

ISSN (E): 2320-3862 ISSN (P): 2394-0530 NAAS Rating: 3.53 JMPS 2018; 6(5): 137-140 © 2018 JMPS Received: 21-07-2018 Accepted: 24-08-2018

Rafiqul Hasan Khan Department of Pharmacy, BRAC University, Dhaka, Bangladesh

Nura Ahmed Department of Pharmacy, BRAC University, Dhaka, Bangladesh

Amina Ferdous Chowdhury Department of Pharmacy, BRAC University, Dhaka, Bangladesh

Shakera Islam Keya Department of Pharmacy, BRAC University, Dhaka, Bangladesh

Mahbuba Mohoshina Runa Department of Pharmacy, BRAC University, Dhaka, Bangladesh.

Monika Nasrin Munni Department of Pharmacy, BRAC University, Dhaka, Bangladesh

Kazi Nuruddin Al Masud Department of Pharmacy, BRAC University, Dhaka, Bangladesh

Correspondence Kazi Nuruddin Al Masud Department of Pharmacy, BRAC University, Dhaka, Bangladesh

Phytochemical screening of Goniothalamus sesquipedalis

Rafiqul Hasan Khan, Nura Ahmed, Amina Ferdous Chowdhury, Shakera Islam Keya, Mahbuba Mohoshina Runa, Monika Nasrin Munni and Kazi Nuruddin Al Masud

Abstract

This study was conducted to perform the phytochemical screening of *Goniothalamus sesquipedalis*. The plant belongs to the Annonaceae family and locally used analgesic agent. This study provides a scientific basis for the use of *Goniothalamus sesquipedalis* in traditional medicine. The plant was extracted using Ethanol. The extract was subjected to phytochemical screening. The phytochemical screening of *Goniothalamus sesquipedalis* seeds showed the presence of alkaloids, flavonoids, tannins, phlobatannin, glycosides, sterol and tannins but showed absence of quinines, phenol and resin.

Keywords: methanol, Goniothalamus sesquipedalis, phytochemical screening

Introduction

Medicinal plants are an important part of traditional medicines. Traditional medicines are a set of knowledge, practices and techniques. Indeed, indigenous theory, experience and belief in diverse cultures can be used to diagnose and prevent other diseases in addition to maintaining health. Traditional medicines use leaves or roots for various diseases. The main source of 87% of medicines used for all kinds of human diseases in nature. About 25% of Medicaid Drug Approximately 80% of people in developing countries rely on traditional medicines for their well-being ^[1]. In this study, we focus on traditional plants used in Bangladesh, such as antioxidants, antimicrobials and cytotoxic agents to find new sources of medicines for effective and effective treatment of the disease. We discovered the unknown pharmacological effects of these plants. Phytochemical investigations show that 15% ^[2]. Based on the several activities provided by the other species of *Goniothalamus* genus, *Goniothalamus sesquipedalis* plant has been chosen to perform the phytochemical screening.

Materials and Methods

Plant collection and identification

The seeds of *Goniothalamus sesquipedalis* was collected in the month of October 2017 from Chittagong, Bangladesh. After that, its verification (Verification code number: 42930) was done by the National Herbarium of Bangladesh (NHB), Mirpur, Dhaka by submitting plant sample.

Chemicals

Potassium mercuric iodide, Iodine in potassium iodide, Dragendorff's reagent, Saturated picric acid, Tannic acid, Neutral ferric chloride, Sodium nitroprusside, Glacial acetic acid, Concentrated H₂SO₄, Molise's reagents, CuSO₄, Alcoholic KOH, Basic lead acetate, Acetic anhydride, NaOH, Chloroform, NaNO₂, Methanol, Folin-Ciocalteu reagent, Na₂CO₃ were used.

Preparation of plant extract

After the washing the leaves with clean water the leaves were shade dried for several days and were grounded finely as a granular particle with a high power grinding machine. About 400gm of grounded leave powder of *Goniothalamus sesquipedalis* which was drenched in 2L of ethanol for 14 days period in a room temperature (22-25°C) with random agitation. After 14 days of soaking, the substances of the bottle were emptied out first to filter them by using

Whitman filter paper (pore size 100nm). The filtrate was concentrated by using rotary evaporator (Heidolph) at 30°C temperature with a rotation speed of 100rpm up to form the concentrated Ethanolic extract.

Phytochemical screening

The crude extract of *Goniothalamus sesquipedalis* was used for phytochemical screening to identify its chemical compound present in its leaves.

Procedure of extract preparation for screening

2-3 grams of dried ethanol extract was mixed with 50ml ethanol in a 100ml of conical flask. After that, that flask was labeled properly closing with cotton plugs and kept still for 1 to 2 hours. Later, the mixture was filtered through Whitman filter paper. Collected filtrates were used for phytochemical screening by following the standard process ^[6, 7]. The following qualitative tests were performed sequentially:

Tests for Alkaloids

Mayer's test

A few drops of Mayer's reagent (Potassium mercuric iodide solution) was added in 1ml of seed extract. If cream color precipitation form then it will contain the presence of alkaloids.

Wagner's test

In 1ml of seed extract was added with the same amount of Wagner's reagent (Iodine in potassium iodide). If reddish brown color precipitation form then it will point to the presence of alkaloids.

Dragendorff's reagent test

2ml of Dragendorff's reagent was added in 1ml of seed extract and later dilute HCL of 2ml was added in that solution. If orange color precipitation forms then it will confirm the presence of alkaloids.

Hager's test

A few drops of Hager's reagent (Saturated picric acid solution) was added in 2ml of seed extract. If bright yellow color precipitation form then it will point to the presence of alkaloids.

Tannic acid test

A few drops of tannic acids was added in 1ml of seed extract. If yellow-brown colored precipitation form then it will point to the presence of alkaloids.

FeCl₃ test

About 1-2 ml extract was mixed with a little amount of neutral ferric chloride solution in drop wise. If cream yellow precipitation forms then it will point to the presence of alkaloids.

Tests for glycosides

Legal's Test

Addition of alkaline sodium nitroprusside and pyridine in extract solution results in the formation of cherry red color then it will confirm to the presence of glycosides.

Keller Mililani test

At first 1ml of glacial acetic acid was mixed-up with 1 ml of extract and cooled. After that 2-3 drops of ferric chloride was mixed and 2ml of concentrated H_2SO_4 was added carefully in

sideways of test tube walls. If reddish brown colored ring at the junction of two layers form then it will point to the presence of glycosides.

Concentrated H₂SO₄ test

1ml of Concentrated H_2SO_4 was added in 1ml of seed extract and kept still for 2 minutes. If reddish color precipitate form then it will point to the presence of glycosides.

Molise's test

In seed extract around 2-3 drops of Molise's reagents was added. Later, a few drops of concentrated H_2SO_4 was added properly. If reddish purple colored ring at the junction of two layers form then it will point to the presence of glycosides.

Test for phlobatannin

At first 2-3ml of 10% HCl was added in 10ml of seed extract in a boiling test tube which was boiled for 5-6 minutes. If red color precipitate occurs then it will point to the presence of phlobatannin.

Test for resins

3-4ml of the CuSO₄ solution was mixed-up with seed extract which was shaken vigorously for 1-2 minutes and allowed to discrete. If green color precipitate occurs then it will point to the presence of resins.

Test for quinines

Alcoholic KOH solution was added in seed extract. If color ranging from red to blue occur then it will point to the presence of quinines.

Test for Saponin

In the test tube 5ml of the extract was taken and shake vigorously to get a stable froth. 5-6 drops of olive oil were added into frothing solution. If the emulsion is formed then it will point to the presence of saponin.

Tests for phenols

Pelagic acid test

A few drops of 5% (w/v) glacial acetic acid was added in seed extract. After that 5% (w/v) NaNO₂ solution was added. If muddy brown color form then it will point to the presence of phenols.

Phenol test

1ml of the FeCl₃ solution was added in 2ml of seed extract. If the development of intense color form then it will point to the presence of phenols.

Tests for Tannins

Ferric chloride test

A few drops of $FeCl_3$ was added in seed extract. If blackish color precipitate form then it will point to the presence of tannins.

Lead acetate test

A few drops of basic lead acetate was added in 1-2ml of seed extract. If bulky red color precipitate form then it will point to the presence of tannins.

Alkaline reagent test

A few drops of sodium hydroxide solution was added in seed extract. If red color form then it will point to the presence of tannins.

Tests for flavonoids Lead-acetate test

A few drops of basic lead acetate solution was added in 1-2ml of seed extract. If reddish brown color precipitate form then it will point to the presence of flavonoids.

FeCl₃ test

A few drops of neutral ferric chloride solution was added in 1-2ml of seed extract. If the deposition of blackish red color precipitate form then it will point to the presence of flavonoids.

Alkaline reagent test

A few drops of sodium hydroxide was added in 1-2ml of seed extract. If yellowish red color occurs then it will point to the presence of flavonoids.

Test for sterols

Libermann-Burchard test

A few drops of acetic anhydride solution was mixed with 1-2ml of seed extract. After that, a few drops of concentrated H_2SO_4 was given beside the test tube walls in the mixture. If reddish brown color ring at the junction of two layers occur then it will point to the presence of sterols.

Salkowski test

5ml of chloroform was added in 1-2ml of seed extract. After that, 1ml of concentrated H_2SO_4 was put beside the test tube walls. If the reddish color in the lower layer occurs then it will point to the presence of sterols.

Result and Discussion

Phytochemical screening

Serial Number	Class of compound	Result
1	Alkaloid	+
2	Glycoside	+
3	Phlobatannin	++++
4	Resin	-
5	Quinone	-
6	Phenol	-
7	Tannin	+++++
8	Flavonoids	++
9	Sterol	+++

 Table 1: Phytochemical screening of Goniothalamus sesquipedalis

 Leaf

Note: (+) = presence in a single method test, (++) = presence experimented in two methods, (+++) = presence experimented in three methods, (++++) = presence experimented in four methods, (+++++) = presence experimented in five methods and (-) = absence.

Discussion

The phytochemical screening of *Goniothalamus sesquipedalis* showed the presence of alkaloids, flavonoids, glycosides, phlobatannin, sterol and tannins whereas showing the absence of Quinone, phenol and resin.

Conclusion

The phytochemical analysis showed the presence of several phytochemicals which can be further isolated using compound isolation. The Phytochemical analysis showed *Goniothalamus sesquipedalis* has different phytochemicals which can be isolated and study in the further experiments.

References

1 Khatun A, Rahman M, Haque T, Rahman Md. M, Akter

M, Akter S, *et al.* Cytotoxicity Potentials of Eleven Bangladeshi Medicinal Plants. The Scientific World Journal, 2014, 1-7. doi:http://dx.doi.org/10.1155/2014/ 913127.

- 2 Verpoorte R. Pharmacognosy in the new millennium: lead finding and biotechnology. J Pharm Pharmacol. 2000; 52:253-262.
- 3 Ahmed F, Rahman S, Ahmed N, Hossain M, Biswas A, Sarkar S, *et al.* Evaluation of *Neolamarckia cadamba* (Roxb.) Bosser leaf extract on glucose tolerance in glucose-induced hyperglycemic mice. Afr J Tradit Complement Altern Med. 2011; 8(1):79-81. Retrieved from https://www.ncbi.nlm.nih.gov/pubmed/22238487
- 4 Keya MA, Rahman AHMM. Angiosperm Diversity at the Village Sabgram of Bogra, Bangladesh with Emphasis on Medicinal Plants. American Journal of Plant Biology. 2017; 2(1):25-34.
- 5 Huq AKMM, Jamal JA, Stanslas J. Ethnobotanical, Phytochemical, Pharmacological, and Toxicological Aspects of *Persicaria hydropiper* (L.) Delarbre. PMC. US National Library of Medicine National Institutes of Health, 2014.
- 6 Kokate CK. Practical Pharmacognosy. (4th Ed). New Delhi, India: Vallabh Prakashan Publication, 1999.
- 7 Evans WC. Trease and Evans Pharmacognosy. (14th Ed.). Singapore: Harcourt Brace and company, Asia Pvt. Ltd, 1997.
- 8 Pietta A, Sionetti P, Mauri P. Antioxidant activity of selected medicinal plants. Agric Food Chem. 1998; 46:4487-4490.
- 9 Velioglu YS, Mazza G, Gao YL, Oomah BD. Antioxidant activity and totalphenolics in selected fruits, vegetables and grain products. J Agric Food Chem. 1998; 46:4113-4117.
- 10 Harbertson JF, Spayd S. Measuring Phenolics in the Winery. American Journal of Enology and Viticulture, 2006, 57(3).
- 11 Skerget M, Kotnik P, Hadolin M, Hras A, Simonic M, Knez Z. Phenols, proanthocyanidins, flavones and flavonols in some plant materials and their antioxidant activities. Food chemistry. 2005; 89:191-198.
- 12 Majhenik, *et al.* Antioxidant and antimicrobial activity of guarana seed extracts, Food chemistry. 2007; 10:1016.
- 13 Bozin B, Mimica-Dukic N, Samojlik I, Goran A, Igic R. Phenolics as antioxidants in garlic (Allium sativum L., Alliaceae). Food Chem. 2008; 111:925-929.
- 14 Karagözler AA, Erdag B, Emek YC, Uygum DA. Antioxidant activity and proline content of leaf extracts from Dorystoechas hastata. Food Chem. 2008; 111:400-407.
- 15 Brand-Williams W, Cuvelier ME, Berset C. Use of a free radical method to evaluate antioxidant activity. LWT -Food Science and Technology. 1995; 28(1):25–30. https: //doi.org/10.1016/S0023-6438(95)80008-5
- 16 Choi SJ, Kim JK, Kim HK, Harris K, Kim CJ, et al. 2,4-Di-tert-butylphenol from sweet potato protects against oxidative stress in PC12 cells and in mice. Journal of Medicinal Food. 2013; 16(11):977–83. https://doi.org/10. 1089/jmf.2012.2739
- 17 Desmarchelier C, Mongelli E, Coussio J, Ciccia G. Evaluation of the *in vitro* antioxidant activity in extracts of *Uncaria tomentosa* (Willd.) DC. Phytotherapy Research. 1997; 11(3):254–256. https://doi.org/10.1002/ (SICI)1099-1573(199705)11:3<254::AID-PTR76>3.0.CO;2-5

Journal of Medicinal Plants Studies

- 18 Bondet V, Brand-Williams W, Berset C. Kinetics and Mechanisms of Antioxidant Activity using the DPPH. Free Radical Method. LWT - Food Science and Technology. 1997; 30(6):609-615. https://doi.org/10. 1006/FSTL.1997.0240
- 19 Pandey G. Antioxidant and Antibacterial Activities of Leaf Extract of Achyranthes aspera Linn. (Prickly Chaff Flower). European Journal of Medicinal Plants. 2014; 4:695-708. https://doi.org/10.9734/EJMP/2014/8786.
- 20 Beaulah AG, Sadiq MA, Santhi JR. Antioxidant and Antibacterial activity of Achyranthes Aspera: An *in vitro* study. Der Pharma Chemica. 2011; 3(5):255-262. Retrieved from http://derpharmachemica.com/archive. html
- 21 Rishikesh. Phytochemical and Pharmacological Investigation of *Achyranthes Aspera* Linn. Scholars Academic Journal of Pharmacy. 2013; 2:74-80.
- 22 Priya CL, Kumar G, Loganathan K, Rao B. Antioxidant Activity of *Achyranthes Aspera* linn stem Extracts, Pharmacology online. 2010; 2:228-237.
- 23 Benariba N, Djaziri R, Bellakhdar W, Belkacem N, Kadiata M, *et al.* Phytochemical screening and free radical scavenging activity of Citrullus colocynthis seeds extracts. Asian Pacific Journal of Tropical Biomedicine. 2013; 3(1):35-40. http://doi.org/10.1016/S22211691(13) 60020-9.