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Trypanocide activity of *Castela coccinea* Griseb. extracts

[Actividad tripanocida de extractos de *Castela coccinea* Griseb.]

María L. MARTÍNEZ^{a*}, María NOEL CAMPAGNA^a, María S. RATTI^a, Isabel NOCITO^b, Esteban SERRA^b, Susana GATTUSO^a
and Martha A. GATTUSO^{a*}

^aFarmacobotánica. Fac. Cs. Bioq. y Farm. UNR. Rosario, Argentina. ^bParasitología. Fac. Cs. Bioq. y Farm. UNR. Rosario, Argentina.

Abstract

Castela coccinea Griseb is a small tree that belongs to Simaroubaceae family. This work shows that this plant extracts present *in vitro* a strong activity against *Trypanosoma cruzi* epimastigotes. Particularly, wood dichloromethane extract displayed the most intensive trypanocide effect. A fraction of this extract, purified by column chromatography over Silica-gel, was enriched in coumarins, phenolic acids and alkaloids. This fraction was even more effective with an IC₅₀ of 15 µg/mL. Our results suggest that alkaloids could be the responsible compounds for this activity. The 8-amino-6-methoxyquinoline group was detected by GC-MS, even though the whole molecule could not be identified yet. On the other hand, all extracts have exhibited cytotoxicity against the human cell line K-562, at high concentrations. However, the selectivity index of wood dichloromethane extract has been 7.5.

Keywords: *Castela*; Simaroubaceae; Trypanocide activity; Cytotoxicity.

Resumen

Castela coccinea Griseb es un árbol de pequeño porte que pertenece a la familia Simaroubaceae. En el presente trabajo los extractos de esta planta muestran una fuerte actividad *in vitro* contra epimastigotes de *Trypanosoma cruzi*. El extracto diclorometánico de madera es el que presenta mayor efecto tripanocida. Una fracción de este extracto, purificada por cromatografía en columna, está enriquecida en cumarinas, ácidos fenólicos y alcaloides. Esta fracción es aún más activa con una CI₅₀ = 15 µg/mL. Nuestros resultados sugieren que los alcaloides son los responsables de esta actividad. Más aún, se ha detectado la presencia del grupo 8-amino-6-metoxiquinolona por cromatografía gaseosa acoplada a un espectrómetro de masa, pero todavía la molécula entera no ha sido identificada. Por otro lado, todos los extractos mostraron actividad citotóxica contra la línea celular humana K-562, a altas concentraciones. Sin embargo, el índice de selectividad para el extracto diclorometánico de la madera es de 7.5.

Palabras Clave: *Castela*; Simaroubaceae; Actividad tripanocida; Citotoxicidad.

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*Contactos | Contacts: Facultad Cs. Bioquímicas y Farmacéuticas. Suipacha 531 – S2002LRK – Rosario, Argentina. E-mails: mlmartin@fbioyf.unr.edu.ar; marthagattuso@fibertel.com.ar



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INTRODUCTION

Castela coccinea Griseb. mainly grows in South America and belongs to the Simaroubaceae family. This family has played a prominent role in folk medicine because of its anthelmintic and antiamoebic properties (Polonsky, 1973). Chemical examination of these plants has been carried out by several groups resulting in the isolation of quassinoids (Ogura et al., 1977; Grieco et al., 1999; Jiwajinda et al., 2001; Jacobs et al., 2007), alkaloids (Sanchez and Comin, 1971; Ogura et al., 1978; Ouyang et al., 1995; Readell et al., 2003) and terpenoids (Sherman et al., 1980). In recent years, attention has been focused on quassinoids, bitter principles of the family, as several of them have shown promising antitumor, antiviral, antimalarial, antileukemic and antifeedant properties (Polonsky, 1985).

Bourdy et al. has demonstrated that leaf extracts of *Castela coccinea* Griseb display strong activity against *Plasmodium falciparum*, which is the etiological agent of the most severe form of malaria (Bourdy et al., 2004). Malaria has low incidence in South America. On the other hand, South America is the endemic zone for Chagas, another parasitic disease caused by *Trypanosoma cruzi*. Although the incidence of Chagas has declined over the past 20 years mainly due to the efforts made towards the control and elimination of the vector, American trypanosomiasis remains a sickness difficult to eradicate (Moncayo, 2003). Current therapy is based on two drugs, nifurtimox and benznidazole, which have been proved effective against the acute phase of the disease. Otherwise, their effectiveness in the chronic stage is controversial. Additionally, serious side effects have been reported in about 50 % of treated patients (Castro et al., 2006). Therefore, it still remains a compelling need to develop safer and more efficient drugs, as well as new approaches to overcome the drawbacks of this chemotherapy strategy.

The aim of this work is to examine anti-trypanosomal activity of *Castela coccinea* Griseb extracts. Cell viability was also evaluated after treating cells with different extracts of *Castela coccinea* Griseb to investigate the behaviour in biological systems, because it is well-known that many Simaroubaceae plants show cytotoxic activity (Among others, Cuendet and Pezzutto, 2004; Guo et al., 2005; Lopez Saez et al., 2008).

MATERIALS AND METHODS

Plant Material

Castela coccinea Griseb. (Simaroubaceae) was collected from Argentina: Santa Fe Province, General Department Obligado, Berna Locality in December 2004 and October 2007 and was authenticated by Luis Oakley, Departamento de Biología y Recursos Naturales, Universidad Nacional de Rosario (UNR). A voucher specimen was deposited in the Herbarium of SF referring Pensiero 6943 to material collected in 2004 and in the Herbarium of UNR referring Oakley et al. 053 to material collected in 2007.

Chemicals

1,1-diphenyl-2-picryldrazyl (DPPH), dimethyl sulfoxide (DMSO), 2-aminoethyl diphenylborinate and anisaldehyde were purchased from Sigma Chemicals, St. Louis, MO, USA. The culture media Eagle's minimum essential medium (MEM), Dulbecco's modified Eagle's medium (DMEM), trypsin, penicillin G, streptomycin were purchased from Gibco BRL (Gaithersburg, MD, USA). All other chemicals and reagents used were of the highest commercially available purity.

Preparation of extracts and partial purification

The bark (100 g) and the wood (100 g) of *Castela coccinea* Griseb were extracted three times with ethanol (EtOH) or dichloromethane (DCM) at room temperature and then concentrated under reduce pressure yielding the residues: **Bark_{EtOH}** (2.0 g), **Wood_{EtOH}** (2.0 g), **Bark_{DCM}** (1.24 g) and **Wood_{DCM}** (0.93 g). Each of them was resuspended in the corresponding solvent or in DMSO.

Wood_{DCM} extract was subjected to chromatography on a Silica gel-60 column (40 cm length, 2 cm i.d.) equilibrated with hexane and eluted with a stepwise gradient of hexane and ethyl acetate. 90 fractions of 5 mL each were collected and analyzed by TLC aluminium sheets. On the basis of the TLC profile the similar fractions were pooled into 5 fractions: **I**, **II**, **III**, **IV** and **V**.

Trypanocide assay

Epimastigotes from CL Brener and Y strains were cultured in modified liver infusion tryptose (LIT) medium supplemented with 10% fetal bovine serum (FBS) at 28 ± 0.5 °C in 24 wells culture flasks (Camargo, 1964). Cultures ($3 - 4 \times 10^6$ parasites/mL)

where incubated with increasing amounts of each extract. The parasite's growth was monitored by counting them in a Neubauer chamber. Percentage of inhibition was calculated as the *ratio* between parasite's growth in the presence or absence of each extract, after 72 h of culture. IC₅₀ was obtained by plotting the percentage of inhibition against extract concentration. Each experiment was performed in triplicate and trypanocide drug benznidazole (BZL) was used as positive control.

Cytotoxicity assay

Human myelogenous leukaemia K-562 cell line was cultured in DMEM supplemented with penicillin G (100 units/mL), streptomycin (100 µg/mL) and 10-15% fetal bovine serum in a 5% CO₂ humidified incubator at 37 °C.

The extracts were first dissolved in DMSO and then in MEM. Confluent cultures were treated with medium containing the extracts at concentrations ranging from 1 up to 450 µg/mL or the solvent alone (control) for 24, 48 and 72 h. Each drug concentration was tested in quadruplicate. Final concentration of DMSO in the test medium and controls was 1%. After the indicated treatments, cells were removed from culture flasks by treatment with trypsin.

The effect of the DMSO 1% was analyzed in all experimental procedures and there was no significant difference between the solvent and the control on all the measured parameters.

Cell death was determined by incubating with 0.4% Trypan Blue, and then the stained cells were counted in a Neubauer haemocytometer. The results were expressed as the percentage of cell death relative to solvent-treated control incubations and the IC₅₀ values were calculated using non-linear regression analysis (percentage of cell death vs. concentration). *In vitro* cytotoxicity effect was calculated as the *ratio* between cell's growth in the presence or absence of each extract, after 72 h of culture.

Morphological features of the cells (variation in size or nuclear/cytoplasmic ratio, presence of granules) were monitored by light microscopy.

General experimental procedures

The extracts were subjected to thin layer chromatography (TLC) examination on aluminium sheets pre-coated with silica gel 60 F 254 (Merck). Mobile phase **I** Toluene: ethylacetate: formic acid (9:3:0.7), **II**: ethylacetate: Acetic Acid: Formic acid: water (10:1.1:1.1:2.6), **III** Toluene: ether (1:1

saturated with 10% acetic acid). The chromatograms were observed first without chemical treatment, under UV 254 and UV 365 nm light, and then using the spray reagents: NP/PEG, Dragendorff, KOH 5%, Liebermann-Burchard reagent or FeCl₃ 3% (Wagner et al., 2001). TLC sheet was also immersed in a 0.4 mM DPPH methanolic solution for 10 s.

GC-MS analysis was carried out on PERKIN ELMER Gas Chromatograph attached to a TURBOMASS Mass Spectrometer. SE-30, 25 m x 0.22 mm de ID column was used for fraction IV separation. Helium was used as the carrier gas and kept at 1 mL/min. Oven temperature was programmed at 100 °C (5 min) to 310 °C at a rate of 10 °C/min (10 min). The mass-scanning was done by electron impact (EI) source with electron energy set to 70 eV and mass range *m/z* 50-450.

RESULTS

Cytotoxicity

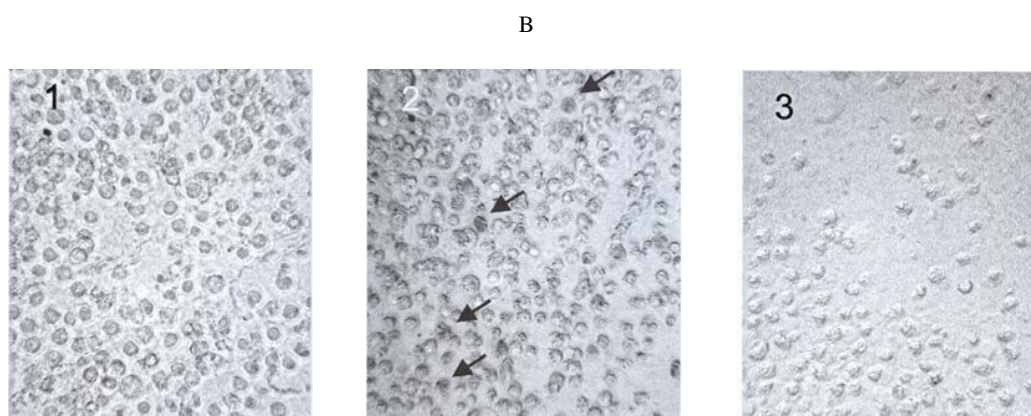
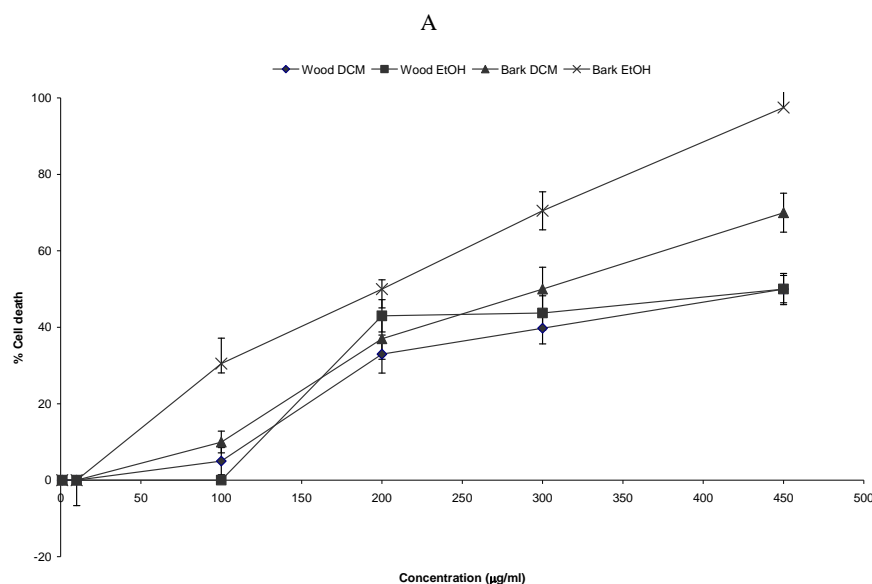
The *in vitro* cytotoxic effect was determined using the haemocytometer to estimate the total number of viable and non-viable cells and was also evaluated by Trypan-blue exclusion method (Fig. 1A). All extracts have shown significant cell death at the highest dosage of 0.45 mg/mL. IC₅₀ was estimated from graphical interpolation. Among the tested fractions, Bark_{EIOH} extract has shown the strongest inhibition with an IC₅₀ of 0.20 mg/mL. IC₅₀ for wood extracts was 0.45 mg/mL. Microscopically detectable alterations were observed in the cell morphology. When cells were incubated with bark extract granulation and vacuolization in the cytoplasm were observed, as well as irregular borders and a lower number of cells were detected (Fig. 1B).

Antitrypanosomal activity of the extracts

Ethanollic and DCM extracts of *Castela coccinea* Griseb bark and wood were screened for antiprotozoal activity against *Trypanosoma cruzi* epimastigotes (Fig. 2). Wood_{DCM} extract showed the strongest activity with an IC₅₀ = 60 µg/mL. BZL was used as positive control and showed an IC₅₀ = 10 µg/mL. Cytotoxicity did not correlate with antiprotozoal activity. Wood extracts were more selective against the parasites than against mammalian cells. This was demonstrated by their Selectivity Index (SI = IC₅₀ for K-562 cells/ IC₅₀ antiprotozoal activity) values > 1. Wood_{DCM} SI has been 7.5.

Figure 1. Effect of *Castela coccinea* Griseb extracts on inhibition of the leukemic cell line K-562.

A. *In vitro* cytotoxicity effect calculated as the ratio between cell's growth in the presence or absence of each extract, after 72 h of culture. **B.** Photomicrographs **1)** Cells incubated with solvent alone (Control) **2)** Bark_{EtOH} (100 µg/mL) treated cells and staining with Trypan blue. Death staining cells are shown with an arrow. It was observed a lower number of cells, irregular borders and cytoplasmic granules. **3)** Bark_{EtOH} (200 µg/mL) treated cells without staining. A very low number of cells and abundant granules were observed.



Fractionation and phytochemical screening of Wood_{DCM} extract

The Wood_{DCM} extract was purified by column chromatography. When tested against the parasite, Fraction **I**, the high non-polar fraction, showed an IC₅₀ value of 185 µg/mL, while fraction **III** exhibited an IC₅₀ value of 60 µg/mL. The highest activity was concentrated at fraction **IV** (Fig. 3), and this fraction also had activity against DPPH.

Phytochemical analysis by TLC has demonstrated the presence of alkaloids and terpenoids in fraction **III** and alkaloids, phenolic acids and coumarin in fraction **IV**. When this fraction (**IV**) was subjected to GC-MS,

scopoletin and an 8-amino-6-methoxyquinoline group were detected, however the identity of the whole alkaloid molecule could not be determined. Despite scopoletin being the main component of fraction **IV** (Fig. 4), it did not show any trypanocide activity up to 6 µg/mL. On the other hand, the phenolic acids detected in fraction **IV** were also detected in fraction **V** (the most polar fraction). It is a very interesting fact that this last fraction had not trypanocide activity at all. Therefore, trypanocide activity would not be caused by effect of these phenolic acids. On the contrary, they were responsible for DPPH scavenging activity.

Figure 2. *In vitro* trypanocide activity of *Castela coccinea* Griseb extracts expressed as percentage of parasite's growth inhibition..

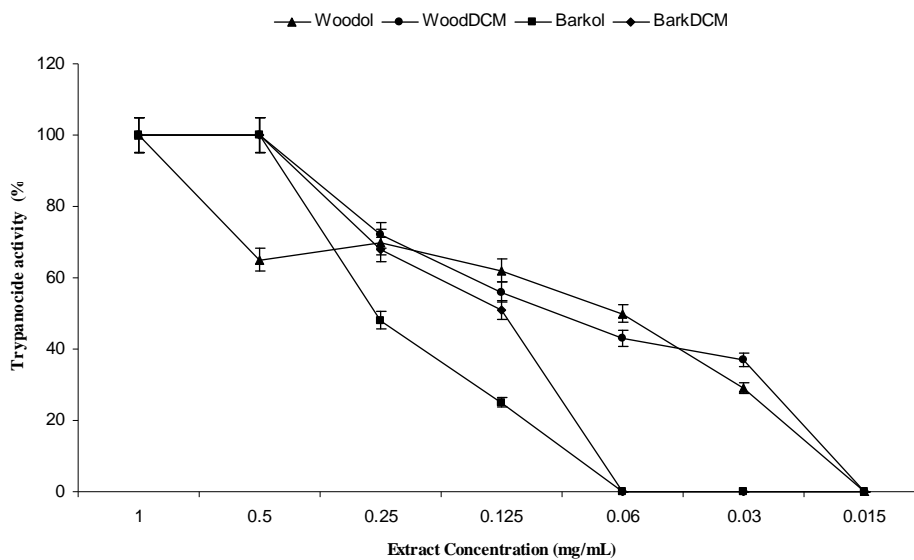


Figure 3. *In vitro* trypanocide activity of different wood DCM fractions expressed as percentage of parasite's growth inhibition.

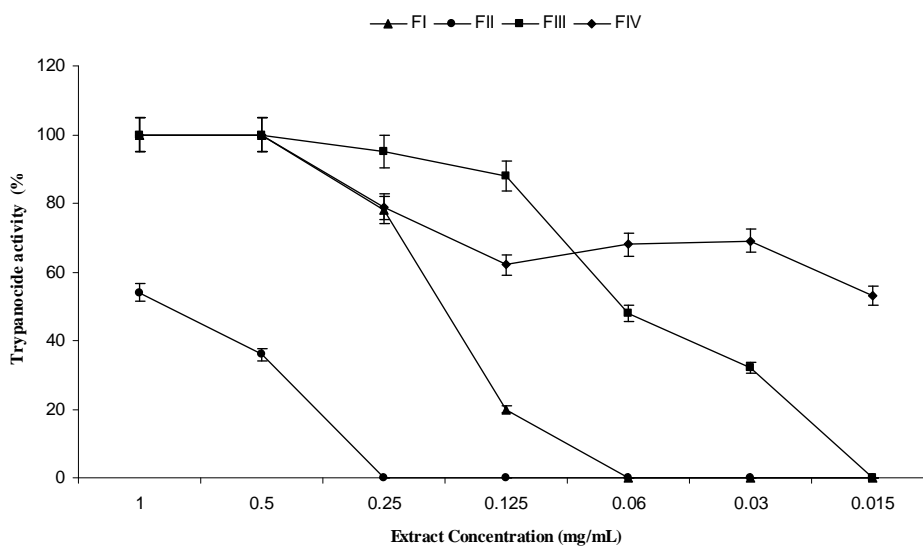
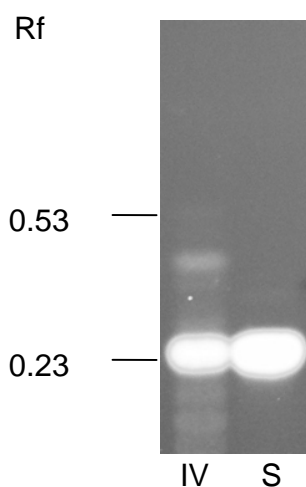


Table 1. *In vitro* trypanocide activity and cytotoxicity of *Castela coccinea* Griseb extracts.

	Cytotoxicity IC ₅₀ (µg/mL)		Selectivity index ^a
	<i>T. cruzi</i>	K-562	
Wood _{ol}	81	450	5.5
FI	185	-	-
FII	-	-	-
FIII	60	-	-
FIV	15	-	-
FV	-	-	-
Wood _{DCM}	60	450	7.5
Bark _{ol}	220	200	0.9
Bark _{DCM}	168	300	1.78

(a) Selectivity index was determined by the ratio of the IC₅₀ value on K-562 cells to the IC₅₀ value on *T. cruzi*.

Figure 4. Analysis of fraction IV by TLC. TLC plate was stained with NP-PEG. Fraction IV exhibited eight to ten blue fluorescent zones from the start up to Rf 0.53. Solvent system III. S: commercial scopoletin 0.1% in ethanolic solution (5 µL).



DISCUSSION

This work showed that *Castela coccinea* Griseb presents a strong trypanocide activity and particularly wood extracts displayed the most intensive effect. On the other hand, all extracts have exhibited cytotoxicity at high concentrations. However, Wood_{DCM} was more selective against parasites than against mammalian cells with a selectivity index of 7.5. A fraction of this extract enriched in coumarins, phenolic acids and alkaloids was more effective against *Trypanosoma cruzi* with an IC₅₀ of 15 µg/mL.

Several coumarins and indol or quinoline alkaloids exhibit remarkable parasitaemia suppression against different strains of *Plasmodium* or *Trypanosoma* (among others Phillipson, 1999; Camacho, 2004; Muhammad et al., 2003 and 2004; Argotte-Ramos et al., 2006; Huang et al., 2006; Fournet et al., 2007; Bringmann, 2007; Frederich, 2008). Our results suggest that alkaloids could be responsible for this activity. The 8-amino-6-methoxyquinoline group was detected but the whole molecule could not be identified yet. Interestingly, compounds containing this group, as for example, primaquine (CAS N° 63-45-6) are already being used to produce radical cure and prevent relapse of vivax and oval malaras following treatment with blood schizontocide (Dongre et al., 2005; Musonda et al., 2007; Kaur et al., 2007).

The fraction with the highest trypanocide activity also showed antioxidant capacity. It has been proven that *Trypanosoma* is sensitive to oxidative stress, in particular by reactive oxidant species produced during the immune response of the host. The fact that this fraction both show radical scavenging and kill parasites suggests that these activities should be associated with two different molecules present in the extract. The detected antioxidant compounds were phenolic acids, also present in the most polar fraction without show trypanocide activity. However, the possibility that the same molecule works as a radical scavenger in mammalian cells and as a radical inducer in parasites could not be completely miss-considered.

The screening of natural products provides the chance to discover new molecules of unique structure with high activity and selectivity, which can be further optimized by semi- or fully synthetic procedures. Alkaloids are one of the most fascinating natural products, providing many drugs for human use (Phillipson et al., 1999; Kayser et al., 2003). The results of this work show that *Castela coccinea* Griseb extracts are potentially a rich source of bioactive compounds and the search of these entities is carried out. A preliminary phytochemical study is done but further studies are needed as this plant is unknown from the phytochemical point of view.

CONCLUSION

Castela coccinea Griseb has not been previously studied for their trypanocidal effect. Wood_{DCM} showed strong activity against *Trypanosoma cruzi* with an IC₅₀ of 15 µg/mL. Our results suggest that alkaloids could be responsible for this activity. Interestingly, although the whole molecule identity could not be assessed, an 8-amino-6-methoxyquinoline group was detected. A bioguided fractionation is in progress to identify the active compounds.

REFERENCES

- Argotte-Ramos R, Ramirez-Avila G, Rodriguez-Gutierrez Mdel C, Ovilla-Munoz M, Lanz-Mendoza H, Rodriguez MH, Gonzalez-Cortazar M, Alvarez L. 2006. Antimalarial 4-phenylcoumarins from the stem bark of *Hintonia latiflora*. *J Nat Prod* 69:1442-1444.
- Bourdy G, Oporto P, Gimenez A, Deharo E. 2004. A search for natural bioactive compounds in Bolivia through a multidisciplinary approach Part IV. Evaluation of the antimalarial activity of plants used by Isoceño-Guaraní Indians. *J Ethnopharmacol* 93:269-277.
- Bringmann G, Brun R, Kaiser M, Neumann S. 2007. Synthesis and antiprotozoal activities of simplified analogs of naphthylisoquinoline alkaloids. *Eur J Med Chem* 43:32-42.
- Camacho MR, Phillipson D, Croft S, Yardley V, Solis P. 2004. *In vitro* antiprotozoal and cytotoxicity activities of some alkaloids, quinines, flavonoids and coumarins. *Planta Med* 70:70-72.
- Camargo EP. 1964. Growth and differentiation in *T. cruzi*. I. Origin of metacyclic trypanosomes in liquid medium. *Rev Med Trop Sao Paulo* 6:220-224.
- Castro JA, Montalto de Mecca M, Bartel LC. 2006. Toxic side effects of drugs used to treat Chagas' disease (American trypanosomiasis). *Hum Exp Toxicol* 25:471-479.
- Cuendet M, Pezzuto JM. 2004. Antitumor activity of bruceantin: An old drug with new promise. *J Nat Prod* 67(2):269-272.
- Dongre V, Karmuse P, Nimbalkar M, Singh D, Kumar A. 2005. Application of GC-EI-MS for the identification and investigation of positional isomer in primaquine, an antimalarial drug. *J Pharm Biomed Anal* 39:111-116.
- Frederich M, Tits M, Angenot L. 2008. Potential antimalarial activity of indole alkaloids. *Trans R Soc Trop Med Hyg* 102(1):11-19.
- Fournet A, Ferreira ME, Rojas de Arias, Guy I, Guinaudeau H, Heinzen H. 2007. Phytochemical and antiprotozoal activity of *Ocotea lancifolia*. *Fitoterapia* 78:382-384.
- Grieco P, Haddad J, Piñeiro-Nuñez M, Huffman J. 1999. Quassinoid constituents from the twigs and thorns of *Castela polyandra*. *Phytochemistry* 50:637-645.
- Guo Z, Vangapandu S, Sindelar RW, Walker LA, Sindelar RD. 2005. Biologically active quassinoids and their chemistry: potential leads for drug design. *Curr Med Chem* 12:173-190.
- Huang F, Tang LH, Yu LQ, Ni YC, Wang QM, Nan FJ. 2006. *In vitro* potentiation of antimalarial activities by daphnetin derivatives against *Plasmodium falciparum*. *Biomed Environ Sc* 19(5):367-370.
- Jacobs H, Simpson D, Reynolds W. 2007. Quassinoids and a coumarin from *Castela macrophylla* (Simaroubaceae). *Biochem Syst Ecol* 35:42-44.
- Jiwajinda S, Santisopasri V, Murakami A, Hirai N, Ohigashi H. 2001. Quassinoids from *Eurycoma longifolia* as plant growth inhibitors. *Phytochemistry* 58:959-962.
- Kaur K, Patel SR, Patil P, Jain M, Khan SI, Jacob MR, Ganesan S, Tekwanib BL, Jaina R. 2007. Synthesis, antimalarial, antileishmanial, antimicrobial, cytotoxicity, and methemoglobin (MetHB) formation activities of new 8-quinolinamines. *Bioorg Med Chem* 15:915-930.
- Kayser O, Kiderlen AF, Croft SL. 2003. Natural products as antiparasitic drugs. *Parasitol Res* 90: S55-S62.
- López Sáez J y Pérez Soto J. 2008. Etnofarmacología y actividad biológica de *Quassia amara* (Simaroubaceae): Estado de la cuestión. *Bol Latinoam Caribe Plant Med Aromat* 7(5):234-246.
- Moncayo A. 2003. Chagas' disease: current epidemiological trends after the interruption of vectorial and transfusional transmission in the Southern. *Mem Inst Oswaldo Cruz* 98:577-591.
- Musonda C, Littl S, Yardley V, Chibalea K. 2007. Application of multicomponent reactions to antimalarial drug discovery. Part 3: Discovery of aminoxazole 4-aminoquinolines with potent antiplasmodial activity in vitro. *Bioorg Med Chem Lett* 17:4733-4736.

- Muhammad I, Dunbar DC, Khan SI, Tekwani BL, Bedir E, Takamatsu S, Ferreira D, Walker LA. 2003. Antiparasitic alkaloids from *Psychotria klugii*. *J Nat Prod* 66(7):962-967.
- Muhammad I, Bedir E, Khan SI, Tekwani BL, Khan IA, Takamatsu S, Pelletier J, Walker LA. 2004. A new antimalarial quassinoid from *Simaba orinocensis*. *J Nat Prod* 67:772-777.
- Ogura M, Cordell GA, Kinghorn AD, Fransworth NR. 1977. Potential anticancer agents VI. Constituents of *Ailanthus excelsa*. *Lloydia* 40:579.
- Ogura M, Cordell GA, Fransworth NR. 1978. Alkaloid constituents of *A. excelsa*. *Lloydia* 41:166.
- Ouyang Y, Mitsunaga K, Koike K and Ohmoto T. 1995. Alkaloids and quassinoids of *Brucea mollis* var. *tonkinensis*. *Phytochemistry* 30(4):911-913.
- Phillipson JD. 1999. New drugs from nature – It could be yew. *Phytother Res* 13:2-8.
- Polonsky J. 1973. Quassinoid bitter principle. *Fortschr Chem Org Naturst* 30:101.
- Polonsky J. 1985. Quassinoid bitter principle—II. *Fortschr Chem Org Naturst* 47:237.
- Readel K, Seigler D, Young D. 2003. 5-methoxycanthin-6-one from *Leitmeria floriana* (Simaroubaceae). *Biochem Syst Ecol* 31:167-170.
- Sanchez E and Comin J. 1971 Two new carbonile alkaloids from *Aeschrion crenata*. *Phytochemistry* 10:2155-2159.
- Sherman MM, Borris RP, Ogura M, Cordell GA, Fransworth NR. 1980. 3S, 24S, 25-trihydroxytirucall-7-ene from *A. excelsa*. *Phytochemistry* 19:1499.
- Wagner H, Bladt S, Zgainiki E. 2001. *Plant Drug Analysis*. Berlin, Springer, pp 360-364.

