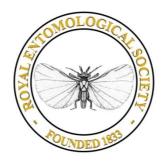
Royal Entomological Society



HANDBOOKS FOR THE IDENTIFICATION OF BRITISH INSECTS

To purchase current handbooks and to download out-of-print parts visit:

http://www.rovensoc.co.uk/publications/index.htm

http://www.royensoc.co.uk/publications/index.htm



This work is licensed under a <u>Creative Commons</u> <u>Attribution-NonCommercial-ShareAlike 2.0 UK:</u> <u>England & Wales License.</u>

Copyright © Royal Entomological Society 2013

MYCETOPHILIDAE

(Bolitophilinae, Ditomyiinae, Diadocidiinae, Keroplatinae, Sciophilinae and Manotinae) DIPTERA, NEMATOCERA

A. M. Hutson, D. M. Ackland and L. N. Kidd



ROYAL ENTOMOLOGICAL SOCIETY OF LONDON

Editor: M. G. Fitton

MYCETOPHILIDAE

(Bolitophilinae, Ditomyiinae, Diadocidiinae, Keroplatinae, Sciophilinae and Manotinae) DIPTERA, NEMATOCERA

By

A. M. Hutson

Department of Entomology British Museum (Natural History) London SW7 5BD

D. M. Ackland

Coniston House New Street Deddington Oxon OX5 4SP

and

L. N. Kidd

8 Central Avenue Greenfield Oldham OL3 7DH

1980 ROYAL ENTOMOLOGICAL SOCIETY OF LONDON The aim of the *Handbooks* is to provide illustrated identification keys to the insects of Britain, together with concise morphological, biological and distributional information. The series also includes a *Check list of British insects*.

Each handbook should serve both as an introduction to a particular group of insects and as an identification manual.

Details of handbooks currently available, and an order form, can be obtained from the Royal Entomological Society, 41 Queen's Gate, London SW7 5HU.

World List abbreviation: Handbk Ident. Br. Insects

© Royal Entomological Society of London, 1980

First published 1980 by the Royal Entomological Society of London, 41 Queen's Gate, London SW7 5HU.

Printed by G. Donald & Company Limited, Osiers Road, London SW18 1NL

Contents

									- 4	uge
Introduction										5
Relationships with other fa	milies									5
Immature stages										6
General behaviour and hab	oitats									7
Associations with fungi										8
Lists of fungi and other ha	bitats v	with a	associ	ated N	Aycet	ophili	idae			10
Parasites and disease .					٠.	٠.				14
The fossil record										14
Collecting and preserving										15
Rearing Diptera from fu	ngi									15
Preparation of specimen										15
Morphology										16
Notes on the keys										20
Acknowledgements .										21
Check list										21
Key to subfamilies .										25
Subfamily Bolitophilinae										26
Subfamily Ditomylinae .										28
Subfamily Diadocidiinae										28
Subfamily Keroplatinae										29
Subfamily Sciophilinae .										40
Subfamily Manotinae .										69
References										70
Figures										73
Index										107

Introduction

The Fungus-gnats (Mycetophilidae) are a large family of terrestrial Nematocera, usually with a compact hump-backed appearance (fig. 1), long coxae and well developed tibial spurs. Most species are rather small, but some may reach a length of about 10mm. They are generally some combination of brown, black and yellow, rarely strikingly patterned, but sometimes brightly coloured like Hymenoptera. The family ranges throughout all the major land areas of the world (as well as on many oceanic islands) from northern Greenland in the north to Tierra del Fuego and the Crozet Islands in the south. The Mycetophilids of the tropics have been relatively poorly studied and although the fauna there may not be as representative of the whole family as in the temperate regions, it is certainly rich in species, most of which still await description. About 3400 species have been described.

The basic diagnostic characters of the family are as follows: usually three ocelli, median ocellus sometimes reduced or absent; antennae elongate and placed near the middle of the height of the eye with 11-17 (usually 16) generally distinctly separated segments; thorax distinctly arched, with no suture dividing the mesonotum; coxae long; legs fairly long with tibial spurs usually well developed; other bristles on legs and thorax often strong; venation with Rs usually simple, sometimes forked very close to the base forming a "small cell" over the r-m cross-vein, sometimes forked nearer the tip of the wing; median and posterior forks usually present, one or more of branch veins sometimes incomplete or absent; discal cell absent; anterior veins usually more strongly developed than posterior (post-radial) veins.

About 450 species in seven subfamilies are recorded from Britain. About half of these species belong to the single subfamily Mycetophilinae and are not included in this work, which gives keys to species and figures of male genitalia for the other 204 species (included in the subfamilies Bolitophilinae, Ditomyiinae, Diadocidiinae, Keroplatinae, Sciophilinae and Manotinae).

Relationships with other families

The Sciaridae, formerly considered a subfamily of the Mycetophilidae, but currently given separate family status, have a similar appearance and agree in most of the above listed characters. The British Mycetophilidae and Sciaridae are most closely related to the Scatopsidae and Cecidomyiidae. The Scatopsidae (and its related families Synneuridae and Canthyloscelidae) have large eyes that meet or almost meet above the antennae, tibial spurs usually reduced or absent and uniform pubescence on the thorax (no distinct rows of bristles). They are mostly small stout black flies, sometimes with some yellow. Most of the Cecidomyiidae are small fragile species with an eye-bridge and no ocelli, wing venation reduced to two postradial veins reaching the wing margin, very short first tarsal segment, no tibial spurs. Some Cecidomyiidae (in the subfamily Lestremiinae) have ocelli, wing venation similar to Sciaridae and a long first tarsal segment.

The Sciaridae are readily separable from the Mycetophilidae in having the eyes produced over the antennae forming an eye-bridge and in having shorter coxae. The included species are rather uniform in general appearance with characteristic wing venation, generally dark colouration and usually a lack of strong bristles. A few paler species, e.g. *Phytosciara flavipes* Meigen, may be mistaken for Mycetophilidae on superficial examination. The females of some species are wingless and with degeneration of the wings there may be modifications of the thoracic structure and reduction of eyes (including loss of the eye-bridge). Even the halteres may be lost. No apterous Mycetophilidae are known, but the known females of two genera

(Baeopterogyna Vockeroth and Moriniola Matile) and the female of one Macrocera species from the south Indian Ocean, are brachypterous. The apterous and brachypterous forms may be confusing, but associated males and larvae will leave no doubt as to the correct family placement.

British Sciaridae have been little studied, but recent systematic studies by Tuomikoski (1966a) and Steffan (e.g. 1966) might facilitate a better classification of our own fauna. While there may be sufficient grounds for giving the Sciaridae separate family status, many of the groups here treated as subfamilies of the Mycetophilidae are equally well defined and discrete enough for some authors to consider them too as separate families. Whether or not it is fully justified to give the Sciaridae special separate status, it is convenient and practical to do so until we have greater knowledge of their morphology and biology.

Immature stages

Eggs (fig. 2) are usually oval, white and opalescent, the opalescence due to the chorion being raised into longitudinal rows of tubercles. The larva emerges with the aid of a small chitinous egg-burster.

Larvae (fig. 3) are usually slender, white and with a distinct dark head capsule. The head capsule is complete and non-retractile, with the mandibles opposed and in a horizontal plane. It is light coloured in a few species, such as the cave-dwelling Speolepta leptogaster Winnertz and the polymycetophagous Mycetophila fungorum Degeer. The epicranial plates meet below the head in Keroplatinae, Sciophilinae and Mycetophilinae. Antennae are generally very reduced and obscure, but are well developed in Bolitophilinae. The abdomen is of at least 8 segments, has no tergal plates, no prolegs and no anal lobes (although small anal papillae are present in Keroplatinae and the posterior spiracles of Ditomyiinae are on small spinelike processes). Most larvae that live internally in fungi (especially Mycetophilinae) have some development of locomotory aids in the form of creeping welts, but these are poorly developed in Bolitophilinae and absent in the web-spinning Sciophilinae and Keroplatinae. The Keroplatinae also tend to be relatively large and vermiform. while Phronia (Mycetophilinae) are hemiovoid and either covered with slime or with a protective limpet-like shield made up of local debris. Epicypta (Mycetophilinae) has a similar protective case. The larvae of Keroplatinae are apneustic, the rest are varieties of hemipneustic and may vary from instar to instar, e.g. in Bolitophila and Brachypeza the first instar is metapneustic, 2nd and 3rd are propneustic and the 4th (last) instar is peripneustic. While usually white, some larvae may be coloured from dull vellow to bright green.

The evaluation of characters to establish a satisfactory style of description of larvae was presented by Lastovka (1971) and this has been largely followed by Trifourkis (1977).

The pupa (fig. 4) is obtect adecticous and with sufficient of the adult characters apparent for identification with the family. The web-spinners tend to pupate in or by the web, with or without a cocoon. Those developing in soft fungi leave the fungus and pupate in nearby soil or under bark, etc., sometimes with a loose cocoon. Others may pupate in situ in hard fungi or non-fungus media with or without a cocoon, some cocoons being very substantial. The pupal stage is usually very short and hence has been rather neglected, but particularly in those species that build a substantial cocoon the pharate adult may remain in the cocoon for some time awaiting suitable emergence conditions.

General behaviour and habitats

Fungus-gnats are ubiquitous and form an important part of the Dipterous fauna of many habitats. However, little is recorded of the behaviour of adults and much remains to be learned of their larval habitats. Adults are found, often in large numbers, in moist dark places, especially steep damp overhanging banks in woods. chiffs, rivers, the mouths of caves, culverts and tunnels, the root systems of fallen trees, etc. Some prefer more open situations and a few prefer the higher foliage of broad-leaved trees. Most species are probably crepuscular and have their peak of activity around dusk with a lesser peak around dawn. They rest during the daylight period, but are more prone to disturbance during bright, moist conditions. Many species are attracted to light; Malaise traps, pitfall and sticky traps and windows can all produce a variety of species including many that, presumably because of their activity rhythms, are not taken by conventional sweeping, etc. Males of a few species, e.g. in the genera Mycetophila and Phronia, sometimes form dancing swarms. Some species are attracted to flowers, particularly in the Keroplatinae, and genera such as Antlemon and Asindulum have elongate mouthparts presumably for feeding on nectar. Species of Macrocera and Mycomya are frequently to be seen resting in fairly exposed situations on vegetation, but published suggestions that they feed at flowers need confirmation. Seepage from trees, honeydew, some odoriferous rotting vegetation, etc. may attract others. Many of the species that breed in fungi may be found around their hosts.

In Britain, probably the most diverse fauna is found during June. There is then a lull before the autumn flush associated with the peak appearance of fungus fruiting bodies. During this autumn peak Mycetophilidae form an important part of the Dipterous fauna, being extremely numerous when most other flies are on the decline. While far more abundant in these periods, Mycetophilidae (especially Bolitophilinae and Mycetophilinae) can be found throughout the year, except for the severest cold spells of winter. In winter they may collect in heavy concentrations in patches of evergreen vegetation, such as ivy or a conifer. Many species hibernate in caves (see the journal *Hypogean Fauna*, published by the British Cave Research Association), cellars, outhouses, etc., under bark and in hollow plant stems.

About one third of the species of Mycetophilidae recorded in Britain have been reared. Of these c.75% are associated with the fruiting bodies of fungi, including Myxomycetes, c.20% are associated with rotting wood and 5% with other habitats. Because a large amount of work has gone into the rearing of Diptera from fungi. including major efforts in Britain by Edwards (1925), Buxton (1961) and Trifourkis (1977), in Germany by Eisfelder (1954, 1955) and Plassmann (1969, 1971a), in Hungary by Dely-Draskovits (1974) and in Finland by Hackman & Meinander (1979) it seems likely that a smaller proportion of the as yet unreared groups are associated with fungi. Preliminary analysis suggests that probably no more than 60% of the British fauna is closely associated with fungal fruiting bodies. Of the species that have been associated with fungi, about 60% occur inside the body of the fungus, while the rest live on the (under) surface. Again this proportion may not be accurate, because of the notoriously low success rate in attempts to rear larvae that occur inside the fungus. Two species are known to produce gall-like formations on fungi. Some of the species that are found on the outside of fungi are equally common on non-fungus substrates, such as the Keroplatinae and some Mycomya spp. Some of the species that feed on fungus material, particularly those found in the later stages of decay of the fungus, also occur in other habitats, e.g. Docosia gilvipes Haliday.

Most of the species that are associated with rotting wood live on the surface or under bark—only a few penetrate the wood. Probably most of these are feeding on fungal hyphae. The figure of 5% for the forms known to breed in other habitats may

be the most serious inaccuracy. The figure covers mosses and liverworts, bird and mammal nests, accumulations of rotting plant material, in the tunnels made by earthworms, in grass tussocks, on the walls of caves. The larval pabulum of many groups, including some quite large groups with very common species, is hardly known and it is probable that many of these occur in as yet uninvestigated habitats. For instance, for a short period in the early spring *Boletina gripha* emerges in enormous numbers from areas as diverse as Kent woodland and Westmorland moorland. It is active for most of the summer in smaller numbers, probably passing through a series of overlapping generations—but its larva is unknown. Possibly many such non-sporophore feeders feed on mycelia in the substrate.

Along with this wide diversity of habitat goes a range of diet from vegetarian to carnivorous. All those species feeding inside fungus or wood are assumed to be vegetarians, feeding on fungal tissues. Most of those feeding on the outside of fungi are probably feeding on fungal spores. However, some of the latter and others that live under stones, logs, in grassland, etc. (particularly in the Keroplatinae) are, at least in part, predaceous on small animals that they trap in webs. The webs of many species have droplets of oxalic acid deposited on them by the larvae and these quickly kill any small animal that comes into contact. In New Zealand and Australia, larvae of the genus Arachnocampa drop a number of vertical filaments each with regularly spaced sticky mucous drops. These droplets catch small insects. The other extraordinary feature of this larva is that it has a controllable luminescence at the ends of its Malpighjan tubules. The larvae often occur in caves and are sometimes so numerous that the filaments are lit up by the luminescence of the larvae and the sight has become a tourist attraction (Richards, 1960). There has been discussion on the possibility of establishing such a colony as a "natural phenomenon" in a tourist cave in England. Some South American cave species may use vertical filaments in the same way, but are not so spectacular. These species and Arachnocampa belong to the Keroplatinae and several other species of this group, including some of our own, demonstrate a limited degree of luminosity. Cave dwelling species of Macrocera have been described as starting life as scavengers, but quickly become predaceous using a similar technique to Arachnocampa, while others spin a large loose web in places where they might expect to find prey, e.g. near bat guano (Peck and Russell, 1976). Such species should be searched for in Britain, where we have so far only recorded Speolepta leptogaster Winn. as regularly breeding in caves. Speolepta is extremely common and is probably a scavenger, eating any organic matter, such as algae, fungus or animal remains, that occur within its territory. The most outstanding predatory larva is Planarivora insignis Hickman, which is an endoparasite of terrestrial planarians in Tasmania (Hickman, 1965).

The species that occur in bird and mammal nests, accumulations of rotting vegetable matter and perhaps also some species that seem to prefer fungi in an advanced state of decay may better be termed saprophagous than fungivorous.

Most species overwinter in the larval stage. Some species appear to be univoltine, but many have a series of overlapping generations.

Associations with fungi

The host specificity of those species of Mycetophilidae that breed in fungi is still poorly understood. Workers who have studied this question in some detail have found different preferences. The poor success rate so far achieved in rearing attempts has been an unfortunate limitation on host data.

Sporophores of the larger fungi are particularly common in summer, building up to a maximum in late autumn. Their infestation with Mycetophilidae remains fairly

constant from June to October (Russell-Smith, 1979). There is then a decrease in infestation rate in November at the time of the decrease in actual numbers of sporophores. The effect of rainfall on infestation is slight, but dry periods may **slightly reduce** infestation, while very wet fungi become stodgy and less attractive to flies. The highest infestation rates occur in the Amanitaceae, where over 90% of sporophores may be infested. Boletaceae and Russulaceae may reach infestation rates of over 80%. Cortinariaceae and Tricholomataceae over 70% and Polyporaceae in the lower 60%'s. However, within the Polyporaceae infestation rate be very high in some genera such as Coriolus and Bjerkandera. Similarly some scacra of the other families show consistently low infestation rates. Within genera the infestation rate varies greatly from species to species; thus although the average infestation rate for Russula species is high, this may range from 30% to 90% according to species. A major problem in assessing the importance of various groups of fungi is the widely differeing results of different studies. For instance, Edwards (1925) reared Ditomyia fasciata Meigen easily and in numbers from what he considered its host fungus, Polystictus [= Coriolus] versicolor (L. ex Fr.) Quél. Buxton (1961) reared this species from only 1 of 16 samples of C. versicolor, but also from 3 of nine samples of Fomes ulmarius (Sow.) Fr., 1 of 1 Daedalia biennis (Bull) Ouel and Polyporus picipes Fr. and 1 of 8 samples of Bjerkandera adusta (Wild.) Fr. He did not find it in any of 11 samples of Polyporus squamosus (Huds. ex Fr.) Fr. Plassmann (1969) reared it from Trametes [= Coriolus] versicolor as well as Inonotus radiatus (Sow.) Fr. and Leptoporus amorphus Fr. Trifourkis (1977) did not find it at all in 70 samples of Polyporaceae including 15 samples of Coriolus versicolor, 8 B. adusta and 2 P. squamosus. The reason for this variation is uncertain: human factors, such as techniques of rearing and the length of study, are doubtless important, but many natural features may also be involved, such as climate (dry/wet; windy/sheltered), geography (general distribution, altitude) ecology (forest type, composition of tree species, their density, age, etc). The whole problem is further complicated by the difficulties of identifying Mycetophilid larvae, together with the taxonomic and nomenclatural problems of identifying fungi.

Despite such problems, data are accumulating that enable some statements on the host preferences of Mycetophilidae. Although most of the species reared from fungi are restricted to that medium, very few are known to be very host-specific. A few appear to be monophagous (restricted to one species of fungus) such as Mycetophila cingulum Meigen on Polyporus squamosus, Trichonta falcata Lundström on Stereum hirsutum (Willd. ex Fr.) and Trichonta vernalis Landrock on Calocera cornea (Batsch. ex Fr.) Fr., Bolitophila hybrida Meigen is extremely common in Paxillus involutus (Batsch. ex Fr.) Fr., but has been recorded from a variety of other fungi. Similarly B. saundersii Curtis is usually found in Hypholoma fasciculare (Huds. ex Fr.) Quél, but occasionally occurs in other fungi.

Some species appear to be stenophagous (restricted to a few closely related fungi) such as Bolitophila cinerea Meigen on a variety of Hypholoma and Pholiota spp., Sciophila buxtoni Freeman from a variety of tough lignicolous Polypores such as species of Coriolus, Pseudotrametes and Daedaleopsis. Cordyla fasciata Meigen, Exechia nigroscutellata Landrock and Mycetophila alea (Laffoon) are all restricted to Russulaceae, while Exechia separata Lundström and Mycetophila signatoides Dziedzicki are restricted to Boletaceae.

Sciophila lutea Macquart, Docosia gilvipes Haliday and Rondaniella dimidiata Meigen are extreme examples of polyphagous species, in fact D. gilvipes also breeds in the nests of birds and animals, but too little is known about the real preferences of many species to decide whether they might more correctly be termed oligophagous (having a preference for a limited variety of fungi that presumably share some features). Certainly many are restricted to soft ground fungi, such as most of the

Mycetophilinae, while others prefer a broad spectrum of lignicolous fungi (such as *Dynatosoma fuscicorne* Meigen and *Mycetophila ornata* Stephens). For these species other factors such as phenology, habitat or density of fungi may be more important in limiting host choice.

The development of a soft fungal sporophore may be separated into three phases: I, fungus fresh with no signs of decay; II, decaying through age and damage (mainly due to the activity of fungivores); III, semi-liquid state through advanced state of decay and histolysis of fungal tissues. Most Mycetophilidae attack at stages I to II, but species such as Mycetophila fungorum Degeer will continue to develop in deliquescing sporophores with little regard to the species. Hackman & Meinander (1979) recognise four stages of development. Tough aerial sporophores tend to desiccate and wither or become friable rather than rot; in these late stages they are unsuitable for Mycetophilidae.

Naturally most effort has been put into rearing from the larger Basidiomycetes. Most of these belong to the "Hymenomycetes" (including all agarics, boletes, polypores, coral fungi, hedgehog fungi, dry rots, etc.) and even some of the smaller fungi in this group, such as *Marasmius*, have been shown to support Mycetophilidae. The "Gasteromycetes" (Puff-balls, earthballs, stinkhorns, etc.) and the phragmobasidiomycetes (jelly fungi) have not proved so suitable—the former less so than the latter. The other major group of fungi, the Ascomycetes, have received little attention from workers apart from Buxton (1961), who made a special effort to check them. He had limited success, but found a few previously unreared species. Little has been written on the role of other fungi and Myxomycetes, although some Mycetophilinae (Buxton, 1954) and the rare *Manota* (Chandler, 1978) have been associated with the latter. More work on these obscure fungi may produce interesting results. Probably many species feed on hyphae in the ground.

Trifourkis (1977) gives a thorough review of the available records for the 39 Mycetophilid species that he reared. A similar survey of the records for other species would be an invaluable base-line for future research. A summarised list for British species has been published by Chandler (1979a). A summary list of fungi and other habitats with the species reared from them is presented here for the groups included in this volume. Discussion on the associations of Mycetophilidae (and other Diptera) and fungi and the problems of defining and categorising these associations are given in Eisfelder (1954, 1955), Buxton (1961), Trifourkis (1977) and Hackman & Meinander (1979).

Lists of fungi and other habitats with associated Mycetophilidae

There follows a systematic list of the genera of fungi referred to in the text. This largely follows Pegler (1973) and Henderson, Orton & Watling (1969) and is the system adopted in a guide to British fungi by Kibby (1979). The associated Mycetophilidae (excluding Mycetophilinae) are listed on the right side of the page.

ASCOMYCOTINA

Discomycetes
Pezizales
HUMARIACEAE
Aleuria

Docosia gilvipes

PEZIZACEAE

Peziza

Pyrenomycetes

Sphaeriales

XYLARIACEAE

Bulgaria

Hypoxylon Ustulina

Xvlodon

BASIDIOMYCOTINA

Holobasidiomycetes

"Hymenomycetes"

Agaricales

BOLETACEAE

Boletus

PAXILLACEAE

Paxillus

PLEUROTACEAE

Pleurotus

TRICHOLOMATACEAE

Armillaria

Clitocybe

Collybia

Flammulina

Marasmius

Tricholoma

Xeromphalina

CORTINARIACEAE

Cortinarius

Hebeloma

Pholiota

Ripartites

Rozites

STROPHARIACEAE

Hypholoma

COPRINACEAE

Coprinus

Lacrymaria

AGARICACEAE

Agaricus

Docosia gilvipes

Sciophila hirta

Sciophila lutea, Rondaniella

Sciophila lutea, Docosia gilvipes

Apolephthisa

Bolitophila cinerea, maculipennis, rossica, hybrida, Sciophila lutea, Coelophthinia thoracica, Rondaniella, Docosia gilvipes

Bolitophila saundersii. maculipennis.

pseudohybrida, hybrida.

Mycomya marginata, Sciophila lutea,

Rondaniella

Bolitophila maculipennis

Bolitophila tenella, cinerea, Sciophila hirta, Rondaniella, Docosia gilvipes

Bolitophila saundersii, glabrata, pseudohybrida, hydrida, Docosia gilvipes

Bolitophila cinerea, Sciophila hirta, lutea,

Leia bimaculata

Bolitophila cinerea

Bolitophila cinerea

Bolitophila saundersii, pseudohybrida,

hybrida, Docosia gilvipes

Bolitophila fumida

Bolitophila tenella, ? hybrida

Bolitophila cinerea, saundersii, Leia

bimaculata

Bolitophila tenella. cinerea. hybrida.

Sciophila hirta. lutea. Phthinia

winnertzi, Rondaniella

Bolitophila pseudohybrida

Bolitophila hybrida

Bolitophila tenella, cinerea, saundersii, Mycomya wankowiczii, Docosia gilvipes

Bolitophila hybrida, Docosia gilvipes

Bolitophila cinerea

Mycomya prominens, Neoempheria

Bolitophila cinerea. pseudohybrida.

hvbrida

AMANITACEAE Amanita

RUSSULACEAE Lactarius

Russula

Aphyllophorales POLYPORACEAE

> Bjerkandera Coriolus

Daedaleopsis
Daedalia
Fomes
Grifola
Heterobasidion
Heteroporus
Lenzites
Meripilus
Piptoporus
Polyporus

Polystictus Poria

Pseudotrametes Tyromyces

HYMENOCHAETACEAE

Inonotus Phellinus

GANODERMATACEAE

Ganoderma

THELOPHORACEAE

Thelophora

SPARASSIDACEAE

Sparassis

STEREACEAE Stereum

Chondostereum

MERULIACEAE
Merulius
Phlebia

Bolitophila cinerea, saundersii, hybrida, Docosia gilvipes

Bolitophila hybrida, Sciophila hirta, lutea, Docosia gilvipes

Bolitophila pseudohybrida, Sciophila lutea, Rondaniella, Leia bimaculata, Docosia gilvipes

Ditomyia, Keroplatus, Cerotelion, Neoempheria, Leptomorphus

Rondaniella, Docosia gilvipes

Orfelia (s.s.), Mycomya marginata, Sciophila buxtoni, hirta, lutea, Acnemia nitidicollis, Rondaniella, Ectrepesthoneura hirta, Docosia gilvipes

Sciophila buxtoni Sciophila hirta, lutea Sciophila rufa, Rondaniella Sciophila lutea, Rondaniella

Rondaniella Sciophila lutea Sciophila hirta Sciophila lutea Docosia gilvipes

Bolitophila occlusa, Sciophila rufa,

Rondaniella, Docosia gilvipes

Docosia gilvipes

Mycomya marginata, wankowiczii, ?wrzesniowskyii, ?Monoclona, Apolephthisa, Tetragoneura

Sciophila buxtoni, lutea

Bolitophila occlusa, Tetragoneura

Ditomyia

Mycomya winnertzi, ?Sciophila ochracea,

lutea

Mycomya winnertzi, Sciophila hirta

Mycomya prominens

Mycomya marginata, Rondaniella

Mycomya cinerascens, marginata, wankowiczii, Sciophila hirta, lutea, Coelosia tenella, Rondaniella, Cerotelion lineatus

Mycomya marginata

Cerotelion lineatus, Mycomya marginata Mycomya marginata, Apolephthisa

CONIOPHORACEAE

Serpula Cerotelion lineatus

CORTICIACEAE

Peniophora Diadocidia

CANTHARELLACEAE

Cantharellus Sciophila hirta

GOMPHACEAE

Gomphus Sciophila lutea

HYDNACEAE

Hydnum Sciophila interrupta, lutea, Coelophthinia

Dacrymycetales
CALOCERACEAE

Calocera Leia bimaculata

"Gasteromycetes"

Phallales

PHALLACEAE

Phallus Mycomya wankowiczii

Sclerodermatales

SCLERODERMATACEAE

Scleroderma Docosia gilvipes

Phragmobasidiomycetes

Auriculariales

AURICULARIACEAE

Auricularia Mycomya marginata, duplicata, Sciophila

hirta, Docosia gilvipes, Cerotelion

lineatus

Tremellales

TREMELLACEAE

Sebacina Mycomya marginata

Below is a list of the other habitats mentioned in the text of the keys with the fungus-gnats that have been associated with them. The rotting wood category covers a variety of situations including living on the surface, under bark, with or without encrusting fungi, on the ground or aerial branches, etc. There is little doubt that the larvae of many species reared from rotting wood feed on fungi in the wood.

Rotting wood Diadocidia spp, Macrocera spp, Macro-

rryncha, Keroplatus, Cerotelion, Platyura, Orfelia, Mycomya, Syntemna, Acnemia, Monoclona, Phthinia, Apolephthisa, Boletina flaviventris,

Ectrepesthoneura, Manota.

Turf, grass tussocks Macrocera spp, Orfelia spp

Under boulders Orfelia
Worm tunnels Orfelia

Mosses & liverworts Orfelia, ?Neuratelia, Gnoriste, Boletina

spp

Caves, etc Macrocera spp, Speolepta

Mammal & bird nests Leia bifasciata, piffardi, Docosia gilvipes,

carbonaria, fumosa

Parasites and disease

Records of Hymenopterous parasites are scarce and many are based on rather loose associations of Mycetophilid and Hymenopteran. The few catalogues of relevance to the British fauna are, in the main, old and unreliable and the extraction of records is time consuming. There are, however, some groups which appear to be closely associated with Mycetophilidae. Thus, in the Ichneumonidae the Oxytorinae (= Microleptinae) and Orthocentrinae are usually associated with fungus-gnats. Several Braconidae (Ichneumonoidea), Proctotrupidae and Diapriidae (Proctotrupoidea) have been reared. The families Pteromalidae, Eulophidae and possibly Encyrtidae (all Chalcidoidea) may contain Mycetophilid parasites, as may Eucoilidae and Figitidae (Cynipoidea), Hyperparasites of the family Ceraphronidae (Ceraphronoidea) have been reared from Sciaridae and may occur on Mycetophilidae. Trifourkis (1977) lists many rearing records and Masner (1968) has recently reviewed the Proctotrupidae associated with Mycetophilidae.

A thorough survey of the available records would provide valuable data for those studying either the parasites or the flies. Since many of these records may not be very reliable every attempt should be made to check the identifications and to get further records by careful rearing. Richards' (1977) introduction to the order Hymenoptera gives keys to families and access to relevant literature. Askew (1971) includes general information on the biology of parasitic Hymenoptera.

Almost nothing is known or readily accessible about other parasites and pathogens. Mycetophilidae have been experimentally infected with iridescent virus, but there are no records of such infection in the wild. Madwar (1937) found nematodes in *Diadocidia*.

Parasitic fungi are probably not uncommon, but a superficial survey of recent reviews of the subject produced no records from Mycetophilidae. Large numbers of adult Mycetophilidae (*Rymosia fasciata* Meigen) attacked by the entomophagous fungus *Verticillium ?lucanii* have been found on the walls of a cave in Wales (A. M. Hutson, unpublished).

Steinhaus (1963) may be useful as an introduction to general studies of diseases and pathogens.

The fossil record

Mycetophilidae form a very large proportion of the fossil Diptera, particularly from amber, but the literature is very scattered—there is no catalogue and little review of literature. Rohdendorf (1964 [1974]) has given an overall review of our knowledge of fossil Diptera, However, in a subject which is very prone to alternative interpretation of specimens, age of origin and of unreliability of published descriptions and figures, he has (perhaps justifiably) relied mainly on the material available to him in Russia. According to Rohdendorf the oldest Diptera are described from the Upper Triassic (c.200 million years before present) and over half of these are recognisable as closely related to Mycetophilidae, but in two groups that are now extinct. There is little material from early and late Jurassic, but a good sample from Middle Jurassic (mostly from Karatau-a deposit which is usually considered to be of rather later origin than this). The main group from the Triassic is strongly represented in (early and) middle Jurassic along with several other groups. Few data have been published for the Cretaceous period—material from this period is now available, but has not been studied in detail yet. By the Tertiary, from which a large number of species have been described, all the early groups have been replaced by the currently recognised groups (discussed in this handbook as subfamilies). Among the material described from this period it is interesting to note the presence of groups that are now very small and rare, such as the Manotinae and Lygistorrhininae. Other authors who have published on fossil Mycetophilidae include Loew, Meunier, Handlirsch, Edwards and Hennig (e.g. Hennig, 1954).

This necessarily short review deals with a subject that is greatly in need of more investigation—a synthesis of the available information and literature on Mycetophilidae would provide valuable background studies for an assessment of contemporary forms.

Collecting and preserving

The section on general behaviour and habitats gives an idea of where to find Mycetophilidae. Sweeping sites mentioned there and any nearby low vegetation will produce many specimens, which can be supplemented by a variety of traps. Further details of techniques and equipment for collecting can be obtained from Stubbs & Chandler (1979) and Cogan & Smith (1974). Careful rearing of larvae to adults provides valuable data on life-cycles and some suggestions for rearing from fungi follow.

Rearing Diptera from fungi

Collected fungi should be kept isolated, either as individual sporophores or as accurately identified conspecific samples, to prevent cross-contamination. A plastic box can be used as a rearing cage; the box should be ventilated but proofed against oviposition by flies from the outside, and containing some fine sterilized peat. The peat will absorb some of the products of decay of the fungus and serve as a pupation site for the larvae. A high humidity should be maintained by frequent spraying with non-chlorinated water. The depth of the peat can be varied from shallow to deep in the box, providing a moisture gradient for the larvae to choose their pupation site. Decomposed fungi should be removed after the larvae have emerged, but the box should be kept intact and regularly inspected for emergences for at least six months. Fungi of various ages should be collected and their condition noted since some Mycetophilidae prefer older, even deliquescent fungi. Other features, such as fungal size, consistency, surface area, substrate, position are also worth noting as these may be factors limiting the choice of host fungus of apparently polyphagous flies. Odour and chemistry may be more difficult to record.

If detailed information on host choice is required, detailed records of emergence should be kept for each sample. If associations of immature stages are required, some of the fungus should be examined at the time of collection and examples of each larval type present separated and preserved. When adult flies emerge the associated pupal exuvium should be collected and this will usually have the larval head capsule attached. Remember that one sporophore may support several species of Mycetophilid.

Supplementary information can be obtained by the use of emergence traps over individual sporophores or groups of sporophores. It is also possible to trap those flies visiting specific sporophores.

Preparation of specimens

All specimens collected should be carefully labelled with locality (including grid reference), date, collector and any available information on ecology.

Adults collected by sweeping or trapping may be stored satisfactorily in 70-80% alcohol; many characters, particularly the genitalia and many of the smaller bristles are easier to see in this medium. However, some workers dislike alcoholic material

and there is a danger of the alcohol evaporating. It is probably easier to review and extract data from a collection that is pinned, although a pinned collection is more prone to damage or attack by pests. If material is stored in spirit the label should be large enough not to move when the tubes are in transit, as movement will damage the specimens. The tube should be filled completely with alcohol, a plug of cotton wool or equivalent placed in the end and the tube then inverted in a larger wide-mouthed jar of alcohol. This allows easy topping-up of alcohol levels and the jars can be used for grouping species, genera, etc.

For pinning Mycetophilidae a micropin should be pushed through the thorax, either laterally (i.e. through the pleurae) or vertically (i.e. down through the mesonotum). Care should be taken not to damage any of the taxonomic characters on both sides of the thorax. The micropin should then be fixed into a small strip of *Polyporus*, Plastozote or equivalent. Alternatively the specimen may be glued on its side to the tip of a card triangle, with the legs protected by the card. A stainless steel pin should then be used to support these mounted specimens and their labels. It is useful to mount all specimens uniformly, e.g. facing the left, to facilitate comparison.

It may be necessary to remove the genitalia and clear them in order to see the structures clearly. Cut off the end of the abdomen, preferably at about the junction of the 5th and 6th segment and drop it into 10% Caustic Potash (KOH). Many specimens will clear in about an hour, but others will need much longer, and they can be left for up to 24 hours without harm. Then wash in distilled water and transfer to Glacial Acetic Acid for 5-30 minutes. They can remain in this medium for a day or so without damage. If possible they should then be stored in glycerine in small corked tubes on the same pin as the rest of the specimen. The cork can be sealed with paraffin-wax or nail varnish, etc. This method enables the specimen to be examined from any angle without distortion, but has the disadvantage that, since the specimens usually have to be removed from the tube for examination, they are not so readily viewed as when mounted on slides. If mounting on slides is preferred the genitalia should be transferred from Glacial Acetic Acid to Clove Oil for half an hour and then mounted in Canada Balsam. They can be mounted separately on standard 8×2.5 cm (= 3×1 in) glass slides remembering to label as the rest of the specimen and number both parts so that they can be individually associated. A preferable alternative is to mount the genitalia under a small coverslip onto a small piece of clear rigid perspex which can be pinned with the rest of the specimen. So long as they are mounted on clear perspex they can be examined from above and below, with transmitted or direct lighting, but specimens frequently get distorted. If genitalia are mounted it may sometimes be necessary to separate some parts, e.g. dististyle, aedeagus, tergite 9, etc., from the main part of the genitalia, but this should be avoided if at all possible.

Larvae and pupae should be stored in alcohol or mounted on slides.

If specimens have a special association, e.g. male and female taken in copulation or larvae and pupae associated with reared adults, ensure that this is clear from the labels.

Morphology

This section can be used as an introduction to the external morphology of the family but is primarily intended to discuss and illustrate all the structures used in the keys that follow. The naming of parts is still in a state of flux, particularly the structure of the thoracic sclerites, wing veins and genitalia. The differences in nomenclature of these parts is largely due to various views on the homologies of the

structures. The aim of this work is to facilitate identification of specimens and to encourage work on the biology of an ecologically important group of Diptera. To this end the terminology has been kept as simple as possible while adopting a system that is a compromise between earlier works on the family, particularly in Britain (e.g. Edwards, 1925), the introductory *Handbook* to the Diptera (Oldroyd, 1970) and such modern opinion as is now considered undisputed. In the venation the use of "Median fork" and "posterior fork" is an attempt to avoid conflicting opinions. The most recent review of Mycetophilid morphology is by Vockeroth (1980) in a work that has made a serious attempt to discuss and unify the nomenclature of the structures of Diptera.

HEAD (figs. 5-17). Generally convex behind, but closely adpressed to the thorax, occasionally (Manotinae) more flat behind. Top of head lower than the level of the front of the thorax. Antennae (figs. 5-8) arising at the middle height of the head, occasionally (Manotinae) higher. Antennae consisting of distinct scape, pedicel and multisegmented flagellum. The flagellum usually consists of 14 segments, sometimes less (to 9 in some Cordyla), occasionally more (one extra in some Keroplatinae). Flagellum sometimes laterally flattened, almost serrate (some Keroplatinae), very elongate in Macrocera (Keroplatinae). Flagellar segments usually covered in dense short hairs, sometimes with stouter or longer hairs, the arrangement of hairs usually constant for all flagellar segments, but sometimes varying (e.g. Macrocera). Segments usually distinct, but divisions difficult to distinguish towards apex in some Macrocera. Length of antennae varying from little longer than head to several times the length of the body. Some sexual dimorphism in length (e.g. Macrocera) or bristling (e.g. Bolitophila saundersii).

Ocelli usually three, middle one usually reduced, sometimes absent. Lateral ocelli varying from within their own diameter of the mid-line of the head to immediately adjacent to the eye margin. Middle ocellus usually set a little in front of lateral ocelli, sometimes all three in line across head. Postvertex usually with bristles, sometimes of two kinds. Eyes usually reniform being emarginated around the base of antennae, but always well separated from each other. Eyes with inter-ommatidial hairs, sometimes short or sparse. Clypeus usually short, slightly elongate in Asindulum and Macrorrhyncha (Keroplatinae), very elongate in Gnoriste (Sciophilinae). Maxillary palps usually of five segments, but first (basal) two segments often very short and obscured. Number of palpal segments reduced in some genera, e.g. Keroplatus in which only one segment is clearly visible (but actually three segments present). Labrum usually small, but elongate in Antlemon, Macrorrhyncha, Asindulum and Gnoriste. Prementum usually very short and broad, elongate in Antlemon, Macrorrhyncha and Asindulum; labellae one or two segmented, small in Antlemon and most other genera, but large and fleshy in Asindulum and Macrorrhyncha.

THORAX (figs. 18-23). Usually stout and deep, more delicate in such genera as Bolitophila, Coelophthinia, Phthinia and Speolepta, but still with hump-backed appearance in lateral view. Pronotum reduced anteriorly, usually remaining as well developed lateral prontal lobes, which are more or less divided into an anterior and posterior pronotum. Below the pronotum and separating it from the front coxae is the propleuron (or prosternum or proepisternum), a small sclerite which may not be well separated from the pronotum. These sclerites carry bristles of various lengths and strengths and their position and number is of importance in some groups. Just behind the pronotum, on a weakly sclerotised area, lies the prothoracic spiracle, which has soft hairs completely surrounding the aperture, but may also have a row of stronger bristles associated with it.

The mesonotum is usually broad with the traces of a prescutal suture evident at sides at the humeri. The surface may be densely covered with fine short bristles sometimes with some longer bristles, or may have a pair of median bare stripes that

start at the front and taper away towards the rear of the mesonotum and a pair of lateral bare stripes that usually start at about the level of the remains of the prescutal suture and run back to the rear of the mesonotum. These bare stripes are occasionally very narrow. The arrangement and naming of the remaining bristles is illustrated in fig. 19. The scutellum may bear a small number of large bristles, or these may be interspersed with small bristles, or the larger bristles may be absent. The postnotum may bear bristles and the arrangement of these bristles varies.

Behind the prothorax and fore-coxa is the mesepisternum, which is usually clearly divided into an upper anepisternum and a lower katepisternum (or sternopleuron). Behind the mesepisternum and below the wing base is the pteropleuron (or mesepimeron). Behind this is the pleurotergite (or laterotergite). The pleurotergite is usually large and bulbous; it lies on either side of the postnotum (or mediotergite or mesophragma) and between the wing base and the metathoracic spiracle and haltere. Between the sclerite that bears the haltere and the hind coxa is the hypopleuron (or metepisternum). Bristles may be present towards the top of the anepisternum, the anterior margin of the katepisternum, the pteropleuron, the pleurotergite and the anterior edge of the hypopleuron. Some of the bristles may be extremely small and fine. Other bristles are present in some groups, but their taxonomic value has not been fully investigated.

LEGS (figs. 24-25): Coxae long and broad, modified only in some Mycomya spp. with a dense brush of setae on the inner anterior surface at the distal end of the forecoxa or with a spur of variable length at the same site on the mid coxa. Front tibia with a preapical brush on the anterior surface and a single apical spur. Mid and hind tibiae usually with a pair of apical spurs, but these may be reduced or absent in some Keroplatinae. Mid-tibia of some Sciophilinae has a sensory pit usually surrounded by dense soft hairs on the dorsal surface. Hind tibia sometimes with an apical comb of bristles on the posterior side. Tibial bristles variable in size and arrangement, fine tibial setae arranged irregularly or in distinct rows for at least part of the length of the tibia. Tarsi usually simple, 2nd and 3rd (and occasionally 1st or 4th segments swollen beneath on the fore-legs of the females of some species. Tarsal claws often simple, more frequently with teeth or with pectinate basal lobe. Empodium present or absent, pulvilla absent.

WINGS (figs. 46-49). Usually clear, sometimes patterned. Pattern usually involves wing tip and/or area of r-m. Heavily sclerotised areas of wing base variably blackened or pale (e.g. axillary sclerite above distal end of squama in Syntemna spp.) Further investigation of this area would be fruitful. Surface of wing membrane more or less covered with microtrichia, macrotrichia or some of each. However, these terms are not used accurately in this family. Macrotrichia are articulated hairs while microtrichia are unarticulated hair-like outgrowths of the cuticle. The term microtrichia as used in Mycetophilidae undoubtedly includes small macrotrichia, particularly in the Sciophilinae. An examination of the distribution of true microand macrotrichia may help towards a better classification of this subfamily. For the purpose of this work microtrichia are relatively small structures, not clearly distinguishable at less than 10× magnification, while macrotrichia are larger obvious wing hairs. Where there is any possibility of confusion other characters are available. Both macrotrichia and microtrichia are present in Ditomyiinae, Diadocidiinae, some Macrocera, Sciophilinae, etc. Macrotrichia are usually directed away from the wing base (decumbent), but sometimes they are erect or directed back towards the wing base (reflexed). Microtrichia are arranged randomly or tending to form distinct rows, especially towards apex of the Median and posterior forks (Mycetophilinae).

Vein Sc is absent, short or long, ending in Costa (Sc1 present), in Radius (Sc1 absent) or ending free. R1 simple, R2 + 3 absent, R4 + 5 usually simple, if R4

differentiated it is usually short, either near the base of Rs enclosing the "small cell" (some Sciophilinae) or ending near to or beyond the apex of R1 (Bolitophilinae, Keroplatinae, Ditomyiinae); long in Ditomyiinae, short in others. Usually a short cross-vein connecting Rs with the veins of the Median fork (r-m), but this cross-vein replaced by a short fusion in Keroplatinae. Veins of Median fork connected to veins posterior fork by a posterior cross-vein (Ditomyiinae, Keroplatinae, Diadocidinae, most Bolitophilinae), by a short fusion (some Bolitophilinae) or connected only at wing base in other subfamilies. In those groups in which the Median fork connects to the posterior fork the basal cell enclosed may be partly divided by a trace of the basal extension of the Median vein (e.g. Bolitophilinae, some Ditomyiinae and Keroplatinae). One of the veins of the Median or posterior fork sometimes weak or absent or disconnected at base. The posterior fork set back to base of wing in Ectrepesthoneura (similar to Sciaridae). Anal vein variable in length or absent. Macrotrichia always present on Radial veins, present or absent on Sc and postradial veins. One or two weak axillary veins or folds, never with macrotrichia, but with a black spot on the fork in some Leia spp. and Rondaniella. Anal area of wing usually rounded, but somewhat right-angled in Macrocera or virtually absent in some long-winged species (e.g. Bolitophilia, Phthinia).

ABDOMEN. Generally narrow at base and broadest near middle, broadest towards apex in some Keroplatinae. Any patterning of abdomen often more strongly developed in male. Usually 8 unmodified pre-genital segments in male, reduced to seven in some Sciophilinae and Ditomyiinae and 6 in some Sciophilinae. Male genitalia partly rotated in *Diadocidia* and some Sciophilinae (this rotation often responsible for the reduction in the number of unmodified pre-genital segments). Female with 7 unmodified pre-genital segments. Some further sexual dimorphism in bristling of abdomen, particularly obvious in dense patches of setae on sides of male abdomen of *Orfelia modesta*.

Male genitalia (figs. 50-53) with tergite 9 usually s. ongly modified and with a single or paired accessory plate (cerci) at the distal end in some Ditomyiinae and Sciophilinae (notably *Mycomya* and *Boletina*). Below tergite 9 is a pair of soft lobes (tergite 10) above a single lobe—these form the proctiger which surrounds the anus. The pair of basistyles usually form the bulk of the genitalia, but are of limited taxonomic use (although the apodemes from the basistyle are characteristic in some groups, e.g. *Bolitophila*). The dististyles are articulated at the apex of the basistyles and these are frequently very complex and are of wide application in the taxonomy. The aedeagus and parameres are of great value in the identification of many groups, but can normally only be seen in cleared specimens.

The ovipositor (figs. 54-61) is relatively simple, but provides useful features at the species level in most groups. In many groups there is insufficient closely associated material for females to be reliably described. In some groups where association of males and females seems satisfactory and where identification otherwise relies heavily on male genitalia some descriptive notes on the ovipositor have been given as an aid to identification. Tergite 8 is unmodified, 9 usually reduced, 10 weak and membranous or absent. Cerci usually two-segmented, sometimes only one-segmented. The shape and arrangement of the segments of the cerci are often useful features for identification. Sternum 8 is usually lengthened and very variable in shape. It is frequently partly divided medially forming two lateral lobes. Incrassations around the median division may be characteristic. The apical margin may carry spines or occasionally scales. Sternite 9 reduced. Usually two spermathecae, which vary in size, shape and sclerotisation.

Notes on the keys

This is a large and diverse family, some species show a notable range of variation and imperfections are not uncommon. It has not been possible to allow for all anomalies in these keys. It is therefore not expected that people unfamiliar with the family will always be able to identify isolated specimens reliably. Potential users of this work are recommended to have a variety of specimens available to demonstrate the range of variation in important characters. This can be done either by building up a quantity of material before attempting identification or by reference to a recognised named collection.

The keys are largely based on the work of Edwards (1925). Into this framework has been incorporated additional species, additional characters and revised opinions of relationships. However, the information included here has been very restricted and so there is little doubt that the extra background information in Edwards' work will ensure that it remains very useful. After Edwards' death the family received relatively little attention in Europe until in recent years such authors as Burghele-Balacesco, Chandler, Gagné, Hackman, Lastovka, Matile, Plassmann, Stackelberg, Tuomikoski and the present authors have made taxonomic contributions. Work on the Russian fauna has been published by Ostroverkhova (=Plotnikova) and Stackelberg and in Japan by Okada, Saigusa and Sasakawa. Details of the Nearctic fauna are accessible from Vockeroth (1980) and Laffoon (1965). In the keys to small genera it is hoped that sufficient information will be available for students to be able to recognise the fact if they have a species that does not appear in the key. Similarly it is hoped that sufficient information is presented for taxonomists working on a broader front to recognise possible misinterpretations of species in Britain. In the larger genera, it is not always possible to ensure this, but the greater number of characters in a longer key may help. The figures of male genitalia should confirm identification of that sex, although these may not be sufficient for the separation of closely related groups of species. In most groups the females also show fairly reliable characters of the ovipositor but they have not all been discussed in much detail here. This is partly because the association of female with known males is not always reliable. The most obvious gap here is the large genus Mycomya, in which of 33 species, the females of 9 species are unknown, the females of several species are uncertainly associated and several groups have so far proved inseparable. Thus no key to Q Mycomya is provided, the key being divided primarily by the strong secondary sexual characters present in some males.

The keys are not necessarily phylogenetic, but are arranged for the convenience of identification of the British fauna as known.

Localities are given by county. In most cases the county names have not been converted to the current county district names and there may even be overlap where both names are given. Distribution is summarised for some commonly recorded species. Phenology is given by months of occurrence (in the form of Roman numerals), but this may include some records of distinctive larvae (e.g. Speolepta) or bred material, which may spread the dates of occurrence beyond the natural range of the adults. Host fungi are given by genus, but this tells little of the habitat preference and host range that a thorough analysis by host species would give. No thorough search of the literature for faunistic records has been attempted—most of the records included here are of material examined by current workers on the family. The other published records should be confirmed where possible.

Original draft keys were produced by L. N. Kidd. These were checked and revised or rewritten by A. M. Hutson. The introduction and notes preceding each key were prepared by A. M. Hutson. The figures were mainly prepared by D. M. Ackland, but A. M. Hutson prepared those for the introduction, for *Neoempheria* and the

Sciophilini, and those for *Diadocidia* were taken from Laštovka and Matile (1972). The cover illustration and fig. 1 are by Stephen Falk.

Acknowledgements

Copies of parts of the manuscript in various stages of preparation were distributed to P. J. Chandler (U.K.), L. Matile (Paris) and J. R. Vockeroth (Ottawa) and we are grateful for their comments and for additional information. These Dipterists have also been very generous with specimens at their disposal. Many other entomologists have been most helpful in providing information and loan of specimens, including many types. Among these should be mentioned C. Dahl (Lund), R. Danielsson (Lund), R. J. Gagné (Washington), W. Hackman (Helsinki), J. Ismay (Oxford), A. Kaltenbach (Vienna), P. Laštovka (Prague), B. Lindeberg (Helsinki), E. Plassmann (Senckenberg), A. Stackelberg (Leningrad), J. L. Steylik (Brno), W. Tobias (Senckenberg).

Geoffrey Kibby, Department of Entomology, British Museum (Natural History), has provided considerable help with fungus questions.

Check list

The following check list corrects and updates that in Kloet & Hincks (1976). The subfamily Mycetophilinae is not included in the keys that follow and is therefore omitted from this check list. A total of 204 species in 47 genera are included here.

MYCETOPHILIDAE

BOLITOPHILINAE

MESSALA Curtis, 1836 S. BOLITOPHILA s.s. basicornis (Mayer, 1951) cinerea Meigen, 1818 saundersii (Curtis, 1836) spinigera Edwards, 1925 tenella Winnertz, 1863

S. CLIOPISA Enderlein, 1936 bimaculata Zetterstedt, 1838 dubia Siebke, 1863 disjuncta Loew, 1869 fumida Edwards, 1941 glabrata Loew, 1869 hybrida (Meigen, 1804) fusca Meigen, 1818 maculipennis Walker, 1836 occlusa Edwards, 1913 pseudohybrida Landrock, 1912 triangulata Edwards, 1941 rossica Landrock, 1912

DIADOCIDIINAE DIADOCIDIA Ruthé, 1831 ferruginosa (Meigen, 1830) spinosula Tollet, 1948 valida Mik, 1874

DITOMYIINAE

DITOMYIA Winnertz, 1846 fasciata (Meigen, 1818)

SYMMERUS Walker, 1848
PLESIASTINA Winnertz, 1852
annulatus (Meigen 1830)
zonata Stephens, 1838
ferrugineus Walker, 1848
vittata (Walker, 1856)

KEROPLATINAE

MACROCERINI

MACROCERA Meigen, 1803 anglica Edwards, 1925 angulata Meigen, 1818 aterrima Stackelberg, 1945 bipunctata Edwards, 1925 grandis: Edwards 1913, nec Lundstroem, 1912 centralis Meigen, 1818 crassicornis Winnertz, 1863 estonica Landrock, 1924 fasciata Meigen, 1804 fascipennis Staeger, 1840 fastuosa Loew, 1869 longibrachiata Landrock, 1917 lutea Meigen, 1804 maculata Meigen, 1818 parva Lundstroem, 1914

phalerata Meigen. 1818 propleuralis Edwards, 1941 pusilla Meigen, 1830 ? multicincta Curtis, 1837 stigma Curtis, 1837 stigmoides Edwards, 1925 tusca Loew, 1869 vittata Meigen. 1830 ? dorsalis Curtis, 1837 zetterstedti Lundstroem, 1914

KEROPLATINIMACRORRHYNCHA Winnertz,

1846 flava Winnertz, 1846 rostrata: (Edwards, 1913), nec (Zetterstedt, 1851)

ASINDULUM Latreille, 1805 nigrum Latreille, 1805

KEROPLATUS Bosc, 1792 testaceus Dalman, 1818

CEROTELION Rondani, 1856 humeralis (Zetterstedt, 1850) lineatus (Fabricius, 1775) laticornis (Meigen, 1818)

PLATYURA Meigen, 1803 marginata Meigen, 1804 atrata (Fabricius, 1805) rufipes Meigen, 1818

ORFELIA Costa, 1857 PLATYURA: auctt., nec Meigen, 1803

S. ISONEUROMYIA Brunetti, 1912 semirufa (Meigen, 1818) unicolor (Walker, 1856) vitripennis (Walker, 1856)

S. MONOCENTROTA Edwards, 1925 lundstroemi (Edwards, 1925)

S. NEOPLATYURA Malloch, 1928 biumbrata (Edwards, 1913) flava (Macquart, 1826) modesta (Winnertz, 1863) nigricauda (Strobl, 1893)

S. ORFELIA s.s. discoloria (Meigen, 1818) fasciata (Meigen, 1804) nemoralis (Meigen, 1818) flavipes (Meigen, 1818) nigricornis (Fabricius, 1805) antica (Walker, 1856)

pallida (Staeger, 1840) aestivalis (Winnertz, 1863) tristis (Lundstroem, 1911) unicolor (Staeger, 1840)

S. PYRATULA Edwards, 1929 perpusilla (Edwards, 1913) zonata (Zetterstedt, 1855) concisa (Walker, 1856)

S. RUTYLAPA Edwards, 1929 ruficornis (Zetterstedt, 1851) pectinifera (Edwards, 1913)

S. URYTALPA Edwards, 1929 atriceps (Edwards, 1913) macrocera (Edwards, 1913) ochracea (Meigen, 1818) dorsalis (Staeger, 1840) mycetophiloides (Walker, 1856) nigriceps (Walker, 1856)

ANTLEMON Loew, 1871 servulum (Walker, 1837)

SCIOPHILINAE

MYCOMYINI MYCOMYA Rondani, 1856 britteni Kidd, 1955 cinerascens (Macquart, 1826) circumdata (Staeger, 1840) lucorum (Winnertz, 1863) clavigera (Lundstroem, 1912) collini Edwards, 1941 digitifera Edwards, 1925 duplicata Edwards, 1925 exigua (Winnertz, 1863) ? nigra: (Walker, 1837), nec (Macquart, 1826) fimbriata (Meigen, 1818) cingulata: (Walker, 1837), nec (Meigen, 1804) affinis: (Dziedzicki, 1885), nec (Staeger, 1840) flava (Winnertz, 1863) ruficollis: (Walker, 1856), nec (Zetterstedt, 1852) flavicollis (Zetterstedt, 1852) fuscata (Winnertz, 1863) hyalinata (Meigen, 1830) incisurata (Zetterstedt, 1838) ? annulata: (Walker, 1948), ? nec (Meigen, 1818) kingi Edwards, 1941 lambi Edwards, 1941 marginata (Meigen, 1818) maura (Walker, 1856) melanoceras Edwards, 1924 nigricornis: (Lundstroem, 1909), nec (Zetterstedt, 1852)

ornata (Meigen, 1818) parva (Dziedzicki, 1885) pectinifera Edwards, 1924 nigricornis: (Dziedzicki, 1885), preocc. prominens (Lundstroem, 1913) punctata (Meigen, 1804) rosalba Hutson, 1979 tenuis (Walker, 1856) trilineata (Zetterstedt, 1838) ferruginea: (Walker, 1837), ? nec (Meigen, 1818) trivittata (Zetterstedt, 1838) marginata (Dziedzicki, 1885), preocc. tumida (Winnertz, 1863) vittiventris (Zetterstedt, 1852) wankowiczii (Driedzicki, 1885) winnertzi (Dziedzicki, 1885) fasciata: (Walker, 1856), nec

NEOEMPHERIA Osten-Sacken, 1878 bimaculata (von Roser, 1840) lineola (Meigen, 1818) pictipennis (Haliday, 1833) winnertzi Edwards, 1913 pictipennis: (Winnertz, 1863), nec (Haliday, 1833)

SCIOPHILINI

EUDICRANA Loew, 1869 nigriceps (Lundstroem, 1909)

(Zetterstedt, 1838)

? wrzesniowskii (Dziedzicki, 1885)

ALLOCOTOCERA Mik, 1886 pulchella (Curtis, 1837)

LEPTOMORPHUS Curtis, 1831 walkeri Curtis, 1831

POLYLEPTA Winnertz, 1863 guttiventris (Zetterstedt, 1852) borealis: (Walker, 1856), nec (Zetterestedt, 1852) undulata Winnertz, 1863 splendida: Verrall, 1901, nec Winnertz, 1863

NEURATELIA Rondani, 1856 ANACLINIA Winnertz, 1863 nemoralis (Meigen, 1818) elongatus (Walker, 1848) nigricornis Edwards, 1941

ANACLILEIA Meunier, 1904 PARANEUROTELIA Landrock, 1911 dispar (Winnertz, 1863) SYNTEMNA Winnertz, 1863 LOEWIELLA Meunier, 1894 hungarica (Lundstroem, 1912) nitidula Edwards, 1925 stylata Hutson, 1979

PARATINIA Mik, 1874 sciarina Mik, 1874

PHTHINIA Winnertz, 1863 humilis Winnertz, 1863 winnertzi Mik. 1869

AZANA Walker, 1856 anomala (Staeger, 1840) scatosoides Walker, 1856

MEGALOPELMA Enderlein, 1911 nigroclavatum (Strobl, 1910) jenkinsoni (Edwards, 1913)

SCIOPHILA Meigen, 1818 LASIOSOMA Winnertz, 1863 adamsi Edwards, 1925 buxtoni Freeman, 1956 cliftoni Edwards, 1925 fenestella var. Edwards, 1913 fenestella Curtis, 1837 fridolini Stackelberg, 1943 geniculata Zetterstedt, 1838 hirta Meigen, 1818 interrupta (Winnertz, 1863) limbatella Zetterstedt, 1852 sharpi Edwards, 1913 lutea Macquart, 1826 f. analis (Winnertz, 1863) nigronitida Landrock, 1925 nigra (Landrock, 1912), nec Macquart, 1826 nonnisilva Hutson, 1979 ochracea Walker, 1856 plurisetosa Edwards, 1921 quadriterga Hutson, 1979 rufa Meigen, 1830 varia (Winnertz, 1863)

ACNEMIA Winnertz, 1863 amoena Winnertz, 1863 nitidicollis (Meigen, 1818) defecta (Walker, 1856) longipes Winnertz, 1863

MONOCLONA Mik, 1886 rufilatera (Walker, 1837) halterata (Staeger, 1840) unicornuta (Dziedzicki, 1884)

GNORISTINI SPEOLEPTA Edwards, 1925 leptogaster (Winnertz, 1863) COELOPHTHINIA Edwards, 1941 thoracica (Winnertz, 1863)

COELOSIA Winnertz, 1863 flava (Staeger, 1840) ? flava (Walker, 1837) silvatica Landrock, 1918 tenella (Zetterstedt, 1852)

DZIEDZICKIA Johannsen, 1909 marginata (Dziedzicki, 1885)

PALAEODOCOSIA Meunier, 1904 alpicola (Strobl, 1894) flava (Edwards, 1913) janickii (Dziedzicki, 1923)

GNORISTE Meigen, 1818 bilineata Zetterstedt, 1852 longirostris Siebke, 1874

GRZEGORZEKIA Edwards, 1941 collaris (Meigen, 1818)

APOLEPHTHISA Grzegorzek, 1885 subincana (Curtis, 1837) melanoceras (Walker, 1856)

BOLETINA Staeger, 1840 basalis (Meigen, 1818) brevicornis Zetterstedt, 1852 digitata Lundstroem, 1914 dispecta Dziedzicki, 1885 dubia (Meigen, 1804) analis (Meigen, 1818) inermis Lundstroem, 1912 flaviventris Strobl, 1894 reuteri Lundstroem, 1906 gripha Dziedzicki, 1885 griphoides Edwards, 1925 groenlandica Staeger, 1845 lundstroemi Landrock, 1912 moravica Landrock, 1912 nasuta (Haliday, 1839) lundbecki; Edwards, 1913, nec Lundstroem, 1912 nigricans Dziedzicki, 1885 nigrofusca Dziedzicki, 1885 nitida Grzegorzek, 1885 pallidula Edwards, 1925 flaviventris: Landrock, 1924, nec Strobl, 1894 pectinunguis Edwards, 1932 plana Walker, 1856 rejecta Edwards, 1941 sciarina Staeger, 1840 silvatica Dziedzicki, 1885 trispinosa Edwards, 1913 trivittata (Meigen, 1818)

SYNAPHA Meigen, 1818 EMPALIA Winnertz, 1863 fasciata Meigen, 1818 paradoxa (Edwards, 1913) vitripennis (Meigen, 1818) finalis (Walker, 1856)

LEUNI

RONDANIELLA Johannsen, 1909 dimidiata (Meigen, 1804) terminalis (Meigen, 1818) elegans (Winnertz, 1863) variegata (Winnertz, 1863)

LEIA Meigen, 1818 GLAPHYROPTERA Winnertz. 1863 arsona Hutson, 1978 bifasciata Gimmerthal, 1846 bilineata (Winnertz, 1863) trimaculata (Strobl, 1910) strobli Landrock, 1925 bimaculata (Meigen, 1804) octomaculata Curtis, 1837 f. fasciola Meigen, 1818 crucigera Zetterstedt, 1838 cylindrica (Winnertz, 1863) fascipennis Meigen, 1818 flavicornis Meigen, 1818 piffardi Edwards, 1925 subfasciata (Meigen, 1818) winthemi Lehmann, 1822

MEGOPHTHALMIDIA Dziedzicki, 1889

crassicornis (Curtis, 1837) helvola (Walker, 1856) valida (Walker, 1856)

TETRAGONEURA Winnertz, 1846 sylvatica (Curtis, 1837) compressa (Walker, 1856)

ECTREPESTHONEURA
Enderlein, 1911
MEUNIERIA Johannsen, 1909,
preocc.
colyeri Chandler, 1979
hirta (Winnertz, 1846)
aliena (Walker, 1856)
pubescens (Zetterstedt, 1860)

DOCOSIA Winnertz, 1863 carbonaria Edwards, 1941 fumosa Edwards, 1925 fuscipes (Roser, 1840) gilvipes (Walker, 1856) moravica Landrock, 1916

villosa Landrock, 1912

pallipes Edwards, 1941 sciarina (Meigen, 1830) pubescens (Walker, 1837) basalis (Walker, 1856) valida Winnertz, 1863 setosa Landrock, 1916 sp. indet. MANOTINAE MANOTA Williston, 1896 unifurcata Lundstroem, 1913

Key to subfamilies

The only subfamily that does not occur in Britain is the Lygistorrhininae. This subfamily contains a single tropical genus in which Rs arises at the base of the wing and the proboscis is very long. General works of relevance to the European fauna are Landrock (1926-1927), Edwards (1925), Bei-Bienko (1969). Many species also occur in the Nearctic fauna, which is covered by Vockeroth (1980) and Laffoon (1965).

1	Posterior fork and stem of Median fork connected by a cross-vein or fused for a short distance (figs. 46, 62-74)
	Veins of median and posterior forks separate to wing base (figs. 47-49, 75-87)
2	R4 present and long, generally at least half as long as R5; Sc short and ending free (figs. 65-66)
_	R4 less than half as long as R5, sometimes weak or absent; Sc almost always long and ending distinctly in Costa (fig. 46)
3	Media and Radius fused for a short distance (figs. 46, 67-74) KEROPLATINAE (p.29)
_	Cross-vein <i>r-m</i> present (figs. 62-64)
4	Posterior cross-vein and r-m in line across wing; Media without a basal section (fig. 64)
_	Posterior cross-vein well before r-m; Media with basal section distinct (figs. 62-63) BOLITOPHILINAE (p.26)
5	Prothorax with distinct long, strong bristles; head convex behind; orbital bristles not
,	forming a conspicuous projecting row; antennae inserted at about middle of height of
	head (figs. 11-12); stem of median fork present, either the anterior or posterior
	branch of median fork may be disconnected at the base, but never both together6
	Prothorax without strong bristles; head flat or slightly concave behind, with a row of
	projecting orbital bristles which are more or less curved backwards; antennae inserted
	near top of head (fig. 13); stem of median fork absent, the branches present as
	detached veins in the distal part of the wing
6	Microtrichia of wings irregularly arranged; Sc usually long (figs. 47-49) (shorter in
U	Tetragoneura and Megophthalmidia (Leiini), which have irregular tibial setae and
	lateral ocelli remote from eye margin); lateral ocelli usually far from eye margin
	(fig. 17) (except in Eudicrana (Sciophilini), Docosia and some Leia (Leijni) in which
	tibial setae are irregular and Sc long); fine tibial setae usually irregularly arranged
	(fig. 32, 34, 40-41) (except in Mycomyiini, in which Sc long, R4 present); R4 present
	(figs. 47-48) (Mycomyilni and some groups in other tribes) or absent (fig. 49):
	pleurotergum bare (Mycomyiini and some groups in other tribes) or haired (figs. 20,22)
	SCIOPHILINAE (p.40)
	Microtrichia of wings in more or less definite lines, particularly near posterior wing
	margin (not so clear in <i>Trichonta</i> and <i>Phronia</i>); Sc short (longer in <i>Trichonta</i>);
	lateral ocelli touching eye margin; fine setae of tibia in regular rows; R4 absent;
	pleurotergum haired
	(N.B. the Mycetophilinae are not included in this volume).
	(11.2. the injectophimme are not metaded in this volume).

Subfamily Bolitophilinae

As currently understood this subfamily contains the single Holarctic genus *Bolitophila* with about 60 species. *Arachnocampa* from New Zealand and Australia was thought to belong here, but is now generally regarded as a genus of Keroplatinae. Other suggestions of the occurrence of the group in Australia await confirmation.

All the species that have been reared have been found in soft fruiting bodies of fungi.

Genus **Bolitophila** Meigen (figs. 6, 14, 24, 54-55, 62-63, 88-101)

This is currently separated into two subgenera (or even genera), but this division is probably artificial. The separation is based on one obvious character, is not supported by any other character in the adults, and recent examination of larvae (Trifourkis, 1977) produces no evidence for a natural division in the genus. They are all drab, grey-brown, elongate species with a wing length of about 5.5mm (4-7mm).

The subgenus *Bolitophila* comprises about 20 species, of which 12 are known from Europe. They breed mainly in fungi related to *Hypholoma* (Strophariaceae) or *Pholiota* (Cortinariaceae). The larvae are gregarious. The adults of many species frequently hibernate in caves and cellars. About 40 species are included in the subgenus *Cliopisa* Enderlein. About 20 of these are European breeding mainly in Agaricales, less frequently in Polypores and Boleti.

Stackelberg (1969a) includes most of the European species. See also Hutson & Kidd (1971, 1974), Chandler (1978), Matile (1976), Plassmann (1971b, 1975).

Key to species.

- 4 Stigma faint, grey; pubescence of σ antennae barely as long as diameter of segments; segments 2 and 3 of fore tarsus of female swollen, 2 a little longer than 3; σ genitalia fig. 90; Q ovipositor, fig. 54, with apex of sternum 8 evenly tapered cinerea Meigen Very common and widespread. i-xii. Mainly from Hypholoma and Pholiota, but also from Agaricus, Amanita, Armillaria, Boletus, Collybia, Flammulina, Hebeloma, Lacrymaria, Marasmius.
- 5 No dark spot over r-m (fig. 62); dististyle of σ genitalia with prominent apical teeth directed inwards (fig. 91); segments 2 and 3 of \$\times\$ fore tarsus swollen and of equal

- Cumbria, Yorks; Scotland: Argyll, Ayr, Dumfr, Inverness, Perth. v-vii, x
 Anterior vein of posterior fork very faint at base, but traceable; mesonotum uniform, unstriped; dististyle of σ genitalia rounded apically with the apical point at right angles to the inner surface of dististyle (fig. 95); ovipositor with sternum 8 very truncate, last segment of cerci elongate oval, broadest at its mid point
- Scotland: Inverness. vi. Xeromphalina.
- 9 Wings with dark spot over r-m. 10

 Wings without a dark spot over r-m. 11
- Dististyle of or genitalia with a single beak-like apical tooth (fig. 97); segments of Q cerci distinctly longer than broad......bimaculata Zetterstedt Scotland: Inverness. v.
- 11 Mesonotum usually shining; postnotum orchreous with a black apical spot; posterior branch of posterior fork strongly curved at tip; dististyle of or genitalia with a pair of weak apical points (fig. 98); Q front tarsal segments 2, 3 and 4 more or less equally short and swollen; Q Sternum 8 rounded apically, terminal segment of cerci round, basal segment nearly quadrate with a shallowly concave apexglabrata Loew Beds, Berks, Herts, Notts, Yorks. viii-x. Clitocybe.

- 13 Posterior branch of posterior fork normal; dististyles of o genitalia with a single weak apical point (fig. 100); Q sternum 8 tapered evenly; terminal segment of cerci very

small and round, penultimate segment only shallowly concave apically

Subfamily Ditomyiinae

A small subfamily of less than 100 species in about eight genera, found in the Palaearctic, Nearctic, Neotropical and Australasian regions. Stackelberg (1969b) and Zaitzev (1978) include keys to the western Palaearctic species, Saigusa (1973) has additional species and genera from Japan.

Key to genera

- Eyes rounded; R4 much longer than R4 + 5 (fig. 65); an episternum and postnotum bare......DITOMYIA Winnertz (p.28)

Genus Symmerus Walker

(Figs. 9, 66, 102)

A Holarctic genus of 13 species, two of which occur in Europe. A single species is recorded from Britain. Munroe (1974) includes a revision of the genus, excluding the species described by Saigusa (1973).

Postnotal bristles brown; wing length 5-7mm; σ genitalia (fig. 102) with Sternum 9 not markedly constricted at mid length, the lateral margins straight; σ cerci covered with short pale setae
 Derby, Devon, Gloucs, Hants, Heref, Herts, Hunts, Kent, Lancs, Oxon, Staffs, Yorks; Wales: Glamorgan, Montgomery; Scotland: Argyll. ν-νiii. In rotting logs.

Genus **Ditomyia** Winnertz (Figs. 65, 103)

A Holarctic and Neotropical genus of about 5 species, two of which occur in Europe. One species recorded from Britain.

Subfamily Diadocidiinae

Apart from the genus *Diadocidia*, a genus of one species from Chile is included in this subfamily. Two other southern hemisphere genera have been included in this

subfamily, but are usually included in the separate family Sciaridae. Lastovka & Matile (1972) revised the Holarctic species of *Diadocidia*. See also Hutson & Kidd (1974), Chandler (1978).

Genus **Diadocidia** Ruthé (Figs. 10, 26-27, 64, 288-290)

A genus recorded from the Palaearctic, Nearctic, Neotropical and Australasian regions with less than ten described species. All three species recorded from Europe occur in Britain. The larvae may be found in mucous tubes under rotting logs and have been associated with the fungus *Peniophora*. The adults are fairly stout yellowish-brown flies with greyish wings.

Two subgenera have been recognised by Lastovka & Matile (1972) in the Holarctic fauna.

Key to subgenera and species

Subfamily Keroplatinae

A large subfamily of worldwide distribution. Larvae are web-spinners on the underside of hard fungi, under bark, rotting wood, in caves and even in worm-holes in pasture. They are more or less predacious, eating small invertebrates that get caught in the sticky strands of their webs, which are liberally supplied with drops of clear fluid containing oxalic acid. In species in caves and deep forest, the sticky strands may be suspended vertically to catch flying prey. The most spectacular cases of this are the cave "glow-worms" of New Zealand and Australia, Arachnocampa spp. One Tasmanian species, Planarivora insignis Hickman, has become an internal parasite of terrestrial planarians. The adults of many species feed at flowers.

This is generally regarded as a primitive and diverse subfamily and this has

resulted in the description of a large number of small genera and subgenera, but it is only when the world fauna as a whole is better known that the relative status of these groups can be properly evaluated. In the British fauna this particularly affects Orfelia and its relatives and for the purposes of this handbook these groups have been treated in a very conservative fashion, based on the earlier works of Edwards. Some of the subgenera of Orfelia have already been raised to generic rank and others may follow suit; for uniformity, they have been kept as subgenera here, but have been included in the following generic key. Matile is currently investigating the generic classification of this group.

Stackelberg (1969c and 1969d) discusses most of the European species. See also Hutson & Kidd (1974), Chandler (1978).

Key to genera and subgenera

1	Posterior fork slightly approximated near its base, then divergent (fig. 67); anal angle of wing angular; tibial bristles absent; anepisternum with some hairs
_	MACROCERA Meigen (p.31) Posterior fork divergent from base; anal angle of wing rounded; tibial bristles usually present, even if small.
2	Mouth parts elongate, at least as long as head; pleurotergite bare; R4 ending in Costa3 Mouthparts not elongate
3	Labellae small, but prementum and labrum elongate (fig. 16); a few prospiracular hairs present; postnotum bare; anepisternum bare; tibial setae irregular
	Labellae greatly enlarged and fleshy (fig. 15)
4	Prospiracular bristles absent; a patch of short hairs on anepisternum; postnotum bare; Anal vein nearly reaching wing margin; claws large with several basal spines; Costa not
_	extending beyond R5; Sc long, ending in Costa ASINDULUM Latreille (p.34) Prospiracular bristles present; anepisternum bare; postnotum with a few short setae; Anal vein not nearly reaching wing margin; claws small, at most with one basal spine; Costa extending beyond R5; Sc short and ending free (fig. 68)
_	MACRORRHYNCHA Winnertz (p.34)
5	Palpi reduced, with only one swollen visible segment (but actually 3 segments present); antennae stout and strongly laterally compressed (fig. 8)
-	Palpi normal, with 3-4 distinctly visible segments (but actually five segments present); antennae not strongly lateral compressed
6	R4 ending in R1 (fig. 46), pleurotergites hairy KEROPLATUS Bosc (p.34)
7	R4 ending in Costa (fig. 69); pleurotergites bare CEROTELION Rondani (p.35) R4 ending in R1; Media with a distinct fold like basal extension (fig. 70); pleurotergites and postnotum bare; empodium present
_	R4 ending in Costa; basal extension of Media indistinguishable; empodium absent8
8	Pleurotergites hairy; each tibia with a single (posterior) spur, anterior spurs absent; at most only a few posteroventral bristles on hind tibia; no prospiracular hairs; postnotum bare; mesonotum with bare stripes; anepisternum with very short hairs above; Anal vein almost reaching wing margin (fig. 71); branches of Median and posterior forks bare; tibial setae irregular at base, in more regular rows towards tip
_	Pleurotergites bare; mid and hind tibia with a pair of spurs, although the anterior spur may be minute (Rutylapa spp, fig. 38); usually some anterior and dorsal bristles on hind tibia (Orfelia Costa s.l.)
9	Branches of median and posterior forks with close-set small macrotrichia; postnotum usually bare (one or two small hairs sometimes present in <i>Pyratula</i>); mesonotum
	with bare stripes (sometimes inconspicuous in <i>Isoneuromyia</i> and <i>Neoplatyura</i>) (fig. 19); tibial bristles well developed (fig. 39)

10 Prospiracular bristles present; tibial setae somewhat irregular, but tending to form distinct rows towards the apex (fig. 33); anepisternum usually bare; Anal vein strong and reaching wing margin (fig. 73)......ORFELIA (NEOPLATYURA Malloch) (p.36) Tibial setae in regular rows throughout, all rows alike; Anal vein strong and almost reaching wing margin (fig. 72); an episternum bare ORFELIA (ISONEUROMYIA Brunetti) (p.36) 12 Anal vein strong and reaching wing margin.... ORFELIA (URYTALPA Edwards) (p.37) Anal vein abbreviated, not reaching wing margin ORFELIA (PYRATULA Edwards) (p.37) 13 Prospiracular bristles absent; about 6 rows of tibial setae much more closely set than others, appearing as conspicuous black lines; anepisternum bare; anterior spur on mid and hind tibia well developed; Anal vein weak ORFELIA (ORFELIA Costa) (p.38) Prospiracular bristles present; all rows of tibial setae alike, anepisternum usually with a few short hairs; anterior spur of hind and mid tibia minute; Anal vein strong ORFELIA (RUTYLAPA Edwards) (p.38)

Genus Macrocera Meigen

(Figs. 5, 67, 104-125)

A worldwide genus of about 150 species, that is sometimes put with about 4 other small genera into a separate tribe of the Keroplatinae. About 45 species occur in the western Palaearctic. The adults are distinctive in usually having very long fine antennae and they normally rest with the wings slightly spread. They may occur at flowers. Immature stages are poorly known, but they have been reared from clumps of turf, on rotting wood and on cave walls. The larvae are predaceous, although some start as scavengers.

Key to species

1	Wings with only microtrichia
2	Wings quite unspotted, or at most with a faint cloud in the base of the posterior fork 3
	Wings with at least one small spot around base of Rs and/or at wing tip
_	Sc ending above tip of basal cell
4	Thorax with mesonotum striped; a markedly smaller species, about 3mm long; σ antennae about 1.5× length of body, Q antennae slightly longer than body; wing length 3.5mm; σ genitalia (fig. 104)pusilla Meigen
	Herts, Hunts, Yorks. viii-ix.
_	Thorax uniformly ochreous except for a pair of small dark spots on front margin of mesonotum and an ill-defined dark pleural stripe; a relatively larger species, about 4mm long; σ antennae about $3 \times$ body length, Q antennae about $1.5 \times$ body length; wing length 4-4.5mm; σ genitalia (fig. 105)
5	Abdomen with black apical bands; mesonotum with three shining red-brown stripes; large elongate species, about 8mm; wing length 6.5mm; or genitalia (fig. 106)
	longibrachiata Landrock Devon. viii.
_	Abdomen almost unicolorous or with pale apical bands6
6	Abdominal tergites with conspicuous pale apical bands
_	Abdomen almost unicolorous yellow or black

7	Lateral stripes of mesonotum largely obsolete, but their front ends turned down and connected with a dark brown patch on and above the notopleuron; median stripe much widened at front margin; propleurae blackish; vein R1 thickened at tip (but to a lesser extent than vittata); wing length about 5.0mm; c unknown propleuralis Edwards
_	Devon. vi. Lateral stripes well developed and well separated from dark pleural markings; vein R1
•	not thickened at tip
8	Antennae with conspicuous pale rings; median thoracic stripe much widened at front margin; propleurae black; wing length 5-5.5mm; o genitalia (fig. 107)
	Cumbria, Gloucs, Herts. vi-x.
-	Antennae at most faintly ringed; median thoracic stripe with no more than normal widening at front margin; propleurae yellow
9	Posterior fork usually clear, occasionally with a slight trace of a spot in the basal constriction; thoracic bristles strong, scutellar bristles about twice as long as depth of scutellum; Costa usually extended to about 1/3 of distance from R5 to Median fork; or tergites 1-6 pale apically; or genitalia (fig. 108) with two strong spines at apex of dististyle
	Berks, Bucks, Chesh, Cornwall, Devon, Hants, Heref, Herts, Hunts, Kent, Lancs, Lincs, London, Middx, Notts, Staffs, Suffolk, Surrey, Sussex, Westmorland,
	Yorks; Wales: Brecon, Caerns, Flint, Merioneth; Scotland: Aberdeen, Argyll,
	Inverness, Rhum; Ireland: Down, Kerry, Wicklow. v-x. [N.B. Tergite 9 may be truncate or distinctly bilobed. This may be a complex of species.]
_	A more or less distinct spot in the basal constriction of the posterior fork; thoracic
	bristles weaker, scutellar bristles barely as long as depth of scutellum; Costa extended beyond end of R5 to at most about 1/4 of the distance to Median vein. At least of tergites 6 and 7 completely dark; of genitalia (fig. 109) without apical spines to dististle
10	Berks, Cumbria, Kent, Lancs, Oxon, Sussex; Wales: Gwynnedd. vii-x. or antennae more than twice as long as body; wing length 5-7mm
11	or antennae not much longer than body length; wing length 4-5.5mm
	Berks, Bucks, Chesh, Derby, Devon, Dorset, Hants, Heref, Hunts, Kent, Lancs, Lincs, Norfolk, London, Staffs, Suffolk, Surrey, Sussex, Warwick, Westmorland, Yorks; Wales: Brecknock, Glamorgan, Merioneth; Scotland: Argyll, Arran, Banff,
_	Inverness, Sutherland; Ireland: Clare, Laois, Wicklow. v-ix. Mesonotum uniformly yellowish; thoracic bristles yellowish and weak; basal extension
	of Media strong and distinct, dividing the basal cell into two; vein R1 not thickened at tip; wing length 5-6mm; or genitalia (fig. 111)
12	R1 thickened at tip; R4 usually short and almost vertical; wing length about 5mm; or genitalia (fig. 112)
	Scotland: Inverness, Ross & Crom; Ireland: Kerry. vii, x. R1 not thickened at tip; R4 longer, curving into Costa
13	or antennal flagellum with first seven segments well differentiated, the other segments
13	uniformly narrow, covered with bristles and difficult to differentiate (fig. 5); or dististyle with stout bristles confined to basal third (fig. 113); wing length 4-5.5mm
	Derby, Hants, Herts, Lincs, Norfolk, Westmorland, Yorks; Wales: Flint,
	Gwynedd; Scotland: Aberdeen, Inverness, Perth, Ross & Crom, Sutherland;
	Ireland: Laois, v-viii.
_	or antennal flagellum with only the first six segments well differentiated; or dististyle with strong bristles on basal three-quarters; or genitalia (fig. 114) var. of parva Lundström
	Scotland: A hordeen Invernees Porth vii

14	Wings with tip clear
1.6	Wings with tip dark (faintly so in maculata)
15	into posterior fork and towards Costa, a small cloud at base of Rs and on Costa at
	tip of R1; wing length 4.5-6mm; σ genitalia (fig. 115)
	Berks, Cambs, Chesh, Cumbria, Devon, Derby, Gloucs, Hants, Heref, Herts, Kent,
	Lancs, Lincs, Middx, Norfolk, Oxon, Suffolk, Surrey, Sussex; Wales: Brecknock,
	Flint, Glamorgan, Gwent, Gwynedd; Scotland: Aberdeen, Moray, Sutherland;
	Ireland: Kildare, Wexford, Wicklow. v-ix.
_	Abdominal segments with pale basal bands; a small dark cloud at base of R5 and over
	base of Rs, no dark Costal spot; wing length about 6mm; or genitalia (fig. 116)bipunctata Edwards
	Cumbria, Lancs. vii.
16	Abdomen largely yellow, the tergites with black apical bands; wings with central fascia
	completely crossing wing
_	Abdomen unicolourous dark or with the apices of the segments paler
17	Larger species; thorax dark; R4 long; wing markings darker without spot in base of
	posterior fork; wing length 7mm; or genitalia (fig. 117)
	Devon. viii.
_	Smaller species; thorax light; wing markings paler, but with a spot near base of posterior fork; wing length 4.5-6mm; σ genitalia (fig. 118)angulata Meigen
	Bucks, Chesh, Cumberland, Derby, Devon, Dorset, Essex, Hants, Heref, Herts,
	Kent, Lincs, Middx, Norfolk, Notts, Oxon, Suffolk, Sussex, Westmorland, Yorks;
	Wales: Brecknock, Gwynedd; Ireland: Kerry. v-ix.
18	Wings with central fascia completely crossing wing; a separate dark spot on wing
	margin between veins of posterior fork; mesonotum unmarked; wing length about
	5.5mm; o genitalia (fig. 119)
_	Wings with the central fascia incomplete; dark spots usually present at ends of veins of
	posterior fork, but not between them; wing markings often very faint; mesonotum
	with three blackish stripes; wing length 4-5.5mm; or genitalia (fig. 120)
	3 4 3 4 7
	maculata Meigen
10	Cambs, Herts, Somerset, Suffolk, Surrey. v-x.
19	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \(\sigma\) genitalia
19	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; or genitalia (fig. 121)zetterstedti Lundström
19	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \(\sigma\) genitalia (fig. 121)
19 	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; or genitalia (fig. 121)
_	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \(\sigma\) genitalia (fig. 121)
_	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \(\sigma\) genitalia (fig. 121)
_	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \(\sigma\) genitalia (fig. 121)
_	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; or genitalia (fig. 121)
_	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \$\sigma\$ genitalia (fig. 121)
_	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; or genitalia (fig. 121)
_ 20	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \$\sigma\$ genitalia (fig. 121)
_ 20	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \(\sigma\) genitalia (fig. 121)
_ 20	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \(\sigma\) genitalia (fig. 121)
	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \(\sigma\) genitalia (fig. 121)
_ 20	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \(\sigma\) genitalia (fig. 121)
	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \$\sigma\$ genitalia (fig. 121)
	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \$\sigma\$ genitalia (fig. 121)
	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \$\sigma\$ genitalia (fig. 121)
	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \(\sigma\) genitalia (fig. 121)
	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \$\sigma\$ genitalia (fig. 121)
	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \(\sigma\) genitalia (fig. 121)
	Cambs, Herts, Somerset, Suffolk, Surrey. v-x. R4 absent; wings clear; wing length 4-4.5mm, a small montane species; \$\sigma\$ genitalia (fig. 121)

Cambs, Chesh, Devon, Hants, Heref, Kent, Lancs, Lincs, Middx, Norfolk, Notts, Suffolk, Surrey, Sussex, Westmorland, Yorks; Wales: Brecknock, Merioneth; Scotland: Aberdeen, Arran, Banff, Inverness, Moray, Ross & Crom; Ireland: Clare, Dublin, Kerry, Laois, Wicklow. v-ix.

Genus Asindulum Latreille

(Figs. 15, 131)

A Holarctic genus of one European and Russian species, one Israeli and one Nearctic species. The biology is unknown and the adults are rare. Matile (1975) revised the Palaearctic species.

Genus Macrorrhyncha Winnertz

(Figs. 68, 132)

A holarctic genus of two Nearctic species and eight Palaearctic species, of which only one is known from Britain. Other species may well occur, but were not found in about 100 specimens examined in this study. Adults can be found at umbellifer flowers and have been bred from rotting wood. Matile (1975) revised the Palaearctic species.

Genus Keroplatus Bosc

(Figs. 46, 128)

At present this genus contains four European, four Nearctic and two African species. Other species may belong in this genus, but their status needs review. Keroplatus included groups that are now regarded as separate genera, such as Cerotelion Rondani. Too many areas remain unexamined in the light of more recent taxonomic opinion for any meaningful indication of the distribution and number of species on a world scale. However, this and its closely related genera form a fairly small and distinct group. Apart from the four recognised European species a number of other species have been described in this genus, but these species have remained unrecognised so far through inadequate description.

The adults may be found around rotting logs. The larvae live under a large irregular, flat, mucilaginous net on the underside of logs, usually with polyporaceous fungi. They feed partly on the fungus and partly on other animals. They are flattened and somewhat luminous. The pupa develops in a distinct dry cocoon.

 A large pale brown species with brown markings towards the leading edge and tip of the wings; abdomen uniform pale brown, sometimes with darker brown basal bands or lateral patches; wing length 6-8.5mm; or genitalia (fig. 128) testaceus Dalm Berks, Hants, Oxon, Sussex, Warwick, vi-x, Under rotting logs, usually with Polyporaceous fungi. Has also been recorded from Stereum, Serpula and Auricularia.

Genus Cerotelion Rondani

(Figs. 8, 69, 126-127)

A small genus of four Palaearctic, one Nearctic and an uncertain number of species from elsewhere in the world (see Keroplatus). Adults may be found around rotting logs and have been found flying around and hanging onto spiders' webs. The cylindrical larva lives in a net under logs, particularly in association with Polyporaceous fungi. From an irregular, 3-dimensional net with a small mucous area for the larva's protection, it forages partly on fungal spores and partly as a predator. The pupa develops under a dense net, but without a distinct cocoon. Matile (1969) discusses the European species.

Key to species

- 1 Scutellum with a more or less single row of bristles, some distinctly stronger than others; shaded area of wing-tip distinctly demarcated, without a clear spot, at most narrowly connected to the cloud proximal to the apex of the posterior fork; central wing spot distinct (above base of median fork); hind coxae at most with the tip blackish; or tarsal claws small and inconspicuous, Q slightly larger, both with a single basal Beds, Berks, Derby, Hants, Heref, Herts, Kent, Lancs, London, Surrey, v-x. Under rotting logs, especially encrusted with Polyporaceous or other encrusting fungi. Sometimes in houses possibly associated with rv-rot fungus Merulius lacrymans.
- Scutellum with a broad band of dense uniformly short bristles; shaded area of wing tip diffuse and with a clear spot at the tip of R5 and connected along wing margin to large shaded area proximad of the apex of posterior fork; central wing cloud faint or absent; hind coxae blackish at base on outer surface; or tarsal claws very large, pectinate for their whole length, Q claws smaller with three or four basal teeth; wing length 4.5-6.5mm; or genitalia (fig. 127)humeralis Zetterstedt Somerset. [This record is based on a single specimen in the collection of A. E. J. Carter which could not be found during the present studyl.

Genus Platyura Meigen

(Figs. 70, 129)

One western and one eastern Palaearctic species and 7 Nearctic species. This genus was formerly known under the generic name Apemon Johannsen. Adults occur in a wide variety of habitats but are never common. The larvae live under logs in a large web and are voracious predators. The pupa develops in a thick mass of web but without a distinct cocoon.

- A large dark species; Sc extending beyond base of Rs; head, thorax and abdomen black, abdomen often with reddish basal bands to tergites 3-5; wings with apical and posterior areas slightly smokey, anterior part yellowish, usually with a slightly darker patch at base of Median fork; wing length 6.5-9mm; or genitalia (fig. 129)
 - marginata Meigen Bucks, Hants, Heref, Gloucs, Kent, Lancs, Middx, Staffs, Surrey, Sussex; Wales: Gwynedd, Montgomery. v-viii. Under rotting logs.

Genus Monocentrota Edwards

(Figs. 71,130)

One Palaearctic species, three African and possibly one Indian and one Neotropical species. This genus has been considered a subgenus of *Orfelia* Costa and possibly does not merit exclusion from that genus any more or less than many of the subgenera still included in it. Its biology is unknown.

Genus Orfelia Costa

This is the genus that was known as *Platyura* for a very long time. It is a large worldwide genus, split into about 20 subgenera. Some groups that have been included as subgenera, such as *Monocentrota* Edwards, are now regarded as separate genera and others of the groups here regarded as subgenera may be (or sometimes have been) similarly elevated. The Nearctic, many of the Neotropical and indeed some of the European species have not yet been assigned to subgenera. Thus comments on the distribution and status of the subgenera must be regarded as provisional. Six subgenera are included here and these cover all the subgenera recorded from Europe. The key to subgenera is included in the generic key on p. 30.

The adults of some species have been found on flowers, particularly Umbelliferae, and some occur in rather more open, dry habitats than most Mycetophilidae. The larvae live in webs under bark of logs or under fallen wood itself, under boulders, in or under moss, among grass roots below the soil surface and in worm-burrows in pasture. The net is irregular, usually flat but may invade cavities. The threads of the net are well supplied with drops of fluid containing oxalic acid, which attacks any animals that contact it. There is a central main tube of thread that the larva moves along and from which it forages to animals trapped by the net.

Subgenus Isoneuromyia Brunetti

(Figs. 72,133)

The subgenus was based on an Oriental species. Most of the 12 or so species currently included are from the Oriental region with others from New Zealand, Neotropical region, Australia, West Indies and Europe. Nothing is known of their breeding biology.

Subgenus Neoplatyura Malloch (Figs. 33, 73, 134-137)

(Figs. 33, 73, 134-137)

This subgenus was based on a Nearctic species and includes a number of species from most parts of the world. Four species are recorded from Europe and all occur in Britain.

Key to species

Subgenus Pyratula Edwards

(figs. 39, 138-139)

A small group of two European species. Their life history is unknown. They are small dark species.

Key to species

Subgenus Urytalpa Edwards (Figs. 18-19, 140-142)

Apart from the three European species included here, only one Oriental species has been included in this subgenus. Largish species, variable in colour.

Key to species

- Costa reaching only 1/4 or less of distance from tip of R5 to tip of anterior branch of Median fork; wings often with a small faint grey spot at tip; thorax dark (σ) to pale with or without stripes (Q); wing length 5-6.5mm; σ genitalia (fig. 142)

Subgenus Rutylapa Edwards

(Figs. 38, 143)

Four Ethiopian, five Oriental and one European species have been included in this subgenus. Nothing is known of their life history.

A medium sized species with orange thorax; abdominal tergites dark (Φ) or yellow with dark apical and lateral borders, the apical band tending to develop slightly along the mid-dorsal line (Φ); wing length about 4mm; Φ genitalia (fig. 143)
 ruficornis Zetterstedt

Hants, Oxon. vii.

Subgenus Orfelia Costa (Figs. 56-57, 74, 144-150)

About 20 species are recorded in this subgenus from the Palaearctic, Nearctic, Neotropical and Oriental regions. One European species has recently been found in New Zealand, but is probably the result of an introduction. Most of the European species can only be identified satisfactorily by reference to the male genitalia. Most of the other characters given in the following key should be treated with caution and the females of the yellow species have not been properly identified for lack of suitably associated males. Also isolated females of nigricornis may be quite orange and run to fasciata; no way has yet been found for confidently separating the females of these two species.

Key to species

- - Berks, Cambs, Chesh, Devon, Essex, Hants, Herts, Kent, Lancs, London, Middx,

Norfolk, Oxon, Surrey, Sussex, Wilts; Wales: Brecknock; Scotland: Dumfries, Inverness, Kincardine, Perth; Ireland: Armagh, Cork, Kerry, Kildare, Wicklow. v-vii. Wings with preapical dark band; palpal segments more elongate, especially terminal segment; o dististyles much longer than broad, divided into two styles; Q cerci 3 Subapical wing cloud clearly demarcated, dark; mesonotum usually with some yellow around border of three more or less confluent black stripes; Q mesonotum may be more orange with or without vague stripes; halteres usually orange; wing length 5-6mm; σ genitalia (fig. 145) nigricornis Fabricius Cambs, Chesh, Hants, Herts, Kent, Lancs, London, Sussex; Wales: Gwent; Scotland: Aberdeen, Inverness. v-vii. Subapical wing cloud not clearly demarcated from rest of wing, which is slightly smokey; body almost entirely black; halteres usually dark; wing length 4-5mm; or genitalia (fig. 146).....tristis Lundström Berks, Essex, Hants, Herts, Kent, London, Oxon, Surrey, Suffolk, Yorks. vi-viii. Anterior spur of mid-tarsus reduced, 1/4 to 1/3 length of posterior spur; wings with a very faint cloud at tip; or abdomen uniformly dark or tergites with narrow pale apical bands, ♀ abdomen dark or light; wing length 3.5-4.5mm; ♂ genitalia (fig. 147); Devon, Dorset, Hants, Lancs, Surrey; Ireland: Waterford. vi-vii. 5 or genitalia (fig. 148) with upper style sinuous and haired only at base, lower style narrow and bare, a tooth-like process on inner side of basistyle; of abdominal tergites yellow with black basal bands; Q cerci more than three times as long as broad at base; wings with distinct subapical dark band; wing length 4.5-5.5mm fasciata Meigen Beds, Berks, Dorset, Essex, Hants, Heref, Herts, London, Middx, Surrey (?Q: Kent, Sussex); Ireland; Wicklow, vi-viii, x. Moulds under wet bark. or genitalia with upper style less sinuous and with bristles at or near tip, lower style broad with long strong bristles along inner surface, no process on basistyle; Q cerci less than twice as long as broad at base (this last character may not be reliable as Wings usually with distinct preapical dark band; or abdomen usually unbanded, yellow; or genitalia (fig. 149) with hairs on upper style stronger, blacker and placed at tip; wing length 4-5mmunicolor Staeger Berks, Cambs, Chesh, Cumbria, Essex, Hants, Herts, Kent, Lancs, Sussex; Ireland: Armagh, Wicklow; (?Q: Surrey, Caerns, Kildare). vi-viii. Under Coriolus.

> Genus Antlemon Haliday (figs. 16, 151)

so conspicuous and well removed from tip; wing length 3.5-4mm

vi-vii.

Wings with preapical band pale to absent; σ abdomen with broad black bands at base of tergites, sometimes all dark; σ genitalia (fig. 150) with hairs on upper style not

Hants, Suffolk; Ireland: Wexford; (?Q: Bucks, Chesh, Herts, Surrey; Waterford).

A Palaearctic genus of three species. The adults may be found at flowers, particularly Umbelliferae. Immature stages are unknown. Apart from the elongate mouthparts this genus could easily be mistaken for an *Orfelia* and might run to subgenus *Pyratula* in these keys. A. halidayi has a much longer proboscis (about twice the length of the front coxa) and brevimanum differs in details of the genitalia. Matile (1977) has revised the genus and includes the British species and brevimanum in the subgenus Antlemonopsis Tollet.

 Small black or dark brown species; palps well developed; proboscis about same length as front coxa; wing length 2.5-3.5mm; or genitalia (fig. 151) only with very small

..... discoloria Meigen

Subfamily Sciophilinae

The subfamily Sciophilinae contains about one third of the British fauna. The subfamily is divided into 5 tribes, of which four are represented in the British fauna. The other tribe, the Metanepsiini, is only known so far from one genus in tropical Africa and Malaysia.

The two tribes Mycomyiini and Sciophilini are fairly easy to define, with certain exceptions noted in the key, but the other two tribes are not so clearly differentiated since they are diverse and contain a number of genera that show characters that do not clearly agree with the normal pattern of these groups. A review of this group may be required to provide a better classification for these genera.

The life histories of relatively few species of the Sciophilinae are known. Species have been reared from in or on the surface of fungi, under bark, in rotten wood, on liverworts, mosses and in the nests of birds and mammals. The species that live on fungi and under bark, etc, inhabit a small mucilaginous tube supported by retaining threads. Some species cover this with a dry irregular web and such webs may be large and communal. Some of the nest inhabitants entirely free-living. Those living on fungi feed mainly on fungal spores, while those under bark and in rotting wood are probably feeding on fungi associated with the substrate. The species on liverworts and mosses appear to be feeding on these green plants, a habit not known elsewhere in the family. The nest inhabitants are probably saprophagous.

Records of adult activity are limited to the few observations of species occurring at flowers. These records are scattered through the subfamily.

At the time of writing, a revision of the Palaearctic species of *Docosia* is in preparation by Lastovka and Matile. This will add many species to the known fauna of this genus. Other revisions of the genera *Ectrepesthoneura* (by Chandler, 1979) and *Syntemna* (by Hutson, 1979) are published, but there are no other revisions recently published that add substantially to the European fauna. In the same paper, Hutson also discusses the synonymy of various taxa of this subfamily.

Key to tribes

- Fine tibial setae irregularly arranged, at most slightly linearly arranged at apex (fig. 32); ocelli three (fig. 12) (except in *Eudicrana* in which the two ocelli are contiguous with the eye margin; in *Dziedzickia* and *Palaeodocosia* the middle ocellus may be reduced or even absent, but the tibial setae are irregular); empodium usually present. . 2
- Wings without macrotrichia; postnotum bare (except Coelophthinia which has no macrotrichia and an elongate sense organ on the dorsal surface of mid-tibia in both sexes)
 3

R1 short, usually little if any longer than r-m which is long and nearly horizontal;
 Median fork usually longer than its stem; Sc long or short (figs. 49, 85-87)

[N.B. R1 is rather long in *Rondaniella*, when the Median fork is hardly longer than its stem, and in *Docosia*, in which Sc is short, *r-m* is nearly horizontal.]

Tribe Mycomyini

A large worldwide tribe of about ten genera, the two largest and most widespread occurring in Britain. The tribe contains some of the more striking sexual dimorphisms in the family, shown in a simple fashion by the mid-coxal spur of the males of some of our *Mycomya* species. In other genera, such as *Echinopodium*, of South America, these outgrowths are very elaborate, while in the African *Syndocosia*, the wing tip of the male is curiously modified. Larvae of this group have usually been found in rotting wood, under bark or in Basidiomycete fungi, but the larva of one African genus, *Viridivora*, feeds on liverworts and mosses. Adult behaviour is virtually unrecorded.

The two genera included here are the only ones recorded from the Holarctic region.

Key to genera

- Wings usually with conspicuous markings; Costa usually continued at least a short distance beyond tip of R5, which does not quite reach wing tip; usually a more or less distinct, often vein-like, fold between R5 and anterior branch of Median fork; eyes not or scarcely emarginatedNEOEMPHERIA Osten-Sacken (p.45)

Genus Mycomya Rondani

(Figs. 17, 25, 31, 75, 152-183)

A genus of about 80 Palaearctic, 60 Nearctic and 80 Neotropical species, with smaller numbers in the other regions of the southern hemisphere. In the males of many species a long spur is developed on the inner apex of the mid-coxa and this curves up towards the body (fig. 31). The males of one group of species have obvious patches of dense setae on the fore-coxae. These features are never seen in the females. The spurs are not the result of a natural grouping, but are such an ideal key character that a key specifically for males is provided. Without these characters the females are not easy to separate, the females of many species are unknown or uncertainly associated and it has not been possible to find sufficient reliable distinction between some of the known species. For these reasons no guide to females is included here.

The purpose of the male adornments is unknown and adult activity is poorly recorded—they are usually found resting on leaves in woodland with their wings slightly divaricate. A few species have been bred from larvae in or on fungi or associated with fungi on dead wood. Those species that are known spin webs and the webs may be communal.

Again, exceptions to some of the characters in the key are to be expected in this genus and identification of many species can only be confirmed by reference to the male genitalia. It is likely that the female ovipositor will provide reliable characters, but little has been published and preliminary examination suggests some difficulties.

Key to species (males)

1	Mid-coxa with a spur (fig. 31)
_	Mid-coxa without a spur
2	Base of posterior fork below or before base of stem of Median fork; postnotum with
	or without bristles
_	Base of posterior fork well beyond base of stem of Median fork; postnotum with one
	to three bristles at tip
3	Coxal spur about half the length of coxa; wing length 3.5-5mm; genitalia (fig. 152)
	incisurata Zetterstedt
	Berks, Bucks, Chesh, Cornwall, Cumbria, Dorset, Essex, Hants, Heref, Herts, Kent,
	Lancs, Notts, Staffs, Suffolk, Surrey, Sussex, Westmorland, Worcs, Yorks; Wales:
	Brecknock, Merioneth, Montgomery; Scotland: Aberdeen, Arran, Inverness, Moray,
	Perth; Ireland: Antrim, Clare, Galway, Kerry, Kildare, Killarney, Wicklow. v-x.
_	Coxal spur more than half and usually more than total length of coxa4
4	Basitarsus of fore-leg shorter than tibia
_	Basitarsus of fore-leg at least as long as tibia (occasionally shorter in winnertzi)7
5	Thorax all yellow; Sc curved into R1 (Sc1 absent); wing length 3.5-4.5mm; genitalia
	(fig. 153)flavicollis Zetterstedt
	Devon, Dorset, Hants, Herts, Oxon, Surrey; Ireland: Kildare. vi-x.
-	Thorax dark, dorsum heavily dusted; Sc1 present and ending in Costa
6	Small species, wing length 3.5-4mm; basitarsus of fore-leg distinctly shorter than tibia;
	tarsal claws with one small tooth; genitalia (fig. 154) exigua Winnertz
	Hants, Herts, Lancs, Oxon, Suffolk, Sussex, Yorks; Scotland: Aberdeen, Arran,
	Perth, Sutherland; Ireland: Kerry. v-vi.
_	Larger species, wing length 5.5-6mm; basitarsus of fore-leg almost as long as tibia;
	tarsal claws with two distinct teeth; genitalia (fig. 155)
_	Scotland: Inverness, Perth. vi.
7	Scutellum with two bristles (winnertzi and wankowiczii, couplet 14, normally have four
	scutellar bristles, but may have less)
_	Scutellum normally with four bristles (cinerascens, couplet 9, normally has two bristles,
8	but may have four)
0	Sc1 usually absent, sometimes present or represented by a stump-vein from Sc2
9	Pleurae darkened, particularly lower half of katepisternum; mesonotal stripes more
,	or less fused; abdominal segments with yellow hind margins; wing length 4-5mm,
	genitalia (fig. 156)
	Beds, Berks, Cambs, Chesh, Cornwall, Cumbria, Devon, Dorset, Hants, Heref,
	Herts, Hunts, Kent, Lancs, Notts, Oxon, Surrey, Sussex, Worcs, Yorks:
	Wales: Dyfed, Flint, Merioneth, Monmouth, Montgomery; Scotland; Arran,
	Inverness, Moray, Nairn; Ireland; Kerry, Wicklow, iii-xi, From Stereum.
_	Pleurae yellow, the lower half of the katepisternum very obscurely darkened;
	mesonotal stripes distinct and separate; abdomen yellow with black dorsal stripe on
	segments 1-5, sixth segment black with yellow triangular mark at base; genitalia

	Leitrim, Tipperary, Wicklow. ii-xii. From Auricularia, Chondostereum, Coriolus, Merulius, Phlebia, Pleurotus, Poria, Sebacina, Sparassis, Stereum. Also from fungoid wood.
	No cloud over small cell of wing
12	Abdominal tergites 1-5 yellow with a black basal dorsal line, segments 6 and 7 black;
12	wing length 5.5-6mm; genitalia (fig. 161) with lobes of tergum 8 bearing 20 or more
	hairs on posterior marginpunctata Meigen
	Scotland: ?Glasgow, Inverness (Eigg Is). v-viii.
_	Abdominal pattern different, either uniform dark or with pale transverse bands; each
	lobe of tergum 8 with at most about 10 hairs on its posterior margin
13	Tergites uniformly dark or with narrow apical pale bands; no lateral sternal processes to genitalia
_	Tergites with broad pale apical bands; lateral sternal processes to genitalia present 15
14	Sc1 often not reaching Costa (sometimes completely absent); median dorsal process of
•	genitalia (fig. 162) swollen and bulbous; wing length 4.5-6mm
	hyalinata Meigen
	Berks, Chesh, Cumbria, Hants, Lancs, Norfolk, Notts, Shrop, Somerset, Sussex,
	Yorks; Wales: Gwynedd; Scotland: Arran, Inverness, Kincardine; Ireland: Armagh. iv-x.
_	Sc1 complete (1 specimen); median dorsal process of genitalia (fig. 163) evenly tapered
	from base; wing length about 5mmrosalba Hutson
	N. Yorks. vi.
15	Genitalia (fig. 164) with lateral dorsal processes outwardly concave, without inner basal
	protuberance; wing length 4.5-7mmwinnertzi Dziedzicki
	Chesh, Cornwall, Cumbria, Derby, Devon, Dorset, Hants, Herts, Kent, Lancs,
	Oxon, Worcs, Yorks; Wales: Gwynedd, Powys; Scotland: Aberdeen, Inverness, Nairn,
	Perth; Ireland: Antrim, Clare, Kerry, Killarney, Leitrim, Roscommon, Westneath.
	iii-xii. On Ganoderma, Phellinus.
_	Genitalia (fig. 165) with lateral dorsal processes outwardly convex, a small protuberance
	on inner side near base; wing length 5-5.5mm wankowiczii Dziedzicki
	Beds, Chesh, Cumbria, Hants, Herts, Notts, Suffolk; Scotland: Arran; Ireland:
	Wicklow. v-x. On Poria and Stereum on fallen birch branches. Also on Hypholoma
	and Phallus.
16	Genitalia (fig. 166) with two or three very long lateral bristles; wing length 3-4mm
	tenuis Walker
	Berks, Cambs, Chesh, Cornwall, Durham, Hants, Herts, Lancs, Middx, Notts,
	Suffolk, Sussex, Yorks; Wales: Brecknock, Merioneth; Scotland: Aberdeen, Argyll,
	Arran, Inverness, Moray. iii-x.
_	Genitalia (fig. 167) without such bristles; wing length 4.5-5mmduplicata Edwards
	Chesh, Essex, Hants, Lancs, Notts, Oxon, Surrey, Sussex; Scotland: Inverness,
	Kincardine. v-x. From Auricularia.
17	Front coxa with a dense brush-like patch of fine setae at the tip towards the inner
	side (fig. 25); postnotum with a few bristles
	Front coxae without obvious patch of fine setae; postnotum bare
18	Sc completely bare; claws with three teeth; thorax entirely dark and heavily grey-dusted,
	at most with the scutellum sometimes obscurely yellowish; abdomen darker than
	in other species of this group; wing length 5-5.5mm; genitalia (fig. 168)
	fuscata Winnertz
	Scotland: Inverness, Moray, Ross & Cromarty. viii.
_	Sc with macrotrichia; claws with only one or two small teeth
19	Mesonotal stripes more or less separate; scutellum yellow, postnotum and pleurotergites
	yellow on upper half
_	Mesonotum uniformly dark, with at most the shoulders yellow
20	Mesonotum somewhat shining; wing length 5-6mm; genitalia (fig. 169)
	ornata Meigen
	Lancs, Sussex; Scotland: ?Glasgow, Inverness, Moray. v, viii-ix.
_	Mesonotum dusted; wing length 5-6mm; genitalia (fig. 170) prominens Lundström
	Cumberland, Chesh, Derby, Dorset, Herts, Kent, Lancs, Lincs, Sussex, Warwick,

Hants, Herts, Kent, Lancs, Middx, Shrop, Surrey, Sussex, Worcs, Yorks; Wales: Brecknock, Caerns; Scotland: Aberdeen, Arran, Inverness; Ireland: Down, Kerry,

21	Yorks; Wales: Caerns; Scotland: Inverness, Moray, Perth; Ireland: Antrim. iii-ix. From Agaricaceae and Thelophora. Also rotten wood.
21	Scutellum dark; postnotum and pleurotergites entirely dark; abdominal tergites broadly yellow posterolaterally; shoulders usually yellow; genitalia (fig. 171) with parameter moderately long, flattened and densely pubescent along their inner edge; wing length 4.5-6mm
	Avon, Devon, Hants, Herts, Sussex, Yorks; Scotland: Argyll, Ayr, Inverness, Perth. iv-vi, ix.
_	Scutellum more or less yellow; rest of thorax entirely dark grey dusted; abdominal tergites with posterior margins only indistinctly pale; genitalia (fig. 172) with parameres very short and sickle-shaped, bare; wing length 5-5.5mm
22	Base of posterior fork approximately below or before base of stem of Median fork. Scl generally present and reaching Costa
_	Base of posterior fork well beyond base of stem of Median fork; Sc1 absent30
23	Scutellum with four bristles
_	Scutellum with two bristles
24	Abdomen all dark
_	Abdomen banded; Sc with macrotrichia
25	Sc bare; wing length 4.5-5mm; genitalia (fig. 173)melanoceros Edwards Scotland: Inverness, Moray, Perth. ix.
_	Sc with macrotrichia; wing length about 4.5mm; genitalia (fig. 174) pectinifera Edwards Cornwall. ix.
26	Abdominal tergites with broad basal yellow markings
_	Abdominal tergites with apical yellow markings
27	Base of posterior fork below or just beyond base of stem of Median fork; stem of Median fork almost as long as its upper branch; yellow tergal bands divided by a mid dorsal black stripe; wing length 4.5-5mm; genitalia (fig. 175) circumdata Staeger Berks, Dorset, Herts, Hants, Notts, Worcs; Scotland: Arran; Ireland: Kildare. y-yi, ix.
_	Base of posterior fork distinctly before base of stem of Median fork; stem of Median fork barely half as long as its upper branch; wing length 5-6mm
	?wrzesniowskii Dziedzicki Hants; Wales: Gwynedd; Scotland: Arran. v-vii. onPoria under fallen birch branch.
	[N.B. All these records are based on females. Since Dziedzicki's original specimer appears to have been destroyed and no other male has been traced in any collection, the identity of the British specimens must be questioned and no figure of the genitalia is given here. Any possible male should be compared with Dziedzicki's figures (1885: Tab VII, figs. 1-2).]
28	Scutellum and pleurae dark; thorax practically all uniform dark; Sc1 sometimes very short and not reaching Costa; wing length 4.5mm; genitalia (fig. 176)
	Oxon; Scotland: Inverness, Moray, Nairn. v-vi.
_	Scutellum yellow, pleurae largely yellow; mesonotum with shoulders extensively yellow; Sc1 reaching Costa; wing length 4-4.5mm; genitalia (fig. 177) parva Dziedzicki Hants, Kent, Oxon, Suffolk; Ireland: Kildare. vi-viii, x.
29	Sc with macrotrichia: Sc2 about opposite base of small cell; postnotum and pleurotergites uniformly dark; wing length about 4mm; genitalia (fig. 178)
	Bucks, y. collini Edwards
-	Sc bare; Sc2 well beyond the base of small cell; postnotum and pleurotergites yellow basally, black apically; wing length about 5mm; genitalia (fig. 179)
	Vorker Seetland Inverses Stirling vii viii
30	Yorks; Scotland: Inverness, Stirling. vii-viii. All branches of Median and posterior forks bare; wing length 3.5-4.5mm; genitalia

Dunbarton, Moray, Perth, Stirling. vii. ix-x.

Chesh, Lancs, Norfolk, Notts, Surrey, Yorks; Wales: Merioneth; Scotland:

Branches of Median and posterior forks with macrotrichia, although sometimes scarce

	on posterior fork
31	Transfer and the first and the first and the part aproved
	bands; wing length 4-4.5mm; genitalia (fig. 181)
	Beds, Hants, Herts, Kent, Oxon, Staffs, Suffolk, Sussex; Wales: Merioneth;
	Scotland: Perth; Ireland: Antrim, Clare, Down, Galway. iv-vii, ix.
_	At least scutellum yellow
32	Adbominal tergites pale basally or all dark; wing length 3-4mm; genitalia (fig. 182)
	fimbriata Meigen
	Chesh, Gloucs, Hants, Lancs, Norfolk, Oxon, Staffs, Sussex, Yorks; Wales:
	Brecknock, Merioneth; Scotland: Inverness, Perth, St. Kilda; Ireland: Wicklow. v-ix.
_	Abdominal tergites yellow, more or less darkened basally; wing length 3.5-4mm;
	genitalia (fig. 183)
	Berks, Chesh, Devon, Dorset, Hants, Heref, Kent, Lancs, Oxon, Sussex, Yorks;
	Wales: Merioneth: Scotland: Inverness: Ireland: Mayo v-x.

Genus Neoempheria Osten-Sacken

(Figs. 184-187)

A genus of over 100 species most of which are tropical, particularly Neotropical. About 8 species are known in Europe and four are recorded from Britain. The adults of most species (including all the species recorded from Britain) have patterned wings and are found in damp woodland. Larvae may be found in webs on fungi or on rotting wood.

Key to species

Tribe Sciophilini

The Sciophilini include species with a wide range of general appearance, arranged in about twenty genera. Most of these genera contain few species: in Britain Sciophila has 17 species, but none of the others has more than three. The genus Baeopterogyna Vockeroth is the only genus known from the Holarctic that has not been recorded from Britain; it is similar to Neuratelia Rondani, but lacks distinct tibial setae, the tibial spurs are reduced to about equal to the width of the tibia, anepisternum, pleurotergite and katepisternum with short hairs, wing macrotrichia short and sparse, female brachypterous.

The adults of a few species may be found at flowers. The larvae have been reared from fungi or rotting wood, where they may construct webs, and from moss.

Key to genera

1	Ocelli two, contiguous with eye margins; postnotum almost covered with small fine hairs as well as the usual large bristles apically; R5 straight; Median and posterior forks both complete; Costa slightly extended beyond end of R5; Sc2 well beyond middle of Sc
_	Ocelli three, lateral ocelli not contiguous with eye margins, although sometimes (e.g. Paratinia, Phthinia) quite close
2	Base of posterior fork distinctly proximal to that of Media; hind tibia without a distinct apical comb (fig. 43)
_	Base of posterior fork distinctly distal to that of Media, or this fork absent; hind tibia often with distinct apical comb (fig. 42); Postnotum hairy
3	Postnotum hairy, at least towards the sides at tip; pleurotergites hairy
_	Postnotum completely bare8
4	R5 sinuous; base of anterior branch of Median fork weak or absent; wings usually unmarked; Usually grey-brown species
-	R5 straight; wing with dark markings; anterior branch of Median fork complete or very narrowly interrupted at base; yellowish species
5	Costa distinctly produced beyond end of R5; base of anterior branch of Median fork faint but traceable
_	Costa only produced slightly beyond end of R5; base of anterior branch of Median fork absent; Sc2 well before middle of Sc; R4 absent (fig. 77)
6	Anterior branch of Median fork arising only slightly beyond posterior fork; Sc2 before
	middle of Sc, faint or absent; R4 absent
7	middle of Sc, distinct; R4 frequently present (fig. 78) POLYLEPTA Winnertz: (p.48) Sc2 before middle of Sc; wing marking restricted to near apex of R1 and R5; anterior
	branch of median fork very narrowly interrupted at base; Costa slightly produced beyond R5
_	Sc2 beyond middle of Sc; bold wing markings covering most of wing tip and with smaller marking over base of R5 and r-m; Median fork complete; Costa ending at tip of R5
	(fig. 76)
8	Pleurotergites bare; Sc ending in Costa; body long and slender
9	Slender species; fore basitarsus very much longer than tibia, often more than twice as long; posterior branch of posterior fork sinuous; median and posterior forks
_	complete
	posterior fork more or less straight when present
10	Median fork complete
_	Only two major veins posterior to the radius, with a short remnant vein on the wing margin between them (? anterior branch of Median fork absent, posterior branch

present but weak at base; anterior branch or posterior fork widely detached at base, posterior branch present and complete); Sc very short, ending free; macrotrichia decumbent; an episternum haired; r-m long and horizontal

Sc2 well beyond base of Rs; macrotrichia erect or reflexed towards wing base;

12 Posterior fork complete; anepisternum with small hairs (fig. 20); macrotrichia

decumbentSCIOPHILA Meigen (p.51)

No posterior fork, only a simple vein present; an episternum bare; Sc2 before Rs......13

Macrotrichia reflexed (fig. 48); R4 usually present MONOCLONA Mik (p.51)

Genus Eudicrana Loew

(Fig. 188)

A genus of about 10 species most of which are Neotropical. The single Nearctic species may be the same as the European species, but the only other Holarctic species, described from Japan, differs in having distinct markings in the middle of the wing as well as at the tip. The life-history is unknown.

An elongate species with wing tip dark; thorax pale with three distinct stripes; abdomen banded; wing length about 7mm; or genitalia (fig. 188) nigriceps Lundström Scotland: Inverness, vi.

Genus Allocotocera Mik

(Fig. 189)

A genus of about 12 species most of which occur in the Neotropical, Australasian and New Zealand regions. One species occurs throughout the Palaearctic and in the Nearctic. Adults may be found on Umbelliferous flowers or in damp woods. Immature stages are unknown.

A small yellow species; wings with a dark cloud around apex of cell R1; abdomen of σ with segments 1-5 black apically, segments 6 and 7 entirely black, genitalia (fig. 189) yellow; Q abdomen with only segment 6 completely black; wing length 3-3.5mm

Wales: Gwynedd, Powys; Scotland: Inverness; Ireland: Kerry, (?i) vi-viii.

Genus Leptomorphus Curtis

(Figs. 76, 190)

This genus includes about 15 species from most parts of the world, but about half from the Nearctic. In the Palaearctic one species occurs in Europe (including Britain), one from Europe to Japan and two only in Japan.

The larvae are quite common in webs on bark growing fungi (e.g. Poria spp.) on fallen branches. The larval skin remains attached to the tail of the pupa, which hangs free, head downwards.

Large yellowish species; wings with broad cloud at tip, cloud over base of Rs and r-m, and below posterior fork; R4 absent (fig. 76); thorax yellow with three dark dorsal stripes, sometimes confluent; abdomen yellow with more or less distinct black apical bands, segment 7 entirely black in male; wing length 5.5-8mm; or genitalia (fig. 190)

..... walkeri Curtis Cambs. Gloucs, Hants, Heref, Herts, Kent, Lancs, Middx, Oxon, Staffs, Suffolk, Surrey. vi-x. On Poria, etc.

Genus Anaclileia Meunier

(Fig. 191)

A small genus known from two species in Europe and three undescribed species in the Nearctic region. Life history unknown. Dark species, similar to *Neuratelia*, but smaller.

Genus Neuratelia Rondani

(Figs. 43, 77, 192-193)

N. nemoralis Meigen occurs commonly throughout the Holarctic. Five other species are known from the Palaearctic (including one from Japan) and 13 from the Nearctic. Single species are also known from the Oriental and Neotropical regions. A key including most of the Palaearctic species has been published by Matile (1974).

The adults are mainly rather elongate species, usually dark brown or black, with yellow legs. Laffoon (1965) states that the larvae occur in moss, but I have not been able to find a source for this and Matile (1974) says that the larva is unknown.

Key to species

- 1 Antennae with first flagellar segment largely yellow; wing length 4-5mm; or genitalia (fig. 192) without spines on basistyle and with pointed dististyle
 - Avon, Berks, Cambs, Chesh, Cumbria, Derby, Dorset, Hants, Heref, Herts, Kent, Lancs, Middx, Staffs, Surrey, Sussex, Yorks; Wales: Gwynedd, Powys; Scotland: Aberdeen, Arran, Inverness, Perth, Rannoch, Stirling; Ireland: Antrim, Clare, Kerry, Killarney, Laois, Leitrim, Tipperary, Wicklow, v-vi.

Genus Polylepta Winnertz

(Figs. 32, 35, 78, 194)

Four Palaearctic species, three Nearctic and one Oriental species. Of the European species, splendida Winnertz is distinct in having a dark wing tip and quite different genitalia; zonata Zetterstedt is probably a synonym of guttiventris since the venational characters used to separate these two species are not reliable; borealis Lundström is very similar to guttiventris, but differs in details of the genitalia, particularly in the curved inner dististyle, and in the uniform dark abdomen. The males of both guttiventris and borealis have the base of the mid-tibia swollen and the outer surface covered in fine pale setae, as in figs. 32 and 35 (information lacking for other species). They are both fairly elongate species. Nothing is known of the life history of this genus.

 Gwynedd, Powys; Scotland: Inverness, Moray, Perth, Sutherland; Ireland: Wicklow. v-x.

[N.B. 4 damaged 99 from Kent are very pale and may be a second species or a pale form of this species—described as *flava* Grzegorzek.]

Genus Syntemna Winnertz (Figs. 47, 195-197)

An Holarctic genus of about 14 species, 8 of which occur in the Palaearctic and 10 in the Nearctic regions. Most species occur in the north of their region and there is a strong Holarctic element. Specimens are uncommon in collections and are usually taken singly. Very few females have been collected. The male genitalia are the only reliable characters so far found for separating species, and so, while the female ovipositor presents good (if smaller) differences, there have been problems in associating them with males. The Palaearctic species have been revised by Hutson (1979). The three species recorded in Britain differ from most other species in having the combination of R4 present, wing macrotrichia extending towards wing base well beyond Median fork, σ front tarsus without ventral brush of fine hairs, hairs of dorsum of thorax and scutellum very pale. The females of five species are known and four of these have swollen front tarsi. They are all small dark species with a wing length of about 3.5mm.

Behaviour of adults is unrecorded, but two species have been reared from rotting wood. Larvae and pupae are undescribed.

Key to species

- Palps dark; mesonotum uniform dark; stem of posterior fork with many macrotrichia extending back towards origin of vein; 15-30 macrotrichia on upper surface of basal cell.
- 2 Spot above distal end of squama pale; Anal vein with 8-12 macrotrichia; pedical and at least 3 basal flagellar segments of antennae usually yellow; mesonotal bristles usually shorter, finer, more yellow; or Tergite 2 distinctly pale; or genitalia (fig. 196); ? Q with lateral lobes of Sternum 8 somewhat pointed, longer than broad

Genus **Azana** Walker (Fig. 198)

A small genus of two species in Europe and single species in each of Canary

Islands, North America, Africa and Ceylon. No details of life history are known.

Genus Megalopelma Enderlein

(Fig. 199)

A small genus with a single species widespread in the western Palaearctic, two species in the Nearctic and about four in the Neotropical region. No details of life history are known.

 Small species with uniform dark thorax and abdomen; legs pale; stem of haltere pale, knob black; stem of Median fork very short; stem of posterior fork very long; R4 present or absent; wing length 3.5-4mm; or genitalia (fig. 199)

Devon, Shrop, Sussex, Worcs, Wilts; Wales: Denbigh; Scotland: Moray; Ireland: Dublin, Kerry. v-ix.

Genus Acnemia Winnertz

(Figs. 42, 200-202)

Another widespread genus with few species. Apart from the three species included here, two others have been described from Europe, but are poorly documented. There are also three Nearctic species, five Neotropical, one Ethiopian and one Oriental species.

One species has been reared from a pupa found under rotting wood. Adults have been found at flowers. The adults rest with the wings slightly divaricate (as in *Sciophila*). They are small species.

Key to species

- 2 Ocelli in a very shallow triangle, the middle one placed a little forwards; palps yellowish; antennae uniform dark; a small species, wing length 2.5-3.5mm; Tegenitalia (fig. 201); Sternum 8 rather pointed at apex......mitidicollis Meigen Berks, Cambs, Chesh, Devon, Hants, Heref, Herts, Kent, Lancs, Norfolk, Notts, Shrop, Staffs, Surrey, Sussex, Warwick, Westmorland, Yorks; Wales: Brecknock, Merioneth; Scotland: Aberdeen, Inverness, Moray, Perth; Ireland: Armagh, Kerry, Wicklow. iii-xi.

Sussex; Wales: Gwynedd; Ireland: Wicklow. x.

Genus Monoclona Mik

(Figs. 48, 203)

A principally New World genus with five species in the Nearctic and about eight in the Neotropic. Four species are described from Europe, but it is likely that they all belong to the single species included here, which also occurs in North America.

It has been reared from rotten wood attacked by fungus (?Poria sp.). The larva lives in a mucilaginous tube and pupation takes place in a dry silken cocoon in a crevice of the wood.

Genus Sciophila Meigen

(Figs. 20-21, 204-220)

This is by far the largest genus of this tribe, with 17 of the 30 or more Palaearctic species occurring in Britain. About 25 species occur in North America, although a number of them are shared with the Palaearctic. Very few species have been described from the southern hemisphere, but a few species are known from South America and single species from the Ethiopian, Oriental and Australasian regions.

Very few useful structural characters apart from the male genitalia have been found to separate the species of this genus and so colour is used rather widely. Some common species, such as S. lutea Macquart and hirta Meigen, are so variable that they cannot readily be fitted definitively into such a key; other species are so little known that it is difficult to know how reliable the characters for their separation are. The male genitalia seem to be reliable and are illustrated, the female genitalia show some differences, but have not been thoroughly investigated and deserve further examination. Females of many species were not available in the present study. This key in particular should be used with caution.

Adults rest with their wings only partially overlapping. The larvae of most species are to be found on fungi, particularly hard bark fungi. They live on the undersurface of the fungus in a mucilaginous tube and with a dry silk web. They probably feed mainly on fungal spores. They pupate in a slight cocoon in a crevice in the fungus or in the bark of the host tree. Some species may be associated with rotting wood.

Key to species

1	Microtrichia of wings absent, except at extreme base, macrotrichia obvious
_	Microtrichia covering wing membrane between macrotrichia
2	Antennal flagellum, head and abdomen black
_	Base of antennal flagellum, head and abdomen ochreous or red4
3	Thorax black; Sc2 before base of Rs; hind coxae slightly darkened; wing length about
	5mm; o genitalia (fig. 204)limbatella Zetterstedt
	Scotland: Inverness. vii.
_	Thorax reddish: Sc2 beyond base of Rs: all coxae pale: wing length 4-4 5mm; or genitalia

_	Smaller ochreous species; front tibia without antero-dorsal row of bristles; Sc2 above base of Rs; wing length 3.5-4.5mm; or genitalia (fig. 207)ochracea Walker Cambs, Hunts. v-vi. From "?Fomes" on old Plum tree, cocoons on Cherry tree;
5	probably Phellinus sp. Anterior branch of posterior fork widely interrupted at base; hind femora black at tip and on ventral surface near base; Sc2 ending just before base of Rs; thorax uniform dark; antennal flagellum dark; wing length about 2.5mm; or genitalia (fig. 208)
	Cambs, Hants, Suffolk. ix. Hydnum.
_	Posterior fork complete
6	Hind femur entirely yellow7
_	Hind femur with dark apex, body all black
7	At most first flagellar segment yellowish
8	At least first two flagellar segments pale; thorax at least partly pale
	Bucks, Cambs, Chesh, Essex, Greater Manchester, Hants, Heref, Herts, Hunts,
	Kent, Lancs, London, Surrey, Worcs; Wales: Gwynedd; Scotland: Inverness;
	Ireland: Down, Dublin. iv-xi. From Armillaria, Auricularia, Bulgaria, Cantharellus, Collybia, Coriolus, Lactarius, Lenzites, Pholiota, Stereum; especially Polyporaceae.
_	Sc on or after base of Rs9
9	Larger species, wing length 3.5-4.5mm; thorax light (typical var.) to dark (var. analis
	Winnertz); o genitalia (fig. 210)
	London, Norfolk, Surrey, Sussex, W. Yorks; Scotland: Angus; Ireland: Antrim.
	iii-x. From Boletus, Collybia, Coriolus, Daedalia, Gomphus, Grifola, Heteroporus,
	Hydnum, Hypoxylon, Lactarius, Meripilus, Phellinus, Pholiota, Pleurotes,
	Pseudotrametes, Russula, Stereum, Ustulina.
10	Smaller species, wing length 2.5-3.5mm; thorax dark
10	dististyle very short, emarginate apically; tergite 9 shallowly emarginated apically, with two long bristles on prominent tubercles near the lateral margins
	fridolini Stackelberg
	Sussex, Yorks. vi, ix. Hypopleuron bare; wing length about 3mm; σ genitalia (fig. 212) with dististyle short
	with a dorso-apical posteriorly directed process and two strong latero-apical spines;
	tergite 9 with an obtusely pointed apexcliftoni Edwards
	lo; Clifton coll.—no further data.
11	Dorsum of thorax completely orange
12	Larger species, wing length 3.5-4.5mm; & genitalia (fig. 210) with tergite 9 shallowly
	emarginate apically with two long bristles on tubercles near lateral margins; latero-
	ventral process of dististyle longer than broad, with lateral as well as two terminal
	bristles; parameres prominent and broadly bifurcate at tiplutea Macquart See couplet 9 for details of distribution, etc.
_	Smaller species, wing length c.3mm; or genitalia (fig. 213) with tergite 9 evenly rounded
	apically with c.12 long bristles; latero-ventral process of dististyle short and broad with
	only two terminal bristles; parameres inconspicuousplurisetosa Edwards
12	?Chesh; Scotland: Arran. viii-ix. Smaller species, wing length less than 3.5mm
13	Larger species, wing length over 3.5mm
14	Length of antennal flagellar segments 2 and 3 about three times width; or genitalia
	(fig. 214) with tergite 9 broadly truncate; dististyle reduced, latero-ventral process with
	only two long bristles; parameres very narrow, finely pointed and simple
_	Length of antennal flagellar segments 2 and 3 about twice width; σ genitalia (fig. 215)
	with tergite 9 with a distinctly triangular apex; latero-ventral process of dististyle
	with long bristles all along one edge; parameres short with rounded apex and small

- or genitalia (fig. 216) with tergite 9 deeply and rectangularly emarginate, a row of strong bristles along the margin of the emargination and a brush of strong hairs on the apico-lateral lobes; parameres inconspicuous, ? not bifurcate.....varia Winnertz ?Bristol: Scotland: Moray. ix-x.
 - [N.B. only one σ seen. In this specimen the genitalia are badly preserved and it is possible that the strongly bristled "apico-lateral lobes" are not part of tergite 9. In this case T9 is simply truncate apically.]
- Hind femur broadly black at tip and on ventral surface near base; hypopleuron bare...18
- 17 Sc2 before base of Rs; wing length 3-4mm; or genitalia (fig. 209) with dististyle rounded without posteriorly directed process; tergite 9 evenly rounded........... hirta Meigen See couplet 8 for details of distribution, etc.

- Hind coxa dark at tip and base only; Sc2 not beyond base of Rs; wing length about 4mm; or genitalia (fig. 220)......geniculata Zetterstedt Scotland: Arran. ix.

Genus Phthinia Winnertz

(Figs. 36, 221-223)

Apart from the two species included here, the Palaearctic has a third European species that is inadequately described and a fourth species in Japan. The genus is also widely recorded from the rest of the world with three species in North America, three in South America, one in New Zealand and one in Australia. The adults are elongate medium sized species which occur around rotting wood from which they have been reared. The postnotum is only sparsely haired laterally.

Key to species

- Fore-leg with basitarsus nearly twice as long as tibia; body not so elongate; Median fork
 narrowing to a point at base as usual; Anal vein curving down at tip; anal lobe

[N.B. 1 or specimen of *P. humilis* from Crowborough, Sussex (coll. F. Jenkinson) has rather different genitalia from all other specimens (fig. 223). However, no other differences were found and the genitalia are very similar to *humilis* compared with other *Phthinia* spp. and so it is considered inadvisable to describe it as a separate species until further material is available.]

Genus Paratinia Mik (Fig. 224)

A very small genus of two dark Palaearctic species and a pale species that may be a synonym of the single Nearctic species. Life history unknown, but the adult of at least the British species would appear to be nocturnal.

Tribe Gnoristini

Like the Sciophilini, this tribe includes genera showing a wide range of general appearance. About 15 genera are currently recognised, but some of these are being studied with a view to dividing them into a number of smaller genera (e.g. Vockeroth, 1980). Most of the genera are small—in the Holarctic region only Boletina includes more than ten species, but other larger genera occur in the southern hemisphere. Only one Holarctic genus does not occur in Britain: this is Hadroneura, known from two European and three Nearctic species. In the following key to genera Hadroneura would run to Dziedzickia. Dziedzickia may be a complex of genera and only when this has been examined can the status of Hadroneura be qualified; meanwhile the European species of Hadroneura differ from Dziedzickia in the genitalia and in having a rather more elongate lower part of head.

Again there are biological data for only very few species. The activity of adults is virtually unknown. The larvae have been found in or on fungi, decaying wood, liverworts and mosses. The biology of *Speolepta leptogaster* Winnertz, an inhabitant of underground cavities, is better studied than most (Matile, 1970).

The distinctions between this tribe and the next may prove difficult without some experience. Some of the more likely sources of error are accounted for in the key to genera.

Key to genera

3	Sc not reaching Costa, either ending in R, or Sc1 ending free
	Sc ending in Costa
4	Sc ending free, seventh segment of abdomen quite large (σ and Q); tibial bristles weak; pleurotergites bare
_	Sc ending in R; seventh abdominal segment small and usually retracted, at least in the male; tibial bristles well developed; pleurotergites hairy
5	Stem of Median fork at least twice as long as r-m; R4 usually present
	DZIEDZICKIA Johannsen (p.57)
_	Stem of Median fork very short, varying from slightly shorter than r - m to about 1.25 \times as long; R4 absent (fig. 80)
6	Proboscis (clypeus) very elongate, much longer than rest of head (fig. 11)
_	Proboscis (clypeus) shorter than head7
7	Sc2 present and well beyond middle of Sc
_	Sc2 near middle of Sc or absent
8	Base of posterior fork well beyond base of stem of Median fork (fig. 81); lateral ocelli
	less than their own width from eye margin; empodium well developed; Sc at most
	with one or two setae; tibial bristles tending to form rows towards apex; seventh
	abdominal segment small and retractedSYNAPHA Meigen (p.58)
_	Base of posterior fork more or less opposite base of stem of Median fork; lateral ocelli
	at least their own width from eye margin; empodium rudimentary or absent; Sc
	setulose; tibial bristles irregular throughout; seventh abdominal segment larger
	GRZEGORZEKIA Edwards (p.58)
9	R4 present; Sc2 absent (fig. 83); Sc setulose; tergite 9 of σ with terminal row of strong
	spines
	. [N.B. Rondaniella (Leiini) may run to here. In contrast to the genera considered
	here, it has a distinct dark wing tip. See page 64.]
_	R4 absent; Sc2 usually present (fig. 84); Sc bare (except <i>plana</i> and sometimes <i>dubia</i>);
	tergite 9 of σ without a terminal row of spines (sometimes with short combs on
	apex)
	apex)BULETINA Staeger (P-37)

and the Court of the sending in Done Call and the Court

Genus Coelophthinia Edwards

(Fig. 225)

A small genus of one Palaearctic and one Nearctic species, both of which may be synonymous. One species has been reared from fungi.

Small species, wing length 3-4mm; head and antennal flagellum dark, scape and pedicel yellow; mesonotum yellow with three dark stripes; halteres with yellow stem and black knob; abdomen dark; σ genitalia (fig. 225) yellow; Q genitalia darker, but the terminal segment of cerci yellow and broadly rounded apically

Genus Coelosia Winnertz

(Figs. 79, 227-229)

A genus of about ten species in the Holarctic region and a few species in the Neotropical and Oriental regions. One species has been associated with fungus.

Key to species

1 Thorax entirely yellow; Costa extending to less than half distance from R5 to anterior vein of Median fork; postradial veins bare; palps, base of antennae, halteres and legs yellow; wing length 3.5-5mm; or abdomen yellow except for dark tergite 6,

- - 2 Costa extending almost three-quarters of distance from R5 to anterior vein of Median fork; dingy species, palps, base of antennae, knob of halteres and legs all darkened; wings slightly greyish, all veins dark; postradial veins bare, but slightly seamed with dark brown, particularly at base of posterior fork; wing length 3.5-5mm; or genitalia (fig. 228); Q terminal segment of cerci small and oval silvatica Landrock Herts, Kent, Oxon; Scotland: Moray, ii-iii, ix.

Genus Speolepta Edwards (Figs. 37, 226)

A small genus of a single Palaearctic species and one unnamed Nearctic species. A second Palaearctic species, recently described from a single specimen, is undoubtedly a synonym. S. leptogaster Winnertz is very common in underground cavities and has occasionally been found outside this habitat—even at light traps. It is the only British fly that approaches a troglodytic way of life, but is perhaps more correctly termed a troglophile.

The larvae are common on the walls and roof of damp caves, mines, tunnels, cellars, etc. They are probably equally common in small rock fissures, etc. They are white with a pale head capsule and live in a slime-tube suspended with the aid of supporting threads. The tube is probably an overlapping roll of mucilaginous silk and not a sealed tube and the larvae can partly extrude from any part of the length of the tube to forage on the substrate. It probably feeds largely on algae and other available organic matter. The pupa hangs head downwards without a cocoon. The adults are not as common as might be expected from the numbers of larvae and it might be that the larvae have a protracted development while the adults are short-lived. It is not known to what extent the species is seasonal within the cave environment, but more adults have been collected in May and June than at other times of year.

An elongate grey-brown species, wing length 4-4.5mm; R4 present or absent; tibial spurs short, especially on fore and mid-legs; σ mid-tibia slightly swollen near base with an inconspicuous sense organ on dorsal surface; σ genitalia (fig. 226); Q sternum 8 with a broadly truncate apex, terminal segment of cerci small and oval

Cornwall, Cumbria, Derby, Devon, Durham, Gloucs, Kent, Lancs, Oxon, Somerset, Surrey, Yorks; Wales: Clwyd, Dyfed, Gwent, Merioneth, Mid-Glamorgan, Powys; Scotland: Inverness (Eigg Is.), Renfrew; Ireland: Clare, Fermanagh, Sligo, Tipperary. i, iv-xi. In caves, etc.

Genus Dziedzickia Johannsen

(Fig. 230)

This may cover a complex of genera, but as currently understood most species occur in the New World with one Palaearctic species (the type-species of the genus) and one African species. Species of *Hadroneura* would run to here, but differ in structure of the genitalia. Other species currently included in *Dziedzickia* do not demonstrate all the features used in this generic key. Life history unknown.

Genus Palaeodocosia Meunier

(Figs. 80, 231-233)

Apart from the Baltic Amber species upon which this genus was based, the three species included here are the only species that have so far been included in the genus. Some Nearctic species currently included in *Dziedzickia* belong with species of this genus, but a study of a wider range of species is necessary to establish a better understanding of the genera in this complex. The British species all have a largely yellow mesonotum with a pair of black lateral patches. While the last two species in the key seem quite discrete in British material, a Nearctic species (vittata Coquillett) shows intermediate characters and it may be that further material will suggest that these species should be synonymised. Life history unknown.

Key to species

- 1 Abdomen mainly yellow, the posterior margins of the first five and practically the whole of the sixth and seventh tergites black; Sc reaching base of Rs; veins of Median and posterior forks bare; wing length 4.5-5.5mm; σ genitalia (fig. 233)
- Abdomen mainly blackish-brown, with moderately broad yellow posterior margins; Sc ending well before base of Rs (fig. 80); veins of Median and posterior forks with macrotrichia.
- 2 Scutellum pale; mesonotum with dark lateral stripes not extending inwards beyond intra-alar bristles; fine bristles of mesonotum dark; wing length 4mm; ♂ genitalia (fig. 232) with tergite 9 uniformly hairy, hairs on distal margin not unusually stout, sternum 8 not noticeably produced and with fewer long bristles; ♀ not examined
 - Hants, Lancs; Wales: Caerns; Scotland: Inverness, vi-viii.

Bucks, Cambs, Chesh, Herts, Hants, Kent; Wales: Gwynedd; Scotland: Perth; Ireland: Killarney. v, vii, ix-x.

Genus Gnoriste Meigen

(Figs. 11, 234-235)

An Holarctic genus of 8 species and possibly another species in South America. Four species are known from Europe and a fifth species has been described from Japan. All species are quite large and have a characteristic very long proboscis. Chandler (1976) provides a key to species. The activity of adults is unrecorded. One species has been reared from larvae found in mosses.

Key to species

- 1 Ground colour of mesonotum yellow, with distinct black stripes, sometimes fused; proboscis (fig. 11) half body length or less (less than three times length of front coxae); σ with posterior claw of all legs stunted; wing length 7-8mm; σ genitalia (fig. 235); σ wings usually dark tipped, Q clear..... bilineata Zetterstedt Scotland; Inverness, Perth, Stirling, vi.

Genus Synapha Meigen (Figs. 81-82, 236-237)

A rather more widespread genus than most of the genera of this tribe, including the two Palaearctic species included here, three in the Nearctic region, about 10 in each of New Zealand and South America and a single species in Australia. However some of these may not properly belong to this genus. Adults have been found at flowers, but the breeding biology is unknown.

Key to species

Genus Grzegorzekia Edwards (Fig. 238)

Apart from the single species included here, known from the Western Palaearctic, a second species has been described from Austria, but may be a synonym. Adult activity is unrecorded. The larvae have been found on damp rotton wood either on

the surface or suspended in a web-like structure into which they rapidly retreat when disturbed. The webs may be close to each other. Pupation takes place on the surface of the wood; there is no cocoon.

— Medium large species, wing length about 5.5mm; prothorax light yellow, rest of thorax shining black; wings with the tip slightly darkened and a slight dark cloud over the small cell; vein R5 sinuous; abdominal tergites dark with pale basal bands in σ, bands reduced or absent in Θ; σ genitalia (fig. 238), Q terminal segment of cerci large and yellow, sternum 8 truncate with about 4 strong bristles at apex

......collaris Meigen

Cambs, Gloucs, Hants, Suffolk, Warwick. iv-vii, ix.

Genus Apolephthisa Grzegorzek

(Figs. 83, 239)

This genus comprises one Palaearctic and one unidentified Nearctic species. Adult activity unknown. The larvae live under bark or on bark-growing fungi, in a mucilaginous tube anchored with lateral threads. There is no cocoon.

Medium sized species, wing length 3-4mm; head, thorax and abdomen black; halteres yellow; legs yellow with the hind femur more or less dark tipped; pleurotergites haired; postradial veins with macrotrichia; base of anterior branch of posterior fork weak or absent (fig. 83); σ genitalia (fig. 239) with a row of large spines on the apex of tergite 9; apex of Q abdomen distinctly tapered, last segment of cerci elongate oval, penultimate segment about 1.5 times as long as broad subincana Curtis Beds, Berks, Chesh, Cornwall, Essex, Hants, Heref, Herts, Lancs, Shrop, Staffs, Suffolk, Sussex, Yorks; Wales: Gwynedd; Scotland: Arran, Inverness, Midlothian, Moray, Perth; Ireland: Down, Kerry, Wicklow. iii-x. From Phlebia, Xylodon and under bark of oak (Quercus).

Genus **Boletin**a Staeger (Figs. 7, 28-30, 58-59, 84, 240-262)

The largest genus of this tribe, with about 100 species, all from the northern hemisphere but including about 10 species from the Oriental region. Although principally species of wooded streams, members of this genus are found in a wider variety of habitats and distribution than most genera of the family. Their ability to thrive in open moorland, etc. has enabled some species to colonise the more barren areas of Britain and its islands. Wetlands, which are generally poor habitats for Mycetophilidae, will frequently produce at least a species of *Boletina*. This is perhaps a result of the use of diverse breeding sites, since while some species develop in the more normal habitats such as rotting wood, others develop in mosses and liverworts. The breeding biology of most species is unknown and other species may occur in unexpected situations.

The following key includes 23 species. One other species, silvatica Dziedzicki, was included in the British fauna by Edwards (1941) based on a single σ from Gloucestershire which should be in the British Museum (Natural History). We have not been able to find this specimen and no other British specimens are known to us, so it has been omitted from this work. It would run to the end of the key (to the sciarina-group) and differs from other species by the structure of the genitalia. The genitalia are figured in Landrock (1929) and Ostroverkhova & Stackelberg (1969).

There have been various attempts to separate species or groups of species as separate genera or subgenera, but none has so far found general acceptance. The use

of more obvious characters, such as the presence or absence of pleurotergal bristles, does not provide a natural division. A species such as *flaviventris* Strobl may exhibit a combination of characters and a general appearance that make it quite distinct from the rest of the British fauna, but a thorough investigation of the whole genus is necessary before any groupings can be satisfactorily delimited.

Key to species

1	Pleurotergites with bristles, at least near the ridge
2	Sc sparcely setose
	Sc bare
3	club-shaped, other claws unmodified; or genitalia (fig. 240); Q sternum 8 not deeply divided medially (fig. 59)
	Oxon, Suffolk, Surrey, Sussex, Westmorland, Yorks; Wales: Brecon, Caerns, Flint, Merioneth; Scotland: Argyll, Inverness, Midlothian, Moray; Ireland: Wicklow. v-x.
_	Sc2 normally absent; wing length 4-5.5mm; σ with posterior claw of anterior tarsus club-shaped, anterior claws of all legs greatly enlarged and with a comb of fine teeth on the underside (fig. 30), other claws similar but shorter; σ genitalia (fig. 241); ∇ sternum 8 deeply divided medially (fig. 58); (see also couplet 6)
	Midlothian, Moray, Perth, Sutherland; Ireland: Antrim, Leix, Mayo, Monaghan, Westmeath. iv-ix.
4	Sc2 normally absent
_	Sc2 present
5	Abdomen with yellowish terminal bands on tergites 2-4; scape, pedicel and first two flagellar segments ochreous, rest dark; Median fork weakly developed and with
	few macrotrichia; wing length 3-3.5mm; σ with posterior claw of front tarsus short and club-shaped, all other claws unmodified; σ genitalia (fig. 242); φ sternum 8 with four very strong bristles
_	Abdomen entirely or mostly blackish; at most base of first flagellar segment yellow;
6	Median fork well developed, densely beset with macrotrichia except at base
_	R5 nearly straight; shoulders and pleurae not heavily dusted, more or less shining; wing length 3.5-4.5mm; σ tarsal claws unmodified; σ genitalia (fig. 243)
7	Thorax uniformly shining black without dusting
_	Thorax greyish dusting at least on shoulders
8	Abdominal sternites 2-4 all yellow, tergites 2-4 extensively yellow; wing length about 4mm; σ posterior claw of front tarsus short and club-shaped; all other claws unmodified; σ genitalia (fig. 244) with proximal comb of cerci rudimentary
	Hants, Somerset; Ireland: Clare, Wicklow. v-vii, ix-x.
9	Abdominal'segments 2-4 not yellow
-	Hants; Ireland: Kerry. v-vi. Wing length 3.5-4mm; all σ tarsal claws short and rounded; σ genitalia (fig. 246) with

	cerci broader, the distal comb with more numerous and slender teethdispecta Dziedzick
10	Hants, Heref, Kent, Somerset; Wales: Montgomery; Scotland: Argyll. vi, vii, ix-xi First two segments of antennal flagellum yellow; wing length about 4.5mm; σ with
	conspicuous projecting horn on face below antennae; σ with posterior tarsal claws of front legs club-shaped, all others unmodified; σ genitalia (fig. 247); Q sternum Q
	short with a broad straight apical margin
_	Only first segment of antennal flagellum yellow; wing length about 4.5mm; or withou facial horn; all tarsal claws large and broad with very fine pectinations; or genitalia
	(fig. 248) pectinunguis Edwards Scotland: Inverness. vi.
11	[N.B. B. dubia may run to here, but differs by characters given in couplets 3 and 6. Costa scarcely reaching beyond R5 (fig. 84); r-m long and almost in line with R5; wing
	length 4.5-5.5mm; σ tarsal claws large and unmodified (fig. 28); σ genitalia (fig. 249); σ sternum 8 with a row of spinous bristles at apex trivittata Meiger Berks, Cambs, Chesh, Cumberland, Derby, Dorset, Gloucs, Hants, Heref, Kent,
	Lancs, Lincs, Norfolk, Notts, Shrop, Staffs, Suffolk, Westmorland, Yorks; Wales, Caerns, Denbigh, Flint, Glam, Merioneth; Scotland: Aberdeen, Arran, Inverness,
	Moray, Nairn, Perth, Ross & Crom, Sutherland, Tayside: Ireland: Antrim, Clare, Down, Kerry, Kildare, Laois, Leitrim, Sligo, Wicklow. v-vii, ix-x.
_	Costa reaching distinctly beyond R5; r-m shorter, oblique and not approaching alignment with R5.
12	Sc2 normally absent; Sc bare, ending well before base of Rs; Anal vein with macrotrichia shoulders and large lateral triangles on tergites 2-4 of abdomen yellow; setae at aper of tibia almost in rows; or tarsal claws unmodified; wing length 4-4.5mm; or genitalia (fig. 250) with cerci small and without combs of spines; Q ovipositor very attenuate
	Berks, Cambs, Chesh, Hants, Herts, Lancs, Westmorland, Yorks; Wales: Caerns,
	Merioneth; Scotland: Dumfries, Inverness, Moray, Perth, Sutherland; Ireland. Wicklow. v-vi, ix. Decaying wood.
_	Sc2 present; Sc reaching to approximately base of Rs, usually bare; Anal vein usually bare; σ genitalia usually with well developed cerci bearing combs of spines;
13	Q ovipositor short and contracted
_ 14	Antennae all black, or at most with first flagellar segment partly yellow
14	particularly in \circ ; macrotrichia on entire posterior fork and its stem; wing length 4.5-5mm; all \circ tarsal claws large, blunt and very finely pectinate; \circ genitalia (fig. 251); \circ sternum 8 well developed and rather pointed with about 8 strong bristles apically
	basalis Meigen
	Berks, Chesh, Derby, Devon, Gloucs, Hants, Heref, Herts, Kent, Lancs, Oxon, Shrop, Somerset, Surrey; Wales: Merioneth; Scotland: Arran, Inverness, Midlothian, Moray, Perth; Ireland: Antrim, Down, Killarney. v-vi, viii-ix.
_	Thorax and abdomen all black; macrotrichia on posterior fork restricted to outer half of
	the fork veins; wing length 4-5mm; all or tarsal claws unmodified; or genitalia (fig.

15 Hind coxa distinctly black, usually yellow at extreme tip; macrotrichia well developed on all postradial veins, including Anal vein; Anal vein very long; setae occasionally present on Sc; wing length 4.5-5mm; σ tarsal claws all large, blunt and very finely pectinate, anterior claws slightly smaller than posterior; σ genitalia (fig. 253); Q sternum 8 well developed and rounded with about 6 strong bristles apically

Scotland: Inverness, Perth. vi. groenlandica Staeger

Hind coxa not distinctly black, but may be somewhat darkened; Anal veing bare and short as usual; wing length less than 4mm; or tarsal claws small, not pectinate......16
 [It is only possible to satisfactorily identify these last nine species (the sciarina-group) by

reference to the male genitalia, noting particularly the number and size of cercal combs, shape of dististyles, ventral process of basistyle and parameres. The characters given for nigrofusca and moravica should be used with caution, details of the claws are difficult to detect. Note that trispinosa and usually gripha have darkened hind coxae. The first flagellar segment of brevicornis is usually vellowish.

- 16 Macrotrichia of posterior fork continuous from wing margin to just before fork (occasionally also in *nigricans* and *moravica*); wing length 3-3.5mm; all σ tarsal claws broad and rounded with an indistinct apical spur; σ genitalia (fig. 254)

Tribe Leiini

A tribe of less than 20 genera, most of which contain few species. Three genera that occur in the Holarctic have not been found in Britain. These are Novakia, Greenomyia and Neoclastobasis. Novakia Strobl is very similar to Tetragoneura Winnertz, but differs in having vein r-m extended back almost to the wing base and is therefore several times longer than R1, in having a short fusion of R1 and R4 + 5 just beyond r-m (Tetragoneura may show this fusion, but usually has a short R4 present enclosing a very narrow small cell), in having the dorsum of the thorax uniformly covered in short hairs, while Tetragoneura has two narrow median anterior bare stripes and narrow lateral bare stripes, and in having the antennae short and stout, about as long as the head, while they are more than twice as long in

Tetragoneura. Novakia is known from one Palaearctic, one Nearctic and one Neotropical species. Novakia has recently been associated with genera that are here included in the Sciophilini (Papavero, 1978). Greenomyia Brunetti is similar to Leia, but the ocelli are remote from the eye margin, the Median and posterior forks are complete (anterior vein of posterior fork slightly interrupted in one species) and the whole of the wing tip is usually dark. In some ways it is like Rondaniella, but Sc2 is present, the Costa does not extend beyond R5, r-m is double the length of R1. Greenomyia is known from 4 Palaearctic, 2 Oriental and one undescribed Nearctic species. Neoclastobasis and Clastobasis were considered as a separate tribe, Clastobasini, but this group is now regarded as belonging to the Leiini. Neoclastobasis is known from two Palaearctic species and has the whole of the wing tip dark, the anterior vein of the posterior fork neither connected at the base nor reaching the wing margin, the posterior vein of the Median fork not reaching the wing margin, lateral ocelli remote from the eye margin, palps very long—about as long as the antennae. Matile (1978) provides a key to genera.

One other genus, Allactoneura Enderlein, which has been reared from stored products in the Old World tropics and has probably been spread around the tropics by human agency, may appear in Britain. With its lack of strong prothoracic bristles, flattened back to the head, distinct orbital bristles and antennae inserted above the middle of the head, it may run to the subfamily Manotinae. However, it is generally regarded as belonging to the Leiini. Apart from the above characters the dorsum of the thorax, both surfaces of the abdomen and the upper surface of the anterior wing veins are covered with scales. r-m is remarkable in having a long horizontal section that fades away towards the wing base, while the connection with the Median veins is through a short vertical section. The veins of the posterior fork are separate to the wing base. The larvae are probably saprophagous rather than fungivorous.

Little is recorded of the adult activity of the members of this tribe, but some genera, such as *Leia*, may be found in drier habitats and higher above the ground than is usual in the family. Species have been reared from a variety of habitats, including fungi, fungus infected wood, other vegetable matter and the nests of birds and mammals. Some tropical species have been associated with stored products.

Key to genera

- Sc ending free or in R
 R1 over twice as long as r-m, which is rather oblique; Sc2 absent; anterior branch of Median fork often detached at base; lateral ocelli far from eye margin; Costa produced beyond apex of R5; stem of Median fork much longer than r-m; apex of wing dark; anterior branch of posterior fork disconnected at base, apparently arising from wing base as a fold in the wing membrane
 RONDANIELLA Johannsen (p.64)
- R1 shorter than r-m, which is horizontal (fig. 49); Sc2 present; anterior branch of Median fork complete at base or almost so; lateral ocelli rather variable in position, but usually close to eye margin; Costa ending at apex of R5; stem of Median fork not longer than r-m; wings usually with a subapical fascia; anterior branch of posterior fork usually narrowly disconnected at base, but originating well beyond wing base

Genus Rondaniella Johannsen

(Figs. 34, 263)

A very small genus. Within Europe, apart from the single species included here, a second species has been recognised at various times, but this is generally regarded as a synonym. Outside Europe there is one Nearctic species that occurs as far south as Mexico and one Oriental species. While the Oriental species would seem to be a valid species, we can see no reliable differences between North American and European specimens and the status of these two species needs investigations.

Adult activity is unrecorded, but the species has been bred frequently from a variety of fungi.

Genus **Leia** Meigen (Figs. 41, 49, 60-61, 264-272)

This is by far the largest genus of the tribe, with over 130 species distributed throughout the world. The greater number of species are described from South America, but the genus is well represented in the northern hemisphere with about 20 species in Europe.

The adults may be found in the foliage of trees as well as lower vegetation and are one of the groups that frequently occurs on windows. Species have been bred from fungi, from decaying wood and from the nests of birds and mammals. One immigrant species is included in the key, although still only known from one record.

Key to species

1 Halteres black; wings with clouds over base of cell R5, below posterior fork and before apex of wing; thorax orange with black marks at base of wings and centre of posterior margin of thorax; wing length 3.5-4mm; or genitalia (fig. 264)..... arsona Hutson London. vi. In rotting root ginger (Zingiber).

_	Halteres white
2	A distinct dark spot present at base of cell R5
_	No dark spot at base of cell R5 (but see piffardi Edwards, couplet 8)4
3	Without an isolated dark spot at the tip of R5; a fairly broad, diffuse preapical fascia
	crossing wing; dorsum of thorax with at most reduced lateral stripes; scutellum all
	yellow; wing length 3.5-4.5mm; or genitalia (fig. 265); terminal segment of female
	cercus large and setose bifasciata Gimmerthal
	Chesh, Hants, Somerset, Yorks. iv-v, vii-viii. From nest of Red Squirrel (Sciurus
	vulgaris); also under bark of oak (Quercus).

- Abdomen with black bands on tergites; hind femora with distinct black apical bands...6
 First abdominal tergite all orange; no distinct black mark at base of fork of axillary veins; frequently with small weak cloud below posterior fork; wing length 4-5mm; or genitalia (fig. 267); terminal segment of Q cercus elongate, deeply sunk into penultimate segment and with a dark patch at its base (fig. 60).... fascipennis Meigen Bucks, Cambs, Chesh, Cumberland, Devon, Dorset, Essex, Gloucs, Hants, Heref, Herts, Hunts, Kent, Lancs, Middx, Oxon, Somerset, Surrey, Sussex, Yorks; Wales: Merioneth, Monmouth, Powys; Scotland: Argyll, Arran, Inverness, Perth; Ireland:
- First abdominal tergite narrowly black at tip; a distinct black mark at base of fork
 of axilliary veins; no cloud below posterior fork; wing length 4-5mm; or genitalia
 (fig. 268); terminal segment of 9 cercus small rounded and yellow (fig. 61)
 - Berks, Derby, Essex, Gloucs, Heref, Hunts, Kent, Oxon, Suffolk, Surrey; Scotland: Perth, Ross & Crom; Ireland: Wicklow. iv-ix.

[N.B. This species may be a synonym of L. subfasciata Meigen.]

Down, Dublin, Galway, Leitrim, Louth, Waterford. v-x.

- - 8 Colour very variable, from largely black thorax and abdomen to largely pale, even in the palest specimens (var. fasciola Meigen) bands on abdominal tergites usually broader in middle than at sides (especially tergite 2); tergite 1 distinctly angled at apex; no clouds on wing apart from the subapical fascia; wing length 4-6mm; ♂ genitalia (fig. 271); ♀ sternum 8 evenly convex dorsolaterally...... bimaculata Meigen Beds, Berks, Cambs, Chesh, Dorset, Essex, Hants, Heref, Herts, Kent, London, Middx, Norfolk, Surrey, Sussex, Warwick, Yorks; Wales: Merioneth; Ireland: Down. v-x. Calocera, Collybia, Hebeloma, Russula (decaying).

- [N.B. There are very few intermediates between typical bimaculata and the variety fasciola. In the British Museum (Natural History) 18 specimens have an all black mesonotum without a trace of differentiable stripes, 28 have an all pale mesonotum without a trace of stripes and six are intermediate in showing a variable amount of striping on the mesonotum. These six include specimens bred from a sample that includes all dark specimens.]

Genus Megophthalmidia Dziedzicki

(Figs. 22-23, 44, 273)

About four species are known in this genus in the western Palaearctic and one in Japan. One is known from North America and about seven from South America. Most Holarctic species are darker and smaller than the species recorded here. Nothing is known of their life history.

- Stout orange species, with short dense black bristles; wing length 3-4mm; antennae stout; mouthparts not noticeably elongate; wings yellowish with darkened apex; posterior fork before base of stem of Median fork; σ genitalia (fig. 273)
 - Cambs, Cornwall, Devon, Hants, Heref, Middx, Somerset, Staffs, Surrey, Sussex;

 Ireland: Down. vi-ix.

Genus **Tetragoneura** Winnertz (Figs. 12, 45, 86, 274)

A widespread genus of about 60 species, but only four species described from the Palaearctic region. Over half the described species are from the Neotropical region, 14 from New Zealand, about 10 in the Nearctic region and a few from Australia. Of the Palaearctic species, two occur in Europe and two in Japan. The single species occuring in Britain is one of the most common woodland species. The only reliable characters so far published for separating the Palaearctic species are in the male genitalia.

The larvae live in a mucilaginous tube among bark encrusting fungi on small fallen branches. A slight dry cocoon is spun for pupation.

— Small black species, wing length 2.5-3mm; head, and thorax and abdomen black with white bristles; pedicel of antennae orange, rest black; haltere yellow; base of hind and mid-coxae and ventral surface of femora dark, rest of legs yellow (these colour characters slightly variable); 2nd palpal segment slightly swollen; R4 present or base of Rs fused with R1; \(\sigma\) mid-tibia without sensory area; \(\sigma\) genitalia (fig. 274)

Berks, Chesh, Cumberland, Devon, Dorset, Essex, Gloucs, Hants, Heref, Herts, Kent, Lancs, Middx, Norfolk, Notts, Oxon, Shrop, Suffolk, Surrey, Sussex, Warwick, Westmorland, Wilts, Worcs, Yorks; Wales: Brecknock, Caerns, Denbigh, Flint, Merioneth; Scotland: Banff, Inverness, Moray; Ireland: Down, Galway, Kerry, Killarney, Wicklow. v-x. From Poria.

Genus Ectrepesthoneura Enderlein

(Figs. 40, 85, 275-277)

This is an Holarctic genus of six European and an uncertain number of Nearctic species. Two species have been described from Japan, but the published details strongly suggest that they belong to the family Sciaridae. Some authors (e.g. Vockeroth, 1980) have suggested that this genus is not differentiable from Tetragoneura, but in the Palaearctic the two groups are distinct and so they are separated here in the absence of any firm synonymy. The inclusion of four recently described European species in this genus might enable a more valid re-appraisal of the status of this genus. Chandler (1979b) discusses the genus, including some of the Nearctic species and provides a key to the European species published so far. Most of the species are only satisfactorily separated by reference to the male genitalia, which are very distinct, but E. gracilis Edwards is distinct in being a more elongate species with slight clouds at the wing tips, across the middle of the wing and behind the posterior fork. E. gracilis also has the post-radial veins strongly developed, anal vein bare, R5 somewhat sinuous, R1 about twice length of r-m, etc. E. referta Plassmann is very similar to the species discussed below. One other species was not seen during the present study, while its description was awaiting publication by Plassmann. This is another genus in which so few specimens have been seen of most of the species that it has not been possible to test the reliability of many of the characters included here. They are all small, dark species with a wing length of 2.5-3.5mm and a sensory area near the base of the mid-tibia in the male.

Two species have been reared from dead wood.

Key to species

1 Femora without black markings at base on ventral surface; hind femora only weakly darkened at apex; Costa not reaching beyond half distance from R5 to apex of anterior branch of Median fork; bare stripes of mesonotum broader than the areas of bristles that separate them; 1st and 2nd visible palpal segments swollen, but not greatly, 1st shorter than 2nd (penultimate) segment; preapical comb of front tibia pale; tibial spurs pale; thorax and abdomen uniformly black; peucel and base of 1st flagellar segment pale, rest dark; anterior bristles of hind tibia divided into basal and apical groups, basal group of larger bristles with a distinctly large dark last bristle, apical group of uniformly weaker bristles; or genitalia (fig. 275)

Berks, Cambs, Devon, Hants, Heref, Herts, Kent, Lancs, Middlx, Norfolk, Oxon, Surrey, Sussex; Scotland: Perth; Ireland: Down. v-x. From dead wood, sometimes associated with encrusting fungi. e.g. Coriolus.

Hants, Sussex; Scotland: Dunbarton, vi-vii,

Genus Docosia Winnertz

(Figs. 87, 278-286)

A genus of about 25 Palaearctic, 15 Nearctic and 2 Neotropical species. The Palaearctic species are being revised by Lastovka & Matile (in prep.). All species are dark with pale hairs and with a similar general appearance to species of *Ectrepesthoneura* and *Tetragoneura*. Adult activity is unrecorded and most species are generally uncommon on the wing.

One species has been reared from a wide variety of fungi. On one occasion a single specimen of this species emerged from the nest of a Short-tailed Vole (Microtus agrestis), but it is possible that the larva did not develop there, but only pupated there. This species differs markedly from the other British species, especially in the female ovipositor. Another species has been reared from the disused nests of a variety of birds. In some cases these nests had been taken over for the winter by small rodents. A third species has been bred from fungi and from rotting wood. The larvae of the fungus feeders are unusual in the Sciophilinae in living actually inside the fungus and usually in fungi that are in an advanced state of decay. The larvae in birds' nests are unusual in being free-living, without a mucilaginous tube and are believed to be saprophagous. Pupation takes place in a dense tough cocoon covered with dirt particles.

Key to species

- 4 Hind femur dark at base and tip only; apical half of wing slightly darkened; hairs on dorsum of thorax long and white; about 9 dorsal and 15-16 anterior bristles on hind tibia; wing length about 3.5mm; ♂ genitalia (fig. 281).....setosa Landrock Yorks; Wales: Powys; Scotland: Perth. v.
- Hind femur with base, apex and entire ventral edge dark; wings quite clear; hairs on dorsum of thorax short and slightly yellow; 7 dorsal and 13-14 anterior bristles on hind tibia; wing length about 3.5mm; ♥ genitalia (fig. 280)......sp. indet.

	Yorks; Wales: Powys. vi.
	[This species is to be described in a forthcoming revision of the genus by Lastovka et
	al.]
5	Hind femur completely black6
	Hind femur at least partly yellow
6	Entire wing slightly brown tinged, especially Costal region; coxae ochreous; bristles
	of dorsum of thorax short and darker, supra-alar bristles especially dark; wing
	length 2.5-3.5mm; or genitalia (fig. 282) fumosa Edwards
	Chesh, Cornwall, Gloucs, Herts, Oxon, Surrey, Sussex. iv-ix. From nest of Aves:
	Corvus (Corvidae), "finch" (Fringillidae), Prunella (Prunellidae), Turdus (Turdidae).
_	Wings clear; coxae black; bristles of dorsum of thorax very pale and long, with white
	supra-alar bristles; wing length 2.5-3.5mm; or genitalia (fig. 283)
	carbonaria Edwards
	Oxon, Norfolk, Suffolk. iv-v.
7	Coxae largely dark; pleurotergal hairs very short; hind femur with tip and whole of
	ventral surface dark, the dark area sometimes extended so that pale area incon-
	spicuous; wing length 3.5-4.5mm; & genitalia (fig. 284) sciarina Meigen
	Berks, Cambs, Devon, Gloucs, Hants, Heref, Herts, Kent, Suffolk, Surrey, Wilts;
	Wales: Caerns, Powys; Scotland: Perth. iv-vi.
_	Coxae yellow; pleurotergal hairs long as usual
8	Hind femur at most with slightly blackened tip; Sc usually with at least one macro-
	trichia; wing length 3-4mm; o genitalia (fig. 285) pallipes Edwards
	Beds, Hunts, Norfolk, Suffolk; Wales: Glamorgan, Scotland: Inverness, Moray.
	vii-ix.
—	Hind femur darkened at base beneath and at tip; Sc bare; wing length 3.5-4mm; or
	genitalia (fig. 286) moravica Landrock
	Berks, Devon, Heref, Herts, Lancs, Oxon, Norfolk, Suffolk; Wales: Flint, Powys.

Subfamily Manotinae

v-vii.

A small subfamily of four genera, only one of which occurs in the Palaearctic. Tuomikoski (1966b) has discussed the relationships of this subfamily and the genera included in it. *Allactoneura* was included when the subfamily was first described, but is now considered to belong to the Leiini (Sciophilinae). Should this genus occur in Britain, possibly in tropical plant products, it will probably run to this subfamily in the key, but it is discussed under Leiini (p.62).

Genus Manota Williston

(Figs. 13, 287)

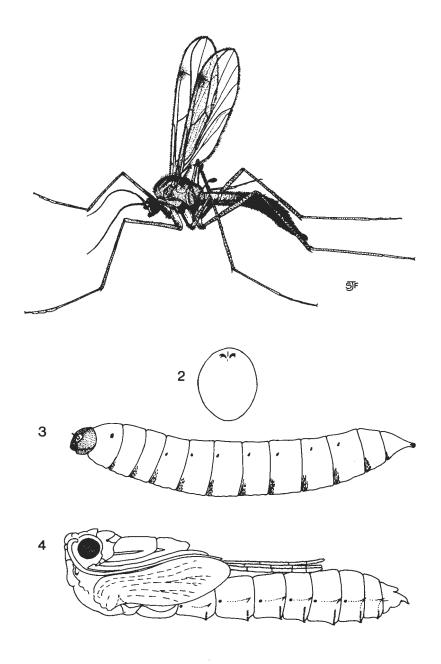
A genus of about 15 species including one Palearctic species and an unnamed Nearctic species. Most of the described species are from Africa, but many species remain to be described from here and elsewhere in the tropics. In temperate areas it is rarely found and there are few records of the European species. It has been collected at light and by sweeping the lower branches of a larch tree (*Larix europea*). A single specimen is recorded (Chandler, 1978) as emerging from rotten beech wood bearing a Myxomycete, but any association requires confirmation.

References

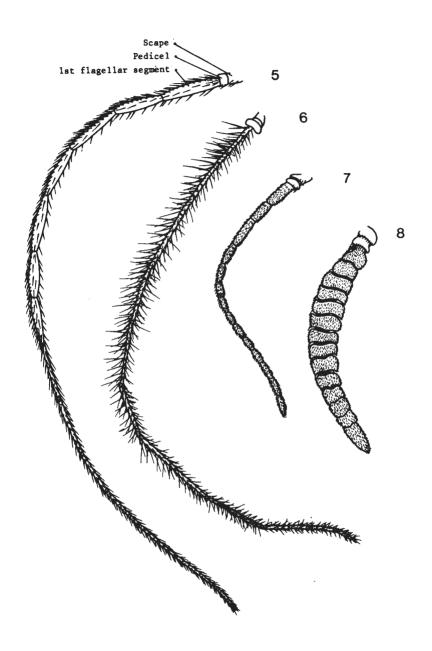
- ASKEW, R. R. 1971. Parasitic Insects. 316pp, Heinemann, London.
- BEĬ-BIENKO, G. Ya. (ed) 1969. Diptera, Fleas. Part 1. Keys to the insects of the European part of the U.S.S.R. V. Opred. faune SSSR 100: 1-807. [In Russian].
- BUXTON, P. A. 1954. British Diptera associated with fungi. 2. Diptera bred from Myxomycetes. *Proc. R. ent. Soc. Lond.* (A) 29: 163-171.
- ——1961. British Diptera associated with fungi. III. Flies of all families reared from about 150 species of fungi. *Entomologist's mon. Mag.* 96: 61-94.
- CHANDLER, P. J. 1976. Gnoriste longirostris Siebke (Diptera, Mycetophilidae) new to Britain: its synonymy with G. groenlandica Lundbeck established *Proc. Brit. ent. nat. Hist. Soc.* 1976: 36-41.
- ----1978. Notes on British Fungus Gnats of the subfamilies Ditomyiinae, Bolitophilinae, Diadocidiinae, Manotinae and Keroplatinae (Dipt., Mycetophilidae). *Entomologist's mon. Mag.* 113: 31-44.
- ——1979a. Association with Fungi. p. 199-211 in Stubbs, A. E. & Chandler. P. J. (eds) 1979 q.v.
- ——1979b. The European and eastern Nearctic fungus-gnats in the genus *Ectrepesthoneura* (Mycetophilidae). Syst. ent. 5: 27-41.
- COGAN, B. H. & SMITH, K. G. V. 1974. Insects. *Instructions for Collectors* no. 4a. 169pp, British Museum (Natural History), London.
- DELY—DRASKOVITS, A. 1974' Systematische und ökologische untersuchungen an den in Ungarm als Schädlinge der Hutpilze auftretenden Fliegen VI. Mycetophilidae (Diptera). Folia ent. hung. 27(1): 29-41.
- DZIEDZICKI, H. 1885. II. Przyczynek do fauny owadów dwuskrzydlych. Rodzaje nowe: Hertwigia, nov. Gen.; Eurycera, nov. Gen. i gatunki rodzajów: *Boletina*, *Sciophila*. *Pam. fizyogr.* 5: 164-194.
- EDWARDS, F. W. 1925. British Fungus-Gnats (Diptera, Mycetophilidae). With a revised Generic Classification of the Family. *Trans. ent. Soc. Lond.* 1924: 505-670.
- ---1941. Notes on British Fungus-Gnats (Dipt., Mycetophilidae). Entomologist's mon. Mag. 77: 21-32, 67-82.
- EISFELDER, I. 1954. Beiträge zur Kenntnis der Fauna in höheren Pilzen. Z. Pilzk. 16: 1-12. ——1955. Die häufigsten Pilzbewohner. Z. Pilzk. 19: 12-20.
- HACKMAN, W. & MEINANDER, M. 1979. Diptera feeding as larvae on macrofungi in Finland. Annls zool. fenn. 16: 50-83.
- HENDERSON, D. M., ORTON, P. D. & WATLING, R. 1969. British Fungus Flora. Agarics and Boleti: Introduction. 58pp. HMSO, Edinburgh.
- HENNIG, W. 1954. Flügelgeäder und System der Diptera unter Berucksichtigung der aus dem Mesozoikum beschreibenen Fossilien. Beitr. Ent. 4: 245-388.
- HICKMAN, V. V. 1965. On *Planarivora insignis* gen. et sp. n. (Diptera: Mycetophilidae), whose larval stages are parasitic in land Planarians. *Pap. Proc. R. Soc. Tasm.* 99: 1-8.
- HUTSON, A. M. 1979. Notes on Sciophilinae (Dipt., Mycetophilidae) with a revision of Palaearctic Syntemma Winnertz Entomologist's Mon. Mag. 114: 131-145.
- --- & KIDD, L. N. 1971. Notes on British Bolitophilinae including three species new to Britain (Dipt., Mycetophilidae). *Entomologist* 104: 219-226.
- ————1974. Notes on British Bolitophila, Diadocidia and Macrocera (Diptera, Mycetophilidae). Entomologist's mon. Mag. 110: 27-39.
- KIBBY, G. G. 1979. Mushrooms and toadstools, a field guide. Oxford 256pp.
- KLOET, G. S. & HINCKS, W. D. 1976. A check list of British Insects. Diptera and Siphonaptera. *Handbk. Ident. Br. Insects* 11(5): I-139.
- LAFFOON, J. L. 1965. Family Mycetophilidae (Fungivoridae). *In Stone*, A., Sabrosky, C. W., Wirth, W. W., Foote, R. H. & Coulson, J. R. A catalog of the Diptera of America north of Mexico. *Agric. Handb. Forest Serv. U.S.* No. 276: iv + 1-1696.
- LANDROCK, K. 1926-27. 8. Fungivoridae (Mycetophilidae). Fliegen palaearkt. Reg. II(1): 1-195.
- LAŠTOVKA, P. 1971. A study on the last instar larvae of some Czechoslovak *Mycetophila* (Diptera, Mycetophilidae). *Acta Univ. Carol.*, *Biologica* 1970: 137-176.
- ——& MATILE, L. 1972. Révision des *Diadocidia* Holarctiques. (Dipt. Mycetophilidae). *Annls Soc. ent. Fr.* (N.S.) 8(1): 205-223.

- MADWAR, S. 1937. Biology and Morphology of the immature stages of Mycetophilidae (Diptera, Nematocera). *Phil. Trans. R. Soc. B*, 227: 1-110.
- MASNER, L. 1968. The Fungus Gnats (Dipt., Mycetophiloidea) as potential hosts of Proctotrupid wasps (Hym., Proctotrupoidea). Acta ent. bohemoslavaca 65: 464-466.
- MATILE, L. 1969. Note sur les genres Keroplatus et Cerotelion et description de Cerotelion racovitzai n.sp. (Dipt., Mycetophilidae). Bull. Soc. ent. Fr. 74: 82-86.
- --- 1970. Les Diptères Cavernicoles. Annls Spéléol. 25(1): 179-222.
- ——1974. Notes sur les Mycetophilidae (Diptera) de la Fauna de France. III. Le genre Neuratelia. *Entomologiste*, 30(1): 26-33.
- ——1975. Révision des Asindulum et des Macrorrhyncha de la région Paléarctique (Dipt., Mycetophilidae). Annls Soc. ent. Fr. (N.S.) 11(3): 491-515.
- ——1976. Notes sur les Mycetophilidae (Diptera) de la faune de France. IV. Le genre Bolitophila. 1: sous-genre Bolitophila s. str. (1re partie). Entomologiste 32(6): 235-244.
- ——1977. Révision des Keroplatinae du genre Antlemon (Dipt., Mycetophilidae). Annls Soc. ent. Fr. (N.S.) 13(4): 639-649.
- ——1978. Description d'un *Neoclastobasis* nouveau de Hongrie et remarques sur divers Leijni (Diptera: Mycetophilidae). *Folia ent. hung.* N.S. 31(1): 167-172.
- MUNROE, D. D. 1974. The systematics, phylogeny, and zoogeography of Symmerus Walker and Australosymmerus Freeman (Diptera: Mycetophilidae: Ditomyiinae) *Mem. ent. Soc. Can.* 92: 1-183pp.
- OLDROYD, H. 1970. Diptera. 1. Introduction and key to families. *Handbk Ident. Br. Insects* 9(1): 1-104.
- OSTROVERKHOVA, G. P. & STACKELBERG, A. A. 1969 24. Fam. Mycetophilidae (Fungivoridae). pp.265-320 in Bei-Bienko, 1969 (q.v.).
- PAPAVERO, N. 1978. A Catalogue of the Diptera of the Americas south of the United States. 19E Family Mycetophilidae (Fungivoridae) 78pp. Sao Paulo.
- PECK, S. B. & RUSSELL, D. R. 1976. Life history of the fungus gnat *Macrocera nobilis* in American caves (Diptera: Mycetophilidae). Can Ent. 108: 1235-1241.
- PEGLER, D. N. 1973. The Polypores. Bull. Br. mycol. Soc. 7(1) suppl.: 1-43.
- PLASSMANN, E. 1969. Die Fungivoriden des Naturschutzparkes Hoher Vogelsberg (Ein Beitrag zur Biologie, Ökologie und Taxonomie der Fungivoriden, sowie zur kenntnis der Larven). 163pp., Inaugral Dissertation, Giessen.
- ——1971a. Uber die Fungivoriden-fauna (Diptera) des Naturparkes Hoher Vogelsberg. Oberhess. naturw. Z. 38: 53-87.
- ——1971b. Die Pilzmückengattung Messala (Dipt., Fungivoridae). Ent. Z. 81: 164-173.
- ——1975. Revision der europäischen Arten der Pilzmücken-gattung Bolitophila Meigen (Dipt: Mycetophilidae). Entomologica scand. 6: 145-157.
- RHODENDORF, B. B. 1964. Historical development of Diptera. *Trudy paleont. Inst.* 100: 1-311. [In Russian; English translation by Moore & Thiele, published Alberta 1974, 360pp].
- RICHARDS, A. M. 1960. Observations on the New Zealand Glow-worm Arachnocampa luminosa (Skuse) 1890. Trans. R. Soc. N.Z. 88: 559-574.
- RICHARDS, O. W. 1977. Hymenoptera. Introduction and keys to families. *Handbk. Ident. Br. Insects* 6(1): 1-100.
- RUSSELL-SMITH, A. 1979. A study of fungus flies (Diptera: Mycetophilidae) in beech woodland. *Ecological Entomology*, 4: 355-364.
- SAIGUSA, T. 1973. A systematic study of the Mycetophilidae of Japan (Diptera). Part 1. A revision of the subfamily Ditomyiinae. Sieboldia 4(3): 167-200.
- STACKELBERG, A. A. 1969a. 17 Family Bolitophilidae. Pp.247-257 in Bef-Bienko, 1969 (q.v.). ——1969b. 18. Fam. Ditomyiidae. P.257 in Bef-Bienko, 1969 (q.v.).
- --- 1969c. 19. Fam. Ceroplatidae (Keroplatidae). Pp. 258-261 in Bel-Bienko, 1969 (q.v.).
- ——1969d. 21. Fam. Macroceridae. Pp.261-264 in Bel-Bienko, 1969 (q.v.).
- STEFFAN, W. A. 1966. A generic revision of the family Sciaridae (Diptera) of America north of Mexico. *Univ. Calif. Publs Ent.* 44: 1-77.
- STEINHAUS, E, A, (ed.) 1963. Insect Pathology, and advanced treatise. 2 vols: 661 & 689pp. London.
- STUBBS, A. E. & CHANDLER, P. J. (eds.) 1979. A Dipterists' Handbook. *Amat. Ent.* 15: i-ix, 1-255.

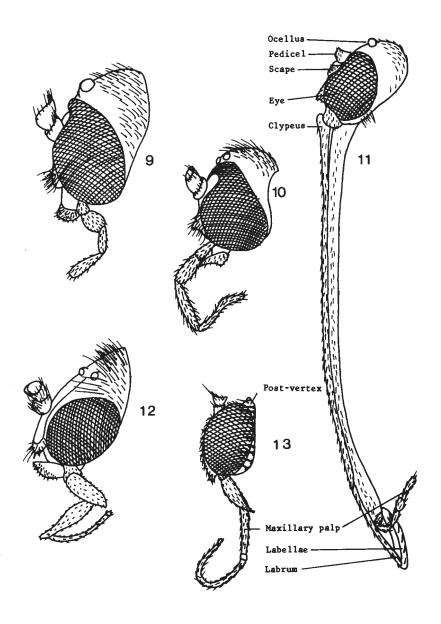
- TRIFOURKIS, S. 1977. The bionomics and Taxonomy of the larval Mycetophilidae and other fungicolous Diptera. PhD Thesis, North East London Polytechnic, 732pp. (unpublished).
- TUOMIKOSKI, R, 1966a. Zur Kenntnis der Sciariden (Dipt.) Finnlands. Suomal. eläin-ja kasvit. Seur. van. eläin Julk, 21(4): 1-164.
- ---1966b. On the subfamily Manotinae Edws (Dipt., Mycetophilidae). Suom. hyont. Aikak. 32(3): 211-223.
- VOCKEROTH, J. R. ?1980. in Manual to the Families and Genera of Nearctic Diptera. Volume 1. Ottawa (in press).
- ZAITZEV, A. I. 1978. Flies of the family Ditomyiidae (Diptera) from the U.S.S.R. Ent. Obozr., 57: 668-676.



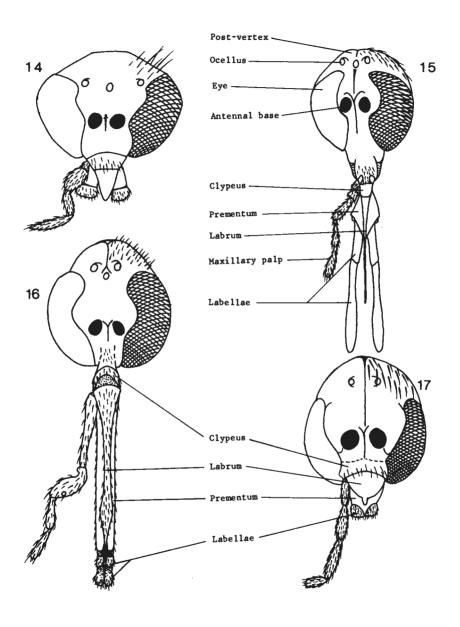
Figs 1-4. 1, adult Q Bolitophila cinerea (drawn by S. Falk). 2, egg of unidentified Mycetophilid. 3, 4th instar larva of Bolitophila cinerea. 4, pupa of B. occlusa.



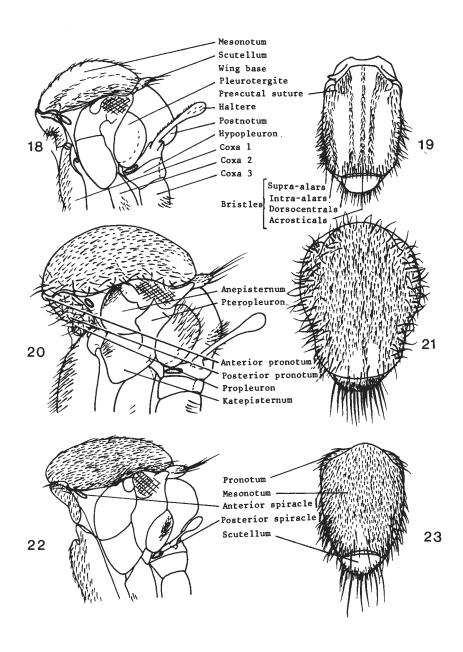
Figs. 5-8. Antennae. 5, Macrocera parva σ . 6, Bolitophila saundersii σ . 7, Boletina basalis σ 8, Cerotelion lineatus σ .



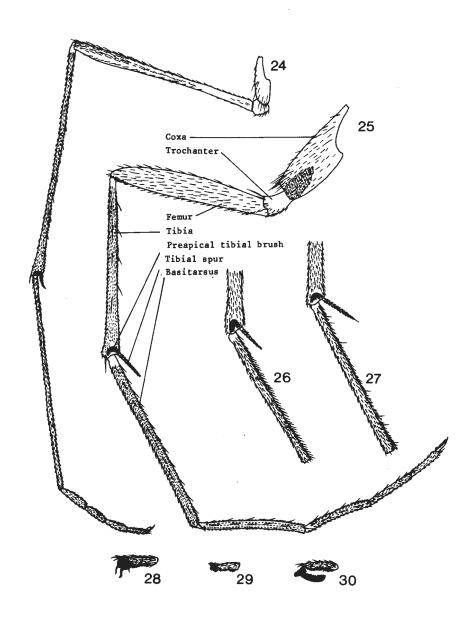
Figs. 9-13. Heads, lateral view. 9, Symmerus annulatus σ . 10, Diadocidia ferruginosa σ . 11, Gnoriste bilineata σ . 12, Tetragoneura sylvatica σ . 13, Manota defecta σ (Caribbean species).



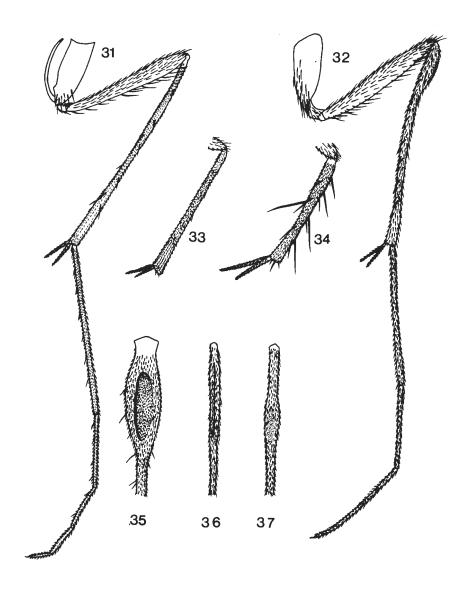
Figs. 14-17. Heads, anterior view. 14, Bolitophila hybrida \heartsuit . 15, Asindulum nigrum \circlearrowleft . 16, Antlemon servulum \circlearrowleft . 17, Mycomya incisurata \heartsuit .



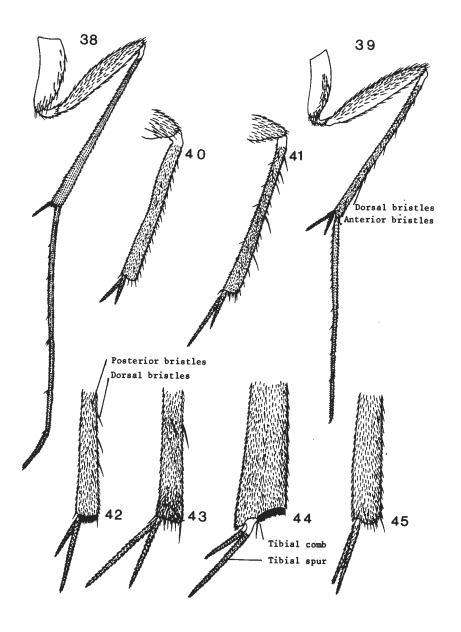
Figs. 18-23. Thorax (left: lateral view; right: dorsal view). 18-19, Orfelia (Urytalpa) atriceps & 20-21, Sciophila rufa & 22-23, Megophthalmidia crassicornis &.



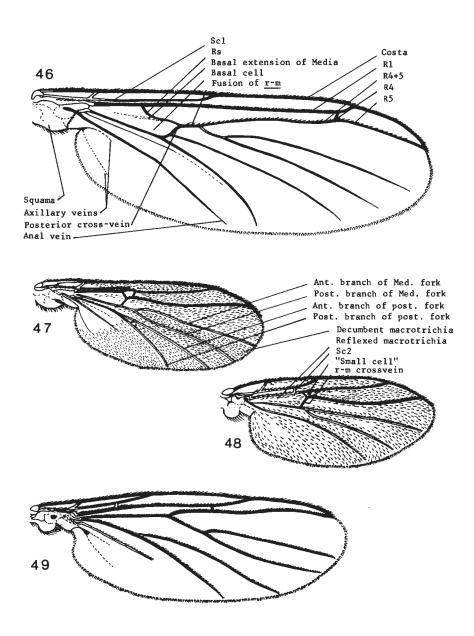
Figs. 24-30. Fore legs. 24, Bolitophila saundersii Q. 25, Mycomya fuscata σ . 26, Diadocidia ferruginosa σ . 27, D. spinosula σ . 28, Boletina trivittata σ . 29, B. gripha σ . 30, B. důbia σ . (Figs. 24-27 anterior view; figs. 28-30 posterior view of 5th tarsal segment).



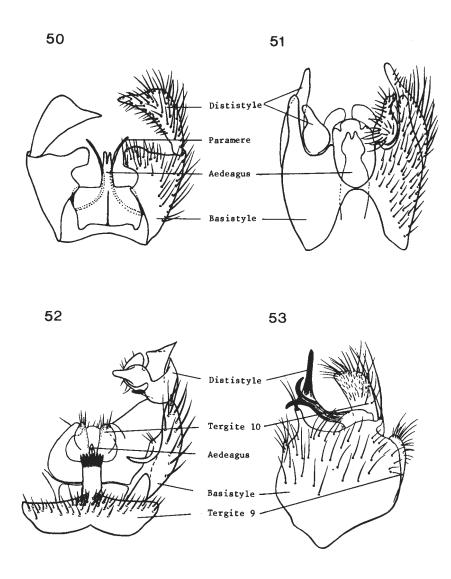
Figs. 31-37. Mid-legs. 31, Mycomya cinerascens σ . 32, Polylepta guttiventris σ . 33, Orfelia (Neoplatyura) flava σ . 34, Rondaniella dimidiata φ . 35, Polylepta guttiventris σ . 36, Phthinia humilis σ . 37, Speolepta leptogaster σ . (Figs. 31-34 anterior view; figs. 35-37 dorsal view of base of tibia).



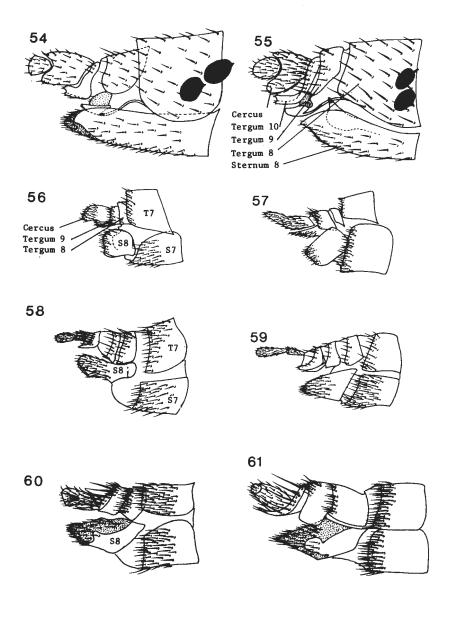
Figs. 38-45. Hind legs. 38, Orfelia (Rutylapa) ruficornis 9. 39, Orfelia (Pyratula) zonata σ . 40, Ectrepesthoneura hirta σ . 41, Leia piffardi σ . 42, Acnemia nitidicollis σ . 43, Neuratelia nemoralis σ . 44, Megophthalmidia crassicornis σ . 45, Tetragoneura sylvatica σ . (Figs. 38-41 anterior view; figs. 42-45 posterior view).



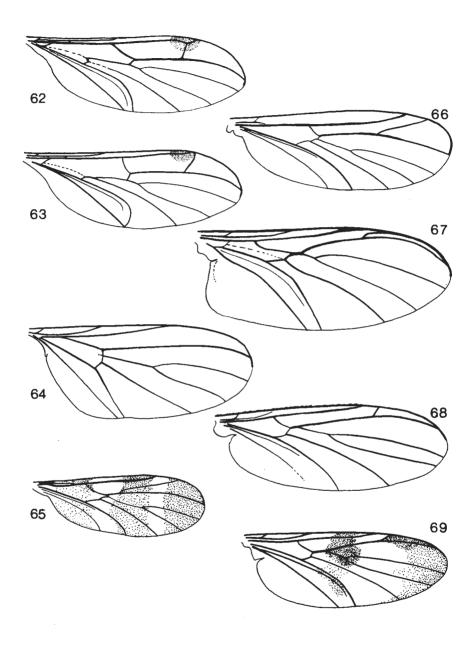
Figs. 46-49. Wings. 46, Keroplatus testaceus \circ . 47, Syntemna stylata \circ . 48, Monoclona rufilatera \circ . 49, Leia bimaculata \circ . (See also figs. 62-87).



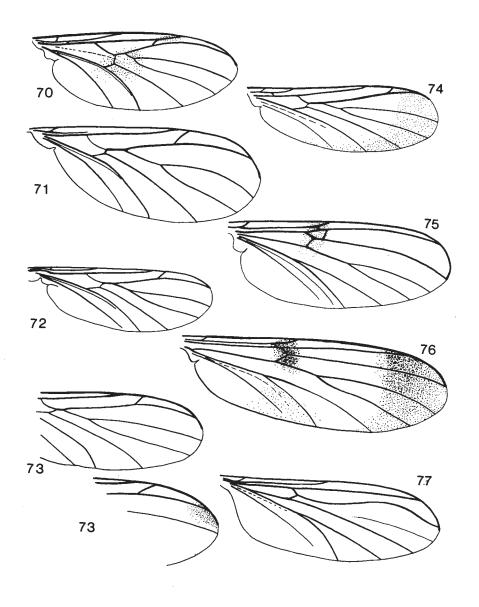
Figs. 50-53. σ genitalia. 50, Bolitophila pseudohybrida (dorsal, tergite 9 removed). 51, Orfelia (Urytalpa) atriceps (ventral). 52, Syntemna nitidula (dorsal). 53, Leia subfasciata (lateral).



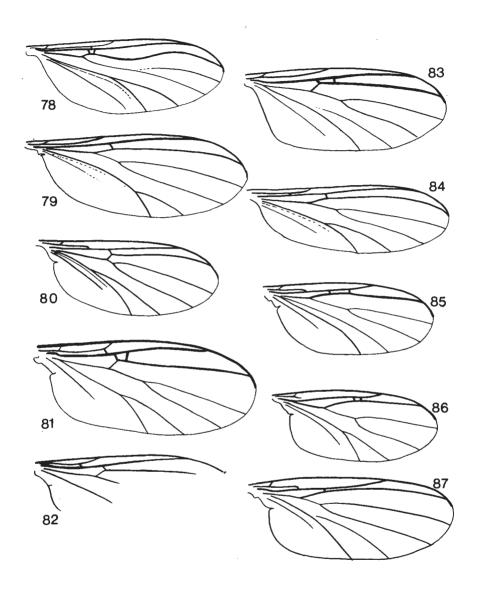
Figs. 54-61. \heartsuit ovipositor (lateral). 54, Bolitophila cinerea. 55, B. saundersii. 56, Orfelia (O.) nemoralis. 57, O. (O.) tristis. 58, Boletina dubia. 59, B. plana. 60, Leia fascipennis. 61, L. crucigera.



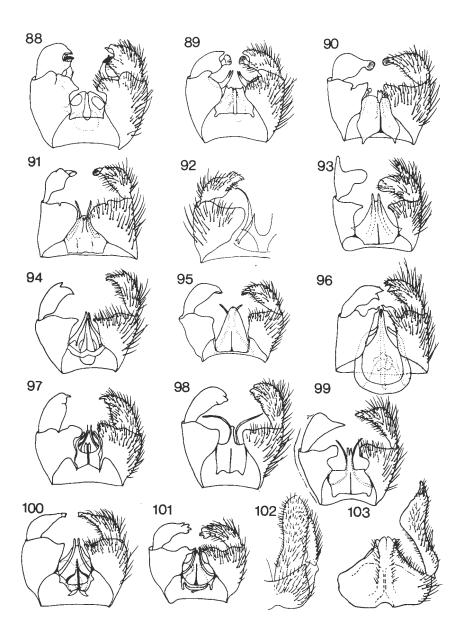
Figs. 62-69. Wings. 62, Bolitophila saundersii. 63, B. occlusa. 64, Diadocidia ferruginosa. 65, Ditomyia fasciata. 66, Symmerus annulatus. 67, Macrocera vittata. 68, Macrorrhyncha flava. 69, Cerotelion lineatus.



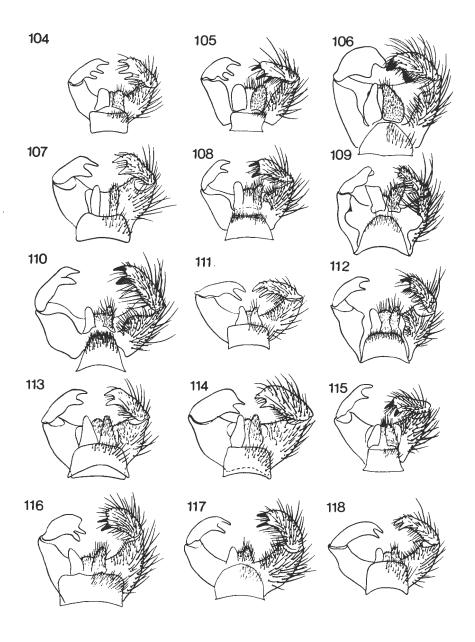
Figs. 70-77. Wings. 70, Platyura marginata. 71, Monocentrota lundstroemi. 72, Orfelia (Isoneuromyia) semirufa. 73, O. (Neoplatyura) modesta. 73a, tip of wing of O. (N.) flava. 74, O. (s.s.) nemoralis. 75, Mycomya marginata. 76, Leptomorphus walkeri. 77, Neuratelia nemoralis.



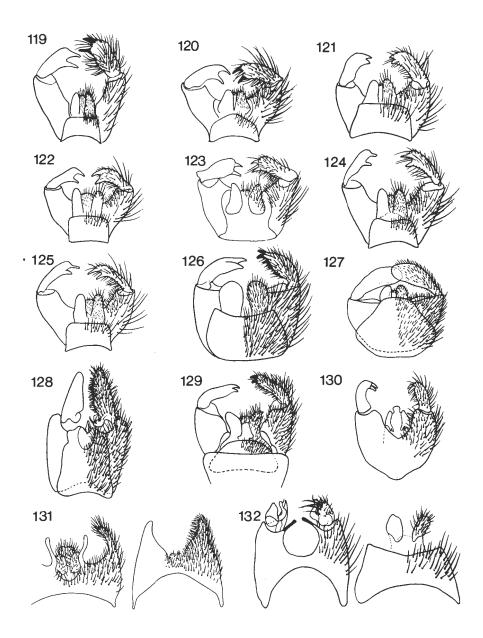
Figs. 78-87. Wings. 78, Polylepta guttiventris. 79, Coelosia tenella. 80, Palaeodocosia janickii. 81, Synapha vitripennis. 82, S. fasciata (anterior part of wing only). 83, Apolephthisa subincana. 84, Boletina trivittata. 85, Ectropesthoneura hirta. 86, Tetragoneura sylvatica. 87, Docosia sciarina.



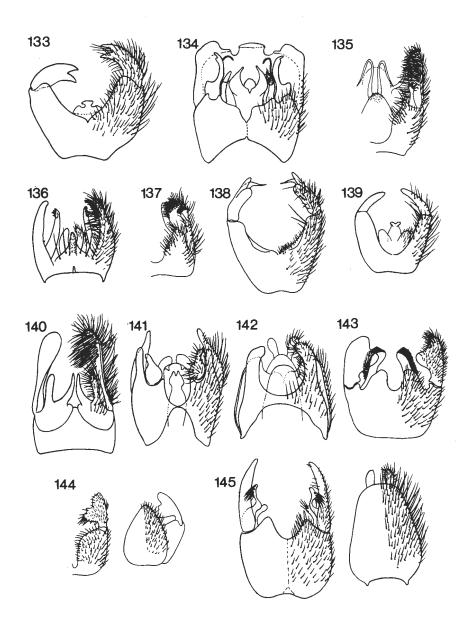
Figs. 88-103. σ genitalia. 88-103, Bolitophila spp, (dorsal, tergite 9 removed). 88, B. tenella. 89, B. basicornis. 90, B. cinerea. 91, B. saundersii. 92, B. spinigera. 93, B. occlusa. 94, B. dubia. 95, B. fumida. 96, B. maculipennis. 97, B. bimaculata. 98, B. glabratà. 99, B. pseudohybrida. 100, B. hybrida. 101, B. rossica. 102, Symmerus annulatus (dorsal). 103, Ditomyia fasciata (dorsal).



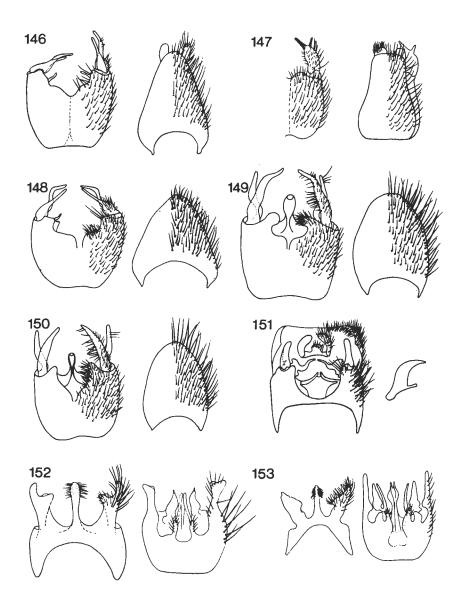
Figs. 104-118. σ genitalia, Macrocera spp. (dorsal). 104, M. pusilla, 105, M. anglica. 106, M. longibrachiata. 107, M. crassicornis. 108, M. fasciata. 109, M. tusca. 110, M. vittata. 111, M. lutea. 112, M. aterrima. 113, M. parva. 114, var. of M. parva. 115, M. centralis. 116, M. bipunctata. 117, M. fastuosa. 118, M. angulata.



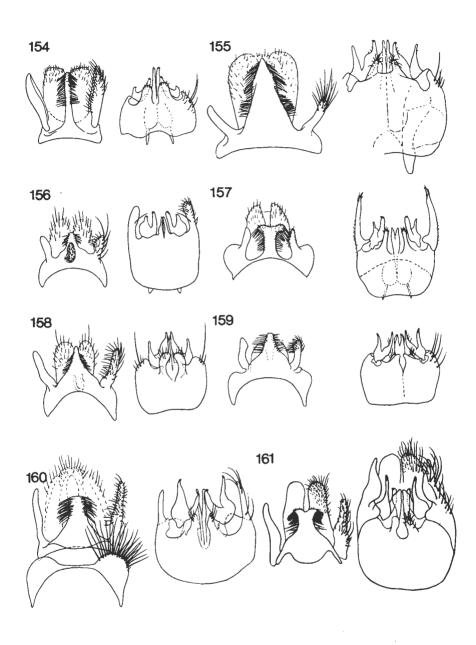
Figs. 119-132. σ genitalia. 119-125, Macrocera spp. (dorsal). 119, M. fascipennis. 120, M. maculata. 121, M. zetterstedti. 122, M. phalerata. 123, M. estonica. 124, M. stigma. 125, M. stigmoides. 126, Cerotelion lineatus (dorsal). 127, C. humeralis (dorsal). 128, Keroplatus testaceus (dorsal). 129, Platyura marginata (dorsal). 130, Monocentrota lundstroemi (ventral). 131, Asindulum nigrum (left: dorsal; right: ventral). 132, Macrorryyncha flava (left: dorsal; right: ventral).



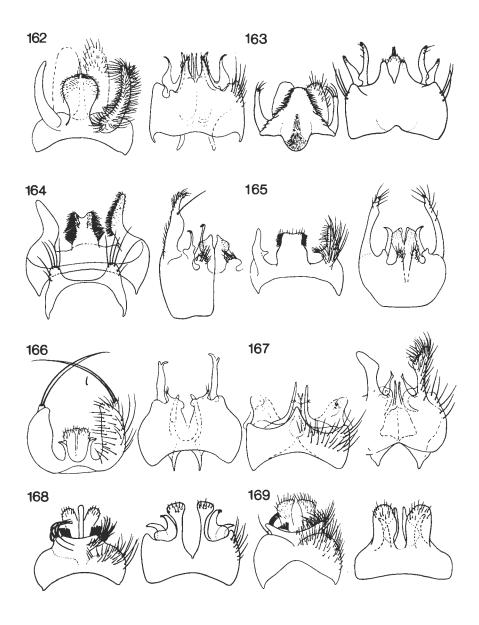
Figs. 133-145. σ genitalia, Orfelia spp. (ventral; figs. 144-145 with tergite 9). 133, O. (Isoneuromyia) semirufa. 134, O. (Neoplatyura) biumbrata. 135, O. (N.) flava. 136, O. (N.) nigricauda. 137, O. (N.) modesta. 138, O. (Pyratula) zonata. 139, O. (P.) perpusilla. 140, O. (Urytalpa) macrocera. 141, O. (U.) atriceps. 142, O. (U.) ochracea. 143, O. (Rutylapa) ruficornis. 144, O. (Orfelia) nemoralis. 145, O. (O.) nigricornis.



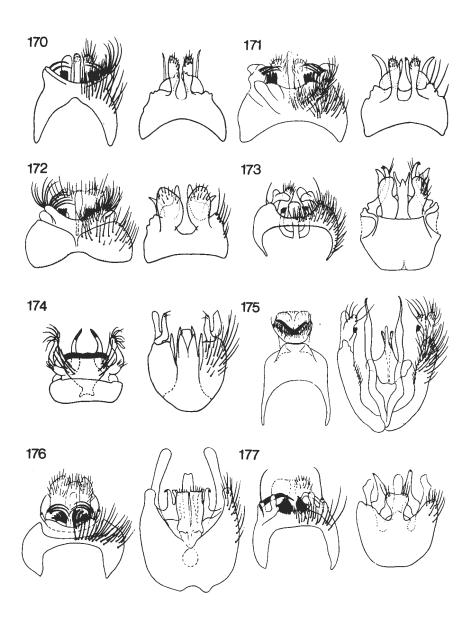
Figs. 146-153. © genitalia. 146-150, Orfelia (Orfelia) spp. (left: ventral; right: tergite 9). 146, O. (O.) tristis. 147, O. (O.) pallida. 148, O. (O.) fasciata. 149, O. (O.) unicolor. 150, O. (O.) discoloria. 151, Antlemon servulum (left: ventral; right: lateral view of dististyle). 152, Mycomya incisurata (left: dorsal; right: ventral). 153, M. flavicollis.



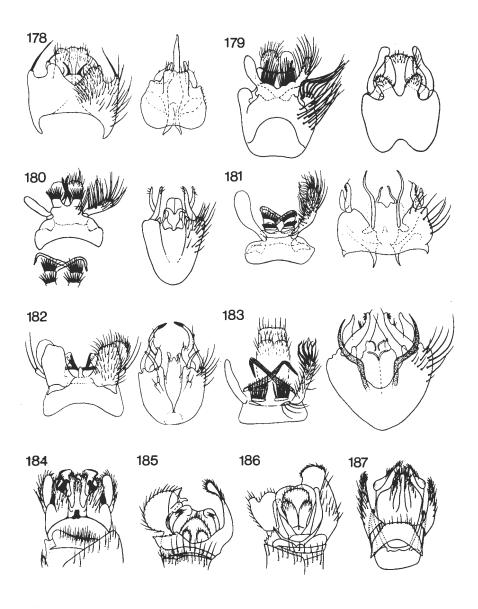
Figs. 154-161. σ genitalia, Mycomya spp. (left: dorsal; right: ventral). 154, M. exigua. 155, M. clavigera. 156, M. cinerascens. 157, M. britteni, 158, M. kingi. 159, M. trivittata. 160, M. marginata. 161, M. punctata.



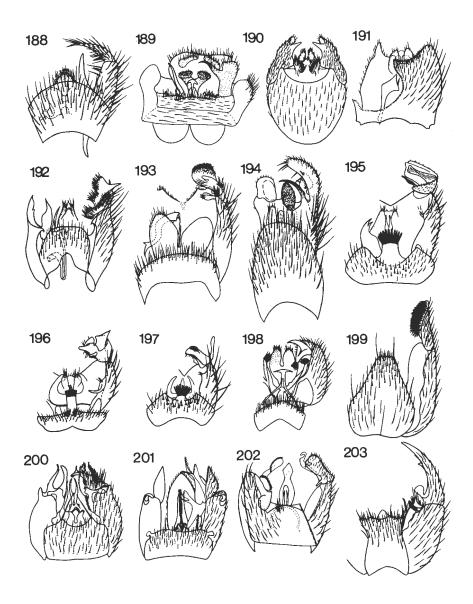
Figs. 162-169. genitalia, Mycomya spp. (left: dorsal; right: ventral). 162, M. hyalinata. 163, M. rosalba. 164, M. winnertzi. 165, M. wankowiczii. 166, M. tenuis. 167, M. duplicata. 168, M. fuscata. 169, M. ornata.



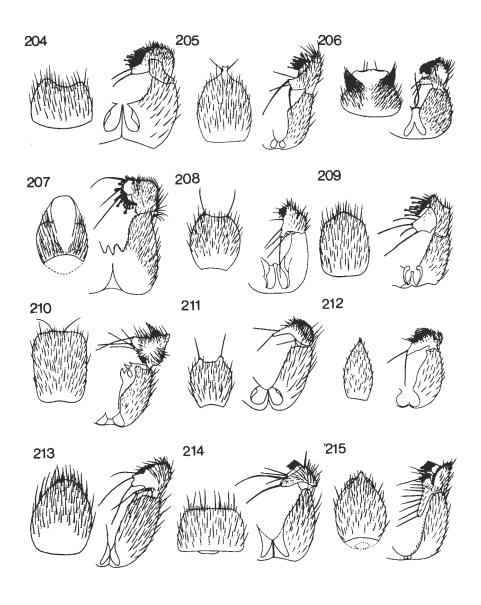
Figs. 170-177. or genitalia, Mycomya spp. (left: dorsal; right: ventral). 170, M. prominens. 171, M. tumida. 172, M. lambi. 173, M. melanoceras. 174, M. pectinifera. 175, M. circumdata. 176, M. digitifera. 177, M. parva.



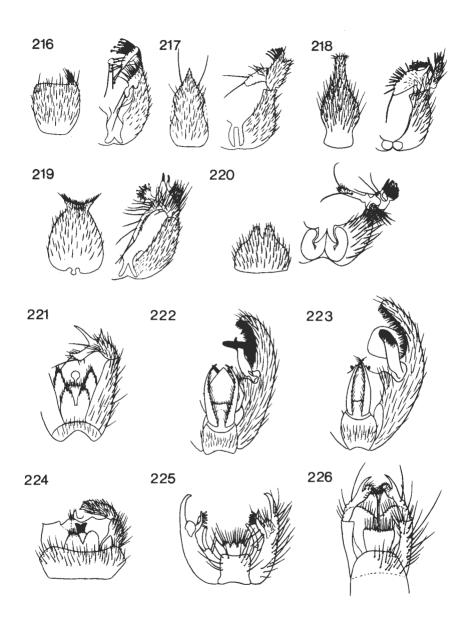
Figs. 178-187. σ genitalia. 178-183, Mycomya spp. (left: dorsal; right: ventral). 178, M. collini. 179, M. vittiventris. 180, M. flava. 181, M. maura. 182, M. fimbriata. 183, M. trilineata. 184-187, Neoempheria spp. (dorsal). 184, N. lineola. 185, N. bimaculata. 186, N. pictipennis. 187, N. winnertzi.



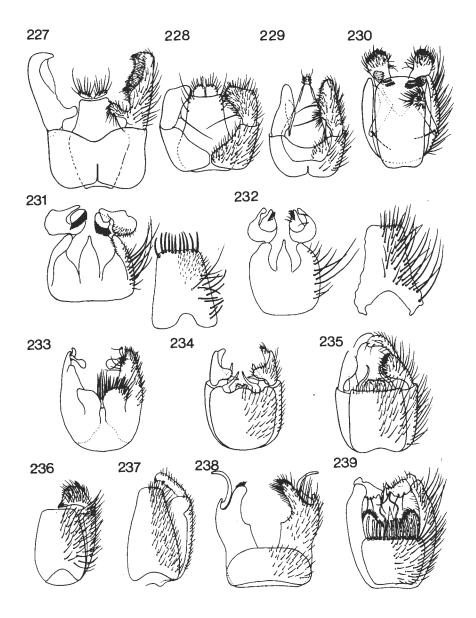
Figs. 188-203. O genitalia. 188, Eudicrana nigriceps (dorsal). 189, Allocotocera pulchella (ventral). 190, Leptomorphus walkeri (ventral). 191, Anaclileia dispar (dorsal, part of tergite 9 removed). 192, Neuratelia nemoralis (dorsal). 193, N. nigricornis (dorsal). 194, Polylepta guttiventris (dorsal). 195, Syntemna hungarica (dorsal). 196, S. nitidula (dorsal). 197, S. stylata (dorsal). 198, Azana anomala (dorsal). 199, Megalopelma nigroclavatum (dorsal). 200, Acnemia amoena (dorsal). 201, A. nitidicollis (dorsal). 202, A. longipes (dorsal). 203, Monoclona rufilatera (dorsal).



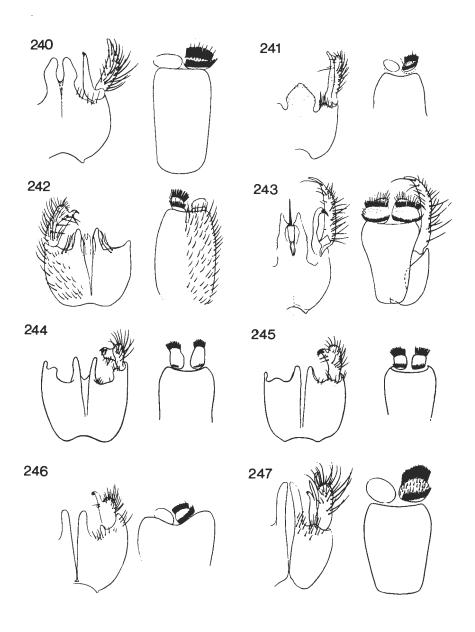
Figs. 204-215. σ genitalia, Sciophila spp. (left: tergite 9; right: ventral). 204, S. limbatella. 205, S. buxtoni. 206, S. rufa. 207, S. ochracea. 208, S. interrupta. 209, S. hirta. 210, S. lutea. 211, S. fridolini. 212, S. cliftoni. 213, S. plurisetosa. 214, S. quadriterga. 215, S. fenestella.



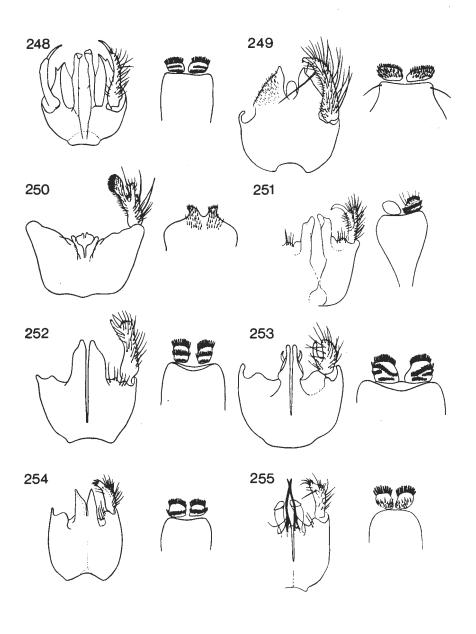
Figs. 216-226. σ genitalia. 216-220, Sciophila spp. (left: tergite 9; right: ventral). 216, S. varia. 217, S. nonnisilva. 218, S. adamsi. 219, S. nigronitida. 220, S. geniculata. 221-223, Phthinia spp. (dorsal). 221, P. winnertzi. 222, P. humilis. 223, var. of P. humilis. 224, Paratinia sciarina (dorsal). 225, Coelophthinia thoracica (dorsal). 226, Speolepta leptogaster (dorsal).



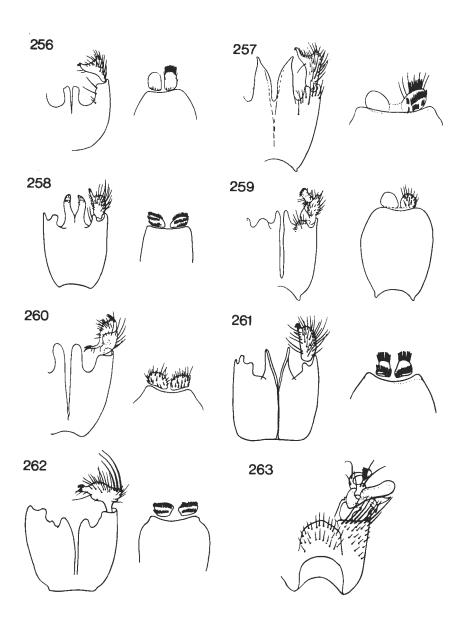
Figs. 227-239. σ genitalia (dorsal; figs. 231-232 with tergite 9 on right). 227, Coelosia flava, 228, C. sylvatica. 229, C. tenella. 230, Dziedzickia marginata. 231, Palaeodocosia janickii. 232, P. alpicola. 233, P. flava. 234, Gnoriste longirostris. 235, G. bilineata. 236, Synapha fasciata. 237, S. vitripennis. 238, Grzegorzekia collaris. 239, Apolephthisa subincana.



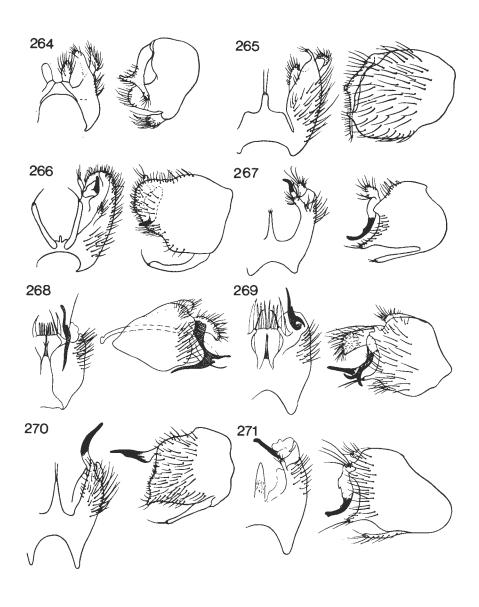
Figs. 240-247. σ genitalia, Boletina spp. (left: ventral; right: tergite 9). 240, B. plana. 241, B. dubia. 242, B. pallidula. 243, B. villosa. 244, B. nitida. 245, B. rejecta. 246, B. dispecta. 247, B. nasuta.



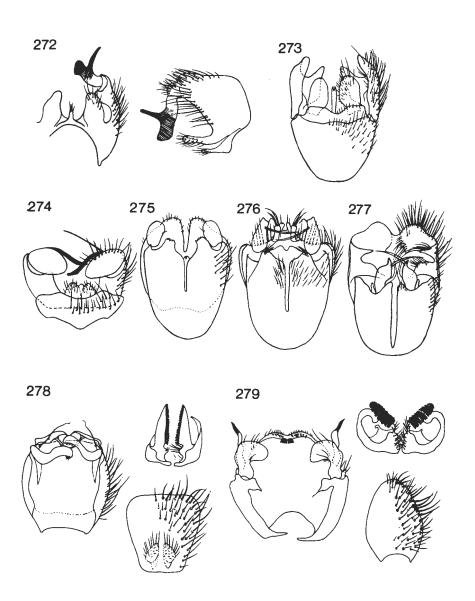
Figs 248-255. σ genitalia, Boletina spp. (left: ventral; right: tergite 9). 248, B. pectinunguis. 249, B. trivittata. 250, B. flaviventris. 251, B. basalis. 252, B. digitata. 253, B. groenlandica. 254, B. nigrofusca. 255, B. moravica.



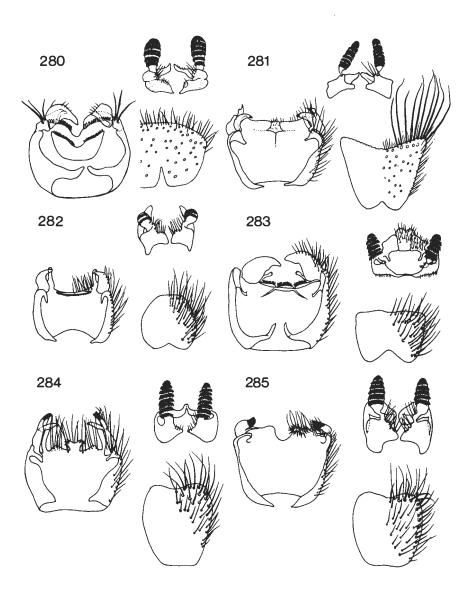
Figs. 256-263. σ genitalia. 256-262, Boletina spp. (left: ventral; right: tergite 9). 256, B. brevicornis. 257, B. gripha. 258, B. griphoides. 259, B. lundstroemi. 260, B. nigricans. 261, B. sciarina. 262, B. trispinosa. 263, Rondaniella dimidiata (dorsal).



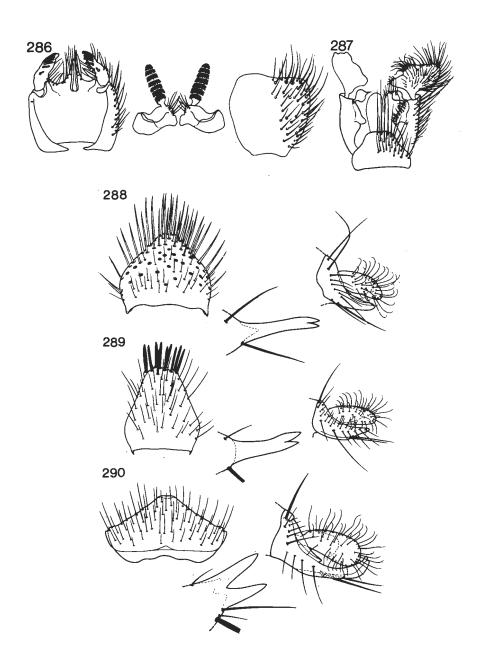
Figs. 264-271. σ genitalia, Leia spp. (left: ventral; right: lateral). 264, L. arsona. 265, L. bifasciata, 266, L. winthemi. 267, L. fascipennis. 268, L. crucigera, 269, L. subfasciata. 270, L. cylindrica. 271, L. bimaculata.



Figs. 272-279. σ genitalia. 272, Leia piffardi (left: ventral; right: lateral). 273, Megophthalmidia crassicornis (dorsal). 274, Tetragoneura sylvatica (dorsal). 275-277, Ectrepesthoneura spp. (ventral). 275, E. pubescens. 276, E. hirta. 277, E. colyeri. 278-279, Docosia spp. (left: dorsal view of genital capsule; above right: parameres; below right: tergite 9). 278, D. gilvipes. 279, D. fuscipes.



Figs. 280-285. Docosia spp., σ genitalia (left: dorsal view of genital capsule; above right: parameres; below right: tergite 9). 280, D. sp. indet. (Lastovka in prep.). 281, D. setosa. 282, D. fumosa. 283, D. carbonaria. 284, D. sciarina. 285, D. pallipes.



Figs. 286-290. Docosia moravica, σ genitalia (left: dorsal view of genital capsule; centre: parameres; right: tergite 9). 287, Manota unifurcata σ (dorsal). 288-290, Diadocidia spp. (left: σ tergite 9; centre: apex of σ dististyle; right: Θ ovipositor). 288, D. ferruginosa. 289, D. spinosula. 290, D. valida.

INDEX

Principal references are given first. Bold type indicates a page with an illustration. Synonyms are in italics.

Acnemia, 50; 12, 13, 23, 47, 80, 96. adamsi (Sciophila), 53; 23, 98. Adidocidia, 29. aestivalis (Orfelia), 22. affinis (Mycomya), 22. Agaricaceae, 11, 14. Agaricus, 11, 26, 27, 28. Aleuria, 10, 68. aliena (Ectrepesthoneura), 24. Allactoneura, 63, 69. Allocotocera, 47; 23, 46, 96. alpicola (Palaeodocosia), 57; 24, 99. Amanita, 12, 26, 27, 28, 68. Amanitaceae, 9, 12. amoena (Acnemia), 50; 23, 96. Anaclileia, 48; 23, 46, 96. Anaclinia, 23. analis Meigen (Boletina), 24. analis Winnertz (Sciophila), 52; 23. anglica (Macrocera), 31; 21, 88. angulata (Macrocera), 33; 21, 88. annulata (Mycomya), 22. annulatus (Symmerus), 28; 21, 75, 84, 87. anomala (Azana), 50; 23, 96. antica (Orfelia), 22. Antlemon, 39; 7, 17, 22, 30, 76, 91. Antlemonopsis, 39. Apolephthisa, 59; 12, 13, 24, 55, 86, 99. Arachnocampa, 8, 29. Armillaria, 11, 26, 52, 64, 68. arsona (Leia), 64; 24, 103. Ascomycetes, 10. Asindulum, 34; 7, 17, 22, 30, 76, 89. aterrima (Macrocera), 32; 21, 88. atrata (Platyura), 22. atriceps (Orfelia), 38; 22, 77, 82, 90. Auricularia, 13, 35, 43, 52, 68. Auriculariaceae, 13. Azana, 49; 23, 47, 96.

Baeopterogyna, 6, 46.
basalis Meigen (Boletina), 61; 24, 74, 101.
basalis Walker (Docosia), 25.
basicornis (Bolitophila), 26; 21, 87.
Basidiomycetes, 10.
bifasciata (Leia), 65; 13, 24, 103.
bilineata Winnertz (Leia), 24.
bilineata Zetterstedt (Gnoriste), 58; 24, 75, 99.
bimaculata Meigen (Leia), 65; 11, 12, 13, 24, 66, 81, 103.
bimaculata Roser (Neoempheria), 45; 23, 95.
bimaculata Zetterstedt (Bolitophila), 27; 21, 87.
bipunctata (Macrocera), 33; 21, 88.

birds, 7, 13, 66, 69. biumbrata (Orfelia), 37; 22, 90. Bjerkandera, 9, 12, 64,68. Boletaceae, 9, 11. Boletina, 59; 8, 13, 19, 24, 54, 55, 74, 78, 83, 86, 100-102. Boletus, 11, 26, 27, 28, 52, 55, 64, 68. Bolitiphila, 26; 6, 9, 11, 12, 17, 19, 21, 73, 74, 76, 78, 82, 83, 84, 87. Bolitophilinae, 26; 7, 19, 21, 25. borealis Lundström (Polylepta), 48. borealis Walker (Polylepta), 23. Brachypeza, 6. brevicornis (Boletina), 62: 24, 102. brevimanum (Antlemon), 39. britteni (Mycomya), 42; 22, 92. Bulgaria, 11, 52. buxtoni (Sciophila), 51; 9, 12, 23, 97. Calocera, 9, 13, 65. Caloceraceae, 13. Cantharellaceae, 13. Cantharellus, 13, 52. carbonaria (Docosia), 69; 13, 24, 106. caves, 7, 13, 56. Cecidomyiidae, 5. centralis (Macrocera), 33; 21, 88. Cerotelion, 35; 12, 13, 22, 30, 34, 74, 84, 89. Chondostereum, 12, 43. cinerascens (Mycomya), 42; 12, 22, 79, 92. cinerea (Bolitophila), 26; 9, 11, 12, 21, 73, 83, 87. cingulata (Mycomya), 22. circumdata (Mycomya), 44; 22, 94. Clastobasis, 63. clavigera (Mycomya), 42; 22, 92. cliftoni (Sciophila), 52; 23, 97. Cliopisa, 26; 21. Clitocybe, 11, 27, 28, 68. Coelophthinia, 55; 11, 13, 17, 24, 40, 54, 98. Coelosia, 55; 12, 24, 54, 86, 99. collaris (Grzegorzekia), 59; 24, 99.

collini (Mycomya), 44; 22, 95.

compressa (Tetragoneura), 24.

colyeri (Ectrepesthoneura), 67; 24, 104.

Coriolus, 9, 12, 39, 43, 50, 51, 52, 64, 67, 68.

Collybia, 11, 26, 52, 65.

concisa (Orfelia), 22.

Conjophoraceae, 13.

Coprinus, 11, 28, 68. Cordyla, 9, 17,

Cortinariaceae, 9, 11, 26.

Cortinarius, 11, 26, 28.

Coprinaceae, 11.

Corticiaceae, 13.

Corvus, 69. crassicornis Curtis (Megophthalmidia), 66; 24, 77, 80, 104. crassicornis Winnertz (Macrocera), 32; 21, 88. crucigera (Leia), 65; 24, 83, 103. cylindrica (Leia), 65; 24, 103.

Daedaleopsis, 9, 12, 51. Baedalia, 9, 12, 52. defecta (Acnemia), 23. Diadocidia, 29; 13, 14, 19, 20, 21, 28, 75, 78, 84, 106. Diadocidiinae, 28; 18, 19, 21, 25. digitata (Boletina), 61; 24, 101. digitifera (Mycomya), 44; 22, 94. dimidiata (Rondaniella), 64; 9, 24, 79, 102. discoloria (Orfelia), 39; 22, 91. disjuncta (Bolitophila), 21. dispar (Anaclileia), 48; 23, 96. dispecta (Boletina), 61; 24, 100. Ditomyia, 28; 9, 12, 21, 84, 87. Ditomyiinae, 28; 6, 18, 19, 21, 25. Docosia, 68; 7, 9, 10, 11, 12, 13, 24, 40, 41, 64, 86, 104-106. dorsalis Curtis (Macrocera), 22. dorsalis Staeger (Orfelia), 22. dubia Meigen (Boletina), 27; 21, 78, 83, 100. dubia Siebke (Bolitophila), 27; 21, 87. duplicata (Mycomya), 43; 13, 22, 93. Dynatosoma, 10. Dziedzickia, 57; 24, 40, 54, 55, 99.

Echinopodium, 41.
Ectrepesthoneura, 67; 12, 13, 19, 24, 40, 64, 68, 80, 86, 104.
elegans (Rondaniella), 24.
elongatus (Neuratelia), 23.
Empalia, 24.
Epicypta, 6.
estonica (Macrocera), 33; 21, 89.
Eudicrana, 47; 23, 40, 46, 96.
Exechia, 9.
exigua (Mycomya), 42; 22, 92.

fasciata Meigen (Ditomyia), 28; 9, 21, 84, 87.
fasciata Meigen (Macrocera), 32; 21, 88.
fasciata Meigen (Orfelia), 39; 22, 38, 91.
fasciata Meigen (Synapha), 58; 24, 86, 99.
fasciata Walker (Mycomya), 23.
fasciola (Leia), 65; 24, 66.
fascipennis Meigen (Leia), 65; 24, 83, 103.
fascipennis Staeger (Macrocera), 33; 21, 89.
fastuosa (Macrocera), 33; 21, 88.
fenestella (Sciophila), 53; 23, 97.
ferruginea (Mycomya), 23.
ferrugineus (Symmerus), 21.
ferruginosa (Diadocidia), 29; 21, 75, 78, 84, 106.

fimbriata (Mycomya), 45; 22, 95. finalis (Synapha), 24. Flammulina, 11, 26. flava Edwards (Palaeodocosia), 57; 24, 99. flava Grzegorzek (Polylepta), 49. flava Macquart (Orfelia), 37; 22, 79, 85, 90. flava Staeger (Coelosia), 56; 24, 99. flava Walker (Coelosia), 24. flava Winnertz (Mycomya), 44; 22, 95. flava Winnertz (Macrorrhyncha), 34; 22, 84, 89, flavicollis (Mycomya), 42; 22, 91. flavicornis (Leia), 24. flavipes (Orfelia), 22. flaviventris Landrock (Boletina), 24. flaviventris Strobl (Boletina), 61: 13. 24, 60, 101. Fomes, 9, 12, 51, 52, 64. fridolini (Sciophila), 52; 23, 97. fumida (Bolitophila), 27; 11, 21, 87. fumosa (Docosia), 69; 13, 24, 106. fusca (Bolitophila), 21. fuscata (Mycomya), 43; 22, 78, 93. fuscipes (Docosia), 68; 24, 104.

Ganoderma, 12, 43, 52. Ganodermataceae, 12. geniculata (Sciophila), 53; 23, 98. gilvipes (Docosia), 68; 7, 9, 10, 11, 12, 13, 24, 104. glabrata (Bolitophila), 27; 11, 21, 87. Glaphyroptera, 24. Gnoriste, 58; 13, 17, 24, 55, 75, 99. Gnoristini, 54; 23, 40. Gomphaceae, 13. Gomphus, 13, 52. gracilis (Ectrepesthoneura), 67. grandis (macrocera), 21. grass, 7, 13. Greenomyia, 62-63. Grifola, 12, 52, 64. gripha (Boletina), 62; 8, 24, 78, 102. griphoides (Boletiru), 62; 24, 102. groenlandica (Boletina), 61; 24, 101. Grzegorzekia, 58; 24, 55, 99. guttiventris (Polylepta), 48; 23, 79, 86, 96.

Hadroneura, 54, 57.
halterata (Monoclona), 23.
Hebeloma, 11, 26, 27, 65.
helvola (Megophthalmidia), 24.
Heterobasidion, 12, 64.
Heteroporus, 12, 52.
hirta Meigen (Sciophila), 52, 53; 11, 12, 13, 23, 51, 97.
hirta Winnertz (Ectrepesthoneura), 67; 12, 24, 80, 86, 104.

lutea Meigen (Macrocera), 32; 21, 88. Humariaceae, 10. humeralis (Cerotelion), 35; 22, 89. Lygistorrhininae, 15. humilis (Phthinia), 54; 23, 79, 98. hungarica (Syntemna), 49; 23, 96. Macrocera, 31; 6, 7, 8, 13, 17, 18, 19, hyalinata (Mycomya), 43; 22, 93. 21, 30, 74, 84, 88-89. hybrida (Bolitophila), 28; 9, 11, 12, 21, macrocera (Orfelia), 38; 22, 90. 76, 87. Macrocerini, 21. Macrorrhyncha, 34; 13, 17, 22, 30, 84, 89. Hydnaceae, 13. maculata (Macrocera), 33; 21, 89. Hydnum, 13, 52, 55. maculipennis (Bolitophila), 27; 11, 21, 87. Hymenochaetaceae, 12. Hymenoptera, 14. mammals, 7, 13, 66, 68. Hypholoma, 9, 11, 26, 27, 43, 68. Manota, 69; 10, 13, 25, 75, 106. Hypoxylon, 11, 52, 64. Manotinae, 69; 15, 17, 25, 63. Marasmius, 10, 11, 26. incisurata (Mycomya), 42; 22, 76, 91. marginata Dziedzicki (Dziedzickia), 57: 24, 99. inermis (Boletina), 24. Inonotus, 9, 12. marginata Dziedzicki (Mycomya), 23. interrupta (Sciophila), 52; 13, 23, 97. marginata Meigen (Mycomya), 42; 11, 12, Isoneuromyia, 36; 22, 30, 31, 85, 90. 13, 22, 85, 92. marginata Meigen (Platyura), 35; 22, 85, 89. maura (Mycomya), 45; 22, 95. janickii (Palaeodocosia), 57; 24, 86, 99. jenkinsoni (Megalopelma), 23. Megalopelma, 50; 23, 47, 96. Megophthalmidia, 66; 24, 63, 77, 80, 104. Keroplatinae, 29; 6, 7, 8, 17, 18, 19, 21, 25. melanoceras Edwards (Mycomya), 44: Keroplatini, 22. 22, 94. melanoceras Walker (Apolephthisa), 24. Keroplatus, 34; 12, 13, 17, 22, 30, 81, 89. kingi (Mycomya), 42; 22, 92. Meripilus, 12, 52. Meruliaceae, 12. Lacrymaria, 11, 26. Merulius, 12, 43. Lactarius, 12, 28, 52, 68. Messala, 21. lambi (Mycomya), 44; 22, 94. Metanepsiini, 40. Larix, 69. Meunieria, 24. Microtus, 68. Lasiosoma, 23. laticornis (Cerotelion), 22. modesta (Orfelia), 37; 22, 85, 90. Leia, 64; 11, 12, 13, 19, 24, 63, 80, 81, Monocentrota, 36; 22, 30, 85, 89. Monoclona, 51; 12, 13, 23, 47, 81, 96. 82, 83, 103-104. Leiini, 62; 24, 41, 55. moravica Landrock (Boletina), 62; 24, 101. Lenzites, 12, 52. moravica Landrock (Docosia), 69; 24, 106. Lepista, 68. Moriniola, 6. leptogaster (Speolepta), 56; 8, 23, 79, 98. moss, 7, 13. Leptomorphus, 47; 12, 23, 46, 85, 96. multicincta (Macrocera), 22. Mycetophila, 6, 7, 9, 10. Leptoporus, 9. Mycetophilinae, 6, 7, 10, 18, 25. limbatella (Sciophila), 51; 23, 97. lineatus (Cerotelion), 35; 12, 13, 22, mycetophiloides, 22, Mycomya, 41; 7, 11, 12, 13, 18, 19, 20, 74, 84, 89. 22, 76, 78, 79, 85, 91-95. lineola (Neoempheria), 45; 23, 95. liverwort, 13. Mycomyini, 41; 22, 40. Loewiella, 23. Myxomycetes, 10, 69. longibrachiata (Macrocera), 31; 21, 88. longipes (Acnemia), 50; 23, 96. nasuta (Boletina), 61; 24, 100. longirostris (Gnoriste), 58; 24, 99. nemoralis Meigen (Neuratelia), 48; 23, lucorum (Mycomya), 22. 80, 85, 96. lundbecki (Boletina), 24. nemoralis Meigen (Orfelia), 38; 22, 83, lunstroemi Edwards (Monocentrota), 85, 90. 36; 22, 85, 89. Neoclastobasis, 62, 63. lunstroemi Landrock (Boletina), 62; Neoempheria, 45; 11, 12, 20, 23, 41, 95, Neoplatyura, 36; 22, 30, 31, 79, 85, 90. 24, 102. lutea Macquart (Sciophila), 52, 53; 9, 11, Neuratelia, 48; 13, 23, 46, 80, 85, 96. 12, 13, 23, 51, 97. nigra Macquart (Sciophila), 23.

nigra Walker (Mycomya), 22. nigricans (Boletina), 62; 24, 102. nigricauda (Orfelia), 37; 22, 90. nigriceps Lundström (Eudicrana) 47; 23, 96. nigriceps Walker (Orfelia), 22. nigricornis Dziedzicki (Mycomya), 23. nigricornis Edwards (Neuratelia). 48: 23, 96. nigricornis Fabricius (Orfelia), 39; 22, 38, 90, nigricornis Lundström (Mycomya), 22. nigroclavatum (Megalopelma), 50; 23, 96. nigrofusca (Boletina), 62; 24, 101. nigronitida (Sciophila), 53; 23, 98. nigrum (Asindulum), 34; 22, 76, 89. nitida (Boletina), 60; 24, 100. nitidicollis (Acnemia) 50; 12, 23, 80, 96. nitidula (Ectrepesthoneura), 49: 23, 82, 96. nonnisilva (Sciophila), 53; 23, 98. Novakia, 62, 63.

occlusa (Bolitophila), 27; 12, 21, 73, 84, 87. ochracea Meigen (Orfelia), 38; 22, 90. ochracea Walker (Sciophila), 52; 12, 23, 97. octomaculata (Leia), 24. Orfelia, 36, 38; 12, 13, 19, 22, 30, 31, 39, 77, 79, 80, 82, 83, 85, 90-91. ornata (Mycomya), 43; 23, 93.

Palaeodocosia, 57; 24, 40, 55, 86, 99. pallida (Orfelia), 39; 22, 91. pallidula (Boletina), 60; 24, 100. pallipes (Docosia), 69; 25, 106. paradoxa (Synapha), 24. Paraneurotelia, 23. Paratinia, 54; 23, 40, 46, 98. parva Dziedzicki (Mycomya), 44; 23, 94. parva Lundström (Macrocera), 32; 21, 74, 88. Paxillaceae, 11. Paxillus, 9, 11, 27, 28. pectinifera Edwards (Mycomya), 44; 23, 94. pectinifera Edwards (Orfelia), 22. pectinunguis (Boletina), 26; 24, 101. Peniophora, 13, 29. perpusilla (Orfelia), 37; 22, 90. Peziza, 11, 68. Pezizaceae, 11. phalerata (Macrocera), 33; 22, 89. Phallaceae, 13. Phallus, 13, 43. Phellinus, 12, 43, 52. Phlebia, 12, 43, 59. Pholiota, 9, 11, 26, 28, 52, 53, 64. Phronia, 7 Phthinia, 53; 11, 13, 17, 19, 23, 46, 79, 98. Pica, 66. pictipennis (Neoempheria), 45; 23, 95. piffardi (Leia) 66; 13, 24, 65, 80, 104.

Piptoporus, 12, 68. plana (Boletina), 60; 24, 55, 83, 100. Planarivora, 8, 29. Platyura, 35; 13, 22, 30, 36, 85, 89. Pleurotaceae, 11. Pleurotus, 11, 43, 52, 64. plurisetosa (Sciophila), 52; 23, 97. Polylepta, 48; 23, 46, 79, 86, 96. Polyporaceae, 9, 12, 28, 35, 52. Polyporus, 9, 12, 27, 51, 64, 68. Polystictus, 9, 12, 68. Poria, 12, 43, 44, 47, 51, 66. prominens (Mycomya), 43; 11, 12, 23, 94. propleuralis (Macrocera), 32; 22. Prunella, 69. pseudohybrida (Bolitophila), 27; 11, 12, 21, 82, 87. Pseudotrametes, 9, 12, 51, 52. pubescens Walker (Docosia), 25, pubescens Zetterstedt (Ectrepesthoneura), 67; 24, 104. pulchella (Allocotocera), 47; 23, 96. punctata (Mycomya), 43; 23, 92. pusilla (Macrocera), 31; 22, 88. Pyratula, 37; 22, 30, 31, 39, 80, 90. quadriterga (Sciophila), 52; 23, 97. referta (Ectrepesthoneura), 67. rejecta (Boletina), 60; 24, 100. reuteri (Boletina), 24. Ripartites, 11, 27. Rondaniella, 64; 9, 11, 12, 19, 24, 41, 55, 63, 79, 102. rosalba (Sciophila), 43; 23, 93. rossica (Bolitophila), 28; 11, 21, 87. rostrata (Macrorrhyncha), 22. Rozites, 11, 28. rufa (Sciophila), 51; 12, 23, 77, 97. ruficollis (Mycomya), 22. ruficornis (Orfelia), 38; 22, 80, 90. rufilatera (Monoclona), 51; 23, 81, 96. rufipes (Platyura), 22. Russula, 9, 12, 27, 52, 64, 65, 68. Russulaceae, 9, 12, Rutylapa, 38; 22, 30, 31, 80, 90. Rymosia, 14. saundersii (Bolitophila), 27; 9, 11, 12, 21, 74, 78, 83, 84, 87. Scatopsidae, 5. scatopsoides (Azana), 23. Sciaridae, 5, 19, 67. sciarina Meigen (Docosia), 69; 25, 86. 106. sciarina Mik (Paratinia), 54; 23, 98. sciarina Staeger (Boletina), 62; 24, 102. Sciophila, 51; 9, 11, 12, 13, 33, 46, 47, 50, 77, 97-98. Sciophilinae, 40; 6, 18, 19, 22, 25.

Sciophilini, 45; 20, 23, 40, 65.

Sciurus, 66. tristis (Orfelia), 39; 22, 83, 91. Scleroderma, 13, 68. trivittata Meigen (Boletina), 61; 24, Sclerodermataceae, 13. 78, 86, 101. trivittata Zetterstedt (Macrocera), 42; 23, 92. Sebacina, 13, 43. semirufa (Orfelia), 36; 22, 85, 90. tumida (Mycomya), 44; 23, 94. Serpula, 13, 35. Turdus, 66, 69. servulum (Antlemon), 40; 22, 76, 91. tusca (macrocera), 32; 22, 88. setosa (Docosia), 68; 25, 106. Tyromyces, 12, 27. sharpi (Sciophila), 23. silvatica Dziedzicki (Boletina), 24, 59. undulata (Polylepta), 23. silvatica Landrock (Coelosia), 56; 24, 99. unicolor Staeger (Orfelia), 39; 22, 91. Sparassidaceae, 12. unicolor Walker (Orfelia), 22. Sparassis, 12, 43, 64. unicornuta (Monoclona), 23. Speolepta, 56; 6, 8, 13, 17, 20, 23, 54, unifurcata (Manota), 69; 25, 106. 55**, 79, 98.** Urytalpa, 37; 22, 31, 77, 82, 90. spinigera (Bolitophila), 27; 21, 87. Ustulina, 11, 52, 68. spinosula (Diadocidia), 29; 21, 78, 106. splendida (Polylepta), 23, 48. valida Mik (Diadocidia), 29: 21, 106. Stereaceae, 12. valida Walker (Megophthalmidia), 24. Stereum, 12, 35, 42, 43, 52, 56, 64. valida Winnertz (Docosia), 25. varia (Sciophila), 53; 23, 98. stigma (Macrocera), 33; 22, 89. stigmoides (Macrocera), 33; 22, 89. variegata (Rondaniella), 24. strobli (Leia), 24. Verticillium, 14. Strophariaceae, 11, 26. villosa (Boletina), 60; 24, 100. stylata (Syntemna), 49; 23, 81, 96. Viridivora, 41. vitripennis Meigen (Synapha), 58; 24, subfasciata (Leia), 65; 24, 82, 103. subincana (Apolephthisa), 59; 24, 86, 99. 86, 99. sylvatica (Tetragoneura), 66; 24, 75, 80, vitripennis Walker (Orfelia), 22. 86, 104. vittata Coquillett (Palaeodocosia), 57. Symmerus, 28; 21, 75, 84, 87. vittata Meigen (Macrocera), 32; 22, 84, 88. Synapha, 58; 24, 55, 86, 99. vittata Walker (Symmerus), 21. Syndocosia, 41. vittiventris (Mycomya), 44; 23, 95. Syntemna, 49; 13, 18, 23, 40, 46, 81, 82, 96. walkeri (Leptomorphus), 47; 23, 85, 96. wankowiczii (Mycomya), 43; 11, 12, 13, 23, tenella Winnertz (Bolitophila), 26; 11, 12, 42, 94. winnertzi Dziedzicki (Mycomya), 43; 12, tenella Zetterstedt (Coelosia), 56; 24, 23, 42, 93. winnertzi Edwards (Neoempheria), 45; 23. 86, 99. 95. tenuis (Mycomya), 43; 23, 93. terminalis (Rondaniella), 24. winnertzi Mik (Phthinia), 53; 11, testaceus (Keroplatus), 35; 22, 81, 89. 23, 98. Tetragoneura, 66; 12, 24, 62, 63, 64, 67, winthemi (Leia), 65; 24, 103. 68, 75, 80, 86, 104. wood, 7, 13. Thelophora, 12, 44. wrzesniowskii (Mycomya), 44; 12, 23. Thelophoraceae, 12. thoracica (Coelophthinia), 55; 11, 24, 98. Xeromphalina, 11, 27. Trametes, 9. Xylariaceae, 11. Xylodon, 59. Tremellaceae, 13. triangulata (Bolitophila), 21. zetterstedti (Macrocera), 33; 22, 89. Tricholoma, 11, 27, 28, 68. Tricholomataceae, 9, 11, 27. Zingiber, 64. Trichonta, 9. zonata Stephens (Symmerus), 21. zonata Zetterstedt (Orfelia), 37; 22, 80, 90. trilineata (Mycomya), 45; 23, 95. trimaculata (Leia), 24. zonata Zetterstedt (Polylepta), 48. trispinosa (Boletina), 62; 24, 102.

