



Barcham

The Tree Specialists

Ely, 10th July 2019

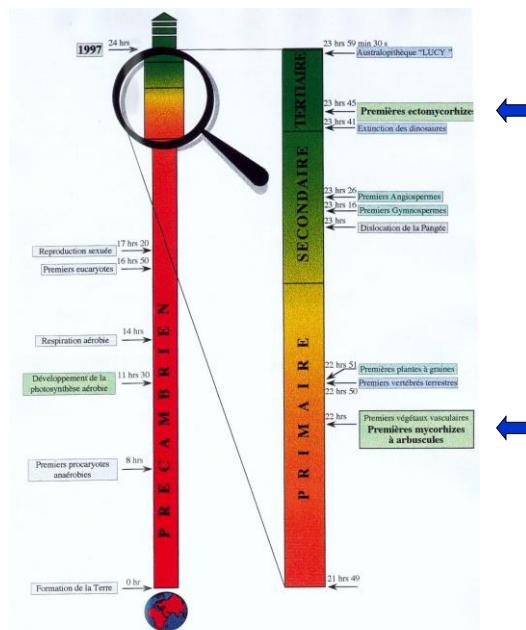
The Mycorrhizal World

Lucio Montecchio

De Rebus Plantarum, Spin-off of the University of Padova

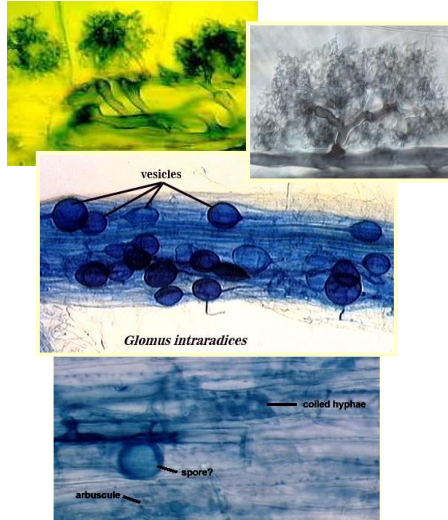
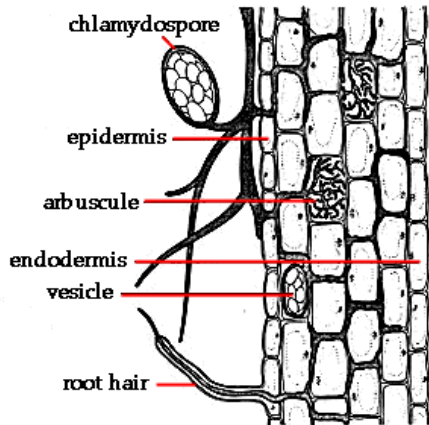


A very old cooperation



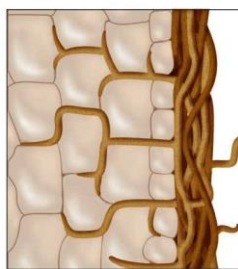
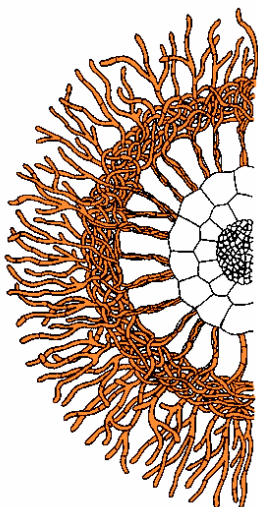
Endomycorrhizae

Endo = inside (the tip)



Ectomycorrhizae

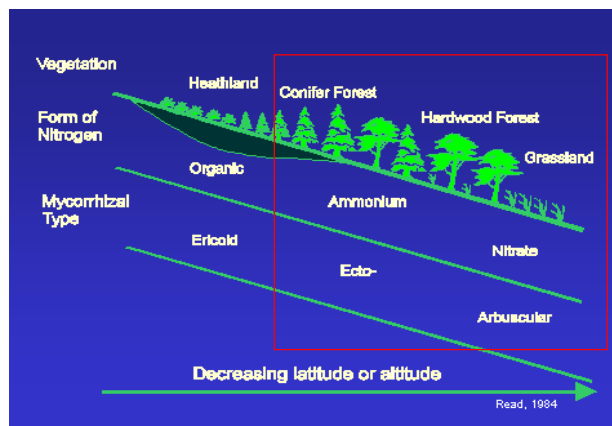
Ecto = outside /the tip)



Many mycorrhizal types, and sub-types

	Plant	Fungus
Arbuscular mycorr.	All taxa	Glomeromycetes
Ectomycorrhizae	Angiosperms, Gymnosperms	Basidiomycetes, Ascom.
Ectendomycorrhizae	<i>Pinaceae</i>	Basidiom., Ascom.
Arbutoids	Ericales	Basidiom.
Monotropoids	Monotropoids	Basidiom.
Ericoids	Ericales, Briophytes	Ascom.
Orchidoids	<i>Orchidaceae</i>	Basidiom.

In trees, mainly ectomycorrhizae and arbuscular mycorrhizae



Truffles: From ancient times to the first scientific studies

- **Pliny the Elder** (70 b.c.) thinks they are soil modifications.
- **Ray** (1700) observes spores inside truffles
- **De Borchii** (1780) demonstrates that spores produce mycelium
- **Vittorio Pico** (1787): truffles are different. First steps into mycorrhizal mycology.

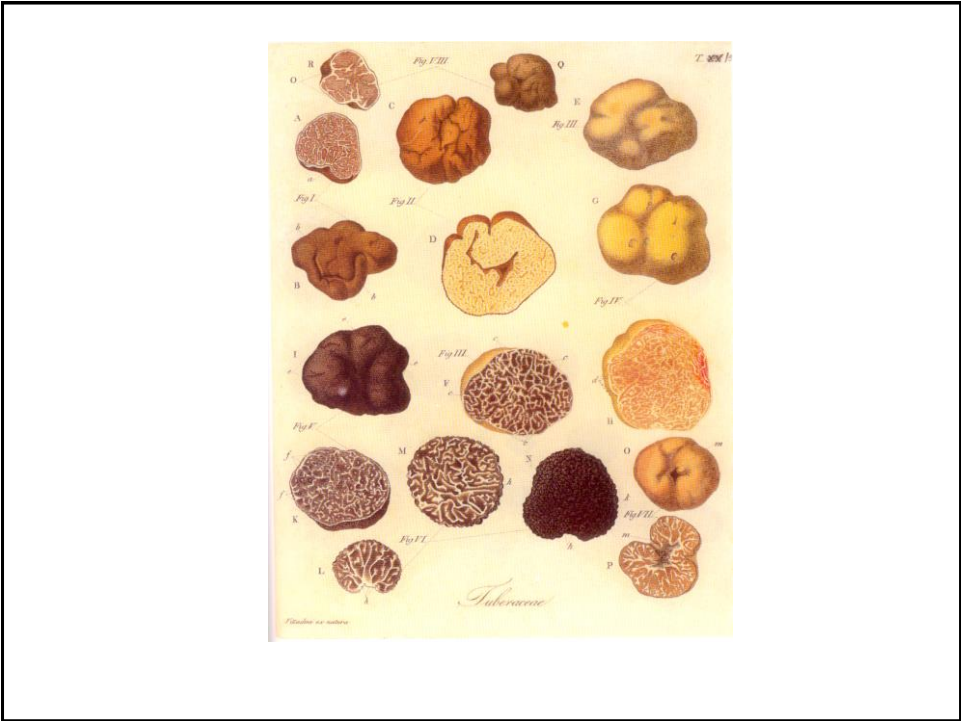
Carlo Vittadini



1831: «roots uptake nutrients from the fungal mantle».

He publishes the first modern descriptions of truffle species:

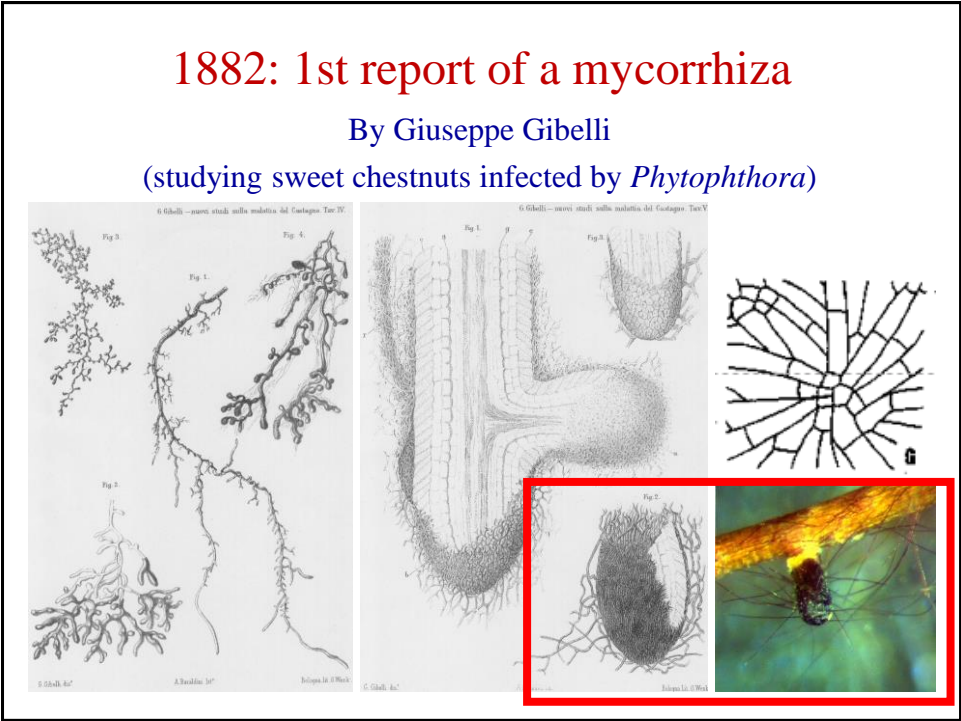
Tuber aestivum, borchii, brumale, excavatum, ferrugineum, foetidum, macrosporum, maculatum, melanosporum, mesentericum, nitidum, (microsporum, oligosporum).



1882: 1st report of a mycorrhiza

By Giuseppe Gibelli

(studying sweet chestnuts infected by *Phytophthora*)



1885: from *symbiosis* to *mutualism*



Albert Bernhard Frank:
«Mycorrhiza» = fungus + root tip):

The fungus uptakes water and salts from the soil and transfer them to the plant.

The plant gives the fungus root exudates.

From mutualism to symbiosis

2008: a behavioural *continuum* ...

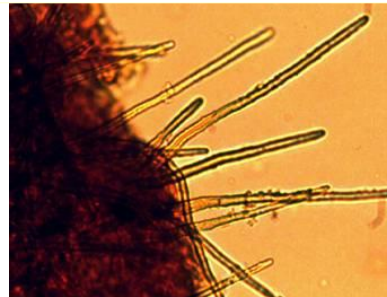
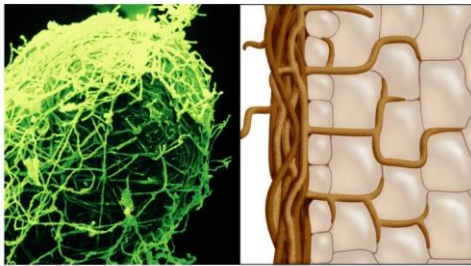
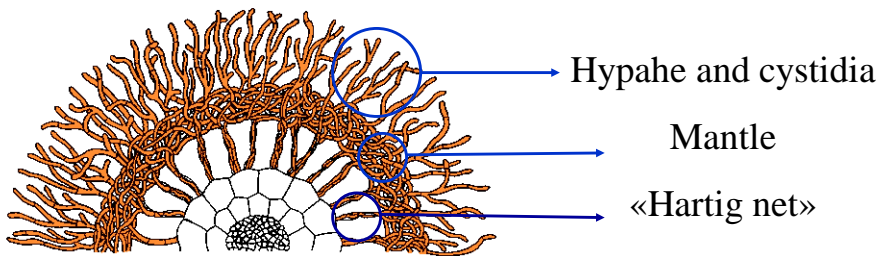
... sometimes to parasitism

Entoloma saepium/Rosa e Prunus spp. (Götsche, 1972; Agerer e Waller, 1993)

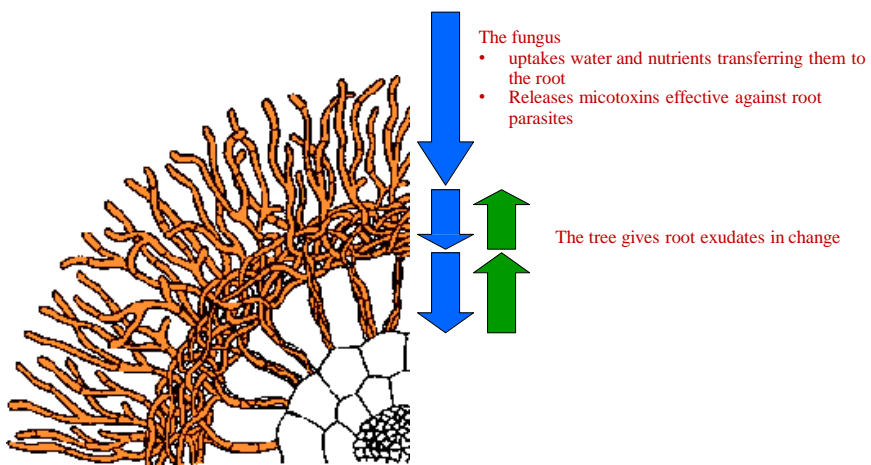
Hygrophorus penarius/faggio (Di Marino, Montecchio, Agerer, 2008)

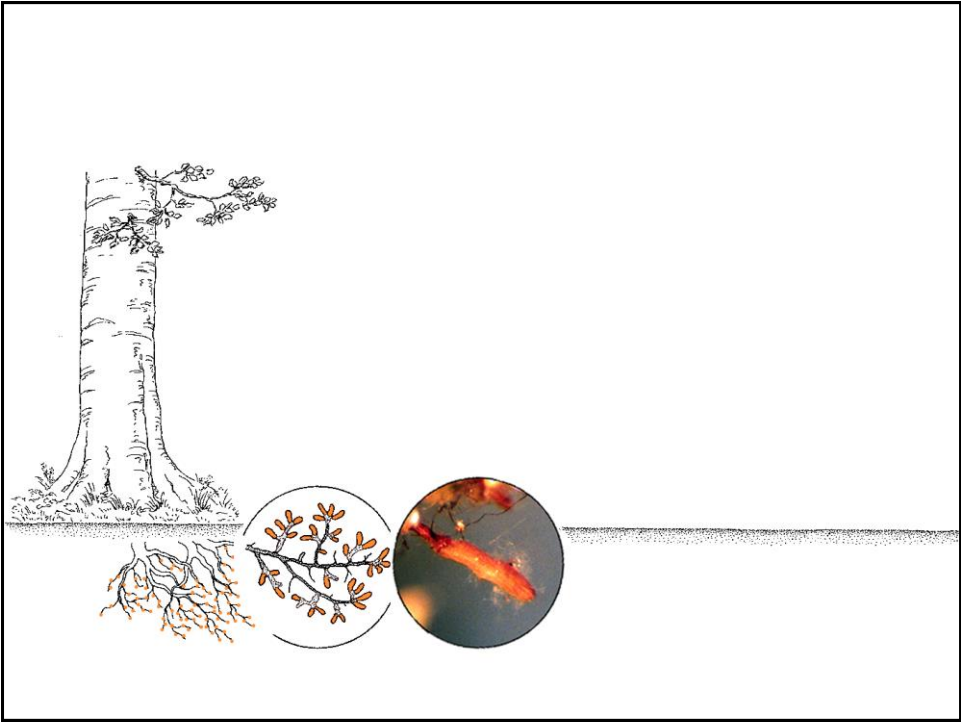
... likely related to species involved, age, genotypes, their
physiological status, environmental conditions.

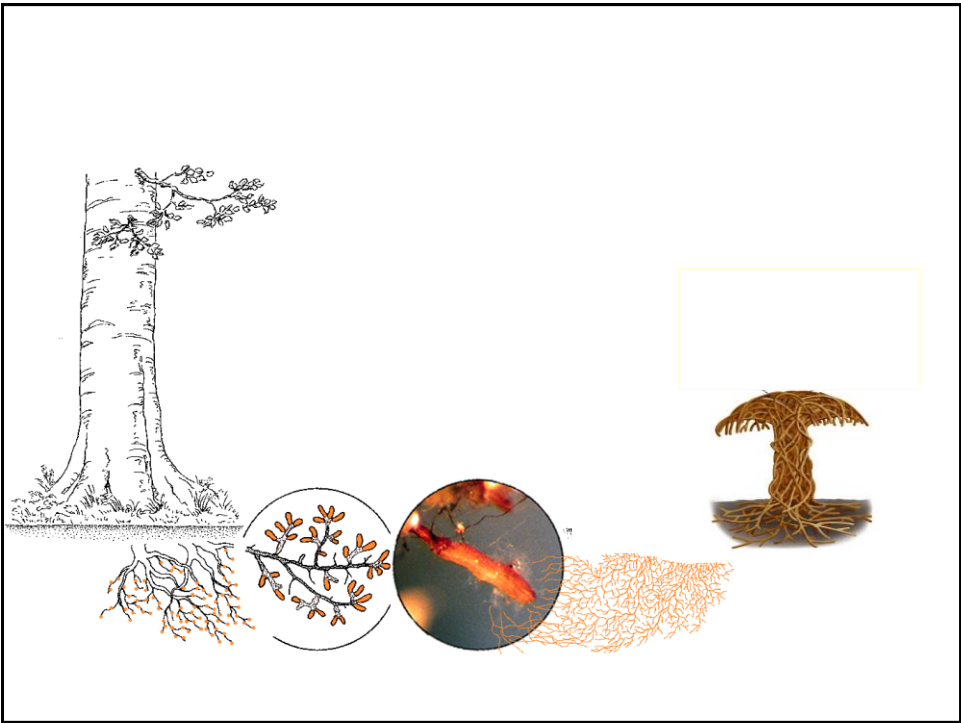
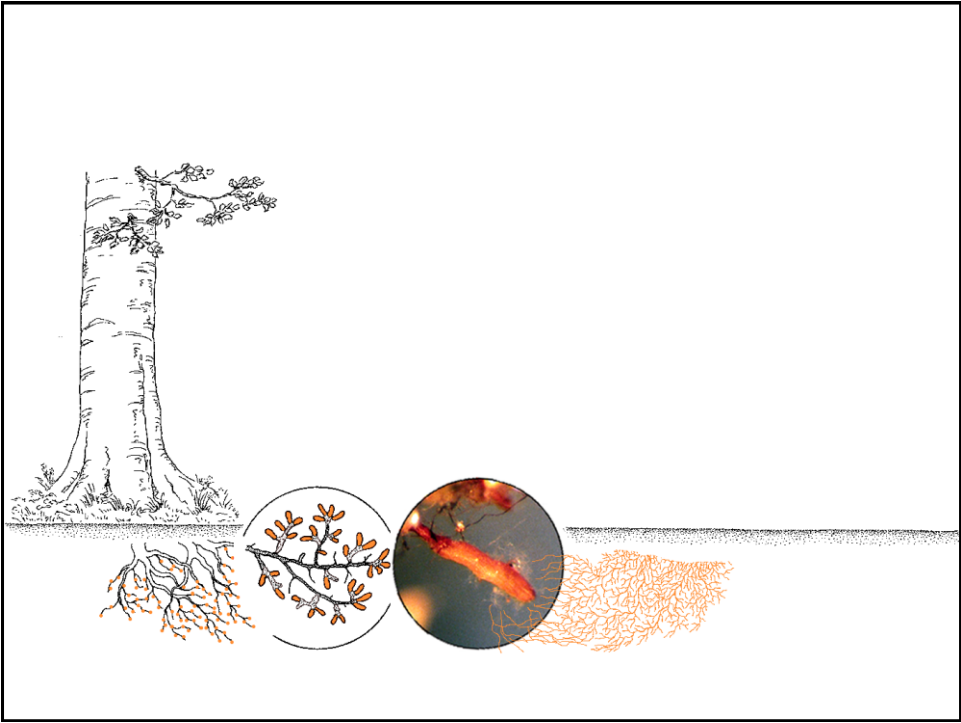
Anatomy of an ectomycorrhiza



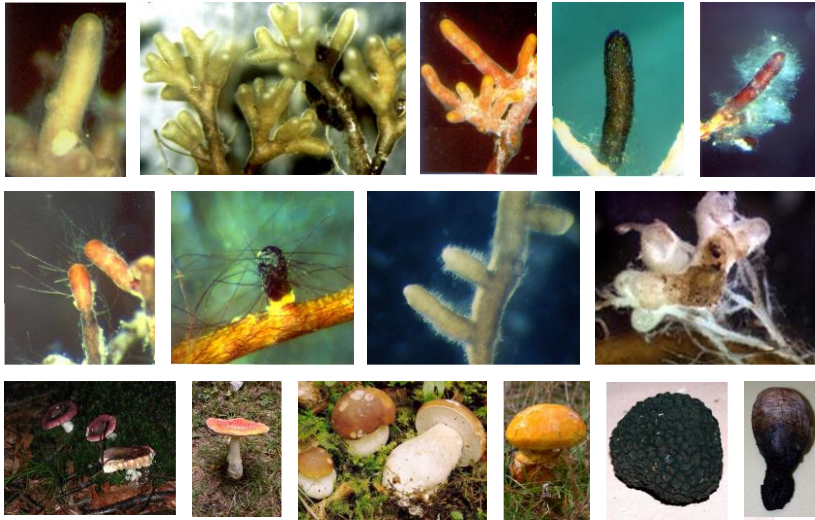
Ectomycorrhizae are
exactly **at the tree/soil interface.**







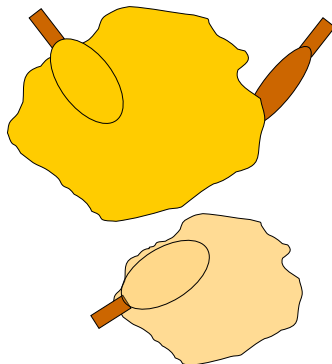
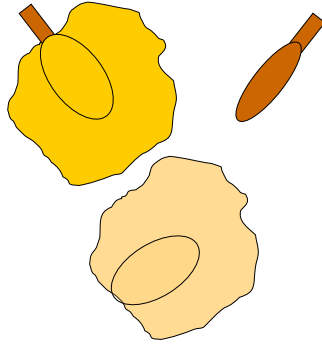
One tree, many fungi



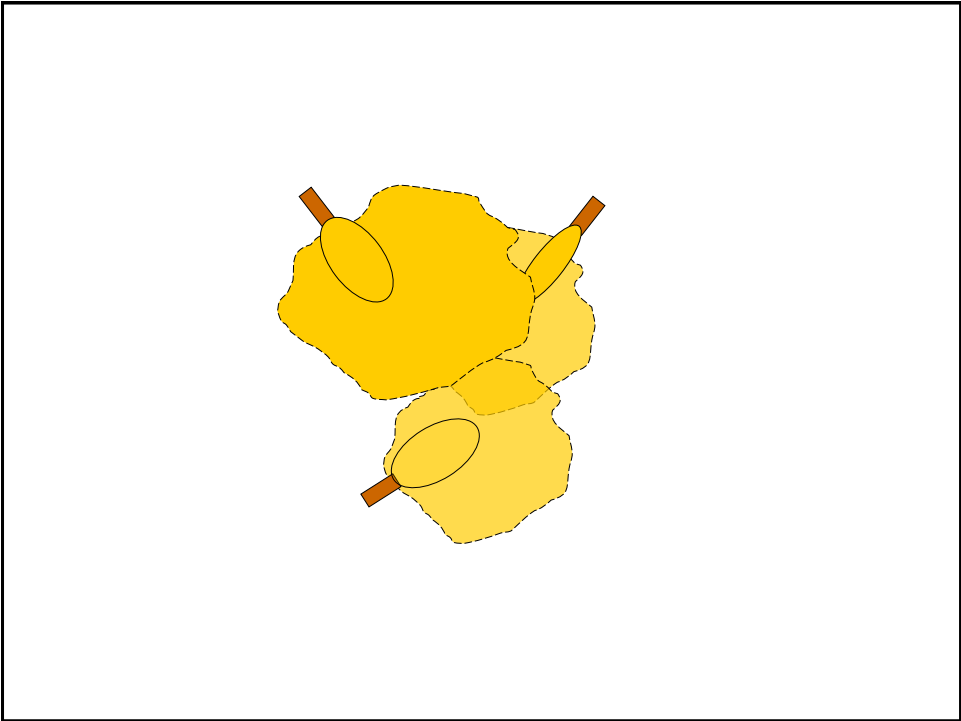
A dynamic composition driven by competition



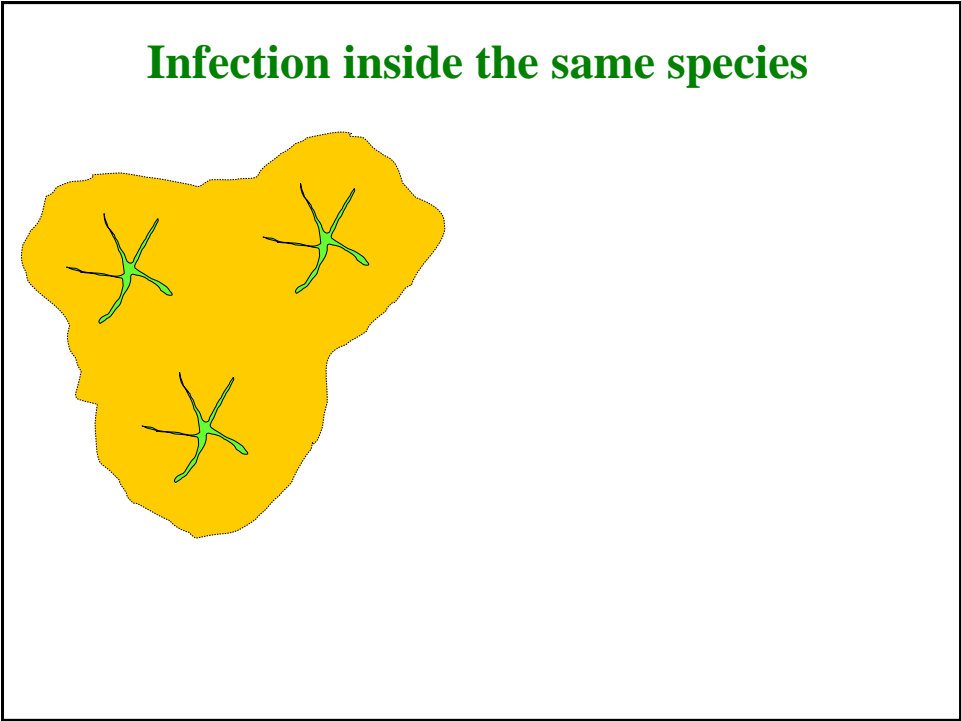
Competition and infection



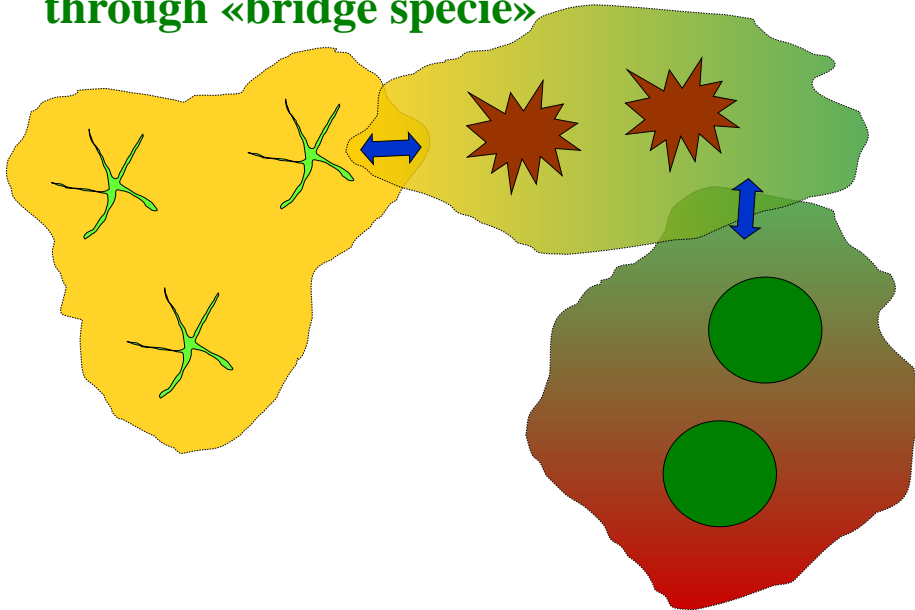
The best performing fungus spreads to the closest tips



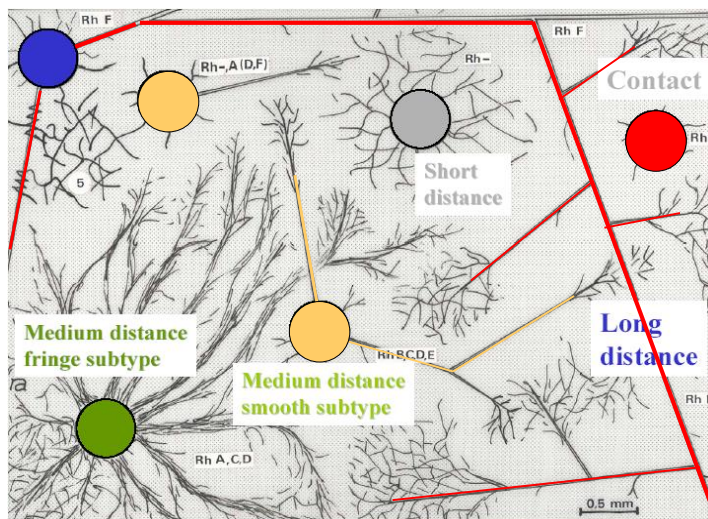
Infection inside the same species



Infection between different species, also through «bridge specie»



Exploration types: different species, similar benefits



Change with depth

(humus, pH, humidity, etc.)

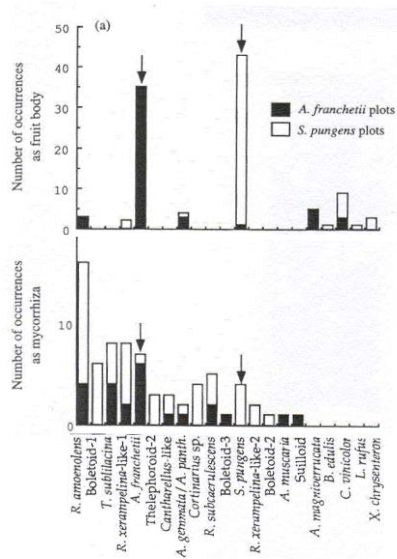
	O			E1			E2			EB		B1			B2			C		
	1	2	3	1	2	3	1	2	3	1	2	1	2	3	1	2	3	1	2	3
<i>Tylospora</i> sp.	I	I		I	*		I	*		I	I								III	
<i>Corinarius</i> sp.	I	I					IV	I		I	I	II	II		I	I	I	I	I	
<i>P. reticulatum</i>	III	IV		III	IV		II	II		I				I				III	I	
<i>Piloderma</i> sp. JS 15686														I				I		
<i>Inocybe</i>																				
<i>P. byssinum</i>		I																		
<i>T. submollis</i>	I	I		I																
<i>P. fallax</i>	I	I			I	I	I													
<i>H. olivaceoalbus</i>		I			I		I													
<i>R. decolorans</i>		IV			IV		I			*										
<i>Dermocybe</i> sp.	*	*		I			I	I		I										
<i>Tomentoloid</i>					*			I												
<i>L. utilis</i>							II	I	II		II									
<i>Piloderma</i> sp. 2							I	I	I	I	I	I	I		II					
<i>Piloderma</i> sp. 3										I	I			I						
<i>Piloderma</i> sp. 1											I	I	I		I					
<i>S. luteus</i>								I		III	III	II		IV	I			I		
Non ID nr. 15														I						
Non ID nr. 12															I					
<i>Wilcoxina</i>																		I		
<i>R. adusta</i>																		IV		
<i>T. portentosum</i>																		III		

Presenza dei diversi taxa negli orizzonti di un suolo podzolico in una foresta di conifere nel nord della Svezia gli orizzonti sono indicati dalle lettere O, E, B e C, sono riportati per ciascun orizzonte i dati ricavati da 3 campioni. La abbondanza di ciascuna specie è indicata attraverso i seguenti simboli: *=<1%; I=1-25%; II=26-50%; III=51-75%; IV=76-100%. (da Rosling e coll. 2003)

Consistenza del consorzio

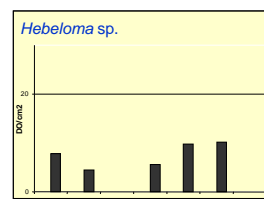
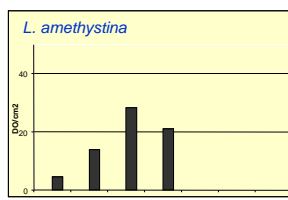
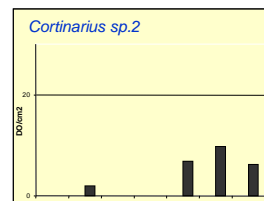
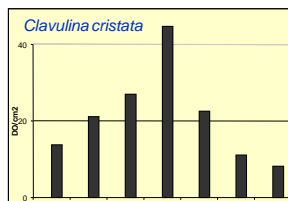
Monitoraggio degli sporocarpi:
imprecisa corrispondenza

Riconoscimento da campioni radicali:
Maggior accuratezza

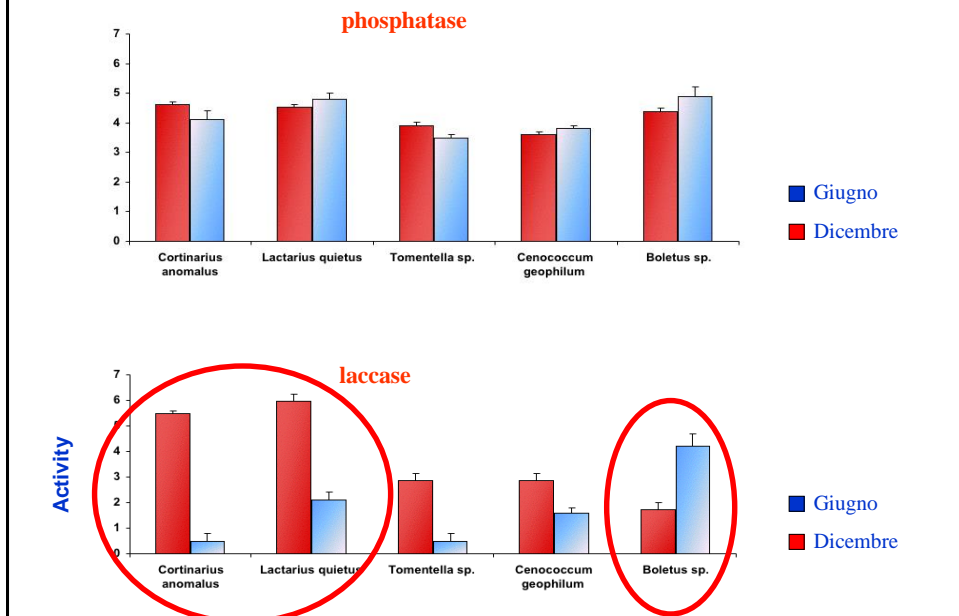


Change in time

Morfotipi	M	A	M	G	L	A	S	O	N	D
<i>Cortinarius anomalus</i>	✓	✓	✓	✓	✓			✓	✓	✓
<i>Lactarius quietus</i>	✓	✓	✓	✓	✓				✓	✓
<i>Byssocorticiium atrovirens</i>	✓	✓	✓	✓	✓					
<i>Cenococcum geophilum</i>				✓	✓			✓	✓	✓
<i>Tomentella sp.</i>				✓	✓			✓	✓	✓
<i>Piloderma sp.</i>				✓						
<i>Hebeloma sp.</i>										✓
<i>Laccaria amethystina</i>										✓
<i>Russula nigricans</i>										✓
<i>Clavulina cristata</i>	✓	✓	✓							✓
<i>Boletus sp.</i>		✓	✓	✓	✓					✓



Performances change with time, too.



How to assess a mycorrhizal community?

A sampling method to describe the Norway spruce ectomycorrhizal community at plant level

L. MONTECCHIO & L. SCATTOLIN

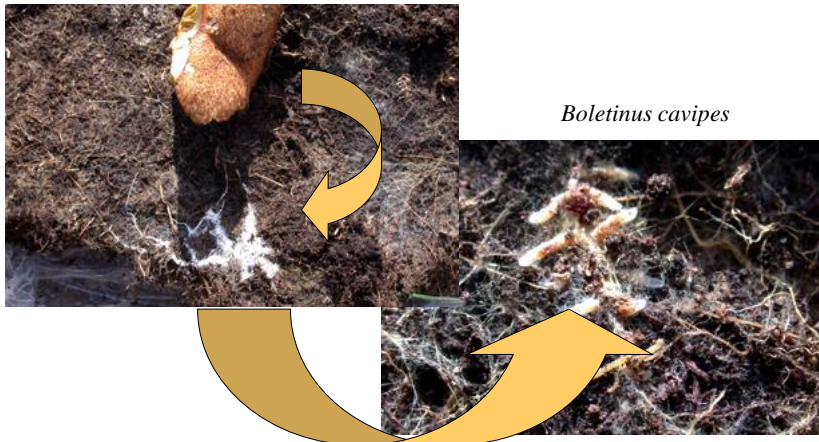
Dipartimento Territorio e Sistemi Agro-Forestali, Università degli Studi di Padova, Italy

of EM to be observed in every root core can be reduced from 10 to 4, 7, 5 and 9 in Ma, Pa, Ga and *pleres* in Fo, were shown to be more important, probably because of their distribution strategy.

Table VI. EM relative abundances (RA) in four trees in Pa site, considering 10 tips and 6 tips observed in each root core. In the right column: observed vs. expected RA, Chi-square = 29.64001, df = 1, $P < 0.001805$. In evidence: $(O - E)^2/E$ values > 1.63959 ($=8$).

	10 tips/root core (E)				6 tips/root core (O)				$(O - E)^2/E$
	Tree								
	1	2	3	4	1	2	3	4	
<i>A. muscaria</i>	10.706	16.972	2.777	8.164	11.944	14.930	5.000	7.777	0.018
<i>A. byssoides</i>	18.964	0.000	1.759	0.925	19.166	0.000	2.083	1.388	0.231
<i>C. geophilum</i>	54.108	41.382	35.300	34.943	56.041	33.750	42.777	52.847	9.172
<i>E. granulatus</i>	0.520	1.521	7.083	22.184	0.694	1.666	6.250	1.527	19.233
<i>H. velutipes</i>	0.000	6.967	21.180	3.042	0.000	2.777	5.555	3.472	0.060
<i>H. rufescens</i>	0.595	4.768	2.546	3.349	0.000	4.861	2.777	3.055	0.025
<i>H. olivaceoalbus</i>	0.000	0.000	2.546	4.117	0.000	0.000	2.777	5.000	0.189
<i>I. appendiculata</i>	3.645	1.388	3.842	0.000	3.125	2.083	3.472	0.000	0.000
<i>L. badiusanguineus</i>	1.041	2.979	3.888	5.456	0.694	4.375	2.777	4.444	0.187
<i>L. deterrimus</i>	0.520	0.925	2.546	0.000	0.694	1.388	1.388	0.000	0.000
<i>P. nigra</i>	4.166	6.539	6.388	2.232	2.500	12.152	10.833	2.083	0.009
<i>R. ochroleuca</i>	5.729	16.554	10.138	15.583	5.138	22.013	14.305	18.402	0.509

Only when a fruitbody is visible,
it's easy to detect its mycorrhiza

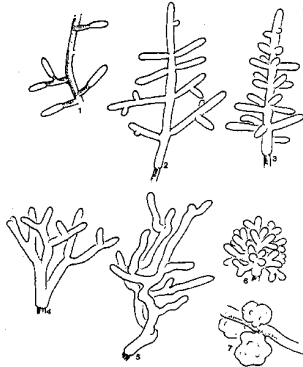


Usually, identification is not easy

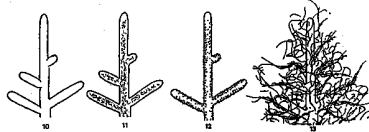
- Only about 500 species described worldwide in detail.
- Taxonomy needs skill and time, and money.



Mycorrhizal types

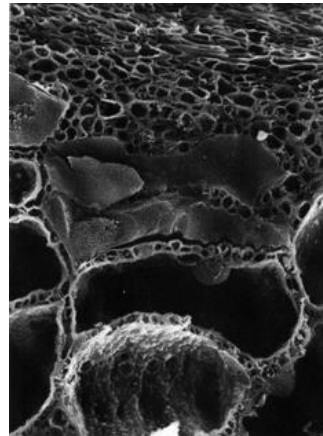
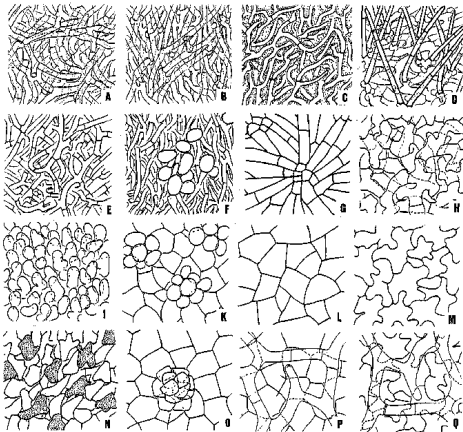


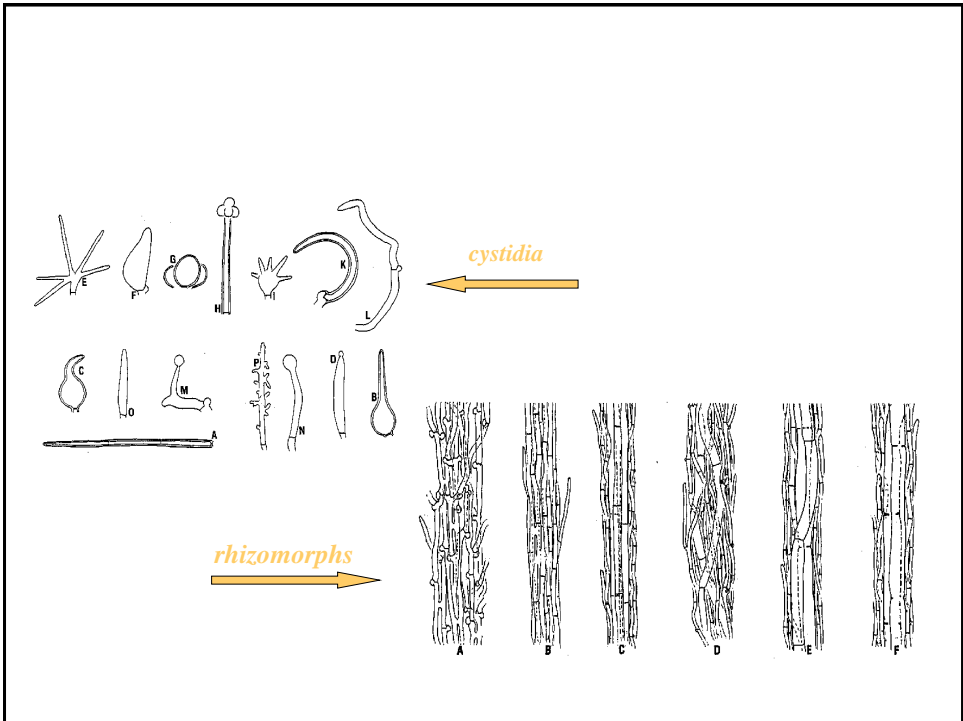
Figs. 1-7: Type of ramification - 1: Simple + unramified. - 2: Monopodial piram. - 3: Mesopodial-pyramidal. - 4: Dichotomous. - 5: Irregularly piram. dichotomous-like. 6: Coraloid. - 7: Tubercle-like.



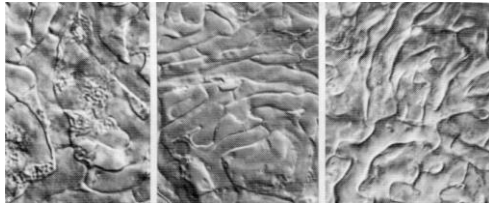
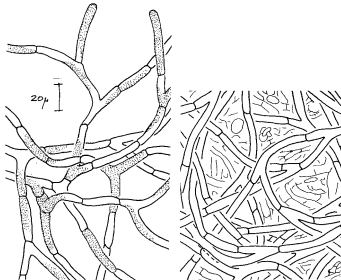
Figs. 10-13: Features of mantle surface. - 10: Smooth. - 11: Reticulate. - 12: Grainy (wart). - 13: Woolly.

Mantle and Hartig net

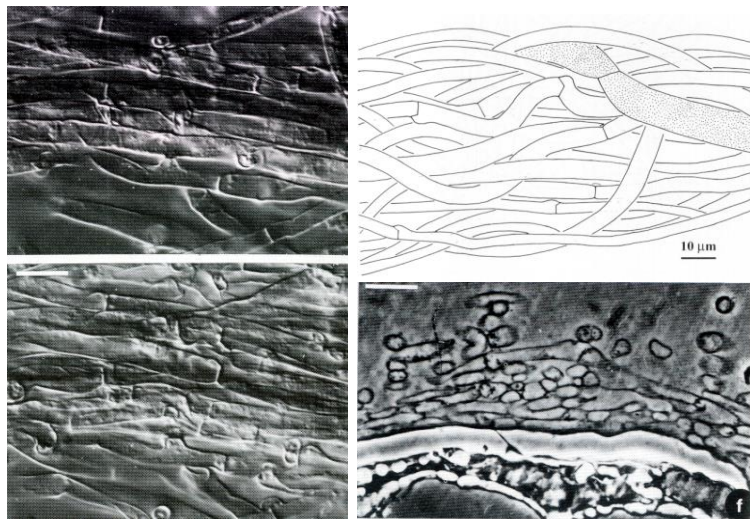
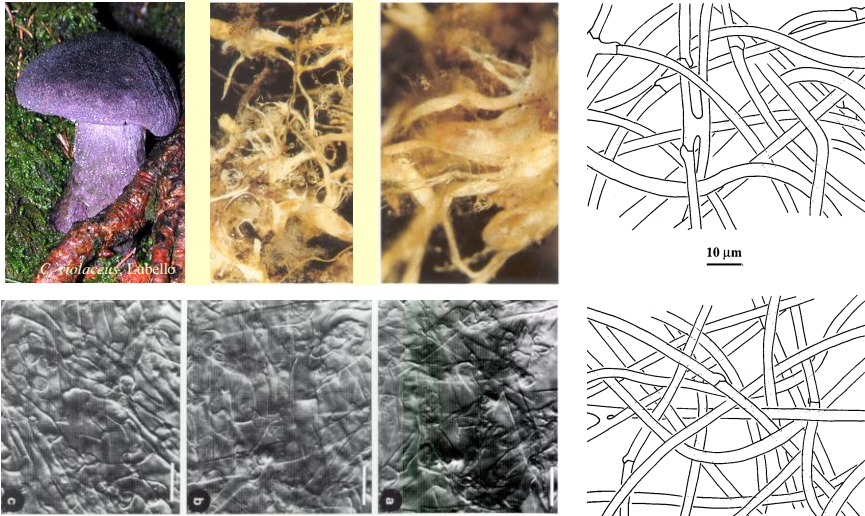




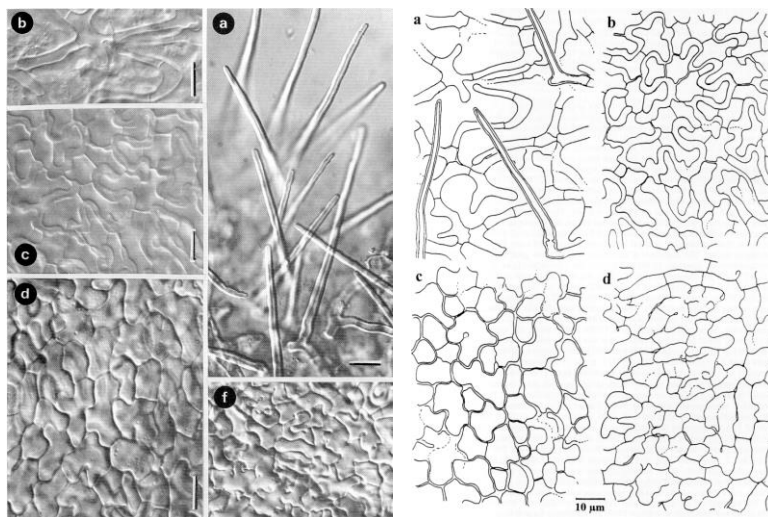
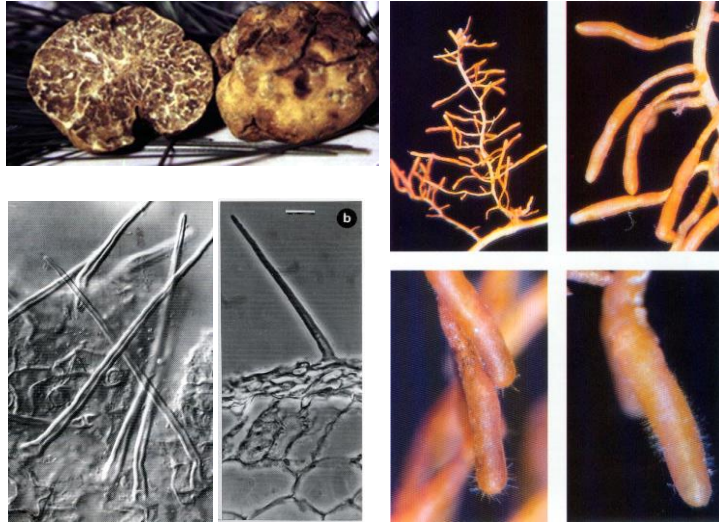
Main features in *Leccinum lepidum*



Main features in *Cortinarius* sp.

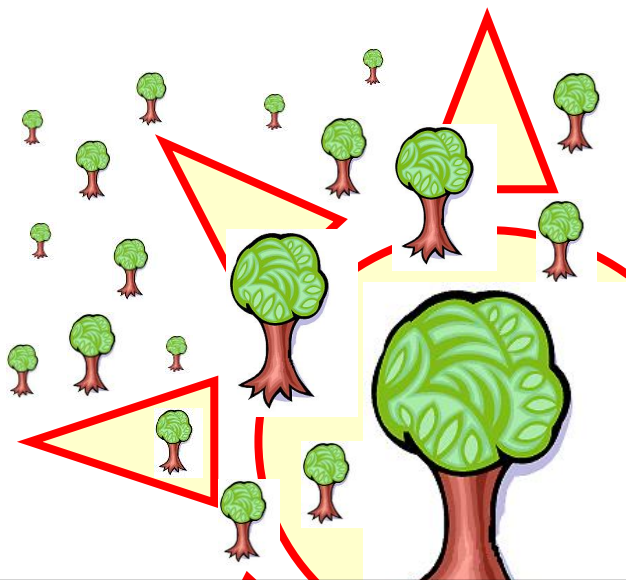


Main features in *Tuber* sp.

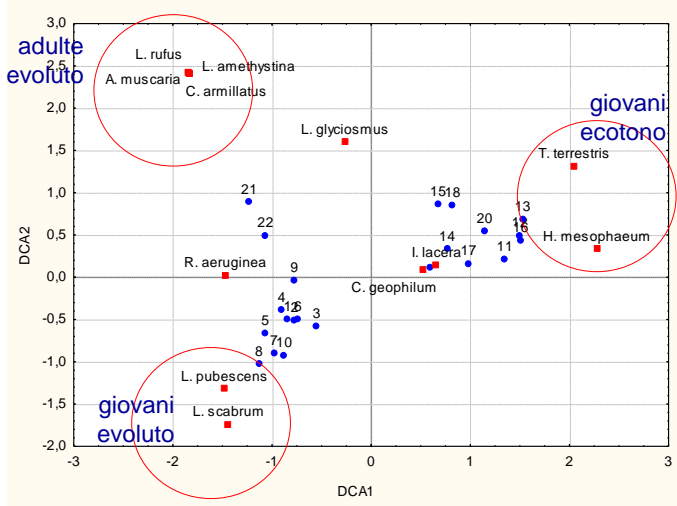


A bit of ecology

Pioneer trees need pioneer symbionts

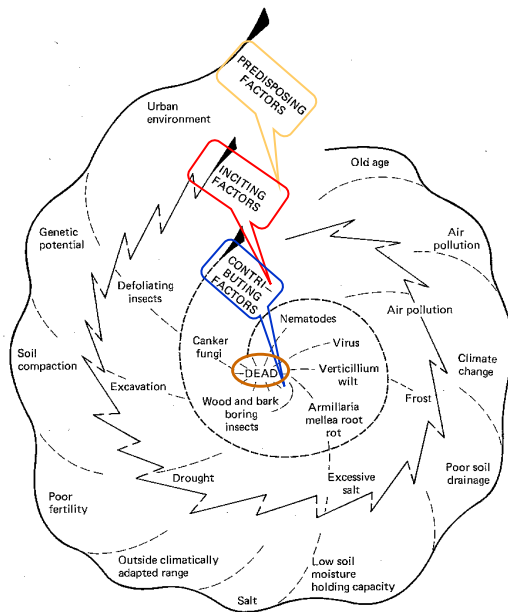


T. terrestris e *H. mesophaeum*:
Very good in non-forest soils



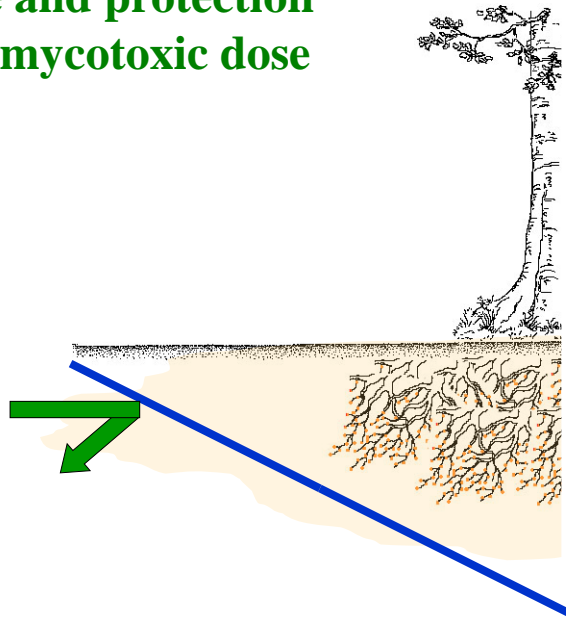
Mycorrhizal symbioses and trees' decline

Trees decline



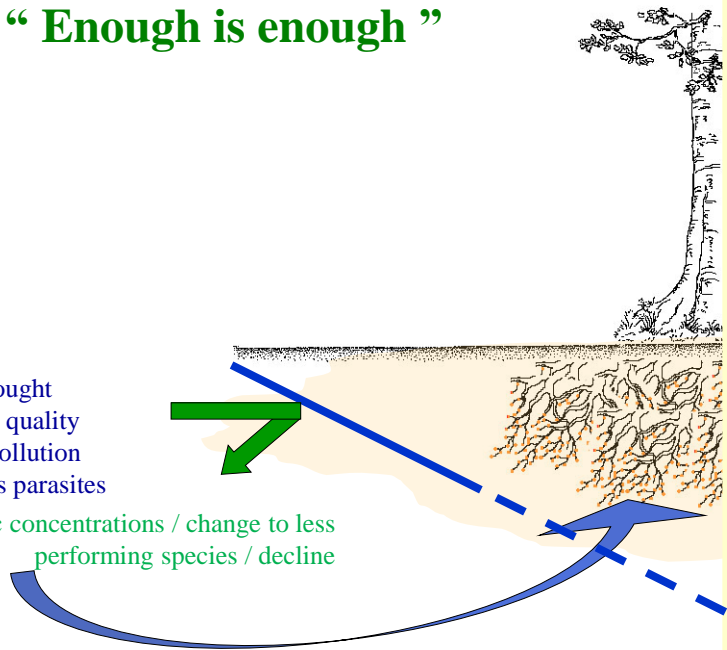
Tree care and protection until the mycotoxic dose

Drought
Water quality
Soil pollution
Root tips parasites



“ Enough is enough ”

Drought
Water quality
Soil pollution
Root tips parasites
Fungitoxic concentrations / change to less performing species / decline



An overview of oak decline in the Mediterranean region

Main declining oak species in the Mediterranean region

Common oak (*Quercus robur*)
Turkey oak (*Quercus cerris*)
Hungarian oak (*Quercus frainetto*)
Downy oak (*Quercus pubescens*)
Holm oak (*Q. ilex*)
Cork oak (*Q. suber*).



Oak decline show symptoms common to all forest trees decline

«Forest» decline is documented in Europe since the 1940s.

Since the eighties it has been reported also on Oak species (or was it a matter of knowledge and experience in survey?).

Symptoms are mainly associated to soil-related physiological stressors (drought, compaction, fertility, assolation, root up-take).

- 1) Root uptake deficiencies.
- 2) Canopy transparency increases due to yellowing, wilting, fall of leaves.
- 3) Annual internodes shorten.
- 4) Epicormic twigs appear along the trunk, downwards to the collar.

5) Human-assisted practices speed up the process: mainly intensive sylviculture, pasture, land-reclamation.

6) Endophytic fungi commonly present in all healthy trees turn to parasitism (i.e. *Biscogniauxia*, *Hypoxylon*, *Diplodia*, *Collybia*).

7) Known pests and pathogens infect declining trees causing root rots, bark cracks and wood decay (i.e. *Phytophthora*, *Armillaria*, *Ganoderma*, *Phellinus*)

8) Fall and substitution by «stressors-tolerant» genotypes or species, not necessarily trees (i.e. Holm- and Cork oak with *Cistus*).



Visible symptoms are related to insufficient water uptake





Leaves far from the roots dies and epicormic twigs appear



Endophytic mutualists turn to a parasitic behaviour



Root and wood parasites compromise tree stability

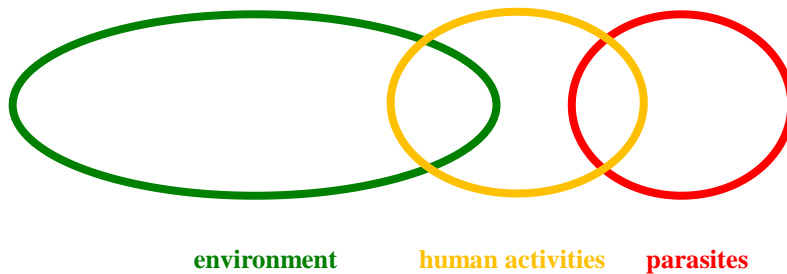


Declining trees fall down giving room to different genotypes or species



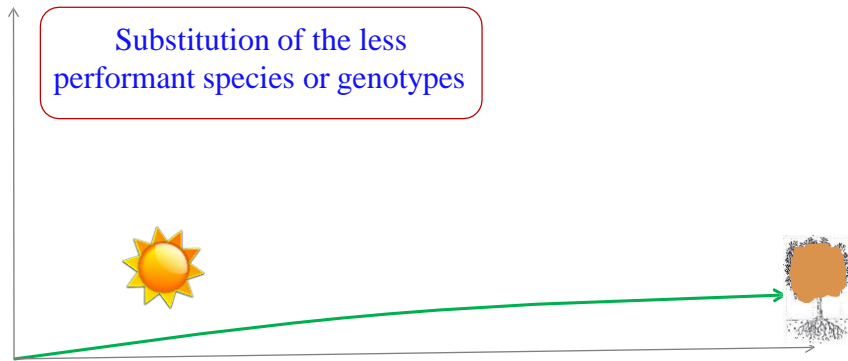
What «Decline» is?

The result of three consecutive, overlapping causes



«Environment»: well known ecological dynamics

Substitution of the less performant species or genotypes



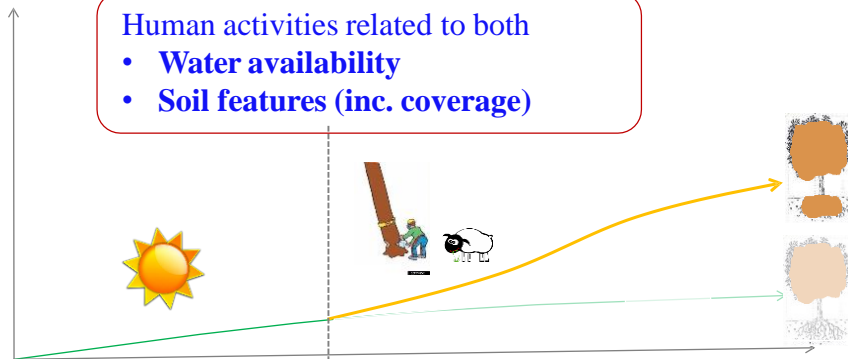
Long-term Environmental factors

Low resilience to long-term climate changes
Senescence

Human activities compromising «soil health»

Human activities related to both

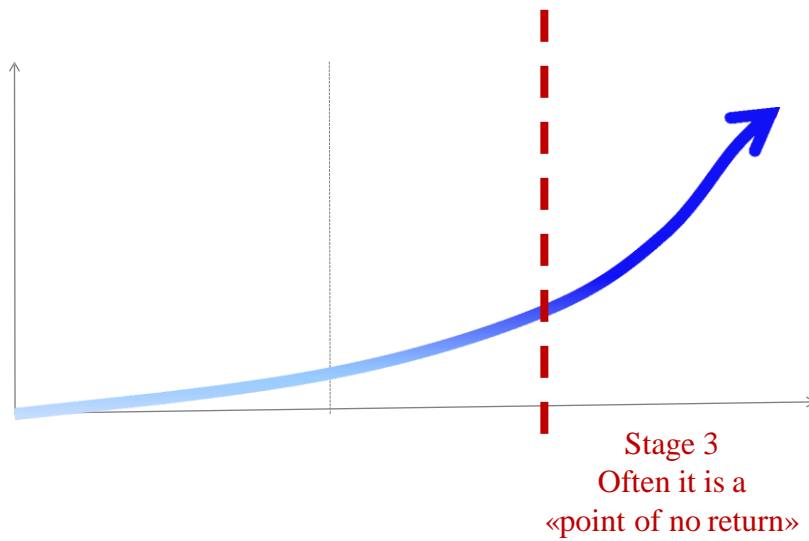
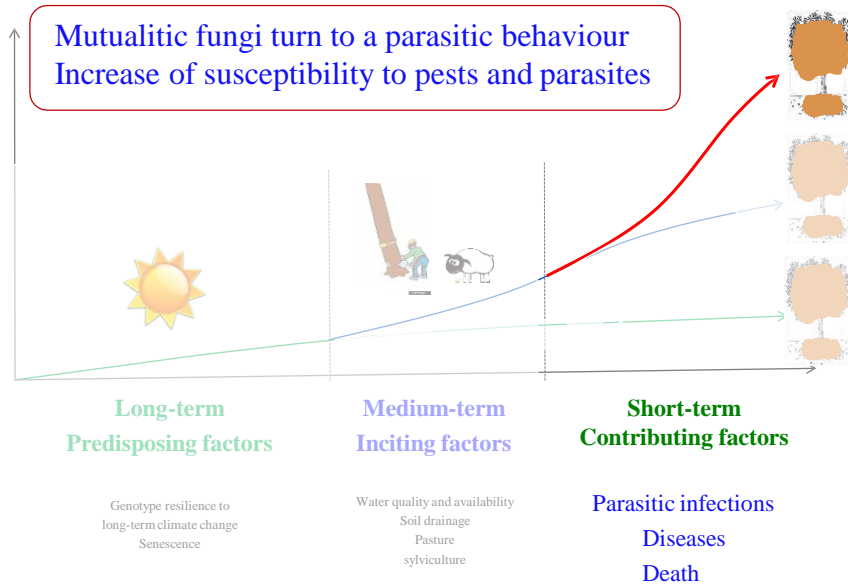
- Water availability
- Soil features (inc. coverage)

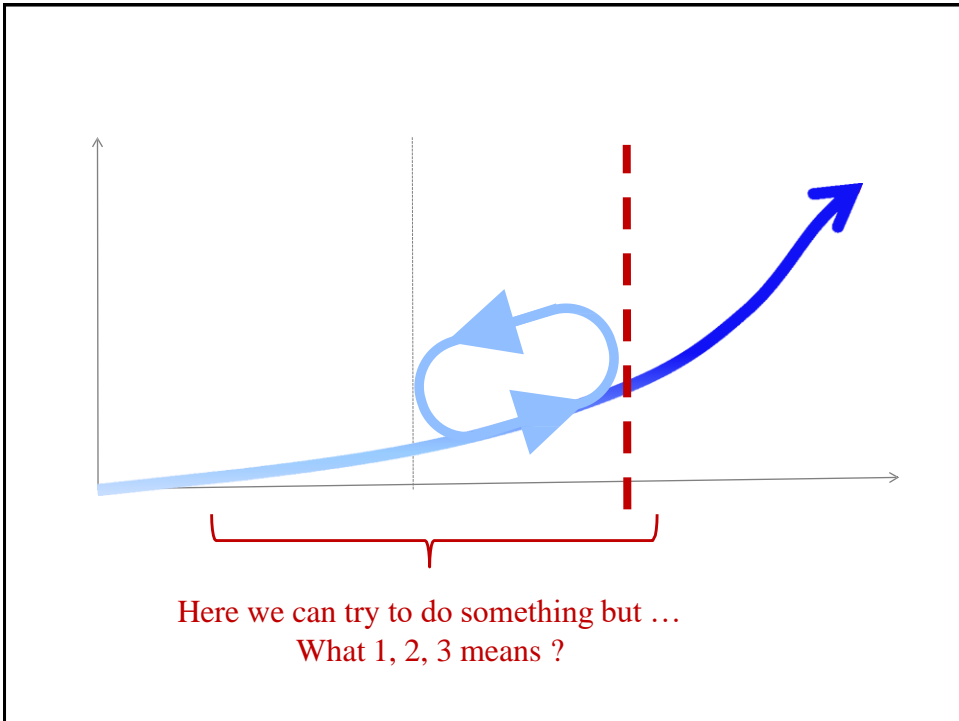


Medium-term Inciting factors

Water quality and availability
Soil drainage
Pasture, grazing
Agriculture, Sylviculture

Pests and parasites as final effect on already stressed trees

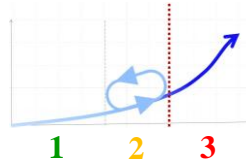




Forest owners and managers need to quantify decline.
«1 or Low» «2 or Medium» «3 or High» usually are nonobjective,
mainly depending on previous experience and ability.

«How much declining is my declining oak»

The complex block contains text, a speech bubble, and illustrations. At the top, there is a bolded sentence followed by a line of text in quotes. Below this is a green speech bubble containing another line of text. At the bottom, there is a cartoon illustration of a doctor in a white coat and stethoscope, pointing his right index finger upwards. To the left of the doctor are two icons: a thermometer and a sad face emoji with a blue forehead.



Decline can be measured through the composition of the ectomycorrhizal community.

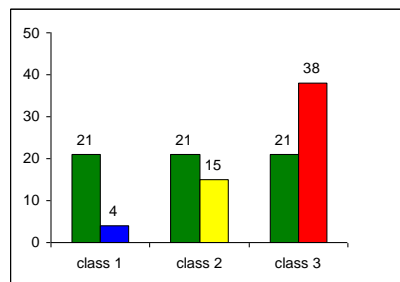
**Changes in ectomycorrhizal diversity
in a declining *Quercus ilex* coastal forest**

Lucio Montecchio, Roberto Causen, Sergio Rossi and Sergio Mutto Accordi

**Pedunculate oak decline: thinning effects
on the ectomycorrhizal exploration groups**

Elena Mosca¹, Linda Scattolin¹, Jean Garbaye², Sergio Mutto Accordi¹, Lucio Montecchio¹
¹Università degli Studi di Padova, Dipartimento Te.S.A.F., V.le dell'Università 16, I-35020
 Legnaro, Italy; ²INRA, UMR1136 Interactions Arbres/Microorganismes, F-54280
 Champenoux, France

Few, peculiar species can say where we are



At least 1 among the «blue» ones?

At least 1 among the yellows?

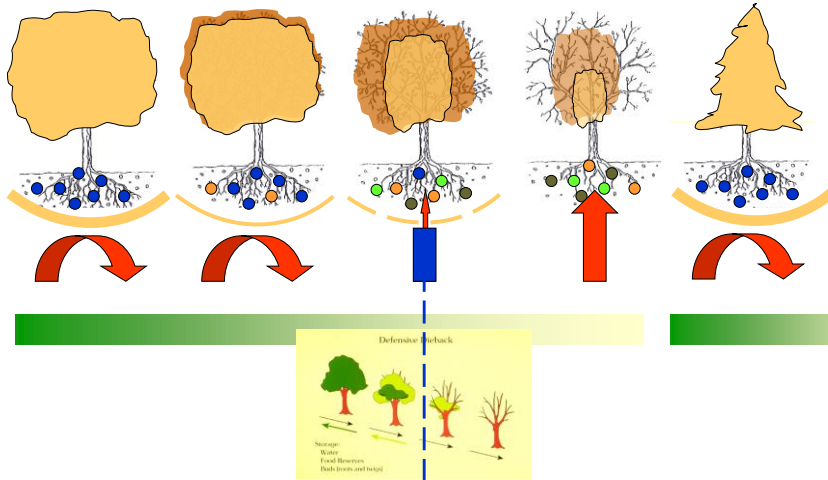
At least 1 among the reds?

Not a problem!

Do something

Too late, sorry

**Decline of both fungi and trees.
Species substitution**



**Tree species decline
Change in forest composition**



Sweet Chestnut Ink Disease

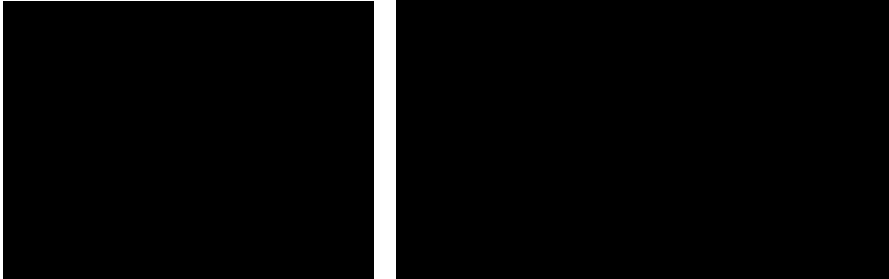
cause, symptoms and control

What CID is?

A lethal disease caused by *Phytophthora* (*cambivora* and *cinnamomi*), a soil-borne parasite.

Compacted soils, poor drainage and temporary films of water allow the parasite to move to the root tips by means of its **flagellated spores** (rotating like a propeller).

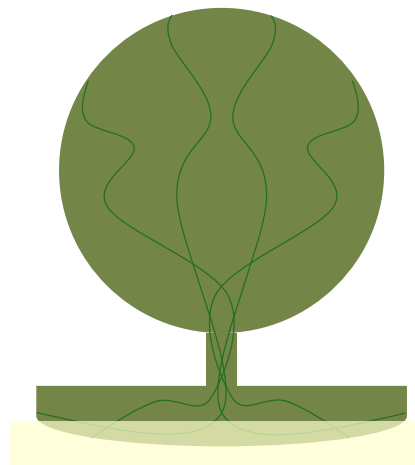
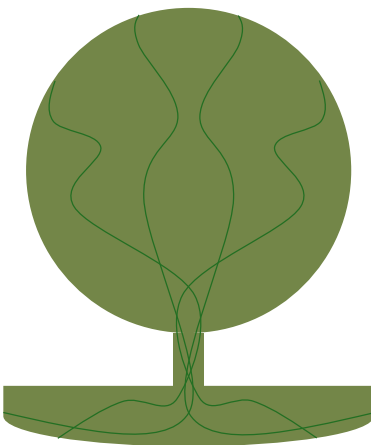
Flagellate spores need water to swim

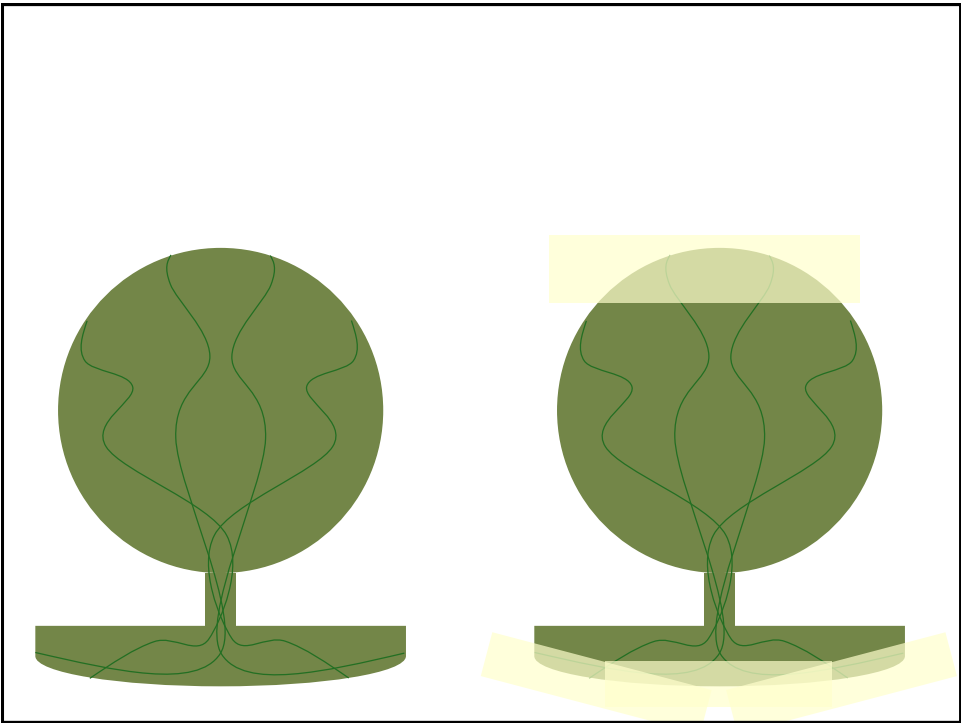
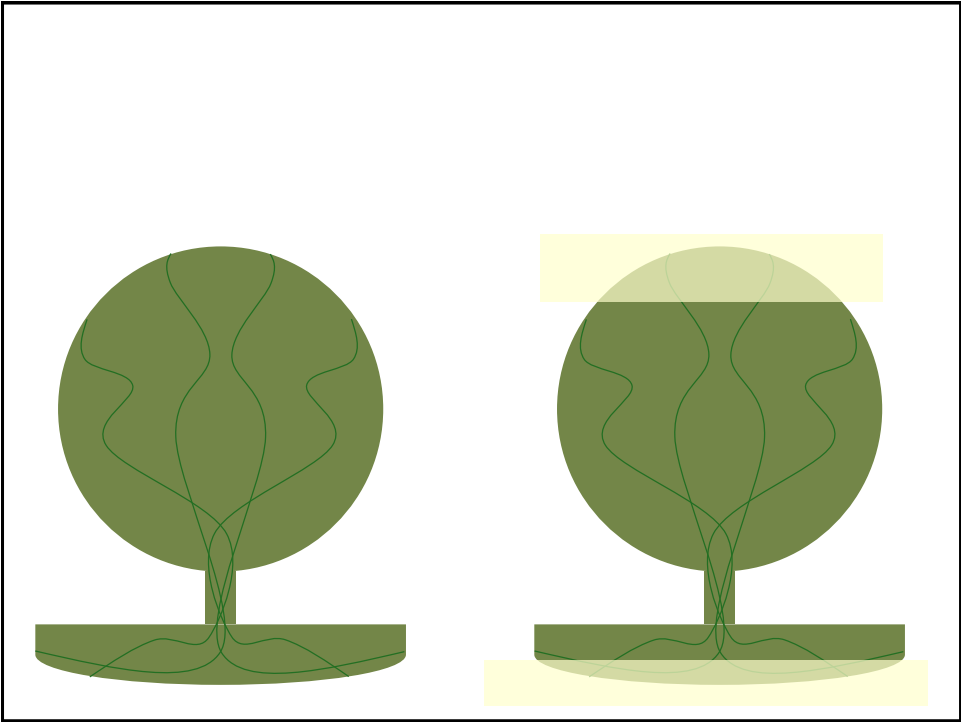


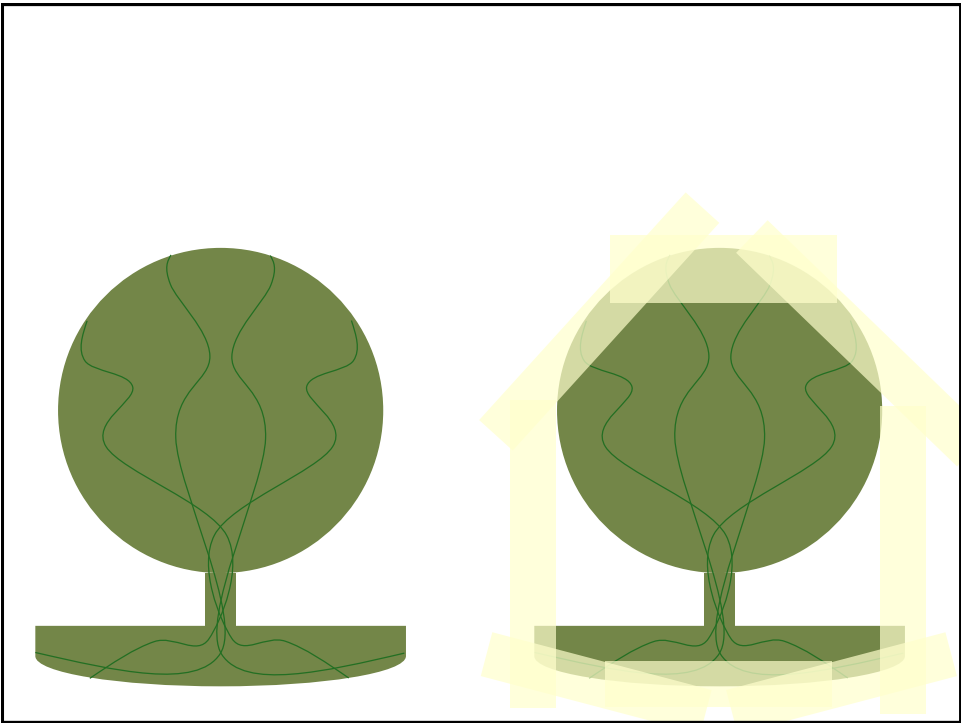
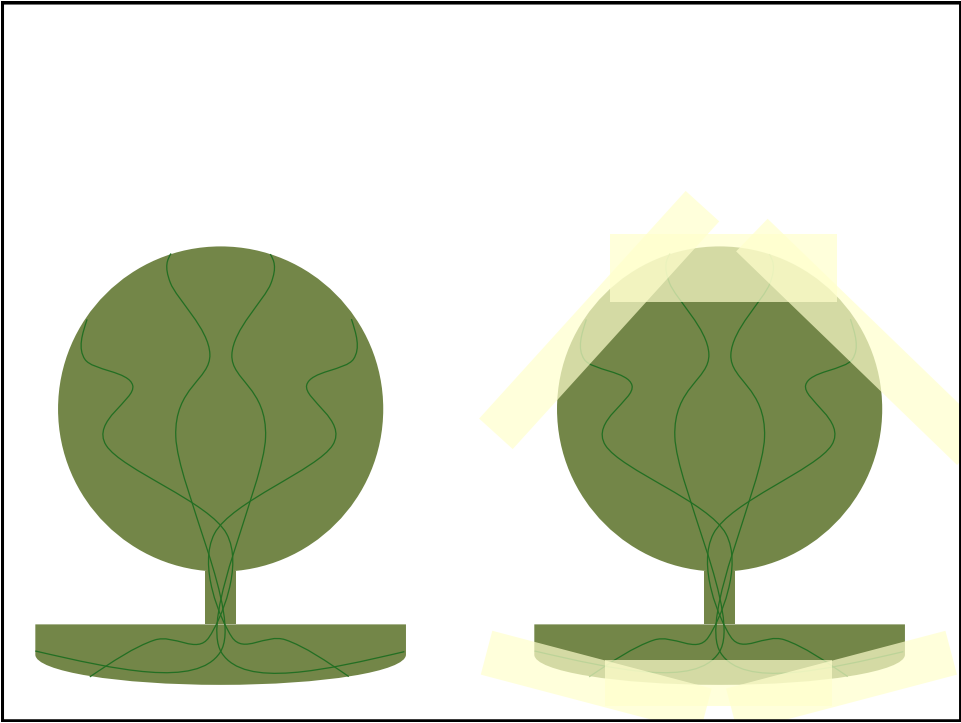
The Training Tree
Pubblicato il 28 ott 2016

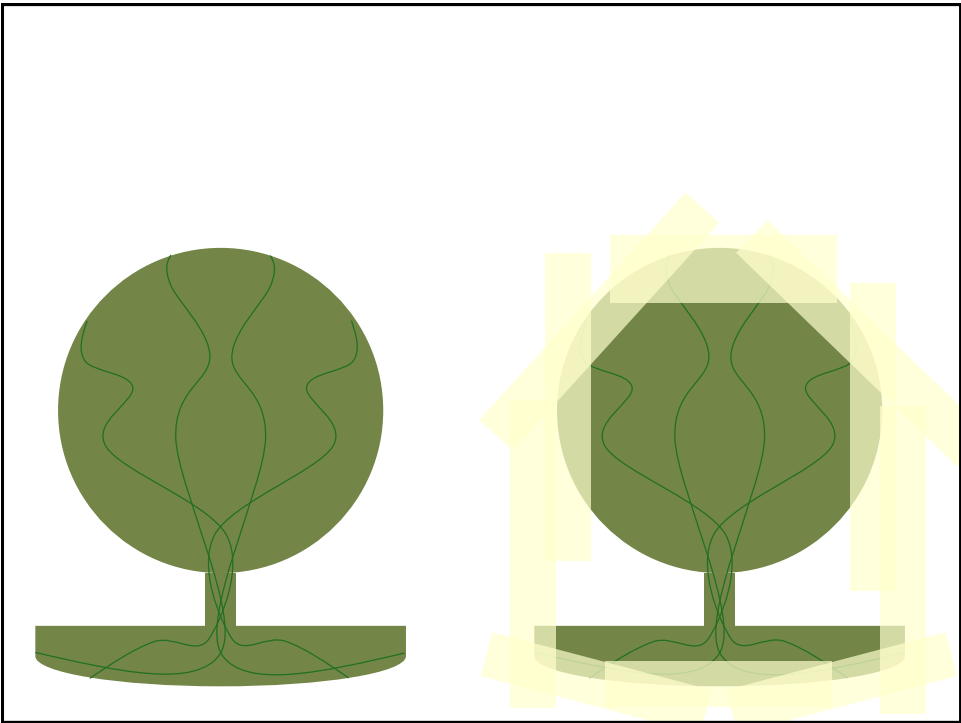
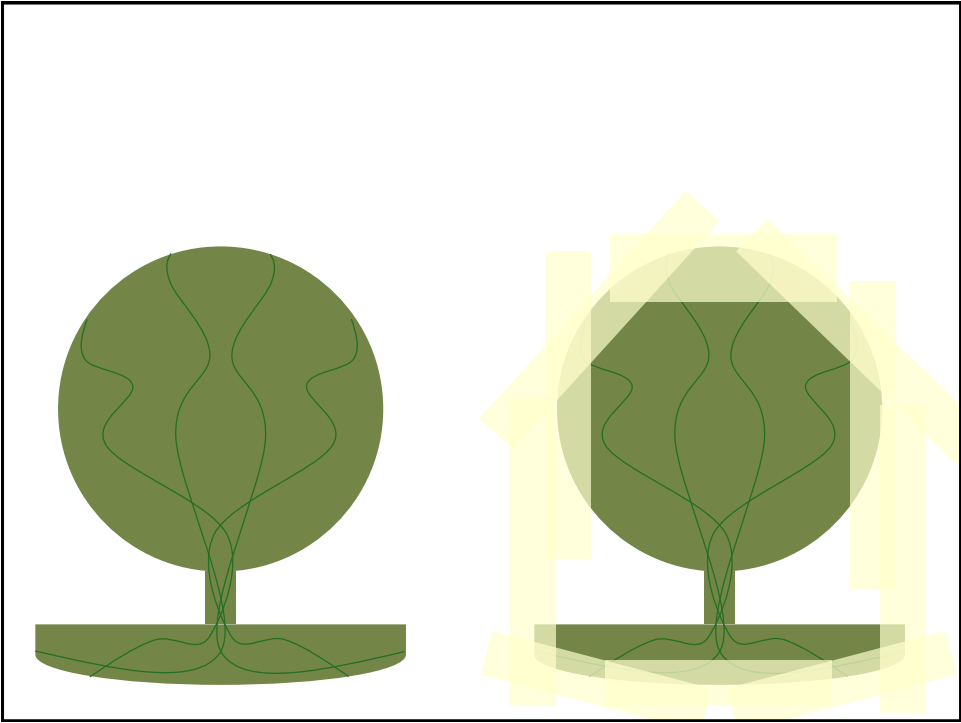
Phytophthora species are not true fungi. They are Oomycetes, Kingdom **Chromista**.

Lack of rootlets, lack of canopy
Macroscopic symptoms are related to a
insufficient nutritional uptake.



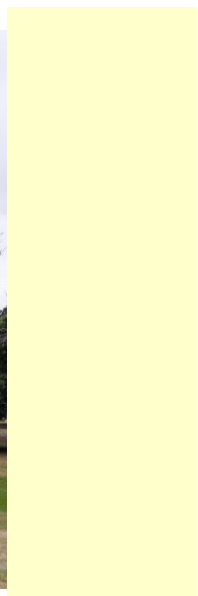








Three different stages of CID
Greenwich Park, October 2016



Three different stages of CID
Greenwich Park, October 2016



Three different stages of CID
Greenwich Park, October 2016

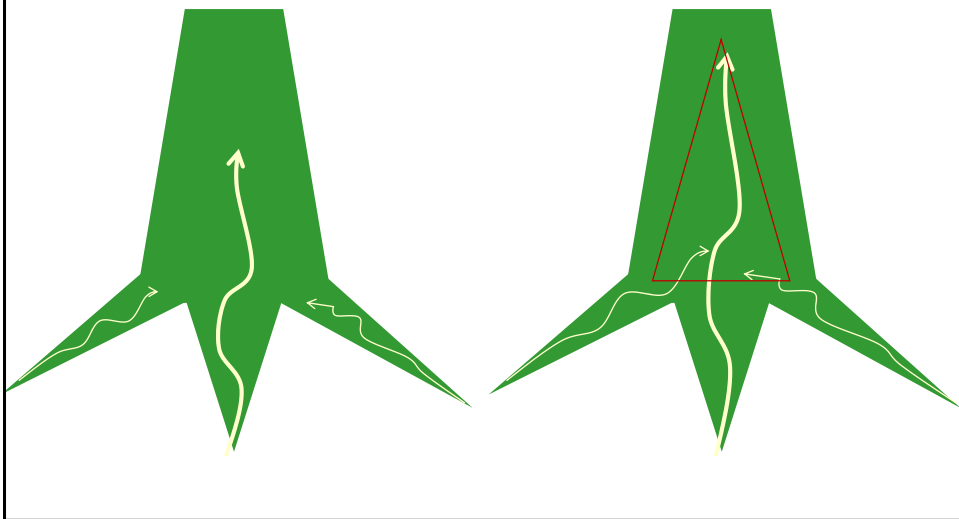
Early stages **above-ground** symptoms:
vertical cracks and necroses rising from main roots,
sometimes leaking tannins («ink», a tree reaction).

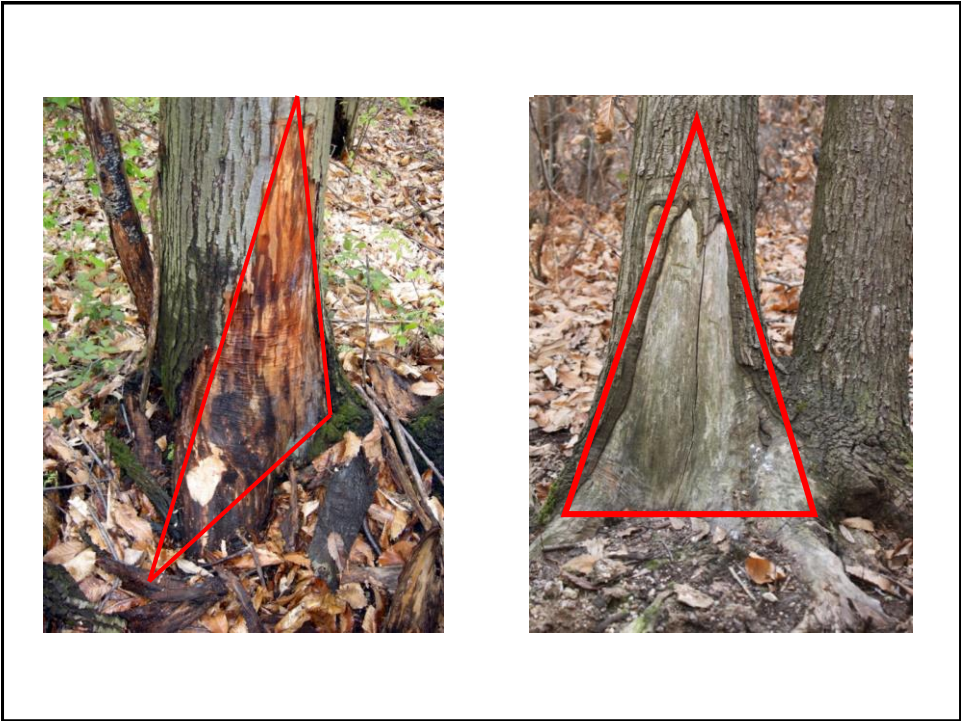




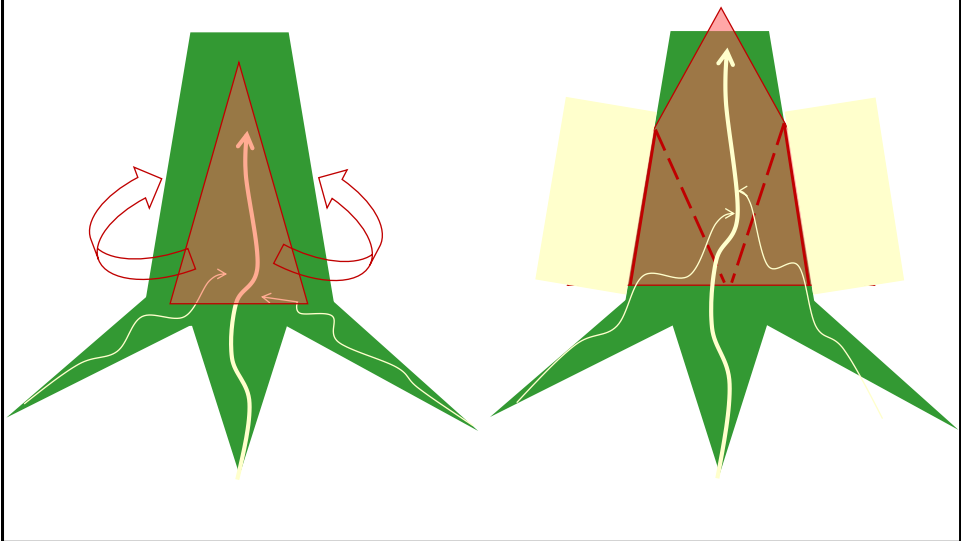
Tannins leak mainly in warm, mediterranean climates

Then, individual necroses merge in a bigger, (usually) triangular one.

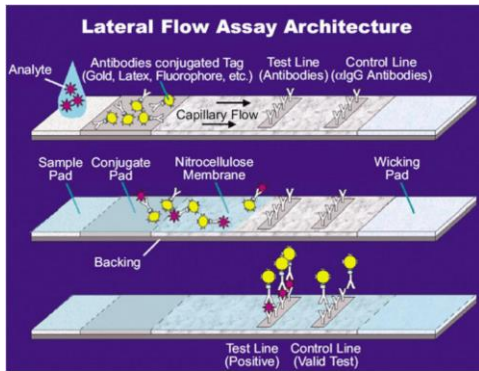




Once the necroses girdles the base, uptake stops and the whole canopy dries out.



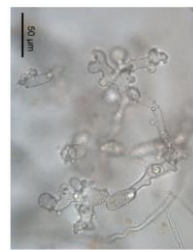
Diagnosis: is it easy?
to genus level, yes. Through lateral flow tests



An illustration, by NASA, of a lateral flow assay.[1]



Diagnosis: is it easy?
to genus level, yes. Through lateral flow tests
to species level, no. We need a good lab and well trained staff



Sequences producing significant alignments:

Select: All None Selected:0

Alignments Download GenBank Graphics Distance tree of results

Description	Max score	Total score	Query cover	E value	Ident	Accession
<input type="checkbox"/> Phytophthora cinnamomi strain CBS 144.22 cytochrome oxidase subunit I (cox1) gene_part	717	1130	84%	0.0	100%	KC609419.1



**Early stages below-ground symptoms:
infected, still non symptomatic chestnuts show a peculiar mycorrhizal
community**

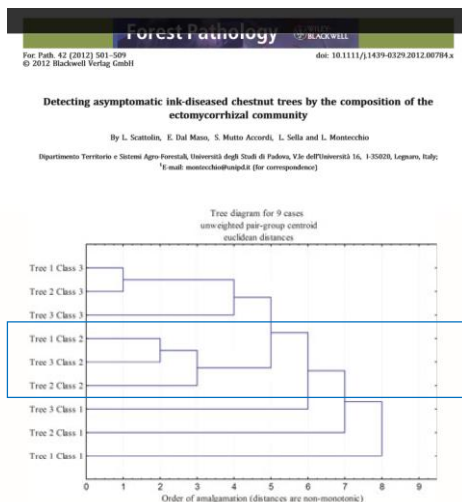


Fig. 4. Cluster analysis dendrogram, based on the Euclidean distances from the tree centroids using the data acquired in October 2008 (trees 1, 2 and 3).

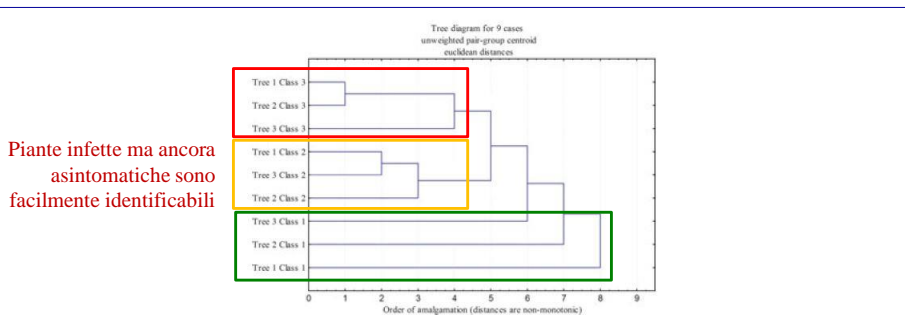
For. Path. 42 (2012) 501-509
© 2012 Blackwell Verlag GmbH

doi: 10.1111/j.1439-0329.2012.00784.x

**Detecting asymptomatic ink-diseased chestnut trees by the composition of the
ectomycorrhizal community**

By L. Scattolin, E. Dal Maso, S. Mutto Accordi, L. Sella and L. Montecchio

Dipartimento Territorio e Sistemi Agro-Forestali, Università degli Studi di Padova, V.le dell'Università 16, I-35020, Legnaro, Italy;
¹E-mail: montecchio@unipd.it (for correspondence)



Piante infette ma ancora
asintomatiche sono
facilmente identificabili

Fig. 4. Cluster analysis dendrogram, based on the Euclidean distances from the tree centroids using the data acquired in October 2008 (trees 1, 2 and 3).

Prevention: forecasting and survey

Large-scale fuzzy rule-based prediction for suitable chestnut ink disease sites: a case study in north-east Italy

By E. Dal Maso¹ and L. Montecchio^{1,2}

Large-scale fuzzy prediction of chestnut ink disease

7

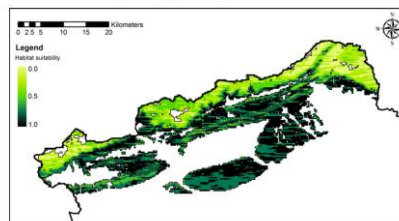


Fig. 4. Predicted spatial habitat suitability for chestnut ink disease in the study area. Different colours represent different levels of predicted probability.

Containment

In urban forestry pesticides effective against *Phytophthora* are not allowed.

Integrated pest management (IPM) strategies to:

- Improve soil health, drainage and composition (addition of organic substance)
- Biostimulate trees to increase their natural defences (trunk injection)

Control of CID through trunk injection

ARBORICULTURAL JOURNAL, 2017
<https://doi.org/10.1080/03071375.2017.1345538>



Check for updates

An enhanced trunk injection formulation of potassium phosphite against chestnut ink disease

Elisa Dal Maso^{a,b}, Jonathan Cocking^{b,c} and Lucio Montecchio^{a,b}

^aDipartimento Territorio e Sistemi Agro-Forestali, Università degli Studi di Padova, Legnaro, Italy; ^bPAN/De Rebus Plantarum, Spinoff of the University of Padova, Padova, Italy; ^cJCA Ltd, Halifax, UK

ABSTRACT

Chestnut ink disease, caused by *Phytophthora cinnamomi* and *P. cambivora*, is one of the most destructive diseases affecting *Castanea sativa*. Currently, disease control requires careful integrated chemical and agronomic measures. Trunk injection with potassium phosphite was proven to be effective in reducing symptoms but little is known about the ideal formulation. In this research, potassium phosphite at different concentrations and with some other bio stimulants was injected into sweet chestnuts which had been inoculated with a local strain of *P. cinnamomi*. The most effective formulation, potassium phosphite mixed with a micronutrient solution, was then tested with a preventive approach; as a result, the trunk injection treatments completely ceased the development of the disease *in planta* in most cases. In order to consider the importance of each component of the solution, potassium phosphite and the bio stimulant elements were tested *in vitro* singly and in an isobolographic analysis of interactions. The results broaden the knowledge base on endotherapeutic treatments as an effective measure for the management of chestnut ink disease with potential for many other diseases.

KEYWORDS

Phytophthora cinnamomi; *Castanea sativa*; xylematic injection; micronutrients; isobolographic analysis

Potassium salts of phosphorous acid vs. Chestnut Ink Disease

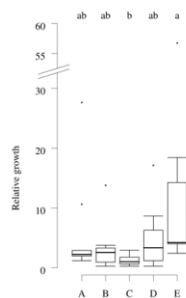
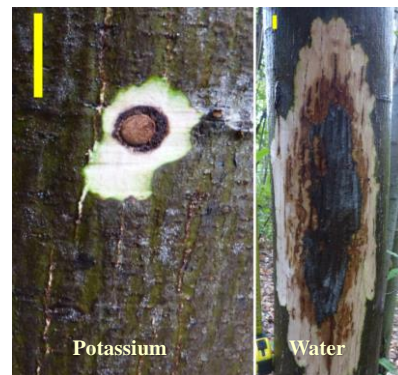
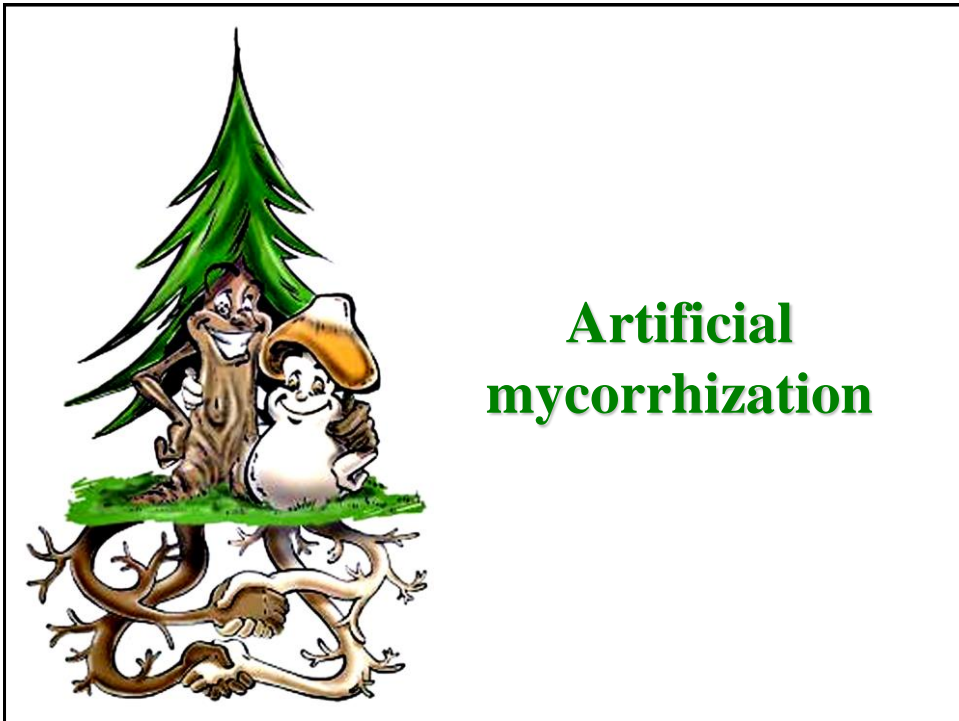


Figure 22. Differences in the relative increase of the necrotic areas after 50 days from the treatments. A = Potassium phosphite 35 %; B = Potassium phosphite 70 %; C = Potassium phosphite 35 % plus micronutrient solution 0.1 %; D = Potassium phosphite 35 % plus alicin solution 20%; E = Control.



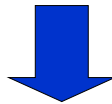
Curative
 Injection 21 days after infection.
 End of trial 50 days after injection (= 71).

Preventative (bar=2cm)
 Injection 27 days before infection.
 End of trial 50 days after infection (=77).



Benefits

Nitrogen and potassium uptaken easily
Increase of useful bacterial communities
Biocontrol of root parasites



Quicker growth
Higher resistance to transplant stresses
Higher resistance to root parasites

When it's useful (not in a natural forest soil, of course)

- Non-pioneer species on poor soils.
- Edible fruit bodies production.



It's possible
(1887: Frank)

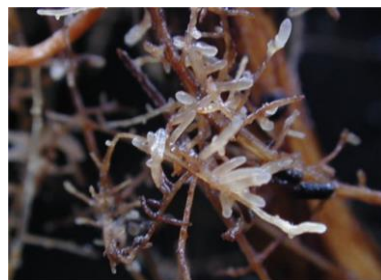


Artificial inoculation

- Transplanting close to a mycorrhized tree («proximity»)
- Sowing into naturally infected soil
- Mixing carpophores into soil
- **Mixing mycelium into soil**

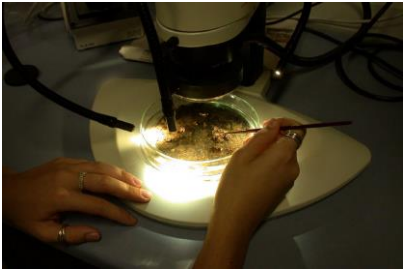
The fungus must be ...

- A pioneer species (easy to isolate and grow far from living tips).
- Able to maintain his abilities for a long time, also when growing in artificial substrates for weeks.
- Compatible with the nursery conditions and environment.
- Compatible with the destination tree and environment



Laccaria laccata

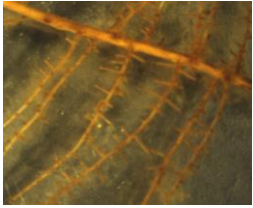
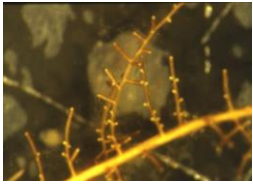
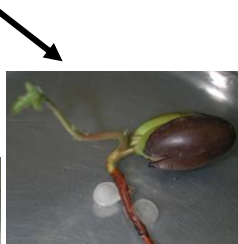
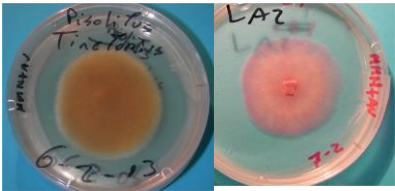
Production

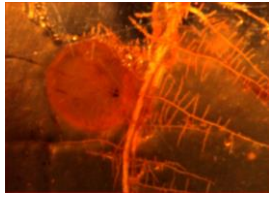


Ad hoc labs and skillful technicians



Preliminary investigations *in vitro*

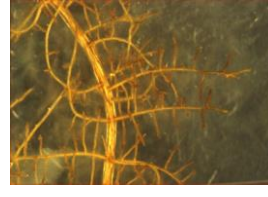




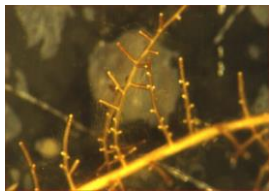
Laccaria amethystina LA3



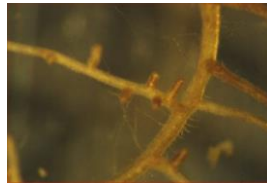
Laccaria bicolor LB1



Laccaria laccata LL2



Laccaria laccata LL3



Laccaria laccata LL4



Paxillus involutus PI-I

Production of pure strains of the selected species



Different methods



Artificial mycorrhization



Adjustment of the growing methods:

- Disinfection of substrates, pots, seed.
- Composition e pH of the soils
- Decrease fertilization
- Avoid fungicides
- Avoid herbicides
- Decrease watering
- Survey, survey, survey



Hygiene !!!





Laccaria laccata: on pinus mugo 12 months after



Laccaria bicolor fruiting
from the bottom of a tray!



Perspectives

- Long term plans (i.e. mix of non-antagonistic species)
- Local strains
- Quality control and certification



Certification: not easy



R. fibula: not mycorrhizal



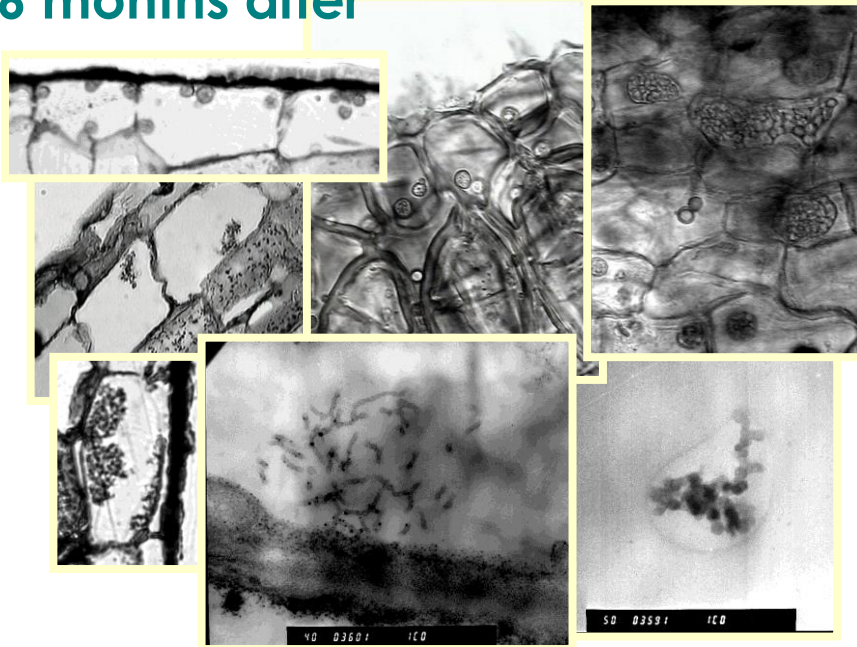
H. crustuliniforme: yes

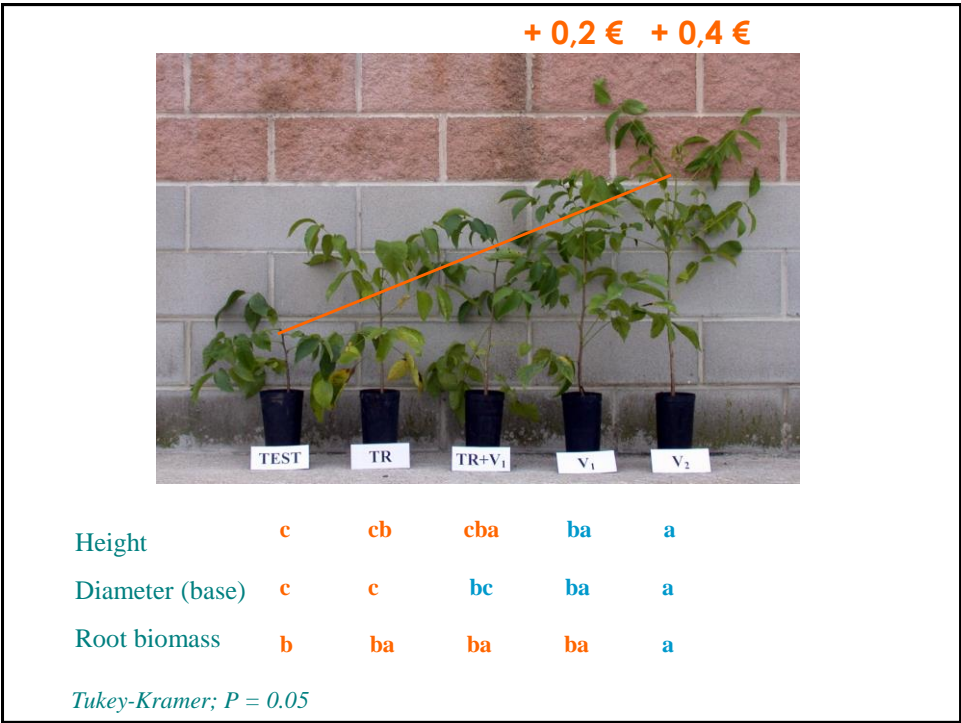
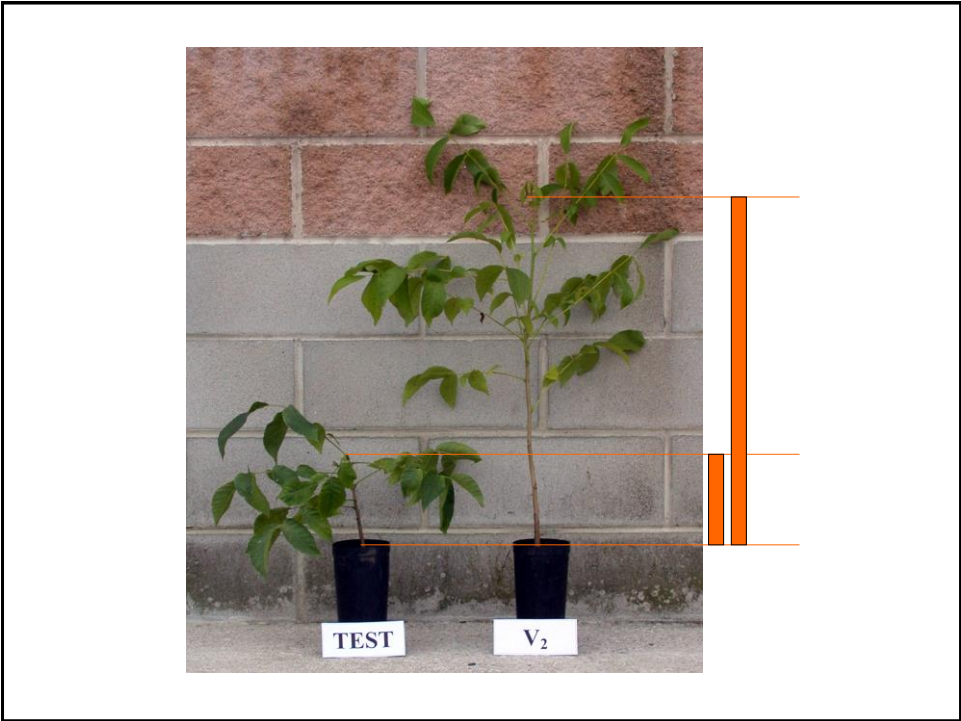
Artificial mycorrhization on *Juglans regia*

5 treatments x 100 replicates.

- Control
- *T. harzianum*
- *T. harzianum* + AM dose a
- AM dose a
- AM dose 2a

6 months after





TRUFFLES!



White truffles



Black truffles

9 species allowed to collect in Italy

- | | |
|----------------------------|---|
| 1) Tartufo bianco pregiato | (<i>Tuber magnatum</i> Pico) |
| 2) Tartufo bianchetto | (<i>Tuber borchii</i> Pico) |
| 3) Tartufo nero pregiato | (<i>Tuber melanosporum</i> Vitt.) |
| 4) Tartufo scorzone | (<i>Tuber aestivum</i> Vitt.) |
| 5) Tartufo uncinato | (<i>Tuber uncinatum</i> Chat.) |
| 6) Tartufo brumale | (<i>Tuber brumale</i> Vitt.) |
| 7) Tartufo moscato | (<i>T. brumale</i> var. <i>moschatum</i> De Ferry) |
| 8) Tartufo mesentericum | (<i>Tuber mesentericum</i> Vitt.) |
| 9) Tartufo macrosporum | (<i>Tuber macrosporum</i> Vitt.) |

TUBER MAGNATUM PICO





Q. robur

P. alba



Tuber magnatum

TUBER BORCHII



3. PIANTE SIMBIONTI AUTOCTONE IN VENETO

Q. pubescens

Q. ilex

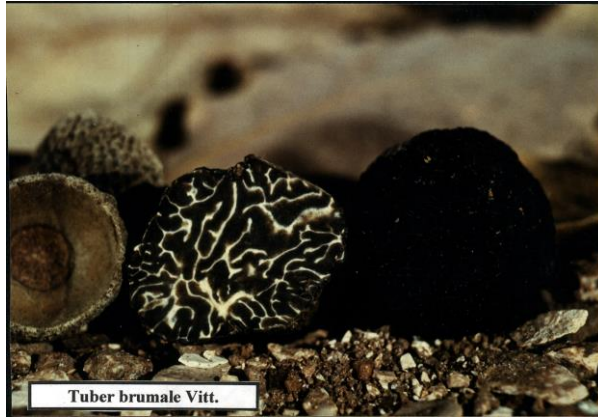


Tuber borchii

TUBER MELANOSPORUM



TUBER BRUMALE



TUBER MOSCHATUM



TUBER UNCINATUM Chatin



C. avellana

O. carpinifolia



Tuber aestivum

TOXIC !!!



Tuber rufum



Balsamia vulgaris



Choiromyces meandriformis

Collection



