



Article – special issue

Doi 10.5943/mycosphere/si/1b/13

Copyright © Guizhou Academy of Agricultural Sciences

***Botryosphaeriaceae*: Current status of genera and species**

Dissanayake AJ^{1,3,*}, Phillips AJL^{2,*,#}, Li XH^{1,#}, Hyde KD³

¹ Institute of Plant and Environment Protection, Beijing Academy of Agriculture and Forestry Sciences, Beijing 100097, People's Republic of China.

² University of Lisbon, Faculty of Sciences, Biosystems and Integrative Sciences Institute (BioISI), Campo Grande, 1749-016 Lisbon, Portugal.

³ Center of Excellence in Fungal Research, Mae Fah Luang University, Chiang Rai 57100, Thailand.

* These authors contributed equally to this work.

Corresponding authors: AJL Phillips, XH Li.

Dissanayake AJ, Phillips AJL, Li XH, Hyde KD 2016 – *Botryosphaeriaceae*: Current status of genera and species. *Mycosphere* 7(7), 1001–1073, Doi 10.5943/mycosphere/si/1b/13

Abstract

The *Botryosphaeriaceae* comprises endophytes, saprobes and plant pathogens. Recently, some taxa in *Botryosphaeriaceae* have undergone nomenclatural changes. This paper represents the first comprehensive overview of *Botryosphaeriaceae* since 2013. Twenty three genera and 187 species of *Botryosphaeriaceae* are listed alphabetically, brief notes for species in each genus are provided. Information is provided for the location of ex-type isolates, new host records and geographical distribution of species. Available DNA sequences from ex-type isolates are listed in a table and multilocus phylogenetic trees for major genera in *Botryosphaeriaceae* are provided.

Key words – Botryosphaeriales – phylogeny – systematics – taxonomy

Introduction

Theissen & Sydow (1918) introduced the *Botryosphaeriaceae* for three genera, namely, *Botryosphaeria*, *Phaeobotryon* and *Dibotryon*. Over the years the family and genera have undergone several taxonomic revisions and updates. Introduction of new genera, together with the addition of separate names for sexual and asexual genera resulted in at least 78 genera within this family. In addition, the type genus (*Botryosphaeria*) was subject to several changes that broadened the concept resulting in a diverse range of morphologies, especially in the asexual states, associated with a single sexual genus. Indeed the concept of the sexual state incorporated taxa with ascospores that could be hyaline or coloured, septate or aseptate. As a result, the genus *Botryosphaeria* and the family *Botryosphaeriaceae* became unmanageable and confused.

In a phylogenetic approach based on the 28S rRNA gene Crous et al. (2006) revealed 10 lineages within *Botryosphaeriaceae*. They considered these lineages to represent individual genera that could be recognised on the morphology of their sexual morphs. The existing anamorph genera names were used for most of the genera but new names were introduced for *Neofusicoccum* and *Pseudofusicoccum*. However, some of the clades could not be fully resolved, notably the one comprising *Diplodia/Lasiodiplodia/Tiarosporella*. Nevertheless, that study provided a stable base for future work on systematics of the family.

Phillips et al. (2008) resolved the status of species with brown ascospores, introduced or re-instated a further five genera in *Botryosphaeriaceae* and clarified the status of *Diplodia* and *Lasiodiplodia*. Damm et al. (2007) showed that *Aplosporella* resides in *Botryosphaeriaceae*, Rojas et al. (2008) determined that *Endomelanconiopsis* also resides there and Phillips & Alves considered *Melanops* to be yet another genus. Liu et al. (2012) used a phylogenetic approach together with a study of herbarium specimens and introduced two further genera namely *Botryobambusa* and *Cophinforma* bringing the total number of genera known from culture to 21.

Wikee et al. (2013) re-instated *Phyllostictaceae* for *Phyllosticta*, Slippers et al. (2013) introduced *Saccharataceae* for *Saccharata*, *Aplosporellaceae* for *Aplosporella* and *Melanopsaceae* for *Melanops*. Phillips et al. (2013) provided comprehensive descriptions, notes and phylogenies for the 17 genera and 110 species known from culture that remained in the family. Since then 6 new genera and 85 new species/species combinations have been introduced (Table 1). In this paper we provide an updated list of all genera and species known from culture in the *Botryosphaeriaceae* together with relevant data on types and ex-type cultures. We also include updated information on hosts and known geographic distribution. Backbone trees for the larger genera (*Botryosphaeria*, *Diplodia*, *Dothiorella/Spencermartinsia*, *Lasiodiplodia* and *Neofusicoccum*) are included as well as an overview tree of the Botryosphaeriales showing the currently accepted limits of the *Botryosphaeriaceae*. In preparing this paper we included novel taxa and taxonomic changes published up to and including 31 October 2016 as well as those published in this issue of *Mycosphere*.

Materials and methods

The genera and species included in Phillips et al. (2013) was used as the starting point. Genera and species not included in that paper, or introduced since 2013 were found in searches of MycoBank and the mycological literature in general. All names were checked for validity, and for the existence of cultures and relevant sequences in GenBank before they were included in the current list. All genera and species known from culture are listed in Table 1. The USDA fungal database (Farr & Rossman 2016) was used to gather information on host association and geographic distribution. Additional new disease reports, distribution and hosts were retrieved from recent publications.

Phylogenetic analysis

Sequences of internal transcribed spacer (ITS), translation elongation factor (*tef1- α*) and 28S large subunit ribosomal gene (LSU) were download from NCBI GenBank (<https://www.ncbi.nlm.nih.gov/genbank/>). An overview backbone phylogenetic tree for the order Botryosphaeriales was constructed from ITS, *tef1- α* and LSU. Separate phylogenetic trees of the larger genera (*Botryosphaeria*, *Diplodia*, *Dothiorella/Spencermartinsia*, *Lasiodiplodia* and *Neofusicoccum*) were constructed from combined ITS and *tef1- α* sequences.

Single gene regions were aligned with Clustal X1.81 (Thompson et al. 1997) and combined with BioEdit v. 7.0.9.0 (Hall 1999). Further alignment of the sequences was done using default settings of MAFFT v. 7 (Kato & Toh 2008; <http://mafft.cbrc.jp/alignment/server/>) and manually adjusted where necessary using BioEdit v. 7.0.9.0 (Hall 1999). Maximum Parsimony analysis (MP) was performed with PAUP (Phylogenetic Analysis Using Parsimony) v. 4.0b10 (Swofford 2002). Gaps were treated as missing data and ambiguously aligned regions were excluded. Trees were inferred using the heuristic search option with Tree Bisection Reconnection branch swapping and 1000 random sequence additions. Maxtrees was set at 1000, branches of zero length were collapsed and all multiple parsimonious trees were saved. Descriptive tree statistics for parsimony (tree length, consistency index, retention index, rescaled consistency index, and homoplasy index) were calculated for trees generated under different optimality criteria.

The best model of evolution for each gene region was determined using MRMODELTEST v. 2.2 (Nylander 2004) and maximum likelihood analyses were performed in RAXML GUI v. 0.9b2 (Silvestro & Michalak 2010). The RAXML analyses were run with a rapid bootstrap analysis

Table 1 GenBank accession numbers of genera and species in *Botryosphaeriaceae* and representative genera and species in order Botryosphaeriales. Type species are given in bold typeface. Isolates and sequence accession numbers of *Botryosphaeriaceae* genera and species introduced since Phillips et al. (2013) are italicized.

Species	Cultures	ITS	<i>tefl-a</i>	LSU
<i>Botryosphaeriaceae</i>				
<i>Alanphillipsia aloecicola</i>	CBS 138896	KP004444	N/A	KP004472
<i>Alanphillipsia aloeigena</i>	CBS 136408	KF777137	N/A	KF777193
<i>Alanphillipsia aloes</i>	CBS 136410	KF777138	N/A	KF777194
<i>Alanphillipsia aloetica</i>	CBS 136409	KF777139	N/A	KF777195
<i>Alanphillipsia euphorbiae</i>	CBS 136411	KF777140	N/A	KF777196
<i>Barriopsis archontophoenicis</i>	MFLUCC 14-1164	KX235306	KX235305	KX235307
<i>Barriopsis fusca</i>	CBS 174.26	EU673330	EU673296	DQ377857
<i>Barriopsis iraniana</i>	IRAN 1448C	FJ919663	FJ919652	KF766318
<i>Barriopsis iraniana</i>	IRAN1449C	FJ919665	FJ919654	N/A
<i>Barriopsis tectonae</i>	MFLUCC 12-0381	KJ556515	KJ556516	N/A
<i>Botryobambusa fusicoccum</i>	MFLUCC 11-0143	JX646792	JX646857	JX646809
<i>Botryobambusa fusicoccum</i>	MFLUCC 11-0657	JX646793	JX646858	JX646810
<i>Botryosphaeria agaves</i>	MFLUCC 11-0125	JX646791	JX646856	JX646808
<i>Botryosphaeria agaves</i>	MFLUCC 10-0051	JX646790	JX646855	JX646807
<i>Botryosphaeria auasmontanum</i>	CMW 25413	KF766167	EU101348	KF766332
<i>Botryosphaeria corticis</i>	CBS 119047	DQ299245	EU017539	EU673244
<i>Botryosphaeria corticis</i>	ATCC 22927	DQ299247	EU673291	EU673245
<i>Botryosphaeria dothidea</i>	CBS 115476	AY236949	AY236898	DQ377852
<i>Botryosphaeria dothidea</i>	CBS 110302	AY259092	AY573218	EU673243
<i>Botryosphaeria fabicerciana</i>	CBS 127193	HQ332197	HQ332213	N/A
<i>Botryosphaeria fabicerciana</i>	CMW 27108	HQ332200	HQ332216	N/A
<i>Botryosphaeria fusispora</i>	MFLUCC 10-0098	JX646789	JX646854	JX646806
<i>Botryosphaeria fusispora</i>	MFLUCC 11-0507	JX646788	JX646853	JX646805
<i>Botryosphaeria minutispermata</i>	GZCC 16-0013	KX447675	KX447678	N/A
<i>Botryosphaeria minutispermata</i>	GZCC 16-0014	KX447676	KX447679	N/A
<i>Botryosphaeria ramosa</i>	CBS 122069	EU144055	EU144070	N/A
<i>Botryosphaeria scharifii</i>	CBS 124703	JQ772020	JQ772057	N/A
<i>Botryosphaeria scharifii</i>	CBS 124702	JQ772019	JQ772056	N/A
<i>Botryosphaeria sinensia</i>	CGMCC 3.17722	KT343254	KU221233	N/A
<i>Botryosphaeria sinensia</i>	CGMCC 3.17724	KT343256	KU221234	N/A
<i>Cophinforma atrovirens</i>	MFLUCC 11-0425	JX646800	JX646865	JX646817
<i>Cophinforma atrovirens</i>	MFLUCC 11-0655	JX646801	JX646866	JX646818
<i>Cophinforma mamane</i>	CBS 117444	KF531822	KF531801	DQ377855
<i>Diplodia africana</i>	CBS 120835	EF445343	EF445382	N/A
<i>Diplodia africana</i>	CBS 121104	EF445344	EF445383	N/A
<i>Diplodia agrifolia</i>	CBS 132777	JN693507	JQ517317	N/A
<i>Diplodia agrifolia</i>	UCROK1429	JQ411412	JQ512121	N/A
<i>Diplodia alatafructa</i>	CBS 124931	FJ888460	FJ888444	N/A
<i>Diplodia alatafructa</i>	CBS 124933	FJ888478	FJ888446	N/A
<i>Diplodia allocellula</i>	CBS 130408	JQ239397	JQ239384	JQ239410
<i>Diplodia allocellula</i>	CBS 130410	JQ239399	JQ239386	JQ239412
<i>Diplodia bulgarica</i>	CBS 124254	GQ923853	GQ923821	N/A
<i>Diplodia bulgarica</i>	CBS 124135	GQ923852	GQ923820	N/A
<i>Diplodia corticola</i>	CBS 112549	AY259100	AY573227	AY928051
<i>Diplodia corticola</i>	CBS 112546	AY259110	DQ458872	EU673262
<i>Diplodia crataegicola</i>	MFLU 15-1311	KT290244	KT290248	N/A
<i>Diplodia cupressi</i>	CBS 168.87	DQ458893	DQ458878	EU673263

Species	Cultures	ITS	<i>tef1-a</i>	LSU
<i>Diplodia cupressi</i>	CBS 261.85	DQ458894	DQ458879	EU673264
<i>Diplodia estuarina</i>	CMW41231	KP860831	KP860676	N/A
<i>Diplodia estuarina</i>	CMW41230	KP860830	KP860675	N/A
<i>Diplodia fraxinii</i>	CBS 136010	KF307700	KF318747	N/A
<i>Diplodia galiicola</i>	MFLU 15-1310	KT290245	KT290249	N/A
<i>Diplodia insularis</i>	CBS 140350	KX833072	KX833073	N/A
<i>Diplodia intermedia</i>	CBS 124462	GQ923858	GQ923826	N/A
<i>Diplodia intermedia</i>	CBS 124134	HM036528	GQ923851	N/A
<i>Diplodia malorum</i>	CBS 124130	GQ923865	GQ923833	N/A
<i>Diplodia malorum</i>	CBS 112554	AY259095	DQ458870	N/A
<i>Diplodia mutila</i>	CBS 112553	AY259093	AY573219	AY928049
<i>Diplodia mutila</i>	CBS 230.30	DQ458886	DQ458869	AY928049
<i>Diplodia neojuniperi</i>	CPC 22753	KM006431	KM006462	N/A
<i>Diplodia olivarum</i>	CBS 121887	EU392302	EU392279	N/A
<i>Diplodia olivarum</i>	CBS 121886	EU392297	EU392274	N/A
<i>Diplodia pseudoseriata</i>	CBS 124906	EU080927	EU863181	N/A
<i>Diplodia pseudoseriata</i>	CBS 124907	EU080922	EU863179	N/A
<i>Diplodia quercivora</i>	CBS 133852	JX894205	JX894229	N/A
<i>Diplodia rosacearum</i>	CBS 141915	KT956270	KU378605	N/A
<i>Diplodia rosulata</i>	CBS 116470	EU430265	EU430267	DQ377896
<i>Diplodia rosulata</i>	CBS 116472	EU430266	EU430268	DQ377897
<i>Diplodia sapinea</i>	CBS 393.84	DQ458895	DQ458880	DQ377893
<i>Diplodia sapinea</i>	CBS 109725	DQ458896	DQ458881	EU673270
<i>Diplodia scrobiculata</i>	CBS 118110	AY253292	AY624253	N/A
<i>Diplodia scrobiculata</i>	CBS 109944	DQ458899	DQ458884	EU673268
<i>Diplodia scrobiculata</i>	CBS 113423	DQ458900	DQ458885	EU673267
<i>Diplodia seriata</i>	CBS 112555	AY259094	AY573220	AY928050
<i>Diplodia seriata</i>	CBS 119049	DQ458889	DQ458874	EU673266
<i>Diplodia subglobosa</i>	CBS 124133	GQ923856	GQ923824	N/A
<i>Diplodia tsugae</i>	CBS 418.64	DQ458888	DQ458873	DQ377867
<i>Dothiorella acacicola</i>	CBS 141295	KX228269	KX228376	KX228320
<i>Dothiorella americana</i>	CBS 128309	HQ288218	HQ288262	N/A
<i>Dothiorella americana</i>	CBS 128310	HQ288219	HQ288263	N/A
<i>Dothiorella brevicollis</i>	CBS 130411	JQ239403	JQ239390	JQ239416
<i>Dothiorella californica</i>	CBS 141587	KX357188	KX357211	N/A
<i>Dothiorella capri-amissi</i>	CMW 25403	EU101323	EU101368	N/A
<i>Dothiorella capri-amissi</i>	CMW 25404	EU101324	EU101369	N/A
<i>Dothiorella casuarini</i>	CBS 120688	DQ846773	DQ875331	N/A
<i>Dothiorella casuarini</i>	CBS 120690	DQ846774	DQ875333	N/A
<i>Dothiorella dulcispinae</i>	CBS 130413	JQ239400	JQ239387	JQ239413
<i>Dothiorella iberica</i>	CBS 115041	AY573202	AY573222	AY928053
<i>Dothiorella iberica</i>	CBS 113188	AY573198	EU673278	EU673230
<i>Dothiorella iranica</i>	IRAN1587C	KC898231	KC898214	N/A
<i>Dothiorella longicollis</i>	CBS 122068	EU144054	EU144069	N/A
<i>Dothiorella longicollis</i>	CBS 122066	EU144052	EU144067	N/A
<i>Dothiorella moneti</i>	MUCC505	EF591920	EF591971	EF591937
<i>Dothiorella moneti</i>	MUCC 507	EF591922	EF591973	EF591939
<i>Dothiorella neclivorem</i>	DAR80992	KJ573643	KJ573640	N/A
<i>Dothiorella oblonga</i>	CMW 25407	EU101300	EU101345	N/A
<i>Dothiorella omnivora</i>	CBS 140349	KP205497	KP205470	N/A
<i>Dothiorella omnivora</i>	CBS 188.87	EU673316	EU673283	N/A
<i>Dothiorella parva</i>	IRAN1579C	KC898234	KC898217	N/A
<i>Dothiorella parva</i>	IRAN1585C	KC898235	KC898218	N/A

Species	Cultures	ITS	<i>tef1-a</i>	LSU
<i>Dothiorella pretoriensis</i>	CBS 130404	JQ239405	JQ239392	JQ239418
<i>Dothiorella pretoriensis</i>	CBS 130403	JQ239406	JQ239393	JQ239419
<i>Dothiorella prunicola</i>	CBS 124723	EU673313	EU673280	EU673232
<i>Dothiorella rhamnii</i>	MFLUCC 14-0902	KU246381	N/A	KU246382
<i>Dothiorella santali</i>	MUCC 509	EF591924	EF591975	EF591941
<i>Dothiorella santali</i>	MUCC 508	EF591923	EF591974	EF591940
<i>Dothiorella sarmentorum</i>	IMI 63581b	AY573212	AY573235	AY928052
<i>Dothiorella sarmentorum</i>	CBS 115038	AY573206	AY573223	DQ377860
<i>Dothiorella sempervirentis</i>	IRAN1583C	KC898236	KC898219	N/A
<i>Dothiorella sempervirentis</i>	IRAN1581C	KC898237	KC898220	N/A
<i>Dothiorella striata</i>	ICMP16824	EU673320	EU673287	EU673240
<i>Dothiorella striata</i>	ICMP16819	EU673320	EU673287	EU673239
<i>Dothiorella symphoricarposicola</i>	MFULCC 13-0497	KJ742378	KJ742381	N/A
<i>Dothiorella symphoricarposicola</i>	MFLUCC 13-0196	KU234782	KU234796	N/A
<i>Dothiorella tectonae</i>	MFLUCC12-0382	KM396899	KM409637	N/A
<i>Dothiorella thailandica</i>	CBS 133991	JX646796	JX646861	JX646813
<i>Dothiorella thripsita</i>	BRIP 51876	FJ824738	KJ573639	N/A
<i>Dothiorella ulmacea</i>	CBS 138855	KR611881	KR611910	KR611899
<i>Dothiorella uruguayensis</i>	CBS 124908	EU080923	EU863180	N/A
<i>Dothiorella vidmadera</i>	DAR78992	EU768874	EU768881	N/A
<i>Dothiorella vidmadera</i>	DAR78993	EU768876	EU768882	N/A
<i>Dothiorella vinea-gemmae</i>	DAR81012	KJ573644	KJ573641	N/A
<i>Endomelanconiopsis endophytica</i>	CBS 120397	EU683656	EU683637	EU683629
<i>Endomelanconiopsis endophytica</i>	CBS 122550	EU683664	EU683645	EU683634
<i>Endomelanconiopsis microspora</i>	CBS 353.97	EU683655	EU683636	EU683628
<i>Eutiarosporella africana</i>	CBS 133854	KC769956	KC769852	KC769990
<i>Eutiarosporella africana</i>	CBS 135850	KC769957	KC769853	KC769991
<i>Eutiarosporella dactylidis</i>	MFLUCC 13-0276	KM978944	KP031694	KM978949
<i>Eutiarosporella dactylidis</i>	MFLUCC 15-0915	KU246378	N/A	KU246380
<i>Eutiarosporella darliae</i>	DAR 82491	KP309793	KP309805	N/A
<i>Eutiarosporella darliae</i>	DAR 82493	KP309786	KP309807	N/A
<i>Eutiarosporella pseudodarliae</i>	DAR 82489	KP309796	KP309797	N/A
<i>Eutiarosporella pseudodarliae</i>	DAR 82490	KP309794	KP309800	N/A
<i>Eutiarosporella tritici</i>	CBS 118719	KC769961	KF531809	DQ377941
<i>Eutiarosporella tritici-australis</i>	DAR 82485	KP309788	KP309799	N/A
<i>Eutiarosporella tritici-australis</i>	DAR 82486	KP309787	KP309804	N/A
<i>Eutiarosporella urbis-rosarum</i>	CBS 130405	JQ239407	JQ239394	JQ239420
<i>Eutiarosporella urbis-rosarum</i>	CBS 130406	JQ239408	JQ239395	JQ239421
<i>Lasiodiplodia avicenniae</i>	CMW41467	KP860835	KP860680	N/A
<i>Lasiodiplodia avicenniae</i>	LAS199	KU587957	KU587946	N/A
<i>Lasiodiplodia brasiliense</i>	CMM 4015	JX464063	JX464049	N/A
<i>Lasiodiplodia brasiliense</i>	CMM 2185	KC484800	KC481530	N/A
<i>Lasiodiplodia bruguierae</i>	CMW41470	KP860832	KP860677	N/A
<i>Lasiodiplodia bruguierae</i>	CMW41614	KP860833	KP860678	N/A
<i>Lasiodiplodia caatinguensis</i>	CMM1325	KT154760	KT008006	N/A
<i>Lasiodiplodia caatinguensis</i>	IBL352	KT154759	KT154753	N/A
<i>Lasiodiplodia citricola</i>	CBS 124707	GU945354	GU945340	N/A
<i>Lasiodiplodia citricola</i>	CBS 124706	GU945353	GU945339	N/A
<i>Lasiodiplodia crassispora</i>	CBS 118741	DQ103550	EU673303	DQ377901
<i>Lasiodiplodia crassispora</i>	WAC 12534	DQ103551	DQ103558	N/A
<i>Lasiodiplodia crassispora</i>	CBS 110492	EF622086	EF622066	N/A
<i>Lasiodiplodia euphorbicola</i>	CMM3609	KF234543	KF226689	N/A
<i>Lasiodiplodia euphorbicola</i>	IBL329	KT247490	KT247492	N/A

Species	Cultures	ITS	<i>tef1-a</i>	LSU
<i>Lasiodiplodia exigua</i>	CBS 137785	KJ638317	KJ638336	N/A
<i>Lasiodiplodia exigua</i>	BL184	KJ638318	KJ638337	N/A
<i>Lasiodiplodia gilanensis</i>	CBS 124704	GU945351	GU945342	N/A
<i>Lasiodiplodia gilanensis</i>	CBS 124705	GU945352	GU945341	N/A
<i>Lasiodiplodia gonubiensis</i>	CBS 115812	AY639595	DQ103566	N/A
<i>Lasiodiplodia gonubiensis</i>	CBS 116355	AY639594	DQ103567	N/A
<i>Lasiodiplodia gravistriata</i>	CMM 4564	KT250949	KT250950	N/A
<i>Lasiodiplodia gravistriata</i>	CMM 4565	KT250947	KT266812	N/A
<i>Lasiodiplodia hormozganensis</i>	CBS 124709	GU945355	GU945343	N/A
<i>Lasiodiplodia hormozganensis</i>	CBS 124708	GU945356	GU945344	N/A
<i>Lasiodiplodia iraniensis</i>	CBS 124710	GU945346	GU945334	N/A
<i>Lasiodiplodia iraniensis</i>	CBS 124711	GU945347	GU945335	N/A
<i>Lasiodiplodia laeliocattleyae</i>	CBS 167.28	KU507487	KU507454	N/A
<i>Lasiodiplodia lignicola</i>	CBS 134112	JX646797	KU887003	N/A
<i>Lasiodiplodia macrospora</i>	CMM3833	KF234557	KF226718	N/A
<i>Lasiodiplodia mahajangana</i>	CBS 124927	FJ900597	FJ900643	N/A
<i>Lasiodiplodia mahajangana</i>	CBS 124925	FJ900595	FJ900641	N/A
<i>Lasiodiplodia margaritacea</i>	CBS 122519	EU144050	EU144065	N/A
<i>Lasiodiplodia margaritacea</i>	CBS 122065	EU144051	EU144066	N/A
<i>Lasiodiplodia mediterranea</i>	CBS 137783	KJ638312	KJ638331	N/A
<i>Lasiodiplodia mediterranea</i>	ALG36	KJ638314	KJ638333	N/A
<i>Lasiodiplodia missouriana</i>	CBS 128311	HQ288225	HQ288267	N/A
<i>Lasiodiplodia missouriana</i>	CBS 128312	HQ288226	HQ288268	N/A
<i>Lasiodiplodia parva</i>	CBS 456.78	EF622083	EF622063	KF766362
<i>Lasiodiplodia parva</i>	CBS 494.78	EF622084	EF622064	N/A
<i>Lasiodiplodia plurivora</i>	CBS 120832	EF445362	EF445395	N/A
<i>Lasiodiplodia plurivora</i>	CBS 121103	AY343482	EF445396	N/A
<i>Lasiodiplodia pontae</i>	CMM1277	KT151794	KT151791	N/A
<i>Lasiodiplodia pseudotheobromae</i>	CBS 116459	EF622077	EF622057	N/A
<i>Lasiodiplodia pseudotheobromae</i>	CBS 447.62	EF622081	EF622060	N/A
<i>Lasiodiplodia pyriformis</i>	CMW 25414	EU101307	EU101352	N/A
<i>Lasiodiplodia pyriformis</i>	CMW 25415	EU101308	EU101353	N/A
<i>Lasiodiplodia rubropurpurea</i>	CBS 118740	DQ103553	EU673304	N/A
<i>Lasiodiplodia rubropurpurea</i>	WAC 12536	DQ103554	DQ103572	N/A
<i>Lasiodiplodia subglobosa</i>	CMM3872	KF234558	KF226721	N/A
<i>Lasiodiplodia subglobosa</i>	CMM 4046	KF234560	KF226723	N/A
<i>Lasiodiplodia thailandica</i>	CPC 22795	KJ193637	KJ193681	N/A
<i>Lasiodiplodia theobromae</i>	CBS 164.96	AY640255	AY640258	EU673253
<i>Lasiodiplodia theobromae</i>	CBS124.13	DQ458890	DQ458875	N/A
<i>Lasiodiplodia theobromae</i>	CBS 111530	EF622074	EF622054	N/A
<i>Lasiodiplodia theobromae</i>	CAA 006	DQ458891	DQ458876	N/A
<i>Lasiodiplodia venezuelensis</i>	CBS 118739	DQ103547	EU673305	N/A
<i>Lasiodiplodia venezuelensis</i>	WAC 12540	DQ103548	DQ103569	N/A
<i>Lasiodiplodia viticola</i>	CBS 128313	HQ288227	HQ288269	N/A
<i>Lasiodiplodia viticola</i>	CBS 128315	HQ288228	HQ288270	N/A
<i>Macrophomina phaseolina</i>	CBS 227.33	KF951627	KF952000	DQ377906
<i>Macrophomina phaseolina</i>	CBS 162.25	KF531826	KF951996	DQ377905
<i>Marasasiomyces karoo</i>	CBS 118718	KF531828	KF531807	DQ377939
<i>Mucoharknessia cortaderiae</i>	CPC 19974	KM108374	N/A	KM108401
<i>Mucoharknessia cortaderiae</i>	CPC 22208	KM108375	N/A	KM108402
<i>Mucoharknessia anthoxanthii</i>	MFLUCC 15-0904	KU246377	N/A	KU246379
<i>Neodeightonia licuriensis</i>	COAD1780	KP165429	KP165430	N/A
<i>Neodeightonia palmicola</i>	MFLUCC 10-0822	HQ199221	N/A	HQ199222

Species	Cultures	ITS	<i>tef1-a</i>	LSU
<i>Neodeightonia palmicola</i>	MFLUCC 10-0823	HQ199224	N/A	HQ199225
<i>Neodeightonia phoenicum</i>	CBS 122528	EU673340	EU673309	EU673261
<i>Neodeightonia phoenicum</i>	CBS 169.34	EU673338	EU673307	EU673259
<i>Neodeightonia rattanica</i>	MFLUCC 15-0712	KX646357	KX646360	KX646352
<i>Neodeightonia rattanica</i>	MFLUCC 15-0313	KX646358	KX646361	KX646353
<i>Neodeightonia rattanicola</i>	MFLUCC 15-0319	KX646359	KX646362	KX646354
<i>Neodeightonia subglobosa</i>	CBS 448.91	EU673337	EU673306	DQ377866
<i>Neodeightonia subglobosa</i>	MFLUCC 11-0163	JX646794	JX646859	JX646811
<i>Neofusicoccum algeriense</i>	CBS 137504	KJ657702	KJ657715	N/A
<i>Neofusicoccum algeriense</i>	ALG9	KJ657704	KJ657721	N/A
<i>Neofusicoccum andinum</i>	CBS 117453	AY693976	AY693977	N/A
<i>Neofusicoccum andinum</i>	CBS 117452	DQ306263	DQ306264	N/A
<i>Neofusicoccum arbuti</i>	CBS 116131	AY819720	N/A	N/A
<i>Neofusicoccum arbuti</i>	CBS 117090	AY819724	KF531791	N/A
<i>Neofusicoccum australe</i>	CMW 6837	AY339262	AY339270	KF766367
<i>Neofusicoccum australe</i>	CMW 6853	AY339263	AY339271	N/A
<i>Neofusicoccum batangarum</i>	CBS 124924	FJ900607	FJ900653	N/A
<i>Neofusicoccum batangarum</i>	CBS 124923	FJ900608	FJ900654	N/A
<i>Neofusicoccum braziliense</i>	CMM1338	JX513630	JX513610	N/A
<i>Neofusicoccum cordaticola</i>	CBS 123634	EU821898	EU821868	N/A
<i>Neofusicoccum cordaticola</i>	CBS 123635	EU821903	EU821873	N/A
<i>Neofusicoccum cryptoaustrale</i>	CMW23785	FJ752742	FJ752713	N/A
<i>Neofusicoccum eucalypticola</i>	CBS 115679	AY615141	AY615133	N/A
<i>Neofusicoccum eucalypticola</i>	CBS 115766	AY615143	AY615135	N/A
<i>Neofusicoccum eucalyptorum</i>	CBS 115791	AF283686	AY236891	N/A
<i>Neofusicoccum eucalyptorum</i>	CMW 10126	AF283687	AY236892	N/A
<i>Neofusicoccum grevilleae</i>	CBS 129518	JF951137	N/A	N/A
<i>Neofusicoccum hellenicum</i>	CERC1947	KP217053	KP217061	N/A
<i>Neofusicoccum kwambonambiense</i>	CBS 123639	EU821900	EU821870	N/A
<i>Neofusicoccum kwambonambiense</i>	CBS 123641	EU821919	EU821889	N/A
<i>Neofusicoccum lumnitzerae</i>	CMW41469	KP860881	KP860724	N/A
<i>Neofusicoccum lumnitzerae</i>	CMW41228	KP860882	KP860725	N/A
<i>Neofusicoccum luteum</i>	CBS 110299	AY259091	AY573217	AY928043
<i>Neofusicoccum luteum</i>	CBS 110497	EU673311	EU673277	N/A
<i>Neofusicoccum macroclavatum</i>	CBS 118223	DQ093196	DQ093217	N/A
<i>Neofusicoccum macroclavatum</i>	WAC 12446	DQ093197	DQ093218	N/A
<i>Neofusicoccum mangiferae</i>	CBS 118531	AY615185	DQ093221	N/A
<i>Neofusicoccum mangiferae</i>	CBS 118532	AY615186	DQ093220	N/A
<i>Neofusicoccum mangroviorum</i>	CMW41365	KP860859	KP860702	N/A
<i>Neofusicoccum mangroviorum</i>	CMW42481	KP860848	KP860692	N/A
<i>Neofusicoccum mediterraneum</i>	CBS 121718	GU251176	GU251308	N/A
<i>Neofusicoccum mediterraneum</i>	CBS 121558	GU799463	GU799462	N/A
<i>Neofusicoccum nonquaesitum</i>	CBS 126655	GU251163	GU251295	N/A
<i>Neofusicoccum nonquaesitum</i>	PD 301	GU251164	GU251296	N/A
<i>Neofusicoccum oculatum</i>	CBS 128008	EU301030	EU339509	N/A
<i>Neofusicoccum oculatum</i>	MUCC 286	EU736947	EU339511	N/A
<i>Neofusicoccum parvum</i>	CMW 9081	AY236943	AY236888	AY928045
<i>Neofusicoccum parvum</i>	CBS 110301	AY259098	AY573221	N/A
<i>Neofusicoccum pennatisporum</i>	MUCC 510	EF591925	EF591976	N/A
<i>Neofusicoccum protearum</i>	STE-U 4361	AF196295	N/A	N/A
<i>Neofusicoccum protearum</i>	STE-U 1775	AF452539	N/A	N/A
<i>Neofusicoccum ribis</i>	CBS 115475	AY236935	AY236877	N/A
<i>Neofusicoccum ribis</i>	CBS 121.26	AF241177	AY236879	N/A

Species	Cultures	ITS	<i>tef1-a</i>	LSU
<i>Neofusicoccum umdonicola</i>	CBS 123645	EU821904	EU821874	N/A
<i>Neofusicoccum umdonicola</i>	CBS 123646	EU821905	EU821875	N/A
<i>Neofusicoccum ursorum</i>	CMW 24480	FJ752746	FJ752709	N/A
<i>Neofusicoccum viticlavatum</i>	CBS 112878	AY343381	AY343342	N/A
<i>Neofusicoccum viticlavatum</i>	CBS 112977	AY343380	AY343341	N/A
<i>Neofusicoccum vitifusiforme</i>	CBS 110887	AY343383	AY343343	N/A
<i>Neofusicoccum vitifusiforme</i>	CBS 110880	AY343382	AY343344	N/A
<i>Neoscytalidium dimidiatum</i>	CBS 145.78	KF531816	KF531795	DQ377922
<i>Neoscytalidium dimidiatum</i>	CBS 499.66	KF531820	KF531798	DQ377925
<i>Neoscytalidium dimidiatum</i>	CBS 251.49	KF531819	KF531797	DQ377923
<i>Neoscytalidium dimidiatum</i>	UTHSCSA DI 14-340	KM357894	N/A	KM357895
<i>Neoscytalidium dimidiatum</i>	IP127881	AY819727	N/A	DQ377925
<i>Neoscytalidium dimidiatum</i>	CBS 135275	KF571862	N/A	N/A
<i>Neoscytalidium novaehollandiae</i>	CBS 122071	EF585540	EF585580	KF766374
<i>Neoscytalidium novaehollandiae</i>	CBS 122610	EF585536	EF585578	N/A
<i>Neoscytalidium orchidacearum</i>	MFLUCC 12-0533	KU179865	N/A	KU179864
<i>Phaeobotryon cupressi</i>	CBS 124700	FJ919672	FJ919661	N/A
<i>Phaeobotryon cupressi</i>	IRAN 1458C	FJ919671	FJ919660	N/A
<i>Phaeobotryon mamane</i>	CBS 122980	EU673332	EU673298	EU673248
<i>Phaeobotryon mamane</i>	CPC 12442	EU673333	EU673299	DQ377899
<i>Phaeobotryon negundinis</i>	MFLUCC 15-0436	KU820970	KU853997	KU820971
<i>Phaeobotryon negundinis</i>	CAA 797	KX061513	KX061507	N/A
<i>Phaeobotryon rhois</i>	CFCC 89662	KM030584	KM030598	KM030591
<i>Phaeobotryon rhois</i>	CFCC 89663	KM030585	KM030599	KM030592
<i>Pseudofusicoccum adansoniae</i>	CBS 122055	EF585523	EF585571	N/A
<i>Pseudofusicoccum adansoniae</i>	WAC 12689	EF585534	EF585567	EF585554
<i>Pseudofusicoccum ardesiacum</i>	CBS 122062	EU144060	EU144075	N/A
<i>Pseudofusicoccum ardesiacum</i>	WAC 13294	GU172405	GU172437	N/A
<i>Pseudofusicoccum artocarp</i>	CPC 22796	KM006452	KM006483	N/A
<i>Pseudofusicoccum kimberleyensis</i>	CBS 122058	EU144057	EU144072	N/A
<i>Pseudofusicoccum kimberleyensis</i>	CBS 122059	EU144056	EU144071	N/A
<i>Pseudofusicoccum olivaceum</i>	CBS 124939	FJ888459	FJ888437	N/A
<i>Pseudofusicoccum olivaceum</i>	CBS 124940	FJ888462	FJ888438	N/A
<i>Pseudofusicoccum stromaticum</i>	CBS 117448	AY693974	AY693975	DQ377931
<i>Pseudofusicoccum stromaticum</i>	CBS 117449	DQ436935	DQ436936	DQ377932
<i>Pseudofusicoccum violaceum</i>	CBS 124936	FJ888474	FJ888442	N/A
<i>Pseudofusicoccum violaceum</i>	CBS 124937	FJ888458	FJ888440	N/A
<i>Sakireeta madreeya</i>	CBS 532.76	KC769960	KM108427	DQ377940
<i>Sardiniella urbana</i>	CBS 141580	KX379674	KX379675	KX379676
<i>Sardiniella urbana</i>	BL180	KX379677	KX379678	KX379679
<i>Sardiniella urbana</i>	BL181	KX379680	KX379681	KX379682
<i>Spencermartinsia alpina</i>	CGMCC 3.18001	KX499645	KX499651	N/A
<i>Spencermartinsia citricola</i>	ICMP16828	EU673323	EU673290	N/A
<i>Spencermartinsia citricola</i>	CMP16827	EU673322	EU673289	N/A
<i>Spencermartinsia mangiferae</i>	IRAN1584C	KC898221	KC898204	N/A
<i>Spencermartinsia mangiferae</i>	IRAN1545C	KC898223	KC898206	N/A
<i>Spencermartinsia plurivora</i>	IRAN1557C	KC898225	KC898208	N/A
<i>Spencermartinsia plurivora</i>	IRAN1537C	KC898226	KC898209	N/A
<i>Spencermartinsia rosulata</i>	CBS 121760	EU101290	EU101335	N/A
<i>Spencermartinsia rosulata</i>	CMW 25392	EU101293	EU101338	N/A
<i>Spencermartinsia viticola</i>	CBS 117009	AY905554	AY905559	DQ377873
<i>Spencermartinsia westrale</i>	DAR80529	HM009376	HM800511	N/A
<i>Spencermartinsia westrale</i>	DAR80530	HM009377	HM800512	N/A
<i>Spencermartinsia yunnana</i>	CGMCC 3.17999	KX499643	KX499649	N/A

Species	Cultures	ITS	<i>tef1-a</i>	LSU
<i>Spencermartinsia yunnana</i>	CGMCC 3.18000	KX499644	KX499650	N/A
<i>Sphaeropsis citrigena</i>	ICMP 16812	EU673328	EU673294	EU673246
<i>Sphaeropsis citrigena</i>	ICMP 16818	EU673329	EU673295	EU673247
<i>Sphaeropsis eucalypticola</i>	CBS 133993	JX646802	JX646867	JX646819
<i>Sphaeropsis eucalypticola</i>	MFLUCC 11-0654	JX646803	JX646868	JX646820
<i>Sphaeropsis porosa</i>	CBS 110496	AY343379	AY343340	DQ377894
<i>Sphaeropsis porosa</i>	CBS 110574	AY343378	AY343339	DQ377895
<i>Sphaeropsis variabilis</i>	CMW 25420	EU101313	EU101358	N/A
<i>Sphaeropsis variabilis</i>	CMW 25421	EU101314	EU101359	N/A
<i>Sphaeropsis visci</i>	CBS 122526	EU673324	EU673292	N/A
<i>Sphaeropsis visci</i>	CBS 186.97	EU673325	EU673293	DQ377868
<i>Tiarosporella paludosa</i>	CPC 22701	KM108378	N/A	KM108404
<i>Tiarosporella paludosa</i>	CPC 22702	KM108379	N/A	KM108405
Aplosporellaceae				
<i>Aplosporella javeedii</i>	CMW 38166	KC769939	KC769847	KC769980
<i>Aplosporella prunicola</i>	CBS 121167	KF766147	N/A	JX681071
<i>Aplosporella yalgorensis</i>	MUCC 511	EF591927	EF591978	EF591944
<i>Bagnisiella examinans</i>	CBS 551.66	KF766148	GU349056	KF766316
Melanopsaceae				
<i>Melanops sp.</i>	CBS 118.39	FJ824771	FJ824776	DQ377856
<i>Melanops tulasnei</i>	CBS 116805	FJ824769	FJ824774	FJ824764
Phyllostictaceae				
<i>Phyllosticta citricarpa</i>	CBS 102374	FJ824767	FJ538376	DQ377877
<i>Phyllosticta minima</i>	CBS 111635	KF766215	KF766433	EU754194
<i>Phyllosticta parthenocissi</i>	CBS 111645	FJ824766	EU683653	DQ377876
<i>Phyllosticta philoprina</i>	CBS 616.72	KF766171	KF766402	KF766341
<i>Phyllosticta podocarp</i>	CBS 111647	KF766217	KF766434	KF766383
Planistromellaceae				
<i>Kellermania crassispora</i>	CBS 131714	KF766175	KF766406	KF766345
<i>Kellermania dasylirionis</i>	CBS 131715	KF766177	KF766408	KF766347
<i>Kellermania macrospora</i>	CBS 131716	KF766178	KF766409	KF766348
<i>Kellermania nolinae</i>	CBS 131717	KF766180	KF766411	KF766350
<i>Kellermania plurilocularis</i>	CBS 131719	KF766181	KF766412	KF766351
<i>Kellermania yuccifoliorum</i>	CBS 131726	KF766185	KF766416	KF766355
Saccharataceae				
<i>Saccharata capensis</i>	CBS 122693	KF766224	EU552095	KF766390
<i>Saccharata kirstenboschensis</i>	CBS 123537	KF766225	N/A	FJ372409
<i>Saccharata proteae</i>	CBS 115206	KF766226	KF766438	DQ377882
Septorioidaceae				
<i>Septoriooides pini-thunbergii</i>	CBS 473.91	KF251243	N/A	KF251746
<i>Septoriooides strobi</i>	CBS141443	KT884699	KT884713	KT884685
<i>Septoriooides strobi</i>	CBS141444	KT884700	KT884714	KT884686
<i>Septoriooides strobi</i>	CBS141445	KT884702	KT884716	KT884688
<i>Septoriooides strobi</i>	V-3-11-1	KT884703	KT884717	KT884689
Outgroup				
<i>Lecanosticta acicola</i>	LNPV 252	JX901755	JX901639	JX901844

of a random starting tree and 1000 ML bootstrap replicates. The search strategy was set to rapid bootstrapping with one thousand non-parametric bootstrapping iterations using the general time reversible model (GTR) with a discrete gamma distribution. The best scoring trees were selected with a final likelihood values.

Posterior probabilities (PP) were determined by Markov Chain Monte Carlo sampling (BMCMC) in MrBayes v. 3.0b4 (Ronquist & Huelsenbeck 2003). MrModeltest v. 2.3 (Nylander 2004) was used to carry out statistical selection of best-fit model of nucleotide substitution and was incorporated into the analysis. Six simultaneous Markov chains were run for 1 000 000 generations and trees were sampled every 100th generation. The 2000 trees representing the burn-in phase of the analyses, were discarded and the remaining 8000 trees used for calculation of posterior probabilities (PP) in the majority rule consensus tree. The strains used in this study are listed in Table 1 with details of type cultures and sequence data.

Results and Discussion

A complete list of all genera and species in the *Botryosphaeriaceae* that are known from culture is provided in Table 1. Where possible the ex-type isolate together with one further isolate are included. Also included in Table 1 are representative isolates of other families in the Botryosphaeriales.

The family Botryosphaeriaceae

Presently 23 genera are included the family (Fig. 1). Some doubts remain about the limits of the family, especially regarding the position of *Pseudofusicoccum*. Slippers et al (2013) considered that *Pseudofusicoccum* lies in a separate clade at the base of the family while Phillips et al. (2013) placed it as sister to *Endomelanconiopsis* within *Botryosphaeriaceae*. Although in Fig. 1 we show it to be sister to *Phyllosticta*, and thus within *Phyllostictaceae*, this tree is based on only 3 loci and thus is not as robust as the trees of Slippers et al (2013) or Phillips et al. (2013). Therefore at this stage we consider that *Pseudofusicoccum* is a genus within *Botryosphaeriaceae* even though its position remains uncertain. Apart from *Pseudofusicoccum* the positions of the remaining 22 genera are similar to previously published phylogenies of *Botryosphaeriaceae*.

Genera and species

1. *Alanphillipsia* Crous & M.J. Wingf., *Persoonia* 31: 197 (2013), MycoBank MB805816.

Alanphillipsia was introduced to accommodate species that are morphologically aplosporella-like but the conidia have a hyaline outer layer (Crous et al. 2013, 2014). Five species are currently known and all have been reported from the Western Cape Province of South Africa. The sexual morph has not been reported for any species.

Type species – *Alanphillipsia aloes* Crous & M.J. Wingf. 2013.

1.1. *Alanphillipsia aloicola* Crous, *Persoonia* 33: 217 (2014), MycoBank MB810590.

Sexual morph not reported. See Crous et al. (2014) for illustrations and descriptions of asexual morph.

Type – South Africa, Western Cape Province, Clanwilliam, Ramskop, *Aloe* sp. (*Asphodelaceae*), Sept. 2013, M.J. Wingfield (holotype CBS H-21978, culture ex-type CBS 138896).

Host – *Aloe* sp. (Crous et al. 2014).

Distribution – South Africa (Crous et al. 2014).

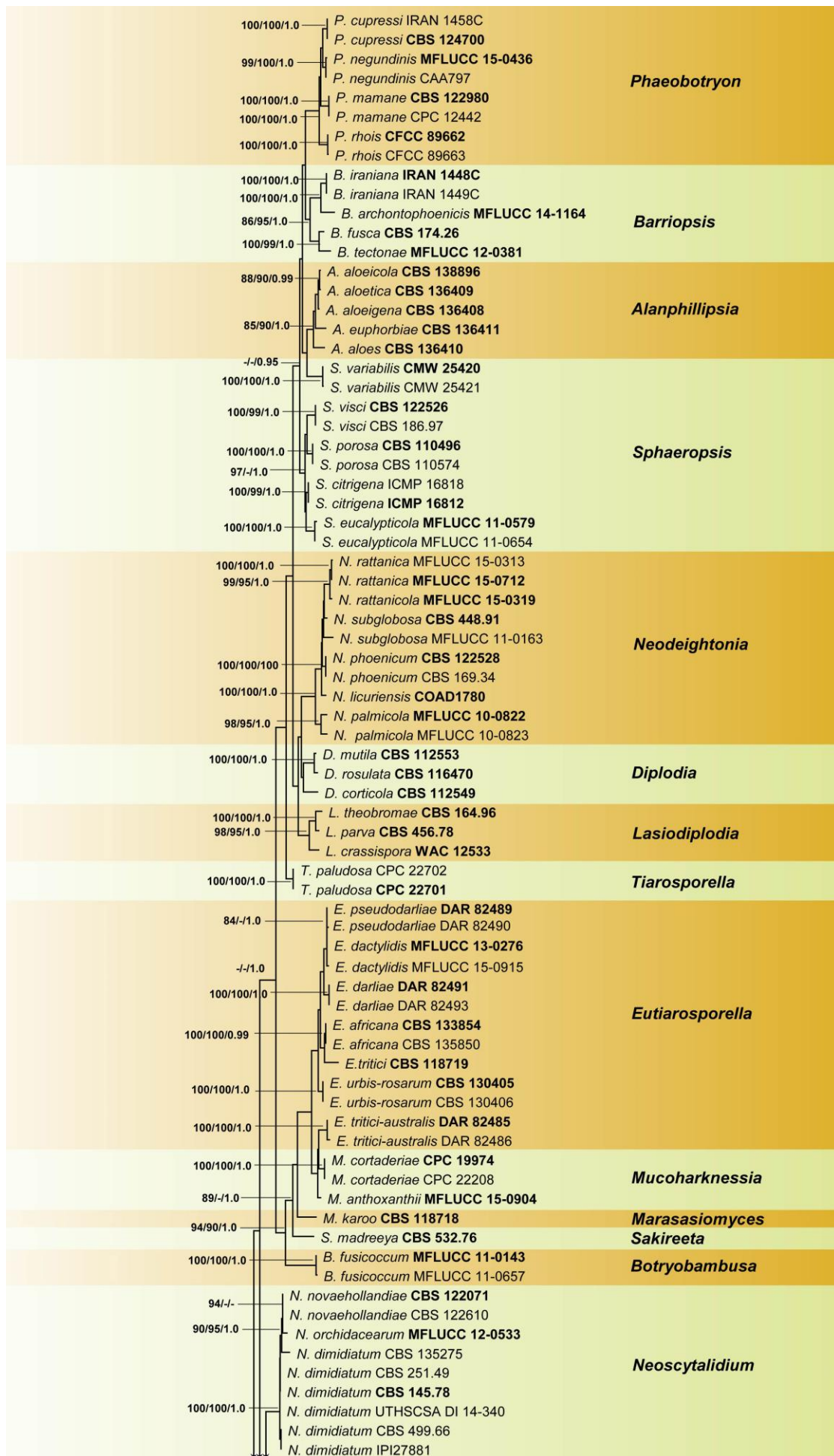
1.2. *Alanphillipsia aloeigena* Crous & M.J. Wingf., *Persoonia* 31: 199 (2013), MycoBank MB805818.

Sexual morph not reported. See Crous et al. (2013) for illustrations and descriptions of asexual morph.

Type – South Africa, Namakwaland, Goegap Nature Reserve, on leaves of *Aloe melanacantha* (*Asphodelaceae*), 26 Sept. 2012, M.J. Wingfield (holotype CBS H-21419, culture ex-type CPC 21286 = CBS 136408).

Host – *Aloe melanacantha* (Crous et al. 2013).

Distribution – South Africa (Crous et al. 2013).



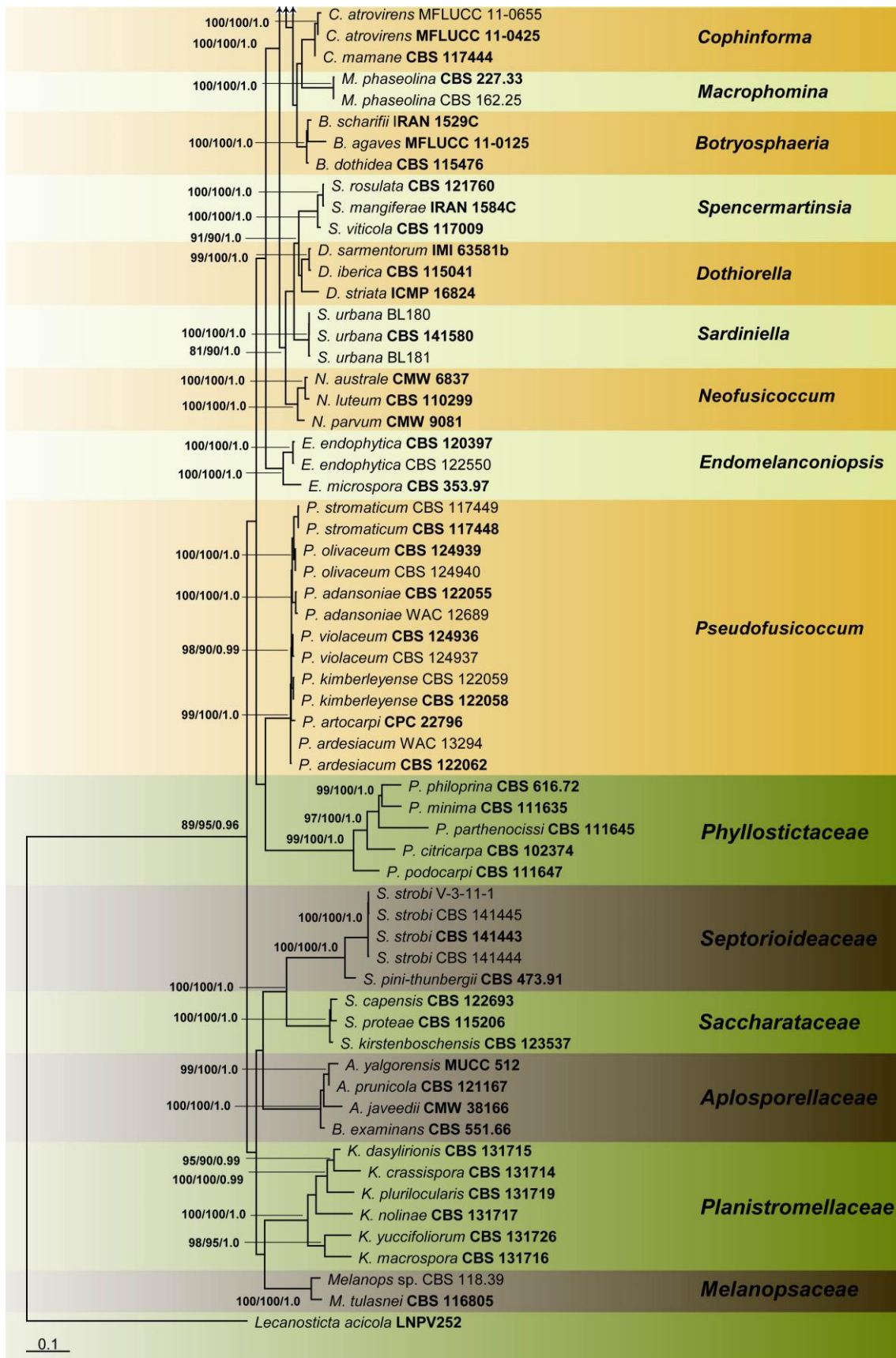


Fig. 1 – Phylogram generated from Maximum Likelihood analysis based on combined ITS, *tef1-a* and LSU dataset of Botryosphaerales. Bootstrap support values for maximum likelihood (ML), maximum parsimony (MP) greater than 75% and Bayesian posterior probabilities above 0.90 are indicated near the nodes. The ex-type strains are in bold and the tree is rooted with *Lecanosticta acicola* (LNPV252).

1.3. *Alanphillipsia aloes* Crous & M.J. Wingf., *Persoonia* 31: 197 (2013), MycoBank MB805817. Sexual morph not reported. See Crous et al. (2013) for illustrations and descriptions of asexual morph.

Type – South Africa, Western Cape Province, Clanwilliam, on dark lesions of dying *Aloe dichotoma* (*Asphodelaceae*), Sept. 2012, M.J. Wingfield (holotype CBS H-21418, cultures ex-type CPC 21298 = CBS 136410).

Host – *Aloe dichotoma* (Crous et al. 2013).

Distribution – South Africa (Crous et al. 2013).

1.4. *Alanphillipsia aloetica* Crous, *Persoonia* 31: 201 (2013), MycoBank MB805819.

Sexual morph not reported. See Crous et al. (2013) for illustrations and descriptions of asexual morph.

Type – South Africa, Eastern Cape, Uitenhage, on *Aloe* sp. (*Asphodelaceae*), 14 July 2012, P.W. Crous (holotype CBS H-21420, culture ex-type CPC 21110, 21109 = CBS 136409).

Host – *Aloe* sp. (Crous et al. 2013).

Distribution – South Africa (Crous et al. 2013).

1.5. *Alanphillipsia euphorbiae* Crous & M.J. Wingf., *Persoonia* 31: 203 (2013), MycoBank MB805821.

Sexual morph not reported. See Crous et al. (2013) for illustrations and descriptions of asexual morph.

Type – South Africa, Western Cape Province, Kirstenbosch Botanical Garden, on leaves of *Euphorbia* sp. (*Euphorbiaceae*), Sept. 2012, M.J. Wingfield (holotype CBS H-21421, culture ex-type CPC 21629, 21628 = CBS 136411).

Host – *Aloe* sp. (Crous et al. 2013).

Distribution – South Africa (Crous et al. 2013).

2. *Barriopsis* A.J.L. Phillips, A. Alves & Crous, *Persoonia* 21: 39 (2008), MycoBank MB511712.

Barriopsis comprises four species. Sexual and asexual morphs have been reported within the genus. The pale brown, 1-septate conidia that are striate when immature are unique in the *Botryosphaeriaceae*.

Type species – *Barriopsis fusca* (N.E. Stevens) A.J.L. Phillips, A. Alves & Crous 2008.

2.1. *Barriopsis archontophoenicis* Konta, Boonmee & K.D. Hyde, 7: 924 (2016), Facesoffunginumber FOF02097.

Sexual morph and asexual morph illustrations and descriptions in Konta et al. (2016a, this volume). This species forms the sexual morph in culture after long periods of incubation (up to 6 months).

Type – Thailand, Chiang Mai Province, Mushroom Research Centre, on *Archontophoenix alexandrae* (*Arecaceae*), 15 August 2014, S. Konta (holotype MFLU 15–0015, culture ex-type MFLUCC 14–1164, MUCL 55901).

Host – *Archontophoenix alexandrae* (Konta et al. 2016a).

Distribution – Thailand (Konta et al. 2016a).

2.2. *Barriopsis fusca* (N.E. Stevens) A.J.L. Phillips, A. Alves & Crous, *Persoonia* 21: 39 (2008), MycoBank MB511713.

Sexual morph illustrations and descriptions are available in Liu et al. (2012) and Phillips et al. (2008, 2013). The only available culture of *B. fusca* (CBS 174.26, ex-type) has lost its ability to

sporulate. According to Stevens (1926) the asexual morph is lasiodiplodia-like with hyaline conidia that become dark-brown and septate with irregular longitudinal striations.

Type – Cuba, Herradura, on twigs of *Citrus* sp. (*Rutaceae*), 15 Jan. 1925, N.E. Stevens (holotype BPI 599052, culture ex-type CBS 174.26).

Host – *Citrus* sp. (Stevens 1926).

Distribution – Cuba (Stevens 1926), Florida (Shear 1923).

2.3. ***Barriopsis iraniana*** Abdollahz., Zare & A.J.L. Phillips, *Persoonia* 23: 4 (2009), MycoBank MB513235.

Sexual morph not reported. See Abdollahzadeh et al. (2009), Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Iran, Hormozgan Province, Minab, Hajikhademi, on twigs of *Mangifera indica* (*Anacardiaceae*), 27 Feb. 2007, J. Abdollahzadeh & A. Javadi (holotype IRAN 13939F, culture ex-type IRAN 1448C = CBS 124698).

Hosts – *Citrus* sp., *Mangifera indica*, *Olea* sp. (Abdollahzadeh et al. 2009).

Distribution – Iran (Abdollahzadeh et al. 2009).

2.4. ***Barriopsis tectonae*** Doilom, L.A. Shuttlew. & K.D. Hyde, *Phytotaxa* 176: 84 (2014), MycoBank MB808202.

Sexual morph and asexual morph have been reported. See Doilom et al. (2014) for illustrations and descriptions.

Type – Thailand, Phayao Province: Muang District, on dead branch of *Tectona grandis* (*Lamiaceae*), 12 March 2012, M. Doilom (holotype MFLU 14-0024, culture ex-type MFLUCC 12-0381 = CBS 137786).

Host – *Tectona grandis* (Doilom et al. 2014).

Distribution – Thailand (Doilom et al. (2014).

3. ***Botryobambusa*** Phook., J.K. Liu & K.D. Hyde, *Fungal Diversity* 57: 166 (2012), MycoBank MB801313.

Botryobambusa comprises a single species, which is known in both the sexual and asexual morph.

Type species – *Botryobambusa fusicoccum* Phook., J.K. Liu & K.D. Hyde 2012.

3.1. ***Botryobambusa fusicoccum*** Phook., J.K. Liu & K.D. Hyde, *Fungal Diversity* 57: 166 (2012), MycoBank MB801314.

Sexual morph and asexual morph have been reported. See Liu et al. (2012), Phillips et al. (2013) for illustrations and descriptions.

Type – Thailand, Lampang Province, Jae Hom District, Mae Yuag Forestry Plantation, on dead culms of *Bambusa* sp. (*Poaceae*), 19 Aug. 2010, R. Phookamsak (holotype MFLU 11-0179, culture ex-type CBS 134113 = MFLUCC 11-0143).

Host – *Bambusa* sp. (Liu et al. 2012).

Distribution – Thailand (Liu et al. 2012).

4. ***Botryosphaeria*** Ces. & De Not., *Comm. Soc. crittog. Ital.* 1: 211 (1863), MycoBank MB635.

Type genus of *Botryosphaeriaceae*. *Botryosphaeria* comprises ten species (Fig. 2). Sexual and asexual morphs have been reported in the genus.

Type species – *Botryosphaeria dothidea* (Moug. : Fr.) Ces. & De Not. 1863.

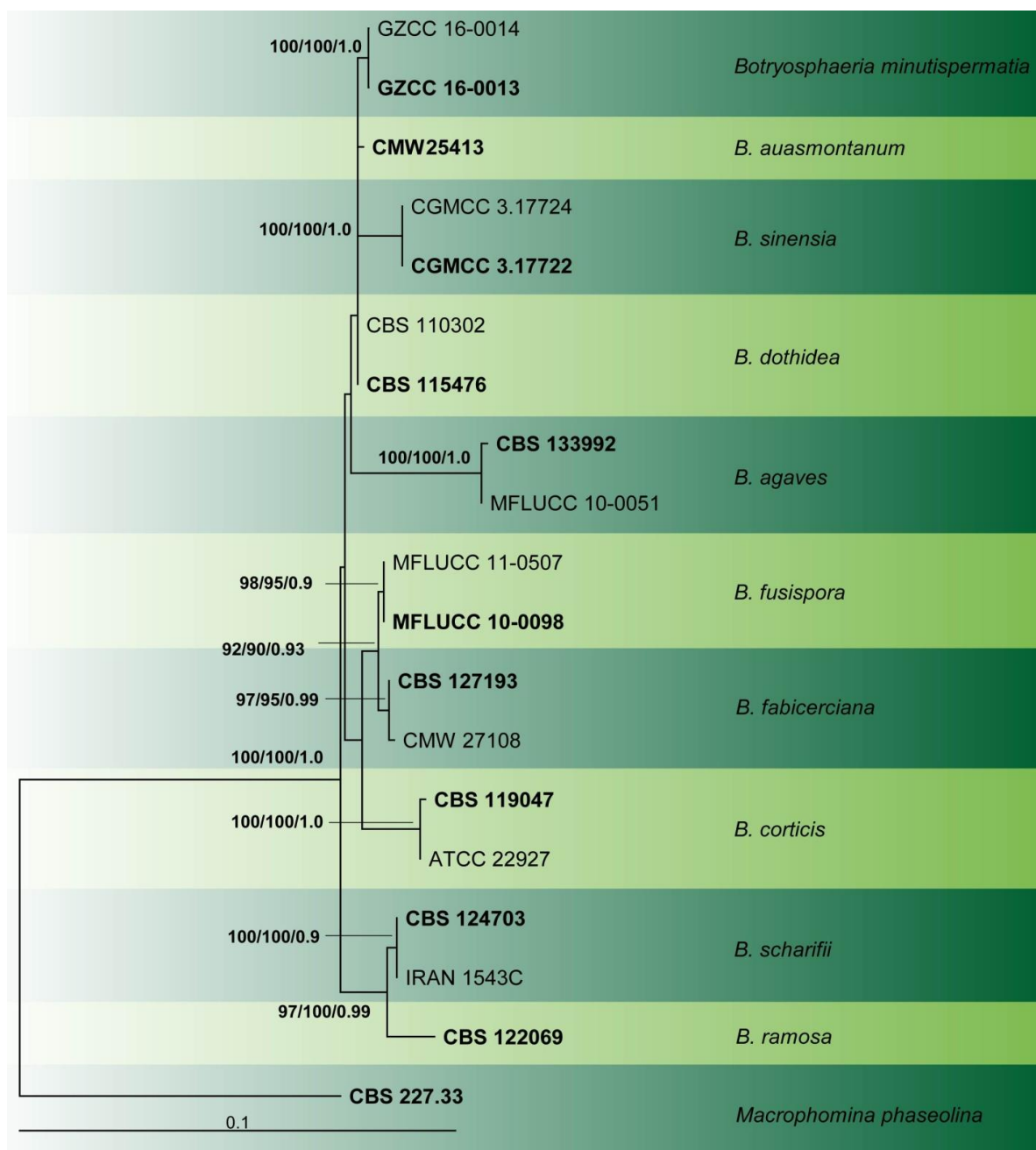


Fig. 2 – Phylogram generated from Maximum Likelihood analysis based on combined ITS and *tef1- α* dataset of *Botryosphaeria*. Bootstrap support values for maximum likelihood (ML), maximum parsimony (MP) greater than 75 % and Bayesian posterior probabilities greater than 0.90 are indicated near the nodes. The ex-type strains are in bold and the tree is rooted with *Macrophomina phaseolina* (CBS 227.33).

4.1. *Botryosphaeria agaves* (Henn.) E.J. Butler, *Annls mycol.* 9: 415 (1911), MycoBank MB119799.

See Liu et al. (2012) and Phillips et al. (2013) for illustrations and descriptions of sexual morph. Asexual morph not reported.

Type – Tanzania, Zanzibar, on leaves of *Agave sisalana*, Zimmerman, holotype presumably lost (not in B). Thailand, Chiang Rai Province, Doi Tung, on living and dead leaves of *Agave* sp. (*Asparagaceae*), 16 Jun. 2010, R. Phookamsak (neotype designated in Phillips et al. (2013) MFLU 11-0161, culture ex-neotype MFLUCC 11-0125 = CBS 133992).

Host – *Agaves* sp. (Liu et al. 2012).

Distribution – Thailand (Liu et al. 2012).

4.2. *Botryosphaeria auasmontanum* F.J.J. Van der Walt, Slippers & G.J. Marais, *Persoonia* 33: 162 (2014), MycoBank MB518721.

Sexual morph not reported. See Slippers et al. (2014) for illustrations and descriptions of asexual morph.

Type – Namibia, Windhoek, *Acacia mellifera* (*Fabaceae*), Feb. 2006, F.J.J. van der Walt & J. Roux (holotype PREM 59632, culture ex-type CBS 121769).

Host – *Acacia mellifera* (Slippers et al. 2014).

Distribution – Namibia (Slippers et al. 2014).

4.3. *Botryosphaeria corticis* (Demaree & Wilcox) Arx & E. Müll., *Beitr. Kryptfl. Schweiz* 11: 42 (1954), MycoBank MB293807.

Sexual morph and asexual morph have been reported. See Phillips et al. (2013) for illustrations and descriptions.

Type – USA, North Carolina, Atkinson, *Vaccinium corymbosum* (*Ericaceae*), 14 Feb. 1940, J.B. Demaree (holotype BPI 598729), New Jersey, Hammonton, on cankered stems of *V. corymbosum*, May 2005, P.V. Oudemans (epitype CBS H-19706, cultures ex-epitype CBS 119047, CBS 119048).

Hosts – *Vaccinium* species including *V. ashei*, *V. corymbosum*, *V. tenellum* and *V. virgatum* (Phillips et al. 2006, Wright & Harmon 2010).

Distribution – USA (Florida, Georgia, Maryland, Mississippi, New Jersey, North Carolina) (Phillips et al. 2006, Wright & Harmon 2010).

4.4. *Botryosphaeria dothidea* (Moug. : Fr.) Ces. & De Not., *Comm. Soc. crittog. Ital.* 1: 212 (1863), MycoBank MB183247.

Sexual morph and asexual morph reported. See Phillips et al. (2013) for illustrations and descriptions.

Type – France, *Rosa* sp., 1823, Fries ex Mougeot. Herbarium S (neotype of *Sphaeria dothidea* designated by Slippers et al. 2004a). Switzerland, Ticino, Crocifisso, *Prunus* sp., Oct. 2000, B. Slippers (epitype PREM 57372 designated by Slippers et al. (2004a), culture ex-epitype CBS 115476). Italy, on branches of *Aesculus* sp., P.A. Saccardo, PAD, (neotype of asexual morph designated by Crous & Palm 1999).

Hosts – According to USDA fungal database, this species has been reported in 171 host genera belonging to 75 families. However, the validity of many of these records is questionable since they were made prior to the re-evaluation of *B. dothidea* by Slippers et al. (2004a) who established a modern and reliable concept for this species. Studies subsequent to Phillips et al. (2013) confirm the following hosts: *Acer platanoides* in China (Wang et al. 2015), *Artemisia* sp. in China (Cosoveanu et al. 2016), *Camellia sinensis* in China (Jayawardena et al. 2016), *Cupressus sempervirens* in Iran (Mohammadi et al. 2014), *Juglans regia* in California, China (Chen et al. 2014a, Li et al. 2016b), Kiwifruit in China (Zhou et al. 2015), *Malus domestica* in Turkey, Serbia (Vasic et al. 2013, Turkolmez et al. 2016), *Mangifera indica* in Brazil (Marques et al. 2013b, Rabari et al. 2016), ornamental trees in Western Balkans (Zlatkovic et al. 2016), *Phoenix dactylifera* in Iran (Mohammadi 2014), *Pistacia vera* in California, Iran (Chen et al. 2014b, Mohammadi et al. 2014), *Prunus serrulata* in China (Yan et al. 2016), *Psidium guajava* in Brazil (Nogueira Junior et al. 2016), *Pyrus* sp. in China (Zhai et al. 2014), Tobacco in China (Bian et al. 2015), *Vaccinium corymbosum* in China (Xu et al. 2015b), *Vitis vinifera* in Algeria, China, Italy, Tunisia, Turkey (Yan et al. 2013, Ammad et al. 2014, Chebil et al. 2014, Akgul et al. 2015, Carlucci et al. 2015).

Distribution – Worldwide.

4.5. *Botryosphaeria fabicerciana* (S.F. Chen, Pavlic, M.J. Wingf. & X.D. Zhou) A.J.L. Phillips & A. Alves, *Stud. Mycol.* 76: 77 (2013), MycoBank MB805457.

Sexual morph not reported. See Chen et al. (2011), Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – China, Fujian Province, from senescing twigs of an unknown *Eucalyptus* sp. (*Myrtaceae*), Aug. 2007, M.J. Wingfield (holotype PREM 60449, culture ex-type CBS 127193).

Hosts – *Eucalyptus* sp. (Chen et al. 2011), *Mangifera indica* (Nogueira Junior et al. 2016, Marques et al. 2013b).

Distribution – China (Chen et al. 2011), Brazil (Marques et al. 2013b, Nogueira Junior et al. 2016).

4.6. *Botryosphaeria fusispora* Boonmee, J.K. Liu & K.D. Hyde, *Fungal Diversity* 57: 171 (2012), MycoBank MB 801319.

Sexual morph and asexual morph reported. See Liu et al. (2012) for illustrations and descriptions.

Type – Thailand, Chiang Rai, Doi Tung, on dried bark of *Entada* sp. (*Fabaceae*), 10 Jun. 2009, S. Boonmee (holotype MFLU 10-0028, culture ex-type MFLUCC 10-0098).

Hosts – *Caryota* sp., *Entada* sp. (Liu et al. 2012).

Distribution – Thailand (Liu et al. 2012).

4.7. *Botryosphaeria minutispermata* Ariyawansa, K.D. Hyde & Z.Y. Liu, *Phytotaxa* 275: 40 (2016), Facesoffunginumber FoF02393.

Sexual morph and asexual morph reported. See Ariyawansa et al. (2016) for illustrations and descriptions.

Type – China, Guizhou Province, Guiyang City, Huaxi District, Guizhou Academy of Agricultural Sciences garden, on dead wood, 14 December 2015, HA Ariyawansa (holotype GZAAS 16-0009, culture ex-type GZCC 16-0013).

Host – Unknown host (Ariyawansa et al. 2016).

Distribution – China (Ariyawansa et al. 2016).

4.8. *Botryosphaeria ramosa* (Pavlic, T.I. Burgess & M.J. Wingf.) A.J.L. Phillips & A. Alves, *Stud. Mycol.* 76: 77 (2013), MycoBank MB805458.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions.

Type – Australia, Western Australia, Bell Gorge, on *Eucalyptus camaldulensis* (*Myrtaceae*), Jul. 2006, T.I. Burgess (holotype PREM 59846, culture ex-type CBS 122069).

Host – *Eucalyptus camaldulensis* (Pavlic et al. 2008).

Distribution – Australia (Pavlic et al. 2008).

4.9. *Botryosphaeria sharifii* Abdoll., Zare & A.J.L. Phillips, *Mycologia* 105: 213 (2013), MycoBank MB564800

Sexual morph not reported. See Abdollahzadeh et al. (2013) for illustrations and descriptions of asexual morph.

Type – Iran, Tehran, on fruits of *Mangifera indica* (*Anacardiaceae*) imported from Pakistan, Aug. 2006, J. Abdollahzadeh (holotype IRAN 14275F, culture ex-type CBS 124703).

Host – *Mangifera indica* (Abdollahzadeh et al. 2013).

Distribution – Iran, Pakistan (Abdollahzadeh et al. 2013).

4.10. *Botryosphaeria sinensia* Y.P. Zhou, Y. Zhang ter. *Phytotaxa* 245: 043 (2016), MycoBank MB813802.

Sexual morph and asexual morph have been reported. See Zhou et al. (2016) for illustrations and descriptions.

Type – China, Henan Province, Puyang, Qingfeng county, on twigs of *Populus* sp. (*Salicaceae*), 11 Nov. 2014 (holotype HMAS 246714!, culture ex-type CGMCC 3.17722).

Hosts – *Juglans regia*, *Morus* sp., *Populus* sp. (Zhou et al. 2016).

Distribution – China (Zhou et al. 2016).

5. *Cophinforma* Doilom, J.K. Liu & K.D. Hyde, *Fungal Diversity* 57: 174 (2012), MycoBank MB801315.

This genus comprises two species. Sexual morph and asexual morph have been reported within the genus.

Type species – *Cophinforma atrovirens* (Mehl & Slippers) A. Alves & A.J.L. Phillips 2013.

5.1. *Cophinforma atrovirens* (Mehl & Slippers) A. Alves & A.J.L. Phillips, *Stud. Mycol.* 76: 80 (2013), MycoBank MB805459.

Although Phillips et al. (2013) stated that the sexual morph has not been reported, Liu et al. (2012) described and illustrated the sexual morph of *C. eucalypti*, which Phillips et al. (2013) regarded as a synonym of *C. atrovirens*. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – South Africa, Mpumalanga Province, Mawewe Nature Reserve, from an asymptomatic branch of *Pterocarpus angolensis* (*Fabaceae*), Dec. 2005, J.W.M. Mehl & J. Roux (holotype PREM 60341, culture ex-type CBS 124934, paratype PREM 60342, culture ex-papatype CBS 124935).

Hosts – *Eucalyptus* sp. (Liu et al. 2012), *Eucalyptus urophylla* (Xu et al. 2015a), *Pterocarpus angolensis* (Mehl et al. 2011).

Distribution – South Africa (Mehl et al. 2011), Thailand (Liu et al. 2012), Venezuela (Xu et al. 2015a).

5.2. *Cophinforma mamane* (D.E. Gardner) A.J.L. Phillips & A. Alves, *Stud. Mycol.* 76: 80 (2013), MycoBank MB805460.

Sexual morph and asexual morph reported. See Gardner (1997) for illustrations and Phillips et al. 2013 for descriptions of asexual morph.

Type – USA, Hawaii, Hawaii Island, Hawaii Volcanoes National Park, Kipuka Ki, on bark of a swollen branch of *Sophora chrysophylla* (*Fabaceae*), 1 May 1996, D.E. Gardner (holotype BISH 644614, no cultures extant; isotype BISH 737731, no cultures extant; paratypes BPI 737732, BPI 737733, no cultures extant).

Hosts – *Sophora chrysophylla* (Gardner 1997), *Mangifera indica* (Marques et al. 2013b), *Vitis vinifera* (Correia et al. 2013).

Distribution – Brazil (Correia et al. 2013, Marques et al. 2013b), USA (Hawaii) (Gardner 1997).

6. *Diplodia* Fr., in Montagne, *Annls Sci. Nat., Bot., sér. 2*, 1: 302 (1834), MycoBank MB8047.

This genus comprises 26 species (Fig. 3). Sexual and asexual morphs have been reported.

Type species – *Diplodia mutila* (Fr. : Fr.) Fr. 1849.

6.1. *Diplodia africana* Damm & Crous, *Mycologia* 99: 671 (2008), MycoBank MB501323.

Sexual morph not reported. See Damm et al. (2007), Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – South Africa, Western Cape Province, Paarl, from wood section close to pruning wound of *Prunus persica* (*Rosaceae*), 10 Jun. 2004, U. Damm (holotype CBS H-19843, culture ex-type CBS 120835, additional culture STE-U 6289).

Hosts – *Juniperus phoenicea* (Alves et al. 2014), *Prunus persica* (Damm et al. 2007).

Distribution – Italy (Alves et al. 2014), South Africa (Damm et al. 2007).

6.2. *Diplodia agrifolia* S.C. Lynch & Eskalen, *Mycologia* 105: 135 (2012), MycoBank MB800443.

Sexual morph not reported. See Lynch et al. (2013), Phillips et al. 2013 for illustrations and descriptions of asexual morph.

Type – USA, California, San Diego County, Mataguay Scout Camp, on cankered branch of *Quercus agrifolia* (*Fagaceae*), 23 Feb. 2010, S.C. Lynch & A. Eskalen (holotype BPI 884095, culture ex-type CBS 132777 = ATCC MYA-4895 = UCROK 732).

Hosts – *Quercus agrifolia*, *Q. kelloggii* (Lynch et al. 2013).

Distribution – USA (Lynch et al. 2013).

6.3. *Diplodia alatafructa* Mehl & Slippers, *Mycologia* 103: 542 (2011), MycoBank MB513498.

Sexual morph not reported. See Mehl et al. (2011), Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – South Africa, Mpumalanga Province, Sudwala Caves area, from a stem wound on *Pterocarpus Angolensis* (*Fabaceae*), Dec. 2005, J.W.M. Mehl & J. Roux (holotype PREM 60337, culture ex-type CBS 124931).

Host – *Pterocarpus angolensis* (Mehl et al. 2011).

Distribution – South Africa (Mehl et al. 2011).

6.4. *Diplodia allocellula* Jami, Gryzenh., Slippers & M.J. Wingf., *Cryptog. Mycol.* 33: 256 (2012), MycoBank MB564140

Sexual morph not reported. See Jami et al. (2012), Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – South Africa, Gauteng Province, Pretoria, from branch of *Acacia karroo* (*Fabaceae*) Nov. 2009, M. Gryzenhout & F. Jami (holotype PREM 60701, culture ex-type CBS 130408).

Host – *Acacia karroo* (Jami et al. 2012).

Distribution – South Africa (Jami et al. 2012).

6.5. *Diplodia bulgarica* A.J.L. Phillips, J. Lopes & Bobev, *Persoonia* 29: 33 (2012), MycoBank MB19632.

Sexual morph not reported. See Phillips et al. (2012, 2013) for illustrations and descriptions of asexual morph.

Type – Bulgaria, Plovdiv, on dead twigs of *Malus sylvestris* (*Rosaceae*), 2005, S.G. Bobev (holotype CBS H-20189, culture ex-type CBS 124254).

Host – *Malus* sp. (Phillips et al. 2012).

Distribution – Bulgaria, Iran (Phillips et al. 2012).

6.6. *Diplodia corticola* A.J.L. Phillips, A. Alves & J. Luque, *Mycologia* 96: 603 (2004), MycoBank MB488568.

Sexual morph and asexual morph reported. See Phillips et al. 2013 for illustrations and descriptions.

Type – Portugal, Beira Littoral, Requeixo near Aveiro, on dead branches of *Quercus suber* (*Fagaceae*), Feb. 2002, A. Alves (holotype LISE 94839, culture ex-type CBS 112549).

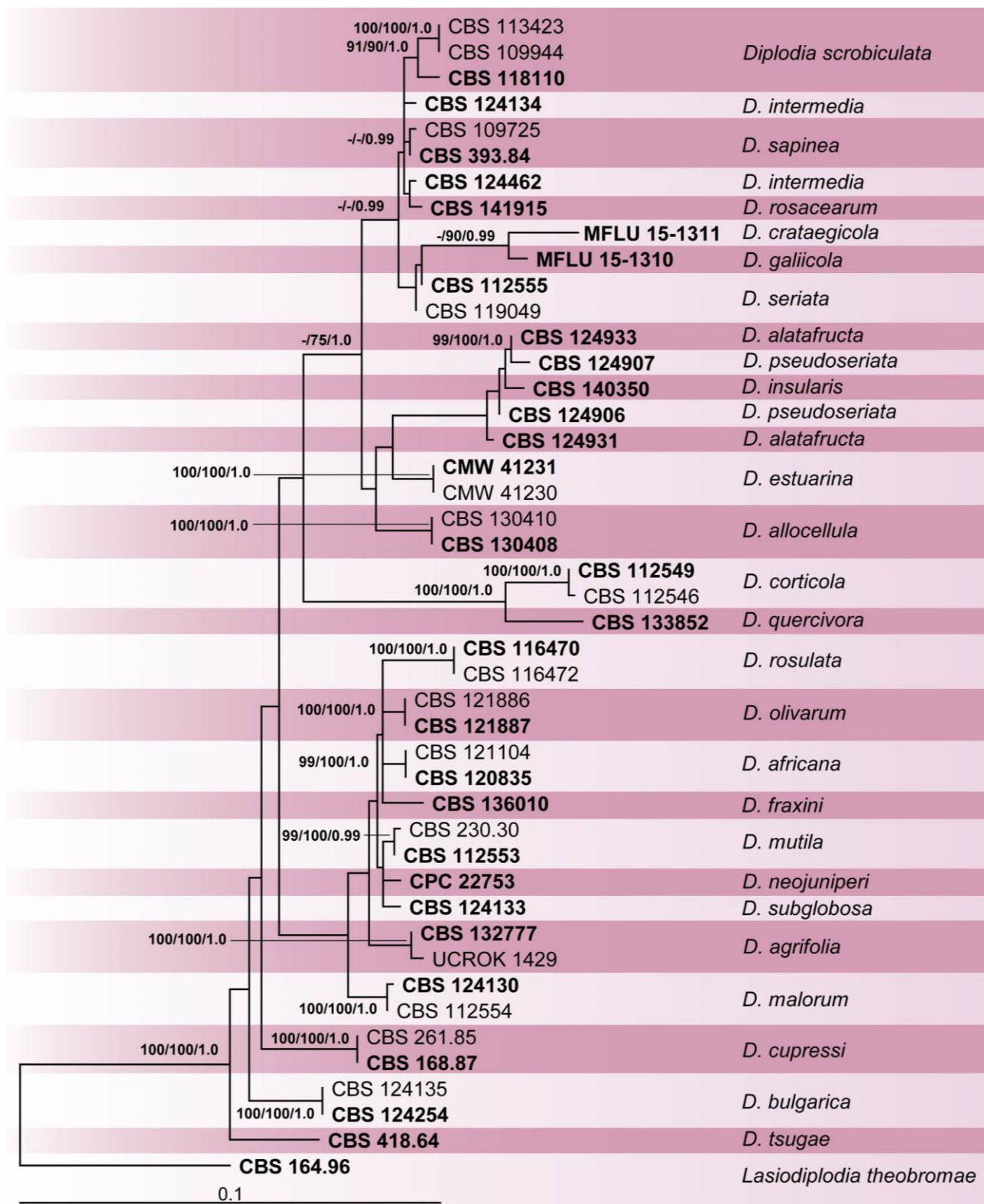


Fig. 3 – Phylogram generated from Maximum Likelihood analysis based on combined ITS and *tefl-α* dataset of *Diplodia*. Bootstrap support values for maximum likelihood (ML), maximum parsimony (MP) greater than 75 % and Bayesian posterior probabilities above 0.90 are indicated near the nodes. The ex-type strains are in bold and the tree is rooted with *Lasiodiplodia theobromae* (CBS 164.96).

Hosts – *Quercus* sp. (Alves et al. 2004, Linaldeddu et al. 2014, Acimovic et al. 2016), *Vitis vinifera* (Úrbez-Torres et al. 2010, Carlucci et al. 2015).

Distribution – Italy (Carlucci et al. 2015, Linaldeddu et al. 2014), Portugal, Spain, Tunisia, USA (Alves et al. 2004, 2014), California (Úrbez-Torres et al. 2010), Maine (Acimovic et al. 2016).

6.7. *Diplodia crataegicola* Dissanayake, Camporesi & K.D. Hyde, *Fungal Diversity* 75: 51 (2015), Facesoffunginumber FoF00885.

Sexual morph not reported. See Ariyawansa et al. (2015) for illustrations and descriptions of asexual morph.

Type – Italy, Province of Forlì-Cesena [FC], Passo del Barbotto–Mercato Saraceno, on dead branch of *Crataegus* sp. (*Rosaceae*), 3 November 2012, E. Camporesi (holotype MFLU 15-1311, cultures ex-type MFLUCC 15–0648, KUMCC15-0075, GZCC 15-0002).

Host – *Crataegus* sp. (Ariyawansa et al. 2015).

Distribution – Italy (Ariyawansa et al. 2015).

6.8. *Diplodia cupressi* A.J.L. Phillips & A. Alves, *Fungal Diversity* 23: 9 (2006), MycoBank MB510136.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Israel, Bet Dagan, dried culture from cankered stems of *Cupressus sempervirens* (*Cupressaceae*), 1986, Z. Solel (holotype IMI 303475, culture ex-type CBS 168.87).

Hosts – *Cupressus* and *Juniperus* sp. (Alves et al. 2006, 2014, Solel et al. 1987).

Distribution – Cyprus, Greece, Israel, Italy, Morocco, South Africa, Tunisia, USA (de Wet et al. 2009, Alves et al. 2006, 2014, Solel et al. 1987).

6.9. *Diplodia estuarina* J.A Osorio, Jol. Roux & Z.W. de Beer, *Fungal Biology* (available online, <http://dx.doi.org/10.1016/j.funbio.2016.09.004>, 2016), MycoBank MB812009.

Sexual morph not reported. See Osorio et al. (2016) for illustrations and description of asexual morph.

Type – South Africa, Kwazulu Province, St. Lucia and Richards Bay, from asymptomatic branches of *Avicennia marina* (*Acanthaceae*), J.A. Osorio & Jol. Roux (holotype PREM 61247, culture ex-type CBS 139666).

Hosts – *A. marina* and *R. mucronata* (Osorio et al. 2016).

Distribution – South Africa (Osorio et al. 2016).

6.10. *Diplodia fraxinii* (Fr. : Fr.) Fr., *Summa Veg. Scand.* 2: 417. 1849. MycoBank MB247549.

Sexual morph not reported. See Alves et al. (2014) for illustrations and descriptions of asexual morph.

Type – Portugal, Monte da Caparica, on dead twigs of *Fraxinus angustifolia* (*Oleaceae*), 14 March 2013, Antonio Deidda (neotype LISE 96134, MBT176183, culture ex-neotype CBS 136010). Cascais, on dead twigs of *F. angustifolia*, 13 April 2013, Antonio Deidda (designated as morphotype A, culture CBS 136012 = CAD010).

Host – *Fraxinus* sp. (Alves et al. 2014).

Distribution – Italy, Netherlands, Portugal (Alves et al. 2014).

6.11. *Diplodia galiicola* Dissanayake, Camporesi & K.D. Hyde, *Fungal Diversity* 75: 54 (2015), Facesoffunginumber FoF00884.

Sexual morph not reported. See Ariyawansa et al. (2015) for illustrations and descriptions of asexual morph. Linaldeddu et al. (2016c) consider this species a synonym of *D. seriata*.

Type – Italy, Province of Forlì-Cesena [FC], Strada San Zeno, Galeata, on dead stem of *Galium* sp. (*Rubiaceae*), 30 October 2013, E. Camporesi, IT 1495 (holotype MFLU 15-1310, culture ex-type MFLUCC 15–0647).

Host – *Galium* sp. (Ariyawansa et al. 2015).

Distribution – Italy (Ariyawansa et al. 2015).

6.12. ***Diplodia insularis*** Linaldeddu, A. Alves & A.J.L. Phillips, *Mycosphere* 7: 968 (2016), MycoBank MB818231, Facesoffunginumber FoF 02607.

Sexual morph not reported. See Linaldeddu et al. (2016c this volume) for illustrations and descriptions of the asexual morph.

Type – Italy, Santa Maria Island, isolated from a branch canker of *Pistacia lentiscus* (*Anacardiaceae*), 7 November 2013, Benedetto T. Linaldeddu, (holotype LISE 96309, culture ex-type CBS 140350).

Hosts – *Eriobotrya japonica*, *Fraxinus angustifolia*, *Pistacia lentiscus* (Linaldeddu et al. 2016c this volume).

Distribution – Caprera, Santa Maria, Sardinia (Italy) and Castellón (Spain) (Linaldeddu et al. 2016c this volume).

6.13. ***Diplodia intermedia*** A.J.L. Phillips, J. Lopes & Bobev, *Persoonia* 29: 33 (2012), MycoBank MB19633.

Sexual morph and asexual morph reported. See Phillips et al. (2013) for illustrations and descriptions.

Type – Portugal, Setúbal, Monte da Caparica, dead twigs of *Malus sylvestris* (*Rosaceae*), Mar. 2006, A.J.L. Phillips (holotype CBS H-20190, culture ex-type CBS 124462).

Hosts – *Cydonia*, *Malus* (Phillips et al. 2012).

Distribution – Portugal (Phillips et al. 2012).

6.14. ***Diplodia malorum*** Fuckel, *Jb. nassau. Ver. Naturk.* 23-24: 395 (1870), MycoBank MB246351

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Germany, Rhineland, on *Malus* sp. (*Rosaceae*), 1870, J. Fuckel, Fuckel, Fungi rhenani N° 1706 (holotype in G, isotypes K and M). Portugal, Setúbal, Monte da Caparica, *Malus sylvestris*, Feb. 2006, A.J.L. Phillips (epitype CBS H-201888, culture ex-epitype CBS 124130).

Hosts – *Malus* sp. (Phillips et al. 2012), *Populus alba* (Alves et al. 2014).

Distribution – Germany, Portugal (Phillips et al. 2012), Italy (Alves et al. 2014).

6.15. ***Diplodia mutila*** (Fr. : Fr.) Fr., *Summa Veg. Scand.* 2: 417 (1849), MycoBank MB201741.

Sexual morph and asexual morph reported. See Phillips et al. (2013), Alves et al. (2014) for illustrations and descriptions.

Type – France, Ardenne, Sedan, on bark of *Populus nigra* (*Salicaceae*), date unknown, Montagne (isotype K 99664). PORTUGAL, Beira Litoral, Aveiro, *Populus alba*, 2012, A. Alves, (epitype LISE 96136, culture ex-epitype CBS 136014).

Hosts – Recent studies subsequent to Phillips et al. (2013) confirm the following hosts: *Chamaecyparis lawsoniana*, *Fraxinus excelsior*, *Fraxinus ornus*, *Ilex* sp., *Persea americana*, *Phoenix dactylifera*, *Populus alba*, *Prunus salicina*, *Taxus baccata*, *Vitis vinifera* (Alves et al. 2014). *Aesculus hippocastanum* (Zlatkovic et al. 2016), *Cupressus arizonica* (Zlatkovic et al. 2016), *Malus domestica* (Úrbez-Torres et al. 2016), *P. halepensis* (Zlatkovic et al. 2016), *Phoenix dactylifera* (Mohammadi et al. 2014), *Vitis vinifera* (Carlucci et al. 2015).

Distribution – British Columbia (Úrbez-Torres et al. 2016), England, Italy, Portugal, South Africa, USA (Alves et al. 2014), Italy (Carlucci et al. 2015), Iran (Mohammadi et al. 2014), Montenegro, Serbia (Zlatkovic et al. 2016).

6.16. *Diplodia neojuniperi* T. Trakunyingcharoen, L. Lombard & Crous, *Persoonia* 34: 92 (2014), MycoBank MB810168.

Sexual morph not reported. See Trakunyingcharoen et al. (2015b) for illustrations and descriptions of asexual morph.

Type – Thailand, Chiang Mai province, on leaf of *Juniperus chinensis* (*Cupressaceae*), Feb. 2012, T. Trakunyingcharoen (holotype CBS H-21932, culture ex-type CBS 138652).

Host – *Juniperus chinensis* (Trakunyingcharoen et al. 2015b).

Distribution – Thailand (Trakunyingcharoen et al. 2015b).

6.17. *Diplodia olivarum* A.J.L. Phillips, Frisullo & Lazzizzera, *Fungal Diversity* 31: 67 (2008), MycoBank MB511402.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Italy, Puglia, Lecce, Scorrano, Basco Belvedere, on rotting drupes of *Olea europaea* (*Oleaceae*), Dec. 2004, S. Frisullo (holotype CBS H-19914, culture ex-type CBS 121887).

Hosts – *Ceratonia siliqua* (Alves et al. 2014), *Olea europaea* (Lazzizzera et al. 2008), *Quercus coccifera* (Alves et al. 2014).

Distribution – Italy (Lazzizzera et al. 2008, Alves et al. 2014), Spain (Gramaje et al. 2012), Tunisia (Alves et al. 2014).

6.18. *Diplodia pseudoseriata* C.A. Pérez, Blanchette, Slippers & M.J. Wingf., *Fungal Diversity* 41: 63 (2009), MycoBank MB513545.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Uruguay, Paysandu, Guaviyu, isolated from asymptomatic twig of *Blepharocalyx salicifolius* (*Myrtaceae*), Aug. 2006, C. Pérez (holotype PREM 60264, culture ex-type CBS 124906 (ex-type)).

Hosts – *Acca sellowiana*, *Blepharocalyx salicifolius*, *Eugenia uniflora*, *Eugenia involucrata*, *Hexachlamis edulis*, *Myrceugenia euosma*, *Myrciaria tenella*, *Myrcianthes cisplatensis* (Pérez et al. 2010), *Fraxinus angustifolia* (Alves et al. 2014).

Distribution – Italy (Alves et al. 2014), Uruguay (Pérez et al. 2010).

6.19. *Diplodia quercivora* Linald. & A.J.L. Phillips, *Mycologia* 105: 1269 (2013), MycoBank MB801757.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Tunisia, Tabarka, isolated from branch cankers of *Quercus canariensis* (*Fagaceae*), 20 Sep. 2006, B.T. Linaldeddu (holotype LISE 96110, culture ex-type CBS 133852).

Hosts – *Quercus canariensis* (Linaldeddu et al. 2013), *Quercus virginiana* (Dreaden et al. 2014).

Distribution – Tunisia (Linaldeddu et al. 2013), USA (Dreaden et al. 2014).

6.20. *Diplodia rosacearum* S. Giambra, A. Alves, J. Armengol & S. Burruano, *Mycosphere* 7: 983 (2016), MycoBank MB818575.

Sexual morph not reported. See Giambra et al. (2016) for illustrations and description of the asexual morph.

Type – Italy, Sicily, S. Maria di Gesù, isolated from a branch canker of *Eriobotrya japonica* (*Rosaceae*), March 2015, S. Giambra, (holotype LISE 96310, culture ex-holotype CBS 141915).

Host – *Eriobotrya japonica* (Giambra et al. 2016).

Distribution – Italy (Giambra et al. 2016).

6.21. *Diplodia rosulata* Gure, Slippers & Stenlid, *Mycol. Res.* 109: 1010 (2005), MycoBank MB344348.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Ethiopia, Southeastern Oromia, Gambo, Munessa-Shashamane Forest Enterprise, from seeds of *Prunus africana* (*Rosaceae*), 20 Jul. 2001, A. Gure (holotype CBS H-12357, culture ex-type CBS 116470).

Host – *Prunus africana* (Gure et al. 2005).

Distribution – Ethiopia (Gure et al. 2005).

6.22. *Diplodia sapinea* (Fr.) Fuckel, *Jb. nassau. Ver. Naturk.* 23-24: 393 (1870), MycoBank MB146913.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Sweden, Suecia Smaland, Femsjo, on *Pinus* sp. (*Pinaceae*), E. Fries, Scleromyceti Sueciae Exsiccati No 126, *Sphaeria sapinea* Fries, lectotype: B, isotypes: G, K, E, UPS, C, BR, FH. The Netherlands, Gelderland, Schovenhorst, Putten, Pinetum, on cones of *Pinus nigra*, June 1984, H.A. van der Aa. (Epitype CBS H-18340, culture ex-epitype CBS 393.84).

Host – Mostly in *Pinus* sp. (Alves et al. 2014, Adamson et al. 2015, Zlatkovic et al. 2016).

Distribution – Worldwide.

6.23. *Diplodia scrobiculata* J. de Wet, Slippers & M.J. Wingf., *Mycol. Res.* 107: 562 (2003), MycoBank MB372427.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – USA, Wisconsin, Jackson County, *Pinus banksiana* (*Pinaceae*), 1987, M.A. Palmer, (holotype PREM 57461, culture ex-type CBS 118110).

Hosts – *Olea europaea*, *Arbutus unedo* (Alves et al. 2014), *Pinus banksiana*, *P. resinosa*, and *P. greggii* (de Wet et al. 2003).

Distribution – Europe (France, Italy), Mexico and, USA (California, Minnesota, Wisconsin) (de Wet et al. 2003, Alves et al. 2014).

6.24. *Diplodia seriata* De Not., *Micr. Ital., Dec.* 4: no. 6 (1842), MycoBank MB180468.

Sexual morph and asexual morph reported. See Phillips et al. (2013), Alves et al. (2014) for illustrations and descriptions.

Type – Italy, on dead stems of *Jasminium* sp., 18 Aug. 1837, De Notaris (holotype HERB RO). Portugal, Montemor-o-Novo, on dead stems of *Vitis vinifera* (*Vitaceae*), 31 Jul. 1997, A.J.L. Phillips, (epitype CBS H-19809, culture ex-epitype CBS 112555).

Hosts – Plurivorous.

Distribution – Worldwide.

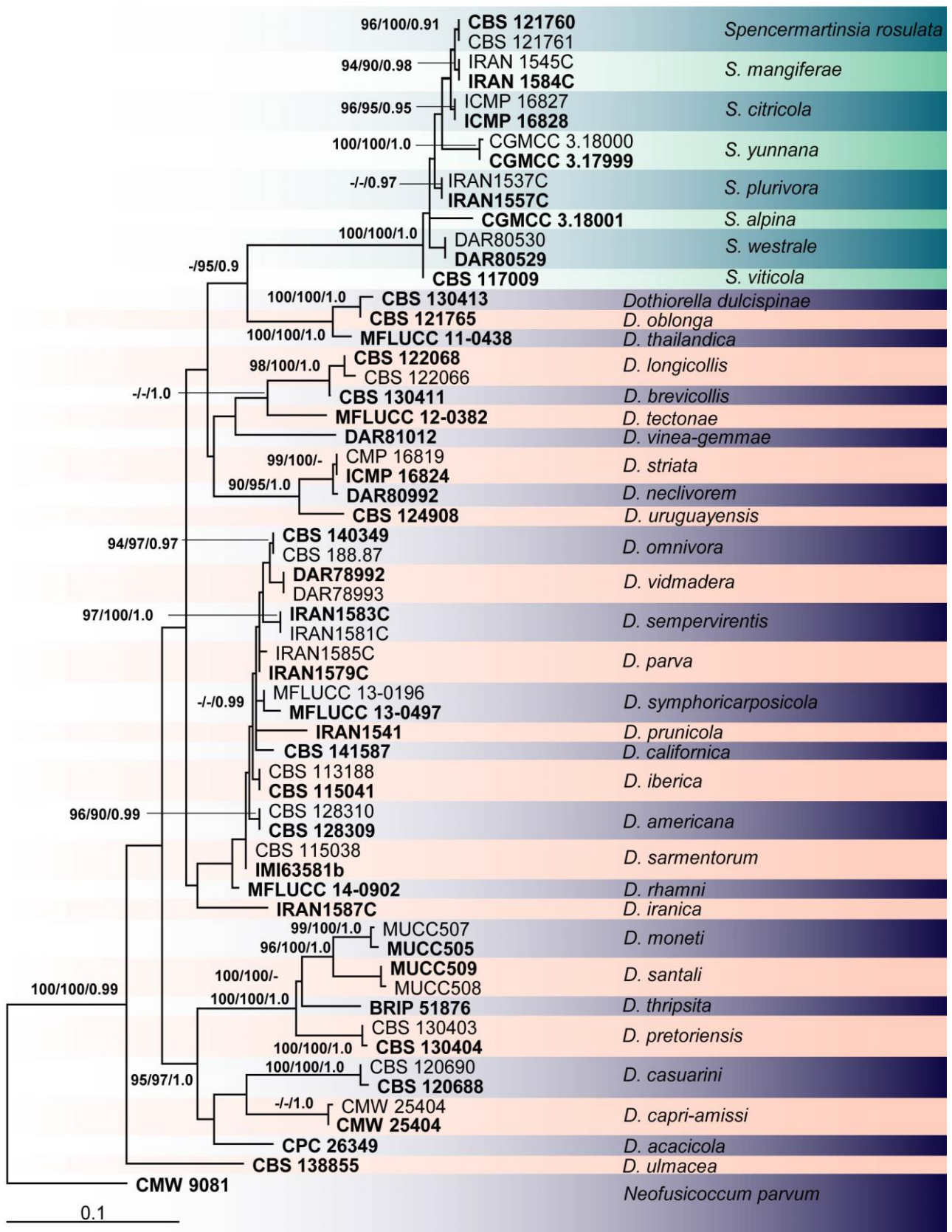


Fig. 4 – Phylogram generated from Maximum Likelihood analysis based on combined ITS and *tef1- α* dataset of *Dothiorella* and *Spencermartinsia*. Bootstrap support values for maximum likelihood (ML), maximum parsimony (MP) greater than 75 % and Bayesian posterior probabilities above 0.90 are indicated near the nodes. The ex-type strains are in bold and the tree is rooted with *Neofusicoccum parvum* (CMW 9081).

6.25. *Diplodia subglobosa* A.J.L. Phillips, Deidda & Linald., *Fungal Diversity* 67: 152 (2014), MycoBank MB806049.

Sexual morph not reported. See Alves et al. (2014) for illustrations and descriptions of asexual morph.

Type – Spain, Cataluña, *Lonicera nigra*, (no date), J. Luque (holotype LISE 96135, culture ex-type CBS 124133). Other cultures: Spain, Cataluña, *Fraxinus excelsior* (*Oleaceae*), (no date), J. Luque (CBS 124132). Italy, Sicily, *Fraxinus ornus*, 2006, A. Sidoti (CBS 124131); *Fraxinus excelsior*, (no date) B. Slippers (CMW7776).

Hosts – *Fraxinus* sp., *Lonicera nigra* (Alves et al. 2014).

Distribution – Italy and Spain (Alves et al. 2014).

6.26. *Diplodia tsugae* (A. Funk) A.J.L. Phillips & A. Alves, *Persoonia* 29: 35 (2012), MycoBank MB801409.

Sexual morph and asexual morph reported. See Funk (1964) for illustrations and Phillips et al. 2013, for descriptions.

Type – Canada, British Columbia, near Coola (Snootli Creek), on branches of *Tsuga heterophylla* (*Pinaceae*), 11 Sep. 1963, A. Funk (holotype DAVFP 15485. Lake Cowichan, 1 Nov. 1962, A. Funk, isotype CBS H-6790, culture ex-isotype CBS 418.64 = IMI 197143).

Host – *Tsuga heterophylla* (Funk 1964).

Distribution – Canada (British Columbia) (Funk 1964).

7. *Dothiorella* Sacc., *Michelia* 2: 5 (1880), MycoBank MB8098.

This genus comprises 30 species (Fig. 4). Sexual morph and the asexual morph have been reported within the genus. The number of species described in this genus has doubled since 2013 when Phillips et al (2013) included 12 species names and 16 unnamed lineages. With this increase in the number of species the phylogenetic separation between *Dothiorella* and *Spencermartinsia* has become less distinct. Nevertheless, we continue to regard these as two genera pending a detailed re-evaluation.

Type species – *Dothiorella pyrenophora* Sacc., *Michelia* 2: 5. 1880.

7.1. *Dothiorella acacicola* Crous & M.J. Wingf., *Persoonia* 36: 354 (2016), MycoBank MB817032.

Sexual morph not reported. See Crous et al. (2016) for illustrations and descriptions of asexual morph.

Type – France, La Réunion, S21°12'47.6" E55°36'48.7", on leaves of *Acacia mearnsii* (*Fabaceae*), 8 Mar. 2015, P.W. Crous & M.J. Wingfield (holotype CBS H-22607, culture ex-type CPC 26349 = CBS 141295).

Host – *Acacia mearnsii* (Crous et al. 2016).

Distribution – France (Crous et al. 2016).

7.2. *Dothiorella americana* Urb.-Torr., Peduto & Gubler, *Fungal Diversity* 52: 184 (2012), MycoBank MB519956.

Sexual morph not reported. See Úrbez-Torres et al. (2012) for illustrations and descriptions of asexual morph.

Type – USA, Missouri, Purdy, on diseased interspecific grape cultivar Vignoles (Ravat51), R.K. Striegler & G.M. Leavitt (holotype UCD2252MO, culture ex-type CBS 128309).

Host – *Vitis* sp. (Úrbez-Torres et al. 2012).

Distribution – USA (Missouri) (Úrbez-Torres et al. 2012).

7.3. *Dothiorella brevicollis* Jami, Gryzenh., Slippers & M.J. Wingf., *Cryptog. Mycol.* 33: 256 (2012), MycoBank MB564142.

Sexual morph not reported. See Jami et al. (2012) for illustrations and descriptions of asexual morph.

Type – South Africa, Gauteng Province, Pretoria, from healthy wood section of *Acacia karroo* (*Fabaceae*), Nov. 2009, F. Jami (holotype PREM 60704, culture ex-type CBS 130411).

Host – *Acacia karroo* (Jami et al. 2012).

Distribution – South Africa (Gauteng Province) (Jami et al. 2012).

7.4. *Dothiorella californica* D.P. Lawr. & Trouillas, *Fungal Biology* (available online, <http://dx.doi.org/10.1016/j.funbio.2016.09.005>, 2016), MycoBank MB817293.

Sexual morph not reported. See Lawrence et al. (2016) for illustrations and descriptions of asexual morph.

Type – USA, California: Napa County, 38.4367° N, 122.4028° W, near Oakville 294 CA, 47 m.a.s.l., isolated from discolored wood of *Umbellularia californica* (*Lauraceae*) November 2004, F.P. Trouillas (holotype BPI 910162, culture ex-type CBS 141587).

Host – *Umbellularia californica* (Lawrence et al. 2016).

Distribution – California (Lawrence et al. 2016).

7.5. *Dothiorella capri-amissi* F.J.J. van der Walt, Slippers & G.J. Marais, *Persoonia* 33: 162 (2014), MycoBank MB518723.

Sexual morph not reported. See Slippers et al. (2014) for illustrations and descriptions of asexual morph.

Type – South Africa, Prieska, from *Acacia erioloba* (*Fabaceae*), Feb. 2005, F.J.J. van der Walt & G.J. Marais (holotype PREM 59626, culture ex-type CBS 121878).

Host – *Acacia erioloba* (Slippers et al. 2014).

Distribution – South Africa (Slippers et al. 2014).

7.6. *Dothiorella casuarini* J. de Wet, Slippers & M.J. Wingf., *Mycologia* 101: 505 (2009), MycoBank MB510856.

Sexual morph not reported. See de Wet et al. (2009) for illustrations and descriptions of asexual morph.

Type – Australia, Canberra, Cotter River, on *Casuarina* sp. (*Casuarinaceae*), 2000, M.J. Wingfield (holotype PREM 59650, culture ex-type CBS 120688).

Host – *Casuarina* sp. (de Wet et al. 2009).

Distribution – Australia (Canberra) (de Wet et al. 2009).

7.7. *Dothiorella dulcispinae* Jami, Gryzenh., Slippers & M.J. Wingf., *Cryptog. Mycol.* 33: 256 (2012), MycoBank MB564141.

Sexual morph not reported. See Jami et al. (2012) for illustrations and descriptions of asexual morph.

Type – South Africa, Gauteng Province, Pretoria, from die-back wood section of *Acacia karroo* (*Fabaceae*), Nov. 2009, F. Jami (holotype PREM 60706, culture ex-type CBS 130413).

Hosts – *Acacia karroo* (Jami et al. 2012), *Acacia mellifera* (Slippers et al. 2014).

Distribution – Namibia (Slippers et al. 2014), South Africa (Gauteng Province) (Jami et al. 2012).

7.8. *Dothiorella iberica* A.J.L. Phillips, J. Luque & A. Alves, *Mycologia* 97: 524 (2005), MycoBank MB344530.

Sexual morph and asexual morph reported. See Phillips et al. (2013) for illustrations and descriptions.

Type – Spain, Zaragoza province, Aragon, Tarazona, on dead twigs of *Quercus ilex* (*Fagaceae*), Dec. 2002, J. Luque (holotype LISE 94944, culture ex-type CBS 115041).

Hosts – Recent studies subsequent to Phillips et al. (2013) confirm the following hosts: *Acer opalus* (Dissanayake et al. 2016), *Citrus sinensis* (Adesemoye et al. 2014), *Ostrya* sp. (Pavlic-Zupanc et al. 2015), *Pistacia* sp. (Chen et al. 2014b), *Prunus dulcis* (Doll et al. 2015), *Quercus agrifolia* (Lynch et al. 2014).

Distribution – Recent studies subsequent to Phillips et al. (2013) confirm the following distribution: California (Adesemoye et al. 2014, Chen et al. 2014b, Doll et al. 2015, Lynch et al. 2014), Italy (Pavlic-Zupanc et al. 2015, Dissanayake et al. 2016).

7.9. *Dothiorella iranica* Abdollahz., Zare & A.J.L. Phillips, *Persoonia* 32: 4 (2014), MycoBank MB803988.

Sexual morph not reported. See Abdollahzadeh et al. (2014) for illustrations and descriptions of asexual morph.

Type – Iran, Golestan Province, Gorgan (Agriculture Research Center), on twigs of *Olea europea* (*Oleaceae*), June 2007, A. Javadi (holotype IRAN 16253F, culture ex-type IRAN 1587C = CBS 124722).

Hosts – *Olea europea* (Abdollahzadeh et al. 2014), *Paliurus* sp. (Hyde et al. 2016)

Distribution – Iran (Abdollahzadeh et al. 2014), Italy (Hyde et al. 2016)

7.10. *Dothiorella longicollis* Pavlic, T.I. Burgess & M.J. Wingf., *Mycologia* 100: 859 (2008), MycoBank MB512053.

Sexual morph not reported. See Pavlic et al. (2008) for illustrations and descriptions of asexual morph.

Type – Australia, Western Australia, Tunnel Creek National Park, on healthy branches of *Lysiphyllum cunninghamii* (*Fabaceae*), Jul. 2006, T.I. Burgess (holotype PREM 59485, culture ex-type CBS 122068).

Hosts – *Lysiphyllum cunninghamii*, *Terminalia* sp. (Pavlic et al. 2008)

Distribution – Australia (Pavlic et al. 2008).

7.11. *Dothiorella moneti* K. Taylor, Barber, G.E. Hardy & T.I. Burgess, *Mycol. Res.* 113: 342 (2009), MycoBank MB511825.

Sexual morph not reported. See Taylor et al. (2009) for illustrations and descriptions of asexual morph.

Type – Australia, Western Australia, Yalgorup National Park, from healthy stem of *Acacia rostellifera* (*Fabaceae*), Jun. 2005, K.M. Taylor (holotype PERTH 07692978, culture ex-type MUCC 505 = WAC 13154).

Host – *Acacia rostellifera* (Taylor et al. 2009).

Distribution – Australia (Taylor et al. (2009).

7.12. *Dothiorella neclivorem* W.M. Pitt & J.R. Úrbez-Torres, *Australas. Pl. Path.* 44: 49 (2015), MycoBank MB808287.

Sexual morph not reported. See Pitt et al. (2015) for illustrations and descriptions of asexual morph.

Type – Australia, Pokolbin, New South Wales, berries of Chardonnay, November 2008, N. Wunderlich (holotype DAR80992, culture ex-type DAR80992).

Host – *Vitis vinifera* (Pitt et al. 2015).

Distribution – Australia (Pitt et al. 2015).

7.13. *Dothiorella oblonga* F.J.J. Van der Walt, Slippers & G.J. Marais, *Persoonia* 33: 163 (2014), MycoBank MB518719.

Sexual morph not reported. See Slippers et al. (2014) for illustrations and descriptions of asexual morph.

Type – South Africa, Pretoria, Ditholo, *Acacia mellifera*, May 2006, F.J.J. van der Walt & R.N. Heath (holotype PREM 59628, culture ex-type CBS 121765); Ditholo, *A. mellifera* (paratype PREM 59629, culture ex-paratype CBS 121766). Namibia, Rundu, *A. mellifera*, Feb. 2006, F.J.J. van der Walt & J. Roux (paratype PREM 59627, culture ex-paratype CBS 121764).

Host – *Acacia mellifera* (Slippers et al. 2014).

Distribution – Namibia, South Africa (Slippers et al. 2014).

7.14. *Dothiorella omnivora* Linaldeddu, Deidda & Scanu, *Eur J Plant Pathol* 146: 272 (2016), MycoBank MB812898.

Sexual morph not reported. See Linaldeddu et al. (2016a) for illustrations and descriptions of asexual morph.

Type – Italy, Aritzo, on cankered branch of *Corylus avellana* (*Betulaceae*), 09 July 2008, B.T. Linaldeddu (holotype LISE 96304, culture extype CBS 140349).

Hosts – *Chamaecyparis lawsoniana*, *Corylus avellana*, *Fraxinus excelsior*, *Juglans regia*, *Ostrya carpinifolia*, *Thuja occidentalis*, *Vitis vinifera* (Linaldeddu et al. 2016a).

Distribution – Australia, Bosnia and Herzegovina, France, Iran, Italy, Serbia (Linaldeddu et al. 2016a).

7.15. *Dothiorella parva* Abdollahz., Zare & A.J.L. Phillips, *Persoonia* 32: 5 (2014), MycoBank MB803989.

Sexual morph not reported. See Abdollahzadeh et al. (2014) for illustrations and descriptions of asexual morph.

Type – Iran, Ardabil Province, Ardabil (Fandoghlo Forest Park), on twigs of *Corylus avellana*, July 2007, J. Abdollahzadeh & A. Javadi (holotype IRAN 14264F, culture ex-type IRAN 1579C = CBS 124720).

Hosts – *Corylus avellana* (Abdollahzadeh et al. 2014), *Ostrya carpinifolia* (Pavlic-Zupanc et al. 2015).

Distribution – Iran (Abdollahzadeh et al. 2014), Italy (Pavlic-Zupanc et al. 2015).

7.16. *Dothiorella pretoriensis* (Jami, Gryzenh., Slippers & M.J. Wingf.) Abdollahz. & A.J.L. Phillips, *Stud. Mycol.* 76: 108 (2013), MycoBank MB803995.

Sexual morph not reported. See Jami et al. (2012) for illustrations and descriptions of asexual morph.

Type – South Africa, Gauteng, Pretoria, from wood of *Acacia karroo* (*Fabaceae*) with die-back symptoms, Nov. 2009, F. Jami (holotype PREM 60709, culture ex-type CBS 130404).

Host – *Acacia karroo* (Jami et al. 2012).

Distribution – South Africa (Jami et al. 2012).

7.17. *Dothiorella prunicola* A.J.L. Phillips & Abdollahz., *Persoonia* 32: 6 (2014), MycoBank MB803991.

Sexual morph not reported. See Abdollahzadeh et al. (2014) for illustrations and descriptions of asexual morph.

Type – Portugal, Algarve Province, on twigs of *Prunus dulcis* (*Rosaceae*), June 2007, E. Diogo (holotype IRAN 16252F, culture ex-type IRAN 1541C = CBS 124723).

Host – *Prunus dulcis* (Abdollahzadeh et al. 2014).

Distribution – Portugal (Abdollahzadeh et al. 2014).

7.18. *Dothiorella rhamni* Wanasinghe, Bulgakov, E.B.G. Jones & K.D. Hyde, *Fungal Diversity* 78: (2016), Facesoffunginumber FoF01668.

Sexual morph not reported. See Li et al. (2016a) for illustrations and descriptions of asexual morph.

Type – Russia, Rostov region, Oktyabrsky District, near natural sanctuary “Persianovskaya steppe”, Khoruli hollow, ravine grove (47.5006484° E, 40.1385927° N), on *Rhamnus cathartica* (*Rhamnaceae*), 26 April 2014, T.S. Bulgakov (holotype MFLU 15–3541, culture ex-type MFLUCC 14–0902).

Host – *Rhamnus cathartica* (Li et al. 2016a).

Distribution – Russia (Li et al. 2016a).

7.19. *Dothiorella santali* K. Taylor, Barber & T.I. Burgess, *Mycol. Res.* 113: 345 (2009), MycoBank MB511828.

Sexual morph not reported. See Taylor et al. (2009) for illustrations and descriptions of asexual morph.

Type – Australia, Western Australia, Yalgorup National Park, from healthy stem of *Santalum acuminatum* (*Santalaceae*), Jun. 2005, K.M. Taylor (holotype PERTH 07693028, culture ex-type MUCC 509 = WAC 13155).

Host – *Santalum acuminatum* (Taylor et al. 2009).

Distribution – Australia (Taylor et al. 2009).

7.20. *Dothiorella sarmentorum* (Fr.) A.J.L. Phillips, A. Alves & J. Luque, *Mycologia* 97: 522 (2005), MycoBank MB501403.

Sexual morph and asexual morph reported. See Phillips et al. (2013) for illustrations and descriptions.

Type – England, Warwickshire, on *Ulmus* sp. (*Ulmaceae*), Aug. 1956, E.A. Ellis, (holotype and culture ex-type IMI 63581b [as *Othia spiraeae*]).

Hosts – Plurivorous. See Dissanayake et al. (2016) for host details (identifications are supported by molecular data of post-2005).

Distribution – Worldwide, See Dissanayake et al. (2016) for distribution of *D. sarmentorum*.

7.21. *Dothiorella sempervirentis* Abdollahz., Zare & A.J.L. Phillips, *Persoonia* 32: 6 (2014), MycoBank MB803987.

Sexual morph not reported. See (Abdollahzadeh et al. 2014) for illustrations and descriptions of asexual morph.

Type – Iran, Golestan Province, Gorgan (City Park), on twigs and cones of *Cupressus sempervirens* (*Cupressaceae*), Aug. 2006, M.A. Aghajani (holotype IRAN 14265F, culture ex-type IRAN 1583C = CBS 124718).

Host – *Cupressus sempervirens* (Abdollahzadeh et al. 2014).

Distribution – Iran (Abdollahzadeh et al. 2014).

7.22. *Dothiorella striata* A.J.L. Phillips & Abdollahz., *Persoonia* 32: 7 (2014), MycoBank MB803990.

Sexual morph not reported. See (Abdollahzadeh et al. 2014) for illustrations and descriptions of asexual morph.

Type – New Zealand, Northland, Kerikeri, Collins Orchard, Inlet Road, on twigs of *Citrus sinensis* (*Rutaceae*), Sept. 2006, S.R. Pennycook, P.R. Johnston & B.C. Paulus (holotype PDD 92029, culture ex-type ICMP 16824 = CBS 124731).

Host – *Citrus sinensis* (Abdollahzadeh et al. 2014).

Distribution – New Zealand (Abdollahzadeh et al. 2014).

7.23. *Dothiorella symphoricarposicola* W.J. Li, J.K. Liu & K.D. Hyde, *Cryptogamie Mycologie*, 35: 257 (2014), Facesoffunginumber FOF000024.

Sexual morph not reported. See (Li et al. 2014) for illustrations and descriptions of asexual morph.

Type – Italy, Forlì-Cesena [FC], Berleta-Santa Sofia, on dead bark of *Symphoricarpos* sp. (*Caprifoliaceae*), 18 May 2013, Erio Camporesi, IT-1075 (holotype MFLU 14-0217, cultures ex-type MFLUCC 13-0497 and MFLUCC 13-0498); Spinello-Santa Sofia, on dead bark of *Symphoricarpos* sp., 28 June 2013, Erio Camporesi IT-1075A (paratype MFLU 14-0297).

Hosts – *Laburnum alpinum*, *Laurus nobilis*, *Sambucus nigra*, *Symphoricarpos* sp. (Li et al. 2014, Dissanayake et al. 2016).

Distribution – Italy (Li et al. 2014, Dissanayake et al. 2016).

7.24. *Dothiorella tectonae* Doilom, L.A. Shuttleworth, & K.D. Hyde, *Phytotaxa* 233: 001 (2015), Facesoffunginumber FoF00165.

Sexual morph not reported. See Doilom et al. (2015) for illustrations and descriptions of asexual morph.

Type – Thailand, Phayao Province, Muang District, on dead branch of *Tectona grandis* (*Lamiaceae*), 12 March 2012, M. Doilom (holotype MFLU 14-0272, culture ex-type MFLUCC 12-0382, MUCL 55409).

Host – *Tectona grandis* (Doilom et al. 2015).

Distribution – Thailand (Doilom et al. 2015).

7.25. *Dothiorella thailandica* Abdollahz., A.J.L. Phillips & A. Alves, *Stud. Mycol.* 76: 110 (2013), MycoBank MB805461.

Sexual morph not reported. See Liu et al. (2012) for illustrations and descriptions of asexual morph.

Type – Thailand, Chiang Rai Province, Doi Pui, on dead bamboo culm (*Poaceae*), 1 Sep. 2011, D.Q. Dai (holotype MFLU 12-0751, culture ex-type MFLUCC 11-0438 = CBS 133991).

Host – Bamboo (Liu et al. 2012).

Distribution – Thailand (Liu et al. 2012).

7.26. *Dothiorella thripsita* R.G. Shivas & D.J. Tree, *Persoonia* 22: 169 (2009), MycoBank MB513166.

Sexual morph not reported. See Shivas et al. (2009) for illustrations and descriptions of asexual morph.

Type – Australia, Queensland, Tallegalla, on dead stems and phyllodes of *Acacia harpophylla* (*Fabaceae*), Mar. 2008, D.J. Tree & C.E.C. Tree (holotype BRIP 51876, culture ex-type BRIP 51876).

Host – *Acacia harpophylla* (Shivas et al. 2009).

Distribution – Australia (Shivas et al. 2009).

7.27. *Dothiorella ulmacea* Crous & R.K. Schumach., *Sydowia* 67: 100 (2015), MycoBank MB812527.

Sexual morph not reported. See Crous et al. (2015b) for illustrations and descriptions of asexual morph.

Type – Germany, on a twig of *Ulmus laevis* (*Ulmaceae*), 1 Mar. 2014, leg. R. K. Schumacher (holotype CBS H-22257, culture ex-type CBS 138855); other specimen and culture on twigs of *Ulmus laevis*, 9 July 2014, leg. R. K. Schumacher (CBS H-22282, culture CPC 24945 = CBS 140005).

Host – *Ulmus laevis* (Crous et al. 2015b).

Distribution – Germany (Crous et al. 2015b).

7.28. *Dothiorella uruguayensis* (C.A. Pérez, Blanchette, Slippers & M.J. Wingf.) Abdollahz. & A.J.L. Phillips, *Stud. Mycol.* 76: 110 (2013), MycoBank MB803999.

Sexual morph not reported. See Pérez et al. (2010) for illustrations and descriptions of asexual morph.

Type – Uruguay, Paysandu, Tres Bocas, endophytic on twigs of *Hexachlamis edulis*, Aug. 2006, C.A. Pérez (holotype PREM 60268, culture ex-type CBS 124908).

Host – *Hexachlamis edulis* (Pérez et al. 2010).

Distribution – Uruguay (Pérez et al. 2010).

7.29. *Dothiorella vidmadera* W.M. Pitt, Úrbez-Torres & Trouillas, *Fungal Diversity Res. Ser.* 61: 216 (2013), MycoBank MB803533.

Sexual morph and asexual morph reported. See Pitt et al. (2013b) for illustrations and descriptions of asexual morph and Li et al. (2016a) for illustrations and descriptions of sexual morph.

Type – Australia, EdenValley, South Australia, *Vitis vinifera* (*Vitaceae*) W.M. Pitt & A. Loschiavo, (holotype DAR78992, culture ex-type DAR78992).

Hosts – *Fraxinus ornus* (Li et al. 2016a), *Robinia pseudoacacia* (Hyde et al. 2016), *Vitis vinifera* (Pitt et al. 2013b).

Distribution – Australia (Pitt et al. 2013b), Italy (Li et al. 2016a), Russia (Hyde et al. 2016).

7.30. *Dothiorella vinea-gemmae* W.M. Pitt & J.R. Úrbez-Torres, *Australas. Pl. Path.* 44: 51 (2015), MycoBank MB808288.

Sexual morph not reported. See Pitt et al. (2015) for illustrations and descriptions of asexual morph.

Type – Australia, Pokolbin, New South Wales, *Vitis vinifera* (*Vitaceae*), November 2008, N. Wunderlich (holotype, DAR81012, culture ex-type, DAR81012).

Host – *Vitis vinifera* (Pitt et al. 2015).

Distribution – Australia (Pitt et al. 2015).

8. *Endomelanconiopsis* E.I. Rojas & Samuels, *Mycologia* 100: 770 (2008), MycoBank MB511837. *Endomelanconiopsis* comprises only two species and only the asexual morph has been reported within the genus.

Type species – *Endomelanconiopsis endophytica* E.I. Rojas & Samuels 2008.

8.1. *Endomelanconiopsis endophytica* E.I. Rojas & Samuels, *Mycologia* 100: 770 (2008), MycoBank MB511838.

Sexual morph not reported. See Rojas et al. (2008) for illustrations and descriptions of asexual morph.

Type – Panama, Nombre de Dios, isolated from leaves of *Theobroma cacao* (*Malvaceae*), 2000, E. Rojas, L. Mejía & Z. Maynard (holotype BPI 878370, culture ex-type CBS 120379).

Hosts – *Carapa guianensis* (Ferreira et al. 2015), *Heisteria concinna*, *Theobroma cacao* (Rojas et al. 2008)

Distribution – Brazil (Ferreira et al. 2015), Panama (Rojas et al. 2008).

8.2. *Endomelanconiopsis microspora* (Verkley & Aa) E.I. Rojas & Samuels, *Mycologia* 100: 770 (2008), MycoBank MB511839.

Sexual morph not reported. See Verkley & van der Aa (1997) for illustrations and descriptions of asexual morph.

Type – Papua New Guinea, Central Province, 22 km E of Port Moresby, Varirata National Park near Varirata Lookout, soil in dry secondary forest with *Casuarina* and *Eucalyptus*, and conglomerate rock outcrops, 23 Oct. 1995, A. Aptroot, H.A. van der Aa (holotype CBS H-12183, culture ex-type CBS 353.97).

Substrate – Soil (Verkley & van der Aa 1997).

Distribution – Papua New Guinea (Verkley & van der Aa 1997).

9. *Eutiarosporella* Crous, *Phytotaxa* 202: 85 (2015), MycoBank MB811248

This genus was introduced for tiarosporella-like fungi with long conidiomatal necks and holoblastic conidiogenesis (Crous et al. 2015a). The genus comprises seven species. Both sexual morph and asexual morph have been reported within the genus.

Type species – *Eutiarosporella tritici* (B. Sutton & Marasas) Crous 2015.

9.1. *Eutiarosporella africana* (Jami, Gryzenh., Slippers & M.J. Wingf.) Crous, *Phytotaxa* 202: 85 (2015), MycoBank MB811249.

Sexual morph not reported. See Jami et al. (2014) for illustrations and descriptions of asexual morph.

Type – South Africa, Gauteng Province, Pretoria, from healthy wood section of *Celtis africana* (*Cannabaceae*), Nov. 2011, F. Jami & M. Gryzenhout (holotype PREM 60866, culture ex-type CBS 133854).

Host – *Celtis Africana* (Jami et al. 2014).

Distribution – South Africa (Jami et al. (2014).

9.2. *Eutiarosporella dactylidis* (Thambug., Camporesi & K.D. Hyde) Dissanayake, Camporesi & K.D. Hyde, *Fungal Diversity* 78: (2016), Facesoffunginumber FoF00334

Sexual morph and asexual morph reported. See Thambugala et al. (2014) for illustrations and descriptions.

Type – Italy, Teodorano – Meldola (province of Forlì-Cesena [FC]), on dead leaves of *Dactylis glomerata* (*Poaceae*), 15 December 2012, Erio Camporesi (holotype MFLU 14-0580, cultures ex-type MFLUCC 13-0276 = ICMP 20383).

Hosts – *Arrhenatherum eliatum*, *Dactylis glomerata* (Thambugala et al. 2014), *Avenella flexuosa* (Li et al. 2016a)

Distribution – Italy (Thambugala et al. 2014, Li et al. 2016a).

9.3. *Eutiarosporella darliae* E. Thynne, M.C. McDonald, M. Evans, H. Wallwork, S. Neate & P.S. Solomon, *Australas. Pl. Path.* 44: 533 (2015), MycoBank MB811116.

Sexual morph not reported. See Thynne et al. (2015) for illustrations and descriptions of asexual morph.

Type – Australia, Queensland, isolated from *Triticum aestivum* (*Poaceae*), 2012, QLD DPI, (holotype and culture ex-type DAR 82491).

Host – *Triticum aestivum* (Thynne et al. 2015).

Distribution – Australia (Thynne et al. 2015).

9.4. *Eutiarosporella pseudodarliae* E. Thynne, M.C. McDonald, M. Evans, H. Wallwork, S. Neate & P.S. Solomon, *Australas. Pl. Path.* 44: 534 (2015), MycoBank MB 811117.

Sexual morph not reported. See Thynne et al. (2015) for illustrations and descriptions of asexual morph.

Type – Australia, Queensland, isolated from *Triticum aestivum* (*Poaceae*), 2011, QLD DPI (holotype and culture ex-type DAR 82489).

Host – *Triticum aestivum* (Thynne et al. 2015).

Distribution – Australia (Thynne et al. 2015).

9.5. *Eutiarosporella tritici* (B. Sutton & Marasas) Crous, *Phytotaxa* 202: 85 (2015), MycoBank MB811250.

Sexual morph not reported. See (Crous et al. 2006) for illustrations and descriptions of asexual morph.

Type – South Africa: Free State Province: Heilbron, on *Triticum aestivum* (*Poaceae*), 18 Jan. 1973, W.F.O. Marasas (holotype PREM 44966, isotype IMI 186786, culture ex-type CBS 118719).

Host – *Triticum aestivum* (Sutton & Marasas 1976).

Distribution – South Africa (Sutton & Marasas 1976).

9.6. *Eutiarosporella tritici-australis* E. Thynne, M.C. McDonald, M. Evans, H. Wallwork, S. Neate & P.S. Solomon, *Australas. Pl. Path.* 44: 532 (2015), MycoBank MB 811115.

Sexual morph not reported. See Thynne et al. (2015) for illustrations and descriptions of asexual morph. In our families tree (Fig. 1) *Eutiarosporella tritici-australis* appears to be more closely related to the genus *Mucoharknessia*. Based on ITS sequence comparison, Thynne et al. (2015) noted that some of their isolates cluster with *M. cortaderiae*. However, the morphological description of *M. cortaderiae* is in contrast with the isolates obtained from white grain disorder of wheat (Thynne et al. 2015). On account of their morphology, Thynne et al. (2015) concluded that the white grain disorder isolates reside in *Eutiarosporella* and not *Mucoharknessia*. Subsequently, Li et al. (2016a) introduced a new species in *Mucoharknessia*; *M. anthoxanthii* with morphology similar to *Eutiarosporella*. For these reasons, we consider that the position of *E. tritici-australis* in *Eutiarosporella* is tenable and needs to be re-assessed.

Type – Australia, South Australia, isolated from *Triticum aestivum* (*Poaceae*), 2012, SARDI (holotype and culture ex-type DAR 82485).

Host – *Triticum aestivum* (Thynne et al. 2015).

Distribution – Australia (Thynne et al. 2015).

9.7. *Eutiarosporella urbis-rosarum* (Jami, Gryzenh., Slippers & M.J. Wingf.) Crous, *Phytotaxa* 202: 85 (2015), MycoBank MB811251.

Sexual morph not reported. See Jami et al. (2012) for illustrations and descriptions of asexual morph.

Type – South Africa: Free State Province, Bloemfontein, healthy wood of *Vachellia karroo* (*Fabaceae*), June 2008, M. Gryzenhout (holotype PREM 60698, culture ex-type CBS 130405).

Host – *Vachellia karroo* (Jami et al. 2012).

Distribution – South Africa (Jami et al. 2012).

10. ***Lasiodiplodia*** Ellis & Everh., *Bot. Gaz.* 21: 92 (1896), MycoBank MB8708.

This genus comprises 31 species (Fig. 5). Both sexual asexual morph have been reported within the genus. Phillips et al. (2013) and Slippers et al. (2014) have shown that morphology is not a reliable character for species differentiation and species can be recognized only from combined ITS and *tefl-α* sequence data. Cruywagen et al. (2016) suggest that hybridization between *Lasiodiplodia* species is widespread and further suggest that some of the currently recognized species may in fact be hybrids, e.g. *L. viticola*, *L. missouriana*, *L. laeliocattletae* and *L. brasiliense*. Nevertheless, in the following list they are treated as distinct species until hybridization is confirmed.

Type species – *Lasiodiplodia theobromae* (Pat.) Griff. & Maubl., *Bull. trimest. Soc. Mycol. Fr.* 25: 57. 1909.

10.1. ***Lasiodiplodia avicenniae*** J.A. Osorio, Jol. Roux & Z.W. de Beer, *Fungal Biology* (available online, <http://dx.doi.org/10.1016/j.funbio.2016.09.004>, 2016), MycoBank MB812010.

Sexual morph not reported. See Osorio et al. (2016) for illustrations and description of asexual morph.

Type – South Africa, Kwazulu-Natal Province, Beachwood and Isipingo, from asymptomatic branches of *Avicennia marina* (*Acanthaceae*), J.A Osorio & Jol. Roux (holotype PREM 61249, cultures ex-type CBS 139670).

Host – *Avicennia marina* (Osorio et al. 2016).

Distribution – South Africa (Osorio et al. 2016).

10.2. ***Lasiodiplodia brasiliense*** M.S.B. Netto, M.W. Marques & A.J.L. Phillips, *Fungal Diversity* 67: 134 (2014), MycoBank MB807525.

Sexual morph not reported. See Netto et al. (2014) for illustrations and descriptions of asexual morph.

Type – Brazil, Pernambuco, Farm Dan, on *Mangifera indica* stems (*Anacardiaceae*), 2010, M.W. Marques (holotype URM 85580, ex-type CMM4015 = URM7118).

Hosts – *Carica papaya*, *Mangifera indica* (Netto et al. 2014), *Cocos nucifera* (Rosado et al. 2016), *Tectona grandis* (Doilom et al. 2015), *Vitis vinifera* (Correia et al. 2016a).

Distribution – Brazil (Netto et al. 2014, Correia et al. 2016a, Rosado et al. 2016), Thailand (Doilom et al. 2015).

10.3. ***Lasiodiplodia bruguierae*** J.A. Osorio, Jol. Roux & Z.W. de Beer, *Fungal Biology* (available online, <http://dx.doi.org/10.1016/j.funbio.2016.09.004>, 2016), MycoBank MB812011.

Sexual morph not reported. See Osorio et al. (2016) for illustrations and descriptions.

Type – South Africa, Kwazulu-Natal Province, Mlalazi Nature Reserve, Mtunzini, from asymptomatic branches of *Bruguiera gymnorrhiza* (*Rhizophoraceae*), J.A Osorio & Jol. Roux (holotype PREM 61248, culture ex-type CBS 139669).

Host – *Bruguiera gymnorrhiza* (Osorio et al. 2016).

Distribution – South Africa (Osorio et al. 2016).

10.4. ***Lasiodiplodia caatinguensis*** I.B.L. Coutinho, F.C.O Freire, C.S. Lima & J.E. Cardoso, *Plant Pathology* (available online, Doi: 10.1111/ppa.12565, 2016), MycoBank MB815122.

Sexual morph not reported. See Coutinho et al. (2016) for illustrations and descriptions of asexual morph.

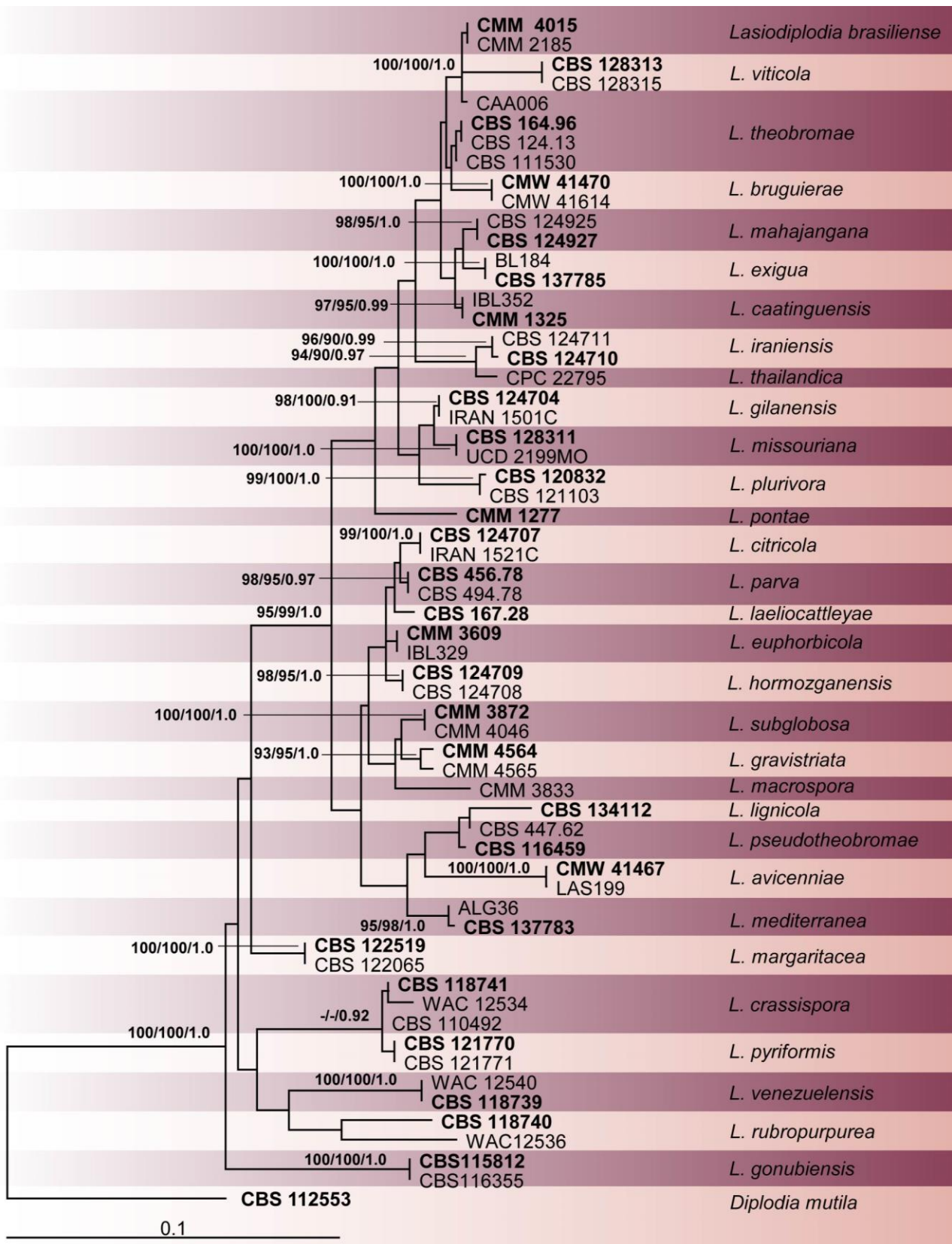


Fig. 5 – Phylogram generated from Maximum Likelihood analysis based on combined ITS and *tef1- α* dataset of *Lasiodiplodia*. Bootstrap support values for maximum likelihood (ML), maximum parsimony (MP) greater than 75 % and Bayesian posterior probabilities above 0.90 are indicated near the nodes. The ex-type strains are in bold and the tree is rooted with *Diplodia mutila* (CBS 112553).

Type – Brazil, Itarema, Ceara, trunk canker of *Citrus sinensis* (*Rutaceae*), October of 2013, I. B. L. Coutinho & J. S. Lima (holotype VIC42498, culture ex-type CMM1325).

Host – *Citrus sinensis* (Coutinho et al. 2016).

Distribution – Brazil (Coutinho et al. 2016).

10.5. *Lasiodiplodia citricola* Abdollahz., Javadi & A.J.L. Phillips, *Persoonia* 25: 4 (2010), MycoBank MB16777.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Iran, Gilan Province, Chaboksar, on twigs of *Citrus* sp. (*Rutaceae*), Jun. 2007, J. Abdollahzadeh & A. Javadi (holotype IRAN 14270F, cultures ex-type IRAN 1522C = CBS 124707).

Hosts – *Citrus* sp. (Abdollahzadeh et al. 2010), *Juglans regia* (Chen et al. 2013a, 2014a), *Prunus persica* (Chen et al. 2013b), *Pistacia vera* (Chen et al. 2014b), *Vitis vinifera* (Carlucci et al. 2015).

Distribution – Iran (Abdollahzadeh et al. 2010), Italy (Carlucci et al. 2015), USA (California) (Chen et al. 2013a, b, 2014a, b).

10.6. *Lasiodiplodia crassispora* T.I. Burgess & Barber, *Mycologia* 98: 425 (2006), MycoBank MB500235.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Australia, Western Australia, Kununurra, from canker on *Santalum album* (*Santalaceae*), Dec. 2003, T.I. Burgess (holotype MURU 407, cultures ex-type WAC 12533 = CMW 14691).

Hosts – *Eucalyptus urophylla* (Pérez et al. 2010), *Mangifera indica* (Marques et al. 2013a), *Pterocarpus angolensis* (Mehl et al. 2011), *Santalum album* (Burgess et al. 2006), *Vitis vinifera* (Úrbez-Torres et al. 2010, van Niekerk et al. 2010, Correia et al. 2016a).

Distribution – Australia (Western Australia) (Burgess et al. 2006), Brazil (Marques et al. 2013a, Correia et al. 2016a), South Africa (van Niekerk et al. 2010, Mehl et al. 2011), Uruguay (Pérez et al. 2010), USA (California) (Úrbez-Torres et al. 2010).

10.7. *Lasiodiplodia euphorbiicola* A.R. Machado & O.L. Pereira, *Fungal Diversity* 67: 238 (2014), MycoBank MB804872.

Synonym – *Lasiodiplodia marypalmae* (as *marypalme*) Netto, M.W. Marques, A.J.L. Phillips & M.P.S. Câmara, *Fungal Diversity* 67: 136 (2014), MycoBank MB807526.

Sexual morph not reported. See Machado et al. (2014a) for illustrations and descriptions of asexual morph. Sequences of *L. euphorbicola* were not available to Netto et al. (2014) when they described *L. marypalmae* as a new species. In the phylogenetic analysis of Machado et al. (2014a) the ex-type isolate of *L. marypalmae* nested with the ex-type of *L. euphorbicola* in a well-supported clade and for this reason they considered them to be synonyms.

Type – Brazil, Colatina, Espírito Santo, collar and root rot of *Jatropha curcas* (*Euphorbiaceae*), 2011, A. R. Machado & O. L. Pereira, (holotype VIC39109, culture ex-type CMM3609).

Hosts – *Carica papaya* (Netto et al. 2014), *Jatropha curcas* (Machado et al. 2014a), *Manihot esculenta* (Machado et al. 2014b), *Vitis vinifera* (Correia et al. 2016a).

Distribution – Brazil (Machado et al. 2014a, b, Netto et al. 2014), Italy (Correia et al. 2016a).

10.8. *Lasiodiplodia exigua* Linaldeddu, Deidda & A.J.L. Phillips, *Fungal Diversity* 71: 207 (2015), MycoBank MB808355.

Synonym – *Lasiodiplodia americana* S.F. Chen, G.Q. Li & T.J. Michailides, *Mycologia* 107: 1 (2015), MycoBank MB810934.

Sexual morph not reported. See Linaldeddu et al. (2015) for illustrations and descriptions of asexual morph. Rodríguez-Galvéz et al. (2016) showed that *L. americana* and *L. exigua* are phylogenetically indistinguishable and since *L. exigua* is the older name it takes precedence.

Type – Tunisia, Nabeul, isolated from a branch canker of *Retama raetam* (*Fabaceae*), 27 June 2012, Benedetto T. Linaldeddu (holotype LISE 96302, culture ex-type CBS 137785).

Hosts – *Pistacia vera*, *Retama raetam* (Chen et al. 2015, Linaldeddu et al. 2015).

Distribution – Tunisia (Linaldeddu et al. 2015), USA (Chen et al. 2015).

10.9. *Lasiodiplodia gilanensis* Abdollahz., Javadi & A.J.L. Phillips, *Persoonia* 25: 5 (2010), MycoBank MB16778.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Iran, Gilan Province, Rahimabad-Garmabdost, on twigs of unknown woody plant, Jun. 2007, J. Abdollahzadeh & A. Javadi (holotype IRAN 14272F, cultures ex-type IRAN 1523C = CBS 124704).

Host – *Pistacia vera* (Chen et al. 2014b).

Distribution – Iran (Abdollahzadeh et al. 2010), USA (Chen et al. 2014b).

10.10. *Lasiodiplodia gonubiensis* Pavlic, Slippers & M.J. Wingf., *Stud. Mycol.* 50: 318 (2004), MycoBank MB500079.

Sexual morph and asexual morph have been reported. See Pavlic et al. (2004) for illustrations and descriptions of the asexual morph and Trakunyingcharoen et al. (2015a) for illustrations and descriptions of the sexual morph.

Type – South Africa, Eastern Cape Province, Gonubie, isolated from *Syzygium cordatum* (*Myrtaceae*), Jul. 2002, D. Pavlic (holotype PREM 58127, culture ex-type CBS 115812).

Hosts – *Phyllanthus emblica* (Trakunyingcharoen et al. 2015a), *Syzygium cordatum* (Pavlic et al. 2004, 2007).

Distribution – South Africa (Pavlic et al. 2004, 2007), Thailand (Trakunyingcharoen et al. 2015a).

10.11. *Lasiodiplodia gravistriata* M.S.B. Netto & M.P.S. Camara, *Fungal Biology* (available online, <http://dx.doi.org/10.1016/j.funbio.2016.07.006>, 2016) MycoBank MB816925.

Sexual morph not reported. See Netto et al. (2016) for illustrations and descriptions of asexual morph.

Type – Brazil, Minas Gerais, Coracao de Jesus, on *Anacardium humile* stems (*Anacardiaceae*), 2013, M. S. B. Netto (holotype URM 89942, culture ex-type CMM 4564).

Host – *Anacardium humile* (Netto et al. 2016).

Distribution – Brazil (Netto et al. 2016).

10.12. *Lasiodiplodia hormozganensis* Abdollahz., Zare & A.J.L. Phillips, *Persoonia* 25: 6 (2010), MycoBank MB16779.

Sexual morph not reported. See Abdollahzadeh et al. (2010) and Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Iran, Hormozgan Province, Rodan, on twigs of *Olea* sp. (*Oleaceae*), Jun. 2007, J. Abdollahzadeh & A. Javadi holotype IRAN 14271F, cultures ex-type IRAN 1500C = CBS 124709).

Hosts – *Bougainvillea spectabilis* (Li et al. 2015), *Carica papaya* (Netto et al. 2014), *Mangifera indica* (Abdollahzadeh et al. 2010, Marques et al. 2013a, Al-sadi et al. 2013), *Olea* sp.

(Abdollahzadeh et al. 2010), *Phoenix dactylifera*, *Citrus* (Al-sadi et al. 2013, 2014), *Vitis vinifera* (Correia et al. 2016a).

Distribution – Brazil (Marques et al. 2013a, Netto et al. 2014), China (Li et al. 2015), Iran (Hormozgan Province) (Abdollahzadeh et al. 2010), Italy (Correia et al. 2016a), Oman (Al-sadi et al. 2014), UAE (Al-sadi et al. 2013).

10.13. *Lasiodiplodia iraniensis* Abdollahz., Zare & A.J.L. Phillips, *Persoonia* 25: 8 (2010), MycoBank MB16780.

Synonym – *Lasiodiplodia jatrophiicola* A.R. Machado & O.L. Pereira, *Fungal Diversity* 67: 239 (2014), MycoBank MB804869.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph. Machado et al. (2014a) considered *L. jatrophiicola* as morphologically and phylogenetically distinct from *L. iraniensis*. However, as pointed out by Phillips et al. (2013), morphology is not a suitable character for distinguishing species in *Lasiodiplodia*. Rodríguez-Galvéz et al. (2016) showed that these two species cannot be separated on the basis of ITS and *tefl-α* sequence data.

Type – Iran, Hormozgan Province, Bandar Abbas, Geno mountain, on twigs of *Salvadora persica* (*Salvadoraceae*), Mar. 2007, J. Abdollahzadeh & A. Javadi (holotype IRAN 14268F, cultures ex-type IRAN 1520C = CBS 124710).

Hosts – *Caryota mitis* (Zhu et al. 2015), *Citrus* sp., *Eucalyptus* sp., *Jatropha curcas* (Machado et al. 2014a), *Juglans* sp., *Mangifera indica*, *Salvadora persica*, *Terminalia catapa* (Abdollahzadeh et al. 2010, Marques et al. 2013a, Sakalidis et al. 2011, Al-sadi et al. 2013, Li et al. 2015), *Vitis vinifera* (Correia et al. 2016a).

Distribution – Australia (Sakalidis et al. 2011), Brazil (Machado et al. 2014a, Marques et al. 2013a), China (Li et al. 2015, Zhu et al. 2015), Iran (Hormozgan & Golestan Provinces) (Abdollahzadeh et al. 2010), Italy (Correia et al. 2016a), UAE (Al-sadi et al. 2013).

10.14. *Lasiodiplodia laeliocattleyae* (Sibilia) A. Alves, *Fungal Biology* (available online, <http://dx.doi.org/10.1016/j.funbio.2016.06.004>, 2016), MycoBank MB815697.

Basionym – *Diplodia cattleyae* Sibilia, *Boll. R. Staz. Patolog. Veget. Roma*, N.S. 7: 433 (1927). MycoBank MB268837.

Synonym – *Lasiodiplodia egyptiaca* A.M. Ismail et al., *Australas. Plant Path.* 41: 655 (2012), MycoBank MB564516.

Sexual morph not reported. See Rodríguez-Galvéz et al. (2016) for illustrations and description of the asexual morph. Rodríguez-Galvéz et al. (2016) revealed that *L. laeliocattleyae* is phylogenetically indistinguishable from *L. egyptiaca* and therefore they correspond to the same species.

Type – Italy, from living leaves and pseudobulbs of the cultivated orchid *Laeliocattleya*, 1927, C. Sibilia (holotype location not known, culture ex-type of *Diplodia laeliocattleyae* CBS 167.28).

Hosts – *Laeliocattleya*, *Mangifera indica* (Ismail et al. 2012, Marques et al. 2013), *Cocos nucifera* (Rosado et al. 2016).

Distribution – Brazil (Marques et al. 2013, Rosado et al. 2016), Egypt (Ismail et al. 2012), Italy, Peru (Rodríguez-Galvéz et al. 2016).

10.15. *Lasiodiplodia lignicola* (Ariyaw., J.K. Liu & K.D. Hyde) A.J.L. Phillips, A. Alves & Abdollahz., *Stud. Mycol.* 76: 120 (2013), MycoBank MB805462.

Basionym – *Auerswaldia lignicola* Ariyawansa, J.K. Liu & K.D. Hyde, *Fungal Diversity* 57: 161. 2012.

Sexual morph and asexual morph reported. See Liu et al. (2012) for illustrations and descriptions of sexual morph (as *Auerswaldia lignicola*) and Phillips et al. (2013) for illustrations and descriptions of asexual morph. In Phillips et al. (2013) and other phylogenetic studies, the phylogenetic position

of *L. lignicola* was unclear. Cruywagen et al. (2016) re-sequenced *tefl-α* of the ex-type strain and this sequence (KU887003) was used in the analyses in the present paper, which clearly places this species in *Lasiodiplodia*.

Type – Thailand, Chiang Rai Province, Muang District, Bandu, on dead wood, 30 Sep. 2011, A.D. Ariyawansa (holotype MFLU 12–0750, cultures ex-type MFLUCC 11-0435 = CBS 134112).

Hosts – Dead wood of unknown host (Liu et al. 2012).

Distribution – Thailand (Liu et al. 2012).

10.16. *Lasiodiplodia macrospora* A.R. Machado & O.L. Pereira, *Fungal Diversity* 67: 240 (2014), MycoBank MB804871.

Sexual morph not reported. See Machado et al. (2014a) for illustrations and descriptions of asexual morph.

Type – Brazil, Colatina, Espírito Santo, Collar and root rot of *Jatropha curcas* (*Euphorbiaceae*), 2011, A. R. Machado & O. L. Pereira (holotype VIC39111, culture ex-type CMM3833).

Host – *Jatropha curcas* (Machado et al. 2014a).

Distribution – Brazil (Machado et al. 2014a).

10.17. *Lasiodiplodia mahajangana* Begoude, Jol. Roux & Slippers, *Mycol. Progr.* 9: 110 (2010), MycoBank MB514012.

Sexual morph not reported. See Didier Begoude et al. (2010) for illustrations and descriptions of asexual morph.

Type – Madagascar, Mahajanga, isolated from healthy branches of *Terminalia catappa* (*Combretaceae*), Oct. 2007, J. Roux (holotype PREM 60288 holotype, culture ex-type CBS 124925).

Hosts – *Euphorbia ingens* (van der Linde et al. 2011), *Terminalia catappa* (Didier Begoude et al. 2010).

Distribution – Madagascar (Didier Begoude et al. 2010), South Africa (van der Linde et al. 2011).

10.18. *Lasiodiplodia margaritacea* Pavlic, T.I. Burgess & M.J. Wingf., *Mycologia* 100: 860 (2008), MycoBank MB512052.

Sexual morph not reported. See Pavlic et al. (2008) for illustrations and descriptions of asexual morph.

Type – Australia, Western Australia, Tunnel Creek Gorge, on *Adansonia gibbosa* (*Malvaceae*), Jul. 2006, T.I. Burgess (holotype PREM 59844, culture ex-type CBS 122519).

Host – *Adansonia gibbosa* (Pavlic et al. 2008).

Distribution – Australia (Pavlic et al. 2008).

10.19. *Lasiodiplodia mediterranea* Linaldeddu, Deidda & Berraf-Tebbal, *Fungal Diversity* 71: 207 (2015), MycoBank MB808356.

Sexual morph not reported. See Linaldeddu et al. (2015) for illustrations and descriptions of asexual morph.

Type – Italy, Bortigiadas, isolated from a branch canker of *Quercus ilex* (*Fagaceae*), June 2004, Benedetto T. Linaldeddu (holotype LISE 96303, culture ex-type CBS 137783).

Hosts – *Citrus sinensis*, *Quercus ilex*, *Vitis vinifera* (Linaldeddu et al. 2015, Andolfi et al. 2016).

Distribution – Algeria, Italy (Linaldeddu et al. 2015, Andolfi et al. 2016).

10.20. *Lasiodiplodia missouriana* Urb.-Torr., Peduto & Gubler, *Fungal Diversity* 55: 181 (2012), MycoBank MB519954.

Sexual morph not reported. See Úrbez-Torres et al. (2012) for illustrations and descriptions of asexual morph.

Type – USA, Saint James, on *Vitis vinifera* × *V. labrusca* hybrid cv. Catawba (*Vitaceae*), Jun. 2006, R.K. Striegler & G.M. Leavitt (holotype UCD2193MO, culture ex-type CBS 128311).

Host – *Vitis* sp. (Úrbez-Torres et al. 2012).

Distribution – USA (Úrbez-Torres et al. 2012).

10.21. *Lasiodiplodia parva* A.J.L. Phillips, A. Alves & Crous, *Fungal Diversity* 28: 9 (2008), MycoBank MB510942.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Colombia, Dep. Meta, Villavicencio, cassava field soil, 1978, O. Rangel (holotype CBS H-19915, culture ex-type CBS 456.78).

Hosts – Cassava-field soil, *Terminalia* sp. (Begoude et al. 2011), *Theobroma cacao* (Alves et al. 2008), *Vitis vinifera* (Correia et al. 2013).

Distribution – Colombia, Sri Lanka (Alves et al. 2008), Cameroon (Begoude et al. 2011), Brazil (Correia et al. 2013).

10.22. *Lasiodiplodia plurivora* Damm & Crous, *Mycologia* 99: 674 (2008), MycoBank MB501322.

Sexual morph not reported. See Damm et al. (2007) for illustrations and descriptions of asexual morph.

Type – South Africa, Western Cape Province, Stellenbosch, from V-shaped necrotic lesion of *Prunus salicina* (*Rosaceae*), May 2004, U. Damm (holotype CBS H-19844, culture ex-type CBS 120832).

Hosts – *Prunus salicina*, *Vitis vinifera* (Damm et al. 2007).

Distribution – South Africa (Western Cape Province) (Damm et al. 2007).

10.23. *Lasiodiplodia pontae* F.C.O. Freire, I.B.L. Coutinho, C.S. Lima & J.E. Cardoso, *Plant Pathology* (available online, Doi: 10.1111/ppa.12565, 2016), MycoBank MB815121.

Sexual morph not reported. See Coutinho et al. (2016) for illustrations and descriptions of asexual morph.

Type – Brazil, Pío IX, Piauí, necrotic canker on *Spondias purpurea* (*Anacardiaceae*), October 2009, J. S. Lima & F. C. O. Freire (holotype VIC42947, culture ex-type CMM1277).

Host – *Spondias purpurea* (Coutinho et al. 2016).

Distribution – Brazil (Coutinho et al. 2016).

10.24. *Lasiodiplodia pseudotheobromae* A.J.L. Phillips, A. Alves & Crous, *Fungal Diversity* 28: 8 (2008), MycoBank MB510941.

Sexual morph reported by Tennakoon et al. (2016). Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Costa Rica, San Carlos, on *Gmelina arborea* (*Lamiaceae*), J. Carranza-Velazquez (holotype CBS H-19916, culture ex-type CBS 116459).

Hosts – Recent studies subsequent to Phillips et al. (2013) confirm the following hosts: *Acacia mangium*, *Citrus aurantium*, *Coffea* sp., *Gmelina arborea*, *Rosa* sp. (Alves et al. 2008, Castro-

Medina et al. 2014), *Bougainvillea spectabilis* (Li et al. 2015), *Carica papaya* (Netto et al. 2014), *Cocos nucifera* (Rosado et al. 2016), *Eucalyptus grandis* (Pillay et al. 2013), *Jatropha curcas* (Machado et al. 2014a), *Juglans regia* China (Li et al. 2016b), *Mangifera indica* (Sandoval-Sanchez et al. 2013, Marques et al. 2013a), *Manihot esculenta* Brazil (Machado et al. 2014b), *Persea americana*, *Hevea brasiliensis* (Trakunyingcharoen et al. 2015b), *Schizolobium parahyba* (Mehl et al. 2014), *Tectona grandis* (Doilom et al. 2015), *Vitis vinifera* (Correia et al. 2013, Correia et al. 2016a, Dissanayake et al. 2015a).

Distribution – Brazil (Marques et al. 2013a, Machado et al. 2014a, b, Correia et al. 2013, Netto et al. 2014, Rosado et al. 2016), China (Dissanayake et al. 2015a, Li et al. 2015, 2016), Costa Rica, Netherlands, Suriname, Zaire (Alves et al. 2008), Ecuador (Mehl et al. 2014), Italy (Correia et al. 2016a), Mexico (Sandoval-Sanchez et al. 2013), South Africa (Pillay et al. 2013), Thailand (Doilom et al. 2015, Trakunyingcharoen et al. 2015b), Venezuela (Castro-Medina et al. 2014).

10.25. *Lasiodiplodia pyriformis* F.J.J. Van der Walt, Slippers & G.J. Marais, *Persoonia* 33: 163 (2014), MycoBank MB518722.

Sexual morph not reported. See Slippers et al. (2014) for illustrations and descriptions of asexual morph.

Type – Namibia, Dordabis, from *Acacia mellifera* (*Fabaceae*), Feb. 2006, F.J.J. van der Walt & J. Roux (holotype PREM 59633, culture ex-type CBS 121770).

Host – *Acacia mellifera* (Slippers et al. 2014).

Distribution – Namibia (Slippers et al. 2014).

10.26. *Lasiodiplodia rubropurpurea* T.I. Burgess, Barber & Pegg, *Mycologia* 98: 431 (2006), MycoBank MB500236.

Sexual morph not reported. See Burgess et al. 2006) for illustrations and descriptions of asexual morph.

Type – Australia, Queensland, Tully, from canker on *Eucalyptus grandis* (*Myrtaceae*), May 2003, T.I. Burgess (holotype MURU 409, culture ex-type CBS 118740).

Host – *Eucalyptus grandis* (Burgess et al. 2006).

Distribution – Australia (Queensland) (Burgess et al. 2006).

10.27. *Lasiodiplodia subglobosa* A.R. Machado & O.L. Pereira, *Fungal Diversity* 67: 241 (2014), MycoBank MB804870.

Sexual morph not reported. See Machado et al. (2014a) for illustrations and descriptions of asexual morph.

Type – Brazil, Jaíba, Minas Gerais, Collar and root rot of *Jatropha curcas* (*Euphorbiaceae*), 2011, A. R. Machado & O. L. Pereira (holotype VIC39112, culture ex-type CMM3872).

Hosts – *Carya illinoensis* (Poletto et al. 2016), *Jatropha curcas* (Machado et al. 2014a).

Distribution – Brazil (Machado et al. 2014a, Poletto et al. 2016).

10.28. *Lasiodiplodia thailandica* T. Trakunyingcharoen, L. Lombard & Crous, *Persoonia* 34: 95 (2014), MycoBank MB810169.

Sexual morph not reported. See Trakunyingcharoen et al. (2015b) for illustrations and descriptions of asexual morph.

Type – Thailand, Chiang Mai province, on twigs of *Mangifera indica* (*Anacardiaceae*), May 2012, T. Trakunyingcharoen (holotype CBS-H 21933, culture ex-type CBS 138760).

Host – *Mangifera indica* (Trakunyingcharoen et al. 2015b).

Distribution – Thailand (Trakunyingcharoen et al. 2015b).

10.29. *Lasiodiplodia theobromae* (Pat.) Griffon & Maubl., *Bull. Soc. mycol. Fr.* 25: 57 (1909), MycoBank MB188476.

Sexual and asexual morph reported but the connection between the two morphs has not been proven conclusively (Phillips et al. 2013). See Phillips et al. (2013) for illustrations and descriptions.

Type – Ecuador, on *Theobroma cacao*, Lagerheim, holotype not found, and presumably lost. Papua New Guinea, Madang, Jais Aben, from unidentified fruit along coral reef coast, No. 1995, A. Aptroot (neotype CBS H-21411, culture ex-neotype CBS 164.96).

Hosts – Wide host range.

Distribution – Widely distributed in tropical and subtropical regions.

10.30. *Lasiodiplodia venezuelensis* T.I. Burgess, Barber & Mohali, *Mycologia* 98: 432 (2006), MycoBank MB500237.

Sexual morph not reported. See Burgess et al. (2006) for illustrations and descriptions of asexual morph.

Type – Venezuela, Estado Portuguesa, Acarigua, from wood of living *Acacia mangium* (*Fabaceae*), Oct. 2003, S. Mohali (holotype MURU 413, culture ex-type CBS 118739).

Host – *Acacia mangium* (Burgess et al. 2006).

Distribution – Venezuela (Burgess et al. 2006).

10.31. *Lasiodiplodia viticola* Urb.-Torr., Peduto & Gubler, *Fungal Diversity* 55: 183 (2012), MycoBank MB519966.

Sexual morph not reported. See Úrbez-Torres et al. (2010) for illustrations and descriptions of asexual morph.

Type – USA, Arkansas, Altus, on interspecific hybrid grape Vignoles cv. Ravat 51R, D. Cartwright & W. D. Gubler (holotype UCD2553AR, cultures ex-type UCD2553AR = CBS 128313).

Host – *Vitis* hybrids (Úrbez-Torres et al. 2010).

Distribution – USA (Arkansas and Missouri) (Úrbez-Torres et al. 2010).

11. *Macrophomina* Petr., *Annls mycol.* 21: 314 (1923), MycoBank MB8814.

This genus comprises only one species. Sexual morph not reported.

Type species – *Macrophomina phaseolina* (Tassi) Goid., *Annali Sper. agr.* N.S. 1: 457. 1947.

11.1. *Macrophomina phaseolina* (Tassi) Goid., *Annali Sper. agr.* N.S. 1: 457 (1947), MycoBank MB300023.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Holotype apparently lost. Cultures: Niger, *Vigna minima*, M. Ndiaye, CPC 11052, 11070. Senegal, soil, M. Ndiaye, CPC 11079, 11085, 11106, 11108. Uganda, *Eucalyptus* sp. (*Myrtaceae*), Jan. 1925, CBS 162.25; Unknown, *Zea mays*, Jun. 1933, S.F. Ashby, CBS 227.33.

Host – Plurivorous.

Distribution – Cosmopolitan.

12. *Marasasiomyces* Crous, *Phytotaxa* 202: 86 (2015), MycoBank MB811252.

This genus was introduced to accommodate tiarosporella-like fungi and comprises only one species. Sexual morph not reported.

Type species – *Marasasiomyces karoo* (B. Sutton & Marasas) Crous 2015.

12.1. *Marasasiomyces karoo* (B. Sutton & Marasas) Crous, *Phytotaxa* 202: 86 (2015), MycoBank MB811253.

Sexual morph not reported. See Sutton & Marasas (1976) and Crous et al. 2006 for illustrations and descriptions of asexual morph.

Type – South Africa. Cape Province: Colesberg, on dead stems of *Erioccephalus* sp. (*Asteraceae*), Feb. 1971, W.F.O. Marasas (holotype PREM 44967, isotype IMI 186782, culture ex-type CBS 118718).

Hosts – *Erioccephalus* sp., *Nestlera* sp., *Tribulus terrestris* (Sutton & Marasas 1976).

Distribution – South Africa (Sutton & Marasas 1976).

13. *Mucoharknessia* Crous, R.M. Sánchez & Bianchin., *Phytotaxa* 202: 86 (2015), MycoBank MB811254.

This genus was introduced for species resembling *Harknessia* with conidia bearing mucoid appendages. *Mucoharknessia* comprises two species. Only the asexual morph has been reported within the genus.

Type species – *Mucoharknessia cortaderiae* Crous, R.M. Sánchez & Bianchin 2015.

13.1. *Mucoharknessia cortaderiae* Crous, R.M. Sánchez & Bianchin., *Phytotaxa* 202: 86 (2015), MycoBank MB811255.

Sexual morph not reported. See Crous et al. (2015a) for illustrations and descriptions of asexual morph.

Type – Argentina. Buenos Aires Province, Punta Alta, 38°47'27,6"S 62°6'48,6"W, on leaves of *Cortaderia selloana* (*Poaceae*), 29 Mar. 2011, F.E. Anderson (holotype BBB, isotype CBS H-21853, culture ex-isotype CBS 131032).

Host – *Cortaderia selloana* (Crous et al. 2015a).

Distribution – Argentina (Crous et al. 2015a).

13.2. *Mucoharknessia anthoxanthii* Dissanayake, Camporesi & K.D. Hyde, *Fungal Diversity* 78: 19 (2016), Facesoffungi number FoF01651.

Sexual morph not reported. See Li et al. (2016a) for illustrations and descriptions of asexual morph.

Type – Italy, Province of Forlì-Cesena [FC], Passo delle Forche - Galeata, on dead stem of *Anthoxanthum odoratum* (*Poaceae*), 24 November 2012, Erio Camporesi (holotype MFLU 15–3477, culture ex-type MFLUCC 15–0904).

Host – *Anthoxanthum odoratum* (Li et al. 2016a).

Distribution – Italy (Li et al. 2016a).

14. *Neodeightonia* C. Booth, *Mycol. Pap.* 119: 17 (1970), MycoBank MB3450.

This genus comprises six species. Both sexual and the asexual morphs have been reported within the genus.

Type species – *Neodeightonia subglobosa* Booth, in Punithalingam 1970.

14.1. *Neodeightonia licuriensis* A. R. Machado & O. L. Pereira, *Cryptog. Mycol.* 36: 121 (2015), MycoBank MB810662.

Sexual morph not reported. See Adamcik et al. (2015) for illustrations and descriptions of asexual morph.

Type – Brazil, along highway between the cities Castro Alves and Santa Terezinha, Bahia, on necrotic lesions on the rachis and leaves of *Syagrus coronata* (*Arecaceae*), 2010, O. L. Pereira, (holotype VIC42826, culture ex-type COAD1780).

Host – *Syagrus coronate* (Adamcik et al. 2015).

Distribution – Brazil (Adamcik et al. 2015).

14.2. *Neodeightonia palmicola* J.K. Liu, Phook. & K.D. Hyde, *Sydowia* 62: 268 (2010), MycoBank MB518804.

Sexual morph and asexual morph reported. See Liu et al. (2012) for illustrations and descriptions.

Type – Thailand, Chiang Rai, Muang District, Khun Korn Waterfall, on dead leaves of *Arenga westerhoutii* (*Arecaceae*), 18 Dec. 2009, Jian-Kui Liu (holotype MFLU10 0407, culture ex-type MFLUCC10 0822 = CBS 136074).

Host – *Arenga westerhoutii* (Liu et al. 2012).

Distribution – Thailand (Liu et al. 2012).

14.3. *Neodeightonia phoenicum* A.J.L. Phillips & Crous, *Persoonia* 21: 43 (2008), MycoBank MB511708.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Spain, Catalonia, Tarragona, Salou, on *Phoenix* sp. (*Arecaceae*), F. Garcia (holotype CBS H-20108, culture ex-type CBS 122528).

Host – *Phoenix* sp. (Phillips et al. 2008, Ligoixigakis et al. 2013).

Distribution – Spain, USA (California) (Phillips et al. 2008), Greece (Ligoixigakis et al. 2013).

14.4. *Neodeightonia rattanica* Konta & K.D. Hyde, *Mycosphere* 7: 953 (2016), Facesoffunginumber FoF02237.

Sexual morph and asexual morph reported. See Konta et al. (2016b) for illustrations and descriptions.

Type – Thailand, Phang-Nga, on dead rachis of *Calamus* sp. (*Arecaceae*), 6 December 2014, S. Konta (holotype MFLU 15-1443, culture ex-type MFLUCC 15-0712).

Host – *Calamus* sp. (Konta et al. 2016b).

Distribution – Thailand (Konta et al. 2016b).

14.5. *Neodeightonia rattanicola* Konta & K.D. Hyde, *Mycosphere* 7: 954 (2016), Facesoffunginumber FoF02238.

Sexual morph and asexual morph reported. See Konta et al. (2016b) for illustrations and descriptions.

Type – Thailand, Phang-Nga, on dead rachis of *Calamus* sp. (*Arecaceae*), 6 December 2014, S. Konta (holotype MFLU 15-0294, culture ex-type MFLUCC 15-0319).

Host – *Calamus* sp. (Konta et al. 2016b).

Distribution – Thailand (Konta et al. 2016b).

14.6. *Neodeightonia subglobosa* C. Booth, *Mycol. Pap.* 119: 19 (1970), MycoBank MB318601.

Sexual morph and asexual morph reported. See Phillips et al. (2013) for illustrations and descriptions.

Type – Sierra Leone, Njala (Kori), on dead culms of *Bambusa arundinacea* (*Poaceae*), 17 Aug. 1954, F.C. Deighton (holotype IMI 57769(f), culture ex-type CBS 448.91).

Host – *Bambusa arundinacea* (Punithalingam 1970).

Distribution – Sierra Leone (Punithalingam 1970).

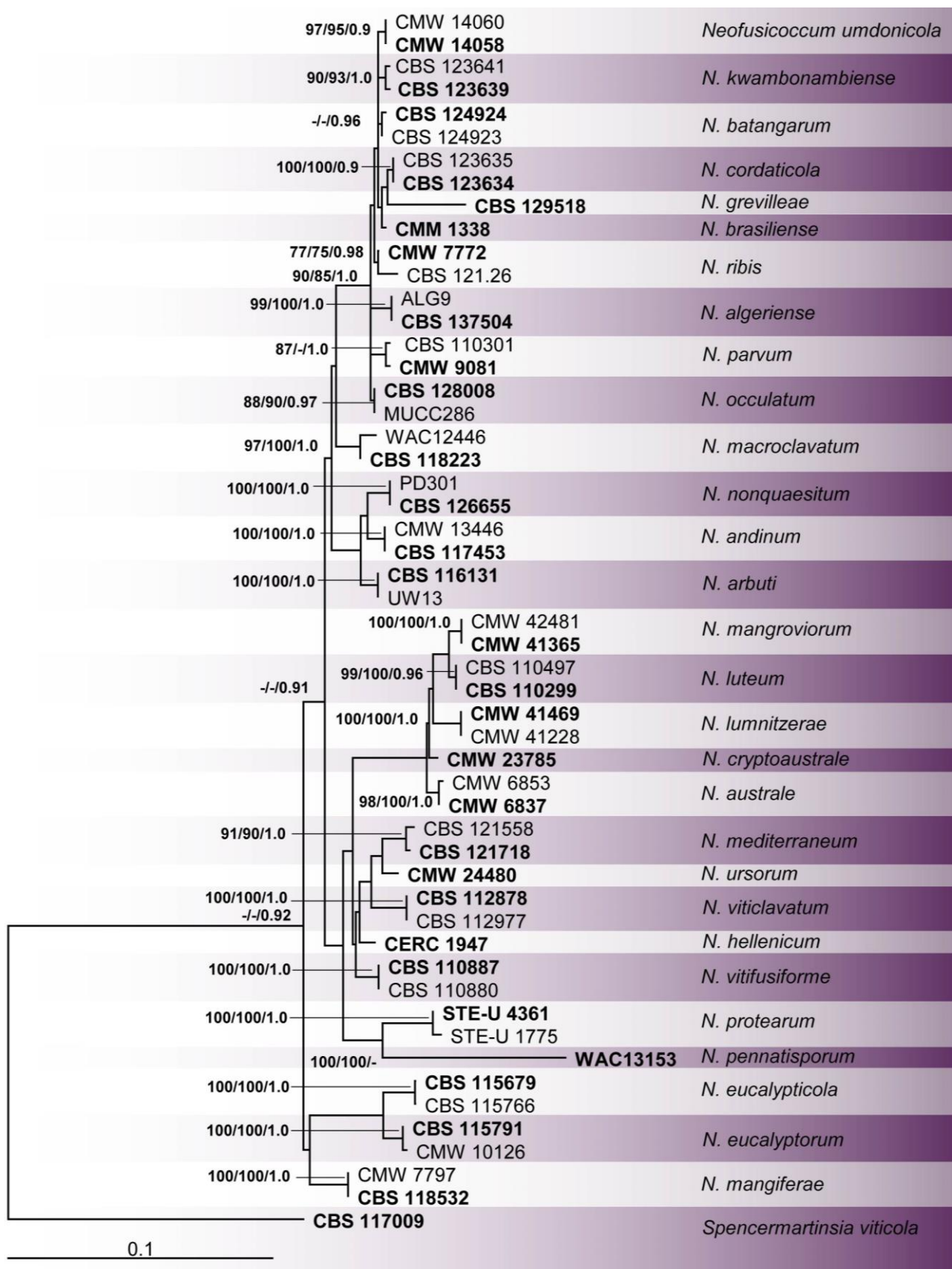


Fig. 6 – Phylogram generated from Maximum Likelihood analysis based on combined ITS and *tef1- α* dataset of *Neofusicoccum*. Bootstrap support values for maximum likelihood (ML), maximum parsimony (MP) greater than 75 % and Bayesian posterior probabilities greater than 0.90 are indicated near the nodes. The ex-type strains are in bold and the tree is rooted with *Spencermartinsia viticola* (CBS 117009).

15. *Neofusicoccum* Crous, Slippers & A.J.L. Phillips, *Stud. Mycol.* 55: 247 (2006), MycoBank MB500870.

This genus comprises 29 species (Fig. 6). Both sexual and asexual morphs have been reported within the genus.

Type species – *Neofusicoccum parvum* (Pennycook & Samuels) Crous, Slippers & A.J.L. Phillips, *Stud. Mycol.* 55: 248. 2006.

15.1. *Neofusicoccum algeriense* Berraf-Tebbal & A.J.L. Phillips, *Fungal Diversity* 53: 423 (2014), MycoBank MB808496.

Sexual morph not reported. See Berraf-Tebbal et al. (2014) for illustrations and descriptions of asexual morph. Lopes et al. (2016) consider this species as a synonym of *N. parvum*.

Type – Algeria. Ain-Benian, on trunks and branches of *Vitis vinifera* (*Vitaceae*), May 2013, A. Berraf-Tebbal (holotype LISE 9630, culture ex-type CBS 137504).

Host – *Vitis vinifera* (Berraf-Tebbal et al. 2014).

Distribution – Algeria (Berraf-Tebbal et al. 2014).

15.2. *Neofusicoccum andinum* (Mohali, Slippers & M.J. Wingf.) Mohali, Slippers & M.J. Wingf., *Stud. Mycol.* 55: 247 (2006), MycoBank MB500871.

Sexual morph not reported. See Mohali et al. (2006) for illustrations and descriptions of asexual morph.

Type – Venezuela, Mérida Morph, Merida, Mucuchies (3140 m), Cordillera of Los Andes, on branches of *Eucalyptus* sp. (*Myrtaceae*), Feb. 2003, S. Mohali (holotype PREM 58238, culture ex-type CBS 117453).

Host – *Eucalyptus* sp. (Mohali et al. 2006).

Distribution – Venezuela (Mohali et al. 2006).

15.3. *Neofusicoccum arbuti* (D.F. Farr & M. Elliott) Crous, Slippers & A.J.L. Phillips, *Stud. Mycol.* 55: 248 (2006), MycoBank MB500872.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – USA, Washington, King Co., Seattle, Magnolia Bluffs, isolated from cankers of *Arbutus menziesii* (*Ericaceae*), Oct. 2003, collected by M. Elliott, isolated by A. Rossman (holotype BPI 843970, culture ex-type CBS 116131).

Hosts – *Arbutus menziesii* (Pacific madrone) (Farr et al. 2005, Mcgregor et al. 2016), *Vaccinium* sp. (Espinoza et al. 2009).

Distribution – USA, Canada (Farr et al. 2005), Chile (Espinoza et al. 2009), North America (Mcgregor et al. 2016).

15.4. *Neofusicoccum australe* (Slippers, Crous & M.J. Wingf.) Crous, Slippers & A.J.L. Phillips, *Stud. Mycol.* 55: 248 (2006), MycoBank MB500873.

Sexual morph and asexual morph reported. See Phillips et al. (2013) for illustrations and descriptions.

Type – Australia, Victoria, Batemans Bay, *Acacia* sp. (*Fabaceae*), M.J. Wingfield (holotype PREM 57589, culture ex-type CMW 6837).

Hosts – *Acacia karroo* (Jami et al. 2015), *Eucalyptus grandis*, *Syzygium cordatum* (Pillay et al. 2013), *Mangifera indica* (Ismail et al. 2013), *Olea europaea* (Triki et al. 2015), *Pistacia vera* (Chen et al. 2014b), *Quercus robur* (Barradas et al. 2013), *Vaccinium* (Castillo et al. 2013), *Vitis vinifera* (Berraf-Tebbal et al. 2014, Pitt et al. 2013a, Valencia et al. 2015, Billones-Baaijens et al. 2015).

Distribution – Algeria (Berraf-Tebbal et al. 2014), Australia (Pitt et al. 2013a), Chile (Valencia et al. 2015), Italy (Ismail et al. 2013, New Zealand (Billones-Baaijens et al. 2015), Portugal (Barradas et al. 2013), South Africa (Jami et al. 2015), South Africa (Pillay et al. 2013), Spain (Castillo et al. 2013), Tunisia (Triki et al. 2015), USA (Chen et al. 2014b).

15.5. *Neofusicoccum batangarum* Begoude, Jol. Roux & Slippers, *Stud. Mycol.* 76: 137 (2013), MycoBank MB514013.

Sexual morph not reported. See Didier Begoude et al. (2010) for illustrations and descriptions of asexual morph.

Type – Cameroon, Kribi, Beach, isolated from healthy branches of *Terminalia catappa* (*Combretaceae*), Dec. 2007, D. Begoude & J. Roux, a dry culture on pine needles (holotype PREM 60285, culture ex-type CBS 124924).

Hosts – Prickly Pear Cactus (Conforto et al. 2016), *Schinus terebinthifolius* (Shetty et al. 2011), *Terminalia catappa* (Didier Begoude et al. 2010).

Distribution – Brazil (Conforto et al. 2016), Cameroon (Didier Begoude et al. 2010), USA (Shetty et al. 2011).

15.6. *Neofusicoccum brasiliense* M.W. Marques, A.J.L. Phillips & M.P.S. Câmara, sp. nov., MycoBank MB819515.

“*Neofusicoccum brasiliense*” M.W. Marques, A.J.L. Phillips & M.P.S. Câmara, *Fungal Diversity* 61: 201 (2013), nom. inval.

Sexual morph not reported. Full description and diagnosis in Marques et al. (2013b).

Marques et al. (2013b) introduced *Neofusicoccum brasiliense* but did not designate a holotype and thus the name was invalid. To validate the name a suitable specimen has been deposited as follows.

Type – Brazil, Pernambuco, Petrolina, Lote 1195, on *Mangifera indica* stems (*Anacardiaceae*), 2010, M.W. Marques (holotype URM 89948, culture ex-type CMM1338).

Host – *Mangifera indica* (Marques et al. 2013b).

Distribution – Brazil (Marques et al. 2013b).

15.7. *Neofusicoccum cordaticola* Pavlic, Slippers & M.J. Wingf., *Mycologia* 100: 643 (2008), MycoBank MB512498.

Sexual morph not reported. See Pavlic et al. (2009) for illustrations and descriptions of asexual morph.

Type – South Africa, Kwazulu-Natal Province, Sodwana Bay, on symptomless branches and leaves, dying branches and pulp of ripe fruits of *Syzygium cordatum* (*Myrtaceae*), Mar. 2002, D. Pavlic, (holotype PREM 60066, culture ex-type CBS 123634).

Host – *Syzygium cordatum* (Pavlic et al. 2009).

Distribution – South Africa (Pavlic et al. 2009).

15.8. *Neofusicoccum cryptoaustrale* Pavlic, Maleme, Slippers & M.J. Wingf., *Persoonia* 31: 271 (2013), MycoBank MB512477.

Sexual morph not reported. See Crous et al. (2013) for illustrations and descriptions of asexual morph.

Type – South Africa, Gauteng Province, Pretoria, from branches and leaves of living Eucalyptus trees (*Myrtaceae*), May 2005, H.M. Maleme (holotype PREM 59817, culture ex-type CBS 1122813); Gauteng Province, Eucalyptus trees, H.M. Maleme (paratype PREM 59818, culture ex-paratype CMW 20738, MycoBank MB512477).

Host – *Eucalyptus* sp. (Crous et al. 2013).

Distribution – South Africa (Crous et al. 2013).

15.9. *Neofusicoccum eucalypticola* (Slippers, Crous & M.J. Wingf.) Crous, Slippers & A.J.L. Phillips, *Stud. Mycol.* 55: 248, MycoBank MB500874.

Sexual morph and asexual morph reported. See Slippers et al. (2004b) for illustrations and descriptions.

Type – Australia, Victoria, Orbost, on *Eucalyptus grandis* (*Myrtaceae*), 2001, M.J. Wingfield (holotype PREM 57848, culture ex-type CBS 115679).

Host – *Eucalyptus* sp. (Slippers et al. 2004b, Burgess et al. 2006).

Distribution – Australia (Slippers et al. 2004b).

15.10. *Neofusicoccum eucalyptorum* (Crous, H. Sm. ter. & M.J. Wingf.) Crous, Slippers & A.J.L. Phillips, *Stud. Mycol.* 55: 248. 2006. MycoBank MB500875.

Sexual morph and asexual morph reported. See Smith et al. (2001) for illustrations and descriptions.

Type – of sexual morph: South Africa, Mpumalanga, Sabie, *Eucalyptus grandis*, 1995, H. Smith, holotype PREM 56603; of asexual morph: South Africa, Mpumalanga, Sabie, *E. grandis*, 1995, H. Smith, holotype PREM 56604. The ex-type isolate was not designated in the original publication and could not be traced. Slippers et al. (2004b) regarded CBS 115791 as representative.

Hosts – *Blepharocalyx salicifolius* (Pérez et al. 2010), *Eucalyptus* spp. (Burgess et al. 2006, Smith et al. 2001, Slippers et al. 2004b, Pérez et al. 2010, De la Mora-Castaneda et al. 2014), *Myrceugenia glaucescens*, *Myrrhimum atropurpureum* var. *octandrum*, *Syzygium cordatum* (Pillay et al. 2013).

Distribution – Australia (Slippers et al. 2004b), Mexico (De la Mora-Castaneda et al. 2014), South Africa (Smith et al. 2001, Pillay et al. 2013), Uruguay (Pérez et al. 2010).

15.11. *Neofusicoccum grevilleae* Crous & R.G. Shivas, *Persoonia* 26: 117 (2011), MycoBank MB560162.

Sexual morph not reported. See Crous et al. (2011) for illustrations and descriptions of asexual morph.

Type – Australia, Queensland, Brisbane, on leaves of *Grevillea aurea* (*Proteaceae*), 14 Jul. 2009, P.W. Crous & R.G. Shivas (holotype CBS H-20578, culture ex-type CBS 129518).

Host – *Grevillea aurea* (Crous et al. 2011).

Distribution – Australia (Crous et al. 2011).

15.12. *Neofusicoccum hellenicum* S.F. Chen, G.Q. Li & T.J. Michailides, *Mycologia* 107: 1 (2015), MycoBank MB810935.

Sexual morph not reported. See Crous et al. (2011) for illustrations and descriptions of asexual morph.

Type – Greece, Thessaloniki, from twigs of one *Pistacia vera* cultivar Aegina (*Anacardiaceae*), 23 Oct 1997, T.J. Michailides (holotype CSFF2010, culture ex-type CERC1947 = CFCC50067).

Host – *Pistacia vera* (Crous et al. 2011).

Distribution – Greece (Crous et al. 2011).

15.13. *Neofusicoccum kwambonambiense* Pavlic, Slippers & M.J. Wingf., *Mycologia* 100: 643 (2008), MycoBank MB512499.

Sexual morph not reported. See Pavlic et al. (2009) for illustrations and descriptions of asexual morph.

Type – South Africa, Kwazulu-Natal Province, Kwambonambi, on symptomless branches and leaves, dying branches and pulp of ripe fruits of *Syzygium cordatum* (*Myrtaceae*), Mar 2002, D. Pavlic (holotype PREM 60067, culture ex-type CBS 123639).

Hosts – *Acacia karroo*, *Celtis Africana* (Jami et al. 2014), *Eucalyptus grandis* (Pillay et al. 2013), *Fragaria ananassa* (Lopes et al. 2014), *Syzygium cordatum* (Pavlic et al. 2009, Pillay et al. 2013, Pavlic-Zupanc et al. 2015), *Vitis vinifera* (Abreo et al. 2013).

Distribution – Brazil (Lopes et al. 2014), South Africa (Pavlic et al. 2009, Pillay et al. 2013, Jami et al. 2014, Pavlic-Zupanc et al. 2015), Uruguay (Abreo et al. 2013).

15.14. *Neofusicoccum lumnitzeriae* J.A. Osorio, Jol. Roux & Z.W. de Beer, *Fungal Biology* (available online, <http://dx.doi.org/10.1016/j.funbio.2016.09.004>, 2016), MycoBank MB812012.

Sexual morph not reported. See Osorio et al. (2016) for illustrations and description.

Type – South Africa, Kwazulu-Natal Province: Kosi Bay, from asymptomatic branches of *Lumnitzera racemosa* (*Combretaceae*) J.A Osorio & Jol. Roux. (holotype PREM 61251, culture ex-type CBS 139674).

Host – *Lumnitzera racemosa* (Osorio et al. 2016).

Distribution – South Africa (Osorio et al. 2016).

15.15. *Neofusicoccum luteum* (Pennycook & Samuels) Crous, Slippers & A.J.L. Phillips, *Stud. Mycol.* 55: 248 (2006), MycoBank MB500876.

Sexual morph and asexual morph reported. See Phillips et al. (2013) for illustrations and descriptions.

Type – New Zealand, Bay of Plenty, Te Puke, No 1 Road, DSIR Research Orchard, from lesions on ripe fruit of *Actinidia deliciosa* (*Actinidiaceae*), 6 Oct. 1982, S.R. Pennycook (holotype PDD 45400, culture ex-type of asexual morph PDDCC 8004). Portugal, Estremadura, Oeiras, Quinta do Marquês, on cane of *Vitis vinifera* cv. Galego Dourado (*Vitaceae*), Mar. 1996, A.J.L. Phillips (holotype of sexual morph LISE 94070, culture ex-type CBS 110299).

Hosts – Recent studies subsequent to Phillips et al. (2013) confirm the following hosts: *Citrus* sp. (Adesemoye et al. 2014), *Erica arborea* (Linaldeddu et al. 2015), *Vitis vinifera* (Billones-Baaijens et al. 2015, Chebil et al. 2014).

Distribution – Recent studies subsequent to Phillips et al. (2013) confirm the following distribution: California (Adesemoye et al. 2014), Italy (Linaldeddu et al. 2015), New Zealand (Billones-Baaijens et al. 2015), Tunisia (Chebil et al. 2014).

15.16. *Neofusicoccum macroclavatum* (T.I. Burgess, Barber & G.E. Hardy) T.I. Burgess, Barber & G.E. Hardy, *Stud. Mycol.* 55: 248 (2006), MycoBank MB500877.

Sexual morph not reported. See Burgess et al. (2005) for illustrations and descriptions of asexual morph.

Type – Australia, Western Australia, Denmark, from wood of living *Eucalyptus globulus* (*Myrtaceae*), Oct. 2002, T.I. Burgess (holotype MURU 400, culture ex-type CBS 118223).

Hosts – *Eucalyptus globulus*, *E. saligna* (Burgess et al. 2005), *Vitis vinifera* (Billones-Baaijens et al. 2013).

Distribution – Australia (Burgess et al. 2005), New Zealand (Billones-Baaijens et al. 2013).

15.17. *Neofusicoccum mangiferae* (Syd. & P. Syd.) Crous, Slippers & A.J.L. Phillips, *Stud. Mycol.* 55: 248 (2006), MycoBank MB500878.

Sexual morph not reported. See Slippers et al. (2005) for illustrations and descriptions of asexual morph.

Type – India, Lucknow, on *Mangifera indica* (*Anacardiaceae*), F. Bahadur (E.J. Butler 1724), 22 Oct. 1908 (holotype HCIO Cultures linked to the type could not be located and probably do not exist. Slippers et al. (2005) regarded the following as representatives: CBS 118531, CBS 118532.

Hosts – *Mangifera indica* (Slippers et al. 2005, Ni et al. 2012, Serrato-Diaz et al. 2014), *Vitis vinifera* (Dissanayake et al. 2015b).

Distribution – Australia, India (Slippers et al. 2005), China (Dissanayake et al. 2015b), Puerto Rico (Serrato-Diaz et al. 2014), Taiwan (Ni et al. 2012).

15.18. *Neofusicoccum mangroviorum* J.A. Osorio, Jol. Roux & Z.W. de Beer, *Fungal Biology* (available online, <http://dx.doi.org/10.1016/j.funbio.2016.09.004>, 2016), MycoBank MB814641.

Sexual morph not reported. See Osorio et al. (2016) for illustrations and description.

Type – South Africa, Kwazulu-Natal Province, Beachwood, Kosi Bay, Mtunzini, Mgazana, from asymptomatic branches of *Avicennia marina*, J.A Osorio & Jol. Roux (holotype PREM 61305, culture ex-type CBS 140738).

Host – *Avicennia marina*, *Bruguiera gymnorrhiza*, *Lumnitzera racemosa* and *Rhizophora mucronata* (Osorio et al. 2016).

Distribution – South Africa (Osorio et al. 2016).

15.19. *Neofusicoccum mediterraneum* Crous, M.J. Wingf. & A.J.L. Phillips, *Fungal Planet*, 19: 2 (2007), MycoBank MB504461.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Greece, Rhodes, Rhodos Palace Hotel parking lot, on branches and leaves of *Eucalyptus* sp., 12 Jun. 2006, collected by P.W. Crous, M.J. Wingfield & A.J.L. Phillips (holotype CBS H-19921, culture ex-type CBS 121718).

Hosts – Recent studies subsequent to Phillips et al. (2013) confirm the following hosts: *Vitis vinifera* (Berraf-Tebbal et al. 2014), *Diospyros kaki* (Palou et al. 2013), *Juglans regia* (Chen et al. 2014a), *Mangifera indica* (Krishnapillai & Wijeratnam 2015), *Olea europaea* (Úrbez-Torres et al. 2013), *Pistachia vera* (Michailides & Morgan 2016, Chen et al. 2014b).

Distribution – Recent studies subsequent to Phillips et al. (2013) confirm the following distribution: Algeria (Berraf-Tebbal et al. 2014), California (Chen et al. 2014a, b, Michailides & Morgan 2016, Úrbez-Torres et al. 2013), India (Krishnapillai et al. 2015), Spain (Palou et al. 2013).

15.20. *Neofusicoccum nonquaesitum* Inderb., Trouillas, Bostock & Michailides, *Mycologia* 102: 1360 (2010), MycoBank MB518135.

Sexual morph not reported. See Inderbitzin et al. (2010) for illustrations and descriptions of asexual morph.

Type – USA, California, Napa County, St Helena, on cankered branch of *Umbellularia californica* (*Lauraceae*), 12 Nov. 2004, F.P. Trouillas (holotype UC1946389, culture ex-type CBS 126655).

Hosts – *Malus domestica* (Rooney-Latham & Soriano 2016), *Sequoiadendron giganteum* (Rooney-Latham et al. 2012), *Umbellularia californica*, *Prunus dulcis* (Inderbitzin et al. 2010), *Vaccinium corymbosum* (Espinoza et al. 2009, Pérez et al. 2014).

Distribution – Chile (Espinoza et al. 2009, Pérez et al. 2014), North America (Rooney-Latham et al. 2012), USA (California) (Inderbitzin et al. 2010, Rooney-Latham & Soriano 2016).

15.21. *Neofusicoccum occulatum* Sakalidis & T.I. Burgess, *Mol. Phylog. Evol.* 60: 340 (2011), MycoBank MB518777.

Sexual morph not reported. See Sakalidis et al. (2011) for illustrations and descriptions of asexual morph.

Type – Australia, Queensland, Karanda, symptomless branches of *Eucalyptus grandis* hybrid (*Myrtaceae*), Mar 2002, T.I. Burgess (holotype MURU467, culture ex-type CBS 128008).

Host – *Eucalyptus* (Sakalidis et al. 2011).

Distribution – Australia (Sakalidis et al. 2011).

15.22. *Neofusicoccum parvum* (Pennycook & Samuels) Crous, Slippers & A.J.L. Phillips, *Stud. Mycol.* 55: 248 (2006), MycoBank MB500879.

Sexual morph and asexual morph reported. See Phillips et al. (2013) for illustrations and descriptions.

Type – New Zealand, Bay of Plenty, Te Puke, No 3 Road, Baldwin Orchard, on small dead branch of *Populus nigra* (*Salicaceae*), 17 Dec. 1981, S.R. Pennycook (holotype PDD 45438, culture ex-type PDDCC 8003 = ATCC 58191).

Hosts – Plurivorous.

Distribution – Worldwide.

15.23. *Neofusicoccum pennatisporum* K. Taylor, Barber & T.I. Burgess, *Mycol. Res.* 113: 346 (2009), MycoBank MB511826.

Sexual morph and asexual morph reported. See Taylor et al. (2009) for illustrations and descriptions. Morphologically and phylogenetically this species is divergent and its position within *Neofusicoccum* needs to be re-evaluated.

Type – Australia, Western Australia, Yalgorup National Park, from healthy stem of *Allocasuarina fraseriana* (*Casuarinaceae*), Jun. 2005, K.M. Taylor, holotype PERTH 07693044. WAC 13153 = MUCC 510 (ex-type).

Host – *Allocasuarina fraseriana* (Taylor et al. 2009).

Distribution – Western Australia (Taylor et al. 2009).

15.24. *Neofusicoccum protearum* (Denman & Crous) Crous, Slippers & A.J.L. Phillips, *Stud. Mycol.* 55: 249 (2006), MycoBank MB500880.

Sexual morph and asexual morph reported. See Denman et al. (2003) for illustrations and descriptions.

Type – Of sexual morph: South Africa, Western Cape, Porterville, Baanbreek Farm, on stems of *Protea magnifica* (*Proteaceae*), 27 Jul. 1997, S. Denman (holotype PREM 57329, cultures ex-type STE-U 4361 = CPC 4361).

Hosts – *Protea* sp., *Leucadendron* sp. (Denman et al. 2000, 2003, Marinowitz et al. 2008), *Santalum acuminatum* (Taylor et al. 2009).

Distribution – Australia, Portugal (continental and Madeira), South Africa, Spain (Tenerife), USA (Hawaii) (Denman et al. 2000, 2003, Marinowitz et al. 2008, Taylor et al. 2009).

15.25. *Neofusicoccum ribis* (Slippers, Crous & M.J. Wingf.) Crous, Slippers & A.J.L. Phillips, *Stud. Mycol.* 55: 249 (2006), MycoBank MB500881.

Sexual morph and asexual morph reported. See Slippers et al. (2004b) for illustrations and descriptions.

Type – USA, New York, Geneva, on *Ribes vulgare*, 1911, J.G. Grossenbacher & B.M. Duggar (holotype CUP-A (F.Col. 3408), culture ex-type CBS 115475).

Hosts – More than 250 hosts are listed in Farr & Rossman (2016) but many of the reports were published before the concept of *N. ribis* was clarified by Slippers et al. (2004a) and thus are not reliable. Recent studies subsequent to Phillips et al. (2013) confirm the following hosts: *Hevea brasiliensis* (Ngobisa et al. 2013), *Mallus domestica* (Jurick et al. 2013), *Psidium guajava* (Nogueira Junior et al. 2016).

Distribution – Brazil (Nogueira Junior et al. 2016), Malaysia (Ngobisa et al. 2013), Pennsylvania (Jurick et al. 2013).

15.26. *Neofusicoccum umdonicola* Pavlic, Slippers & M.J. Wingf., *Mycologia* 100: 644 (2008), MycoBank MB512500.

Sexual morph not reported. See Pavlic et al. (2009) for illustrations and descriptions of asexual morph.

Type – South Africa, Kwazulu-Natal Province, Kosi Bay from symptomless branches and leaves, dying branches and pulp of ripe fruits of *Syzygium cordatum* (*Myrtaceae*), Mar. 2002, D. Pavlic (holotype PREM 60068, culture ex-type CBS 123645).

Hosts – *Schizolobium parahyba* (Mehl et al. 2014), *Syzygium cordatum* (Pavlic et al. 2009, Pavlic-Zupanc et al. 2015).

Distribution – Ecuador (Mehl et al. 2014), South Africa (Pavlic et al. 2009).

15.27. *Neofusicoccum ursorum* Pavlic, Maleme, Slippers & M.J. Wingf., *Persoonia* 31: 271 (2013), MycoBank MB512478.

Sexual morph not reported. See Crous et al. (2013) for illustrations and descriptions of asexual morph.

Type – South Africa, Gauteng Province, Pretoria, from branches and leaves of living Eucalyptus trees (*Myrtaceae*), May 2005, H.M. Maleme (holotype PREM 59815, culture ex-type CBS 122811).

Host – *Eucalyptus* sp. (Crous et al. 2013).

Distribution – South Africa (Crous et al. 2013).

15.28. *Neofusicoccum viticlavatum* (Van Niekerk & Crous) Crous, Slippers & A.J.L. Phillips, *Stud. Mycol.* 55: 249 (2006), MycoBank MB500882.

Sexual morph not reported. See van Niekerk et al. (2004) for illustrations and descriptions of asexual morph.

Type – South Africa, Western Cape Province, Stellenbosch, on *V. vinifera* (*Vitaceae*), 2002, F. Halleen (holotype CBS H-7755, culture ex-type CBS 112878).

Host – *Vitis vinifera* (van Niekerk et al. 2004).

Distribution – South Africa (Western Cape Province) (van Niekerk et al. 2004).

15.29. *Neofusicoccum vitifusiforme* (Van Niekerk & Crous) Crous, Slippers & A.J.L. Phillips, *Stud. Mycol.* 55: 249 (2006), MycoBank MB500883.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – South Africa, Western Cape Province, Stellenbosch, on *V. vinifera* (*Vitaceae*), 2002, J.M. van Niekerk (holotype CBS H-7756, culture ex-type CBS 110887).

Hosts – *Acacia karroo* (Jami et al. 2013), *Eucalyptus corticosa* (as *N. corticosae*) (Summerell et al. 2006), *Eucalyptus* sp., *Eucalyptus camaldulensis*, *Eucalyptus diversicolor*, *E. pauciflora*,

Eucalyptus marginata, *Eucalyptus rubida*, *Eucalyptus viminalis* (as *D. eucalypti*) (Barber et al. 2005, Taylor et al. 2009, Sutton 1980), *Juglans regia* (Chen et al. 2014a), *Olea europaea* (Lazzizzera et al. 2008, Úrbez-Torres et al. 2013), *Pistachia vera* (Chen et al. 2014b), *Prunus armeniaca*, *Prunus persica*, *Prunus salicina* (Damm et al. 2007), *Schizolobium parahyba* (Mehl et al. 2014), *Vaccinium corymbosum* (Kong et al. 2010), *Vitis vinifera* (van Niekerk et al. 2004, Úrbez-Torres et al. 2012, Mondello et al. 2013).

Distribution – Australia (Sutton 1980, Barber et al. 2005, Summerell et al. 2006, Taylor et al. 2009), China (Kong et al. 2010), Italy (Lazzizzera et al. 2008, Mondello et al. 2013), South Africa (van Niekerk et al. 2004, Damm et al. 2007, Jami et al. 2013, Mehl et al. 2014), USA (Úrbez-Torres et al. 2012, 2013, Chen et al. 2014a, b).

16. *Neoscytalidium* Crous & Slippers, *Stud. Mycol.* 55: 244. 2006, MycoBank MB500868.

This genus comprises three species. The sexual morph has not been reported for any species.

Type species – *Neoscytalidium dimidiatum* (Penz.) Crous & Slippers.

Phillips et al. (2013) incorrectly stated that *N. hyalinum* is the type species of *Neoscytalidium*. The oldest epithet is *dimidiatum* based on *Torula dimidiata* (1882). Huang et al. (2016) corrected this error and placed *N. dimidiatum* as type species of the genus.

16.1. *Neoscytalidium dimidiatum* (Penz.) Crous & Slippers, *Stud. Mycol.* 55: 244 (2006), MycoBank MB500869.

Basionym – *Torula dimidiata* Penz., *Michelia* 2: 466. 1882 (basionym; hyphomycete synanamorph).

Synonym – *Neoscytalidium hyalinum* (C. K. Campb. & J. L. Mulder) AJL Phillips, Groenewald & Crous, *SIM* 76: 148 (2013). See Huang et al. (2016).

Sexual morph not reported. See Crous et al. (2006) for illustrations and description of asexual morph.

Type – Holotype of basionym apparently lost. Since *N. hyalinum* and *N. dimidiatum* are considered to be synonyms, the isotype and ex-isotype culture of *N. hyalinum* have been used as the standard for this species. Lectotype: United Kingdom, sole of human foot, 20 Nov. 1973, C.K. Campbell (isotype CBS H-7745, culture ex-isotype CBS 145.78).

Hosts – Recent studies subsequent to Phillips et al. (2013) confirm the following hosts: *Jatropha curcas* (Machado et al. 2014a), *Manihot esculenta* (Machado et al. 2014b), *Vitis vinifera* (Correia et al. 2016b).

Distribution – Recent studies subsequent to Phillips et al. (2013) confirm the following distribution: Brazil (Machado et al. 2014a, b, Correia et al. 2016b).

16.2. *Neoscytalidium novaehollandiae* Pavlic, T.I. Burgess & M.J. Wingf., *Mycologia* 100: 862 (2008), MycoBank MB512103.

Sexual morph not reported. See Pavlic et al. (2008) for illustrations and descriptions of asexual morph.

Type – Australia, Western Australia, Bell Gorge, on *Crotalaria medicaginea* (*Fabaceae*), Jul. 2006, T.I. Burgess (holotype PREM 60069, culture ex-type CBS 122071).

Hosts – *Acacia synchronica*, *Adansonia gibbosa*, *Crotalaria medicaginea*, *Grevillia agrifolia* (Pavlic et al. 2008), *Mangifera indica*, *Ficus carica* (Ray et al. 2010).

Distribution – Australia (Pavlic et al. 2008, Ray et al. 2010).

16.3. *Neoscytalidium orchidacearum* S. K. Huang, N. Tangthirasunun, J. C. Kang & K. D. Hyde, *Mycobiology* 44: 79 (2016).

Sexual morph not reported. See Huang et al. (2016) for illustrations and descriptions of asexual morph.

Type – Thailand, Sukhotai, on dead leaves of Orchid (*Orchidaceae*), 5 Aug 2012; S Hongsanan (holotype MFLU 13-0294, cultures ex-type MFLUCC 12-0533 and GZUCC 15113001).

Host – Orchid (Huang et al. 2016).

Distribution – Thailand (Huang et al. 2016).

17. *Phaeobotryon* Theiss. & Syd., *Ann. Mycol.* 13: 664 (1915), MycoBank MB3892.

This genus comprises four species. Sexual and asexual morphs have been reported within the genus. No cultures of the type species are extant.

Type species – *Phaeobotryon cercidis* (Cooke) Theiss. & Syd., *Ann. Mycol.* 13: 664. 1915.

17.1. *Phaeobotryon cupressi* Abdollahz., Zare & A.J.L. Phillips, *Persoonia* 23: 6 (2009), MycoBank MB513236.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Iran, Golestan Province, Gorgan, City Park, on twigs of *Cupressus sempervirens* (*Cupressaceae*), 15 Aug. 2006, M.A. Aghajani (holotype IRAN 13940F, cultures ex-type IRAN 1455C = CBS 124700).

Hosts – *Cupressus sempervirens* (Abdollahzadeh et al. 2009, Zlatkovic et al. 2016), *Juniperus scopulorum* (Alves et al. 2013).

Distribution – Iran (Abdollahzadeh et al. 2009), USA (Alves et al. 2013), Western Balkans (Zlatkovic et al. 2016).

17.2. *Phaeobotryon mamane* Crous & A.J.L. Phillips, *Persoonia* 21: 45 (2008), MycoBank MB506581.

Sexual morph and asexual morph reported. See Phillips et al. (2008) for illustrations and descriptions.

Type – USA, Hawaii, Manna Koa Park, Saddle Road, on stems of *Sophora chrysophylla* (*Fabaceae*), Jul. 2005, W. Gams (holotype CBS H-20109, culture ex-type CBS 122980).

Host – *Sophora chrysophylla* (Phillips et al. 2008).

Distribution – USA (Hawaii) (Phillips et al. 2008).

17.3. *Phaeobotryon negundinis* Daranagama, Bulgakov & K.D. Hyde, *Mycosphere* 7: 936 (2016), Facesoffunginumber FoF01916.

Sexual morph not reported. See Daranagama et al. (2016) for illustrations and descriptions of asexual morph.

Type – Russia, Rostov region, Rostov-on-Don city, Botanical Garden of Southern Federal University, Higher Park, on dying and dead twigs and branches of *Acer negundo* L. (*Sapindaceae*), 05 March 2014, T.S. Bulgakov (holotype MFLU 16-0475, culture ex-type MFLUCC 15-0436).

Hosts – *Acer negundo*, *Ligustrum vulgare*, *Forsythia* × *intermedia* (Daranagama et al. 2016).

Distribution – Russia (Daranagama et al. 2016).

17.4. *Phaeobotryon rhois* C.M. Tian, X.L. Fan & K.D. Hyde, *Phytotaxa* 205: 95 (2015), MycoBank MB811599, Facesoffunginumber FOF00596.

Sexual morph not reported. See Fan et al. (2015) for illustrations and descriptions of asexual morph.

Type – China, Ningxia Province: Yinchuan City, Yongning County, Wanghong Town, Nongsheng Village, 38°22'97.02" N, 106°21'44.89" E, alt. 1101 m, on twigs and branches of *Rhus typhina*

(Anacardiaceae), 26 July 2013, Xin-lei Fan & Hong Fan (holotype BJFC-S1007, cultures ex-type CFCC 89662 = CCTCC AF2014017).

Host – *Rhus typhina* (Fan et al. 2015).

Distribution – China (Fan et al. 2015).

18. *Pseudofusicoccum* Mohali, Slippers & M.J. Wingf., *Stud. Mycol.* 55: 249 (2006), MycoBank MB500884.

This genus comprises seven species. The sexual morph has not been reported for any species. The phylogenetic position of this genus has been questioned (Slippers et al. 2013) and it is still uncertain if it resides within *Botryosphaeriaceae*. However, until its position has been definitively resolved we continue to regard it as a genus within *Botryosphaeriaceae*.

Type species – *Pseudofusicoccum stromaticum* (Mohali, Slippers & M.J. Wingf.) Mohali, Slippers & M.J. Wingf., *Stud. Mycol.* 55: 249. 2006.

18.1. *Pseudofusicoccum adansoniae* Pavlic, T.I. Burgess & M.J. Wingf., *Mycologia* 100: 855 (2008), MycoBank MB512048.

Sexual morph not reported. See Pavlic et al. (2008) for illustrations and descriptions of asexual morph.

Type – Australia, Western Australia, Derby, on *Adansonia gibbosa* (*Malvaceae*), Jul. 2006, T.I. Burgess (holotype PREM 59841, culture ex-type CBS 122055).

Hosts – *Adansonia gibbosa*, *Acacia synchronica*, *Eucalyptus* sp., *Ficus opposita* (Pavlic et al. 2008), *Adansonia gregorii*, *Grevillea agrifolia* (Sakalidis et al. 2011), *Cassia fistula* (Trakuningcharoen et al. 2015b), *Dimocarpus longan* (Trakuningcharoen et al. 2015b), *Hevea brasiliensis* (Trakuningcharoen et al. 2015a), *Jatropha podagrica* (Sharma et al. 2013), *Senna siamea* (Trakuningcharoen et al. 2015b), *Tectona grandis* (Doilom et al. 2015).

Distribution – Australia (Pavlic et al. 2008, Sakalidis et al. 2011), India (Sharma et al. 2013), Thailand (Doilom et al. 2015, Trakuningcharoen et al. 2015a, b).

18.2. *Pseudofusicoccum ardesiacum* Pavlic, T.I. Burgess & M.J. Wingf., *Mycologia* 100: 858 (2008), MycoBank MB512051.

Sexual morph not reported. See Pavlic et al. (2008) for illustrations and descriptions of asexual morph.

Type – Australia, Western Australia, Mount Hardman, Great Northern Highway, on *Adansonia gibbosa* (*Malvaceae*), Jul. 2006, T.I. Burgess (holotype PREM 59843, culture ex-type CBS 122062).

Hosts – *Adansonia gibbosa*, *Eucalyptus* sp. (Pavlic et al. 2008), *Caesalpinia pulcherrima*, *Veitchia merrillii* (Trakuningcharoen et al. 2015b).

Distribution – Australia (Pavlic et al. 2008), Thailand (Trakuningcharoen et al. 2015b).

18.3. *Pseudofusicoccum artocarpi* T. Trakuningcharoen, L. Lombard & Crous, *Persoonia* 34: 96 (2014), MycoBank MB810167.

Sexual morph not reported. See Trakuningcharoen et al. (2015b) for illustrations and descriptions of asexual morph.

Type – Thailand, Chiang Mai province, on twigs of *Artocarpus heterophyllus* (*Moraceae*), May 2012, T. Trakuningcharoen (holotype CBS H-21935, culture ex-type CBS 138655).

Host – *Artocarpus heterophyllus* (Trakuningcharoen et al. 2015b).

Distribution – Thailand (Trakuningcharoen et al. 2015b).

18.4. *Pseudofusicoccum kimberleyensis* Pavlic, T.I. Burgess & M.J. Wingf., *Mycologia* 100: 857 (2008), MycoBank MB512049.

Sexual morph not reported. See Pavlic et al. (2008) for illustrations and descriptions of asexual morph.

Type – Australia, Western Australia, Tunnel Creek National Park, on *Acacia synchronica* (*Fabaceae*), Jul. 2006, T.I. Burgess (holotype PREM 59842, culture ex-type CBS 122058).

Hosts – *Adansonia gibbosa*, *Acacia synchronica*, *Eucalyptus* sp., *Ficus opposita* (Pavlic et al. 2008).

Distribution – Western Australia (Pavlic et al. 2008).

18.5. *Pseudofusicoccum olivaceum* Mehl & Slippers, *Mycologia* 103: 537 (2011), MycoBank MB513501.

Sexual morph not reported. See Mehl et al. (2011) for illustrations and descriptions of asexual morph.

Type – South Africa, Mpumalanga Province, Kruger National Park, Pretoriuskop, on an asymptomatic branch of *Pterocarpus angolensis* (*Fabaceae*), Sep. 2005, J. Roux (holotype PREM 60328, culture ex-type CBS 124939).

Host – *Pterocarpus angolensis* (Mehl et al. 2011).

Distribution – South Africa (Mehl et al. 2011).

18.6. *Pseudofusicoccum stromaticum* (Mohali, Slippers & M.J. Wingf.) Mohali, Slippers & M.J. Wingf., *Stud. Mycol.* 55: 249, MycoBank MB500885.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – Venezuela, Portuguesa Morph, Acarigua, Smurfit Company, on branches of *Eucalyptus urophylla* (*Myrtaceae*), Feb. 2003, S. Mohali (holotype PREM 58237, culture ex-type CMW 13366, other representative cultures CBS 117448, CBS 117449).

Hosts – *Eucalyptus* sp. (Mohali et al. 2006), *Acacia mangium* (Mohali et al. 2006), *Mangifera indica* (Marques et al. 2012, 2013b).

Distribution – Brazil (Marques et al. 2012, 2013b), Venezuela (Mohali et al. 2006).

18.7. *Pseudofusicoccum violaceum* Mehl & Slippers, *Mycologia* 103: 541 (2011), MycoBank MB513500.

Sexual morph not reported. See Mehl et al. (2011) for illustrations and descriptions of asexual morph.

Type – South Africa, Mpumalanga Province, Mawewe Nature Reserve, on an asymptomatic branch of *Pterocarpus angolensis* (*Fabaceae*), Dec. 2005, J.W.M. Mehl & J. Roux (holotype PREM 60333, culture ex-type CBS 124936).

Hosts – *Pterocarpus angolensis* (Mehl et al. 2011), *Tinospora cordifolia* (Mishra et al. 2012).

Distribution – India (Mishra et al. 2012), South Africa (Mehl et al. 2011).

19. *Sakireeta* Subram. & K. Ramakr., *Journal of the Indian Botanical Society* 36: 83 (1957).

This genus comprises only one species. The sexual morph has not been reported.

Type species – *Sakireeta madreeya* Subram. & K. Ramakr 1957.

19.1. *Sakireeta madreeya* Subram. & K. Ramakr., *Journal of the Indian Botanical Society* 36: 84 (1957).

Synonym – *Tiarosporella madreeya* (Subram. & K. Ramakr.) Nag Raj, *Canadian Journal of Botany* 51: 2470 (1974) [1973].

Sexual morph not reported. See Crous et al. (2015a) for illustrations and descriptions of asexual morph.

Type – India, Madras, Choolai, on dead culm of *Aristida setacea* (*Poaceae*), 27 Sept. 1951, K. Ramakrishnan (holotype MUBL 631); Kurukshetra Univ., undetermined grass host, July 1976, R.S. Mehrotra, CBS H-21854, culture CBS 532.76.

Host – *Aristida setacea* (Crous et al. 2015a).

Distribution – India (Crous et al. 2015a).

20. *Sardiniella* Linaldeddu, A. Alves & A.J.L. Phillips, *Mycosphere* 7: 900 (2016), MycoBank MB817511.

Sardiniella was introduced to accommodate species that are morphologically similar to *Diplodia* and *Dothiorella/Spencermartinsia*. Phylogenetically *Sardiniella* is closely related to *Neofusicoccum* and *Dothiorella/Spencermartinsia*. This genus comprises a single species. The sexual morph has not been reported.

Type species – *Sardiniella urbana* Linaldeddu, A. Alves & A.J.L. Phillips 2016.

20. 1. *Sardiniella urbana* Linaldeddu, A. Alves & A.J.L. Phillips, *Mycosphere* 7: 900 (2016), MycoBank MB817512.

Sexual morph not reported. See Linaldeddu et al. (2016b) for illustrations and descriptions of asexual morph.

Type – Italy, Sassari, isolated from a branch canker of *Celtis australis* (*Cannabaceae*), 9 September 2013, Benedetto T. Linaldeddu (holotype LISE 96308, culture ex-type CBS 141580).

Host – *Celtis australis* (Linaldeddu et al. 2016b)

Distribution – Italy (Linaldeddu et al. 2016b)

21. *Spencermartinsia* A.J.L. Phillips, A. Alves & Crous, *Persoonia* 21: 51 (2008), MycoBank MB511762.

This genus comprises eight species (Fig. 4). Sexual morph and asexual morphs have been reported within the genus. Phylogenetically *Spencermartinsia* is very closely related to *Dothiorella*. Morphologically the two can be differentiated only in the sexual morphs; ascospores of *Spencermartinsia* bear hyaline apiculi, which are absent in *Dothiorella*. With the addition of more species, the distinction between these two genera has become more tenuous. Nevertheless, in this paper we continue to regard them as separate pending further study.

Type species – *Spencermartinsia viticola* (A.J.L. Phillips & J. Luque) A.J.L. Phillips, A. Alves & Crous, *Persoonia* 21: 51. 2008.

21.1. *Spencermartinsia alpina* Y. Zhang ter. & M. Zhang, sp. nov., MycoBank MB 819517.

“*Spencermartinsia alpina*” Y. Zhang ter. & M. Zhang, *Mycosphere* 7: 945 (2016), nom. inval.

Zhang et al. (2016) introduced *Spencermartinsia alpina* as a new species but did not explicitly designate a holotype and thus the name was invalid. To validate the name the holotype is cited below. The sexual morph was not reported. Full description and diagnosis in Zhang et al. (2016).

Type – China, Yunnan Province, Chuxiong City, Zixi Mountain Forest Park, from thick branch of dead young tree of *Platycladus orientalis* (*Cupressaceae*), 16 Dec 2014, leg. W. He & J.R. Wu, det. Y. Zhang (holotype HMAS254733, culture ex-type CGMCC 3.18001).

Host – *Platycladus orientalis* (Zhang et al. 2016).

Distribution – China (Zhang et al. 2016).

21.2. *Spencermartinsia citricola* A.J.L. Phillips & Abdollahz., *Persoonia* 32: 7 (2014), MycoBank MB803992.

Sexual morph not reported. See Abdollahzadeh et al. (2014) for illustrations and descriptions of asexual morph.

Type – New Zealand, Northland, Kerikeri, Collins Orchard, Inlet Road, on twigs of *Citrus sinensis* (*Rutaceae*), Sept. 2006, S.R. Pennycook, P.R. Johnston and B.C. Paulus (holotype PDD92023, cultures ex-type ICMP16828 = CBS124729).

Host – *Citrus sinensis* (Abdollahzadeh et al. 2014).

Distribution – New Zealand (Abdollahzadeh et al. 2014).

21.3. *Spencermartinsia mangiferae* Abdollahz., Javadi & A.J.L. Phillips, *Persoonia* 32: 9 (2014), MycoBank MB803993.

Sexual morph not reported. See Abdollahzadeh et al. (2014) for illustrations and descriptions of asexual morph.

Type – Iran, Hormozgan Province, Bandar Abbas (Hajiabad-Siaho), on twigs of *Mangifera indica* (*Anacardiaceae*), Mar. 2007, J. Abdollahzadeh & A. Javadi (holotype IRAN 14266F, cultures ex-type IRAN 1584C = CBS 124727).

Host – *Mangifera indica* (Abdollahzadeh et al. 2014).

Distribution – Iran (Abdollahzadeh et al. 2014).

21.4. *Spencermartinsia plurivora* Abdollahz., Javadi & A.J.L. Phillips, *Persoonia* 32: 9 (2014), MycoBank MB803994.

Sexual morph not reported. See Abdollahzadeh et al. (2014) for illustrations and descriptions of asexual morph.

Type – Iran, Khuzestan Province, Dezful (Safiabad Citrus Research Centre), on twigs of *Citrus* sp. (*Rutaceae*), Nov. 2006, J. Abdollahzadeh & A. Javadi (holotype IRAN 14267F, cultures ex-type IRAN 1557C = CBS 124724).

Host – *Citrus* sp. (Abdollahzadeh et al. 2014).

Distribution – Iran (Abdollahzadeh et al. 2014).

21.5. *Spencermartinsia rosulata* F.J.J. Van der Walt, Slippers & G.J. Marais, *Persoonia* 33: 164 (2014), MycoBank MB518724.

Sexual morph not reported. See Slippers et al. (2014) for illustrations and descriptions of asexual morph.

Type – Namibia, Windhoek, from *Acacia karroo* (*Fabaceae*), Feb. 2006, F.J.J. van der Walt & J. Roux (holotype PREM 59622, culture ex-type CBS 121760).

Host – *Acacia* sp. (Slippers et al. 2014).

Distribution – Namibia, South Africa (Slippers et al. 2014).

21.6. *Spencermartinsia viticola* (A.J.L. Phillips & J. Luque) A.J.L. Phillips, A. Alves & Crous, *Persoonia* 21: 51 (2008), MycoBank MB511763.

Sexual morph and asexual morph reported. See Phillips et al. (2013) for illustrations and descriptions.

Type – Spain, Catalonia, Vimbodí, near the Monastery of Poblet, on pruned canes of *Vitis vinifera* cv. Garnatxa Negra (*Vitaceae*), Aug. 2004, J. Luque & S. Martos (holotype LISE 95177, culture ex-type CBS 117009).

Hosts – *Acacia karroo* (Jami et al. 2013, 2014), *Celtis africana* (Jami et al. 2014), *Citrus* sp. (Adesemoye & Eskalen 2011, Inderbitzin et al. 2010), *Gymnosporia buxifolia* (Jami et al. 2014), *Populus cathayana* (Zhang et al. 2009), *Poniciana gilliesii* (Phillips et al. 2008), *Prunus persica* and *P. salicina* (Damm et al. 2007), *Vitis vinifera* (de Wet et al. 2009, Luque et al. 2005, Qiu et al. 2011, Úrbez-Torres et al. 2007, Pitt et al. 2013b, Diaz et al. 2013, Valencia et al. 2015).

Distribution – Australia (Qiu et al. 2011, Pitt et al. 2013b), Chile (Diaz et al. 2013), China (Zhang et al. 2009, Valencia et al. 2015), France (Phillips et al. 2008), South Africa (Damm et al. 2007, de Wet et al. 2009, Jami et al. 2013, 2014), Spain (Luque et al. 2005) and USA (Adesemoye & Eskalen 2011, Inderbitzin et al. 2010, Úrbez-Torres et al. 2007).

21.7. *Spencermartinsia westrale* W.M. Pitt, J.R. Úrbez-Torres & Trouillas, *Australas. Pl. Path.* 44: 48 (2015), MycoBank MB808286.

Sexual morph not reported. See Pitt et al. (2015) for illustrations and descriptions of asexual morph.

Type – Australia, Upper Swan, Western Australia, discarded canes of *Vitis vinifera* (*Vitaceae*), November 2009, F.P. Trouillas (holotype, DAR80529, culture ex-type, DAR80529).

Host – *Vitis vinifera* (Pitt et al. 2015).

Distribution – Australia (Pitt et al. 2015).

21.8. *Spencermartinsia yunnana* Y. Zhang ter. & M. Zhang, sp. nov, MycoBank MB 819518.

“*Spencermartinsia yunnana*” Y. Zhang ter. & M. Zhang, *Mycosphere* 7: 946 (2016), nom. inval.

Zhang et al. (2016) introduced *Spencermartinsia yunnana* as a new species but did not explicitly designate a holotype and thus the name was invalid. To validate the name the holotype is cited below. The sexual morph not reported. Full description and diagnosis in Zhang et al. (2016).

Type – China, Yunnan Province, Chuxiong City, Zixi Mountain Forest Park, from dead branch of *Camellia* sp. (*Theaceae*), 16 Dec 2014, leg. W. He & J.R. Wu, det. Y. Zhang (holotype HMAS254734, culture ex-type CGMCC 3.17999).

Hosts – *Acer buergerianum*, *Camellia* sp., *Poncirus trifoliata*, *Ternstroemia gymnanthera* (Zhang et al. 2016).

Distribution – China (Zhang et al. 2016).

22. *Sphaeropsis* Sacc., *Michelia* 2: 105. 1880, MycoBank MB9992.

This genus comprises five species. Sexual and asexual morphs have been reported within the genus.

Type species – *Sphaeropsis visci* (Alb. & Schwein.) Sacc., *Michelia* 2: 105. 1880.

22.1. *Sphaeropsis citrigena* (A.J.L. Phillips, P.R. Johnst. & Pennycook) A.J.L. Phillips & A. Alves, *Stud. Mycol.* 76: 158 (2013), MycoBank MB805463.

Sexual morph and asexual morph reported. See Phillips et al. (2008) and Phillips et al. (2013) for illustrations and descriptions.

Type – New Zealand, Northland, Kerikeri, Davies Orchard (#2), Inlet Road, on recently dead bark-covered twigs of *Citrus sinensis* (*Rutaceae*), 6 Sep. 2006, S.R. Pennycook, P.R. Johnston & B.C. Paulus (holotype PDD 89238, culture ex-type ICMP 16812).

Host – *Citrus sinensis* (Phillips et al. 2013).

Distribution – New Zealand (Phillips et al. 2013).

22.2. *Sphaeropsis eucalypticola* (Doilom, J.K. Liu, & K.D. Hyde) A.J.L. Phillips, *Stud. Mycol.* 76: 158 (2013), MycoBank MB805464.

Asexual morph not reported. See Liu et al. (2012) and Phillips et al. (2013) for illustrations and descriptions of sexual morph.

Type – Thailand, Chiang Rai Province, Muang District, on dead twig of *Eucalyptus* sp. (*Myrtaceae*), 8 Aug. 2011, M. Doilom (holotype MFLU 12-0753, culture ex-type MFLUCC 11-0579 = CBS 133993).

Hosts – *Eucalyptus* sp. (Liu et al. 2012), *Tectona grandis* (Doilom et al. 2015).

Distribution – Thailand (Liu et al. 2012, Doilom et al. 2015).

22.3. *Sphaeropsis porosa* (Van Niekerk & Crous) A.J.L. Phillips & A. Alves, *Stud. Mycol.* 76: 159 (2013), MycoBank MB805465.

Sexual morph not reported. See Phillips et al. (2013) for illustrations and descriptions of asexual morph.

Type – South Africa, Western Cape Province, Stellenbosch, on *Vitis vinifera* (*Vitaceae*), 2002, J.M. van Niekerk (holotype CBS H-12039, culture ex-type CBS 110496).

Host – *Vitis vinifera* (van Niekerk et al. 2004).

Distribution – South Africa (Western Cape Province) (van Niekerk et al. 2004).

22.4. *Sphaeropsis variabilis* F.J.J. Van der Walt, Slippers & G.J. Marais, *Persoonia* 33: 164 (2014), MycoBank MB518720.

Sexual morph not reported. See Slippers et al. (2014) for illustrations and description of asexual morph. In our families tree (Fig. 1) *S. variabilis* appears to be more closely related to *Alanphillipsia* than to *Sphaeropsis*. In the phylogenies presented by Slippers et al. (2014) *S. variabilis* was not clearly aligned within *Sphaeropsis*, but they did not include any species of *Alanphillipsia* in their phylogenies. Furthermore, Slippers et al. (2014) did not include any details of paraphyses, which are an important character that distinguishes *Sphaeropsis* from *Diplodia*. For these reasons we consider that the position of *S. variabilis* in *Sphaeropsis* is tenable and needs to be re-assessed.

Type – Namibia, Windhoek, from *Acacia karroo* (*Fabaceae*), Feb. 2006, F.J.J. van der Walt & J. Roux (holotype PREM 59637, culture ex-type CBS 121774).

Host – *Acacia* sp. (Slippers et al. 2014).

Distribution – Namibia, South Africa (Slippers et al. 2014).

22.5. *Sphaeropsis visci* (Alb. & Schwein.) Sacc., *Michelia* 2: 105. (1880), MycoBank MB281898.

Sexual morph and asexual morph reported. See Phillips et al. (2008) and Phillips et al. (2013) for illustrations and descriptions.

Type – Germany, on *Viscum album* (*Santalaceae*), Albertini & Schweinitz, holotype could not be located and presumably lost. Ukraine, National Nature Park ‘Svjatie Gory’, on branches of *Viscum album*, 10 Mar. 2007, Á. Akulov (neotype CWU (MYC) AS 2271, cultures ex-neotype CBS 122526, CBS 122527).

Hosts – *Viscum album* (Sutton 1980, Phillips et al. 2008, Varga et al. 2014, Poczai et al. 2015, Zlatkovic et al. 2016).

Distribution – Austria, Czechoslovakia, Egypt, Romania (Sutton 1980), Ukraine (Phillips et al. 2008), Hungary (Poczai et al. 2015), Serbia, Luxemburg (Zlatkovic et al. 2016).

23. *Tiarosporella* Höhn, *Ber. Deutsch. Bot. Ges.* 37: 159. (1919), MycoBank MB10233.

This genus comprises only one species. The sexual morph has not been reported.

Type species: *Tiarosporella paludosa* (Sacc. & Fiori ex P. Syd.) Höhn 1919.

23.1. *Tiarosporella paludosa* (Sacc. & Fiori) Höhn., *Berichte der Deutschen Botanischen Gesellschaft* 37: 159 (1919).

Basionym – *Neottiospora paludosa* Sacc. & Fiori, *Hedwigia Beiblätter* 38: 137 (1899).

Sexual morph not reported. See Crous et al. (2015a) for illustrations and descriptions of asexual morph.

Type – Germany, Berlin, Zahlendorf, on leaves of *Eriophorum polystachium* (Cyperaceae), Oct. 1895, P. Sydow (holotype in FH, isotype DAOM 130546). Netherlands, Winterswijk in the Korenburgerveen, Latitude 51.990133, Longitude 6.664013, on *Trichophorum cespitosum* subsp. *germanicum*, 28 Apr. 2013, W. Quaedvlieg (epitype CBS H-21855 MBT200481, culture ex-epitype CBS 138577).

Hosts – *Eleocharis palustris*, *Trichophorum cespitosum* (Crous et al. 2015a).

Distribution – Germany, Netherlands (Crous et al. 2015a).

Acknowledgements

We are grateful to the CARS-30, Youth Foundation of Beijing Academy of Agriculture and Forestry Sciences (number QNJJ 201515). K.D. Hyde thanks the Chinese Academy of Sciences, project number 2013T2S0030, for the award of Visiting Professorship for Senior International Scientists at Kunming Institute of Botany. AJL Phillips acknowledges support from Biosystems and Integrative Sciences Institute (BioISI, FCT/UID/ Multi/04046/2013).

References

- Abdollahzadeh J, Javadi A, Mohammadi Goltapeh E, Zare R, Phillips AJL. 2010 – Phylogeny and morphology of four new species of *Lasiodiplodia* from Iran. *Persoonia* 25, 1–10.
- Abdollahzadeh J, Javadi A, Zare R, Phillips AJL. 2014 – A phylogenetic study of *Dothiorella* and *Spencermatinsia* species associated with woody plants in Iran, New Zealand, Portugal and Spain. *Persoonia* 32, 1–12.
- Abdollahzadeh J, Mohammadi Goltapeh E, Javadi A, Shams-Bakhsh M, Zare R, et al. 2009 – *Barriopsis iraniana* and *Phaeobotryon cupressi*: two new species of the *Botryosphaeriaceae* from trees in Iran. *Persoonia* 23, 1–8.
- Abdollahzadeh J, Zare R, Phillips AJL. 2013 – Phylogeny and taxonomy of *Botryosphaeria* and *Neofusicoccum* species in Iran, with description of *Botryosphaeria scharifii* sp. nov. *Mycologia* 105, 210–220.
- Abreo E, Martinez S, Bettucci L, Lupo S. 2013 – Characterization of *Botryosphaeriaceae* species associated with grapevines in Uruguay. *Australasian Plant Pathology* 42, 241–249.
- Acimovic SG, Harmon CL, Bec S, Wyka S, Broders K, et al. 2016 – First report of *Diplodia corticola* causing decline of red oak (*Quercus rubra*) trees in Maine. *Plant Disease* 100, 649–650.
- Adamcik S, Cai L, Chakraborty D, Chen XH, Cotter HVT, et al. 2015 – Fungal Biodiversity profiles 1-10. *Cryptogamie Mycologie* 36, 121–166.
- Adamson K, Klavina D, Drenkhan R, Gaitnieks T, Hanso M. 2015 – *Diplodia sapinea* is colonizing the native Scots pine (*Pinus sylvestris*) in the northern Baltics. *European Journal of Plant Pathology* 143, 343–350.
- Adesemoye AO, Eskalen A. 2011 – First report of *Spencermatinsia viticola*, *Neofusicoccum austral* & *N. parvum* causing branch canker of citrus in California. *Plant Disease* 95, 770.
- Adesemoye AO, Mayorquin JS, Wang DH, Twizeyimana M, Lynch SC, et al. 2014 – Identification of species of *Botryosphaeriaceae* causing bot gummosis in citrus in California. *Plant Disease* 98, 55–61.
- Akgul DS, Savas NG, Teker T, Keykubat B, Mayorquin JS. 2015 – Fungal trunk pathogens of sultana seedless vineyards in Aegean region of Turkey. *Phytopathologia Mediterranea* 54, 380–393.

- Al-Sadi AM, Al-Ghaithi AG, Al-Fahdi N, Al-Yahyai R. 2014 – Characterization and pathogenicity of fungal pathogens associated with root diseases of citrus in Oman. *International Journal of Agriculture and Biology* 16, 371–376.
- Al-Sadi AM, Al-Wehaibi AN, Al-Shariqi RM, Alhammadi MS, Alhosni IA, et al. 2013 – Population genetic analysis reveals diversity in *Lasiodiplodia* species infecting date palm, citrus, and mango in Oman and the UAE. *Plant Disease* 97, 1363–1369.
- Alves A, Barradas C, Phillips AJL, Correia A. 2013 – Diversity of *Botryosphaeriaceae* species associated with conifers in Portugal. *European Journal of Plant Pathology* 135, 791–804.
- Alves A, Correia A, Luque J, Phillips AJL. 2004 – *Botryosphaeria corticola*, sp. nov. on *Quercus* species, with notes and description of *Botryosphaeria stevensii* and its anamorph, *Diplodia mutila*. *Mycologia* 96, 598–613.
- Alves A, Correia A, Phillips AJL. 2006 – Multi-gene genealogies and morphological data support *Diplodia cupressi* sp. nov., previously recognized as *D. pinea* f. sp. *cupressi*, as a distinct species. *Fungal Diversity* 23, 1–15.
- Alves A, Crous PW, Correia A, Phillips AJL. 2008 – Morphological and molecular data reveal cryptic speciation in *Lasiodiplodia theobromae*. *Fungal Diversity* 28, 1–13.
- Alves A, Linaldeddu BT, Deidda A, Scanu B, Phillips AJL. 2014 – The complex of *Diplodia* species associated with *Fraxinus* and some other woody hosts in Italy and Portugal. *Fungal Diversity* 67, 143–156.
- Ammad F, Benchabane M, Toumi M, Belkacem N, Guesmi A, et al. 2014 – Occurrence of *Botryosphaeriaceae* species associated with grapevine dieback in Algeria. *Turkish Journal of Agriculture and Forestry* 38, 865–876.
- Andolfi A, Basso S, Giambra S, Conigliaro G, Piccolo SL, et al. 2016 – Lasiolactols A and B produced by the grapevine fungal pathogen *Lasiodiplodia mediterranea*. *Chemistry & Biodiversity* 13, 395–402.
- Ariyawansa HA, Hyde KD, Jayasiri SC, Buyck B, Cui YY, et al. 2015 – Fungal Diversity Notes 111–252 Taxonomic and phylogenetic contributions to fungal taxa. *Fungal Diversity* 75, 27–274.
- Ariyawansa HA, Hyde KD, Liu JK, Wu SP, Liu ZY. 2016 – Additions to Karst Fungi 1: *Botryosphaeria minutispermata* sp. nov., from Guizhou Province, China. *Phytotaxa* 275, 035–044.
- Barber PA, Burgess TJ, Hardy GESTJ, Slippers B, Keane PJ, et al. 2005 – *Botryosphaeria* species from Eucalyptus in Australia are pleoanamorphic, producing *Dichomera* synanamorphs in culture. *Mycological Research* 109, 1347–1363.
- Barradas C, Correia A, Alves A. 2013 – First report of *Neofusicoccum australe* and *N. luteum* associated with canker and dieback of *Quercus robur* in Portugal. *Plant Disease* 97, 560.
- Begoude BAD, Slippers B, Wingfield MJ, Roux J. 2011 – The pathogenic potential of endophytic *Botryosphaeriaceae* fungi on *Terminalia* species in Cameroon. *Forest Pathology* 41, 281–292.
- Berraf-Tebbal A, Guereiro MA, Phillips AJL. 2014 – Phylogeny of *Neofusicoccum* species associated with grapevine trunk diseases in Algeria, with description of *Neofusicoccum algeriense* sp. nov. *Phytopathologia Mediterranea* 53, 416–427.
- Bian CH, Miao P, Kang YB. 2015 – First report of canker disease of flue-cured tobacco caused by *Botryosphaeria dothidea* in China. *Plant Disease* 99, 890–890.
- Billones-Baaijens R, Jones EE, Ridgway HJ, Jaspers MW. 2013 – Virulence affected by assay parameters during grapevine pathogenicity studies with *Botryosphaeriaceae* nursery isolates. *Plant Pathology* 62, 1214–1225.
- Billones-Baaijens R, Ridgway HJ, Jones EBG, Jaspers MV. 2015 – Spatial distribution of *Neofusicoccum* species within a rootstock mother vine indicates potential infection pathways. *European Journal of Plant Pathology* 141, 267–269.

- Burgess TI, Barber PA, Hardy GESTJ. 2005 – *Botryosphaeria* spp. Associated with eucalypts in Western Australia, including the description of *Fusicoccum macroclavatum* sp. nov. *Australasian Plant Pathology* 34, 557–567.
- Burgess TI, Barber PA, Mohali S, Pegg G, Beer W de, et al. 2006 – Three new *Lasiodiplodia* spp. from the tropics, recognized based on DNA sequence comparisons and morphology. *Mycologia* 98, 423–435.
- Carlucci A, Cibelli F, Lops F, Raimondo ML. 2015 – Characterization of *Botryosphaeriaceae* species as causal agents of trunk disease on grapevines. *Plant Disease* 99, 1678–1688.
- Castillo S, Borrero C, Castano R, Rodríguez A, Avilés M. 2013 – First report of canker disease caused by *Neofusicoccum parvum* and *N. australe* on blueberry bushes in Spain. *Plant Disease* 97, 1112–1112.
- Castro-Medina F, Mohali SR, Úrbez-Torres JR, Gubler WD. 2014 – First report of *Lasiodiplodia pseudotheobromae* causing trunk cankers in *Acacia mangium* in Venezuela. *Plant Disease* 98, 686–686.
- Chebil S, Fersi R, Yakoub A, Chenenaoui S, Chattaoui M, et al. 2014 – First report of *Botryosphaeria dothidea*, *Diplodia seriata*, and *Neofusicoccum luteum* associated with canker and dieback of grapevines in Tunisia. *Plant Disease* 98, 420–420.
- Chen SF, Fichtner E, Morgan DP, Michailides TJ. 2013a – First report of *Lasiodiplodia citricola* and *Neoscytalidium dimidiatum* causing death of graft union of English walnut in California. *Plant Disease* 97, 993–993.
- Chen SF, Li G, Liu F, Michailides TJ. 2015 – Novel species of *Botryosphaeriaceae* associated with shoot blight of pistachio. *Mycologia* 107, 780–792.
- Chen SF, Morgan DP, Hasey JK, Michailides TJ. 2013b – First report of *Lasiodiplodia citricola* associated with stem canker of peach in California, USA. *Journal of Plant Pathology* 95, 659–659.
- Chen SF, Morgan DP, Hasey JK, Anderson K, Michailides TJ. 2014a – Phylogeny, morphology, distribution, and pathogenicity of *Botryosphaeriaceae* and *Diaporthaceae* from English walnut in California. *Plant Disease* 98, 636–652.
- Chen SF, Morgan DP, Michailides TJ. 2014b – *Botryosphaeriaceae* and *Diaporthaceae* associated with panicle and shoot blight of pistachio in California, USA. *Fungal Diversity* 67, 157–179.
- Chen SF, Pavlic D, Roux J, Slippers B, Xie YJ, et al. 2011 – Characterization of *Botryosphaeriaceae* from plantation-grown Eucalyptus species in South China. *Plant Pathology* 60, 739–751.
- Conforto C, Lima NB, Garcete-Gomez JM, Camara MPS, Michereff S. 2016 – First report of cladode brown spot in cactus prickly pear caused by *Neofusicoccum batangarum* in Brazil. *Plant Disease* 100, 1238–1239.
- Correia KC, Camara MPS, Barbosa MAG, Sales R, Agusti-Brisach C, et al. 2013 – Fungal trunk pathogens associated with table grape decline in Northeastern Brazil. *Phytopathologia Mediterranea* 52, 380–387.
- Correia KC, Silva MA, de Moraes MA Jr, Armengol J, Phillips AJL, et al. 2016a – Phylogeny, distribution and pathogenicity of *Lasiodiplodia* species associated with dieback of table grape in the main Brazilian exporting region. *Plant pathology* 65, 92–103.
- Correia KC, Silva MA, Netto MSB, Vieira WAS, Câmara MPS, et al. 2016b – First report of grapevine dieback caused by *Neoscytalidium hyalinum* in Brazil. *Plant Disease* 100, 213–213.
- Cosoveanu A, Hernandez M, Iacomi-Vasilescu B, Zhang X, Shu S, et al. 2016 – Fungi as endophytes in Chinese *Artemisia* spp: juxtaposed elements of phylogeny, diversity and bioactivity. *Mycosphere* 7, 102–117.
- Coutinho IBL, Freire FCO, Lima CS, Lima JS, Goncalves FJT, et al. 2016 – Diversity of genus *Lasiodiplodia* associated with perennial tropical fruit plants in northeastern Brazil. *Plant Pathology* Doi: 10.1111/ppa.12565.

- Crous PW, Groenewald JZ, Shivas RG, Edwards J, Seifert KA, et al. 2011 – Fungal Planet Description Sheets: 69–91. *Persoonia* 26, 108–156.
- Crous PW, Müller MM, Sánchez RM, Giordano L, Bianchinotti MV, et al. 2015a – Resolving *Tiarosporella* spp. allied to *Botryosphaeriaceae* and *Phacidiaceae*. *Phytotaxa* 202, 073–093.
- Crous PW, Palm ME. 1999 – Reassessment of the anamorph genera *Botryodiplodia*, *Dothiorella* and *Fusicoccum*. *Sydowia* 51, 167–175.
- Crous PW, Schumacher RK, Wingfield MJ, Lombard L, Giraldo A, et al. 2015b – Fungal Systematics and Evolution: FUSE 1. *Sydowia* 67, 81–118.
- Crous PW, Slippers B, Wingfield MJ, Rheeder J, Marasas WFO, et al. 2006 – Phylogenetic lineages in the *Botryosphaeriaceae*. *Studies in Mycology* 55, 235–253.
- Crous PW, Wingfield MJ, Guarro J, Cheewangkoon R, van der Bank M, et al. 2013 – Fungal Planet description sheets: 154–213. *Persoonia* 31, 188–296.
- Crous PW, Wingfield MJ, Richardson DM, Le Roux JJ, Strasberg D, et al. 2016 – Fungal Planet description sheets: 400–468. *Persoonia* 36, 316–458.
- Crous PW, Wingfield MJ, Schumacher RK, Summerell BA, Giraldo A, et al. 2014 – Fungal Planet description sheets: 281–319. *Persoonia* 33, 212–292.
- Cruywagen EM, Slippers B, Roux J, Wingfield MJ. 2016 – Phylogenetic species recognition and hybridization in *Lasiodiplodia*: A case study on species from baobabs. *Fungal Biology* <http://dx.doi.org/10.1016/j.funbio.2016.07.014>.
- Damm U, Crous PW, Fourie PH. 2007 – *Botryosphaeriaceae* as potential pathogens of *Prunus* species in South Africa, with descriptions of *Diplodia africana* and *Lasiodiplodia plurivora* sp. nov. *Mycologia* 99, 664–680.
- Daranagama DA, Thambugala KM, Campino B, Alves A, Bulgakov TS, et al. 2016 – *Phaeobotryon negundinis* sp. nov. (Botryosphaeriales) from Russia. *Mycosphere* 7, 933–941.
- De la Mora-Castaneda JG, Cibrian-Tovar D, Perez-Vera OA. 2014 – *Neofusicoccum eucalyptorum* (= *Botryosphaeria eucalyptorum*) and *N. parvum*: pathogens in eucalyptus plantations in Mexico. *Revista Chapingo Serie Ciencias Forestales Y Del Ambiente* 20, 187–197.
- Denman S, Crous PW, Groenewald JZ, Slippers B, Wingfield BD, et al. 2003 – Circumscription of *Botryosphaeria* species associated with *Proteaceae* based on morphology and DNA sequence data. *Mycologia* 95, 294–307.
- Denman S, Crous PW, Taylor JE, Kang J-C, Pascoe I, et al. 2000 – An overview of the taxonomic history of *Botryosphaeria*, and a re-evaluation of its anamorphs based on morphology and ITS rDNA phylogeny. *Studies in Mycology* 45, 129–140.
- Diaz GA, Auger J, Besoain X, Bordeu E, Latorre BA. 2013 – Prevalence and pathogenicity of fungi associated with grapevine trunk diseases in Chilean vineyards. *Ciencia e Investigación Agraria* 40, 327–339.
- Didier Begoude BA, Slippers B, Wingfield MJ, Roux J. 2010 – *Botryosphaeriaceae* associated with *Terminalia catappa* in Cameroon, South Africa and Madagascar. *Mycological Progress* 9, 101–123.
- Dissanayake AJ, Camporesi E, Hyde KD, Phillips AJL, Fu CY, et al. 2016 – *Dothiorella* species associated with woody hosts in Italy. *Mycosphere* 7, 51–63.
- Dissanayake AJ, Zhang W, Li XH, Zhou Y, Hyde KD, et al. 2015b – First report of *Neofusicoccum mangiferae* associated with grapevine dieback in China. *Phytopathologia Mediterranea* 54, 414–419.
- Dissanayake AJ, Zhang W, Liu M, Chukeatirote E, Yan JY, et al. 2015a – *Lasiodiplodia pseudotheobromae* causes pedicel and peduncle discolouration of grapes in China. *Australasian Plant Disease* 10, 21.
- Doilom M, Shuttleworth L, Roux J, Chukeatirote E, Hyde KD. 2014 – *Barriopsis tectonae* sp. nov. a new species of *Botryosphaeriaceae* from *Tectona grandis* (teak) in Thailand. *Phytotaxa* 176, 081–091.

- Doilom M, Shuttleworth LA, Roux J, Chukeatirote E, Hyde KD. 2015 – *Botryosphaeriaceae* associated with *Tectona grandis* (teak) in Northern Thailand. *Phytotaxa* 233, 1–26.
- Doll DA, Rolshausen PE, Pouzoulet J, Michailides TJ. 2015 – First report of *Dothiorella iberica* causing trunk and scaffold cankers of almond in California. *Plant Disease* 99, 1185–1185.
- Dreaden TJ, Black AW, Mullerin S, Smith JA. 2014 – First report of *Diplodia quercivora* causing shoot dieback and branch cankers on live oak (*Quercus virginiana*) in the United States. *Plant Disease* 98, 282.
- Espinoza JG, Briceño EX, Chávez ER, Úrbez-Torres JR, Latorre BA. 2009 – *Neofusicoccum* sp. associated with stem canker & dieback of blueberry in Chile. *Plant Disease* 93, 1187–1194.
- Fan XL, Hyde KD, Liu JK, Liang YM, Tian CM. 2015 – Multigene phylogeny and morphology reveal *Phaeobotryon rhois* sp. nov. (Botryosphaeriales, Ascomycota). *Phytotaxa* 205, 90–98.
- Farr DF, Elliot M, Rossman AY, Edmonds RL. 2005 – *Fusicoccum arbuti* sp. nov. causing cankers on Pacific madrone in western North America with notes on *Fusicoccum dimidiatum*, the correct name for *Scytalidium dimidiatum* and *Nattrassia mangiferae*. *Mycologia* 97, 730–741.
- Farr DF, Rossman AY. 2016. Fungal databases, systematic mycology and microbiology laboratory, ARS, USDA. (SMML database). <https://nt.ars-grin.gov/fungaldatabases/>. Accessed 21 September 2016.
- Ferreira MC, Almeida V, Mariana de L, Zani CL, Junior PAS, et al. 2015 – Molecular phylogeny, diversity, symbiosis and discover of bioactive compounds of endophytic fungi associated with the medicinal Amazonian plant *Carapa guianensis* (Meliaceae). *Biochemical Systematics and Ecology* 59, 36–44.
- Funk A. 1964 – *Botryosphaeria tsugae* sp. nov, causing dieback of western hemlock in British Columbia. *Canadian Journal of Botany* 42, 769–775.
- Gardner DE. 1997 – *Botryosphaeria mamane* sp. nov. associated with witches'- brooms on the endemic forest tree *Sophora chrysophylla* in Hawaii. *Mycologia* 89, 298–303.
- Giambra S, Piazza G, Alves A, Mondello V, Berbegal M, et al. 2016 – *Botryosphaeriaceae* species associated with diseased loquat trees in Italy and description of *Diplodia rosacearum* sp. nov. *Mycosphere* 7, 978–989.
- Gramaje D, Agustí-Brisach C, Pérez-Sierra A, Moralejo E, Olmo D, et al. 2012 – Fungal trunk pathogens associated with wood decay of almond trees on Mallorca (Spain). *Persoonia* 28, 1–13.
- Gure A, Slippers B, Stenlid J. 2005 – Seed-borne *Botryosphaeria* spp. from native *Prunus* and *Podocarpus* trees in Ethiopia, with a description of the anamorph *Diplodia rosulata* sp. nov. *Mycological Research* 109, 1005–1014.
- Hall TA. 1999 – BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series* 41, 95–98.
- Huang SK, Tangthirasunun N, Phillips AJL, Dai DQ, Wanasinghe DN, et al. 2016 – Morphology and phylogeny of *Neoscytalidium orchidacearum* sp. nov. (*Botryosphaeriaceae*). *Mycobiology* 44, 79–84.
- Hyde KD, Hongsanan S, Jeewon R, Bhat DJ, McKenzie EHC, et al. 2016 – Fungal diversity notes 367–491: taxonomic and phylogenetic contributions to fungal taxa. *Fungal Diversity* 80, 1–270.
- Inderbitzin P, Bostock RM, Trouillas FP, Michailides TJ. 2010 – A six locus phylogeny reveals high species diversity in *Botryosphaeriaceae* from California almond. *Mycologia* 102, 1350–1368.
- Ismail AM, Cirvilleri G, Lombard L, Crous PW, Groenewald JZ, et al. 2013 – Characterisation of *Neofusicoccum* species causing mango dieback in Italy. *Journal of Plant Pathology* 95, 549–557.
- Ismail AM, Cirvilleri G, Polizzi G, Crous PW, Groenewald JZ, et al. 2012 – *Lasiodiplodia* species associated with dieback disease of mango (*Mangifera indica*) in Egypt. *Australasian Plant Pathology* 41, 649–660.

- Jami F, Slippers B, Wingfield MJ, Gryzenhout M. 2013 – Greater *Botryosphaeriaceae* diversity in healthy than associated diseased *Acacia karroo* tree tissues. *Australasian Plant Pathology* 42, 421–430.
- Jami F, Slippers B, Wingfield MJ, Gryzenhout M. 2014 – *Botryosphaeriaceae* species overlap on four unrelated, native South African hosts. *Fungal Biology* 118, 168–179.
- Jami F, Slippers B, Wingfield MJ, Loots MT, Gryzenhout M. 2015 – Temporal and spatial variation of *Botryosphaeriaceae* associated with *Acacia karroo* in South Africa. *Fungal Ecology* 15, 51–62.
- Jami F, Slippers B, Wingfield MJ, Gryzenhout M. 2012 – Five new species of the *Botryosphaeriaceae* from *Acacia karroo* in South Africa. *Cryptogamie Mycologie* 33, 245–266.
- Jayawardena RS, Li XH, Xu W, Yan JY, Li HL, et al. 2016. First Report of *Botryosphaeria dothidea* causing leaf necrosis of *Camellia sinensis* in Fujian Province, China. *Plant Disease* 100, 854–854.
- Jurick WM, Vico I, Gaskins VL, Janisiewicz WJ, Peter KA. 2013 – First report of *Neofusicoccum ribis* causing postharvest decay of apple fruit from cold storage in Pennsylvania. *Plant Disease* 97, 999–999.
- Katoh K, Toh H. 2008 – Recent developments in the MAFFT multiple sequence alignment program. *Briefings in Bioinformatics* 9, 276–285.
- Kong CS, Qiu XL, Yi KS, Yu XF, Yu L. 2010 – First report of *Neofusicoccum vitifusiforme* causing blueberry blight of blueberry in China. *Plant Disease* 94, 1373.
- Konta S, Hongsanan S, Phillips AJL, Jones EBG, Boonmee S, et al. 2016b – *Botryosphaeriaceae* from palms in Thailand II - two new species of *Neodeightonia*, *N. rattanica* and *N. rattanicola* from *Calamus* (rattan palm). *Mycosphere* 7, 950–961.
- Konta S, Phillips AJL, Bahkali AH, Jones EBG, Eungwanichayapant DP, et al. 2016a – *Botryosphaeriaceae* from palms in Thailand - *Barriopsis archontophoenicis* sp. nov, from *Archontophoenix alexandrae*. *Mycosphere* 7, 921–932.
- Krishnapillai N, Wijeratnam RSW. 2015 – First report of *Neofusicoccum mediterraneum* causing stem end rot on Karuthakolumban mangoes. *Plant Disease* 99, 1858–1858.
- Lawrence DP, Hand FP, Gubler WD, Trouillas FP. 2016 – *Botryosphaeriaceae* species associated with dieback and canker disease of bay laurel in northern California with the description of *Dothiorella californica* sp. nov. *Fungal Biology* doi: 10.1016/j.funbio.2016.09.005.
- Lazzizzera C, Frisullo S, Alves A, Phillips AJL. 2008 – Morphology, phylogeny and pathogenicity of *Botryosphaeria* and *Neofusicoccum* species associated with drupe rot of olives in southern Italy. *Plant Pathology* 57, 948–956.
- Li GJ, Hyde KD, Zhao RL, Hongsanan S, Abdel-Aziz FA, et al. 2016a – Fungal diversity notes 253–366: taxonomic and phylogenetic contributions to fungal taxa. *Fungal Diversity* 78: 1–237.
- Li GQ, Arnold RJ, Liu FF, Li JQ, Chen SF. 2015 – Identification and Pathogenicity of *Lasiodiplodia* Species from *Eucalyptus urophyllaxgrandis*, *Polyscias balfouriana* and *Bougainvillea spectabilis* in Southern China. *Journal of Phytopathology* 163, 956–967.
- Li GQ, Liu FF, Li JQ, Liu QL, Chen SF. 2016b – Characterization of *Botryosphaeria dothidea* and *Lasiodiplodia pseudotheobromae* from English Walnut in China. *Journal of Phytopathology* 164, 348–353.
- Li W, Liu J, Bhat DJ, Camporesi E, Xu J, et al. 2014 – Introducing the novel species, *Dothiorella symphoricarposicola*, from snowberry in Italy. *Cryptogamie Mycology* 35, 257–270.
- Ligoxigakis EK, Markakis EA, Papaioannou IA, Typas MA. 2013 – First report of palm rot of *Phoenix* spp caused by *Neodeightonia phoenicum* in Greece. *Plant Disease* 97, 286–286.
- Linaldeddu BT, Alves A, Phillips AJL. 2016b – *Sardiniella urbana* gen. et sp. nov., a new member of the *Botryosphaeriaceae* isolated from declining *Celtis australis* trees in Sardinian streetscapes. *Mycosphere* 7, 893–905.

- Linaldeddu BT, Deidda A, Scanu B, Franceschini A, Alves A, et al. 2016a – Phylogeny, morphology and pathogenicity of *Botryosphaeriaceae*, *Diatrypaceae* and *Gnomoniaceae* associated with branch diseases of hazelnut in Sardinia (Italy). *European Journal of Plant Pathology* Doi: 10.1007/s10658-016-0912-z.
- Linaldeddu BT, Deidda A, Scanu B, Franceschini A, Serra S, et al. 2015 – Diversity of *Botryosphaeriaceae* species associated with grapevine and other woody hosts in Italy, Algeria and Tunisia, with descriptions of *Lasiodiplodia exigua* and *Lasiodiplodia mediterranea* sp. nov. *Fungal Diversity* 71, 201–214.
- Linaldeddu BT, Franceschini A, Alves A, Phillips AJL. 2013 – *Diplodia quercivora* sp. nov.: a new species of *Diplodia* found on declining *Quercus canariensis* trees in Tunisia. *Mycologia* 105, 1266–1274.
- Linaldeddu BT, Maddau L, Franceschini A, Alves A, Phillips AJL. 2016c – *Botryosphaeriaceae* species associated with lentisk dieback in Italy and description of *Diplodia insularis* sp. nov. *Mycosphere* 7, 962–977.
- Linaldeddu BT, Scanu B, Maddau L, Franceschini A. 2014 – *Diplodia corticola* and *Phytophthora cinnamomi*: the main pathogens involved in holm oak decline on Caprera island (Italy). *Forest Pathology* 44, 191–200.
- Liu JK, Phookamsak R, Doilom M, Wikee S, Li YM, et al. 2012 – Towards a natural classification of Botryosphaeriales. *Fungal Diversity* 57, 149–210.
- Lopes A, Phillips AJL, Alves A (2016) – Mating type genes in the genus *Neofusicoccum*: mating strategies and usefulness in species delimitation. *Fungal Biology* Doi 10.1016/j.funbio.2016.08.011
- Lopes UP, Zambolim L, Pinho DB, Barros AV, Costa H, et al. 2014 – Postharvest rot and mummification of strawberry fruits caused by *Neofusicoccum parvum* and *N. kwambonambiense* in Brazil. *Tropical Plant Pathology* 39, 178–183.
- Luque J, Martos S, Phillips AJL. 2005 – *Botryosphaeria viticola* sp. nov. on grapevines: a new species with a *Dothiorella* anamorph. *Mycologia* 97, 1111–1121.
- Lynch SC, Eskalen A, Zambino PJ, Mayorquin JS, Wang DH. 2013 – Identification and pathogenicity of *Botryosphaeriaceae* species associated with coast live oak (*Quercus agrifolia*) decline in southern California. *Mycologia* 105, 125–140.
- Lynch SC, Zambino PJ, Scott TA, Eskalen A. 2014 – Occurrence, incidence and associations among fungal pathogens and *Agrilus auroguttatus*, and their roles in *Quercus agrifolia* decline in California. *Forest Pathology* 44, 62–74.
- Machado AR, Pinho DB, de Oliveira SAS, Pereira OL. 2014b – New occurrences of *Botryosphaeriaceae* causing black root rot of *cassava* in Brazil. *Tropical Plant Pathology* 39, 464–470.
- Machado AR, Pinho DB, Pereira OL. 2014a – Phylogeny, identification and pathogenicity of the *Botryosphaeriaceae* associated with collar and root rot of the biofuel plant *Jatropha curcas* in Brazil, with a description of new species of *Lasiodiplodia*. *Fungal Diversity* 67, 231–247.
- Marincowitz S, Groenewald JZ, Wingfield MJ, Crous PW. 2008 – Species of *Botryosphaeriaceae* occurring on *Proteaceae*. *Persoonia* 21, 111–118.
- Marques MW, Lima NB, de Moraes MA Jr, Barbosa MAG, Souza BO, et al. 2013a – Species of *Lasiodiplodia* associated with mango in Brazil. *Fungal Diversity* 61, 181–193.
- Marques MW, Lima NB, de Moraes MA Jr, Michereff SJ, Phillips AJL, et al. 2013b. *Botryosphaeria*, *Neofusicoccum*, *Neoscytalidium* and *Pseudofusicoccum* species associated with mango in Brazil. *Fungal Diversity* 61, 195–208.
- Marques MW, Lima NB, Michereff SJ, Câmara MPS, Souza CRB. 2012 – First report of mango dieback caused by *Pseudofusicoccum stromaticum* in Brazil. *Plant Disease* 96, 144.
- Mcgregor RR, Sakalidis ML, Hamelin RC. 2016 – *Neofusicoccum arbuti*: a hidden threat to *Arbutus menziesii* characterized by widespread latent infections and a broad host range. *Canadian Journal of Plant Pathology* 38, 70–81.

- Mehl JWM, Slippers B, Roux J, Wingfield MJ. 2011 – *Botryosphaeriaceae* associated with *Pterocarpus angolensis* (kiaat) in South Africa. *Mycologia* 103, 534–553.
- Mehl JWM, Slippers B, Roux J, Wingfield MJ. 2014 – *Botryosphaeriaceae* associated with dieback of *Schizolobium parahyba* trees in South Africa and Ecuador. *Forest Pathology* 44, 396–408.
- Michailides TJ, Morgan DP. 2016 – Association of *Botryosphaeria* panicle and shoot blight of Pistachio with injuries of fruit caused by Hemiptera insects and birds. *Plant Disease* 100, 1405–1413.
- Mishra A, Gond SK, Kumar A, Sharma VK, Verma SK, et al. 2012 – Season and tissue type affect fungal endophyte communities of the Indian medicinal plant *Tinospora cordifolia* more strongly than geographic location. *Microbial Ecology* 64, 388–398.
- Mohali SR, Slippers B, Wingfield MJ. 2006 – Identification of *Botryosphaeriaceae* from Eucalyptus, *Acacia* and *Pinus* in Venezuela. *Fungal Diversity* 25, 103–125.
- Mohammadi H, Kazemi S, Farahmand H. 2014 – *Phaeoacremonium* and *Botryosphaeriaceae* species associated with cypress (*Cupressus sempervirens* L.) decline in Kerman province (Iran). *Phytopathologia Mediterranea* 53, 27–39.
- Mohammadi H. 2014 – *Phaeoacremonium* spp and *Botryosphaeriaceae* spp associated with date palm (*Phoenix dactylifera* L) decline in Iran. *Journal of Phytopathology* 162, 575–581.
- Mondello V, Lo PS, Conigliaro G, Alfonzo A, Torta L, et al. 2013 – First report of *Neofusicoccum vitifusiforme* and presence of other *Botryosphaeriaceae* species associated with *Botryosphaeria* dieback of grapevine in Sicily (Italy). *Phytopathologia Mediterranea* 52, 388–396.
- Netto MSB, Assuncao IP, Lima GSA, Marques MW, Lima WG, et al. 2014 – Species of *Lasiodiplodia* associated with papaya stem-end rot in Brazil. *Fungal Diversity* 67, 127–141.
- Netto MSB, Lima WG, Correia KC, Da Silva CFB, Thon M, et al. 2016 – Analysis of phylogeny, distribution, and pathogenicity of *Botryosphaeriaceae* species associated with gummosis of *Anacardium* in Brazil, with a new species of *Lasiodiplodia*. *Fungal Biology* <http://dx.doi.org/10.1016/j.funbio.2016.07.006>.
- Ngobisa AICN, Abidin MAZ, Wong MY, Wan NM. 2013 – *Neofusicoccum ribis* associated with leaf blight on rubber (*Hevea brasiliensis*) in Peninsular Malaysia. *Plant Pathology Journal* 29, 10–16.
- Ni HF, Yang HR, Chen RS, Rueyfen L, Tinghsuan H. 2012 – New *Botryosphaeriaceae* fruit rot of mango in Taiwan: identification and pathogenicity. *Botanical Studies* 53, 467–478.
- Niekerk JM van, Bester W, Halleen F, Crous PW, Fourie PH. 2010 – First report of *Lasiodiplodia crassispora* as a pathogen of grapevine trunks in South Africa. *Plant Disease* 94, 1063.
- Niekerk JM van, Crous PW, Groenewald JZ, Fourie PH, Halleen F. 2004 – DNA phylogeny, morphology and pathogenicity of *Botryosphaeria* species on grapevines. *Mycologia* 96, 781–798.
- Nogueira Junior AF, Fischer IH, Braganca CAD, Massola Jr NS, Amorim L. 2016 – Identification of *Botryosphaeriaceae* species that cause styler-end rot of guavas and characterisation of the disease monocycle. *European Journal of plant pathology* 144, 271–287.
- Nylander JAA. 2004 – MrModeltest 2.0. Program distributed by the author. Evolutionary Biology Centre, Uppsala University.
- Osorio JA, Crous CJ, Beer ZWD, Wingfield MJ, Roux J. 2016 – Endophytic *Botryosphaeriaceae*, including five new species, associated with mangrove trees in South Africa. *Fungal Biology* doi:10.1016/j.funbio.2016.09.004J.
- Palou L, Montesinos-Herrero C, Besada C, Taberner V. 2013 – Postharvest fruit rot of Persimmon (*Diospyros kaki*) in Spain caused by *Lasiodiplodia theobromae* and *Neofusicoccum* spp. *Journal of Phytopathology* 161, 625–631.
- Pavlic D, Slippers B, Coutinho TA, Gryzenhout M, Wingfield MJ. 2004 – *Lasiodiplodia gonubiensis* sp. nov., a new *Botryosphaeria* anamorph from native *Syzygium cordatum* in South Africa. *Studies in Mycology* 50, 313–322.

- Pavlic D, Slippers B, Coutinho TA, Wingfield MJ. 2007 – *Botryosphaeriaceae* occurring on native *Syzygium cordatum* in South Africa and their potential threat to Eucalyptus. *Plant Pathology* 56, 624–636.
- Pavlic D, Slippers B, Coutinho TA, Wingfield MJ. 2009 – Molecular and phenotypic characterization of three phylogenetic species discovered within the *Neofusicoccum parvum/N. ribis* complex. *Mycologia* 101, 636–647.
- Pavlic D, Wingfield MJ, Barber P, Slippers B, Hardy GESTJ, et al. 2008 – Seven new species of the *Botryosphaeriaceae* from baobab and other native trees in Western Australia. *Mycologia* 100, 851–866.
- Pavlic-Zupanc D, Pisčur B, Slippers B, Wingfield M, Jurc D. 2015 – Molecular and morphological characterization of *Dothiorella* species associated with dieback of *Ostrya carpinifolia* in Slovenia and Italy. *Phytopathologia Mediterranea* 54, 222–231.
- Pérez CA, Wingfield MJ, Slippers B, Altier NA, Blanchette RA. 2010 – Endophytic and canker-associated *Botryosphaeriaceae* occurring on non-native Eucalyptus and native *Myrtaceae* trees in Uruguay. *Fungal Diversity* 41, 53–69.
- Pérez SF, Merino-Gergichevich C, Guerrero JC. 2014. Detection of *Neofusicoccum nonquaesitum* causing dieback and canker in highbush blueberry from Southern Chile. *Journal of Soil Science and Plant Nutrition* 14, 581–588.
- Phillips AJL, Alves A, Abdollahzadeh J, Slippers B, Wingfield MJ, et al. 2013 – The *Botryosphaeriaceae*: genera and species known from culture. *Studies in Mycology* 76, 51–167.
- Phillips AJL, Alves A, Pennycook SR, Johnston PR, Ramaley A, et al. 2008 – Resolving the phylogenetic and taxonomic status of dark-spored teleomorph genera in the *Botryosphaeriaceae*. *Persoonia* 21, 29–55.
- Phillips AJL, Lopes J, Abdollahzadeh J, Bobev S, Alves A. 2012 – Resolving the *Diplodia* complex on apple and other *Rosaceae* hosts. *Persoonia* 29, 29–38.
- Phillips AJL, Oudemans PV, Correia A, Alves A. 2006 – Characterization and epitypification of *Botryosphaeria corticis*, the cause of blueberry cane canker. *Fungal Diversity* 21, 141–155.
- Pillay K, Slippers B, Wingfield MJ, Gryzenhout M. 2013 – Diversity and distribution of co-infecting *Botryosphaeriaceae* from *Eucalyptus grandis* and *Syzygium cordatum* in South Africa. *South African Journal of Botany* 84, 38–43.
- Pitt WM, Huang R, Steel CC, Savocchia S. 2013a – Pathogenicity and epidemiology of *Botryosphaeriaceae* species isolated from grapevines in Australia. *Australasian Plant Pathology* 42, 573–582.
- Pitt WM, Úrbez-Torres JR, Trouillas FP. 2013b – *Dothiorella vidmadera*, a novel species from grapevines in Australia and notes on *Spencermartinsia*. *Fungal Diversity* 61, 209–219.
- Pitt WM, Úrbez-Torres JR, Trouillas FP. 2015 – *Dothiorella* and *Spencermartinsia*, new species and records from grapevines in Australia. *Australasian Plant Pathol* 44, 43–56.
- Poczai P, Varga I, Hyvonen J. 2015 – Internal transcribed spacer (ITS) evolution in populations of the hyperparasitic European mistletoe pathogen fungus, *Sphaeropsis visci* (*Botryosphaeriaceae*): The utility of ITS2 secondary structures. *Gene* 558, 54–64.
- Poletto T, Gonzatto Maciel C, Muniz M, Blume E, et al. 2016 – First report of stem canker caused by *Lasiodiplodia subglobosa* on *Carya illinoensis* in Brazil. *Plant Disease* 100, 1016–1017.
- Punithalingam E. 1969 – Studies on Sphaeropsidales in culture. *Mycological Papers* 119, 1–24.
- Qiu Y, Steel CC, Ash GJ, Savocchia S. 2011 – Survey of *Botryosphaeriaceae* associated with grapevine decline in the Hunter Valley and Mudgee grape growing regions of New South Wales. *Australasian Plant Pathology* 40, 1–11.
- Rabari V, Rakhshiyi P, Patel P, Thaker V. 2016 – First Report of *Botryosphaeria dothidea* on *Mangifera indica* L in Gujarat. *Journal of Phytopathology* 164, 286–289.

- Ray JD, Lanoiselet V, Burgess TI. 2010 – First record of *Neoscytalidium dimidiatum* and *Neoscytalidium novaehollandiae* on *Mangifera indica* and *Ficus carica* in Australia. *Australasian Plant Disease Notes* 5, 48–50.
- Rodríguez-Galvéz E, Guerrero P, Barradas C, Crous PW, Alves A. 2016 – Phylogeny and pathogenicity of *Lasiodiplodia* species associated with dieback of mango in Peru. *Fungal Biology* <http://dx.doi.org/10.1016/j.funbio.2016.06.004>.
- Rojas EI, Herre EA, Mejía LC, Arnold AE, Chaverri P, et al. 2008 – *Endomelanconiopsis*, a new anamorph genus in the *Botryosphaeriaeae*. *Mycologia* 100, 760–775.
- Ronquist F, Huelsenbeck JP. 2003 – MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19, 1572–1574.
- Rooney-Latham S, Soriano MC. 2016 – First Report of *Neofusicoccum nonquaesitum* Causing Branch Dieback of Apple in California. *Plant Disease* 100, 1012–1012.
- Rooney-Latham S, Tidwell TE, Blomquist CL, Peek KS. 2012 – First report of *Neofusicoccum nonquaesitum* causing branch cankers on giant sequoia (*Sequoiadendron giganteum*) in North America. *Plant Disease* 96, 905.
- Rosado AWC, Machado AR, Freire FCO, Pereira OL. 2016 – Phylogeny, identification, and pathogenicity of *Lasiodiplodia* associated with postharvest stem-end rot of coconut in Brazil. *Plant Disease* 100, 561–568.
- Sakalidis ML, Hardy GESTJ, Burgess TI. 2011 – Use of the genealogical sorting index (GSI) to delineate species boundaries in the *Neofusicoccum parvum*-*Neofusicoccum ribis* species complex. *Molecular Phylogenetics and Evolution* 60, 333–344.
- Sandoval-Sanchez M, Nieto-Angel D, Sergio SIJ, Téliz-Ortiz D, Orozco-Santos M, et al. 2013 – Fungi associated to stem-end rot and dieback of mango (*Mangifera indica* L.). *Agrocienca* 47, 61–73.
- Serrato-Diaz LM, Rivera-Vargas L, French-Monar RD. 2014 – First Report of *Neofusicoccum mangiferae* Causing Rachis Necrosis and Inflorescence Blight of Mango (*Mangifera indica*) in Puerto Rico. *Plant Disease* 98, 570–571.
- Sharma R, Kulkarni G, Shouche YS. 2013 – *Pseudofusicoccum adansoniae* isolated as an endophyte from *Jatropha podagrica*: new record for India. *Mycotaxon* 123, 39–45.
- Shear CL, Stevens NE, Wilcox MS. 1923 – *Botryosphaeria* and *Physalospora* in the Eastern United States. *Mycologia* 17, 98–107.
- Shetty KG, Minnis AM, Rossman AY, Jayachandran K. 2011 – The Brazilian peppertree seed-borne pathogen, *Neofusicoccum batangarum*, a potential biocontrol agent. *Biological Control* 56, 91–97.
- Shivas RG, Tree DJ, Tan YP, Ballard EL. 2009 – *Dothiorella thripsita*. *Fungal Planet* 32, *Persoonia* 22, 168–169.
- Silvestro D, Michalak I. 2010 - RaxmlGUI: a graphical front-end for RAXML. Available from: <http://sourceforge.net/projects/raxmlgui/> (accessed 25 September 2016)
- Slippers B, Boissin E, Phillips AJL, Groenewald JZ, Wingfield MJ, et al. 2013 – Phylogenetic lineages in the Botryosphaerales: A systematic and evolutionary framework. *Studies in Mycology* 76, 31–49.
- Slippers B, Crous PW, Denman S, Coutinho TA, Wingfield BD, et al. 2004a – Combined multiple gene genealogies and phenotypic characters differentiate several species previously identified as *Botryosphaeria dothidea*. *Mycologia* 96, 83–101.
- Slippers B, Fourie G, Crous PW, Coutinho TA, Wingfield BD, et al. 2004b – Speciation and distribution of *Botryosphaeria* sp. on native and introduced Eucalyptus trees in Australia and South Africa. *Studies in Mycology* 50, 343–358.
- Slippers B, Johnson GI, Crous PW, Coutinho TA, Wingfield BD, et al. 2005 – Phylogenetic and morphological re-evaluation of the *Botryosphaeria* species causing diseases of *Mangifera indica*. *Mycologia* 97, 99–110.
- Slippers B, Roux J, Wingfield MJ, Van der Walt FJJ, Jami F, et al. 2014 – Confronting the constraints of morphological taxonomy in the *Botryosphaerales*. *Persoonia* 33, 155–168.

- Smith DR, Stanosz GR. 2001 – Molecular and morphological differentiation of *Botryosphaeria dothidea* (anamorph *Fusicoccum aesculi*) from some other fungi with *Fusicoccum* anamorphs. *Mycologia* 93, 505–515.
- Solel Z, Madar Z, Kimchi M, Golan Y. 1987 – *Diplodia* canker of cypress. *Canadian Journal of Plant Pathology* 9, 115–118.
- Stevens NE. 1926 – Two species of *Physalospora* on citrus and other hosts. *Mycologia* 18, 206–217.
- Summerell BA, Groenewald JZ, Carnegie AJ, Summerbell RC, Crous PW. 2006 – Eucalyptus microfungi known from culture *Alysidiella*, *Fusculina* and *Phlogicylindrium* genera nova, with notes on some other poorly known taxa. *Fungal Diversity* 23, 323–350.
- Sutton BC, Marasas WFO. 1976 – Observations on *Neottiosporina* and *Tiarosporella*. *Transactions of the British Mycological Society* 67, 69–76.
- Sutton BC. 1980 – The Coelomycetes: Fungi imperfecti with pycnidia, acervuli and stromata. Commonwealth Mycological Institute, Kew, Surrey, England.
- Swofford DL. 2002 – PAUP* 4.0: phylogenetic analysis using parsimony (*and other methods). Sinauer Associates, Sunderland.
- Taylor K, Barber PA, Hardy GESTJ, Burgess TI. 2009 – *Botryosphaeriaceae* from tuart (*Eucalyptus gomphocephala*) woodland, including descriptions of four new species. *Mycological Research* 113, 337–353.
- Tennakoon DS, Phillips AJL, Phookamsak R, Ariyawansa HA, Bahkali AH, et al. 2016 – Sexual morph of *Lasiodiplodia pseudotheobromae* (*Botryosphaeriaceae*, Botryosphaerales, Dothideomycetes) from China. *Mycosphere* 7, 990–1000.
- Thambugala KM, Daranagama DA, Camporesi E, Singtripop C, Liu ZY, et al. 2014 – Multi-locus phylogeny reveals the sexual state of *Tiarosporella* in *Botryosphaeriaceae*. *Cryptogam Mycol* 35, 359–367.
- Theissen F, Sydow H. 1918 – Vorentwürfe zu den Pseudosphaerales. *Annales Mycologici* 16, 1–34.
- Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F, Higgins DG. 1997 – The Clustal X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Research* 24, 4876–4882.
- Thynne E, McDonald MC, Evans M, Wallwork H, Neate S, et al 2015 – Re-classification of the causal agent of white grain disorder on wheat as three separate species of *Eutiarosporella*. *Australasian Plant Pathology* 44, 527–539.
- Trakunyingcharoen T, Cheewangkoon R, To-anun C. 2015a – Phylogenetic study of the *Botryosphaeriaceae* species associated with avocado and para rubber in Thailand. *Chiang Mai Journal of Science* 42, 104–116.
- Trakunyingcharoen T, Lombard L, Groenewald JZ, Cheewangkoon R, To-Anun C, et al. 2015b – Caulicolous Botryosphaerales from Thailand. *Persoonia* 34, 87–99.
- Triki MA, HadjTaieb SK, Cheffi M, Gharbi Y, Rhouma A. 2015 – First report of dieback of Olive trees caused by *Neofusicoccum australe* in Tunisia. *Journal of Plant Pathology* 97, 212–212.
- Turkolmez S, Dervis S, Ciftci O, Serçe CU. 2016 – First Report of Canker and Dieback Caused by *Botryosphaeria dothidea* on Apple in Turkey. *Plant Disease* 100, 1237–1238.
- Úrbez-Torres JR, Boule J, O'Gorman DT. 2016 – First Report of *Diplodia seriata* and *D. mutila* Causing Apple Dieback in British Columbia. *Plant Disease* 100, 1243–1244.
- Úrbez-Torres JR, Gubler WD, Luque J. 2007 – First report of *Botryosphaeria iberica* and *B. viticola* associated with grapevine decline in California. *Plant Disease* 91, 772.
- Úrbez-Torres JR, Peduto F, Rooney-Latham S, Gubler WD. 2010 – First report of *Diplodia corticola* causing grapevine (*Vitis vinifera*) cankers and trunk cankers and dieback of canyon live oak (*Quercus chrysolepis*) in California. *Plant Disease* 94, 785.
- Úrbez-Torres JR, Peduto F, Striegler RK, Urrearomero KE, Rupe JC, et al. 2012 – Characterization of fungal pathogens associated with grapevine trunk diseases in Arkansas and Missouri. *Fungal Diversity* 52, 169–189.

- Úrbez-Torres JR, Peduto F, Vossen PM, Krueger WH, Gubler WD. 2013 – Olive twig and branch dieback: etiology, incidence, and distribution in California. *Plant Disease* 97, 231–244.
- Valencia D, Torres C, Camps R, López E, Celisdiez JL, et al. 2015 – Dissemination of *Botryosphaeriaceae* conidia in vineyards in the semiarid Mediterranean climate of the Valparaiso Region of Chile. *Phytopathologia Mediterranea* 54, 394–402.
- van der Linde JA, Six DL, Wingfield MJ, Roux J. 2011 – *Lasiodiplodia* species associated with dying *Euphorbia ingens* in South Africa. *Southern Forests* 73, 165–173.
- Varga I, Poczai P, Cernak I, Hyvönen J. 2014 – Application of direct PCR in rapid rDNA ITS haplotype determination of the hyperparasitic fungus *Sphaeropsis visci* (*Botryosphaeriaceae*). *Springerplus* 3, 569.
- Vasic M, Duduk N, Vico, Ivanovic MS. 2013 – First Report of *Botryosphaeria dothidea* Causing white rot of apple fruit in Serbia. *Plant Disease* 97, 1659–1659.
- Verkley GJM, Aa HA van der. 1997 – *Endomelanconium microsporium*, a new coelomycete isolated from soil in Papua New Guinea. *Mycologia* 89, 967–970.
- Wang X, Li YX, Dong HX, Jia XZ, Zhang XY. 2015 – First Report of *Botryosphaeria dothidea* causing canker of *Acer platanoides* in China. *Plant Disease* 99, 1857–1857.
- Wet J de, Burgess T, Slippers B, Preisig O, Wingfield BD, et al. 2003 – Multiple gene genealogies and microsatellite markers reflect relationships between morphotypes of *Sphaeropsis sapinea* and distinguish a new species of *Diplodia*. *Mycological Research* 107, 557–566.
- Wet J de, Slippers B, Preisig O, Wingfield BD, Tsopelas P, et al. 2009 – Molecular and morphological characterization of *Dothiorella casuarinae* sp. nov. and other *Botryosphaeriaceae* with diplodia-like conidia. *Mycologia* 101, 503–511.
- Wikee S, Lombard L, Nakashima C, Motohashi K, Chukeatirote E, et al. 2013 – A phylogenetic re-evaluation of *Phyllosticta* (*Botryosphaeriales*). *Studies in Mycology* 76, 1–29.
- Wright AF, Harmon PF. 2010 – Identification of species in the *Botryosphaeriaceae* family causing stem blight on southern highbush blueberry in Florida. *Plant Disease* 94, 966–971.
- Xu C, Wang C, Ju L, Zhang R, Biggs AR, et al. 2015a – Multiple locus genealogies & phenotypic characters reappraise causal agents of apple ring rot in China. *Fungus Dive* 71, 215–231.
- Xu C, Zhang H, Zhou Z, Hu T, Wang S. 2015b – Identification and distribution of *Botryosphaeriaceae* species associated with blueberry stem blight in China. *European Journal of Plant Pathology* 143, 737–752.
- Yan JY, Xie Y, Zhang W, Wang, Liu JK, et al. 2013 – Species of *Botryosphaeriaceae* involved in grapevine dieback in China. *Fungal Diversity* 61, 221–236.
- Yan YC, Zhang ZX, Song YJ, Deng DF, Liu ZY. 2016 – First report of *Prunus serrulata* stem canker caused by *Botryosphaeria dothidea* in China. *Plant Disease* 100, 858–858.
- Zhai L, Zhang M, Lv G, Chen X, Jia N, et al. 2014 – Biological and molecular characterization of four *Botryosphaeria* species isolated from pear plants showing stem wart and stem canker in China. *Plant Disease* 98, 716–726.
- Zhang M, He W, Wu JR, Zhang Y. 2016 – Two new species of *Spencermartinsia* (*Botryosphaeriaceae*, *Botryosphaeriales*) from China. *Mycosphere* 7, 942–949.
- Zhang R, Guo X, Sun G, Tang M, Gleason ML. 2009 – *Dothiorella viticola* on *Populus cathayana* in China: a new record. *Mycotaxon* 109, 129–135.
- Zhou Y, Dou Z, He W, Zhang X, Zhang Y. 2016 – *Botryosphaeria sinensia* sp nov., a new species from China. *Phytotaxa* 245, 43–50.
- Zhou Y, Gong G, Cui Y, Zhang DX, Chang XL, et al. 2015 – Identification of *Botryosphaeriaceae* species causing kiwifruit rot in Sichuan Province, China. *Plant Disease* 99, 699–708.
- Zhu H, Qin WQ, Liu L, Yan W, Ca OCG. 2015 – First report of leaf spot of fishtail palm (*Caryota mitis*) caused by *Lasiodiplodia jatrophiicola* in China. *Plant Disease* 99, 1038–1038.
- Zlatkovic M, Keca N, Wingfield MJ, Jami F, Slippers B. 2016 – *Botryosphaeriaceae* associated with the die-back of ornamental trees in the Western Balkans. *Antonie Van Leeuwenhoek International of General Molecular Microbiology* 109, 543–564.