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Phyto-pharmacological review of genus manilkara

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Abstract

Manilkra is a pantropical genus in the family Sapotaceae. Many species from this genus are economically important and yield quality timber, edible fruits and useful latex. The plants from Manilkara species are extensively distributed all over tropical and semitropical areas like Africa, Australia, Asia, Madagascar and Latin America, as well as islands such as Pacific and in the Caribbean. Phytochemical studies of different species showed the presence of triterpenoids, phenolic compounds and saponins as major phytoconstituents which were correlated with various pharmacological activities. Because of high diversity in chemical constituents and their biological activities, genus Manilkara can serve as a potential medicinal resource for drug discovery. The present review includes traditional use, phytopharmacological studies carried out on few medicinal plants from genus Manilkara. As a promising source of new bioactive compounds. The present review can provide baseline for future research studies.

Keywords: Manilkara, traditional use, phytochemicals, pharmacological potentials

1. Introduction

Medicinal plants and their derivatives play major role in treatment of diseases in spite of advances in chemical technology. Recently, there has been tremendous evolution in plant derived drugs in the world due to the realization of toxicity associated with synthetic drugs. People around the world are increasingly aware of the fact that drug from natural sources are safer. Therefore, there has been a tremendous increase in demand for drugs derived from plants.

The Genus Manilkara Adans. (Sapotaceae) is consisting of around 79 species, widely spread over major tropical regions of the world (30 South and Central American, 35 African and 14 Southeast Asian). It is mainly found in wet tropical forest, but most of the African species exist in dry forest, while many Asian species are present in coastal areas and on limestone. According to Pennington's (1991) classification, Sapotaceae consists of five tribes, Manilkara belongs to one of the three subtribes of Manilkarinae of the tribe Mimusoepae and is characterised by having perianth parts in multiples of three; a calyx of two whorls of three sepals, six petals, six stamens and six staminodes or absent; fruit indehiscent; seed scar usually elongate; basiventral^[1].

The various parts of the plants of genus Manilkara have been used in folklore medicine and ethnomedicine for therapy of many human ailments^[2]. These bioactivities are mainly due to the existence of certain classes of secondary metabolites like triterpenoids, flavonoids, saponins and so on^[3].

Some plants from the genus Manilkara have received considerable interest due to their extremely high economical, medicinal and nutritional values. Only seven species such as *M. zapota*, *M. hexandra*, *M. kauki*, *M. obovata* (synonym - *M. argentea*), *M. subseria*, *M. bidentata*, *M. pellegriniana* from Manilkara genus have been scientifically investigated for their pharmacological activity and phytochemical constituents. The most popular, widely utilized and highly investigated species in this genus are *M. zapota* and *M. hexandra* and other species in the genus are less unexplored^[4, 5].

Present review includes list of traditional uses, isolated phytochemicals and pharmacological activities carried out on crude extracts, fractions and isolated compounds from different species of genus Manilkara. We aim to provide a way forward for future research studies mainly on unknown species from Manilkara.

2. Traditional uses and geographical distribution

Details of traditional uses and distribution of plants from genus Manilkara and related references are given in Table 1.

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Table 1: Geographical Distribution and Traditional Uses

Species	Distribution	Traditional use	References
<i>M. Hexandrum</i>	Upper Gangetic plain, central and south India, Gujarat and Rajasthan, Vidarbha region of Maharashtra, and Madhya Pradesh	Fruit-aphrodisiac, useful in leprosy, extract of the leaves is given for vaginal discharges (Ayurveda), cures biliousness, good for the heart Bark-aphrodisiac, tonic to the body and heart, cures vomiting, bronchitis, appetiser Seeds-cure ulcers, opacities in the cornea (Yunani)	[2, 69, 70]
<i>M. Kauki</i>	A native of Malay peninsula, occasionally planted in Indian gardens	Root and bark-astringent, given in infantile diarrhoea. Leaves--treatment of Beri-Beri, tumour, Milk (latex)-used in inflammation of the ear and in conjunctivitis. Seeds-used in ophthalmia, employed internally as a tonic, febrifuge. Treatment of leprosy, thirst, delirium and disorder of the many secretions, used as anthelmintic	[2]
<i>M. zapota</i>	Widely seen in India, Southern Mexico, Caribbean and Central America as well as in South East Asia. <i>M. zapota</i> fruit is very popular in Malaysia, Thailand, Singapore, Cambodia and Indonesia	Fruit -reduces inflammation and pain in gastritis, anti-spasmodic agent Fruits and crushed seeds – have diuretic property helps in preventing oedema, prevent kidney and bladder stones formation. Latex -useful as a material for filling tooth cavities. Bark -- tea made from bark to treat dysentery, constipation and piles, Bark and fruit decoction is used for fevers and diarrhoea.	[5,65]
<i>M. obovata</i> Synonym- <i>M. argentea</i>	Widely distributed in Africa	Useful for treatment of cardiovascular ailments. Bark-African countries bark preparations are used for treatment of stomach troubles, skin and mucosal problems, Root-preparation from roots can be used as laxative	[15, 50, 71]
<i>M. subseria</i>	Widely found on the sand bank of eastern Brazil.	Used medicinally in Brazil also as food and timber	[2, 33]
<i>M. bidentata</i>	Widely found in northern South America, Central America and the Caribbean	Used to treat many diseases. Stem - to cure dysentery and nausea; Fruits - to treat constipation Leaves- treatment of paralysis of limbs	[3, 72]

3. Pharmacological activity

Various crude extracts and few isolated compounds from different parts of plants from genus *Manilkara* have been widely screened for their biological activity. According to the literature, plants of the genus *Manilkara* are potential sources of antibacterial, antioxidant, hepatoprotective, anticancer, hypolipidemic, hypoglycemic and antiarthritic activity.

3.1 Antioxidant activity

Kaneria *et al* reported the strong antioxidant activity of methanol and acetone extract of leaves with high phenolic content from 2 different studies supports the idea of direct correlation between phenolic content and strong antioxidant activity of zapota leaves [6, 7]. The cold ethanolic extract of leaves and Isolated compound myricetin-3-o- α -L-rhamnoside from crude methanolic extract of leaves were found to be having good antioxidant activity [8, 9]. Methanolic extract flowers showed higher dose dependent percentage inhibition in 2, 2-diphenyl-1-picrylhydrazyl scavenging assay than aqueous extract with IC₅₀ values 1.97 μ g/ml and 4.22 μ g/ml respectively [10]. Among ethanolic and methanolic pulp and peel extract of sapota, ethanolic peel extract showed the presence of highest flavonoid and phenolic content with maximum multiple radical scavenging activity with least IC₅₀ value [11]. Ethanolic bark extract of *M. zapota* was found to be good antioxidant at different concentration [12]. Aqueous methanolic (80%) fruit and seed extract of *M.*

hexandra subjected to six different antioxidant assays pointed that *M. hexandra* fruits as better source of antioxidants [13].

Suman and Sanjib carried out four different antioxidant assays on different fractions of leaves of *M. hexandra* and concluded that methanolic extract fraction has strong antioxidant activity with high content of phenolics and flavonoids [14].

Akosung *et al* reported the xanthenes and coumarins for the first time in *Manilkara*. Also isolated new friedelane triterpene lacefriedelic acid and the new prenylated xanthone lacexanthone from extracts of leaves and roots of *M. obovata* showed promising antioxidant activity which is beyond the activity measured for BHA [15].

The stem barks and leaves of *M. kauki* reported the good antioxidant activity with highest amount of phenolic compounds and flavonoids compare to fruits seeds and woods [16].

3.2 Antiurease activity

The isolated compound canophyllol and lupeol from a twig extract of *M. obovata* show better urease inhibition compare to thiourea. It may find promising application in food industry [15].

3.3 Elastase and tyrosinase inhibition

Isolated compound myricetin-3-o- α -L-rhamnoside from methanolic extract of *Zapota* leaves reported moderate elastase and tyrosinase inhibition activities [9]. Study

conducted by Sirinada and Chanya showed the significant anti tyrosinase activity of ethyl acetate extract of *M. kauki* stem [16].

3.4 Microbiological activity

Aqueous extract of the leaves of *M. zapota* was noticeably more potent in hindering the growth of against 10 Gram positive, 12 Gram negative bacteria and fungus *Candida tropicalis* [17]. Ethyl acetate extract of both stem bark and leaves of *M. zapota* were found to be good antibacterial/antifungal agent against tested pathogenic Gram positive and negative bacteria and fungi such as *Aspergillus flavus*, *Fusarium* sp and *Vasianfactum* sp [18]. Methanolic extract of *M. zapota* leaves was effective against five bacterial strains namely *Bacillus subtilis*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Salmonella typhimurium* and *Enterobacter aerogenes* and acetone extract of Zapota leaves has highest inhibitory effect on *K. pneumoniae* [6, 7]. Mulvaney *et al* screened ethanolic leaves extract of zapota, its fractions and isolated components for antimicrobial and anthelmintic activity against *Candida albicans*, *Trichophyton rubrum*, *staphylococcus aureus* and *Strongyloides venezuelensis* which indicated *M. zapota* as natural drug for the treatment for microbial and strongyloidiasis drug resistant infections [19].

Among different solvent fractions, acetone, ethyl acetate fractions of *M. zapota seed* showed higher zone of inhibition against *S. aureus*, *E. Coli*, *Haemophilus* sp, *Yersinia* sp, *B. subtilis*, and *Enterobacter faecalis* by agar well diffusion method compare standard drugs whereas aqueous fraction showed inhibition only against *Haemophilus* sp [20].

Acetone extract of seeds of *M. zapota* showed significant antibacterial activity in disc diffusion and broth dilution methods [21].

The antimicrobial study of aqueous root extract of *M. zapota* against *Staphylococcus aureus* and *Escherichia coli* revealed that root can be used to fight infections against both Gram positive and negative bacteria [22]. Methanolic flower extract *M. zapota* reported maximum zone of inhibition against *B. Subtilis* (29mm) followed by *S. aureus*, *P. aeruginosa* and *S. Typhi* (27.5mm) and aqueous extract showed maximum zone of inhibition for *S. typhi* (28.5 mm) followed by *S. aureus*, *B. subtilis* and *P. aeruginosa* [10]. Rakholiya *et al* reported that fruit peel of *M. zapota* showed good antifungal activity and more active against Gram negative bacteria and. So it has been proved to be a good source of natural antimicrobial compound [23].

Evaluation of antibacterial activity of methanol: water (7:3) extract of stem bark of *M. hexandra* was carried out by disk diffusion method. The sample showed growth inhibitory activity against *Staphylococcus aureus* at ug/ml (7mm) and 150 ug/ml (8mm) and against *Bacillus subtilis* (7 mm) [24].

The optimized topical gel prepared from methanolic and hydroalcoholic leaf extracts of *M. hexandra* showed good antibacterial activity against the strain *Klebsiella pneumoniae* [25]. Study conducted by Mahida *et al.* showed methanol extract of *M. hexandra* effectively inhibited the growth of multidrug resistant *Staphylococcus aureus* and *S. epidermidis* [26]. The different studies revealed that acetone seed extract, methanolic extract of leaves, roots, aqueous extracts of leaves, aerial parts of *M. hexandra*, were found to be having good antimicrobial activity [27-31].

Antifungal activity of some plant extracts was tested against the *Solanum melongena* damping off pathogen, *Pythium*

aphanidermatum in PDA cultures. Among that cold water extracts of *M. Kauki* showed (78.8%) inhibitory effect [32].

Fernandes *et al* reported hexanic extract from fruits of *M. subseria* showed good antibacterial activity against *staphylococcus aureus* ATCC25923 and weak cytotoxicity in Vero cells, may be due to high amount of beta and alpha amyryn caproates and caprylates [33].

3.5 Anticancer activity

The methanolic extract *M. zapota* fruit (MESF) showed good cytotoxicity *in vitro* by affecting cell viability in NALM6, human and mouse breast cancer cell lines such as EAC, MCF7 and T47D. This study stated that Zapota fruit induces cytotoxicity in cancer cells by activating intrinsic pathway of apoptosis and *in vivo* Ehrlich ascites carcinoma model showed significant inhibition of tumour growth and a three-fold increase in life span of tumour bearing animals compared to untreated tumour mice. This study revealed that zapota fruit can be used as potential cancer preventing agent [34].

Activity-guided fractionation of a methanol extract from the fruit of *M. zapota* resulted in the isolation of two new antioxidants, in which 4-O-galloyl chlorogenic acid displayed cytotoxicity in the HCT-116 and SW-480 human colon cancer cell lines with IC₅₀ values of 154 and 134 µM, respectively [35].

Two new polyhydroxylated pentacyclic triterpenes from *M. pellegriniana* were investigated for cytotoxic effect on SPC212 (human mesothelioma), A549 (human non-small cell lung cancer (NSCLC)), HepG2 (hepatocarcinoma), DLD-1 (colorectal adenocarcinoma) cell lines.

Amongst compound 1β, 2α, 3α, 5, 19α, 24-hexahydroxyurs-11(12), 20(30)-dien-28-oic acid was active against all cell lines and showed promising cytotoxic effect towards SPC212 lung cancer cells with IC₅₀ value of 0.52 µM [36].

Antitumor effect of ethyl acetate extract of *M. zapota* fruits (EEFM) was carried out against Ehrlich ascites carcinoma (EAC) in Swiss albino mice at the doses of 50 and 100 mg/kg body weight. Treatment with EEFM at 100 mg/kg body weight indicated a significant increase in the survival time, decrease in the viable tumour cell count, weight gain in the EAC tumour hosts, improvement in the altered haematological parameters, like haemoglobin content, RBC and WBC count of the tumour bearing mice. It also restored altered biochemical (SALP and SGOT) parameters during tumour progression. In same study, EEFM showed potent cytotoxicity in the brine shrimp lethality bioassay, against brine shrimp nauplii (*Artemia salina*) with LC₅₀ of 3.06 µg/mL, against standard ampicillin trihydrate which showed LC₅₀ of 7.21µg/ml. This study indicated the both antitumour and cytotoxic potential of the zapota fruit [37].

Tan and his co-workers studied the apoptosis-inducing activity of water extract of *M. zapota* leaf in HepG2 cells. This study revealed that leaf water extract causes early apoptosis in hepatocellular carcinoma (HepG2 cells) via modulation of multiple signalling pathways like intrinsic mitochondrial pathways leads suppression of metastasis. All the data recorded indicated that *M. zapota* leaf water extract has noteworthy apoptotic potentials via the regulation of ERK1/2/Akt1/JNK1 transcriptional activity [38].

The ethyl acetate extract of stem bark of *M. zapota* showed significant antitumour efficacy in Ehrlich ascites carcinoma model. Intraperitoneal administration of this extract reduced viable EAC cells, increased the survival time, and restored

altered haematological parameters [39].

In another study Rashid *et al* used same model to evaluate *in vivo* antitumor activity of ethyl acetate extract of the leaves of *M. zapota* and isolated compound Erythrodiol. Leaf extract increased the life span, weight of the tumour bearing mice, decreased the tumour bearing cell and improved the altered haematological parameters. Erythrodiol reduced the viable tumour cell count by 70.8% in comparison with the untreated control [40].

3.6 Anti-inflammatory, antipyretic, Analgesic activity and antinociceptive activity

The petroleum ether fraction of ethanolic extracts of *M. zapota* leaves showed maximum body temperature lowering effect (36.86 °C) at 4th hr and the ethyl acetate fraction showed anti-inflammatory effect [41]. In another study Ethanol extract and its fractions showed prominent peripheral antinociceptive activity and significant central analgesic activity at 400 mg /kg [42].

Crude methanolic extract of bark and ethyl acetate extract of *M. zapota* leaves has good anti-inflammatory activity [43, 44].

The acetone fraction of *M. hexandra* seed containing the crude saponin mixture possessed a significant inhibitory activity against LPS (Lipo-Poly Saccharide)-induced nitric oxide method indicating a significant anti-inflammatory activity [45].

Study conducted on *M. bidentata* resin extract and isolated fractions for anti-inflammatory activities showed decrease in IL-8 and IL-1 β pro-inflammatory cytokines, suggested the use of bidentata resin extract as an anti-inflammatory and anti-aging substance for pharmaceutical and cosmetic industries [3].

The evaluation of analgesic activity of zapota leaves was carried out by using acetic acid-induced writhing test in mice concluded that *M. zapota* leaves possess significant analgesic activity and supported its traditional use as analgesic [46]. The ethanolic extracts of *M. zapota* leaves showed analgesic activity at doses of 200 mg/kg [47]. Another study ethanolic extract of whole plant of *M. zapota* showed dose dependent analgesic activity, suggested the benefits of whole plant of zapota for treating pain and inflammation [48].

3.7 Hepatoprotective activity

In two different studies the crude ethanol extract of bark and cold ethanolic leaf extract of *M. zapota* were tested *in vivo* CCL4 liver damage in rats hepatoprotective model. Results indicated the restoration of altered serum marker enzymes, total bilirubin, total protein and liver weight when compared to standard. These studies suggested that hepatoprotective activity bark and leaves may be due to their antioxidant potential and presence of flavonoids, carotenoids and ascorbic acid [8, 12].

Administration of lyophilized zapota fruit extract in CCL4 intoxicated rats resulted in prominent dose-dependent reversal of elevated serum biomarkers, bilirubin and abnormal lipid profile at (250 and 500 mg/kg proved its hepatoprotective and lipid-lowering effects [49]. Investigation of hepatoprotective activity of methanolic and petroleum ether extracts of *M. obovata* seed against CCL4 induced hepatic damage in rats reported their significant antioxidant and hepatoprotective activities [50].

3.8 Hypocholesterolaemic effect

Aqueous extract of *M. zapota* leaves showed a significant decrease in cholesterol level close to that of atorvastatin [51].

3.9 Hypoglycemic activity

Alcoholic and aqueous extracts of the *M. zapota* leaves showed the significant decrease in the blood glucose level in hyperglycaemia induced rats but not in normal rats compared to Metformin [51].

The hypoglycaemic activity of petroleum ether extracts of leaves and methanol extracts of seeds of *M. zapota* was tested in alloxan induced diabetes model. The oral glucose tolerance test after 15, 30, 60, 90 and 120 min of administering 2mg/body weight of glucose noted the decrease in blood glucose level in a promising way, indicated hypoglycaemic potency of the leaves and seeds of *M. zapota* [52].

Anti hyperglycaemic activity of 50% hydroalcoholic bark extract of *M. hexandra* out by administering 250 mg/kg or 500 mg/kg body weight of extract orally to the streptozotocin induced diabetic rats once daily for 21 days. Result showed that extract has significant antidiabetic activity and good hypolipidemic activity in diabetic conditions [53].

Another study Nimbekar *et al* investigated the hypoglycaemic activity of the methanolic extracts of *M. hexandra* on normoglycemic and alloxan induced diabetic rats. Methanolic extract of *M. hexandra* at (400mg/kg body weight) showed effective hypoglycaemic activity supports the folklore use in the management of type II diabetes [54].

3.10 Antidiarrhoeal activity

Ganguly *et al* tested antidiarrhoeal activity of crude ethanolic extracts of *M. zapota* leaves and it's different fractions such as petroleum ether, carbon tetra chloride and ethyl acetate by castor oil-induced diarrhoeal model indicated the significant antidiarrhoeal activity of ethanol extract (200 and 400 mg/kg) by inhibition of defecation by 53.57 and 60.71%, respectively compared with that of loperamide 71.42% [42].

3.11 Anti-arthritis Activity

Investigation of *M. zapota* ethanolic leaf extract for anti-arthritis activity using *in vitro* protein denaturation inhibition model showed good anti arthritic activity as compared to standard drug acetyl salicylic acid at concentration of 100 mcg/ml and 250 mcg/ml [55].

In another *in vitro* antiarthritic evaluation of methanolic and hydroalcoholic leaf extracts of *M. hexandra* by protein denaturation method and proteinase inhibition method, both the extract inhibited the denaturation of bovine albumin and proteinase enzyme in dose dependent manner against standard drug diclofenac sodium suggested that activity is due to their ability to control production of auto-antigen [56].

3.12 Corneal wound healing

Ethanolic extract of the leaves of *M. zapota* showed corneal wound healing activity on Bovine eyes culture / organ culture suggested that may be due to the occurrence of bioflavonoids and its antioxidant properties [57].

3.13 Immunostimulatory activity

The polysaccharides obtained from *M. hexandra* bark was

tested for Immunostimulatory effect showed dose dependent effect may be due to stimulating macrophage function [58].

3.14 Antifertility Activity

The aqueous powdered drug (2gm/body weight) of seeds of *M. hexandra* showed marked decrease in sperm count in albino male rats in an experiment and proved to be good anti fertility agent [59].

3.15 Antiulcer Activity

Effects of the flavonoid rich fraction of ethyl acetate extract (extract A3) of stem bark of *M. hexandra*, was investigated in experimental animals for ethanol induced, ethanol-indomethacin induced and pylorus ligated gastric ulcers. On administration, extract A3 inhibited the formation of gastric lesions induced by ethanol and pylorus ligation, decreased the ethanol induced vascular permeability, reduced the lipid peroxidation in experimental animals. There was increased mucus production and glycoprotein content in animals pre-treated with extract indicated the protective action of the extract A3 [60].

Another study ether, ethyl acetate and aqueous fractions of acetone extract showed significant inhibition of HCl-ethanol induced gastric ulcer. Amongst ethyl acetate fraction found to be very effective. It sped up the healing of chronic gastric ulcer induced by acetic acid (100mg/kg) and showed protective action in cysteamine -induced duodenal ulcer by reduction in total lesion area and score for intensity [61].

3.16 Inhibition of SARS-CoV-2 main protease activity

Metabolic profiling of secondary metabolites of methanolic leaf extract and ethyl acetate bark extract of *M. hexandra* using LC-HRESIMS (Liquid chromatography-high resolution electrospray ionization mass ionization spectrometry) leads to elucidation of eighteen compounds. Amongst major components were polyphenolics, flavonoids and triterpenes. Flavonoids like rutin, myricitrin, mearnsitrin, and quercetin 3-o- β -D-glucoside showed considerable SARS-CoV-2 main protease inhibitor activity. Amongst rutin showed highest inhibition. It can be promising lead against SARS-CoV-2 pandemic [62].

3.17 Pesticidal Activity

Fernandes *et al* reported that hexane soluble fraction from ethanolic fruit extract of *M. subseria* and its triterpenoids were active against cotton pest *Dysdercus peruvians* [63]. Insecticidal nano emulsion prepared from polar fraction of fruits of *M. subsericea* (5%) showed mortality in *D. peruvians* [64].

3.18 Formulation of fruit extract for cosmeceutical application

Nano emulsion prepared using ethyl acetate peel extract as active constituent was subjected to accelerated stability and cytotoxicity study and it indicated the extract as non-toxic and safe to be used as active ingredient in cosmeceutical application against skin aging [65].

Methanolic extract and its six isolated phenylethanoids from *M. zapota* seeds were screened for antioxidant activity using DPPH assay. Methanolic crude extract exhibited significantly high antioxidant values (IC₅₀ 8.50 μ g/mL) and phenylethanoid compounds showed moderate antioxidant activities with (IC₅₀ 62.52-70.20) mM compared to standard BHA (butylated hydroxy anisole) with (IC₅₀ 44.20 mM) [66].

4. Phytochemistry of manilkara

Chemical investigation of different plants of Manilkra species have led to the isolation and characterisation of many primary and secondary metabolites. These chemical constituents can be categorised as triterpenoids, saponins, phenolic compounds, volatile compounds, phytosterols, saturated and unsaturated fatty acids and others.

4.1 Triterpenoids

Many triterpene esters and pentacyclic triterpenoids have been identified in Manilkara species. In study conducted by Fernandes *et al*, the mixture containing the triterpenes β -amyryn acetate (76.3%) and α -amyryn acetate (23.7%) showed inhibition of acetylcholinesterase activity and suggested that these two compounds as chemical markers of *M. subseria* [67].

4.2 Phenolic Compounds

Phenolic compounds such as simple phenols, phenolic acids, flavonoids, coumarins and xanthenes are present in genus Manilkara. The most common flavonoids present in this genus are quercetin, quercitol, kaempferol, ferulic acid, chlorogenic acid and catechin. (Table 2 and 3, 33-96 and 173-185).

4.3 Saponins

Few saponins including pentacyclic triterpenoid saponin Manilkoraside was isolated from stem bark of *M. zapota*. It showed potent inhibitive effects on HL-60 and HT-29 and moderate activity on A 549, A 431 and MCF-7 cell lines [68]. There was isolation of novel saponin along with other 2 saponins from *M. hexandra* seeds [45].

4.4 Phytosterols

β -sitosterol, stigmasterol, α -spinosterol and other few phytosterols have been reported from Manilkra species. (Table 3, 113-123).

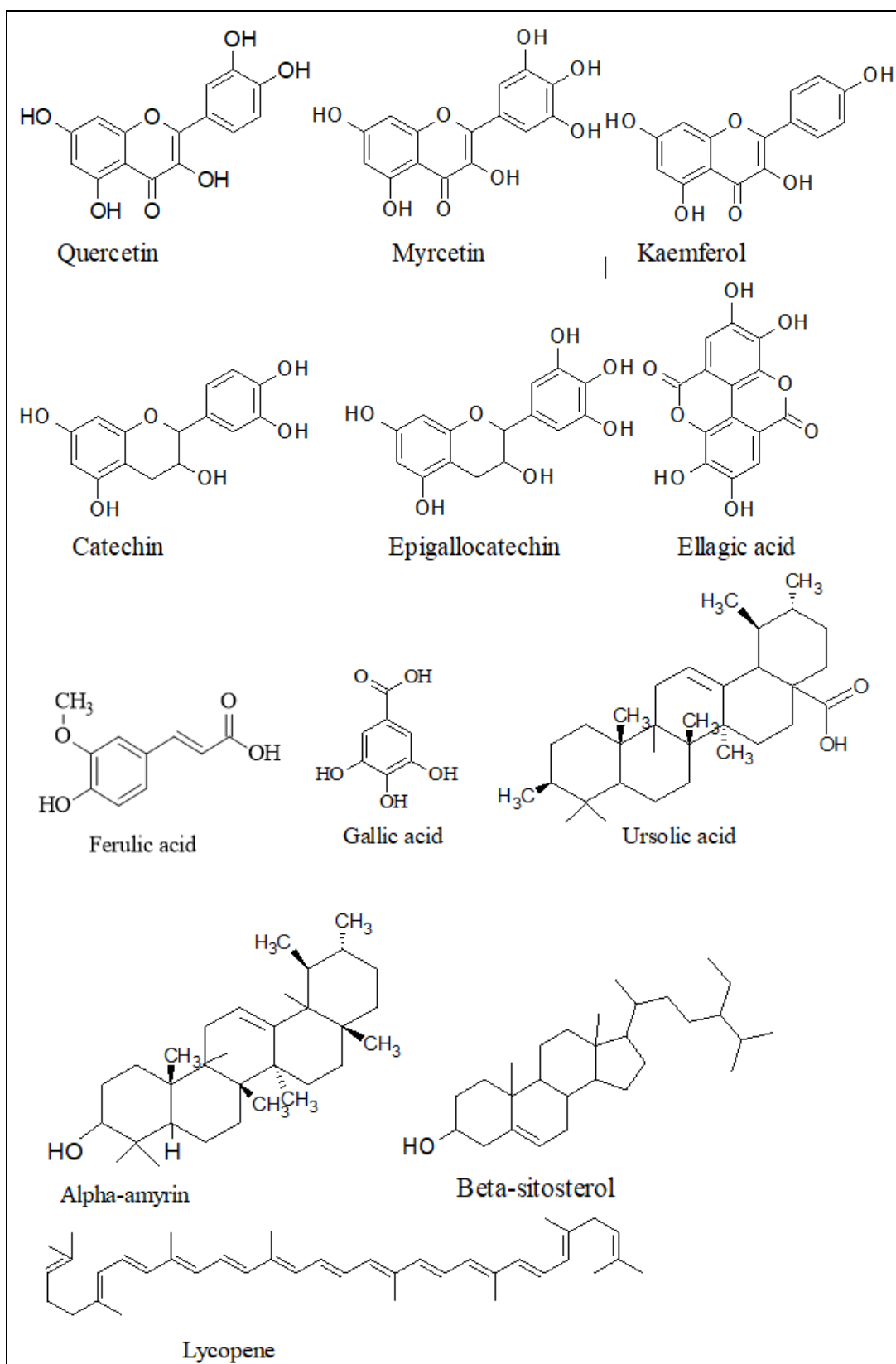
4.5 Volatile compounds

34 volatile compounds were identified from leaves of *M. subseria*. (Table-3,127-165).

4.6 Fatty Acids

Many saturated and unsaturated fatty acids have been isolated from Manilkara species. (Table-3, 97-112).

Other phytoconstituents and isolated compound from different species of genus Manilkra are given in Table 3.

Table 2: Some important chemical compounds present in Genus Manilkara**Table 3:** Bioactive Constituents from Manilkara

SI NO	COMPOUND	SPECIES	PART USED	REFERENCE
Triterpenoids				
1	Erythrodiol	<i>M. zapota</i>	Leaf	[35]
2	α -amyrin	<i>M. hexandra</i>	Roots	[71]
3	β - amyrin	<i>M. hexandra</i> <i>M. zapoata</i>	Roots seeds	[61, 71]
4	α -amyrin acetate	<i>M. subseria</i> <i>M. hexandra</i>	Fruits Fruits	[20, 62, 72, 73]

5	β - amyirin acetate	<i>M. subseria</i> <i>M. pellegriniana</i>	Fruits Stem bark	[20, 31, 62, 72]
6	Alpha amyirin caprolates & caprylates	<i>M. subseria</i>	Fruits	[20]
7	Beta amyirin caprolates & caprylates	<i>M. subseria</i>	Fruits	[20]
8	Pomoleic acid	<i>M. subseria</i> , <i>M. zapota</i>	Fruits Leaves	[72]
9	Oleanolic Acid	<i>M. subseria</i> <i>M. obovate</i> <i>M. zapota</i>	Leaves Root, stem bark, Seeds	[11, 46, 61]
10	Obovatol	<i>M. subseria</i> <i>M. obovata</i>	Zapota Root, stem bark, leaves	[72, 74]
11	Ursolic acid	<i>M. subseria</i> <i>M. obovata</i> <i>M. hexandra</i> <i>M. pellegriniana</i>	Fruits Twig Fruits Stem bark	[11, 31, 62, 71]
12	1 β -hydroxyeucaphic acid	<i>M. pellegriniana</i>	Stem bark	[31]
13	Lacefriedelic acid	<i>M. obovata</i>	Leaves	[11]
14	Friedelin	<i>M. obovata</i>	Twig	[11]
15	Friedelanol	<i>M. obovata</i>	Stem bark	[11]
16	Canophyllol	<i>M. obovata</i>	Twig	[11]
17	Canophyllic acid	<i>M. obovata</i>	Leaves	[11]
18	Ethylcanophyllate	<i>M. obovata</i>	Root, stem bark, leaves	[74]
19	Lupeol	<i>M. obovata</i> <i>M. zapota</i>	Twig Seeds	[11, 61]
20	Betulinic acid	<i>M. subseria</i> <i>M. zapota</i> <i>M. obovata</i>	Root Seeds Root	[11, 61, 62]
21	Maslinic acid	<i>M. obovata</i>	Twig	[11]
22	Taraxerol	<i>M. hexandra</i> <i>M. pellegriniana</i>	Leaves, Roots Stem bark	[73, 75, 31]
23	Taraxeryl acetate,	<i>M. hexandra</i> <i>M. pellegriniana</i>	Bark Stem bark	[75, 31]
24	α -amyirin cinnamate	<i>M. hexandra</i>	Bark	[75]
25	3 β -O-acetyl- α -amyirin	<i>M. hexandra</i> <i>M. bidentata</i>	Seeds Resin	[41, 75]
26	3 β -O-trans cinnamyl- α -amyirin	<i>M. bidentata</i>	Resin	[41]
27	3 β -O-trans cinnamyl lupeol	<i>M. bidentata</i>	Resin	[41]
28	3 β ,6 α ,19 α ,23-tetrahydroxyurs-12-en-28-oic acid 1	<i>M. argentea</i>	Stem bark	[31]
29	1 β -hydroxyeucaphic acid	<i>M. pellegriniana</i>	Stem bark	[31]
30	2 α -hydroxy-ursolic acid	<i>M. argentea</i>	Stem bark	[31]
31	2 α , 3 α , 19 α ,20 β , 23-pentahydroxyurs-12-en-28-oic acid,	<i>M. pellegriniana</i>	Stem bark	[31]
32	1 β , 2 α , 3 α , 5, 19 α , 24-hexahydroxyurs-11(12), 20(30)-dien-28-oic acid	<i>M. pellegriniana</i>	Stem bark	[31]
Phenolic Compounds				
33	D-quercitol,	<i>M. subseria</i> , <i>M. zapota</i> <i>M. hexandra</i>	Seeds Leaves, seeds Leaves, seeds	[6, 60, 73]
34	Methyl chlorogenate	<i>M. zapota</i> ,	Fruits	[30]
35	Myricetin	<i>M. subseria</i> , <i>M. zapota</i>	Leaves Leaves	[72, 76]
36	Myricitrin	<i>M. zapota</i> , <i>M. subseria</i>	Fruits Leaves	[30, 72]
37	Dihydromyricetin	<i>M. zapota</i>	Fruits, leaves	[30,76]
38	Myricetin-3-O- α -L-rhamnoside	<i>M. zapota</i>	Seeds, leaves,	[6, 46]
39	Myricetin-3-O- β -D-glucopyranoside	<i>M. zapota</i>	Leaves	[17]
40	Quercetin	<i>M. hexandra</i> , <i>M. subseria</i> <i>M. zapota</i>	Seeds, fruits Leaves Seeds, fruits	[9, 30, 72, 75]
41	Quercitrin	<i>M. zapota</i> <i>M. subseria</i>	Fruits Leaves	[30, 72]
42	Dihydroquercetin	<i>M. hexandra</i>	Seeds	[75]
43	Chlogeneic acid	<i>M. zapota</i>	Leaves	[17, 76]
44	Methyl 4-galloylchlorogenate	<i>M. zapota</i>	Fruits	[30]
45	4-O-galloylchlorogenic acid	<i>M. zapota</i>	Fruits	[30]
46	Catechin	<i>M. zapota</i> , <i>M. hexandra</i>	Leaf, fruits, seeds Fruits	[30, 72, 77]
47	Epicatechin,	<i>M. zapota</i>	Fruits	[30]
48	Gallocatechin,	<i>M. zapota</i>	Fruits	[30]
49	Gallic acid,	<i>M. zapota</i>	Fruit, seed	[60, 77]
50	Apigenin-7-O- α -L-rhamnoside,	<i>M. zapota</i>	Leaves	[46]

51	Leucodelphinidin,	<i>M. zapota</i>	Fruits	[78]
52	Leucocyanidin	<i>M. zapota</i>	Fruits	[78]
53	Leucopelargonidin	<i>M. zapota</i>	Fruits	
54	Saccharose	<i>M. zapota</i>	Leaves, seeds	[6]
55	Caffeic acid	<i>M. zapota</i>	Leaves	[46]
56	Trans-cinnamic	<i>M. zapota</i>	Fruit	[79]
57	Ellagic acid	<i>M. zapota</i>	Fruit	[79]
58	Catechol	<i>M. hexandra</i>	Fruits, seed	[9]
59	P-hydroxybenzoic acid	<i>M. hexandra</i> , <i>M. zapota</i>	Fruits Fruits	[9,79]
60	Kaempferol	<i>M. subseria</i> , <i>M. zapota</i> <i>M. hexandra</i>	Leaves Fruits Fruits	[9,72,77]
61	5-caffeoyl quinic acid conjugates	<i>M. zapota</i>	Fruit	[80]
62	Catechin	<i>M. subseria</i> , <i>M. zapota</i> <i>M. hexandra</i>	Leaves Fruits, seeds Fruits, seeds	[9,30,60,79]
63	Salicylic acid,	<i>M. zapota</i> , <i>M. hexandra</i>	Leaves Fruit, seeds	[9,76]
64	3,4-Dihydroxybenzoic acid	<i>M. zapota</i>	Fruit pulp, seeds	[60]
65	Vanillic acid	<i>M. zapota</i> , <i>M. hexandra</i>	Leaves seeds	[9,76]
66	Gallic acid	<i>M. zapota</i> <i>M. hexandra</i> <i>M. zapota</i>	Immature fruits, leaves Seeds, fruits fruit	[9, 30, 76, 79]
67	Caffeic acid	<i>M. zapota</i>	Leaves	[46, 76]
68	Coumaric acid	<i>M. hexandra</i>	Leaves and fruits	[9]
69	Ferulic acid	<i>M. zapota</i> , <i>M. hexandra</i> <i>M. zapota</i>	Leaves Fruits, seeds fruits	[9, 76, 79]
70	Chlorogenic acid	<i>M. zapota</i>	Leaves, Immature fruits	[30, 76, 79]
71	Syringic acid	<i>M. subseria</i> <i>M. zapota</i>	Leaves Leaves	[76]
72	Afzelechin	<i>M. zapota</i>	Leaves	[76]
73	Epicatechin	<i>M. zapota</i> <i>M. hexandra</i>	Leaves Seeds, fruits	[9, 76]
74	Epigallocatechin	<i>M. zapota</i> , <i>M. hexandra</i>	Leaves Fruit, seeds	[9, 76]
75	flavanol	<i>M. hexandra</i>	Fruit, seeds	[9]
76	Flavan-3-ol	<i>M. hexandra</i>	Fruit, seeds	[9]
77	Laricitrin	<i>M. zapota</i>	Leaves	[76]
78	Myricetin-3-O-rhamnoside	<i>M. zapota</i>	Leaves	[76]
79	Laricitrin-3-O-rhamnoside	<i>M. zapota</i>	Leaves	[76]
80	Prodelphinidin B	<i>M. zapota</i>	Leaves	[76]
81	2-Hydroxybenzaldehyde	<i>M. zapota</i>	Leaves	[76]
82	Guaiacol	<i>M. zapota</i>	Leaves	[76]
83	Pyroglutamic acid	<i>M. zapota</i>	Leaves	[76]
84	Threonic acid	<i>M. zapota</i>	Leaves	[76]
85	Vanillin	<i>M. zapota</i>	Leaves	[76]
86	7,9- di-tert-butyl-1-oxaspiro [4.5] deca-6,9- diene-2,8-dione.	<i>M. hexandra</i>	Stem bark	[21]
87	3-Hydroxycoumarin	<i>M. zapota</i>	Leaves	[17]
88	Mearnstrin	<i>M. zapota</i>	Leaves	[17]
89	Germanicol	<i>M. zapota</i>	Leaves	[17]
90	Germanicol acetate	<i>M. zapota</i>	Leaves	[17]
Saponins				
91	Manilkoraside	<i>M. zapota</i>	Stem bark	[63]
92	Aridanin	<i>M. hexandra</i> <i>M. obovata</i>	Stem bark Stem bark	[11, 81]
93	1 β, 2α, 3β, 19α-tetrahydroxyursolic acid 28-O-beta -D-glucopyranoside	<i>M. hexandra</i>	Stem bark	[81]
94	Saponin 1 (16-a-hydroxyprotobassic acid)	<i>M. hexandra</i>	Seeds	[40]
95	Saponin 2 (16-a-hydroxyprotobassic acid)	<i>M. hexandra</i>	Seeds	[40]
96	Saponin 3(protobassic acid)	<i>M. hexandra</i>	Seeds	[40]
Fatty acids (saturated and unsaturated)				
97	Oleic acid	<i>M. zapota</i>	Leaves	[46]
98	Ethyl oleate	<i>M. bidentata</i>	Seeds	[82]
99	α-linolenic acid	<i>M. zapota</i>	Leaves	[46]
100	Linoleic acid,	<i>M. zapota</i>	Leaves	[46]
101	Lupeol acetate	<i>M. zapota</i>	Leaves	[46]
102	Oleanolic acid	<i>M. zapota</i> <i>M. obovata</i>	Seeds, Leaves Stem bark	[46, 61]

103	Lupeol-3-acetate	<i>M. zapota</i>	Leaves	[46, 6]
104	Palmitic acid	<i>M. zapota</i>	Leaves, fruit, seeds	[46, 60]
105	Capric acid	<i>M. zapota</i>	Leaves	[46]
106	Stearic acid	<i>M. zapota</i>	Leaves	[46, 83]
107	Linoleic acid	<i>M. zapota</i>	Leaves	[46]
108	Oleic acid	<i>M. zapota</i>	Fruits, seeds	[83]
109	Myristic acid	<i>M. zapota</i>	Fruit, seed	[83]
110	Garbanzol	<i>M. argentea</i>	Stem bark	[31]
111	3,5,7,4',5'-pentahydroxyflavan	<i>M. argentea</i>	Stem bark	[31]
112	(+)-proto-quercitol	<i>M. argentea</i>	Stem bark	[31]
Phyto sterols				
113	β -sitosterol	<i>M. zapota</i>	Leaves, stem bark	[46, 75, 81]
114	Stigmasterol	<i>M. zapota</i>	Leaves	[46]
115	A mixture of stigmasterol and b-sitosterol	<i>M. obovata</i> <i>M. zapota</i>	Twig Seeds	[11,61]
116	Stigmasterol-3-O- β -D-glucoside	<i>M. obovata</i>	Twig	[11]
117	b-sitosterol-3-O- β -D-glucoside	<i>M. obovata</i>	Leaves	[11]
118	Stigmasterol-3-O- β -D-glucopyranoside	<i>M. zapota</i>	Seeds	[61]
119	β -D-glucoside of β -sitosterol	<i>M. hexandra</i>	Seeds	[73]
120	α -spinosterol	<i>M. hexandra</i>	Bark, seed	[75]
121	3 α - spinasterol	<i>M. pellegriniana</i>	Stem bark	[31]
122	Spinasterone	<i>M. pellegriniana</i>	Stem bark	[31]
123	α -spinasterol glucoside	<i>M. argentea</i>	Stem bark	[31]
Carotenoid				
124	β -carotene	<i>M. zapota</i>	Fruits	[84]
125	Lycopene	<i>M. zapota</i>	Fruits	[85]
126	α -tocopherol (Vitamin E)	<i>M. zapota</i>	Fruits	[84]
Volatile compounds essential oils				
127	Hexanol	<i>M. subseria</i>	Leaves	[72]
128	Trans-2-decenal	<i>M. subseria</i>	Leaves	[72]
129	3-hexen-1-ol	<i>M. subseria</i>	Leaves	[72]
130	Octanal	<i>M. subseria</i>	Leaves	[72]
131	Heptanol	<i>M. subseria</i>	Leaves	[72]
132	Heptanal	<i>M. subseria</i>	Leaves	[72]
133	2-heptanone	<i>M. subseria</i>	Leaves	[72]
134	(3E)-3-hexenyl acetate	<i>M. subseria</i>	Leaves	[72]
135	Beta-ocimene	<i>M. subseria</i>	Leaves	[72]
136	Octanol	<i>M. subseria</i>	Leaves	[72]
137	Linalool oxide	<i>M. subseria</i>	Leaves	[72]
138	2-nonanone	<i>M. subseria</i>	Leaves	[72]
139	Linalool	<i>M. subseria</i>	Leaves	[72]
140	Terpineol	<i>M. subseria</i>	Leaves	[72]
141	Pentacosane	<i>M. subseria</i>	Leaves	[72]
142	Methyl salicylate	<i>M. subseria</i>	Leaves	[72]
143	Heptacosane	<i>M. subseria</i>	Leaves	[72]
144	Safranal	<i>M. subseria</i>	Leaves	[72]
145	Squalene	<i>M. subseria</i>	Leaves	[72]
146	Beta-cyclocitral	<i>M. subseria</i>	Leaves	[72]
147	Nonacosane	<i>M. subseria</i>	Leaves	[72]
148	Geraniol	<i>M. subseria</i>	Leaves	[72]
149	Beta-damascenone	<i>M. subseria</i>	Leaves	[72]
150	Beta-damascone	<i>M. subseria</i>	Leaves	[72]
151	Beta-caryophyllene	<i>M. subseria</i>	Leaves	[72]
152	Farnesene	<i>M. subseria</i>	Leaves	[72]
153	Caryophyllene oxide	<i>M. subseria</i>	Leaves	[72]
154	1,2,3-Propanetriol	<i>M. zapota</i>	Fruits, Seeds	[83]
155	5-Methyl-2(3H) furanone	<i>M. zapota</i>	Fruits, Seeds	[83]
156	Octadecanoic acid	<i>M. hexandra</i>	Stem bark	[21]
157	Hexadecanoic acid	<i>M. bidentata</i> <i>M. subseria</i> <i>M. hexandra</i>	Seed Leaves and fruits Stem bark	[82, 69, 21]
158	Hexadecanoic acid ethyl esters	<i>M. subseria</i> <i>M. bidentata</i>	Fruits Seed	[69, 82]
159	Octadecanoic acid ethyl esters	<i>M. subseria</i>	Fruits	[69]
160	Eicosene	<i>M. subseria</i>	Leaves	[72]
161	Heneicosane	<i>M. subseria</i>	Leaves	[72]
162	Phytol	<i>M. subseria</i>	Leaves	[72]
163	Docosene	<i>M. subseria</i>	Leaves	[72]
164	Tricosane	<i>M. subseria</i>	Leaves	[72]
165	Untriacontane	<i>M. subseria</i>	Leaves	[72]

Carboxylic And Dicarboxylic Acid				
166	Succinic acid	<i>M. zapota</i>	Leaves	[76]
167	Malic Acid	<i>M. zapota</i>	Leaves	[76]
168	Adipic acid	<i>M. zapota</i>	Leaves	[76]
169	3-Oxadipic acid	<i>M. zapota</i>	Leaves	[76]
170	Cinnamic acid	<i>M. hexandra</i>	Leaves	[71]
171	Trans-cinnamic acid	<i>M. zapota</i>	Fruit	[79]
172	Formic acid	<i>M. zapota</i>	Fruit, seed	[83]
Coumarins				
173	Isocallophyllic acid	<i>M. obovata</i>	Leaves	[11]
174	Calophyllolide	<i>M. obovata</i>	Leaves	[11]
175	3-methoxyphenol	<i>M. obovata</i>	Twig	[11]
176	Calaustralin	<i>M. obovata</i>	Twig	[11]
177	Callophyllic acid	<i>M. obovata</i>	Stem bark	[74]
178	Obovacoumaric acid	<i>M. obovata</i>	Stem bark	[74]
Xanthones				
179	Lacexanthone	<i>M. obovata</i>	Root	[11]
180	Gerontoxanthone A	<i>M. obovata</i>	Roots	[11]
181	6-deoxyjacareubin	<i>M. obovata</i>	Twig	[11]
182	1,3-dihydroxy-2-methoxyxanthone	<i>M. obovata</i>	Stem Bark	[11]
183	1,2-dihydroxy-3-methoxyxanthone	<i>M. obovata</i>	Stem bark	[11]
184	2-hydroxyxanthone	<i>M. obovata</i>	Roots	[11]
185	Norathyriol	<i>M. zapota</i>	Leaves	[76]
Phenylethanoid				
186	2-(4-Hydroxyphenethyl) tetratriacontanoate	<i>M. zapota</i>	Seeds	[61]
187	2-(4-Hydroxyphenethyl) tetracosanoate	<i>M. zapota</i>	Seeds	[61]
188	2-(4-Hydroxyphenethyl) docosanoate	<i>M. zapota</i>	Seeds	[61]
189	2-(4-Hydroxyphenethyl) eicosanoate	<i>M. zapota</i>	Seeds	[61]
190	2-(4-Hydroxyphenethyl) octadecenoate	<i>M. zapota</i>	Seeds	[61]
191	2-(4-Hydroxyphenethyl) hexadecanoate	<i>M. zapota</i>	Seeds	[61]
Carbohydrates				
192	Rhamnose	<i>M. hexandra</i> <i>M. zapota</i>	Seed Leaves	[73, 76]
193	Glucose	<i>M. hexandra</i> <i>M. zapota</i>	Seeds Leaves	[73, 76]
194	Xylose,	<i>M. hexandra</i>	Seeds	[73]
195	Arabinose	<i>M. hexandra</i>	Seeds	[73]
196	Sucrose	<i>M. zapota</i>	Leaves	[76]
Minerals				
197	Iron, copper, zinc, calcium, potassium	<i>M. zapota</i>	Fruits	[77]
Amino Acids				
198	Aspartic acid and glutamic acid, lysine, proline, hydroxyproline	<i>M. zapota</i> <i>M. hexandra</i>	Fruit Fruit	[86, 78, 73]
Hydrocarbons				
199	n-hexadecane	<i>M. zapota</i>	Leaves	[46]
200	n-tricontane	<i>M. zapota</i>	Leaves	[46]
201	n-octacosane	<i>M. zapota</i>	Leaves	[46]
202	n-docosane	<i>M. zapota</i>	Leaves	[46]
203	n-tetracosane	<i>M. zapota</i>	Leaves	[46]
204	Hentriacontane	<i>M. zapota</i>	Leaves	[71]
Others				
205	Shikimic acid	<i>M. zapota</i>	Leaves	[76]
206	3-O-Galloylquinic acid	<i>M. zapota</i>	Leaves	[76]
207	Esculetin	<i>M. zapota</i>	Leaves	[76]
208	Quinic acid	<i>M. zapota</i>	Leaves	[76]
209	Hydroquinone glucuronide	<i>M. zapota</i>	Leaves	[76]
210	Glyceraldehyde	<i>M. zapota</i>	Fruit, seed	[83]
211	3-Glucogallic acid	<i>M. zapota</i>	Leaves	[76]
212	3-p-Coumaroylquinic acid	<i>M. zapota</i>	Leaves	[76]
213	2-Propyl tetrahydropyran-3-ol	<i>M. bidentata</i>	Seeds	[87]
214	2-Furancarboxyaldehyde, (hydroxymethyl)-	<i>M. bidentata</i>	Seeds	[87]
215	5-methyl-2-furancarboxyaldehyde	<i>M. bidentata</i>	Seeds	[87]
Glycosides				
216	Cyanogenetic glycosides	<i>M. zapota</i>	Latex	[88]

5. Conclusion

This review highlighted the phytopharmacological findings of different plant extracts from genus Manilkra. Quite majority of the species remain unknown or scientifically unexplored.

Thus, it justifies the need for the advanced research studies on Manilkra species on their therapeutic potential. It will pave the way for future drug design.

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7. Conflict of interest

We declare that the authors have no conflict of interest

8. References

1. Armstrong KE. A revision of the Asian-Pacific species of *Manilkara* (Sapotaceae). *Edinburgh Journal of Botany*. 2013 Mar;70(1):7-56.
2. Kirtikar KR, Basu BD. *Indian Medicinal Plants*. 2nd ed. Dehradun, India: International Book Distributors; c2006. p. 1494-1498.
3. Rhourri-Frih B, Renimel I, Chaimbault P, André P, Herbet G, Lafosse M. Pentacyclic triterpenes from *Manilkara bidentata* resin. Isolation, identification and biological properties. *Fitoterapia*. 2013;88:101-108.
4. Mishra N, Pareek A. Traditional Uses, Phytochemistry and Pharmacology of *Mimusops hexandra* Roxb. *Advances in Pharmaceutical and Ethnomedicines*. 2014;2(2):32-35.
5. Milind PP. Chickoo: A Wonderful Gift from Nature. *International Journal of Advance Research, Ideas and Innovations in Technology*. 2015;6(4):544-550.
6. Kaneria M, Baravalia Y, Vaghasiya Y, Chanda S. Determination of Antibacterial and Antioxidant Potential of Some Medicinal Plants from Saurashtra Region, India. *Indian Journal of Pharmaceutical Sciences*. 2009;71(4):406-412.
7. Kaneria M, Chanda S. Evaluation of antioxidant and antimicrobial properties of *Manilkara zapota* L. (chiku) leaves by sequential Soxhlet extraction method. *Asian Pacific Journal of Tropical Biomedicine*. 2012;2(3):S1526-533.
8. Islam ME, Parvin MS, Islam MR, Islam MS, Hasan SMR. Antioxidant Activity of the Ethanol Extract of *Manilkara zapota* Leaf. *Journal of Scientific Research*. 2011;4(1):193.
9. Rao GV, Sahoo MR, Madhavi MS, Mukhopadhyay T. Phytoconstituents from the leaves and seeds of *Manilkara zapota* Linn. *Der Pharmacia Lettre*. 2014;6(2):69-73.
10. Priya P, Shoba FG, Parimala M, Sathya J. Antioxidant and antibacterial properties of *Manilkara zapota* (L.) Royen flower. *International Journal of Pharmaceutical and Clinical Research*. 2014;6(2):174-178.
11. Gomathy K, Baskar R, Kumaresan K. Comparison of antioxidant potential in pulp and peel extracts of *Manilkara zapota* (L.) P. Royen. *African Journal of Biotechnology*. 2013;12(31):4936-4943.
12. Islam MR, Parvin MS, Hasan MR, Ekramul M. *In Vitro* And *In Vivo* Antioxidant Activity of Ethanolic Extract of *Manilkara zapota* Bark. *Journal of Global Pharma Technology*. 2010;2(11):23-30.
13. Parikh B, Patel VH. Quantification of phenolic compounds and antioxidant capacity of an underutilized Indian fruit: Rayan [*Manilkara hexandra* (roxb.) Dubard]. *Food Science and Human Wellness*. 2017;6(1):10-19.
14. Dutta S, Ray S. Evaluation Of *In Vitro* Free Radical Scavenging Activity of Leaf Extract Fractions of *Manilkara hexandra* (Roxb) *dubard* in Relation to Total Phenolic Contents. *International Journal of Pharmacy and Pharmaceutical Sciences*. 2015;7(10):296-301.
15. Akosung E, Kenmogne SB, Lobe Songue J, Longue Ekon JP, Lateef M, Ngeufa Happi E, et al. Bioactive constituents from *Manilkara obovata* (Sabine & G. Don) J.H. Hemsl. *Natural Product Research*. 2021;35(22):4347-4356.
16. Srisupap S, Chaicharoenpong C. *In vitro* antioxidant and antityrosinase activities of *Manilkara kauki*. *Acta Pharmaceutica*. 2021;71(1):153-162.
17. Nair R, Chanda S. Antimicrobial Activity of Terminalia catappa, *Manilkara zapota* and *Piper betel* Leaf Extract. *Indian Journal of pharmaceutical sciences*. 2008;70(3):390-393.
18. Osman MA, Aziz MA, Habib MR, Karim MR. Antimicrobial Investigation on *Manilkara zapota* (L.) P. Royen. *International Journal of Drug Development & Research*. 2011;3(1):185-190.
19. Mourão Mulvaney LC, Xavier-Júnior FH, Rodrigues AM, Stien D, Allegretti SM, Garcia VL. Antimicrobial and anthelmintic activities of the ethanolic extract, fractions and isolated compounds from *Manilkara zapota* LP Royen (Sapotaceae). *Journal of Pharmacy and Pharmacology*. 2021;73(3):377-387.
20. Shanmugapriya K, Saravana PS, Payal H, Mohammed SP, Bennai W. A comparative study of antimicrobial potential and phytochemical analysis of *Artocarpus heterophyllus* and *Manilkara zapota* seed extracts. *Journal of Pharmacy Research*. 2011;4(8):2587-2589.
21. Kothari V, Seshadri S. *In vitro* antibacterial activity in seed extracts of *Manilkara zapota*, *Anona squamosa* and *Tamarindus indica*. *Biological Research*. 2010;43(2):165-168.
22. Bhargavi S, Kanakaiah B, Sowmya DK, Ravi B, Nama S. An evaluation of the antibacterial activity of root extracts of *Manilkara Zapota* against *Staphylococcus Aureus* and *Escherichia Coli*. *International Journal of Phytopharmacology*. 2013;4(3):171-173.
23. Rakholiya K, Kaneria M, Chanda S. Inhibition of microbial pathogens using fruit and vegetable peel extracts. *International Journal of Food Sciences and Nutrition*. 2014;65(6):733-739.
24. Monisha SI, Vimala JR. Extraction, Identification and Pharmacological Evaluation of Phyto-Active Compound in *Manilkara Hexandra* (roxb.) *Dubard* Stem Bark. *Biosciences Biotechnology Research Asia*. 2018;15(3):687-698.
25. Pingili D, Amminbavi D, Awasthi A, Khan FM. Formulation, evaluation and *in vitro* antibacterial screening of herbal gel containing *Manilkara hexandra* (roxb.) *dubard* leaf extract. *International Journal of Pharmaceutical Sciences and Research*. 2018;9(2):702-707.
26. Mahida Y, Mohan JS. Screening of plants for their potential antibacterial activity against *Staphylococcus* and *Salmonella* spp. *Indian Journal of Natural Products and Resources*. 2007;6:301-305.
27. Patel K, Ali AK, Nair N, Kothari V. *In vitro* Antibacterial Activity of *Manilkara hexandra* (Sapotaceae) Seed Extracts and Violacein against multidrug resistant *Streptococcus mutans*. *Journal of Natural Remedies*. 2015;15(1):1-11.
28. Parekh J, Chanda S. *In vitro* screening of antibacterial activity of aqueous and alcoholic extracts of various Indian plant species against selected pathogens from Enterobacteriaceae. *African Journal of Microbiology*

- Research. 2007; 1(6):92-99.
29. Sumitra C, Jigna P. Assessment of Antimicrobial Potential of *Manilkara hexandra* Leaf. *Pharmacognosy Journal*. 2010; 2(12):448-455.
 30. Bharvad PB, Nayak AR, Patel NK, Mohan JSS. Screening Of Crude Root Extracts of Some Indian Plants for Their Antibacterial Activity. *Journal of Pure and Applied Sciences*. 2011;19:14-18.
 31. Sisodiya D and Shrivastava P. Antimicrobial activity of *Euphorbia thymifolia* (L.) and *Manilkara hexandra* (roxb). *International Journal of Current Advanced research*. 2018;7(2A):9660-9663.
 32. Narayana B, Sivaprakasam MK, Jeyarajan R. Antifungal activity of some plant extracts. *Indian Journal of Forestry*. 1994;17(1):10-4.
 33. Fernandes CP, Corrêa AL, Lobo JFR, Caramel OP, De Almeida FB, Castro ES, *et al.* Triterpene Esters and Biological Activities from Edible Fruits of *Manilkara subsericea* (Mart.) Dubard, Sapotaceae. *BioMed Research International*. 2013;2013:1-7.
 34. Srivastava M, Hegde M, Chiruvella KK, Koroth J, Bhattacharya S, Choudhary B, *et al.* Sapodilla Plum (*Achras zapota*) Induces Apoptosis in Cancer Cell Lines and Inhibits Tumor Progression in Mice. *Scientific Report*. 2015;4(1):1-9.
 35. Ma J, Luo X-D, Protiva P, Yang H, Ma C, Basile MJ, *et al.* Bioactive Novel Polyphenols from the Fruit of *Manilkara zapota* (Sapodilla). *Journal of Natural product*. 2003;66(7):983-986.
 36. Mogue LDK, Ango PY, Fotso GW, Mapitse R, Kapche DWFG, Karaosmanoğlu O, *et al.* Two new polyhydroxylated pentacyclic triterpenes with cytotoxic activities from *Manilkara pellegriniana* (Sapotaceae). *Phytochemistry Letters*. 2019;31:161-165.
 37. Khalek MA, Khatun Z, Rowshanul Habib M, Rezaul Karim M. Antitumor activity of *Manilkara zapota* (L.) fruits against Ehrlich ascites carcinoma in mice. *Biologija*. 2015;61(3-4):145-152.
 38. Tan BL, Norhaizan ME, Chan LC. *Manilkara zapota* (L.) P. Royen Leaf Water Extract Induces Apoptosis in Human Hepatocellular Carcinoma (HepG2) Cells via ERK1/2/Akt1/JNK1 Signalling Pathways. *Evidence-Based Complementary and Alternative Medicine*. 2018;2018:1-17.
 39. Osman MA, Rashid MM, Aziz MA, Habib MR, Karim MR. Inhibition of Ehrlich ascites carcinoma by *Manilkara zapota* L. stem bark in Swiss albino mice. *Asian Pacific Journal of Tropical Biomedicine*. 2011;1(6):448-451.
 40. Rashid MM, Hossain MI, Osman MA, Aziz MA, Habib MR, Karim MR. Evaluation of antitumor activity of *Manilkara zapota* leaves against Ehrlich ascites carcinoma in mice. *Environ. Environmental and Experimental Biology*. 2014;12:131-135.
 41. Ganguly A, Mahmud ZA, Uddin MMN, Rahman SA. In-vivo anti-inflammatory and anti-pyretic activities of *Manilkara zapota* leaves in albino Wistar rats. *Asian Pacific Journal of Tropical Disease*. 2013;3(4):301-307.
 42. Ganguly A, Al Mahmud Z, Kumar Saha S, Abdur Rahman SM. Evaluation of antinociceptive and antidiarrhoeal properties of *Manilkara zapota* leaves in Swiss albino mice. *Pharmaceutical Biology* 2016;54(8):1413-1419.
 43. Hossain H, Jahan F, Howlader SI, Dey SK, Hira A, Sarkar RP. Evaluation of Anti-inflammatory Activity and Total Flavonoids Content of *Manilkara zapota* (Linn.) Bark. *International Journal of Pharmaceutical and Phytopharmacological Research*. 2012;2:35-39.
 44. Konuku K, Karri KC, Gopalakrishnan VK, Hagos Z, Kebede H, Naidu TK, *et al.* Anti-Inflammatory Activity of *Manilkara zapota* Leaf Extract. *International Journal of Current Pharmaceutical Research*. 2017;9(4):130-134.
 45. Eskander JY, Haggag EG, El-Gindi MR, Mohamedy MM. A novel saponin from *Manilkara hexandra* seeds and anti-inflammatory activity. *Medicinal Chemistry Research*. 2014;23(2):717-724.
 46. Manirujjaman, Sultana F, Chowdhury M, Hossain M, Imran-UI-Haque M. *In Vivo* Assay of Analgesic Activity of Methanolic and Petroleum Ether Extracts of *Manilkara zapota* leaves. *British Journal of Pharmaceutical Research*. 2013 Oct 24;4:186-191.
 47. Jain PK, Soni P, Upmanyu N, Shivhare Y. Evaluation of Analgesic Activity of *Manilkara zapota* (Leaves). *Bangladesh Journal of Pharmacology*. 2011;1:14-17.
 48. Khan A. Evaluation of Analgesic and Anti-Inflammatory Activity of Whole Plant Extract of *Manikara zapota* Linn. *World Journal of Pharmacy and Pharmaceutical Sciences*. 2016;5(7):881-192.
 49. Alrashood ST, Al-Asmari AK, Alotaibi AK, Manthiri RA, Rafatullah S, Hasanato RM, *et al.* Protective effect of lyophilized sapodilla (*Manilkara zapota*) fruit extract against CCl4-induced liver damage in rats. *Saudi Journal of Biological Sciences*. 2020;27:2373-2379.
 50. Okafor SN, Okonkwoand TJ, Okoye TC. Evaluation of the antioxidant and hepatoprotective activities of *Manilkara obovata* seed extract in murine models. *IOSR Journal of Applied Chemistry*. 2015;8:114-117.
 51. Abdel Monem A, Meselhy M, Mossa M, Shazly A, Fayek N. Chemical and biological study of *Manilkara zapota* (L.) Van Royen leaves (Sapotaceae) cultivated in Egypt. *Pharmacognosy Research*. 2012;4(2):85-91.
 52. Paul SR, Hakim ML. *In vivo* hypoglycemic study of *Manilkara zapota* leave and seed extracts. *Bangladesh Journal of Pharmacology*. 2015;10(1):246-250.
 53. Das T, Das B, Saha D, Mishra SB. Anti-Hyperglycemic Activity of Hydro-Alcoholic Bark Extract of *Manilkara hexandra* (Roxb) In Streptozotocin Induced Diabetic Rats. *International Journal of Pharmacy and Pharmaceutical Sciences*. 2006;8(4):185-188.
 54. Nimbekar TP, Katolkar PP, Patil AT. Effects of *Manilkara hexandra* on blood glucose levels of normal and Alloxan induced diabetic rats. *Research Journal of Pharmacy Technology*. 2012;5(3):367-368.
 55. Singh M, Soni P, Upmanyu N, Shivhare Y. *In-vitro* Anti-arthritis Activity of *Manilkara zapota* Linn. *Asian Journal of Pharmacy and Technology*. 2011;1(4):123-124.
 56. Pingili D, Awasthi A, Amminbavi D. Assessment of *in vitro* antiarthritic activity of *Manilkara hexandra* (roxb.) Dubard leaf extract. *Annals of Phytomedicine*. 2016;5(2):152-155.
 57. SS. Phytochemical screening and in-vitro corneal wound healing activity of the leaves of *Manilkara zapota* (l) van royen var. Pkm1 in newer herbal drug development. *World Journal of Pharmaceutical Research*. 2017;6(9):639-647.
 58. Periasamy G, Kumar AS, Prameela R, Kishorekumar K, Gnananath K. Stimulation of immune system function by polysaccharides of *Manilkara hexandra* (roxb.) bark *International Journal of Pharmacy and Pharmaceutical*

- Sciences. 2012;4(3):430-432.
59. Gopalkrishnan B, Ringmichon C, Chachad D. Antifertility activity of *Manilkara hexandra* (roxb.) *Dubard* seed extract on male albino rats. *International Journal of Applied Biology and Pharmaceutical Technology*. 2016;7(3):71-76.
 60. Shah M, Goswami S, Santani D. Effect of *Manilkara hexandra* (roxb.) *Dubard* against experimentally-induced gastric ulcers. *Phytotherapy Research*. 2004;18(10):814-818.
 61. Modi KP, Lahiri SK, Goswami SS, Santani DD, Shah MB. Evaluation of antiulcer potential of *Mimusops hexandra* in experimental gastro duodenal ulcers. *Journal of Complementary and Integrative Medicine*. 2012;9(1):1-12.
 62. Abd El-Mordy FM, El-Hamouly MM, Ibrahim MT, El-Rheem GA, Aly OM, Abd El-kader AM, *et al.* Inhibition of SARS-CoV-2 main protease by phenolic compounds from *Manilkara hexandra* (roxb.) *Dubard* assisted by metabolite profiling and in silico virtual screening. *RSC Advances*. 2020;10(53):32148-32155.
 63. Fernandes CP, Xavier A, Pacheco JPF, Santos MG, Mexas R, Ratcliffe NA, *et al.* Laboratory evaluation of the effects of *Manilkara subsericea* (Mart.) *Dubard* extracts and triterpenes on the development of *Dysdercus peruvianus* and *Oncopeltus fasciatus*. *Pest Management Science*. 2013;69(2):292-301.
 64. Fernandes C, De Almeida F, Silveira A, Gonzalez M, Mello C, Feder D, *et al.* Development of an insecticidal nanoemulsion with *Manilkara subsericea* (Sapotaceae) extract. *Journal of Nanobiotechnology*. 2014;12(1):22.
 65. Shafii ZA. Phytochemical and Antioxidant Properties of *Manilkara zapota* (L.) P Royen Fruit Extracts and its Formulation for Cosmeceutical Application. *Asian Journal of Plant Science*. 2017;7(3):29-41.
 66. Fomani M, Nougua A, Toze F, Ndom J, Waffo A, Wansi J. Bioactive Phenylethanoids from the Seeds of *Manilkara zapota*. *British Journal Pharmaceutical Research*. 2015;8(5):1-5.
 67. Fernandes CP, Corrêa AL, Cruz RAS, Silva-Filho MV, Santos MG, De Brito MA, *et al.* Anticholinesterasic Activity of *Manilkara subsericea* (Mart.) *Dubard* Triterpenes. *Latin American Journal of Pharmacy*. 2011;30(8):1631-1634.
 68. Awasare S, Bhujbal S, Nanda R. *In Vitro* Cytotoxic Activity of Novel Oleanane Type of Triterpenoid Saponin From Stem Bark of *Manilkara zapota* Linn. *Asian Journal of Pharmaceutical and Clinical Research*. 2012;5:183-188.
 69. Attar SK, Thakur NS, Patel HF, Singh ND, Makawana AI, Leua HN, *et al.* Underutilized fruit *Manilkara hexandra* (Khirmi). *Rastriya Krishi*. 2016;11(1):17-18.
 70. Gopalkrishnan B, Shimpi S, CLR. Stem bark of *Manilkara hexandra* (roxb.) *Dubard* - pharmacognosy. *World Journal of Pharmacy Pharmaceutical Science*. 2014;3(2):2503-2511.
 71. Akosung E, Djouaka Bavoua JL, Tabekoueng GB, Stammer H-G, Frese M, Kapche Wabo Fotso GD, *et al.* Antibacterial constituents from the roots, stem bark and leaves of *Manilkara obovata* (Sabine & G. Don) J H. Hemsl. (Sapotaceae). *Phytochemistry Letters*. 2021;44:55-61.
 72. Anjali, Garg V, Dhiman A *et al.* genus *Manilkara*: An update. *Journal of Pharma Innovation*. 2018;7(1):316-318.
 73. Misra G, Mitra CR. *Mimusops hexandra*-III. Constituents of root, leaves and mesocarp. *Phytochemistry*. 1968;7(12):2173-2176.
 74. De Almeida F, Fernandes C, Romao W, Vanini G, Costa H, França H, *et al.* Secondary metabolites from leaves of *Manilkara subsericea* (Mart.) *Dubard*. *Pharmacognosy Magazine*. 2015;11(44):533.
 75. Mitra CR, Misra G. *Mimusops hexandra*-I.: Constituents of fruit and seed. *Phytochemistry* 1965;4(2):345-348.
 76. Misra G, Mitra CR. *Mimusops Hexandra*-II.: Constituents of bark and seed. *Phytochemistry* 1966;5(3):535-538.
 77. Islam S, Alam MB, Ann HJ, Park JH, Lee SH, Kim S. Metabolite Profiling of *Manilkara zapota* L. Leaves by High-Resolution Mass Spectrometry Coupled with ESI and APCI and *In Vitro* Antioxidant Activity, α -Glucosidase, and Elastase Inhibition Assays. *International Journal of Molecular Sciences*. 2020;22(1):132.
 78. Singh JP, Kaur A, Shevkani K, Singh N. Composition, bioactive compounds and antioxidant activity of common Indian fruits and vegetables. *Journal of Food Science and Technology*. 2016;53(11):4056-4066.
 79. Mathew AG, Lakshminarayana S. Polyphenols of immature sapota fruit. *Phytochemistry*. 1969;8(2):507-509.
 80. Can-Cauich CA, Sauri-Duch E, Betancur-Ancona D, Chel-Guerrero L, González-Aguilar GA, Cuevas-Glory LF, *et al.* Tropical fruit peel powders as functional ingredients: Evaluation of their bioactive compounds and antioxidant activity. *Journal of Functional Foods*. 2017;37:501-506.
 81. Pontes PV, Moreira RFA, Trugo LC, Maria CABD. The content of chlorogenic acids in tropical fruits. *Journal of the Science of Food and Agriculture*. 2002;82(10):1177-1181.
 82. Srivastava M, Singh J. A New Triterpenoid Saponin from *Mimusops hexandra*. *International Journal of Pharmacognosy*. 1994;32(2):197-200.
 83. Powder-George YL, Mohammed FK. GC-MS analysis of the bioactive phytoconstituents of various organic crude extracts from the seed kernels of *Manilkara bidentata* (balata) collected in Trinidad, W.I. *Natural Product Research*. 2018;32(3):358-361.
 84. Shafii ZA. Phytochemical and Antioxidant Properties of *Manilkara zapota* (L.) P Royen Fruit Extracts and its Formulation for Cosmeceutical Application. *Asian Journal of Plant Science*. 2017;7:29-41.
 85. Charoensiri R, Kongkachuichai R, Suknicom S, Sungpuag P. Beta-carotene, lycopene, and alpha-tocopherol contents of selected Thai fruits. *Food Chemistry*. 2009;113(1):202-207.
 86. Silva LMR Da, Figueiredo EAT De, Ricardo NMPS, Vieira IGP, Figueiredo RW De, Brasil IM, *et al.* Quantification of bioactive compounds in pulps and by-products of tropical fruits from Brazil. *Food Chemistry*. 2014;143:398-404.
 87. Hall NT, Smoot JM, Knight RJ, Nagy S. Protein and amino acid compositions of ten tropical fruits by gas-liquid chromatography. *Journal of Agriculture Food Chemistry*. 1980;28(6):1217-1221.
 88. Mahajan RT, Badgujar SB. Phytochemical Investigations of some Laticiferous Plants belonging to Khandesh Region of Maharashtra: *Ethnobotanical Leaflets*. 2008;(1):1145-1152.