



Research with strain collections: The advantage of strain multitudes using the example of diagnostics of fungal infections

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ECCO XL, 28th September, 2022



Mucorales are everywhere

- Basal filamentous fungi
- Ubiquitous saprobes (e.g. soil, hay; fruits - post harvest pathogen)

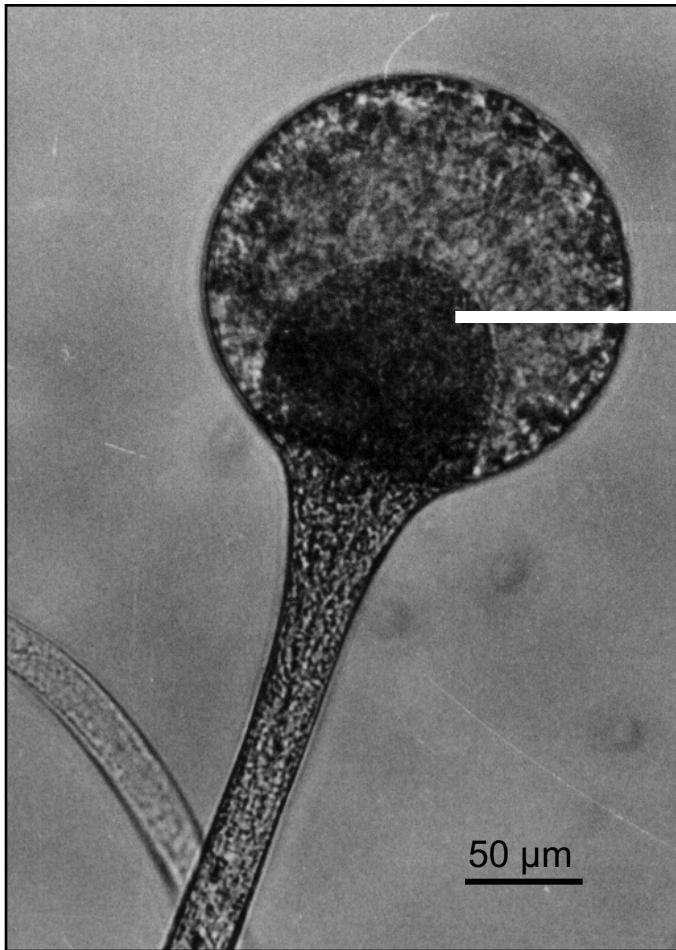


Photo by: Don Edwards

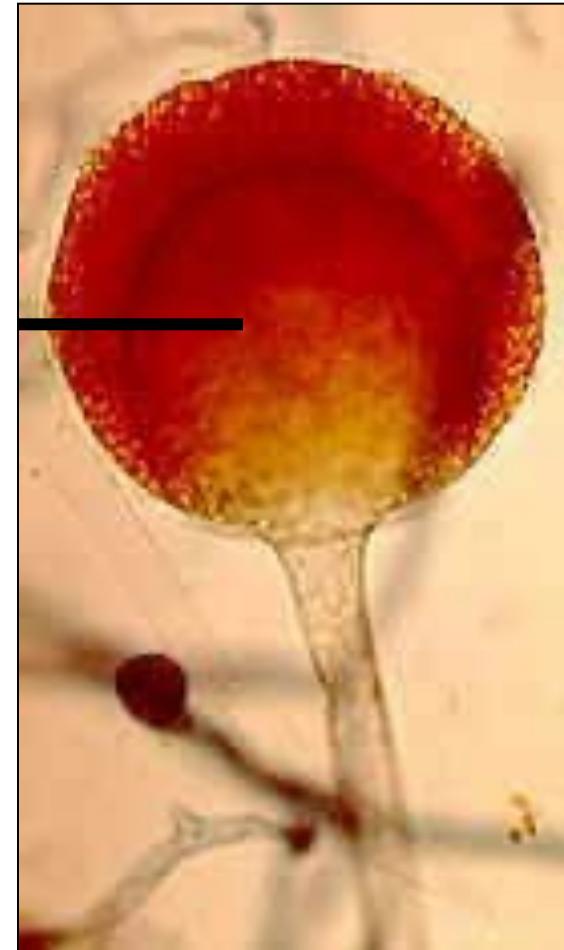


Photo courtesy: Grit Walther

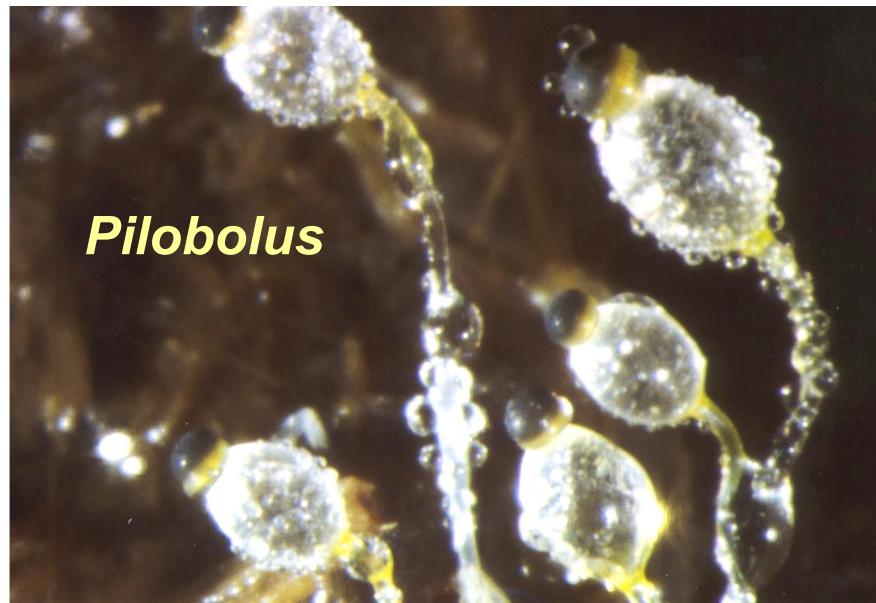
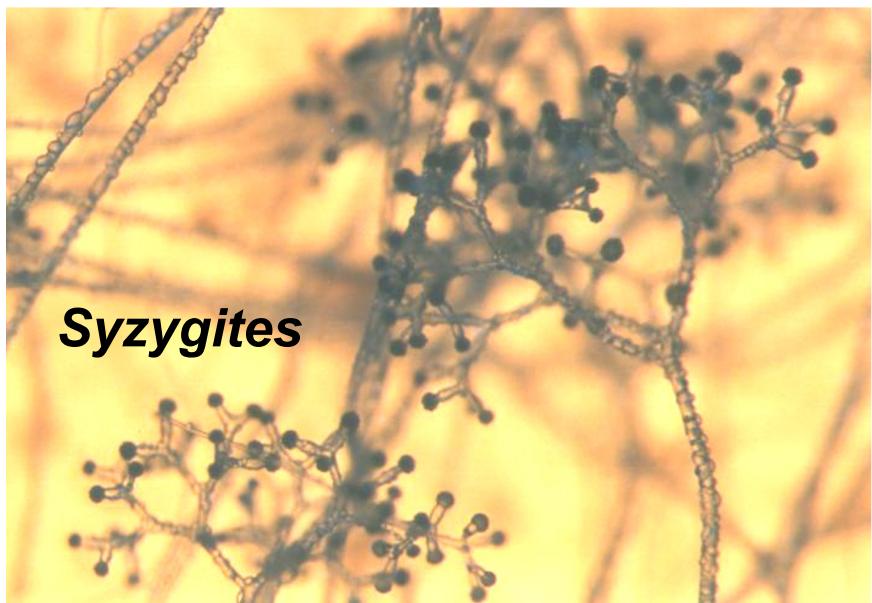
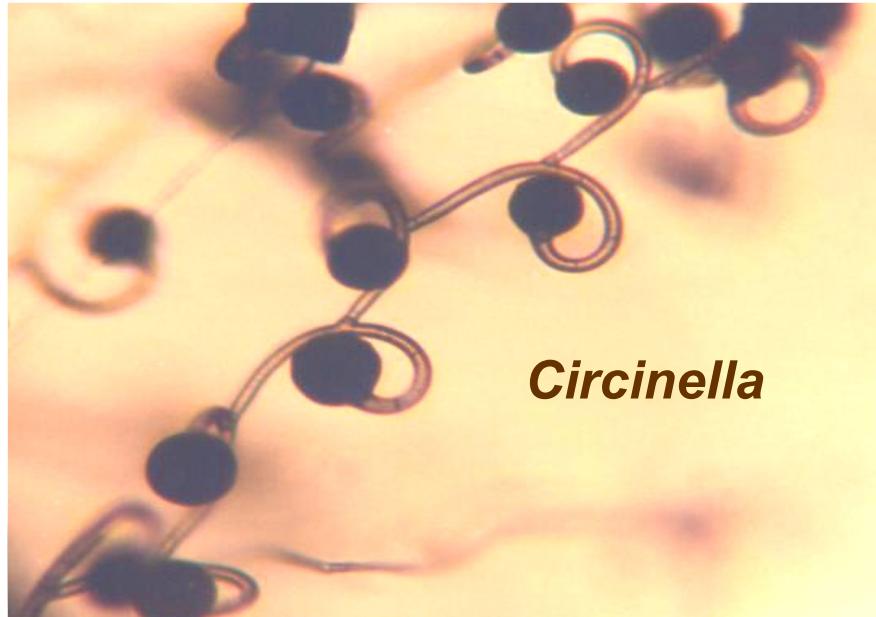
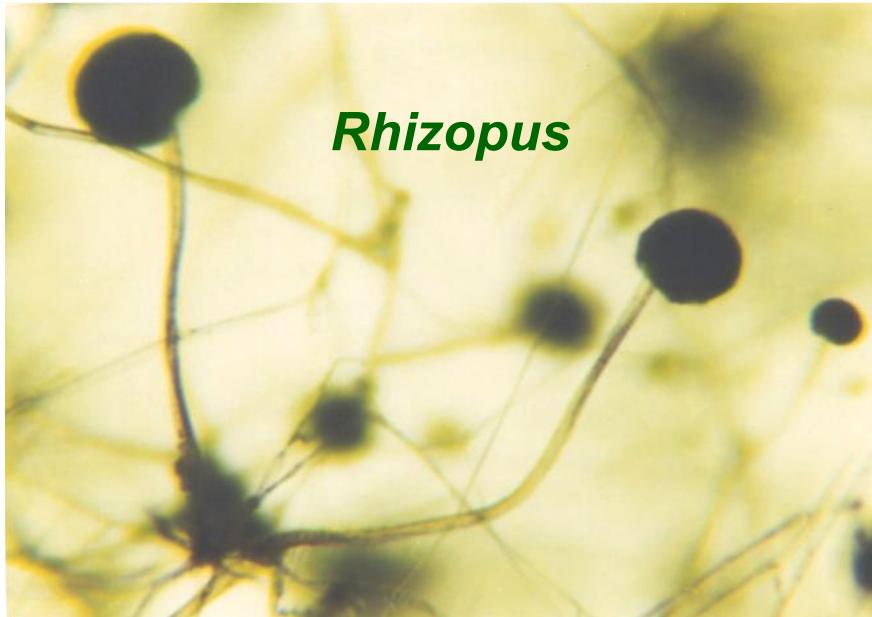
The trade-mark of the Mucorales is a swollen extension of the sporangiophore called a columella, which protrudes like a balloon into the sporangium.



Columella
a bulbous
vesicle at the
sporangiophore apex

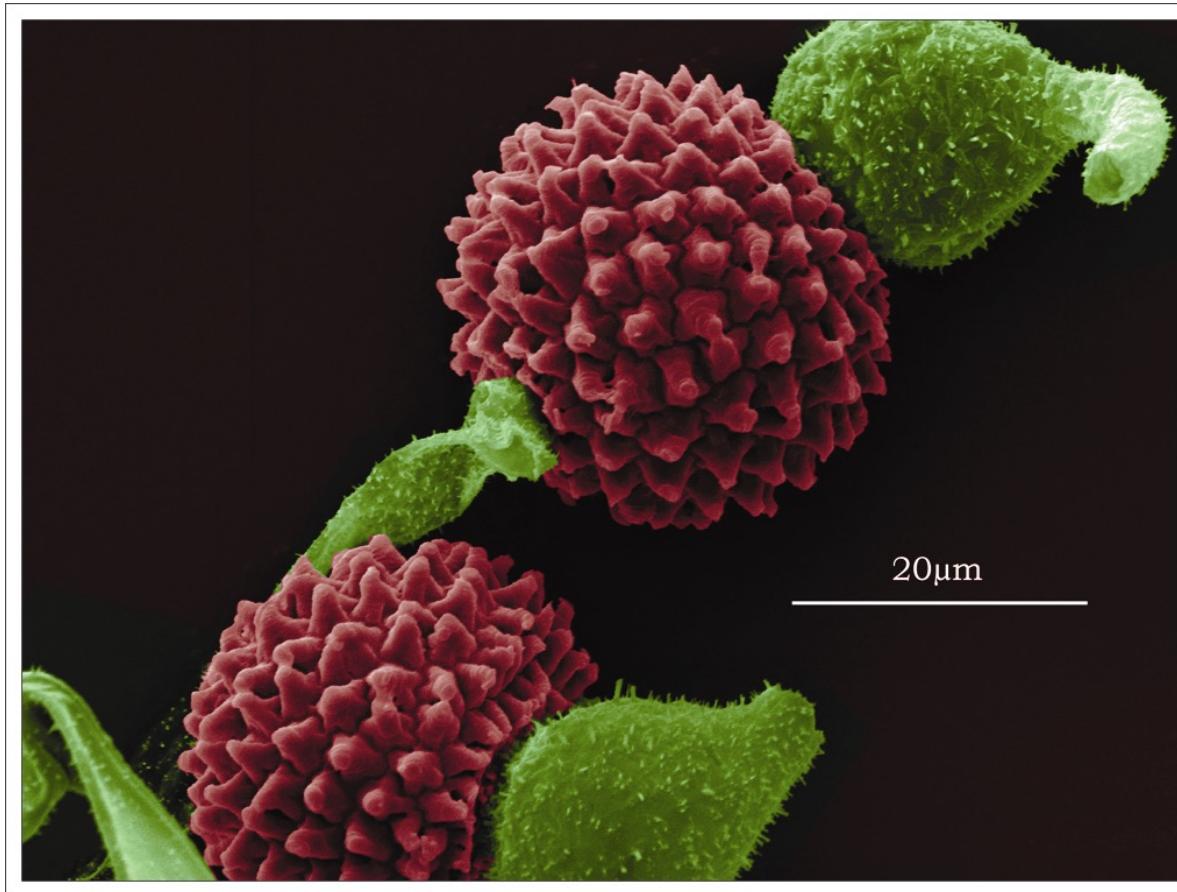


Mucorales propagate asexually *via* mitospores produced endogenously in sporangia



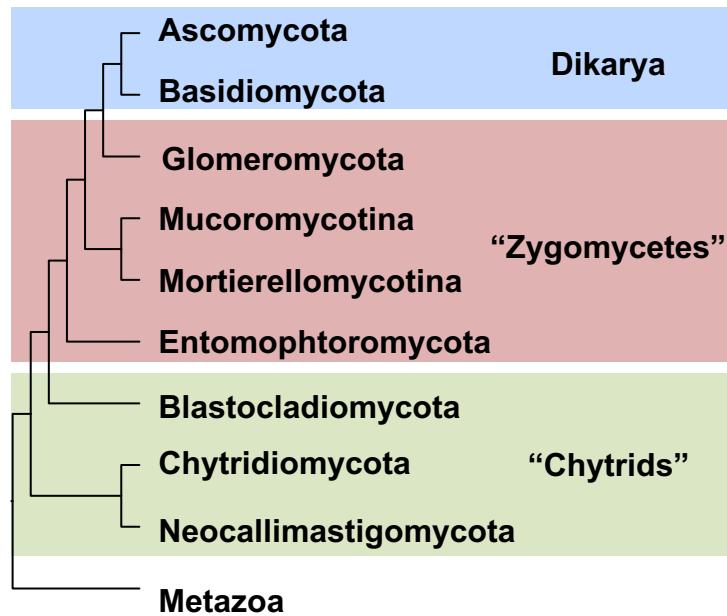
Sexual Interaction – Gametangiogamy

Conjugation of non-differentiated, coenocytic gametangia leads to the formation of zygotes (zygospores), which are associated with a pair of two yoke-shaped suspensors



Mucor moellerii (ex: *Zygorhynchus moellerii*)

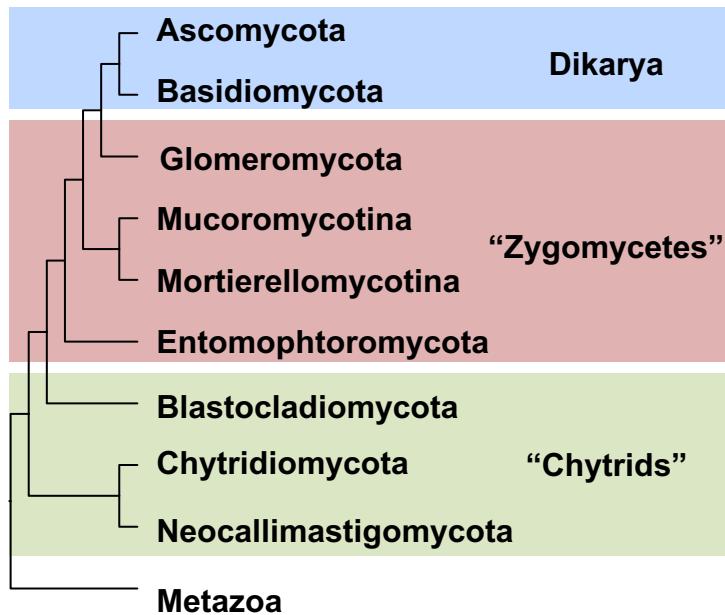
Mucorales



- Food production (Asian cuisine)
- Tempeh, soy sauce etc.



Mucorales



- Ancient terrestrial filamentous fungi
- Mainly ubiquitous saprotrophs
- Food production (Asian cuisine)

Invasive infections	Cases per year
Aspergillosis	200,000
Candidiasis	400,000
Mucormycosis	10,000

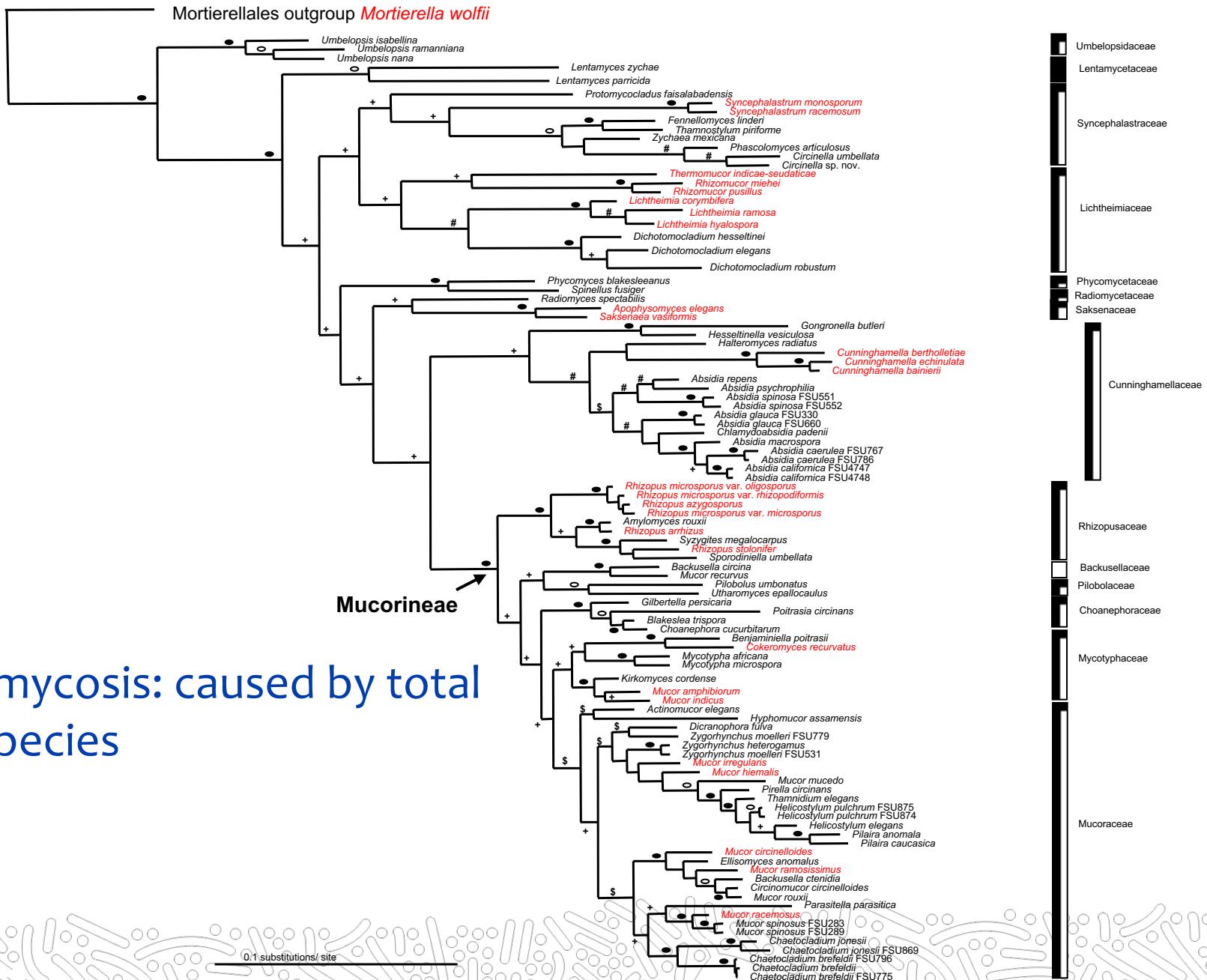


Lidor & Nunley, 1997, N Engl J Med



Petrikkos et al. 2012, CID

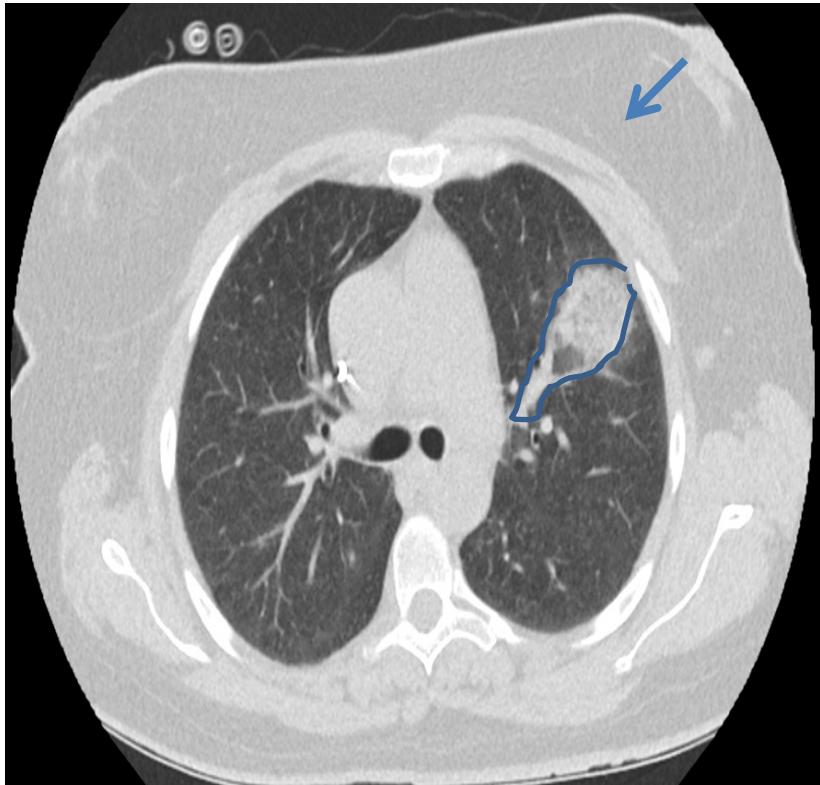
- Emerging fungal infections
- COVID-19 associated
- Fast progress, high mortality rates
- Reliable diagnostics is mandatory for therapy



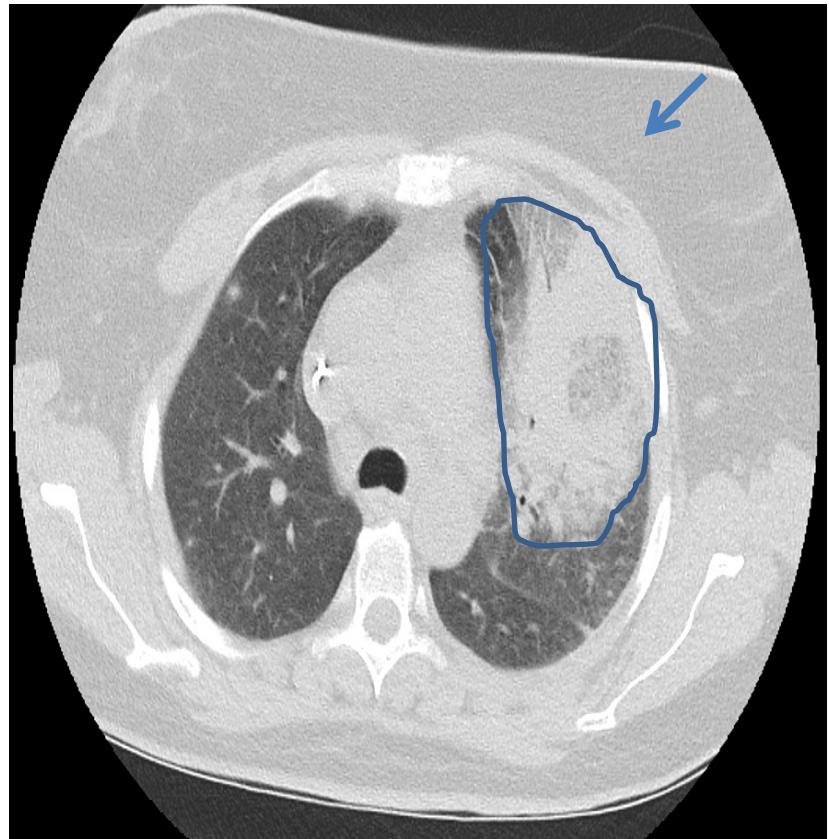
Mucormycosis: caused by total
of 26 species

Mucormycosis: pulmonary infection

Female patient: 38 yrs., admitted to UKJ, KIM II, Int. Oncology, Sept. 2017



$t = 0$



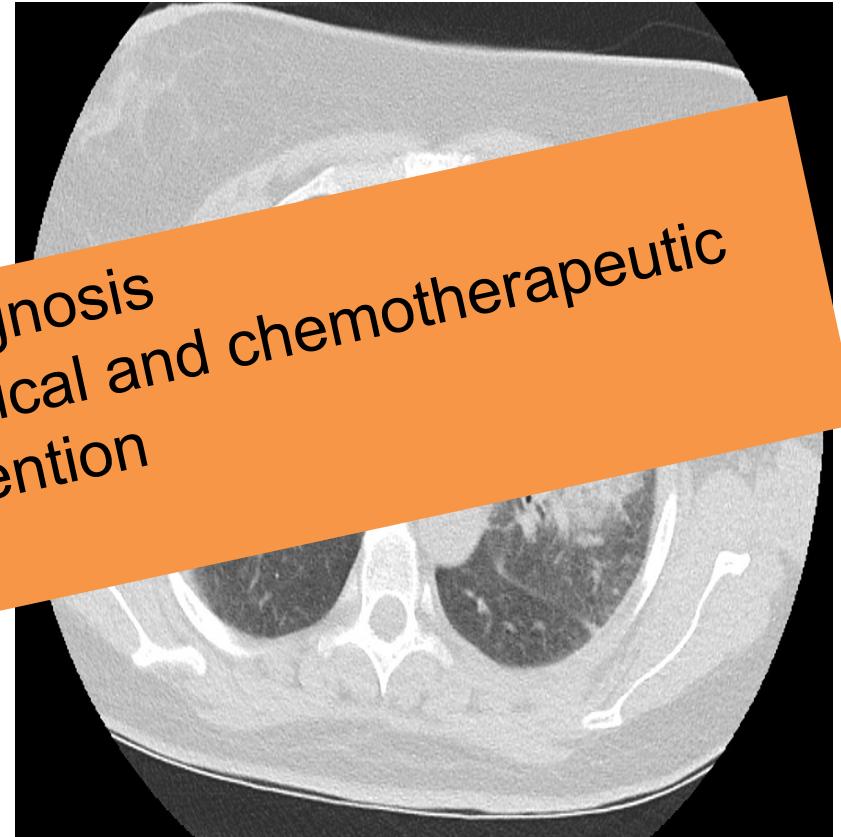
$t = 1 \text{ week later} - > 5x \text{ augmentation of lesion}$

Mucormycosis: pulmonary infection

Female patient: 38 yrs., admitted to UKJ, Sept. 2017



t = 0



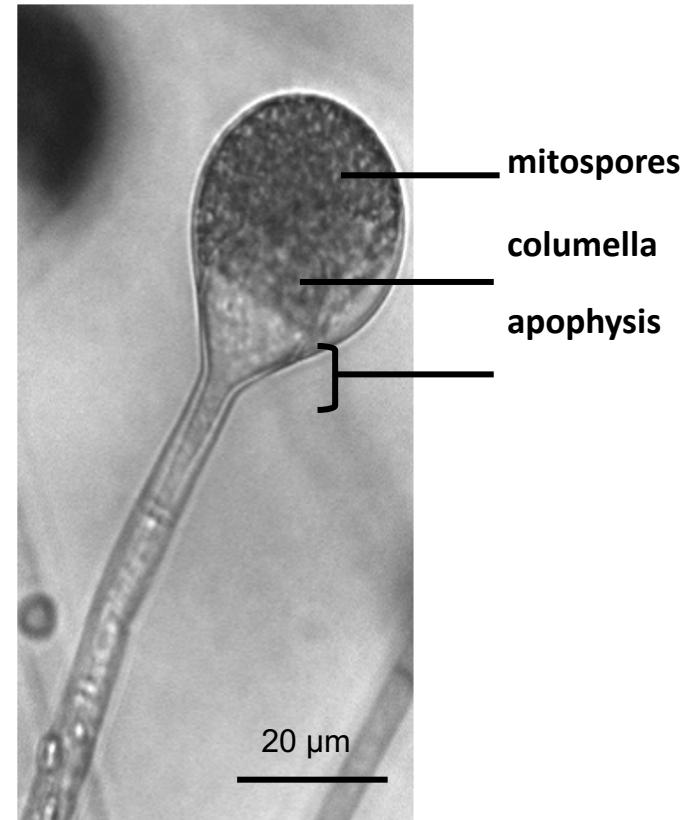
t = 1 week later

Bad prognosis
Survival just with thoraxsurgical and chemotherapeutic intervention

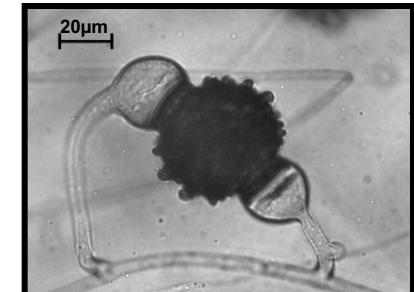
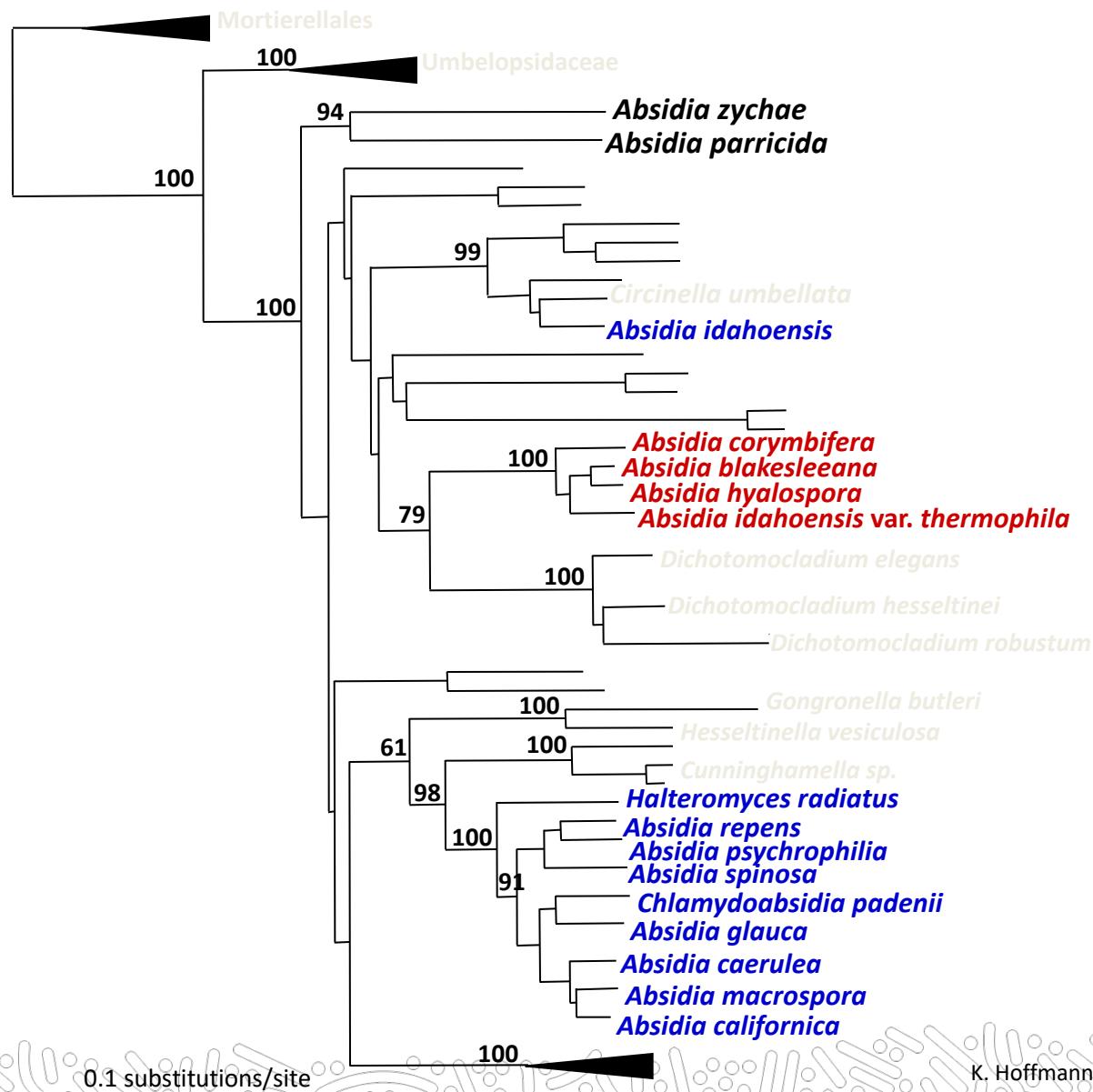
Absidia corymbifera: 1st species reported as causative agent of mucormycosis

- *Absidia corymbifera* (Cohn)
Sacc. & Trotter, in Saccardo,
Syll. fung. (Abellini) 21: 825
(1912)
- Basionym: *Mucor corymbifer*
Cohn in Lichtheim, *Z. klin. Med.* 7: 149 (1884)

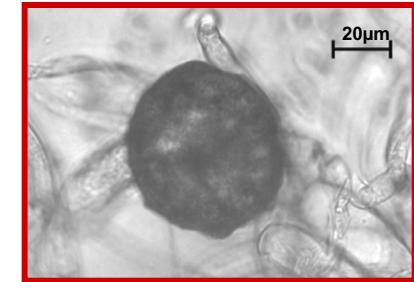
-pyriform, apophysate sporangia



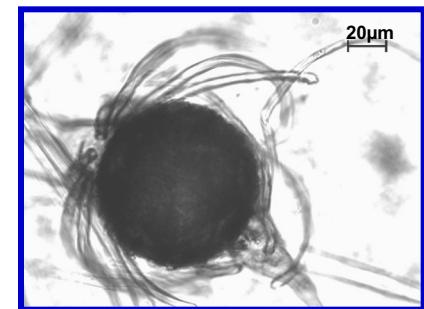
Absidia: polyphyletic origin



Zygospores: *Mucor*-like, papillate

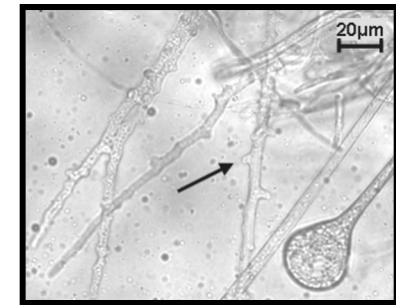
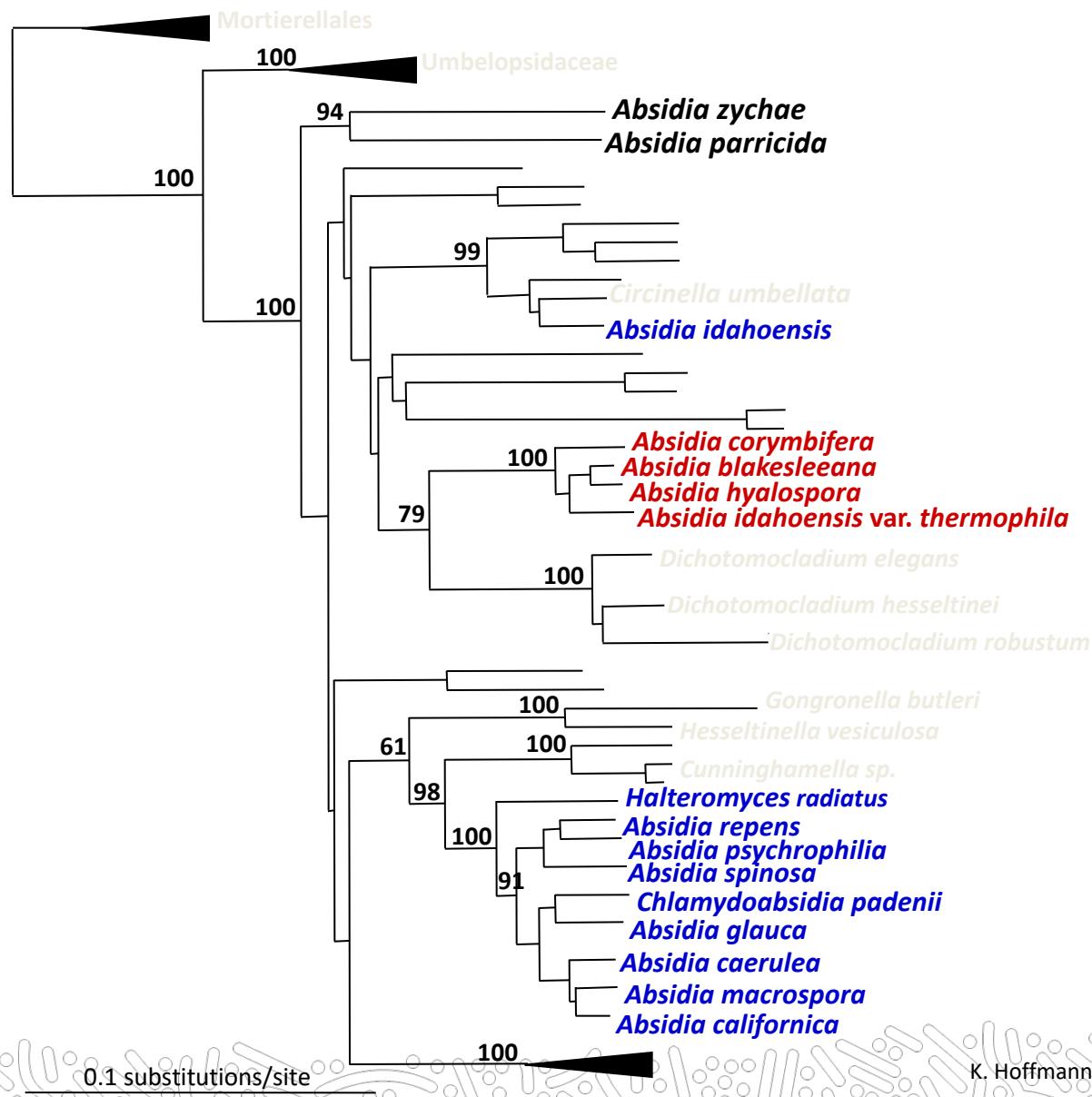


Zygospores: naked, equatorial ring

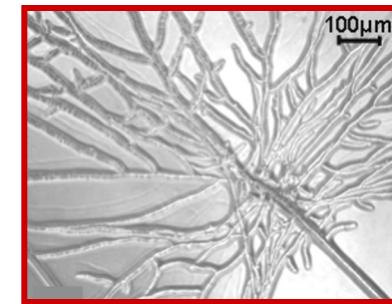


Zygospores: finger-like appendages

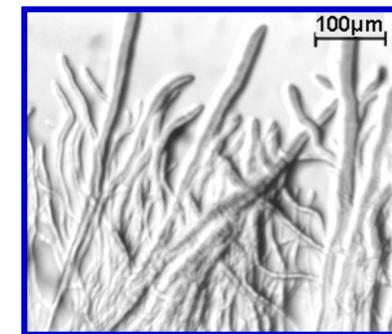
Absidia: polyphyletic origin



Substrate mycelium: sucker-like

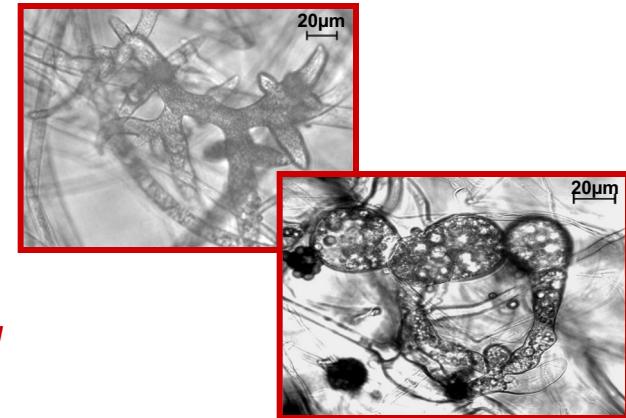
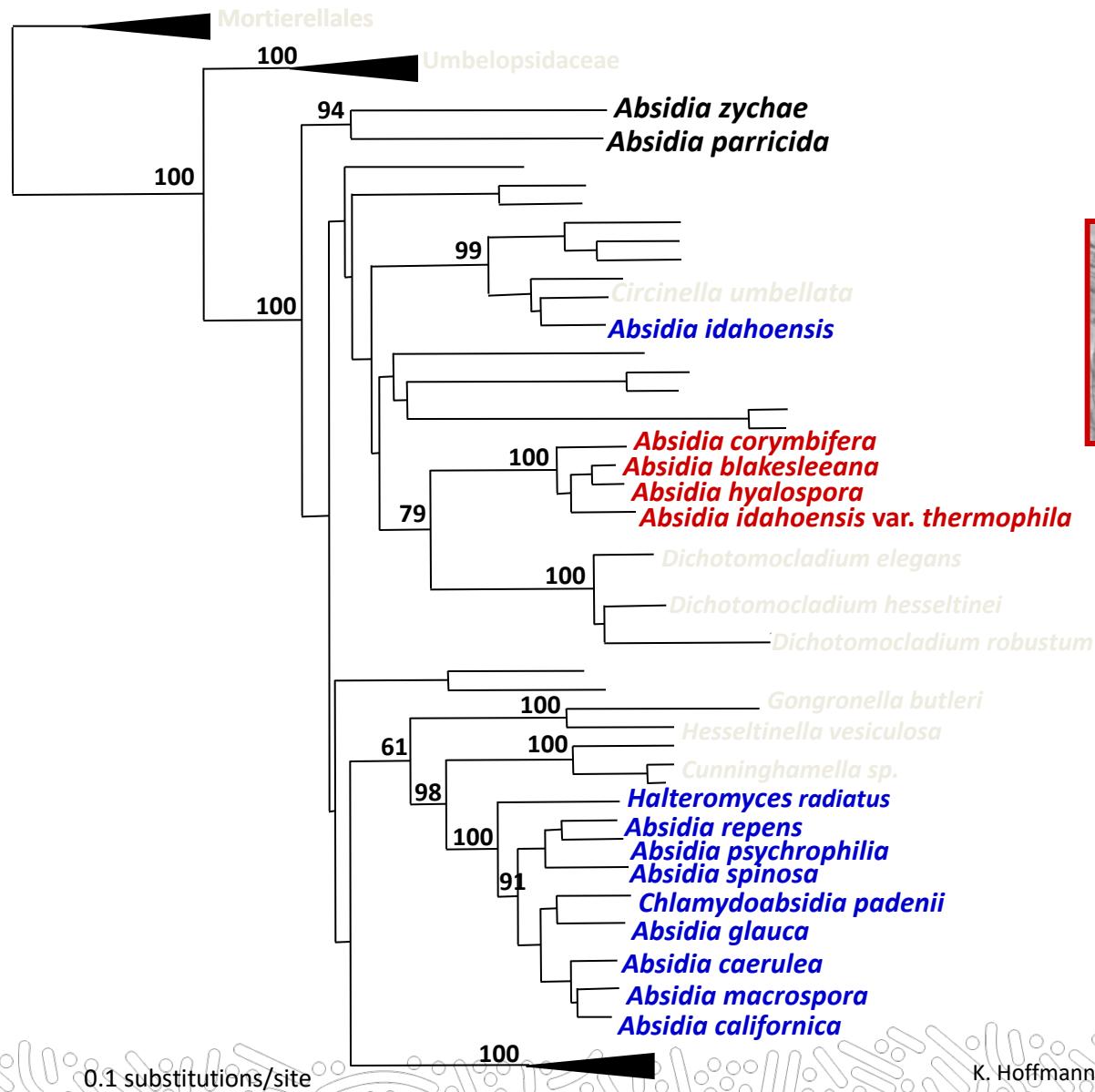


Substrate mycelium: tree-like



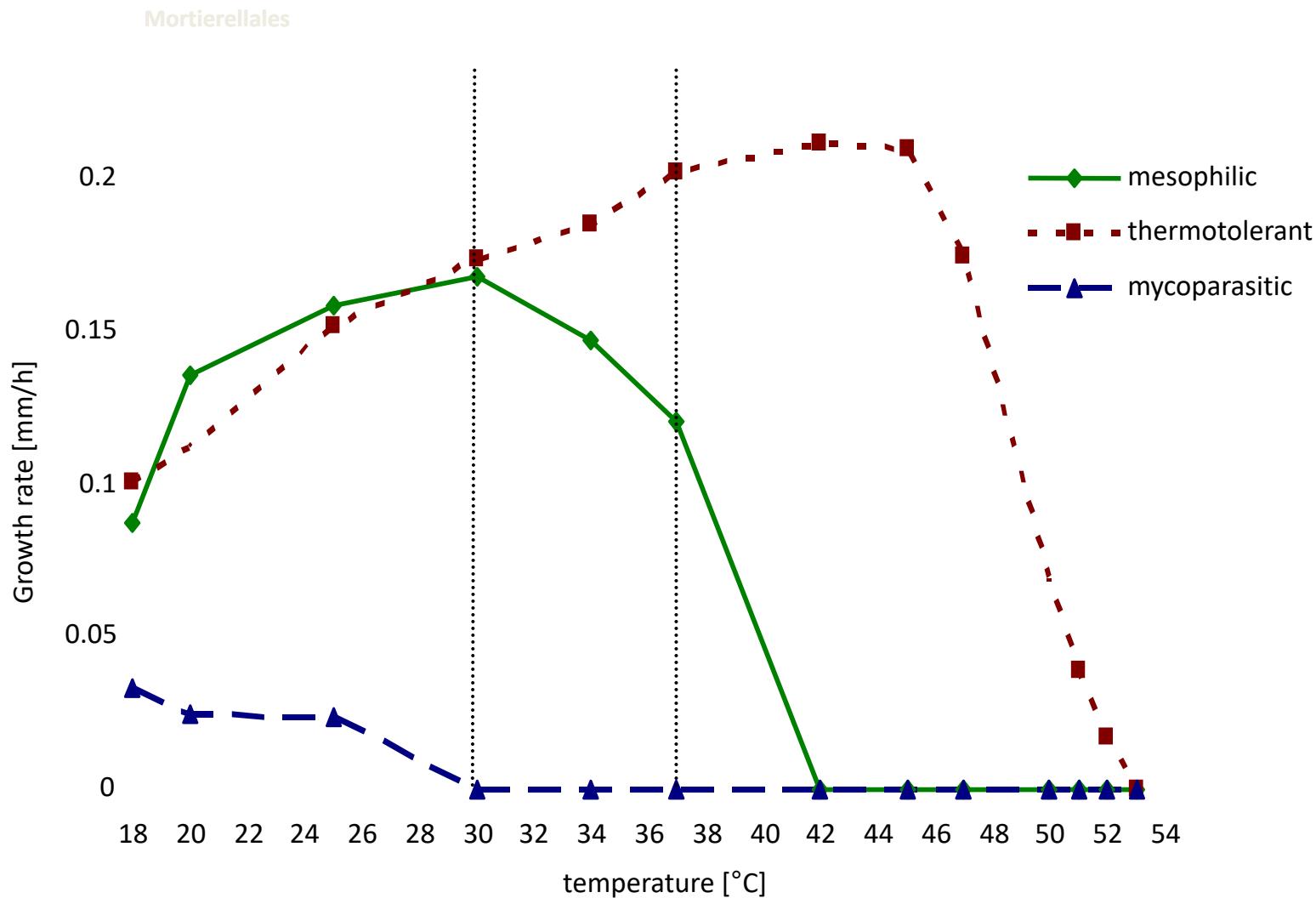
Substrate mycelium: straight

Absidia: polyphyletic origin



Giant cells: species dependent antler-like or bulbous; abundant

Absidia: growth physiology



Signature sequences – Motifs of the nuc ITS 1

	3'-end 18S rDNA	5'-end of ITS1 region
<i>Absidia coerulea</i> FSU 767 (+)	GGATCATTA	CAAATGGCGAGGTGGACGATTATCCGACCCTTCAC
<i>Absidia coerulea</i> FSU 786 (-)	GGATCATTA	CAAATGGCGAGGTGGACGATTATCCGACCCTTCAC
<i>Absidia cylindrospora</i> FSU 309	GGATCATTA	CAAATGGCGAGGTGGACGATTATCCGACCCTTCAC
<i>Absidia cylindrospora</i> FSU 906	GGATCATTA	CAAATGCCGGGGTGGCTCTCTCGGGGAGCTTCT
<i>Absidia glauca</i> FSU 329 (+)	GGTACATTA	CAATGGCBBBBAAAGGCTTGCCCTCTCCCCAAATC
<i>Absidia glauca</i> FSU 330 (-)	GGATCATTA	CAATGGCBBBBAAAGGCTTGCCCTCTCCCCAAATC
<i>Absidia glauca</i> FSU 659 (+)	GGTACATTA	CAATGGCBBBBAAAGGCTTGCCCTTTCCCCAAAT
<i>Absidia glauca</i> FSU 660 (-)	GGTACATTA	CAATGGCBBBBAAAGGCTTGCCCTTTCCCCAAAT
<i>Absidia glauca</i> FSU 661 (+)	GGTACATTA	CAAATGGCBBBBAGAAGGTTGCCCTTTCCCCAAAT
<i>Absidia glauca</i> FSU 662 (-)	GGTACATTA	CAAATGGCBBBBGAAGACTTGCACTTCTCCCCAAAT
<i>Absidia orchidis</i> FSU 1608 (-)	GGTACATTA	CAAATGGCBBBBGAAGATTATACGACCCTTCAC
<i>Absidia orchidis</i> FSU 323 (-)	GGATCATTA	CAAATGTAAGAGTGGATAAGTATAACGAGATATGCT
<i>Absidia repens</i> FSU 939	GGTACATTA	CAAATGCCCTCTGGTTCTTTGGGAGGAGGGCA
<i>Absidia spinosa</i> FSU 550	GGTACATTA	CAGATGTGGGGTGGCCATAGGTCCTTACTTTTAA
<i>Absidia spinosa</i> FSU 551	GGTACATTA	CAAATGGATTGGTTGCTATACGGTAATCCGATTTCARWKGBVD
<i>Absidia parricida</i> FSU 547	GGATCATTA	<u>CATGTTTATGACTGAGCAATTCTCTGAAAAGAGGA</u>
<i>Absidia parricida</i> FSU 917	GGTACATTA	<u>CATGTTTATGACTGAGCAATTCTCTGAAAAGAGGG</u>
<i>Absidia blakesleeana</i> FSU 2684 (+)	GGTACATTA	<u>CTGAGAGGT</u> CTCTTCTGTTCTCCAGTATGTGCAAT
<i>Absidia corymbifera</i> [AB054042]*	GGATCATTA	<u>CTGAGAGGT</u> CTAAAAAACCACTAGTTGGGGTCTCT
<i>Absidia corymbifera</i> FSU 787 (+)	GGTACATTA	<u>CTGAGAGGT</u> AAAAACCGCATAGCGGGATATTCCCT
<i>Absidia corymbifera</i> FSU 788 (-)	GGTACATTA	<u>CTGAGAGGT</u> AAAACCACTAGCGGGAAATCTTCCCT
<i>Absidia corymbifera</i> FSU 938 (+)	GGTACATTA	<u>CTGAGAGGT</u> TATTAAGCCTTGGCGGGTTGGCTAAC

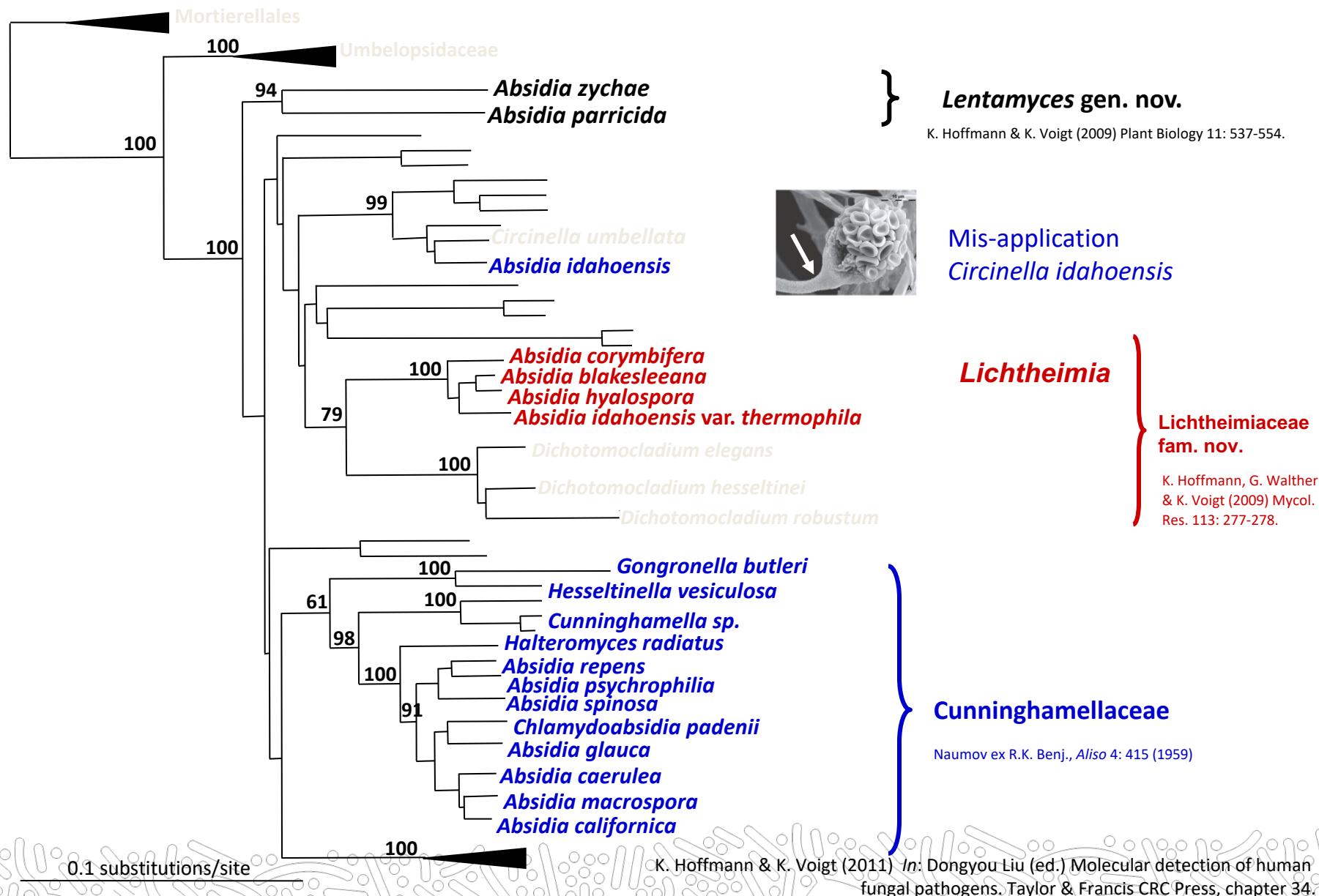
* ... obtained from The National Centre for Biotechnology Information at www.ncbi.nlm.nih.gov

mesophilic

mycoparasitic

thermotolerant

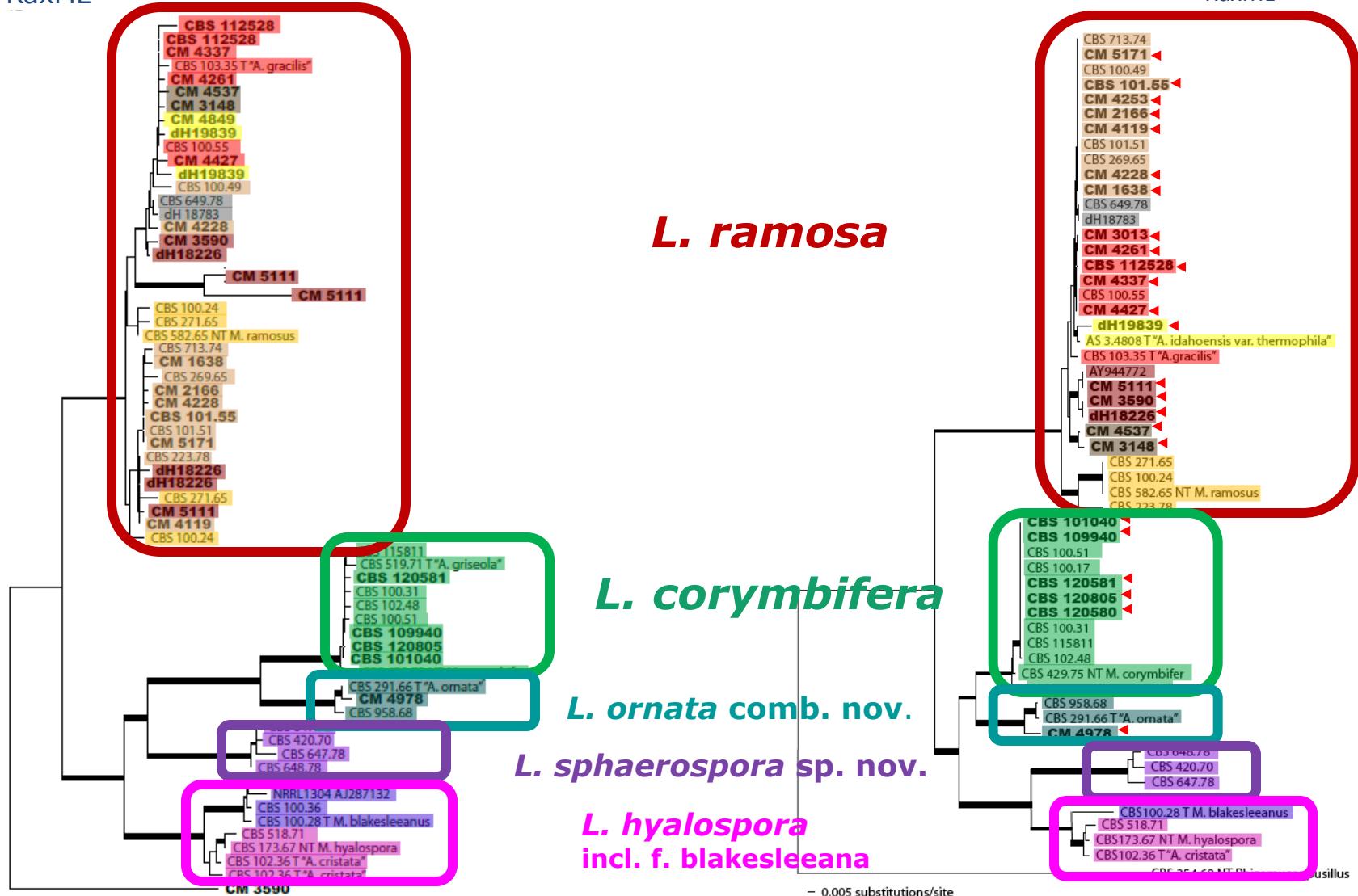
Lichtheimia: the introduction of Lichtheimiaceae fam. nov.



Actin, Nj
Bt: Nj;
RaxML

Phylogenetic species recognition

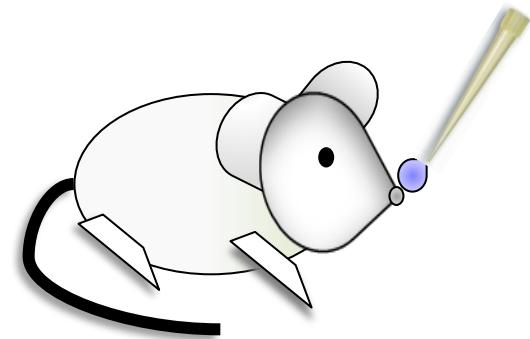
ITS, Nj
Bt: Nj;
RaxML



Establishment of a mouse model

Isolate	Designation in strain	Designation in this study	Source
		collection	
<i>L. corymbifera</i>	JMRC:FSU:09	Reference	Environme
	682	strain	nt
<i>L. corymbifera</i>	JMRC:FSU:10	LCJ5	Environme
	164		nt
<i>L. corymbifera</i>	JMRC:FSU:10	LCJ3	Human
	061		
<i>L. corymbifera</i>	JMRC:FSU:10	LCJ9	Human
	240		
<i>L. ramosa</i>	JMRC:FSU:11	LCJ22	Human
	788; Luo <i>et al.</i>		
	Schulze <i>et al.</i> (2017) Ketoacidosis alone does not predispose to mucormycosis by <i>Lichtheimia</i> in a murine pulmonary infection model. <i>Virulence</i> . (25)		
<i>L. ramosa</i>	JMPC:FSU:06	LCJ17	Environme

Intranasal infection



L. corymbifera type strain

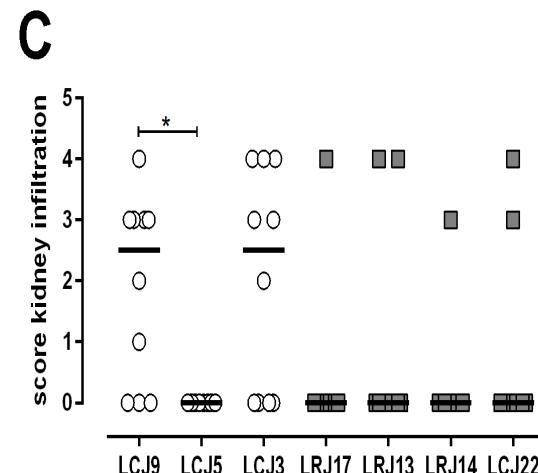
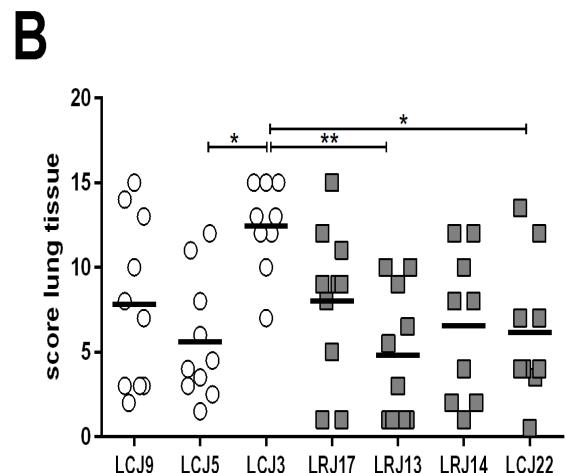
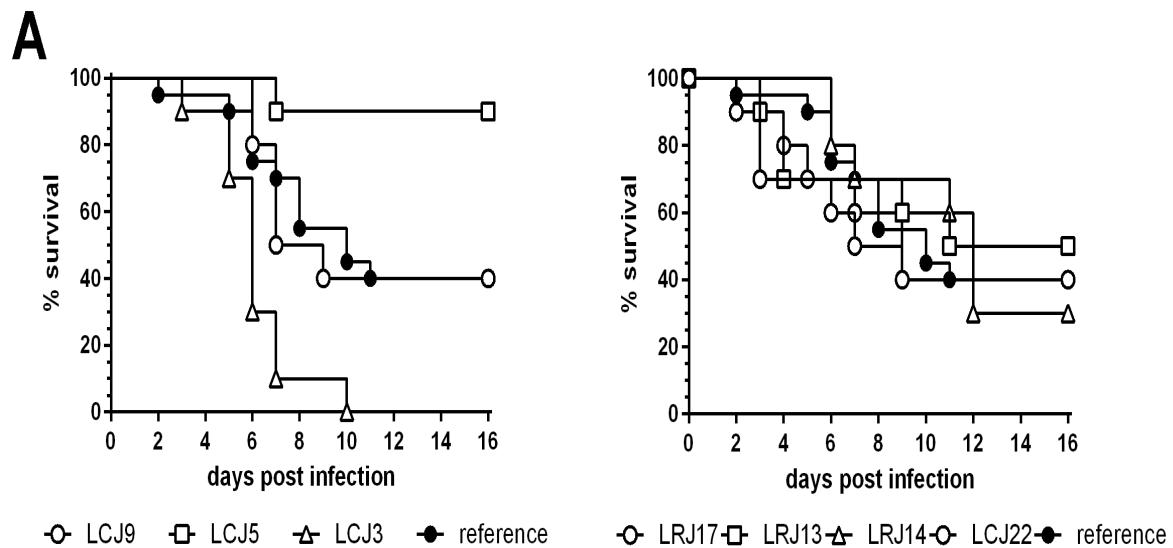
- average and reproducible virulence potential (clinical species)
- average *in vitro* phenotype (growth, stress resistance)
- Mice: Immunocompetent vs. streptozotocin vs. cortisol
- n=5-10 mice/group/experiment

Strain specific virulence of different *Lichtheimia* isolates in a mouse model of pulmonary infection

(A) Survival of immunocompromised female CD-1 mice intranasally infected with 5×10^6 spores of *L. corymbifera* and *L. ramosa* strains. Combined data from two independent experiments ($n=5$ mice/group/experiment). $**p < 0.01$, determined by Log-rank (Mantel-Cox) test.

(B) Analysis of histological scores from lung and kidneys

(C) ($n=8-10$ mice/group). Statistical significance was calculated using one-way ANOVA with a Tukey's multiple comparison test, $*p < 0.05$ and $**p < 0.01$.



Summary

- Proper diagnostics requires many isolates
- Growth kinetics is indicative for the systematics of *Lichtheimia*
- Isolates of *Lichtheimia* differ in virulence in mouse
- Haploid genomes are common, diploid genomes are exceptional in *Lichtheimia* due to decreased stress tolerance and virulence



Thank you

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Hyang Burm Lee



Thank you

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