
Two new species of *Stachybotrys*, and a key to the genus

Umpava Pinruan¹, Eric H.C. McKenzie^{2*}, E.B. Gareth Jones¹ and Kevin D. Hyde³

¹National Centre for Genetic Engineering and Biotechnology, Biotech Central Research Unit, 113 Phahonyothin Road, Klong 1, Klongluang, Pathumthani, Thailand, 12120

²Landcare Research, Private Bag 92170, Auckland, New Zealand

³Centre for Research in Fungal Diversity, Department of Ecology & Biodiversity, The University of Hong Kong, Pokfulam Road, Hong Kong SAR, PR China

Pinruan, U., McKenzie, E.H.C., Jones, E.B.G. and Hyde, K.D. (2004). Two new species of *Stachybotrys*, and a key to the genus. *Fungal Diversity* 17: 145-157.

Stachybotrys palmae sp. nov. found on decaying petioles of *Licuala longicalycata* in a peat swamp forest, Thailand, and *S. cordylines* sp. nov. found on decaying leaves of *Cordyline banksii* in New Zealand are described and illustrated. The new species are compared with other species in the genus, and a key is provided to all accepted species of *Stachybotrys*. A new name, *S. thermotolerans*, is proposed for *S. ramosa* Udaiyan, a later homonym of *S. ramosa* Dorai & Vittal. A list of accepted *Stachybotrys* species with references is provided.

Key words: anamorphic fungi, hyphomycetes, monocotyledonous plants, palm fungi.

Introduction

We are studying fungi occurring on monocotyledonous plants (e.g. Hyde *et al.*, 2002; McKenzie *et al.*, 2002; Yanna *et al.*, 2002; Zhou and Hyde, 2002). In this paper we describe two new species of *Stachybotrys*. The first species was found in Thailand on decaying petioles of *Licuala longicalycata* Furtado, a tropical palm species. The second species occurred on *Cordyline banksii* Hook. f., a New Zealand species of *Laxmanniaceae*. *Stachybotrys* species are saprobes, common in soil (Ellis, 1971, 1976), decaying plant material (Whitton *et al.*, 2001) and wild fruits (Tang *et al.*, 2003) and have also been recorded on submerged wood in mangroves (Maria and Sridhar, 2003). They produce single-celled conidia aggregated in slimy heads. The genus is worldwide in distribution although some species are restricted to the tropics and subtropics. More than 50 species of *Stachybotrys* are accepted, including those species formerly included in the genus *Memnoniella*. The new species are described and compared with similar species.

*Corresponding author: Eric McKenzie; e-mail: McKenzieE@landcareresearch.co.nz

Materials and methods

Decaying fronds of *Licuala longicalycata* were collected from Sirindhorn Peat Swamp Forest, Narathiwat, Thailand, and decaying leaves of *Cordyline banksii* were collected from the Waitakere Ranges, Auckland, New Zealand. The decaying tissues were returned to the laboratory and incubated in damp chambers. Single spore isolates of both fungi were obtained. Microscopic measurements for *S. palmae* were taken from specimens mounted in water while those for *S. cordylines* were taken from specimens mounted in lactophenol.

Taxonomy

Stachybotrys palmae Pinruan, **sp. nov.** (Figs. 1-5)

Etymology: referring to its association with palms.

Conidiophora macronematosa, mononematosa, solitaria vel fasciculata, eramosa, erecta, recta vel paulo flexuosa, laevia, 2-5 septata, brunnea, pallidae ad apicem, crassitunicata, (80-)110-230 μm longa, 6.3-10 μm crassa. *Cellulae conidiogenae* monophialidicae, discretiae, 5-7 in verticillo dispositae, clavatae, 11-12.5 \times 6-7.5 μm , laevae, hyalinae. *Conidia* in massis globosis aggregata, ellipsoidea, hyalina, verrucosa, 10-15 \times 5-7.5 μm ; apice et basi truncata.

Conidiophores macronematous, mononematous, single or in groups, unbranched, erect, straight or slightly flexuous, smooth, 2-5 septate, brown, apical cell hyaline, thick-walled, (80-)110-230 μm long, 6.3-10 μm wide. *Conidiogenous cells* monophialidic, discrete, determinate, clavate, smooth, hyaline, forming a whorl of 5-7 at the apex of the conidiophores, 11-12.5 μm long, 6-7.5 μm thick in the broadest part. *Conidia* aggregated in pale cream slimy heads, ellipsoidal or boat-shaped, truncate at the base and apex, hyaline, verrucose, non-septate, 10-15 \times 5-7.5 μm (\bar{x} = 13 \times 7 μm , n = 20).

Habitat: Saprobic on *Licuala longicalycata*.

Distribution: Thailand.

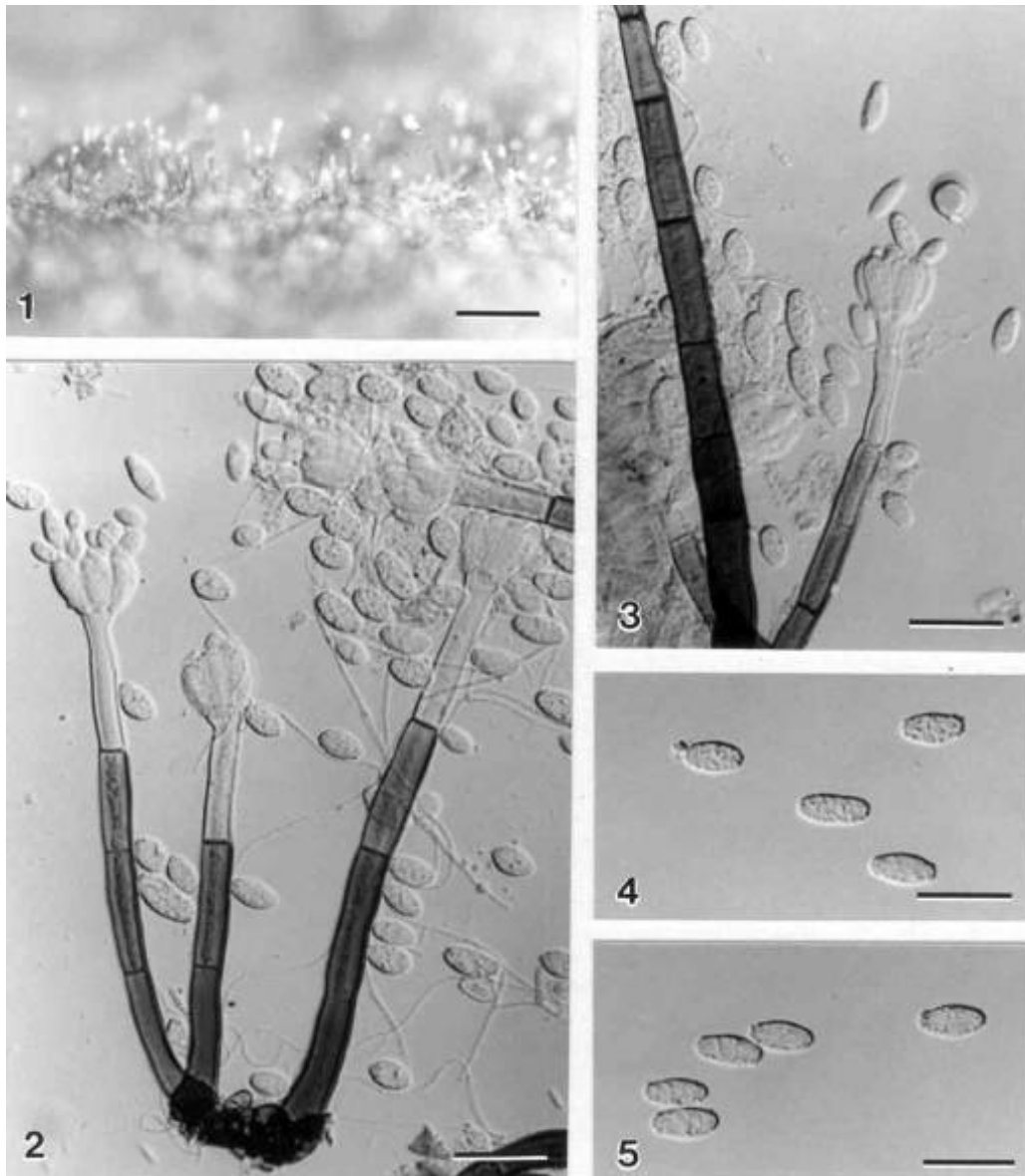
Material examined: THAILAND, Narathiwat, Sirindhorn Peat Swamp Forest, on decaying rachis of *Licuala longicalycata* Furtado, 12 May 2001, U. Pinruan (Wah 35) [BIOTEC Bangkok Herbarium (BBH), **holotype**].

Notes: *Stachybotrys palmae* is one of only five species of *Stachybotrys* that produce hyaline conidia. Of these, *S. palmae* is the only species with rough-walled conidia. Those of the other four species (*S. bambusicola* Rifai, *S. bisbyi* (Sriniv.) G.L. Barron, *S. guttulispota* Muhsin & Al-Helfi, *S. palmijunci* Rifai) are smooth-walled.

Stachybotrys cordylines McKenzie, **sp. nov.** (Figs. 6-13)

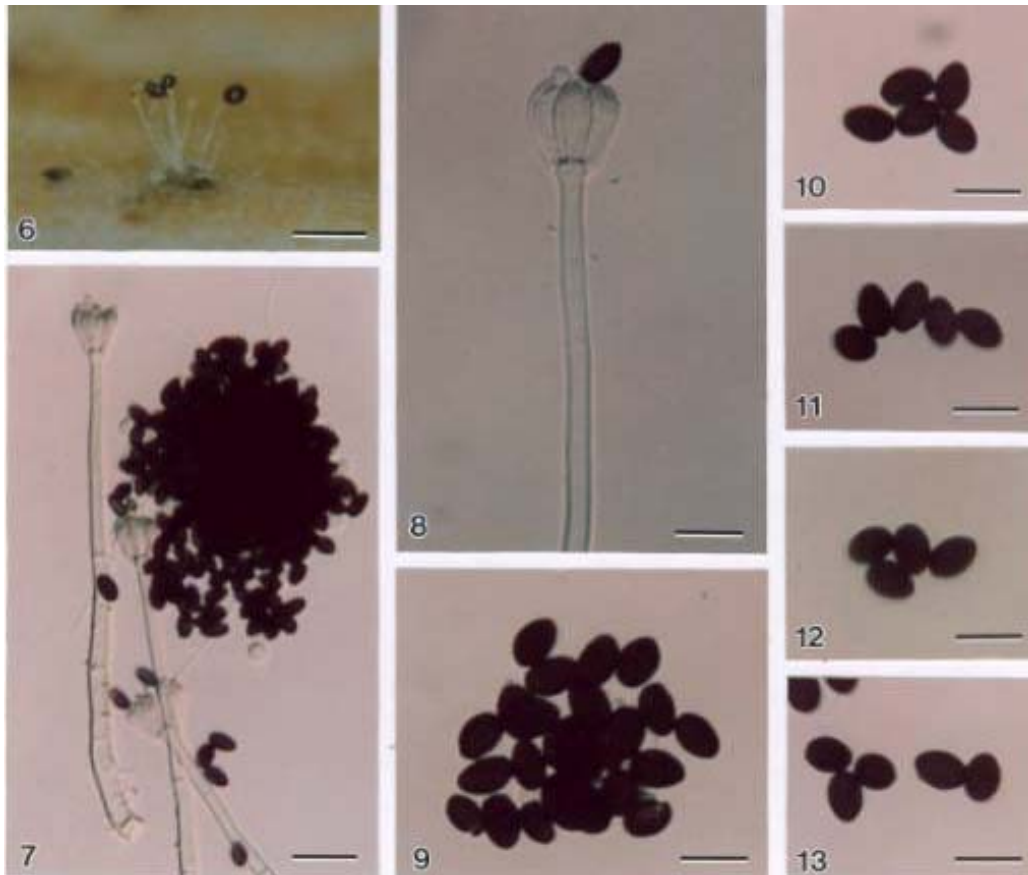
Etymology: referring to the host genus *Cordyline*.

Conidiophora macronematosa, mononematosa, solitaria, eramosa, erecta, recta vel paulo flexuosa, laevia, interdum granulis magnis tecta, 4-7 septata, hyalina, crassitunicata, 95-



Figs. 1-5. Light micrographs of *Stachybotrys palmae* (from holotype). **1.** Colonies on substratum. **2, 3.** Conidiophores with pale apical cell. **4, 5.** Conidia. Bars: 1 = 200 μm ; 2-5 = 20 μm .

160 μm longa, basi 5.8-8 μm , prope apicem 3-4.2 μm , apice inflata 3.5-5.1 μm diam. *Cellulae conidiogena*e monophialidicae, discretae, 7-8 in verticillo dispositae, clavatae, 11-14(-16) \times 3.8-5.4 μm , laevae, hyalinae. *Conidia* in massis globosis aggregata, ellipsoidea vel obovoidea, olivacea, rugulosa, eseptata, 7-8.3 \times 3.2-5.1 μm .



Figs. 6-13. Light micrographs of *Stachybotrys cordylinis* (from holotype). **6.** Colonies on substratum. **7, 8.** Conidiophores. **9-13.** Conidia. Bars: 6 = 150 μm ; 7 = 20 μm ; 8-13 = 10 μm .

Conidiophores macronematous, mononematous, single, unbranched, erect, straight or slightly flexuous, smooth or sometimes covered with a few large granules near the base, 4-7 septate, hyaline, thick-walled, 95-160 μm long, 5.8-8 μm thick near the base, tapering to 3-4.2 μm near the apex, slightly enlarged at the apex to 3.5-5.1 μm thick and bearing a whorl of 7-8 phialides. *Conidiogenous cells* monophialidic, discrete, clavate, 11-14(-16) μm long, 3.8-5.4 μm thick in the broadest part, smooth, hyaline. *Conidia* aggregated in slimy, black, glistening heads, ellipsoid or obovoid, olive green, rugulose, non-septate, 7-8.3 \times 3.2-5.1 μm (\bar{x} = 7.6 \times 4.5 μm , n = 20).

Habitat: Saprobic on *Cordyline banksii*.

Distribution: New Zealand.

Material examined: NEW ZEALAND, Auckland, Waitakere Ranges, near Spraggs Bush, on dead leaves of *Cordyline banksii* Hook. f., 4 Aug 2003, R.E. Beever [PDD 78085, **holotype**]. Culture ex holotype [ICMP 15219].

The conidia of *S. cordylines* are similar in size and shape to those of *S. albipes*. However, the latter fungus has smooth conidia, whereas those of *S. cordylines* are rugulose. It differs from *S. chartarum* by having hyaline conidiophores; those of *S. chartarum* are dark olivaceous towards the apex.

***Stachybotrys thermotolerans* McKenzie, nom. nov.**

≡ *Stachybotrys ramosa* Udaiyan, Journal of Economic and Taxonomic Botany 15: 641, 1991[1992], nom. illegit., non Dorai & Vittal, Transactions of the British Mycological Society 87: 642, 1986[1987].

Notes: Udaiyan (1991) described several new species of hyphomycetes that were isolated from water-cooling towers. The *Stachybotrys* was stated to have an optimum temperature for growth of 43°C. Udaiyan (1991) called this fungus a ‘thermophile’, but gave insufficient data to equate to the definition of a thermophile (Mouchacca, 1997).

Discussion

Memnoniella and *Stachybotrys* have been considered distinct genera (e.g., Ellis, 1971, 1976). However, the only difference between these two genera is that the conidia are in long, dry chains in *Memnoniella* while they are in slimy masses in *Stachybotrys*. Some authors have considered this is not a valid generic distinction and have suggested the two genera be combined under the older name of *Stachybotrys* (e.g. Smith, 1962; Carmichael *et al.*, 1980). This is supported by the molecular results of Haugland *et al.* (2001). All accepted species of *Stachybotrys* and *Memnoniella* are included in the following key. Synonyms have mainly followed those listed by Jong and Davis (1976). However, their synonymy of *S. sinuatophora* Matsush. with *S. nephrospora* Hansf. has not been accepted. A recently collected specimen (PDD 77554, on dead stems of *Abelmoschus esculentus*, Fiji) matched the description of *S. sinuatophora*. The conidiophores are distinctly sinuous, and quite unlike those of *S. nephrospora*. The four remaining species of *Memnoniella* (*M. indica*, *M. leprosa*, *M. longistipitata* and *M. levispora*) should probably be transferred to *Stachybotrys*, following molecular analysis.

Key to the accepted species of *Stachybotrys*

- | | | |
|----|---|----|
| 1. | Conidia when mature roughened, or surface ridged or banded, or with delicate striations | 2 |
| 1. | Conidia when mature smooth | 39 |
| 2. | Conidial surface ridged, banded, striate or rugulose | 3 |
| 2. | Conidia rough-walled | 6 |

3.	Conidia cylindrical, with delicate, oblique striations, 11-16 × 4-6 μm <i>S. cylindrospora</i>	
3.	Conidia ellipsoid	4
4.	Conidia with a ridged surface, 12-13 × 5-5.5 μm <i>S. virgata</i>	
4.	Conidia less than 12 μm long	5
5.	Conidia with a ridged or banded surface, 7-12 × 4-6 μm <i>S. chartarum</i>	
5.	Conidia with a rugulose surface, 7-8.3 × 3.2-5.1 μm <i>S. cordylinea</i>	
6.	Conidiophores synnematos	7
6.	Conidiophores mononematous	8
7.	Conidia subglobose, 7-12 μm diam., apex black, base pale <i>Memnoniella leprosa</i>	
7.	Conidia globose, 4-5.5 μm diam <i>S. stilboidea</i>	
8.	Conidia produced in dry chains, sometimes also in slimy heads	9
8.	Conidia aggregated in slimy heads	14
9.	Conidia regularly of two kinds; catenate and globose, 5-5.5 μm diam.; non-catenate and cylindrical, 7-10 × 3.5-5 μm <i>S. zuckii</i>	
9.	Conidia mainly of only one kind	10
10.	Conidiophores regularly branched, tapering to a slender, pointed apex; conidia globose, 5-6 μm diam. <i>Memnoniella indica</i>	
10.	Conidiophores not tapering towards apex	11
11.	Conidiophores (170-)260-460(-610) μm long, unbranched; conidia catenate and globose or subglobose, 5.8-8.5 × 6.3-8.3 μm; rarely producing cylindrical or ovoid conidia in slimy heads, 10.6-11.9 × 4.8-5.7 μm <i>Memnoniella longistipitata</i>	
11.	Conidiophores shorter	12
12.	Conidia globose to subglobose, 3-5 μm diam.; rarely producing cylindrical conidia in slimy heads, 7-9 × 3-5 μm <i>S. echinata</i>	
12.	Conidia larger	13
13.	Conidia globose, 5-7 μm diam <i>S. subsimplex</i>	
13.	Conidia globose, 7-8 μm diam <i>Stachybotrys</i> sp. (Haugland <i>et al.</i> , 2001)	
14.	Conidia mainly globose or subglobose	15
14.	Conidia mainly of other shapes	20
15.	Conidia more than 10 μm diam	16
15.	Conidia mostly less than 10 μm diam	17
16.	Conidia 11-12 μm diam <i>S. sphaerospora</i>	
16.	Conidia (15.4-)21-25.2(-28) μm diam <i>S. nilagirica</i>	
17.	Conidiophores regularly sympodially branched; conidia 4.5-8 μm diam. <i>S. globosa</i>	
17.	Conidiophores not regularly sympodially branched	18

Fungal Diversity

18. Conidia 5-6 μm diam.....	<i>S. microspora</i>
18. Conidia more than 6 μm diam	19
19. Conidia 6-8 μm diam.; conidiophores (40-)70-90 μm long.....	<i>S. ruwenzoriensis</i>
19. Conidia 7.5-10.5 \times 7-10.5 μm ; conidiophores 58-272 μm long.....	<i>S. kapiti</i>
20. Conidia reniform or curved	21
20. Conidia not reniform	25
21. Conidia tightly reniform, (11-)13-15(-15.5) \times (10.5-)12-14 μm	<i>S. nephrodes</i>
21. Conidia less than 10 μm wide.....	22
22. Conidiophores distinctly sinuous, branched; conidia 8-12 \times 6-7 μm	<i>S. sinuatophora</i>
22. Conidiophores not sinuous	23
23. Conidia strictly kidney-shaped, 10-13.5 \times 6-9.5 μm ; phialides 10-14 \times 5-6 μm ; conidiophores smooth, 86-137 μm long.....	<i>S. reniverrucosa</i>
23. Conidia more variable, not strictly kidney-shaped	24
24. Conidia reniform or comma-shaped, 8-12 \times 4-6(-7) μm ; phialides smooth, 10-12 \times 5-6 μm ; conidiophores smooth or roughened, up to 400 μm long.....	<i>S. nephrospora</i>
24. Conidia ovoid to reniform, 9-12 \times 4.5-8 μm ; phialides smooth or verruculose, 10-21 \times 4-7 μm ; conidiophores smooth, up to 190 μm long.....	<i>S. oenanthes</i>
25. Conidia usually more than 10 μm long.....	26
25. Conidia usually less than 10 μm long.....	32
26. Conidia more than 8 μm wide	27
26. Conidia less than 8 μm wide.....	28
27. Conidia ellipsoid or obovoid, 10-15 \times 9.5-11(-12.5) μm ; conidiophores 80-235 μm long	<i>S. verrucispora</i>
27. Conidia ellipsoid to broadly ellipsoid, 14.5-19 \times 8-11.5 μm ; conidiophores 54-75 μm long	<i>S. waitakere</i>
28. Conidia cylindrical or ellipsoid; conidiophores sometimes branched	29
28. Conidia ellipsoid or navicular; conidiophores not branched	30
29. Conidia cylindrical, (10-)11-13(-15) \times (3.5-)4-4.5(-5.3) μm ; conidiophores up to 320 μm long.....	<i>S. freycinetiae</i>
29. Conidia ellipsoid or cylindrical, 10-19 \times 5-6.5 μm ; conidiophores 120-260 μm long	<i>S. xanthosomae</i>
30. Conidia hyaline, ellipsoid or navicular, 10-15 \times 5-7.5 μm ; conidiophores smooth, 80-230 μm long	<i>S. palmae</i>
30. Conidia dark-coloured	31

31.	Conidiophores warted, up to 100 µm long; conidia ellipsoid, 12-13 × 5-5.5 µm	<i>S. virgata</i>	
31.	Conidiophores smooth, up to 180 µm long; conidia ellipsoid, 10-15 × 6-8 µm	<i>S. kampalensis</i>	
32.	Conidia more than 5 µm wide		33
32.	Conidia usually less than 5 µm wide		34
33.	Conidia ellipsoid or ovoid, 7.5-10(-14) × 5-7 µm; conidiophores up to 270 µm long	<i>S. dichroa</i>	
33.	Conidia ellipsoid, 7-9 × 6 µm; conidiophores 45-50 µm long	<i>S. klebahnii</i>	
34.	Conidia biguttulate, oblong, (5.5-)6-8 × 3-4 µm	<i>S. queenslandica</i>	
34.	Conidia without guttules		35
35.	Conidiophores often branched, up to 130 µm long; conidia cylindrical or ellipsoid, 6.5-9.5 × 2-3.5 µm	<i>S. breviuscula</i>	
35.	Conidiophores unbranched or rarely branched		36
36.	Conidiophores smooth, up to 200 µm long; conidia ellipsoid, 3-6 × 2.5-3.5 µm; phialides 7-11 × 3-4 µm	<i>S. parvispora</i>	
36.	Conidiophores less than 100 µm long		37
37.	Conidia ovoid or ellipsoid, 5-8 × 2.5-3.5 µm; phialides 5.5-11 × 2.5-3 µm; conidiophores verrucose, 63-95 µm long	<i>S. mangiferae</i>	
37.	Conidia usually wider		38
38.	Conidia ellipsoid, 7-9 × 3.5-4.5 µm; phialides 9-11 × 4-5 µm; conidiophores smooth, hyaline, 48-85 µm long	<i>S. zeae</i>	
38.	Conidia ellipsoid or cylindrical, 7-9 × 3-6 µm; phialides 7.5-11 × 3 µm; conidiophores smooth or verrucose, pale brown, darker towards apex, 48-94 µm long	<i>S. suthepensis</i>	
39.	Conidia produced in dry chains, subglobose, 4-6 µm diam	<i>Memnoniella levispora</i>	
39.	Conidia aggregated in slimy heads		40
40.	Conidia reniform or curved		41
40.	Conidia not reniform or curved		43
41.	Conidiophores not proliferating or branched; conidia 4.5-7 × 3-4.5 µm	<i>S. renisporoides</i>	
41.	Conidiophores proliferating or branched		42
42.	Conidiophores proliferating through apex; conidia 5.5-7 × 4-5 µm	<i>S. proliferata</i>	
42.	Conidiophores sympodially proliferating or branching; conidia 5.2-7 × 3.5-5.2 µm	<i>S. renispora</i>	
43.	Conidia usually more than 14 µm in length		44
43.	Conidia usually less than 14 µm in length		46
44.	Conidia broadly ellipsoid, 16-28 × 12-17 µm, with a basal apiculus	<i>S. theobromae</i>	
44.	Conidia smaller, without an apical apiculus		45

Fungal Diversity

45. Conidia cylindrical-clavate or ellipsoid, pale brown, 14.5-17.5 × 6.5-11 μm *S. cannae*
 45. Conidia ellipsoid, hyaline, 13.8-18.4 × 4-5.8 μm *S. palmijunci*
46. Conidia fusiform or limoniform47
 46. Conidia of other shapes48
47. Conidia limoniform or fusiform, hyaline, pink in mass, 8-14 × 6-9 μm or 10-16 × 3-6 μm .
 *S. bisbyi*
 47. Conidia fusoid, cylindrical or ellipsoid, grey to dark grey, black in mass, 7-10.2 × 2.5-3.5
 μm..... *S. havanensis*
48. Conidia mostly more than 6 μm wide49
 48. Conidia mostly less than 6 μm wide50
49. Conidia subglobose or ellipsoid, dark brown, 7-9 × 6-7 μm; conidiophore sympodially
 branched *S. ramosa*
 49. Conidia obovoid, rarely ellipsoid, hyaline or pale pink, 10-15.5 × 6.5-8 μm
 *S. bambusicola*
50. Conidia cylindrical, 8.8-12 × 2-2.4 μm *S. longispora*
 50. Conidia mainly other shapes, wider.....51
51. Conidia distinctly biguttulate, 9-12 × 3.5-5 μm *S. guttulispora*
 51. Conidia not guttulate52
52. Conidiophores often sympodially branched; conidia ellipsoid, 8.5-15.5 × 3.5-5 μm;
 phialides 8.5-12 × 5-7 μm; conidiophores up to 68 μm long *S. thermotolerans*
 52. Conidiophores not usually sympodially branched53
53. Conidia ovoid, (4-)7-9(-12) × (3-)5-6(-7) μm; phialides 9-16 × 3-5 μm; conidiophores
 usually smooth, hyaline, up to 250 μm long.....
 *Melanopsamma pomiformis* (anamorph *S. albipes*)
 53. Conidia of other shapes, mostly less than 5 μm wide54
54. Conidia cylindrical or subcylindrical, occasionally rough-walled, 7-11 × (2.5-)3.5-5 μm;
 phialides 8-13 × 3.2-5 μm; conidiophores grey to black, 70-120 μm long... *S. yunnanensis*
 54. Conidia of other shapes55
55. Conidia navicular, dark brown, 6-9 × 3-4 μm; phialides 8-13 × 3-4 μm; conidiophores
 subhyaline to pale brown, up to 60 μm long..... *S. sansevieriae*
 55. Conidia of other shapes56
56. Conidia ellipsoid, olivaceous or dark, 5.9-10.2 × 2.2-5.2 μm; conidiophores hyaline
 *S. lunzinensis*
 56. Conidia broadly ellipsoid to obovoid, blackish green, 8-10.5 × 4-5.5 μm; phialides 8-11 ×
 4-6 μm; conidiophores hyaline at base, darker towards apex, which is sometimes
 verrucose *S. chlorohalonata*

Accepted *Stachybotrys* (and *Memnoniella*) species

- S. albipes* (Berk. & Broome) S.C. Jong & E.E. Davis, Mycotaxon 3: 425 (1976) (teleomorph = *Melanopsamma pomiformis* (Pers.) Sacc.).
≡ *Sporocybe albipes* Berk. & Broome, Annals and Magazine of Natural History 8: 19 (1871).
- S. bambusicola* Rifai, Transactions of the British Mycological Society 47: 270 (1964).
- S. bisbyi* (Sriniv.) G.L. Barron, Mycologia 56: 315 (1964).
≡ *Hyalostachybotrys bisbyi* Sriniv., Journal of the Indian Botanical Society 37: 341 (1958).
- S. breviscula* McKenzie, Mycotaxon 41: 180 (1991).
- S. cannae* Bat., Anais da Sociedade de Biologia de Pernambuco 15: 394 (1957).
- S. chartarum* (Ehrenb.) S. Hughes, Canadian Journal of Botany 36: 812 (1958).
≡ *Stilbospora chartarum* Ehrenb., Sylvae Mycologicae Berolinenses: 9 (1818).
- S. chlorohalonata* B. Andersen & Thrane, Mycologia 95: 1228 (2003).
- S. cordylines* McKenzie, Fungal Diversity 17: 146 (2004).
- S. cylindrospora* C.N. Jensen, Cornell University Agricultural Experiment Station Bulletin 315: 496 (1912).
- S. dichroa* Grove, Journal of Botany, London 24: 201 (1886).
- S. echinata* (Rivolta) G. Sm., Transactions of the British Mycological Society 45: 392 (1962).
≡ *Penicillium echinatum* Rivolta, Dei Parassiti Vegetali: 451 (1873).
- S. freycinetiae* McKenzie, Mycotaxon 41: 183 (1991).
- S. globosa* P.C. Misra & S.K. Srivast., Transactions of the British Mycological Society 78: 556 (1982).
- S. guttulispota* Muhsin & Al-Helfi, Sydowia 34: 133 (1981).
- S. havanensis* Mercado & J. Mena, Acta Botánica Cubana 55: 2 (1988).
- M. indica* T.S.K. Prasad, Asha & Bhat, Mycotaxon 85: 341 (2003).
- S. kampalensis* Hansf. Proceedings of the Linnean Society 155: 45 (1943).
- S. kapiti* Whitton, McKenzie & K.D. Hyde, New Zealand Journal of Botany 39: 493 (2001).
- S. klebahnii* Burnhard, Phytopathologische Zeitschrift 1: 314 (1930).
- M. leprosa* R.F. Castañeda, Fungi Cubenses (La Habana): 10 (1986).
- M. levispora* Subram., Journal of the Indian Botanical Society 33: 40 (1954).
- S. longispota* Matsush., Icones Microfungorum a Matsushima Lectorum: 145 (1975).
- M. longistipitata* D.W. Li, Chin S. Yang, Vesper & Haugland, Mycotaxon 85:

- 254 (2003).
- S. lunzinensis* Svilv., Zentralblatt für Bakteriologie, Parasitenkunde und Infektionskrankheiten, Abt. II, 103: 182 (1941).
- S. mangiferae* P.C. Misra & S.K. Srivast., Transactions of the British Mycological Society 78: 556 (1982).
- S. microspora* (B.L. Mathur & Sankhla) S.C. Jong & E.E. Davis, Mycotaxon 3: 448 (1976).
 ≡ *S. atra* var. *microspora* B.L. Mathur & Sankhla, Scientia and Culture 32: 93 (1966).
- S. nephrodes* McKenzie, Mycotaxon 41: 185 (1991).
- S. nephrospora* Hansf., Proceedings of the Linnean Society, London 155: 45 (1943).
- S. nilagirica* Subraman., Proceedings of the Indian Academy of Science, B 46: 330 (1957).
- S. oenanthes* M.B. Ellis, Mycological Papers 125: 29 (1971).
- S. palmae* Pinruan, Fungal Diversity 17: 146 (2004).
- S. palmijunci* Rifai, Reinwardtia 8: 537 (1974).
- S. parvispora* S. Hughes, Mycological Papers 48: 74 (1952).
- S. proliferata* K.G. Karand., S.M. Kulk. & Patw., Biovigyanam 18: 79 (1992).
- S. queenslandica* Matsush., Matsushima Mycological Memoirs 6: 40 (1989).
- S. ramosa* Dorai & Vittal, Transactions of the British Mycological Society 87: 642 (1986).
- S. renispora* P.C. Misra, Mycotaxon 4: 161 (1976).
- S. renisporoides* K.G. Karand., S.M. Kulk. & Patw., Biovigyanam 18: 79 (1992).
- S. reniverrucosa* Whitton, McKenzie & K.D. Hyde, New Zealand Journal of Botany 39: 496 (2001).
- S. ruwenzoriensis* Matsush., Matsushima Mycological Memoirs 4: 17 (1985).
- S. sansevieriae* G.P. Agarwal & N.D. Sharma, Journal of the Indian Botanical Society 53: 78 (1974).
- S. sinuatophora* Matsush., Microfungi of the Solomon Islands and Papua-New Guinea: 61 (1971).
- S. sphaerospora* Morgan-Jones & R.C. Sinclair, Mycotaxon 10: 372 (1980).
- S. stilboidea* Munjal & J.N. Kapoor, Mycopathologia et Mycologia Applicata 39: 121 (1969).
- S. subsimplex* Cooke, Grevillea 12: 33 (1883).
- S. suthepensis* Photita, Lumyong, K.D. Hyde & McKenzie, Cryptogamie Mycologie 24: 149 (2003).
- S. theobromae* Hansf., Proceedings of the Linnean Society, London 155: 45 (1943).

- S. thermotolerans* McKenzie, Fungal Diversity 17: 149 (2004).
S. verrucispora Matsush., Matsushima Mycological Memoirs 4: 18 (1985).
S. virgata Krzemien. & Badura, Acta Societatis Botanicorum Poloniae 23: 759 (1954).
S. waitakere Whitton, McKenzie & K.D. Hyde, New Zealand Journal of Botany 39: 497 (2001).
S. xanthosomae Mercado & J. Mena, Acta Botánica Cubana 55: 4 (1988).
S. yunnanensis H.Z. Kong, Mycotaxon 62: 427 (1997).
S. zaeae E.B.G. Jones & Karr, Mycotaxon 4: 510 (1976).
S. zuckii K. Matsush. & Matsush., Matsushima Mycological Memoirs 8: 53 (1995).

Acknowledgements

This project is supported by Thailand research grant BRT R_145008, and New Zealand Foundation for Research, Science and Technology. We are grateful to Graduate School, Chiang Mai University, Saisamorn Lumyong, Ruud Valyasevi, and Morakot Tanticharoen for continued support, and to Manetr Boonyanant and his staff, for research facilities at the Sirindhorn Field and Nature Study Center, Narathiwat.

References

- Carmichael, J.W., Kendrick, W.B., Connors, I.L. and Sigler, L. (1980). *Genera of Hyphomycetes*. University of Alberta Press, Edmonton.
- Ellis, M.B. (1971). *Dematiaceous Hyphomycetes*. Commonwealth Mycological Institute, Kew, UK.
- Ellis, M.B. (1976). *More Dematiaceous Hyphomycetes*. Commonwealth Mycological Institute, Kew, UK.
- Haugland, R.A., Vesper, S.J. and Harmon, S.M. (2001). Phylogenetic relationships of *Memnoniella* and *Stachybotrys* species and evaluation of morphological features for *Memnoniella* species identification. Mycologia 93: 54-65.
- Hyde, K.D., Yanna, Pinnoi, A. and Jones, E.B.G. (2002). *Goidanichiella fusiforma* sp. nov. from palm fronds in Brunei and Thailand. Fungal Diversity 11: 119-122.
- Jong, S.C. and Davis, E.E. (1976). Contribution to the knowledge of *Stachybotrys* and *Memnoniella* in culture. Mycotaxon 3: 409-485.
- Maria, G.L. and Sridhar, K.R. (2003). Diversity of filamentous fungi on woody litter of five mangrove plant species from the southwest coast of India. Fungal Diversity 14: 109-126.
- McKenzie, E.H.C., Pinnoi, A., Wong, M.K.M., Hyde, K.D. and Jones, E.B.G. (2002). Two new hyaline *Chalara* species and a key to species described since 1975. Fungal Diversity 11: 129-139.
- Mouchacca, J. (1997). Thermophilic fungi: biodiversity and taxonomic status. Cryptogamie, Mycologie 18: 19-69.
- Smith, G. (1962). Some new and interesting species of micro-fungi. III. Transactions of the British Mycological Society 45: 387-394.

Fungal Diversity

- Tang, A.M.C., Hyde, K.D. and Corlett, R.T. (2003). Diversity of fungi on wild fruits in Hong Kong. *Fungal Diversity* 14: 165-185.
- Udaiyan, K. (1991)[1992]. Some interesting hyphomycetes from the industrial water cooling towers of Madras. *Journal of Economic and Taxonomic Botany* 15: 427-647.
- Whitton, S.R., McKenzie, E.H.C. and Hyde, K.D. (2001). Microfungi on the *Pandanaceae*: *Stachybotrys*, with three new species. *New Zealand Journal of Botany* 139: 489-499.
- Yanna, Ho, W.H. and Hyde, K.D. (2002). Fungal succession on fronds of *Phoenix hanceana* in Hong Kong. *Fungal Diversity* 10: 185-211.
- Zhou, D.Q. and Hyde, K.D. (2002). Fungal succession on bamboo in Hong Kong. *Fungal Diversity* 10: 213-227.

(Received 4 March 2004; accepted 15 June 2004)