ICCC-12 Conference 2010 Florianópolis SC – Brazil September 2010

New species and populations in *Fusarium*: examples from the tropics

Ludwig H. Pfenning Associate Professor, Plant Pathology Department Universidade Federal de Lavras, Brazil Iudwig@ufla.br





Problems and constraints with regard to information on tropical species, populations and plant diseases

- 1. Data on geographical distribution of species and diversity in the tropics are more related to research activities than to facts;
- 2. Many regions are absolutely under-investigated. This is true also for the genus *Fusarium*;

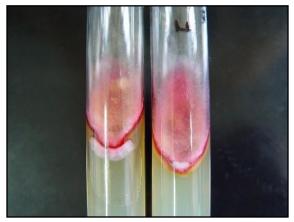


Problems and constraints with regard to information on tropical species, populations and plant diseases

- 3. Causal agents of diseases are not always the same in every country, region or continent;
- 4. Names used in older reports, like *Fusarium moniliforme* or *Fusarium roseum*, are no longer in use, and stand for a large diversity of different forms and populations.



Fusarium is a typical anamorph form-genus, with species sharing morphological markers





The culture



Microconidia



Macroconídia



Mono- and polyphialides

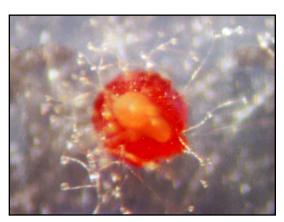


Chlamydospores



Telomorphs of *Fusarium* species

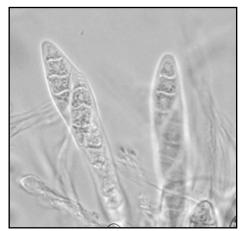










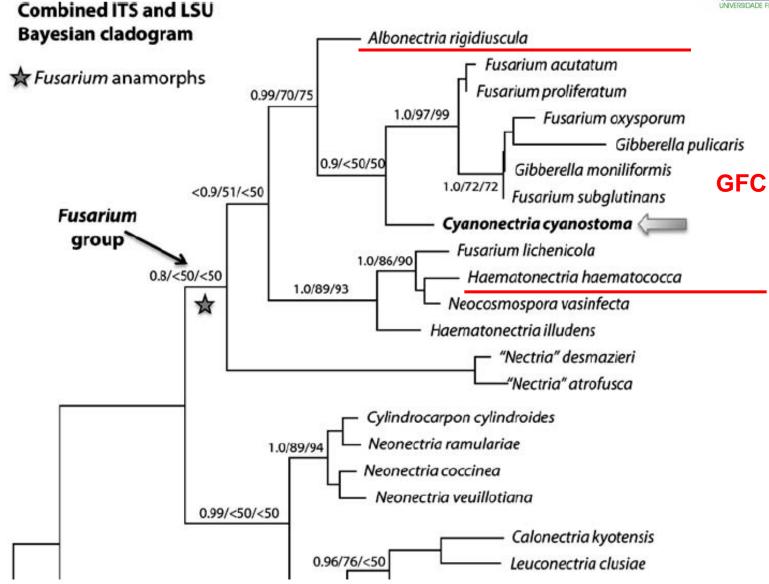


Gibberella

Haematonectria

Albonectria







Why is Fusarium important?

- Biology and ecology
- Economical importance
- Model for species concepts and molecular characterization

Plant pathogen

Soil fungus

→ soil and rhizosphere, root rots

Vascular fungus

 \rightarrow wilts, dieback

Endophyte

 \rightarrow plants, seeds, etc.

Toxin producer

→ contamination of plant products

Human pathogen

 \rightarrow nosocomial infection



Examples of species with mostly tropical and subtropical distribution :

- Fusarium lateritium, Fusarium stilboides
- Fusarium guttiforme, Fusarium ananatum GFC
- Fusarium mangiferae, F. sterilihyphosum GFC
- Fusarium decemcellulare Albonectria rigidiuscula

Form-species, a high diversity of typically tropical populations, many phylogenetic or biological species:

- Fusarium oxysporum
- Fusarium solani Haematonectria / Neocosmospora



Species Concepts

- Morphological Species Concept MSC
 Population shares morphological characters
- Biological Species Concept BSC
 Population delimited by reproductive barrier mating population
- Phylogenetic Species Concept PSC
 Population delimited by concordance in groupings generated based on DNA sequences clades



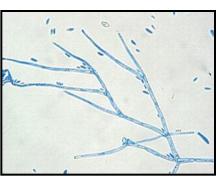
Morphological Species

Fusarium solani

Morphological markers:
macro- and microconidia frequent
chlamydospores frequent
monophialides in the aerial mycelium are long

Many host plants some *formae speciales* Also clinical

→ Species Complex







Biological Species

Biological Species Concept - BSC

Population of individuals that breed and form fertil descendents

Population is delimited by a reproductive barrier mating population or breeding population

Ex. Gibberella fujikuroi Complex



Phylogenetic Species

Phylogenetic Species Concept - PSC

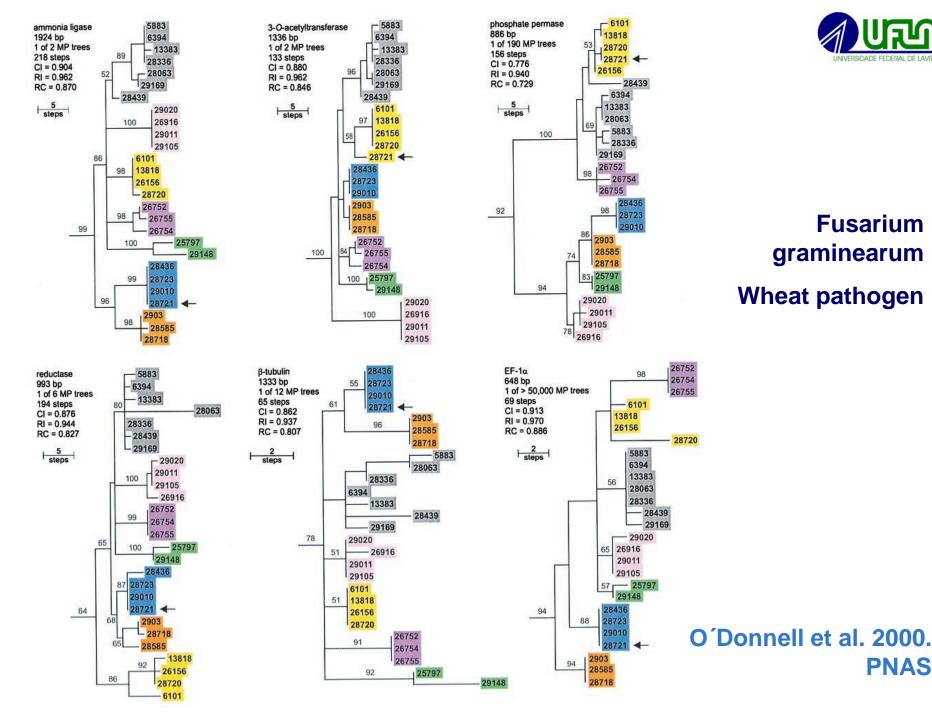
Population delimited by concordance in groupings generated based on DNA sequences - clades

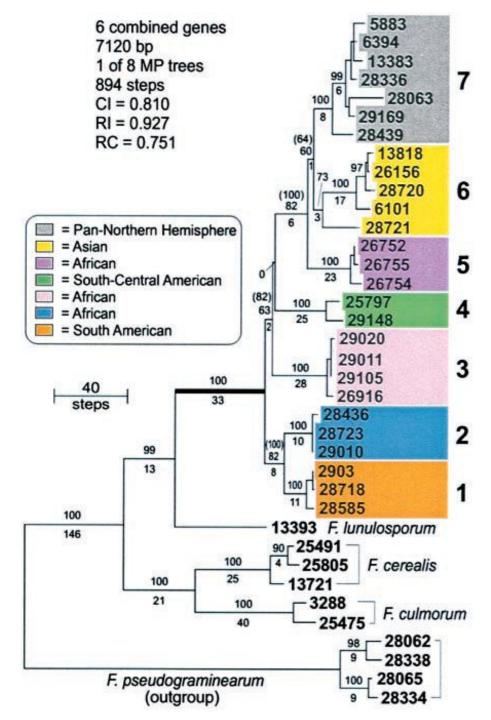
Population of individuals that share apomorphic characters

p.ex. substitution of nucleotides etc.

Geneological Concordance Phylogenetic Species Recognition

Taylor et al. 2000. Fungal Genetics Biology







O'Donnell et al. 2000. PNAS

ICCC-12 Session 7B Species and Populations in Fusarium





Available online at www.sciencedirect.com



Fungal Genetics and Biology 41 (2004) 600-623



www.elsevier.com/locate/yfgbi

Genealogical concordance between the mating type locus and seven other nuclear genes supports formal recognition of nine phylogenetically distinct species within the *Fusarium* graminearum clade

Kerry O'Donnell,^{a,*} Todd J. Ward,^a David M. Geiser,^b H. Corby Kistler,^c and Takayuki Aoki^d

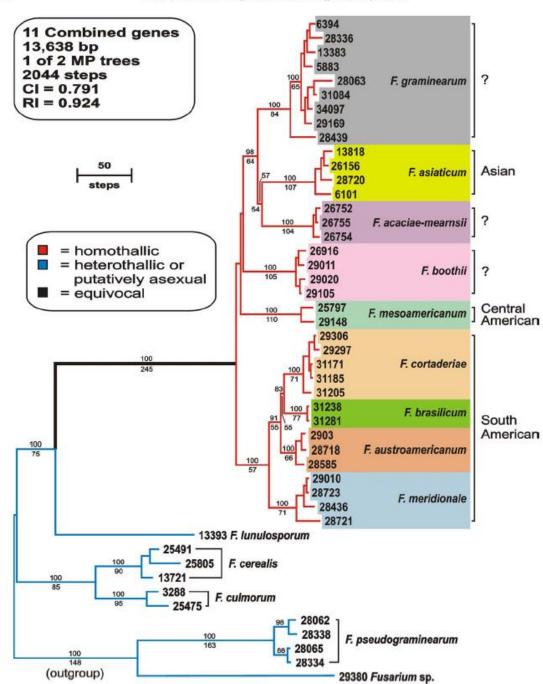
^a Microbial Genomics and Bioprocessing Research Unit, National Center for Agricultural Utilization Research, US Department of Agriculture, Agricultural Research Service, Peoria, IL 61604, USA

^b Department of Plant Pathology, The Pennsylvania State University, University Park, PA 16802, USA

^c Cereal Disease Laboratory, US Department of Agriculture, Agricultural Research Service, 1551 Lindig Street, St. Paul, MN 55108, USA

^d Genetic Diversity Department, National Institute of Agrobiological Sciences (NIAS), 2-1-2 Kannondai, Tsukuba, Ibaraki 305-8602, Japan





O'Donnell et al. 2004. FGB



Examples of tropical crops which develop diseases, caused by *Fusarium* species :

- i. Fruit crops:Mango, pineapple, banana, passionfruit
- ii. Palm trees: oil palm, date palm, canary palm
- iii. Coffee and Cocoa
- iv. Spices, others:

 Black pepper, tobacco, vanilla, cotton
- v. Monocots: Sugarcane, rice, sorghum, maize



Examples of tropical crops which develop diseases, caused by *Fusarium* species :

- i. Fruit crops:Mango, pineapple, banana, passionfruit
- ii. Palm trees: oil palm, date palm, canary palm
- iii. Coffee and Cocoa
- iv. Spices, others:

 Black pepper, tobacco, vanilla, cotton
- v. Monocots: Sugarcane, rice, sorghum, maize



Etiology of Mango Malformation in Brazil

Mangifera indica L. Healthy tree



Symptoms of malformation







Vegetative - shoot



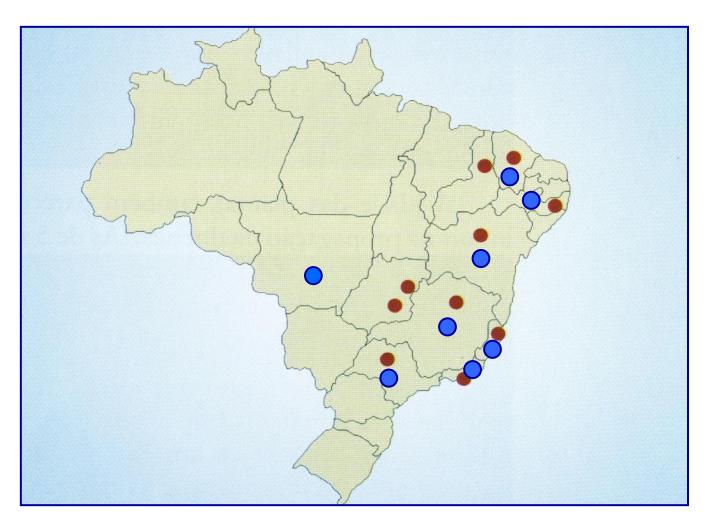
Objectives

- □ To identify the causal agents of mango malformation disease in Brazil
 - Methods AFLP analysis, Gene Sequencing, Morphological Characterization, Pathogenicity Test
- □ To induce the sexual stage for the causal agents Methods Identification of mating type (PCR) and crossing
- □ To develop a PCR-based method for the detection and identification of the causal agents
 - Methods primer design, tests for specificity and sensitivity



Disease Distribution in Brazil

- Lima 2006, 59 isolates, 8 states
- Cunha et al 2000





Pathogen x Endophyte's isolation

Malformed tissue Fusarium only

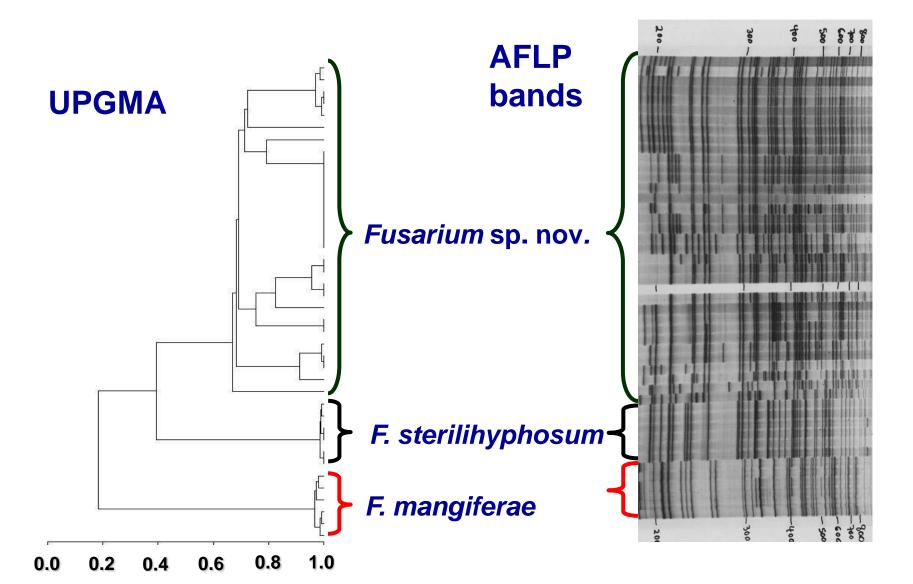


Asymptomatic tissue about 15 species





AFLP analysis of *Fusarium* from mango





Sequence analysis of genes encoding β -tubulin and translation elongation factor 1- α

Primers tub2 – T1 and T2 - O'Donnell et al., 1998, Mycologia

Primers tef1 – EF1 and EF2 - O'Donnell et al., 1998, PNAS

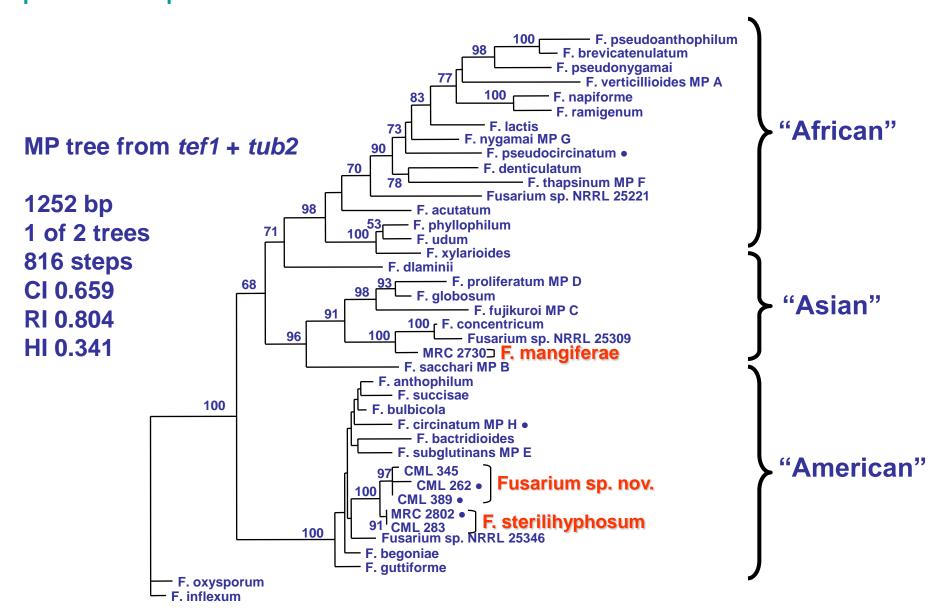
Sequencing – MegaBACE 500, both directions

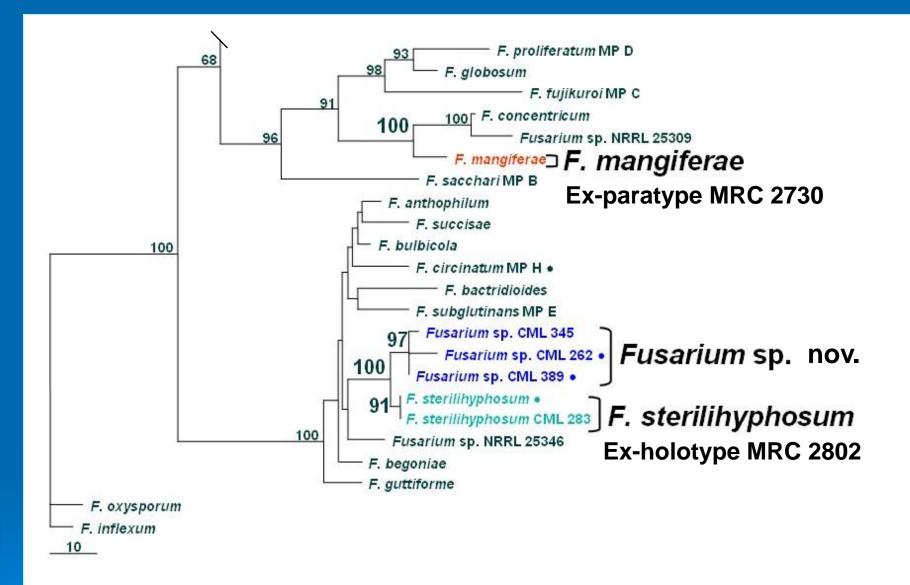
Analysis – Maximum Parsimony

Program – PAUP 4 version beta 10

ICCC-12 Session 7B Species and Populations in Fusarium







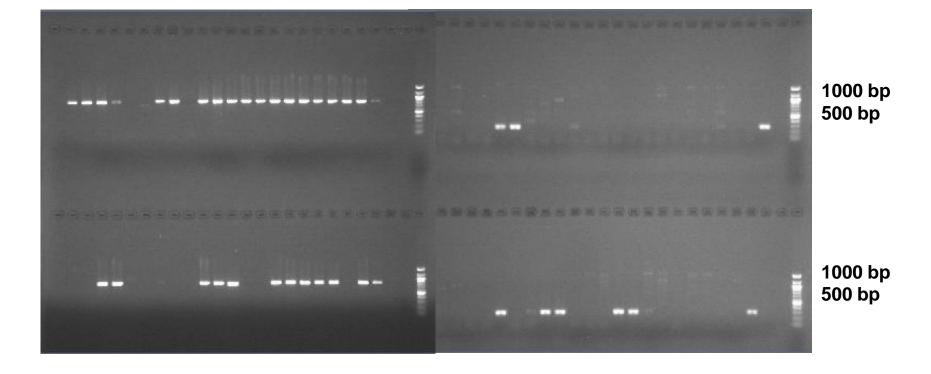
Maximum Parsimony Tree of Combined tub2 and tef1



Crossing

Identification of mating type primers according to Steenkamp et al. (2000)

MAT-1





Fertile crosses only to Fusarium sp. nov.





Evidence from morphology, AFLP fingerprint, phylogeny and crossing

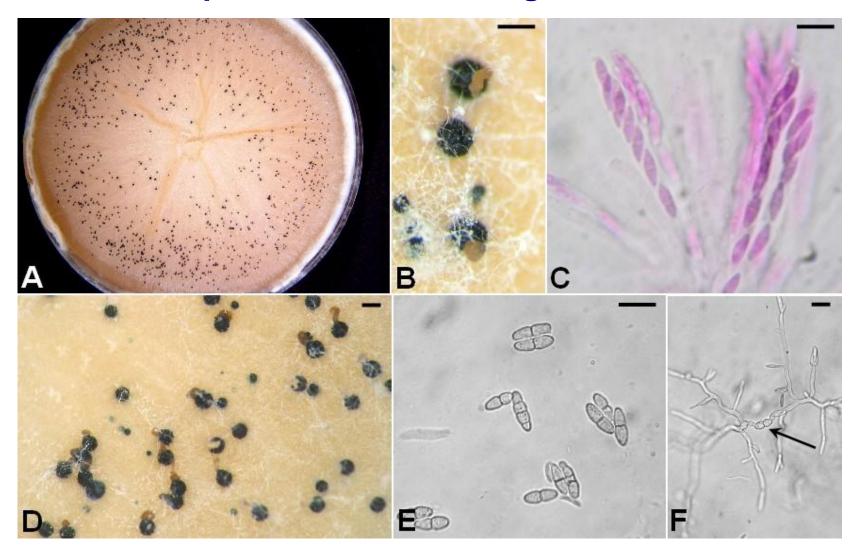
F. mangiferae - cosmopolitan, but not found in Brazil

F. sterilihyphosum - South Africa and Brazil

Gibberella sp. nov. - Brazil

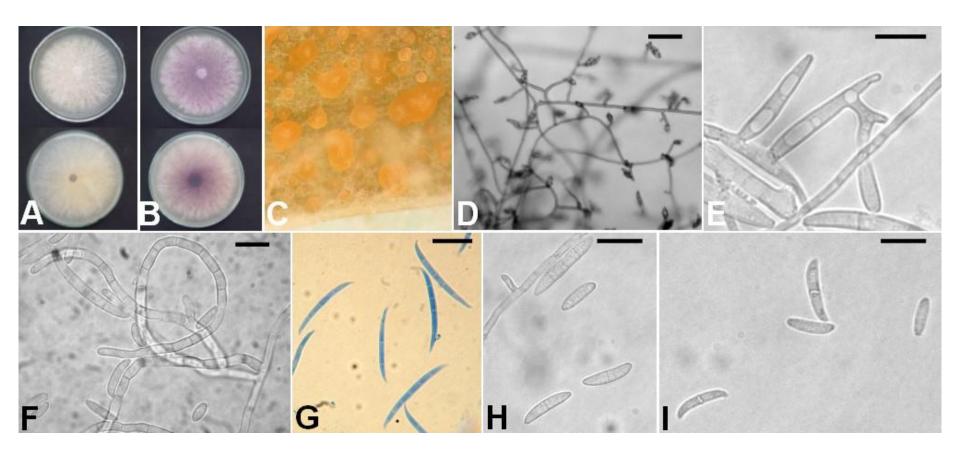


Gibberella sp. nov. Lima, Pfenning and Leslie





Gibberella anam. nov. Lima, Pfenning and Leslie





Pathogenicity



Control Fusarium sp. nov.







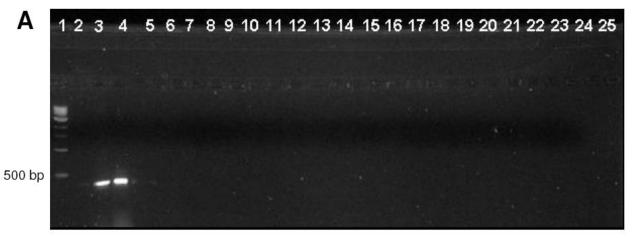
F. sterilihyphosum



PCR detection of the causal agents of mango malformation

Ц	Primer Sets designed from the terr sequences
	Fbra – Fusarium sp. nov. and F. sterilihyphosum (~380 bp)
	Fman – F. mangiferae (~217 bp)

- □ Specificity pure DNA from mango pathogens and endophytes
- □ Sensitivity DNA dilutions from 20 ng to 1 fg
- Pathogens' detection in shoot, leaf, bud, flower, fruit peel mango tissue
- □ Inoculation according to Freeman et al. (1999)





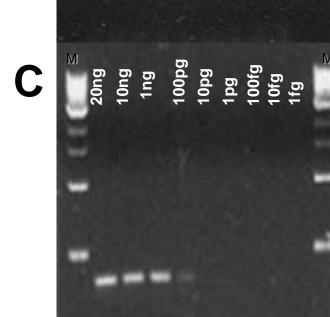
Specificity

- A Fbra primer set (~380 bp)
- B Fman primer set (~217 bp)

- 1. Marker
- 2. Control water
- 3. Fusarium sp. nov. (DNA)
- 4. F. sterilihyphosum (DNA)
- 5. F. mangiferae (DNA)
- 6. F. proliferatum
- 7. F. sacchari
- 8. F. subglutinans
- 9. F. pseudocircinatum
- 10. F. oxysporum
- 11. F. semitectum
- 12. F. decemcellulare
- 13. Neocosmospora sp.
- 14. Phomopsis mangifera
- 15. Cladosp. cladosporioides
- 16. Epiccocum purpurascens
- 17. Fusiccocum mangiferum
- 18. Alternaria alternata
- 19. Coll. gloeosporioides
- 20. Aspergillus niger
- 21. Penicillium sp.
- 22. Pestalotiopsis sp.
- 23. Chalara fimbriata
- 24. Lasiodiplodia sp.
- 25. Phoma sp.



B



Sensitivity

A. Fbra - Fusarium sp. nov.

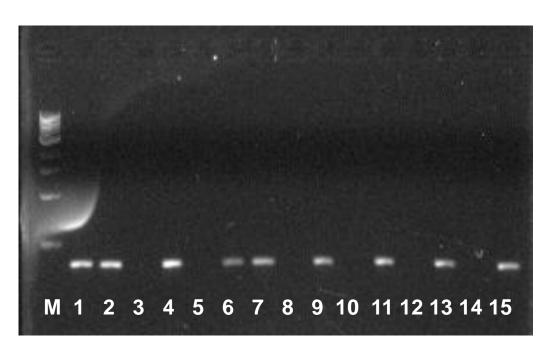
B. Fbra - F. sterilihyphosum

C. Fman - F. mangiferae



Detection of Fusarium sp. nov. and F. sterilihyphosum

Fbra primer set

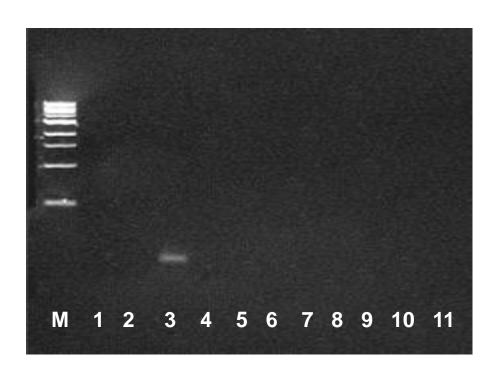


- 1. Fusarium sp. nov. (DNA)
- 2. F. sterilihyphosum (DNA)
- 3. F. mangiferae (DNA)
- 4. Naturally infected panicle
- 5. Asymptomatic inflorescence
- 6. Malformed plant inoculated with Fusarium sp. nov.
- 7. Malformed plant inoculated with F. sterilihyphosum.
- 8 and 9. Asymptomatic plant
- 10 and 11. Fruit peel
- 12 and 13. Fruit peduncle
- 14 and 15. Mango leaf



Detection of F. mangiferae

Fman primer set



- 1. Fusarium sp. nov. DNA
- 2. F. sterilihyphosum DNA
- 3. F. mangiferae DNA
- 4. Naturally infected panicle
- 5. Asymptomatic inflorescence
- 6. Malformed plant inoculated with Fusarium sp. nov.
- 7. Malformed plant inoculated with F. sterilihyphosum
- 8. Asymptomatic plant
- 9. Fruit peel
- 10. Fruit peduncle
- 11. Mango leaf



Conclusions

- □ The main causal agent of mango malformation disease in Brazil corresponds to a new Fusarium species within the Gibberella fujikuroi species complex
- ☐ The teleomorphic stage of Fusarium sp. nov. can be induced in the laboratory
- □ Fusarium sp. nov. and F. sterilihyphosum are the only known causal agents of the disease in Brazil and can be detected through PCR



Most important disease of this crop, restricted to South America?

Fusarium subglutinans f.sp. ananas

Fusarium guttiforme

Morphology, Specificity, tub and tef genes Limited number of isolates

Nirenberg & O'Donnell 1998



Gibberella fujikuroi complex - GFC, but no teleomorph known so far



Fusarium associated with pineapple

Symptomatology





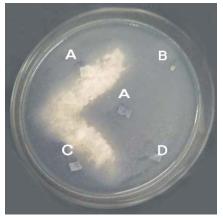






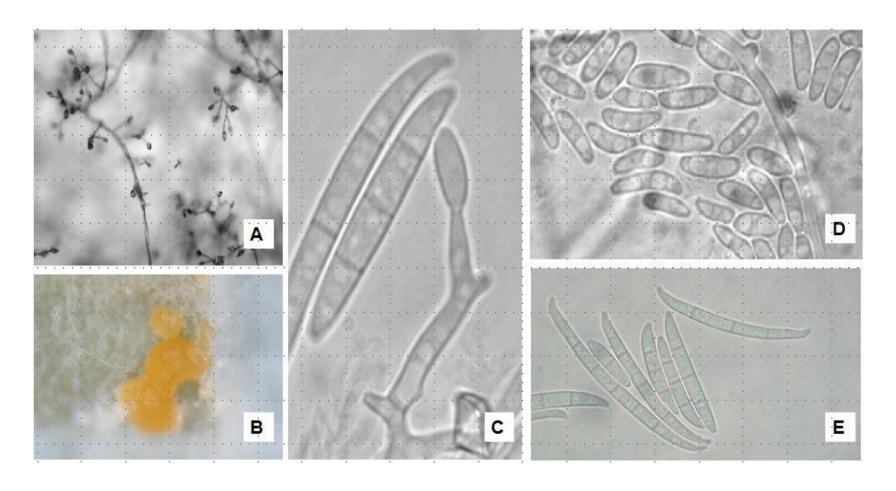
- ☐ Collection of isolates from all over Brazil
- Morphological markers
- □ Pathogenicity tests
- □ VCG analysis
- □ AFLP fingerprint
- □ Phylogenetic analysis of *tub* and *tef* genes





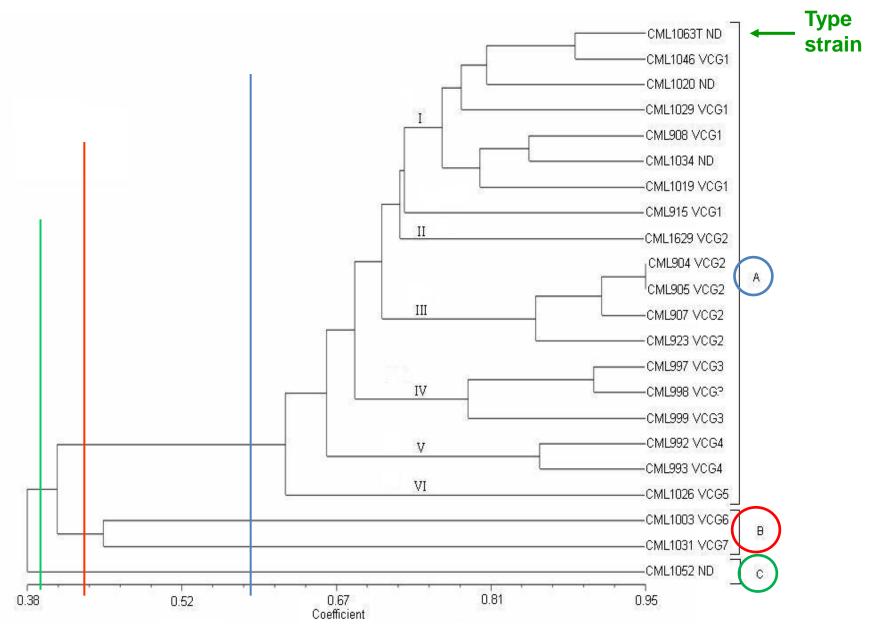


Morphology of *Fusarium guttiforme*



Groups evidenced by VCG and AFLP fingerprint analysis





ICCC-12 Session 7B Species and Populations in Fusarium

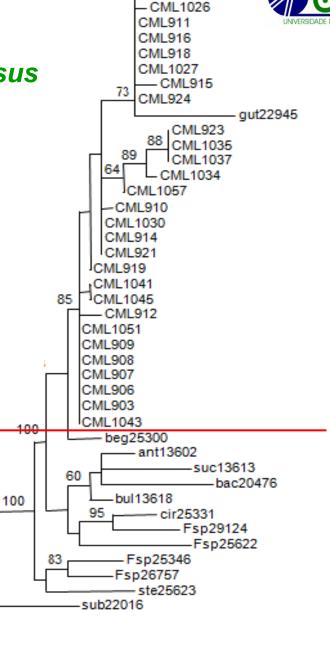
Fusariosis of Pineapple - *Ananas comosus*

tub2 and tef1

F. guttiforme and species in the American clade of GFC

CML - Coleção Micológica de Lavras

NRRL - National Center for Agricultural Utilization Research



CML905

inf20433 oxy22902



Conclusions

	F. guttiforme confirmed as the main pathogen of Fusarium wilt and resinosis of pineapple in Brazil.
	High genetic diversity observed in the population is consistent with the hypothesis that it is near the center of origin, and the possibility of the occurrence of sexual reproduction in species cannot be excluded.
	The observation of two divergent AFLP groups with similarity below 40% in comparison with the main group suggests that there may be distinct populations causing wilt in pineapple in Brazil.
Pe	erspectives and future work
	Further collection, including native species of <i>Ananas</i>

Metabolite profiling

☐ Crossing experiments

□ And



FUNGAL BIOLOGY 114 (2010) 515-527







Fusarium ananatum sp. nov. in the Gibberella fujikuroi species complex from pineapples with fruit rot in South Africa

Adriaana JACOBS^{a,*}, Pieter Schalk VAN WYK^b, Walter F. O. MARASAS^c, Brenda D. WINGFIELD^d, Michael J. WINGFIELD^a, Teresa A. COUTINHO^a

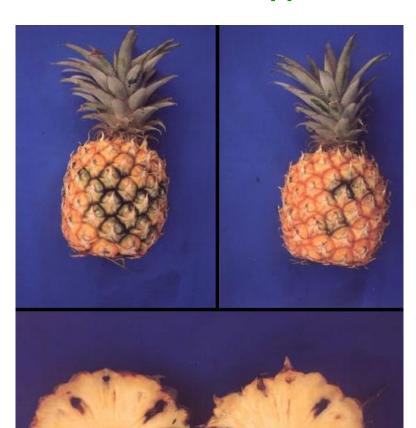
^aDepartment of Microbiology and Plant Pathology, Forestry and Agricultural Biotechnology Institute (FABI), University of Pretoria, Pretoria, South Africa

^bSoygro, PO Box 457, Hartswater, South Africa

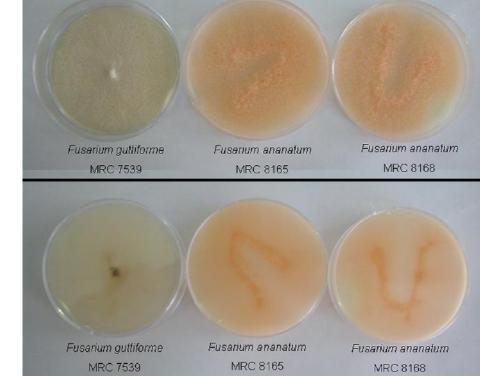
^cForestry and Agricultural Biotechnology Institute (FABI), University of Pretoria, Pretoria, South Africa

^dDepartment of Genetics, Forestry and Agricultural Biotechnology Institute (FABI), University of Pretoria, Pretoria, South Africa





Symptoms
Fusarium ananatum
Black spot



Culture morphology



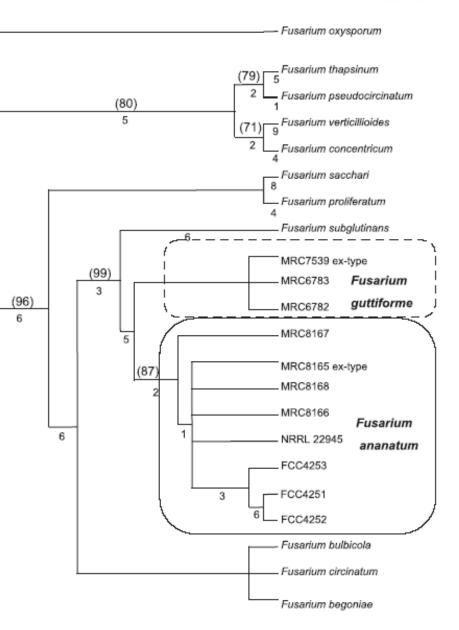
Phylogenetic tree of F. ananatum and related species produced using parsimony of tef-1a

Informative characters 53

CI = 0.6737

RI = 0.8510

g1 = -0.476216





Panama Disease, Fusariosis, wilt disease Banana - *Musa* spp.

Fusarium oxysporum f. sp. cubense

Teleomorph unknown

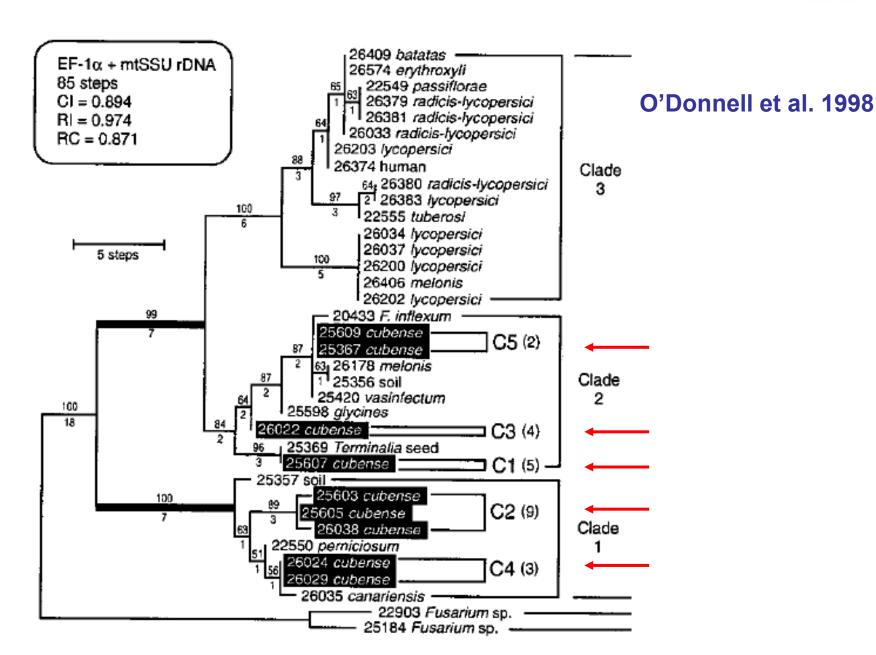
Four races ??

- Polyphyletic
- Distinct evolutionary origin
- ... Five species (?), maybe more



Panama disease of banana





ICCC-12 Session 7B Species and Populations in Fusarium



Panama disease of banana

APPLIED AND ENVIRONMENTAL MICROBIOLOGY, July 2009, p. 4770–4781 0099-2240/09/\$08.00+0 doi:10.1128/AEM.00370-09 Copyright © 2009, American Society for Microbiology. All Rights Reserved.

Vol. 75, No. 14

Evolutionary Relationships among the *Fusarium oxysporum* f. sp. *cubense* Vegetative Compatibility Groups[∇]

Gerda Fourie,1* E. T. Steenkamp,1 T. R. Gordon,2 and A. Viljoen1,3

Department of Microbiology and Plant Pathology, Forestry and Agricultural Biotechnology Institute (FABI), University of Pretoria, Pretoria 0002, South Africa¹; Department of Plant Pathology, University of California, Davis, California 95616²; and Department of Plant Pathology, University of Stellenbosch, Private Bag X1, Matieland 7602, South Africa³

Received 16 February 2009/Accepted 21 May 2009

.... clearly showed that ability of F. oxy. cub

1 to cause disease on banana has emerged multiple times, independently

2 to cause disease to a specific banana cultivar is also a polyphyletic trait



Panama disease of banana

Disease caused by several pathogens referred to as *Fusarium oxysporum* f. sp. *cubense*

Ploetz 2006

- Fusarium oxysporum f. sp. cubense represents at least five distinct clonal lineages
- Races of F. oxysporum f. sp. cubense may belong to different species
- Concept of races is not consistent
- The formae speciales concept is not phylogenetically informative



Fusariosis of Passion fruit

Passiflora spp. - Disease complex

Causal agents:

- 1 Fusarium oxysporum f. sp. passiflorae Wilt
- 2 Fusarium solani
 root rot, stem rot and canker
 develop to wilting and dieback
 present also as an endophyte

Ploetz 2003

Nunes & Albuquerque 1995, Pará, Brazil Fusarium solani - Nectria haematococca

Nirenberg & Brielmeyer-Liebetanz 1996, Germany Fusarium striatum – Haematonectria ipomoeae



Fusariosis of Passion fruit

Wilt syndrom in the field





Photos: Jeferson M. Dariva



Fusariosis of Passion fruit

F. oxysporum f.sp. passiflorae



Fusarium solani





Photos: Jeferson M. Dariva

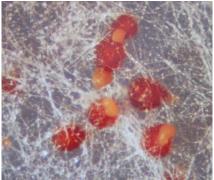


Morphological markers within the form species Fusarium solani

- Growth rate of colony, pigmentation
- Ramification of the conidiophore
- Format, septation and size of microand macroconidia
- Specificity, Others?

Telomorph – *Haematonectria*

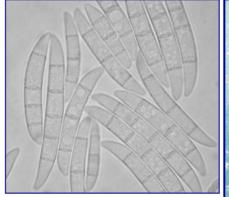
- Homothallic + heterothallic spp.
- Ascospores

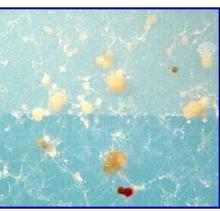














Form species Fusarium solani - Section Martiella

Named species, populations, strains etc.:

Fusarium ambrosium India

Fusarium illudens Nova Zelandia – tel Nectria illudens

Fusarium martii-phaseoli USA, Phaseolus vulgaris

Fusarium striatum (homothallic) Panama – tel Haematonectria ipomoeae

Fusarium sp. Guiania

Fusarium sp. Venezuela

Nectria borneensis Indonesia

Nectria plagianthi Nova Zelandia

Neocosmospora africana

N. vasinfecta

N. ornamentata (homothallic)

(O'Donnell 2000)

ICCC-12 Session 7B Species and Populations in Fusarium



Named formae speciales and mating populations in the Fusarium solani species complex - FSSC

1	F solf. sp.	batatas	MP-II	Ipomoea batatas

2 F sol f. sp. cucurbitae race 1 MP-I Cucurbita spp.

3 F sol f. sp. cucurbitae race 2 MP-V Cucurbita spp.

4 F sol f. sp. glycines Glycine max

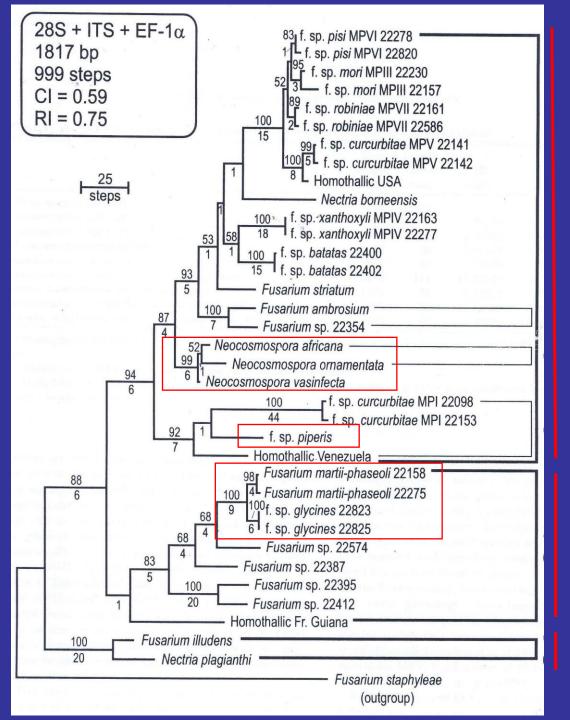
5 F sol f. sp. mori MP-III Morus alba

6 F sol f. sp. piperis Piper nigrum

7 F sol f. sp. pisi MP-VI Pisum sativum

8 F sol f. sp. robiniae MP-VII Robinia spp.

9 F sol f. sp. xanthoxyli MP-IV Xanthoxylum sp.





Strict consensus cladogram

Clade 3

Asian

Africa

S-American

Clade 2 - SDS pathogens soy beans

S-American

O'Donnell 2000

Clade 1
New Zealand





SDS - Sudden death syndrome of soybean

Causal agent: Fusarium solani f.sp. glycinis

Mycoscience (2005) 46:162–183 DOI 10.1007/s10267-005-0235-y © The Mycological Society of Japan and Springer-Verlag Tokyo 2005

FULL PAPER

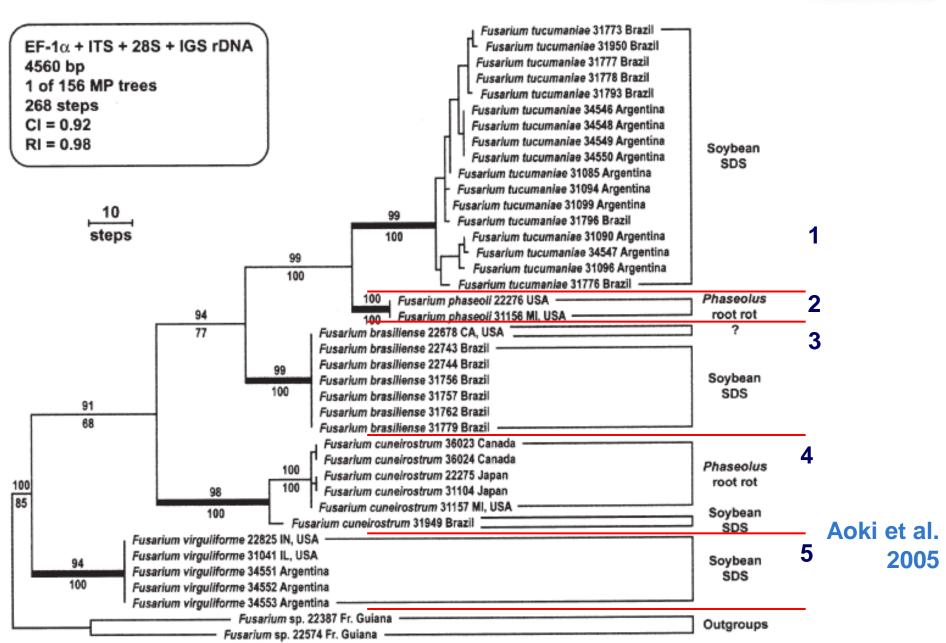
Takayuki Aoki · Kerry O'Donnell María Mercedes Scandiani

Sudden death syndrome of soybean in South America is caused by four species of *Fusarium*: *Fusarium brasiliense* sp. nov., *F. cuneirostrum* sp. nov., *F. tucumaniae*, and *F. virguliforme*

Fusarium solani on soybean and common bean Morphology; pathogenicity; ITS, 28S, IGS, EF-1 α 5 distinct species, only a few isolates from Brazil were included

SDS - Sudden death syndrome of soybean





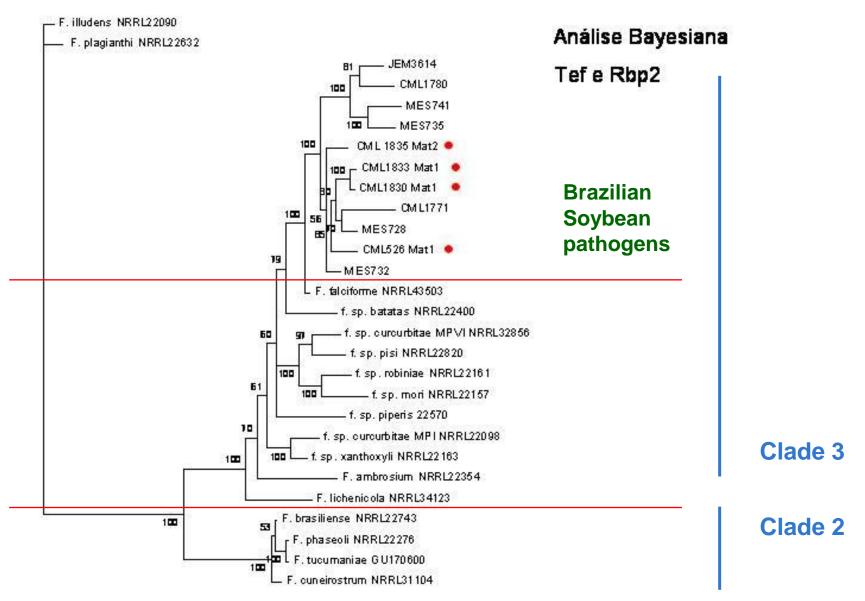


Etiology of Sudden death syndrome of soybean in Brazil

Objectives

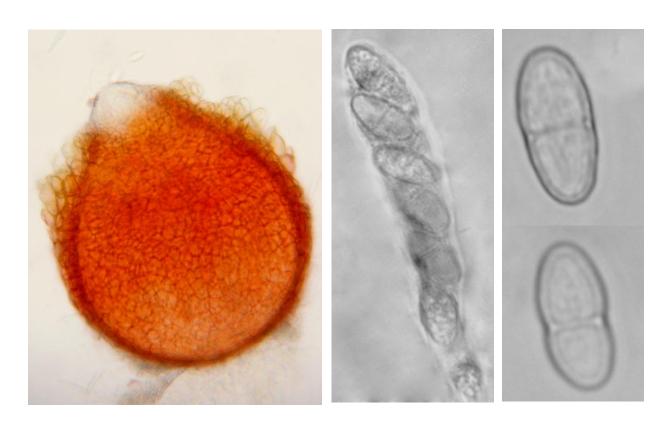
- □ To identify the causal agents of Sudden death syndrome of soybean in Brazil
 - Methods Gene Sequencing, Morphological Characterization, Pathogenicity Test
- □ To identify biological species mating populations Methods Identification of mating type (PCR) and crossing





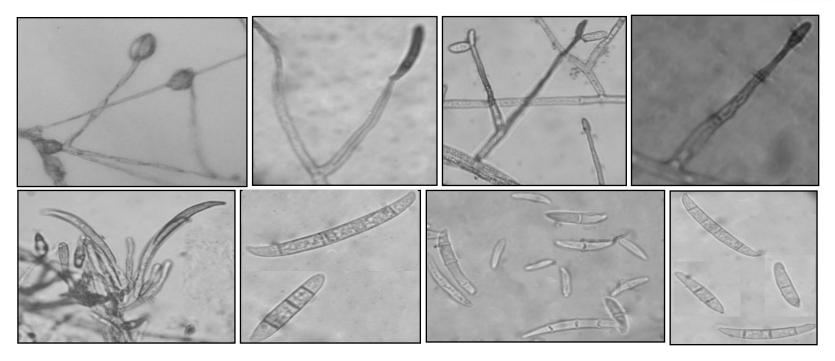


Sexual reproduction in the soybean sudden death syndrome pathogen

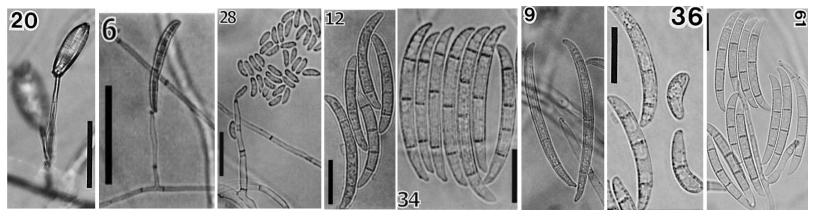


Morphological characters





Brazilian isolates



Aoki et al; 2003, 2005



Piper nigrum - Black Pepper Fusarium solani f. sp. piperis

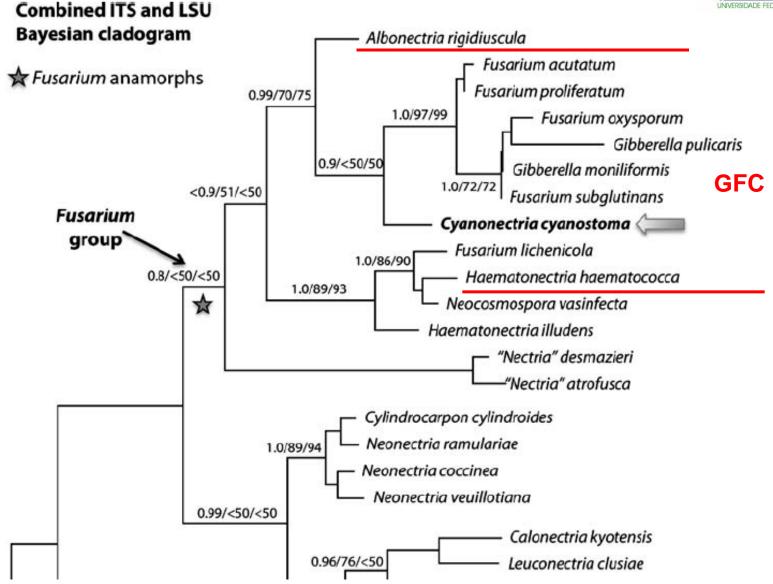
Most important disease of black pepper in Brazil

Is supposed to be a distinct species within FSSC



Photos: Lahyre I. Gomes





ICCC-12 Session 7B Species and Populations in Fusarium



Cocoa - Theobroma cacau

Fusarium decemcellulare Albonectria rigidiuscula

Disease: cushion gall



ICCC-12 Session 7B Species and Populations in Fusarium



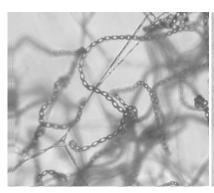


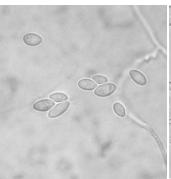


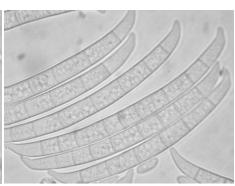




Fusarium decemcellulare Teleomorph Albonectria rigidiuscula

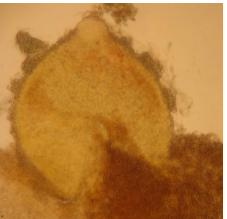


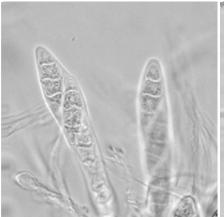


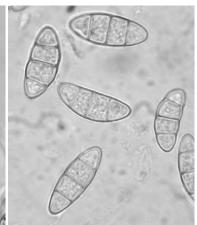














Characterization of populations - approaches

- Morphology Msc
- AFLP fingerprint
- Sequencing and Phylogenetic analysis PSC
- ☐ Sexual Compatibility, mating populations BSC
- Vegetative Compatibility, vegetative compatibility groups

Phylogenetic Classification System only monophyletic taxa are accepted



Characterization of populations and species

The Ideal World – El Mundo Ideal – O Mundo Ideal

MSC = BSC = PSC



Biological Resource Centers:

Gateway to biodiversity and services to innovation in biotechnology

Our research activities:

- Impact of agriculture on soil fungi diversity
- Diversity and specificity of fungal endophytes in crops and plants of natural vegetation
- Metabolite profiling for chemotaxonomy and bioprospection
- Species concepts and characterization of populations of plant pathogens, for the development of PCR-based protocols for diagnostics and quarantine porposes



Biological Resource Centers:

Gateway to biodiversity and services to innovation in biotechnology





01/12/1998

CCT 4776

nd

fruto

campo cultivado

ORIGEM

maracujá (Passiflora edulis)

Data de acesso

Substrato / hospedeiro

Histórico

Cultivar

Habitat

Especificação





Acknowledgements

Sarah S. Costa and Virginia G. Elizei, PhD students

Kedma S. Matos and Erica Sfalsin, Master students

Edson L. Rezende, Ana Karla F. Machado, Culture Collection CML

Lucas M. Abreu, Pos-Doc

Cristiano S. Lima - UFRPE, Recife PE, Brazil

José Aires Ventura - INCAPER, Vitória ES, Brazil

Jeferson M. Dariva - Unimontes, Janaúba MG, Brazil

John F. Leslie - KSU, Manhattan, Kansas, USA







