

Section 4

Discussion and Conclusions

During this study, over 1100 collections have been examined including 70 type collections. One hundred and fourteen taxa including ten new species are distinguished and are placed in the genus *Septoria*. The distribution and listing of species by host plant family as a result of this study are to be found in Appendices A and B. Most of the collections examined are currently held in herbaria within Australia and no systematic collecting has been undertaken by the author. Comparison with type material or authentic material available has been invaluable in establishing the identity of a number of species previously misidentified. Keys are presented for species on hosts in the families Apiaceae, Asteraceae, Caryophyllaceae, Fabaceae, Mimosaceae and Poaceae.

Five taxa still need to be redispersed to other genera based on non-pycnidial conidiomatal morphology, these being *S. paeoniae* var *berolinensis* on *Paeonia*, *S. pisi* on *Pisum*, *S. selenophomoides* on various hosts in the Orchidaceae, *S. transversalis* on *Aspidistra* and *S. unedonis* on *Arbutus*. In the absence of examination of the type collection those taxa are placed under their currently available name in *Septoria*. In addition, six taxa recognised in this study are at present unnamed since there are either several potential names that appear to be available or the available collections do not fit an established taxon on that host or on a closely related host in the family. These include *Septoria* sp. aff. *associata* on *Carduus*, *Septoria* sp. aff. *carthamicola* on *Carthamus*, *Septoria* sp. aff. *cocoina* on *Arecastrum* and “Kentia”, *Septoria* sp. cf. *noli-tangere* on *Impatiens*, *Septoria* sp. on *Lathyrus* and *Septoria* sp. on various hosts including *Boronia*, *Coleonema*, *Hedera*, *Lonicera*, *Ligustrum*, *Prunus*, *Rosa* and *Stephanotis*. This last taxon is regarded as having a possible saprophytic or endophytic mode of existence from its occurrence on hosts in several plant families and being associated with dead, dying or incubated leaf tissue, and, being almost impossible to separate morphologically on any of the hosts. At this time, any attempt to apply a name to this taxon is fraught with difficulty, several names being available that could be applicable to the taxon based on a brief glance at Saccardo’s *Sylloge Fungorum* where several species described from languid or dried leaves with very similar morphological characters are to be found.

In Section 1 two hundred and thirty records and reports of species of *Septoria* occurring in Australia were presented. In addition to the one hundred and fourteen taxa recognised in this study, eighty one records of *Septoria* are unconfirmed on the basis of lack of herbarium collections being available or, in several cases, a misinterpretation of the literature. This large number of unconfirmed records serves to emphasise the need for specimens to be placed in systematic reference collections for future study. In the ever increasing international trade in agricultural commodities, quarantine decisions are made on accurate knowledge of pest organisms within the country. Published records without the availability of reference material on which such records are based can only hamper the decision-making process.

Ten species of *Septoria* dealt with in this study have already been redisposed to other genera or have been reclassified in this treatment. Those previously redisposed are *S. avenae*, *S. nodorum*, *S. chenopodii* and *S. atriplicis* which have been transferred to the genus *Stagonospora*, *S. lepidospermatis* transferred to *Clypeopycnis* (Sutton & Pascoe 1989) and *S. martiniana* transferred to *Cytostagonospora* (Sutton & Swart 1986). In this treatment a further four species are transferred to other genera based on type examination or redisposed to other genera. *Septoria azaleae* is transferred to *Phloeospora* based on the acervular nature of the conidioma and enteroblastic percurrent conidiogenesis, *Septoria martiniae* is transferred to *Septocytia* based on the multi-locular nature of the conidioma and holoblastic sympodial conidiogenesis, *Septoria thelymitrae* is transferred to *Selenophoma* based on the enteroblastic percurrent conidiogenesis and lunate aseptate conidia. The un-named species of *Septoria* reported on *Matthiola incana* is identified as *Ascochyta matthiolae* based on the non-proliferating enteroblastic conidiogenesis and the mostly 1-septate conidia.

Septoria lagenophorae is recognised as a hyperparasite, occurring mainly in association with rusts but occasionally other fungi. Such a mode of existence is not unusual, there being several genera of hyperparasitic fungi. The only other species of *Septoria* described as a possible hyperparasite is *S. ficariaeoides*, described originally from rusts of the Ranunculaceae and possibly reported by Jorstad (1967). The possible conspecificity of *S. lagenophorae* and *S. ficariaeoides* is still to be investigated.

The typification and history of the development of taxonomic concepts, and current morphological criteria used to distinguish *Septoria* have been discussed previously (Section 1). All taxa accepted in this study as species of *Septoria* are based on the presence of pycnidial conidiomata and filiform septate conidia. Morphologically similar species on hosts in different families are retained as separate

species at present. Considerable variation in conidiogenesis is observed across the taxa studied but is accepted as being intrinsic to the current concept of the genus. Based on the premise of Minter (1987) and the recent confirmation by Verkley (1998a, 1998b) of the plasticity of conidiogenesis and that enteroblastic non-progressive, enteroblastic percurrent and sympodial holoblastic conidiogenesis are merely differences in development and do not represent taxonomic characters at the generic level, the use of conidiogenesis at that level is now regarded as an interim measure in the systematics of anamorphic fungi.

In this study, the primary criterion for delimitation of species has been scrutiny of named taxa at the host plant family level. Conidial width has been determined to be one of the more stable characters that can be used when delimiting species and has been used in the construction of keys to recognised species in many families. Examination of a large number of collections has shown that variation in conidium width is small and varies less as the conidium gets narrower. Over the large number of species and collections examined, variation of 0.5 μ m appears to be normal for most conidia measured, with variation up to 1 μ m being more common as conidia get wider. Secondary characters such as length and septation have also proven to be useful particularly for separating several species in the Poaceae, such as *S. halophila* (1-septate) from *S. passerinii* (1-3 septate), and *S. bromi* (2-septate) from *S. calamagrostidis* (3-4 septate).

Sutton (1980) suggested that three subgroupings within *Septoria* could possibly be distinguished based on conidiogenesis as follows: (i) mainly holoblastic sympodial species as in the type species *S. cytisi* e.g. *S. chrysanthemella*, (ii) species with phialidic conidiogenesis e.g. *S. apiicola* and *S. tritici*, and (iii) simple holoblastic species without proliferation e.g. *S. adanensis*. However, Farr (1992) noted that both sympodial and percurrent proliferation of conidiogenous cells could be found in the type species *S. cytisi*.

The Australian species of *Septoria* recognised in this study have exhibited variation in mode of conidiogenesis and five groups based on observed proliferation of the conidiogenous locus can be recognised. These groupings are presented below. Within the recognised groupings, species are arranged in alphabetical order.

Group I: Holoblastic (non-proliferating)

<i>S. adanensis</i>	<i>S. confluens</i>	<i>S. hydrocotyles</i>	<i>S. schizeilematis</i>
<i>S. aesculi</i>	<i>S. crataegi</i>	<i>S. ixodiae</i>	<i>S. silybi</i>
<i>S. agropyrina</i>	<i>S. cryptica</i>	<i>S. lamiicola</i>	<i>S. tabacina</i>
<i>S. anaxaea</i>	<i>S. cymbopogonis</i>	<i>S. lavandulae</i>	<i>S. wahlenbergii-australiensis</i>
<i>S. anenomes</i>	<i>S. cucurbitacearum</i>	<i>S. obesa</i>	<i>S. williamsiae</i>
<i>S. antirrhini</i>	<i>S. cymbopogonis</i>	<i>S. paradisi</i>	<i>Septoria</i> sp. aff. <i>cathamicola</i>
<i>S. betae</i>	<i>S. dianthi</i>	<i>S. passifloricola</i>	<i>Septoria</i> sp. aff. <i>cocoina</i>
<i>S. betulae</i>	<i>S. erigerontis</i>	<i>S. perforans</i>	<i>Septoria</i> sp. cf. <i>noli-tangere</i>
<i>S. canberrica</i>	<i>S. ficariae</i>	<i>S. phyllodorum</i>	
<i>S. carthami</i>	<i>S. grampianensis</i>	<i>S. podolepidis</i>	
<i>S. centellae</i>	<i>S. helichrysicola</i>	<i>S. roemeriana</i>	

Group II: Holoblastic Sympodial +/- Percurrent Proliferation

<i>S. aciculosa</i>	<i>S. galinsogae</i>	<i>S. lycopersici</i>	<i>S. sambucina</i>
<i>S. armeriae</i>	<i>S. gerberae</i>	<i>S. macalpinei</i>	<i>S. sonchi</i>
<i>S. australiae</i>	<i>S. goodeniicola</i>	<i>S. malvicola</i>	<i>S. stellariae</i>
<i>S. centaureae</i>	<i>S. hardenbergiae</i>	<i>S. menyanthes</i>	<i>S. suaedae-australis</i>
<i>S. cerastii</i>	<i>S. helianthi</i>	<i>S. menyanthicola</i>	<i>S. tetraethecae</i>
<i>S. chrysanthemella</i>	<i>S. humuli</i>	<i>S. minima</i>	<i>S. thuemeniana</i>
<i>S. citri</i>	<i>S. lactucae</i>	<i>S. passerinii</i>	<i>S. urens</i>
<i>S. colensoi</i>	<i>S. lamentana</i>	<i>S. petroselini</i>	<i>S. verbenae</i>
<i>S. convolvuli</i>	<i>S. lepidii</i>	<i>S. phlogis</i>	<i>S. vignae</i>
<i>S. divaricata</i>	<i>S. linicola</i>	<i>S. ribis</i>	<i>Septoria</i> sp. (various hosts)
<i>S. exotica</i>	<i>S. lobeliae</i>	<i>S. rubi</i>	

Group III: Enteroblastic (non-proliferating)

<i>S. apiicola</i>	<i>S. elymi</i>	<i>S. polygonati</i>	<i>S. varia</i>
<i>S. aureocorona</i>	<i>S. halophila</i>	<i>S. polygonorum</i>	<i>S. violae</i> f. <i>odoratae</i>
<i>S. bromi</i>	<i>S. hydrocotylicola</i>	<i>S. triseti</i>	<i>Septoria</i> sp. aff. <i>associata</i>
<i>S. calmagrostidis</i>	<i>S. gaurina</i>	<i>S. tritici</i>	<i>Septoria</i> sp. (<i>Lathyrus</i>)
<i>S. cyclaminis</i>	<i>S. lagenophorae</i>	<i>S. urticae</i>	

Group IV: Enteroblastic Percurrent

<i>S. gladioli</i>	<i>S. silenicola</i>	<i>S. zeicola</i>
<i>S. pisoniae</i>	<i>S. sisymbrii</i>	

Group V: Holoblastic (non-proliferating) + Enteroblastic (non-proliferating)

<i>S. geranii</i>

The forty three Australian species placed in Group II belong in the genus *Septoria* sensu stricto as defined by Sutton (1980) and emended by Farr (1992). All these species have conidiogenesis which has been observed to be holoblastic with sympodial proliferation and in several cases enteroblastic percurrent proliferation as well.

Forty one species have been placed in Group I where conidiogenesis is observed to be holoblastic without proliferation. One Australian species in this group (*S. ixodiae*) has a teleomorph currently identified as *Mycosphaerella ixodiae* and one other species, *S. crataegi* has a reported *Mycosphaerella* teleomorph (Jørstad 1965) suggesting that the placement in *Septoria* (of some species at least) found in this group is appropriate at present.

The nineteen species placed into Group III have enteroblastic conidiogenesis without proliferation. Whether the situation observed under the light microscope is one of cryptic percurrent proliferation or simple phialidic conidiogenesis without proliferation is yet to be shown but phialidic conidiogenesis may prove to be the basis of a segregate group as suggested by Sutton (1980) and Verkley (1998b). At the limit of the light microscope, the observation of thickening and the presence of a collarete seem to indicate proliferation but it requires electron microscopy to truly elucidate the process involved.

Only in three plant families is there observed consistent conidiogenesis of the species recognised there. The species in the Menyanthaceae (*S. menyanthes* and *S. menyanthicola*) and Polemoniaceae (*S. divaricata* and *S. phlogis*) exhibit holoblastic conidiogenesis with sympodial proliferation. All three species in the Ranunculaceae (*S. anenomes*, *S. ficariae* and *S. williamsiae*) exhibit holoblastic conidiogenesis without proliferation. In all other plant families the species are placed into several of the groups.

There still remains the possibility of segregation of *Septoria* into recognisable groups based on conidiogenesis with sympodial and percurrent proliferation, simple holoblastic and simple enteroblastic (phialidic) groups providing the framework. One species, *S. geranii*, exhibits both non-proliferating enteroblastic and holoblastic conidiogenesis (Group V). Further study is required to ascertain if proliferation in this species does occur which would place the species in *Septoria* sensu stricto.

The five species placed in Group IV, which exhibit only enteroblastic percurrent proliferation, could easily be accommodated within the genus *Stagonospora* as defined by Sutton (1980) but, given the

difficulty in now defining these two genera using conidiogenesis as a basis for generic separation, the placement of these species in *Septoria* is open to interpretation. The placement of *Septoria nodorum* and *S. avenae* into *Stagonospora* is based on the knowledge of their respective teleomorphs. In the anamorphic state both of those species have only been observed to be holoblastic without proliferation and in the absence of knowledge of any teleomorphic connection would be placed into *Septoria* (Group I) as defined here. Association with a teleomorph is lacking for nearly all species in this study but where it is recognised it does not correlate with the conidiogenesis observed in the anamorph. In this study five species of *Mycosphaerella* are described with their anamorphs including *M. aureocorona* sp. nov. associated with *Septoria aureocorona* on *Acacia*. The anamorph-teleomorph association is summarised in Table 4.1.

Table 4.1: Australian species of *Septoria* and their *Mycosphaerella* teleomorphs related to conidiogenesis

Teleomorph	Anamorph	Conidiogenesis
<i>Mycosphaerella</i> sp. (<i>Centella</i>)	<i>S. centellae</i>	Holoblastic (non-proliferating)
<i>Mycosphaerella graminicola</i>	<i>S. tritici</i>	Enteroblastic (non-proliferating)
<i>Mycosphaerella suaedae-australis</i>	<i>S. suaedae-australis</i>	Holoblastic (sympodial)
<i>Mycosphaerella ixodiae</i>	<i>S. ixodiae</i>	Holoblastic (non-proliferating)
<i>Mycosphaerella aureocorona</i>	<i>S. aureocorona</i>	Enteroblastic (non-proliferating)

Since Sutton (1980), the recognition of variation in conidiogenesis in many species has appeared to make the genus heterogenous. However, the realisation that this variation is natural and intrinsically developmental has led to the situation where anateleomorphic species are consequently difficult to place in any genus with absolute certainty. The differences between pycnidial genera such as *Stagonospora* and *Septoria*, with hyaline multi-septate conidia is negligible at the anamorphic level. The current reliance on small differences in conidial morphology (bacilliform in *Stagonospora*) and, as at Sutton (1980), percurrent conidiogenesis in *Stagonospora* vs. sympodial holoblastic conidiogenesis in *Septoria*, is a separation now in considerable doubt with enteroblastic percurrent proliferation now known from the type species, *S. cytisi*. This leaves only minor conidial differences and known teleomorphic connections in *Leptosphaeria* (with its segregates) and *Mycosphaerella* as the distinguishing features. However, the problem of separating morphologically close anamorphic

genera is not new in the study of the anamorphic fungi and finding a workable generic concept is the current challenge.

The correlation with teleomorph genera can also present further complications as there can often be difficulty separating apparently indistinguishable teleomorphs, one example of which is *Mycosphaerella fijiensis* and *M. musicola* which, although capable of being distinguished at the molecular level, at the morphological level can only be distinguished by their anamorphs and the symptoms they produce on banana leaves (Samuels & Seifert 1995). The morphologically similar anamorphic genera *Drechslera*, *Bipolaris* and *Exserohilum* (formerly all placed in *Drechslera*) are found to be separable based on certain characters such as conidial germination patterns, hilum characters and the quite morphologically separate teleomorphs *Pyrenophora*, *Cochliobolus* and *Setosphaeria* (Alcorn 1988). The research into *Drechslera* was driven by a need by both mycologists and plant pathologists working with the genus (which includes plant pathogens common on many cultivated Poaceae) to identify anamorphs due to the rare occurrence of the teleomorphs in nature.

A similar situation now exists in *Septoria* and other septorioid genera. The use of conidiogenesis as a generic separation character is now shown to be mostly unworkable, and has left few characters to separate morphologically similar genera, particularly where the teleomorph is unknown. Conidiomatal morphology, conidiogenesis, conidial morphology and host specificity are still the only characters which can be used to recognise anamorphs at the generic level. Sutton (1996) suggested that groupings according to conidiogenous events and conidiomatal structure offered the prospect of a more rational means of separation of taxa within *Mycosphaerella* at the sub-generic level due to the gross heterogeneity exhibited by the genus. Thirty two anamorphic genera connected to *Mycosphaerella* (including *Septoria*) are keyed out by von Arx (1983), and twenty two anamorphs are listed by Sivanesan (1984). However, that does not offer an immediate solution for anamorphic recognition. The gathering of other taxonomic characters such as correct interpretation of conidiogenesis based on ultrastructure, conidial germination and teleomorph production *in-vitro*, appear to be possible areas for further research.

In Australia the species of *Septoria* recognised occur on hosts in fifty five plant families (Appendix A). The largest numbers of species occur on hosts in the families Asteraceae and Poaceae, two of the largest and most cosmopolitan families. There are no species as yet confirmed on hosts in the family Myrtaceae, the report of a species of *Septoria* on *Leptospermum* in Western Australia being unconfirmed due to lack of an available collection. The reasons for the absence of *Septoria* spp. on

Eucalyptus and the Myrtaceae generally in Australia probably relate to the origins of the genus. *Eucalyptus* in particular appears to be of Australian origin and evolved in isolation, being not very closely related to the shrubby myrtaceous elements of the flora although they share a common ancestry (Barlow 1981). Several other pathogenic genera such as *Mycosphaerella* and the anamorphic *Sonderhenia* and *Kirramyces* (now *Phaeophleospora*) have obviously evolved with *Eucalyptus* and appear to occupy the niche of *Septoria*. On the myrtaceous genus, *Callistemon*, is found *Lecanosticta gaubae*, an acervular genus with brown verrucose conidia being produced sympodially and percurrently (analogous to *Phloeospora*) with a teleomorph currently placed as *Mycosphaerella gaubae* v. Arx & O. Constaninescu, but appears to be a candidate for the genus *Eruptio* (Barr 1996).

In contrast, there have been four species of *Septoria* described from *Acacia*, another conspicuous member of the Australian flora. Only one species of *Septoria* has been described from *Acacia* outside Australia, this being the curious example of *Septoria acaciae*, described from commercial plantings of *A. paradoxa* (given as *A. armata*) in Denmark but not been collected from its host in Australia (Sutton & Pascoe 1987). The recognition of *S. anaxaea* on *Senecio* spp. in Australia is also curious since it has not been reported elsewhere from outside Italy (the type locality) since its description. Also there is as yet no species of *Septoria* known from the family Proteaceae in Australia, or in South Africa where the family is well represented. The spread of taxa across broad groupings of plants shows that thirty one taxa occur on native plants, thirty two on introduced ornamental plants, twenty on food and fibre crop plants, and twenty seven on weeds. Twenty five of the recognised species are regarded as endemic, fifteen of these being already known before this study, occurring on native plant hosts and distinct from species already recognised on those genera or families elsewhere throughout the world. An additional ten new species have been recognised on native hosts as a result of this study.

The Australian flora is derived from immigration on several fronts (Barlow 1981). It is suggested that a subtropical migration from South Africa via India and Madagascar may have persisted until the middle-late Cretaceous. After this route was broken by continental drift there still existed a southern migratory route from South America via Antarctica until the Oligocene when forests of *Nothofagus*, Proteaceae and Myrtaceae were in existence. At the beginning of the Tertiary the Gondwanan flora in existence was derived from these migrations. Under conditions of geographical separation, the flora differentiated from the existing original Gondwanan stock from the early Tertiary (when Australia separated from Antarctica) until its contact with the Sunda plate in the Miocene. The subsequent introduction of the Indomalayan flora occurred during the late Tertiary. In addition, this contact with

the Indomalayan region allowed for immigration of a number of typically northern temperate genera via the uplifted mountain systems in Malaya and New Guinea; some of these genera include *Hydrocotyle*, *Ranunculus* and *Viola* (Barlow 1981, Burbridge 1960).

Two species of *Septoria* recognised in this study show some interesting mycogeographical features. The recognition of the occurrence of *S. halophila* on species of *Hordeum* and *Poa* (Poaceae) across much of Australia suggests that it occupies a niche in the Southern Hemisphere similar to that of *S. passerinii* in the Northern Hemisphere. *Septoria apiicola* presents a much more confused picture, the original type host being *Apium australe*, a species with a Southern Hemisphere distribution along with *A. prostratum*, found in Australia and New Zealand. The conidial width found in *A. prostratum* collections as well as those given by authors for the type collection of *S. apiicola* is 1-2 μ m, which is narrower than most authors give for *S. apiicola* found on cultivated *Apium* spp. (2-2.5 μ m). This suggests that two taxa are involved which although cross-infective can be distinguished morphologically. Given that the cultivated *Apium* spp. come from northern temperate areas it is more likely that the name *S. apiicola* should be used for the narrow spored taxon with a southern temperate distribution and possibly *S. apii* for the wider spored taxon found commonly on cultivated *Apium* spp.

Most of the species of *Septoria* recognised in this study are temperate in distribution, occurring across the cool southern temperate to northern warm temperate regions of Australia. Few species are recognised from the tropical regions, none being recorded from the tropical areas of northern Queensland, and the only species recorded at present from the Northern Territory being *Septoria lactucae*. This probably reflects the difficulty and, hence, lack of systematic collecting in these areas. Shaw (1984) lists only sixteen species of *Septoria* for Papua New Guinea and at least thirteen of these are from cultivated crop or ornamental plants, only one record of a *Septoria* sp. occurring on a native grass (*Polytoca macrophylla* Benth.), and *Septoria australiae* on *Viola betonicifolia*. *Viola betonicifolia* is known throughout Australia and extends into Malesia and Asia. However, the records of plant parasitic fungi throughout the Indomalayan region are few and are usually biased toward crop plants, very little having been published on the occurrence of native host pathogens.

Species such as *S. gaurina* on *Oenothera* (Onagraceae) and *S. sambucina* on *Sambucus* (Sambucaceae) appear to have been introduced from the U.S.A. where they occur on their native hosts and are quite distinct from their European counterparts. Some introduced species have extended their host range onto native host plants, two examples of which are *S. tritici* which has been identified

on the native grass hosts *Danthonia* and *Dichelachne* and, *S. stellariae* on *Drymaria*. In essence, the species of *Septoria* recognised in Australia reflect in part the origins of its vascular plant flora, having both endemic and introduced elements.

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APPENDIX A : Distribution of Australian species of *Septoria* by host family

Host Family	No. of <i>Septoria</i> Species	Host genera	New <i>Septoria</i> Species	No. Endemic Species of <i>Septoria</i>
Aizoaceae	1	1		1
Apiaceae	6	11		1
Araliaceae	1	1		
Arecaeae	1	2		
Asclepiadaceae	1	1		
Asteraceae	22	22	2	5
Balsaminaceae	1	1		
Betulaceae	1	1		
Brassicaceae	2	3		
Campanulaceae	2	2	1	1
Cannabidaceae	1	1		
Caprifoliaceae	1	1		
Caryophyllaceae	4	5		
Chenopodiaceae	2	2		1
Convolvulaceae	1	1		
Cucurbitaceae	1	3		
Ericaceae	1	1		
Euphorbiaceae	1	1		
Fabaceae	4	6		1
Geraniaceae	2	3		1
Goodeniaceae	1	1		1
Grossulariaceae	1	1		
Hippocastanaceae	1	1		
Iridaceae	1	1		
Lamiaceae	2	2		
Liliaceae	2	2		
Linaceae	1	1		
Malvaceae	1	1		
Menyanthaceae	2	2	1	1
Mimosaceae	4	1		4
Myoporaceae	1	1		
Nyctaginaceae	1	1	1	1
Oleaceae	1	1		
Onagraceae	1	1		
Orchidaceae	1	3		
Paeoniaceae	1	1		
Passifloraceae	1	1		
Plantaginaceae	1	1		1
Plumbaginaceae	1	1		
Poaceae	12	25	3	3
Polemoniaceae	2	1		
Polygonaceae	1	1		
Primulaceae	1	1		
Ranunculaceae	3	3	1	1
Rosaceae	5	5		
Rubiaceae	1	1		
Rutaceae	2	4		

Host Family	No. of <i>Septoria</i> species	Host Genera	New <i>Septoria</i> species	No. Endemic species of <i>Septoria</i>
Sambucaceae	1	1		
Scrophulariaceae	3	3	1	1
Solanaceae	2	2		1
Thymeleaceae	1	1		
Tremandraceae	1	1		
Urticaceae	1	1		
Verbenaceae	1	1		
Violaceae	2	2		

APPENDIX B : Australian species of *Septoria* listed by host family

AIZOACE

Carpobrotus aequilaterus *S. confluens*

APIACEAE

Apium graveolens *S. apiicola*
A. garaveolens var. *rapaceum* *S. apiicola*
A. prostratum *S. apiicola*
Centella asiatica *S. centellae*, *S. hydrocotyles*,
S. hydrocotylicola
Coriandrum sativum *S. petroselini*
Hydrocotyle acutiloba *S. hydrocotyles*, *S. hydrocotylicola*
H. hirta *S. hydrocotyles*
H. laxiflora *S. hydrocotyles*, *S. hydrocotylicola*
H. pedicellosa *S. hydrocotyles*
Petroselinum crispum *S. petroselini*
Schizeilema fragoseum *S. schizeilematis*

ARALIACEAE

Hedera helix *Septoria* sp.

ARECACEAE

Arecastrum romanzoffianum *Septoria* sp. aff. *S. cocoina*
Howea sp. *Septoria* sp. aff. *S. cocoina*

ASCLEPIADACEAE

Stephanotis floribunda *Septoria* sp.

ASTERACEAE

Actites megalocarpa *S. sonchi*
Arctotheca calendula *S. perforans*
Carduus tenuiflorus *Septoria* sp. aff. *S. associata*
Carthamus tinctorius *S. carthami*, *Septoria* sp. aff. *S. carthamicola*
Centaurea cyanea *S. centaureae*
Chrysanthemum indicum *S. adanensis*, *S. chrysanthemella*, *S. obesa*
C. leucanthemum *S. minima*
C. morifolium *S. adanensis*, *S. obesa*
Chrysanthemum sp. *S. chrysanthemella*, *S. minima*
Conyza albida *S. erigerontis*
C. bonariensis *S. erigerontis*
Duchesnea indica/ *Frommeella duchesnae* *S. lagenophorae*
Galinsoga parviflora *S. galinsogae*
Gerbera jamesonii *S. gerberae*
Helianthus annuus *S. helianthi*
H. argophyllus *S. helianthi*
Helichrysum ramosissimum *S. helichrysicola*
Hypochaeris radicata/ *Puccinia* *S. lagenophorae*
H. glabra/ *Puccinia* *S. lagenophorae*
Ixodia achilleioides *S. ixodiae*
I. alata *S. ixodiae*
Lactuca sativa *S. lactucae*
L. serriola *S. lactucae*
Lagenophora billardieri/ *Puccinia* *S. lagenophorae*
Lagenophora sp./ *Puccinia* *S. lagenophorae*

<i>Leucanthemum maximum</i>	<i>S. obesa</i>
<i>Olearia argophylla</i>	<i>S. paradisi</i>
<i>O. stellulata</i>	<i>S. paradisi</i>
<i>Podolepis jaceoides</i>	<i>S. podolepidis</i>
<i>P. neglecta</i>	<i>S. podolepidis</i>
<i>Senecio glomeratus</i> x <i>minimus</i>	<i>S. anaxaea</i>
<i>S. gunnii</i>	<i>S. anaxaea</i>
<i>S. quadridentatus</i>	<i>S. anaxaea</i>
<i>S. vagans</i>	<i>S. anaxaea</i>
<i>Senecio</i> sp.	<i>S. anaxaea</i>
<i>Silybum marianum</i>	<i>S. silybi</i>
<i>Sonchus oleraceus</i>	<i>S. silybi</i>
BALSMINACEAE	
<i>Impatiens</i> sp.	<i>Septoria</i> sp. aff. <i>S. noli-tangere</i>
BETULACEAE	
<i>Betula papyrifera</i>	<i>S. betulae</i>
<i>B. pendula</i>	<i>S. betulae</i>
<i>B. platyphylla</i> var. <i>japonica</i>	<i>S. betulae</i>
<i>B. pubescens</i>	<i>S. betulae</i>
<i>Betula</i> sp.	<i>S. betulae</i>
BRASSICACEAE	
<i>Cardaria draba</i>	<i>S. lepidii</i>
<i>Sinapis arvensis</i>	<i>S. sisymbrii</i>
<i>Sisymbrium officinale</i>	<i>S. sisymbrii</i>
CAMPANULACEAE	
<i>Pratia purpurascens</i>	<i>S. lobeliae</i>
<i>Wahlenbergia gracilentata</i>	<i>S. wahlenbergii-australiensis</i>
<i>W. stricta</i>	<i>S. wahlenbergii-australiensis</i>
CANNBIDACEAE	
<i>Humulus lupulus</i>	<i>S. humuli</i>
CAPRIFOLIACEAE	
<i>Lonicera caprifolium</i>	<i>Septoria</i> sp.
CARYOPHYLLACEAE	
<i>Cerastium glomeratum</i>	<i>S. cerastii</i>
<i>Dianthus barbatus</i>	<i>S. dianthi</i>
<i>D. caryophyllus</i>	<i>S. dianthi</i>
<i>Drymaria diandra</i>	<i>S. stellariae</i>
<i>Silene gallica</i>	<i>S. silenicola</i>
<i>Stellaria media</i>	<i>S. stellariae</i>
CHENOPODIACEAE	
<i>Beta vulgaris</i> ssp. <i>cicla</i>	<i>S. betae</i>
<i>B. vulgaris</i> ssp. <i>vulgaris</i>	<i>S. betae</i>
<i>Suaeda australis</i>	<i>S. suaetae-australis</i>
CONVOLVULACEAE	
<i>Convolvulus arvensis</i>	<i>S. convolvuli</i>
CUCURBITACEAE	
<i>Citrullus vulgaris</i>	<i>S. cucurbitacearum</i>
<i>Cucumis sativus</i>	<i>S. cucurbitacearum</i>

<i>Cucurbita maxima</i>	<i>S. cucurbitacearum</i>
<i>Cucurbita pepo</i>	<i>S. cucurbitacearum</i>
<i>Cucurbita</i> sp.	<i>S. cucurbitacearum</i>
ERICACEAE	
<i>Arbutus unedo</i>	<i>S. unedonis</i>
EUPHORBIACEAE	
<i>Euphorbia parvicaruncula</i>	<i>S. thuemeniana</i>
<i>E. peplus</i>	<i>S. thuemeniana</i>
FABACEAE	
<i>Brachysema sericeum</i>	<i>S. hardenbergiae</i>
<i>Hardenbergia violacea</i>	<i>S. hardenbergiae</i>
<i>Lathyrus odoratus</i>	<i>Septoria</i> sp.
<i>Pisum sativum</i>	<i>Septoria pisi</i>
<i>Psoralea adscendens</i>	<i>S. hardenbergiae</i>
<i>Vigna lanceolata</i>	<i>S. vignae</i>
<i>V. unguiculata</i> ssp. <i>sesquipedalis</i>	<i>S. vignae</i>
<i>V. unguiculata</i> ssp. <i>unguiculata</i>	<i>S. vignae</i>
GERANIACEAE	
<i>Erodium crinitum</i>	<i>S. canberrica</i>
<i>Geranium antrorsum</i>	<i>S. geranii</i>
<i>G. homenum</i>	<i>S. geranii</i>
<i>G. neglectum</i>	<i>S. geranii</i>
<i>G. pilosum</i>	<i>S. geranii</i>
<i>G. solanderi</i>	<i>S. geranii</i>
<i>Geranium</i> sp.	<i>S. geranii</i>
<i>Pelargonium australe</i>	<i>S. canberrica</i>
<i>Pelargonium</i> sp. (? <i>rodneyanum</i>)	<i>S. canberrica</i>
GOODENIACEAE	
<i>Goodenia ovata</i>	<i>S. goodeniicola</i>
GROSSULARIACEAE	
<i>Ribes nigrum</i>	<i>S. ribis</i>
<i>Ribes rubrum</i>	<i>S. ribis</i>
<i>Ribes</i> sp.	<i>S. ribis</i>
HIPPOCASTANACEAE	
<i>Aesculus hippocastaneum</i>	<i>S. aesculi</i>
IRIDACEAE	
<i>Gladiolus</i> sp. cult.	<i>S. gladioli</i>
LAMIACEAE	
<i>Lamium amplexicaule</i>	<i>S. lamiicola</i>
<i>Lavandula vera</i>	<i>S. lavandulae</i>
<i>Lavandula</i> sp.	<i>S. lavandulae</i>
LILIACEAE	
<i>Aspidistra elatior</i> var. <i>variegata</i>	<i>S. transversalis</i>
<i>Polygonatum</i> sp.	<i>S. polygonati</i>
LINACEAE	
<i>Linum usitatissimum</i>	<i>S. linicola</i>

MALVACEAE

*Malva neglecta**S. malvicola*

MENYANTHACEAE

*Nymphoides crenata**S. menyanthis**N. exiliflora**S. menyanthicola**Villarsia exaltata**S. menyanthicola**Villarsia* sp.*S. menyanthicola*

MIMOSACEAE

*Acacia harpophylla**S. phyllodiorum**A. myrtifolia**S. grampianensis**A. retinodes**S. aureocorona**A. saligna**S. aureocorona**A. verniciflua**S. lamentana**Acacia* sp. (? *suaveolens*)*S. aureocorona**Acacia* sp.*S. phyllodiorum*

MYOPORACEAE

*Myoporum debile**S. colensoi**M. insulare**S. colensoi**M. montanum**S. colensoi**M. viscosum**S. colensoi*

NYCTAGINACEAE

*Pisonia grandis**S. pisoniae*

OLEACEAE

*Ligustrum ovalifolium**Septoria* sp.*Ligustrum* sp.*Septoria* sp.

ONAGRACEAE

Oenothera spp.*S. gaurina*

ORCHIDACEAE

Bulbophyllum sp.*S. selenophomoides**Cattleya* sp.*S. selenophomoides**Dendrobium attenuatum**S. selenophomoides**D. biggibum**S. selenophomoides**D. canaliculatum**S. selenophomoides**D. discolor**S. selenophomoides**D. kingianum**S. selenophomoides**D. semifuscum**S. selenophomoides**D. schneiderae**S. selenophomoides**D. smilliae**S. selenophomoides**D. speciosum**S. selenophomoides**Dendrobium* sp.*S. selenophomoides*

PAEONIACEAE

*Paeonia lactiflora**S. paeoniae* var. *berolinensis**P. officinalis**S. paeoniae* var. *berolinensis*

PASSIFLORACEAE

*Passiflora edulis**S. passifloricola**P. edulis* x *flavicarpa**S. passifloricola**P. flavicarpa**S. passifloricola**P. quadrangularis**S. passifloricola*

PLANTAGINACEAE

<i>Plantago debilis</i>	<i>S. varia</i>
<i>P. gaudichaudiana</i>	<i>S. varia</i>
<i>P. hispidus</i>	<i>S. varia</i>
<i>P. varia</i>	<i>S. varia</i>

PLUMBAGINACEAE

<i>Armeria</i> sp. cult.	<i>S. armeriae</i>
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POACEAE

<i>Agrostis capillaris</i>	<i>S. triseti</i>
<i>Aira caryophylla</i>	<i>S. elymi</i>
<i>Avena fatua</i>	<i>S. calamagrostidis</i>
<i>A. ludoviciana</i>	<i>S. calamagrostidis</i>
<i>A. sativa</i>	<i>S. calamagrostidis</i>
<i>A. sterilis</i>	<i>S. calamagrostidis</i>
<i>Avena</i> sp.	<i>S. calamagrostidis, S. tritici</i>
<i>Briza maxima</i>	<i>S. elymi</i>
<i>Bromus arenarius</i>	<i>S. bromi</i>
<i>B. hordeaceus</i>	<i>S. bromi</i>
<i>B. molliformis</i>	<i>S. bromi</i>
<i>B. racemosus</i>	<i>S. bromi</i>
<i>Capillepedium spicigerum</i>	<i>S. capillepedii</i>
<i>Cymbopogon refractus</i>	<i>S. cymbopogonis</i>
<i>Danthonia caespitosa</i>	<i>S. tritici</i>
<i>Deyeuxia quadriseta</i>	<i>S. bromi</i>
<i>Dichelachne micrantha</i>	<i>S. tritici</i>
<i>D. rara</i>	<i>S. tritici</i>
<i>D. sciurea</i>	<i>S. calamagrostidis, S. tritici</i>
<i>Digitaria ciliaris</i>	<i>S. capillepedii</i>
<i>Echinopogon caespitosus</i>	<i>S. calamagrostidis</i>
<i>Ehrharta longiflora</i>	<i>S. triseti</i>
<i>Elymus scabrus</i>	<i>S. agropyrina, S. elymi</i>
<i>E. scabrus</i> var. <i>pleurinerve</i>	<i>S. agropyrina</i>
<i>Eulalia tricuspada</i>	<i>S. capillepedii</i>
<i>Glyceria</i> sp.	<i>S. bromi</i>
<i>Holcus lanatus</i>	<i>S. tritici</i>
<i>Hordeum glaucum</i>	<i>S. halophila, S. passerinii</i>
<i>H. leporinum</i>	<i>S. halophila, S. passerinii</i>
<i>H. murinum</i>	<i>S. halophila, S. passerinii</i>
<i>Lolium rigidum</i>	<i>S. tritici</i>
<i>Lophochloa pumila</i>	<i>S. triseti</i>
<i>Paspalum distichum</i>	<i>S. capillepedii</i>
<i>Phragmites australis</i>	<i>S. cryptica</i>
<i>Poa annua</i>	<i>S. halophila</i>
<i>Poa</i> sp.	<i>S. halophila</i>
<i>Triticum aestivum</i>	<i>S. tritici</i>
<i>Zea mays</i>	<i>S. zeicola</i>

POLEMONIACEAE

<i>Phlox drummondii</i>	<i>S. phlogis, S. divaricata</i>
<i>P. paniculata</i>	<i>S. phlogis</i>

POLYGONACEAE

<i>Polygonum lapathifolium</i>	<i>S. polygonorum</i>
<i>Polygonum</i> sp. (? minus)	<i>S. polygonorum</i>

PRIMULACEAE

Cyclamen sp.*S. cyclaminis*

RANUNCULACEAE

*Anemone nemorosa**S. anenomes**Clematis aristata**S. williamsiae**Ranunculus lappaceus**S. ficariae**Ranunculus* sp.*S. ficariae*

ROSACEAE

*Cratageus monogyna**S. cratagei**Fragaria x ananassa**S. aciculosa**Prunus dulcis**Septoria* sp.*Rosa* sp. cult*Septoria* sp.*Rubus fruticosus**S. rubi**R. hillii**S. rubi**R. idaeus**S. rubi**R. loganobaccus**S. rubi**R. moluccanus**S. rubi**R. parvifolius**S. rubi**R. rosifolius**S. rubi**R. vulgaris**S. rubi**R. ulmifolius**S. rubi**R. ursinus**S. rubi**Rubus* x cult.*S. rubi*

RUBIACEAE

Galium sp. (? *aparine*)*S. urens*

RUTACEAE

*Boronia muelleri**Septoria* sp.*Citrus aurantium**S. citri**C. limon**S. citri**C. sinensis**S. citri**C. paradisi**S. citri**C. limonia**S. citri**Citrus* sp.*S. citri**Coleonema* sp.*Septoria* sp.*Correa* sp.*S. citri*

SCROPHULARIACEAE

*Antirrhinum majus**S. antirrhini**Derwentiana derwentiana**S. macalpinii**D. derwentiana* ssp. *derwentiana**S. exotica**Hebe imperialis**S. exotica**H. speciosa**S. exotica**Hebe* x *speciosa**S. exotica**Hebe* x *veronica**S. exotica*

SOLANACEAE

*Lycopersicon esculentum**S. lycopersici**Nicotiana rosulata**S. tabacina**N. suaveolens**S. tabacina**N. velutina**S. tabacina**Nicotiana* sp.*S. tabacina*

THYMELEACEAE

Daphne sp.*S. roemeriana*

URTICACAE

*Urtica incisa**U. urens**S. urticae**S. urticae*

VERBENACEAE

*Verbena officinalis**S. verbenae*

VIOLACEAE

*Hymenanchera dentata**Viola betonicifolia**V. caleyana**V. hederacea**V. odorata**S. australiae**S. australiae**S. australiae**S. australiae**S. violae* f. *odoratae*

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