often catenulate and sometimes had radiating hyphae, the occurrence of chlamydospores in some isolates and the inability to induce selfing in A2 isolates; from *P. megasperma* by the sterility of most isolates, the production of chlamydospores by some isolates and a markedly higher optimum and a slightly higher maximum temperature for growth.

*Phytophthora gibbosa* is known only from a very limited area in the southwest of WA around Scott River where it has been found associated with dying native vegetation on seasonally wet sites. *Phytophthora gregata* has a scattered distribution in WA. It was recovered from native vegetation in the Northern sandplains around Lancelin, from native vegetation on seasonally wet sites further south around Busselton, Nannup and Scott River (together with *P. gibbosa*), and from 1 yr old dying *Pinus radiata* on seasonally wet sites in the Donnybrook Sunland plantations. *Phytophthora gregata* also occurs in southeastern Australia in wet native *Eucalyptus* forests and in swamps in Victoria, and was associated with dying alpine heathland vegetation in Tasmania. The known distribution of *P. litoralis* comprises areas along the south coast of WA and isolated sites c. 60–80 km inland around Wilga and Borden where it was recovered from river systems and dying native vegetation on wet sites. The distribution of *P. thermophila* in WA is quite scattered, ranging from the northern Sandplains, the Perth area and rehabilitated mine-sites in the northern Jarrah forest around Dwellingup, to the southwest around Nannup, Boyup Brook, Manjimup and Pemberton in the Southern Jarrah Forest and further east to Tambellup in the Great Southern. It was most often associated with dying native vegetation on both low and high disease impact sites. In addition it was also found widespread in river systems across the southwest of WA (<http://www.fishingforphytophthora.murdoch.edu.au>).

## DISCUSSION

Four new *Phytophthora* species and one informally designated taxon within ITS Clade 6, *P. thermophila*, *P. litoralis*, *P. gregata* and *P. gibbosa* and *P. taxon paludosa*, have been described from natural ecosystems in Australia based on differences in morphology and sequence data of three gene regions. The phylogenies of the ITS and HSP90 gene regions are congruent, with each species residing in highly supported terminal clades. The mitochondrial *cox1* gene shows considerable intraspecific variation, and while isolates from a single species form a single lineage, the support is not as strong as in the nuclear gene regions. In their overview of ITS Clade 6, Brasier et al. (2003a)

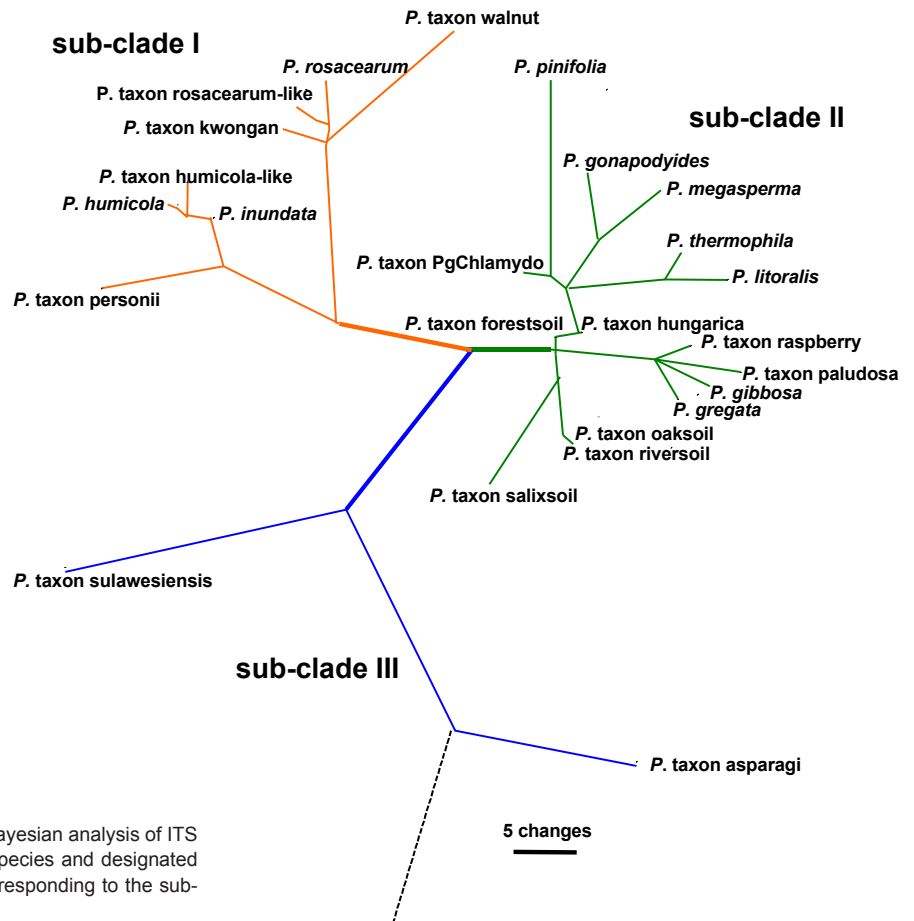
recognised three sub-clades comprising three described species (*P. gonapodyides*, *P. megasperma*, *P. humicola*) and 10 phenotypic taxa. Now, in total, there are 10 species and 15 designated taxa, which is getting closer to the estimation of Brasier (2009) that the number of extant species in Clade 6 is between 28 and 84.

### Radiation within Clade 6

Brasier et al. (2003a) introduced several new taxa in sub-clade II; *P. taxon PgChlamydo* and *P. taxon salixsoil*, previously included in *P. gonapodyides*, *P. taxon forestsoil*, *P. taxon riversoil* and *P. taxon oaksoil* from natural ecosystems in Europe, and *P. taxon raspberry* from Australia and Sweden. To date none of these taxa has been described; however, based on ITS sequence, the *P. taxon raspberry* isolates from Australia are now considered as *P. gregata*. With the addition of *P. pinifolia* (Durán et al. 2008), *P. taxon hungarica* and the four new species and one taxon from the current study, sub-clade II has expanded significantly (Fig. 12). However, while individual taxa reside in strongly supported terminal clades, sub-clade II is still predominantly characterised by multiple short branches with weak bootstrap and Bayesian support for higher level clustering. This tree topology supports recent radiation from an ancestral type and average genetic distance across the sub-clade is only 1.9%. *Phytophthora pinifolia* is an exception within sub-clade II as it is 14 bp different from the closest taxon, *P. taxon PgChlamydo*, and differs from other taxa by a genetic distance of 2.2–3.3% (Fig. 12). The inclusion of the new taxa has increased the structure within sub-clade II and two distinct clusters can be distinguished. *Phytophthora thermophila* and *P. litoralis* are sister species to each other and more distantly to *P. gonapodyides* and *P. megasperma* (Fig. 12). These four species share a common ancestor. Similarly, *P. gregata*, *P. gibbosa* and *P. taxon paludosa*, and isolates of *P. taxon raspberry* from Europe, form a new cluster within sub-clade II which also share a common ancestor (Fig. 12). *Phytophthora taxon salixsoil* differs from other taxa by a genetic distance of 1.8–3.5% in ITS region and is consistently placed in a basal position in all parsimony, distance and Bayesian analyses (also in HSP90 and *cox1* phylogenies). The other European taxa are poorly resolved and differ from each other by 2–7 bp.

Although there are relatively small differences between many sub-clade II taxa, including the newly described species, ITS sequences presented in this study were singular with the exception of 2–3 ambiguous sites in *P. litoralis* and *P. taxon PgChlamydo*. These data concur with previous observations where no evidence for reticulation was found, even though radiation appeared to be recent and the taxa shared niches (Brasier et al. 2003a). However, this is an extreme oversimplification of the observations from Australian surveys in native vegetation and waterways. We have described here several discernable species from among a huge array of putative hybrids all containing numerous single base pair polymorphisms (unpubl. data). In phylogenetic analyses these putative hybrids fall within the *P. thermophila/P. litoralis* cluster and characterisation studies are currently underway.

In 2003, sub-clade I consisted of *P. humicola* and three undescribed taxa: *P. sp. O-group* (later described as *P. inundata* by Brasier et al. 2003b), *P. sp. apple-cherry* (later described as *P. rosacearum* by Hansen et al. 2009b) and *P. taxon walnut*. Several new taxa have now been recognised within sub-clade I: *P. taxon personii*, *P. taxon humicola-like*, *P. taxon kwongan* and *P. taxon rosacearum-like*. Brasier et al. (2003a) characterised this group as having relatively long branch lengths thought to be indicative of ancient divergence; and indeed there is 18–28 bp difference (up to 4.5%) between taxa in the two main branches (Fig. 12). However, within the two branches



**Fig. 12** Radial phylogenetic tree generated after Bayesian analysis of ITS rDNA sequences, showing relationships between species and designated taxa in Clade 6 of *Phytophthora*. The branches corresponding to the sub-clades are coloured accordingly.

there are now additional taxa separated by smaller genetic distances representing more recent divergence. Sub-clade III is represented by *P. taxon asparagi* (Saude et al. 2008) and *P. taxon sulawesiensis*, which are distantly related to each other and the other taxa within ITS Clade 6.

When discussing the role of geographic isolation in speciation we are hampered by uncertainty about the origin of taxa due to considerable anthropogenic movement. Often the same taxa can be observed across several continents in both hemispheres and the true origin of the taxa is obscured. *Phytophthora gonapodyides*, *P. taxon salixsoil* and *P. taxon PgChlamydo* are ubiquitous in water bodies in Europe and the northwestern USA (Jung et al. 1996, Brasier et al. 2003a, Nechwatal & Mendgen 2006, Hansen et al. 2009a, Reeser et al. 2011) and appear to have a global distribution, excluding the tropics (Zeng et al. 2009). However, there are some distribution limits; *P. taxon PgChlamydo* was not found in Alaska (Reeser et al. 2011) and in WA, *P. gonapodyides* has never been isolated and *P. taxon PgChlamydo* and *P. taxon salixsoil* were only rarely recovered from native flora, but not from streams. Other species have a more limited current distribution: *P. thermophila* and *P. litoralis* are only known from Australia where they occupy the same niche as *P. gonapodyides* and *P. taxon salixsoil* in the Northern Hemisphere. *Phytophthora taxon forestsoil* and *P. taxon riversoil* are only recorded from Europe (Brasier et al. 2003a, Bakonyi et al. 2009) and *P. taxon hungarica* has been recorded from Alaska and Europe (Adams et al. 2009, Bakonyi et al. 2009). *Phytophthora taxon oaksoil* has been found in Europe and in mountain stream surveys in Oregon (Reeser et al. 2011). New taxa close to *P. gonapodyides*, *P. taxon salixsoil* and *P. taxon PgChlamydo* have been recently found in North American surveys (Hulvey et al. 2010, Reeser et al. 2011). As more sequence data become available and new species are described, a clearer picture of natural species distribution will emerge.

Seven of the nine taxa in sub-clade I are found in WA (*P. humicola* was only reported from Taiwan and *P. taxon walnut* only from California). *Phytophthora inundata* is widely distributed, commonly isolated in stream surveys and infrequently associated with dying vegetation (Stukely et al. 2007); and many unique isolates related to *P. rosacearum* have been isolated from dry kwongan heathlands. Until more information becomes available from environmental surveys world-wide, the current distribution and variation suggests either an Australian origin or substantial post-introduction radiation within Western Australia.

#### Evolutionary and ecological implications

Based on data from 12 taxa in ITS Clade 6, Brasier et al. (2003a, b) postulated a tendency towards inbreeding or breakdown of sexual reproduction. Only seven years later and another 13 taxa richer, this hypothesis is reinforced. Of the 15 known taxa in sub-clade II, seven including *P. gibbosa*, *P. gregata* and *P. taxon paludosa* are homothallic, three are self-sterile and silent A1s, one is self-sterile and partially silent A1 (*P. litoralis*), one is self-sterile or sporadically self fertile (*P. thermophila*) and three are fully sterile (Table 4). Of the eight known taxa in sub-clade I, five are homothallic, one is heterothallic (*P. inundata*) and two are fully sterile (Table 4). Also *P. taxon asparagi* from sub-clade III is homothallic; no data are available for *P. taxon sulawesiensis* and hence it is excluded from further discussion. Overall, 13 taxa (54 %) are homothallic and 10 taxa (42 %) are sterile while only one species, *P. inundata*, is heterothallic (Table 4). This is in contrast to the 89 *Phytophthora* species described so far from the other nine ITS clades of which 72 % are homothallic, 25 % are heterothallic and only 3 % are sterile.

When considering the influence of natural selection on sexual reproduction in *Phytophthora* one has to be conscious of oospores not only providing new genotypes via recombination, but also acting as the most enduring resting structures for

survival in unfavourable environmental conditions, such as drought and hot or cold temperatures. Homothallism provides resting structures, but limits a species' ability for recombination and hence its adaptability to changing environmental conditions. Sterile taxa are essentially evolutionary dead-ends, having abandoned both the potential for sexual recombination and the potential of forming long-term resting structures. For Clade 6 *Phytophthora*s to follow this evolutionary path there must have been an advantage in this life strategy within their ecological niche.

In general, one of the most important selective influences on *Phytophthora* pathogens is the host plant (Brasier & Hansen 1992). In the tropical rainforests, high spatial diversity of the flora requires, from any pathogen, the potential for rapid adaptation to new host genotypes or even new host species via genetic recombination. In order to avoid falling back in the perpetual arms race with new host genotypes rapid adaptation is required, and this is usually best achieved by sexual recombination (Lane 2009). Therefore, heterothallism with its intrinsic tendency towards outcrossing is the most successful sexual strategy for tropical *Phytophthora*s, e.g. *P. botryosa*, *P. capsici*, *P. cinnamomi*, *P. colocasiae*, *P. megakarya*, *P. nicotianae* and *P. palmivora*. Due to the rare occurrence of both mating types simultaneously infecting the same host tissue, oospore formation in nature for heterothallic species is certainly much less common than for homothallic species. However, in the constantly warm and wet tropical environment, the conditions for asexual multiplication and spread via zoospores are continuously favourable and oospores are not required as long-term resting structures. The occasional mating will guarantee sexual recombination and adaptability. In contrast, in temperate and Mediterranean climates, *Phytophthora* species must be able to survive alternating low winter temperatures and long dry periods in summer. Here, homothallism, a compromise between the need for adaptability via sexual recombination and the regular and abundant production of resting structures for long-term survival in a dormant state, is by far the most common sexual breeding system, as reflected by *P. austrocedrae*, *P. cactorum*, *P. citricola*, *P. ilicis*, *P. multivora*, *P. nemorosa*, *P. plurivora*, *P. pseudosyringae* and *P. quercina*. In accordance with this simplified macroclimatic distribution of breeding systems, none of the 13 homothallic ITS Clade 6 taxa, except *P. humicola* which was recovered from Citrus orchards in Taiwan (Ko & Ann 1985), and none of the 10 sterile Clade 6 taxa, have ever been recorded from wet tropical climates (Erwin & Ribeiro 1996, Brasier et al. 2003a, Zeng et al. 2009).

Brasier et al. (2003a, b) proposed the sexual degeneration in Clade 6 may be connected to their aquatic habitat. Although sporadically recorded from forest soils, nine of the 10 sterile Clade 6 taxa are commonly found in rivers, lakes or water-logged soils and swamps (Table 4) which provide constantly wet conditions enabling continuous asexual multiplication and spread via zoospores. Within the freshwater aquatic environment, multiple *Phytophthora*, *Pythium*, *Achlya* and *Saprolegnia* species with high inoculum levels (Hallett & Dick 1981, Yamak et al. 2002, Nechwatal et al. 2008, Hwang et al. 2009, Reeser et al. 2011) are competing for the same substrate, i.e. freshly fallen green leaves and/or fine or coarse roots and root collars of riparian woody species (Brasier et al. 2003b, Jung & Blaschke 2004). The predominantly aquatic sterile taxa of Clade 6 have undergone an intense selection towards a life strategy focused on rapid and abundant asexual multiplication via zoospores in order to compete with the multitude of other oomycetes in freshwater ecosystems. Accordingly, oospores are not required as long-term resting structures and an allocation of resources into their production would be wasteful. Interestingly, in mountainous

streams in Hainan sterile isolates of the homothallic *P. insolita* from ITS Clade 9 are commonly found (Zeng et al. 2009).

Without sexual recombination these sterile aquatic Clade 6 *Phytophthora*s also lose the ability to generate new genotypes. Significant genetic variation of phenotypic characters such as virulence, host range and growth rates in F1 progenies is observed for heterothallic outcrossing species and has also been shown for predominantly inbreeding homothallic species like *P. cactorum*, *P. medicaginis*, *P. sojae* or *P. syringae* (Boccas 1972, Erwin & Ribeiro 1996, MacIntyre & Elliott 1974). Many of the F1 progenies were less fit than the parents, showing poor growth and sporulation or low pathogenicity. The higher variation of behavioural characters provided by sexual as compared to asexual reproduction would lead to progenies less adapted to the specific and stable environmental conditions of freshwater ecosystems than the parental genotype. Since sporangial production and aggressiveness are highly negatively correlated characters, as shown for *P. infestans* (Caten 1970), the aquatic Clade 6 taxa have presumably quit the arms race and co-evolution with potential host plants in favour of outnumbering the waterborne inoculum of competitors. In *P. litoralis* and *P. thermophila* this is achieved by chains of nested and extended proliferating sporangia, and the frequent germination of cysts by releasing secondary zoospores (diplanetism) or by forming microsporangia. In addition, *P. litoralis* is using the cytoplasm remaining in the sporangiophore after the formation of the primary terminal sporangia for the production of secondary lateral sporangia.

Ephemeral streams and some habitats, particularly in the shallow water zone along the banks of bigger rivers, lakes and swamps, periodically dry out. Thus, taxa within this niche must form short-term resting structures to survive until the water returns or they will become locally extinct. Four of the six self-sterile Clade 6 taxa commonly associated with streams, i.e. *P. litoralis*, *P. thermophila*, *P. taxon PgChlamydo* and *P. taxon personii*, the part-heterothallic *P. inundata* and the homothallic *P. taxon humicola*-like produce chlamydospores after several days at 15–20 °C in liquid culture (Table 4). Stimulation of chlamydospore production in water at temperatures too high or too low for growth or infection is known for *P. cactorum*, *P. citrophthora*, *P. lateralis*, *P. nicotianae* and *P. palmivora* and seems to be a stress-related phenomenon associated with high CO<sub>2</sub> concentrations at sufficiently high oxygen concentrations (Blackwell 1943, Englander & Roth 1980, Darmono & Parke 1990, Erwin & Ribeiro 1996). Interestingly, in Europe the recently described *P. gallica* from ITS Clade 10 also occurs in seasonally flooded natural ecosystems, is sterile and forms chlamydospores (Jung & Nechwatal 2008). These examples of convergent evolution of the same life strategy under similar ecological conditions, in taxa from phylogenetically distant clades, illustrate the adoption of such a life strategy for survival in aquatic habitats. Since, as a trade-off, these taxa have relinquished the potential for sexual reproduction, adaptation to changing environmental conditions can only be achieved by spontaneous mutations or parasexuality, i.e. heterokaryon formation, hyphal anastomosis and zoospore fusion within or between species (somatic hybridisation). Mutations create new variation, but their occurrence is rare and most often deleterious. In the absence of sexual reproduction the combination of different beneficial mutations in the same individual of a sterile population can only occur via somatic fusion if vegetative compatibility permits. In *Phytophthora* the extent of parasexual processes and their role in generating phenotypic variation on which natural selection can work is still poorly understood (Kuhn 1981, Brasier 1992).

Within *P. inundata*, the only heterothallic Clade 6 species known to date, a notable proportion of intraspecific A1 × A2 pairings

Table 4 Habitats and phenotypic characters of *Phytophthora* taxa from ITS Clade 6.

Taxon and source of data	Habitats <sup>1</sup>	Maximum temp. (°C)	Optimum temp. (°C)	Breeding behaviour <sup>2</sup>	Mean oogonia diam (µm)	Antheridia type <sup>3</sup>	Chlamydospores <sup>4</sup>	Hyphal swellings <sup>4</sup>	Sporangial proliferation <sup>5</sup>
<b>Sub-clade I</b>									
<i>P. inundata</i> <sup>6,7,8</sup> ( <i>P. sp.</i> O-group)	Aq, Rip, Hor	35–38	28–30	HETA1 csf, HET A2 SS, PARHET A2 csf or SS	40.1	A	++ g, [rad] <sup>9</sup>	++ g, ang, el, [cat, rad] <sup>9</sup>	Int ne <sup>8</sup>
<i>P. humicola</i> <sup>9</sup>	Hor	38	28	HO	43.8 <sup>6</sup> , 39 <sup>9</sup>	P/a	–	+++ g, cat, rad	Int e, [ext]
<i>P. taxon humicola-like</i> <sup>8</sup>	Hea	na	na	HO	41.0	P	++ g, [irr, rad]	++ g, el, cat [rad]	Int ne, ext
<i>P. taxon personii</i> <sup>8</sup>	Aq, Rip, Hor	na	na	Sterile	–	–	+++ g, [irr, rad]	+++ g, el, [cat], rad	Int ne, [ext]
<i>P. taxon walnut</i> <sup>6</sup>	Hor	38	28	Sterile	–	–	–	na	na
<i>P. rosacearum</i> <sup>6,10</sup> ( <i>P. sp.</i> apple-cherry)	Hor	36 <sup>10</sup> , 38 <sup>6</sup>	28 <sup>6</sup> , 30 <sup>10</sup>	HO	37.5 <sup>6</sup> , 29–34 <sup>10</sup>	P/A	– <sup>10</sup>	na	Int ne, ext
<i>P. taxon rosacearum-like</i> <sup>8</sup>	Hea	na	na	HO	na	na	na	na	na
<i>P. taxon kwongan</i> <sup>8</sup>	Hea	na	na	HO	na	na	na	na	na
<b>Sub-clade II</b>									
<i>P. taxon salixsoil</i> <sup>6,11,12</sup>	Aq, Rip	37 <sup>11</sup> , 38 <sup>6</sup>	30 <sup>11</sup> , 33 <sup>6</sup>	SS silent A1	–	–	–	(+), g, [cat]	Int ne, [ext]
<i>P. taxon PgChlamydo</i> <sup>6,7,13</sup>	Aq, Rip, For, Am, Nur	36–37	25–30 <sup>6</sup>	SS, silent A1	–	–	+++ g, irr	+++ g, cat	Int ne, [ext]
<i>P. taxon forestsoil</i> <sup>6,14</sup>	For <sup>6</sup> , Rip <sup>14</sup>	33 <sup>6</sup> , < 33 <sup>14</sup>	25–28	HO	42.3 <sup>6</sup> , 37.7 <sup>14</sup>	P/a	–	+ g, [rad]	Int ne, ext
<i>P. taxon hungarica</i> <sup>14</sup>	Rip	< 33	25	HO	41.4	P/a	–	–	Int ne, ext
<i>P. megasperma</i> <sup>6,9,12</sup>	Rip, For, Hor, Nur, Ag	25–34	20–25	HO	43.5–60.5 <sup>6</sup>	P, P/a or P/A	–	+++ g, ang, el, cat	Int ne
<i>P. gonapodyides</i> <sup>6,7</sup>	Aq, Rip, For, Am, Nur	34–37	25–28	SS, silent A1	–	–	–	(+), g	Int ne, [ext]
<i>P. thermophila</i> <sup>12</sup>	Aq, Rip, For	35–< 37	32.5	Sterile or partially or sporadically self fertile	31.1	P, mul	(++) g (rad)	+++ g, ang, el, cat, [rad]	Int ne
<i>P. litoralis</i> <sup>12</sup>	Aq, Rip, Hea	32.5–< 35	30	Sterile or SS silent A1	–	–	(++) g [rad]	+++ g, ang, el, cat, [rad, agg]	Int ne, ext, sec
<i>P. pinifolia</i> <sup>15,16</sup>	Pla, Can	30	25	Sterile	–	–	–	++ g, rad	–, [cad]
<i>P. taxon oaksoil</i> <sup>6,14</sup>	Aq, Rip	33	25	Sterile	–	–	–	–	na
<i>P. taxon riversoil</i> <sup>6</sup>	Rip	na	na	Sterile?	–	–	–	na	na
<i>P. taxon paludosa</i> <sup>12</sup>	Rip	32.5	25	HO	33.3	P/a	–	++ g, el, cat	Int e, ext
<i>P. gibbosa</i> <sup>12</sup>	For, Pla	32.5–< 35	30	HO	38.1	A	–	++ g, el	Int e, ext
<i>P. gregata</i> <sup>12</sup>	Rip, For, Hea	32.5–< 35	25 [30]	HO or potentially self fertile, csf, AB	36.8	P/a	–	+++ g, ang, el, cat, agg	Int ne
<i>P. taxon raspberry (Europe)</i> <sup>6</sup>	Hor	35	28	HO	46.8	P/A	–	na	na
<b>Sub-clade III</b>									
<i>P. taxon asparagi</i> <sup>17</sup>	Ag	< 30	25	HO	25–30	A	–	na	Int e, ext

na, not available; ( ) feature is only present in some isolates of a taxon; [ ] feature is rare.

<sup>1</sup> Aq, aquatic; Rip, riparian or wetland soils; Hea, heathland soils; For, forest soils; Pla, forest plantations; Can, canopy; Hor, woody horticultural crops; Am, amenity trees; Nur, nurseries; Ag, agricultural crops.<sup>2</sup> HO, homothallic; HET, normally A1 or A2 heterothallic; PARHET, partially A1 or A2 heterothallic; fails to mate in many combinations; SS, self sterile in single culture; csf, chimerically self fertile in single culture. SS Silent A1, self sterile but able to induce gametangial formation in an A2 type of another species (see Brasier et al. 2003b); sterile, no oogonia produced in single culture, or in pairings with other isolates of its own or A1 and A2 tester strains of other taxa; AB, high abortion rate (> 80 %).<sup>3</sup> P, paragonous; A, amphigynous; P/A, equal paragonous and amphigynous; P/a, predominantly paragonous; p/a, predominantly amphigynous; mul, some oogonia with 2-3 antheridia.<sup>4</sup> –, none; (+) rare in some isolates; ++, frequent; g, globose to subglobose; irr, irregular; ang, angular; el, elongated; cat, catenulate; rad, radiating hyphae; agg, hyphal aggregations.<sup>5</sup> Int, internal proliferation; n, nested; e, extended; ext, external proliferation; –, no proliferation; sec, secondary production of lateral sporangia; cad, caducous.<sup>6</sup> Brasier et al. 2003a; <sup>7</sup> Brasier et al. 2003b; <sup>8</sup> Jung & Stukely unpubl.; <sup>9</sup> Erwin & Ribeyro 1996; <sup>10</sup> Hansen et al. 2009b; <sup>11</sup> Nechwatal & Mendgen 2006; <sup>12</sup> this study; <sup>13</sup> Hansen et al. 2009a; <sup>14</sup> József Bakonyi & Zoltán Á. Nagy, pers. comm.; <sup>15</sup> Durán et al. 2008; <sup>16</sup> Durán et al. 2010;<sup>17</sup> Saude et al. 2008.



failed to produce gametangia, indicative of a degenerating compatibility system (Brasier et al. 2003b). As with other introduced heterothallic *Phytophthora* species, it is thought this disruption is caused by the anthropogenic spread of this species beyond its native geographical distribution leading to reproductive isolation of the A1 and A2 mating types (Brasier 1992, Erwin & Ribeiro 1996). Taxa within Clade 6 that are self-sterile, may have evolved silent A1 mating types in a similar manner. However, homothallic species, i.e. *P. cactorum*, *P. heveae*, *P. katsurae* and *P. sojae*, are also able to stimulate oogonia formation in A1 and/or A2 isolates of the heterothallic species *P. nicotianae* and *P. palmivora*, respectively (Brasier 1972, Ko 1980). Hence, it can only be speculated whether the self-sterile silent A1 taxa have kept the genetic 'memory' of a heterothallic A1 ancestor or of a homothallic ancestor able to stimulate A2 strains of heterothallic *Phytophthora* species. Since the self-sterile and partially silent A1 species *P. gonapodyides* and *P. littoralis* most likely share an ancestor with the homothallic *P. megasperma* and the self-sterile or inconsistently homothallic *P. thermophila*, a homothallic ancestor of this cluster within sub-clade II is most plausible. This is supported by the finding of sterile isolates of the homothallic *P. insolita* in Taiwan which were able to induce selfing in A2 isolates of *P. nicotianae* (Ann & Ko 1994). In a later study, Ho et al. (2002) demonstrated occasional production of oogonia in aged cultures of some sterile isolates of *P. insolita*.

The disrupted mating system observed for *P. gregata* may reflect 'evolution in action' with the end point being sterility. While in the closely related *P. gibbosa*, abortion rates between 16 and 37 % were following simple Mendelian ratios, the observed average abortion rate of 85 % for *P. gregata* was far beyond Mendelian segregation of homozygous recessive lethals. Five of the eight isolates studied produced hardly any viable oospores, and some isolates produced oospores only in chimaeric patches or in paired cultures with A1 and A2 tester strains. Most likely, the extensive breakdown of the sexual system is caused by the accumulation of lethal mutations, repulsion linkages or chromosomal aberrations (Rutherford & Ward 1985, Brasier 1992). High abortion rates of oogonia are also known from other homothallic *Phytophthora* species. In some races of *P. megasperma* f. sp. *glycinea* oospore abortion rates reach 90 % (Rutherford & Ward 1985). Also the majority of oospores aborted within 14–21 d in all isolates of *P. pseudosyringae* and *P. psychrophila* examined (Jung et al. 2002, 2003). Despite the disruption of its breeding system, *P. gregata* is a successful species with a high genetic variability and a wide distribution across Australia. Since most isolations of *P. gregata* have come from native vegetation on permanently or seasonally wet sites the apparent evolution of this species towards sterility may follow similar rules of natural selection as in predominantly aquatic Clade 6 taxa. The role of the largely non-functional oospores as survival structures for adverse environmental conditions has presumably been taken over by the abundantly formed, dense and often quite large hyphal aggregations. These resemble stromata formed by *P. ramorum* beneath the cuticle of infected leaves (Moralejo et al. 2006). Recently, Brasier et al. (2010) also reported the formation of stromata in agar cultures of *P. lateralis* isolates from Taiwan.

Most Clade 6 taxa including those commonly occurring in waterways in boreal climates such as *P. gonapodyides* and *P. taxon salixsoil* (Reeser et al. 2011) have high optimum and maximum temperatures for growth (Brasier et al. 2003a, b). Also the four new species described in this study are 'high temperature' taxa, with *P. thermophila* having an optimum and maximum of 33 and 35 °C, respectively. Brasier et al. (2003a) suggested either a tropical origin of the ancestors of Clade 6

or an eco-physiological adaptation to litter breakdown as possible explanations. Due to the almost complete absence of heterothallism within Clade 6 and a lack of records of Clade 6 taxa from the tropics, a tropical origin seems unlikely. We favour the hypothesis of the high temperature optimum and tolerance being an adaptation to a warm microhabitat such as the littoral zone of water bodies where temperatures particularly in summer often exceed 30 °C due to a combination of factors like shallow depth and slow flow of the water, a dark colour of the muddy ground and intense litter breakdown.

### Host range and pathogenicity

*Phytophthora gibbosa*, *P. gregata*, *P. littoralis* and *P. thermophila* have been recovered from rhizosphere soil of dying plants from four native species (*Banksia grandis*, *Eucalyptus marginata*, *Xanthorrhoea gracilis* and *X. preissii*), *Acacia pycnantha* (endemic to eastern Australia), from several unidentified species of the genera *Banksia*, *Eucalyptus*, *Grevillea*, *Hakea* and *Patersonia*, and also from the exotic plantation timber species *Pinus radiata*. Disease expression is often low impact or only associated with scattered individual plant deaths. In WA, all four new *Phytophthora* species appear to be opportunistic pathogens associated with sporadic but severe mortality on wet sites or following episodic favourable conditions such as unseasonably heavy rain or flooding. Similarly, *P. gonapodyides* and *P. inundata* episodically cause root and collar rot and aerial cankers in various tree species in Europe during extremely wet periods; *Fagus sylvatica*, *Q. robur* and *Alnus glutinosa* (*P. gonapodyides*; Jung et al. 1996, Jung & Blaschke 2004, Brown & Brasier 2007) and *Aesculus*, *Salix* and olive trees (*P. inundata*; Sanchez-Hernandez et al. 2001, Brasier et al. 2003b). However, the pathogenicity of all four new *Phytophthora* species and *P. taxon paludosa* is untested and their host ranges are unknown.

### Implications for management

Significant public and private resources have been applied to the investigation and management of *Phytophthora* Dieback (caused by *P. cinnamomi*) in natural ecosystems in WA for more than three decades (Shearer & Tippett 1989, Shearer & Smith 2000). Extensive vegetation health survey has been in continuous operation in the WA jarrah (*E. marginata*) forest since 1978. Mapping of the extent of the disease, based on shadowless colour aerial photography, is verified by the testing of soil and root samples for the presence of *Phytophthora* (Strelein et al. 2006). Dieback-free areas are then quarantined to minimise the risk of spread of the pathogen into priority areas.

Recent re-examination of a large number of these *Phytophthora* isolates of species other than *P. cinnamomi*, and sequencing of their ITS gene regions to confirm identity, has revealed a surprisingly diverse number of new records and undescribed taxa (Burgess et al. 2009; this study). The majority of the soil and root samples from which these *Phytophthoras* were isolated were associated with dying native vegetation. It will be prudent, therefore, for land managers to apply the precautionary principle in managing all of these soilborne *Phytophthora* taxa in natural ecosystems, regardless of their present known impact.

While a particular *Phytophthora* taxon may not now be causing visible damage at a site where it is long-established (or endemic), it may cause very significant damage when first introduced to a new site – as a result of a different suite of plant species being present, and of subtle differences in the interaction of environmental and site factors. The devastating effect of the introduction of *P. cinnamomi* on WA native vegetation has been well documented. The implications of climate change must also be considered, whereby conditions more conducive to

the pathogens and to disease expression may be created (e.g. increasingly frequent summer rainfall events). The presence of a swarm of Clade 6 hybrids in WA natural ecosystems (unpubl. data) adds a new dimension of urgency to the need to prevent the further transfer of *Phytophthoras* between sites.

Collectively, all of these *Phytophthora* taxa should now be regarded as a threat to native ecosystems, and they need to be managed in the same way as *P. cinnamomi* in terms of minimising their further spread. This can be achieved simply by implementing proven local dieback hygiene practices such as restricting activities to dry soil conditions, and ensuring that vehicles and footwear are thoroughly clean when moving from an infested to a clean area. This may be especially important for the *Phytophthora* taxa that are not yet widespread or common.

### Conclusions

Clade 6 is arguably the most unusual and distinctive major clade in *Phytophthora*. Predominantly aquatic Clade 6 species have developed a highly successful ecological strategy as rapid colonisers of leaf litter and opportunistic pathogens of woody plants under flooded conditions while other species are aggressive soilborne pathogens of agricultural and horticultural crops with either wide (e.g. *P. megasperma*), moderately wide (e.g. *P. rosacearum*) or narrow (e.g. *P. taxon asparagi*) host ranges. Most extraordinarily for Clade 6 *Phytophthoras*, the recently described *P. pinifolia* has evolved towards an aerial lifestyle, producing caducous non-proliferating sporangia and causing an epidemic needle blight in *Pinus radiata* plantations in Chile (Durán et al. 2008). However, since *P. pinifolia* is a clonal introduction in Chile (Durán et al. 2010) its ecological niche in its unknown centre of origin may be different. Within Clade 6, the high number of taxa, their wide variation of habitats, lifestyle and breeding systems (heterothallic, homothallic, silent A1, potentially or sporadically self fertile, and fully sterile) and their tendency to hybridise may necessitate a rethinking of species boundaries and species recognition for this clade.

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