

MDPI

Review

# Medical Application of Substances Derived from Non-Pathogenic Fungi Aspergillus oryzae and A. luchuensis-Containing Koji

Hiroshi Kitagaki 📵

Graduate School of Advanced Health Sciences, Saga University, Saga 840-8502, Japan; ktgkhrs@cc.saga-u.ac.jp

Abstract: Although most fungi cause pathogenicity toward human beings, dynasties of the East Asian region have domesticated and utilized specific fungi for medical applications. The Japanese dynasty and nation have domesticated and utilized *koji* fermented with non-pathogenic fungus *Aspergillus oryzae* for more than 1300 years. Recent research has elucidated that *koji* contains medicinal substances such as Taka-diastase, acid protease, *koji* glycosylceramide, *kojic* acid, oligosaccharides, ethyl-α-d-glucoside, ferulic acid, ergothioneine, pyroglutamyl leucine, pyranonigrin A, resistant proteins, deferriferrichrysin, polyamines, *Bifidobacterium*-stimulating peptides, angiotensin I-converting enzyme inhibitor peptides, 14-dehydroergosterol, beta-glucan, biotin, and citric acid. This review introduces potential medical applications of such medicinal substances to hyperlipidemia, diabetes, hypertension, cardiovascular and cognitive diseases, chronic inflammation, epidermal permeability barrier disruption, coronavirus disease 2019 (COVID-19), and anti-cancer therapy.

Keywords: medical application; koji; fermentation; non-pathogenic fungus; medicinal substances



Citation: Kitagaki, H. Medical Application of Substances Derived from Non-Pathogenic Fungi Aspergillus oryzae and A. luchuensis-Containing Koji. J. Fungi 2021, 7, 243. https://doi.org/ 10.3390/jof7040243

Academic Editors: Katsuhiko Kitamoto and Yujiro Higuchi

Received: 27 February 2021 Accepted: 19 March 2021 Published: 24 March 2021

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

### 1. Introduction

Fungi in general cause many infectious pathogenicities in human beings, including *Aspergillus*, *Candida*, *Cryptococcus*, and *Mucor* species [1]. However, several fungi have been utilized as medicines across the world since the prehistoric ages. Fungi are eukaryotic microbes that have complex biosynthesis pathways and various morphologies. They have unique biosynthesis pathways and thus produce unique substances that are not contained in mammals, plants, or prokaryotic bacteria [2].

Within fungi, mushrooms have attracted attention in terms of their medical potential since the prehistoric ages [3], possibly because of their visibility without microscopes. For example, hoelen, which is a *Wolfiporia cocos* mushroom proliferating on the roots of pine trees, has been used in East Asian countries, including China and Japan, as a medicine for more than 1800 years [4]. Moreover, a *Ganoderma lucidum* mushroom is described in Shennong Ben Cao Jing, which was written in 25–220 A.D. in China, as being able to increase the lifespan [5].

The beta-glucan contained in *Schizophyllum commune*, Schizophyllan, and *Lentinula edodes*, Lentinan, has been registered and utilized as immunoactivating medicines to treat cervical, stomach, and colon cancer [6]. However, medical applications of fungi which do not form mushrooms had not been described until the modern age, possibly because they are invisible without microscopes and microscopes only prevailed across the world in the 18–19th centuries.

The use of a red fungus *Monascus* in China is one of these examples [7]. The health benefits of red rice *koji* has been recognized in China for more than approximately 700 years. Its medical use was first described in Compendium of Materia Medica, which was written in 1498 A.D., indicating that it clears the blood and supports digestion of foods in the intestine. Dr. Endo elucidated, in 1979, that monacolin K produced by *Monascus ruber* has

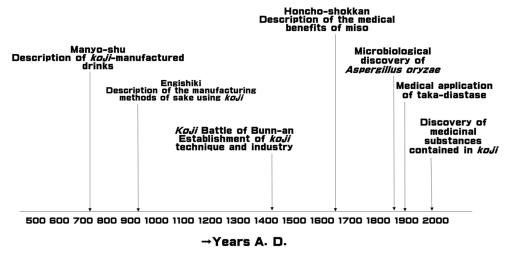
J. Fungi **2021**, 7, 243 2 of 14

a hypocholesterolemic effect [8]. Moreover, it also contains medicinal substances such as monascin [9], ankaflavin [10], and ergosterol [11].

Moreover, the fungus *Penicillium camemberti* or *roqueforti*, belonging to ascomycetes, has been used to manufacture blue cheese in France for centuries. In 1929, Sir Fleming found that a fungus, *Penicillium rubens*, kills pathogenic bacteria [12]. Later, penicillin, which was identified as a substance that kills pathogenic bacteria, and related antibiotics were utilized to eradicate bacterial infectious diseases across the world.

Tempeh, which is an Indonesian traditional fermented food lasting for more than 400 years, and produced by fermenting soybeans with *Rhizopus oligosporus* or *R. oryzae*, has several health benefits such as improvement of cognitive function, gut immunity, intestinal microbial flora, hyperlipidemia, and anemia [13].

Since the earliest historical age of Japan, Japanese people have isolated fungus Aspergillus oryzae, belonging to ascomycetes, and domesticated and utilized it to produce koji as a catalyst of starch in rice. During the process that lasted for 1300 years, non-pathogenic strains were genetically selected by specialized technicians of koji. Indeed, miso is described in Taihoritsuryo, which is one of the earliest legal codes announced by the emperor Monmu in 701, A.D. (Figure 1). The koji drink amazake is described in Manyoshu, which is a collection of poems of the imperial court written in 731–733 A.D. (Figure 1). A battle between the specialized technicians and the Muromachi government is described in 1444, A.D. (Koji Battle of Bunn-an). Therefore, it is evident from the literature that specialized technicians have genetically selected and maintained the strains, and A. oryzae strain presently used to produce *koji* is considered to be non-pathogenic. Ryukyu people in Okinawa Islands also domesticated Aspergillus luchuensis. Therefore, koji that uses these fungi are generally regarded as safe (GRAS) by the FDA and is used in most Japanese fermented foods and drinks, including miso, soy sauce, sake, amazake, vinegar, kurozu, fermented barley extract, and shochu as saccharifyer of starch contained in steamed rice, barley, or soybeans [14]. Japanese people have long recognized the health benefits of these fermented foods. Indeed, Honcho-shokkan describing the medical benefits of foods in Japan written in 1695, A.D. describes that dietary intake of koji supports the digestion of foods, decreases occlusion, and smoothens the blood flow. Yojo-kun (regimen sanitatis for longevity of Japan), written by the Japanese scholar Ekken Kaibara in 1712, A.D. mentions that the dietary intake of miso is gentle for the body and compensates for the functions of the gastrointestinal tract. Furthermore, the average life span of Japanese people is one of the longest in the world, and Japanese cuisines contain many fermented foods, most of which contain koji [15]. It could thus be hypothesized that the substances contained in Japanese fermented foods and/or koji are beneficial for health. However, medical knowledge on this non-pathogenic fungus, A. oryzae, is possibly limited because of its modest medicinal effect and is still under study at present.



**Figure 1.** History of technologies and research on *koji* in Japan.

J. Fungi **2021**, 7, 243 3 of 14

Recent studies in Japan have elucidated that *koji* contains many substances that have potential medical applications, and this review summarizes such studies. This novel knowledge will pave the way for medical applications of non-pathogenic fungus *A. oryzae* in the future.

#### 2. Taka-Diastase and Acid Protease

A Japanese scholar, Jokichi Takamine, applied a patent describing a starch-degrading enzyme preparation derived from *koji* and designated it as Taka-diastase in 1911 [16]. It has been widely used as a starch digestant to treat stomach upset, stomachache, heartburn, and overeating throughout the world.

Recently, the dietary intake of the acid proteases contained in *Aspergillus oryzae* or *A. luchuensis* was shown to increase intestinal *Bifidobacterium* and *Lactobacillus* spp. [17,18]. This could be attributed to the remnant of enzymatic activity when they pass through the stomach and the small intestine. Therefore, acid protease could be a new generation of prebiotics.

# 3. Koji Glycosylceramide

Glycosylceramide is a sphingolipid consisting of a sphingoid base, fatty acid, and a sugar moiety (Figure 2). It is contained in various species, including plants and fungi. Sphingolipids have versatile biological functions, such as signal transduction [19], enhancement of the immune system [20], and raft formation [21]. Their structures have great variations, depending on the structure and type of the sphingoid base, fatty acid, and sugar moiety. The characteristic structure of A. oryzae is N-2'-hydroxyoctadecanoyl-L-O-β-D-glucopyranosyl-9-methyl-4,8-sphingadienine, N-2'-hydroxyoctadecanoyl-L-O-β-Dglucopyranosyl-4,8-sphingadienine, N-2'-hydroxy-3'-octadecenoyl-L-O-β-D-glucopyranosyl-9-methyl-4,8-sphingadienine, N-2'-hydroxyhexadecanoyl-L-O-β-D-glucopyranosyl-4,8sphingadienine, N-2'-hydroxyicosanoyl-L-O-β-D-glucopyranosyl-4,8-sphingadienine, N-2'-hydroxyicosanoyl-L-O-β-D-glucopyranosyl-4,8-sphingadienine, and the corresponding molecules in which sugar moiety is substituted for galactose instead of glucose [22–26]. It is contained in koji and Japanese fermented foods at 1–4 mg/serving [27]. Most of the glycosylceramide in koji is considered not to be degraded by enzymes in the small intestine, and passes through to the large intestine [28]. The dietary intake of the glycosylceramide purified from koji at a concentration of 1% w/w of diet in mice increases intestinal Blautia coccoides, Bacteroides sartorii, and Hathewaya histolytica in mice. Since B. coccoides is considered to be a beneficial microbe, koji glycosylceramide could be considered as a new type of prebiotics. Also, feeding of glycosylceramide at a concentration of 1% w/w of diet to obese mice increases bile acids and lowers liver cholesterol in obese mice [29]. Since liver cholesterol is one of the causes of hyperlipidemia, koji glycosylceramide could potentially be medically utilized to treat hyperlipidemia.

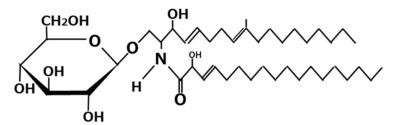


Figure 2. Chemical structure of glycosylceramide.

Furthermore, glycosylceramide purified from *koji* increases the expression of the genes involved in tight junctions and ceramide delivery in normal human epidermal keratinocytes [30]. The dietary intake of the glucosylceramide derived from konjac extract improves transepidermal water loss (TEWL) in hairless mice with sodium dodecyl sulfate (SDS)-induced skin roughness [31]. Since the glycosylceramide contained in *koji* contains

J. Fungi **2021**, 7, 243 4 of 14

the same species of glucosylceramide contained in konjac [23], the dietary intake of *koji* might improve TEWL. Therefore, the dietary intake of *koji* or the topical administration of *koji* to the skin might be beneficial for skin disorders.

## 4. Kojic Acid

A. oryzae produces kojic acid (Figure 3), which functions as a competitive and reversible inhibitor of animal and plant polyphenol oxidases, xanthine oxidase, and some amino acid oxidases, and is used as an food additive for preventing enzymatic browning or in cosmetic preparations as a skin-lightening agent [32]. Therefore, *koji* contains kojic acid at the concentration of 0–9.5 mg/g, and the topical administration of kojic acid at 50  $\mu$ g/mL inhibits browning of the skin [33]. Although some researchers have reported the occurrence of thyroid adenomas when continuously administered, it is generally recognized that the consumption of kojic acid at levels normally found in food is not a concern for safety.

Figure 3. Chemical structure of Kojic acid.

## 5. Oligosaccharides

Rice, barley, wheat, and soy contain various oligosaccharides consisting of glucose, fructose, xylulose, galactose, maltose, and arabinose [34]. For example, starch consists of  $\alpha$ -1,4-linked glucose polymers, with  $\alpha$ -1,6-linked branches [35]. Xylan is a polymer consisting of  $\beta$ -1,4-linked xylulose [36]. The genome of A. oryzae contains abundant genes of glycolytic enzymes [37]. For example,  $\alpha$ -amylase degrades starch and generates various lengths of  $\alpha$ -1,4-linked glucose polymers randomly with  $\alpha$ -1,6-linked branches [38]. These glucose oligomers are degraded by pancreatic amylases to glucose, and the resultant glucose is absorbed in the small intestine [39]. However, since pancreatic enzymes do not degrade oligosaccharides which contain unique structures derived from starch or includes xylulose or arabinose, such oligosaccharides are not degraded in the small intestine, instead passing through to the large intestine and, as a result, are assimilated by intestinal microbes. Indeed, koji manufactured from rice and sake manufactured using koji contain various oligosaccharides including isomalto-pligosaccharide containing  $\alpha$ -1,2-bond [40] and glucooligosaccharide with adjacent  $\alpha$ -1,6 branches at the non-reducing end derived from starch (Figure 4) [41] and shoyu manufactured from rice koji and soy contains oligosaccharides consisting of galactose, arabinose, and galacturonic acid [42]. It has been reported that the dietary ingestion of oligosaccharides increases intestinal Bifidobacterium [43]. Moreover, the dietary ingestion of oligosaccharide is beneficial for ulcerative colitis and minimal hepatic encephalopathy [44,45]. Furthermore, Shoyu polysaccharide produced by koji enzymes from soy alleviates perennial allergic rhinitis in humans [43] and promotes iron absorption in rats and humans [46].

J. Fungi **2021**, 7, 243 5 of 14

Figure 4. Chemical structure of resistant gluco-oligosaccaride.

Koji-fermented barley also contains oligosaccharides consisting of glucose, xylose, and arabinose that stimulate the growth of lactic acid bacteria and Bifidobacterium [47]. Since Bifidobacterium infantis ameliorates intestinal mucositis in rats [48] and Bifidobacterium adolescentis extract has an anti-proliferative effect on human colon cancer cell lines [49], it is considered that the dietary ingestion of koji-fermented foods and drinks (especially those manufactured from raw rice, barley, wheat, and soy) has potential medical applications to treat such patients.

### 6. Ethyl- $\alpha$ -D-Glucoside

Ethyl- $\alpha$ -D-glucoside (Figure 5), as well as  $\alpha$ -glyceryl glucoside, ethyl- $\alpha$ -maltoside, and ethyl- $\alpha$ -isomaltoside, are formed from ethanol or glycerol and glucose, maltose, or isomaltose by the action of the transglycosidase produced by *koji* [50,51]. For example, the concentration of ethyl- $\alpha$ -d-glucoside is reported to be 0.33% w/w [50]. The dietary intake of the ethyl- $\alpha$ -D-glucoside (1% w/w) contained in sake and sake lees, the remnant of sake production, improves epidermal permeability barrier disruption by UVB irradiation in hairless mice [52]. It also upregulates collagen I and fibroblast growth factor I and VII in cultured human dermal fibroblasts [53]. Moreover, the topical application of ethyl- $\alpha$ -D-glucoside increases the intercellular lipid content, accelerates the differentiation of corneocytes, and reduces the thickness of the epidermis, thus improving the functions of the stratum corneum in murine epidermis [54]. Therefore, the dietary intake of sake or sake lees leads to the intake of ethyl- $\alpha$ -D-glucoside, thus resulting in improvement of epidermal permeability barrier disruption.

**Figure 5.** Chemical structure of ethyl- $\alpha$ -D-glucoside.

# 7. Ferulic Acid

Ferulic acid is ester-bound to carbohydrates (Figure 6) in cereals [55], and is formed by koji enzymes during sake brewing [56]. Ferulic acid is contained in sake at a concentration of 1–6 mg/L [57]. The dietary intake of ferulic acid lowers the lipid levels in hyperlipidemic diabetic rats [58]. The oral administration of 30 mg/kg/day ferulic acid or ethyl ferulate provides protective effects toward retinal degeneration [59]. Furthermore, the dietary intake of ferulic acid is protective toward the toxicity of the  $\beta$ -amyloid peptide, which causes Alzheimer's disease [60]. Therefore, the dietary intake of koji leads to the intake

J. Fungi **2021**, 7, 243 6 of 14

of ferulic acids, thus resulting in amelioration of dyslipidemia, retinal degeneration, and Alzheimer's disease.

Figure 6. Chemical structure of ferulic acid.

## 8. Ergothioneine

Ergothioneine is a derivative of amino acid that has a strong antioxidative activity (Figure 7) [61,62]. *Koji* fermented with *A. oryzae* is reported to contain  $59.8 \pm 20.4 \,\mu\text{g/g}$  ergothioneine [62]. Feeding of ergothioneine at a concentration of 8 mg/kg body weight to mice protected against cisplatin-induced neuronal injury and enhances cognition [63].

Figure 7. Chemical structure of ergothioneine.

Since skin cells and tissue can absorb ergothioneine, and it is accumulated in peripheral skin cells, it is considered to contribute to the antioxidant activity of skin cells and tissue [64].

Functional variants of the ergothioneine transporter, OCTN cation transporter genes, are associated with ulcerative colitis, Crohn's disease, and autoimmune thyroid disease [65]. Therefore, it was hypothesized that the dietary intake of *koji* leads to the intake of ergothioneine and may contribute to the amelioration of these diseases.

# 9. Pyroglutamyl-Leucine

Peptides generated by proteases in *koji* are further modified during sake brewing (Figure 8). The pyroglutamyl-leucine contained in sake is one such example of a peptide, and is contained in sake at a concetration of 10–15 mg/L [66]. It has been shown to attenuate hepatitis and colitis in animal models [66].

Figure 8. Chemical structure of pyroglutamyl-leucine.

### 10. Pyranonigrin A

It has been reported that *koji* contains some antioxidant substances [67,68]. Later, one such antioxidant substance was identified as pyranonigrin A (Figure 9) [69]. Pyranonigrin A is a potential main protease inhibitor of SARS-CoV-2 and might be utilized to prevent infection of coronavirus disease 2019 (COVID-19) [70]. Therefore, the dietary intake of

J. Fungi **2021**, 7, 243 7 of 14

*koji* and related fermented products leads to the intake of pyranonigrin A, resulting in the prevention of cardiovascular diseases and COVID-19.

Figure 9. Chemical structure of pyranonigrin-A.

#### 11. Resistant Proteins

Most proteins in rice are degraded during sake brewing by the proteases contained in *koji*. However, nondigestable proteins, especially prolamin accumulated in the protein bodies of the endosperm of rice [71], remain in the sake lees, the remnant of sake brewing. Nondigestable proteins are called resistant proteins. As a result, sake lees contain a high content of resistant proteins. These resistant proteins are also resistant to human pancreatic proteases, and thus resistant proteins pass through the stomach and the small intestine and reaches the large intestine. As a result, they inhibit the absorption of lipids in the intestine, and improve intestinal microbial flora [72]. Therefore, the dietary intake of sake lees leads to the intake of resistant proteins, thus resulting in improvement of hyperlipidemia.

## 12. Deferriferrichrysin

*Koji* and its fungus *A. oryzae* contain a low-molecular-weight iron chelating peptide, deferriferrichrysin. Since it functions as a natural antioxidant [73], it might be beneficial for skin protection and prevention of inflammation.

# 13. Polyamines

During the manufacturing of *koji*, one of polyamines, agmatine (Figure 10), is produced at 3.1–8.7 mmol/L [74,75]. Polyamines have anti-inflammatory effects through the regulation of aberrant DNA methylation and decreased incidence of colon tumors, although accelerated the growth of established tumors [76]. Therefore, dietary intake of *koji* may lead to intake of polyamines, thus resulting in preventing the effects of colon tumors at a certain concentration.

$$H_2N$$
 $N$ 
 $NH_2$ 
 $NH_2$ 

Figure 10. Chemical structure of agmatine.

# 14. Bifidobacterium-Stimulating Peptides

Proteases contained in *koji* acts on rice proteins and gives rise to various peptides. Certain peptides containing glutamate, serine, and alanine in *koji* were shown to increase *Bifidobacteria*, *B. longum*, *B. adolescentis*, and *B. breve* [77]. Therefore, *koji* is considered to improve the intestinal microbe.

# 15. Angiotensin I-Converting Enzyme Inhibitor Peptides

The renin–angiotensin system increases blood pressure. Angoitensin I-converting enzyme catalyzes the conversion of Ang I to Ang II, which eventually generates AT1R and causes proinflammatory effects [78,79]. Sake utilizing *koji* contains the angiotensin I-converting enzyme inhibitor peptides Val-Tyr, His-Tyr, Arg-Phe, Val-Trp, and Tyr-Trp. These peptides have been shown to be antihypertensive in spontaneously hypertensive

J. Fungi **2021**, 7, 243 8 of 14

rats [80,81]. Therefore, the dietary intake of sake and *koji*-fermented products leads to the intake of these peptides, thus resulting in anti-hypertensive effects.

## 16. 14-Dehydroergosterol

It was found that *Aspergillus luchuensis mut. kawachii* produces 14-dehydroergosterol (Figure 11) as an anti-inflammatory substance that induces tolerogenic dendritic cells [82]. Moreover, *koji* extract containing 14-dehydroergosterol produced by *Aspergillus luchuensis mut. kawachii* has been shown to improve skin moisture, TEWL, and skin wrinkles in humans in randomized, double-blind, controlled trial [83].

**Figure 11.** Chemical structure of 14-dehydroergosterol.

#### 17. Beta-Glucan

Beta-glucan (Figure 12) was first identified as a medical component of *Agaricus blazeii* from the observations of lower incidence of cancers and infections of virus and bacteria, as well as the increased life span within native Americans [84]. Later, it was found that beta-glucan contained in the fungus is an immunostimulant and activates macrophages through dectin1 and CR3 (CD11b/CD18) [85,86]. It also ameliorates allergies and reduces the triglyceride and cholesterol levels in the blood [87]. *Aspergillus* spp. contains  $\beta$ -(1 $\rightarrow$ 3)-glucan as its cell wall component (Figure 11), and  $\beta$ -(1 $\rightarrow$ 3)-glucan of *Aspergillus fumigatus*, a close relative of *A. oryzae*, activates dectin-1 [88]. Since *koji* is manufactured using *A. oryzae*, it is considered to contain beta-glucan.

Figure 12. Chemical structure of beta-glucan.

The administration of beta-glucan is considered to improve the quality of life of cancer patients and is useful as a complementary or adjuvant therapy and immunomodulatory agent in cancer patients in combination with other cancer treatments [89,90]. Therefore, the content of beta-glucan in *koji* is desired to be investigated.

#### 18. Biotin

Lack of biotin (Figure 13) is known to causes skin disorders such as dermatitis, hair loss, neuritis, and susceptibility to infections [91]. It is also severely teratogenic in rodents. Since *Aspergillus oryzae* used for production of *koji* has a biosynthetic pathway of biotin [92] and *koji* contains 1.01 mg/100 g biotin [93], dietary intake of *koji* might contribute to the prevention of these disorders.

J. Fungi **2021**, 7, 243 9 of 14

Figure 13. Chemical structure of biotin.

#### 19. Citric Acid

A. luchuensis, which has been used for manufacturing awamori and shochu in Okinawa prefecture and Kyusyu island, produces citric acid (Figure 14) and koji fermented with A. luchuensis contains citric acid at approximately 1800 mg/L [94]. Therefore, koji manufactured using A. luchuensis contains citric acid (Figure 12). The dietary intake of citric acid (2 g/L) in drinking water inhibits the development of cataracts, proteinuria, and ketosis in diabetic rats [95]. Moreover, in a mouse model of systemic inflammation, dietary intake of citric acid (1–2 g/kg) has been shown to decrease brain lipid peroxidation and inflammation, liver damage, and DNA fragmentation in lipopolysaccharide-treated mice [96]. Therefore, fermented foods containing koji manufactured with A. luchuensis might have beneficial health effects toward diabetic patients and chronic inflammation, which should be a target of future study.

Figure 14. Chemical structure of citric acid.

# 20. Potential Risk of Koji

A. oryzae in general produces aflatoxin and cyclopiazonic acid [97]. Ingestion of aflatoxin causes an acute hepatic necrosis [98]. Cyclopiazonic acid shows immunosuppressive activity at low doses and causes necrosis of various tissues [99]. However, since A. oryzae used for manufacture of koji has been long succeeded in Japan, it has lost functional genes to produce aflatoxin [100] or acquired a new gene to produce less toxic cyclopiazonic acid [101]. Also, A. luchuensis has lost the functional gene to synthesize ochratoxin A [102].

It should be noted that some strains produce trace amounts of certain mycotoxins including aspirochlorin and csypyrones [97] and it is necessary to monitor the concentrations of these substances.

#### 21. Conclusions

In conclusion, *koji* fermented with *A. oryzae* or *A. luchuensis* and *koji*-fermented products contain abundant substances that have the potential for use in medical applications (Figure 15). It is desired that the medical benefits of these substances be verified in human studies and applied to practical medical use. More research is desired to medically apply *koji* to cure diseases across the world in the future.

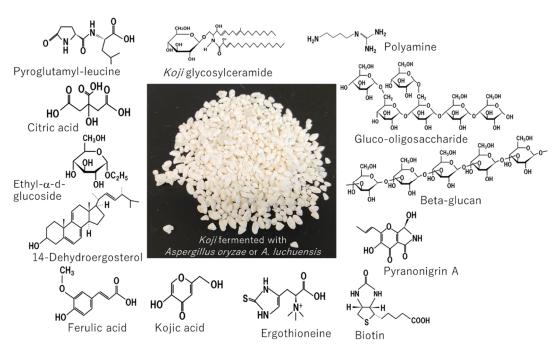


Figure 15. Medicinal substances contained in nonpathogenic Aspergillus-fermented Koji.

Funding: This research was funded by JSPS KAKENHI grant number 20K05809.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

#### References

- Rippon, J.W. Pathogenesis and epidemiology of opportunistic mycotic infections: A review. Am. J. Med. Technol. 1977, 43, 226–228. [PubMed]
- 2. Keller, N.; Turner, G.; Bennett, J. Fungal secondary metabolism—From biochemistry to genomics. *Nat. Rev. Microbiol.* **2005**, *3*, 937–947. [CrossRef] [PubMed]
- 3. Wasser, S.P. Current findings, future trends, and unsolved problems in studies of medicinal mushrooms. *Appl. Microbiol. Biotechnol.* **2011**, *89*, 1323–1332. [CrossRef]
- 4. Wang, Y.-Z.; Zhang, J.; Zhao, Y.-Z.; Li, T.; Shen, T.; Li, J.-Q.; Li, W.-Y.; Liu, H.-G. Mycology, cultivation, traditional uses, phytochemistry and pharmacology of Wolfiporia cocos (Schwein.) Ryvarden et Gilb.: A review. *J. Ethnopharmacol.* 2013, 147, 265–276. [CrossRef] [PubMed]
- 5. Cör, D.; Knez, Ž.; Knez Hrnčič, M. Antitumour, antimicrobial, antioxidant and antiacetylcholinesterase effect of *Ganoderma Lucidum* terpenoids and polysaccharides: A review. *Molecules* **2018**, *23*, 649. [CrossRef]
- 6. Zhang, Y.; Kong, H.; Fang, Y.; Nishinari, K.; Phillips, G.O. Schizophyllan: A review on its structure, properties, bioactivities and recent development. *Bioact. Carbohydr. Diet. Fibre* **2013**, *1*, 53–71. [CrossRef]
- 7. Lin, Y.L.; Wang, T.H.; Lee, M.H.; Su, N.W. Biologically active components and nutraceuticals in the *Monascus*-fermented rice: A review. *Appl. Microbiol. Biotechnol.* **2008**, 77, 965–973. [CrossRef] [PubMed]
- 8. Endo, A. Monacolin K, a new hypocholesterolemic agent produced by *Monascus* species. *J. Antibiot.* **1979**, 32, 852–854. [CrossRef] [PubMed]
- 9. Akihisa, T.; Tokuda, H.; Ukiya, M.; Kiyota, A.; Yasukawa, K.; Sakamoto, N.; Kimura, Y.; Suzuki, T.; Takayasu, J.; Nishino, H. Anti-tumor-initiating effects of Monascin, an azaphilonoid pigment from the extract of *Monascus pilosus* fermented rice (red-mold rice). *Chem. Biodivers.* **2005**, *2*, 1305–1309. [CrossRef]
- 10. Hsu, W.H.; Lee, B.L.; Huang, Y.C.; Hsu, Y.W.; Pan, T.M. Ankaflavin, a novel Nrf-2 activator for attenuating allergic airway inflammation. *Free Radic. Biol. Med.* **2012**, 53, 1643–1651. [CrossRef]
- 11. Kohama, K.; Tanaka, T.; Sakamoto, M.; Dai, H.; Tsuge, K.; Kawaguchi, S.; Ozeki, Y.; Fukami, Y.; Kitagaki, H. Identification of ergosterol as the anti-inflammatory substance contained in Monascus anka and M. pilosus. *J. Brew. Soc. Jpn.* **2020**, *115*, 1–9.

12. Fleming, A. On the antibacterial action of cultures of a *Penicillium*, with special reference to their use in the isolation of *B. influenzæ*. *Br. J. Exp. Pathol.* **1929**, *10*, 226–236. [CrossRef]

- 13. Ahnan-Winarno, A.D.; Cordeiro, L.; Winarno, F.G.; Gibbons, J.; Xiao, H. Tempeh: A semicentennial review on its health benefits, fermentation, safety, processing, sustainability, and affordability. *Compr. Rev. Food Sci. Food Saf.* **2021**, 20, 1717–1767. [CrossRef] [PubMed]
- 14. Kitagaki, H.; Kitamoto, K. Breeding research on sake yeasts in Japan: History, recent technological advances, and future perspectives. *Annu. Rev. Food Sci. Technol.* **2013**, *4*, 215–235. [CrossRef]
- 15. Asano, M.; Nakano, F.; Nakatsukasa, E.; Tsuduki, T. The 1975 type Japanese diet improves the gut microbial flora and inhibits visceral fat accumulation in mice. *Biosci. Biotechnol. Biochem.* **2020**, *84*, 1475–1485. [CrossRef]
- 16. Takamine, J. Amylolytic Enzym. U.S. Patent 991561A, 9 May 1911.
- 17. Yang, Y.; Iwamoto, A.; Kumrungsee, T.; Okazaki, Y.; Kuroda, M.; Yamaguchi, S.; Kato, N. Consumption of an acid protease derived from *Aspergillus oryzae* causes bifidogenic effect in rats. *Nutr. Res.* **2017**, 44, 60–66. [CrossRef]
- 18. Yang, Y.; Kumrungsee, T.; Kuroda, M.; Yamaguchi, S.; Kato, N. Feeding *Aspergillus* protease preparation combined with adequate protein diet to rats increases levels of cecum gut-protective amino acids, partially linked to *Bifidobacterium* and *Lactobacillus*. *Biosci. Biotechnol. Biochem.* **2019**, *83*, 1901–1911. [CrossRef]
- 19. Hannun, Y.; Obeid, L. Sphingolipids and their metabolism in physiology and disease. *Nat. Rev. Mol. Cell Biol.* **2018**, 19, 175–191. [CrossRef]
- 20. Heung, L.J.; Luberto, C.; Del Poeta, M. Role of sphingolipids in microbial pathogenesis. *Infect. Immun.* 2006, 74, 28–39. [CrossRef]
- 21. Simons, K.; Ikonen, E. Functional rafts in cell membranes. Nature 1997, 387, 569-572. [CrossRef]
- 22. Tani, Y.; Amaishi, Y.; Funatsu, T.; Ito, M.; Itonori, S.; Hata, Y.; Ashida, H.; Yamamoto, K. Structural analysis of cerebrosides from *Aspergillus* fungi: The existence of galactosylceramide in A. oryzae. *Biotechnol. Lett.* **2014**, *36*, 2507–2513. [CrossRef]
- 23. Hirata, M.; Tsuge, K.; Jayakody, L.N.; Urano, Y.; Sawada, K.; Inaba, S.; Nagao, K.; Kitagaki, H. Structural determination of glucosylceramides in the distillation remnants of shochu, the Japanese traditional liquor, and its production by *Aspergillus kawachii*. *J. Agric. Food Chem.* **2012**, *60*, 11473–11482. [CrossRef]
- 24. Takahashi, K.; Izumi, K.; Nakahata, E.; Hirata, M.; Sawada, K.; Tsuge, K.; Nagao, K.; Kitagaki, H. Quantitation and structural determination of glucosylceramides contained in sake lees. *J. Oleo Sci.* **2014**, *63*, 15–23. [CrossRef] [PubMed]
- 25. Hamajima, H.; Fujikawa, A.; Yamashiro, M.; Ogami, T.; Kitamura, S.; Tsubata, M.; Tan, S.; Matsunaga, H.; Sawada, K.; Kumagai, S.; et al. Chemical analysis of the sugar moiety of monohexosylceramide contained in *koji*, Japanese traditional rice fermented with *Aspergillus. Fermentation* **2016**, *2*, 2. [CrossRef]
- 26. Fujino, Y.; Ohnishi, M. Structure of cerebroside in Aspergillus oryzae. Biochim. Biophys. Acta 1976, 486, 161–171. [PubMed]
- 27. Sakamoto, M.; Sakatani, M.; Ferdouse, J.; Hamajima, H.; Tsuge, K.; Nishimukai, M.; Yanagita, T.; Nagao, K.; Mitsutake, S.; Kitagaki, H. Development of a quantitative method for the contents of glycosylceramide contained in Japanese foods brewed with *koji* and its application. *J. Brew. Soc. Jpn.* **2017**, *112*, 655–662.
- 28. Hamajima, H.; Matsunaga, H.; Fujikawa, A.; Sato, T.; Mitsutake, S.; Yanagita, T.; Nagao, K.; Nakayama, J.; Kitagaki, H. Japanese traditional dietary fungus *koji* Aspergillus oryzae functions as a prebiotic for *Blautia coccoides* through glycosylceramide: Japanese dietary fungus *koji* is a new prebiotic. *SpringerPlus* **2016**, *5*, 1321. [CrossRef] [PubMed]
- 29. Hamajima, H.; Tanaka, M.; Miyagawa, M.; Sakamoto, M.; Nakamura, T.; Yanagita, T.; Nishimukai, M.; Mitsutake, S.; Nakayama, J.; Nagao, K.; et al. *Koji* glycosylceramide commonly contained in Japanese traditional fermented foods alters cholesterol metabolism in obese mice. *Biosci. Biotechnol. Biochem.* **2019**, *83*, 1514–1522. [CrossRef]
- 30. Miyagawa, M.; Fujikawa, A.; Nagadome, M.; Kohama, K.; Ogami, T.; Kitamura, S.; Kitagaki, H. Glycosylceramides purified from the Japanese traditional non-pathogenic fungus *Aspergillus* and *koji* increase the expression of genes involved in tight junctions and ceramide delivery in normal human epidermal keratinocytes. *Fermentation* **2019**, *5*, 43. [CrossRef]
- 31. Uchiyama, T.; Nakano, Y.; Ueda, O.; Mori, H.; Nakashima, M.; Noda, A.; Ishizaki, C.; Mizoguchi, M. Oral intake of glucosylceramide improves relatively higher level of transepidermal water loss in mice and healthy human subjects. *J. Health Sci.* **2008**, *54*, 559–566. [CrossRef]
- 32. Burdock, G.A.; Soni, M.G.; Carabin, I.G. Evaluation of health aspects of *kojic* acid in food. *Regul. Toxicol. Pharmacol.* **2001**, 33, 80–101. [CrossRef]
- 33. Da Costa, J.P.; Rodrigues, A.P.D.; Farias, L.H.S.; Frade, P.C.R.; Da Silva, B.J.M.; Nascimento, J.L.M.D.; Silva, E.O. Biological effects of kojic acid on human monocytes in vitro. *Biomed. Pharmacother.* **2018**, *101*, 100–106. [CrossRef]
- 34. Henry, R.J. The carbohydrates of barley grains. J. Inst. Brew. 1988, 94, 71–78. [CrossRef]
- 35. Okuda, M. Rice used for Japanese sake making. Biosci. Biotechnol. Biochem. 2019, 83, 1428–1441. [CrossRef]
- 36. Bastawde, K.B. Xylan structure, microbial xylanases, and their mode of action. *World J. Microbiol. Biotechnol.* **1992**, *8*, 353–368. [CrossRef] [PubMed]
- 37. Machida, M.; Asai, K.; Sano, M.; Tanaka, T.; Kumagai, T.; Terai, G.; Kusumoto, K.-I.; Arima, T.; Akita, O.; Kashiwagi, Y.; et al. Genome sequencing and analysis of *Aspergillus oryzae*. *Nature* **2005**, *438*, 1157–1161. [CrossRef] [PubMed]
- 38. Nigam, P.; Singh, D. Enzyme and microbial systems involved in starch processing. *Enzyme Microb. Technol.* **1995**, 17, 770–778. [CrossRef]
- 39. Robyt, J.F.; French, D. The action pattern of porcine pancreatic alpha-amylase in relationship to the substrate binding site of the enzyme. *J. Biol. Chem.* **1970**, 245, 3917–3927. [CrossRef]

40. Baba, S.; Okuri, Y.; Fukuzawa, M.; Iida, T.; Kobayashi, I.; Imai, K. Generation of oligosaccharides from steamed rice by enzymes of Aspergillus oryzae. *J. Brew. Soc. Jpn.* **1974**, *69*, 844–846.

- 41. Honda, C.; Katsuta, R.; Yamada, M.; Kojima, Y.; Mamiya, A.; Okada, N.; Kawamura, T.; Totsuka, A.; Shindo, H.; Hosaka, M.; et al. Novel glucoamylase-resistant gluco-oligosaccharides with adjacent α-1, 6 branches at the non-reducing end discovered in Japanese rice wine, sake. *Carbohydr. Polym.* **2021**, 251, 116993. [CrossRef]
- 42. Kobayashi, M.; Matsushita, H.; Shioya, I.; Nagai, M.; Tsukiyama, R.; Saito, M.; Yamamoto, K. Quality of life improvement with soy sauce ingredients, Shoyu polysaccharides, in perennial allergic rhinitis: A double-blind placebo-controlled clinical study. *Int. J. Mol. Med.* 2004, 14, 885–889. [CrossRef] [PubMed]
- 43. Bouhnik, Y.; Vahedi, K.; Achour, L.; Attar, A.; Salfati, J.; Pochart, P.; Marteau, P.; Flourié, B.; Bornet, F.; Rambaud, J.C. Short-chain fructo-oligosaccharide administration dose-dependently increases fecal bifidobacteria in healthy humans. *J. Nutr.* **1999**, 129, 113–116. [CrossRef] [PubMed]
- 44. Ishikawa, H.; Matsumoto, S.; Ohashi, Y.; Imaoka, A.; Setoyama, H.; Umesaki, Y.; Tanaka, R.; Otani, T. Beneficial effects of probiotic bifidobacterium and galacto-oligosaccharide in patients with ulcerative colitis: A randomized controlled study. *Digestion* **2011**, *84*, 128–133. [CrossRef] [PubMed]
- 45. Malaguarnera, M.; Greco, F.; Barone, G.; Gargante, M.P.; Malaguarnera, M.; Toscano, M.A. *Bifidobacterium longum* with fructo-Oligosaccharide (FOS) treatment in minimal hepatic encephalopathy: A randomized, double-blind, placebo-controlled study. *Dig. Dis. Sci.* **2007**, *52*, 3259. [CrossRef] [PubMed]
- 46. Kobayashi, M.; Nagatani, Y.; Magishi, N.; Tokuriki, N.; Nakata, Y.; Tsukiyama, R.; Tsuji, K. Promotive effect of Shoyu polysaccharides from soy sauce on iron absorption in animals and humans. *Int. J. Mol. Med.* **2006**, *18*, 1159–1163. [CrossRef]
- 47. Furuta, Y.; Hokazono, R.; Takashita, H.; Omori, T.; Ishizaki, A.; Sonomoto, K. Growth stimulator of lactic acid bacteria and Bifidobacteria in by-product of barley shochu. *Seibutsu-kogaku* **2007**, *85*, 161–166.
- 48. Mi, H.; Dong, Y.; Zhang, B.; Wang, H.; Peter, C.; Gao, P.; Fu, H.; Gao, Y. *Bifidobacterium infantis* ameliorates chemotherapy-induced intestinal Mucositis via regulating T cell immunity in colorectal cancer rats. *Cell. Physiol. Biochem.* **2017**, 42, 2330–2341. [CrossRef] [PubMed]
- 49. Lee, D.K.; Jang, S.; Kim, M.J.; Kim, J.H.; Chung, M.J.; Kim, K.J.; Ha, N.J. Anti-proliferative effects of *Bifidobacterium adolescentis* SPM0212 extract on human colon cancer cell lines. *BMC Cancer* **2008**, *8*, 310. [CrossRef]
- 50. Imanari, T.; Zenzo Tamura, Z. The identification of α-ethyl glucoside and sugar-alcohols in sake. *Agric. Biol. Chem.* **1971**, 35, 321–324. [CrossRef]
- 51. Kojima, Y.; Honda, C.; Kobayashi, I.; Katsuta, R.; Matsumura, S.; Wagatsuma, I.; Takehisa, M.; Shindo, H.; Hosaka, M.; Nukada, T.; et al. Transglycosylation forms novel glycoside ethyl α-maltoside and ethyl α-isomaltoside in sake during the brewing process by α-glucosidase A of *Aspergillus oryzae*. *J. Agric. Food Chem.* **2020**, *68*, 1419–1426. [CrossRef]
- 52. Hirotsune, M.; Haratake, A.; Komiya, A.; Sugita, J.; Tachihara, T.; Komai, T.; Hizume, K.; Ozeki, K.; Ikemoto, T. Effect of ingested concentrate and components of sake on epidermal permeability barrier disruption by UVB irradiation. *J. Agric. Food Chem.* **2005**, 53, 948–952. [CrossRef]
- 53. Bogaki, T.; Mitani, K.; Oura, Y.; Ozeki, K. Effects of ethyl-α-d-glucoside on human dermal fibroblasts. *Biosci. Biotechnol. Biochem.* **2017**, *81*, 1706–1711. [CrossRef]
- 54. Nakahara, M.; Mishima, T.; Hayakawa, T. Effect of a sake concentrate on the epidermis of aged mice and confirmation of ethyl alpha-D-glucoside as its active component. *Biosci. Biotechnol. Biochem.* **2007**, *71*, 427–434. [CrossRef] [PubMed]
- 55. Lan, W.; Yang, Y.; Zhonghu, H.; Desen, W.; Aihua, L.; Yong, Z. Determination of phenolic acid concentrations in wheat flours produced at different extraction rates. *J. Cereal Sci.* **2013**, *57*, 67–72.
- 56. Uno, T.; Itoh, A.; Miyamoto, T.; Kubo, M.; Kanamaru, K.; Yamagata, H.; Yasufuku, Y.; Imaishi, H. Ferulic acid production in the brewing of rice wine (sake). *J. Inst. Brew.* **2009**, *115*, 116–121. [CrossRef]
- 57. Hasizume, K.; Ito, T.; Shimohashi, M.; Ishizuka, T.; Okuda, M. Ferulic acid and ethyl ferulate in sake: Comparison of levels between sake and mirin and analysis of their sensory properties. *Food Sci. Tech. Res.* **2013**, *19*, 705–809. [CrossRef]
- 58. Balasubashini, M.S.; Rukkumani, R.; Menon, V.P. Protective effects of ferulic acid on hyperlipidemic diabetic rats. *Acta Diabetol.* **2003**, *40*, 118–122. [CrossRef] [PubMed]
- 59. Kohno, M.; Musashi, K.; Ikeda, H.O.; Horibe, T.; Matsumoto, A.; Kawakami, K. Oral administration of ferulic acid or ethyl ferulate attenuates retinal damage in sodium iodate-induced retinal degeneration mice. *Sci. Rep.* **2020**, *10*, 8688. [CrossRef]
- 60. Yan, J.-J.; Cho, J.-Y.; Kim, H.-S.; Kim, K.-L.; Jung, J.-S.; Huh, S.-O.; Suh, H.-W.; Kim, Y.-H.; Song, D.-K. Protection against β-amyloid peptide toxicity *in vivo* with long-term administration of ferulic acid. *Br. J. Pharmacol.* **2001**, *133*, 89–96. [CrossRef]
- 61. Halliwell, B.; Cheah, I.K.; Tang, R.M.Y. Ergothioneine—A diet-derived antioxidant with therapeutic potential. *FEBS Lett.* **2018**, 592, 3357–3366. [CrossRef] [PubMed]
- 62. Horie, Y.; Goto, A.; Imamura, R.; Itoh, M.; Ikegawa, S.; Ogawa, S.; Higashi, T. Quantification of ergothioneine in *Aspergillus oryzae*-fermented rice bran by a newly-developed LC/ESI-MS/MS method. *LWT* **2020**, *118*, 108812. [CrossRef]
- 63. Song, T.Y.; Chen, C.L.; Liao, J.W.; Ou, H.C.; Tsai, M.S. Ergothioneine protects against neuronal injury induced by cisplatin both in vitro and in vivo. *Food Chem. Toxicol. Assoc.* **2010**, *48*, 3492–3499. [CrossRef]
- 64. Markova, N.G.; Karaman-Jurukovska, N.; Dong, K.K.; Damaghi, N.; Smiles, K.A.; Yarosh, D.B. Skin cells and tissue are capable of using L-ergothioneine as an integral component of their antioxidant defense system. *Free Radic. Biol. Med.* **2009**, *46*, 1168–1176. [CrossRef]

65. Peltekova, V.D.; Wintle, R.F.; Rubin, L.A.; Amos, C.I.; Huang, Q.; Gu, X.; Newman, B.; Van Oene, M.; Cescon, D.; Greenberg, G.; et al. Functional variants of OCTN cation transporter genes are associated with Crohn disease. *Nat. Genet.* **2004**, *36*, 471–475. [CrossRef] [PubMed]

- 66. Kiyono, T.; Hirooka, K.; Yamamoto, Y.; Kuniishi, S.; Ohtsuka, M.; Kimura, S.; Park, E.Y.; Nakamura, Y.; Sato, K. Identification of pyroglutamyl peptides in Japanese rice wine (sake): Presence of hepatoprotective pyroGlu-Leu. *J. Agric. Food Chem.* **2013**, *61*, 11660–11667. [CrossRef] [PubMed]
- 67. Kitagaki, H. Transition of DPPH-scavenging ability during sake brewing. J. Brew. Soc. Jpn. 2003, 98, 589–593. [CrossRef]
- 68. Saigusa, N.; Ohba, R. Effects of *koji* production and Saccharification time on the antioxidant activity of amazake. *Food Sci. Technol. Res.* **2007**, *13*, 162–165. [CrossRef]
- 69. Miyake, Y.; Ito, C.; Itoigawa, M.; Osawa, T. Isolation of the antioxidant pyranonigrin-A from rice mold starters used in the manufacturing process of fermented foods. *Biosci. Biotechnol. Biochem.* **2007**, *71*, 2515–2521. [CrossRef]
- 70. Rao, P.; Shukla, A.; Parmar, P.; Rawal, R.M.; Patel, B.; Saraf, M.; Goswami, D. Reckoning a fungal metabolite, pyranonigrin A as a potential main protease (M<sup>pro</sup>) inhibitor of novel SARS-CoV-2 virus identified using docking and molecular dynamics simulation. *Biophys. Chem.* **2020**, *264*, 106425. [CrossRef]
- 71. Kizaki, Y.; Inoue, Y.; Okazaki, N.; Kobayashi, S. Isolation and determination of protein bodies (PB-I, PB-II) in polished rice endosperm. *J. Brew. Soc. Jpn.* **1991**, *86*, 293–298. [CrossRef]
- 72. Watanabe, T. Ingredients in "Sake Cake" contribute to health and beauty. J. Brew. Soc. Jpn. 2012, 107, 282–291. [CrossRef]
- 73. Todokoro, T.; Fukuda, K.; Matsumura, K.; Irie, M.; Hata, Y. Production of the natural iron chelator deferriferrichrysin from *Aspergillus oryzae* and evaluation as a novel food-grade antioxidant. *J. Sci. Food Agric.* **2016**, *96*, 2998–3006. [CrossRef] [PubMed]
- 74. Kobayashi, K.; Watanabe, S. Changes in polyamine contents of miso *koji* during *koji* cultivation. *J. Brew. Soc. Jpn.* **2017**, 112, 140–146.
- 75. Akasaka, N.; Kato, S.; Kato, S.; Hidese, R.; Wagu, Y.; Sakoda, H.; Fujiwara, S. Agmatine production by *Aspergillus oryzae* is elevated by low pH during solid-state cultivation. *Appl. Environ. Microbiol.* **2018**, *84*, e00722-18. [CrossRef]
- 76. Soda, K.; Kano, Y.; Chiba, F.; Koizumi, K.; Miyaki, Y. Increased polyamine intake inhibits age-associated alteration in global DNA methylation and 1,2-dimethylhydrazine-induced tumorigenesis. *PLoS ONE* **2013**, *8*, e64357. [CrossRef] [PubMed]
- 77. Hosoyama, H.; Osawa, M.; Hamano, M. Bifidobacterium-stimulating substance in rice bran *koji. J. Jpn. Soc. Food Sci. Technol.* **1991**, 38, 940–944. [CrossRef]
- 78. Wang, D.; Chai, X.Q.; Magnussen, C.G.; Zosky, G.R.; Shu, S.H.; Wei, X.; Hu, S.S. Renin-angiotensin-system, a potential pharmacological candidate, in acute respiratory distress syndrome during mechanical ventilation. *Pulm. Pharmacol. Ther.* **2019**, *58*, 101833. [CrossRef] [PubMed]
- 79. Chappell, M.C. Biochemical evaluation of the renin-angiotensin system: The good, bad, and absolute? *Am. J. Physiol. Heart Circ. Physiol.* **2016**, *310*, H137–H152. [CrossRef] [PubMed]
- 80. Saito, Y.; Wanezaki, K.; Kawato, A.; Imayasu, S. Antihypertensive effects of peptide in sake and its by-products on spontaneously hypertensive rats. *Biosci. Biotechnol. Biochem.* **1994**, *58*, 812–816. [CrossRef]
- 81. Saito, Y.; Wanezaki, K.; Kawato, A.; Imayasu, S. Structure and activity of angiotensin I converting enzyme inhibitory peptides from sake and sake lees. *Biosci. Biotechnol. Biochem.* **1994**, *58*, 1767–1771. [CrossRef]
- 82. Ano, Y.; Ikado, K.; Shindo, K.; Koizumi, H.; Fujiwara, D. Identification of 14-dehydroergosterol as a novel anti-inflammatory compound inducing tolerogenic dendritic cells. *Sci. Rep.* **2017**, *7*, 13903. [CrossRef]
- 83. Sugihara, Y.; Ikushima, S.; Miyake, M.; Kirisako, T.; Yada, Y.; Fujiwara, D. Improvement of skin conditions by ingestion of *Aspergillus kawachii* (*Koji*) extract containing 14-dehydroergosterol in a randomized, double-blind, controlled trial. Clinical, cosmetic and investigational dermatology. *Clin. Cosmet. Investig. Dermatol.* 2018, 11, 115–124. [CrossRef]
- 84. Vetvicka, V.; Vannucci, L.; Sima, P.; Richter, J. Beta glucan: Supplement or drug? From laboratory to clinical trials. *Molecules* **2019**, 24, 1251. [CrossRef]
- 85. Brown, G.D.; Taylor, P.R.; Reid, D.M.; Willment, J.A.; Williams, D.L.; Martinez-Pomares, L.; Wong, S.Y.; Gordon, S. Dectin-1 is a major beta-glucan receptor on macrophages. *J. Exp. Med.* **2002**, *196*, 407–412. [CrossRef] [PubMed]
- 86. Di Renzo, L.; Yefenof, E.; Klein, E. The function of human NK cells is enhanced by β-glucan, a ligand of CR3 (CD11b/CD18). *Eur. J. Immunol.* **1991**, 21, 1755–1758. [CrossRef] [PubMed]
- 87. Anderson, J.W.; Story, L.; Sieling, B.; Chen, W.J.; Petro, M.S.; Story, J. Hypocholesterolemic effects of oat-bran or bean intake for hypercholesterolemic men. *Am. J. Clin. Nutr.* **1984**, *40*, 1146–1155. [CrossRef]
- 88. Steele, C.; Rapaka, R.R.; Metz, A.; Pop, S.M.; Williams, D.L.; Gordon, S.; Kolls, J.K.; Brown, G.D. The beta-glucan receptor dectin-1 recognizes specific morphologies of *Aspergillus fumigatus*. *PLoS Pathog.* **2005**, *1*, e42. [CrossRef] [PubMed]
- 89. Ostadrahimi, A.; Esfahani, A.; Asghari Jafarabadi, M.; Eivazi Ziaei, J.; Movassaghpourakbari, A.; Farrin, N. Effect of beta glucan on quality of life in women with breast cancer undergoing chemotherapy: A randomized double-blind placebo-controlled clinical trial. *Adv. Pharm. Bull* **2014**, *4*, 471–477.
- 90. Ostadrahimi, A.; Ziaei, J.E.; Esfahani, A.; Jafarabadi, M.A.; Movassaghpourakbari, A.; Farrin, N. Effect of beta glucan on white blood cell counts and serum levels of IL-4 and IL-12 in women with breast cancer undergoing chemotherapy: A randomized double-blind placebo-controlled clinical trial. *Asian Pac. J. Cancer Prev.* 2014, 15, 5733–5739. [CrossRef] [PubMed]
- 91. León-Del-Río, A. Biotin in metabolism, gene expression, and human disease. Journal of inherited metabolic disease. *J. Inherit. Metab. Dis.* **2019**, 42, 647–654. [CrossRef]

92. Tanabe, Y.; Maruyama, J.I.; Yamaoka, S.; Yahagi, D.; Matsuo, I.; Tsutsumi, N.; Kitamoto, K. Peroxisomes are involved in biotin biosynthesis in *Aspergillus* and *Arabidopsis*. *J. Biol. Chem.* **2011**, *286*, 30455–30461. [CrossRef]

- 93. Kurahashi, A.; Oguro, Y. Ingredients in koji amazake. J. Brew. Soc. Jpn. 2017, 112, 668-674.
- 94. Ochi, H.; Yamamoto, H.; Takayama, K.; Mizutani, M. Study on citric acid productivity by liquid barley *koji*. *Rep. Miyazaki Prefect*. *Ind. Technol. Cent. Miyazaki Prefect*. *Food R D Cent*. **2013**, *56*, 87–89.
- 95. Nagai, R.; Nagai, M.; Shimasaki, S.; Baynes, J.W.; Fujiwara, Y. Citric acid inhibits development of cataracts, proteinuria and ketosis in streptozotocin (type 1) diabetic rats. *Biochem. Biophys. Res. Commun.* **2010**, *393*, 118–122. [CrossRef]
- 96. Abdel-Salam, O.M.; Youness, E.R.; Mohammed, N.A.; Morsy, S.M.; Omara, E.A.; Sleem, A.A. Citric acid effects on brain and liver oxidative stress in lipopolysaccharide-treated mice. *J. Med. Food* **2014**, *17*, 588–598. [CrossRef]
- 97. Rank, C.; Klejnstrup, M.L.; Petersen, L.M.; Kildgaard, S.; Frisvad, J.C.; Held Gotfredsen, C.; Ostenfeld Larsen, T. Comparative chemistry of *Aspergillus oryzae* (RIB40) and *A. flavus* (NRRL 3357). *Metabolites* **2012**, 2, 39–56. [CrossRef]
- 98. Massey, T.E.; Stewart, R.K.; Daniels, J.M.; Liu, L. Biochemical and molecular aspects of mammalian susceptibility to aflatoxin B1 carcinogenicity. *Proc. Soc. Exp. Biol. Med.* **1995**, 208, 213–227. [CrossRef] [PubMed]
- 99. Burdock, G.A.; Flamm, W.G. Safety assessment of the mycotoxin cyclopiazonic acid. *Intern. J. Toxicol.* **2000**, 19, 195–218. [CrossRef]
- 100. Kiyota, T.; Hamada, R.; Sakamoto, K.; Iwashita, K.; Yamada, O.; Mikami, S. Aflatoxin non-productivity of *Aspergillus oryzae* caused by loss of function in the aflJ gene product. *J. Biosci. Bioeng.* **2011**, *111*, 512–517. [CrossRef] [PubMed]
- 101. Kato, N.; Tokuoka, M.; Shinohara, Y.; Kawatani, M.; Uramoto, M.; Seshime, Y.; Fujii, I.; Kitamoto, K.; Takahashi, S.; et al. Genetic safeguard against mycotoxin cyclopiazonic acid production in *Aspergillus oryzae*. *ChemBioChem* **2011**, *12*, 1376–1382. [CrossRef]
- 102. Yamada, O.; Takara, R.; Hamada, R.; Hayashi, R.; Tsukahara, M.; Mikami, S. Molecular biological researches of Kuro-*Koji* molds, their classification and safety. *J. Biosci. Bioeng.* **2011**, *112*, 233–237. [CrossRef] [PubMed]