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Wood-inhabiting fungi in southern China. 4. Polypores from Hainan Province

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Extensive surveys on wood-rotting fungi were recently carried out in Hainan Province, southern China. Around 2500 specimens of poroid wood-inhabiting fungi were collected during ten field trips, and 235 polypores were identified. Four species, *Grammothelopsis asiatica* Y.C. Dai & B.K. Cui, *Inonotus latemarginatus* Y.C. Dai, *Perenniporia hattorii* Y.C. Dai & B.K. Cui and *Wrightoporia austrosinensis* Y.C. Dai, are described and illustrated as new. Of the 235 species, 99 were found from the province for the first time.

Introduction

Hainan Province, located between 18°10´–20°10´N and 108°37´–111°05´E, is an island in southern China. It has both lowland area in the northeast and mountainous terrain in the central and southwestern regions, and the highest altitude is 1867 m a.s.l. in the central area. The annual mean temperature is 19.7 °C, and the annual precipitation is 2650 mm. Most parts of the island have a subtropical to tropical vegetation. The plant flora is well known, and 3324 seed plants, belonging to 1237 genera and 206 families, are reported (Wu 1994). However, the fungal flora is poorly known; 305 agarics were recorded previously from the province (Bi *et al*. 1997), and 136 polypores were found mostly during more or less recent inventories (Zhao 1989, Zhao & Zhang 1992, Dai *et al*. 2004, 2009a, 2009b, Yuan & Dai 2008, Xiong & Dai 2008, Yu & Dai 2009, Zhou *et al*. 2009, Cui *et al*. 2010, 2011a, 2011b, Dai 2010). Considering the abundance of woody species in the study area, the polypore flora is far from completely known.

Several mycological investigations dealing with many new species from Hainan were published recently (Zhang *et al*. 2008a, 2008b, 2009, Ma *et al*. 2008a, 2008b). The present paper belongs to a series of studies devoted to the fungal diversity of the Hainan Province. By support from the National Natural Science Foundation of China and the Ministry Science and Technology of China, ten field investigations on wood-decaying fungi were made in the Hainan Province during 2002–2009, and around 2500 specimens of poroid wood-inhabiting fungi

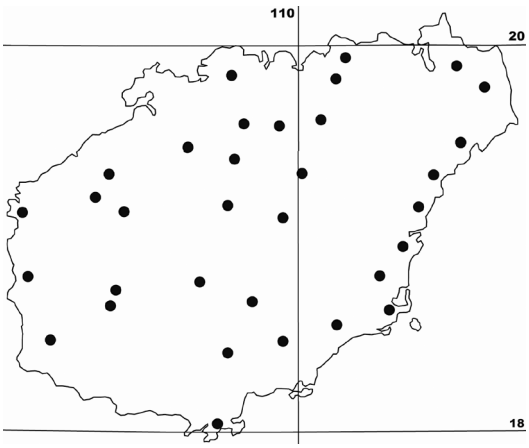

Fig. 1. Locations (black dots) of the investigated areas in Hainan Province.

were collected from 25 localities (Fig. 1). 235 polypores were identified after laboratory analysis, including four new species, and the present paper is a summary of these inventories.

Material and methods

The present study is based mostly on materials collected by the authors, and the studied specimens are deposited in the herbaria of the Beijing Forestry University (BJFC) and the Institute of Applied Ecology of the Chinese Academy of Sciences (IFP). A few specimens previously collected from the province and deposited in the herbarium of the Institute of Microbiology, the Chinese Academy of Sciences (HMAS), were also studied. All the materials were examined under the microscope Nikon 80i. The drawings were made with the aid of a drawing tube. Microscopic features, measurements, and drawings were made from slide preparations stained with Cotton Blue and Melzer's reagent. Spores were measured from sections cut from the tubes; in presenting spore size data 5% of the measurements were excluded from each end of the range, and these are shown in parentheses. The abbreviations include IKI (Melzer's reagent, with IKI– = inamyloid), KOH (5% potassium hydroxide), and CB (Cotton Blue; CB+ = cyanophilous; CB(+) = weakly cyanophilous; CB– = acyanophilous). Additional abbreviations include *L* (mean spore length; arithmetic average of all spores), *W* (mean spore width; arithmetic average of all spores), *Q* (variation in the *L*/*W* ratios between the specimens studied), and n (number of spores measured from a given number of specimens). The special color terms follow Petersen (1996).

New taxa

Grammothelopsis asiatica Y.C. Dai & B.K. Cui, *sp. nova* (Fig. 2)

MycoBank no.: MB 518961.

Carpophorum, resupinatum. Facies pororum cremea; pori angulati vel rotundi, 3–4 per mm. Systema hypharum dimiticum, hyphae generatoriae fibulatae, hyphae skeletales IKI–, hyphae skeletales subiculi 2–4.5 µm in diam. Sporae ellipsoideae, IKI–, CB– vel CB(+), 10.5–13 × 5.4–6 µm.

Type: China. Yunnan Province, Mengla County, Xishuangbanna Botanical Garden, on fallen bamboo, 31.X.2009 *Cui 8336* (holotype BJFC; isotype IFP).

Etymology: *asiatica* (Lat.), referring to Asia.

Fruitbody. Basidiocarps annual, resupinate, adnate, corky, without odour or taste when fresh, becoming corky upon drying, up to 9 cm long, 1 cm wide, and 0.6 mm thick at centre. Sterile margin narrow, cream, less than 1 mm wide. Pore surface cream; pores round to angular, 3–4 per mm; dissepiments thin, entire to slightly lacerate. Subiculum very thin, cream, corky, azonate, less than 0.1 mm thick. Tubes concolorous with the pore surface, corky, less than 0.5 mm long.

Hyphal structure. Hyphal system dimitic; generative hyphae with clamp connections; skeletal hyphae IKI–, CB– to CB(+), tissue unchanged in KOH.

Subiculum. Generative hyphae infrequent, hyaline, thin-walled, rarely branched, 1.6–3.2 µm in diam; skeletal hyphae dominant, hyaline, thick-walled with a narrow lumen, unbranched, interwoven, 2–4.5 µm in diam.

Tubes. Generative hyphae infrequent, hyaline, thin-walled, rarely branched, 1.4–3 µm in diam; skeletal hyphae dominant, hyaline, thick-

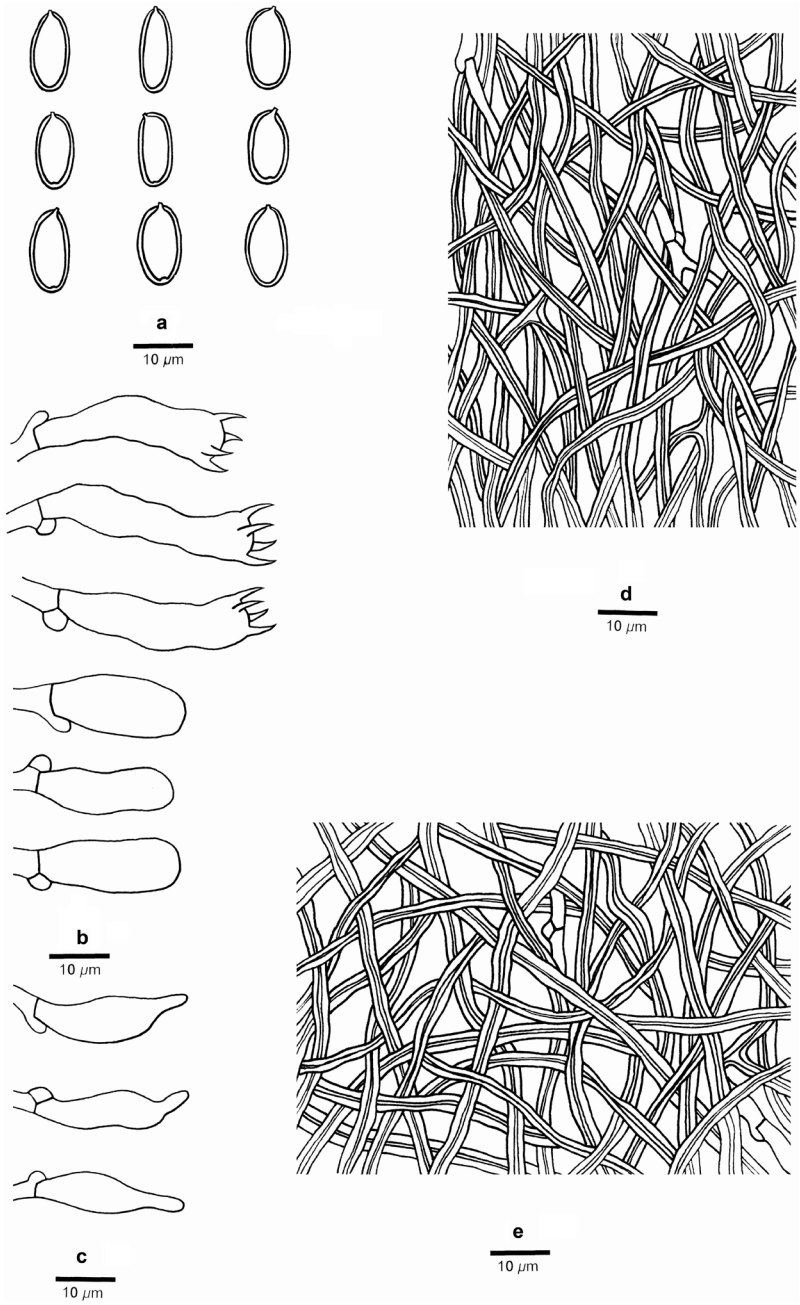

Fig. 2. Microscopic structures of *Grammothelopsis asiatica* (drawn from the holotype). — **a**: Basidiospores. — **b**: Basidia and basidioles. — **c**: cystidioles. — **d**: Hyphae from trama. — **e**: Hyphae from subiculum.

walled with a narrow lumen to subsolid, rarely branched, interwoven, 1.8–4.2 μm in diam. Cystidia absent, cystidioles present, fusoid to subulate, 18–25 × 6–9 μm; basidia clavate, with four sterigmata and a basal clamp connection at the base, 22–34 × 6–10 μm; basidioles in shape similar to basidia, but distinctly smaller.

Spores. Basidiospores ellipsoid to oblong ellipsoid, hyaline, thick-walled, smooth, IKI–, CB(+), (9–)10.5–13(–15) × (5–)5.4–6(–6.7) μm, L = 12.14 μm, W = 5.72 μm, Q = 2.1–2.2 (n = 60/2).

Additional specimens examined (paratypes). **China**.

Hainan Province, Lingshui County, Diaoluoshan Forest Park, on fallen bamboo, 30.V.2008 *Dai 9881* (BJFC & IFP). Changjiang County, Bawangling Nat. Res., on fallen bamboo, 9.XI.2009 *Dai 11588* (BJFC, IFP & H).

A total of five species in *Grammothelopsis* have been reported, four of them, *G. bambusicola*, *G. incrustata*, *G. neotropica* and *G. puiggarii*, were found in Central and South America (David & Rajchenberg 1985, Rajchenberg & Wright 1987, Ryvarden & de Meijer 2002, Robledo & Ryvarden 2007), and *G. macrospora* was recorded in tropical Africa (Ryvarden & Johansen 1980). *Grammothelopsis asiatica* represents the first record of the genus in Asia.

Grammothelopsis asiatica is similar to *G. bambusicola*, which also was found on bamboo (Ryvarden & de Meijer 2002). The type material of *G. bambusicola* was studied, its skeletal hyphae and spores are strongly dextrinoid, and it has dendrohyphidia at dissepiment edge. In addition, its basidiospores are broadly ellipsoid, strongly cyanophilous, (11–)11–13.5(–15) × (7–)7.8–9(–10) μm, L = 12.56 μm, W = 8.32 μm, Q = 1.51 (n = 30/1). In contrast, *G. asiatica* has indextrinoid skeletals and spores, dendrohyphidia are absent at dissepiment edges, and the spores are ellipsoid to oblong ellipsoid, weakly cyanophilous.

Grammothelopsis incrustata and *G. neotropica* have indextrinoid basidiospores, but both have distinctly larger basidiospores (16–22 × 6–8 μm and 18–20 × 7–8 μm, Robledo & Ryvarden 2007). *Grammothelopsis macrospora* differs from *G. asiatica* by having large pores (1–2 per mm), abundant dendrohyphidia, broadly ellipsoid and strongly dextrinoid basidiospores (15–20 × 7.5–11 μm; Robledo & Ryvarden 2007). *Grammothelopsis puiggarii* is distinguished from *G. asiatica* by larger pores (1–2 per mm), broadly ellipsoid and strongly dextrinoid basidiospores (17–20 × 10–12 μm; Robledo & Ryvarden 2007).

Other specimen studied. *Grammothelopsis bambusicola*. **Brazil**. Parana State, Colombo, on rotten bamboo (O 70521).

Inonotus latemarginatus Y.C. Dai, *sp. nova* (Fig. 3)

MycoBank no.: MB 518962.

Carpophorum annuum, applanatum, solitarium. Facies pororum electrinum vel luteolum; pori rotundi, 4–6 per mm. Systema hypharum monomiticum, hyphae septatae sine fibulis, hyphae contexti 4–8 μm in diam. Hyphae setoideae crassitunicatae, acutae, 8–15 μm in diam. Sporae subglobosae, IKI–, CB(+), 7.1–8.7 × 6.2–7.8 μm.

Type: China. Hainan Province, Baoting County, Qixianling National Park, on rotten angiosperm wood, 28.V.2008 *Dai 9758* (IFP).

Etymology: *latemarginatus* (Lat.), referring to the wide sterile margin.

Fruitbody. Basidiocarps annual, pileate, single, soft corky and without odour or taste when fresh, becoming corky to brittle and very light when dry. Pileus applanate, semicircular, projecting up to 8 cm long, 10 cm wide, and 1 cm thick at base; margin obtuse. Pileal surface matted-tomentose to velutinate, indistinctly concentrically zonate, fuscous to brick from base toward to margin when fresh, becoming fuscous and shrunken when dry; margin curry-yellow when fresh, becoming date-brown when dry. Pore surface curry-yellow to buff-yellowish from base towards margin when fresh, becoming snuff-brown when bruised, cinnamon-buff when dry, sterile margin distinct, up to 5 mm wide; pores round, 4–6 per mm; dissepiments thin, matted, slightly lacerate. Context soft and spongy when fresh, becoming brittle to fibrous, shrunken and yellowish brown when dry, up to 5 mm thick, upper layer with a distinct fuscous crust; tube layer concolorous to context, tubes hard corky to brittle, up to 5 mm long.

Hyphal structure. Hyphal system monomitic; all septa without clamp connections, IKI–, CB–; tissue darkening but otherwise unchanged in KOH.

Context. Generative hyphae hyaline to yellowish brown, thin- to slightly thick-walled with a wide lumen, more or less straight, occasionally branched, 4–8 μm in diam, hyphae at upper crust reddish brown, thick-walled, strongly agglutinated, 4–6 μm in diam. Hyphoid setae absent.

Tubes. Generative hyphae hyaline to yellowish, thin- to fairly thick-walled with a wide lumen, straight, rarely branched, more or less subparallel along the tubes, 3–5 μm in diam.

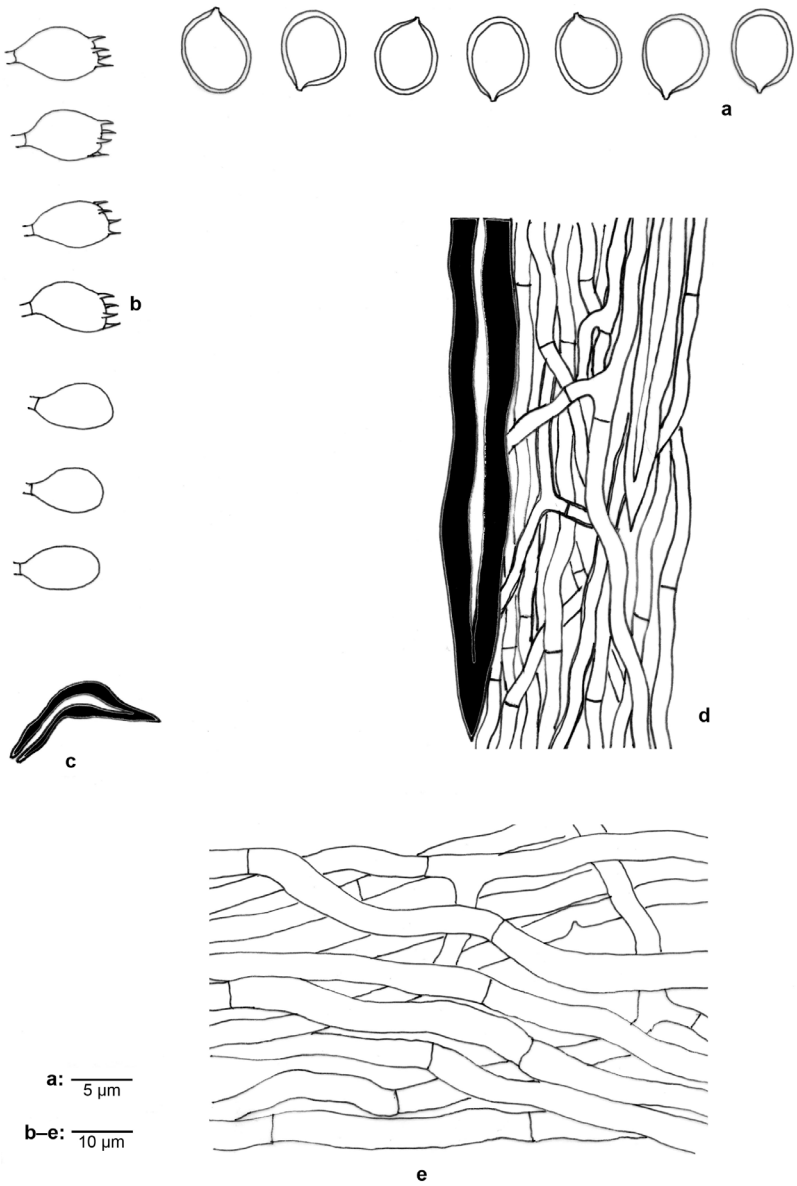

Fig. 3. Microscopic structures of *Inonotus latemarginatus* (drawn from the holotype). — **a**: Basidiospores. — **b**: Basidia and basidioles. — **c**: Hymenial setae. — **d**: Hyphae from trama. — **e**: Hyphae from context.

Hyphoid setae occasionally present, conspicuous, thick-walled with a wide to narrow lumen, nonseptate, tapering to a point, more or less parallel along the tubes, up to 200 um long, 8–15 μm in diam in widest part (n = 10/1). Hymenial setae very rarely present, ventricose to hooked, 16–18 × 6–9 μm. Cystidia and cystidioles absent; basidia barrel-shaped, 10–12 × 7–9 μm, and basidioles similar to basidia.

Spores. Basidiospores subglobose, pale yellowish, fairly thick-walled, smooth, IKI–, CB(+), (7–)7.1– 8.7(–9.3) × (5.9–)6.2–7.8(–8.8) μm, L = 7.85 μm, W = 7.05 μm, Q = 1.11 (n = 32/1).

Inonotus latemarginatus is similar to *I. quercustris*, which was originally described from Louisiana (Blackwell & Gilbertson 1985), and later found in Argentina (Urcelay & Rajchenberg 1999). However, *I. quercustris* differs from *I. latemarginatus* by ungulate basidiocarps (20 × 16 × 10 cm), larger pores (3–5 per mm in USA, 2 per mm in Argentina), abundant hyphoid setae in trama, distinctly longer basidia (18–23 ×

9–11 μm), and especially ellipsoid basidiospores (9–10 × 6–8 μm in USA, 8–10 × 6–7 μm in Argentina).

Inonotus hemmesii, *I. ochroporus* and *I. rodwayi* are pileate species with broadly ellipsoid to subglobose basidiospores and presence of hyphoid setae in tube trama, so they may be confused with *I. latemarginatus*. However, *I. hemmesii* has ungulate basidiocarps (30 × 20 × 15 cm), its hyphoid setae are narrower (7–10 μm wide), and its basidiospores are ellipsoid to ovoid (7–9 × 5.5–7 μm; Gilbertson & Ryvarden 2002). *Inonotus ochroporus* has ungulate basidiocarps (40 × 30 × 30 cm) with duplex context, and its basidiospores are smaller (5.8–7 × 4.9–6.2 μm; Cui *et al*. 2007). *Inonotus rodwayi* has larger pores (2–4 per mm), its basidiospores are ellipsoid and narrower (7–8.5 × 5.2–6.5 μm; Dai & Wu 2002), and its hyphoid setae project into tubes.

Perenniporia hattorii Y.C. Dai & B.K. Cui, *sp. nova* (Fig. 4)

MycoBank no.: MB 518963.

Carpophorum annuum, resupinatum vel effuso-reflexum. Facies pororum cremea vel bubalina; pori rotundi vel angulati, 3–5 per mm. Systema hypharum dimiticum, hyphae generatoriae fibulatae, hyphae skeletales subiculi 2.5–4.8 μm in diam. Sporae hyalinae, ellipsoideae vel truncatae, amyloideae, CB+, 9.8–12.7 × 5.8–7.2 μm.

Type: China. Hainan Province, Ledong County, Jianfengling Nature Reserve, on fallen angiosperm branch, 3.IX.2008 *Dai 10285* (holotype BJFC; isotype IFP).

Etymology: *Hattorii* (Lat.), in honor of the Japanese mycologist Dr. Tsutomu Hattori.

Fruitbody. Basidiocarps annual, resupinate to effused-reflexed, adnate, soft corky to corky, without odour or taste when fresh, becoming corky to fragile upon drying, up to 15 cm long, 4 cm wide, and 1.2 mm thick at centre. Sterile margin cream to cream buff, up to 1 mm wide. Pore surface cream to buff when fresh, becoming cinnamon-buff when bruised, pale yellowish brown upon drying; pores round to angular, 3–5 per mm; dissepiments thin, entire. Subiculum thin, cream to buff, corky, azonate, up to 0.2 mm thick. Tubes concolorous with the pore surface, corky to fragile, up to 1 mm long.

Hyphal structure. Hyphal system dimitic; generative hyphae with clamp connections; skeletal hyphae weakly amyloid, CB+, tissue unchanged in KOH.

Subiculum. Generative hyphae infrequent, hyaline, thin-walled, rarely branched, 1.5–3.4 μm in diam; skeletal hyphae dominant, hyaline, thick-walled with a narrow lumen to subsolid, occasionally branched, interwoven, 2.5–4.8 μm in diam.

Tubes. Generative hyphae infrequent, hyaline, thin-walled, rarely branched, 1.4–3.2 μm in diam; skeletal hyphae dominant, hyaline, thick-walled with a narrow lumen to subsolid, occasionally branched, interwoven, 2.2–4.5 μm in diam. Cystidia absent; fusoid cystidioles present, 18–25 × 6–9 μm; basidia barrel-shaped, with four sterigmata and a basal clamp connection at the base, 20–32 × 10–15 μm; basidioles pyriform, distinctly smaller than basidia.

Spores. Basidiospores ellipsoid, truncate, hyaline, thick-walled, smooth, weakly amyloid, CB+, (9–)9.8–12.7(–14) × (4.5–)5.8–7.2(–8) μm, L = 11.3 μm, W = 6.48 μm, Q = 1.74–1.75 (n = 90/3).

Additional specimens examined (paratypes). **China**. Hainan Province, Ledong County, Jianfengling Nature Reserve, on fallen angiosperm trunk, 4.IX.2008 *Dai 10315 & 10318* (BJFC & IFP).

Perenniporia hattorii is characterized by an annual growth habit, amyloid and cyanophilous skeletal hyphae, ellipsoid, truncate and amyloid basidiospores.

Perenniporia narymica is a common species in the northern hemisphere (Gilbertson & Ryvarden 1987, Ryvarden & Gilbertson 1994, Núñez & Ryvarden 2001). It also has an annual growth habit, cream to pale yellowish pores, amyloid skeletal hyphae, so it may be confused with *P. hattorii*. However, *P. narymica* has acyanophilous skeletal hyphae, smaller and non truncate, inamyloid basidiospores (4.3–5.5 × 3–3.6 μm; Dai *et al*. 2002), and the hyphae dissolve in KOH.

Perenniporia amylodextrinoidea resembles *P. hattorii* by the resupinate basidiocarps, cream to buff pore surface with pores 3–5 per mm. amyloid skeletal hyphae. However, its basid-

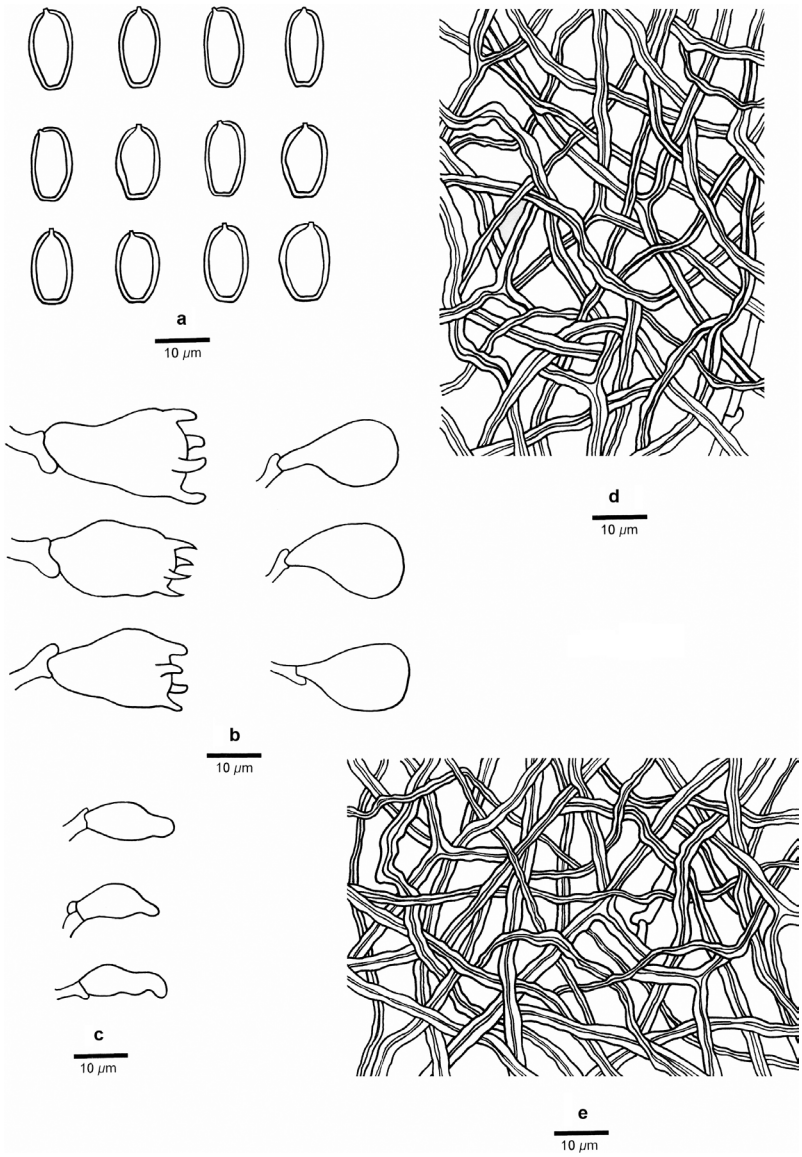

Fig. 4. Microscopic structures of *Perenniporia hattorii* (drawn from the holotype). — **a**: Basidiospores. — **b**: Basidia and basidioles. — **c**: Cystidioles. — **d**: Hyphae from trama. — **e**: Hyphae from context.

iospores are dextrinoid and distinctly smaller (4.5–5.5 × 3–3.5 µm; Gilbertson & Ryvarden 1987).

Wrightoporia austrosinensis Y.C. Dai, *sp. nova* (Fig. 5)

MycoBank no.: MB 518964.

Carpophorum, resupinatum, contextum cremeum. Facies pororum alba vel cremea; pori angulati vel rotundi, 1–3 per mm. Systema hypharum dimiticum, hyphae generatoriae fibulatae, hyphae skeletales non dextrinoideae, hyphae skeletales subiculi 1.7–2 µm in diam. Cystidia hymenii abundantia. Sporae ellipsoideae, hyalinae, asperae, amyloideae, 3–3.2 × 2–2.4 µm.

TYPE: China. Hainan Prov., Changjiang County, Bawangling Nat. Res., on rotten wood of *Pinus*, 8.XI.2009 *Dai 11579* (holotype IFP; isotype BJFC).

ETYMOLOGY: *austrosinensis* (Lat.), referring to southern China.

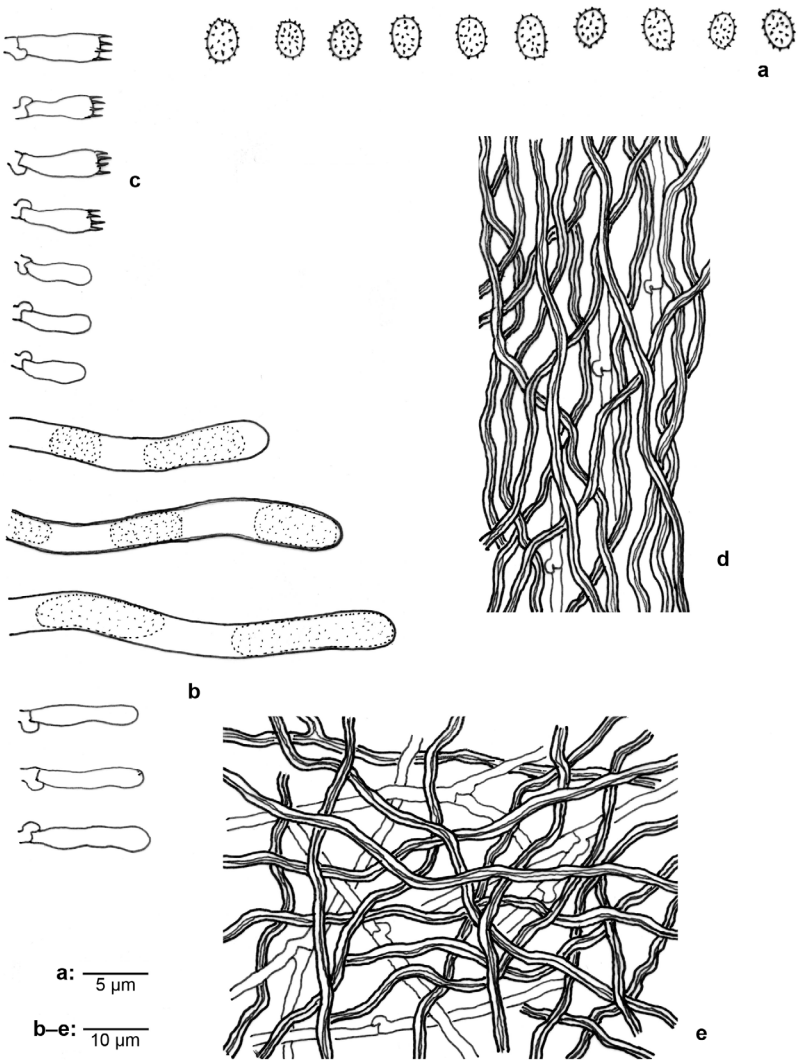

Fig. 5. Microscopic structures of *Wrightoporia austrosinensis* (drawn from the holotype). — **a**: Basidiospores. — **b**: Cystidia and gloeocystidia. — **c**: Basidia and basidioles. — **d**: Hyphae from trama. — **e**: Hyphae from context.

FRUITBODY. Basidiocarps annual, resupinate, soft and cottony when fresh, very difficult to separate from substrate, without odour or taste, cottony when dry, up to 30 cm long, 10 cm wide, 2 mm thick at centre. Pore surface white to cream when fresh, becoming cream to pale brownish upon drying; sterile margin distinct, white, cottony, up to 4 mm wide; pores round to angular, 1–3 per mm, dissepiments thin, entire to slightly lacerate. Context white, very thin. Tubes cream, cottony, up to 2 mm long.

HYPHAL STRUCTURE. Hyphal system dimitic; generative hyphae with clamp connections; skeletal hyphae IKI–, CB–, tissue unchanged in KOH.

CONTEXT. Generative hyphae hyaline, thin-walled, frequently branched, 1.9–2 μm in diam; skeletal hyphae dominant, hyaline, thick-walled with a narrow lumen to subsolid, rarely branched, interwoven, 1.7–2 μm in diam.

TUBES. Generative hyphae frequent, hyaline, thin-walled, rarely branched, 1.8–2.6 μm in diam; skeletal hyphae dominant, hyaline, thick-walled with a narrow lumen to subsolid, rarely branched, flexuous, loosely interwoven, 1.8–2.1 μm in diam. Cystidia clavate to tubular, thin- to slightly thick-walled, originated from trama, embedded or projecting into hymenium, 35–90 × 4–11 μm; gloeocystidia clavate, thin-walled, 17–27 × 5–8 μm; basidia subclavate, with a

basal clamp connection and four sterigmata, 8.5–10 × 2.9–3.5 μm; basidioles in shape similar to basidia, but slightly smaller.

Spores. Basidiospores ellipsoid, hyaline, thin-walled, echinulate, amyloid, CB–, (2.9–)3–3.2(–3.7) × 2–2.4(–2.5) μm, L = 3.11 μm, W = 2.2 μm, Q = 1.41 (n = 35/1).

Wrightoporia subrutilans and *W. austrosisnensis* have similar basidiospores, but the former has smaller pores (4–5 per mm), strongly dextrinoid skeletals, and lack of cystidia (Gilbertson & Ryvarden 1987). In the macro-morphology, *W. austrosinensis* resembles *W. lenta* because of the white and tough basidiocarps. However, *W. lenta* has distinctly larger basidiospores (5.3–7 × 4.8–5.8 μm, measured from *Dai 1506*) and strongly dextrinoid skeletals.

Wrightoporia straminea and *W. tropicalis* both have thick-walled cystidia. The former species is distinguished from *W. austrosisnensis* by resupinate to effused-reflexed basidiocarps with grayish orange pore surface, smaller pores (6–8 per mm), and lack of gloeocystidia (Hattori 2008). *Wrightoporia tropicalis* differs from *W. austrosisnensis* by perennial and woody hard basidiocarps, smaller pores (6–8 per mm), and dextrinoid skeletals (Cui & Dai 2006).

Checklist

In the following an alphabetical list (according to genera) of the polypores is given, and the authors of the scientific names are according to the Authors of Fungal Names (http://www.indexfungorum.org/AuthorsOfFungalNames.htm). Substrates are omitted because most species were collected on unnamed angiosperm wood. The concept of polypores circumscribed here is in a wide sense, including poroid wood-decaying fungi in Basidiomycota.

Abortiporus biennis (Bull.) Singer, *Dai 10026*.
Abundisporus roseoalbus (Jungh.) Ryvarden, *Dai 9334*, *9534*, *9624*, *10834*.
Amauroderma elmerianum Murrill, *Dai 4345*.
Amauroderma perplexum Corner, *Dai 10811*.
Amauroderma preussii (Henn.) Steyaert, *Dai 9552*, *9553*, *9554*, *9557*, *9566*, *9571*.
Amauroderma rude (Berk.) Torrend, *Dai 7896*.
Amauroderma rugosum (Blume & T. Nees) Torrend, *Cui 5275*, *6185*, *Dai 7862*, *7872*, *7972*.
Amauroderma subresinosum (Murrill) Corner, *Cui 6189*, *Dai 7801*, *9522*, *10307*.
Amylosporus campbellii (Berk.) Ryvarden, *Dai 7802*, *7803*.
Antrodia sordida Ryvarden & Gilb., *Dai 10827*, *10830*.
Antrodiella duracina (Pat.) I. Lindblad & Ryvarden, *Cui 5158*, *6391*, *Dai 9693*.
Antrodiella liebmannii (Fr.) Ryvarden, *Cui 5166*, *Dai 9270A*, *9821*.
Antrodiella zonata (Berk.) Ryvarden, *Dai 4360*, *4481*, *7821*, *9939*.
Aurificaria indica (Massee) D.A. Reid, *Cui 6473*.
Bjerkandera adusta (Willd.) P. Karst., *Cui 5198*, *5376*, *Dai 10328*.
Bondarzewia podocarpi Y.C. Dai & B.K. Cui, *Dai 9261*, *10660*, *10662*, *10669*.
Ceriporia crassitunicata Y.C. Dai & Sheng H. Wu, *Cui 5243*.
Ceriporia lacerata N. Maek., Suhara & R. Kondo, *Dai 9501*, *9613*.
Ceriporia mellita (Bourdot) Bondartsev & Singer, *Dai 9667*.
Ceriporia reticulata (Hoffm.) Domański, *Dai 9453*.
Ceriporia spissa (Schwein. *ex* Fr.) Rajchenb., *Dai 2913*, *6984*, *7000*, *7878*.
Ceriporia viridans (Berk. & Broome) Donk, *Cui 5230*, *5231*, *Dai 9995*, *10833*.
Ceriporiopsis mucida (Pers.) Gilb. & Ryvarden, *Cui 5183*.
Ceriporiopsis resinascens (Romell) Domański, *Cui 6589*, *6594*, *Dai 9264*.
Cinereomyces vulgaris (Fr.) Spirin, *Dai 4497*, *7830*, *7831*, *9376*, *9650*, *9658*, *9677*, *9797*, *9807*, *9888*, *9985*.
Coltricia sideroides (Lév.) Teng, *Dai 9238*, *9239*, *9240*.
Coltriciella dependens (Berk. & M.A. Curtis) Murrill, *Cui 6526*, *Dai 4440*, *7938*.
Coltriciella subglobosa Y.C. Dai, *Wei 5020*.
Coltriciella tasmanica (Cleland & Rodway) D.A. Reid, *Dai 11589*.
Coriolopsis aspera (Jungh.) Teng, *Cui 6256*, *Dai 9333*, *9448*, *9472*, *9496*, *9515*, *9516*, *9541*, *9575*, *9669*, *10737*, *10786*, *10789*, *10799*.
Coriolopsis byrsina (Mont.) Ryvarden, *Dai 10788*.
Coriolopsis caperata (Berk.) Murrill, *Dai 9575*, *9500*.
Coriolopsis glabro-rigens (Lloyd) Núñez & Ryvarden, *Dai 7894*.
Coriolopsis polyzona (Pers.) Ryvarden, *Cui 6231*, *Dai 7798*, *9468*, *9486*, *9495*, *9514*, *9573*, *10816*.
Coriolopsis retropicta (Lloyd) Teng, *Dai 9870*.
Coriolopsis sanguinaria (Klotzsch) Teng, *Dai 4590*, *7954*, *9314*, *9350*, *9362*, *9943*, *9980*, *9990*, *10303*.
Coriolopsis strumosa (Fr.) Ryvarden, *Dai 4582*.
Coriolopsis telfarii (Klotzsch) Ryvarden, *HMAS 30136*.
Cyclomyces fuscus Fr., *Dai 9975*, *10016*.
Cyclomyces setiporus (Berk.) Pat., *Cui 6243*, *6249*, *Dai 9252*, *9355*, *9865*.
Cyclomyces tabacinus (Mont.) Pat., *Dai 4365*, *4455*, *4460*, *4480*, *4509*, *9837*, *9845*, *9958*, *9963*, *10309*, *10317*.
Daedalea incana (P. Karst.) Sacc. & D. Sacc., *Cui 6422*, *Dai 9343*.
Daedaleopsis purpurea (Cooke) Imazeki & Aoshima, *Cui 6481*, *6514*, *Dai 9872*.
Datronia mollis (Sommerf.) Donk, *Dai 10772*.

Dictyopanus pusillus (Pers. *ex* Lév.) Singer, *Dai 4479*.
Earliella scabrosa (Pers.) Gilb. & Ryvarden, *Cui 6236*, *Dai 4565*, *4571*, *4615*, *9469*, *9488*, *9510*, *9549*, *9574*, *9620*, *9803*, *9869*, *10329*, *10344*, *10797*.
Echinoporia hydnophora (Berk. & Broome) Ryvarden, *Dai 7962*, *9600*, *9744*, *9809*, *9960*.
Elmerina cladophora (Berk.) Bres., *Cui 5194*, *5461*.
Erastia salmonicolor (Berk. & M.A. Curtis) Niemelä & Kinnunen, *Cui 6673*, *Dai 7892*.
Favolaschia pustulosa (Jungh.) Kuntze, *Cui 5417*, *5423*, *5427*.
Favolaschia volkensii (Bres.) Henn., *Dai 4489*.
Fistulina hepatica (Schaeff.) With., *Dai 7814*, *9703*.
Flabellophora superposita (Berk.) G. Cunn., *Dai 10284*.
Flavodon flavus (Klotzsch) Ryvarden, *Cui 6188*, *6195*, *6281*, *Dai 4575*, *9452*, *9547*, *9551*, *9621*.
Fomitiporia bannaensis Y.C. Dai, *Cui 5300*, *Dai 7978*, *9911*.
Fomitiporia ellipsoidea B.K. Cui & Y.C. Dai, *Dai 4019*, *9251*, *9278*, *9293*, *9967*, *10311*.
Fomitiporia pusilla (Lloyd) Y.C. Dai, *Cui 5211*.
Fomitiporia robusta (P. Karst.) Fiasson & Niemelä, *Dai 10881*.
Fomitiporia texana (Murrill) Nuss, *HMAS 30866*.
Fomitopsis carnea (Blume & T. Nees) Imazeki, *HMAS 7518*.
Fomitopsis feei (Fr.) Kreisel, *Cui 5178*, *Dai 7822*, *7838*, *7947*, *9268*, *9838*.
Fomitopsis pinicola (Sw.) P. Karst., *Dai 11583*.
Fomitopsis rhodophaea (Lév.) Imazeki, *Dai 9306*.
Fomitopsis spraguei (Berk. & M.A. Curtis) Gilb. & Ryvarden, *Dai 9260*.
Funalia gallica (Fr.) Bondartsev & Singer, *Dai 9718*, *9720*, *10356*, *10741*, *10814*.
Ganoderma australe (Fr.) Pat., *Cui 6245*, *Dai 4347*, *9465*, *9519*, *9715*, *10041*.
Ganoderma boninense Pat., *Dai 9564*.
Ganoderma calidophilum J.D. Zhao *et al*., *Cui 6393*.
Ganoderma gibbosum (Nees) Pat., *Dai 10795*.
Ganoderma multipileum Ding Hou, *Cui 5445*, *5445*, *6186*.
Ganoderma nigrolucidum (Lloyd) D.A. Reid, *Dai 10774*.
Ganoderma shangsiense J.D. Zhao, *Cui 6434*, *Dai 10326*.
Ganoderma sinense J.D. Zhao *et al*., *Dai 4559*, *9271*.
Ganoderma tropicum (Jungh.) Bres., *Dai 4346*, *4581*, *4585*, *7805*, *9447*, *9521*, *9523*, *9524*, *10042*.
Gloeophyllum imponens (Ces.) Teng, *Dai 4383*, *9593*, *9862*, *9988*.
Gloeophyllum striatum (Swartz) Murrill, *Dai 1191a*.
Gloeophyllum trabeum (Pers.) Murrill, *Cui 6234*, *Dai 4466*, *9694*.
Gloeoporus dichrous (Fr.) Bres., *Dai 9276*, *9512*, *9652*, *9657*, *9685*, *9916*.
Gloeoporus thelephoroides (Hook.) G. Cunn., *Dai 9266*, *9482*, *9622*.
Grammothele fulgio (Berk. & Broome) Ryvarden, *Cui 6655*, *Dai 4419*, *4520*, *4600*.
Grammothele lineata Berk. & M.A. Curtis, *Cui 6668*, *Dai 9646*, *9716*, *9723*, *9670*, *9716*, *9723*.
Grammothelopsis asiatica Y.C. Dai & B.K. Cui, *Dai 9881*, *11588*.
Haddowia longipes (Lév.) Steyaert, *HMAS 29743*.
Hapalopilus flavus B.K. Cui & Y.C. Dai, *Cui 6414*.
Haploporus alabamae (Berk. & Cooke) Y.C. Dai & Niemelä, *Dai 4363*, *9324*, *9746*, *10778*.
Haploporus papyraceus (Schwein.) Y.C. Dai & Niemelä, *Dai 9962*.
Heterobasidion ecrustosum Tokuda, T. Hatt. & Y.C. Dai, *Dai 9358*, *9731*, *9804*, *9840*.
Hexagonia apiaria (Pers.) Fr., *Cui 6447*, *6467; Dai 9714*, *9732* , *10364*, *10784*.
Hexagonia glabra (P. Beauv.) Ryvarden, *Dai 10991*.
Hexagonia tenuis (Hook) Fr., *Cui 6200*, *6248*, *6258*, *Dai 4567*, *7843*, *7879*, *9513*, *9540*, *9618*, *9952*, *10350*, *10801*.
Humphreya coffeata (Berk.) Steyaert, *HMAS 37912*.
Hyphodontia flavipora (Cooke) Sheng H. Wu, *Dai 4448*, *4522*, *4523*, *4545*, *4611*, *7880*, *7886*, *7891*, *7911*, *9637*, *9651*, *10002*, *10762*.
Hyphodontia latitans (Bourdot & Galzin) Ginns & M.N.L. Lefebvre, *Dai 9499*, *9499*, *9647*, *9662*, *9648*, *9649*, *9655*, *9659*, *9661*, *9662*, *9663*, *9664*, *10743*, *10749*.
Hyphodontia taiwaniana Sheng H. Wu, *Wu 0809-172*.
Hyphodontia tropica Sheng H. Wu, *Dai 7942*, *9279*, *9449*, *9704*, *9705*, *9712*, *9844*, *9849*, *10802*.
Inonotus acutus B.K. Cui & Y.C. Dai, *Cui 5400*.
Inonotus chrysomarginatus B.K. Cui & Y.C. Dai, *Cui 6689*.
Inonotus hainanensis H.X. Xiong & Y.C. Dai, *Dai 9259*.
Inonotus latemarginatus Y.C. Dai, *Dai 9758*.
Inonotus patouillardii (Rick) Imazeki, *Dai 4608*.
Inonotus perchocolatus Corner, *Dai 7939*.
Inonotus rickii (Pat.) D.A. Reid, *Dai 10359*.
Irpex lacteus (Fr.) Fr. *s. lato, Cui 6436*, *Dai 4504*, *9684*.
Irpex vellereus Berk. & Broome, *Dai 7832*, *9833*.
Junghuhnia nitida (Pers.) Ryvarden, *Cui 5443*.
Junghuhnia pseudominuta H.S. Yuan & Y.C. Dai, *Dai 9360*, *Yuan 4105*.
Laccocephalum hartmannii (Cooke) Núñez & Ryvarden, *HMAS 54101*.
Laetiporus sulphureus (Bull.) Murrill, *Cui 5442*, *5448*, *5488*.
Lenzites acuta Berk., *Dai 10358*, *10832*.
Lenzites vespacea (Pers.) Pat., *Cui 5447*, *Dai 7916*, *9839*, *9874*, *10287*.
Leucophellinus hobsonii (Cooke) Ryvarden, *Dai 10733*, *10790*.
Lignosus hainanensis B.K. Cui & Y.C. Dai, *Dai 10670*.
Lignosus rhinocerotis (Cooke) Ryvarden, *HMAS 30036*.
Lindtneria flava Parmasto, *Dai 10800*.
Lindtneria trachyspora (Bourdot & Galzin) Pilát, *Dai 7819*.
Megasporoporia cystidiolophora B.K. Cui & Y.C. Dai, *Dai 9906*.
Megasporoporia hexagonoides (Speg.) J.E. Wright & Rajchenb., *Cui 5179*, *Dai 7834*, *9281*.
Megasporoporia major (G.Y. Zheng & Z.S. Bi) Y.C. Dai, *Cui 6592*, *Dai 9915*, *9933*, *10779*.
Megasporoporia minuta Y.C. Dai & X.S. Zhou, *Cui 6425*, *Dai 10863*.
Megasporoporia setulosa (Henn.) Rajchenb., *Dai 4373*, *9783*, *9785*.
Microporus affinis (Blume & Nees) Kuntze, *Dai 4350*, *4371*, *7826*, *7858*, *9603*, *9751*, *9829*.
Microporus vernicipes (Berk.) Kuntze, *Dai 4374*, *4410*.
Microporus xanthopus (F.) Pat., *Cui 5488*, *Dai 7833*, *7897*, *9283*.
Nigrofomes melanoporus (Mont.) Murrill, *Dai 4396*, *4403*,

7899, *9602*, *9674*, *9802*, *9802*, *10839*.
Nigroporus vinosus (Berk.) Murrill, *Dai 4396*, *4461*, *7920*, *9301*, *9681*, *9880*, *10763*, *10775*.
Oligoporus sericeomollis (Romell) Bondartseva, *Dai 9337*.
Oxyporus corticola (Fr.) Ryvarden, *Dai 7871*.
Oxyporus ginkgonis Y.C. Dai, *Cui 6476*, *Dai 9707*, *10794*.
Oxyporus subulatus Ryvarden, *Cui 6477*.
Parmastomyces mollissimus (Maire) Pouzar, *Dai 9263*, *9325*.
Perenniporia corticola (Corner) Decock, *Dai 4592*.
Perenniporia hattorii Y.C. Dai & B.K. Cui, *Dai 10285*, *10315*, *10318*.
Perenniporia latissima (Bres.) Ryvarden, *Cui 6595*, *Dai 9255*, *9368*, *9926*.
Perenniporia medulla-panis (Jacq.) Donk, *Dai 9326*, *9328*, *9363*, *9739*, *10780*, *10781*.
Perenniporia narymica (Pilát) Pouzar, *Cui 5425*, *Dai 9267*.
Perenniporia ochroleuca (Berk.) Ryvarden, *Dai 9588*, *9597*.
Perenniporia subacida (Peck) Donk, *Dai 7816*.
Perenniporia subadusta (Z.S. Bi & G.Y. Zheng) Y.C. Dai, *Dai 10661*.
Perenniporia tephropora (Mont.) Ryvarden, *Cui 6283*, *6290*, *Dai 4613*, *4618*, *7796*, *7827*, *9462*, *9483*, *9506*, *9518*, *9675*, *9728*, *9986*, *10751*, *10760*.
Phaeolus schweinitzii (Fr.) Pat., *Cui 6566*.
Phellinidium lamaënse (Murrill) Y.C. Dai, *Dai 4468*, *4546*, *7872*, *9250*, *9353*, *9368*, *9374*.
Phellinidium noxium (Corner) Bondartseva & S. Herrera, *Cui 6308*, *Dai 9299*, *9528*, *9530*, *10009*, *10292*.
Phellinus baumii Pilát, *Cui 6631*, *Dai 10841*.
Phellinus cesatii (Bres.) Ryvarden, *Cui 5405*.
Phellinus chryseus (Lév.) Ryvarden, *Dai 10842*.
Phellinus collinus Y.C. Dai & Niemelä, *Dai 7866*.
Phellinus contiguus (Pers.) Pat., *Dai 9747*, *9861*.
Phellinus discipes (Berk.) Ryvarden, *Cui 5416*, *Dai 4381*, *4429*, *7867*, *10286*, *10313*.
Phellinus durissimus (Lloyd) Roy, *Cui 5235*.
Phellinus fastuosus (Lév.) Ryvarden, *Cui 5342*, *5494*, *Dai 9241*.
Phellinus ferreus (Pers.) Bourdot & Galzin, *Dai 7870*, *7922*, *7924*, *9332*, *9349*, *9856*, *9860*, *10769*, *10874*.
Phellinus ferruginosus (Schrad.) Pat., *Cui 6648*.
Phellinus gilvus (Schwein.) Pat., *Cui 6253*, *Dai 4355*, *4364*, *4432*, *7836*, *7841*, *7863*, *7901*, *7941*, *9572*, *9660*, *9666*, *10813*.
Phellinus glaucescens (Petch) Ryvarden, *Dai 7860*, *9371*.
Phellinus inermis (Ellis & Everhart) G. Cunn., *Cui 6474*, *Dai 9527*, *9531*, *9805*, *10000*.
Phellinus kanehirae (Yasuda) Ryvarden, *Cui 5155*, *5336*, *Dai 9262*, *9370*.
Phellinus lloydii (Cleland) G. Cunn., *Dai 9243*, *9316*, *9625*, *9766*, *10805*, *10809*, *10838*.
Phellinus merrillii (Murrill) Ryvarden, *Cui 5192*, *Dai 9642*.
Phellinus minisporus B.K. Cui & Y.C. Dai, *Dai 9632*.
Phellinus pachyphloeus (Pat.) Pat., *Dai 9248*, *10341*.
Phellinus pullus (Berk. & Mont.) Ryvarden, *Dai 4436*, *7955*, *9313*, *9627*.
Phellinus rhabarbarinus (Berk.) G. Cunn., *Dai 4389*, *4423*, *4606*, *7873*, *10876*.
Phellinus rimosus (Berk.) Pilát, *Dai 4384*, *9247*, *9310*.
Phellinus senex (Nees & Mont.) Imazeki, *Cui 5486*, *Dai 7979*.
Phellinus torulosus (Pers.) Bourdot & Galzin, *Cui 5167*, *5441*, *Dai 4430*.
Phellinus tricolor (Bres.) Kotl., *Dai 4399*, *9819*, *10858*.
Phellinus umbrinellus (Bres.) Ryvarden, *Cui 5247*, *Dai 9319*, *9373*.
Phellinus wahlbergii (Fr.) A.D. Reid, *Dai 9339*, *9340*.
Phylloporia hainaniana Y.C. Dai & B.K. Cui, *Cui 5160*, *Dai 9460*.
Phylloporia pectinata (Klotsch) Ryvarden, *Dai 7958*, *9245*, *9246*, *9249*, *9253*, *9257*, *9536*, *9823*.
Phylloporia ribis (Schumach.) Ryvarden, *Cui 6497*, *Dai 10831*.
Phylloporia weberiana (Bres. & Henn. *ex* Sacc.) Ryvarden, *Dai 9242*.
Physisporinus xylostromatoides (Bres.) Y.C. Dai, *Dai 4601*.
Polyporus arcularius (Batsch) Fr., *Cui 6450*, *9507*, *9636*, *9721*.
Polyporus dictyopus Mont., *Cui 5269*, *5452*, *6456*, *6484*, *6497*.
Polyporus grammocephalus Berk., *Dai 7799*, *7849*, *7869*, *7893*, *7898*, *7905*, *7915*, *9576*, *9695*, *9754*, *10029*, *10749b*.
Polyporus jiangenglingensis (G.Y. Zheng) H.D. Zheng & P.G. Liu, *HMIGD 14403*.
Polyporus leprieurii Mont., *Dai 10791*.
Polyporus mikawai Lloyd, *Cui 5387*, *6247*, *Dai 10770*.
Polyporus minor Z.S. Bi. & G.Y. Zheng, *Dai 7847*.
Polyporus moluccensis (Mont.) Ryvarden, *Cui 5267*, *Dai 4352*, *4511*, *4540*, *9384*.
Polyporus mori (Pollini) Fr., *Dai 7842*, *7946*.
Polyporus philippinensis Berk., *Dai 7959*, *7961*, *9354*, *10301*, *10849*.
Polyporus pumilus Y.C. Dai & Niemelä, *Cui 5464*.
Polyporus squamosus (Huds.) Fr., *Yang 740*.
Polyporus tenuiculus (Beauv.) Fr., *Cui 6390*, *Dai 4540*, *9755*.
Polyporus virgatus Berk. & M.A. Curtis, *HMAS 7622*.
Postia alni Niemelä & Vampola, *Dai 9601*, *10840*, *10854*.
Postia fragilis (Fr.) Jülich, *Dai 9708*, *9710*.
Pouzaroporia subrufa (Ellis & Dearn.) Vampola, *Dai 9743*.
Protomerulius caryae (Schwein.) Ryvarden, *Dai 4549*, *9582*, *9846*.
Protomerulius efibulatus Y.C. Dai & Y.L. Wei, *Dai 9322*, *9378*, *9851*.
Pseudofavolus cucullatus (Mont.) Pat., *Dai 10325*, *10771*.
Pycnoporus sanguineus (L.) Murrill, *Cui 6242*, *Dai 7795*, *7835*, *9459*, *9471*, *9476*, *9479*, *9494*, *9497*, *9505*, *9623*, *9679*, *9683*, *9722*.
Pyrofomes albomarginatus (Lév.) Ryvarden, *Dai 9369*, *9828*.
Pyrrhoderma sendaiense (Yasuda) Imazeki, *Dai 7957*, *10340*.
Rigidoporus adnatus Corner, *Cui 5485*.
Rigidoporus crocatus (Pat.) Ryvarden, *Cui 5431*.
Rigidoporus hypobrunneus (Petch) Corner, *Cui 6678*, *Dai 7811*, *7824*, *9480*, *9619*.
Rigidoporus lineatus (Pers.) Ryvarden, *Dai 9806*.
Rigidoporus microporus (Sw.) Overeem, *Dai 4541*, *9580*, *9617*, *9640*, *9835*, *9858*, *10316*.
Rigidoporus minutus B.K. Cui & Y.C. Dai, *Cui 5193*, *6549*, *Dai 9798*, *9847*.
Rigidoporus ulmarius (Sowerby) Imazeki, *Cui 5244*, *5438*, *5449*, *Dai 9315*.
Rigidoporus vinctus (Berk.) Ryvarden, *Dai 4612*, *7817*, *7926*, *7927*, *9280*, *9286*, *9331*, *9638*, *9676*, *9765*, *9792*,

9801, *9850*, *9860*, *9955*, *9994*, *10848*.
Skeletocutis alutacea (J. Lowe) Jean Keller, *Dai 9285*, *9307*, *9884*.
Skeletocutis luteolus B.K. Cui & Y.C. Dai, *Dai 9529*, *9542*, *10322*, *10742*.
Skeletocutis nivea (Jungh.) Jean Keller, *Dai 7829*, *7858*, *7881*, *7932*, *9598*, *9822*, *9825*, *9977*.
Skeletocutis stellae (Pilát) Jean Keller, *Dai 10822*, *10825*.
Skeletocutis subvulgaris Y.C. Dai, *Dai 11590*.
Sparsitubus nelumbiformis L.W. Xu & J.D. Zhao, *Cui 5182*, *5217*, *Dai 4417*, *9244*.
Theleporus calcicolor (Sacc. & P. Syd.) Ryvarden, *Cui 5226*, *Dai 7921*, *7969*.
Tinctoporellus epimiltinus (Berk. & Broome) Ryvarden, *Dai 4457*, *9347*, *9993*, *10305*.
Trametes elegans (Spreng.) Fr., *Cui 6217*, *Dai 4576*, *9457*, *9546*, *9555*, *9558*, *9559*, *9569*, *9730*, *9788*, *10745*, *10748*, *10812*, *10855*.
Trametes hirsuta (Wulfen) Pilát, *Cui 6239*, *6241*, *Dai 4566*, *4593*, *7908*, *9477*, *9533*, *9550*, *9561*, *9578*, *9581*.
Trametes manilaensis (Lloyd) Teng, *Cui 6212*, *6240*, *Dai 10747*.
Trametes membranacea (Sw.) Kreisel, *Dai 10983*.
Trametes modesta (Kunze *ex* Fr.) Ryvarden, *Cui 6229*, *Dai 4350*, *4351*, *4450*, *7815*, *9290*, *9492*, *10844*, *10873*.
Trametes orientalis (Yasuda) Imazeki, *Cui 6184*, *Dai 4570*, *7882*, *9445*, *9454*, *9458*, *9461*, *9467*, *9475*, *9478*, *9481*, *9713*, *10735*, *10736*.
Trametes pocas (Berk.) Ryvarden, *Dai 11577*.
Trametes pubescens (Schumach.) Pilát, *Dai 7846*.
Trametes versicolor (L.) Lloyd, *Cui 6440*, *Dai 9485*.
Trichaptum abietinum (Pers.) Ryvarden, *Cui 5415*, *6561*, *Dai 7903*.
Trichaptum brastagii (Corner) T. Hatt., *Dai 4376*, *4515*, *4552*, *10773*.
Trichaptum byssogenum (Jungh.) Ryvarden, *Dai 4587*, *4595*, *9537*, *9594*, *9668*, *9689*, *9692*, *10752*.
Trichaptum durum (Jungh.) Corner, *Dai 4400*, *4470*, *4514*, *7875*, *9843*, *10320*.
Trichaptum imbricatum Y.C. Dai & B.K. Cui, *Cui 5175*, *Dai 9254*.
Trichaptum perenne Y.C. Dai & H.S. Yuan, *Yang 885*.
Trichaptum podocarpi Y.C. Dai, *Dai 4529*, *9356*, *9853*.
Tyromyces chioneus (Fr.) P. Karst., *Dai 9302*.
Whitfordia scopulosa (Berk.) Núñez & Ryvarden, *Cui 6209*, *Dai 10739*.
Wrightoporia aurantipora T. Hatt., *Dai 9671*.
Wrightoporia austrosisnensis Y.C. Dai, *Dai 11579*.
Wrightoporia avellanea (Bres.) Pouzar, *Dai 10826*.
Wrightoporia luteola B.K. Cui & Y.C. Dai, *Cui 5200*, *5229*, *Dai 9297*.
Wrightoporia tropicalis (Cooke) Ryvarden, *Dai 4382*, *4409*, *4490*.

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References

Bi, Z. S., Li, T. H., Zhang, W. M. & Song, B. 1997: [*A preliminary agaric flora of Hainan Province*]. — Guangzhou, Guangdong Higher Education Press. [In Chinese].

Blackwell, M. & Gilbertson, R. L. 1985: A new species of *Inonotus* (Aphyllophorales, Hymenochaetaceae) on oak in Louisiana. — *Mycotaxon* 23: 285–290.

Cui, B. K. & Dai, Y. C. 2006: *Wrightoporia* (Basidiomycota, Aphyllophorales) in China. — *Nova Hedwigia* 83: 159–166.

Cui, B. K., Dai, Y. C. & Knudsen, H. 2007: Two species of Hymenochaetaceae (Basidiomycota, Aphyllophorales) new to China. — *Mycosystem*a 26: 144–147.

Cui, B. K., Du, P. & Dai, Y. C. 2011a: Three new species of *Inonotus* (Basidiomycota, Hymenochaetaceae) from China. — *Mycological Progress* 10: 107–114.

Cui, B. K., Tang, L. P. & Dai, Y. C. 2011b: Morphological and molecular evidences of a new species of *Lignosus* (Polyporales, Basidiomycota) from tropical China. — *Mycological Progress*. [In press].

Cui, B. K., Yuan, H. S. & Dai, Y. C. 2010: *Phylloporia* (Basidiomycota, Hymenochaetaceae) in China. — *Mycotaxon* 113: 171–178.

Dai, Y. C. 2010: Hymenochaetaceae (Basidiomycota) in China. — *Fungal Diversity* 45: 131–343.

Dai, Y. C., Cui, B. K. & Yuan, H. S. 2009a: *Trichaptum* (Basidiomycota, Hymenochaetales) from China with a description of three new species. — *Mycological Progress* 8: 281–287.

Dai, Y. C., Niemelä, T. & Kinnunen, J. 2002: The polypore genera *Abundisporus* and *Perenniporia* (Basidiomycota) in China, with notes on *Haploporus*. — *Annales Botanici Fennici* 39: 169–182.

Dai, Y. C., Wei, Y. L. & Wu, X. L. 2004: Polypores from Hainan Province, 1. — *Journal of Fungal Research* 2: 53–57.

Dai, Y. C. & Wu, S. H. 2002: The finding of *Inonotus rodwayi* in Taiwan. — *Fungal Science* 17: 19–22.

Dai, Y. C., Yu, C. J., Yuan, H. S. & Wu, X. L. 2009b: Polypores from Hainan Province, 2. — *Guizhou Science* 27: 54–58.

David, A. & Rajchenberg, M. 1985: Pore fungi from French Antilles and Guiana. — *Mycotaxon* 22: 285–325.

Gilbertson, R. L. & Ryvarden, L. 1987: *North American polypores*, *2*. — Fungiflora, Oslo.

Gilbertson, R. L. & Ryvarden, L. 2002: A new *Inonotus* (Basidiomycota, Hymenochaetaceae) from Hawaii. —

Mycotaxon 81: 91–94.

Hattori, T. 2008: *Wrightoporia* (Basidiomycota, Hericiales) species and their allies collected from Japan. — *Mycoscience* 49: 56–65.

Ma, J., Zhang, K. & Zhang, X. G. 2008a: Two new *Ellisembia* species from Hainan, China. — *Mycotaxon* 104: 141–145.

Ma, J., Zhang, K. & Zhang, X. G. 2008b: Taxonomic studies of *Corynespora* from Hainan, China. — *Mycotaxon* 104: 153–157.

Núñez, M. & Ryvarden, L. 2001: East Asian polypores, 2. *Polyporaceae s. lato*. — *Synopsis Fungorum* 14: 165–522.

Petersen, J. H. 1996: *Farvekort. The Danish Mycological Society's colour-chart*. — Foreningen til Svampekundskabens Fremme, Greve.

Rajchenberg, M. & Wright, J. E. 1987: Type studies of Corticiaceae and Polyporaceae (Aphyllophorales) described by C. Spegazzini. — *Mycologia* 79: 246–464.

Robledo, G. & Ryvarden, L. 2007: The genus *Grammothelopsis* (Basidiomycota, aphyllophoroid fungi). — *Synopsis Fungorum* 23: 7–12.

Ryvarden, L. & de Meijer, A. A. R. 2002: Studies in neotropical polypores 14. New species from the state of Parana, Brazil. — *Synopsis Fungorum* 15: 34–69.

Ryvarden, L. & Gilbertson, R. L. 1994: European polypores, 2. — *Synopsis Fungorum* 7: 394–743.

Ryvarden, L. & Johansen, I. 1980: *A preliminary polypore flora of East Africa*. — Fungiflora, Oslo.

Urcelay, C. & Rajchenberg, M. 1999: Two North American *Inonotus* (Hymenochaetaceae, Aphyllophorales) found in Argentina. — *Mycotaxon* 72: 417–422.

Wu, T. L. 1994: [*A checklist of flowering plants of Islands and reefs of Hainan and Guangdong Province*]. — Science Press, Beijing. [In Chinese].

Xiong, H. X. & Dai, Y. C. 2008: A new species of *Inonotus* (Basidiomycota, Hymenochaetaceae) from China. — *Cryptogamie Mycologie* 29: 279–283.

Yu, C. J. & Dai, Y. C. 2009: *Funalia gallica* (Basidiomycota, Polyporaceae), a polypore new to China. — *Guizhou Science* 27: 37–39.

Yuan, H. S. & Dai, Y. C. 2008: Two new species of *Junghuhnia* (Basidiomycota, Polyporales), and a key to the species of China. — *Nordic Journal of Botany* 26: 96–100.

Zhang, K., Fu, H. B. & Zhang, X. G. 2009: Taxonomic studies of *Corynespora* from Hainan, China. — *Mycotaxon* 109: 85–93.

Zhang, K., Ma, J. & Zhang, X. G. 2008a: Taxonomic studies of *Sporidesmium* from Hainan, China. — *Mycotaxon* 104: 165–169.

Zhang, K., Ma, J. & Zhang, X. G. 2008b: Two new species of *Corynespora* from Hainan, China. — *Mycotaxon* 104: 159–163.

Zhao, J. D. 1989: The Ganodermataceae in China. — *Bibliotheca Mycologica* 132: 1–176.

Zhao, J. D. & Zhang, X. Q. 1992: The polypores of China. — *Bibliotheca Mycologica* 145: 1–524.

Zhou, X. S., Dai, Y. C. & Wu, X. L. 2009: *Grammothele lineata* (Basidiomycota, Polyporales) new to China. — *Guizhou Science* 27: 40–42.