Novel Methods for Controlling Trichothecene Contamination of Grain and Improving the Climate Resilience of Food Safety and Security Programs



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Impact of Trichothecene Producing Cereal Pathogens

Food Safety

- Inhibition of protein synthesis
- Growth Retardation
- **Reproductive Disorders**
- Immunosuppression
- Feed Refusal Vomiting



Impact of Trichothecene Producing Cereal Pathogens

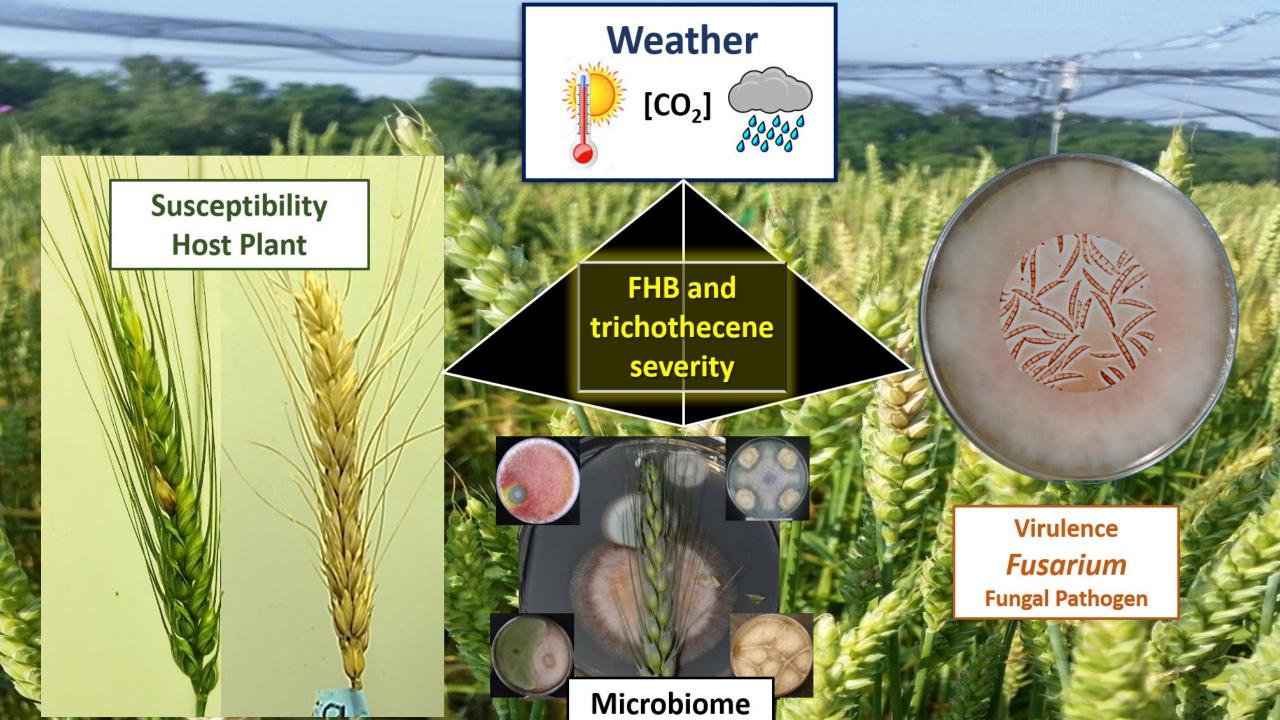
Food Security

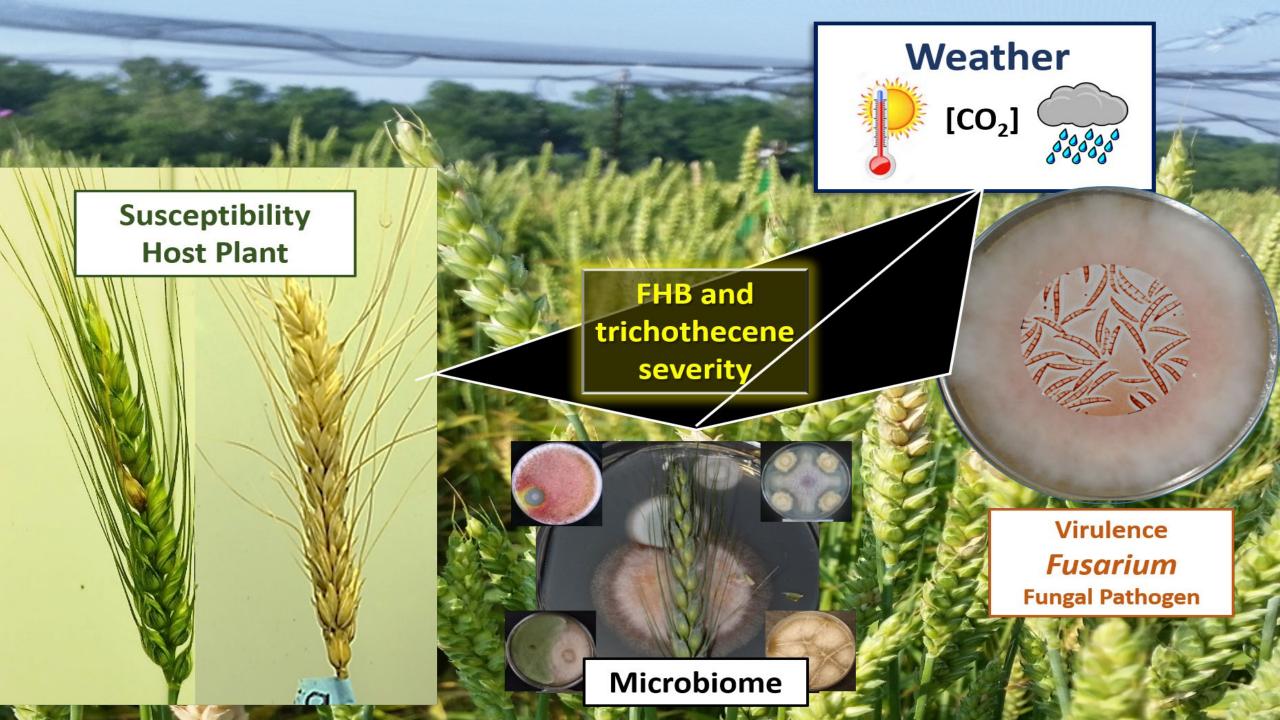
Value of yield forgone:

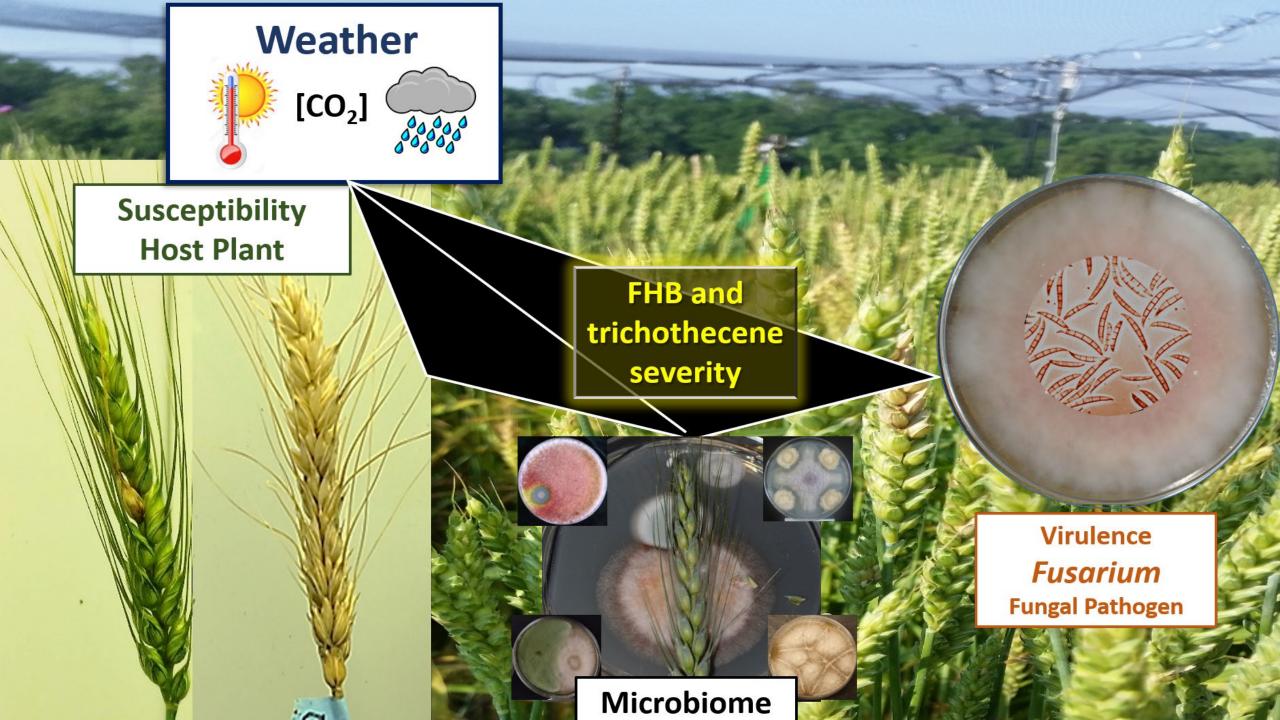
- Wheat:
- Barley:
- Corn:

\$1,176,000,000 \$293,000,000 \$500,000,000 ~ \$2 Billion Annually Loss

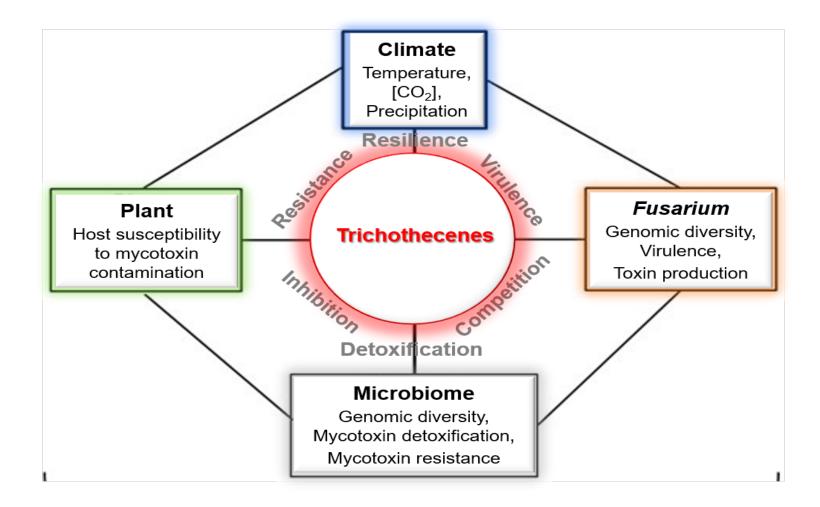
(Mueller et al., 2016; Wilson et al., 2018)



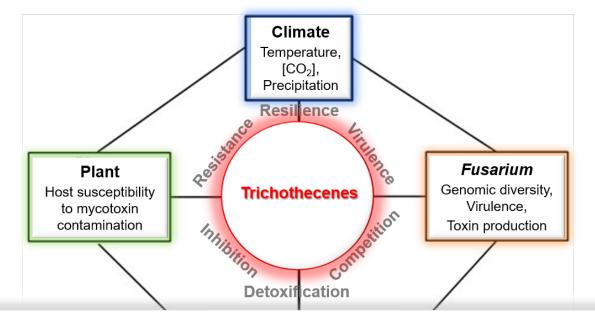




Holistic Ecological Approach to Understanding the Plant-Mycotoxin-Fungal Disease Triangle



Holistic Ecological Approach to Understanding the Plant-Mycotoxin-Fungal Disease Triangle (Microbiome)



Microbiome

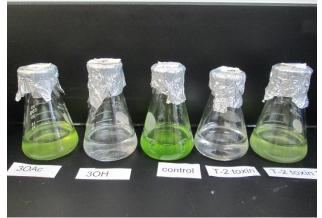
Objective: Identify and characterize microorganisms and microbial genes that can reduce trichothecene contamination of grain-based food and feed.

Trichothecene Resistance Mechanisms

Detoxification mechanisms of trichothecenes:

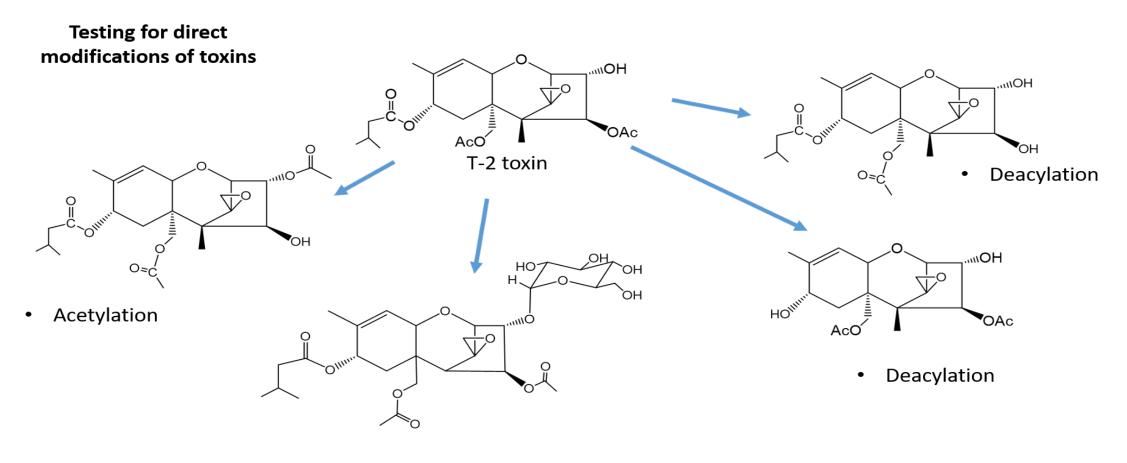
target key structural features tied to toxicity (do not degrade)

- Acetylation
- Glucosylation in plants (allows for sequestration) and also in yeast and fungi
- Deacetylation followed by glucosylation
- Deacylation
- Oxygenation (addition of hydroxyl group)
- Oxidative transformations (3-keto)



Assess phytotoxicity with Chlamydomonas

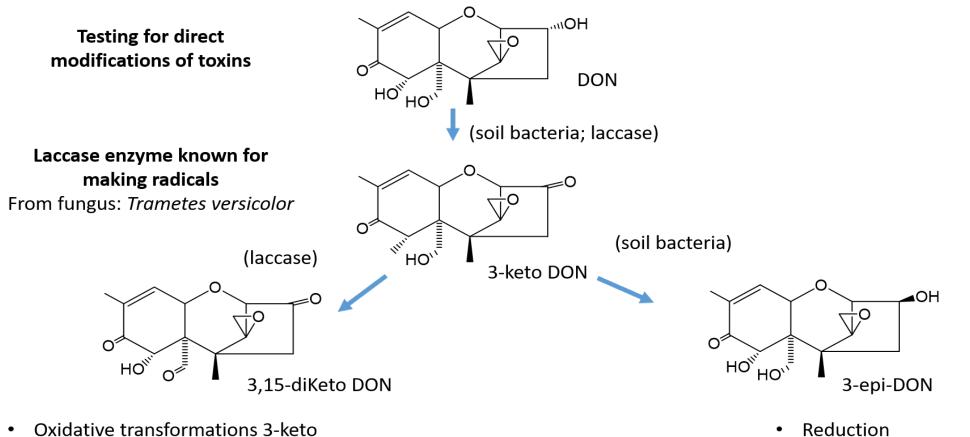
Detoxification of T-2 toxin by *Trichomonascus* and *Blastobotrys* yeast



• Glycosylation

(Kurtzman and McCormick)

Detoxification transformations of DON

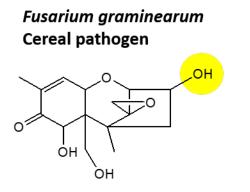


Oxidative transformations 3-keto ٠

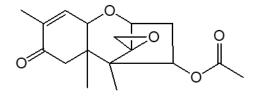
(Bakker and McCormick)

Search for novel trichothecene resistance mechanisms in fungi:

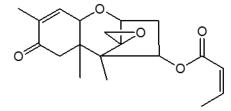
Absence of 3-hydroxyl in non-*Fusarium* trichothecenes indicate the producing fungi have a self-protection mechanism other than the 3-O-acetylation



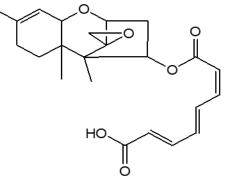
Microcyclospora tardicrescens Apple pathogen



Trichothecium roseum Saprotroph/plant pathogen

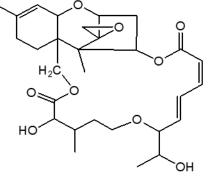


Trichoderma arundinaceum Saprotroph, biocontrol activity



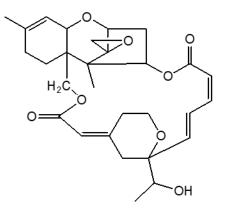


Melon pathogen



Myrothecium roridum

Stachybotrys chartarum Saprotroph, in damp buildings



(Proctor)

Examination of content of trichothecene biosynthetic (*TRI*) genes in trichothecene-producing fungi: some *TRI* genes are unique to *Fusarium*

Fungus	TRI Gene 1	TRI Gene 3	TRI Gene 4	TRI Gene 5	TRI Gene 6	TRI Gene 7	TRI Gene 8	TRI Gene 9	TRI Gene 10	TRI Gene 11	TRI Gene 12	TRI Gene 13	TRI Gene 14	TRI Gene 16	TRI Gene 17	TRI Gene 18	TRI Gene 21	TRI Gene 22	TRI Gene 23	TRI Gene 101
Fusarium graminearum						Ψ						Ψ		Ψ						
Fusarium longipes																				
Fusarium sporotrichioides																				
F. incarnatum-equiseti complex																				
Aspergillus hancockii																				
Beauveria bassiana																				
Cordyceps confragosa																				
Microcyclospora tardicrescens																				
Myrothecium roridum																				
Spicellum ovalisporum																				
Spicellum roseum																				
Stachybotrys chartarum																				
Stachybotrys chlorohalonata																				
Trichoderma brevicompactum																				
Trichoderma arundinaceum																				
Trichothecium roseum																		(5		+ 2 2019

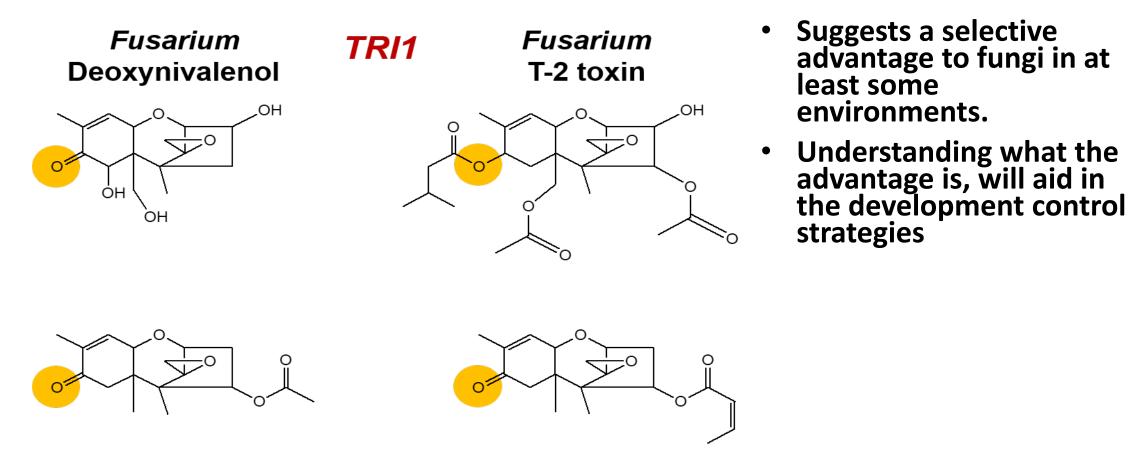
(Proctor et al., 2018)

TRI14 is present in all trichothecene-producing fungi, but is not required for biosynthesis: is TRI14 a resistance gene?

Fungus	TRI Gene 1	TRI Gene 3	TRI Gene 4	TRI Gene 5	TRI Gene 6	TRI Gene 7	TRI Gene 8	TRI Gene 9	TRI Gene 10	TRI Gene 11	TRI Gene 12	TRI Gene 13	TRI Gene 14	TRI Gene 16	TRI Gene 17	TRI Gene 18	TRI Gene 21	TRI Gene 22	TRI Gene 23	TRI Gene 101
Fusarium graminearum						Ψ						Ψ		Ψ						
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Trichoderma arundinaceum																				
Trichothecium roseum																				

Plan: heterologously express *TRI14* in yeast to determine whether it confers trichothecene resistance

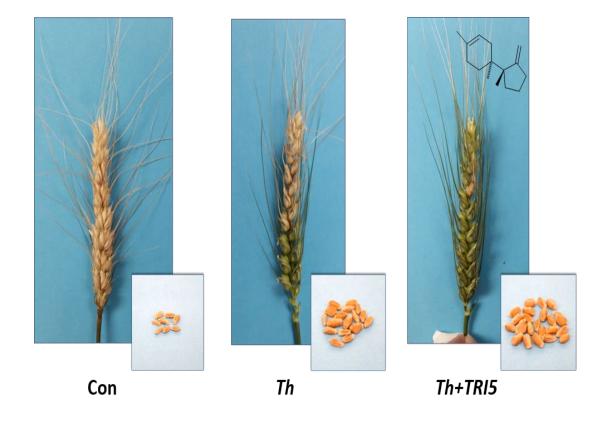
Trichothecene C8 oxygenation has evolved independently at least three times

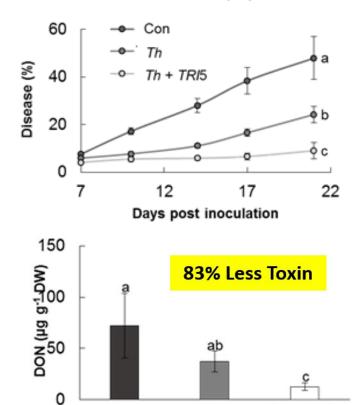


Trichothecium TRI? Microcylospora TRI24

(Proctor et al., 2018)

Trichoderma harzianum + *TRI5* emits the volatile trichodiene which stimulates plant defenses and down regulates DON production d





Th

Con

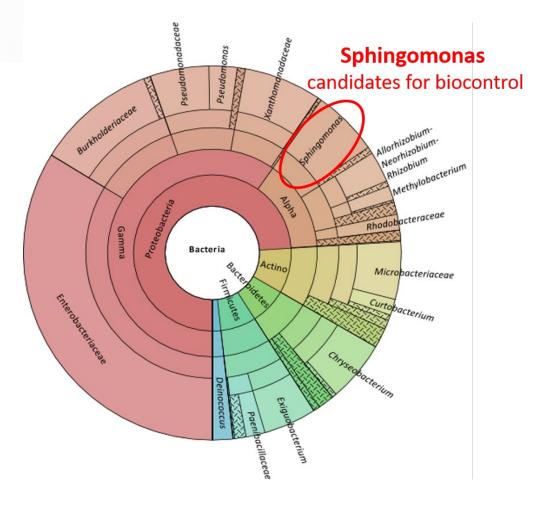


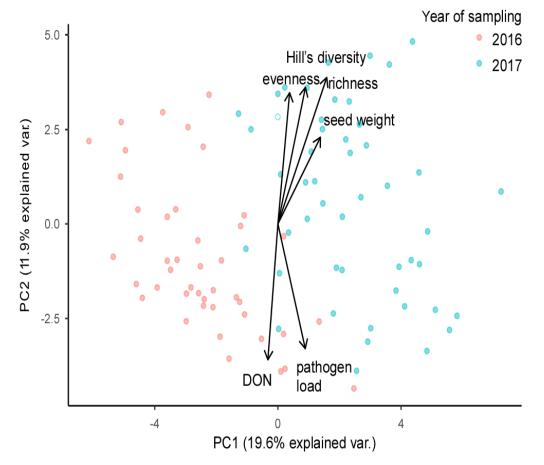
Con= Fg only

(Vaughan)

Th + TRI5

Microbiome characteristics relate significantly to toxin content and pathogen biomass within wheat kernels

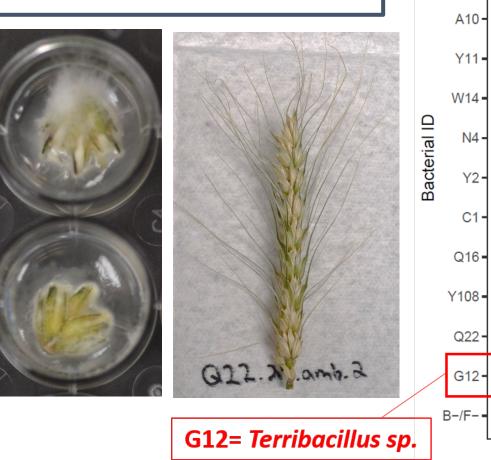


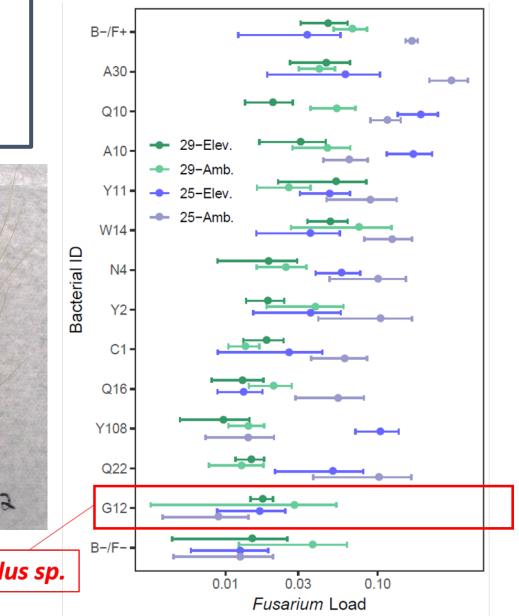


(Bakker and McCormick, 2019)

The efficacy of biocontrol microbes antagonistic to *Fusarium* varies with environmental conditions

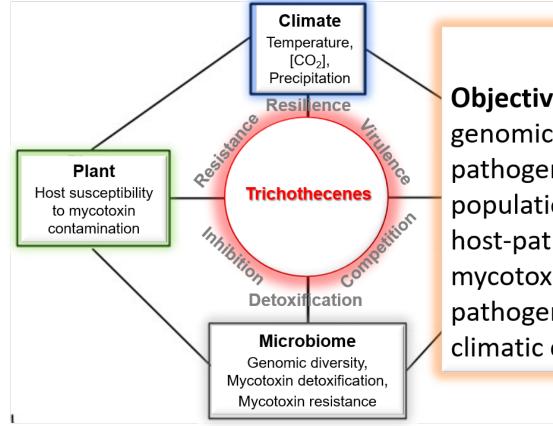






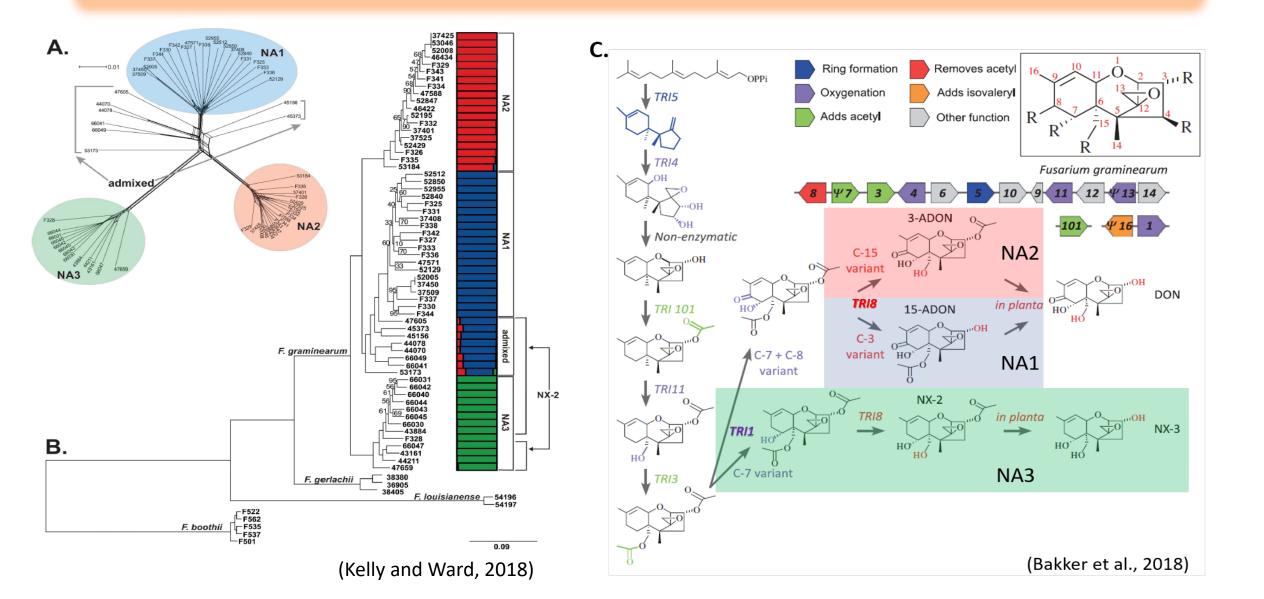
(Whitaker and Bakker, 2019)

Holistic Ecological Approach to Understanding the Plant-Mycotoxin-Fungal Disease Triangle (Fusarium)



Fusarium Objective: Determine the genomic diversity of FHB pathogens and identify population-specific differences in host-pathogen interactions, mycotoxin production, or pathogen fitness under different climatic conditions.

Three North American Populations of F. graminearum

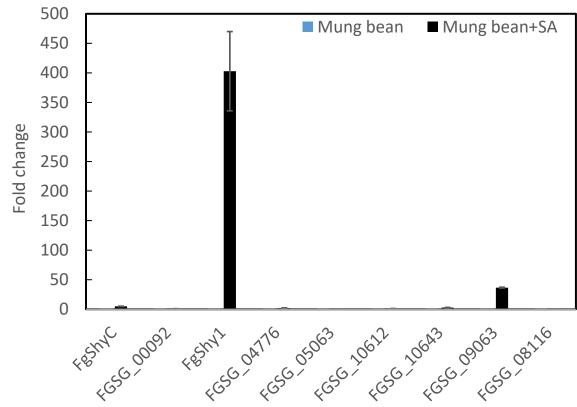


Genomic Signatures of Adaptive Divergence

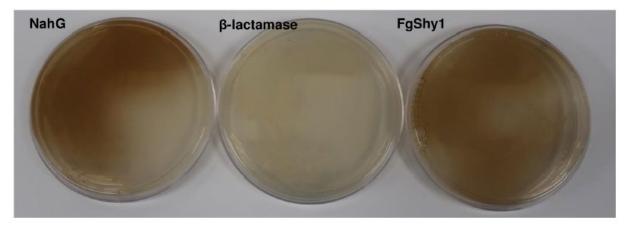
Plant Path (7 loci)	Antagonism (3 loci)	Ecological Adaptation (2 loci)				
Degradative enzymes	Heterokaryon incompatibility	Photolyase				
Adhesin	Chitinases	Cryptochrome				
Trichothecene toxin cluster	Ecp2 effector with a chitinase domain	Perithecial pigment (PKS1)				
	uomani	DEAD DNA/RNA helicase				
		Essential for adaptation to local climate conditions				

(Kelly and Ward, 2018)

Characterization of FgShy1 salicylate hydroxylase



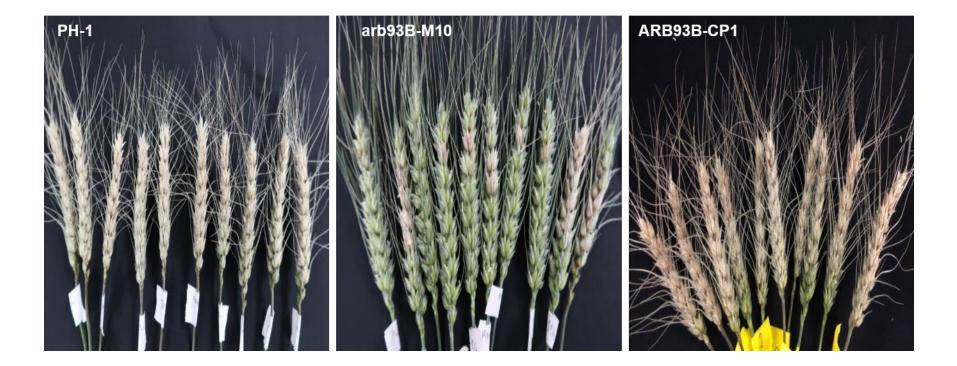
Analysis of salicylate hydroxylase candidates induction gene expression in mung bean liquid medium containing 1mM SA. FgShy1 is highly induced by addition of exogenous SA.



Salicylate Hydroxylase Activity Assay demonstrating salicylate hydroxylase activity of heterologously-expressed FgShy1 in *E.coli.* Development of brown color, indicating conversion of SA to catechol,

(Hao et al., 2019)

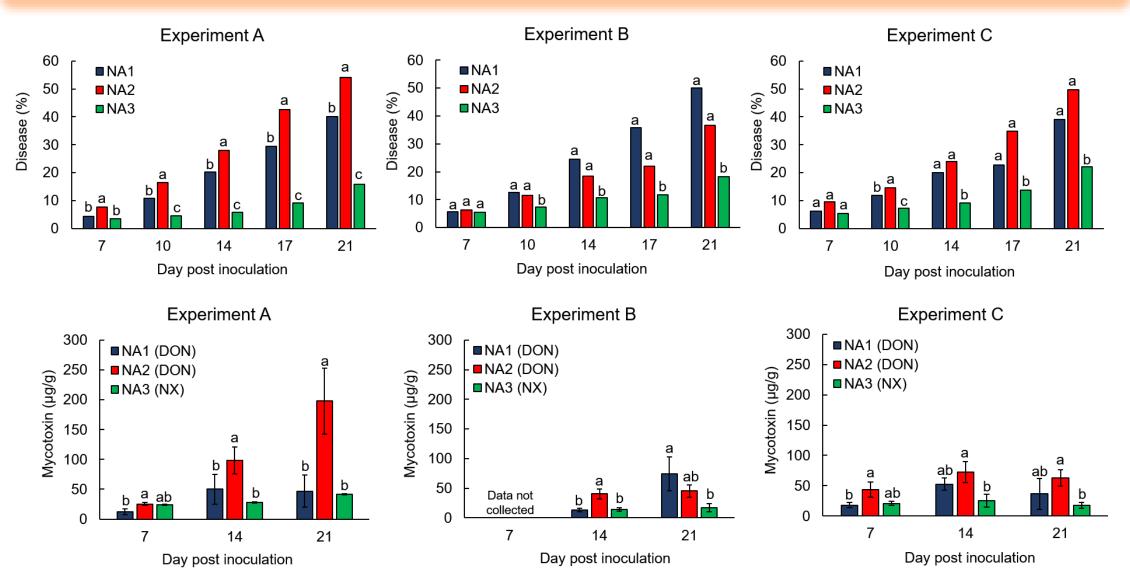
F. graminearum arabinanase (Arb93B) enhances wheat head blight susceptibility by suppressing plant immunity



F. graminearum arabinanase (Arb93B) mutant reduces FHB

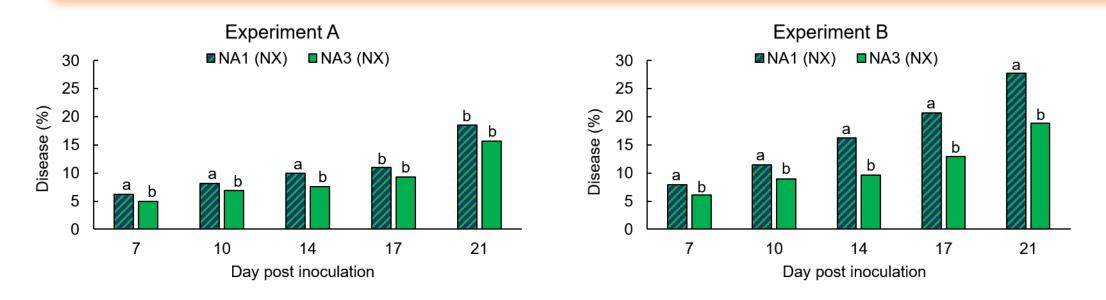
(Hao et al., 2019)

Population Specific Differences in Disease



(Vaughan and Ward et al.)

Population genetic background influences disease development



- NX toxin a contributing factor
- NA1 genetic background is more virulent than NA3

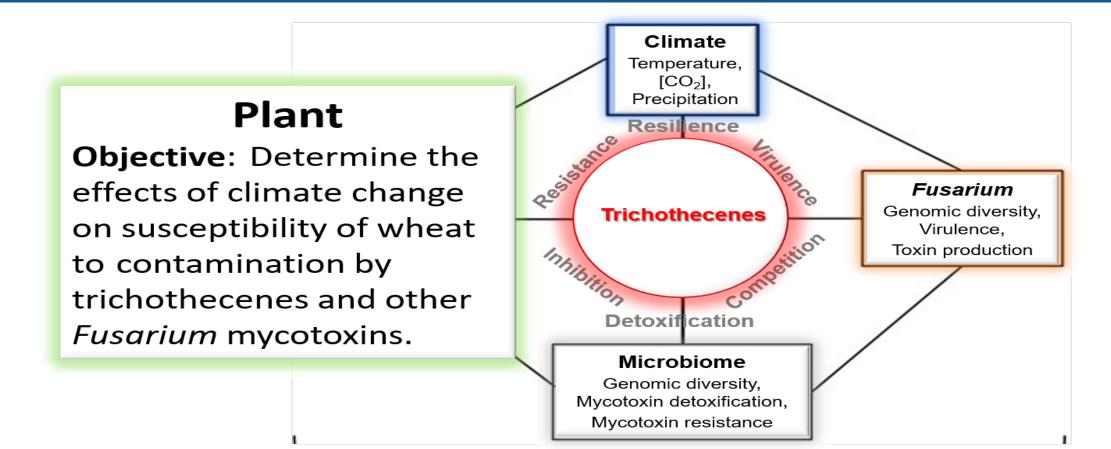
Hypothesis:

1) NA3 has an effector that stimulates plants defenses more strongly

2) NA1 has an effector that down regulates host defenses that NA3 is missing

(Vaughan and Ward et al.)

Holistic Ecological Approach to Understanding the Plant-Mycotoxin-Fungal Disease Triangle (Plant)



Effects of elevated CO₂ level on the metabolic response of resistant and susceptible wheat to *F. graminearum* infection

1x[CO₂] = 400 ppm



2x[CO₂] = 800 ppm

FHB susceptible and resistant Spring Wheat varieties:

- Norm and Alsen
- *F. graminearum* strains:
- 9F1 DON+
- Gz3639 DON+
- Gzt40 DON-

Dip inoculation method

Evaluated 7 pdi

Effects of elevated CO2 level on the metabolic response of resistant and susceptible wheat to F. graminearum infection

Gzt40

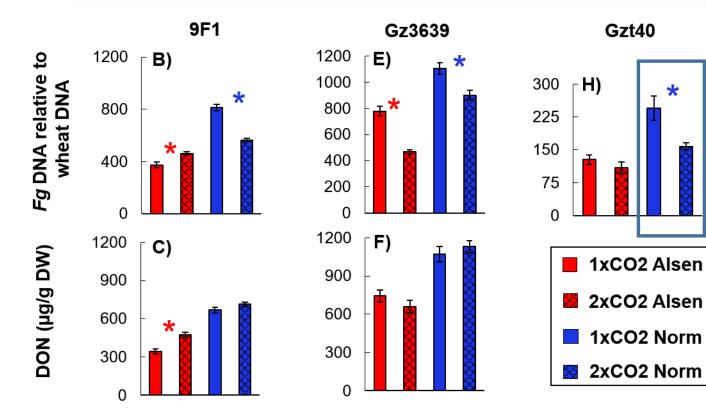
1xCO2 Alsen

2xCO2 Alsen

1xCO2 Norm

*

⊢H)



The effects of elevated CO₂ are dependent on both the *F. graminearum* strain and the wheat variety

Resistant variety Alsen:

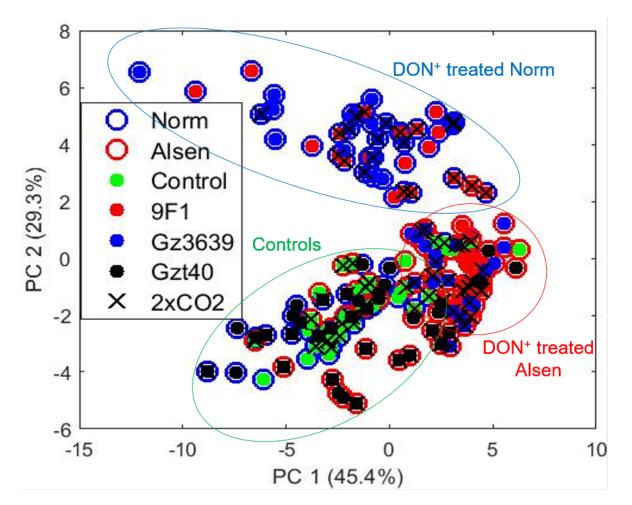
- Effect dependent on Fq strain
- Compromised resistance to 9F1 biomass and DON
- Enhanced resistance Gz3639 biomass but no change in DON

Susceptible variety Norm:

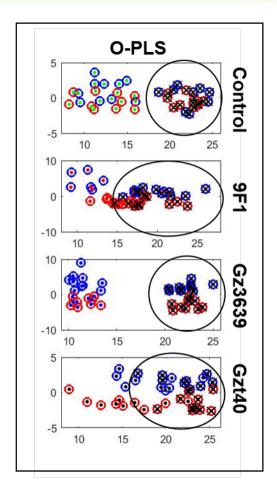
- enhanced resistance to pathogen biomass accumulation
- no change in DON
- DON production per unit *Fg* biomass was increased
- **Enhanced resistance not** dependent on DON

(Cuperlovic-Culf and Vaughan et al., 2018)

Non-Targeted Metabolomic Analysis

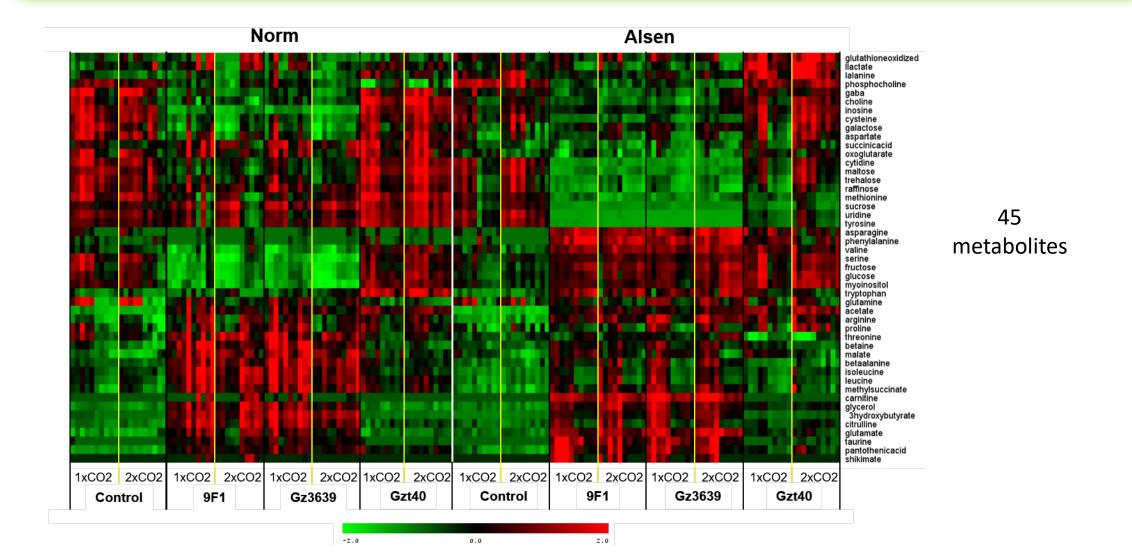


Principal component analysis (PCA) of 1D NMR spectra illustrated variances in the metabolic profiles across all samples



(Cuperlovic-Culf and Vaughan et al., 2018)

Relative Concentration of Targeted Metabolites

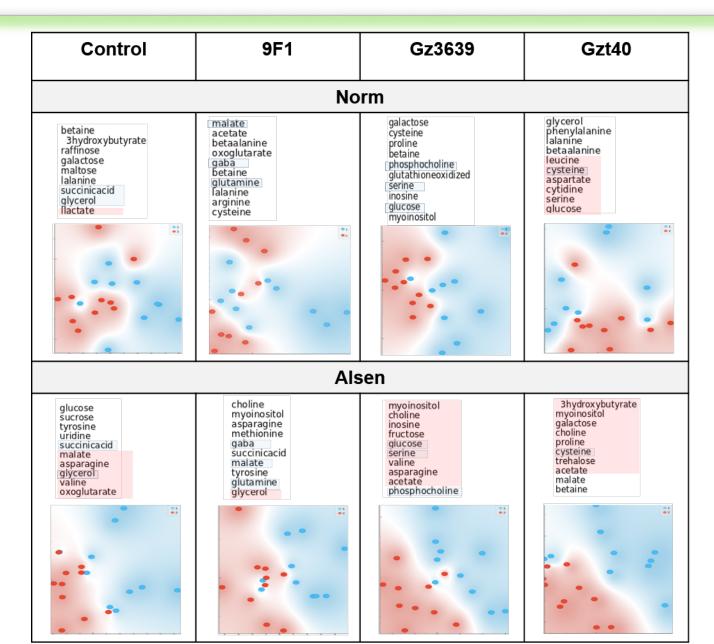


Metabolomic Analysis

- Metabolic pathways that contribute to the primary source of FHB resistance, are upregulated in *Fg* inoculated Alsen relative to Norm at both CO2 concentrations.
- Fhb loci containing varieties will likely remain more resistant to FHB than non Fhb-containing varieties even at elevated CO₂.
- Identification of FHB resistance markers that are not effected by CO₂ concentration differences
 - L-alanine
 - isoleucine
 - hydroxybutarate
 - myoinositol

(Cuperlovic-Culf and Vaughan et al., 2018)

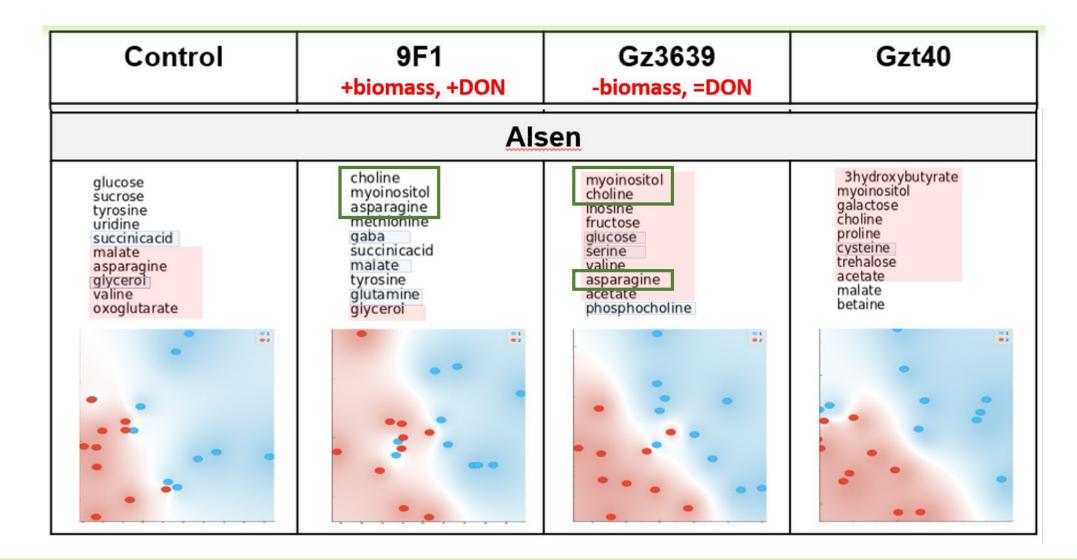
Metabolites with concentration differences at 1xCO2 vs 2xCO2



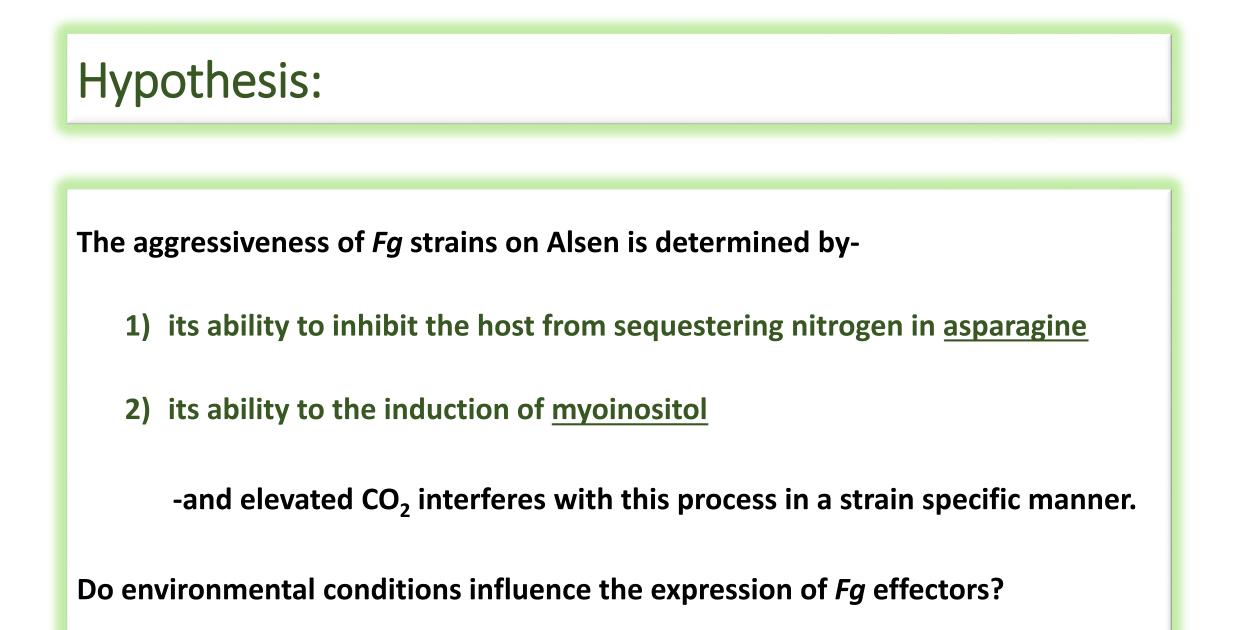
Metabolites with higher concentration in plants at $2xCO_2$ relative to $1xCO_2$

Metabolites that are significantly affected by CO_2 in both Norm and Alsen

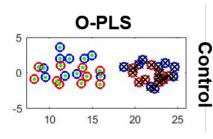
(Cuperlovic-Culf and Vaughan et al., 2018)



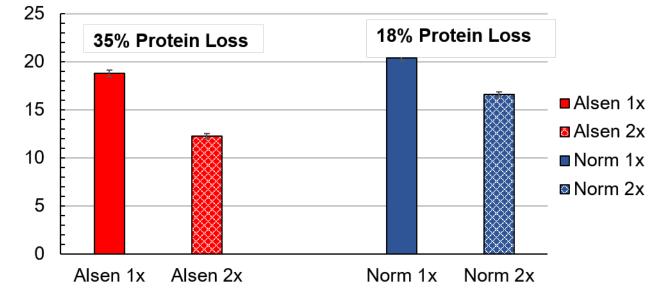
Difference in asparagine, myoinositol and choline are dependent on the infecting Fg strain and may serve as markers for enhanced FHB susceptibility at elevated CO₂



Growth at elevated CO₂ reduces grain protein content of FHB resistant variety significantly more than susceptible variety



Alsen vs. Norm acclimated at elevated [CO₂]

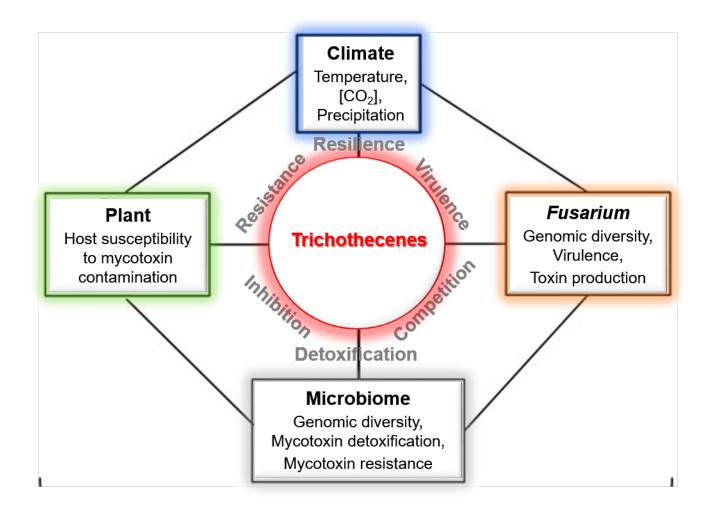


	% Moisture	% Fat	% Protein	% Carbohydrate	% Ash
Alsen T2 400 ppm CO2	$11.03^{\circ}\pm0.08$	$0.58^{AB}\pm\!0.15$	$18.80^{B}\pm0.32$	$77.10^{BC} \pm 1.28$	$2.36^{A} \pm 0.04$
Alsen T2 800 ppm CO2	$11.56^{\text{B}}\pm0.14$	$0.53^{\mathrm{B}}\pm0.10$	$12.26^{D} \pm 0.25$	$83.86^{A}\pm 0.88$	$2.04^{\circ}\pm 0.03$
Norm T2 400 ppm CO2	$12.97^{A}\pm0.13$	$0.87^{ m A} \pm 0.18$	$20.40^{A}\pm0.33$	$74.74^{\circ} \pm 1.82$	$2.53^{A}\pm0.18$
Norm T2 800 ppm CO2	$13.10^{\text{A}} \pm 0.06$	$0.76^{AB} \pm 0.12$	$16.58^{\circ} \pm 0.25$	$78.96^{\text{B}}\pm1.49$	$2.21^{B}\pm 0.05$
	•	•	•	•	(Hay and V/

(Hay and Vaughan)

Future Objectives

Continuation of Holistic Ecological Approach



Future Objectives (2)

Continuation of Holistic Ecological Approach

- **1.** Develop climate resilient control strategies of FHB and DON.
 - Determine the influence of environmental conditions and cultivars on *Fg* secondary metabolites and virulence factors
 - Identify climate resilient biocontrol microorganisms
- 2. Screen microbial communities and identify novel detoxification mechanisms of trichothecenes
- 3. Determine the impact of elevated CO₂ and other abiotic factors on the nutritional quality of FHB moderately resistant parent lines being used in breeding programs.
- 4. Identify the strain specific and variety specific differences that result changes in FHB and DON severity under conditions of abiotic stress
- 5. Evaluate differences in host metabolic responses to *Fg* populations

Future Objectives (3)

New Ideas:

Soliciting input directly from our Stakeholders



National Association of Wheat Growers



Acknowledgements:



Todd Ward



Susan McCormick



Bob Proctor

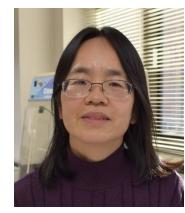


Matt Bakker



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Guixia Hao

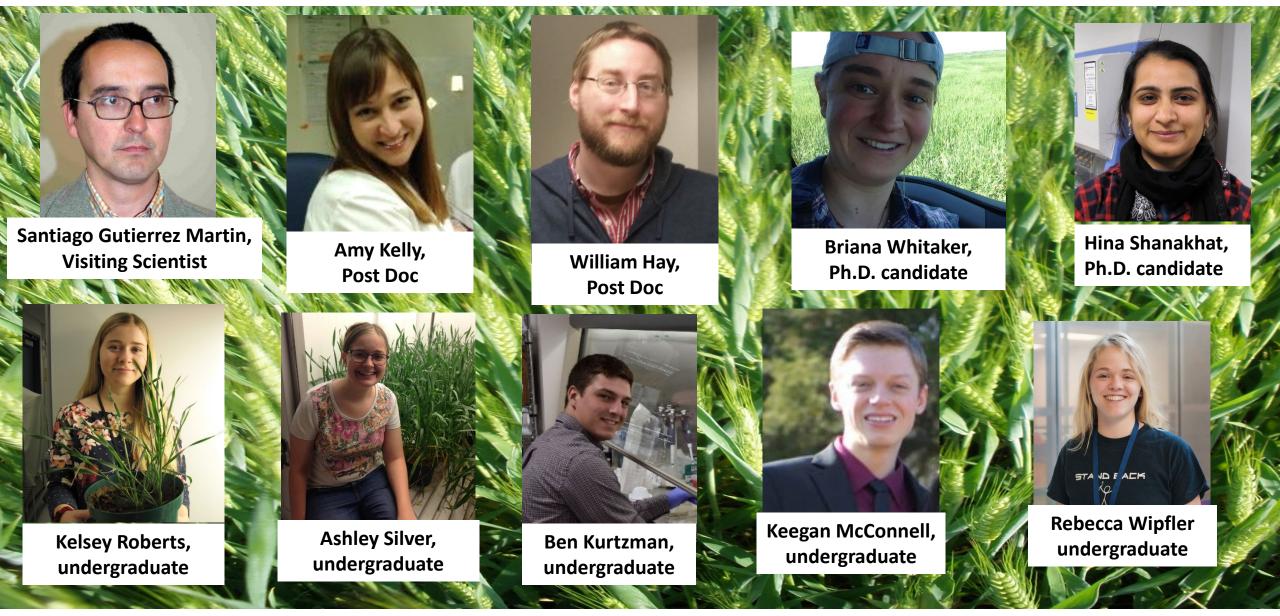


Cletus Kurtzman



Martha Vaughan

Acknowledgements (cont.):



Good things happen when we put our heads together