

Disease and pest resistance of endophyte infected and non-infected drunken horse grass

CHUNJIE LI^{1,2}, XINGXU ZHANG¹, FEI LI¹, ZHIBIAO NAN^{1*} and C. L. SCHARDL²

¹College of Pastoral Agriculture Science and Technology, Lanzhou University; Key Laboratory of Grassland Argo-Ecosystem, Ministry of Agriculture; Gansu Grassland Ecological Research Institute; 730020 Lanzhou, China;

²Department of Plant Pathology, University of Kentucky, Lexington, KY 40546, USA
chunjie@lzu.edu.cn

Abstract

The naturally occurring mutualistic symbiosis of *Neotyphodium gansuense* and drunken horse grass (*Achnatherum inebrians*) was studied previously in China. In this paper, new data on the interactions of endophyte, host and pathogenic fungi, mite and insect pests are presented. Fungal diseases and pests were examined when test plants were grown in pots in a climate chamber or in the field. There were usually no significant ($P>0.05$) differences in the levels of powdery mildew infection (caused by *Blumeria graminis*) under climate chamber conditions; the only exception was that E+ plants had significantly less powdery mildew infection at 50% soil water holding capacity (WHC) than at 30% WHC. There was no significant difference ($P<0.05$) in the levels of infection of E- and E+ plants with leaf spot disease, caused by *Bipolaris sorokiniana*, under field conditions. Regardless of whether grown under growth chamber or field conditions, densities of the mite *Tetranychus cinnabarinus* on E+ grass were significantly ($P<0.05$) lower than that on E- grass. Densities of the bird-cherry-oat aphid (*Rhopalosiphum padi*) on E+ grass were significantly ($P<0.05$) lower than that on E- grass.

Keywords: *Neotyphodium gansuense*, *Achnatherum inebrians*, diseases, pests, resistance, endophytes

Introduction

Neotyphodium and *Epichloë* endophytes have been shown to confer enhanced fitness by increasing drought tolerance, and protecting the host plant against certain nematodes, fungal pathogens, insect herbivores and mammalian herbivores (Siegel *et al.* 1987; Latch 1993; Schardl & Phillips 1997). There is some evidence that colony growth of plant-pathogenic fungi is inhibited by *Neotyphodium* endophytes (White & Cole 1985; Christensen *et al.* 1991; Christensen & Latch 1991; Christensen 1996; Holzmann-Wirth *et al.* 2000), and that disease tolerance or resistance can be imparted by some but not all *Neotyphodium* species (West *et al.* 1989; Gwinn & Gavin 1992; Welty *et al.* 1991; Nan & Li 2000; Wheatley *et al.* 2000; Trevathan 1996; Blank *et al.* 1993; Funk *et al.* 1994; Hume *et al.* 1997).

Deterring herbivores from feeding on the host grass is one of the most beneficial features of endophytes (Siegel *et al.* 1987; Latch 1993; Nan & Li 2004; Popay & Bonos 2005). So far, at least 50 species in 35 genera have been shown to be deterred from feeding on endophyte-infected grasses (Siegel *et al.* 1987; Latch 1993; Popay *et al.* 2000; Nan & Li 2004; Popay & Bonos 2005).

The symbiotic association of *Neotyphodium gansuense* and *Achnatherum inebrians* is distributed widely in China, and can be found under harsh conditions such as arid or semi-arid and alpine or subalpine grasslands (Li *et al.* 2004a,b). Endophyte infected *A. inebrians* has been shown to contain high levels of the ergot alkaloids, ergonovine and lysergic acid amide (Miles *et al.* 1996; Li *et al.* 2006). Dual-culture testing and inoculation of detached leaves have shown that *N. gansuense* can inhibit growth and disease lesion development by some fungal pathogens (Li

2005). Our previous investigation included six diseases of *A. inebrians*: seedling rot (*Alternaria alternata*), rust (*Puccinia stipae-sibiricae*), powdery mildew (*Blumeria graminis*), smut (*Ustilago hypodytes*), leaf spot (*Bipolaris sorokiniana*) and ergot (*Sphacelia* sp.) (Li *et al.* 2003). To explore possibilities of utilising *N. gansuense* for bio-control, this research aimed to determine whether or not endophyte infection reduces the activity of naturally occurring diseases and pests of *A. inebrians*.

Materials and Methods

Seeds of *Achnatherum inebrians* were collected from Sangke grassland, Gansu province, China, and kept under constant 5 °C storage at the Official Forage Seed Testing Center of Ministry of Agriculture, Lanzhou, China.

Endophyte infected seeds were heat treated at a high moisture content to obtain E- seeds (Li 2005). Treated seeds were sown in plastic baskets. Endophyte status was detected by microscopy of samples from 4-week-old seedlings stained with 0.8% aniline blue.

In a climate chamber, E+ or E- plants were transplanted individually into plastic pots, and were watered regularly and weighed to maintain soil moisture content at 30 or 50% WHC; i.e. drought and normal growth conditions, respectively. E+ and E- seedlings were also transplanted into the field to establish trial plots under rain-fed conditions. Incidence of the naturally

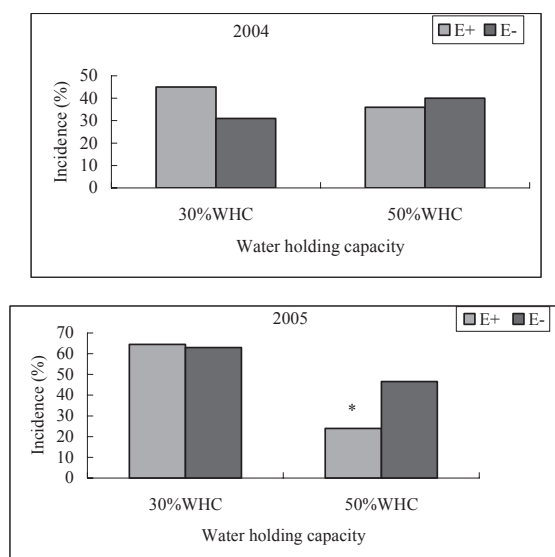


Figure 1 Effects of powdery mildew (*B. graminis*) on E+ and E- *A. inebrians* at 30% and 50% WHC soil moisture content in pot conditions. *Differences are significant between E+ and E- plants ($P<0.05$).

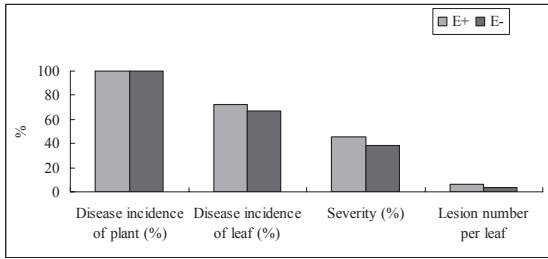


Figure 2 Incidence of E+ and E- *A. inebrians* leaf spot (*B. sorokiniana*) under field conditions.

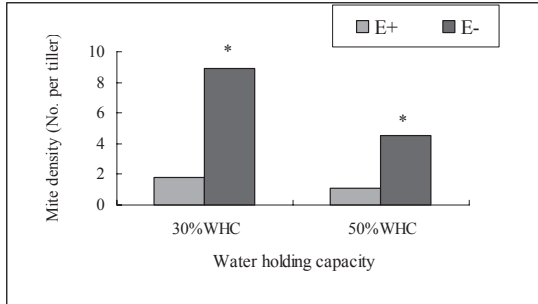


Figure 3 Density of *T. cinnabarinus* on *A. inebrians* under soil moisture content of 30% and 50% water holding capacity in greenhouse pot conditions. *Differences are significant between E+ and E- plants ($P < 0.05$).

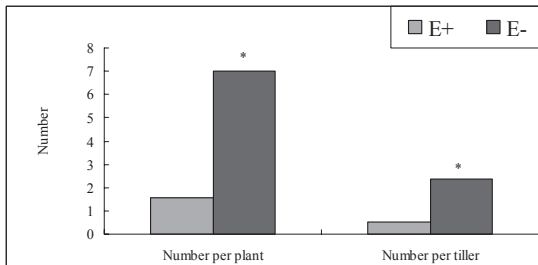


Figure 4 Density of *R. padi* on E+ and E- *A. inebrians* under field conditions. *Differences are significant between E+ and E- plants ($P < 0.05$).

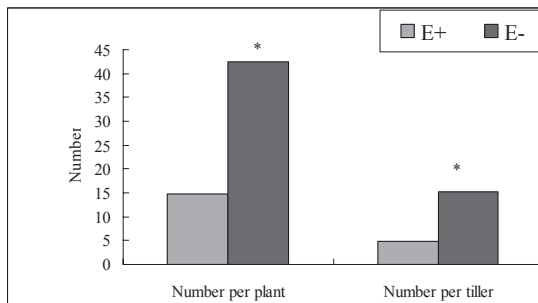


Figure 5 Density of *T. cinnabarinus* on E+ and E- *A. inebrians* under field conditions. *Differences are significant between E+ and E- plants ($P < 0.05$).

occurring diseases powdery mildew (*Blumeria graminis*) and leaf spot (*Bipolaris sorokiniana*), and numbers of mites (*Tetranychus cinnabarinus*) and bird-cherry-oat aphids (*Rhopalosiphum padi*) damage were investigated.

Results

Disease resistance of E+ and E- plants in the climate chamber

In 2004, powdery mildew (*B. graminis*) incidence on E+ and E- *A. inebrians* was similar under a soil moisture content of both 30 and 50% WHC. In 2005, incidence was again similar at 30% WHC but at 50% WHC disease incidence on E+ declined by 49% and was significantly ($P < 0.05$) lower than on E- plants (Fig. 1).

Disease resistance of E+ and E- plants in the field

In the field, the existence of *N. gansuense* did not significantly affect the incidence, severity and leaf lesion number of leaf spot (*B. sorokiniana*) on *A. inebrians* ($P > 0.05$) (Fig. 2).

Pest resistance of E+ and E- plants

In the climate chamber, regardless of drought (30% WHC) or normal (50% WHC) soil conditions, the density of *T. cinnabarinus* on E+ plants was significantly ($P < 0.05$) lower than that on E- plants. Mite levels were highest under drought conditions (Fig. 3).

In the field, the densities of *R. padi* on E+ plants were significantly ($P < 0.05$) lower than that on E- plants (Fig. 4), as was the case for *T. cinnabarinus* (Fig. 5).

Discussion

Epichloë endophytes (*Neotyphodium* and *Epichloë* spp.) may enhance the abilities of grasses to resist pathogens by inhibiting sporulation (Li 2005) and spore germination (Holzmann-Wirth *et al.* 2000), preventing invasion and colonisation, delaying or reducing subsequent lesion development (Nan & Li 2000 2004), and deterring virus-vector insects as well (West *et al.* 1990; Latch 1993). Many studies have shown that pure cultures of many epichloë endophytes inhibit growth of plant-pathogenic fungi (White & Cole 1985; Christensen & Latch 1991; Holzmann-Wirth *et al.* 2000; Siegel & Latch 1989).

In the climate chamber pot trial, effects of endophytes of *A. inebrians* on powdery mildew varied according to the environment (Fig. 1), but in the field neither incidence nor severity of *Bipolaris* leaf spot was affected by endophyte infection (Fig. 2). Other researchers have reported some instances of enhanced disease resistance with *Neotyphodium* or *Epichloë* infection, but not always (Siegel & Latch 1991; Latch 1993; Nan & Li 2004; Popay & Bonos 2005). For example, tall fescue (*Festuca arundinacea*) seedling stem rust (*Puccinia graminis* subsp. *graminicola*) infection was unaffected by *N. coenophialum* infection under glasshouse conditions (Welty *et al.* 1991). Also, *N. coenophialum* did not reduce tall fescue root rot by *Cochliobolus sativus* (Trevathan 1996), root disease by *Pythium aphanidermatum* (Blank *et al.* 1993) or foot rot by *Fusarium oxysporum* and *F. equiseti* (Hume *et al.* 1997). A few studies indicated that endophytes could improve resistance to diseases such as turfgrass blight caused by *Pythium* (Funk *et al.* 1994), purple spot on timothy grass caused by *Cladosporium phlei* (Shimanuki 1987), seedling disease of tall fescue caused by *Rhizoctonia zeae* (Gwinn & Gavin 1992), and dollar spot caused by *Sclerotinia homeocarpa* (Clarke *et al.* 2006). Also, detached leaves of E+ *Elymus cylindricus* had fewer and smaller lesions than those on E- plants 3 days after inoculation with *A. alternata*, *F. avenaceum*, *F. culmorum*, *F. equiseti* and *F. oxysporum* (Nan

& Li 2000). E+ tall fescue plants were more resistant to crown rust (*P. coronata*) under field conditions (West *et al.* 1989) and *N. lolii* infected perennial ryegrass plants were observed to be more resistant to leaf spot (*Pyrenophora semeniperda*) than uninfected plants (Wheatley *et al.* 2000).

Grass disease resistance is influenced by many biotic and non-biotic factors, including variety, pathogen type, environmental conditions, endophyte status and so on (Siegel & Latch 1991; Latch 1993). Our research showed variation of powdery mildew incidence between 30% and 50% WHC soil conditions, and between E+ and E- plants (Figs. 1 & 2). Moreover, different isolates may produce different quantities and types of antifungal substances and some probably confer different agronomic characters on the host plant (Christensen & Latch 1991). Two isolates of *N. uncinatum* and three of four isolates of *Epichloë festucae* gave no inhibition of *D. erythrospila* and *R. zeae* (Christensen 1996).

This is the first report of an evaluation of endophytic grass resistance to pests in China. Our results showed that *Neotyphodium* infection could significantly ($P < 0.05$) reduce densities of *R. padi* and *T. cinnabarinus* on E+ *A. inebrians* compared to E- plants (Fig. 3-5). At least four species of aphids are deterred by grass endophytes: *Diuraphis noxia*, *R. padi*, *Schizaphis graminum* and *Sipha flava* (Siegel *et al.* 1990; Latch 1993; Schardl & Phillips 1997), but only one species of Acarina mites, *Abacarus hystrix* (Li & Nan 2004). This trial shows another mite, *T. cinnabarinus* is deterred by endophytic *A. inebrians*. High levels of two main ergot alkaloids, ergonovine and ergine, are produced in endophytic *A. inebrians* (Miles *et al.* 1996; Li *et al.* 2006). Of the more than 10 known alkaloids associated with grass endophytes, peramine and loline have proved more toxic to insects than have ergot alkaloids and indole diterpenes (Bacon 1995; Schardl & Phillips 1997; Faeth & Bultman 2002). It is possible the very high levels of ergot alkaloids or other substances produced in the interactions between *A. inebrians* and *N. gansuense*, deterred *R. padi* and *T. cinnabarinus*. Future studies to elucidate the chemical basis for invertebrate deterrence would be of considerable practical importance for the use or manipulation of endophytes as bioprotective agents.

ACKNOWLEDGEMENTS

We thank graduate students Xiaoyuan Gou, Jiahui Gao, Pei Tian, Bin Ma and Bin Nie for partial assistance. This research was financially supported by the National High Technology Research and Development Program of China (2004AA244080), Gansu Middle and Young Scientist Foundation (3ZS041-A25-003) and National Nature Science Foundation of China (30070546).

REFERENCES

Bacon, C.W. 1995. Toxic endophyte-infected tall fescue and range grasses. Historic perspectives. *Journal of Animal Science* 73: 861-870.

Blank, C.A.; Gwinn, K.D.; Gavin, A.M. 1993. Tolerance of tall fescue to soilborne pathogens is influenced by *Acremonium coenophialum*. pp. 145-150. *In: Proceedings of the 2nd International Symposium on Acremonium/Grass Interactions.*

Christensen, M.J.; Latch, G.C.M. 1991. Variation among isolates of *Acremonium* endophytes (*A. coenophialum* and possibly *A. typhinum*) from tall fescue (*Festuca arundinacea*). *Mycological Research* 95: 1123-1126.

Christensen, M.J. 1996. Antifungal activity in grasses infected with *Acremonium* and *Epichloë* endophytes. *Australasian*

Plant Pathology 25: 186-191.

Christensen, M.J.; Latch, G.C.M.; Tapper, B.A. 1991. Variation within isolates of *Acremonium* endophytes from perennial ryegrasses. *Mycological Research* 95: 918-923.

Clarke, B.B.; White, J.F.; Hurley, R.H.; Torres, M.S.; Sun, S.; Huff, D.R. 2006. Endophyte-mediated suppression of dollar spot disease in fine fescues. *Plant Disease* 90: 994-998.

Faeth, S.H.; Bultman, T.L. 2002. Endophytic fungi and interactions among host plants, herbivores and natural enemies. pp. 89-123. *In: Multitrophic Level Interactions.* Eds. Tschamfre T.; Hawkins B.A. Cambridge University Press, Cambridge, UK.

Funk, C.R.; Belanger, F.C.; Murphy, J.A. 1994. Role of endophytes in grasses used for turf and soil conservation. pp. 201-208. *In: Biotechnology of Endophytic Fungi of Grasses.* Eds. Bacon C.W.; White J.F. CRC Press, Florida, USA.

Gwinn, K.D.; Gavin, A.M. 1992. Relationship between endophyte infestation level of tall fescue seed lots and *Rhizoctonia zeae* seedling disease. *Plant Disease* 76: 911-914.

Holzmann-Wirth, A.; Dapprich, P.; Eierdanz, S.; Heerz, D.; Paul, V.H. 2000. Anti-fungal substances extracted from *Neotyphodium* endophytes. pp. 65-69. *In: Proceedings of the 3rd International Conference on Harmful and Beneficial Microorganisms in Grassland, Pasture and Turf.*

Hume, D.E.; Quigley, P.E.; Aldaoud, R. 1997. Influence of *Neotyphodium* infection on plant survival of diseased tall fescue and ryegrass. pp. 171-173. *In: Neotyphodium/Grass Interactions.* Eds. Bacon C.W. and Hill N.S. Plenum Press, New York, USA.

Latch, G.C.M. 1993. Physiological interactions of endophytic fungi and their hosts. Biotic stress tolerance imparted to grasses by endophytes. *Agriculture, Ecosystems and Environment* 44: 143-156.

Li, C.J. 2005. Biological and ecological characteristics of *Achnatherum inebrians* / *Neotyphodium* endophyte symbiont. PhD Dissertation, Lanzhou University, China.

Li, C.J.; Gao, J.H.; Ma, B. 2003. Seven diseases of drunken horse grass (*Achnatherum inebrians*) in China (In Chinese with English abstract). *Pratacultural Science* 20 (11): 51-53.

Li, C.J.; Nan, Z.B.; Gao, J.H.; Tian, P. 2004a. Detection and distribution of *Neotyphodium* - *Achnatherum inebrians* association in China. #210. *In: Proceedings of 5th International Symposium on Neotyphodium/Grass Interactions*, Fayetteville, Arkansas.

Li, C.J.; Nan, Z.B.; Volker, H.P.; Dapprich, P.; Liu, Y. 2004b. A new *Neotyphodium* species symbiotic with drunken horse grass (*Achnatherum inebrians*) in China. *Mycotaxon* 90: 141-147.

Li, C.J.; Nan, Z.B.; Schardl, C.L. 2006. Levels and temporal variation of ergot alkaloids in endophyte-infected drunken horse grass, *Achnatherum inebrians*, in China. pp. 203-204. *In: APS, CPS and MSA Joint Meeting Abstracts*, Quebec City, Canada.

Miles, C.O.; Lane, G.A.; Menna, M.E.; Garthwaite, I.; Piper, E.L.; Ball, O.J.P.; Latch, G.C.M.; Allen, J.M.; Hunt, M.B.; Bush, L.P.; Feng, K.M.; Fletcher, I.; Harris, P.S. 1996. High levels of ergonovine and lysergic acid amide in toxic *Achnatherum inebrians* accompany infection by an *Acremonium*-like endophytic fungus. *Journal of Agriculture and Food Chemistry* 44: 1285-1290.

Nan, Z.B.; Li, C.J. 2000. *Neotyphodium* in native grasses in China and observations on endophyte/host interactions. pp. 41-50. *In: Proceedings of 4th International Neotyphodium /Grass Interactions Symposium*, Eds. Volker, H.P. and Dapprich, P.D. Soest Germany.

- Nan, Z.B.; Li, C.J. 2004. Roles of the grass-*Neotyphodium* association in pastoral agriculture systems. *Acta Ecologica Sinica* 24: 605-616.
- Popay, A.J.; Baltus, J.G.; Pennell C.G.L. 2000. Insect resistance in perennial ryegrass infected with toxin-free *Neotyphodium* endophytes. pp. 187-193. *In: Proceedings of 4th International Neotyphodium /Grass Interactions Symposium*, Eds. Volker, H.P. and Dapprich, P.D. Soest, Germany.
- Popay, A.J.; Bonos, S.A. 2005. Biotic responses in endophytic grasses. pp. 163-185. *In: Neotyphodium in Cool-season Grasses*. Eds. Roberts C.A.; West, C.P.; Spiers, D.E. Arkansas, USA.
- Schardl, C.L.; Phillips, T.D. 1997. Protective grass endophytes where are they from and where are they going? *Plant Disease* 81: 430-438.
- Shimanuki, T. 1987. Studies on the mechanisms of the infection of timothy with purple spot disease caused by *Cladosporium phlei* (Gregory) de Vries. *Research Bulletin of the Hokkaido National Agricultural Experiment Station* 148: 1-5.
- Siegel, M.R.; Latch, G.C.M. 1989. Expression of antifungal activity in agar culture by isolates of grass endophytes. *Mycologia* 83: 529-537.
- Siegel, M.R.; Latch, G.C.M.; Bush, L.P.; Fannin, N.F.; Rowan, D.D.; Tapper, B.A.; Bacon, C.W.; Johnson, M.C. 1990. Fungal endophyte-infected grasses: alkaloid accumulation and aphid response. *Journal of Chemical Ecology* 16: 3301-3315.
- Siegel, M.R.; Latch, G.C.M.; Johnson, M.C. 1987. Fungal endophytes of grasses. *Annual Review of Phytopathology* 25: 293-315.
- Trevathan, L.E. 1996. Performance of endophyte-free and endophyte-infected tall fescue seedlings in soil infected with *Cochliobolus sativus*. *Canadian Journal of Plant Pathology* 18: 415-418.
- Welty, R.E.; Barker, R.E.; Azevedo, M.D. 1991. Reaction of tall fescue infected and noninfected by *Acremonium coenophialum* to *Puccinia graminis* subsp. *graminicola*. *Plant Disease* 75: 883-886.
- West, C.P.; Gwinn, K.D. 1993. Role of *Acremonium* in drought, pest and disease tolerances of grasses. pp. 11-30. *In: Proceedings of the 2nd International Symposium on Acremonium/Grass Interactions*.
- West, C.P.; Izeckor, E.; Robbins, R.T.; Gergerich, R.; Mahmood, T. 1990. *Acremonium coenophialum* effects on infestations of barley yellow dwarf virus and soil-borne nematodes and insects in tall fescue. pp. 196-198. *In: Proceedings of International Symposium on Neotyphodium/Grass Interactions*.
- West, C.P.; Turner, K.; Phillips, J.M. 1989. Arkansas report. pp. 23-25. *In: Proceedings of Tall fescue Toxicosis Workshop*.
- Wheatley, W.M.; Nicol, H.I.; Hunt, E.R.; Nikandrow, A.; Cother, N. 2000. An association between perennial ryegrass endophyte, a leafspot caused by *Pyrenophora semeniperda* and preferential grazing by sheep. pp. 71-75. *In: Proceedings of The 3rd International Conference on Harmful and Beneficial Microorganisms in Grassland, Pasture and Turf*.
- White, J.F.; Cole, G.T. 1985. Endophyte-host association in forage grasses. III. *In vitro* inhibition of fungi by *Acremonium coenophialum*. *Mycologia* 77: 487-489.