



The *Gnidia* genus: A review.

Parixit Bhandurge^a, Rajarajeshwari N^b, S Ganapaty^a, Santosh Pattanshetti^b

^a University College of Pharmaceutical Sciences, Andhra University, Visakhapatnam-530003.

^b Visveswarapura Institute of Pharmaceutical Sciences, BSK-II Stage, Bangalore-70

Received:
29th April 2013
Received in revised form:
3rd May 2013
Accepted:
6th May 2013
Available online:
10th May 2013



Online ISSN 2249-622X
<http://www.jbiopharm.com>

ABSTRACT

Gnidia is a member genus of the Thymelaeaceae comprising of about 152 species; several members of which are frequently utilized for a wide variety of ailments like asthma, backache, nightmares, dropsy, boils, sores, induce blistering, treat bruises and burns, constipation, coughs, earache, epilepsy, headache, influenza and fevers, insanity, malaria, measles, pulmonary tuberculosis, poor appetite, smallpox, snake bites, sprains and fractures, tonsillitis, to stabilize heart conditions, stomach and chest complaints, toothache, ulcers and yellow fever and as broad-spectrum purgatives. Several *Gnidia* extracts have also shown antileukemic properties. *Gnidia* elaborates a vast array of biologically active compounds that are chemically diverse and structurally complex. More than 90 compounds have been isolated from different species of *Gnidia*. This review aims at giving an outline of the variety of the secondary metabolites produced by *Gnidia*, as well as their biological activities. The genus *Gnidia* is rich in diterpene esters, coumarins, flavonoids, chromones, lignans, and neolignans. Phytochemical studies on some *Gnidia* species indicated the presence of toxic diterpene esters of daphnane type, which are the main types of plant orthoesters and have remarkable biological activities, such as antineoplastic and cytotoxic. These *gnidia* compounds may be helpful as leads in the synthesis of analogs for treating various ailments.

Keywords: *Gnidia*, Thymelaeaceae, Pharmacological profile, Phytochemistry.

1. INTRODUCTION

Thymelaeaceae family was first established in 1789 by De Jussieu, and currently 45 genera and 800 species are recognized within it. It comprises of many genera with species known to possess toxic, irritant or cocarcinogenic principles which affect animals and humans. It is regarded as a principal source of coumarins and their dimers [1, 2]. The toxic effects of members of this family is due to polyfunctional diterpenoid esters of the daphnane, tigliane and 1-alkyldaphnane type [3]. He *et al.* (2002) reviewed the diversity and biological activities of the structurally unique daphnane-type diterpene derivatives common to the Thymelaeaceae and Euphorbiaceae. Daphnane derivatives are mostly ring C-orthoesters and are more common among Thymelaeaceae than Euphorbiaceae. Daphnetoxin (1) (from several *Daphne* L. species) was the first daphnane-type derivative to be recognized. Later a derivative of daphnetoxin (1), Mezerein (2) was first isolated from *Daphne mezereum* L. It was identified as the poisonous principle responsible for

livestock poisoning by the South African species *Gnidia burchellii* (Meisn.) Gilg (= *Lasiosiphon burchellii* Meisn.) [4]. Toxicity of *Gnidia* species to livestock varies with season and locality. For example, flowering plants of *Gnidia polycephala* (C. A. Mey.) Gilg are apparently more toxic than non-flowering plants [5]. *Gnidia* and other Thymelaeaceae genera have tremendous pharmacological potential, but the useful daphnane orthoesters occur in trace amounts and there still is no large-scale cultivation sufficient for commercial purposes [4].

Linnaeus described *Gnidia*, in the first publication of *Species Plantarum* in 1753. The genus was possibly named after the classical Greek port city 'Cnidus', today known as Cumali. Common names of *Gnidia* are night- and evening-scented bush and 'young-lady-gad-about-at-night', due to the flowers that are fragrant at night [6]. *Gnidia* L., when treated in the broad sense (i.e. including *Lasiosiphon* Fresen is the largest genus in the sub-cosmopolitan family,

*Corresponding author: Rajarajeshwari N| Visveswarapura Institute of Pharmaceutical Sciences, BSK-II Stage, Bangalore-70|

Thymelaeaceae, with 140-160 species occurring in Africa, Arabia, India, Madagascar, and Sri Lanka [3,7,8].

Gnidia can be found in growth forms differing from perennial herbs to shrublets, under-shrubs, shrubs or trees, arising from a woody base or rhizome, and are often ericoid. The bark can vary from a smooth to rough texture with or without lenticels. The branches are slender with alternately positioned, rarely opposite sessile leaves. Several *Gnidia* species are very showy due to their conspicuous flowers. [9-16].

Several species of *Gnidia* are both of medicinal as well as economic importance. Due to the characteristic fibrous bark of Thymelaeaceae, *Gnidia* species are used to tie bundles of wood, thatch and clothing [16]. The flowers of several species of *Gnidia* are employed for dyeing leather [17].

Species of *Gnidia* have been used in the traditional treatments of a variety of medicinal complaints in humans and animals. They have been used to treat a range of conditions in humans including conception and childbirth, asthma, backache, nightmares, dropsy, boils, sores, induce blistering, treat bruises and burns, constipation, coughs, earache, epilepsy, headache, influenza and fevers, insanity, malaria, measles, pulmonary tuberculosis, poor appetite, smallpox, snake bites, sprains and fractures, tonsillitis, to stabilize heart conditions, stomach and chest complaints, toothache, ulcers and yellow fever and as broad-spectrum purgatives [18-25]. Several *Gnidia* extracts have also shown antileukemic properties [4,18,19,26,27]. The Southern Sotho people believe the smoke from burning *Gnidia anthylloides* bewitches people and makes them quarrelsome. Nevertheless, they will use smoke from this plant to treat fevers and bad dreams. In livestock, *Gnidia* species have been used in the treatment of anthrax and botulism. In Madagascar, leaves of *Gnidia gilbertae* Drake are used as a purge to induce vomiting [28]. Crushed roots of *G. kraussiana* are used to make fish poison, and in Madagascar the fibrous barks of *Gnidia* species are used to make rope and twine, paper and ceremonial clothing [28, 29].

Although many *Gnidia* species are used in traditional medicine, severe irritant effects as well as death in humans and animals have been reported, due to the presence of toxic coumarins and diterpene esters [16, 30, 31].

Conflict of Interest: None Declared

1. *Gnidia aberrans* C.H.Wright
2. *Gnidia albosericea* (M.Moss) B.Peterson
3. *Gnidia ambondrombensis* (Boiteau) Z.S. Rogers
4. *Gnidia anomala* Meisn.
5. *Gnidia anthylloides* (L.f.) Gilg
6. *Gnidia apiculata* (Oliv.) Gilg
7. *Gnidia bambutana* Gilg Ledermann ex Engl.
8. *Gnidia baumiana* Gilg
9. *Gnidia baurii* C.H.Wright
10. *Gnidia bojeriana* (Decne. Cambess.) Baill.
11. *Gnidia burchellii* (Meisn.) Gilg
12. *Gnidia burmanii* Eckl. & Zeyh. Meisn.
13. *Gnidia burmanni* Eckl. & Zeyh. Meisn.
14. *Gnidia caffra* (Meisn.) Gilg
15. *Gnidia calocephala* (C.A.Mey.) Gilg
16. *Gnidia caniflora* Meisn.
17. *Gnidia canoargentea* (C.H.Wright) Gilg
18. *Gnidia capitata* L.f.
19. *Gnidia cayleyi* C.H.Wright
20. *Gnidia chapmanii* B.Peterson
21. *Gnidia chrysantha* (Solms) Gilg
22. *Gnidia chrysophylla* Meisn.
23. *Gnidia clavata* Schinz
24. *Gnidia compacta* (C.H.Wright) J.H.Ross
25. *Gnidia conspicua* Meisn.
26. *Gnidia coriacea* Meisn.
27. *Gnidia cuneata* Meisn.
28. *Gnidia danguyana* Leandri
29. *Gnidia decaryana* Leandri
30. *Gnidia decurrens* Meisn.
31. *Gnidia dekindtiana* Gilg
32. *Gnidia denudata* Lindl.
33. *Gnidia deserticola* Gilg
34. *Gnidia dregeana* Meisn.
35. *Gnidia dumicola* S.Moore
36. *Gnidia emini* Engl. & Gilg
37. *Gnidia ericoides* C.H.Wright
38. *Gnidia fastigiata* Rendle
39. *Gnidia flanagani* C.H.Wright
40. *Gnidia foliosa* (H.Pearson) Engl.
41. *Gnidia francisci* Bolus
42. *Gnidia fraterna* (N.E.Br.) E.Phillips
43. *Gnidia fruticulosa* Gilg
44. *Gnidia fulgens* Welw.
45. *Gnidia galpini* C.H.Wright
46. *Gnidia geminiflora* E.Mey. ex Meisn.
47. *Gnidia gilbertae* Drake
48. *Gnidia glauca* (Fresen.) Gilg
49. *Gnidia gnidioides* (Baker) Domke
50. *Gnidia goetzeana* Gilg
51. *Gnidia gossweileri* (S.Moore) B.Peterson
52. *Gnidia gymnostachya* (C.A.Mey.) Gilg
53. *Gnidia harveyana* Meisn.
54. *Gnidia heterophylla* Gilg
55. *Gnidia hibbertioides* (S.Moore) Rogers
56. *Gnidia hirsuta* (L.) Thulin
57. *Gnidia hockii* De Wild.
58. *Gnidia humbertii* (Leandri) Z.S. Rogers
59. *Gnidia humilis* Meisn.
60. *Gnidia imbricata* L.f.

61. <i>Gnidia inconspicua</i> Meisn.	91. <i>Gnidia occidentalis</i> (Leandri) Z.S. Rogers	122. <i>Gnidia scabrida</i> Meisn.	138. <i>Gnidia stenophylloides</i> Gilg
62. <i>Gnidia insignis</i> Compton	92. <i>Gnidia oliveriana</i> Engl. & Gilg	123. <i>Gnidia sericea</i> L.	139. <i>Gnidia strigillosa</i> Meisn.
63. <i>Gnidia involucreta</i> Steud. ex A.Rich.	93. <i>Gnidia oppositifolia</i> L.	124. <i>Gnidia sericocephala</i> (Meisn.) Gilg ex Engl.	140. <i>Gnidia styphelioides</i> Meisn.
64. <i>Gnidia juniperifolia</i> Lam.	94. <i>Gnidia orbiculata</i> C.H.Wright	125. <i>Gnidia setosa</i> Wikstr.	141. <i>Gnidia suavissima</i> Dinter
65. <i>Gnidia kasaiensis</i> S.Moore	95. <i>Gnidia pallida</i> Meisn.	126. <i>Gnidia similis</i> C.H.Wright	142. <i>Gnidia subulata</i> Lam.
66. <i>Gnidia kraussiana</i> Meisn.	96. <i>Gnidia parviflora</i> Meisn.	127. <i>Gnidia singularis</i> Hilliard	143. <i>Gnidia tenella</i> Meisn.
67. <i>Gnidia kundelungensis</i> S.Moore	97. <i>Gnidia parvula</i> Dod	128. <i>Gnidia squarossa</i>	144. <i>Gnidia thesioides</i> Meisn.
68. <i>Gnidia lamprantha</i> Gilg	98. <i>Gnidia pedunculata</i> Beyers	129. <i>Gnidia socotrana</i> (Balf.f.) Gilg	145. <i>Gnidia tomentosa</i> L.
69. <i>Gnidia latifolia</i> (Oliv.) Gilg	99. <i>Gnidia penicillata</i> Licht. ex Meisn.	130. <i>Gnidia somalensis</i> (Franch.) Gilg	146. <i>Gnidia triplinervis</i> Meisn.
70. <i>Gnidia laxa</i> (L.f.) Gilg	100. <i>Gnidia perrieri</i> (Leandri) Z.S. Rogers	131. <i>Gnidia sonderiana</i> Meisn.	147. <i>Gnidia usafuae</i> Gilg
71. <i>Gnidia leipoldtii</i> C.H.Wright	101. <i>Gnidia phaeotricha</i> Gilg	132. <i>Gnidia sparsiflora</i> Bartl. ex Meisn.	148. <i>Gnidia variabilis</i> (C.H.Wright) Engl.
72. <i>Gnidia linearifolia</i> (Wikstr.) B.Peterson	102. <i>Gnidia pinifolia</i> L.	133. <i>Gnidia spicata</i> (L. f.) Gilg	149. <i>Gnidia variegata</i> Gand.
73. <i>Gnidia linearis</i> (Leandri) Z.S. Rogers	103. <i>Gnidia pleurocephala</i> Gilg	134. <i>Gnidia splendens</i> Meisn.	150. <i>Gnidia welwitschii</i> Hiern
74. <i>Gnidia linoidea</i> Wikstr.	104. <i>Gnidia poggei</i> Gilg	135. <i>Gnidia squarrosa</i> (L.) Druce	151. <i>Gnidia wickstroemiana</i> Meisn.
75. <i>Gnidia lucens</i> Lam.	105. <i>Gnidia polyantha</i> Gilg	136. <i>Gnidia stellatifolia</i> Gand.	152. <i>Gnidia woodii</i> C.H.Wright
76. <i>Gnidia macropetala</i> Meisn.	106. <i>Gnidia polycephala</i> Gilg ex Engl.	Table 1: List of Species in <i>Gnidia</i> genus	
77. <i>Gnidia macrorrhiza</i> Gilg	107. <i>Gnidia polystachya</i> P.J.Berghius	2. PHYTOCHEMICAL ASPECTS:	
78. <i>Gnidia madagascariensis</i> (Lam.) Baill.	108. <i>Gnidia propinqua</i> (Hilliard) B.Peterson	This chapter aims at giving an outline of the variety of the secondary metabolites produced by <i>Gnidia</i> , as well as their biological activities. The Thymelaeaceae plants contain coumarins, flavonoids, chromones, lignans, and neolignans [32-34]. Phytochemical studies on the Thymelaeaceae plants due to their widespread uses in medicine have been reported and there are reports on the toxicity of these plants [18]. In the last 70 years, several species of Thymelaeaceae have been the subject of numerous phytochemical studies. Initially, interest may have been due to the marked toxicity of these plants, but the widespread use of some species medicinally has certainly played a part in sustaining this interest [3]. Several genera such as <i>Daphne</i> , <i>Thymelaea</i> , <i>Pimelea</i> , <i>Wikstroemia</i> and <i>Gnidia</i> have been researched upon extensively. The <i>Daphne</i> genus is of prime importance owing to its richness in a variety of different classes of natural products, especially, coumarins [35-37], lignans [35, 38, 39], flavones [38,40,41], daphnane-type diterpene esters [42-45], steroids and guianolides [9]. <i>Gnidia</i> genus has similar chemical constitution as that <i>Daphne</i> . Chemical studies done on some <i>Gnidia</i> species indicated the presence of toxic diterpene esters of daphnane type, which are the main types of plant orthoesters known [46] and have remarkable biological activities, such as	
79. <i>Gnidia meyeri</i> Meisn.	109. <i>Gnidia pulchella</i> Meisn.		
80. <i>Gnidia microcephala</i> Meisn.	110. <i>Gnidia quadrifaria</i> C.H.Wright		
81. <i>Gnidia microphylla</i> Meisn.	111. <i>Gnidia quarrei</i> A.Robyns		
82. <i>Gnidia mollis</i> C.H.Wright	112. <i>Gnidia racemosa</i> Thunb.		
83. <i>Gnidia montana</i> H.Pearson	113. <i>Gnidia razakamalalana</i> Z.S. Rogers		
84. <i>Gnidia multiflora</i> Bartl. ex Meisn.	114. <i>Gnidia rendlei</i> Hiern		
85. <i>Gnidia myrtifolia</i> C.H.Wright	115. <i>Gnidia renniana</i> Hilliard & B.L.Burt		
86. <i>Gnidia nana</i> (L.f.) Wikstr.	116. <i>Gnidia rivae</i> Gilg		
87. <i>Gnidia newtonii</i> Gilg	117. <i>Gnidia robusta</i> B.Peterson		
88. <i>Gnidia nitida</i> Bolus ex C.H.Wright	118. <i>Gnidia robyniana</i> Lisowski		
89. <i>Gnidia nodiflora</i> Meisn.	119. <i>Gnidia rubescens</i> B.Peterson		
90. <i>Gnidia obtusissima</i> Meisn.	120. <i>Gnidia rubrocincta</i> Gilg		
121. <i>Gnidia scabra</i> Thunb.	137. <i>Gnidia stenophylla</i> Gilg		

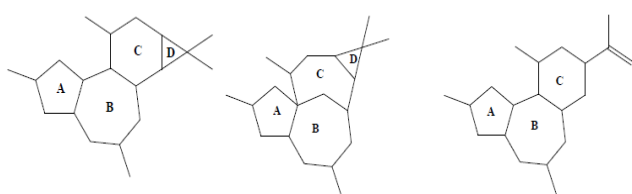
antineoplastic and cytotoxic [23,31,32,47-50]. Furthermore, phytochemical investigations revealed the occurrence of coumarins [48], lignans [51], flavonoids and benzophenone glycosides [25], umbelliferoylflavonoids and spiro-bis- γ -lactone [30].

Diterpenes

Several species of Thymelaeaceae as well as Euphorbiaceae are well known for their interesting physiological and toxic effects. More than 40 years ago, Hecker isolated and finally elucidated the structure of the tumour-promoting phorbol-12,13-diester from the seeds oil of *Croton tiglium* (Euphorbiaceae), which initiated intensive research into the pro-inflammatory and tumour-promoting diterpenes of the two plant families [52]. The chemical nature of toxic diterpenes of Thymelaeaceae has been known only for about thirty years. Although from a large structural variety, they derive all the toxicity from the basic skeletons tigliane, ingenane and daphnane type.

The tigliane, daphnane, and ingenane diterpenes esters are noted for their skin irritant and cocarcinogenic effects [53]. These compounds occur widely in toxic species of the Euphorbiaceae and Thymelaeaceae. However, many of the group, particularly orthoesters of the daphnane series, exhibit potent antileukemic activity. Hence, there has been marked interest in the structural features of this series that are conducive to antitumor activity in contrast to those that are responsible for irritant properties. Representative of the daphnane tumor inhibitors include Gnididin (8) from *Gnidia lamprantha* Gilg., Gnidimacrin (23) from *G. subcordata* (Meissn.) Engl. and Mezerein (2) from *Daphne mezereum* L [48, 50, 54-56].

Explanation on the class of Daphnanes has been emphasized owing to the above stated fact and also because they were found to be much prevalent in species of *Gnidia* and frequently occur as orthoesters. Although there exists large structural variations among these classes, but the toxicity is derived from their basic skeleton [53].



Tiglyane-type

Ingenane-type

Daphnane-type

Tiglanes:

Tiglanes consist of a group of tetracyclic compounds which are present in plants in the form of polyhydroxyl diterpenes. They have a tricyclo[9.3.0.0]tetradecane ring system as tof hatdaphnanes; in addition a gem-dimethyl cyclopropane is fused to the 6-membered ring. They are

found as mixture of many esters from Euphorbiaceae and Thymelaeaceae [53]. The most well-known member is Phorbol (21), occurs naturally in the form of its 12,13-diester and 12,13,20-triester. These trimesters are known as 'cryptic irritants' because they do not exhibit pro-inflammatory activity on mammalian skin unless the C-20 acyl group is removed by hydrolysis; they are potent tumour-promoting agents, inducing susceptibility at levels of carcinogen below the normal threshold [57,58].

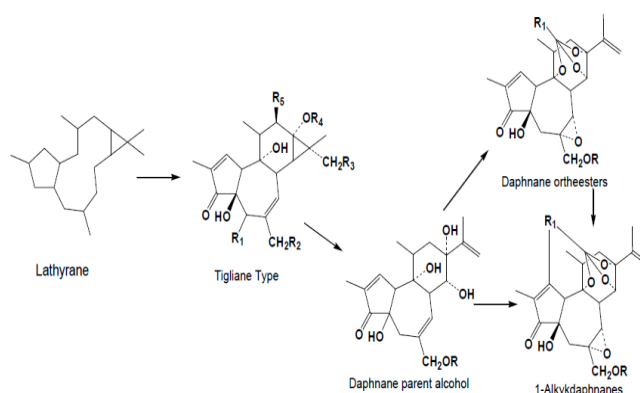
Ingenanes:

Ingenanes represent a structurally unique class of highly oxygenated teracyclic diterpene esters and are made up of Bicyclo[4.4.1]undecane ring system. They possess potent tumour-promoting properties. Ingenol (22) serves as the parent diterpene nucleus from which many of these biologically active esters are derived [59].

Daphnane Diterpenes (Daphnetoxin class diterpenes):

Quite clearly, this class of compounds is characteristic of Thymelaeaceae family. They are found in all the parts of the plant like, root, stem, leaf, fruit, and kernel. Plants from the Thymelaeaceae and the Euphorbiaceae families provide rich sources of daphnane derivatives, which generally occur as ring C-orthoesters. Most of the daphnanes possessing an orthoester function exhibit various physiological activities. They are very irritating to the mucous membranes and are powerful vesicant. This explains the use of bark of some of the species of *Gnidia* as pesticide and fish poison. They have remarkable biological activities, such as cytotoxic [23,32,48,50,60,61], neurotrophic [32, 62], antifertility [63-66], pesticidal [67], antihyperglycemic [68], anti-bladder-hyper-reflexia [69], and anti-HIV activities [70] properties.

Daphnanes are tricyclic diterpenes formed by opening of cyclopropane ring of a 12-deoxyphorbol derivative, followed by reduction of the resulting 13,14-ketol ester to 13 β -isopropenyl moiety. These compounds predominantly occur naturally as orthoesters and are of considerable interest since they possess several biological actions in animals [53, 60].



Based on the presence of oxygen containing functions at the B and C rings, as well as on the substitution pattern of A ring; these daphnane derivatives are classified into four major structural classes: classes 1, 2, 3 and 4 of daphnane derivatives have been outlayed as daphnetoxins, 12-hydroxydaphnetoxins, alkyl daphnanes and resiniferonols, respectively. Classes 1-3 possess a 5 β -OH and 6 α ,7 α -epoxy groups in the seven-membered B ring. Biological activities of structurally similar compounds vary even with small changes such as the length of a carbon chain or the position of an ester group. [4, 71].

▪ **Daphnetoxins**

The compounds of class 1 are based on the absence of C-12 hydroxylation represented by daphnetoxin (1), an orthobenzoate [72]. It is isolated from plants of the Thymelaeaceae, though its analog with an aliphatic orthoester chain, huratoxin (3) [73, 74], which possesses piscicidal activity, was also found in plants of the Euphorbiaceae.

▪ **12 β -hydroxydaphnetoxin**

The daphnane-type orthoesters of class 2 contain a 12 β -OH group, which is either free as in 12 β -hydroxydaphnetoxin (4) [75] or esterified as in 12 β -acetoxyhuratoxin (5) [62, 76], kirkinine (6) [32] and kirkinine D (7) [62].

Mezerein (2) was isolated from *Lasiosiphon burchellii*, which was known to cause poisoning of sheep and other animals in South Africa [75]. It is also known to show potent anti-leukemic property against P-388 and L-1210 leukemia in mice [53]. Structurally similar compound series such as gnididin (8), gniditrin (9), gnidicin (10) has been isolated from *Gnidia lamprantha*, which were found to possess antileukemic and weak carcinogenic effect compared with many phorbol esters [48, 77]. Interestingly, 12-hydroxydaphnetoxin (4), which bears no ester function at C-12 shows neither anti-leukemic nor carcinogenic activity [53, 78]. However its benzoate ester demonstrated activity of the same order of other esters [48]. This affirms the hypothesis that esters at C-12 may act as carrier moiety in processes concerned with cell penetration or selective complex formation [48, 67].

Similar two antileukemic diterpenes having an aliphatic orthoester and a benzoate ester at C-12, gnidilatin (11) and gnidilatidin (12) and their corresponding palmitate analogues were isolated from the same plant. Gnidilatin (11) showed moderate activity while gnidiglaucin (15) and gnidilatidin (12) did not show any activity [23, 48].

▪ **1 α -alkyl daphnanes**

Class 3 is those of the so-called 1 α -alkyl daphnanes which contain an orthoester group incorporating to a macrocyclic bridge from C-1' to C-1 of the daphnane skeleton. Subsequently, C-1 double bond in A ring is

absent and sometimes the keto group is replaced by a benzoate ester at C-3 of ring A [50, 79-84]. These represent a newer group of diterpene esters most of which are isolated from the Thymelaeaceae, and have not yet been found in Euphorbiaceae. Gnidimacrin (23) was the first compound to be encountered in this group and was isolated from *Gnidia subcordata*. Gnidimacrin (23) and its 20-palmitate (24) make part of a subclass possessing 3 β -benzoyloxy group instead of the 3-C carbonyl function. Both compounds were demonstrated to have potent *in vivo* antileukaemic activity. These were the first reported diterpenoids known to contain a novel macrocyclic ring with one terminus at the orthoester carbon [50, 50, 80]. Similar compounds have been isolated from *Pimelea*, *Daphnopsis* and other *Gnidia* species. Daphnopsis factor R₁ (Gnilatmacrin) (25) was isolated from *Daphnopsis racemosa* and *Gnidia kraussiana* [31].

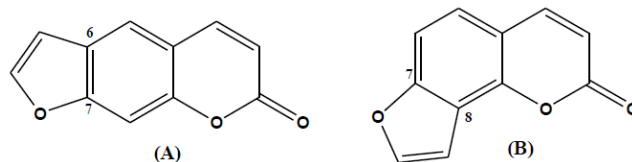
▪ **Resiniferonol:**

Class 4 is that of the resiniferonols, differ from the previous type of orthoester in a way; they possess 6,7-double bonds typical of the phorbol-type esters and the secondary hydroxyl function at C-5 is absent. Resiniferonols have a very narrow distribution in plants, specifically occurring in *Euphorbia* genus [69, 85].

Resiniferatoxin (27) was isolated from *Euphorbia resinifera* and *E. unispina*, based on its extraordinary irritant activity. It is ultrapotent analog of capsaicin, and has attracted special therapeutic interest as an analgesic agent [86].

Coumarins

Generally, coumarins and its derivatives furanocoumarin are found to be present in a plethora of naturally occurring plants. Furanocoumarins are toxic compounds formed by fusion of furan ring to coumarin at C-6 and C-7 position (Linear) (B) for example, psoralen or at C-7 and C-8 position (Angular) (A) for example, angelicin as shown in figure. Even though both linear and angular furanocoumarins are derived from a common 7-hydroxycoumarin precursor (umbelliferone), they differ in their toxicological properties. [86-88].



Coumarins and Furanocoumarins are present either in the free-state or its corresponding glycosides form in nature. These are reported to be present in about 150 different species spreading over to nearly 30 different families, of which a few important ones are *Leguminosae*, *Umbelliferae*, *Solanaceae*, *Oleaceae*, *Rubiaceae*, and *Caprifoliaceae* [87, 86].

Coumarins play a very important role in the chemotaxonomically Thymelaeaceae. More than 40 coumarins with various skeletal patterns have been reported to be isolated from several genera of Thymelaeaceae [18, 89, 90]. They are found in the form of simple coumarins, or as dimers and trimers, or in conjugated form as coumarin glycosides, flavone-coumarin, coumarinolignans etc.

Bicoumarins:

The bicoumarins are a comparatively new class of coumarins and only 29 have been encountered in nature so far. The first of these series of compounds to be isolated was dicoumarol (28) in 1941 [91]. The bicoumarins isolated from plant sources fall into the following categories:

- (i) The two coumarin units are linked at C3-C3' through a methylene group, i.e. dicoumarol (28) [91], gerberinol (29) [92];
- (ii) The two coumarin units are linked by an ether bridge, i.e. daphnoretin (33) [93-112], acetyldaphnoretin (34) [113], edgeworthin (35) [100], candicanin (36) [114], lasiocephalin (37) [115];
- (iii) In this type, two coumarin systems are attached through a carbon-carbon bond, i.e. bicoumol (38) [116], bhubaneswin (39) [117], matsukaze lactone (40) [117, 118], kotanin (41) [119-121], desmethylkotanin (42) [119], orlandin (43) [121], euphorbetin (44) [122-124], isoeuphorbetin (45) [123, 125], ipomopsin (46) [126];
- (iv) Here the two coumarin units are linked through a C-10 alkyl residue (monoterpene unit), i.e. phebalin (48) [127], toddasin/mexolide (49) [128, 129], thamnosin (50) [126, 130-132], and
- (v) where the two coumarin nuclei are linked through 7-methyljuglone unit as in ismailin (51) [133, 134]. In addition, there are two other bicoumarins where the two coumarin units are linked by an ether bridge at C7-C7' as well as a C-C bond at C8-C8' as in gnidicoumarin (52) [47] and in lasioerin (53) [135] the corresponding linkages are through C7-O'' and C6-C6' respectively. In the only naturally occurring bicoumarin - rhamnoside, eriocephaloside (54), the coumarin nuclei are linked through an

ether bridge C4-C7' and the C-C bond is at C3-C8'.

Bhandari *et al* reported isolation of a new coumarin glycoside, Eriocide (55) from *Lasiosiphon eriocephalus*. From the aerial parts of the same taxon Lasiocephalin (37) was isolated, which was the first report of bicoumarin glycoside found in nature. Erioccephaloside (54), isolated from the same taxon, represents a new group of furanobiscoumarins in which the aryl ring of one unit is linked through both oxygen and carbon to the lactone ring of the second unit[1,136]. K. Franke *et al.* (2000), reported a biscoumarin 7,7'-dihydroxy-3,8'-biscoumarin (47), similar to the one with linked between C-3 and C-8' which was previously isolated from *Ipomopsis aggregata* (Polemoniaceae) [30].

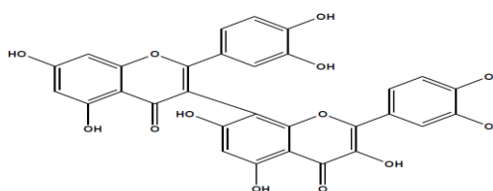
Flavonoids:

Another chemotaxonomically remarkable characteristics of Thymelaeaceae is the presence of diverse flavonoids metabolites such as flavones, flavonols, flavanones, C-glycosylflavones, C-methyl lipophilic flavonoids, less oxygenated flavones, different types of biflavonoids (spiranobiflavonoids, furanobiflavonoids), flavone-coumarin hybrids etc. More than 60 different flavonoids moieties have been reported from this family [18, 89, 90].

Biflavanone:

Biflavonoids are apparently primitive chemical markers in higher plants, since they occur primarily in bryophytes, tracheophytes and gymnosperms. They are, by contrast, far less common in dicotyledonous angiosperms [137]. As compared to the monomeric flavonoids, biflavonoids are limited in structural variation. They are confined largely to oxidative coupling products of chalconaringenin. Biflavonoids are dimers of flavonoids that do not result in cleavage to anthocyanidins on treatment with strong acid. They are products of oxidative coupling mainly of flavones, flavanones and/or aurones and thus, they possess a carbonyl group at C-4 [30, 138]. As a result of this C-4 carbonyl group, the biflavonoids do not produce anthocyanidins upon hydrolysis in strongly acidic environments [139, 140].

The structure of the biflavone, manniflavanone (56) is shown below. This compound has a C-C interflavanil linkage between the C-3 and C-8 carbons on each flavone moiety. Several biflavonoids with C-O-C interflavanil bonds have recently been identified [139-145].



Manniflavanone (56)

Flavone–Coumarin Hybrids

This is comparatively a new class of compounds and only two flavones of this type are known so far. K. Franke *et al.* (2000), reported 6-(8''-Umbelliferyll)-apigenin (57) and 8-(6''-Umbelliferyll)-apigenin (58), from *Gnidia socotrana* (Thymelaeaceae) [30, 146], which represent a new structural type of natural products. However, biscoumarins and flavonoid dimers are frequently described in Thymelaeaceae [147].

Glycosides:

C-Glycosyl Flavonoids:

C-Glycosides are a special type of glycoside since the aglycone is directly attached to carbon 1 of a pyranose ring of a sugar while O-glycosides possess a hemiacetal linkage. Presently over one hundred C-glycosyl flavonoids are known to exist in the plant kingdom. Most C-glycosylflavones are known to occur naturally in pairs as 6- and 8-C-glycosyl isomers. The model compound and one of the first to be isolated was vitexin (64) (8-C-3-D-glucopyranosylapigenin), the structure of which was established by NMR spectroscopy by Horowitz and Gentilini. The 6-C-glycosyl isomer of vitexin (64), named isovitexin (saponaretin) (65), may be formed by an intramolecular rearrangement [148,149].

Xanthone C-glycoside:

It was the first compound with a C-glucosyl residue linked to a polyoxygenated xanthone nucleus was obtained by Wiechowski in 1908 from *Mangifera indica* [34] but the final structure of mangiferin (67) as 2-C--D-glucopyranosyl-1,3,6,7-tetrahydroxanthone was not established until 60 years later [35-41]. Mangiferin (67), being the best-known C-glucosyl xanthone, represents the most important features of all other hitherto known related substances: the glucosyl residue is either attached to carbon 2 or 4 of the xanthone nucleus which is oxygenated in 1, 3, 6 and 5 or 7 positions, respectively.[150]

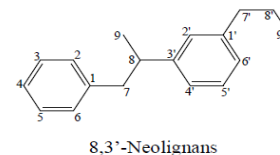
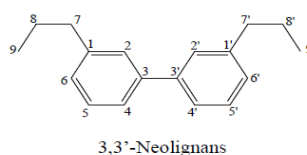
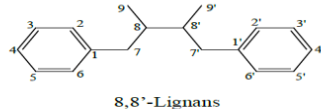
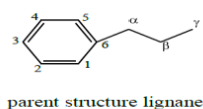
Benzophenone glycosides

Benzophenone glycosides have been isolated from several species of plants kingdom. Ferrari *et al* (2000) reported two benzophenone-C-glycosides, 2,3,4',5,6-pentahydroxybenzophenone-4-C-glucoside (68) and 2,4',6-trihydroxy-4-ethoxy benzophenone-2-O- glucoside (69)

Phenylpropanoic acid glycosides

Franke *et al* (2002) reported 2-O-β-D-glucosyloxy-4-methoxybenzenepropanoic acid (70) and Methyl-2-O-β-D-glucosyloxy-4-methoxybenzenepropanoate (71) from *Gnidia socotrana*.

Lignans



Lignans and neolignans dimers consisting of two phenyl propane monomers linked through C–C or C–O bonds and probably play an important role in defense system of in vascular plants and provide resistance against opportunistic pathogens. They also display several pharmacological activities such as antibacterial, antifungal, antitumour, antimitotic, [151-154].

A variety of lignans and neolignans have been reported from Thymelaeaceae such as furan lignans (lariciresinol (74), taxiresin (75)), furofuran lignans (pinoresinol (76)), dibenzylbutyrolactone lignans (wikstromol (78), kusunokinin (79), matairesinol (80)), diepoxy lignan (macronone (81)), coumarinolignan (daphneticin (82), daphneticin-4''-O-α-D-glucopyranoside (83)), etc. [37, 101 155-160]. Adicardin (84) and Syringin (85) were reported from *Gnidia socotrana* by Franke *et al* (2002) [30].

Spiro-bis-γ-lactone

Murray and Bradshaw (1966) suggested these substances possibly have a role of as carbohydrate storage compounds or as products of a detoxification mechanism that removes phenolic compounds. On the other hand these compounds could represent ascorbigens which are formed by the coupling of ascorbic acid and p-coumaric acid or 3-hydroxy-3-p-hydroxyphenylpropionic acid [161, 162].

K. Franke *et al.* (2000), for the first time reported compounds with the rare spiro-bis-γ-lactone structure 4',6'-Diacetyl-viburnolide A (87), 4',6'-Diacetyl-12-coumaroyl-viburnolide A (88) and Tetracetyl-viburnolide A (89), in the family of Thymelaeaceae [30]. Similar compounds, such as conocarpin, leucodrin and related substances, are typical constituents of Proteaceae [163, 164]. Murray and Bradshaw (1966) discussed a possible role of these substances as carbohydrate storage compounds or as products of a detoxification mechanism that removes phenolic compounds.

Sesquiterpenes (Guaiane-type)

The sesquiterpenes are a class of compounds that have 15 carbon atoms that are formally assembled from 3 isoprene (2-methylbutane) units.

Gnididione (90) is a Furanosquiterpene of the guaiane class, isolated from the stem wood and stem bark of the Kenyan shrub *Gnidia latifolia* by Kupchan and his co-workers. This was the first (and so far, only) member of the guaiane family that contains a furan ring.

Compound	Physical data	Molecular wt. (g/mol)	Molecular formula	Source and Plant part	References
Daphnane Diterpenes					
Daphnetoxin (1)	194-196 ⁰ C	482.52	C ₂₇ H ₃₀ O ₈	<i>G. polystachya</i>	[165]
Mezerein (2)	258-262 ⁰ C	654.71	C ₃₈ H ₃₈ O ₁₀	<i>L. bruchelli</i> <i>G. kraussiana</i>	[53, 71, 75, 166, 167]
12-Hydroxydaphnetoxin (6)	-	498	C ₂₇ H ₃₀ O ₉	<i>L. burchellii</i>	[75, 168- 170]
Gnididin (8)	241.8 ⁰ C	648.73	C ₃₇ H ₄₄ O ₁₀	<i>G. lamprantha</i>	[48, 71, 170]
Gniditrin (9) Syn: Daphne factor P ₁	246.21 ⁰ C	646.27	C ₃₇ H ₄₂ O ₁₀	<i>G. kraussiana</i> <i>G. lamprantha</i>	[31,48, 71, 170, 171]
Gnidicin (10) = Thymeleatoxin, 12-O-Cinnamoyl-5- hydroxy-6,7-epoxy- resiniferonol-9,13,14- orthobenzoate	-	628.66	C ₃₆ H ₃₆ O ₁₀	<i>G. kraussiana</i> <i>L. burchellii</i>	[31, 75,170, 171]
Gnidilatin (11)	234.34 ⁰ C	652	C ₃₇ H ₄₈ O ₁₀	<i>G. latifolia</i> <i>G. kraussiana</i>	[23,31, 71, 170]
Gnidilatidin (12) = odoracin, Yuanhuacine	204-206 ⁰ C	648.29	C ₃₇ H ₄₄ O ₁₀	<i>G. latifolia</i> <i>G. kraussiana</i>	[23] [31]
Gnidilatin-20-palmitate (13)	241.02 ⁰ C	890	C ₅₃ H ₇₈ O ₁₁	<i>G. latifolia</i>	[23, 71, 170]
Gnidilatidin-20-palmitate (14)	242.5 ⁰ C	886	C ₅₃ H ₇₄ O ₁₁	<i>G. latifolia</i>	[23, 31]
Gnidiglaucin (15)	219.8 ⁰ C	590.70	C ₃₂ H ₄₆ O ₁₀	<i>G. glaucus</i>	[23, 170]
Gnidimacrin (23)	172-174 ⁰ C	774.89	C ₄₄ H ₅₄ O ₁₂	<i>G. subcordata</i>	[23, 50, 71, 170]
Gnidimacrin-20-palmitate (24)		1012	C ₆₀ H ₈₄ O ₁₃	<i>G. subcordata</i>	[50, 71]
Compound	Physical data	Molecular wt. (g/mol)	Molecular formula	Source and Plant part	References
Gnilatmacrin (25) = Pimelea factor P ₂ , Daphnopsis factor R ₁	228.8 ⁰ C	638.78	C ₃₇ H ₅₀ O ₉	<i>G. kraussiana</i>	[31]
Excoecariatoxin (16)	215.5 ⁰ C	528.63	C ₃₀ H ₄₀ O ₈	<i>G. kraussiana</i> <i>G. lamprantha</i>	[31] [48, 172]
Pimelea factor P ₁ (17) Wikstrotoxin D	212.8 ⁰ C		C ₃₀ H ₄₄ O ₈	<i>G. lamprantha</i> <i>L. kraussiana</i>	[172]
Montanin (18)	214.2 ⁰ C	560	C ₃₂ H ₄₈ O ₈	<i>G. kraussiana</i>	[31]
Kraussianin (80) = Linimacrin C	242.8 ⁰ C	654.78	C ₃₇ H ₅₀ O ₁₀	<i>G. kraussiana</i>	[31, 173]
5β- dihydroxyresiniferonol- 6α,7α-oxide (81) = Excoecaria factor O ₁	-	525	C ₃₀ H ₃₈ O ₈	<i>L. burchellii</i> <i>G. lamramtha</i>	[174]

Genkwadaphnin (99) = 12-Benzoydaphnetoxin	-	602.63	C ₃₄ H ₃₄ O ₁₀	<i>L. burchellii</i>	[75, 168-170]
Coumarins					
Umbelliferone (24)	230 ⁰ C	162.14	C ₉ H ₆ O ₃	<i>G. polycephala</i>	[22, 175]
Lasiocephalin (28)	217-218 ⁰ C	336.06	C ₁₉ H ₁₂ O ₆	<i>L. eriocephalus</i>	[1,115,176-179]
Gnidicoumarin (43)	355-365 ⁰ C	304.11	C ₁₈ H ₈ O ₅	<i>G. lamprantha</i>	[23, 47, 180]
Lasioerin (44)	350 ⁰ C	304.11	C ₁₈ H ₈ O ₅	<i>L. eriocephalus</i>	[54, 135, 177]
Eriocephaloside (45)	350 ⁰ C	466.09	C ₂₄ H ₁₈ O ₁₀	<i>L. eriocephalus</i>	[136, 159, 177]
Eriocide (46) = 6,8- dihydroxy-7-O-β-D- glucosyloxy-coumarin	350 ⁰ C	518	C ₁₅ H ₁₆ O ₁₀	<i>L. eriocephalus</i>	[166, 177]
7,7'-dihydroxy-3,8'- biscoumarin (47)	-	322.04	C ₁₈ H ₁₀ O ₆	<i>G. socotrana</i> (Leaves and twigs)	[30]
Daphnetin (83) =3,4- dihydroxycoumarin	253-256 ⁰ C	178.14	C ₉ H ₆ O ₄	<i>L. polycephalus</i>	[181]
Compound	Physical data	Molecular wt. (g/mol)	Molecular formula	Source and Plant part	References
Daphnetin-8-β-D- glucoside (84)	263.4 ⁰ C	340.07	C ₁₅ H ₁₆ O ₉	<i>G. polycephala</i>	[181]
Flavonoids					
Manniflavanone (48)	220-225 ⁰ C	590	C ₃₀ H ₂₂ O ₁₃	<i>G. involucrata</i> (Aerial parts)	[25]
6-(8''-Umbelliferyll)- apigenin (49)	360 ⁰ C	453.05	C ₂₄ H ₁₄ O ₈	<i>G. socotrana</i> (Leaves and twigs)	[30]
8-(6''-Umbelliferyll)- apigenin (50)	270-275 ⁰ C	453.05	C ₁₅ H ₁₆ O ₁₀	<i>G. socotrana</i> (Leaves and twigs)	[30]
Genkwain (85) = 4',5- Dihydroxy-7- methoxyflavone	277-279 ⁰ C	284.27	C ₁₆ H ₁₂ O ₅	<i>L. eriocephalus</i>	[136, 177,182]
Genkwain-4'-O-β-D- glucopyranosyl-(1-3) D- xylopyranoside (86)	-	608.17	C ₂₈ H ₃₂ O ₁₅	<i>L. eriocephalus</i>	[182]
Kaempferol-3-O- glucoside (87) =: Astragalin	312.58 ⁰ C	448.38	C ₂₁ H ₂₀ O ₁₁	<i>G. involucrata</i> (Aerial parts)	[25]
Kaempferol-3-(p- coumaroyl)-O-β-D- glucopyranoside(88) =Tiliroside	214- 2160C	594.52	C ₃₀ H ₂₆ O ₁₃	<i>G. kraussiana</i>	[18]
Gnidia biflavonoid 4a (89) (2S,2'R,3R,3'R)-2,2',3,3'- Tetrahydro-3',5,5',7,7'- pentahydroxy-2-(4- hydroxyphenyl)-2'-(3,4,5- trihydroxyphenyl)-[3,8'-bi- 4H-1-benzopyran]-4,4'- dione		590.10	C ₃₀ H ₂₂ O ₁₃	<i>G. involucrata</i> (Aerial parts)	[141]
C- Glycosyl flavonoids					
Vitexin (51) 8-C-β-D- glucopyranosylapigenin	262-264 ⁰ C	432	C ₂₁ H ₂₀ O ₁₀	<i>G. involucrate</i>	[25, 183]

Isovitexin (52) = Saponaretin, 6-C-β-D- glucopyranosylapigenin	244– 246 ⁰ C.	432	C ₂₁ H ₂₀ O ₁₀	<i>G. involucrata</i> (Aerial parts)	[25, 183]
Compound	Physical data	Molecular wt. (g/mol)	Molecular formula	Source and Plant part	References
Isoorientin (93)= <i>Luteolin 6-C-β-D- glucoside</i>	244– 246 ⁰ C.	448	C ₂₁ H ₂₀ O ₁₁	<i>G. involucrata</i> (Aerial parts)	[25, 183]
Yuankanin (94)= Genkwanin-5-O-β-D- primeveroside, 4',5- dihydroxy-6-[(6-O-β-D- xylopyranosyl)-β- glucopyranosyl) oxy]-7- methoxyflavone	301.9 ⁰ C	578.51	C ₂₇ H ₃₀ O ₁₄	<i>G. involucrata</i> (Aerial parts)	[25, 184]
Xanthone C-Glycosides					
Mangiferin (53) = 1,3,6,7-Tetrahydroxy-2- [3,4,5-trihydroxy-6- (hydroxymethyl) oxan-2- yl]xanthen-9-one	271-274 ⁰ C	422.34	C ₁₉ H ₁₈ O ₁₁	<i>G. involucrata</i> (Aerial parts)	[25, 177, 183]
Benzophenone Glycosides					
Mahkoside A (90) = 4,4'- dihydroxy-6- methoxybenzophenone- 2-O-β-D- glucopyranoside	-	422	C ₂₀ H ₂₂ O ₁₀	<i>G. involucrata</i> (Aerial parts)	[25, 185, 186]
2,3,4',5,6- pentahydroxybenzopheno ne-4-C-glucoside (54)	168-171 ⁰ C	424	C ₁₉ H ₂₀ O ₁₁	<i>G. involucrata</i> (Aerial parts)	[25]
2,4',6-trihydroxy-4- methoxybenzophenone-2- O-glucoside (55)	135 ⁰ C	422	C ₂₀ H ₂₂ O ₁₀	<i>G. involucrata</i> (Aerial parts)	[25]
Phenyl propanoid glucosides					
2-O-α-Dglucosyloxy-4- methoxy benzenepropanoic Acid (56)	168– 169 ⁰ C	358	C ₁₆ H ₂₂ O ₉	<i>G. polycephala</i>	[22, 318]
Methyl 2-O-β-D- glucosyloxy-4- methoxybenzenepropan oate (57)	168– 169 ⁰ C	372	C ₁₇ H ₂₄ O ₉	<i>G. polycephala</i>	[22, 318]
Compound	Physical data	Molecular wt. (g/mol)	Molecular formula	Source and Plant part	References
Lignans					
Adicardin (71) = 7-O-beta-D- Apiofuranosyl-(1-6)- beta-D-glucopyranosyl- umbelliferone	201-202 ⁰ C	456.4	C ₂₀ H ₂₄ O ₁₂	<i>G. polycephala</i>	[22, 318]
Syringin (72) = 4-[(1E)-3-Hydroxyprop- 1-en-1-yl]-2,6- dimethoxyphenyl β-D- glucopyranoside	109-111 ⁰ C	372.37	C ₁₇ H ₂₄ O ₉	<i>G. polycephala</i>	[22,136, 177,318]

Gnidifolin (91) = [<i>trans</i> -2(2,4-dihydroxy-3-methoxybenzyl)-3-(4'-hydroxy-3'-methoxybenzyl)butyrolactone]	-	374	C ₂₀ H ₂₂ O ₇	<i>G. latifolia</i>	[51]
Syringaresinol (92) = (2,6-bis (4-hydroxy-3,5-dimethoxyphenyl)-3,7-dioxabicyclo [3.3.0] octane)	183-184 ⁰ C	418.43	C ₂₂ H ₂₆ O ₈	<i>L. eriocephalus</i>	[136, 177, 187]
Spiro-bis-γ-lactone					
4',6'-Diacetyl-viburnolide A (73)	-	568.13	C ₂₅ H ₂₇ O ₁₅	<i>G. socotrana</i> (Leaves and twigs)	[30]
4',6'-Diacetyl-12-coumaroyl-viburnolide A (74)	-	713.17	C ₃₄ H ₃₃ O ₁₇	<i>G. socotrana</i> (Leaves and twigs)	[30]
Tetraacetyl-viburnolide A (75)	219-221 ⁰ C	652.16	C ₂₉ H ₃₂ O ₁₇	<i>G. socotrana</i> (Leaves and twigs)	[30]
Sesquiterpenes (Guanine-type) Guanine Furanosesquiterpenoid					
Gnididione (76)	-	244.29	C ₁₅ H ₁₆ O ₃	<i>G. latifolia</i>	[188- 190]
Isognididione (95)	-	244.29	C ₁₅ H ₁₆ O ₃	<i>G. latifolia</i>	[188- 190]
Compound	Physical data	Molecular wt. (g/mol)	Molecular formula	Source and Plant part	References
Sterols					
β- Sitosterol (96)	136-140 ⁰ C	414.71	C ₂₉ H ₅₀ O	<i>G. kraussiana</i>	[18, 136, 177]
β- Sitosterol-β-Dglucoside (97)	361 ⁰ C	576.8	C ₃₅ H ₆₀ O ₆	<i>G. kraussiana</i> <i>L. eriocephalus</i>	[18, 182]
γ-Pyrone					
Maltol (98) 3-Hydroxy-2-methyl-4H-pyran-4-one	161-162 ⁰ C	126.11	C ₆ H ₆ O ₃	<i>G. kraussiana</i>	[18]
Lipids and Fatty acids					
n-Eicosanol, 1-Glyceryloctacosanoate, n-Octacosanol, Palmitic acid				<i>G. kraussiana</i> <i>L. meissnerianus</i>	[18, 191]
Amino acid					
Alanine, γ-Aminobutyric acid, Arginine, Aspartic acid, Cystine, Glutamic acid, Glycine, Aminoacetic acid, Histidine, Isoleucine, Leucine, Lysine, Methionine, Phenylalanine, Proline, Serine, Threonine, Tyrosine, Valine				<i>L. eriocephalus</i> <i>L. eriocephalus</i>	[192] [192]
Sugars					
Glucose, Maltose, Raffinose, Rhamnose				<i>L. eriocephalus</i>	[192]
Simple organic acids					
Citric acid, Fumaric acid, Malic acid, Malonic acid				<i>L. eriocephalus</i>	[192]

Table: 2: Phytochemical aspects of *Gnidia* species**Biological Aspects:**

Plants of Thymelaeaceae as well as Euphorbiaceae are well known for their interesting physiological activities. Several *Gnidia* and other African Thymelaeaceae species have been used in the traditional treatments of a variety of medicinal complaints in humans and animals. The toxicity of plants in the Thymelaeaceae is well established

for humans as well as for several animal species [3]. The diterpenes esters of tigliane and daphnane type are of violent purgatives, which trigger off, by contact with the skin or mucous membranes, an intense inflammatory reaction [193].

The symptoms of systematic toxicity resulting from ingestion of the plant material include: inflammation of lips, larynx and pharynx, dryness buccal cavity followed by salivation, difficult in swallowing, thirst; rhinitis, edema of eyelids; dizziness, abdominal pain, vomiting, diarrhoea with discharge of blood; albuminuria and blackwater fever, slow respiration, rapid pulse; pale, cold and moist skin; muscular twitching, delirium and drowsiness which can last several days. These symptoms are followed by convulsion and death in 20-25% of cases [194,3].

From Anti-feedant experiments of *Lasiosiphon anthyloides* Meisn., Watt and Breyer-Brandwijk concluded that, the plant in its flowering stage, on ingestion of 4 oz. by sheep or 12 oz. by cattle and horses produced death so rapidly that without any symptoms, apart from acute abdominal pain. Ingestion of smaller amounts produces hemorrhagic gastro-enteritis, pyrexia, inflammatory changes in the intestines, and death. Hutchence fed one pound of *Gnidia ovalifolia* Meisn. leaves daily to cattle and observed development of frothing in the mouth, dullness, absence of rumination, pyrexia, progressive diarrhea, anorexia, progressive weakness, mucous discharge from the eyes, nostrils and vagina, gastroenteritis, hyperemia of

the stomach and intestines, congestion of the kidney and death in fifteen days[3, 195].

The toxicity of plants in the Thymelaeaceae is of considerable economic importance beyond its effects on the livestock industry. Several genera including *Gnidia* and *Lasiosiphon* are used as fish poisons in Europe, Africa, Asia, and the islands of the South Pacific. Several genera are also used as hunting poisons in Africa and Asia. [3].

In France, extracts of *Lasiosiphon kraussiana* Hutch. et Dalz, have been patented for use in the treatment of leprosy [375,376] [3]. Clinical trials are being conducted in China on preparations of *Daphne*, *Gnidia*, *Wikstroemia* and *Pimelea* species that have been reported to have anticancer activity

[46,133,184,186,187,189,212,355,368,431]. The abortifacient and anticancer activities of these products have been shown to be due primarily to the presence of daphnane esters. Plants of this family have been included in several large-scale screening studies [24, 25, 72, 73,161,228] investigating various biological activities.

Esters of diterpene alcohols having the daphnane and tigliane skeleta are reported to be responsible for the irritancy, cocarcinogenicity, abortifacient activity and antileukemic activity of this plant family. This class of compounds has been extensively reviewed [3,87,124-126].

Sr No.	Species	Traditional Use / effects noted
1.	<i>Gnidia spp</i>	Abdominal pain, sore throat, as a purgative, as laxative, wounds healing, burns, snakebites, as molluscicidal agent, as arrow and fish poison[30], Asthma, eye diseases and pthisis, drastic purgative (root), blood purifier (root-bark), cough, headache, toothache, analgesic (Root-wood) [195-198], caused diarrhoea and emphysema [199], Analgesic, Antispasmodic, decongestant, toxic (leaves)[200], Vesicant [201]
2.	<i>G. anthyloides (L.f.) Gilg</i>	Cough [195, 202]
3.	<i>G. buchananii</i>	Bronchitis and abdominal pain(root) [198]
4.	<i>G. burchelli</i> (Meisn.) Gilg.	Caused severe irritation to eyes, nose and skin, dyspnoea. Coughing headache and nausea [5], caused diarrhoea and emphysema [199, 203, 204], Toxic and irritant [204, 205, 317]
5.	<i>Gnidia capitata</i> L. f. (syn. <i>Lasiosiphon capirarus</i>)	Leaves snuffed and also applied as a poultice for toothache[196, 204], Roots Smoke is inhaled for asthma, [206, 204], Anthrax (root)[207, 208], Treats constipation and menstrual pains, gastrointestinal complaints, earache, appetite, burns snake bite, boils and eases child birth [209], used in pregnancy and childbirth [210, 167], toxic and irritant [211].
6.	<i>G. carinata</i> Thunb.	Emetic (Fruit and leaves) [198, 212]
7.	<i>G. chrysantha</i> Gilg	Abortifacient, purgative, as body wash in fever (Root and leaves) [196], toxic and irritant [196]
8.	<i>G. cuneata</i> Meisn. (syn. <i>Lasiosiphon meisneri</i> End.)	Roots used for snakebite and toothache [20,204]
9.	<i>Gnidia danguyana</i> J. Leand.	Anti-Malarial and febrifuge (Aerial parts)[213]

10.	<i>G. genkwa</i>	[62]
11.	<i>G. glabra</i>	Laxative and emetic (root)[214]
12.	<i>G. goetzeana</i> Gilg.	Antitussive (root) [198]
13.	<i>G. gymnostachya</i> Gilg.	Analgesic [215, 204] Analgesic, Antispasmodic, decongestant, toxic (leaves) [215, 204]
14.	<i>G. involucrata</i> Steud.	To reduce the size of the vaginal orifice and as fish poison (roots) [25], Laxative and Vermifuge (root) [25, 214,216], as insecticide, insect repellent and anti-malarial [217]
15.	<i>G. kraussiana</i> Syn: <i>L. kraussiana</i> Hutch. et Dalz.	For chest complaints [26, 202], Stomach ache and enema (Root), Leaves for earache and toothache, Measles, Dropsy, Anorexia, Ulcers (root) [206, 214, 218, 219], Molluscicidal agent (leaves)[205], Arrow and Fish Poison (Leaves and root) [220-222], as an emetic or purgative and also to treat a condition called chidyiso, which is attributed to bewitched food and characterized by gastrointestinal pain [223], Anorexia and Antipsychotic (root) [224], Diarrhoea (stem bark) [225], ear ache, abdominal pain, toothache (leaves) [204], fractures [208], to induce labour, abortifacient, relieves stomach troubles. Fever, menstrual pain, cold and rheumatism [209], used in pregnancy and childbirth; severe gastrointestinal irritation and death of stock [167, 226, 227,317], leprosy (root) [228, 229], Treatment of burns, snake bites, stomach complaints, used to ensure easy birth [230, 26], constipation [193,315]
16.	<i>G. kraussi</i>	Snake bite [231]
17.	<i>G. latifolia</i>	Purgative (root-bark) [198], Antifeedant effects [190, 321]
18.	<i>G. macrorrhiza</i>	Stimulant [232]
19.	<i>G. microcephala</i>	Aphrodisiac (root) [198]
20.	<i>G. polycephala</i> Gilg.	Toxic and irritant [205, 204, 317], Severe irritation to eyes, nose and skin, dyspnoea. Coughing. Headache and nausea [5, 199, 203, 204], constipation (root) [233], for stabilising heart conditions, treatment of tuberculosis and tonsillitis, and ashes are applied onto wounds [22, 234]
21.	<i>G. socotrana</i>	Obstipation (Leaf and fruit) [235]
22.	<i>G. somalensis</i> Gilg.	Tuberculosis (root) [214]
23.	<i>G. stenophylla</i>	Malaria, Acaries, Rabies [236]
24.	<i>G. stenophylloides</i> Gilg.	Antiblennorrhagic (root) [237]
25.	<i>G. vatkeana</i> Engl. Et Gilg.	Purgative, Vesicant (root) [196, 201]
26.	<i>Lasiosiphon spp.</i>	Anticatarrh, Headache, Rubefacient, Wound dressing [197], Eruptive fever, Ophthalmia, Snakebite (root) [196,195], Blood purifier, Sore throat, Tonic (root-bark) [191, 238]
27.	<i>L. anthyloides</i>	Febrifuge, bad dreams [196], Cough, Influenza, Snakebite (root) [196,195, 197, 238], Respiratory tract irritant (root) [201]
28.	<i>L. bipinnatum</i>	Hepatotoxic and respiratory irritant [5, 199], cattle and sheep Poisoning [239]
29.	<i>L. burchellii</i>	Respiratory tract irritant (root) [201]
30.	<i>L. capitatus</i>	Fractures and sprains, Toothache [196, 220], Burns, Aid women in conception, Aid in childbirth, Emetic in anthrax, Enema for backache, Febrifuge, Prevent scalding in febrile patients, Stomachic, Wash for babies, Sore throat (root) Headache (leaf) [196]
31.	<i>L. kraussii</i> Meisn	Arrow, Animal and Fish Poison (Leaves and roots)[195-197, 240], Snake bite, sore throat, washing wounds and bruises [195-197, 238], Burns (leaves) [197], Purgative (leaves and root) [241, 242], for snake bite [231,243]
32.	<i>L. linifolius</i>	Toothache, headache, burns (root), emetic in anthrax (root), purgative and sore throat, for lung inflammations (root)[195,197, 238],
33.	<i>L. meisnerianus</i>	Irritant (root) [201], Blood purifier, respiratory diseases, indolent and callous ulcers [191, 196, 195], Blood purifier, Sore throat, Tonic (root-bark) [191], Sores ophthalmia, disease of sheep and snake bite [191,196, 195, 220, 238,244,]
34.	<i>L. perrieri</i> J. Leand	Anti-Malarial and febrifuge [213]
35.	<i>L. roridus</i> S. Moore	Malaria [196]

Table 3: Biological aspects of *Gnidia* species

Sr No.	Species	Biological Activity
1.	<i>G. burchellii</i> Meisn.	Toxicity studies [203]
2.	<i>G. capitata</i>	Spasmolytic (leaves) [196], Prostaglandin-synthesis inhibition [245], Antibacterial, anthelmintic, brine shrimp toxicity and toxicity studies [246, 247]
3.	<i>G. chrysantha</i> Gilg.	Toxic and irritant [196]
4.	<i>G. cuneata</i> Meisn	<i>In-vitro</i> antiplasmodial activity absent [248]
5.	<i>Gnidia gymnostachya</i> Glig.	Antispasmodic [204, 215]
6.	<i>G. kraussianus</i> Hutch. et Daix.	P-388 lypmhocryptic leukaemia [31,], Death [204, 206], Analgesic, antispasmodic, decongestant, toxic (Roots) [200], Anti-Leprotic (root) [228, 229, 249], natural post harvest grains protectant and insecticidal [250, 251], toxicity studies (roots) [252], potent insecticidal activity [172], molluscicidal activity (leaf, stem and root) [205, 253], <i>In-vitro</i> antiplasmodial activity absent [248], Acaricidal (root) [254], Antibacterial and antifungal [230]
7.	<i>G. macropetala</i>	Prostaglandin-synthesis inhibition [245]
8.	<i>G. socotrana</i>	Antimicrobial activity [235]
9.	<i>G. stenophylla</i>	Anti-malarial (root) [255, 236], antioxidant (root) [256]
10.	<i>G. stenophylloides</i> Gilg.	Antiblennorrhagic (root) [214], toxicity studies and antimalarial [255]
11.	<i>G. subcordata</i>	Antileukemic [4, 50]
12.	<i>Lasiosiphon anthyloides</i> Meisn.	Toxic [195, 257], Respiratory tract irritant (Root) [201], antifeedant [238]
13.	<i>Lasiosiphon burchellii</i> Meisn.	Respiratory tract irritant (Root) [201],
14.	<i>L. meiasnerianus</i> Endl.	Irritant (rootbark) [201]

Table 4: biological activities studies on *Gnidia* species

Sr No.	Plant Part	Traditional Use	Biological Activity
1.	Whole plant	Rubefacient [258, 177], Irritant (bark) [201], Fish poison [259], Insecticide and piscicide [260] vesicant [179], cancers, sore throat, abdominal pain, wounds, burns and snakebites, as molluscicidal, insecticidal, piscicidal, homicidal agents and as arrow poisons [261-264], Rabies [265], tuberculosis [266]	<i>In vitro</i> anticoagulant activity due to Biscoumarin [177, 267], antileprosy [166], antioxidant and anthelmintic activity [268], larvicidal [253, 261, 269], showed good insecticidal, herbicidal and fungicidal activities [270], mild antifungal activity [271, 272], Spasmolytic activity [273, 204], Antidiabetic: potent Amylase and Glucosidase Inhibitor [263, 274-276], antimicrobial [277], Biopesticide [278], Larvicidal [319].
2.	Roots	Indigestion [198], Rabies [260]	Antileukemic [260], Acute toxicity studies [279]
3.	Leaves	Teeth loosening effect Snakebite [221, 177, 280-282], as insecticide, fish poison for blisters, contusions and swellings, back ache and joint pains [77, 260, 263],	Bactericidal effect [283], Irritant [201], Anti-inflammatory [77], Larvicidal [284], Insecticidal and fungicidal [260, 285], Anti-feedant [260, 286], as herbal pesticide [287], Ovicidal [272], pest prevention [288], Antimicrobial [277]
4.	Bark	Arrow and fish poison, for blisters, contusions, swellings and Insecticide [107, 221, 260, 280, 281, 289-291], Diuretic, Sialagogue, Stimulant (bark) [244], Vesicant (bark) [191, 221, 280, 281, 292],	Larvicidal [260], Acute toxicity studies [279], Ovicidal [272], Antimicrobial [277]
5.	Stem	Fish poison [260]	
6.	Root	Rabies [293]	Anti-leukaemia [260]

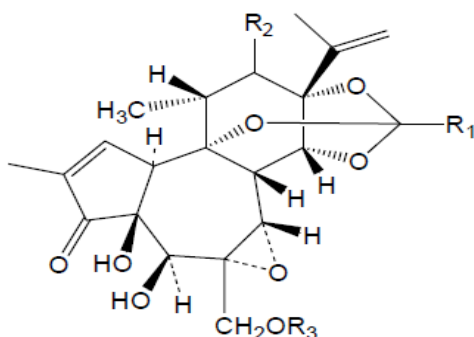
Table 5: Plant part, traditional use and biological activity of *Gnidia* species

Sr. No.	Isolated Constituents	Activity
1.	Diterpene esters of the daphnane type	Antineoplastic activity[23, 31,48, 50, 50], pesticide and insecticidal [294]
2.	Mezerein (2)	Antileukemic activity [53]
3.	Gnididin (8)	Potent antileukemic activity [170]
4.	Gnidicin (10)	Potent antileukemic activity [77, 170]
5.	Gniditrin (9)	Potent antileukemic activity [170], potential cholesterol lowering activity [295]
6.	Excoecariatoxin (16)	Irritant, Piscicidal, Insecticidal [172]
7.	Gnidilatin (11)	Moderate inflammatory cytokines inhibitory activity [296], Piscicidal [297], Moderate Antileukemic activity [170], Moderate inhibitory effects on interleukin 1 (IL-1alpha, IL-1beta) and tumour necrosis factor (TNF-alpha) biosynthesis [313]
8.	Gnidilatin-20 palmitate (13)	Moderate inflammatory cytokines inhibitory activity [296], Substantial Antileukemic activity [297, 170], Moderate inhibitory effects on interleukin 1 (IL-1alpha, IL-1beta) and tumour necrosis factor (TNF-alpha) biosynthesis [313]
9.	Gnidilatidin (12)	Piscicidal [297], No Antileukemic activity [170, 298, 299], Abortifacient [314]
10.	Gnidilatidin-20-palmitate (14)	Substantial Antileukemic activity [297, 170]
11.	Gnidiglaucin (15)	No Antileukemic activity [170]
12.	Lasiocephalin (28)	Toxicity studies [300]
13.	12-hydroxydaphnetoxin (4)	No antileukemic activity due to absence of ester linkage [170], Potent inhibitory effects on interleukin 1 (IL-1alpha, IL-1beta) and tumour necrosis factor (TNF-alpha) biosynthesis [313]
14.	Genkwanin (59)	Antiplasmodial And Cytotoxic [301], Potent inhibitory effects on interleukin 1 (IL-1alpha, IL-1beta) and tumour necrosis factor (TNF-alpha) biosynthesis [313], Antimicrobial [316]
15.	Genkwadaphnin (60)	Potent inhibitory activity on inflammatory cytokines [296], Antileukemic [302]
16.	Syringaresinol (77)	Lymphatic vessel stabilizer [303]
17.	Syringin (85)	Ineffective on cytokine biosynthesis [296]
18.	Kraussianin (26)	Antileukemic [173]
19.	Mahkoside A (72)	Cytotoxicity [185,312]
20.	Mangiferin (67)	Anti-inflammatory, analgesic, antipyretic, antioxidant, immunomodulator, antitumor, antiviral, and anthelmintic and in obesity treatment, antiallergic, Bronchodilatory, antidiabetic, Antiplasmodial Antiamoebic, Cardioprotective, Radioprotective, Lipolytic, Antibacterial and antifungal [304-307], α -amylase inhibitor [275]
21.	Vitexin (64)	α -Glucosidase Inhibitory Activity [308]
22.	Isovitexin (65)	Antioxidant [309], α -Glucosidase Inhibitory Activity [308]
23.	Isoorientin (66)	Hepatoprotective and hypoglycemic [310], Anti-nociceptive, anti-inflammatory and gastroprotective activities, antioxidant, antimicrobial, myolitic activity [311, 316]
24.	Kirkinine (6)	Potent neurotrophic activity [320]

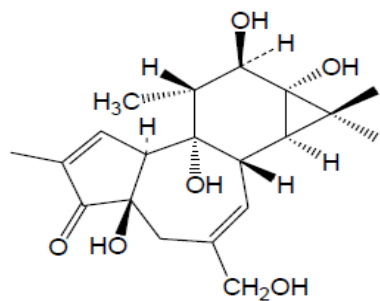
Table: 5: Isolated Constituents and their biological activity of *Gnidia* species

Structures

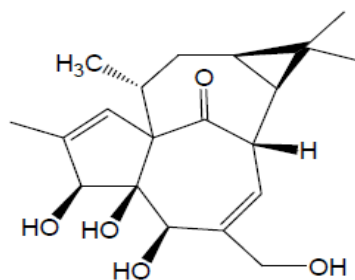
Diterpenes:



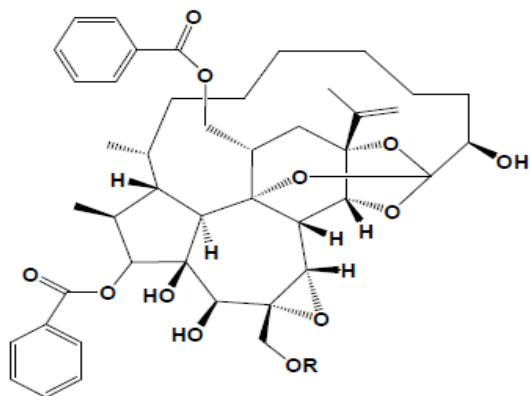
Daphnetoxin	(1) R ₁ = C ₆ H ₅	R ₂ = H	R ₃ = H
Mezerein	(2) R ₁ = C ₆ H ₅	R ₂ = CO(CH=CH) ₂ C ₆ H ₅	R ₃ = H
Huratoxine	(3) R ₁ = (CH=CH) ₂ (CH ₂) ₈ CH ₃	R ₂ = H	R ₃ = H
12-β-hydroxydaphnetoxin	(4) R ₁ = C ₆ H ₅	R ₂ = OH	R ₃ = H
Acetoxyluratoxin	(5) R ₁ = (CH=CH) ₂ (CH ₂) ₈ CH ₃	R ₂ = OCOCH ₃	R ₃ = H
Kirkinine	(6) R ₁ = (CH=CH)(CH ₂) ₁₂ CH ₃	R ₂ = OCOCH ₃	R ₃ = H
Kirkinine D	(7) R ₁ = (CH=CH) ₃ (CH ₂) ₂ CH ₃	R ₂ = OCOCH ₃	R ₃ = H
Gnididin	(8) R ₁ = C ₆ H ₅	R ₂ = CO(CH=CH) ₂ (CH ₂) ₄ CH ₃	R ₃ = H
Gniditrin	(9) R ₁ = C ₆ H ₅	R ₂ = CO(CH=CH) ₃ (CH ₂) ₂ CH ₃	R ₃ = H
Gnidicin	(10) R ₁ = C ₆ H ₅	R ₂ = COCH=CHC ₆ H ₅	R ₃ = H
Gnidilatin	(11) R ₁ = (CH ₂) ₈ CH ₃	R ₂ = COC ₆ H ₅	R ₃ = H
Gnidilatidin	(12) R ₁ = (CH=CH) ₂ (CH ₂) ₄ CH ₃	R ₂ = COC ₆ H ₅	R ₃ = H
Gnidilatin-20-palmitate	(13) R ₁ = (CH ₂) ₈ CH ₃	R ₂ = COC ₆ H ₅	R ₃ = CO(CH ₂) ₁₄ CH ₃
Gnidilatidin-20-palmitate	(14) R ₁ = (CH=CH) ₂ (CH ₂) ₄ CH ₃	R ₂ = COC ₆ H ₅	R ₃ = CO(CH ₂) ₁₄ CH ₃
Gnidiglaucin	(15) R ₁ = (CH ₂) ₈ CH ₃	R ₂ = COCH ₃	R ₃ = H
Excoercariatoxin	(16) R ₁ = (CH=CH) ₂ (CH ₂) ₄ CH ₃	R ₂ = H	R ₃ = H
Pimelea factor P ₁	(17) R ₁ = C ₉ H ₁₉	R ₂ = H	R ₃ = H
Montanin	(18) R ₁ = C ₁₁ H ₂₃	R ₂ = H	R ₃ = H
5β-dihydroxyresiniferonol-6α,7α-oxide	(19) R ₁ = (CH=CH) ₃ (CH ₂) ₂ CH ₃	R ₂ = H	R ₃ = H
Genkwadaphnin	(20) R ₁ = C ₆ H ₅	R ₂ = COCH ₃	R ₃ = H



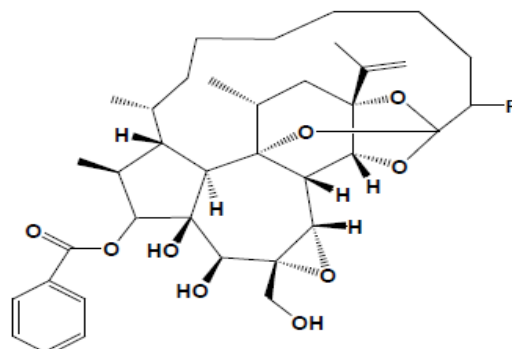
Phorbol (21)



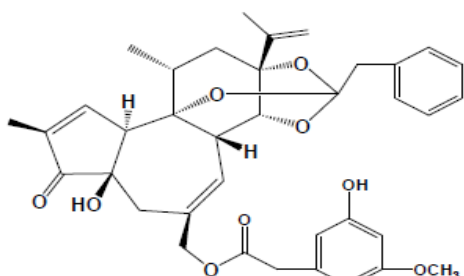
Ingenol (22)



Gnidimacrin (23) R = H
 Gnidimacrin-20-palmitate (24) R = CO(CH₂)₁₄CH₃

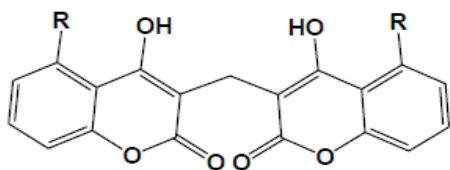


Daphnopsis factor R₁ (Gnilatimacrin) (25) R = H
 Kraussianin (26) R = OH

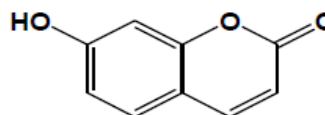


Resiniferotoxin (27)

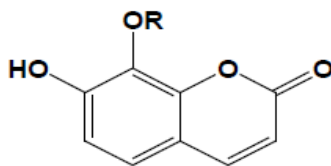
Coumarins:



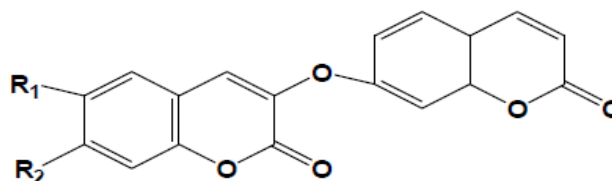
dicoumarol (28), R = H
 Gerberinol (29), R = CH₃



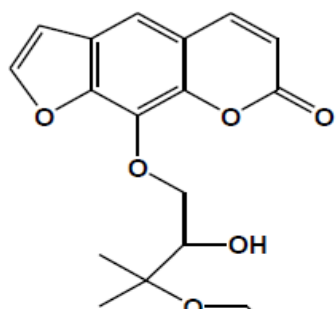
Umbelliferone (30)



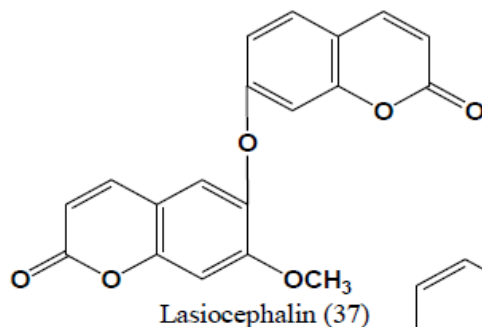
Daphnetin (31) R₁ = H
 Daphnetin-8-β-D-glucoside (32) R₁ = Glu



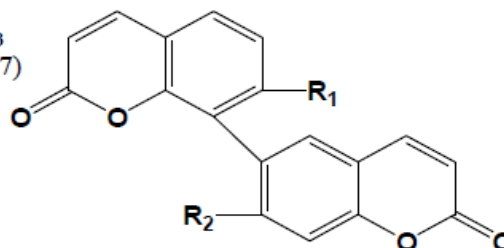
Daphnoretin (33) R₁ = OCH₃ R₂ = OH
 7-acetyldaphnoretin (34) R₁ = OCH₃ R₂ = OCOCH₃
 Edgeworthin (35) R₁ = OH R₂ = OH



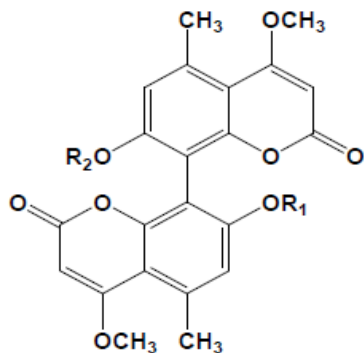
Candicanin (36)



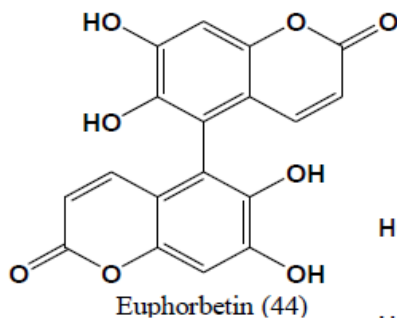
Lasiocephalin (37)



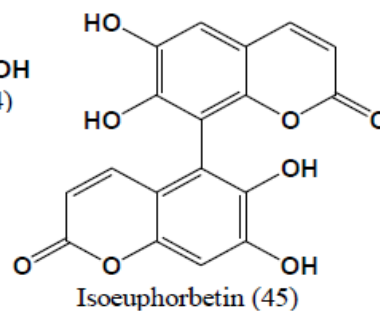
Bicoumol (38) $R_1 = OH$ $R_2 = OH$
 Bhubaneswin (39) $R_1 = OH$ $R_2 = OH$
 Matsukaze lactone (40) $R_1 = OCH_3$ $R_2 = OCH_3$



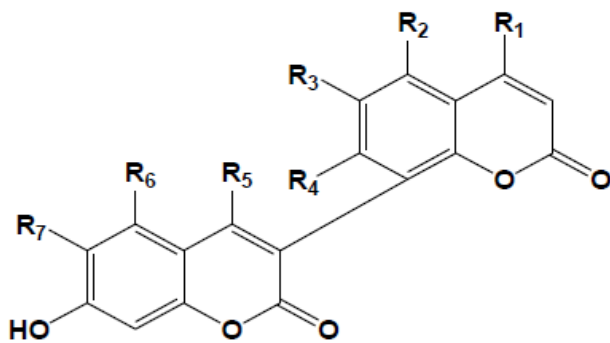
Kotanin (41) $R_1 = CH_3$ $R_2 = CH_3$
 Desmethylkotanin (42) $R_1 = H$ $R_2 = CH_3$
 Orlandin (43) $R_1 = H$ $R_2 = H$



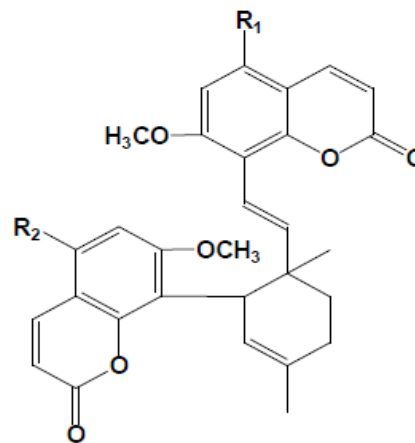
Euphorbetin (44)



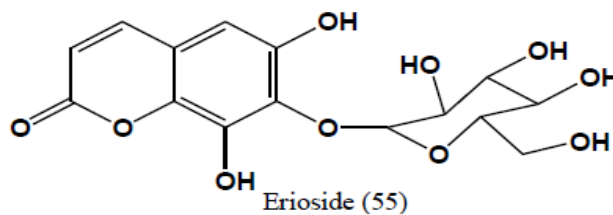
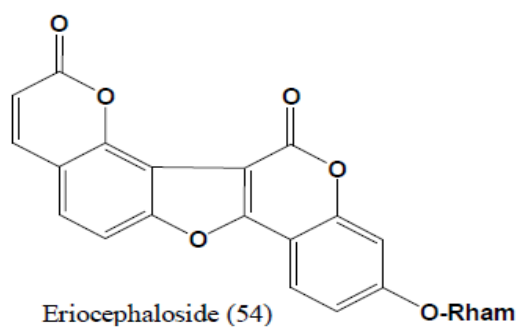
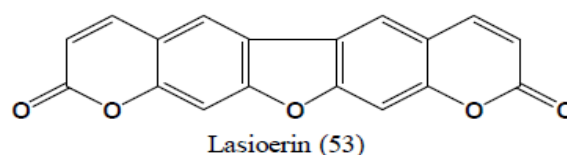
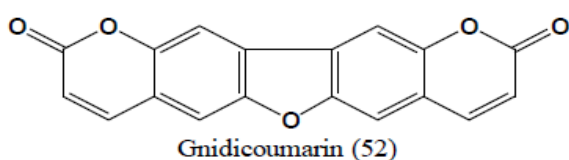
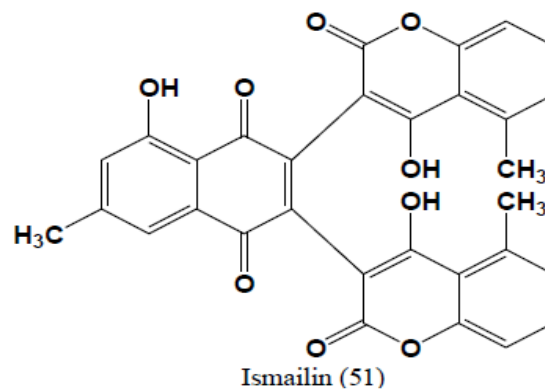
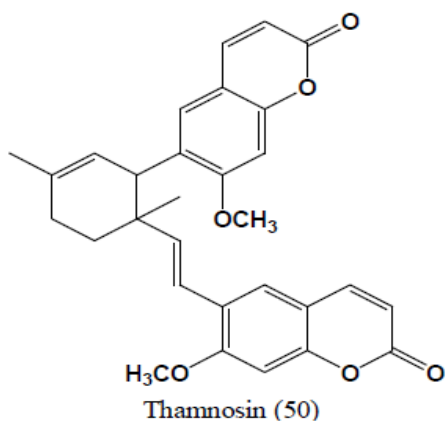
Isoeuphorbetin (45)



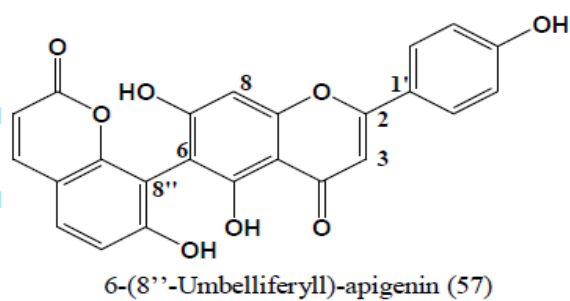
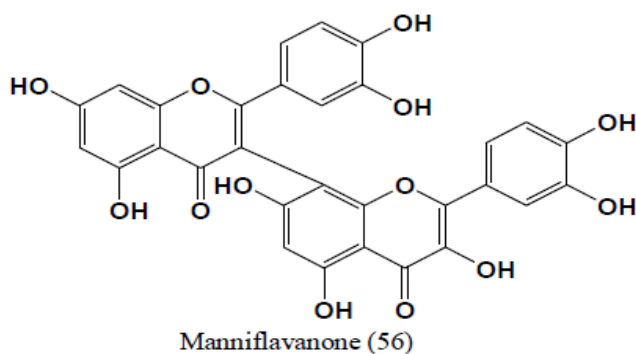
Ipomosin (46) $R_1 = R_2 = R_5 = R_6 = H$
 $R_3 = R_7 = CH_3$
 $R_4 = OH$
 7,7'-dihydroxy-3,8'-biscoumarin (47) $R_3 = R_7 = H$
 $R_2 = R_6 = CH_3$
 $R_1 = R_4 = R_5 = OCH_3$

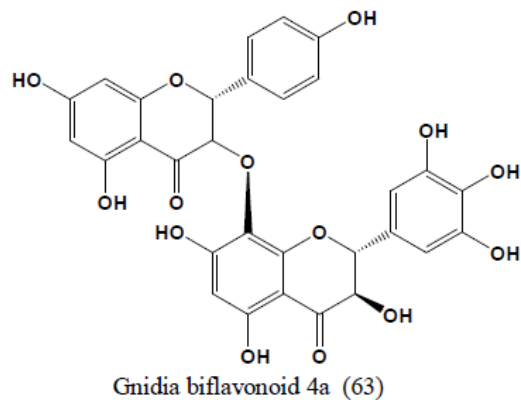
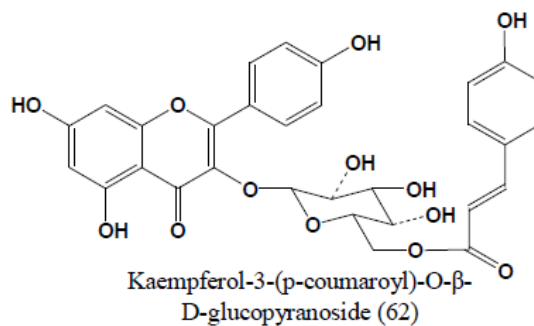
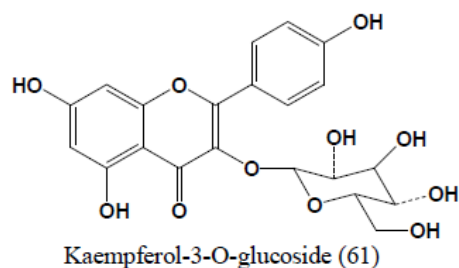
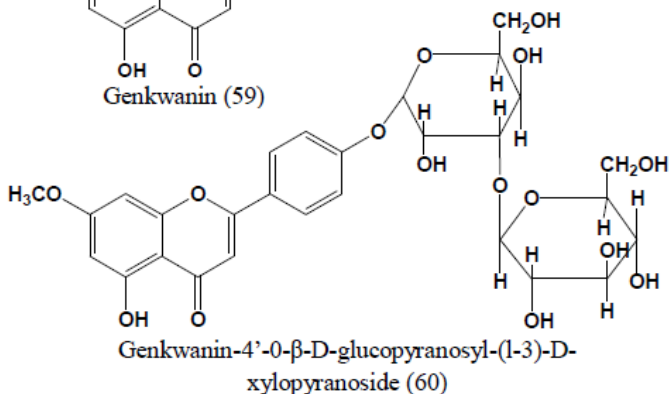
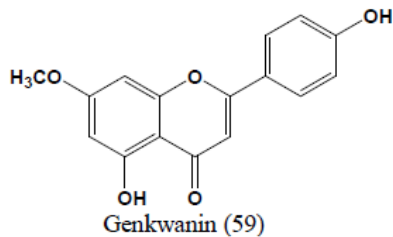
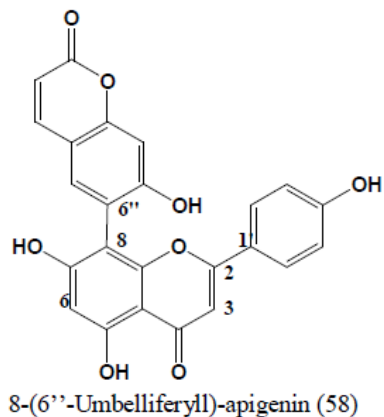


Phebalin (48), $R_1 = R_2 = H$
 Toddasin (49), $R_1 = R_2 = OCH_3$

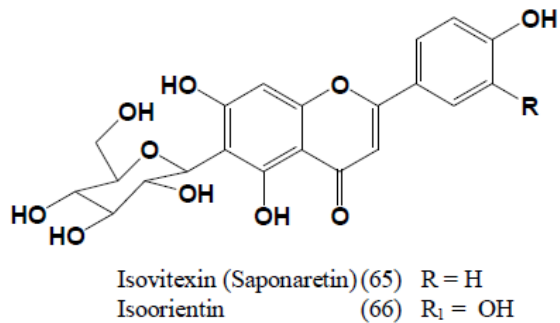
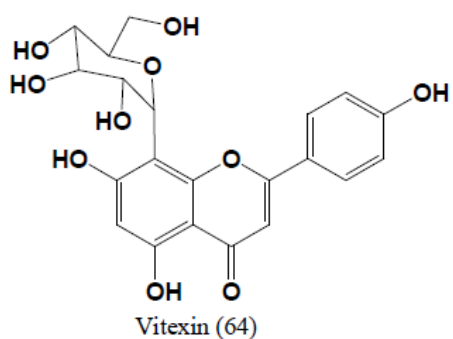


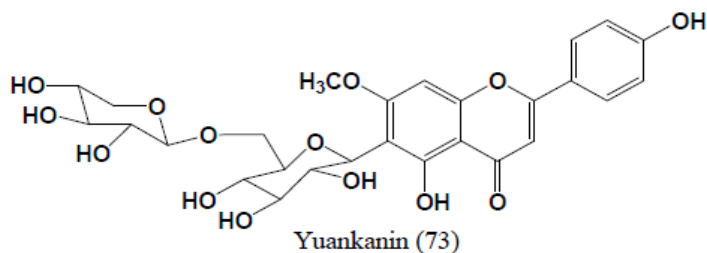
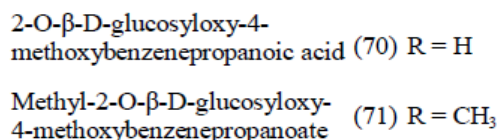
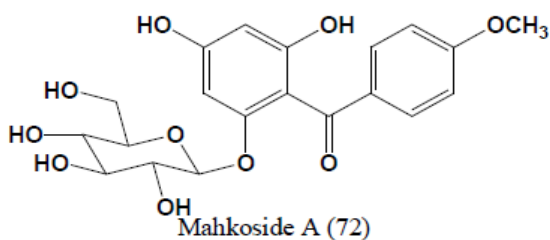
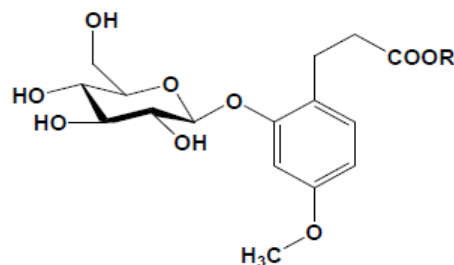
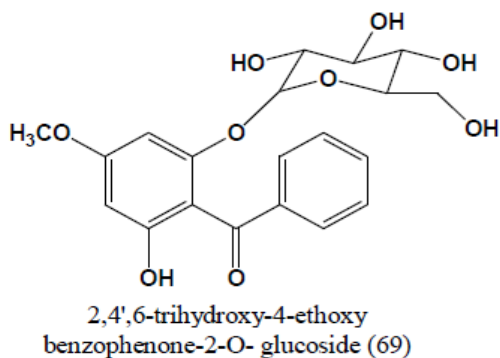
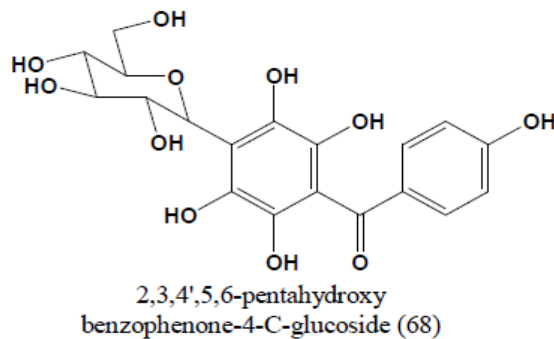
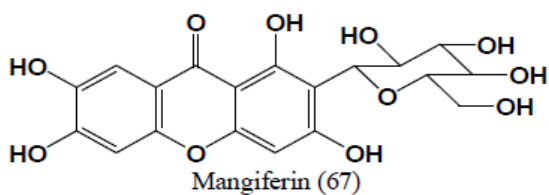
Flavonoids:



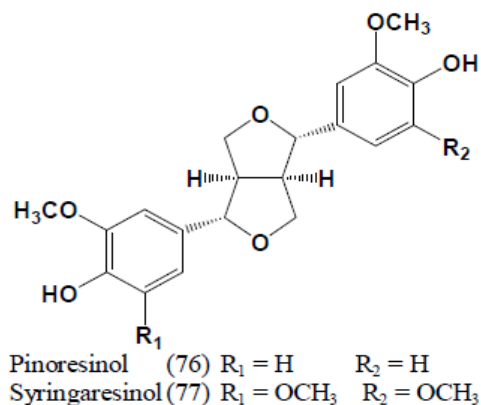
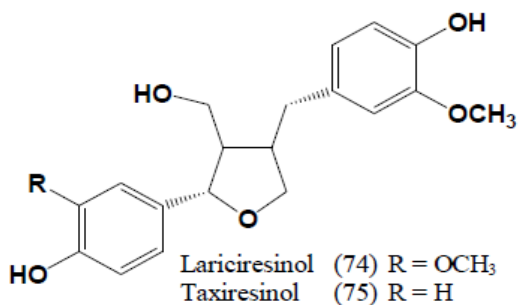


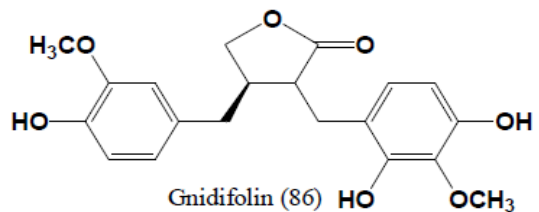
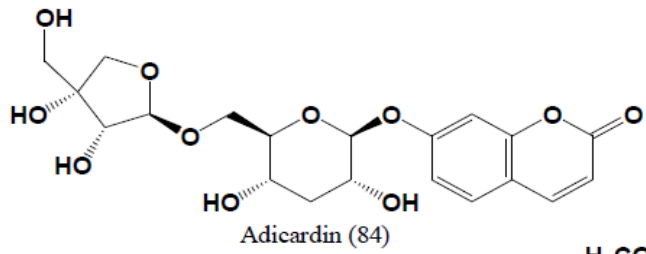
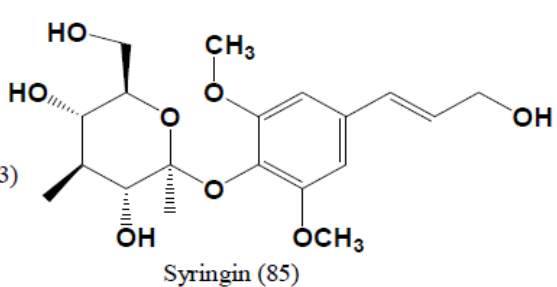
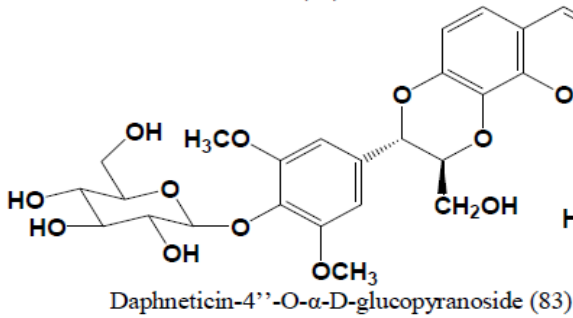
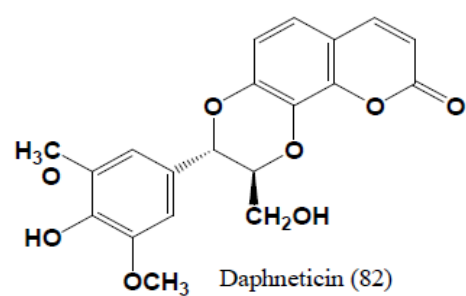
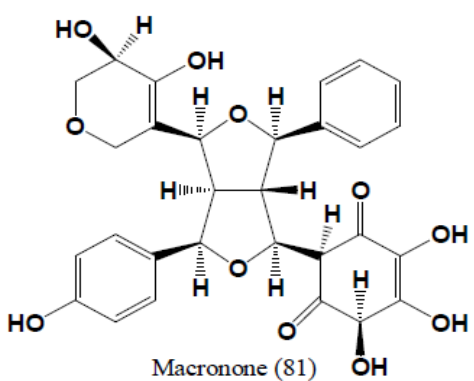
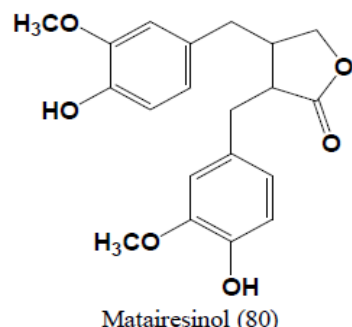
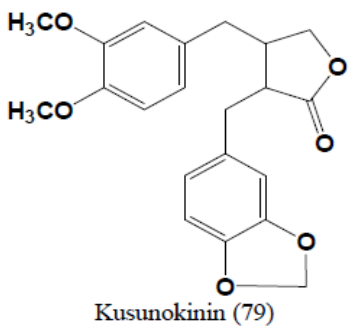
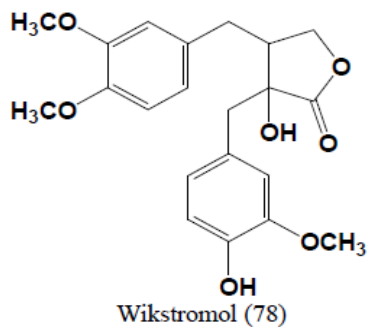
Glycosides:



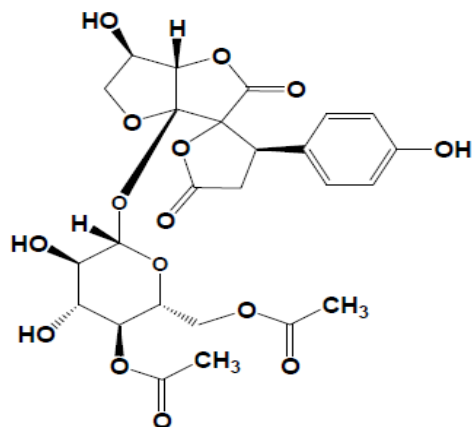


Lignans:

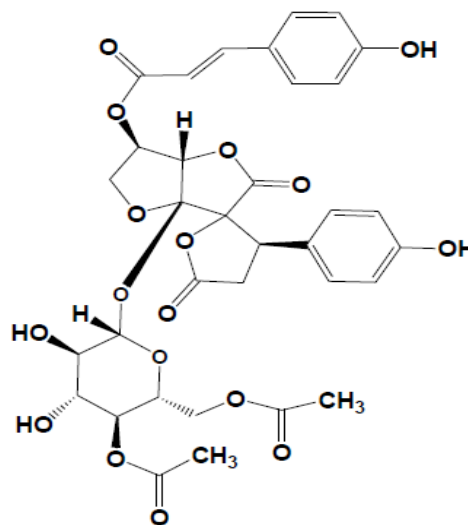




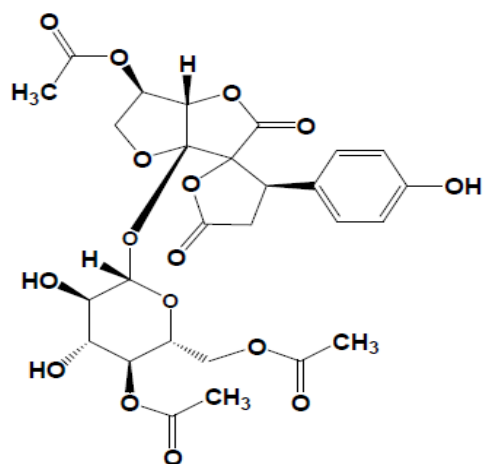
Spiro-bis- γ -lactone:



4',6'-Diacetyl-viburnolide A (73)

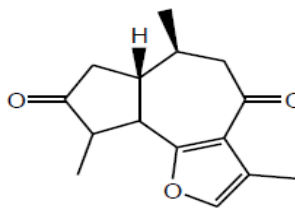


4',6'-Diacetyl-12-coumaroyl-viburnolide A (74)



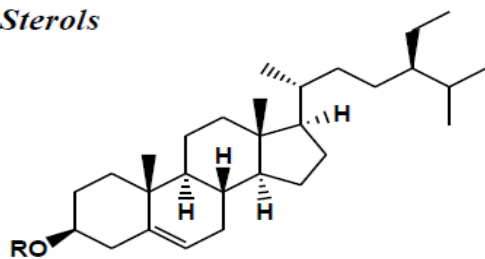
Tetraacetyl-viburnolide A (75)

Sesquiterpenes (Guanine-type):



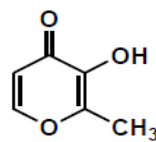
Gnididione (90)

Sterols



β - Sitosterol (96) R = H
 β - Sitosterol- β -Dglucoside (97) R = Glu

γ -Pyrone



Maltol (98)

Acknowledgements:

We would like to thank ICMR for the financial support for this research (21/12/17/09/HSR, dated: 24/06/2010).

References:

1. Hussain H, Hussain J, Al-Harrasi A, Krohn K. The chemistry and biology of bicoumarins. *Tetrahedron*. 2012;68 (report number 969):2553-78.
2. Heywood HV. Flowering plants of the world. London,UK: B.T. Batsford Ltd; 1993.
3. Beaumont AJ, Edwards TJ, Smith FR. Leaf and bract diversity in Gnidia (Thymelaeaceae): patterns and taxonomic value. *Systematics and Geography of Plants*. 2001;71:399-418.
4. He W, Appending G, Van-Puyvelde L, Leysen JE, DeKimpe N. Daphnane-Type Diterpene Orthoesters and their Biological Activities. *Mini Reviews in Medicinal Chemistry*. 2002;2:185-200.
5. Kellerman TS, Coetzer JAW, Naude TW, Botha CJ. Plant Poisonings and Mycotoxicoses of Livestock in Southern Africa. 2 ed. Cape Town: Oxford University Press Southern Africa; 2005;310.
6. Cary M, The Geographic background of Greek and Roman History. London, England: Oxford University Press; 1949.
7. Herber BE, ed. Flowering Plants. Dicotyledons. Malvales, Capparales and Non-betalain Caryophyllales. Berlin: Springer; 2003.
8. Peterson B. Some interesting species of Gnidia. *Botaniska Notiser*. 1959;112(4):465-80.
9. Levyns MR. Thymelaeaceae. Flora of The Cape Peninsula. Juta, Cape Town 1950.
10. Peterson B. Thymelaeaceae. In: Polhill RM, editor. Flora of Tropical East Africa. London, United Kingdom: Crown Agents for Oversea Governments and Administrations; 1978. p. 1-37.
11. Bredenkamp CJ, Beyers JBP. Thymelaeaceae (Dilleniidae-Euphorbiales). In: Liestner OA, editor. Seed plants of Southern Africa: families and genera. Pretoria: Strelitzia 10, National Botanical Institute; 2000. p. 544-6.
12. Hyde M, Wursten B. Gnidia L.: Description of the genus 2002 [17-Feb 2013]. Available from: www.zimbabweflora.co.zw/speciesdata/genus.php?genus%20id=996.
13. Pooley E. Mountain Flowers: A field guide to the flora the Drakensberg and Lesotho. Durban: The Flora Publications Trust; 2003.
14. Beaumont AJ, Edwards TJ, Smith FR. Thymelaeaceae. Goldblatt P, Manning L, editors. Missouri Botanical Gardens: Strelitzia 9: National Botanical Institute; 2000. 676-85 p.
15. Marloth R. The Flora of South Africa: with synoptical tables of the genera of higher plants. Cape Town: Darter Bros. & Co.; 1925.
16. Notten A. Gnidia squarrosa L. (Druce): Kirstenbosch National Botanical Garden; 2004 [cited 2013 17 Feb]. Available from: <http://www.plantzafrica.com/plantefg/gnidiasquar.htm>.
17. Van-Wyk CM, Gericke N. People's Plants. Pretoria: Briza Publications; 2000.
18. Borris RP, Blasko G, Cordell GA. Ethnopharmacologic and Phytochemical studies of Thymelaeaceae. *Journal of Ethnopharmacology*. 1988;24 41-91.
19. Van-Wyk BE, Van-Heerden F, Van-Oudtshoorn B. Poisonous Plants of South Africa. Pretoria: Briza Publications; 2005.
20. Smith A. A contribution to South African Materia Medica: chiefly from plants in use among the natives. 3rd ed. Cape town, South Africa: The Publishing Department; 1895, pp238.
21. Sohni YR, Mutangadura-Mhlanga T, Kale PG. Bacterial Mutagenicity of eight medicinal herbs from Zimbabwe. *Mutation Research / Genetic Toxicology*. 1994;322:133-40.
22. Munkombwe NM, Galebotswe P, Modibesane K, Morebodi N. Phenylpropanoid glycosides of Gnidia polycephala. *Phytochemistry*. 2003;64(8):1401-4.
23. Kupchan SM, Shizuri Y, Summer WC, Haynes H, Leighton AP, et al. Isolation and structural elucidation of new potent antileukemic diterpenoid esters from Gnidia species. *J Org Chem*. 1976;41:3850-3.
24. Khanyile S. Gnidia triplernervis Meisn: South African National Biodiversity Institute; 2006 [cited 2013 17 Feb]. Available from: <http://www.plantzafrica.com/plantefg/gnidiatrip.htm>.
25. Ferrari J, Terreaux C, Sahpaz S, Msonthi JD, Wolfender J-L, et al. Benzophenone glycosides from Gnidia involucrata. *Phytochemistry*. 2000;54(8):883-9.
26. Hutchings A. Zulu Medicinal Plants. An inventory. Pinetown: University of Natal Press; 1996;450.
27. Kupchan SM. Novel natural products with antitumor activity. *Fed Proc*. 1974;33(11):2288-95.
28. Roger ZS. A revision of Malagasy Gnidia (Thymelaeaceae, Thymelaeoideae). *Annals of the Missouri Botanical Garden*. 2009;96:324-68.
29. Peterson B. Flora Zambesiaca. In: Pope GV, Polhill RM, Martins ES, editors. 9. Kew: Royal Botanic Gardens; 2006. p. 85-117.
30. Franke K, Porzel A, Schmidt J. Flavone-coumarin hybrids from Gnidia socotrana. *Phytochemistry*. 2002;61:873-8.
31. Borris RP, Cordell GA. Studies of the Thymelaeaceae II. Actinoplastic principles of Gnidia kraussiana. *J Nat Prod*. 1984;47:270-8.
32. Badawi MM, Handa SS, Farnsworth ADKGCNR. Plant anticancer agents XXVII: antileukemic and cytotoxic constituents of Dirca occidentalis (Thymelaeaceae). *J Pharm Sci*. 1983;72:1285-7.
33. Cottigli F, Loy G, Garau D, Floris C, Casu M, et al. Antimicrobial evaluation of coumarins and flavonoids from the stems of Daphne gnidium L. *Phytomed*. 2001;8:302-5.
34. Yagura T, Shibayama N, Ito M, Honda G, Kiuchi F. Three novel diepoxy tetrahydrochromones from agarwood, artificially produced by intentional wounding. *Tetrahedron letters*. 2005;46(25):4395-8.
35. Li SH, Wu LJ, Gao HY, Chen YH, Li Y. A new dicoumarinoid glycoside from Daphne giraldii. *J Asian Nat Prod Res*. 2005;7:839-42.
36. Riaz M, Malik A. Daphsaifnin, a dimeric coumarin glucoside from Daphne oleoides. *Heterocycles* 2001;55:769-73.
37. Ullah N, Ahmad S, Anis E, Mohammad P, Rabnawaz H, et al. A lignan from Daphne oleoides. *Phytochemistry*. 1999a;50:147-9.
38. Zhang W, Zhang W, Li T, Liu R, Li H, et al. A new flavan from Daphne odora var. atrocaulis. *Fitoterapia*. 2004;75:799-800.
39. Su J, Wu Z, Shen Y, Zhang C, Zhang W. Lignans from Daphne giraldii. *Chem Nat Compd*. 2008;44:648-50.
40. Baba K, Yoshikawa M, Taniguchi M, Kozawa M. Biflavonoids from *Daphne odora*. *Phytochemistry*. 1995;38:1021-26.
41. Liang S, Tang J, Shen YH, Jin HZ, Tian JM, et al. Biflavonoids from *Daphne feddei* and their inhibitory activities against nitric oxide production. *Chem Pharm Bull*. 2008;56:1729-31.
42. Pan L, Zhang XF, Wu HF, Ding LS. A new daphnane diterpene from *Daphne tangutica*. *Chin Chem Lett*. 2006;17:38-40.
43. Taninaka H, Takaishi Y, Honda G, Imakura Y, Yesilada ESE. Terpenoids and aromatic compounds from Daphne oleoides ssp. oleoides. *Phytochemistry* 1999;52:1525-9.
44. Zhan Z, Fan CQ, Ding J, Yue JM. Novel diterpenoids with potent inhibitory activity against endothelium cell HMEC and cytotoxic

- activities from a well-known TCM plant *Daphne genkwa*. *Bioorg Med Chem.* 2005;13:645-55.
45. Zhang Y, Zhang H, Hua S, Ma L, Chen C, et al. Identification of two herbal compounds with potential cholesterol-lowering activity. *Biochem Pharmacol.* 2007;74:940-7.
 46. Liao SG, Chen HD, Yue JM. Plant orthoesters. *Chem Rev.* 2009;30:1092-140.
 47. Kupchan SM, Sweeny JG, Shen TMM-S, Bryan RF. Structure of Gnidia coumarin, a Novel Pentacyclic Dicoumarin from *Gnidia lamprantha*. *J Chem Soc, Chem Commun.* 1975:94-5.
 48. Balandrin MF, Klocke JA, Wurtele EJ, Bollinger WH. Natural plant chemicals: Sources of industrial and medicinal materials. 228(4704), 1154-1160: Science New Series; 1985.
 49. Kupchan SM, Sweeny JG, Baxter RL, Murae T, Zimmerly VA, et al. Gnididin, gniditrin, and gnidicin, novel potent antileukemic diterpenoid esters from *Gnidia lamprantha*. *J Am Chem Soc.* 1975;97:672-3.
 50. Kupchan SM, Shizuri Y, Murae T, Sweeny JG, Haynes R, et al. Gnidimacrin and gnidimacrin 20-palmitate, novel macrocyclic antileukemic diterpenoid esters from *Gnidia subcordata*. *J Am Chem Soc.* 1976;98(18):5719-20.
 51. Bryan RF, Shen MS. Gnidifolin [trans-2-(2,4-dihydroxy-3-methoxybenzyl)-3-(4'-hydroxy-3'-methoxybenzyl)butyrolactone]. *Acta Crystallogr Sect B* 1978;34(327-9).
 52. Hecker E. Co-carcinogenic principles from the seed oil of *Croton tiglium* and from other Euphorbiaceae. *Cancer Res.* 1968;28:2338-49.
 53. Evans FJ, Soper CJ. The tigliane, daphnane, ingenane diterpenes, their chemistry, distribution and biological activities, a review. *Lloydia.* 1978;41(3):193-233.
 54. Kupchan SM, Baxter RL. Mezerein: antileukemic principle isolated from *Daphne mezereum* L. *Science.* 1975;187:652.
 55. Powell RG, Smith-Jr. CR. Antitumour agents from higher plants. In: Swain T, Kleiman R, editors. *The Recent Advances in Phytochemistry.* Illinios: Plenum Publishing Corporation; 1980.
 56. Rajwar S, Khatri P, Patel R, Dwivedi S, Dwivedi A. An Overview on Potential Herbal Anticancer Drugs. *Intr J Res, Pharm Chem.* 2011;1(2):202-10.
 57. Evans FJ. *Naturally occurring phorbol esters.* Boca Raton: CRC Press; 1986.
 58. Kraft AS, Anderson WB. Phorbol esters increase the amount of Ca²⁺, phospholipid-dependent protein kinase associated with plasma membrane. *Nature.* 1983;301(5901):621-3.
 59. Rigby JH, Moore TL, Rege S. Synthetic studies on the ingenane diterpenes. Inter- and intramolecular [6 + 4] tropone-diene cycloaddition reactions. *J Org Chem.* 1986;51 (12):2398-400.
 60. Adolf W, Hecker E. Diterpenoid irritants and co-carcinogens in Euphorbiaceae and Thymalaeceae- Structural relationships in view of their biogenesis. *Isr J Chem.* 1977;16:75-83.
 61. Harborne JB, Baxter H, Moss GP. *Phytochemical Dictionary- A handbook of Bioactive Compounds from plants.* 2 ed. London: Taylor & Francis; 1999.
 62. He W, Cik M, Van-Puyvelde L, Van-Dun J, Appendino G, et al. Neurotrophic and antileukemic daphnane diterpenoids from *Synaptolepis kirkii*. *Bioorg Med Chem.* 2002;10:3245-55.
 63. Hu B, Sha H, Wang C, Yu D, Wu X, et al. Antifertility constituent of the flower *Yuan-Hua* (*Daphne genkwa*) – isolation and structure of yuanhuatine. *Hua Hsueh Hsueh Pao* 1985;43:460-2.
 64. Wang C, Chen Z, Ying B, Zhou B, Liu J, et al. Studies on active principles in the root of *Yuan-hua* (*Daphne genkwa*). II: Isolation and structure of a new antifertile diterpene yuanhuadine. *Hua Hsueh Hsueh Pao.* 1981;39:421-6.
 65. Wang C, Huang H, Xu R, Dou Y, Wu X, et al. Studies on the active principles of *Yuan-Hua* roots. III: Isolation and structure of yuanhuafin. *Hua Hsueh Hsueh Pao.* 1982;40:835-9.
 66. Ying BP, Wang CS, Chou PN, Pan PC, Liu JS. Studies on the active principles of the root of *Yuan-Hua* (*Daphne Genkwa*). I: Isolation and structure of yuanhuacine. *Hua Hsueh Hsueh Pao.* 1977;35:103-8.
 67. Sakata K, Kawazu K, Mitsui T. Piscicidal constituent of *Hura crepitans*. II: Chemical structure of huratoxin. *Agric Biol Chem.* 1971;35:2113-26.
 68. Carney JR, Krenisky JM, Williamson RT, Luo J, Carlson TJ, et al. Maprouneacin, a new daphnane diterpenoid with potent antihyperglycemic activity from *Maprounea africana*. *J Nat Prod.* 1999;62:345-7.
 69. Appendino G, Szallasi A. Euphorbium: modern research on its active principle, resiniferatoxin, revives an ancient medicine. *Life Sci.* 1997;60:681-96.
 70. Asada Y, Sukemori A, Watanabe T, Malla KJ, Yoshikawa T, et al. Stelleralides A–C, novel potent anti-HIV daphnane-type diterpenoids from *Stellera chamaejasme* L. *Org Lett.* 2011;13:2902-7.
 71. Stanoeva E, He W, DeKampe N. Natural and synthetic cage compounds incorporating the 2,9,10-trioxatricyclo[4.3.1.0^{3,8}]decane type moiety. *Bioorg Med Chem.* 2005;13:17-28.
 72. Stout GH, Bakengol WG, Poling M, Hickernell GL. The Isolation and Structure of dephretoxin the poisonous principle of *Daphne* species. *J Am Chem Soc.* 1970;92:1070-1.
 73. Sataka K, Kawazu K, Mitsui T, Masaki M. Structure and Stereochemistry of Huratoxin, a Piscicidal Constituent of *Hura crepitans*. *Tetrahedron Lett.* 1971:1141-4.
 74. Adolf W, Hecker E. On the irritant and cocarcinogenic principles of *Hippomane mancinella*. *Tetrahedron Lett.* 1975;19 1587-90.
 75. Coetzer J, Pieterse MJ. The isolation of 12-hydroxy-daphnetoxin, a degradation product of a constituent of *Lasiosiphon burchellii*. *J South Afr Chem Inst.* 1971;24:241-3.
 76. Niwa M, Takamizawa H, Tatematsu H, Hirata Y. Piscicidal constituents of *Stellera chamaejasme* L. *Chem Pharm Bull.* 1982;30(12):4518-20.
 77. Khare CP. *Indian Medicinal Plants: An Illustrated Dictionary.* Verlag Heidelberg: Springer; 2004, pp. 292.
 78. Nyborg J, La-Cour T. X-ray diffraction study of molecular structure and conformation of mezerein. *Nature.* 1975;257:824-5.
 79. Zayed S, Adolf W, Hecker E. On the active principles of the Thymelaeaceae. I. The irritants and cocarcinogens of *Pimelea prostrata*. *Planta Med.* 1982 45(2):67-77.
 80. Pettit GR, Zou JC, Goswami A, Cragg GM. Antineoplastic agents, 88; *Pimelea prostrata*. *J Nat Prod.* 1983;46(4):563-8.
 81. Hafez A, Adolf W, Hecker E. Active principles of the Thymelaeaceae, III: skin irritant and carcinogenic factors from *Pimelea simplex*. *Planta Med* 1983;49(1):3-8.
 82. Abe F, Iwase Y, Yamauchi T, Kinjo K, Yaga S. Daphnane diterpenoids from the bark of *Wikstroemia retusa*. *Phytochemistry.* 1997;44(4):643-7.
 83. Tyler MI, Howden MEH. 1-Monolinolenoylglycerol specifically potentiates antitumour activity of daphnane orthoesters. *Naturwissenschaften* 1985;72 (7):373-5.
 84. Abe F, Iwase Y, Yamauchi T, Kinjo K, Yaga S, et al. Minor daphnane-type diterpenoids from *Wikstroemia retusa*. *Phytochemistry.* 1998;47(5):833-7.
 85. Hergenbahn M, Adolf W, Hecker E. Resiniferatoxin and other esters of novel polyfunctional diterpenes from *Euphorbia resinifera* and *unispina*. *Tetrahedron Lett.* 1975;16(19):1595-8.

86. Szallasi A, Blumberg PM. Resiniferatoxin, a phorbol-related diterpene, acts as a ultrapotent analog of capsaicin, the irritant constituent in red pepper. *Neurosci.* 1989;30(2):515-20.
87. Kar A. Coumarin and Furanocoumarin Glycosides. 2 ed. New Delhi: New Age International (P) Limited Publishers; 2007.
88. Berebaum M. Patterns of Furanocoumarin Distribution and Insect Herbivory in the Umbelliferae: Plant Chemistry and Community Structure. *Ecology.* 1981;62(5):1254-66.
89. Liao SG, Zhang BL, Wu Y, Yue JM. New phenolic components of *Daphne giraldii*. *Helv Chim Acta.* 2005;88:2873-8.
90. Zhang BB, Dai Y, Liao ZX. Chemical constituents of *Saus-surea eopygmaea*. *Chin J Nat Med.* 2011;9:33.
91. Stamann MA, Huebner CF, Link KP. Studies on the hemorrhagic sweet clover disease. V. Identification and synthesis of the hemorrhagic agent. *J Biol Chem.* 1941;138:513.
92. Link KP. The discovery of dicumarol and its sequels. *Circulation* 1959;19:97-107.
93. Talapatra SK, Mukhopadhyay SK, Talapatra B. Minor coumarins of *Boenninghausenia albiflora*. *Phytochemistry.* 1975 14(3):836-7.
94. Szendrei K, Novak I, Reisch J, Minker E. Acridone alkaloids as constituents in the roots of *Boenninghausenia albiflora* Reichb. *Pharmazie.* 1975;30(11):753-4.
95. Kozawa M, Baha K, Hata MMHNK. Uber die Cumarine der *Boenninghausenia japonica* (Sieb) Nakai. *Chem Pharm Bull.* 1974;22:2746.
96. Basa SC. Natural Bicomarins. *Phytochemistry.* 1988;27(7):1933-41.
97. Komissarenko AN, Kovalev VN. Coronillobiosidol-A cardenolide glycoside from seeds of *Coronilla scorpioides*. *Chem Nat Compd.* 1988;24:619-21.
98. Williams M, Cassidy JM. Potential antitumor agents: a cytotoxic cardenolide from *Coronilla varia* L. *Journal of Pharmaceutical Sciences.* 1976;65(6):912-4.
99. Baba K, Takeuchi K, Hamasaki F, Kozawa M. Chemical studies on constituents of the Thymelaeaceae plants. I. Structures of two new flavans from *Daphne odora* THUNB. *Chem Pharm Bull.* 1986;34:595-602.
100. Majumder PL, Sengupta GC, Dinda BN, Chatterjee A. Edgeworthin, a new bis-coumarin from *Edgeworthia gardneri*. *Phytochemistry.* 1974;13(9):1929-31.
101. Zhuang LG, Sellgmann O, Jurcic K, Wagner H. Constituents of *Daphne tangutica*. *Planta Med.* 1982;45:172-6.
102. Thusoo A, Raina N, Minhaj N, Ahmed SR, Zaman A. Crystalline constituents from *Daphne oleoides*. *Indian J Chem.* 1981;20B:937-8.
103. Handa SS, Kinghorn AD, Cordell GA, Farnsworth NR. Plant anticancer agents. XXVI. Constituents of *Peddiea fischeri*. *J Nat Prod.* 1983 46(2):248-50.
104. Reisch J, Novak I, Szendrei K, Minker E. Chemistry of natural substances. XIX. Additional C3-substituted coumarin derivatives from *Ruta graveolens*: daphnoretin and its methyl ether. *Planta Med.* 1968;16:372-6.
105. Rizk AM, Rimpler H. Isolation of daphnoretin and β -sitosterol- β -d-glucoside from *Thymelea hirsuta*. *Phytochemistry.* 1972;11(1):473-5.
106. Livingston AL, Bickoff EM, Jurd L. Clover Constituents, Isolation of Daphnoretin from Ladino Clover. *J Agr Food Chem.* 1964;12(6):535-8.
107. Torrance SJ, Hoffmann JJ, Cole JR. Wikstromol, antitumor lignan from *Wikstroemia foetida* var. *oahuensis* Gray and *Wikstroemia uva-ursi* Gray (Thymelaeaceae). *J Pharm Sci.* 1979 68(5):664-5.
108. Kato A, Hashimoto Y, Kidohoro M. A new pharmacologically active lignan from *Wikstroemia indica*. *J Nat Prod.* 1979;42:159-62.
109. Lee KH, Tagahara K, Suzuki H, Wu RY, Haruna M, et al. Antitumor Agents. 49. Tricin, Kaempferol-3-O- β -D-Glucopyranoside and (+)-Nortrachelogenin, Antileukemic Principles From *Wikstroemia indica*. *J Nat Prod* 1981;44 (5):530-5.
110. Tandon S, Rastogi RP. Wikstrosin, a tricoumarin from *Wikstroemia viridiflora*. *Phytochemistry.* 1977;16(12):1991-3.
111. Cordell GA. Studies in the Thymelaeaceae I. Nmr Spectral Assignments of Daphnoretin. *J Nat prod.* 1984;47(1):84-8.
112. Phadke CP, Kelkar SL, Wadia MS. A general synthesis of bis[comarinyl] ethers: synthesis of daphnoretin methyl ether. *Synthesis.* 1986:413-5.
113. Chakrabarti R, Das B, Banerji J. Bis-coumarins from *Edgeworthia gardneri* *Phytochemistry.* 1986;25(2):557-8.
114. Bandopadhyay M, Malik SB, Seshadri TR. Candicanin, a novel bicomarinyll derivative from the roots of *Heracleum candicans*. *Tetrahedron Lett.* 1971;12(45):4221-2.
115. Bhattacharya AK, Das SC. Lasiocephalin: a new Coumarin from *Lasiosiphon ericephalus*. *Chem Ind (London).* 1971:885.
116. Spencer RR, Witt SC, Lundin RE, Bickoff EM. Bicoumol, a new bicomarinyll, from ladino clover. *J Agric Food Chem.* 1967:536-8.
117. Basa SC, Das DP, Tripathy RN, Elango V, Shamma M. Bhubaneswin: A New Bicomarins. *Heterocycles* 1984;22(333).
118. Miyazaki T, Mihashi S, Okabayashi K. Studies on the constituents of *Boenninghausenia albiflora* Meissner Var. *Japonica S. Suzuki II. Structure of Matsukaze-Lactone (2).* *Chem Pharm Bull (Tokyo).* 1964 12:1236-9.
119. Büchi G, Klaubert DH, Shank RC, Weinreb SM, Wogan GN. Structure and synthesis of kotanin and desmethylkotanin, metabolites of *Aspergillus glaucus*. *J Org Chem.* 1971;36(8):1143-7.
120. Büchi G, Luk KC, Kobbe B, Townsend JM. Four new mycotoxins from *Aspergillus clavatus* related to tryptoquivaline. *J Org Chem.* 1977;42:244-6.
121. Cutler HG, Crumley FG, Dorner RHCOHRJCJW. Orlandin: a nontoxic fungal metabolite with plant growth inhibiting properties. *J Agric Food Chem.* 1979 27(3):592-5.
122. Dutta PK, Banerjee D, Dutta NL. Euphorbetin: a new bicomarins from *Euphorbia lathyris* L. *Tetrahedron Letters.* 1972;13(7):601-4.
123. Dutta PK, Majumder PC, Dutta NL. Synthetic approaches towards bicomarins: Synthesis of euphorbetin and isoeuphorbetin. *Tetrahedron Lett.* 1975;31(9):1167-70.
124. Sharma DK, Seshadri TR. Oxidative Coupling of Esculetin and Isoscapoletin by $[\text{Fe}(\text{DMF})_3\text{Cl}_2][\text{FeCl}_4]$, Potassium Hexacyanoferrate(III) and Manganese Tris(acetylacetonate). *Indian J Chem.* 1977;15B:939.
125. Dutta PK, Banerjee D, Dutta NL. Isoeuphorbetin, a Novel Bicomarins from *Euphorbia lathyris* Linn. *Indian J Chem.* 1973;11:831.
126. Arisawa M, Kinghorn AD, Cordell GA, Farnsworth NR. Ipomopsin, a New Bicomarins from *Ipomopsis aggregata*. *J Nat Prod.* 1984;47:106.
127. Brown KL, Burfitt AIR, Mathai RCCDHKP. The Constituents of *Phebalium nudum*. III. The Structures of Phebalin and Phebalarin. *AustJ Chem.* 1975;28:1327.
128. Sharma PN, Shoeb A, Kapil RS, Popli SP. Toddasin, a new dimeric coumarin from *Toddalia asiatica*. *Phytochemistry.* 1980;19(6):1258-60.

129. Chakraborty DP, Roy S, Chakraborty A, Mandal AK, Chowdhury BK. Structure and synthesis of mexolide: a new antibiotic dicoumarin from *Murraya exotica* Linn. [syn. *M. paniculata* (L) jack.] *Tetrahedron*. 1980;36(24):3563-4.
130. Kutney JP, Dreyer TIDL. The Structure of Thamnosin. A Novel Dimeric Coumarin System. *J Am Chem Soc*. 1968;90:813.
131. Kutney JP, Dreyer TIDL. Further Studies on Constituents of *Thamnosma montana* Torr. and Frem. The Structure of Thamnosin. A Novel Dimeric Coumarin System. *Tetrahedron* 1970;26:3171.
132. Guise GB, Taylor ERRGSWC. The Chemical Constituents of Australian *Zanthoxylum* Species IV. Two New Coumarins from *Z. suberosum* C.T. White (syn. *Z. dominianum* Merr. and Perry; *Z. ovalifolium* Wight). *Aust J Chem*. 1967;20:2429.
133. Jeffreys JAD, Zakaria M, Waterman PG, Zhong S. A new class of natural product: homologues of juglone bearing 4-hydroxy-5-methyl-coumarin-3-yl units from *Diospyros* species. *Tetrahedron Lett*. 1983;24(10):1085-8.
134. Waterman PG, Zhong S-M, Jeffreys JAD, Zakaria M. Homologues of Juglone with 4-Hydroxy-5-methyl-2-oxo-2H-chromen-3-yl Substituents from *Diospyros* Species. A New Class of Natural Product. *J Chem Res Synop* 21985.
135. Sengupta S, Das SC. Structure of Lasioerin: a Novel Coumarin from *Lasiosiphon eriocephalus* Decne (Thymelaeaceae). *Chem Ind (London)* 1978:954.
136. Bhandari P, Rastogi RP. A novel type of Bicoumarin Rhamnoside from *Lasiosiphon eriocephalus*. *Phytochemistry*. 1981;20(8):2044-47.
137. Harborne JB, Williams CA. The Phytochemical richness of the Iridaceae and its systematic significance. *Annali di Botanica*. 2000;58:43-50.
138. Ferreira D, Slade D, Marais JPI. Flavans and Proanthocyanidins. In: Anderson OM, Markham KR, editors. *Flavonoids: Chemistry, Biochemistry and Applications*. Boca Raton: CRC Press; 2006. pp 553 - 616.
139. Bennie L, Malan E, Coetzee J, Ferreira D. Structure and synthesis of ether-linked proteracacinidin and promelacacinidin proanthocyanidins from *Acacia caffra*. *Phytochemistry*. 2000;53(7):785-93.
140. Bennie L, Coetzee J, Malan E, Ferreira D. Structure and stereochemistry of triflavanoids containing both ether and carbon-carbon interflavanyl bonds. *Phytochemistry*. 2001; 57(6):1023-34.
141. Ferrari J, Terreaux C, Sahpaz S, Msonthi JD, Wolfender J-L, et al. Isolation and on-line LC/CD analysis of 3,8''-linked biflavonoids from *Gnidia involucrata*. *Helv Chim Acta*. 2003;86(8):2768-78.
142. Geiger H, Quinn C. Biflavonoids. In: Harborne JB, Mabry TJ, Mabry H, editors. *The Flavonoids*. London U.K.: Chapman and Hall; 1976. pp 692-742.
143. Bennie L, Coetzee J, Malan E, Das B, Ferreira D. (4-6)-Coupled proteracacinidins and promelacacinidins from *Acacia galpinii* and *Acacia caffra*. *Phytochemistry*. 2002;60(5):521-32.
144. Geiger H, Quinn C. Biflavonoids. In: Harborne JB, Mabry TJ, editors. *The Flavonoids: Advances in Research Since 1980*. London, UK: Chapman & Hall; 1988, pp.99-124.
145. Ferreira D, Slade D, Marais JPI. Bi-, Tri-, Tetra-, Penta-, and Hexaflavonoids. In: Andersen OM, Markham KR, editors. *Flavonoids: Chemistry, Biochemistry and Applications*. Boca Raton: CRC Press; 2006, pp. 1115-33.
146. Valant-Vetschera K, Wollenweber E. Flavones and Flavonols. In: Andersen OM, Markham KR, editors. *Flavonoids: Chemistry, Biochemistry and Applications*. Boca Raton: CRC Press; 2006.
147. Hegnauer R. *Chemotaxonomie der Pflanzen*. Basel: Birkha user Verlag; 1990.
148. Alston RE. Recent Advances of Phytochemistry. In: Marby TJ, editor. 1. New York: Appleton -Century-Crofts; 1968. pp.305-27.
149. Franz G, Grun M. Chemistry, Occurrence and Biosynthesis of C-Glycosyl Compounds in Plants. *Planta Med*. 1983;47:131-40.
150. Mandal S, Joshi PC, Mukherjee K, Chatterjee A. A new Xanthone glycoside from *Lasiosiphon eriocephalus* Dcne. *Indian J Chemistry*. 1994;33B(1):89-91.
151. Attoumbre J, Charlet S, Baltora-Rosset S, Hano C, Grandic SR, et al. High accumulation of dehydrodiconiferyl alcohol-4-(-D-glucoside) in free and immobilized *Linum usitatissimum* cell cultures. *Plant Cell Rep*. 2006;25:859-64.
152. Umezawa T. Lignans In: Higuchi T, editor. *Springer Series in Wood Science, Biochemistry and Molecular Biology of Wood*. Berlin: Springer-Verlag; 1997. pp. 181-194
153. Ayres DC, Loike JD. Lignans-chemical, biological, and clinical properties. Cambridge, UK: Cambridge University Press; 1990.
154. MacRae WD, Towers GHN. Biological activities of lignans. *Phytochemistry*. 1984;23(6):1207-20.
155. Umezawa T, Shimada M. Enantiomeric composition of (-)-pinoresinol, (+)-matairesinol and (+)-wikstromol isolated from *Wikstroemia sikokiana*. *Mokuzai Gakkaishi*. 1996;42:180-5.
156. Umezawa T, Shimada M. Syntheses of (±)-lariciresinols. *Mokuzai Gakkaishi*. 1994;40:231-5.
157. Umezawa T, Isohata T, Kuroda H, Higuchi T, Shimada M. Chiral HPLC and LC-MS analysis of several lignans. In: Kuwahara M, Shimada M, editors. *Biotechnology in pulp and paper industry*. Tokyo: Uni; 1992. pp 507-512.
158. Ullah N, Ahmed S, Muhammad P, Ahmed Z, Nawaz HR, et al. Coumarinolignoid glycoside from *Daphne oleoides*. *Phytochemistry*. 1999 5/10/;51(1):103-5.
159. Bhandari P, Pant P, Rastogi RP. Aquillochin, a coumarinolignan from *Aquilaria agallocha*. *Phytochemistry*. 1982;21(8):2147-9.
160. Ponizo M, Rodriguez FSVB. Some components of *Torvisco* roots. *Daphne gnidium* (Thymelaeaceae). *An Quim*. 1972;68:489-500.
161. Murray AW, Bradshaw RW. Leucoglycodrin, a glycoside from *Leucodendron adscendens*. *Tetrahedron Lett*. 1966;31:3773-7.
162. Glennie CW, Perold GW. Biogenesis of the C-glycoside leucodrin in *Leucadendron argenteum*. *Phytochemistry* 1980;19:1463-6.
163. Kruger PE, Perold GW. Conocarpin, a leucodrin-type metabolite of *Leucospermum conocarpodendron* (L.) Buek. *J Chem Soc, Perkin Trans (C)*. 1970:2127-33.
164. Perold GW, Hodgkinson AJ, Howard AS. Metabolites of Proteaceae. Part V. Reflexin and conocarpic acid from *Leucospermum reflexum* Buek ex Meisner, and the phenoldienone rearrangement of reflexin and conocarpin. *J Chem Soc Perkin 1*. 1972:2450-60.
165. Fuller TC, McClintock E. Angiosperms-Dicotyledons:Thymelaeaceae, Mezereum family. Smith AC, editor. Los Angeles: University of California Press; 1986. 254 p.
166. Bhandari P, Tandon S, Rastogi RP. Eriocide, a new coumarin glucoside from *Lasiosiphon eriocephalus*. *Phytochemistry*. 1980;19:1554-5.
167. Vealea DJH, Furmana KI, Oliver DW. South African traditional herbal medicines used during pregnancy and childbirth *Journal of Ethnopharmacology*. 1992;36:185-91.
168. Coetzer J, Pieterse MJ. The structure of 12-hydroxydaphnetoxin, a poisonous constituent of *Lasiosiphon burchellii*: an X-ray analysis of 12-hydroxydaphnetoxin tribromoacetate. *Acta Cryst*. 1972;B28:620-4.
169. Pieterse MJ. On the isolation of toxic constituents from *Lasiosiphon burchellii* Meisn. *Landbouwet S Afr Agric Sci S Afr Agrochemophys*. 1971;3(4):63-5.
170. Fujita E, Nagao Y. Tumor Inhibitors Having Potential for Interaction with Mercapto Enzymes and/or Coenzymes. *Bioorganic Chemistry*. 1977;6:287-309.

171. Brooks G, Evans AT, Aitken A, Evans FJ, Rizk A-FM, et al. Daphnane Diterpenes of *Thymelaea hirsuta*. *Phytochemistry*. 1990;29(7): 2235-7.
172. Bala AEA, Delorme R, Kollmann A, Kerhoas L, Einhorn J, et al. Insecticidal activity of daphnane diterpenes from *Lasiosiphon kraussianus* (Meisn.) (Thymelaeaceae) roots. *Pesticide Science*. 1999;55:745-50.
173. Bala A, Delorme R, Kollmann A, Kerhoas L, Einhorn J, et al., editors. Isolation and pesticide activity of daphnanes of *Lasiosiphon kraussianus*. Proceedings of ECSOC-4, The Fourth International Electronic Conference on Synthetic Organic Chemistry, <http://www.mdpi.org/ecsoc-4htm>, September 1-30, 2000; Versailles Cedex, France: MDPI, Basel, Switzerland
174. Bala A, Delorme R, Kollmann A, Kerhoas L, Einhorn J, et al., editors. Isolation and pesticide activity of daphnanes of *Lasiosiphon kraussianus*. Proceedings of ECSOC-4, The Fourth International Electronic Conference on Synthetic Organic Chemistry, <http://www.mdpi.org/ecsoc-4htm>, September 1-30, 2000; Versailles Cedex, France: MDPI, Basel, Switzerland
175. Vermeulen NMJ, Pieterse MJ. The isolation of umbelliferone and an active principle from *Arthosolen polycephalus*. The South African Chemical Institute. 1968;21:190-1.
176. Das SC, Sengupta S, Hen W. *Chem Ind*. 1973:792.
177. Sharon M, Gupta A. In vitro Culture of *Lasiosiphon eriocephalus* an Endangered Species. *Acad J Plant Sci*. 2009;2(2):92-6.
178. Heftmann E. *Chromatography*. New York: Reinhold; 1964, p. 592.
179. Viswanathan N, Joshi BS. Toxic constituents of some Indian plants. *Current Sciences*. 1983;52(1):1-8.
180. Shen MS, Bryan RF. Gnidicoumarin. *Acta Crystallographica*. 1975;31B:2907-9. (Chemical Abstracts, 84,68162u).
181. Shen MS, Bryan RF. Gnidicoumarin. *Acta Crystallographica*. 1975;31B:2907-9. (Chemical Abstracts, 84,68162u).
182. Bhandari P, Tandon S, Rastogi RP. Chemical constituents of *Lasiosiphon enriocephalus*. *Planta Medica*. 1981;41:407-10.
183. Peng J, Fan G, Hong Z, Chai Y, Wu Y. Preparative separation of isovitexin and isoorientin from *Patrinia villosa* Juss by high-speed counter-current chromatography. *Journal of Chromatography A*. 2005;1074 111-5.
184. Ragot J, Turbery P, Carreras-Jansou M, Lattes A, Symonds P. Isolement de la 5-primeverosyl genkwanine des recines de *Daphne gnidium*. *Fitoterapia* 1988;59:336-7.
185. Zhang S-Y, Zhang Q-H, Zhao W, Zhang X, Zhang Q, et al. Isolation, characterization and cytotoxic activity of benzophenone glucopyranosides from *Mahkota Dewa* (*Phaleria macrocarpa* (Scheff.) Boerl). *Bioorganic & Medicinal Chemistry Letters*. 2012;22 6862-6.
186. Zhang YB, Xu XJ, Liu HM. Chemical constituents from *Mahkota dewa*. *J Asian Nat Prod Res*. 2006 8(1-2):119-23.
187. Umezawa T. Phylogenetic Distribution of Lignan Producing Plant. *Wood research: bulletin of the Wood Research Institute Kyoto University*. 2003;90:27-110.
188. Kupchan SM, Baxter YSRL, Hynes HR. Gnididione, a new furanosesquiterpene from *Gnidia latifolia*. *Journal of Organic Chemistry*. 1977;42(2):348-50.
189. Buttery CD, Cameron AG, Dell CP, Knight DW. A total synthesis of the guaiane furanosesquiterpene (\pm)-gnididione, a metabolite of *Gnidia latifolia*. *J Chem Soc, Perkin Trans* 1990;1(6):1601-10.
190. Kiptoon JC, Mengers GM, Waiyaki PG. Haematological and Biochemical changes in cattle poisoned by *Gnidia latifolia* Syn *Lasiosiphon latifolius* (Thymelaeaceae). *Toxicology*. 1982;25 129-39.
191. Rogerson H. Chemical examination of the root of *Lasiosiphon meissnerianus*. *American Journal of Pharmacy*. 1911;83:49-56.
192. Dawalkar MP, Kharadi Z, Joshi GV. Studies in free amino acids, sugars and organic acids in the leaves of *Lasiosiphon eriocephalus*. *Naturwissenschaften*. 1958;45(22):547.
193. Brooks G, Evans FJ. Toxic Hazards of Plants from the Families Euphorbiaceae and Thymelaeaceae. Rizk AM, editor. Boca Raton: CRC Press; 1990; pp. 27-63.
194. Von Oettingen WF, editors. *Poisoning – A Guide to Clinical Diagnosis and Treatment*. 2nd ed. Philadelphia: W.B. Saunders Co; 1958, p. 322.
195. Watt JM, Breyer-Brandwijk MG. *The Medicinal and Poisonous Plants of Southern Africa*. Baltimore: William Wood and Co; 1933, pp. 124-7.
196. Watt J, Breyer-Brandwijk MG. *The medicinal and poisonous plants of southern and eastern Africa*. 2 ed. Edinburgh: E. & S. Livingstone Ltd; 1962, pp. 1022-6.
197. Githens TS. *Drug Plants of Africa*. Philadelphia: University of Pennsylvania Press; 1948.
198. Kokwaro JO. *Medicinal Plants of East Africa*. Nairobi: East African Literature Bureau; 1976, pp. 211-2.
199. Botha CJ, Penrith ML. Poisonous plants of veterinary and human importance in southern Africa. *Journal of Ethnopharmacology*. 2008;119 549-58.
200. Felhauer M, Hecker E. Screening of Thymelaeaceae species for irritant, cocarcinogenic and antineoplastic activity. *Planta Medica*. 1986;6:553-4.
201. Mitchell J, Rook A. *Botanical Dermatology: Plants and Plant Products Injurious to the Skin* Greengrass. Vancouver; 1979, pp. 672-6.
202. McGaw LJ, Lall N, Meyer JJM, Eloff JN. The potential of South African plants against Mycobacterium infections. *Journal of Ethnopharmacology*. 2008;119 482-500.
203. Terblanche M, Pieterse MJ, Adelaar TF, Smit JD. Further studies of the toxicology of *Lasiosiphon burchellii* Meisn. (Harpuisbos). *Journal of the South African Veterinary Medical Association* 1966;37:185-9.
204. Hutchings A, Staden Jv. Plants used for stress-related ailments in traditional Zulu, Xhosa and Sotho medicine. Part 1: Plants used for headaches. *Journal of Ethnopharmacology*. 43:89-124.
205. El-Kheir YM, El-Tohami MS. Investigation of moliuscidal activity of certain Sudanese plants used in folk-medicine. III. Macro- and micro-morphology of the leaf, stem, and root of *Gnidia krausstina* Meisn. *Journal of African Medicinal Plants*. 1980;3:57 - 71.
206. Gelfand M, Mavi S, Drummond RB, Ndemera B. *The Traditional Medical Practitioner in Zimbabwe*. Gweru: Mambo Press; 1985, pp. 190-193.
207. Dold AP, Cocks ML. Traditional veterinary medicine in the Alice district of the Eastern Cape Province, South Africa. *South African Journal of Science*. 2001;97:375-9.
208. McGaw LJ, Eloff JN. Ethnoveterinary use of southern African plants and scientific evaluation of their medicinal properties. *Journal of Ethnopharmacology*. 2008;119 559-74.
209. Philander LA. An ethnobotany of Western Cape Rasta bush medicine. *Journal of Ethnopharmacology*. 2011;138:578-94.
210. Hutchings A, Terblanche SE. Observations on the use of some known and suspected toxic Liliiflorae in Zulu and Xhosa medicine. *South African Medical Journal* 1989;75 (2):62-9.
211. Shone DK. *Poisonous Plants of Rhodesia: I. Rhodesia Zambia Malawi*. *Journal of Agricultural Research*. 1966;4:81-94.
212. Pammel LH. *Manual of Poisonous Plants*. Cedar Rapids: The Torch Press; 1911, pp. 642-643.
213. Rasoanaivo P, Petitjean A, Ratsimamanga-Urverg S, Rakoto-Ratsimamanga A. Medicinal plants used to treat malaria in Madagascar. *Journal of Ethnopharmacology*. 1992;37 117-27.

214. Peterson B. Contributions to the knowledge of the flora of Southern Rhodesia. X. Thymelaeaceae. *Botaniska Notiser*. 1958;111:623-31 (Biological Abstracts, 35,8439).
215. Guillardmod AJ. Flora of Lesotho: Cramer. Lehr; 1971, 474. pp.
216. Arnold HL. Poisonous Plants of Hawaii, Rutland. Vermont: Charles E. Tuttle Co; 1968, pp. 58-59.
217. Berhan A, Asfaw Z, Kelbessa E. Ethnobotany of plants used as insecticides, repellents and antimalarial agents in Jabitehnan district, West Gojjam. *Ethiopian Journal of Science* 2006;29(1):87-92.
218. Amusan OG, Sukati NA, Dlamini PS, Sibandze FG. Some Swazi phytomedicines and their constituents. *Afr J Biotech*. 2007;6(3):267-72.
219. Amusan OOG, Dlamini PS, Msonthi JD, Makhubu LP. Some herbal remedies from Manzini region of Swaziland. *Journal of Ethnopharmacology*. 2002;79 109-12.
220. Lewis WH, Elvin-Lewis MPF. *Medical Botany*. New York: John Wiley and Sons; 1977 pp. 20,39,80,121,253,279,323.
221. Hutchinson J, Dalziel JM. *The Useful Plants of West Tropical Africa*. London: Crown Agents for the Colonies; 1948, pp. 42-3.
222. Neuwinger HD. *Plants used for poison fishing in tropical Africa*. *Toxicon*. 2004;44:417-30.
223. Smet PAGMD. Traditional pharmacology and medicine in Africa Ethnopharmacological themes in sub-Saharan art objects and utensils. *Journal of Ethnopharmacology*. 1998;63:1-179.
224. Wild H, Gelfand M. Some native herbal remedies at present in use in Mashonaland. *Central African Journal of Medicine*. 1959;5:292-305.
225. Offiah NV, Makama S, Elisha IL, Makoshi MS, Gotep JG, et al. Ethnobotanical survey of medicinal plants used in the treatment of animal diarrhoea in Plateau State, Nigeria. *BMC Veterinary Research*. 2011;7:36.
226. Gerstner J. Preliminary checklist of Zulu names of plants with short notes. *Banru Studies*. 1941;15:369-83.
227. Hutchings A. A survey and analysis of traditional medicinal plants as used by the Zulu, Xhosa and Sotho. *Bothalia* 1989;19 (1):111-23.
228. Tubery P, inventors Alcohol extract of *Lasiosiphon kraussianus* plant having lymphocytic activity, for use against leprosy. Patent, Fr.M. 6366 (Chemical Abstracts, 74,91170x).1968.
229. Tubery P, inventors Glycoside extract of *Lasiosiphon kraussianus* useful as an anti-leprosy medicament. Patent, Fr.M. 7333. (Chemical Abstracts, 75, 132993331969.
230. Ndhkala AR, Finnie JF, Van-Staden J. Plant composition, pharmacological properties and mutagenic evaluation of a commercial Zulu herbal mixture: *Imbiza ephuzwato*. *Journal of Ethnopharmacology*. 2011;133:663-74.
231. Abbiw D. *Useful Plants of Ghana*. London: Intermediate Technology; 1990 pp. 218-220.
232. Bulldugh CHW, Leary WP. Herbal medicines used by traditional birth attendants in Malawi. *Tropical and Geographical Medicine*. 1982;34:81-5.
233. Van-Wyk B-E, deWet H, Van-Heerden FR. An ethnobotanical survey of medicinal plants in the southeastern Karoo, South Africa. *South African Journal of Botany*. 2008; 74:696-704.
234. Hedberg I, Staugard F. *Traditional Medicinal Plants—Traditional Medicine in Botswana*. Gaborone, Botswana: Ipeleng Publishers; 1989.
235. Mothana RAA, Lindequist U. Antimicrobial activity of some medicinal plants of the island Soqotra. *Journal of Ethnopharmacology*. 2005; 96:177-81.
236. Assefa A, Urga K, Guta M, Mekonene W, Melaku D, et al. In vivo antimalarial activities of plants used in Ethiopian traditional medicine, Delomenna, Southeast Ethiopia. *Ethiopia J Health Sci*. 2007;17(2).
237. Peterson B. Contributions to the knowledge of the flora of Southern Rhodesia. K. Thymelaeaceae. *Botaniska Notiser*. 1958;111:623-31. (Biological Abstracts, 35, 8439).
238. Alexander R. *Lasiosiphon anthylloides* as a poisonous plant. 13th and 14th Reports of the Director of Veterinary Education and Research - Part 1, October 1928. Pretoria : Government Printer and Stationery Office, 1928, pp. 231-239.
239. Penrith ML, Van-Vollenhoven E. Pulmonary and hepatic lesions associated with suspected ganskweek (*Lasiospermum bipinnatum*) poisoning of cattle. *Journal of the South African Veterinary Association*. 1994;65:122-4.
240. Uphof JCT. *Dictionary of Economic Plants*. Winheim: H.R. Engelmann; 1959.
241. Oliver B. *Medicinal Plants in Nigeria*. Ibadan: Nigerian College of Arts, Science and Technology; 1960. pp. 29,69.
242. Tanaka K, Tachi I. Active charcoal. II. Preparation of active carbon by steam process and in a furnace with admission of air Wood Research, Kyoto,12, I-8.1954.
243. Houghton PJ, Osibogun IM. Flowering plants used against snakebite. *Journal of Ethnopharmacology*. 1993;39 1-29.
244. Gathercoal EN, Wirth EH. *Pharmacognosy*. Philadelphia, PA: Lea and Febiger; 1936, pp. 496-497.
245. Jager AK, Hutchings A, Staden Jv. Screening of Zulu medicinal plants for prostaglandin-synthesis inhibitors. *Journal of Ethnopharmacology*. 1996;52:95-100.
246. McGaw LJ, Van-der-Merwe D, Eloff JN. In vitro anthelmintic, antibacterial and cytotoxic effects of extracts from plants used in South African ethnoveterinary medicine. *The Veterinary Journal*. 2007;173:366-72.
247. Aremu AO, Finnie JF, Van-Staden J. Potential of South African medicinal plants used as anthelmintics – Their efficacy, safety concerns and reappraisal of current screening methods. *South African Journal of Botany*. 2012.
248. Clarkson C, Maharaj VJ, Crouch NR, Grace OM, Pillay P, et al. In vitro antiplasmodial activity of medicinal plants native to or naturalised in South Africa. *Journal of Ethnopharmacology*. 2004;92 177-91.
249. Barbosa-Filho JM, do-Nascimento-Júnior FA, de-Andrade-Tomaz AC, de-Athayde-Filho PF, da-Silva MS, et al. Natural products with antileprotic activity. *Brazilian Journal of Pharmacognosy*. 2007;17(1):141-8.
250. Mulungu LS, Ndilahomba B, Nyange CJ, Mwatawala MW, Mwalilino JK, et al. Efficacy of *Chrysanthemum cinerariaefolium*, *Neorautanenia mitis* and *Gnidia kraussiana* against Larger Grain Borer (*Prostephanus truncatus* Horn) and Maize Weevil (*Sitophilus zeamays* Motschulsky) on Maize (*Zea mays* L.) Grain Seeds. *Journal of Entomology*. 2011;8:81-7. Epub July 17, 2010.
251. Chimbe CM, Galley DJ. Evaluation of material from plants of medicinal importance in Malawi as protectants of stored grain against insects. *Crop Protection*. 1996;13(3):289-94.
252. Ebong OO, Nwude N. Toxicity of Methanol Extract of *Lasiosiphon kraussianus* Root. *Planta Med*. 1981;41(3): 267-73.
253. El-Kheir YM, El-Tohami MS. Investigation of molluscicidal activity of certain Sudanese plants used in folk-medicine. II. Molluscicidal activity of the different morphological organs of *Gnidia kraussiana* Meisn "abu gotna" family Thymelaeaceae. *J Trop Med Hyg*. 1979 82(11-12):242-7.
254. Puyelde LV, Geysen D, Ayobangira F-X, Nakizamungu E, Nswimiyimana A, et al. Screening of medicinal plants of Rwanda for Acaricidal activity. *Journal of Ethnopharmacology*. 1985;13:209-15.
255. Nigatu TA, Afework M, Urga K. Effects of *Gnidia stenophylla* Gilg Root Extract in Swiss Albino Mice: Hematological, Biochemical and Histopathological Effects of *Gnidia stenophylla* Gilg Root

- Extract in Swiss Albino Mice: LAP LAMBERT Academic Publishing; 2012.
256. Ibrahim N, Messay G, Berhanu T, Kissi M, Frehiwot T. Anti-Oxidant activity of 80% methanol extracts from *Clerodendron myricoides*, *Satureja punctata*, *Urtica dioica*, *Ajuga remota* and *Gnidia stenophylla*. *Revista CENIC Ciencias Biológicas*. 2010;41:1-7.
257. Alexander R. *Lasiosiphon anthyloides* as a poisonous plant. 1928, pp.231-40.
258. Behl PN, Captain RM, Bedi BMS, Gupta S. Skin-Irritant and Sensitizing Plants Found in India. Behl PN, editor: Department of Dermatology, Irwin Hospital & M.A. Medical College; 1966 pp 149.
259. Anonymus. Larvicidal activity of *Lasiosiphon eriocephalus*: Natural Sciences Repository 2012. Available from: http://triscience.com/Plant/larvicidal-activity-of-lasiosiphon-eriocephalus/doculite_view.
260. Brink M. *Gnidia glauca* (Fresen.) Gilg Wageningen, Netherlands: PROTA; 2009. Available from: http://database.prota.org/PROTAhtml/Gnidia%20glauca_En.htm.
261. Amarajeewa BWRC, Kumar V, editors. Chemistry and mosquito larvicidal activity of *G. glauca*. *Proceedings of the Peradeniya University Research Sessions*; 2007; 12(1): 101-2; Sri Lanka.
262. Dama GY, Bidkar JS, Deore SR, Tare HL. Ethnopharmacognostical survey of the plants available in the Western ghats of India. *Deccan J Natural Products*. 2010;1(3).
263. Ghosh S, Ahire M, Patil S, Jabgunde A, Dusane MB, et al. Antidiabetic Activity of *Gnidia glauca* and *Dioscorea bulbifera*: Potent Amylase and Glucosidase Inhibitors. Evidence-Based Complementary and Alternative Medicine. 2012;1-10.
264. Kareru PG, Kenji GM, Gachanja AN, Keriko JM, Mungai G. Traditional medicines among the Embu and Mbeere peoples of Kenya. *African Journal of Traditional, Complementary and Alternative Medicines*. 2007;4(1):75-86.
265. Teklehaymanot T, Giday M. Ethnobotanical study of medicinal plants used by people in Zegie Peninsula, Northwestern Ethiopia. *Journal of Ethnobiology and Ethnomedicine*. 2007;3(12).
266. Krishnan-Nambiar VP, Sasidharan N, Renuka C, Balagopalan M. Studies on the medicinal plants of Kerla forests. Peechi, Thissur: Kerala Forest Research Institute, 1985.
267. Joshi PC, Mandal S, Das PC, Adhikari P. Preliminary studies (In-vitro) on anticoagulant activity of naturally occurring bis-coumarins , ,(1),. *Aryavaidyan*. 2004;18(1):33-6.
268. Nethravathi HR, Kekuda TRP, Vinayaka KS, Thippeswamy NB, Sudharshan SJ, et al. Studies on antioxidant and anthelmintic activity of *Gnidia glauca* (Fresen) Gilg. *Asian Journal of Bio Science*. 2010 5(1):6-9.
269. Alagesan K, Kumar V, editors. Mosquito Larvicida; constituents of *Piper sylvestre* and *Gnidia glauca*. Symposium on Bioactive Natural Products November 11-15,1996 Kandy, Sri Lanka: Department of Chemistry, University of Peradeniya.
270. Ohigashi H, Takagaki T, Koshimizu K, Watanabe K, Kaji M, et al. Biological activities of plant extracts from Tropical Africa. *African Study Monographs*. 1991;12(4):201-10.
271. Kharde MN, Wabale AS, Adhav RM, Jadhav BD, Wabale AM, et al. Effect of Plant extracts on the Fungal Pathogen Causing Leaf Blight of Tomato in in vitro. *Asian JExpBiol Sci Spl*. 2010:121-3.
272. Javaregowda K, Naik LK. Ovicidal properties of plant extracts against the eggs of teak defoliator, *Hyblaea puera cramer karnataka*. *Journal of Agricultural Science*. 2007;20(2):291-3.
273. Dhar ML, Dhar MM, Dhawan BN, Mehrotra BN, Ray C. Screening of Indian plants for biological activity, Part I. *Indian Journal of Experimental Biology*. 1968;6:232-47.
274. Ghosh S, Patil S, Ahire M, Kitture R, Gurav DD, et al. *Gnidia glauca* flower extract mediated synthesis of gold nanoparticles and evaluation of its chemocatalytic potential. *Journal of Nanobiotechnology*. 2012;10(1):17.
275. Kumar S, Kumar V, Rana M, Kumar D. Enzymes Inhibitors From Plants: An Alternate Approach To Treat Diabetes. *Pharmacognosy Communications*. 2012 2(2):18-33.
276. Bonddupalli A, Kumanan R, Elimalai A, Chinna-Eswaraiah M, Veldi N, et al. A twelve-monthly review on Anti-diabetic plants: Jan-Dec-2011. *International Research journal of Pharmacy* 2012;3(1):77-80.
277. Gupta A, Oza G, Pandey S, Durve A, Bhardwaj S, et al. Antimicrobial Activity of *Lasiosiphon eriocephalus*. *Adv In Biol Res*. 2010;01.
278. Premaratne-Bandara KAN, Balasuriya CA, Kumar V, Thillainathan K, Weerasekera N, editors. Biopesticides from Srilankan plants. Symposium on Bioactive Natural Products November 11-15,1996 Kandy, Sri Lanka: Department of Chemistry, University of Peradeniya.
279. Khadke SS, Pachauri DR, Mahajan SD. An Acute Oral Toxicity Study of *Gnidia glauca* (Fresen.) Gilg. in Albino Rats as per OECD Guideline 425. *International Journal of PharmTech Research*. 2011;3(2):787-91.
280. Chopra RN, Nayar SL, Chopra IC. Glossary of Indian Medicinal Plants. New Delhi: Council of Scientific and Industrial Research; 1956, p. 150.
281. Kirtikar KR, Basu BD. *Indian Medicinal Plants*. 2nd ed. Allahabad, India: Lalit Mohan Basu; 1935;3:2167-71.
282. Brendre VV, Moholkar AL, Joglekar GV. Tooth loosening effect of *Lasiosiphon eriocephalus* (rametha). *J Res Indian Med*. 1972;7(2):76-8.
283. Gupta A, Oza G, Durve A, Sharon M. Bactericidal effect of crude extracts of an endangered plant: *Lasiosiphon eriocephalus* Decne *J Microbiol Biotech Res*. 2012;2 (6):866-70.
284. Chavan SR, Nikam ST. Larvicidal activity of *Lasiosiphon eriocephalus* (Dcne). *Bulletin of the Haffkine Institute*. 1982;10(2):42-4.
285. Javaregowda AD, B H. Management of Rice Gall Midge Using Botanicals and Insecticides Under Rainfed Ecosystem. *Annals of Plant Protection Sciences*. 2012;20(1):4-8.
286. Sundararajan G, Kumuthakalavalli R. Antifeedant activity of aqueous extract of *Gnidia glauca* Gilg. and *Toddalia asiatica* Lam. on the gram pod borer, *Helicoverpa armigera* (Hbn). *J Environ Biol*. 2001 22(1):11-4.
287. Prakash TN, Kaushik BAA. Twelve Harvests: Saga of an organic transformation. *Honey Bee*. 1996:5-6.
288. Prakash TN. Pest preventive measures. *Honey Bee*. 1997:12.
289. Subramanian TV. The insecticidal properties of indigenous vegetable fish poisons. *Journal of the Mysore Agricultural and Experimental Union*. 1932;13:57-60 (Chemical Abstracts, 28,6515).
290. Bussmann RW. Ethnobotany of the Samburu of Mt. Nyiru, South Turkana, Kenya. *Journal of Ethnobiology and Ethnomedicine*. 2006;2:35
291. Sanjeev KK, Sasidharan N. Ethnobotanical observations on the tribals of Chinnar Wildlife Sanctuary. *Ancient Science of Life*. 1997;14(4):1-8.
292. Nevling LI, Jr. A preliminary report on Funifera. *Journal of the Arnold Arboretum*. 1965;46:232-41.
293. Teklehaymanot T, Giday M. Ethnobotanical study of medicinal plants used by people in Zegie Peninsula, Northwestern

- Ethiopia. Journal of Ethnobiology and Ethnomedicine. 2007;3(12).
294. Robert D, Jacques E, Albert K, Ahmed BAE, Paul-Henri D, et al., inventors Composition pesticides, plus particulièrement insecticides, a base De Diterpenes De la Famille Des Daphnanes. 1998. Patent No. 1998024318.
295. Zhang Y, Zhang H, Hua S, Ma L, Liu X, et al. Identification of two herbal compounds with potential cholesterol-lowering activity. *biochemical pharmacology*. 2007;74:940-7.
296. Yesilada E, Taninaka H, Takaishi Y, Honda G, Sezik E, et al. In vitro inhibitory effects of *Daphne oleoides* ssp. *Oleoides* on inflammatory cytokines and activity-guided isolation of active constituents. *Cytokine*. 2001;13(6):359-64.
297. Brink M. *Gnidia latifolia* (Oliv.) Gilg Wageningen. 2009. Available from: http://database.prota.org/dbtw-wpd/exec/dbtwpub.dll?ac=qbe_query&bu=http://database.prota.org/recherche.htm&tn=protab~1&qb0=and&qf0=Species+Code&qf1=Gnidia+latifolia&rf=AfficherWeb.
298. Hall IH, Liou YF, Oswald CB, Lee KH. The effects of genkwadaphnin and gnidilatidin on the growth of P-388, L-1210 leukemia and KB carcinoma cells in vitro. *European Journal of Cancer Clinical Oncology*. 1986;22:45-52.
299. Park B-Y, Min B-S, Ahn K-S, Kwon O-K, Joung H, et al. Daphnane diterpene esters isolated from flower buds of *Daphne genkwa* induce apoptosis in human myelocytic HL-60 cells and suppress tumor growth in Lewis lung carcinoma (LLC)-inoculated mouse model. *Journal of Ethnopharmacology*. 2007; 111:496-503.
300. Khajja BS, Sharma M, Singh R, Mathur GK. Forensic Study of Indian Toxicological Plants as Botanical Weapon (BW): A Review. *J Environment Analytic Toxicol*. 2011;1(4):112.
301. Kraft C, Jennet-Siems K, Siems K, Jakupovic J, Mavi S, et al. In vitro antiplasmodial evaluation of medicinal plants from Zimbabwe. *Phytotherapy Research*. 2003;17:123-8.
302. Kasal R, Lees K-H, Huang H-C. Genkwadaphnin, a potent antileukemic diterpene from *Daphne genkwa*. *Phytochemistry*. 1981;20(2):2592-4.
303. Kajiya K, Ota M, inventors Lymphatic vessel stabilizer patent US 2011/0091584 A1. 2009.
304. Wiechowski-W, Lotos, 1908-56:61, (cited, from, et al. Phytochemical and pharmacological investigations on mangiferin. *Herba polonica*. 2009;55(1):126-39).
305. Singh SK, Sharma VK, Kumar Y, Kumar SS, Sinha SK. Phytochemical and pharmacological investigations on mangiferin. *Herba polonica*. 2009;55(1):126-39.
306. Jensen SR, Schripsema J. Chemotaxonomy and pharmacology of Gentianaceae. In: Struwe L, Albert V, editors.: Cambridge University Press; 2002. p. 573-631.
307. Ahmad A, Pandey S, Sarkar FH. Role of novel Nutraceuticals Garcinol, Plumbagin and Mangiferin in the prevention and therapy of human malignancies: Mechanisms of Anticancer activity. Sarkar FH, editor: Springer; 2012.
308. Yao Y, Cheng X, Wang L, Wang S, Ren G. A Determination of Potential α -Glucosidase Inhibitors from Azuki Beans (*Vigna angularis*) *International Journal of Molecular Science*. 2011;12:6445-51.
309. Delazae A, Gibbons S, Kosari AR, Nazemiyeh H, Modarresi M, et al. Flavone C-glycosides and Cucurbitacin glycosides from *Citrullus colocynthis*. *DARU Journal of Pharmaceutical Sciences*. 2006;14(3):109-14.
310. Aslan M, Orhan DD, Orhan N. Effect of *Gentiana olivieri* on experimental epilepsy models. *Pharmacogn Mag*. 2011 7(28):344-9.
311. Kùpeli E, Aslan M, Gürbüz I, Yesilada E. Evaluation of in vivo Biological Activity Profile of Isoorientin. *Z Naturforsch*. 2004;59c:787-90.
312. Zhang YB, Zhang PY, Dai GF, Liu HM. Study on the Synthesis and Bioactivity of Novel Mahkoside A Derivatives. *Synthetic Communications*. 2007 2007/09/01;37(19):3319-28.
313. Yeilada E, GGC3, Taninaka H, Takaishi Y, Honda G, Sezik E, et al. In vitro inhibitory effects of *Daphne oleoides* ssp. *Oleoides* on inflammatory cytokines and activity-guided isolation of active constituents. *Cytokine*. 2001;13(6):359-64
314. Wang CR, GGR301, Chen ZX, Ying BP, Zhou BN, Liu JS, et al. Studies on the active principles of the root of Yuanhua (*Daphne genkwa*) II. Isolation and structure of a new antifertile diterpene uuanhuadin. *Acta Chem Sin*. 1981;39:421-6.
315. Varga CA, GGR336, Veale DJH. Isihlabezo: Utilization patterns and potential health effects of pregnancy-related traditional herbal medicine. *Soc Sci Med*. 1997;44(7):911-24.
316. Ulubelen A, GGR37, Terem B, Tuzlaci E. Coumarins and flavonoids from *Daphne gnidioides*. *J Nat Prod*. 1986;49(4):693-4.
317. Steyn DG, GGR178. Recent investigations into the toxicity of known and unknown poisonous plants in the Union of South Africa. Onderstepoort: Report of the Director of Veterinary Services, 1931, pp. 707-27. (Chemical Abstracts, 27.5421).
318. Rindl M. Isolation of a glucoside from *Gnidia polycephala* (Januarie Bossie). *Transactions of the Royal Society of South Africa*. 1933;21:239-44.
319. Nethravathi HR, GGR241, Prashith-Kekuda TR, Vinayaka KS, Thippeswamy NB, Sudharshan SJ, et al. Phytochemical Screening And Larvicidal Efficacy Of Extracts Of *Gnidia Glauca* (Fresen) Gilg. *Natural Products*. 2009;5(4).
320. He W, GGR95, Cik M, Lesage A, Van-der-Liden I, De-Kimpe N, et al. Kirkinine, a new daphnane orthoester with potent neurotrophic activity from *Synaptolepis kirkii*. *J Nat Prod*. 2000;63:1185-7.
321. Brink M, GGC2, Dako EGA. *Gnidia latifolia* (Oliv.) Gilg. In: Brink, M. & Achigan-Dako, E.G. (Editors). *Prota 16: Fibres/Plantes à fibres*. 2009;PROTA, Wageningen, Netherlands.

Cite this article as:

Parixit Bhandurge, Rajarajeshwari N, S Ganapaty, Santosh Pattanshetti. The *Gnidia* genus: A review. *Asian Journal of Biomedical and Pharmaceutical Sciences*, 2013, 3: (19), 1-31.